- 1 A common framework for identifying linkage rules across different types of
- 2 interactions.
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- 20 Running headline: Linkage rules in interaction networks
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23 Summary

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25 1. Species interactions, ranging from antagonisms to mutualisms, form the architecture 26 of biodiversity and determine ecosystem functioning. Understanding the rules 27 responsible for who interacts with whom, as well as the functional consequences of 28 these interspecific interactions, is central to predict community dynamics and stability. 29 2. Species traits sensu lato may affect different ecological processes by determining 30 species interactions through a two-step process. First, ecological and life-history traits 31 govern species distributions and abundance, and hence determine species co-32 occurrence and the potential for species to interact. Second, morphological or 33 physiological traits between co-occurring potential interaction partners should match for 34 the realization of an interaction. Here, we review recent advances on predicting 35 interactions from species co-occurrence, and develop a probabilistic model for inferring 36 trait matching. 37 3. The models proposed here integrate both neutral and trait-matching constraints, 38 while using only information about known interactions, thereby overcoming problems 39 originating from under-sampling of rare interactions (i.e. missing links). They can easily 40 accommodate gualitative or guantitative data, and can incorporate trait variation within 41 species, such as values that vary along developmental stages or environmental 42 gradients. 43 4. We use three case studies to show that the proposed models can detect strong trait 44 matching (e.g. predator-prey system), relaxed trait matching (e.g. herbivore-plant

45 system) and barrier trait matching (e.g. plant-pollinator systems).

5. Only by elucidating which species traits are important in each process (i.e. in
determining interaction establishment and frequency), can we advance in explaining
how species interact and the consequences of these interactions for ecosystem
functioning.

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Key words: Trait matching, functional traits, interaction networks, pollination, predation,
herbivory, parasitism, mutualisms, trophic interactions.

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54 Introduction

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Species interactions form the architecture of biodiversity (Bascompte & Jordano 2007). 56 57 There is growing recognition that community structure, stability and functioning depend not only on which species are present in a community, but also on how they interact 58 59 (Tylianakis et al. 2008). Complex networks of biotic interactions such as predation, 60 parasitism and mutualism provide essential information related to conservation 61 (Carvalheiro, Barbosa & Memmott 2008; Tylianakis et al. 2010), community stability and ecosystem functioning (Thompson et al. 2012; Peralta et al. 2014), and evolutionary 62 63 processes (Jacquemyn et al. 2011; Fenster et al. 2015). These insights would be not possible from simple species occurrence data or analysis of pairwise interactions. 64 65 Despite the growing literature describing species interaction networks, we still have a poor understanding of how network structure comes to exist. 66 67 There are few generalizable observations of how species interactions respond to 68

69 environmental changes (Tylianakis et al. 2008). Therefore, understanding what

determines the occurrence of pairwise interactions, and, at a higher level, the structure 70 71 of ecological networks, is a key challenge for ecologists. Overcoming this challenge 72 requires the identification of the mechanisms responsible for who interacts with whom. 73 Natural selection promotes adaptations to increase species efficiency (Castellanos et al. 74 2003). Reciprocal trait adaptations between partners, which have positive demographic 75 consequences, lead to increased interaction strength among co-evolved members 76 (Sargent and Ackerly 2008). Hence, there is a great expectation that incorporating a trait-based approach can help us explain general mechanisms driving pairwise 77 78 interactions. We refer here to traits in a broad sense, comprising adaptations that define 79 organisms in terms of their ecological role, how they interact with the environment and 80 with other species (Díaz & Cabido 2001). Most traits studied so far for predicting 81 species interactions fall into morphological adaptations (e.g. body size), but physiological (e.g. chemical defenses) or behavioral (e.g. diel) adaptations can also 82 83 drive species interactions. Moreover, traits should be precise and measurable attributes 84 of the species. Recent studies indeed suggest that ecological networks of different types (e.g. from antagonistic to mutualistic) could be described from the traits of the 85 86 interacting species (Eklöf et al. 2013). The ability of these methods to predict novel interactions following species invasions or following range shifts is, however, limited. 87 88

Traits are implicated in ecological dynamics at several concatenated levels of
community organization (Fig. 1), and therefore could influence the occurrence of
interactions in multiple ways. Some traits determine species distributions in a multidimensional environmental space, and thus impact co-occurrence in space and time.

93 Since the occurrence of an interaction requires the presence of the two species, traits 94 involved in phenological matching or habitat filtering could constrain interactions. Lifehistory traits impact demography, abundance and biomass, thereby affecting the 95 96 probability of encounter. Then, provided they encounter each other in space and time, 97 the compatibility between traits of the two species (i.e. trait-matching constraints) will also determine whether or not they interact. Finally, the intensity and the impact of an 98 99 interaction will determine the functioning of the network, and also feed back to 100 determine species abundances and dynamics. How efficient an interacting species is on 101 a per capita basis is also likely to be mediated by its behavioural or physiological traits 102 and how these match with those of the other species. Of course, these levels interact 103 with each other through evolutionary processes. Most work to date has focused on 104 morphological trait matching and little, if any, has tackled several of these stages at a 105 time (see the review in Morales-Castilla et al. 2015). Our first objective here is to review 106 what we know about each of these processes and assess their success and limitations 107 at predicting interactions. Our second objective is to propose a way forward to evaluate 108 trait matching in a way that is not confounded with species co-occurrences, and how 109 this can be integrated into a larger framework, from species occurrences to ecosystem 110 functioning.

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112 Traits governing species encounters in space and time.

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114 Habitat filtering constrains the pool of co-occurring species in a region or microhabitat.

115 Sharing habitat-filtering traits, like tolerance to drought or thermal preference, may

116 hence be a prerequisite for two species to interact. Even in large and diffuse networks 117 such as the global planktonic interactome, 18% of the variance in community 118 composition (taxon presence and abundance) can be explained by environment alone. 119 and these co-occurrences can be used successfully to predict interactions between taxa 120 (Lima-Mendez et al. 2015). Microhabitat characteristics can also influence sessile 121 organisms even within close proximity, as shown by interactions among mycorrhizas 122 and plants, where rooting depth could preclude co-occurrence between shallow rooted 123 plants and fungi restricted to lower soil horizons. In fact, the concept of "habitat 124 associations" as a driver of interactions has been pointed to as the sole explanation for 125 these interactions (Zobel & Opik 2014), suggesting that both partners interact simply 126 because they respond independently to different environmental factors.

127

128 At broader spatial scales, species turnover along ecological gradients can also be 129 responsible for a large fraction of network variation in space (Poisot et al. 2012). Range 130 overlap determines the location and the total area over which two species can 131 eventually interact. This can be used to better understand the consequences of range 132 shifts on the local food-web structure (Albouy et al. 2014). Species distribution models in combination with ecological and life-history traits (D'Amen et al. 2015) can be used to 133 134 predict co-occurrence and potential interactions in response to global changes (Albouy 135 et al. 2014, Morales-Castilla et al. 2015).

