ORIGINAL ARTICLE

25-Hydroxyvitamin- D_3 levels are positively related to subsequent cortical bone development in childhood: findings from a large prospective cohort study

A. Sayers • W. D. Fraser • D. A. Lawlor • J. H. Tobias

Received: 20 June 2011 / Accepted: 19 September 2011 / Published online: 12 November 2011 © The Author(s) 2011. This article is published with open access at Springerlink.com

Abstract

Summary In exploring relationships between vitamin D status in childhood and cortical bone, little relationship was observed with plasma concentrations of 25-hydroxyvitamin- D_2 [25(OH) D_2], whereas 25-hydroxyvitamin- D_3 [25(OH) D_3] was positively related to cortical bone mineral content (BMC_C) and cortical thickness, suggesting D_3 exerts a beneficial effect on cortical bone development in contrast to D_2 .

Introduction The study is aimed to determine whether vitamin D status in childhood is related to cortical bone development by examining prospective relationships be-

Electronic supplementary material The online version of this article (doi:10.1007/s00198-011-1813-9) contains supplementary material, which is available to authorized users.

A. Sayers · J. H. Tobias (⊠)
Musculoskeletal Research Unit, School of Clinical Sciences, University of Bristol,
Bristol, UK
e-mail: jon.tobias@bristol.ac.uk

A. Sayers e-mail: adrian.sayers@bristol.ac.uk

W. D. Fraser Norwich Medical School, Faculty of Medicine and Health Sciences, University of East Anglia, Norwich, UK

D. A. Lawlor

MRC Centre for Causal Analyses in Translational Epidemiology, School of Social and Community Medicine, University of Bristol, Bristol, UK

A. Sayers

Musculoskeletal Research Unit, Avon Orthopaedic Centre, Southmead Hospital, Bristol BS10 5NB, UK tween plasma concentrations of $25(OH)D_2$ and $25(OH)D_3$ at 7.6, 9.9 or 11.8 years and peripheral quantitative computed tomography (pQCT) measurements of the midtibia at age 15.5 years, in children from the Avon Longitudinal Study of Parents and Children.

Methods Relationships between vitamin D status and pQCT outcomes were analysed by bootstrap linear regression, adjusted for age, sex, body composition, socioeconomic position and physical activity, in 2,247 subjects in whom all covariates were available. $25(OH)D_3$ was also adjusted for season and $25(OH)D_2$, and $25(OH)D_2$ for $25(OH)D_3$.

Results 25(OH)D₃ was positively related to BMC_C [0.066 (0.009, 0.122), P=0.02], whereas no association was seen with 25(OH)D₂ [-0.008(-0.044,0.027), P=0.7] [beta (with 95% CI) represents SD changes per doubling of vitamin D], P=0.03 for difference in associations of 25(OH)D₂ and 25 $(OH)D_3$ with BMC_C. There were also differences in associations with cortical geometry, since 25(OH)D₃ was positively related to cortical thickness [0.11(0.04, 0.19), P=0.002], whereas no association was seen with $25(OH)D_2$ [-0.04(-0.08,0.009), P=0.1], P=0.0005 for difference. These relationships translated into differences in biomechanical strength as reflected by buckling ratio, which was positively related to $25(OH)D_2$ [0.06(0.01,0.11), P=0.02] indicating less resistance to buckling, but inversely related to 25(OH)D₃ [-0.1(-0.19,-0.02), P=0.03], P=0.001 for difference.

Conclusions In contrast to $25(OH)D_2$, $25(OH)D_3$ was positively related to subsequent cortical bone mass and predicted strength. In vitamin D-deficient children in whom supplementation is being considered, our results suggest that D_3 should be used in preference to D_2 .

Keywords Adolescence \cdot ALSPAC \cdot Cortical bone \cdot pQCT \cdot Vitamin D

Introduction

Severe vitamin D deficiency, caused by reduced sun exposure, leads to osteomalacia (adults)/rickets (children) resulting from defective skeletal mineralisation. Milder vitamin D deficiency, termed 'insufficiency', may also affect skeletal health in the elderly by reducing bone mineral density (BMD) and increasing fracture risk due to secondary hyperparathyroidism, in the absence of mineralisation defects [1]. If also applicable in childhood, vitamin D requirements in children would need to be set to prevent insufficiency rather than vitamin D deficiency and rickets [2]. Vitamin D insufficiency in children may be relatively common. For example, in Maine, USA, 48% of girls aged 9–11 had a total 25-hydroxyvitamin D (25(OH)D) level below the 50 nmol Γ^1 (20 ng m Γ^1) cutoff commonly used to indicate D insufficiency at least once over 3 years [3].

To ensure adequate vitamin D status, recommended dietary allowances of vitamin D have recently been proposed across different age groups including children [4]. However, a recent Cochrane review concluded that vitamin D supplementation in healthy children had limited effects, but more trials are required to confirm the efficacy of supplementation in deficient children [5]. Whereas three studies in children reported modest improvements in bone outcomes following treatment with cholecalciferol (D_3) [6– 8], ergocalciferol (D_2) was without effect in one study [9]. A possible explanation is that D_2 may be less potent than D_3 , since D_3 and its metabolites have a higher affinity than D₂ for hepatic 25-hydroxylase and vitamin D receptors [10]. Furthermore, in one such study, effects of D_3 supplementation on BMD were suggested to be due to changes in lean mass [6], consistent with observations that levels of vitamin D metabolites and sunlight exposure are related to height and body composition [11-13], which are in turn strongly related to bone parameters [14].

Observational studies of the relationship between plasma concentration of total 25(OH)D and bone outcomes in childhood have yielded conflicting findings [15–17]. These differences may have arisen from confounding, which is difficult to adjust based on results of total 25(OH)D levels, since D₂ and D₃ are derived from different sources. For example, as the majority of D_3 is derived from skin synthesis following the action of UVR, 25(OH)D₃ levels are affected by factors influencing sun exposure such as outdoor physical activity which is known to affect bone development [18]. Whereas dietary fish intake and fortification of certain foods contribute to D₃, D₂ is mainly derived from fungi, plants and dietary supplements, implying that dietary patterns affect levels of 25-hydroxyvitamin-D₂ [25 (OH)D₂] and, to a lesser extent, 25-hydroxyvitamin-D₃ [25 (OH)D₃]. This represents another source of confounding since dietary patterns may affect bone development [19],

possibly through coassociation with socioeconomic position (SEP) which is also related to bone development in childhood [20].

