A conformational analysis of Walker motif A [GXXXXGKT (S)] in nucleotide-binding and other proteins

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The sequence GXXXXGKT/S, popularly known as Walker motif A, is widely believed to be the site for binding nucleotides in many proteins. Examination of the crystal structures in the Protein Data Bank showed that about half of the examples having these sequences do not bind or use nucleotides. Data analyses showed 92 different Walker sequences of the variable quartet (XXXX). Ramachandran angles in this segment revealed conformational similarity in the group of 45 proteins, known to bind or utilize nucleotides. The conformations of this segment in other proteins differ widely and it is not known whether they play any role in their functions. A flip of a peptide unit at different locations, with little change in the backbone conformation was noted in nine pairs of these proteins having same Walker sequence. An examination of the immediate neighborhood of the Walker sequence indicates that this region is preceded by a β -strand and followed by an α -helix, resulting in the motif β -W- α , an invariant feature amongst nucleotidebinding proteins.

Keywords: peptide flip/Ramachandran angles/β-turn/Walker motif

Introduction

The motif GXXXXGKT (X, any residue) as a common nucleotide binding fold in the α - and β -subunits of F₁-ATPase, myosin and other ATP-requiring enzymes was first recognized in 1982 by Walker and colleagues (Walker *et al.*, 1982). Since then, this sequence has been found in many proteins that bind nucleotides and thereby gained predictive value for nucleotide binding site in proteins. Crystal structure data of such proteins (Berchtold *et al.*, 1993; Abrahams *et al.*, 1994; Chattopadhyay *et al.*, 2000) indicated that this motif is present in the shape of a loop around nucleotides and utilizes its highly conserved residues of lysine and threonine to bind to their phosphateoxygen atoms. This consensus sequence of GXXXXGKT (S), with serine substituting threonine in some cases, is more popularly known as Walker loop or P-loop (phosphate binding loop).

In view of growing interest in the proteins containing a segment with Walker sequence, the Brookhaven Protein Data Bank (Berman *et al.*, 2000) was searched and 649 polypeptide chains were found to have such a sequence. Many of these proteins do not bind or use nucleotides in their reactions. Therefore, it appeared that the sequence of the variant quartet and the specific loop structure might have a role in

nucleotide binding. To fill the lacunae of information, conformations of the backbone of the peptide fragments of GXXXXGKT (S) were examined using Ramachandran angles. The data analysis in this paper indicates that different foldings are possible for the Walker sequences and only in the nucleotide-binding proteins they have a distinctive loop structure.

Materials and methods

The Ramachandran angles (ϕ, ψ) (Ramachandran *et al.*, 1963; Ramachandran and Sasisekharan, 1968) were computed from the coordinates of atoms available in the Brookhaven Protein Data Bank (Berman *et al.*, 2000). The segment structure similarity was obtained by evaluating the root mean square (r.m.s.) values of the Ramachandran angles. The package of RASMOL (Sayle and Milner-White, 1995) was used to draw the figures.

Results of data analysis

Proteins containing Walker sequences

Search for the sequence GXXXXGKT (S) in the Protein Data Bank (April 2001 release) revealed 649 entries having this sequence, occurring in 395 protein structures with a resolution of 4 Å or better. Out of the 20^4 combinations of sequence possible for the variable region XXXX, only 92 were found to occur, of which 18 had only one entry. The present analysis is limited to these data

The Ramachandran angles of Walker sequence

Groups having more than one entry were examined from the structural viewpoint. The mean and r.m.s. values of the Ramachandran angles ϕ and ψ were computed at the eight residues of the segment. Should the same sequence give the same structure, as is widely believed, the r.m.s. values for a group would be small. Using a liberal upper limit of 40°, dissimilar structures were found to be present in 10 of these groups, as revealed by the high r.m.s. values for some of the Ramachandran angles. Using similarity of the Ramachandran angles as the criterion, these were divided into further sub-groups. The various sequences and location of the segment in the protein of the group thus obtained are given in Table I, along with the PDB code, chain identifier, resolution of the structure and r.m.s. for those groupings with more than one entry (the protein names are not included in Table I owing to the large number of examples; however, they are included in Table II, which gives the selected set). The sub-groups with same sequence are indicated by suffixes A, B and C, to the group number. It can be seen that the r.m.s. values are now reasonably small. Some sequences assume more than one conformation: two for six sequences (005 - GAGALGKT, 012 -GLRSDGKT, 016 - GLPAIGKT, 030 - GATGTGKT, 058 -GTAFEGKS and 077 - GLYRTGKS); three for three sequences (006 - GHVDHGKT, 033 - GPTGVGKT and

Table I. Proteins containing the consensus sequence of GXXXXGKT(S): the location of the segment in the chain, PDB code and resolution of the crystal structure are given

