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A 1024-Channel CMOS Microelectrode Array With 26,400 Electrodes for Recording and Stimulation of Electrogenic Cells In Vitro

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Abstract

To advance our understanding of the functioning of neuronal ensembles, systems are needed to enable simultaneous recording from a large number of individual neurons at high spatiotemporal resolution and good signal-to-noise ratio. Moreover, stimulation capability is highly desirable for investigating, for example, plasticity and learning processes. Here, we present a microelectrode array (MEA) system on a single CMOS die for *in vitro* recording and stimulation. The system incorporates 26,400 platinum electrodes, fabricated by in-house post-processing, over a large sensing area ($3.85 \times 2.10 \text{ mm}^2$) with sub-cellular spatial resolution (pitch of 17.5 µm). Owing to an area and power efficient implementation, we were able to integrate 1024 readout channels on chip to record extracellular signals from a user-specified selection of electrodes. These channels feature noise values of $2.4 \,\mu\text{V}_{rms}$ in the action-potential band (300 Hz-10 kHz) and $5.4 \,\mu\text{V}_{rms}$ in the local-field-potential band (1 Hz-300 Hz), and provide programmable gain (up to 78 dB) to accommodate various biological preparations. Amplified and filtered signals are digitized by 10 bit parallel single-slope ADCs at 20 kSamples/s. The system also includes 32 stimulation units, which can elicit neural spikes through either current or voltage pulses. The chip consumes only 75 mW in total, which obviates the need of active cooling even for sensitive cell cultures.

Index Terms

Extracellular recording and stimulation; high channel count; low noise; low power; microelectrode array (MEA); multirate switched capacitor filter; neural interface; offset compensation; single-slope ADC; switch matrix

I Introduction

EXTRACELLULAR RECORDINGS of the electrical activity of neural and cardiac cell networks in organs such as the brain, the retina, or the heart, can provide a wealth of information about the physiology as well as the pathological degenerations that may cause diseases, such as Parkinson's or Alzheimer's. Microelectrode arrays (MEAs) have been used for a long time for *in vitro* extracellular recordings of electrogenic cell cultures and tissues, such as acute or organotypic brain slices and retinae [1]–[3]. They provide simultaneous multisite recording capability, which is essential to study cellular interconnections and network properties that arise from synchronized cellular activity [4], [5]. However, passive MEAs, which typically include metal electrodes on a glass substrate, are limited in both the number of electrodes (usually less than 300) and the spatial resolution (typically $\geq 30 \ \mu$ m), features that are needed to reconstruct large neural networks at cellular detail.

With CMOS technology, these limitations can be overcome by using multiplexing techniques, which enable access to a large number of closely-spaced electrodes to obtain large sensing areas at high spatial resolution [6]. Moreover, the monolithic integration of recording amplifiers and ADCs, on the same substrate with the electrodes, avoids off-chip parasitics and interference and, at the same time, allows for realizing a large number of recording channels with a low number of connections. Whereas many neural acquisition chips, mostly for *in vivo* applications, have been designed in the last decade (see for example [7]–[14]), only a few implementations of CMOS microelectrode systems have been realized to date. Neural acquisition systems typically interface with arrays or micro-needle probes consisting of only a few hundred electrodes. CMOS MEAs, on the other hand, feature in excess of several thousand recording sites. It is therefore important to provide the capability of recording from such large arrays, while maintaining good signal characteristics. Currently available CMOS MEAs, however, are limited in either spatial resolution [15], [16], noise performance [16]–[18], or readout channel count [15], [19].

One category of CMOS MEAs is based on an active-pixel sensor (APS) architecture [15]– [18]. Since the area for the analog front-end (AFE) amplifier is limited by the pixel size, this scheme results in a tight trade-off between noise, power consumption and spatial resolution. In addition, as all electrodes, even those without significant neural signals, are scanned, the full-frame rate is typically less than 10 kHz [16], [18], due to power constraints. Higher sampling rates are desirable to reconstruct the fast transient of the spike waveforms. In [16] and [18] a subset of the array can be scanned at an increased rate. Nevertheless, the selection is not flexible enough to adapt to complex morphologies or regions of interest.

Another approach is to employ an analog switch matrix [19] to continuously connect a subset of the electrodes to readout units located outside of the sensing area. Each pixel of the

sensing array only contains switches and SRAM cells, leading to a high spatial resolution. The relaxed area constraints for the AFE allow for the implementation of amplifiers with lower noise and anti-aliasing filters. The system proposed in [19] is limited to 126 channels for simultaneous recordings, and, therefore, renders the analysis of large neural networks difficult and time consuming. As an example, ~ 100 subsequent acquisitions are required to cover an array area of $1.8 \times 2.0 \text{ mm}^2$ [20].

In this paper, we present a recently developed CMOS MEA system that further exploits the switch-matrix approach. The system preserves sub-cellular spatial resolution over a large sensing area (8.09 mm²) and features 1024 channels for recording at high temporal resolution (20 kS/s). Despite an eight-fold increase in the channel count with respect to [19], state-of-the-art noise performance has been achieved ($2.4\mu V_{rms}$), owing to an area and power efficient design of the circuitry for amplification and A/D conversion. Moreover, the cross-over distortions and channel-length modulation effects, which had been observed in the design of the stimulation unit in [21], have been largely eliminated. The routing flexibility, provided by the switch-matrix, has been substantially improved, e.g., the high-density blocks can be 5 times larger than those in [19]. Post-CMOS fabrication of electrodes and biocompatible die-bonding and encapsulation have been used to obtain a device that can be handled like a standard MEA dish (see Fig. 1).

This paper is organized in eight sections. Section II presents the system requirements and the proposed architecture. The analog switch-matrix, the readout and the stimulation units are described in Sections III, IV, and V, respectively. Section VI describes the chip implementation and fabrication. Measurement results, including electrophysiological recordings, are given in Section VII. Section VIII compares the chip to the state of the art and concludes the paper.

II System Design

A System Requirements

To enable a broad range of experiments, we aimed at realizing a versatile platform, capable of recording from various *in vitro* and *ex vivo* biological preparations, such as cultured neuronal networks, brain slices, acute retinae and cardiac-cell cultures. High spatial resolution, down to the cellular or sub-cellular level (< 20 μ m), is required to facilitate the task of separating individual signal sources [22], [23]. Such separation is necessary to understand how whole-network properties arise from cellular behavior and inter-cellular connections [5]. In the case of neurons, the cell bodies (somata) have diameters in the range of 5–50 μ m, but the neurites cover a much larger area. As an example, the denditric trees of Purkinje cells extend over several tens or hundreds of μ m [24]. For most cases of neuronal preparations, sub-cellular details of single neurons can be resolved with an electrode pitch of less than 20 μ m [20], [25]. In addition, it is desirable to record simultaneously from distant regions to be able to study interactions between sub-circuits, e.g., in a brain slice, as far as several millimeters away from each other. As a tradeoff with die size, we opted for a rectangular sensing area of ~ 4×2 mm².

The signal levels can vary significantly depending on cell type, distance from the recording electrode, and seal resistance of the cell-electrode cleft [26]. A summary of signal characteristics is reported in Table I. In the case of cardiac myocytes, action potentials (APs) feature amplitudes of up to several tens of mV. In the case of neurons, APs recorded at the soma have amplitudes that are typically in the range 100–500 μ V. In order to also detect low-amplitude spikes from single axons (< 20 μ V[20]), for the readout channels we targeted an input-referred noise of 2 μ V_{rms} in the band 500 Hz–3 kHz, where most spike energy is concentrated [23], corresponding to a thermal noise level of 40 nV/vHz. Further reducing the noise can result in overdesign, at the expense of circuit area or power consumption, since the overall noise performance is limited by the neural background activity and electrode noise (e.g., ~ 80 nV/vHz at 1 kHz, for Pt electrodes with a 25 μ m diameter [26]).

APs have a -3 dB bandwidth typically around 2 kHz [27], with signal content up to 6 kHz. A sampling rate of 20 kS/s is sufficient for most applications. Limiting the recording bandwidth to ~7 kHz reduces the aliased noise from both the electrodes and the circuitry. Local field potentials (LFPs), arising from the synchronized activity of many neurons, can occur simultaneously with APs and exhibit amplitudes of up to a few mV, with frequency components in the range of 1–300 Hz. Therefore, in order to also study LFPs, the readout units must be capable of recording frequencies of a few tens of Hz, while rejecting the large offset and drift of the electrode–electrolyte interface potential (in the range of several hundreds of mV [10], [15]).

Furthermore, versatile electrical stimulation capabilities for precisely and reliably eliciting APs are essential for investigating, for example, mechanisms such as learning and synaptic plasticity in a neural network. Since neurons can be stimulated by either voltage or current signals [21], the availability of both modes is desirable. Typical stimulation pulses have durations of 50–900 μ s, with amplitudes of 0.1–1 V and 50–900 μ A [28]. Finally, to limit the chip-induced heating to less than 2 °C and to avoid active cooling, we aimed for a total power consumption of less than 100 mW.

