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## PAPER



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## A benzothiazole-based new fluorogenic chemosensor for the detection of CN<sup>-</sup> and its realtime application in environmental water samples and living cells<sup>+</sup>

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Since the cyanide ion is used in a wide range of industries and is harmful to both human health and the environment, a number of research efforts are dedicated to creating fluorescence sensors for the detection of cyanide (CN<sup>-</sup>). Herein, for the fluorescence detection of CN<sup>-</sup>, a new highly selective and sensitive sensor 2-(3-(benzo[d]thiazol-2-yl)-4-hydroxybenzylidene)-1H-indene-1,3(2H)-dione (BID) was created by conjugating a benzothiazole moiety with 1H-indene-1,3(2H)-dione. The donor and acceptor components of this hybrid receptor were covalently connected through a double bond. The nucleophilic addition of a cyanide anion to the BID inhibits the intramolecular charge transfer (ICT) transition, resulting in spectral and colour alterations in the receptor. When the solvent polarity was increased from n-hexane to methanol, this molecule exhibited a bathochromic shift in the emission wavelength (610 to 632 nm), suggesting the presence of a solvatochromic action. The sensor BID has shown strong specificity towards CN<sup>-</sup> by interrupting its internal charge transfer (ICT), resulting in a significant change in the UV-vis spectrum and a notable blue shift in the fluorescence emission spectrum. The cyanide anion (CN<sup>-</sup>) is responsible for the optical alterations observed by BID, as opposed to the other anions examined. The detection limit was 5.97 nM, significantly less than the WHO's permitted amount of CN<sup>-</sup> in drinking water. The experimental findings indicate that BID's fluorescence response to CN<sup>-</sup> is pH insensitive throughout a wide pH range of 6.0 to 12.0. The interaction mechanism between the BID and CN<sup>-</sup> ions has been studied by HRMS, <sup>1</sup>H-NMR titration experiments, FT-IR, and DFT, which confirmed the nucleophilic addition of CN<sup>-</sup> on vinylidene and subsequent disturbance of ICT. Additionally, we demonstrated the realtime detection application of CN<sup>-</sup> in environmental water samples and live-cell imaging.

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## Introduction

Ions have a variety of important functions in chemical and biological processes.<sup>1-9</sup> Among them, many anions are naturally occurring, and they have significant roles in essential disciplines, including life science, chemistry, medicine, catalysis, and the environment.<sup>10-14</sup> To this end, for the past few years, anion recognition has become an area of growing concern in the field of environmental and biological sciences. Many types of anions are considered to be extremely dangerous, but one in particular, cyanide (CN<sup>-</sup>), has always been of interest all over the world because of its potency and universal presence in many processes

such as manufacturing of plastic and fiber products, tanning, metallurgy, the petrochemical industry, herbicide production, chelators in water treatment, and silver or gold extraction. Despite its overwhelming application in various fields, CN<sup>-</sup> is still harmful to humans even at a dosage of only 0.5 mg per kilogram.15 Any unintentional discharge of cyanide ions will result in significant environmental risks. The hydrolysis of cyanogenic glycosides in plants such as bitter almonds and bitter cassava as well as cherry, apple, and peach seeds can also create toxic HCN besides industrial disposal. The health effects of excessive exposure to CN<sup>-</sup> include paralysis of the central nervous system, poisoning of the respiratory enzymes, and hemoglobin poisoning. As a result, breathing becomes difficult, and the body's cells begin to die from hypoxia (low oxygen levels). Therefore, a more sensitive and selective approach for detecting hazardous cyanide anions is still greatly needed.16

Various analytical techniques have been applied to determine cyanide ion concentration, including voltammetry, electrochemistry, polarography, and gas chromatography.<sup>17-23</sup>

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#### Paper

Despite their low detection limits, these techniques need sophisticated sample processing, costly equipment, and trained operators. Due to the increased popularity of fluorescent CNsensors, which benefit from excellent selectivity, fast reaction time, and less complex sample preparation, these sensors have advanced significantly in the past several years.<sup>24-37</sup> The reported cyanide fluorescence sensors involved the following mechanistic concepts: coordination, hydrogen bonding interaction, anion-metal affinity, nucleophilic addition reactions, and based on nanotechnology.32,38-52 Hydrogen bonding, anion metal affinity, and coordination sensors all have significant drawbacks, including a lack of selectivity and interference from competing analytes such as fluoride, phosphate, and acetate ions. However, among the numerous chemosensors described, reaction-based sensors (chemodosimeters) offer the benefits of high specificity, outstanding sensitivity, and rapid response time. Due to the nucleophilic nature of cvanide ions, reactionbased approaches often exhibit high selectivity for cyanide detection. These factors provide the groundwork for the development of reaction-based sensors for the detection of cvanide ions. For example, some of the already reported reaction-based cyanide sensors include imines,53,54 amide,55,56 hydrazones,57 deprotonation mechanism of C-H,58 acridinium salts,59 cationic boranes,60 oxazines.61 However, slow response time, requirement of high temperature, need of basic medium, high detection limit, less selectivity, inefficiency in aqueous medium hamper the sensing ability of these chemodosimeters. Nucleophilic-based cyanide fluorescence probes have been created and used widely because of their excellent selectivity and increased sensitivity. In recent times, sono-aided chemical precipitation methods have been developed for the synthesis of ZnS, ZnO and quantum dots (QDs).71,72 It exhibits sensational photocatalytic as well as biological activities.73,74. Moreover, the biological toxicity levels of ZnS QDs were characterised by UVvisible absorption and fluorescence emission spectrum.75