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137 Similar to species distribution in space, species encounter will be determined by the

138 synchrony of their activity periods at different temporal scales (i.e. daily, seasonal,

139 interannual). Mismatch of phenology has been widely called to explain undetected

140 interactions that are not possible to occur (i.e. forbidden interactions; Olesen et al. 2011; 141 Encinas-Viso, Revilla & Etienne 2012; Olito & Fox 2015), that is, species present in the 142 same location that do not interact because they do not overlap in their seasonal activity 143 periods. Phenological overlap during the season has therefore been used as a proxy for 144 interaction probability (Bartomeus et al. 2013). While phenology is usually studied as 145 the timing when species are active during a season (e.g. plant flowering period), daily 146 fluctuations of activity can also be important for defining when interactions among 147 partners can occur. A clear example is the distinction between crepuscular vs. diurnal 148 species (Herrera 2000), but more subtle fluctuations of activity depending on daily 149 temperature may be also relevant (Rader *et al.* 2013). In addition, some species may 150 interact only with partners in a given life-history stage, for example, some 151 ectomycorrhizal fungi may require host trees to be at least several years old and do not 152 interact with seedlings. This highlights the importance and complexity of the temporal 153 constraints on co-occurrence.

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155 Given that species co-occur in space and time, their abundance also determines the 156 frequency at which they will interact (Canard et al. 2014). Abundant species are simply 157 more likely to encounter each other than rare ones. This mechanism has been called 158 neutral because it does not rely on any niche differentiation. Thus, models that use 159 species abundances to predict encounter probabilities have found that abundance alone 160 can explain considerable variance in key aspects of network structure (Vázquez et al. 161 2007: Krishna et al. 2008: Olito & Fox 2015). Abundance is determined primarily by life-162 history traits (e.g. fecundity, longevity, mortality). For plant communities, there is some 163 consensus over which traits relate to abundance or dominance in the community, such

164 as maximal height and position along the slow-fast continuum (e.g. leaf economic 165 spectrum; Wright et al. 2004). Therefore, trait distributions over environmental gradients 166 have been used to predict plant abundance and community structure (Shipley, Vile & 167 Garnier 2006; Laughlin et al. 2012). Similarly, it is possible to relate life-history traits to 168 animal abundances. For instance, species with fast life cycles (usually small, with high 169 reproduction rates and short longevity) tend to be more abundant than large species 170 with slow life histories (White et al. 2007), and large species can decline more rapidly 171 following habitat change (Larsen, Williams & Kremen 2005). As a result, abundance can 172 be largely related to body size and position in the interaction network (Woodward et al. 173 2005). Overall, the relationships between traits, abundance and probability of encounter 174 defines the neutral expectations for interacting. This relationship is complex, for 175 example, because the encounter probability changes both as function of species traits 176 (e.g. landscape use) and as a function of abundance (e.g. through density-dependent 177 foraging).

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179 Trait matching

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181 Trait matching between interacting partners has been identified for a variety of 182 organisms. Plant corolla length and pollinator proboscis length is a classic example 183 (Kritsky 1991). However, most pollinators are quite generalists and while species may 184 have specialized morphology, it does not prevent them from utilizing a diversity of 185 resources (Waser *et al.* 1996). Bird beak size and fruit size has also been shown to be 186 tightly related to dispersal success (Galetti *et al.* 2013). In fishes, predator mouth gap 187 and prey size are also strong determinants of predatory interactions (Cunha & Planas

188 1999). More complex relationships have been found for plants too, with the role of 189 specific leaf area in plant-plant interactions changing from facilitation to competition, 190 depending on resource availability (Gross *et al.* 2009). Trait-matching constraints have 191 been described for most interacting species, ranging from arbuscular mycorrhizas and 192 plants (Chagnon *et al.* 2013) to plants and herbivores (Deraison *et al.* 2015).

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Trait matching between individuals operates in addition to neutral processes to impact
pairwise interactions. Despite advances in these respective fields (e.g. null model
analysis: Vázquez, Chacoff & Cagnolo 2009; trait matching analysis: Dehling *et al.*2014; Spitz, Ridoux & Brind'Amour 2014, Crea *et al.* 2015), we still lack a common
analytical framework with which to evaluate the contribution of species traits to pairwise
interactions, and at the higher level to the structure of interaction networks.

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201 Even though neutral and trait-based null models can predict the general structure of 202 interaction networks, such models often are poor at predicting the occurrence and 203 intensity of individual interactions (Vázquez et al. 2009; Olito & Fox 2015). Such models are useful because they free us from species identities and allow us to detect 204 205 generalities, but there is no guaranty that synthetic network properties do not arise from 206 the wrong reason. Another major problem that may preclude disentangling trait-based 207 processes is that traits could influence interactions directly via trait matching, or 208 indirectly via environmental matching. Hence, even if the variance between neutral and 209 trait matching components is successfully partitioned, this would ignore the fact that 210 some of the 'neutral' variance was generated by species traits via their effect on 211 distribution and abundance (as we outlined in the previous section). Thus, the influence

of abundance versus traits can be seen as a path diagram where traits directly affect
interactions and also affect abundances, which affect interactions (Fig. 1). We propose
a framework that aims to integrate, rather than separate both processes.

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216 A significant challenge before such an analysis can be achieved is to access completely 217 sampled networks with which to validate models. Empirical network data however have 218 inherent uncertainties associated with the way in which they are sampled. Specifically, 219 sampling completeness is rarely achieved when collecting interaction networks (Chacoff 220 et al. 2012, Bartomeus 2013), and hence, some unobserved interactions may indeed 221 occur (i.e. false absence of interactions). This would be less of a problem if the 222 proportion of interactions that are sampled were constant, but this sampling efficiency 223 can vary with local environmental conditions (Laliberté & Tylianakis 2010), species 224 abundance and frequency, and of course, sampling effort. Thus, to truly understand the 225 importance of trait matching for determining species interactions, the absence of an 226 interaction in an empirical dataset cannot be used to infer true absence of that 227 interaction in nature. The nature of the data therefore impedes the direct evaluation of 228 probabilistic models (e.g. Rohr et al. 2010; Crea et al. 2015) and requires methods to 229 estimate absences (Bartomeus 2013) or the development of model fitting procedures 230 based on observed interactions only.

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Another challenge is that null models based on *a priori* rules for interactions have to be
constructed using assumptions of which traits are critical for interaction establishment.
Constructing and interpreting biologically meaningful null models that can isolate the
targeted process to be studied is not an easy task (Vázquez & Aizen 2003). As an

alternative, recent attempts to understand trait matching by statistically modeling
empirical data are promising (e.g., models incorporating imperfect detectability:
Bartomeus 2013; fourth corner analysis: Dehling *et al.* 2014; linear models: GonzálezCastro *et al.* 2015; Dirichlet-multinomial regression: Crea, Ali & Rader 2015), but such
models are still unable to integrate the relative contribution of neutral vs. trait-based
process.