B Chip Architecture

A block diagram illustrating the system architecture is depicted in Fig. 2. The chip features a sensing area of $3.85 \times 2.10 \text{ mm}^2$ with 26,400 electrodes, placed at a pitch of 17.5 μ m (3,265 electrodes/mm²). A matrix of switches, placed below the electrodes, is used to connect an arbitrarily configurable selection of electrodes to 1024 readout channels and 32 stimulation units, all of which are located outside the electrode area.

To adapt to varying experimental requirements, the readout channels provide programmable bandwidth and gain. The full signal band of 1 Hz–6 kHz can be recorded from each channel. Parallel single-slope ADCs, sharing the ramp generator and a 10 bit counter, are used to digitize signals at 20 kS/s and 10 bit resolution. The stimulation circuits, to deliver both voltage and current stimulation pulses, are grouped in two blocks, each comprising 16 units and three 10 bit DACs. By quickly selecting different DAC outputs, complex stimulation patterns with independent bi-phasic or tri-phasic pulses can be generated at each stimulated electrode. Arbitrary waveforms can also be generated, such as sinusoidal waveforms for low-frequency impedance measurements.

A digital core, operating with two clock domains, transfers the readout data off-chip (24 MHz) and receives control settings through an SPI-like interface (up to 50 MHz), used to configure the array, the readout and stimulation units, and to apply stimulation patterns to the DAC inputs. To ensure data validity, both input and output data streams are protected with CRC checksums.

III Switch Matrix

The electrode array is composed of 220×120 pixels. Each pixel includes an electrode, three switches and two SRAM cells. The schematic of a pixel is shown in Fig. 3. Two switches (S₁ and S₂), with dedicated SRAM cells to hold their on/off states, are used to configure the routing path from any specified electrode to the readout and/or stimulation units. S₁ is used to connect the electrode to a signal wire. Six vertical and six horizontal signal wires are used in each pixel for routing, which are shielded by bit lines (BL), word lines (WL), and supply and ground tracks to minimize cross-talks. Neighboring electrodes are connected each to a different line. The availability of more lines per pixel improves the routing capability.

In [19], the horizontal lines extend over the whole width of the array and are common to each pixel in a row, so that only up to six electrodes per row can be addressed simultaneously. In this new design, to further improve the routing flexibility and to reduce the parasitic capacitance, the signal wires were cut into segments, which extend only for a length of 24 pixels (418 μ m length). These segments can be connected through switch S₂ to form a specific path and reach the boundary of the array to connect to readout and/or stimulation units.

This mesh provides high flexibility to adapt the electrode selection to the morphology of biological samples, for example in sparsely distributed sets, at points of interests, or in high-density blocks with a 17.5 μ m resolution. Due to constraints given by the technology, mainly the minimum pitch between metal wires and the number of metal layers, the largest high-density blocks can contain 23 × 23 electrodes, which is 5 times larger than what was possible with the design in [19], where high-density blocks were limited to 6 × 17 electrodes.

Large switch on-resistance can negatively affect the performance of the recording or stimulation. Transmission gates with around 1 k Ω on-resistance were chosen as switches for the given pixel size. All readout channels can be connected to randomly selected electrodes through an average of 4.4 switches. Only in a few configurations, up to ~20 switches are required to route some electrodes. Even in such cases, the switches contribute a noise density of about 18 nV/ \sqrt{Hz} , which is still lower than the targeted noise level of the readout units. The stimulation units can also be directly connected to the electrode through switch S₃ for a low-resistance path. This switch can be activated through an SRAM cell residing at one side of the array.

Custom-made CAD software was developed to design and implement the switch matrix. The performance of different designs was evaluated, in terms of electrode selection flexibility and the shortest paths between electrodes and readout units, using a mathematical graph

representing the array. Wires and switches of the arrays were mapped to nodes and arcs of the graphs respectively. The connectivity between electrodes and readout channels was modeled as "flow" and the number of overall switches used as "cost" in the algorithm. In order to determine the routing paths and readout channels for all selected electrodes, an algorithm similar to the one in [19] was used. A max-flow min-cost problem is solved through Integer Linear Programming. A variety of electrode configurations, such as randomly chosen electrodes, as well as specific electrode patterns, like large contiguous blocks were evaluated.

The physical layout was then automatically generated from the graph representation, starting from a template containing the electrode, the switches, the SRAM cells, and the wires. Based on the configuration of the mathematical graph, a different set of vias and short-track segments was used in each pixel to join the signal wires of adjacent pixels into 24-pixel-long segments, and to provide the wire-to-switch connectivity. A pattern of 24×24 pixels, formed in this way, was replicated to construct the whole array. Fig. 4 shows a subset of the switch-matrix. Including the periphery of the array, a total of 86,000 switches controlled by 59,000 SRAM cells are used.

The same software was used during experiments to configure the state of the switches and to program a configuration into the SRAM cells.

IV Readout

Good gain uniformity across all channels is desired to reconstruct the actual signal amplitudes and cell positions [24], [29]. Closed-loop amplifier topologies were preferred over open-loop solutions to ensure gain uniformity without the need for calibration. To achieve an overall gain of more than 70 dB, three amplification stages have been employed to reduce the area of passive devices. The schematic of a readout channel is shown in Fig. 5. In the first stage, a low-noise amplifier (LNA) provides a gain of 24 dB and high-pass filtering to reject the electrode offset. The second stage is a variable-gain amplifier (VGA), employing a digitally-assisted offset compensation scheme to cancel the output offset of the LNA. Low-pass filtering is implemented in two steps: the VGA limits the noise bandwidth and provides anti-aliasing filtering, whereas a multirate SC filter (SC LPF) further reduces thermal noise and provides precise control over the cutoff frequency.

Fully differential structures were employed for the whole readout chain, to improve rejection of power-supply interference and substrate coupling, and to reduce power consumption in the SC LPF [30]. The area and power breakdown of a readout channel are shown in Fig. 6.

A Low-Noise Amplifier (LNA)

AC coupling is employed to remove the offset and low-frequency drifts of the electrode potential. While alternative solutions to reset [31] or compensate [10] the input offset of the front-end amplifier have been proposed, these solutions introduce step-shaped artifacts in the recorded traces, as an abrupt change of the DC level in the output waveforms is introduced upon reset or compensation. To avoid such artifacts in the signals, continuous-time filtering was preferred for our design. An input capacitance of 1.45 pF, implemented with stacked

poly-poly and MIM capacitors, was chosen for reduced area usage (196 μ m² for both branches) and high input impedance (110 MΩ at 1 kHz). A small input capacitance also reduces the voltage attenuation caused by the capacitive divider formed with the electrodeelectrolyte double layer capacitance [32] (~ 0.2 pF/ μ m² for Pt electrodes [33]). A low highpass cutoff frequency f_{HP} has been obtained with MOS pseudoresistors [34] in parallel with 89 fF capacitors. The frequency f_{HP} is tunable by adjusting the gate voltage (V_{MOSR}) of transistor M_R [15], [35] and can be set as low as 100 mHz. The possibility to tune f_{HP} allows for increasing the dynamic range in experiments in which LFP recording is not required. The switches activated by the *Reset* signal are employed to quickly recover from amplifier saturation after a stimulation pulse [36]. An alternative scheme is offered by input switches, controlled by the *Disconnect* signal, used to disconnect the amplifier prior to stimulation,

A telescopic-cascode OTA, with the common-mode feedback (CMFB) loop using transistors in the triode-region, is employed in order to minimize the number of current branches. All bias currents flow through the input transistors contributing to their transconductance. The reduced output swing is not an issue in this case due to the small signal amplitudes. For a given current budget, the thermal noise of both the input transistors (M_{Pl}) and the active load (M_{NB}) is reduced by increasing the g_{m}/I_D of M_{Pl} . Since operating in weak inversion requires a very large W/L ratio, M_{Pl} were sized for moderate inversion, with a g_m/I_D of 25 V⁻¹, as a tradeoff between transconductance efficiency and area. The noise contribution of M_{NB} was reduced by operating them in strong inversion, with a g_m/I_D set to 3.8 V⁻¹. For the input transistors, a relatively short length of 1.1 μ m was chosen to limit area usage. With this choice the 1/f-noise corner occurs at 300 Hz, the lower limit of the AP signal band. Higher noise levels can be tolerated in the LFP frequency band, due to larger signal amplitudes [10], [15], [37]. Further increasing the gate area of M_{Pl} also results in a larger input capacitance C_{in} for the OTA. The input-referred noise PSD of the closed-loop amplifier, $S_{n, LNA}$, is related to the noise of the OTA, $S_{n, OTA}$, by the relation:

$$S_{n,LNA} \!=\! \left(\frac{C_{1S} \!+\! C_{1F} \!+\! C_{in}}{C_{1S}} \right)^2 \! S_{n,OTA}$$

Therefore a too large C_{in} can degrade the noise in the closed loop configuration [13], [37].

