

2,3-Bifunctionalized Quinoxalines: Synthesis, DNA Interactions and Evaluation of Anticancer, Anti-tuberculosis and Antifungal Activity

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Received: 13 February 2002; in revised form: 8 August 2002 / Accepted: 10 August 2002 / Published: 30 August 2002

Abstract A variety of 2,3-bifunctionalized quinoxalines (**6-14**) have been prepared by the condensation of 1,6-disubstituted-hexan-1,3,4,6-tetraones (**1-4**) with o-phenylenediamine, (R,R)-1,2-diaminocyclohexane and p-nitro-o-phenylenediamine. It is concluded that strong intramolecular N-H---O bonds in the favoured keto-enamine form may be responsible for the minimal biological activities observed in DNA footprinting, anti-tubercular, anti-fungal and anticancer tests with these hyper π -conjugated quinoxaline derivatives. However, subtle alteration by addition of a nitro group affecting the charge distribution confers significant improvements in biological effects and binding to DNA.

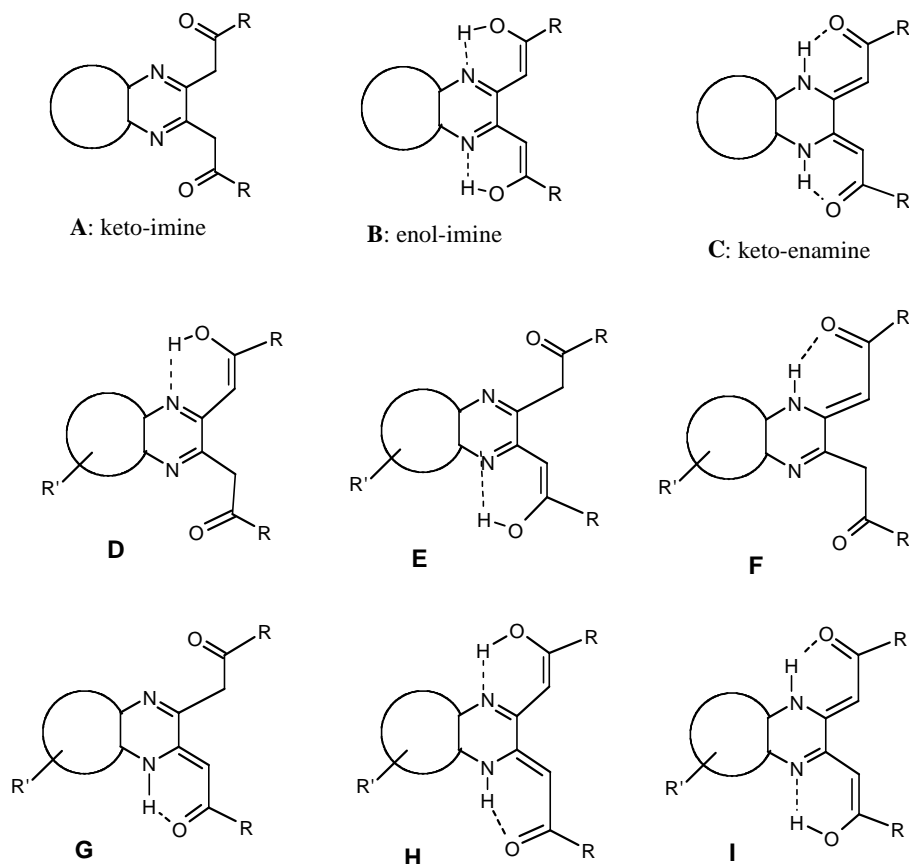
Keywords: Dissymmetric quinoxalines, DNA intercalator, Bayoud, Tuberculosis, Cancer.

Introduction

In recent years, polyfunctionalised quinoxalines have been prepared and studied because of their interesting biological activities [1-5] and DNA interactive behaviour [6-8]. Some act as antidiabetic agents [9], anti-HIV agents [10], or NMDA receptor antagonists [11]. Others can be used in π -conjugated polymer chemistry due to their electron withdrawing properties [12]. In addition, the fluorescence characteristics of 5,8- and 6,7- dimethoxyquinoxaline [13, 14] and their potential as fluoroionophores have recently been described [15].

Previously performed ^1H -, ^{13}C - and ^{15}N -NMR studies have shown that symmetrically substituted quinoxaline ligands **6**, **7** and **10** containing phenyl or cyclohexyl rings lead predominantly to a keto-enamine form with N-H...O intramolecular hydrogen bonds [16]. The existence of this preferred tautomeric form was confirmed by X-ray structural studies of **6** and **10**. Compounds **6** and **7**, containing an extended delocalized π system, exhibited fluorescence properties, while **10** did not. The coexistence of the nine or more tautomeric forms is of particular interest since they present an extended conjugation pathway with a potentially bis-(bidentate) N, O-chelating site (Scheme 1).

Scheme 1: Possible tautomeric forms **A-C** for symmetric quinoxalines **6-13**, and possible tautomeric forms **D-I** for the dissymmetrically substituted quinoxaline derivative **14**.



This type of quinoxaline compound bears a strong resemblance to well-characterised drugs that bind to DNA by intercalation [17]. In general, three major aspects of the binding of drugs to DNA may be expected to influence their biological activity: (i) mode of interaction with the double helix, (ii) sequence specificity of binding, and (iii) kinetics of association/dissociation. Compounds **6-14** represent an attractive model for theoretical and experimental study of intercalators and their medical applications because of the variability in their available forms. Moreover, given the current interest in drug targeting to harmful genes through development of molecules that recognize specific DNA sites [18,19], we became interested in the synthesis of 2,3-disubstituted quinoxaline derivatives having an extended conjugation system. As a first step towards studying possible interactions of **6-14** with DNA, we have carried out *in vitro* anticancer, anti-tubercular and anti-fungal screening, and have examined the DNA interactive properties of the new quinoxalines by DNA footprinting and competition dialysis.