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A final caveat is that most models are constrained to use mean trait values at the species level, neglecting variability among individuals of the same species. However, intraspecific trait variation, which can result from life-history stage, sexual dimorphism, or stochastic, environmental, genetic or epigenetic forces (Bolnick *et al.* 2011), has been shown to affect specific interactions such as competition, as well as overall ecological dynamics (González-Suárez & Revilla 2013).

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250 A probabilistic method for evaluating trait matching

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To overcome the limitations pointed out above, we model the probability of interaction 252 253 among pairs of individuals given their traits, based on a framework developed by Gravel 254 and colleagues (Gravel et al. 2013). The method also has the advantage to build 255 directly on the established theory of ecological network structure (Williams & Martinez, 256 2000; Eklof et al. 2013), by contrast with the above listed methods that are essentially 257 phenomenological. We propose a method to evaluate trait-matching relationships while 258 taking into account abundance of the interacting partners. The fitting procedure uses 259 information about observed interactions only, thereby overcoming problems caused by

260 under-sampling of rare interactions leading to false absences of interactions. The 261 approach implies that sampling effort is enough to adequately describe most true 262 interactions in trait-space and no false positives are recorded (i.e. recording interactions 263 as true when they do not occur). A previous sensitivity analysis however revealed it to 264 be robust to sampling effort (Gravel et al. 2013). The parameters are estimated by 265 maximum likelihood and the fitted model can be used to predict unobserved interactions 266 based on species traits and abundances. Several models, corresponding to different hypotheses, can be fit directly to raw data and accommodate complex trait matching 267 268 response functions to either qualitative or quantitative interaction data. Finally, they can 269 incorporate intraspecific trait variation, avoiding the loss of realism in species with trait 270 values that vary along developmental stages or environmental gradients. In that way we 271 provide a common toolbox to understand trait-matching rules across a variety of 272 interaction types.

273

We are interested in evaluating from empirical data a function describing the probability of an interaction between species *i* and *j* based on their respective sets of traits T_i and T_j . Building upon the model developed by (Gravel *et al.* 2013), we aim to evaluate the parameters of a model that will relate the probability with which an interaction occurs to the set of traits of the two species:

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280 $P(L_{ij} = 1 | T_i, T_j)$ (1)

281

282 Which reads as the probability of observing an interaction *L* between species *i* and *j*

given the traits T_i and T_i . The function describing this probability could take any form. 283 284 For the sake of the example here, we will consider a Gaussian shaped function (i.e. a function that assumes an unimodal relationship between T_i and T_j) to represent the trait-285 286 matching interaction (also termed interaction niche; Williams, Anandanadesan & Purves 287 2010, see below). Other functions, such as a high order polynomial or even regression trees, could be considered as well. The Gaussian function is however convenient 288 289 because it is easy to integrate and further it matches the niche model of network 290 structure (Williams et al. 2010; Eklof et al. 2013).

291

292 Equation 1 could be fit directly to empirical data by maximum likelihood. To do so, the 293 required data should contain information on presence and absence of interactions (e.g. 294 Rohr et al. 2010). The problem we are facing, however, is that records of the true 295 absence of interactions are often not available in most datasets of ecological 296 interactions, and when available, there might be considerable uncertainty in these 297 absences (i.e. false negatives due to insufficient sampling). We therefore derive a likelihood function using Bayes theorem to fit Eq. 1 indirectly, using only information 298 299 about the observed interactions. Parameters are still evaluated by maximum likelihood 300 (using simulated annealing, as described in the supplementary information), but one 301 could eventually develop the method further to compute the posterior distribution of 302 parameters.

303

The data contains information about the traits of species *i* and of species *j* only for observed interactions $L_{ij} = 1$. We consequently revise the problem and model the

probability of observing trait T_i , knowing the trait T_j and the occurrence of the interaction L_{ij} :

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$$309 \quad P(T_i|L_{ij} = 1, T_j) \quad (2)$$

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Which could be interpreted as the probability that we pick trait T_i from the trait distribution we model, given we know there is an interaction between species *i* and *j* and the trait T_j . This equation provides the likelihood for any observation of an interaction based on the traits of the two species. We now use Bayes' theorem, p(A|B)p(B) = p(B|A)p(A), to decompose Eq. 2, yielding the following distribution of the trait of one species, given the trait of the second species and the observation of the interaction:

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319
$$P(T_i|L_{ij} = 1, T_j) = \frac{P(L_{ij} = 1|T_i, T_j) P(T_i)}{P(L_{ij} = 1|T_j)}$$
 (3)

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The first term from the numerator is the trait-matching model, described at Eq. 1. It is the model for which we aim to evaluate parameters. $P(T_i)$ is the probability density function for the trait T_i . It corresponds to the probability of observing this trait in the regional pool. It could be weighted by abundance because the most abundant species are more likely to be sampled. The denominator is the marginal distribution of the traitmatching function, computed as the integral of the numerator over the whole distribution of the trait T_i :

329
$$P(L_{ij} = 1|T_j) = \int_{-\infty}^{\infty} P(L_{ij} = 1|T_i, T_j) * P(T_i) dT_i$$
 (4)

As a side product, the denominator informs us of the generality of the species *j*. This integral might be tricky to compute analytically, depending on the form of Eq. 1 and the distribution of trait T_{i} , but most software offer easy ways to compute it numerically.

335 The model given at Eq. 3 should not be confounded with the more traditional use of the 336 Bayes theorem in statistics. The resulting distribution describes the probability of 337 observing an interaction given a trait, while in statistics, the distribution describes the 338 probability of observing a set of parameters given the data. Here the parameters are 339 estimated by simulated annealing and there is only a single set of parameters yielding 340 the maximum likelihood. True confidence intervals for parameter estimates are hard to 341 evaluate for non-linear models with complex likelihood surfaces, but could nonetheless 342 be evaluated numerically. The Eq. 3 could also be implemented in a Bayesian fitting 343 procedure to obtain a posterior distribution of parameters for Eq. 1 (Eq. 3 being the 344 likelihood of the Bayes theorem), but this would be out of the scope of the current study. 345

The model could be simplified to account only for the effect of abundance (trait distributions) to reveal the importance of the trait-matching constraint. A neutral model in this framework is found when an interaction is equally probable, irrespective of the traits of the two species involved in the interaction (i.e. Eq. 1 is set as a constant). Alternatively, one could want to compare to the situation where interactions are purely determined by trait-matching constraints. In this situation, we consider the distribution of

the trait ($P(T_i)$ uniform within the range of the observed traits. The Eq. 3 remains the same for all three models and could be used to compute the likelihood for each of them. Equations for the pure neutral and trait-matching models, and a multi-trait expansion, as well as all of the R code necessary to perform this analysis are provided in the supplementary material (see Appendix S1 in Supporting Information) and as an R package found at <u>https://github.com/ibartomeus/trait_match</u>.