We examined whether vitamin D status influences cortical bone development in childhood, based on 25(OH) D₂ and 25(OH)D₃ concentrations measured at age 7.6, 9.9 or 11.8, and results of peripheral quantitative computed tomography (pQCT) scans of the mid-tibia performed at age 15.5, in the Avon Longitudinal Study of Parents and Children (ALSPAC). Specifically, we wished to determine (i) whether vitamin D status is related to subsequent cortical bone development, (ii) to what extent are its associations which we find are independent of confounders including physical activity and body composition (based on contemporaneous measures of fat and lean mass by DXA), and (iii) whether there is any evidence that $25(OH)D_3$ has stronger associations with cortical bone development than 25(OH) D_2 , suggesting supplementation regimes based on D_3 are likely to be more effective compared to those using D_2 .

Methods

Study population

All pregnant women resident within a defined part of the former county of Avon in South West England with an expected date of delivery between April 1991 and December 1992 were eligible for recruitment, of whom 14,451 were enrolled [21] (http://www.alspac.bristol.ac.uk). Written informed consent was provided by the mothers, and informed assent was obtained from the children at the time of assessment. Ethical approval was obtained from the ALSPAC Law and Ethics Committee (internal) and the Central and South Bristol Research Ethics Committee (external). Data in ALSPAC is collected by self-completion postal questionnaires sent to main caregivers and the children themselves, by abstraction from medical records, and from examination of the children at research clinics. All children with available data were included in the analyses.

Blood measurements

The primary exposures for this study were circulating concentrations of $25(OH)D_2$ and $25(OH)D_3$ as measured on nonfasting blood samples collected at the age 9.9 research clinic. If no samples were available from the 9.9 clinic, samples from the 11.8 clinic were used, or from the age 7.6 year clinic if neither the 9.9 or 11.8 were available. Following collection samples were immediately spun, frozen and stored at -80° C. Assays were performed in 2010 after a maximum of 12 years in storage with no previous freeze–thaw cycles during this period. 25(OH)D₂,

 $25(OH)D_3$ and deuterated internal standard were extracted from serum samples, following protein precipitation, using Isolute C18 solid phase extraction cartridges. Potential interfering compounds were removed by initial elution with 50% methanol followed by elution of the vitamins using 10% tetrahydrofuran in acetonitrile. Dried extracts were reconstituted prior to injection into a high performance liquid chromatography tandem mass spectrometre in the multiple reaction mode (MRM). The MRM transitions (m/z)used were 413.2>395.3, 401.1>383.3 and 407.5>107.2 for 25(OH)D₂, 25(OH)D₃ and hexa-deuterated(OH)D₃, respectively. Coefficients of variation (CVs) for the assay were <10% across a working range of 1 to 250 ng ml⁻¹ for both 25 (OH)D₂ and 25(OH)D₃. Intact parathyroid hormone [iPTH (1-84)] [1] was measured by electrochemiluminescence immunoassay on an Elecsys 2010 immunoanalyzer (Roche, Lewes, UK). Inter-assay CV was less than 6% from 2 to 50 pmol 1⁻¹. The assay sensitivity (replicates of the zero standard) was 1 pmol 1^{-1} .

pQCT variables

At the age 15.5 research clinic, pQCT scans at the 50% mid-tibia were also performed using the Stratec XCT2000L (Stratec, Pforzheim, Germany). Cortical bone area, cortical bone mineral content (BMC_C), cortical bone mineral density (BMD_C), periosteal circumference, endosteal circumference and cortical thickness were recorded. Strength parametres comprised section modulus, cross-sectional moment of inertia, strength strain index and buckling ratio. A density of $>650 \text{ mg cm}^{-3}$ was used to define cortical bone. Endosteal and periosteal circumference were derived using a circular ring model. 4502 pQCT scans were performed, of which 88 were excluded due to major motion artifacts. Coefficients of variation for pOCT scans, based on 139 subjects scanned a mean of 31 days apart, were 2.7%, 1.3% and 2.9% for BMC_C, BMD_C and cortical bone area, respectively.

Other variables

At 15.5 years research clinics, standing height (mm) was measured using the Harpenden Stadiometer (Holtain, Crymych, Wales, UK), and weight using the Tanita Body Fat Analyzer (model TBF 305; Tanita, Arlington Heights, IL, USA). Whole body DXA scans were performed using a Lunar Prodigy scanner with paediatric scanning software (GE Lunar Prodigy, Madison, WI, USA), providing measures of total body fat and lean mass. Maternal SEP was recorded at 32 weeks gestation by questionnaire and categorised according to the Office of Population Censuses and Surveys. Maternal education was assessed at the same time by questionnaire. Pubertal stage was assessed using a Tanner stage (pubic hair domain) questionnaire completed at age 14.7 years [22]. Moderate and vigorous physical activity was assessed by actigraph accelerometre at age 11, and subsequently found to be related to BMD in ALSPAC [23]. Date of birth and sex was obtained from birth notification, and date of the scan was recorded automatically, allowing age at scan to be calculated.

Statistical analyses

Descriptive statistics show means, standard deviation (SD), medians and lower and upper quartiles. Analyses were performed using seasonally adjusted 25 (OH)D₃, which was modelled according to date of blood sampling using linear regression with trigonometric sine and cosine functions. 25 (OH)D₃ was log_e transformed to reduce heteroscedasticity. The residual was used as the primary 25(OH)D₃ exposure variable in subsequent regression analyses. All analyses were performed on standardised variables, i.e. subtracting the mean and dividing by the SD. To include all participants on whom a $25(OH)D_2$ was assayed, those with a value below the detectable limit of the assay (0.5 ng ml⁻¹) were assigned a binary variable indicating whether an individual was at or below the lower limit, which was used as a covariable in all regression models. No individuals had 25(OH)D₃ below the detectable limit of the assay. Models were checked for linearity by adding higher-order terms into the linear predictor and by comparing the likelihood of nested models.

Further analyses were performed using a nonparametric bootstrap procedure in conjunction with OLS linear regression, based on 5,000 replications. Beta (β) estimates and standard errors were calculated from the mean and SD of the bootstrap distribution, respectively. All P values were calculated using bootstrap means and standard errors, compared to a Z-distribution and 95% percentile confidence intervals calculated. Beta estimates were multiplied by $\log_{e}(2)$ and interpreted as per doubling in 25(OH)D₂ or 25 (OH)D₃. Estimates were calculated separately for males and females, and the difference investigated using a bootstrap Wald test. Combined (male and female) associations were also investigated following adjustment for sex. The difference between the effect of 25(OH)D₂ and 25(OH)D₃ was calculated from the bootstrap replicate distribution, and the P values using a Wald test.