No.	Sequence Segment location	PDB code, (resolution in Å)) chain identifier	Residue r.m.s. (φ,ψ)	Residue r.m.s. (φ,ψ) T (1,6) G (6,2) K (1,2) T (2,1)	
001	GLSGTGKT 248 255	1AQ2 (1.9)	1AYL (1.8)	G (1,1) L (1,1) S (6,1) G (2,1)		
002	GDRQTGKT 169 176	1BMF (2.8) (A,B,C) 1E1Q (2.6) (A,B,C) 1E79 (2.4) (A,B,C) 1MAB (2.8) (A)	1COW (3.1) (A,B,C) 1E1R (2.5) (A,B,C) 1EFR (3.1) (A,B,C) 1NBM (3.0) (A,B,C)	G (7,8) D (7,6) R (6,5) Q (7,11)	T (9,10) G (13,10) K (8,8) T (10,6)	
003	GGAGVGKT 156 163	1BMF (2.8) (D,F) 1E1Q (2.6) (D,E,F) 1E79 (2.4) (D,E,F) 1NBM (3.0) (D,E,F)	1COW (3.1) (D,F) 1E1R (2.5) (D,F) 1EFR (3.1) (D,F)	G (23,11) G (7,19) A (19,18) G (13,17)	V (17,18) G (23,10) K (6,4) T (5,7)	
03A	GGAGVGKT 156 163	1BMF (2.8) (E) 1EFR (3.1) (E)	1COW (3.1) (E)	G (1,2) G (2,0) A (1,1) G (2,2)	V (2,2) G (2,5) K (5,0) T (2,10)	
03B 03C	GGAGVGKT 156 163 GGAGVGKT	1E1R (2.5) (E) 1MAB (2.8) (B)				
004	156 163 GAHALGKT 173 180	1A2F (2.1) 1AA4 (2.1) 1AC8 (2.1) 1AEB (2.1) 1AEF (2.1) 1AEF (2.1) 1AEK (2.1) 1AEK (2.1) 1AEV (2.1) 1AEV (2.1) 1AEV (2.1) 1BEV (2.2) 1BEP (2.2) 1BES (2.0) 1BVA (1.8) (A) 1CCB (2.1) 1CCI (2.4) 1CCK (2.1) 1CCP (2.2) 1CMU (2.1) 1CPE (2.2) 1CMU (2.1) 1CPE (2.2) 1DCC (2.2) 1CCP (2.2) 1	1A2G (2.1) 1AC4 (2.1) 1AEB (2.1) 1AEB (2.1) 1AEG (2.1) 1AEG (2.1) 1AEM (2.1) 1AEM (2.1) 1AEM (2.1) 1AEU (2.1) 1AEU (2.1) 1BEJ (2.4) 1BEM (2.2) 1BEQ (2.1) 1BJ9 (2.2) 1CCA (1.8) 1CCC (2.0) 1CCJ (2.1) 1CCL (2.0) 1CMT (2.1) 1CPD (2.2) 1CPF (2.2) 1CPF (2.2) 1CYF (2.3) 1DJ1 (1.9) (A) 1RYC (1.8) 2CEP (2.2) 2PCB (2.8) (A,C) 3CCP (2.2) 5CCP (2.2)	G (5,8) A (5,4) H (5,7) A (8,5)	L (7,4) G (6,5) K (5,5) T (4,4)	
005	GAGALGKT 173 180	1CCE (2.3)	1CCG (2.1)	G (5,9) A (5,10) G (11,3) A (1,3)	L (2,1) G (2,1) K (4,2) T (1,3)	
005A	GAGALGKT 173 180	1DS4 (2.0) (A) 1DSG (2.5) (A) 1DSP (2.0) (A)	1DSE (2.0) (A) 1DSO (2.0) (A)	G (9,14) A (12,8) G (3,7) A (6,10)	L (13,2) G (4,6) K (3,6) T (5,3)	
06	GHVDHGKT 18 25	1B23 (2.6) (P) 1D8T (2.3) (A,B) 1EFC (2.0) (A,B) 1EXM (1.7) A 1G7T (2.0) (A)	1D2E (1.9) (A–D) 1DG1 (2.5) (G,H) 1EFT (2.5) 1G7S (2.0) (A) 1TUI (2.7) (A,B,C)	G (9,4) H (5,6) V (6,5) D (6,7)	H (8,7) G (8,9) K (7,9) T (6,6)	