B Variable-Gain Amplifier (VGA)

thus preventing saturation.

The offset of the LNA, which is mainly caused by the mismatch in the resistances and the leakage currents of the pseudoresistors, can saturate the amplification chain. Performing high-pass filtering in the second stage with pseudoresistors can result in large harmonic distortion due to their non-linearity at larger signal amplitudes. The distortion can become severe at low frequencies, since the total harmonic distortion (THD) depends on the frequency as THD $\propto 1/[1 + (ff_{HP})^2]$ [12], [38]. To avoid this issue, here we employed instead a DC-coupled amplifier with digitally-assisted offset compensation.

A differential-difference amplifier (DDA) with resistive feedback was used to provide high input impedance in a fully-differential structure. Poly-resistors of 10 k Ω/\Box allowed for high

gain in a small area. The gain can be programmed within the range 0–30 dB, with increments of 6 dB. The DDA is based on a folded-cascode topology (see Fig. 7). An additional input differential-pair in the DDA, driven by a 6 bit DAC, is used to implement channel offset compensation without degrading the PSRR or the CMRR of the VGA. The offset compensation is performed off-chip using a binary search algorithm [10]. At each step of the binary search, the individual bit values of all channels are determined off-chip simultaneously; then, these bit values are programmed into the registers for the compensation is only used to reduce the offset of the readout circuits, whereas the electrode offset and drift are removed by AC coupling, the compensation procedure needs to be applied only once per measurement session.

Since accurate models for the offset contributed by the pseudoresistors in the LNA are not available, the compensation range can be controlled globally by adjusting the shared bias current of the DACs.

An alternative design for the VGA in the second stage, based on the same HPF topology of the LNA, was also implemented for comparison. The THD of the two designs is shown in Fig. 8. In the HPF topology, the distortion becomes severe below 300 Hz, where the THD exceeds –45 dB.

C Switched-Capacitor Low-Pass Filter (SC LPF)

The bandwidth of the VGA is susceptible to variations in process and bias current, and is inversely proportional to the closed-loop gain. To ensure a precise low-pass cutoff frequency, SC filtering is used in the third stage of the amplification chain. A multirate operation scheme was employed that allows for boosting the gain with a reduced capacitance spread, without impacting noise performance or circuit complexity, and led to a compact implementation based on a single OTA. A low clock rate signal (f_F = 60 kHz, 80 kHz, 100 kHz) is used to control the switches in the feedback path to obtain a cutoff frequency around 5 kHz with a small C_{3F}/C_{3I} ratio. The input signal is, instead, sampled at a frequency $f_s = M$ f_F , which can be set higher (M = 1, 2, 4) in order to reduce the kT/C noise of the input switches. The transfer function of the SC LPF is given by

$$H(z) {=} \frac{{\sum\limits_{i = 1}^{M} {{C_{3S}}{z^{ - i}}}}}{{{C_{_{3I}}} {+} {C_{_{3F}}} - {C_{_{3I}}}{z^{ - M}}}}$$

with $z = e^{j2\pi f/f_S}$. By sampling and integrating the input signal *M* times before leaking charge through C_{3F_s} a gain of $f_S C_{3S} f_F C_{3F}$ is obtained. Such a scheme results in a low capacitance spread between C_{3S} , C_{3I} and C_{3F_s} . In this design, $C_{3S} = 4 C_U$, $C_{3I} = 2 C_U$, $C_{3F} = 1 C_U$, for a total of 14 unit capacitors C_U . In contrast, a conventional SC circuit with a clock rate of $M f_F$ would require $2 \times (6M + 1)$ unit capacitors to achieve the same gain, power consumption, cutoff frequency and noise performance (50 C_{IJ_s} for M = 4).

D Analog-to-Digital Converter

Recording spikes with amplitudes of tens of μ V, superimposed on LFPs with amplitudes up to a few mV, requires a resolution of at least 9 bit. A resolution of 10 bit was chosen for the single-slope ADC, as a trade-off between resolution and clock rate. The comparators consist of three gain stages with auto-zeroing. Despite a larger static power consumption compared to dynamic comparators, continuous-time comparators were chosen to avoid large kickback noise, since the ramp signal is shared among 1024 ADCs. A capacitive neutralization technique is used in the first gain stage to further reduce the kickback [39]. The input signal is sampled on capacitors C_{C3} during the (ϕ_{FI} -phase of the SC LPF. During the count phase, a continuous ramp signal is produced by integrating a constant current (I_{RAMP}) onto a 20 pF capacitor (C_{RAMP}). A schematic of the ramp generator is shown in Fig. 9. I_{RAMP} is generated by a current conveyor applying a reference voltage V_R across R_{RAMP} . The upper bound of the ADC range is determined by V_{START} . The lower bound is determined by the final value of the ramp, $V_{RAMP}(T_{RAMP}) = V_{START} - V_R T_{RAMP}(R_{RAMP}C_{RAMP})$ and is, therefore, subject to process and temperature variations in R_{RAMP} and C_{RAMP} , IF V_R is fixed. These variations are eliminated by means of a negative feedback loop. The difference between $V_{RAMP}(T_{RAMP})$ and the target voltage V_{END} is sampled on C_E at the end of each sample frame. The charge on C_E is then transferred onto C_S , shifting the voltage V_R by $(C_E/C_s)[v_{RAMP}(T_{RAMP}) - V_{END}]$. After a few sample frames, $v_{RAMP}(T_{RAMP})$ converges to V_{END} , and the slope of the ramp equals $dv_{RAMP}/dt = V_{END}/T_{RAMP}$. The single-ended ramp at the output of the integrator is converted to a differential signal by a capacitive-feedback amplifier, whose gain can be varied in steps of 0.25 from 0.25 to 2.0 for coarse regulation of the ADC range. The timing diagram of the ADC is shown in Fig. 10. During the ramp phase, the amplifier performs single-ended to differential conversion by setting $\phi_R = 1$ and $\phi_S = 0$. At the end of each sample frame, the amplifier is auto-zeroed by setting $\phi_S = 1$. The output common-mode of the differential ramp is set by a SC CMFB. The simulated power consumption of the ramp generator is 1.14 mW, including the output buffer. The shared counter runs with a 24 MHz clock signal (1200 clock cycles per sample frame). Each count phase lasts 1024 cycles for 10 bit operation. All switching operations in the comparators and ramp generator (auto-zeroing, calibration, CMFB of the output buffer) occur only during the additional 176 clock cycles after the count phases to avoid glitches in the ramp. Gray code is used for the Count signal, to avoid the acquisition of spurious values when the comparator triggers at transitions between two consecutive codes.

V Stimulation Units

Each stimulation unit can be configured to provide either voltage or current stimulation (Fig. 11), as was also done in a previous version [21]. The core of each unit is a class-AB opamp, capable of driving loads as large as 10 nF, while maintaining a low static power consumption. In the voltage mode, the circuit is configured as an inverting amplifier with low output impedance. In the current mode, it is configured as a type II current conveyor. The input resistance R_{IN} can be set to either 20 k Ω or 200 k Ω , for coarse adjustment of the current range. Cascoded transistors in the output branch enhance the output impedance to keep the output current constant in the presence of a varying electrode voltage. A pre-level-shifter at the output of the opamp eliminates the cross-over distortion, which has been

observed in [21]. The stimulation units also include an auto-zeroing scheme for offset compensation. In current mode, low offset is crucial, because an offset current can quickly drive the electrode voltage to either VDD or ground, or can induce undesirable electrochemical processes at the electrodes, whose reaction products, such as oxygen or hydrogen, can harm the cells or tissue samples [27]. Since most components are shared in the two modes, each stimulation unit occupies only 0.055 mm².

VI Chip Implementation

A Layout and Floor Plan

The recording amplifiers and the parallel ADCs are grouped in blocks, each comprising 32 channels and shared logic and bias circuits (see Fig. 12). In each block, the amplifiers are arranged in four rows per stage, to reduce the aspect ratio of the layout and the perimeter/ area ratio of the capacitors. Sensitive analog signals are routed via the top metal (MET4), and shielded from underlying circuits by MET3 planes, which carry supply and reference voltages.

B Bias and Test Structures

Bias currents are routed to each readout block and fed by programmable bias generators. These currents can be varied independently for each stage. In the LNA, power consumption can be traded off with noise levels [9]; in the VGA, it can be used for global tuning of the low-pass corner and to adjust the offset-compensation range. Each readout block can be powered-down independently, to permit long-term continuous monitoring with a small number of electrodes at low power levels.

Off-chip access to the input and output of the recording amplifiers or one stimulation unit is provided by dedicated pads and switches. A voltage buffer can be inserted in front of the output pads to drive off-chip loads. Each recording amplifier can be bypassed to characterize the individual gain stages and the ADC independently.