Minimally adjusted analyses (model 1), which were based on seasonally adjusted $25(OH)D_3$ levels or $25(OH)D_2$, were adjusted for sex, age at pQCT scan, and adjusted for 25(OH) D_2 and seasonally adjusted $25(OH)D_3$, respectively. In model 2, we additionally adjusted for \log_e -transformed fat mass, lean mass and height. In the final model (model 3), we also adjusted for physical activity and social economic factors (maternal or paternal social class, maternal education). Analyses with endosteal circumference were adjusted for periosteal circumference throughout (endosteal adjusted for periosteal circumference). Sensitivity analyses were performed based on model 2 by: (a) adjusting for parathyroid hormone (PTH); (b) restricting those with available puberty information and then, in this subgroup, examining the impact of adjusting for pubertal status (tanner stages IV/V versus earlier stages); and (c) restricting those with 25(OH)D assays collected at age 9.9 years. All analyses were conducted using STATA 11.2(College Station, TX, USA), and data is assumed to be missing at random.

Results

Descriptive analyses

There were 1,709 boys and 1,870 girls with pQCT scans (age 15.5 years), and plasma $25(OH)D_2$ and $25(OH)D_3$ (age 7.6, 9.9 or 11.8 years; see Fig. 1). Those who were included in the analysis were of higher maternal and paternal social

Fig. 1 Summary of data collection

class compared to those who were not. Boys were taller, heavier and had greater lean mass compared to girls, whereas fat mass was higher in girls (Table 1). BMC_C, cortical bone area, periosteal circumference, endosteal circumference and cortical thickness were greater in boys compared to girls, whereas BMD_C was higher in girls. 25 $(OH)D_3$ levels were slightly higher in boys and $25(OH)D_2$ levels slightly higher in girls. PTH levels were slightly higher in girls. There was evidence of weak inverse associations between 25(OH)D₂ and height LM and FM, which appeared somewhat stronger in girls compared to boys, e.g. P=0.06 for gender interaction test for association with height (Table 2). There was little association between $25(OH)D_3$ and height, and LM P>0.75, and weak evidence of an association with FM P=0.06. 25(OH)D₃ was inversely related to PTH, whereas no association was seen for $25(OH)D_2$. There was a very weak association between seasonally adjusted $25(OH)D_3$ and $25(OH)D_2$, r=-0.0298P=0.155, excluding those subjects in whom $25(OH)D_2$ was below the assay detection limit.

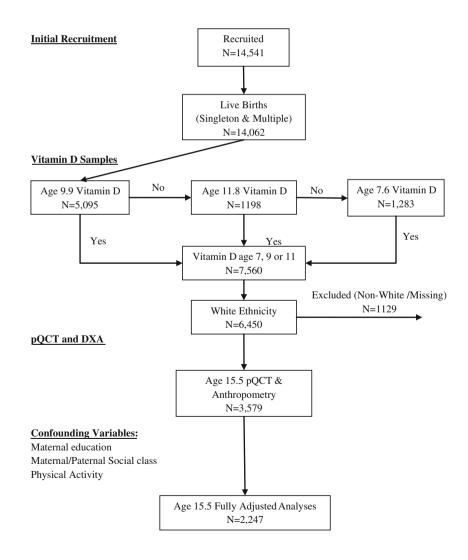


Table 1 Descriptive statistics

			[N (male)=1	[N (male)=1,709; N (female)=1,870]			
			Mean	(SD)	25th	50th	75th
	Age [year]	Male	15.5	(0.26)	15.3	15.4	15.6
		Female	15.5	(0.28)	15.3	15.4	15.6
Anthropometry	Height [cm]	Male	174.3	(7.5)	169.7	174.5	179.4
		Female	164.8	(6.1)	160.7	164.7	168.6
	Weight [kg]	Male	63.5	(11.4)	56.0	61.9	69.3
		Female	58.8	(10.3)	51.9	57.0	63.9
	BMI [kg m ⁻²]	Male	20.8	(3.1)	18.8	20.2	22.2
		Female	21.6	(3.5)	19.3	21.0	23.2
	Fat mass-Total body [kg]	Male	10.8	(7.8)	5.7	8.3	12.9
		Female	18.6	(7.9)	13.2	17.1	22.1
	Lean mass-Total body [kg]	Male	49.8	(6.6)	45.7	49.9	54.1
		Female	37.1	(3.9)	34.5	36.8	39.5
pQCT	BMDC [mg cm ⁻³]	Male	1,074.2	(34.3)	1,053.1	1,077.1	1,099.2
		Female	1,124.6	(22.3)	1,111.2	1,126.3	1,139.8
	BAC [mm ²]	Male	329.1	(46.8)	297.1	329.3	359.6
		Female	275.1	(36.6)	250.0	273.6	298.7
	BMC _C [mg]	Male	353.8	(53.2)	318.8	353.7	388.3
		Female	309.3	(41.0)	281.1	308.0	335.9
	PC [mm]	Male	76.2	(5.3)	72.8	76.1	79.6
		Female	69.5	(4.9)	66.3	69.2	72.6
	EC [mm]	Male	40.9	(5.9)	37.1	40.4	44.1
		Female	37.0	(5.4)	33.6	36.5	39.7
	CT [mm]	Male	5.63	(0.7)	5.2	5.7	6.1
		Female	5.17	(0.6)	4.8	5.2	5.6
Plasma measures	25(OH)D ₃ [ng ml ⁻¹]	Male	24.1	(9.0)	18.1	23.0	28.5
		Female	22.8	(8.2)	17.1	22.1	27.4
	$25(OH)D_2 [ng ml^{-1}]$	Male	1.80	(1.9)	0.5	1.2	2.6
		Female	1.89	(1.9)	0.5	1.4	2.7
	PTH [pmol 1 ⁻¹]	Male	4.53	(1.8)	3.2	4.2	5.5
		Female	5.11	(2.3)	3.5	4.6	6.1

2121

Table shows descriptive characteristics of anthropometric parametres, 50% tibia pQCT parametres, and plasma measures in males and females at age 15.5 years. Statistics are presented as means, SDs, medians, and upper and lower quartiles

Associations between 25(OH)D concentrations and pQCT variables

There was a weak inverse association between $25(OH)D_2$ and BMD_C in both minimally and more fully adjusted analyses, to a similar extent in boys and girls (Table 3). For example, in boys and girls combined, in our most completely adjusted model, a doubling in $25(OH)D_2$ was associated with a 0.05SD decrease in BMD_C . Whereas 25 (OH)D₂ was unrelated to periosteal circumference in minimally adjusted analyses, there was a weak positive association in more fully adjusted models, to a similar degree, in boys and girls. In minimally adjusted analyses, $25(OH)D_2$ was inversely related to cortical bone area, BMC_C, endosteal adjusted for periosteal circumference and cortical thickness in females, but this was not seen after more complete adjustment. There was a positive association between $25(OH)D_2$ levels and buckling ratio in all models, to a similar extent, in boys and girls (Table S1).