The modulation of the conjugation pathway has been studied by replacing either the external phenyl group with a *neo*-pentyl group or a propyl group, or the central aromatic portion with a cyclohexyl moiety. The effects of these substitutions on possible anti-tumour activity and interaction with DNA are reported. We also describe our efforts to extend the methodology to the synthesis of pharmacologically important quinoxalines bearing functional groups at positions 2 and 3 of the quinoxaline ring. The development of a short and convergent approach to the synthesis allows us easy access to the new quinoxaline family **6-14**.

Results and Discussion

Chemistry

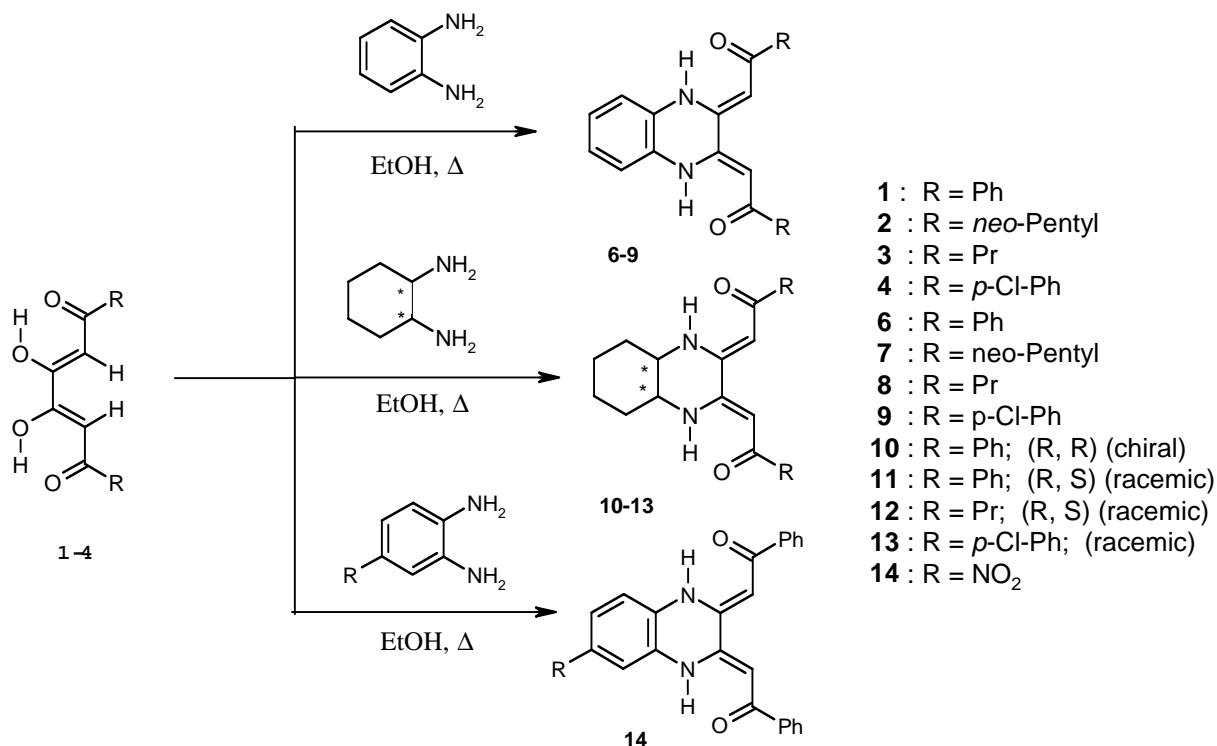
Preparation of 2,3-disubstituted quinoxaline compounds in most cases involves the condensation of diketone [12] or diethyl oxalate compounds [11] with *o*-phenylenediamine. One useful route to 2,3-difunctionalized quinoxalines is the condensation reaction of diketones with various 1,2-diamino-compounds. Disubstituted hexan-1,3,4,6-tetraones have proven to be versatile reagents employed with substituted hydrazines in the synthesis of bipyrazole ligands [12] and have been used for the synthesis of 2,3-difunctionalized quinoxalines **6-8** [13].

A few synthetic methods are available in the literature but none of them are very convenient. Furthermore, hexan-1,3,4,6-tetraone is not commercially available. Accordingly, we have improved the method described by Finar [12]. Our method is shorter (a 1-day reaction at ambient temperature), the yield is excellent (80-85 %) and the method is delightfully simple as well as convenient compared to other methods in the literature.

The procedure has been used for the direct synthesis of several symmetrically substituted quinoxalines **6-13** and one dissymmetrically substituted quinoxaline **14**, all bearing two ketonic arms. The experimental details of the synthesis of compounds **6**, **7** and **10** has been presented elsewhere [16]. For a preparative scale reaction, the commercially available *o*-phenylenediamine was reacted with tetraones **1-4** leading to **6-14**, the products shown by TLC after a 30-60 min reflux in ethanol. Filtration

of compounds **6-14**, followed by washing with absolute ethanol to eliminate unreacted starting materials, gave **6-14** in good yield (70-80%) (Scheme 2).

Scheme 2. Synthesis of quinoxaline derivatives **6-14**



The IR spectrum of the quinoxaline derivative **6** shows one broad, weak absorption band around 3440 cm⁻¹ which may be due to a hydrogen bonded ν(OH) in the end structure or ν(NH) in the enolate/ammonium form. Both C=N imine groups are visible at 1596 and 1563 cm⁻¹ for compound **6**. These bands correlate with protonated imines, which usually exhibit a variable band in the region 1690-1640 cm⁻¹. This compound also shows two weak ν(CO) stretching frequencies at ca. 1735 and 1702 cm⁻¹. From this we can conclude that the symmetric quinoxaline **6** exists in the three principal tautomeric forms **A-C** as shown in Figure 1. In the ¹H-NMR spectrum of **6**, the chemical shifts for all four C(5), C(6), C(7) and C(8) protons coincide at 7.05 ppm, with no coupling to any adjacent atoms. The presence of this singlet peak indicates that the keto-enamine form **C** is preferred over the other forms **A** and **B** in the liquid state. A six-membered delocalized ring can be formed by an intramolecular tautomeric shift of the hydroxyl proton to the nitrogen atom, generating the keto-enamine form **C**. Similar behaviour is equally possible at the second nitrogen centre. This leads to a hyper conjugated quinoxaline derivative that has two identical nitrogen centres since the ligand is highly stabilised by resonance and contains a symmetric charge distribution (Scheme 3).

solid culture media. The percentage inhibition of growth was expressed in terms of the ratio of diameter of the treated mycelium to that of the untreated control (Table 1).