With our interest in CN<sup>-</sup> recognition,<sup>34,62,63</sup> in this work, we employ benzothiazole as a fluorophore to develop and manufacture a turn-on cyanide fluorescence probe based on the nucleophilic addition process. A benzothiazole unit was introduced because of its rigid and planar shape and exceptional photophysical characteristics. The donor and acceptor units were covalently bonded together in this hybrid structure by the use of a double bond. Nucleophilic addition to the vinyl double bond causes a disruption of the intramolecular charge transfer (ICT) transition, which results in alterations in both spectral and colour properties. In the probe (BID), the benzothiazole moiety functions as an electron donor, while dione acts as an electron receiver, generating fluorescence through Intramolecular Charge Transfer (ICT). Benzothiazole and 1H-indene-1,3(2H)-dione are linked by Knoevenagel condensation reaction, which renders increased conjugation and nucleophilic addition site for recognizing cyanide ions. When CN<sup>-</sup> ion was added to an ACN/H<sub>2</sub>O solution containing BID, the fluorescence spectra indicated that the substantial fluorescence amplification occurred at 512 nm and the solution colour changed to vellow under a 365 nm UV light. At the same time, BID's absorption spectra showed a definite peak at 593 nm with a high intensity.

**BID**'s determined detection limit for CN was 5.97 nM, which is quite low. The fast response to  $CN^-$ , strong detecting properties, and ease of setup make this new sensor very promising. The sensor **BID** was used to demonstrate the applicability by detecting  $CN^-$  in water, agricultural goods, using test strips, and live cells imaging. It provides a dependable method for detecting  $CN^-$  in environmental and biological systems.

### **Experimental section**

#### Material and apparatus

The required chemicals were obtained from Sigma Aldrich and Alfa Aesar. Chemicals were utilized in the reaction without further purification. Solvents used for synthesis were LR-grade solvents and were used without further purification. Solvents used in spectroscopy were of HPLC grade. Silica gel 100–200 mesh was used for column chromatography. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were taken in BrukerAscent-400 spectrometer at 25 °C. <sup>1</sup>H NMR and <sup>13</sup>C were recorded in CDCl<sub>3</sub> and DMSO-*d*<sub>6</sub>. Chemical shifts ( $\delta$ ) are given in ppm relative to TMS internal standard. Absorption spectra were recorded on a Shimadzu 3600 spectrophotometer. Emission spectra were recorded on a PerkinElmer LS-55 luminescence spectrometer. Samples for absorption and emission were taken in a quartz cuvette (4 ml volume).

Synthesis of 2-(3-(benzo[d]thiazol-2-yl)-4-hydroxybenzylidene)-1*H*-indene-1,3(2*H*)-dione (BID). The compound 3-(benzo[*d*] thiazol-2-yl)-4 hydroxybenzaldehyde was dissolved in analytical grade EtOH (15 ml). To this crude solution, indane-1,3-dione (1 mmol) and a catalytic amount of piperidine was added. Then, the reaction mixture was allowed to reflux for about three hours. TLC confirmed the completion of the reaction, and the pure compound had been isolated by column chromatography (in 50% ethyl acetate in *n*-hexane). The obtained pure **BID** is orange colour solid. The yield was found to be 32%. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  6.968 (s, 1H), 7.425–7.443 (t, 1H, J = 7.2 Hz), 7.533– 7.552 (t, 1H, J = 7.6 Hz), 7.796 (s, 1H), 7.890-7.896 (t, 5H, J = 2.4 Hz), 8.068–8.087 (d, 1H, J = 7.6 Hz), 8.152–8.171 (d, 1H, J = 7.6Hz), 8.676 (s, 1H), 9.448 (s, 1H). <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 100 MHz): δ 100.98, 117.07, 121.49, 125.16, 129.17, 129.42, 131.43, 135.80, 140.89, 152.47, 166.48, 188.14.

**Optical selectivity and titration experiments.** A stock solution of **BID**  $(2 \times 10^{-5}$ M) was prepared in ACN/H<sub>2</sub>O (7 : 3) system. A quartz optical cell of a 1 cm optical path was used to record absorption and emission spectra. For each addition of CN<sup>-</sup>, the concentration of **BID** was maintained at  $2 \times 10^{-5}$  M. The excitation wavelength used for fluorescence analysis was 320 nm, and we got emissions at 393 nm and 408 nm. Stock solutions of various anions such as F<sup>-</sup>, Cl<sup>-</sup>, Br<sup>-</sup>, I<sup>-</sup>, NO<sub>3</sub><sup>-</sup>, SCN<sup>-</sup>, N<sub>3</sub><sup>-</sup>, ClO<sub>4</sub><sup>-</sup>, H<sub>2</sub>PO<sub>4</sub><sup>-</sup>, AcO<sup>-</sup>, HCO<sub>3</sub><sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, and HSO<sub>4</sub><sup>-</sup> were prepared by dissolving their respective salts in double-distilled water.

### Results and discussion

#### Molecular design, synthesis, and structural characterization

With the dione-vinyl group of **BID**, we intended to use the nucleophilic addition reaction of  $CN^-$  to develop a sensor

capable of tracking CN<sup>-</sup> levels. The β-carbon in the dione-the vinyl group is electron-deficient; thus, CN<sup>-</sup> may easily attack this carbon. Due to the presence of two acceptor units on either side of the vinyl group (dione), the **BID** is expected to exhibit increased reactivity and optical response towards CN<sup>-</sup>. Scheme 1 depicts the synthesis process and molecular structure of sensor **BID**. The sensor **BID** was successfully synthesized in one step using the Knoevenagel condensation of 3-(benzo[*d*]thiazol-2-yl)-4-hydroxybenzaldehyde with 1*H*-indene-1,3(2*H*)-dione in EtOH in the presence of pyridine to generate **BID** in 32% yield. Compound 3-(benzo[*d*]thiazol-2-yl)-4-hydroxybenzaldehyde was prepared according to a reported procedure. The structural identification of the sensor **BID** was confirmed by <sup>1</sup>H NMR, <sup>13</sup>C NMR, and HRMS spectra analysis (Fig. S1–S3†).