Positive associations were observed between $25(OH)D_3$ and cortical bone area and BMC_C in anthropometry adjusted and fully adjusted analyses (Table 4). In all models, $25(OH)D_3$ was positively related to cortical thickness and inversely related to endosteal adjusted for periosteal circumference. For example, in our most fully adjusted model, a doubling in $25(OH)D_3$ was associated with a 0.11 SD increase in cortical thickness. There was also an inverse association between $25(OH)D_3$ and buck-

		Vitamin 2	25(OH)D ₂		Vitamin 2	25(OH)D ₃		P value
		Minimally	y adjusted, N=3,579 (males=1,709)	Minimally	y adjusted, N=3,579 (1	males=1,709)	(D_2D_3)
		Beta	95% CI	P value (sex)	Beta	95% CI	P value (sex)	
Height	Male	-0.026	(-0.072, 0.021)	0.06	-0.070	(-0.169, 0.026)	0.04	0.42
	Female	-0.070	(-0.107, -0.028)		0.056	(-0.016, 0.131)		0.01
	ALL	-0.050	(-0.085, -0.011)		0.000	(-0.061, 0.061)		0.17
Lean mass	Male	-0.021	(-0.059, 0.017)	0.17	-0.027	(-0.112, 0.060)	0.22	0.90
	Female	-0.040	(-0.073, -0.017)		0.034	(-0.012, 0.081)		0.01
	ALL	-0.030	(-0.063, -0.006)		0.007	(-0.040, 0.054)		0.14
Fat mass	Male	-0.017	(-0.066, 0.031)	0.30	-0.048	(-0.160, 0.066)	0.72	0.61
	Female	-0.040	(-0.081, -0.001)		-0.070	(-0.140, -0.003)		0.44
	ALL	-0.030	(-0.069, 0.007)		-0.060	(-0.124, 0.002)		0.40
Ln PTH	Male	-0.010	(-0.064, 0.045)	0.55	-0.260	(-0.367, -0.148)	0.65	0.01
	Female	-0.026	(-0.076, 0.024)		-0.290	(-0.392, -0.189)		0.01
	ALL	-0.019	(-0.064, 0.027)		-0.270	(-0.346, -0.200)		0.01

Table 2 Associations between plasma concentration of $25(OH)D_2$ and $25(OH)D_3$ and anthropometry variables

Table shows associations between plasma concentration of $25(OH)D_2$ and $25(OH)D_3$ and height, total body lean mass, log_e fat mass and log_e parathyroid hormone (PTH), adjusted for sex, age at scan and $25(OH)D_3$ and $25(OH)D_2$ respectively, in 1709 males and 1870 females at age 15.5 years. Beta coefficients represent SD change in height, lean mass, log_e fat mass, and log_e PTH, per doubling of vitamin 25(OH)D₂ and 25 (OH)D₃. 95% confidence intervals are presented for beta coefficients, *P* value (sex) shows the difference in associations between males and females, and *P* value (D₂D₃) is the probability of a difference in associations between 25(OH)D₂ and 25(OH)D₃

ling ratio in both minimally and more fully adjusted analyses (Table S2), suggesting a protective effect on the skeleton since buckling ratio is inversely related to bone strength. These associations tended to be stronger in boys, in whom beta coefficients were two to three times higher than in girls, and P values for gender-specific regression equations were only below the P < 0.05 significance threshold in boys. However, formal gender interaction tests were consistently P >= 0.1, and so evidence that these associations were stronger in boys compared to girls is not compelling.

Subsequently, we compared associations between 25 $(OH)D_2$ and pQCT parametres as shown in Table 3, with associations between 25(OH)3 and pQCT parametres as shown in Table 4. P values for differences in these associations are shown in Table 5, for minimally and more fully adjusted models. In the case of BMD_C and cortical bone area, there was weak evidence of a difference between $25(OH)D_2$ and $25(OH)D_3$ in fully adjusted models, P=0.1and P=0.07, respectively, boys and girls combined (Table 5). For BMC_C, there was moderate evidence of a difference between 25(OH)D₂ and 25(OH)D₃ P<0.05 in all models, boys and girls combined. There was strong evidence of difference between 25(OH)D₂ and 25(OH)D₃ in CT, endosteal adjusted for periosteal circumference and BR, P< 0.001 in minimal and more completely adjusted models, boys and girls combined. Apart from weak evidence of a difference in girls in our anthropometry-adjusted model (P=

0.04), there was no evidence of a difference between 25(OH) D_2 and 25(OH) D_3 with respect to periosteal circumference. No difference was observed for any model in respect of associations between 25(OH) D_2 and 25(OH) D_3 and cross-sectional moment of inertia, section modulus and strength strain index (results not shown).

Sensitivity analyses and exploration of additional models

In view of the biological relationship between vitamin D status and PTH concentrations, we examined whether associations between pQCT parametres and 25(OH)D which we observed were mediated by PTH, but repeating the above analyses including additional adjustment for PTH did not affect the results (see Table S3 for results for buckling ratio, anthropometry-adjusted analyses). In the case of associations between 25(OH)D₂ and buckling ratio, β was attenuated by approximately 15% when restricting analyses to those with complete puberty information, but no further change was seen after adjusting for Tanner stage within this subset. β for the association between 25(OH)D₂ and buckling ratio increased by approximately 50% on restricting analyses to subjects with blood samples at age 9.9, suggesting some associations may be strengthened when vitamin D samples obtained a longer interval before pQCT measurements are excluded. B values were very similar across all groups for associations between 25(OH) D₃ and buckling ratio. We found no evidence of nonline-

letres
paran
pQCT
and
$I)D_2$
25(OH)D
of 25
on c
oncentratior
la c
n plasma con
tions between p
tions
Associa
e 3
Table