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359 We re-analyzed three datasets on different systems ranging from antagonistic to 360 mutualistic interactions to illustrate the overall principle of the method. First, we use data 361 from (Barnes et al. 2008) on the diet of marine fish species. The traits are the individual (log transformed) body size of the predator fish species (M_{pred}) and the individual body 362 size of preys $P(M_{prev})$. We know that larger fish typically feed on smaller ones because 363 364 they must catch and handle the prey with their mouth. The frequency distribution of prey size will indeed influence the distribution of the body mass in the diet of the predator. A 365 366 predator will tend to feed most often on the most abundant preys, which is a neutral component to the interaction probability. The predator does not select from that 367 368 distribution randomly, however, but rather it targets only a specific range (given by Eq.1) ; the niche component). Both the available prey size distribution, $P(M_{prey})$, and the 369 resulting prey size distribution, $P(M_{prey}|L, M_{pred})$, are illustrated in figure 2B for a given 370 371 predator species. The resulting prey distribution has to be somewhere between the regional prey distribution and its preferred prey size. The model therefore integrates 372 373 both neutral and trait-matching constraints.

We consider the following Gaussian function to represent the probability of aninteraction given the size of the predator and the prey:

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378
$$P(L_{ij} = 1 | M_{pred}, M_{prey}) = exp \frac{-(\alpha_0 + \alpha_1 * M_{pred} - M_{prey})^2}{2(\beta_0 + \beta_1 * M_{pred})^2}$$
(5)

379

Where α_0 , α_1 , β_0 and β_1 are fitted parameters describing the linear relationship between 380 the predator size, its optimum ($\alpha_0 + \alpha_1 M_{pred}$) and the <u>range</u> ($\beta_0 + \beta_1 M_{pred}$) of its 381 382 preference function. This formulation considers there is an optimal prey size for the 383 predator and the probability an interaction occurs reduces with any deviation from it 384 (Williams et al. 2010). The optimum also increases linearly with predator size. The same 385 reasoning could also be applied to mutualistic interactions, considering there is an 386 optimal corolla length for a pollinator of a given tongue length. One tricky issue might be 387 to gather information about the prey trait distribution. The distribution of prey traits might 388 be influenced by the interactions if there is a feedback of predators on prey abundance, 389 and in the best situation we need to tease that effect apart. Here we assume that the 390 distribution of the data provides an adequate representation of the distribution of 391 potential prey sizes because of the large number of observed interactions (> 33 000) 392 and their diversity. We thus consider a normal distribution of (log) prey size and 393 computed the average and the standard deviation.

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The predator-prey example provides a case where trait matching is a strong driver of interactions because of a strong predator-prey body size relationship (likelihood = -

21223). The parameters of the fitted model can subsequently be used for predicting
interactions among species that co-occur, but have not been observed to interact (e.g.
due to incomplete sampling) or more interestingly, for species that currently do not cooccur but may do so in the future, for example as a consequence of range shifts under
climate change (Albouy *et al.* 2014) or species invasions.

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403 Next, we use the same models on experimental data on the relationship between 404 grasshopper incisive strength and leaf dry matter content (Deraison et al. 2015). In this 405 case, both traits are species averages. We first find weak trait matching for binary data 406 (who eats whom at the species-level; likelihood = -213; Fig 3A). However, weighting the 407 interactions by consumption frequency removes bias in parameter estimates and the fit 408 of the model is considerably improved (likelihood = -5383). We thus find that strong-409 mandibled grasshoppers prefer plants with higher content of dry matter, as reported in 410 the original paper (Fig 3B).

411

412 The model could also be evaluated using traits measured at the individual level. In the 413 last example, we related pollinator tongue length with plant nectar holder depth in 414 visitation networks from Bartomeus, Vilà & Santamaría (2008). Individual pollinator 415 tongue length was inferred using the allometric relationship with body inter-tegular span 416 within each bee family (Cariveau et al. 2015), while species average flower size was 417 considered for plants. Individual trait data for pollinators allows capture of the inter-418 individual differences when evaluating parameters of trait-matching functions. In 419 addition, this model uses independent information to describe the trait abundance 420 distribution of plant species. In the past examples, abundance was inferred from the

421 network of interactions, but in this case, independent transect measures of percent plant 422 cover in the site are available (Bartomeus et al. 2008). We find that the model can be 423 interpreted as a trait-barrier, where small-tongued individuals cannot access deep 424 flowers, but long-tongued species can access both deep and shallow flowers (likelihood 425 = -705; Fig. 4). However, under such weak constraints (most pollinators can access 426 most plants), abundance is the main determinant of interaction probability. For 427 comparison, using pollinator species trait averages instead of individual values produce 428 a similar model, but with a worst likelihood (-726), indicating that there is a gain from 429 using detailed data when available.

430

431 **Discussion and conclusions**

432 Quantifying the trait-matching relationships across species may help us to understand 433 how networks are structured. For example, the nested structure of plant-pollinator 434 networks may be driven from species abundance (Vázquez et al. 2009) or from barriers 435 to certain interactions (Stang, Klinkhamer & van der Meijden 2006). In contrast, the 436 strong trait-matching observed in plant-herbivore interactions (e.g. plant defenses 437 limiting herbivory for all but a few tolerant species) can produce more modular networks 438 where interactions depart more from the null expectation based solely on abundance 439 (Thébault & Fontaine 2010). Even within plant-pollinator interactions, bird-plant 440 networks are more specialized than insect-plant networks, which is also reflected in 441 their degree of trait-matching (Maglianesi, Böhning-Gaese & Schleuning 2015). Our 442 framework is however limited to pairwise interactions and future work will have to investigate how the distribution of traits in a community constrains the emergent 443 444 network properties. Moreover, trait-matching constraints describe potential interactions,

but may not always reflect realized interactions (Poisot, Stouffer & Gravel 2015). The
future development of a bayesian approach to evaluate the distribution of parameters
will help quantifying the uncertainty of predicted interactions.