#### Solvatochromic properties

BID's UV-vis absorption and photoluminescence properties in increasing polarity solutions have been examined (polarity parameter,  $\Delta f$ , was selected as the solvent polarity metric). As shown in Fig. S4,† compound BID in toluene solvent exhibits two major absorption peaks at 300 nm and 410 nm, which can be assigned to  $\pi$ - $\pi$ \* electron transfer and intramolecular charge transfer (ICT). The polarity of solvents had only a modest influence on the absorption spectra of BID (Fig. 1a) and was essentially unaffected by changes in solvent polarity in ground state energy (excluding DMF and DMSO). Changes in solvent polarity, on the other hand, had a substantial impact on fluorescence spectra (Fig. 1b and Table 1). As the solvent polarity rose, the maximum emission wavelength was redshifted from 610 nm in toluene to 632 nm in CH<sub>3</sub>CN, but the relative emission intensity increased simultaneously. The lowest quantum yield  $(\Phi)$  is calculated in the case of toluene, and the highest  $\Phi$  is found in the case of ethanol solvent. The Lippert-Mattaga plot depicts the correlation between the physical characteristics of the solvent (dielectric constant and refractive index) and the Stokes shift of the compound. The graph of Stokes shift of



Scheme 1 The synthetic route of sensor BID.



Fig. 1 Normalized absorption (a) and emission spectra (b) of BID in various solvents; excitation wavelength is 398 nm.

Table 1 Photophysical data of BID at room temperature

Solvent	$\lambda_{\max}^{a}$ (nm)	$\log E^b$	FWHM <sup>c</sup> (nm)	$\lambda_{\mathrm{em}}^{}d}$ (nm)	$\lambda_{ m s}^{e}$ (cm <sup>-1</sup> )	$\phi^{f}(\mathbf{f})$
		-				
Toluene	408	4.8	74	610	8116.36	0.0407
Chloroform	413	9.7	60	616	7979.30	0.0598
DCM	408	4.8	90	614	8223.15	0.0554
1,4-	405	4.8	80	620	8562.32	0.0504
Dioxane						
Ethanol	408	4.8	66	630	8636.78	0.0650
Methanol	404.5	4.7	70	632	8929.6	0.0530
Ethyl	401.5	4.8	72	612	8566.73	0.0471
acetate						
Acetonitrile	398	4.7	84	618	8818.77	0.0519
DMF	618.5	4.9	40	674	2240.57	0.2574
DMSO	600	4.9	54	682	3079.71	0.3217

<sup>*a*</sup> The absorption maximum in nm. <sup>*b*</sup> Molar extinction coefficient in  $M^{-1}$   $L^{-1}$  cm<sup>-1</sup>. <sup>*c*</sup> FWHM in nm. <sup>*d*</sup> The emission maximum in nm. <sup>*e*</sup> The stoke shift in nm. <sup>*f*</sup> Fluorescence quantum yield.

compound **BID** *vs.* orientation polarizability is depicted in Fig. S5.† **BID** showed a rising trend in Stokes shift value with increasing solvent polarity, indicating the presence of ICT and positive solvatochromism. This is a tendency that can only be expected in solvents that do not form hydrogen bonds. However, polar solvents with the capacity to form hydrogen bonds and other particular interactions (complexation and acid–base chemistry) can cause significant alterations. Thus the shift in optical property trend depends on the solvent polarity and can't be entirely predictable.<sup>64</sup>

#### Sensor BID's optical reaction to the CN- anion

We investigated the optical responsiveness of sensor BID in ACN/H<sub>2</sub>O (7:3, v/v) solution by measuring the changes in absorption and fluorescence emission spectra after adding CN<sup>-</sup> anions (2 equiv.). When the representative anions (10 equiv.) F<sup>-</sup>, Cl<sup>-</sup>, Br<sup>-</sup>, I<sup>-</sup>, OH<sup>-</sup>, AcO<sup>-</sup>, NO<sub>3</sub><sup>-</sup>, SCN<sup>-</sup>, OCl<sup>-</sup>, HCO<sub>3</sub><sup>-</sup>, HS<sup>-</sup>,  $SO_4^{2-}$ ,  $HSO_4^{-}$ ,  $PF_6^{-}$ , and  $N_3^{-}$  were introduced to the sensor solution, there was almost no change in the solution colour or optical spectra (Fig. 2a). The absorption band, on the other hand, was significantly altered once the solution was added with 2 equiv. of cyanide anion. Two major absorption peaks at 300 nm and 398 nm diminished in their absorbance intensity with the spontaneous appearance of a strong absorbance peak at 593 nm. When the concentration of CN<sup>-</sup> ions was increased by 0-2 equiv., the isosbestic spot at 455 nm was found in the UVvis absorption spectra, as shown in Fig. 2b. The absorbance intensity at 593 nm varied with cyanide concentration and was balanced with the addition of 26  $\mu$ M CN<sup>-</sup>. As shown in Fig. 2c, a strong linear correlation exists between the absorbance intensity at 593 nm and the concentration of cyanide, which ranges from 0 to 24  $\mu$ M. The linear equation,  $\gamma = 0.0097x +$ 0.0091, was found from the titration experiment, where y is the absorbance intensity at 593 nm, and x is the concentration of  $CN^{-}$ . The  $R^2$  coefficient of determination is 0.9941, and the error bar is within the reasonable range, indicating that probe BID was capable of accurate cyanide detection quantification.



Fig. 2 (a) UV-vis absorption spectrum of sensor BID ( $1 \times 10^{-5}$  M) in the presence of 10.0 equiv. of different anions in an ACN/H<sub>2</sub>O (7 : 3, v/v) solution; (b) sensor BID ( $1 \times 10^{-5}$  M) absorption spectra in a ACN/H<sub>2</sub>O (7 : 3, v/v) solution after addition of different concentrations of CN<sup>-</sup> anions (0–2.6 equiv.); (c) linear relationship between A<sub>593</sub> nm and the cyanide concentration (0–2.6 equiv.).