Table 1. Antifungal activity of selected quinoxaline compounds **6-9, 11** and **14**.

Compound	Assay	Percent growth inhibition (Concentration, mg/L)			
		(C1)	(C2)	(C3)	Assessment
6	Bayoud	9 (20)	7 (40)	22 (80)	Inactive
7	Bayoud	9 (60)	15 (120)	15 (180)	Inactive
8	Bayoud	17 (60)	17 (120)	19 (180)	Inactive
9	Bayoud	21 (60)	32 (120)	35 (180)	Inactive
11	Bayoud	15 (34)	31 (67)	33 (134)	Inactive
14	Bayoud	29 (18)	31 (36)	51 (72)	Active

The most effective inhibitor is **14**, which produces 51% inhibition of the growth of *Fusarium* in the presence of 72 mg/L. Molecules **6, 11**, and **9** are less effective but produce appreciable growth inhibition at comparable concentrations. Compounds **7** and **8** only inhibit growth weakly and are considered essentially inactive.

The molecules with the best activity contain a phenyl group on each of the pendant arms of the conjugated chain. The replacement of the two phenyl groups by propyl in compound **8** and *neo*-pentyl in **7** results in a drastic loss of antifungal activity. The loss in activity could be due to the loss of the large, rigid character that the phenyl groups confer on the molecule. Hydrophobic molecules with rigid, planar structures such as aromatic rings, have been shown to have the ability to insert into membranes and induce localized permeability changes leading to leakage out of the membrane [21]. Propyl and *neo*-pentyl groups, while also hydrophobic and very easily inserted into the membrane, are much less likely to cause disruption of the lipid packing order. On the other hand, the greater activity seen with the nitroquinoxaline (**14**) versus **6** and **11** cannot be explained by possible cell membrane disturbances as the nitro group is too small to have this type of effect. The enhanced inhibition observed in the presence of **14** is then more likely due to its interaction with some intracellular target. The presence of a strong electron-withdrawing group must alter the nature of the compound in such a way as to promote binding to the target(s).

Evaluation of Anti-tuberculosis Activity in vitro

Six compounds (**6-7, 9-11, 14**) have been evaluated as anti-tuberculosis agents through the TAACF tuberculosis screening program, but only half of them have been shown to inhibit significantly the growth of *Mycobacterium tuberculosis* H₃₇Rv using the Alamar assay at the first level adopted for *in*

in vitro screening. The three compounds **7**, **9** and **14** displayed modest *in vitro* activity (less than 10%). Not surprisingly, none of these quinoxaline derivatives are currently being examined at the *in vivo* stage of the tuberculosis screening program. The anti-tuberculosis data are summarised in Table 2.

Table 2. Antitubercular activity of quinoxaline compounds **6-7, 9-11** and **14**.

Compound	Assay	MIC ($\mu\text{g/mL}$)	% Inhibition	Activity
6	Alamar	> 6.25	0	-
7	Alamar	> 6.25	7	-
9	Alamar	> 6.25	9	-
10	Alamar	> 6.25	0	-
11	Alamar	> 6.25	0	-
14	Alamar	> 6.25	7	-

Evaluation of Anti-tumour Activity *in vitro*

Among a series of eight 2,3-disubstituted quinoxalines bearing a substitution at position 2 of the ring together with various substituents at the C-6, 7 positions, three compounds (numbers **7**, **9** and **10**) were selected by the National Cancer Institute for evaluation of their anticancer activity *in vitro*. The results seemed to confirm that the carboxy or carbethoxy group at positions 2,3 is not helpful, with a few exceptions, in terms of anticancer activity (Table 3).

Table 3. Antitumour activity of selected quinoxalines **7, 9** and **10** *in vitro*

Compound	Concentration	Growth percentages			Assessment
		(Lung) NCI H460	(Breast) MCF7	(CNS) SF-268	
7	0.1 mM	98	86	105	Inactive
9	0.1 mM	85	76	95	Inactive
10	0.1 mM	89	84	100	Inactive

The quinoxaline compounds **7**, **9** and **10** are among the least active substances to have been evaluated as antitumour agents in our laboratories. Accordingly, an effort was initiated to establish a pharmacophore hypothesis to delineate the requirements of the active site via a comprehensive program of analogue synthesis and evaluation of the effects of structural modification(s) on anti-tumour activity of **6**. The chosen strategy was formulated by dissecting the two-dimensional parent structure into three regions. Region I is ring A of the quinoxaline, region II, the substituted acyl functional arms at positions 2 and 3 of quinoxaline, and region III, the phenyl group. We then set out to determine the

resultant *in vitro* and *in vivo* effects of chemical alterations in each region. Neither the unsubstituted A-ring nor the chiral cyclohexyl B-ring of **10** endowed anti-tumour activity. The modulating anti-tumour effect(s) of substituents having different electronegative properties, located at the para-phenyl site comprising the A-ring of region I, were ascertained next. A chloro substituent located at the 4-position on the phenyl ring of **9** generated the higher anti-tumour activity relative to quinoxalines **7** and **10** in all of the tumour lines tested. However, in general *neo*-pentyl or other p-phenyl substituents proved to be unhelpful. We postulate that the strong tendency to form an intramolecular N-H---O bond in the predominant (keto-enamine) form is likely to be responsible for the lack of biological activity observed with these hyper π -conjugated quinoxaline derivatives. If this hypothesis is correct, by modifications of **6** we may be able to modulate the degree of interaction of the compound with DNA.