No.	Sequence	PDB code, (resolution in Å)	chain identifier	Residue	Residue	
	Segment location	TDD code, (resolution III A)		r.m.s. (φ,ψ)	r.m.s. (¢,ψ	
06A	GHVDHGKT 18 25	1AIP (3.0) (A,B,E,F)	1EFU (2.5) (A,C)	G (18,5) H (5,4) V (9,4) D (11,7)	H (2,4) G (5,14) K (10,5) T (3,5)	
)6B	GHVDHGKT 18 25	1ETU (2.9)				
07	GYLVNGKT 1264 271	10MH (2.5) (A) HMY (2.5) 2HMY (2.6) (B) 4MHT (2.7) (A) 6MHT (2.0) (A) 8MHT (2.7) (A)	1FJX (2.2) (A) 1MHT (2.8) (A) 3MHT (2.7) (A) 5MHT (2.7) (A) 7MHT (2.8) (A) 9MHT (2.3) (A)	G (7,4) Y (5,4) L (3,7) V (8,6)	N (9,6) G (9,7) K (10,7) T (9,8)	
)8	GLDAAGKT 24 31	1EOS (2.2) (A) 1HUR (2.0) (A,B) 1RRG (2.4) A,B	1HFV (2.8) (A,B) 1RRF (3.0)	G (4,5) L (9,6) D (7,8) A (3,7)	A (7,9) G (13,9) K (9,9) T (7,6)	
09	GPHGMGKT 56 63	1E2H (1.9) (A,B) 1E2J (2.5) (A,B) 1E2L (2.4) (A,B) 1K13 (2.3) (A,B) 1K16 (2.3) (A,B) 1K18 (2.2) (A,B) 1K1N (2.0) (A,B) 1VTK (2.7) 2VTK (2.8)	1E2I (1.9) (A,B) 1E2K (1.7) (A,B) 1KI2 (2.2) (A,B) 1KI4 (2.3) (A,B) 1KI7 (2.2) (A,B) 1KIM (2.1) (A,B) 1QHI (1.9) (A,B) 2KI5 (1.9) (A,B) 2VITK (2.0)	G (14,4) P (5,6) H (5,5) G (7,12)	M (9,14) G (14,10) K (8,9) T (7,8)	
10	GVRSDGKT 487 494	2VTK (2.8) 1MHY (2.) (D)	3VTK (3.0) 1MHZ (2.7) (D)	G (5,9) V (11,2) R (8,3) S (9,8)	D (2,3) G (9,0) K (14,10) T (1,2)	
11	GESGAGKT 179 186	1B7T (2.5) (A) 1BR2 (2.9) (A–F) 1D0X (2.0) (A) 1D0Z (2.0) (A) 1D1B (2.0) (A) 1DFK (4.2) (A) 1FMV (2.1) (A) 1G8X (2.8) (A,B) 1MMA (2.1) 1MMG (1.9) 1MND (2.6) 1VOM (1.9)	1BR1 (3.5) (A,C,E,G) 1BR4 (3.6) (A,C,E,G) 1D0Y (2.0) (A) 1D1A (2.0) (A) 1D1C (2.3) (A) 1DFL (4.2) (A,B) 1FMW (2.1) (A) 1LVK (1.9) 1MMD (2.0) 1MMN (2.1) 1MNE (2.7) 2MYS (2.8) (A)	G (26,15) E (7,6) S (10,28) G (28,12)	A (10,17) G (20,18) K (18,6) T (6,12)	
12	GLRSDGKT 487 494	1FYZ (2.1) (A,B) 1FZ1 (1.9) (A,B) 1FZ3 (2.0) (A,B) 1FZ5 (2.4) (A,B) 1MMO (2.2) (E)	1FZ0 (2.0) (A,B) 1FZ2 (2.1) (A,B) 1FZ4 (2.3) (A,B) 1FZ7 (1.9) (A,B) 1MTY (1.7) (D,E)	G (2,5) L (4,4) R (4,4) S (8,10)	D (5,4) G (5,5) K (9,8) T (3,2)	
12A	GLRSDGKT 487 494	1MMO (2.2) (D)				
3	GLSGSGKT 248 255	10EN (1.9)				
4	GTAFPGKT 212 219	1QPA (1.8) (A,B)		G (3,3) T (3,1) A (2,2) F (1,1)	P (2,4) G (4,0) K (2,3) T (1,5)	
5	GKVTGGKT 102 109	1STE (2.0)		. (.,.)	1 (1,5)	
16	GLPAIGKT 499 506	1BGX (2.3) (T) 1TAQ (2.4)	1CMW (2.6) (A)	G (4,12) L (16,5) P (2,20) A (12,11)	I (8,13) G (19,13) K (22,15) T (2,10)	
16A	GLPAIGKT 499 506	1QSS (2.3) (A) 1QTM (2.3) (A) 3KTQ (2.3) (A)	1QSY (2.3) (A) 2KTQ (2.3) (A) 4KTQ (2.5) (A)	G (13,10) L (11,2) P (3,8) A (5,20)	I (15,8) G (4,4) K (2,4) T (5,7)	