		Vitamin 25(OH)D ₂	5(OH)D ₂							
		Minimally adj (males=1,709)	Minimally adjusted, N=3,579 (males=1,709)		Anthropometr (males=1,709)	Anthropometry-adjusted, <i>N</i> =3,579 (males=1,709)	62	Anthropometry-, SES- a <i>N</i> =2,247 (males=1,203)	Anthropometry-, SES- and PA-adjusted, N=2,247 (males=1,203)	sted,
		Beta	95% CI	P value (sex)	Beta	95% CI	P value (sex)	Beta	95% CI	P value (sex)
Cortical bone mineral density	Male Female ALL	-0.055 -0.048 -0.051	(-0.099, -0.010) (-0.082, -0.014) (-0.084, -0.017)	0.74	-0.048 -0.046 -0.047	(-0.091, -0.005) (-0.078, -0.014) (-0.079, -0.015)	0.89	-0.049 - 0.048 -0.048	(-0.101, 0.004) (-0.089, -0.008) (-0.087, -0.011)	0.98
Cortical bone area	Male Female ALL	-0.006 - 0.054 -0.033	(-0.055, 0.043) (-0.095, -0.013) (-0.071, 0.006)	0.04	0.013 -0.003 0.004	(-0.021, 0.047) (-0.030, 0.025) (-0.022, 0.031)	0.32	0.006 0.003 0.004	(-0.038, 0.049) (-0.033, 0.039) (-0.029, 0.037)	0.00
Cortical bone mineral content	Male Female ALL	-0.021 - 0.069 - 0.048	(-0.074, 0.031) (-0.113, -0.026) (-0.089, -0.007)	0.05	0.000 -0.015 -0.008	(-0.035, 0.036) (-0.044, 0.015) (-0.036, 0.020)	0.37	-0.007 -0.009 -0.008	(-0.053, 0.039) (-0.047, 0.028) (-0.044, 0.027)	0.93
Periosteal circumference	Male Female ALL	0.018 -0.021 -0.004	(-0.026, 0.062) (-0.061, 0.021) (-0.040, 0.032)	0.08	0.035 0.031 0.033	(0.002, 0.067) (0.000, 0.063) (0.005, 0.060)	0.82	0.037 0.041 0.039	(-0.007, 0.080) (0.003, 0.079) (0.006, 0.075)	0.86
Endosteal adjusted for periosteal circumference	Male Female ALL	0.026 0.052 0.040	(-0.012, 0.063) (0.021, 0.082) (0.011, 0.069)	0.14	0.01 0.029 0.021	(-0.024, 0.044) (0.001, 0.057) (-0.007, 0.047)	0.22	0.017 0.024 0.021	(-0.025, 0.058) (-0.009, 0.059) (-0.010, 0.053)	0.71
Cortical thickness	Male Female ALL	-0.031 - 0.078 - 0.057	(-0.085, 0.025) (-0.123, -0.033) (-0.100, -0.014)	0.07	-0.018 - 0.044 - 0.032	(-0.065, 0.030) (-0.082, -0.006) (-0.069, 0.005)	0.24	-0.029 -0.039 -0.035	(-0.088, 0.029) (-0.089, 0.007) (-0.081, 0.009)	0.73
Table shows associations between plasma concentration of 25(OH)D ₂ and 50% tibial pQCT parametres at age 15.5 years. Beta coefficients represent SD change in pQCT parametre per doubling of vitamin 25(OH)D ₂ . 95% Confidence intervals are presented with respect to the beta coefficients, <i>P</i> value (sex) shows the difference in associations between males and females. Results are also shown for the following adjustments: minimally adjusted=sex and age at scan; anthropometry adjusted=minimally adjusted+height, loge fat mass and lean mass; anthropometry, SES, PA adjusted=anthropometry-adjusted+matemal and paternal social class, maternal education, and physical activity. All analyses were adjusted for vitamin 25(OH)D ₃ .	ween plasma (ifidence inter justments: mi sted+materna	concentration vals are prese nimally adjus and paternal	of 25(OH)D ₂ and 50% nted with respect to th sted=sex and age at st l social class, maternal	tibial pQCT pi e beta coeffici can; anthropor education, and	arametres at a ants, <i>P</i> value netry adjuste I physical ac	age 15.5 years. Beta co (sex) shows the differ d=minimally adjusted tivity. All analyses we	efficients repre ence in associ +height, loge re adjusted for	ssent SD chang ations between fat mass and vitamin 25(O	(JD ₂ and 50% tibial pQCT parametres at age 15.5 years. Beta coefficients represent SD change in pQCT parametre per doubling of respect to the beta coefficients, <i>P</i> value (sex) shows the difference in associations between males and females. Results are also and age at scan; anthropometry adjusted-minimally adjusted+height, loge fat mass and lean mass; anthropometry, SES, PA lass, maternal education, and physical activity. All analyses were adjusted for vitamin 25(OH)D ₃	r doubling of sults are also try, SES, PA

		Vitamin 25(OH)D.	5(OH)D ₃							
		Minimally adj (males=1,709)	Minimally adjusted, <i>N</i> =3,579 (males=1,709)		Anthropometr (males=1,709)	Anthropometry-adjusted, N=3,579 (males=1,709)	62	Anthropon N=2,247 (1	Anthropometry-, SES- and PA-adjusted, $N=2,247$ (males=1,203)	justed,
		Beta	95% CI	P value (sex)	Beta	95% CI	P value (sex)	Beta	95% CI	P value (sex)
Cortical bone mineral density	Male Female ALL	-0.028 0.010 -0.007	(-0.124, 0.066) (-0.054, 0.072) (-0.064, 0.047)	0.52	-0.020 0.015 -0.001	(-0.110, 0.070) (-0.047, 0.077) (-0.052)	0.53	0.018 0.013 0.016	(-0.103, 0.137) (-0.065, 0.089) (-0.054, 0.082)	0.94
Cortical bone area	Male Female ALL	0.062 0.013 0.036	(-0.043, 0.163) (-0.064, 0.087) (-0.028, 0.099)	0.45	0.091 0.006 0.045	(0.023, 0.162) (-0.047, 0.058) (0.003, 0.087)	0.05	0.100 0.031 0.061	(0.015, 0.191) (-0.034, 0.096) (0.008, 0.116)	0.22
Cortical bone mineral content	Male Female ALL	0.057 0.015 0.035	(-0.056, 0.170) (-0.067, 0.093) (-0.034, 0.104)	0.55	0.089 0.008 0.045	(0.019, 0.162) (-0.049, 0.064) (0.002, 0.090)	0.08	0.105 0.034 0.066	(0.014, 0.198) (-0.036, 0.103) (0.009, 0.122)	0.23
Periosteal circumference	Male Female ALL	$\begin{array}{c} 0.002 \\ -0.027 \\ -0.013 \end{array}$	(-0.095, 0.096) (-0.111, 0.054) (-0.077, 0.047)	0.65	$\begin{array}{c} 0.034 \\ -0.038 \\ -0.005 \end{array}$	(-0.039, 0.104) (-0.096, 0.021) (-0.052, 0.039)	0.13	0.004 0.001 0.002	(-0.090, 0.099) (-0.069, 0.068) (-0.056, 0.059)	0.96
Endosteal adjusted for periosteal circumference	Male Female ALL	-0.083 -0.044 -0.062	(-0.161, -0.007) (-0.100, 0.014) (-0.108, -0.015)	0.43	-0.097 -0.036 -0.064	(-0.164, -0.031) (-0.087, 0.018) (-0.105, -0.022)	0.17	-0.127 -0.043 -0.080	(-0.214, -0.045) (-0.106, 0.020) (-0.132, -0.029)	0.13
Cortical thickness	Male Female ALL	0.117 0.054 0.084	(0.006, 0.228) (-0.029, 0.137) (0.014, 0.152)	0.39	0.131 0.054 0.089	(0.039, 0.226) (-0.021, 0.126) (0.031, 0.148)	0.21	0.180 0.061 0.114	(0.061, 0.306) (-0.029, 0.151) (0.041, 0.190)	0.13
Table shows associations between plasma concentration of 25(OH)D ₃ and 50% tibial pQCT parametres at age 15.5 years. Beta coefficients represent SD change in pQCT parametre per doubling of vitamin 25(OH)D ₃ . 95% Confidence intervals are presented with respect to the beta coefficients, <i>P</i> value (sex) shows the difference in associations between males and females. Results are also shown for the following adjustments: minimally adjusted=sex, season of 25(OH)D ₃ measurement and age at scan; anthropometry-adjusted=minimally adjusted=hex, season of 25(OH)D ₃ measurement and age at scan; anthropometry-adjusted=minimally adjusted=hex, season of 25(OH)D ₃ measurement and age at scan; anthropometry-adjusted=minimally adjusted=hex, season of 25(OH)D ₃ measurement and age at scan; anthropometry-adjusted=minimally adjusted=hex season of 25(OH)D ₃ measurement and age at scan; anthropometry-adjusted=height, log ₆ fat mass and lean mass; anthropometry. SES- and PA-adjusted=anthropometry adjusted+maternal and paternal social class, maternal education, and physical activity. All analyses were adjusted for vitamin 25(OH)D ₂	ween plasma c nfidence interv stments: minin A-adjusted=ant	oncentration c als are preser nally adjusted ⁼ thropometry a	of 25(OH)D ₃ and 50% t tied with respect to the =sex, season of 25(OH) djusted+maternal and ₁	tibial pQCT pa beta coefficiel)D ₃ measureme paternal social	rametres at ag ints, <i>P</i> value (int and age at class, matern	e 15.5 years. Beta coef sex) shows the different scan; anthropometry-ar al education, and phys	fficients represe nce in associat djusted=minim tical activity. A	int SD change ions between J ally adjusted+ II analyses we	in pQCT parametre per males and females. Res height, loge fat mass an ere adjusted for vitamin	doubling of ults are also d lean mass; h 25(OH)D ₂