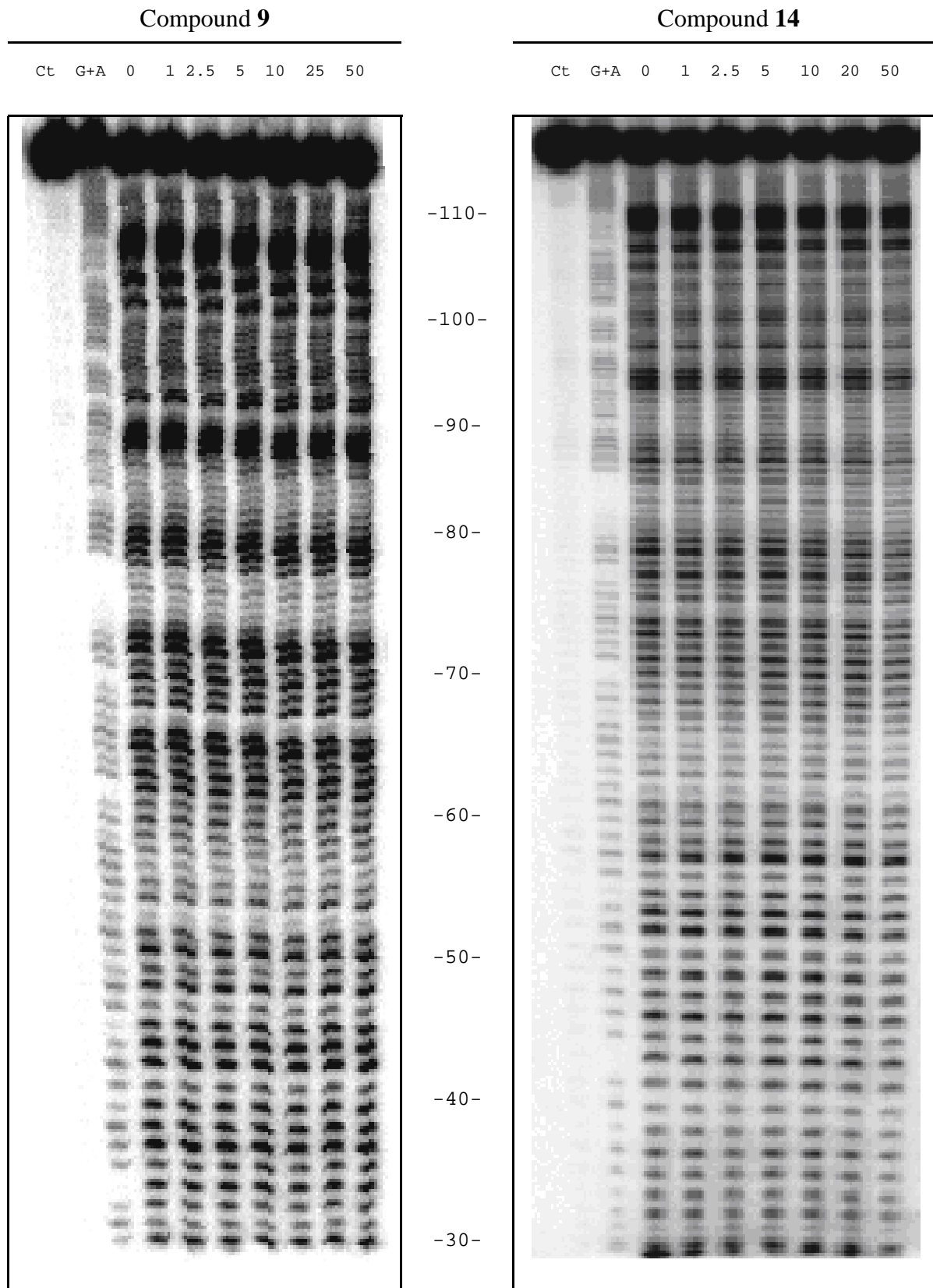
Investigation of binding to DNA

Based on the observations of slight spectral shifts in the presence of nucleic acids (data not shown) we used DNase footprinting techniques [22] to gather information about the dynamic equilibria between the different tautomeric forms and DNA. These experiments serve the dual function of establishing whether significant binding to DNA can be detected by protection from nuclease cleavage, and also detecting whether there are any preferred sites at which these new quinoxaline compounds can bind to natural DNA fragments. For this purpose we have used a number of restriction fragments selected to identify possible binding sites; also to investigate cooperativity of ligand binding. PCR technology was employed to make the labelled DNA substrates [23].

The results yield information that can be used for the design of potential drugs to target critical genes by both 'digital' and 'analogue' sequence readout mechanisms [24]. Scheme 4 shows a DNase I footprinting gel obtained with a 117-mer fragment in the presence of quinoxaline derivative **9** (left panel) and compound **14** (right panel) at the concentrations (μM) shown at the top of each gel lane. The control tracks labelled 'Ct' were not exposed to cleavage by the enzyme. Numbers refer to the nucleotide sequence shown in Scheme 5.

The behaviour of compound **9** is typical of that found for compounds **6**, **7**, **11** and the purified chiral isomer of **11**. There is no evidence of protection of any DNA sequences against nuclease cutting by any of these compounds even at quite high concentrations. We believe that this is due to the predominance and high stability of the keto-enamine form **C** in solution. By contrast, in the case of compound **14** at moderate to high concentrations footprints can easily be seen.