No.	Sequence Segment location	PDB code, (resolution in Å) chain identifier	Residue r.m.s. (φ,ψ)	Residue r.m.s. (φ,ψ)
017	GSQAGGKT 47 54	1WGT (1.9) (A,B)		G (4,1) S (1,1) Q (2,7) A (3,0)	G (9,4) G (5,7) K (4,1) T (2,0)
018	GPESSGKT 66 73	1G18 (3.8) (A) 2REB (2.3)	1G19 (3.0) (A)	G (5,6) P (6,1) E (4,18) S (20,22)	S (22,12) G (27,28) K (33,30) T (12,9)
019	GDVACGKT 12 19	1A2B (2.4) 1DPF (2.0) (A)	1CXZ (2.2) (A)	G (6,5) D (2,6) V (6,2) A (6,10)	C (6,0) G (1,7) K (9,1) T (1,4)
020	GDGGTGKT 7 24	1A2K (2.5) (C,D,E) 1IBR (2.3) (A,C) 1QG2 (2.5) (A) 1RRP (2.9) (A,C)	1BYU (2.1) (A,B) 1QBK (3.0) (C) 1QG4 (2.5) (A,B) 3RAN (2.1) (A–D)	G (12,8 D (6,10) G (8,5) G (4,13)	T (14,5) G (9,13) K (11,4) T (4,6)
021	GDVAVGKT 210 217	1A4R (2.5) (A,B)		G (0,1) D (1,0) V (1,0) A (1,3)	V (4,1) G (3,3) K (3,3) T (2,0)
022	GDGAVGKT 10 17	1AM4 (2.7) (D,E,F) 1DOA (2.6) (A) 1E96 (2.4) (A) 1G4U (2.3) (R) 1HE1 (2.0) (C,D) 2NGR (1.9) (A)	1AN0 (2.8) (A,B) 1DS6 (2.3) (A) 1FOE (2.8) (B,D,F,H) 1GRN (2.1) (A) 1MH1 (1.3)	G (12,7) D (11,12) G (12,11) A (6,27)	V (26,5) G (6,12) K (11,6) T (4,5)
023	GQTSSGKT 86 93	1BG2 (1.8) 3KIN (3.1) (A,C)	2KIN (1.9) (A)	G (13,11) Q (15,6) T (7,4) S (6,14)	S (14,12) G (12,12) K (8,4) T (2,3)
024	GLPARGKT 45 52	1BIF (2.0) 3BIF (2.3) (A)	2BIF (2.4) (A,B)	G (4,3) L (2,2) P (1,5) A (3,4)	R (2,7) G (3,8) K (4,3) T (6,3)
025	GMDLKGKT 206 213	1BVU (2.5) (A-F)		G (16,25) M (26,9) D (12,6) L (4,12)	K (7,5) G (7,11) K (8,9) T (7,7)
026	GDGACGKT 12 19	1CC0 (5.0) (A,C)	1FTN (2.1) 1TX4 (1.6) (B)	G (1,4) D (3,9) G (3,1) A (7,8)	C (3,1) G (2,1) K (1,1) T (1,0)
027	GLHAMGKT 24 31	1CP2 (1.9) (A,B)		G (3,2) L (1,4) H (2,3) A (3,5)	M (3,7) G (8,1) K (3,3) T (1,1)
028 029	GAPANGKT 513 520 GQTGSGKT	1CWV (2.3) (A) 1CZ7 (2.9) (A–D)	2NCD (2.5) (A)	G (5,4)	S (11,16)
,_,	474 481	3KAR (2.3)	2000 (2.0) (A)	Q (4,4) T (3,13) G (11,11)	G (19,13) K (11,9) T (6,4)
030	GATGTGKT 39 46	1D2M (1.9) (A)	1D9Z (3.1) (A)	G (8,3) A (3,5) T (5,6) G (14,7)	T (3,2) G (2,15) K (17,13) T (13,1)
030A	GATGTGKT 39 46	1D9X (2.6) (A)			
031	GPPHSGKT 543 550	1D2N (1.7) (A)	1NSF (1.9)	G (1,1) P (0,1) P (1,1) H (1,2)	S (1,1) G (1,2) K (2,0) T (1,2)
0032	GEQAVGKT 18 25	1D5C (2.3) (A)		H (1,2)	T (1,2)

No.	Sequence Segment location	PDB code, (resolution in Å)	chain identifier	Residue r.m.s. (φ,ψ)	Residue r.m.s. (φ,ψ)
033	GPTGVGKT 57 64	1DO0 (3.0) (A–F) 1E94 (2.8) (E,F) 1G41 (2.3) (A) 1G4B (7.0) (E,F,K,L)	1DO2 (4.0) (A,C) 1G3I (3.4) (A–F) 1G4A (3.0) (E,F)	G (8,8) P (9,10) T (10,11) G (17,24)	V (19,19) G (13,20) K (18,11) T (13,10)
33A	GPTGVGKT 57 64	1DO2 (4.0) (B,D)		G (2,1) P (1,3) T (3,1) G (2,1)	V (1,1) G (1,2) K (1,2) T (1,1)
33B	GPTGVGKT 57 64	1G3I (3.4) (S,T,U,V,W)		G (1,1) P (1,1) T (0,1) G (0,0)	V (0,1) G (1,1) K (0,0) T (0,0)
34	GTEFEGKT 44 51	1DT0 (2.1) (A,B,C)		G (0,1) T (2,1) E (1,1) F (1,2)	E (2,1) G (1,2) K (1,2) T (2,2)
)35	GKGGVGKT 15 22	1F48 (2.3) (A)			
)36	GLQGSGKT 105 112	1FFH (2.0) 2FFH (3.2) (A,B,C) 3NG1 (2.3) A,B	1NG1 (2.0) 2NG1 (2.0)	G (5,7) L (6,4) Q (4,5) G (10,19)	S (19,7) G (9,7) K (3,4) T (4,5)
037	GRPGTGKT 50 57	1FNN (2.0) (A,B)		G (2,1) R (2,4) P (2,2) G (1,7)	T (4,4) G (2,2) K (1,2) T (2,1)
38	GAPVDGKT 116 123	1FS7 (1.6) (A) 1FS9 (2.0) (A)	1FS8 (1.6) (A)	G (2,2) A (3,1) P (2,1) V (1,3)	D (1,2) G (1,0) K (2,0) T (2,2)
39	GVNGVGKT 300 307	1FTS (2.2)			
40	GLDNAGKT 24 31	1FZQ (1.7) (A)			
41	GPSGCGKT 36 43	1G29 (1.9) (1,2)		G (3,2) P (2,1) S (1,3) G (1,2)	C (1,1) G (5,5) K (1,1) T (2,0)
42	GGTGSGKT 178 185	1G6O (2.5) (A,B)		G (1,4) G (8,3) T (2,5) G (4,1)	S (2,2) G (1,3) K (3,1) T (0,3)
143	GPPGLGKT 45 52	1HQC (3.2) (A,B)		G (1,9) P (9,6) P (2,15) G (25,13)	L (5,8) G (8,11) K (16,4) T (0,2)
44	GKGGTGKT 10 17	1HYQ (2.6) (A)			
45	GKVTSGKT 102 109	1JCK (3.5) (B,D)		G (0,0) K (0,0) V (0,0) T (0,0)	S (0,0) G (0,0) K (0,0) T (0,0)
46	GARGCGKT 9 16	1SHK (1.9) (A,B)	2SHK (2.6) (A,B)	G (5,5) A (4,1) R (4,2) G (3,4)	C (7,2) G (2,8) K (7,3) T (4,1)
147	GLDRTGKT 12 19	1TMK (2.1) (A,B) 3TMK (2.0) (A–H)	2TMK (2.4) (A,B)	G (5,1) L (4,9) D (5,14) R (9,8)	T (9,9) G (13,10) K (7,4) T (2,5)