448

449 Parameterized trait-matching functions not only provide a better understanding of the 450 drivers of interactions, but they also allow prediction of novel interactions following 451 deliberate introductions (e.g. of crop species or biological control agents) or 452 unintentional invasions and range shifts (Morales-Castilla et al. 2015). Proxies of trait 453 similarity, like phylogenetic distance, have already been successfully used to predict 454 interactions of exotic species (Pearse & Altermatt 2013) and adding traits has the 455 potential to enhance this approach. Species losses and gains following local and global 456 changes are threatening most ecosystems, and it is simply impossible to measure all potential interactions in the field. Tools are consequently required to assess how the 457 458 interaction network will rewire. We know that exotic species invading a community get 459 easily integrated into the recipient network of interactions (Albrecht et al. 2014), and that 460 after species turnover in a community, the remaining species reshuffle their interactions 461 to adjust to the new composition (Kaiser-Bunbury *et al.* 2010). Our predictive ability in these situations is however still limited. 462

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464 Careful selection of the right set of traits to run the analysis is, however, a critical step.
465 We have seen that traits constraining interactions could potentially comprise all
466 morphological and physiological species characteristics, and hence, are quite specific
467 for each interaction type. A good *a priori* knowledge on the biology of the species and
468 type of interaction involved is needed to select the right trait combinations. For example,

469 we also explored whether body size drives host-parasite relationships using the 470 Tylianakis, Tscharntke & Lewis (2007) dataset, but in this case all models performed 471 poorly because the largest parasitoid is smaller than the smallest host, which allows all 472 types of body size combinations. Alternatively, spurious trait matches could be found 473 when some traits are correlated. For instance, traits like body size correlate 474 allometrically with several other morphological traits (Woodward et al. 2005) and might 475 therefore provide a wrong causal explanation of the interactions. One strong limitation 476 for some interactions, such as fungi and plants, is that the traits governing interactions 477 remain somewhat unclear (Tedersoo et al. 2008; Martínez-García et al. 2015). The 478 challenge for the future will be to determine and quantify the actual traits governing 479 these interactions, including their variability among individuals or genets.

480

Another challenge outlined in Fig. 1 and still unresolved is inferring functioning from a 481 482 network of interactions (Duffy et al. 2009; Thompson et al. 2012). Species interactions 483 are driving several ecosystem processes and functions (e.g. animal pollination, fruit 484 dispersion) as well as energy fluxes (e.g. predation, parasitism). Inferring the function 485 from traits however requires incorporating the interaction efficiency (the per capita 486 strength of a single interaction link; Vázquez et al. 2015), which in turn may be also trait-487 or abundance-mediated, and can depend on the extent of matching (e.g. pollinators with 488 short tongues may be able to visit, but inefficiently pollinate long flower corollas), or on 489 morphological, physiological or behavioural traits (e.g. large pollinators deposit more 490 pollen; Hoehn et al. 2008; Fig 1). Empirical evidence measuring interaction efficiency is 491 still scarce.

493 In conclusion, different traits can inform us about how species form networks of 494 interactions. For some interaction types, like mycorrhizal fungal interactions, traits 495 affecting co-occurrence can be the most relevant for understanding the occurrence of 496 interactions. Conversely, for other interaction types, like those between predators and 497 prey, morphological and physiological traits may be the main determinants of who 498 interacts with whom. Understanding which mechanisms are driving pairwise interactions 499 is key to predict how communities will respond to global change. Interactions regulated 500 by co-occurrence will be more likely to be affected by climate change (e.g. changing 501 phenologies and distributions), while changes in dominance following disturbance may 502 redistribute the interactions in neutral-driven networks. Non-random species extinctions 503 are also expected to affect more drastically interactions regulated by strong trait 504 matching (Larsen, Williams & Kremen 2005). There are still too many unknowns to draw 505 general conclusions about how communities are structured by traits and what 506 implications this has for ecosystem functioning, but we are now armed with appropriate 507 analytical tools to move beyond the mere description of interactions and run predictive 508 analysis of network assembly and dynamics.

509

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515

516 SUPPORTING INFORMATION

- 517 Additional supporting information may be found in the online version of this article.
- 518 Appendix S1: niche, neutral and multi-trait derivations
- 519 Appendix S2: R code as .zip
- 520 Please note: Wiley Blackwell are not responsible for the content or functionality of any
- 521 supporting information supplied by the authors. Any queries (other than missing material) should
- 522 be directed to the corresponding author for the article.
- 523

524 **References:**

- Albouy, C., Velez, L., Coll, M., Colloca, F., Loc'h, F., Mouillot, D. & Gravel, D. (2014)
- 526 From projected species distribution to food-web structure under climate change.
 527 *Global change biology*, **20**, 730–741.
- 528 Albrecht, M., Padrón, B., Bartomeus, I. & Traveset, A. (2014) Consequences of plant
- 529 invasions on compartmentalization and species' roles in plant--pollinator networks.
- 530 Proceedings of the Royal Society of London B: Biological Sciences, **281**,
- 531 20140773.
- 532 Araújo, M.B., Rozenfeld, A., Rahbek, C. & Marquet, P.A. (2011) Using species co-533 occurrence networks to assess the impacts of climate change. *Ecography*, **34**,
- 534 **897–908**.
- 535 Barnes, C., Bethea, D.M., Brodeur, R.D., Spitz, J., Ridoux, V., Pusineri, C., Chase,
- 536 B.C., Hunsicker, M.E., Juanes, F., Kellermann, A. & Others. (2008) Predator and
- 537 prey body sizes in marine food webs: Ecological Archives E089-051. *Ecology*, **89**,

538 881–881.

Bartomeus, I. (2013) Understanding linkage rules in plant-pollinator networks by using
hierarchical models that incorporate pollinator detectability and plant traits. *PloS one*, **8**, e69200.

- Bartomeus, I., Park, M.G., Gibbs, J., Danforth, B.N., Lakso, A.N. &Winfree, R. (2013)
 Biodiversity ensures plant-pollinator phenological synchrony against climate
 change. *Ecology letters*, **16**, 1331–1338.
- 545 Bartomeus, I., Vilà, M. &Santamaría, L. (2008) Contrasting effects of invasive plants in 546 plant–pollinator networks. *Oecologia*, **155**, 761–770.
- 547 Bascompte, J. & Jordano, P. (2007) Plant-Animal Mutualistic Networks: The Architecture 548 of Biodiversity. *Annual review of ecology, evolution, and systematics*, **38**, 567–593.
- 549 Bolnick, D.I., Amarasekare, P., Araújo, M.S., Bürger, R., Levine, J.M., Novak, M.,
- 550 Rudolf, V.H.W., Schreiber, S.J., Urban, M.C. & Vasseur, D.A. (2011) Why
- 551 intraspecific trait variation matters in community ecology. *Trends in ecology*
- 552 &evolution, **26**, 183–192.
- 553 Cadotte, M.W., Carscadden, K. & Mirotchnick, N. (2011) Beyond species: functional
- diversity and the maintenance of ecological processes and services. *The Journal of applied ecology*, **48**, 1079–1087.
- 556 Canard, E.F., Mouquet, N., Mouillot, D., Stanko, M., Miklisova, D. & Gravel, D. (2014)
- 557 Empirical evaluation of neutral interactions in host-parasite networks. *The American*

558 *naturalist*, **183**, 468–479.