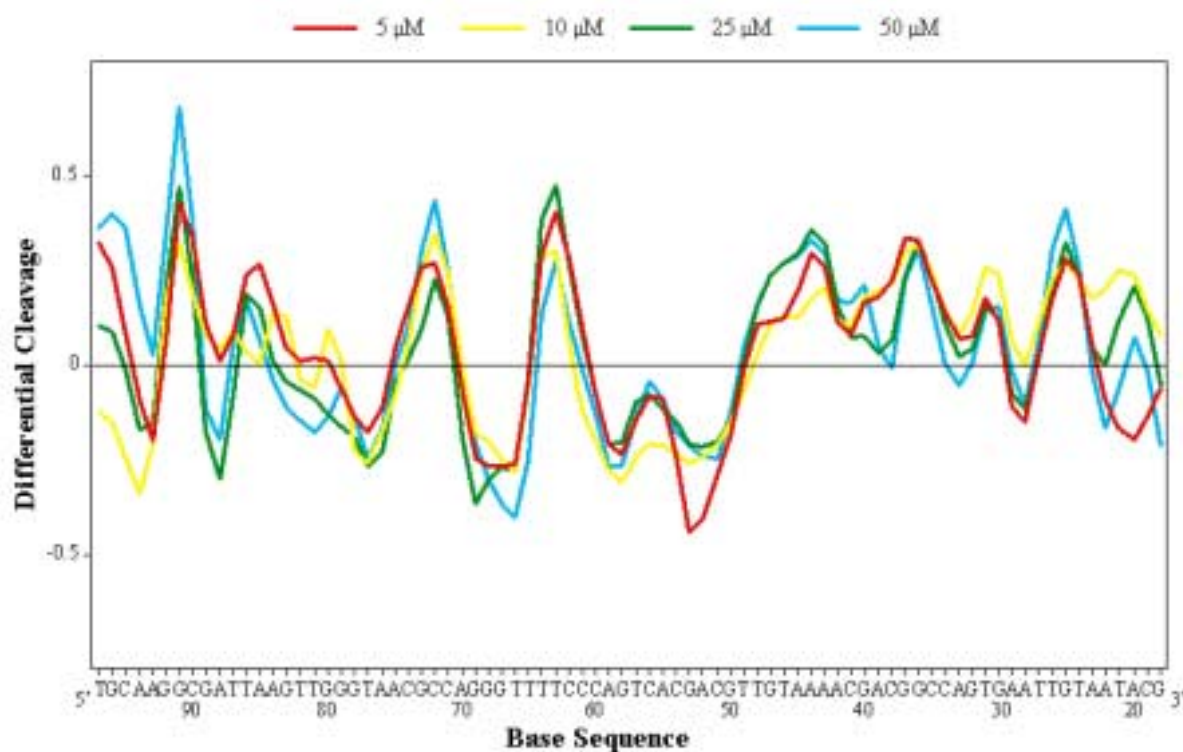
Scheme 4. DNase I footprinting



Differential Cleavage Plot

A differential cleavage plot determined for the nitro-quinoxaline **14** is illustrated in Scheme 5. It reveals binding sites (regions where the enzyme cutting is inhibited) around positions 95, 89, 78, 68, 60, 53 and probably 18, all of which have at least one GC base pair. Enhanced cleavage, generally seen flanking strong binding sites, is evident at several AT-rich clusters, often oligoA-oligoT tracts, near positions 93, 86, 64, 44 and 26. This type of enhanced cleavage is common with many intercalators[1,24]. We cannot explain the apparently clean enhanced peak in the GC-rich region at position 73, unless it reflects some influence of the small AT tract slightly higher up around 76. Compound **14** appears to be endowed with some type of selectivity for binding to GC-containing sequences (Scheme 5).

Scheme 5: Differential cleavage plot determined for the nitro-quinoxaline derivative **14**

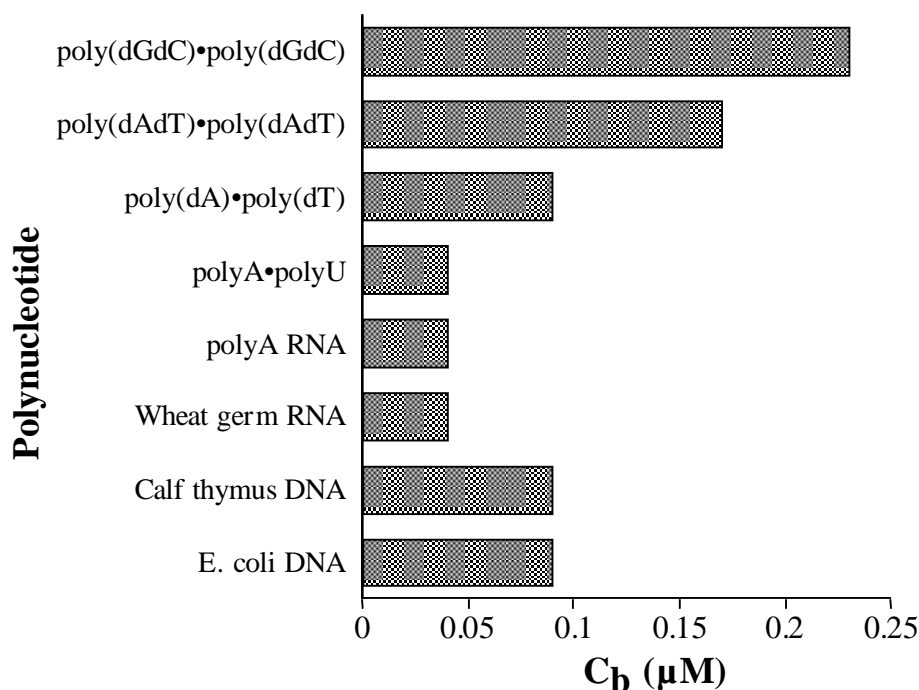


Competition Dialysis

To check the apparent GC-selectivity of compound **14** and to look for evidence of structure-selective binding to nucleic acids we performed competition dialysis experiments [25,26] with **14**, **6**, **9** and **11**. They confirmed that **14** has a clear preference for binding to poly(dGdC), though its estimated