The fluorescence sensing capability of **BID** towards  $CN^-$  in a solution was then investigated. Excitation of probe **BID** at 398 nm resulted in a maximal emission band with low intensity at 625 nm, as illustrated in Fig. 3a. The addition of different anions such as (10 equiv.)  $F^-$ ,  $Cl^-$ ,  $Br^-$ ,  $I^-$ ,  $OH^-$ ,  $AcO^-$ ,  $NO_3^-$ ,  $SCN^-$ ,  $OCl^-$ ,  $HCO_3^-$ ,  $HS^-$ ,  $SO_4^{2-}$ ,  $HSO_4^-$ ,  $PF_6^-$  and  $N_3^-$  to the sensor solution did not affect **BID**'s colour and emission spectra remained nearly unchanged. The only significant change occurred when  $CN^-$  was introduced, resulting in a blue-shifted fluorescence at 512 nm, was the complete disappearnce of the emission band at 625 nm (Fig. 3a). To further examine **BID**'s  $CN^-$  fluorescence sensing characteristics, a series of fluorescence titration studies with  $CN^-$  were performed (Fig. 3b). The cyanide ion inhibited the ICT transition inside the molecule,



Fig. 3 (a) Fluorescence spectra of BID  $(1 \times 10^{-6} \text{ M})$  in the presence of different analytes (10.0 equiv.  $1 \times 10^{-6} \text{ M}$ ); (b) fluorescence emission of probe BID  $(1 \times 10^{-6} \text{ M})$  in ACN/H<sub>2</sub>O (7 : 3, v/v) solution in the presence of a varying concentration of CN<sup>-</sup> from 0 to 1.5  $\mu$ M, inset shows the plot of  $I_{525}$  vs. the concentration of CN<sup>-</sup>; (c) changes in the solution fluorescence color of BID with different analytes under a UV light of 365 nm.

raising the sensor's emission peak at 512 progressively when CN<sup>-</sup> ions were gradually added to the sensor solution. Fluorescence increase at 525 nm of BID was related to CNconcentrations throughout an extensive range from 0 to  $1.5 \mu M$ , as indicated by the correlation coefficient of 0.9947, suggesting that BID was an excellent sensor for the quantitative detection of CN<sup>-</sup>. This whole event corresponds to a fluorescence colour change of **BID** solution from pink to pale yellow. The sensitivity of BID was also investigated fluorometrically. Various solutions of anions were added individually to the BID solutions. Meanwhile, the visual colour changes were observed as shown in Fig (3c). The detection limit (LOD) of sensor BID for CN<sup>-</sup> was determined to be 5.97 nM with a linear range of concentration to be 0–0.5  $\mu$ M (Fig. S6<sup>†</sup>), which is far below the allowable content of CN<sup>-</sup> in drinking water and the results were compared with previously reported chemosensors for CN-(Table S3<sup>†</sup>). The limit of quantitation (LOO) was estimated to be 1.99 nM, which is calculated based on the equation of  $10\sigma/$ slope.

# Effect of competitive anions, pH, reaction time and salt concentrations

A competitive experiment was conducted to illustrate the unique selectivity of **BID** for CN<sup>-</sup>. Anion CN<sup>-</sup> (2.0 equiv.) was applied to sensor **BID** in the presence of other competing analytes (also 2.0 equiv.) evaluated in the optical experiments. Anions coexisting with CN<sup>-</sup> did not affect its fluorescence change, as demonstrated in Fig. 4a. These results indicate that sensor **BID** has good selectivity for the fluorescence detection of cyanide anion in the presence of another competing anion, making it particularly relevant in practical applications for cyanide detection. In addition, the impact of pH on cyanide anion sensing was investigated. Fig. 4b depicts the changes in



Fig. 4 (a) Competitive experiments of the BID + CN<sup>-</sup> system with interfering anions ( $\lambda_{ex} = 398$  nm) in ACN/H<sub>2</sub>O (7 : 3); (b) the fluorescence intensity of BID (1  $\mu$ M) in the absence and presence of CN<sup>-</sup> ( $\lambda_{ex} = 398$  nm) as a function of pH (1.0–12.0); (c) Job's plot for determining the stoichiometry of BID and CN<sup>-</sup> ions; (d) the time-dependent fluorescence spectrum of BID toward CN<sup>-</sup> in ACN/H<sub>2</sub>O (7 : 3).

sensor BID's fluorescence at 525 nm as a function of pH, with and without the addition of CN<sup>-</sup>. It was found that the fluorescence signal of the free sensor BID in the pH range of 1.0 to 12.0 was very weak and was only marginally influenced by H<sup>+</sup> or OH<sup>-</sup>. When CN<sup>-</sup> is present in the pH range of 1.0-12.0, the fluorescence at 525 nm of the sensor BID increases considerably up to pH = 6, and the BID fluorescence response to  $CN^{-}$ between 6.0 and 12.0 was almost pH-independent. Because CNanions are quickly converted to weak acid, HCN and due to the protonation of BID (the guest), the host-guest contact is reduced, and the fluorescence emission of BID is reduced or diminished. The binding mode of BID-CN<sup>-</sup> was examined by Job's plot experiments. Thereby, the complex (BID- $CN^{-}$ ) demonstrates a maximum absorbance in the 0.5 mole fraction. Hence, 1:1 stoichiometry of complexation has been achieved (Fig. 1c). To determine the probe BID's capability for fast cvanide detection, we studied the time courses of the fluorescence intensity increase of probe BID (1  $\times$   $10^{-6}$  M) in the presence of  $CN^{-}$  (2 equiv.) in a mixture of  $ACN/H_2O(7:3, v/v)$  at room temperature (Fig. 4d). Fig. S7<sup>†</sup> illustrates the fluorescence changes in BID and BID-CN- at various salt concentrations demonstrate that the changes in the exhibited fluorescence curve are minor in NaCl solution, showing that the synthesised **BID** and **BID–CN**<sup>-</sup> are highly stable.