C Chip Fabrication

The chip was fabricated in a 0.35 μ m CMOS technology (2P4M). Platinum electrodes were post-processed at wafer level by means of ion beam deposition and etching. In the same step, three Pt-resistors were fabricated on top of the CMOS passivation for use as temperature sensors. To protect the underlying circuits from the saline solution, used as biological medium, and to avoid cell contamination by the aluminum contained in the CMOS process, a multilayer SiO₂/Si₃N₄ passivation stack was deposited by plasma-enhanced chemical vapor deposition (PECVD). Openings in the passivation to the platinum, defining the actual electrode areas (9.3 × 5.4 μ m²), were then obtained in a reactive-ion etching (RIE) step. A shifted-electrode layout [40] was employed to prevent any leakage of aluminum into the biological medium. Fig. 12 shows a micrograph of the chip and close-up view of the fabricated electrodes. The chip was die-bonded on a custom PCB. A polycarbonate ring was used to contain the biological medium and a bio-compatible epoxy was used to encapsulate the bond-wires [19], [36]. In Fig. 13, an SEM image of the chip surface, plated with rat cortical neurons, is shown.

VII Measurements

A Electrical Characterization

The frequency response of one readout channel for four possible gain settings is shown in Fig. 14. The measured maximum gain is 78.3 dB. The spread of the response across all channels was characterized by applying a common signal to all inputs. Owing to closed-loop topologies and SC filtering, very good uniformity in both the gain and the low-pass corner has been obtained, as shown in Fig. 15. The input-referred noise PSD of the readout chain, including the ADC, is shown in Fig. 16. The noise spectral density is 39 nV/vHz at 1 kHz. The noise integrated over the band 1 Hz–10 kHz is 5.9 μ V_{rms}. In the LFP band (1 Hz–300 Hz) the noise amounts to 5.4 μ V_{rms}, whereas in the AP band (300 Hz–10 kHz) it is 2.4 μ V_{rms}. When filtered in the band 500 Hz–3 kHz, for spike detection, the noise is 1.8 μ V_{rms}. The CMRR of the readout was obtained from measurements on all 1024 channels, resulting in an average of 72 dB. The response of the ADC to a 1.1 kHz sine wave is shown in Fig. 17. The ADC achieves an SNDR of 59 dB and an SFDR of 68.9 dB. Kick-back from the comparators on the shared ramp has been observed. The SNDR degrades by a maximum of 8 dB in the worst-case condition, which occurs when all comparators toggle simultaneously.

The performance of the stimulation unit was assessed with typical biphasic waveforms used for eliciting electrical activity in neurons (Fig. 18). Loads as large as 10 nF can be driven in the voltage mode with pulse durations of 250 μ s. In the current mode, the cross-over distortion is eliminated; channel length modulation effects were also reduced with respect to the design in [21]. To quantify the accuracy of the amplitude of the stimulation pulses, the static linearity has been characterized by sweeping an input DC voltage and extracting the residuals of a best-fit line. In the voltage mode, 10 bit linearity within an output range of 3 V was achieved. In the current mode, the linearity is 9 bits within an output range of $\pm 50 \ \mu$ A.

The measured total chip power dissipation is 75 mW at a supply voltage of 3.3 V. Table II shows the breakdown for the different supply domains. The power consumption of the stimulation units is largely dependent on the applied stimuli, due to its class-AB operation. Also, these blocks can be powered down during experiments that do not require stimulation. Since low power dissipation is crucial for the survival of cells cultured on the chip's surface, the temperature increase of the device, filled with PBS and placed inside an incubator, was monitored by means of the on-chip Pt temperature sensors. An increase < 2° C was observed when all channels were powered up (~ 0.2°C for each group of 128 channels), demonstrating that the chip's self-heating is low enough and thus suitable for cell cultures, without requiring active cooling.

B Biological Measurements

The devices were further verified with *in vitro* and *ex vivo* measurements. Portions of acute *ex vivo* rabbit retina were placed on the chip surface, and spontaneous electrical activity was successfully recorded. A raw trace, as recorded by the CMOS MEA without further processing, is shown in Fig. 19. Action potentials from retinal ganglion cells (RGCs) were detected. The recorded samples are marked with dots in the time zoom. Large-scale recordings were performed with cultures of cortical neurons. The neurons were isolated

from rat brain and plated on the surface of MEA chips, which were pre-coated with poly(ethylenimine) and laminin. Spontaneous activity was observed, and APs were detected with a threshold 5.5 times above the noise rms. Fig. 20 shows a portion of the electrode array, with spike amplitudes obtained from simultaneous recordings of two distant high-density patches, consisting of 23×23 and 15×15 electrodes, respectively. The average spike shapes of two neurons with overlapping electrical footprints are also shown. The low-noise characteristics of the recording channels, combined with high spatial resolution, allowed for identifying and separating the individual signal sources. The stimulation capability was verified by applying biphasic voltage pulses (positive first) with a 200 μ s phase duration and peak-to-peak amplitude of 800 mV. Fig. 21 shows an overlay of 26 raw traces, subsequently recorded on an electrode located 288 μ m from the stimulation site. Spikes were reliably elicited ~ 8 ms after the stimulation pulse.

VIII Comparison to State-of-the-Art and Conclusion

The performance of the chip has been summarized and compared to that of other CMOS microelectrode arrays in Table III. Our device achieves state-of-the-art noise characteristics $(2.4 \,\mu V_{\rm rms}$ in the frequency band of APs) while maintaining high spatial resolution (17.5 μm pitch) and low power consumption. Despite the use of closed-loop topologies, which were adopted to ensure high gain uniformity without the need for calibration, each recording amplifiers occupies a very small area (0.033 mm²) and consumes little power (31 μ W). The area and power efficient design of the readout channels allowed for the integration of 1024 of such units, which is about 8 × the channel count of the switch-matrix-based design reported in [19]. This channel count also exceeds that of all neural acquisition ICs reported in literature (e.g., [7]–[14]), for which the integration of only up to 256 channels has been demonstrated. The simultaneous signal acquisition at many recording sites facilitates the reconstruction of interconnections in neural networks. The presented device also features the largest sensing area (8.1 mm²), which permits the simultaneous recording of large patches in distant regions, to investigate long-range interactions between sub-networks. Stimulation units with both voltage and current stimulation capabilities have also been integrated on chip. All these features make the whole MEA system a versatile platform for numerous biological applications. The chip was used to successfully record activity from a variety of biological preparations, validating the suitability of the device for high-throughput electrophysiological measurements.

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References

- Gross GW, Rhoades BK, Azzazy HME, Wu M-C. The use of neuronal networks on multielectrode arrays as biosensors. Biosens Bioelectron. 1995; 10(6):553–567. [PubMed: 7612207]
- [2]. Jimbo Y, Robinson HP, Kawana A. Strengthening of synchronized activity by tetanic stimulation in cortical cultures: application of planar electrode arrays. IEEE Trans Biomed Eng. 1998 Nov; 45(11):1297–304. [PubMed: 9805828]
- [3]. Stett A, et al. Biological application of microelectrode arrays in drug discovery and basic research. Anal Bioanal Chem. 2003 Oct; 377(3):486–95. [PubMed: 12923608]
- [4]. Marblestone AH, et al. Physical principles for scalable neural recording. Front Comput Neurosci. 2013 Jan.7:137. [PubMed: 24187539]
- [5]. Jiang X, Wang G, Lee AJ, Stornetta RL, Zhu JJ. The organization of two new cortical interneuronal circuits. Nat Neurosci. 2013 Feb; 16(2):210–8. [PubMed: 23313910]
- [6]. Hierlemann A, Frey U, Hafizovic S, Heer F. Growing cells atop microelectronic chips: interfacing electrogenic cells in vitro with CMOS-based microelectrode arrays. Proc IEEE. 2011 Feb; 99(2): 252–284.
- [7]. Aziz JNY, et al. 256-channel neural recording and delta compression microsystem with 3D electrodes. IEEE J Solid-State Circuits. 2009 Mar; 44(3):995–1005.
- [8]. Shahrokhi F, Abdelhalim K, Serletis D, Carlen PL, Genov R. The 128-channel fully differential digital integrated neural recording and stimulation interface. IEEE Trans Biomed Circuits Syst. 2010 Jun; 4(3):149–61. [PubMed: 23853339]
- [9]. Wattanapanitch W, Sarpeshkar R. A low-power 32-channel digitally programmable neural recording integrated circuit. IEEE Trans Biomed Circuits Syst. 2011 Dec; 5(6):592–602.
 [PubMed: 23852555]
- [10]. Muller R, Gambini S, Rabaey JM. A 0.013 mm², 5 μW, DC-coupled neural signal acquisition IC with 0.5 V supply. IEEE J Solid-State Circuits. 2012 Jan; 47(1):232–243.
- [11]. Gao H, et al. HermesE: A 96-channel full data rate direct neural interface in 0.13 μm CMOS. IEEE J Solid-State Circuits. 2012 Apr; 47(4):1043–1055.
- [12]. Guo J, Yuan J, Huang J, Law JK-Y, Yeung C-K, Chan M. 32.9 nV/rt Hz 60.6 dB THD dualband micro-electrode array signal acquisition IC. IEEE J Solid-State Circuits. 2012 May; 47(5): 1209–1220.
- [13]. Guo, J., Ng, W., Yuan, J., Chan, M. A 200-channel 10 μW 0.04 mm2 dual-mode acquisition IC for high density MEA. 2013 Symp VLSI Circuits (VLSIC) Dig; 2013. p. C48-C49.
- [14]. Aziz JNY, Genov R, Bardakjian BL, Derchansky M, Carlen PL. Brain-silicon interface for highresolution in vitro neural recording. IEEE Trans Biomed Circuits Syst. 2007 Mar; 1(1):56–62. [PubMed: 23851521]
- [15]. Heer F, Hafizovic S, Franks W, Blau A, Ziegler C, Hierlemann A. CMOS microelectrode array for bidirectional interaction with neuronal networks. IEEE J Solid-State Circuits. 2006 Jul; 41(7): 1620–1629.
- [16]. Berdondini L, et al. Active pixel sensor array for high spatio-temporal resolution electrophysiological recordings from single cell to large scale neuronal networks. Lab Chip. 2009 Sep; 9(18):2644–51. [PubMed: 19704979]
- [17]. Eversmann B, et al. A 128 × 128 CMOS biosensor array for extracellular recording of neural activity. IEEE J Solid-State Circuits. 2003 Dec; 38(12):2306–2317.
- [18]. Eversmann, B., Lambacher, A., Gerling, T., Kunze, A., Fromherz, P., Thewes, R. A neural tissue interfacing chip for in-vitro applications with 32 k recording/stimulation channels on an active area of 2.6 mm². 2011 Proc ESSCIRC; 2011 Sep. p. 211-214.
- [19]. Frey U, et al. Switch-matrix-based high-density microelectrode array in CMOS technology. IEEE J Solid-State Circuits. 2010 Feb; 45(2):467–482.
- [20]. Bakkum DJ, et al. Tracking axonal action potential propagation on a high-density microelectrode array across hundreds of sites. Nat Commun. 2013 Jul.4:2181. [PubMed: 23867868]