binding constant in 200 mM NaCl is quite low at $4.3 \times 10^3 \text{ M}^{-1}$. Its 'specificity index' calculated according to Ren and Chaires [26], comparing the binding to poly(dGdC) against all the other 6 polymers we tested, is 65%. The results are graphed in Scheme 6. Our findings with the other quinoxalines are not shown but can be summarised as follows. Compound **6** interacts with poly(dGdC) almost as tightly as **14** but its selectivity, as evidenced by a lack of observable footprinting, is considerably less. Compound **9** binds weakly to poly(dGdC) and poly(dAdT) but shows no detectable interaction with the other nucleic acids. Compound **10** barely interacts with poly(dGdC) but binds weakly to the other nucleic acids at a modest level which suggests little discrimination between types and sequences. In the light of these observations, the lack of footprinting by the latter three compounds is explicable. However, the preference of **14** vs. **6** for binding to GC-rich sequences in natural DNA can not be explained simply by its affinity for the nucleic acids because **6** binds about as well to the poly(dGdC) but does not protect GC sequences in the DNA from nuclease cleavage. Further investigation of the effects of electron withdrawing groups on ring A of the quinoxaline is warranted to probe this unique binding seen with compound **14** (Scheme 6).

Scheme 6 : Competition dialysis representing interaction between **14** and eight nucleic acids



Conclusions

The 2,3-di-functionalized quinoxalines **6-14** can easily be prepared by condensation of 1,6-disubstituted hexan-1,3,4,6-tetraones **1-4** with the appropriate diamines. These compounds typically form the highly stable keto-enamine tautomer. A number of important points emerge concerning their biological properties and interaction with DNA. The weakly positive results we have recorded, while

not encouraging for purposes of new drug design, confirm that very likely most of these compounds could be used without great risk of toxicity in diverse applications. Based on their fluorescent properties and thermal stability [16], these compounds may be useful as absorption/ fluorescent probes with high quantum yields or as potential chemical precursors to create laser dyes or optic material colorants. The dissymmetric nitroquinoxaline **14**, which can evidently recognise specific DNA sequences, has potential for gene targeting which merits investigation of its capacity to inhibit access of activators or repressors to regulatory sites in DNA controlling gene expression.

Acknowledgments

This work was supported by the Grants from the Ministry of Education of the Kingdom of Morocco (PROTARS No. P1T2/27), the French CNRS (AIN° 98/ 160/ SM) and Willaya of Oujda. We would like to thank Advanced Chemistry Development, Inc., Toronto, Canada for the free on-line nomenclature of organic compounds. We are indebted to TAACF and NCI of USA for biological tests. M.J.W. and T.E. were supported by Cancer Research UK.

Experimental

General

¹H- and ¹³C-NMR spectra were recorded on a Bruker DPX 200 (operating at 200.12 MHz for ¹H, 50.32 MHz for ¹³C), a Varian (operating at 60 MHz for ¹H) or on a Bruker AM 300 (operating at 300.13 MHz for ¹H, at 75.47 MHz for ¹³C and at 30.42 MHz for ¹⁵N) spectrometer. Chemical shifts are listed in ppm and are reported relative to tetramethylsilane (¹H, ¹³C), residual solvent peaks being used as internal standard, or relative to nitromethane (¹⁵N) with external calibration. Complete assignments of the ¹³C spectra required non-decoupled ¹³C-NMR spectra with selective ¹H decoupling. Non-decoupled ¹⁵N-NMR spectra were measured in 15 mm diameter NMR tubes. UV/visible spectra were recorded on a Kontron Uvikon 941 spectrophotometer in dichloromethane solutions. Infrared spectra were obtained on a Bruker IFS28 FTIR spectrometer as KBr pellets. Elemental analyses were performed by the Centre d'Analyse du CNRS (Solaize, France).

Biological Evaluation

The antifungal activity of the compounds was measured against *Fusarium oxysporum* f.sp. *albedinis*. A stock culture of *Fusarium* was isolated from a date palm infected by the vascular fusarirose. The organism was grown aerobically in liquid Czapek medium of the following composition: 2 g/L of NaNO₃, 1 g/L of K₂HPO₄, 0.5 g/L of MgSO₄·7H₂O, 0.01 g/L of FeSO₄·7H₂O and 30 g saccharose/L. The pH was adjusted to 5.4 with citric acid. All liquid cultures were inoculated into 100 mL Erlenmeyer flasks with a narrow collar containing 20 mL of medium at 25°C, in a water

bath, with horizontal rotary agitation at 70 rpm. The growth was followed by measuring the optical density with a Cecil spectrophotometer at a wavelength of 620 nm and by counting the organisms on a cell of Malassez under an optical microscope. Cultures on solid medium were seeded in Petri dishes 50 mm in diameter at 28 °C in a thermostatically controlled drying oven. The growth was followed by counting colonies for yeast and bacteria and by measuring the diameter of the mycelium for *Fusarium*.

Anti-tuberculosis activity assays were provided by the Tuberculosis Antimicrobial Acquisition and Coordinating Facility (TAACF) at the Southern Research Institute, Birmingham, AL, USA. Screening was conducted at 6.25 µg/mL against *Mycobacterium tuberculosis* H37Rv (ATCC 27294) using the Microplate Alamar Blue Assay [27]. *In vitro* anti-cancer activity was measured by the National Cancer Institute against three cell lines, NCI H460 (Lung), MCF7 (Breast), and SF-268 (CNS). DNase footprinting experiments [22-24], differential cleavage plots [22], and competition dialysis studies [25,26] were conducted as previously described.