#### Investigation of sensing mechanism

Further, we decided to investigate the **BID-CN**<sup>-</sup> reaction product to learn more about the sensing mechanism of sensor **BID** for CN<sup>-</sup> exposure. The <sup>1</sup>H NMR spectra data in DMSO- $d_6$  at room temperature before and after cyanide anion addition are shown in Fig. 5a. A peak at 8.75 ppm corresponds to the vinyl proton (Ha), which was eliminated following the addition of 1.0 equiv. CN<sup>-</sup>, whereas a new signal at 5.18 ppm corresponds to the  $\alpha$ -proton  $H_b$  appeared. Additionally, the phenolic proton, which appeared at 9.44 ppm, presented an up-field shift to 8.48 ppm. Meanwhile, the aromatic proton signals from 1*H*indene-1,3(2*H*)-dione exhibited a significant up-field shift compared to those from the **BID**. These outcomes may be



Fig. 5 (a) <sup>1</sup>H NMR spectra (400 Hz, DMSO- $d_6$ ) of sensor BID in the absence and presence of CN<sup>-</sup> (1.0 equiv.). (b) The suggested response mechanism for the sensor BID in conjunction with CN<sup>-</sup>.

explained by BID's nucleophilic addition to CN-, which disrupts conjugation between the benzothiazole framework and the 1H-indene-1,3(2H)-dione group and decreases dione's electron-withdrawing capacity. Furthermore, HRMS analysis verified the formation of the newly created adduct (**BID-CN**<sup>-</sup>) (Fig. S3<sup>†</sup>). According to mass spectral analysis, the peak of BID was found to be 383.0618 m/z. After the one equiv. of CN<sup>-</sup> was added into the BID solution, the BID-CN<sup>-</sup> peak with a mass of 408.0566 m/z appeared, showing conclusively that **BID** : CN<sup>-</sup> is a 1:1 stoichiometric ratio. FT-IR spectra were also used to validate the mechanism of interaction between BID and CN-. As depicted in Fig. S8,† it was noticed that in the presence of CN-(1.0 equiv.), the typical stretching bands for the saturated hydrocarbon (CH) group emerged at 2972 and 2858 cm<sup>-1</sup>, while the distinctive stretching vibration of (C=C) at 1581  $\text{cm}^{-1}$ entirely vanished. Meanwhile, the absorption peak of keto (C= O) group stretching vibrations was moved from 1683 to  $1726 \text{ cm}^{-1}$ .

#### Theoretical calculations

The complex formation between the CN<sup>-</sup> ion and BID was studied further at the molecular level utilizing a density functional theory-based computational method. The density functional theory at the B3LYP/631 G(d) level was utilized effectively to predict photochromism and reaction-based photochromic molecules with high accuracy.<sup>29,64-70</sup> The entire geometry optimizations of the probe BID and its interaction with CN<sup>-</sup> were performed using the aforementioned technique, and its structure is shown in Fig. S9.† The transition energies and relative oscillator strengths of the probable important transitions for the **BID** and **BID** +  $CN^{-}$ , have been found (Table S1<sup>†</sup>). As shown in Fig. 6, HOMO of BID has electron densities that are restricted to the 1H-indene-1,3(2H)-dione group alone, and it has been calculated that there is 3.43 eV of energy difference ( $\Delta E$ ) between the HOMO and LUMO ( $E_{HOMO} = -6.4120$  eV and  $E_{\text{LUMO}} = -2.9777 \text{ eV}$ ). As a result of the ICT that happens in the



Fig. 6 The optimal structures of BID and BID +  $CN^-$  determined by DFT/B3LYP methods.

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excited state, it is discovered that a considerable amount of electron transfer occurs between the benzothiazole unit and the 1*H*-indene-1,3(2*H*)-dione functionality (LUMO). Despite this, when the electron allocation in **BID** +  $CN^-$  is taken into consideration, both the HOMO and LUMO localized entirely inside the 1*H*-indene-1,3(2*H*)-dione functionality, resulting in the electronic excitation occurring in the same chromophore, which severely limits the ICT effect.

#### BID loaded paper-based indicators for CN<sup>-</sup> ion

To further evaluate the chemosensor's utility as a quick approach for detecting CN<sup>-</sup> levels in water samples, paper test strips were impregnated with the chemosensor **BID**  $(2 \times 10^{-3})$ M) using a concentrated acetonitrile solution. After dropping different anion solutions (2 mM) onto a "fluorescence paperbased indicator," the colour change was observed while exposed to a 365 nm UV light. Fig. 7 exhibited that the paperbased fluorescent indicator loaded with BID presented dark pink emission. The fluorescence emission remained constant even after adding various anions as tested in the selectivity study. As expected, it could be observed that the fluorescence was blue after CN<sup>-</sup> was dropped. As predicted, it was apparent that the fluorescence had changed from pink to blue once the CN<sup>-</sup> was dropped. BID paper-based indicator can thus be used for the visual inspection of CN<sup>-</sup> in an easy and straightforward manner.

#### Application in real samples analysis

As a result of worldwide industrial operations and improper waste disposal, many water sources have become severely polluted, endangering aquatic life and the ecosystem. Thus, the development of an effective analytical technique for quantifying  $CN^-$  in environmental samples is imperative. The sensitivity of the **BID** to the detection of  $CN^-$  in real water samples such as tap water, distilled water, and pond water from Vellore Institute of Technology (VIT), Vellore campus, was assessed using the conventional addition approach. The collected water samples were filtered three times with a 0.45 mm membrane filter, then spiked with a particular amount of  $CN^-$ , diluted ten times, and were put to the test using standard procedures (Table 2).