- [21]. Livi P, Heer F, Frey U, Bakkum DJ, Hierlemann A. Compact voltage and current stimulation buffer for high-density microelectrode arrays. IEEE Trans Biomed Circuits Syst. 2010 Dec; 4(6): 372–8. [PubMed: 23850754]
- [22]. Lewicki MS. A review of methods for spike sorting: the detection and classification of neural action potentials. Network. 1998 Nov; 9(4):R53–78. [PubMed: 10221571]
- [23]. Jäckel D, Frey U, Fiscella M, Franke F, Hierlemann A. Applicability of independent component analysis on high-density microelectrode array recordings. J Neurophysiol. 2012 Jul; 108(1):334– 48. [PubMed: 22490552]
- [24]. Frey U, Egert U, Heer F, Hafizovic S, Hierlemann A. Microelectronic system for high-resolution mapping of extracellular electric fields applied to brain slices. Biosens Bioelectron. 2009; 24(7): 2191–2198. [PubMed: 19157842]
- [25]. Fiscella M, et al. Recording from defined populations of retinal ganglion cells using a highdensity CMOS-integrated microelectrode array with real-time switchable electrode selection. J Neurosci Methods. 2012 Oct; 211(1):103–13. [PubMed: 22939921]
- [26]. Guo J, Yuan J, Chan M. Modeling of the cell-electrode interface noise for microelectrode arrays. IEEE Trans Biomed Circuits Syst. 2012 Dec; 6(6):605–13. [PubMed: 23853261]
- [27]. Jochum T, Denison T, wolf P. Integrated circuit amplifiers for multi-electrode intracortical recording. J Neural Eng. 2009 Feb.6(1):012001. [PubMed: 19139560]
- [28]. Wagenaar DA, Pine J, Potter SM. Effective parameters for stimulation of dissociated cultures using multi-electrode arrays. J Neurosci Methods. 2004 Sep; 138(1–2):27–37. [PubMed: 15325108]
- [29]. Holt GR, Koch C. Electrical interactions via the extracellular potential near cell bodies. J Comput Neurosci. 1999; 6(2):169–84. [PubMed: 10333161]
- [30]. Schreier R, Silva J, Steensgaard J, Temes GC. Design-oriented estimation of thermal noise in switched-capacitor circuits. IEEE Trans Circuits Syst I Regul Pap. 2005 Nov; 52(11):2358–2368.
- [31]. Imfeld K, et al. Large-scale, high-resolution data acquisition system for extracellular recording of electrophysiological activity. IEEE Trans Biomed Eng. 2008 Aug; 55(8):2064–73. [PubMed: 18632369]
- [32]. Franks W, Schenker I, Schmutz P, Hierlemann A. Impedance characterization and modeling of electrodes for biomedical applications. IEEE Trans Biomed Eng. 2005 Jul; 52(7):1295–302. [PubMed: 16041993]
- [33]. Robinson DA. The electrical properties of metal microelectrodes. Proc IEEE. 1968; 56(6):1065– 1071.
- [34]. Harrison RR, Charles C. A low-power low-noise CMOS for amplifier neural recording applications. IEEE J Solid-State Circuits. 2003 Jun; 38(6):958–965.
- [35]. Mohseni P, Najafi K. A fully integrated neural recording amplifier with DC input stabilization. IEEE Trans Biomed Eng. 2004 May; 51(5):832–7. [PubMed: 15132510]
- [36]. Heer F, et al. Single-chip microelectronic system to interface with living cells. Biosens Bioelectron. 2007 May; 22(11):2546–2553. [PubMed: 17097869]
- [37]. Venkatraman S, Patten C, Carmena JM. Annu Int Conf IEEE Eng Med Biol Soc. 2009; 2009:2050–3.
- [38]. Yuan S, Johnson LG, Liu CC, Hutchens C, Rennaker RL. Current biased pseudo-resistor for implantable neural signal recording applications. Proc 51st Midwest Symp Circuits Syst. 2008 Aug.:658–661.
- [39]. Figueiredo PM, Vital JC. Low kickback noise techniques for CMOS latched comparators. Proc IEEE Int Symp Circuits and Systems. 2004; 1:I-537–40.
- [40]. Heer F, et al. CMOS microelectrode array for the monitoring of electrogenic cells. Biosens Bioelectron. 2004; 20(2):358–366. [PubMed: 15308242]

Biographies



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After that, he joined the Bio Engineering Laboratory (BEL) at ETH Zurich in Basel as the head of the Circuitry Group. His research interests include analog and mixed-signal integrated-circuit design, with emphases on data converters and biosensor interfaces.



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Alexander Stettler received the Engineering diploma in electrical and communications technology from the Bern University of Applied Sciences, Bern, Switzerland, in 2006. From 2006, he worked Bern University of Applied Sciences as project engineer, where he was responsible for the operation of the cleanroom facility, assisted in research projects and supervised students in semester and diploma theses. In 2009, he received the M.Sc. degree in micro and nanotechnology from the Vorarlberg University of Applied Sciences, Austria.

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David Jäckel received the diploma in electrical engineering from ETH Zurich, Zurich, Switzerland, in 2008. He is currently working towards a Ph.D. degree at the Department of Biosystems Science and Engineering of ETH Zurich in Basel.

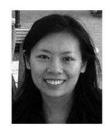
His research interests include signal processing techniques for extracellular neuronal recordings and the combination of intracellular, extracellular, and optical measurements for electrophysiological studies in cultured neuronal networks.



Milos Radivojevic received the Master degree in genetic engineering and biotechnology from the Faculty of Biology, University of Belgrade, Serbia, in 2009. He attended an internship at the Drug Formulation and Delivery Laboratory, Department of Chemistry and Applied Biosciences, ETH Zurich, Switzerland, in 2010. He is currently working towards the Ph.D. degree in neuroengineering at the Bio Engineering Laboratory, in the Department of Biosystems Science and Engineering, ETH Zurich. The focus of his research is investigation of information processing in mammalian neurons and fundamental rules governing learning and memory in neuronal networks.



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Wei Gong received the B.Sc. degree in neuroscience, specialist program, from the University of Toronto, Toronto, ON, Canada, in 2007, and the Master degree in neuroscience from Erasmus University Medical Center, Rotterdam, The Netherlands, in 2011. She is currently working towards the Ph.D. degree at ETH Zurich, Switzerland, in collaboration with Hoffmann-La Roche. Her research focuses on tracing neural activities in hippocampal network from organotypic brain slices, cultivated directly on high-density multi-electrode array (MEA) chips.



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He worked as an interdisciplinary Ph.D. student at the Bio Engineering Laboratory, ETH Zurich, in collaboration with the Neural Circuits Laboratory of Prof. B. Roska, Friedrich Miescher Institute, Switzerland. His Ph.D. work focused mainly on vision, electrophysiology, CMOS-based microelectrode arrays (MEA), data analysis, and modeling of neural network activity.