Synthesis of Compounds 1-4 and 6-14

The key intermediates **1-4** were prepared as exemplified by the synthesis of *1,6-bis-neopentylhexan-1,3,4,6-tetraone* (**2**) [13]. A suspension of NaOMe was prepared by adding sodium metal (23 g, 1 mol) in portions to absolute methanol (500 mL), then a portion of this suspension (27 g, 0.5 mol) was stirred in dry diethyl ether (300 mL) at 0 °C. A mixture of 4,4-dimethyl-2-pentanone (57 g, 0.5 mol) and diethyl oxalate (36.5 g, 0.25 mol) was added dropwise, over 25 min. The mixture was stirred for 12 h until TLC indicated that all of the 4,4-dimethyl-2-pentanone had been consumed. Compound **2** was precipitated and collected by simple filtration, dissolved in aqueous solution and neutralized with glacial acetic acid to yield 42.3 g (60%) of the title compound as a white-yellow precipitate. Mp = 96-98 °C; ¹H-NMR (CDCl₃): δ 15.0 (br s, w_{1/2} = 70 Hz, 2H, OH), 6.30 (s, 2H, =CH), 2.30 (s, 4H, CH₂), 1.0 (s, 18H, CH₃); ¹³C-NMR (CDCl₃): δ 201.5 (C=O), 172.9 (=C-OH), 100.6 (C-H), 53.9 (CH₂-^tBu), 32.2 (CMe₃), 29.9 (CH₃). IR (cm⁻¹) 1604 (s, ν_{C=O}); LRMS (EI): m/z 282 (C₁₆H₂₆O₄, M⁺, 100%).

2,3-Bis-[(2-hydroxy-2-phenyl)ethenyl]quinoxaline (**6**). *o*-Phenylenediamine (108 mg, 1 mmol) and tetraketone **1** (290 mg, 1 mmol) were stirred in refluxing ethanol (50 mL) for 30 min. After cooling the mixture to room temperature, the desired compound was crystallized as analytically pure orange crystals from dichloromethane (293 mg, 80%). Mp = 218-220 °C; ¹H-NMR (CDCl₃): δ 15.0 (br s, w_{1/2} = 8.2 Hz, 2H, H₁), 8.0-7.8 (m, 4H, *m*-H), 7.6-7.4 (m, 6H, *o*-H, *p*-H), 7.10 (m, 4H, H₅₋₇), 6.5 (s, 2H, CH-CO). ¹³C-NMR (CDCl₃): δ 188.9 (C=O), 146.4 (C₂, C₃), 139.3 (*i*-C), 131.7 (*p*-C), 128.6 (*m*-C), 127.2 (*o*-C), 126.3 (C_{4a}, C_{8a}), 125.1 (C₆, C₇), 116.9 (C₅, C₈), 87.5 (CH-CO); ¹⁵N-NMR (CDCl₃): δ -250.3 (d, ¹J_{N-H} = 87 Hz); IR (cm⁻¹): 1591 (s, ν_{C=O}). Anal. calc. (found) for C₂₄H₁₈N₂O₂: C 78.67 (77.71), H 4.95 (4.80), N 7.65 (7.51).

2,3-Bis-[(2-hydroxy-2-neopentyl)ethenyl]quinoxaline (**7**). *o*-Phenylenediamine (108 mg, 1 mmol) and tetraketone **2** (280 mg, 1 mmol) were stirred in refluxing ethanol (50 mL) for 30 min. After cooling, the

yellow precipitate was filtered off and recrystallized from dichloromethane (210 mg, 60%). Mp = 220-222 °C; $^1\text{H-NMR}$ (CDCl_3): δ 14.6 (br s, 2H, H_1), 7.0 (s, 4H, H_{5-7}), 5.6 (s, 2H, CH-CO), 2.3 (s, 4H, CH_2 - ^tBu), 1.0 (s, 18H, Me); $^{13}\text{C NMR}$ (CDCl_3): δ 198.8 (C=O), 144.5 (C_2 , C_3), 126.1 (C_{4a} , C_{8a}), 124.5 (C_6 , C_7), 116.5 (C_5 , C_8), 92.0 (CH-CO), 56.1 (CH_2 - ^tBu), 31.9 [CH_2 -C(CH_3)], 30.1 [CH_2 -C(CH_3)]; $^{15}\text{N-NMR}$ (CDCl_3): δ -254.3 (d, $^1\text{J}_{\text{N-H}} = 87$ Hz); IR (KBr, cm^{-1}): 1595 (s, $\nu_{\text{C=O}}$); Anal. calc. (found) for $\text{C}_{22}\text{H}_{30}\text{N}_2\text{O}_2 \cdot \text{H}_2\text{O}$: C 71.13 (71.87), H 8.41 (8.37), N 7.54 (7.63).

2,3-Bis-[(2-hydroxy-2-propyl)ethenyl]quinoxaline (8). *o*-Phenylenediamine (108 mg, 1 mmol) and tetraketone **3** (226 mg, 1 mmol) were stirred in refluxing ethanol (50 mL) for 60 min. After cooling, the precipitate was filtered off as a brown powder (238 mg, 80%). Mp = 115-117 °C; $^1\text{H-NMR}$ (CDCl_3 , 60 MHz): δ 14.15 (s, 2H, -OH), 7.00 (s, 4H, aromatic CH, $\text{H}_{5,8}$), 5.68 (s, 2H, CH-CO), 2.40 (t, 4H, CH_2 - CH_2CH_3 , $^1\text{J}_{\text{H-H}} = 15$ Hz), 1.60 (m, 4H, CH_2 - CH_2CH_3 , $^1\text{J}_{\text{H-H}} = 8$ Hz), 0.95 (t, 6H, CH_2 - CH_2 - CH_3 , $^1\text{J}_{\text{H-H}} = 4.3$ Hz); IR (cm^{-1}): 3450 (m, ν_{OH}); Anal. calc. (found) for $\text{C}_{18}\text{H}_{22}\text{N}_2\text{O}_2$: C 72.44 (72.60), H 7.37 (7.48), N 9.39 (9.55).

2,3-Bis-[2-hydroxy-2-(4-chlorophenyl)ethenyl]quinoxaline (9). *o*-Phenylenediamine (108 mg, 1 mmol) and tetraketone **4** (362 mg, 1 mmol) were stirred in refluxing ethanol (50 mL) for 60 min. After cooling, the yellow precipitate was filtered off (325 mg, 75%). Mp = 230-232 °C; $^1\text{H-NMR}$ (CDCl_3 , 60 MHz): δ 7.55-8.25 (m, 10H, aromatic CH), 7.15 (s, 4H, $\text{H}_{5,8}$), 6.5 (s, 2H, CH-CO); IR (cm^{-1}): 3500 (f, ν_{OH}); MS for $\text{C}_{24}\text{H}_{16}\text{N}_2\text{O}_2\text{Cl}_2$: m/z calc. = 435.0667, found 435.0666.