Table 4 Associations between plasma concentration of $25(OH)D_3$ and Pqct parametres

Table 5Differences betweenassociations of plasma concen- tration of $25(OH)D_2$ and 25 (OUD) $25(OH)D_2$		Sex	Minimally adjusted <i>P</i> value	Anthropometry- adjusted P value	Anthropometry-, SES- and PA- adjusted <i>P</i> value
(OH)D ₃ with pQCT parametres			N=3,579 (males=1,709)	N=3,579 (males=1,709)	N=2,247 (males=1,203)
	BMD _C	Male	0.62	0.58	0.31
		Female	0.11	0.08	0.16
		All	0.19	0.14	0.10
	BA_C	Male	0.25	0.05	0.07
		Female	0.13	0.77	0.45
		All	0.06	0.10	0.07
Table shows the <i>P</i> value for differences between the associa-	BMC_C	Male	0.22	0.03	0.03
tions of plasma concentration of		Female	0.07	0.46	0.28
$25(OH)D_2$ and $25(OH)D_3$ with		All	0.04	0.04	0.03
50% tibial pQCT parametres at	PC	Male	0.77	0.98	0.53
age 15.5 years (as shown in Tables 3 and 4, respectively).		Female	0.89	0.04	0.30
Results are also shown for the		All	0.80	0.15	0.26
following adjustments: mini-	EC _{PC}	Male	0.01	0.01	0.01
mally adjusted=sex and age at		Female	0.01	0.03	0.07
scan; anthropometry- adjusted=minimally adjusted		All	0.01	0.01	0.01
+height, log _e fat mass and lean	CT	Male	0.02	0.01	0.01
mass; anthropometry-, SES- an		Female	0.01	0.02	0.05
PA-adjusted= anthropometry-		All	0.01	0.01	0.01
adjusted+maternal and paternal social class, maternal education,	BR	Male	0.03	0.03	0.01
and physical activity. All results		Female	0.01	0.01	0.04
are adjusted for 25(OH)D ₂ and D ₃		All	0.01	0.01	0.01

arity of associations between either seasonally adjusted 25 $(OH)D_3$ or 25 $(OH)D_2$ in any of the models fitted.

Discussion

We report by far the largest prospective cohort study of relationships between vitamin D status in childhood and subsequent cortical bone outcomes. 25(OH)D₃ was positively related to BMC_C as measured by pQCT approximately 5 years later, which appeared to be secondary to an increase in CT. This association between 25(OH)D₃ and cortical thickness resulted from a decrease in endosteal expansion, since 25(OH)D₃ showed an equivalent inverse association with endosteal adjusted for periosteal circumference. This relationship may also have led to greater biomechanical strength, in view of the inverse association observed between $25(OH)D_3$ and buckling ratio. The latter relationship may have implications in terms of reduced fracture risk in later life, based on associations between buckling ratio as measured at the hip in elderly populations, and subsequent risk of hip fracture [24, 25]. The effect sizes that we observed were similar in magnitude to that of other important external influences on skeletal development such as fat mass, which we have previously reported to influence cortical bone development [14]. In further analyses, based on the same study sample, we found that a doubling in fat mass was associated with a 0.13 SD increase in cortical thickness (analyses adjusted for age and height), which was similar to that seen for $25(OH)D_3$, of which a doubling was associated with a 0.11 SD increase in cortical thickness.

Identification of 25(OH)D concentrations in childhood associated with optimal outcomes for bone and other health outcomes, and how these might translate into public health recommendations, is a matter of controversy [26]. Arguably, the finding that a doubling in 25(OH)D₃ is associated with a 0.11 SD increase in cortical thickness is not a strong enough effect to justify widespread vitamin D supplementation in childhood. Since >25% of our study population had insufficient total 25(OH)D based on the 20 ng ml⁻¹ cutoff [26], this conclusion is likely to apply to other, predominantly Caucasian, populations with a similarly high prevalence of vitamin D insufficiency based on this definition. This may represent a contrast with early life exposure in utero, when vitamin D status has been suggested to have major long-term influences on subsequent bone development including periosteal growth [27, 28]. On the other hand, $25(OH)D_3$ may have a stronger

association with cortical outcomes in certain subgroups, in whom supplementation may be more justifiable. For example, beta coefficients were generally higher in boys, in whom a doubling in $25(OH)D_3$ was associated with a 0.18 SD increase in CT. Moreover, the magnitude of effects that we observed may have been tempered by aspects of the study design (see 'Limitations' below). Furthermore, whereas observational studies of this nature provide some information as to the likely benefits of vitamin D supplementation in childhood, evidence from randomized controlled trials is required before definitive conclusions can be drawn.