Douglas J. Bakkum received the B.S. degree in mechanical engineering from the University of Wisconsin, Madison, WI, USA, and the M.S. degree in mechanical engineering and the Ph.D. degree in neuroengineering from the Georgia Institute of Technology, Atlanta, GA, USA. His interdisciplinary background spans research in robotics, artificial intelligence, neuroscience, and art.

His current position is a group leader in the Department of Biosystems Science and Engineering at ETH Zurich, Switzerland. He is searching for the fundamental rules neurons use to communicate with each other and how such rules can scale to produce learning, memory, and creativity.



Flavio Heer (S'01–M'13) received the diploma in physics in 2001 and the Ph.D. degree in 2005 from ETH Zurich, Zurich, Switzerland.

From 2005 to 2007, he was a post-doc at the Physical Electronics Lab, ETH Zurich. He worked on circuit design for CMOS-based microelectrode arrays and biochemical sensors. From 2008 to 2012, he worked as a researcher at the Bio Engineering Laboratory, ETH Zurich, Basel, Switzerland. The focus of his research activities was on circuit design and signal processing for electro-chemical impedance spectroscopy. Currently, he is CTO of Zurich Instruments, Zurich, Switzerland.



Andreas Hierlemann (M'04) completed his college education in chemistry at the University of Tübingen (Diploma 1992) and received the Ph.D. degree in physical chemistry in 1996 from the Eberhard-Karls University, Tübingen, Germany.

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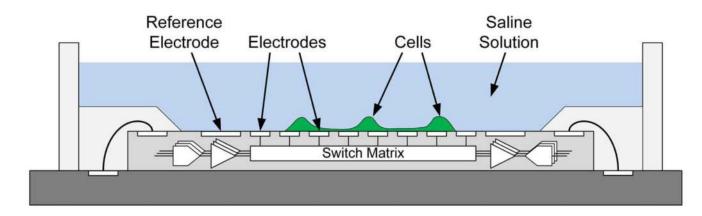
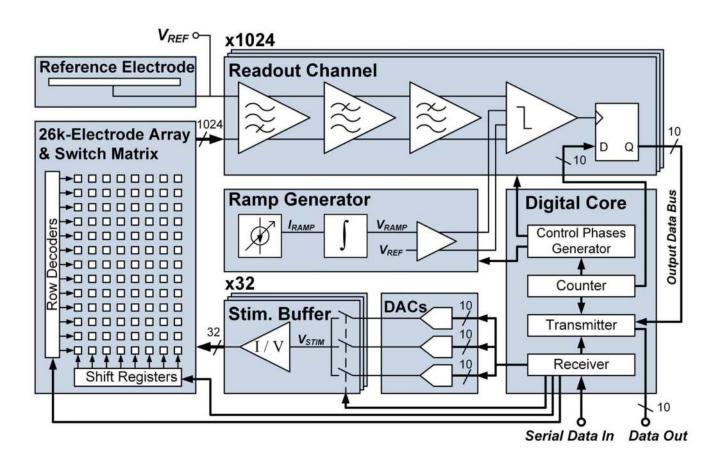


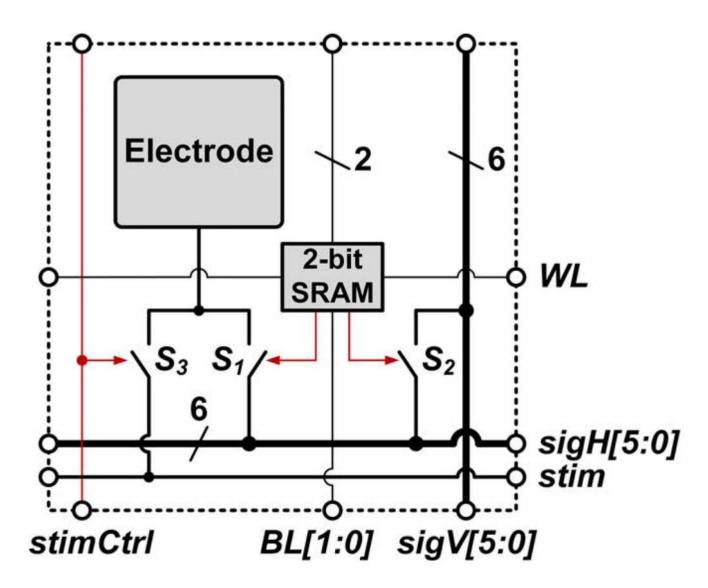


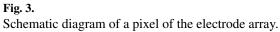
Diagram of the packaged microelectrode array chip (device concept).





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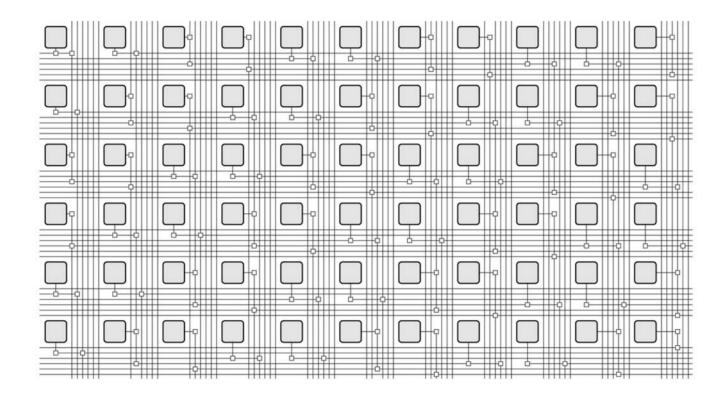
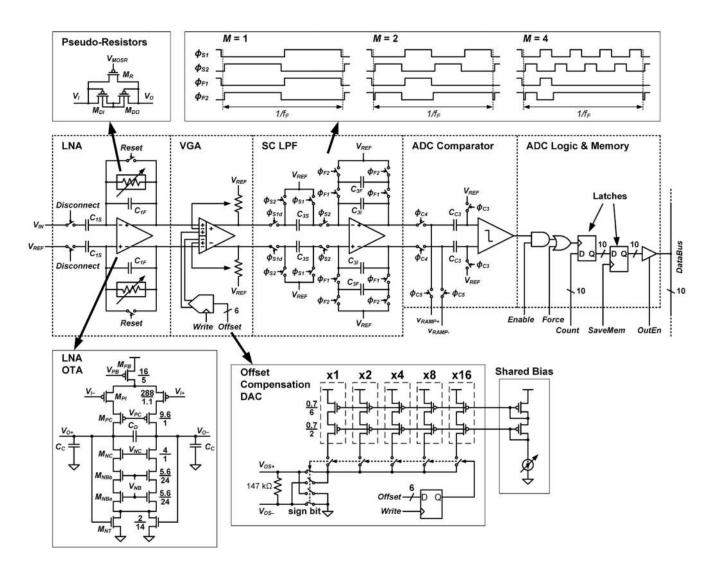


Fig. 4.

Diagram of a subset of the switch-matrix. The larger rectangles represent the electrodes; the smaller rectangular dots represent the switches S_1 and S_2 , used to connect the electrodes to the signal wires and to connect the signal wire segments.





Schematic of a readout channel. The timing diagram for the multirate SC circuit in the third stage is also shown for different values of the multirate factor (M = 1, 2, 4).

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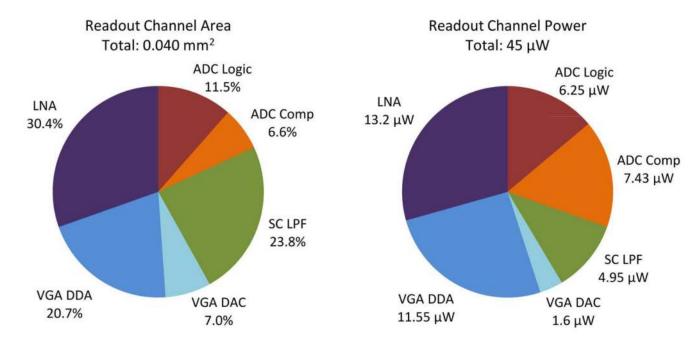


Fig. 6.

Area and power breakdown of a readout channel. The VGA DAC power contribution is for maximum offset compensation applied. The ADC logic includes the digital latches and output buffers, as well as contributions from shared address decoders.

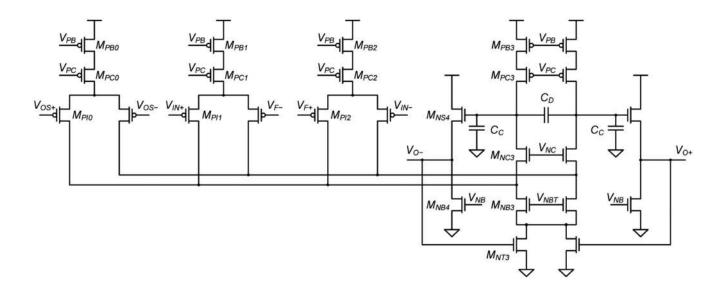


Fig. 7.

Schematic of the DDA in the Variable-Gain Amplifier. An additional input differential pair is used for offset compensation.