Table I. Continued

No.	Sequence Segment location	PDB code, (resolution in Å) chain identifier	Residue r.m.s. (φ,ψ)	Residue r.m.s. (φ,ψ)
048	GNSSVGKT 29 36	1ZBD (2.6) (A)	3RAB (2.0) (A)	G (5,2) N (1,2) S (3,3) S (2,3)	V (3,2) G (1,5) K (1,5) T (4,2)
049	GLEGAGKT 10 17	4TMK (1.9) (A)	5TMP (1.9) (A)	G (3,1) L (2,2) E (3,3) G (2,9)	A (1,9) G (15,7) K (9,9) T (2,3)
050	GAGGVGKS 10 17	121P (1.5) 1CTQ (1.2) (A) 1GNQ (2.5) 1LFD (2.1) (B,D) 1QRA (1.6) (A) 221P (2.3) 5P21 (1.3) 6Q21 (1.9) (A–D)	1BKD (2.8) (R) 1GNP (2.7) 1GNR (1.8) 1Q21 (2.2) 1WQ1 (2.5) (R) 4Q21 (2.0) 621P (2.4) 721P (2.0)	G (6,10) A (13,7) G (9,11) G (10,10)	V (14,13) G (15,9) K (7,11) S (7,7)
051	GADGVGKS	1AGP (2.3)			
052	10 17 GAGESGKS 36 43	1AGR (2.8) (A,D) 1AZT (2.3) (A,B) 1BOF (2.2) 1CJK (3.0) (C) 1CJU (2.8) (C) 1CS4 (2.5) (C) 1FQJ (2.0) (A,D) 1GDD (2.2) 1GG2 (2.4) (A) 1GIL (2.3) 1GOT (2.0) (A) 1TAD (1.7) (A,B,C) 1TND (2.2) (A,B,C)	1AZS (2.3) (C) 1BH2 (2.1) 1CIP (1.5)(A) 1CJT (2.8) (C) 1CJV (3.0) (C) 1CUL (2.4) (C) 1FQK (2.3) (A,C) 1GFI (2.2) 1GIA (2.0) 1GIT (2.6) 1GP2 (2.3) (A) 1TAG (1.8)	G (14,5) A (7,5) G (7,7) E (8,8)	S (9,10) G (10,9) K (7,5) S (6,5)
053	GIVSYGKS 211 218	1AU8 (1.9) (A)	1CGH (1.8) (A)	G (3,1) I (0,0) V (1,3) S (5,2)	Y (1,1) G (1,3) K (6,1) S (2,3)
054	GDGTGGKS 78 85	1CYN (1.8) (A)		3 (3,2)	5 (2,5)
055	GPSGTGKS 8 15	1EX6 (2.3) (A,B) 1GKY (2.0)	1EX7 (1.9) (A)	G (6,5) P (3,5) S (10,6) G (4,6)	T (7,19) G (27,7) K (5,3) S (4,9)
056	GSGGVGKS 10 17	1C1Y (1.9) (A) 1KAO (1.7) 3RAP (2.2) R,S	1GUA (2.0) (A) 2RAP (2.6)	G (5,6) S (5,4) G (5,7) G (7,10)	V (10,3) G (4,4) K (3,4) S (6,6)
057	GDTSDGKS 183 189	1HYL (1.8) (A,B)		G (1,5) D (3,2) T (2,2) S (5,6)	D (5,3) G (2,3) K (4,7) S (6,2)
058	GTAFEGKS 44 51	1ISA (1.8) (A) 1ISC (1.8) (A)	1ISB (1.8) (A)	G (1,2) T (0,0) A (1,1) F (1,0)	E (1,0) G (1,4) K (2,0) S (0,1)
058A	GTAFEGKS 44 51	1ISA (1.8) (B) 1ISC (1.8) (B)	1ISB (1.8) (B)	G (1,2) T (1,1) A (2,2) F (1,2)	E (2,1) G (2,2) K (3,2) S (3,1)
059	GKGGIGKS 9 16	1CP2 (1.9) (A,B) 1FP6 (2.1) (A–D) 1G21 (3.0) (E–H) 1N2C (3.0) (E–H) 2NIB (2.2) (A,B)	1DE0 (2.4) (A,B) 1G1M (2.2) (A,B) 1G5P (2.2) (A,B) 1NIP (2.9) (A,B)	G (17,11) K (14,17) G (16,19) G (12,7)	I (24,21) G (23,13) K (13,11) S (10,12)
059A	GKGGIGKS 9 16	2NIP (2.2) (A,B) 1G20 (2.2) (E,F)		G (8,9) K (13,6) G (8,1) G (2,7)	I (6,14) G (16,2) K (1,4) S (3,5)