559	Cardinale, B.J.,	Duffy, J.E., G	Sonzalez, A.,	Hooper, D.U.,	Perrings, 0	C., Venail, P.,
-----	------------------	----------------	---------------	---------------	-------------	-----------------

- 560 Narwani, A., Mace, G.M., Tilman, D., Wardle, D.A., Kinzig, A.P., Daily, G.C.,
- 561 Loreau, M., Grace, J.B., Larigauderie, A., Srivastava, D.S. & Naeem, S. (2012)
- 562 Biodiversity loss and its impact on humanity. *Nature*, **486**, 59–67.
- 563 Cariveau, D.P., Nayak, G., Bartomeus, I., Zientek, J., Ascher, J. & Winfree, R. (2015)

564 The allometry of bee tongue length an its uses in ecology and evolution. *PLoS One.*565 In review.

566 Carvalheiro, L.G., Barbosa, E.R.M. & Memmott, J. (2008) Pollinator networks, alien

567 species and the conservation of rare plants: Triniaglauca as a case study. *The*568 *Journal of applied ecology*, **45**, 1419–1427.

569 Castellanos, M. C., Wilson, P. & Thomson, J. D. (2003) Pollen transfer by

570 hummingbirds and bumblebees, and the divergence of pollination modes in

571 Penstemon. *Evolution* **57**, 2742-2752.

572 Chacoff, N.P., Vázquez, D.P., Lomáscolo, S.B., Stevani, E.L., Dorado, J. & Padrón, B.

573 (2012) Evaluating sampling completeness in a desert plant–pollinator network. *The*

574	Journal of animal ecology,	81 ,	190–200.
-----	----------------------------	-------------	----------

- 575 Chagnon, P.-L., Bradley, R.L., Maherali, H. & Klironomos, J.N. (2013) A trait-based
- 576 framework to understand life history of mycorrhizal fungi. *Trends in plant science*,
- **18**, 484–491.

578	Crea, C., Ali, R. A. & Rader, R. (2015) A new model for ecological networks using
579	species-level traits. Methods in Ecology and Evolution. In press. doi: 10.1111/2041-
580	210X.12471

581 Cunha, I. & Planas, M. (1999) Optimal prey size for early turbot larvae (Scophthalmus 582 maximus L.) based on mouth and ingested prey size. *Aquaculture*, **175**, 103–110.

583 Dehling, D.M., Töpfer, T., Schaefer, H.M., Jordano, P., Böhning-Gaese, K. &

584 Schleuning, M. (2014) Functional relationships beyond species richness patterns:

585 trait matching in plant–bird mutualisms across scales. *Global ecology and*

biogeography, **23**, 1085–1093.

Deraison, H., Badenhausser, I., Börger, L. & Gross, N. (2015) Herbivore effect traits and
 their impact on plant community biomass: an experimental test using grasshoppers.
 Functional ecology, 29, 650–661.

590 Duffy, J.E., Cardinale, B.J., France, K.E., McIntyre, P.B., Thébault, E. & Loreau, M.

(2007) The functional role of biodiversity in ecosystems: incorporating trophic
complexity. *Ecology letters*, **10**, 522–538.

593 Duffy, J.E., Srivastava, D.S., McLaren, J., Sankaran, M., Solan, M., Griffin, J.,

594 Emmerson, M. & Jones, K.E. (2009) Forecasting decline in ecosystem services

595 under realistic scenarios of extinction. *Biodiversity, ecosystem functioning and*

- 596 human wellbeing: an ecological and economic perspective. Oxford University
- 597 Press, Oxford, British, 60–77.

598 Díaz, S. & Cabido, M. (2001) Vive la difference: plant functional diversity mattersto

600

ecosystem processes. *Trends in ecology & evolution*, **16**, 646–655.

D'Amen, M., Dubuis, A., Fernandes, R.F., Pottier, J., Pellissier, L. & Guisan, A. (2015)

601	Using species richness and functional traits predictions to constrain assemblage
602	predictions from stacked species distribution models. Journal of biogeography, 42,
603	1255–1266.
604	Eklöf, A., Jacob, U., Kopp, J., Bosch, J., Castro-Urgal, R., Chacoff, N.P., Dalsgaard, B.,
605	de Sassi, C., Galetti, M., Guimarães, P.R., Lomáscolo, S.B., Martín González,
606	A.M., Pizo, M.A., Rader, R., Rodrigo, A., Tylianakis, J.M., Vázquez, D.P. & Allesina,
607	S. (2013) The dimensionality of ecological networks. <i>Ecology letters</i> , 16 , 577–583.
608	Encinas-Viso, F., Revilla, T.A. & Etienne, R.S. (2012) Phenology drives mutualistic
609	network structure and diversity. Ecology letters, 15, 198–208.
610	Fenster, C.B., Reynolds, R.J., Williams, C.W., Makowsky, R. & Dudash, M.R. (2015)
611	Quantifying hummingbird preference for floral trait combinations: The role of
612	selection on trait interactions in the evolution of pollination syndromes. Evolution, 69,
613	1113–1127.
614	Gagic, V., Bartomeus, I., Jonsson, T., Taylor, A., Winqvist, C., Fischer, C., Slade, E.M.,
615	Steffan-Dewenter, I., Emmerson, M., Potts, S.G., Tscharntke, T., Weisser, W. &
616	Bommarco, R. (2015) Functional identity and diversity of animals predict ecosystem
617	functioning better than species-based indices. Proceedings of the Royal Society of
618	London B: Biological Sciences, 282, 20142620.
619	Galetti, M., Guevara, R., Côrtes, M.C., Fadini, R., Von Matter, S., Leite, A.B., Labecca,

620	F., Ribeiro, T., Carvalho, C.S., Collevatti, R.G., Pires, M.M., Guimarães, P.R., Jr,
621	Brancalion, P.H., Ribeiro, M.C. & Jordano, P. (2013) Functional extinction of birds
622	drives rapid evolutionary changes in seed size. Science, 340 , 1086–1090.
623	González-Castro, A., Yang, S., Nogales, M. & Carlo, T.A. (2015) Relative importance of
624	phenotypic trait matching and species' abundances in determining plant-avian seed
625	dispersal interactions in a small insular community. <i>AoB plants</i> , 7 .
626	González-Suárez, M. & Revilla, E. (2013) Variability in life-history and ecological traits is
627	a buffer against extinction in mammals. <i>Ecology letters</i> , 16 , 242–251.
628	Gravel, D., Poisot, T., Albouy, C., Velez, L. & Mouillot, D. (2013) Inferring food web
629	structure from predator-prey body size relationships. Methods in ecology and
630	evolution, 4 , 1083–1090.
631	Gross, N., Kunstler, G., Liancourt, P., De Bello, F., Suding, K.N. & Lavorel, S. (2009)
632	Linking individual response to biotic interactions with community structure: a trait-
633	based framework. Functional ecology, 23, 1167–1178.
634	Herrera, C.M. (2000) Flower-to-seedling consequences of different pollination regimes
635	in an insect-pollinated shrub. <i>Ecology</i> , 81 , 15–29.
636	Hoehn, P., Tscharntke, T., Tylianakis, J.M. & Steffan-Dewenter, I. (2008) Functional
637	group diversity of bee pollinators increases crop yield. Proceedings of the Royal
638	Society of London B: Biological Sciences, 275, 2283–2291.
639	Jacquemyn, H., Merckx, V., Brys, R., Tyteca, D., Cammue, B.P.A., Honnay, O. &