In those children in whom vitamin D supplementation is being considered, an important question which follows is which form of vitamin D is the most effective. In contrast to the positive associations between $25(OH)D_3$ and cortical bone outcomes described above, relationships with 25(OH) D_2 were null in the case of BMC_C and cortical thickness. Whereas a weak positive association was present between $25(OH)D_2$ and periosteal circumference, there was a weak inverse association with BMD_C, as well as a weak positive association with buckling ratio suggesting reduced resistance to buckling. Taken together, these findings suggest that, in contrast to 25(OH)D₃, 25(OH)D₂, at these concentrations, is not associated with an overall benefit in terms of future cortical bone development, which may have important implications in terms of the choice of vitamin D supplementation in childhood.

To our knowledge, no previous study has examined the separate relationships between 25(OH)D₂, 25(OH)D₃ and bone outcomes in childhood. Since 25(OH)D₃ makes the major contribution to total 25(OH)D, it is relevant to compare our findings with those from these previous studies based on total 25(OH)D. In a prospective study of 171 girls aged 9-15 years, total 25(OH)D was positively associated with gains in femoral neck BMD over the following 3 years which may have reflected an influence of $25(OH)D_3$ on cortical thickness as we observed [16]. On the other hand, our findings contrast with those of a previous study in which total 25(OH)D was found to be positively related to BMD_C of the radius and tibia in a cross-sectional study based on 193 10- to 12-year-old girls [15]. In terms of previous interventional studies, in a recent study in 20 pairs of peripubertal female twins, D₃ supplements for 6 months led to an increase in tibial cortical bone area due to reduced endosteal expansion as assessed by pQCT [7]. In contrast, in a recent D_2 supplementation trial in 73 girls aged 12-14 years, no effect was observed on pQCT parametres [9]. Although these findings are consistent with our observation of an inverse association between endosteal adjusted for periosteal circumference and 25(OH) D_3 , but not 25(OH) D_2 , to our knowledge, no previous study has directly compared the effect of administering these two forms of vitamin D on cortical bone.

In terms of biological explanations for possible distinct effects of 25(OH)D₂ and 25(OH)D₃ on bone, as suggested by our results, indirect pathways via PTH may be involved. Whereas $25(OH)D_3$ levels are known to be inversely related to PTH, as confirmed here, an equivalent relationship was not seen for $25(OH)D_2$, which is consistent with a previous finding that a large dose of D₃ decreased PTH in the elderly, whereas D_2 was without effect [29]. Any tendency for 25(OH)D₂ and 25(OH)D₃ to differ in respect of their relationships with PTH may be partly due to the fact that D_2 is less potent than D_3 : D_3 and its metabolites have a higher affinity than D₂ for hepatic 25-hydroxylase and vitamin D receptors; D₃ is not directly metabolised to 24(OH)D as is D₂; 25(OH)D₂ has a lower affinity for vitamin D binding protein compared to 25(OH)D₃, leading to faster metabolism and a shorter half life [10]. However, adjusting our analyses for PTH did not attenuate the observed association between 25(OH)D₃ and endosteal adjusted for periosteal circumference, suggesting that differing relationships with PTH are unlikely to explain the distinct associations between $25(OH)D_2$, $25(OH)D_3$ and cortical bone parametres which we observed.

Alternatively, due to the observational nature of this study, these different associations with $25(OH)D_2$ and 25(OH)D₃ may have arisen from confounding. For example, 25(OH)D₃ levels are determined by sun exposure and diet that may be affected by a range of factors including SEP and outdoor physical activity, which may confound relationships with bone outcomes. Although the association between 25(OH)D₃ and endosteal adjusted for periosteal circumference was unaffected by adjusting for observed measurements of these additional factors, unmeasured confounders may be important. For example, D_2 intake is related to consumption of fruits and vegetables, which is positively associated with childhood BMD as measured by DXA [30]. In addition, fruit and vegetable intake is related to a 'prudent' or 'healthy' diet [31], of which intake in pregnancy is positively associated with BMD in subsequent childhood [19].

Limitations

In terms of limitations of this study, our pQCT measurements comprised a single slice, namely, the 50% mid-tibia, which is unable to provide any information about trabecular bone. In the study of 171 girls aged 9–15 years described above, the relationship between baseline total 25(OH)D and subsequent gain in BMD across puberty was particularly strong at the lumbar spine [16] which is rich in trabecular bone. Whereas the present study suggests that 25(OH)D status has minimal effects on cortical bone, it may be that stronger effects exist for trabecular bone which we were unable to evaluate here. A further limitation is the relatively long interval between measurement of 25(OH)D and measurement of cortical bone from pQCT scans, which may have reduced the strength of associations observed between these sets of parametres. Finally, the generalisability of our findings is limited by the fact that the subset of 3,579 subjects forming the basis of the present study is likely to differ in important ways from the original cohort drawn from the general population. For example, maternal social class in the subset on which this paper is based was higher compared with those who were not included (P=0.0001).

In conclusion, we found that in contrast to $25(OH)D_2$, 25 (OH)D₃, as measured in childhood, was positively related to BMC_C, cortical thickness and resistance to buckling as assessed 5 years later. These different associations suggest that supplementation with vitamin D₃ in childhood is likely to prove more beneficial for subsequent cortical bone development compared to vitamin D₂, presumably reflecting important differences between the actions of these two isoforms on bone, the basis of which is currently unclear. Interventional studies are justified in which effects of these two forms of vitamin D are directly compared in the same population, in order to test the conclusions from this observational study, given that we are unable to exclude confounding as a possible explanation for our findings.

Acknowledgements We are extremely grateful to all the families who took part in this study, the midwives for their help in recruiting them and the whole ALSPAC team, which includes interviewers, computer and laboratory technicians, clerical workers, research scientists, volunteers, managers, receptionists and nurses.

The work presented here is funded by a UK Medical Research Council grant (G0701603). The UK Medical Research Council, the Wellcome Trust and the University of Bristol provide core support for ALSPAC. Salary support for AS is provided by Wellcome Trust grant ref. 079960, which also funded the pQCT scans. DAL works in a centre that receives core funds from the UK Medical Research Council (G G0600705) and University of Bristol. No funding body directed the study or interfered with its conduct and interpretation of results; the views presented here are those of the authors and not necessarily any funding body. This publication is the work of the authors who serve as guarantors for the contents of this paper.