No.	Sequence Segment location	PDB code, (resolution in Å) chain identifier	Residue r.m.s. (φ,ψ)	Residue r.m.s. (φ,ψ)
)59B	GKGGIGKS 9 16	1G20 (2.2) (G,H)		G (12,7) K (9,4) G (19,30) G (1,22)	I (6,9) G (18,9) K (2,12) S (4,5)
60	GAVGVGKS 10 17	1HE8 (3.0) (B) 2Q21 (2.2)	1RVD (1.9) (A) 521P (2.6)	G (16,2) A (4,6) V (13,3) G (9,15)	V (13,9) G (8,10) K (10,10) S (8,9)
61	GMVVEGKS 190 197	1DJO (2.0) (A,B) 3PGA (2.) (1 – 4)	1DJP (1.9) (A,B) 4PGA (1.7) (A,B)	G (5,3) M (3,4) V (5,2) V (2,5)	E (9,6) G (10,12) K (6,4) S (5,3)
62	GARGVGKS	421P (2.2)			
063	10 17 GIIAPGKS 539 546	1AHU (2.7) (A,B) 1AHZ (3.3) (A,B) 1E0Y (2.7) (A,B) 1E8G (2.1) (A,B) 1QLT (2.2) (A,B)	1AHV (3.1) (A,B) 1DZN (2.8) (A,B) 1E8F (2.9) (A,B) 1E8H (2.6) (A,B) 1QLU (2.4) (A,B)	G (8,13) I (14,12) I (6,5) A (7,11)	P (8,6) G (6,7) K (5,6) S (5,6)
)64	GAVESGKS 40 47	1VAO (2.5) (A,B) 1AS0 (2.0) 1AS3 (2.4)	2VAO (2.8) (A,B) 1AS2 (2.8)	G (3,1) A (1,4) V (7,3) E (2,3)	S (4,3) G (6,3) K (2,3) S (4,3)
65	GSSGSGKS	1B0U (1.5) (A)		E (2,3)	S (4,3)
66	39 46 GVRFPGKS 313 320	1BAG (2.5)			
067	GGARSGKS 406 413	1C9K (2.2) (A,B,C)	1CBU (2.3) (A,B,C)	G (7,9) G (7,10) A (8,7) R (6,5)	S (5,6) G (5,7) K (7,3) S (6,10)
68	GFAKTGKS 195 202	1CA1 (1.9) 1QMD (2.2) (A,B)	1QM6 (2.5) (A,B)	G (1,3) F (2,1) A (2,3) K (1,1)	T (2,1) G (2,3) K (2,1) S (1,2)
69	GLLEAGKS 174 181	1CD1 (2.6) (A,C)		G (9,4) L (6,3) L (5,5) E (6,12)	A (13,4) G (10,21) K (16,3) S (3,6)
070	GAPGVGKS 10 17	ICLU (1.7) (A) IJAI (1.8)	1JAH (1.8) 821P (1.5)	G (5,2) A (4,3) P (5,2) G (8,3)	V (2,2) G (4,3) K (3,2) S (3,3)
071	GGSCTGKS 434 441	1CLV (2.0) (A) 1TMQ (2.5) (A)	1JAE (1.6) 1VIW (3.0) (A)	G (5,13) G (7,8) S (5,7) C (11,2)	T (5,8) G (8,28) K (21,16) S (14,32)
072	GRSGRGKS 138 145	1HJB (3.0) (C,F) 1IO4 (3.0) (C)	1HJC (2.6) (A,D)	G (8,4) R (11,9) S (12,12) G (14,5)	R (3,6) G (5,11) K (11,3) S (4,3)
73	GPLYLGKS 587 594	1CMX (2.2) (A,C)		G (0,1) P (1,1) L (0,2) Y (2,2)	L (2,3) G (3,1) K (1,2) S (3,0)
74	GLSASGKS 32 39	1D6J (2.0) (A,B)		G (5,2) L (1,2) S (2,1) A (4,7)	S (5,4) G (1,1) K (2,3) S (4,2)
075	GQAMPGKS 105 112	1DDZ (2.2) (A,B)		G (1,1) Q (0,0) A (1,0) M (1,0)	P (1,1) G (1,1) K (0,1) S (1,1)