640	Lievens, B. (2011) Analysis of network architecture reveals phylogenetic
641	constraints on mycorrhizal specificity in the genus Orchis (Orchidaceae). The New
642	phytologist, 192 , 518–528.
643	Kaiser-Bunbury, C.N., Muff, S., Memmott, J., Müller, C.B. & Caflisch, A. (2010) The
644	robustness of pollination networks to the loss of species and interactions: a
645	quantitative approach incorporating pollinator behaviour. Ecology letters, 13, 442-
646	452.

Krishna, A., Guimaraes, P.R., Jr, Jordano, P. & Bascompte, J. (2008) A neutral-niche

theory of nestedness in mutualistic networks. *Oikos*, **117**, 1609–1618.

- Kritsky, G. (1991) Darwin's Madagascan Hawk Moth Prediction. *American Entomologist*, **37**, 206–210.
- Laliberté, E. & Tylianakis, J.M. (2010) Deforestation homogenizes tropical parasitoid-host
 networks. *Ecology*, **91**, 1740-1747.
- Larsen, T.H., Williams, N.M. & Kremen, C. (2005) Extinction order and altered

community structure rapidly disrupt ecosystem functioning. *Ecology letters*, **8**, 538–
547.

- Laughlin, D.C., Joshi, C., van Bodegom, P.M., Bastow, Z.A. & Fulé, P.Z. (2012) A
- 657 predictive model of community assembly that incorporates intraspecific trait

658 variation. *Ecology letters*, **15**, 1291–1299.

- Lima-Mendez, G., Faust, K., Henry, N., Decelle, J. & Colin, S. (2015) Determinants of
- 660 community structure in the global plankton interactome. *Science*.

661	Loreau, M. (1998) Biodiversity and ecosystem functioning: a mechanistic model.
662	Proceedings of the National Academy of Sciences of the United States of America,
663	95 , 5632–5636.

Loreau, M. (2010) Linking biodiversity and ecosystems: towards a unifying ecological
theory. *Philosophical transactions of the Royal Society of London. Series B, Biological sciences*, **365**, 49–60.

667 Maglianesi, M.A., Böhning-Gaese, K. & Schleuning, M. (2015) Different foraging

668 preferences of hummingbirds on artificial and natural flowers reveal mechanisms

structuring plant--pollinator interactions. *The Journal of animal ecology*, **84**, 655–
664.

Martínez-García, L.B., Richardson, S.J., Tylianakis, J.M., Peltzer, D.A. & Dickie, I.A.

672 (2015) Host identity is a dominant driver of mycorrhizal fungal community

673 composition during ecosystem development. *The New phytologist*, **205**, 1565–

674 1576.

Morales-Castilla, I., Matias, M.G., Gravel, D. & Araújo, M.B. (2015) Inferring biotic
interactions from proxies. *Trends in ecology & evolution*, **30**, 347–356.

Olesen, J.M., Bascompte, J., Dupont, Y.L., Elberling, H., Rasmussen, C. & Jordano, P.

678 (2011) Missing and forbidden links in mutualistic networks. *Proceedings of the*

679 Royal Society of London B: Biological Sciences, **278**, 725–732.

680 Olito, C. & Fox, J.W. (2015) Species traits and abundances predict metrics of plant-

681 pollinator network structure, but not pairwise interactions. *Oikos*, **124**, 428–436.

- Pearse, I.S. & Altermatt, F. (2013) Predicting novel trophic interactions in a non-native
 world. *Ecology letters*, **16**, 1088–1094.
- Peralta, G., Frost, C.M., Rand, T.A., Didham, R.K. & Tylianakis, J.M. (2014)
- 685 Complementarity and redundancy of interactions enhance attack rates and spatial
- stability in host-parasitoid food webs. *Ecology*, **95**, 1888–1896.
- Poisot, T., Canard, E., Mouillot, D., Mouquet, N., Gravel, D. & Jordan, F. (2012) The
 dissimilarity of species interaction networks. *Ecology letters*, **15**, 1353–1361.
- 689 Poisot, T., Mouquet, N. & Gravel, D. (2013) Trophic complementarity drives the
- biodiversity–ecosystem functioning relationship in food webs. *Ecology letters*, **16**,
 853–861.
- Poisot, T., Stouffer, D.B. & Gravel, D. (2015) Beyond species: why ecological interaction
 networks vary through space and time. *Oikos*, **124**, 243–251.
- Rader, R., Reilly, J., Bartomeus, I. & Winfree, R. (2013) Native bees buffer the negative
- 695 impact of climate warming on honey bee pollination of watermelon crops. *Global*696 *change biology*, **19**, 3103–3110.
- Rohr, R.P., Scherer, H., Kehrli, P., Mazza, C. & Bersier, L.-F. (2010) Modeling food
- webs: exploring unexplained structure using latent traits. *The American naturalist*, **176**, 170–177.
- Sargent, R.D. & Ackerly, D.D. (2008) Plant-pollinator interactions and the assembly of
- plant communities. *Trends in Ecology and Evolution*, **23**:123-130

702	Shipley, B., Vile, D. & Garnier, E. (2006) From plant traits to plant communities: a
703	statistical mechanistic approach to biodiversity. Science, 314 , 812–814.
704	Spitz, J., Ridoux, V. & Brind'Amour, A. (2014) Let's go beyond taxonomy in diet
705	description: testing a trait-based approach to preypredator relationships. The
706	Journal of animal ecology, 83 , 1137–1148.
707	Stang, M., Klinkhamer, P.G.L. & van der Meijden, E. (2006) Asymmetric specialization
708	and extinction risk in plant-flower visitor webs: a matter of morphology or
709	abundance? <i>Oecologia</i> , 151 , 442–453.
710	Tedersoo, L., Jairus, T., Horton, B.M., Abarenkov, K., Suvi, T., Saar, I. & Kõljalg, U.
711	(2008) Strong host preference of ectomycorrhizal fungi in a Tasmanian wet
712	sclerophyll forest as revealed by DNA barcoding and taxon-specific primers. The
713	New phytologist, 180 , 479–490.
714	Thebault, E. & Loreau, M. (2003) Food-web constraints on biodiversity-ecosystem
715	functioning relationships. Proceedings of the National Academy of Sciences, 100,
716	14949–14954.
717	Thompson, R.M., Brose, U., Dunne, J.A., Hall, R.O., Jr, Hladyz, S., Kitching, R.L.,
718	Martinez, N.D., Rantala, H., Romanuk, T.N., Stouffer, D.B. & Tylianakis, J.M.
719	(2012) Food webs: reconciling the structure and function of biodiversity. Trends in

- *ecology* & *evolution*, **27**, 689–697.
- 721 Thébault, E. & Fontaine, C. (2010) Stability of ecological communities and the
- architecture of mutualistic and trophic networks. *Science*, **329**, 853–856.