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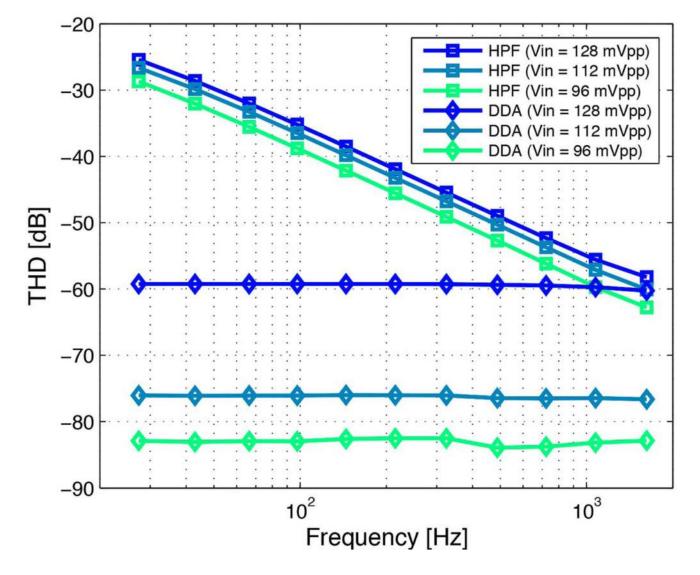
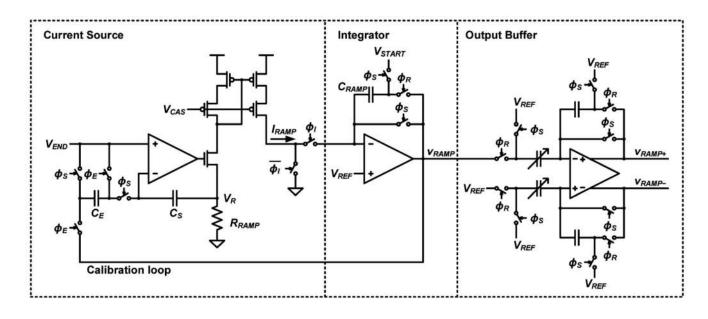
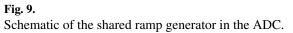


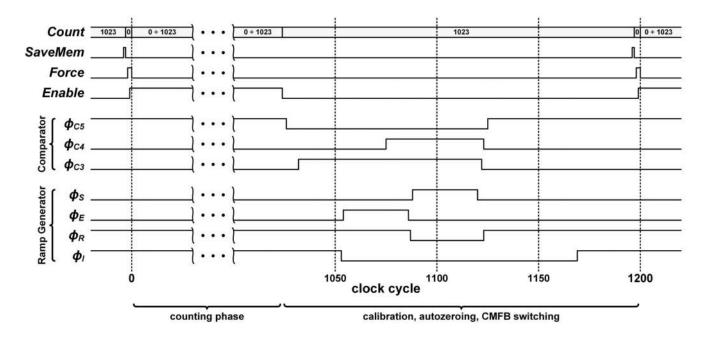
Fig. 8.

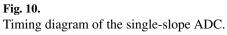
Total harmonic distortion of the VGA used in the second stage of the readout. Comparison between the DDA-based implementation, used in this design, and an high-pass filter (HPF) topology using MOS pseudoresistors.





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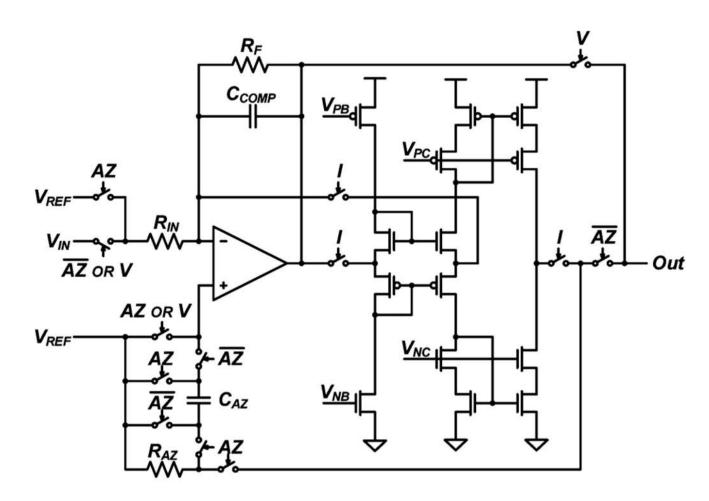


Fig. 11. Schematic of the stimulation unit.

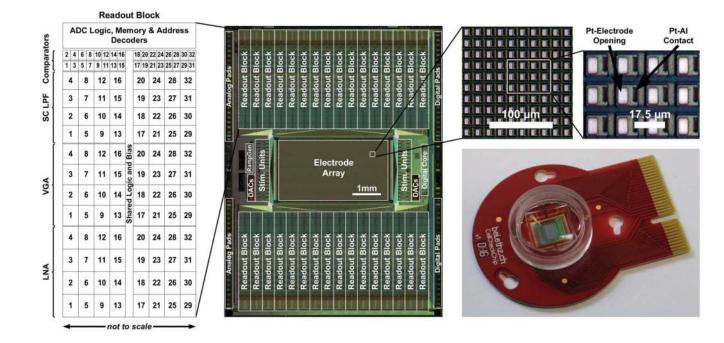


Fig. 12.

Layout of a readout block (32 channels), chip micrograph, close-up view of the electrode array and packaged device. The die size is $7.6 \times 10.1 \text{ mm}^2$

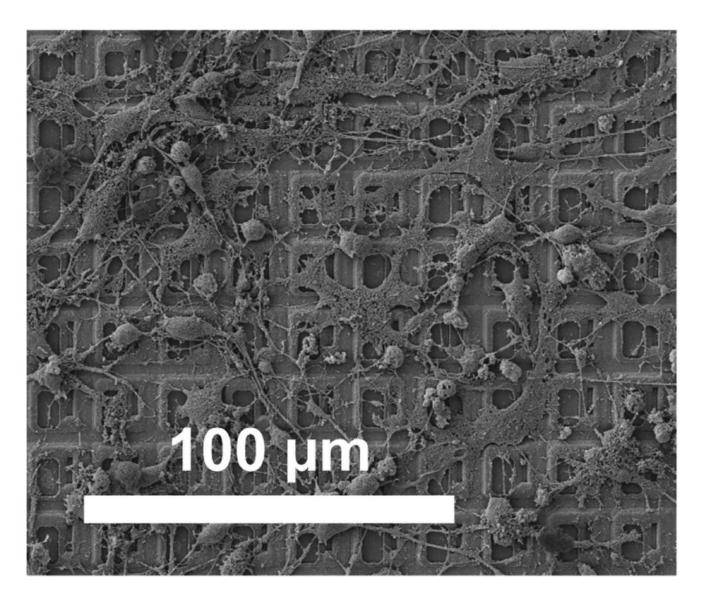
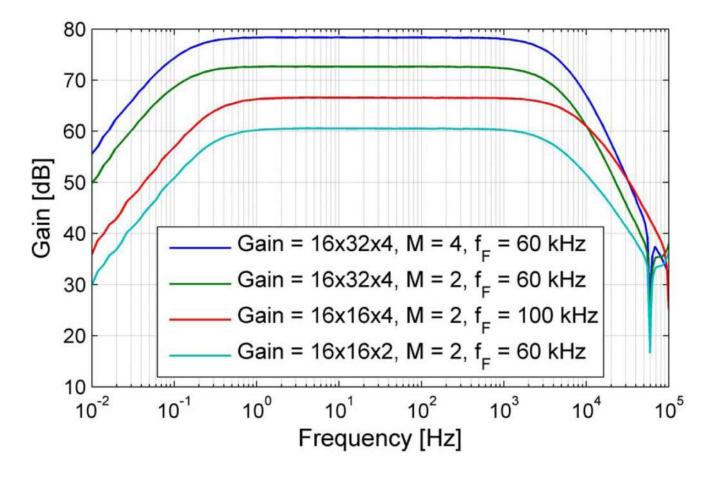


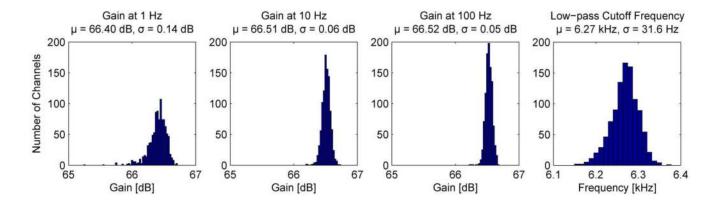
Fig. 13.

SEM image of the chip surface, showing in-house post-processed Pt-electrodes, plated with rat cortical neurons.

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Histograms of the gain (at 1 Hz, 10 Hz and 100 Hz) and the low-pass -3 dB cutoff frequency. A nominal gain of $16 \times 16 \times 4$, with a multirate factor M = 2 and f_F = 100 kHz, was used for this measurement.