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Table I. Continued								
No.	Sequence Segment location	PDB code, (resolution in Å) chain identifier	Residue r.m.s. (φ,ψ)	Residue r.m.s. (φ,ψ)			
076	GVVQPGKS 510 517	1DEE (2.7) (B,D,F)		G (1,1) V (1,1) V (0,2) Q (5,1)	P (2,1) G (1,2) K (1,0) S (2,3)			
)77	GLYRTGKS 45 52	1DG3 (1.8) (A)		Q (3,1)	5 (2,5)			
)77A	45 52 GLYRTGKS 45 52	1F5N (1.7) (A)						
78	GENGIGKS 42 49	1DYW (1.8) (A)	1E3B (1.8) (A)	G (2,2) E (2,1) N (3,2) G (3,3)	I (1,1) G (0,2) K (2,2) S (4,2)			
79	GRPNVGKS 15 22	1EGA (2.4) (A,B)		G (2,0) R (2,3) P (2,1) N (1,3)	V (4,3) G (4,1) K (2,1) S (1,1)			
80	GAAAAGKS 893 900	1EJ6 (3.6) (A)						
081	GEAAVGKS 14 21	1EK0 (1.4) (A)						
182	GSVAVGKS 95 102	1ESM (2.5) (A–D)	1ESN (2.6) (A–D)	G (3,5) S (13,14) V (4,2) A (13,10)	V (3,2) G (8,3) K (6,17) S (8,10)			
083	GAEAAGKS 188 195	1F07 (2.0) (A–D)		G (1,3) A (4,2) E (3,4) A (4,2)	A (4,4) G (4,2) K (2,4) S (5,3)			
84	GQNGSGKS 30 37	1F2T (1.6) (A)	1F2U (1.6) (A,C)	G (13,3) Q (3,12) N (8,4) G (16,14)	S (13,8) G (7,3) K (7,13) S (2,6)			
85	GHVDSGKS 14 21	1F60 (1.6) (A)	1G7C (2.0) (A)	G (0,1) H (5,4) V (3,4) D (0,2)	S (4,6) G (5,3) K (1,1) S (1,3)			
86	GDSGVGKS 27 34	1G16 (1.8) (A–D)	1G17 (2.0) (A,B)	G (3,3) D (3,3) S (1,5) G (4,4)	V (4,2) G (2,3) K (3,2) S (2,2)			
87	GLVSPGKS 951 958	1HQM (3.3) (C)						
88	GESAVGKS 28 35	1HUQ (1.8) (A)						
89	GIPGVGKS 8 15	1NKS (2.5) (A–F)		G (7,2) I (4,3) P (4,7) G (12,11)	V (10,8) G (12,9) K (6,3) S (5,8)			
90	GDVSPGKS 457 464	1QHB (2.3) (A–F)		G (2,2) D (2,2) V (1,2) S (1,1)	P (2,1) G (3,2) K (3,3) S (4,3)			
91	GGNGAGKS 34 41	1QHL (2.2) (A)						
)92	GGSSAGKS 10 17	1QHN (2.7) (A) 1QHX (2.5) (A)	1QHS (2.8) (A) 1QHY (2.6) (A)	G (3,2) G (4,3) S (3,2) S (2,1)	A (1,2) G (4,3) K (3,1) S (4,5)			

For those groups which have more than one entry, structural similarity is brought out by the small r.m.s. values of the Ramachandran angles (ϕ, ψ) , given in the last column.

059 – GKGGIGKS); and four for one sequence (003 – GGAGVGKT). These data implied that highly localized conformational variants are possible in these segments retaining overall structural similarity.

Conformational variants of segments of Walker sequences The next step was the grouping of the conformations irrespective of the sequence of the variable region of the Walker segment. This was done as follows: (1) for those

Table II. The entries selected from Table I regrouped based on their structural similarity [the examples in set VII do not possess any structural similarity, as analyzed using the Ramachandran angles (ϕ, ψ)]