723	Tylianakis, J.M., Didham, R.K., Bascompte, J. & Wardle, D.A. (2008) Global change
724	and species interactions in terrestrial ecosystems. Ecology letters, 11 , 1351–1363.
725	Tylianakis, J.M., Laliberté, E., Nielsen, A. & Bascompte, J. (2010) Conservation of
726	species interaction networks. <i>Biological conservation</i> , 143 , 2270–2279.
727	Tylianakis, J.M., Tscharntke, T. & Lewis, O.T. (2007) Habitat modification alters the
728	structure of tropical host-parasitoid food webs. Nature, 445, 202-205.
729	Vázquez, D.P. & Aizen, M.A. (2003) Null Model Analyses of Specialization in Plant-
730	Pollinator Interactions. Ecology, 84, 2493–2501.
731	Vázquez, D.P., Chacoff, N.P. & Cagnolo, L. (2009) Evaluating multiple determinants of
732	the structure of plant-animal mutualistic networks. <i>Ecology</i> , 90 , 2039–2046.
733	Vázquez, D.P., Melián, C.J., Williams, N.M., Blüthgen, N., Krasnov, B.R. & Poulin, R.
734	(2007) Species abundance and asymmetric interaction strength in ecological
735	networks. <i>Oikos</i> , 116 , 1120–1127.
736	Vázquez, D.P., Morris, W.F. & Jordano, P. (2005) Interaction frequency as a surrogate
737	for the total effect of animal mutualists on plants. <i>Ecology letters</i> , 8 , 1088–1094.
738	Vázquez, D.P., Ramos-Jiliberto, R., Urbani, P. & Valdovinos, F.S. (2015) A conceptual
739	framework for studying the strength of plant-animal mutualistic interactions.
740	<i>Ecology letters</i> , 18 , 385–400.
741	Waser, N.M., Chittka, L., Price, M.V., Williams, N.M. & Ollerton, J. (1996) Generalization
742	in Pollination Systems, and Why it Matters. <i>Ecology</i> , 77 , 1043–1060.

743	White, E.P., Ernest, S.K.M., Kerkhoff, A.J. & Enquist, B.J. (2007) Relationships between
744	body size and abundance in ecology. Trends in ecology & evolution, 22, 323–330.
745	Williams, R.J., Anandanadesan, A. & Purves, D. (2010) The probabilistic niche model
746	reveals the niche structure and role of body size in a complex food web. PloS one,
747	5 , e12092.

Williams, R.J. & Martinez, N.D. (2000) Simple rules yield complex food webs. *Nature*,
404, 180–183.

Winfree, R., W Fox, J., Williams, N.M., Reilly, J.R. & Cariveau, D.P. (2015) Abundance
of common species, not species richness, drives delivery of a real-world ecosystem

752 service. *Ecology letters*, **18**, 626–635.

753 Woodward, G., Ebenman, B., Emmerson, M., Montoya, J.M., Olesen, J.M., Valido, A. &

Warren, P.H. (2005) Body size in ecological networks. *Trends in ecology & evolution*, **20**, 402–409.

756 Wright, I.J., Reich, P.B., Westoby, M., Ackerly, D.D., Baruch, Z., Bongers, F., Cavender-

Bares, J., Chapin, T., Cornelissen, J.H.C., Diemer, M., Flexas, J., Garnier, E.,

Groom, P.K., Gulias, J., Hikosaka, K., Lamont, B.B., Lee, T., Lee, W., Lusk, C.,

759 Midgley, J.J., Navas, M.-L., Niinemets, U., Oleksyn, J., Osada, N., Poorter, H.,

Poot, P., Prior, L., Pyankov, V.I., Roumet, C., Thomas, S.C., Tjoelker, M.G.,

761 Veneklaas, E.J. & Villar, R. (2004) The worldwide leaf economics spectrum.

762 *Nature*, **428**, 821–827.

763 Zobel, M. & Öpik, M. (2014) Plant and arbuscular mycorrhizal fungal (AMF)

- communities--which drives which? *Journal of vegetation science*, **25**, 1133–1140.
- 765 Data Accessibility: All data and R code to perform analysis can be found in the
- 766 appendix and at https://github.com/ibartomeus/trait_match (<u>10.5281/zenodo.46382</u>)

768 Figures:

769 Figure 1



770

771 Figure 1. Species traits may influence the structure of interaction networks in three different ways. 1) Trait-based environmental filtering may determine species 772 773 abundances in space and time, which will affect probability of encounter. 2) Given 774 species co-occurrence, trait matching according to species interaction preferences will 775 shape interaction probability. 3) Species traits might also influence the per capita 776 efficiency and impact of an interaction, and thereby influence network functioning. In 777 addition, 4) emergent properties inherent to the structure of the network will influence network functioning and feedback on community dynamics. 778





781 Figure 2. Illustration of the quantitative framework to evaluate a trait-matching 782 probabilistic function. A) Conceptual representation of a trait-matching constraint. Interactions (in black) are feasible only when both species have traits that are 783 784 compatible. However, we often do not have reliable information on the species that are 785 present, but are not observed to interact (white dots). Dotted lines indicate the trait 786 ranges of compatibility between the species B) Representation of the density function 787 for available body size in the (Barnes et al. 2008) dataset (white bars), the trait-788 matching function (black line) and the observed distribution of prey size for the predator 789 Nototheniops larseni (black bars). C) Representation of the observed interactions (black 790 dots) and the prediction for the maximum likelihood estimate of the trait-matching 791 function (from low probability in red to high probability in white).



Figure 3. Representation of the fitted interaction probability for grasshopper and plant interactions unweighted (A) and weighted (B) by frequency of interaction (from low probability in red to high probability in white). The probability of interaction between a grasshopper and a plant follows a positive relationship between incisive strength and plant leaf dry matter content. Note that the overlapping data in B has been jittered to appreciate the different frequencies of particular interactions. The likelihood for (A) is similar to the neutral model, while much better in (B), indicating that the frequency of interactions must be taken into account to better reveal the trait-matching constraint.

808 Figure 4



809

810 Figure 4. Representation of the fitted interaction probability plant and pollinators

811 weighted by the frequency of interactions (from low probability in red to high probability

in white). Only a few interactions among small tongue sized bees and long corolla depth

813 flowers are realized (red area), while the rest of interactions are explained mainly by

814 abundance.