The standard addition technique was used to calculate the  $CN^-$  concentration, which involved the addition of 2, 4, and 6  $\mu$ M of  $CN^-$  to the above-mentioned water samples. It can be seen from Table 1 that, with satisfactory recovery (97.4–100.2%) and low RSD (0.024–1.37%), the **BID** sensor was able to detect the spiked  $CN^-$ , demonstrating the feasibility and reliability of the sensor **BID** for the determination of  $CN^-$  in a variety of



Fig. 7 Photographs of BID in the presence of various anions and  $CN^{-}$  under a UV lamp at 365 nm.

Table 2 Results of  $\mathsf{CN}^-$  readings in water samples from the environment

Sample	Spiked (µM)	Found $(X^a \pm SD^b) (\mu M)$	Recovery (%)	$\mathrm{RSD}^{c}$ (%)
Tap water	2.0	$1.992\pm0.007$	99.6	0.18
1	4.0	$3.998 \pm 0.007$	99.9	0.22
	6.0	$6.009 \pm 0.003$	100.2	0.36
Distilled water	2.0	$2.003\pm0.002$	100.2	0.34
	4.0	$3.992\pm0.007$	99.8	0.44
	6.0	$6.011 \pm 0.008$	100.2	0.62
Pond water	2.0	$1.948\pm0.004$	97.4	0.60
	4.0	$3.921 \pm 0.006$	98.0	0.51
	6.0	$5.933 \pm 0.005$	98.9	0.78

 $^a$  Mean of three determinations.  $^b$  SD: standard deviation.  $^c$  RSD: relative standard deviation.

environmental water samples. In addition, the obtained results were cross verified with <sup>1</sup>H-NMR based analysis. In this method, peak intensity gradually increases at 5.18 ppm and is used to derive <sup>1</sup>H-NMR calibration plot (Fig S10 and Table S2†).

#### **Bio-imaging application**

Additionally, *Escherichia coli* (*E. coli*) cells were employed as a model system for further use of as-synthesized **BID** on bacterium imaging. As a result, preliminary cytotoxicity testing was performed. As shown in Fig. S11,† there was no discernible reduction in *E. coli* viability when **BID** concentrations were increased to 20  $\mu$ M, supporting **BID's** extremely minimal impact on *E. coli*. Additionally, it was discovered that *E. coli* had a distinct fluorescent red short dot form and that no apparent restriction of *E. coli* growth was seen under the fluorescence microscope. In the other group, **BID** was incubated with *E. coli* 



**Fig. 8** Fluorescent cell imaging: (a) and (b) bright-field image of *E. coli* cells using the green filter of 510 to 560 nm; (c) fluorescent field image of **BID** treated *E. coli* cells; (d) fluorescent field image of **BID** treated *E. coli* cells incubated with  $CN^-$  ions using the yellow filter of 532 to 587 nm.

bacteria for 20 minutes at 37 °C before being incubated with 20 µM of CN<sup>-</sup> for another 20 minutes, resulting in the development of green fluorescence (Fig. 8d). The results showed that the probe was capable of detecting the CN<sup>-</sup> ion in live cells.

## Conclusions

In summary, a new fluorescence sensor BID based on benzothiazole and a 1H-indene-1,3(2H)-dione derivative for detecting CN<sup>-</sup> has been developed. Optical studies revealed that BID has excellent sensitivity and selectivity toward CN<sup>-</sup> while other anions examined have no effect. Under UV light, a dramatic fluorescent color shift from pink to pale yellow was noticed, as well, as a substantial fluorescence increase was observed when CN<sup>-</sup> was added to the sensor BID solution. The sensing mechanism was built on the nucleophilic addition reaction of the sensor with CN<sup>-</sup>. This was also verified by optical studies, <sup>1</sup>H NMR titration, FT-IR spectra, HRMS, and DFT/TD-DFT calculations. The detection limit was determined to be as low as 5.97 nM, and the quick reaction time following the addition of CN<sup>-</sup>, indicates that sensor BID may be utilized as an effective sensor for CN<sup>-</sup> detection. Additionally, the sensor BID was successfully utilized to detect CN<sup>-</sup> in environmental water samples, enabling the sensor's expansion into food industries. Furthermore, the sensor BID responds to CN<sup>-</sup> with fluorescence, allowing image detection and real-time monitoring of  $CN^{-}$  in live *E. coli* cells.

## Conflicts of interest

There are no conflicts to declare.

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## Notes and references

- 1 R. V. Rathod, S. Bera and D. Mondal, Spectrochim. Acta, Part A, 2020, 238, 118419.
- 2 A. Sala, H. Brisset, A. Margaillan, J.-U. Mullot and C. Branger, TrAC, Trends Anal. Chem., 2022, 148, 116536.
- 3 M. Qiao, L. Ding and F. Lv, Coord. Chem. Rev., 2021, 432, 213696.
- 4 H. Gomaa, S. El-Safty, M. A. Shenashen, S. Kawada, H. Yamaguchi, M. Abdelmottaleb and M. F. Cheira, ACS Sustainable Chem. Eng., 2018, 6, 13813-13825.
- 5 I. M. El-Sewify, M. A. Shenashen, A. Shahat, H. Yamaguchi, M. M. Selim, M. M. H. Khalil and S. A. El-Safty, J. Lumin., 2018, 198, 438-448.