(4aR,8aR)-2,3-Bis-[(2-hydroxy-2-phenyl)ethenyl]decahydro-quinoxaline (10). (R,R)-1,2-diaminocyclohexane (114 mg, 1 mmol) and the tetraketone **1** (280 mg, 1 mmol) were stirred in refluxing ethanol (50 mL) for 30 min. After cooling to room temperature, the yellow precipitate was filtered and recrystallized from dichloromethane (260 mg, 70%). Mp = 254-256 °C; $^1\text{H-NMR}$ (CDCl_3): δ 11.30 (br s, 2H, H_1), 8.0-7.8 (m, 4H, *m*-H), 7.6-7.4 (m, 6H, *o*-H, *p*-H), 6.3 (s, 2H, CH-CO), 3.20 (m, 2H, $\text{H}_{4a,8a}$), 1.8 (m, 4H, $\text{H}_{5,8}$), 1.2 (m, 4H, $\text{H}_{6,7}$); $^{13}\text{C-NMR}$ (CDCl_3): δ 190.2 (C=O), 153.2 (C_2 , C_3), 140.0 (*i*-C), 131.4 (*p*-C), 128.5 (*m*-C), 127.1 (*o*-C), 89.0 (CH-CO), 54.7 (C_{4a} , C_{8a}), 29.7 (C_5 , C_8), 24.0 (C_6 , C_7); $^{15}\text{N-NMR}$ (CDCl_3): δ -270.7 (d, $^1\text{J}_{\text{N-H}} = 94$ Hz); IR (cm^{-1}): 1600 (s, $\nu_{\text{C=O}}$); Anal. calc. (found) for $\text{C}_{24}\text{H}_{24}\text{N}_2\text{O}_2 \cdot \text{H}_2\text{O}$: C 73.82 (73.07), H 6.65 (6.08), N 7.17 (6.54).

(±)-(4aR,8aS)-2,3-bis-[(2-hydroxy-2-phenyl)ethenyl]decahydro-quinoxaline (11). (±)-1,2-diaminocyclohexane (342 mg, 3 mmol) and the tetraketone **1** (900 mg, 3 mmol) were stirred in refluxing ethanol (50 mL) for 60 min. After cooling to room temperature, the yellow precipitate was filtered without further purification (780 mg, 70%). Mp > 230 °C; $^1\text{H-NMR}$ (CDCl_3): δ 11.30 (br s, 2H, H_1), 8.0-7.8 (m, 4H, *m*-H), 7.6-7.4 (m, 6H, *o*-H, *p*-H), 6.3 (s, 2H, CH-CO), 3.20 (m, 2H, $\text{H}_{4a,8a}$), 1.8 (m, 4H, $\text{H}_{5,8}$), 1.20 (m, 4H, $\text{H}_{6,7}$); IR (cm^{-1}): 1600 (s, $\nu_{\text{C=O}}$).

(±)-(4*aR*,8*aS*)-2,3-Bis-[(2-hydroxy-2-propyl)ethenyl]decahydro-quinoxaline (**12**). (±)-1,2-diaminocyclohexane (342 mg, 3 mmol) and the tetraketone **3** (700 mg, 3 mmol) were stirred in refluxing ethanol (50 ml) for 60 min. After cooling to room temperature, the white precipitate was filtered off without further purification (730 mg, 80%). Mp = 182-184 °C; ¹H-NMR (CDCl₃): δ 11.15 (br s, 2H, OH), 6.75 (s, 2H, CH-CO), 3.25 (t, 4H, CH₂-CH₂CH₃, ¹J_{H-H} = 14.8 Hz), 2.55-2.15 (m, 4H, CH₂-CH₂CH₃, ¹J_{H-H} = 7.5 Hz), 0.95 (t, 6H, CH₂-CH₂CH₃, ¹J_{H-H} = 7.3 Hz), 1.25-1.85 (m, 10H, C-H, H_{4*a*}-8*a*); IR (cm⁻¹): 1700 (s, ν_{C=O}).

(±)-(4*aR*,8*aS*)-2,3-Bis-[2-hydroxy-2-(4-chlorophenyl)ethenyl]decahydroquinoxaline (**13**). (±)-1,2-diaminocyclohexane (342 mg, 3 mmol) and the tetraketone **4** (1100 mg, 3 mmol) were stirred in refluxing ethanol (50 mL) for 60 min. After cooling to room temperature, the yellow precipitate was filtered off without further purification (1060 mg, 80%). Mp = 176-178 °C; ¹H-NMR (CDCl₃): δ 11.35 (br s, 2H, OH), 7.65-7.35 (m, 8H, aromatic-H), 6.35 (s, 2H, CH-CO), 3.45-1.55 (m, 10H, H_{4*a*}, 8*a*); IR (cm⁻¹): 1600 (s, ν_{C=O}).

2,3-bis-[(2-hydroxy-2-phenyl)ethenyl]-6-nitro-quinoxaline (**14**). *p*-Nitro-*o*-phenylenediamine (459 mg, 3 mmol) and the tetraketone **1** (900 mg, 3 mmol) were stirred in refluxing ethanol (50 mL) for 60 min. After cooling to room temperature, the yellow precipitate was filtered off without further purification (850 mg, 70%). Mp = 178-180 °C; ¹H-NMR (CDCl₃, 60 MHz): δ 7.55-8.45 (m, 10H, aromatic-H), 7.35 (m, 3H, CH, H₅, 7, 8), 7.05 (s, 2H, CH-CO). IR (KBr, cm⁻¹): 1603 (s, ν_{C=O}).

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Sample Availability: Not Available.