Conflicts of interest None.

Open Access This article is distributed under the terms of the Creative Commons Attribution Noncommercial License which permits any noncommercial use, distribution, and reproduction in any medium, provided the original author(s) and source are credited.

References

- 1. Mosekilde L (2005) Vitamin D and the elderly. Clin Endocrinol (Oxf) 62(3):265–281
- Weaver CM (2007) Vitamin D, calcium homeostasis, and skeleton accretion in children. J Bone Miner Res 22(Suppl 2):V45–V49

- Sullivan SS, Rosen CJ, Halteman WA, Chen TC, Holick MF (2005) Adolescent girls in Maine are at risk for vitamin D insufficiency. J Am Dietetic Assoc 105(6):971–974
- 4. Ross AC, Taylor LC, Yaktine AL, Del Valle HB (2010) Committee to review dietary reference intakes for vitamin D and calcium. Institute of Medicine Institute of Medicine
- Winzenberg TM, Powell S, Shaw KA, Jones G (2010) Vitamin D supplementation for improving bone mineral density in children. Cochrane Database Syst Rev 10:CD006944
- El-Hajj Fuleihan G, Nabulsi M, Tamim H et al (2006) Effect of vitamin D replacement on musculoskeletal parameters in school children: a randomized controlled trial. J Clin Endocrinol Metab 91(2):405–412
- 7. Greene DA, Naughton GA (2010) Calcium and vitamin-D supplementation on bone structural properties in peripubertal female identical twins: a randomised controlled trial. Osteoporos Int. Jun 11
- Viljakainen HT, Natri AM, Karkkainen M et al (2006) A positive dose–response effect of vitamin D supplementation on sitespecific bone mineral augmentation in adolescent girls: a double-blinded randomized placebo-controlled 1-year intervention. J Bone Miner Res 21(6):836–844
- Ward KA, Das G, Roberts SA, et al (2010) A randomized, controlled trial of vitamin D supplementation upon musculoskeletal health in postmenarchal females. J Clin Endocrinol Metab. Jul 14
- Houghton LA, Vieth R (2006) The case against ergocalciferol (vitamin D2) as a vitamin supplement. Am J Clin Nutr 84(4):694– 697
- Arunabh S, Pollack S, Yeh J, Aloia JF (2003) Body fat content and 25-hydroxyvitamin D levels in healthy women. J Clin Endocrinol Metab 88(1):157–161
- Parikh SJ, Edelman M, Uwaifo GI et al (2004) The relationship between obesity and serum 1,25-dihydroxy vitamin D concentrations in healthy adults. J Clin Endocrinol Metab 89(3):1196– 1199
- Waldie KE, Poulton R, Kirk IJ, Silva PA (2000) The effects of pre- and post-natal sunlight exposure on human growth: evidence from the southern hemisphere. Early Hum Dev 60(1):35–42
- Sayers A, Tobias JH (2010) Fat mass exerts a greater effect on cortical bone mass in girls than boys. J Clin Endocrinol Metab 95 (2):699–706
- Cheng S, Tylavsky F, Kroger H et al (2003) Association of low 25-hydroxyvitamin D concentrations with elevated parathyroid hormone concentrations and low cortical bone density in early pubertal and prepubertal Finnish girls. Am J Clin Nutr 78(3):485– 492
- Lehtonen-Veromaa MK, Mottonen TT, Nuotio IO, Irjala KM, Leino AE, Viikari JS (2002) Vitamin D and attainment of peak bone mass among peripubertal Finnish girls: a 3-y prospective study. Am J Clin Nutr 76(6):1446–1453
- Tylavsky FA, Ryder KM, Li R et al (2007) Preliminary findings: 25(OH)D levels and PTH are indicators of rapid bone accrual in pubertal children. J Am Coll Nutr 26(5):462–470
- McKay HA, MacLean L, Petit M et al (2005) "Bounce at the bell": a novel program of short bouts of exercise improves proximal femur bone mass in early pubertal children. Br J Sports Med 39(8):521–526
- Cole ZA, Gale CR, Javaid MK et al (2009) Maternal dietary patterns during pregnancy and childhood bone mass: a longitudinal study. J Bone Miner Res 24(4):663–668
- Clark EM, Ness A, Tobias JH (2005) Social position affects bone mass in childhood through opposing actions on height and weight. J Bone Miner Res 20:2082–2089
- Golding J, Pembrey M, Jones R (2001) ALSPAC the Avon Longitudinal Study of Parents and Children: 1. Study methodology. Paediatr Perinat Epidemiol 15:74–87

- 22. Morris N, Udrey J (1980) Validation of a self-administered instrument to assess stage of adolescent development. J Youth Adolesc 9:271–280
- 23. Tobias JH, Steer CD, Mattocks C, Riddoch C, Ness AR (2007) Habitual levels of physical activity influence bone mass in 11 year-old children from the UK: findings from a large population-based cohort. J Bone Miner Res 22:101–109
- 24. Kaptoge S, Beck TJ, Reeve J et al (2008) Prediction of incident hip fracture risk by femur geometry variables measured by hip structural analysis in the study of osteoporotic fractures. J Bone Miner Res 23(12):1892–1904
- 25. Rivadeneira F, Zillikens MC, De Laet CE et al (2007) Femoral neck BMD is a strong predictor of hip fracture susceptibility in elderly men and women because it detects cortical bone instability: the Rotterdam study. J Bone Miner Res 22(11):1781–1790
- Prentice A (2008) Vitamin D deficiency: a global perspective. Nutr Rev 66(10 Suppl 2):S153–S164

- 27. Javaid MK, Crozier SR, Harvey NC et al (2006) Maternal vitamin D status during pregnancy and childhood bone mass at age 9 years: a longitudinal study. Lancet 367(9504):36–43
- Sayers A, Tobias JH (2009) Estimated maternal ultraviolet B exposure levels in pregnancy influence skeletal development of the child. J Clin Endocrinol Metab 94(3):765–771
- 29. Romagnoli E, Mascia ML, Cipriani C et al (2008) Short and longterm variations in serum calciotropic hormones after a single very large dose of ergocalciferol (vitamin D2) or cholecalciferol (vitamin D3) in the elderly. J Clin Endocrinol Metab 93(8):3015–3020
- McGartland CP, Robson PJ, Murray LJ et al (2004) Fruit and vegetable consumption and bone mineral density: the Northern Ireland Young Hearts Project. Am J Clin Nutr 80(4):1019– 1023
- Crozier SR, Robinson SM, Borland SE, Inskip HM (2006) Dietary patterns in the Southampton women's survey. Eur J Clin Nutr 60(12):1391–1399