- 6 I. M. El-Sewify, M. A. Shenashen, A. Shahat, M. M. Selim, M. M. H. Khalil and S. A. El-Safty, Microchem. J., 2018, 139, 24-33.
- 7 S. A. El-Safty, M. Khairy, M. A. Shenashen, E. Elshehy, W. Warkocki and M. Sakai, Nano Res., 2015, 8, 3150-3163.
- 8 S. A. El-Safty and M. A. Shenashen, Sens. Actuators, B, 2013, 183, 58-70.
- 9 M. Khairy, S. A. El-Safty, M. A. Shenashen and E. A. Elshehy, Nanoscale, 2013, 5, 7920-7927.
- 10 V. Amendola and L. Fabbrizzi, Chem. Commun., 2009, 513-531.
- 11 S. Liu, D. Yang, Y. Liu, H. Pan, H. Chen, X. Qu and H. Li, Sens. Actuators, B, 2019, 299, 126937.
- 12 R. M. Duke, E. B. Veale, F. M. Pfeffer, P. E. Kruger and T. Gunnlaugsson, Chem. Soc. Rev., 2010, 39, 3936-3953.
- 13 P. A. Gale, J. T. Davis and R. Quesada, Chem. Soc. Rev., 2017, 46, 2497-2519.
- 14 J. T. Davis, P. A. Gale and R. Quesada, Chem. Soc. Rev., 2020, 49, 6056-6086.
- 15 L. Xue, R. Wang, S. Qi, H. Xu, X. Wang, L. Wu, Q. Yang, J. Du and Y. Li, Talanta, 2021, 225, 122100.
- 16 A. Ishii, H. Seno, and K. Watanabe-Suzuki, O. Suzuki and T. Kumazawa, Determination of Cyanide in Whole Blood by Capillary Gas Chromatography with Cryogenic Oven Trapping, https://pubs.acs.org/doi/pdf/10.1021/ac980498b, accessed 29 August, 2021.
- 17 E. PRandviir and C. E. Banks, TrAC, Trends Anal. Chem., 2015, 64, 75-85.
- 18 B. L. Mitchell, G. A. Rockwood and B. A. Logue, J. Chromatogr. B, 2013, 934, 60-65.
- 19 H.-I. Kang and H.-S. Shin, Anal. Chem., 2015, 87, 975-981.
- 20 T. Suzuki, A. Hioki and M. Kurahashi, Anal. Chim. Acta, 2003, 476, 159-165.
- 21 S. Dadfarnia, A. M. Haji Shabani, F. Tamadon and M. Rezaei, Microchim. Acta, 2007, 158, 159-163.
- 22 M. Shamsipur and H. R. Rajabi, Mater. Sci. Eng., C, 2014, 36, 139-145.
- 23 H. R. Rajabi, M. Shamsipur, A. A. Khosravi, O. Khani and M. H. Yousefi, Spectrochim. Acta, Part A, 2013, 107, 256-262.
- 24 Q. Zou, F. Tao, Z. Xu, Y. Ding, Y. Tian and Y. Cui, Anal. Methods, 2019, 11, 5553-5561.
- 25 M. Kargar, H. R. Darabi, A. Sharifi and A. Mostashari, Analyst, 2020, 145, 2319-2330.
- 26 W.-Z. Zhang, Z.-Z. Chen, X.-J. Han and W.-K. Dong, Spectrochim. Acta, Part A, 2021, 258, 119815.
- 27 Z. Shi, Y. Tu, R. Wang, G. Liu and S. Pu, Dyes Pigm., 2018, 149, 764-773.
- 28 R. Chandra, A. Ghorai and G. K. Patra, Sens. Actuators, B, 2018, 255, 701-711.
- 29 Q. Niu, L. Lan, T. Li, Z. Guo, T. Jiang, Z. Zhao, Z. Feng and J. Xi, Sens. Actuators, B, 2018, 276, 13-22.
- 30 S. Bhardwaj, N. Maurya and A. K. Singh, Sens. Actuators, B, 2018, 260, 753-762.
- 31 K. Deng, L. Wang, Q. Xia, R. Liu and J. Qu, Sens. Actuators, B, 2019, 296, 126645.
- 32 C. Yin, F. Huo, M. Xu, C. L. Barnes and T. E. Glass, Sens. Actuators, B, 2017, 252, 592-599.

- 33 P. Ravichandiran, A. Boguszewska-Czubara, M. Masłyk, A. P. Bella, P. M. Johnson, S. A. Subramaniyan, K. S. Shim and D. J. Yoo, *Dyes Pigm.*, 2020, **172**, 107828.
- 34 S. Munusamy, S. Swaminathan, D. Jothi, V. P. Muralidharan and S. K. Iyer, *RSC Adv.*, 2021, **11**, 15656–15662.
- 35 A. K. Mahapatra, K. Maiti, S. K. Manna, R. Maji,
  C. D. Mukhopadhyay, B. Pakhira and S. Sarkar, *Chem.-Asian J.*, 2014, 9, 3623–3632.
- 36 P. Torawane, S. K. Sahoo, A. Borse and A. Kuwar, *Luminescence*, 2017, **32**, 1426–1430.
- 37 C.-B. Bai, J. Zhang, R. Qiao, Q.-Y. Zhang, M.-Y. Mei, M.-Y. Chen, B. Wei, C. Wang and C.-Q. Qu, *Ind. Eng. Chem. Res.*, 2020, **59**, 8125–8135.
- 38 K. Wang, X. Yu, Y. Zhu, M. Xing, M. Liang, Y. Sun, D. Cao, R. Guan and Z. Liu, *J. Mol. Struct.*, 2018, **1173**, 647–652.
- 39 W.-J. Qu, H.-H. Yang, J.-P. Hu, P. Qin, X.-X. Zhao, Q. Lin, H. Yao, Y.-M. Zhang and T.-B. Wei, *Dyes Pigm.*, 2021, 186, 108949.
- 40 K.-S. Lee, J. T. Lee, J.-I. Hong and H.-J. Kim, *Chem. Lett.*, 2007, **36**, 816–817.
- 41 T.-B. Wei, J.-D. Ding, J.-F. Chen, B.-B. Han, X.-M. Jiang, H. Yao, Y.-M. Zhang and Q. Lin, *New J. Chem.*, 2018, 42, 1271–1275.
- 42 P. R. Lakshmi, P. S. Kumar and K. P. Elango, *Spectrochim. Acta, Part A*, 2020, **229**, 117974.
- 43 L. Wang, Z.-L. Wei, Z.-Z. Chen, C. Liu, W.-K. Dong and Y.-J. Ding, *Microchem. J.*, 2020, **155**, 104801.
- 44 P. S. Kumar, S. Ciattini, C. Laura and K. P. Elango, J. Mol. Liq., 2020, 317, 113970.
- 45 S. Ramesh and S. Kumaresan, *Microchem. J.*, 2021, **169**, 106584.
- 46 Q. Wu, S. Wang, E. Hao and L. Jiao, *Spectrochim. Acta, Part A*, 2021, 247, 119102.
- 47 Y. Wang, J. Wang and Q. Xian, Talanta, 2018, 190, 487-491.
- 48 L. Patra, K. Aich, S. Gharami and T. K. Mondal, *J. Lumin.*, 2018, **201**, 419–426.
- 49 G. Kumar, N. Gupta, K. Paul and V. Luxami, *Sens. Actuators, B*, 2018, **267**, 549–558.
- 50 S. Li, F. Huo, K. Ma, Y. Zhang and C. Yin, *New J. Chem.*, 2021, 45, 1216–1220.
- 51 B. Aydıner, Ö. Şahin, D. Çakmaz, G. Kaplan, K. Kaya, Ü. Ö. Özdemir, N. Seferoğlu and Z. Seferoğlu, *New J. Chem.*, 2020, 44, 19155–19165.
- 52 H. Gomaa, M. A. Shenashen, A. Elbaz, S. Kawada, T. A. Seaf El-Nasr, M. F. Cheira, A. I. Eid and S. A. El-Safty, *J. Colloid Interface Sci.*, 2021, **604**, 61–79.
- 53 B. Yilmaz, M. Keskinates, Z. Aydin and M. Bayrakci, J. Photochem. Photobiol., A, 2022, 424, 113651.