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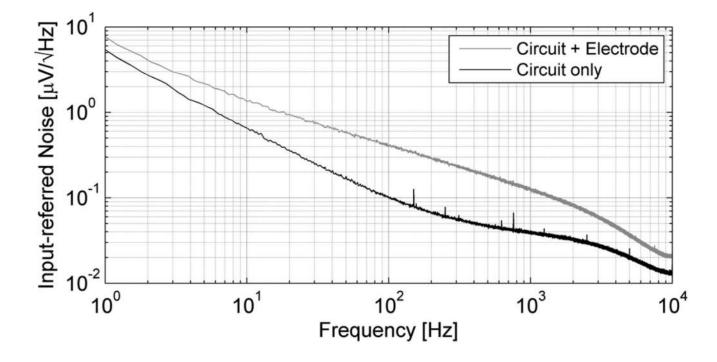
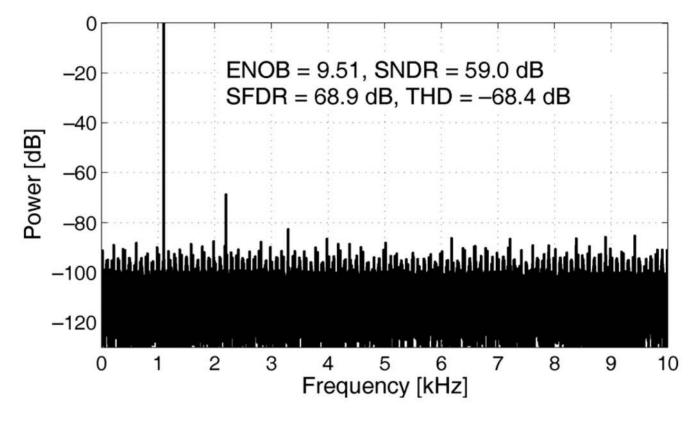
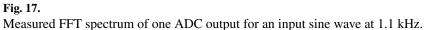


Fig. 16.

Input-referred noise PSD of the full readout chain, including the ADC, with and without electrode contribution.

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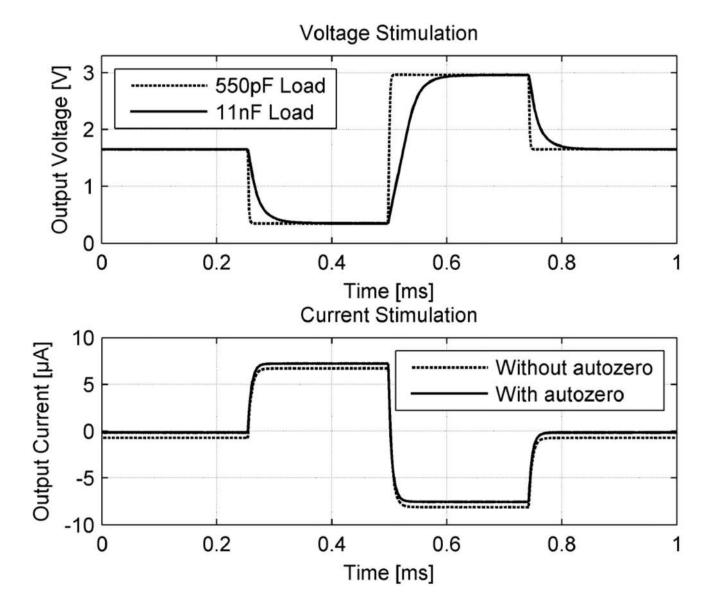


Fig. 18. Biphasic stimulation waveforms in voltage and current mode.

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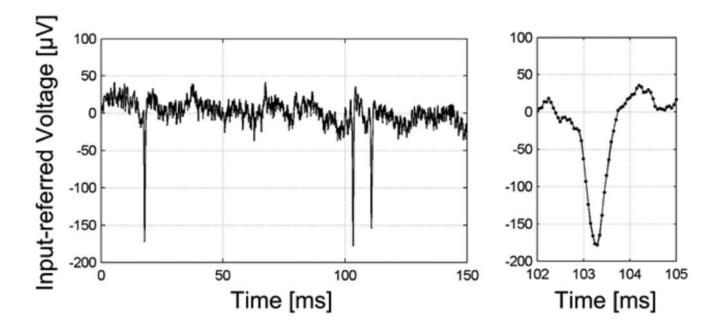


Fig. 19.

Acute *ex vivo* recordings of action potentials from retinal ganglion cells. A raw trace is shown on the left. On the right, a temporal zoom-in of the same trace is shown; recorded measurement values are represented as dots.

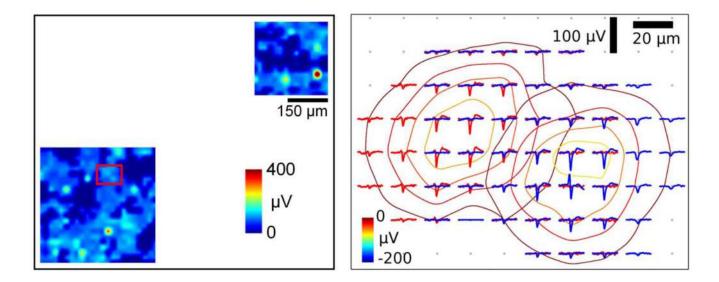


Fig. 20.

Data from cortical neurons, cultured on the chip surface for seven days. Left: Spike amplitude map obtained from simultaneous recordings in two regions of interest. Right: Close-up of the area within the red rectangle recorded at maximum spatial resolution; average spike shapes of individual neurons are shown; contour curves used to identify neighboring neurons represent the amplitude of the spike negative peak.

Stimulation Response

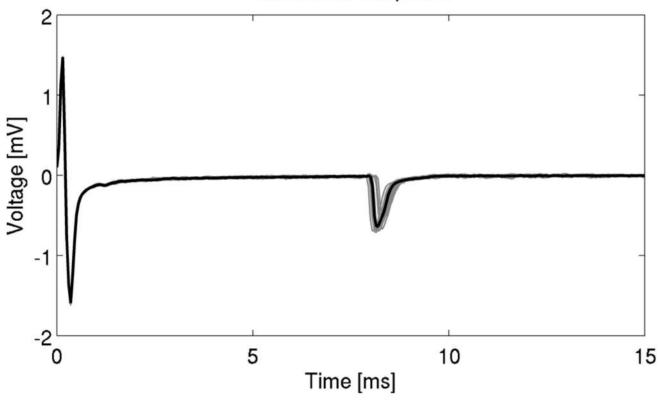


Fig. 21.

Response of a cortical neuron culture to voltage stimulation, recorded at 288 µm from the stimulation electrode. 26 traces of subsequent stimulations and elicited spikes are shown (one trace is shown in black, other traces are in gray).

Table I

Typical Signal Characteristics

Signal Type	Amplitude	Frequency Band
Neuronal Local Field Potentials	< 5 mV	1 Hz – 300 Hz
Neuronal Action Potentials	< 1 mV	300 Hz – 6 kHz
Cardiac Action Potentials	< 50 mV	1 Hz – 1 kHz

Table II

Breakdown of the Measured Power Consumption

Supply	Circuit Blocks	Power [mW]
AVDD 1	LNA, VGA, RampGen	36.8
AVDD 2	SC LPF, ADC Comparators	19
DVDD 1	ADC Memory, Decoders, Logic Buffers	6.4
DVDD 2	Core, I/O	12

Table III

Reference		[18]	[16]	[19]	This work
Туре		Active Pixel	Active Pixel	Switch Matrix	Switch Matrix
Technology		0.5 µm	0.35 µm	0.6 µm	0.35 μm
Supply Voltage		5 V	3.3 V	5 V (3.3 V digital)	3.3 V
Sensing Area		2.60 mm ²	7.13 mm ²	3.50 mm ²	8.09 mm ²
No. Transducers		32768	4096	11016	26400
Pixel Pitch		8.775 μm	42 µm	17.8 μm	17.5 μm
Transducers/mm ²		12987	567	3150	3265
No. Channels		32768	4096	126	1024
A/D Conversion		128 pipeline	off-chip	16 SAR	single-slope
ADC resolution		9 bit	-	8 bit	10 bit
Frame Rate		2.4 kS/s [†]	8 kS/s †	20 kS/s †	20 kS/s
Input Noise	Total	-	$26\mu V_{rms}{}^{\not z}$	2.4 $\mu V_{rms} \ddagger (1 \text{ Hz-10 kHz})$	$5.9 \mu V_{rms} (1 \text{ Hz-}10 \text{ kHz})$
	LFP band	-	-	_	5.4 µV _{rms} (1 Hz-300 Hz)
	AP band	-	-	_	$2.4 \ \mu \ V_{rms} \ (300 \ Hz\text{-}10 \ kHz)$
Stimulation		-	-	V/I	V/I
Total Power		4 W	132 mW	135 mW	75 mW

Comparison to Other CMOS Microelectrode Arrays

 † When reading out all channels.

 \ddagger Recording amplifiers only, excluding ADC.