- 54 A. Pundi, J. Chen, C.-J. Chang, S.-R. Hsieh, M.-C. Lee, C.-H. Chou and T.-D. Way, *Spectrochim. Acta, Part A*, 2021, 262, 120139.
- 55 E. Kandemir, M. Özkütük, B. Aydıner, N. Seferoğlu, H. Erer and Z. Seferoğlu, J. Mol. Struct., 2022, 1249, 131593.
- 56 T. P. Martyanov, A. A. Kudrevatykh, E. N. Ushakov, D. V. Korchagin, I. V. Sulimenkov, S. G. Vasil'ev, S. P. Gromov and L. S. Klimenko, *Tetrahedron*, 2021, 93, 132312.
- 57 X. Lv, J. Liu, Y. Liu, Y. Zhao, M. Chen, P. Wang and W. Guo, Org. Biomol. Chem., 2011, 9, 4954–4958.
- 58 T. Zhu, Z. Li, C. Fu, L. Chen, X. Chen, C. Gao, S. Zhang and C. Liu, *Tetrahedron*, 2020, **76**, 131479.
- 59 Y.-K. Yang and J. Tae, Org. Lett., 2006, 8, 5721-5723.
- 60 T. W. Hudnall and F. P. Gabbaï, *J. Am. Chem. Soc.*, 2007, **129**, 11978–11986.
- 61 S. Sanjabi, J. Keyvan Rad and A. R. Mahdavian, *J. Photochem. Photobiol.*, *A*, 2022, **424**, 113626.
- 62 D. Jothi, S. Munusamy and S. K. Iyer, *J. Photochem. Photobiol.*, *A*, 2021, **420**, 113491.
- 63 D. Jothi, S. Munusamy, S. Sawminathan and S. K. Iyer, *RSC Adv.*, 2021, **11**, 11338–11346.
- 64 S. Nigam and S. Rutan, Appl. Spectrosc., 2001, 55, 362A-370A.
- 65 S. Sawminathan, S. Munusamy, D. Jothi and S. K. Iyer, *ChemistrySelect*, 2021, **6**, 858–864.
- 66 Q. Zou, F. Tao, H. Wu, W. W. Yu, T. Li and Y. Cui, *Dyes Pigm.*, 2019, **164**, 165–173.
- 67 T. Sheshashena Reddy, H. Moon and M.-S. Choi, *Spectrochim. Acta, Part A*, 2021, 252, 119535.
- 68 H. Jain, N. Deswal, A. Joshi, C. N. Ramachandran and R. Kumar, *Anal. Methods*, 2019, **11**, 3230–3243.
- 69 P. Ghorai, K. Pal, P. Karmakar and A. Saha, *Dalton Trans.*, 2020, **49**, 4758–4773.
- 70 G. Shally, V. Kumar, I. Althagafi, A. Kumar, D. Singhal, A. Kumar, R. Gupta and R. Pratap, *New J. Chem.*, 2020, 44, 15559–15566.
- 71 H. R. Rajabi, F. Karimi, H. Kazemdehdashti and L. Kavoshi, J. Photochem. Photobiol., B, 2018, 181, 98–105.
- 72 H. R. Rajabi, R. Naghiha, M. Kheirizadeh, H. Sadatfaraji, A. Mirzaei and Z. M. Alvand, *Mater. Sci. Eng.*, *C*, 2017, 78, 1109–1118.
- 73 F. Karimi, H. R. Rajabi and L. Kavoshi, *Ultrason. Sonochem.*, 2019, 57, 139–146.
- 74 H. R. Rajabi, H. Arjmand, H. Kazemdehdashti and M. Farsi, J. Environ. Chem. Eng., 2016, 4, 2830–2840.
- 75 B. Hemmateenejad, M. Shamsipur, F. Samari and H. R. Rajabi, *J. Iran. Chem. Soc.*, 2015, **12**, 1729–1738.