Sr. No.	Sequence of Walker motif	Segment location	PDB code (chain)	Name of the protein
Set I				
1	GLSGTGKT	248-255	1AYL	Phosphoenol pyruvate kinase
2	GDRQTGKT	169-176	1E79 (A)	F_1 -ATPase α subunit
3	GPESSGKT	66–73	2REB	REC A protein.
4	GLPARGKT	45-52	1BIF	6-Phosphofructo-2-kinase/fructose-2,6-bisphosphatase
5	GQTGSGKT	474-481	3KAR	Kinesin-like protein KAR3
6	GATGTGKT	36–43	1D2M (A)	UvrB protein
7	GPPHSGKT	551-558	1D2N (A)	N-Ethylmaleimide-sensitive fusion protein
8	GPTGVGKT	57-64	1G41 (A)	Heat shock protein HslU
9	GLQGSGKT	105-112	1FFH 1ENN (A)	Signal recognition protein FFH
10 11	G R P G T G K T G L D N A G K T	50–57 24–31	1FNN (A) 1FZQ (A)	CDC6p ADP-ribosylation-like factor
12	GPSGCGKT	36-43	1G29 (1)	Mal K
12	GLDRTGKT	12-19	3TMK (A)	Thymidylate kinase (S.cerevisiae)
13	GLEGAGKT	10-17	4TMK (A)	Thymidylate kinase (<i>E.coli</i>)
15	GPSGTGKS	8-15	1EX7 (A)	Guanylate kinase
16	GSGGVGKS	10-17	1KAO	Small G-protein rap $2a + GDP$
17	GKGGIGKS	8-15	1CP2 (A)	Nitrogenase iron protein
18	GSSGSGKS	39-46	1B0U (A)	Histidine permaease HisP–ATP binding subunit
19	GGARSGKS	6-13	1C9K (A)	Adenosylcobinamide kinase
20	GLYRTGKS	45-52	1F5N (A)	Guanylate binding protein
21	GEAAVGKS	14-21	1EK0 (A)	YPT51
22	GQNGSGKS	30-37	1F2T (A)	Rad50 ABC ATPase
23	GDSGVGKS	27-34	1G16 (A)	Sec4
24	GESAVGKS	28-35	1HUQ (A)	Rab5c
25	GIPGVGKS	8-15	1NKS (A)	Adenylate kinase
26	GGSSAGKS	10-17	1QHX (A)	Chloramphenicol phosphotransferase
27	GHVDHGKT	18-25	1EXM (A)	EF-Tu
28	GPHGMGKT	56-63	1E2K (A)	Thymidine kinase
29	GESGAGKT	179–186	1MMG	Myosin motor domain
30	GDGGTGKT	17–24	1BYU (A)	RAN-GTPase (+ GDP)
31	GDGAVGKT	10-17	1MH1	RAC1
32	GDGACGKT	12–19	1TX4 (B)	Rho A
33	GEQAVGKT	18-25	1D5C (A)	Rab6 + GDP
34	GKGGVGKT	15-22	1F48 (A)	Arsenite transporting ATPase
35	GVNGVGKT	300-307	1FTS	Signal recognition particle receptor
36	GGTGSGKT	178–185	1G60 (A)	Traffic ATPase (+ ADP)
37 38	GARGCGKT	9–16 29–36	1SHK (A) 2DAP (A)	Shikimate kinase Rab3a
38 39	G N S S V G K T G A G G V G K S	10-17	3RAB (A) 1CTQ (A)	P21 Ras
40	GAGGVGKS	40-47	1CIQ (A) 1CIP (A)	Guanine nucleotide binding protein alpha-I
40	GRPNVGKS	15-22	1EGA (A)	GTP binding protein ERA
42	GSVAVGKS	95-102	1ESM (A)	Pantothenate kinase
43	GGAGVGKT	156-163	1E5M (N) 1E79 (D)	F_1 -ATPase β subunit
44	GLDAAGKT	24-31	1HUR (A)	ADP-ribosylation factor-1
45	GQTSSGKT	85-92	1BG2	Kinesin motor domain
Set II	~ ~ ~			
46	GHVDHGKT	18-25	1EFU (A)	EF-Tu
40	GLSASGKS	32–39	1D6J (A)	APS kinase
48	GHVDSGKS	14-21	1F60 (A)	Elongation factor EEF1A
49	GKGGIGKS	9–16	1G20 (G)	Nitrogenase iron protein
50	GLYRTGKS	45-52	1DG3 (A)	Guanylate binding protein-1
Set III				
51 Set III		264-271	(MUT (A)	HhaI methyltransferase
52	G Y L V N G K T	190–197	6MHT(A)	Glutaminase–asparaginase
	GMVVEGKS	190–197	4PGA (A)	Glutaninasc–asparaginasc
Set IV		24.25	1000 (1)	
53	GLHAMGKT	24-31	1CP2 (A)	Nitrogenase iron protein
54	GAEAAGKS	188–195	1F07 (A)	Tetrahydromethanopterin reductase
Set V		105 202	1011	
55	GFAKTGKS	195-202	1CA1	Alpha-toxin
56	GLLEAGKS	174–181	1CD1 (A)	CD1
Set VI				
57	GATGTGKT	39–46	1D9X (A)	UvrB
58	GPPGLGKT	45-52	1HQC (A)	RuvB
Set VII				
59	GGAGVGKT	156-163	1BMF (E)	F_1 -ATPase β subunit (bovine)
60	GGAGVGKT	156–163	1E1R(E)	F_1 -ATPase β subunit (bovine)
			- (/	· · · · · · · · · · · · · · · · · · ·

Table II. Continued								
Sr. No.	Sequence of Walker motif	Segment location	PDB code (chain)	Name of the protein				
61	GGAGVGKT	156-163	1MAB (B)	F ₁ -ATPase β subunit (rat)				
62	GAHALGKT	173–180	2CYP	Cytochrome c peroxidase				
63	GAGALGKT	173–180	1DS4 (A)	Cytochrome c peroxidase				
64	GHVDHGKT	18-25	1ETU	EF-Tu domain 1				
65	GTAFPGKT	212-219	1QPA (A)	Lignin peroxidase				
66	GLRSDGKT	487–494	1MTY (D)	Methane monoxygenase (Mc) ^a				
67	GLRSDGKT	487–494	1MMO (D)	Methane monoxygenase (Mt) ^a				
68	GKVTGGKT	102-109	1STE	Sec2 superantigen				
69	GLPAIGKT	499-506	1BGX (T)	Taq polymerase				
70	GLPAIGKT	499-506	1QSS (A)	Tag Klenow fragment				
71	GSQAGGKT	47–54	1WGT (A)	Wheat germ agglutinin				
72	GMDLKGKT	206-213	1BVU (A)	Glutamate dehydrogenase				
73	GAPANGKT	513-520	1CWV (A)	Invasin				
74	GPTGVGKT	57-64	1DO2 (B)	HslU				
75	GPTGVGKT	57-64	1G3I (S)	HslU protease				
76	GAPVDGKT	116-123	1FS7 (A)	Cytochrome c nitrite reductase				
77	GKGGTGKT	10–17	1HYQ (A)	MinD-1				
78	GKVTSGKT	102-109	1JCK (B)	Sec3 superantigen				
79	GIVSYGKS	211-218	1CGH (A)	Cathepsin G				
80	GDGTGGKS	78-85	1CYN (A)	Cyclophilin B				
81	GDTSDGKS	183-189	1HYL (A)	Collagenase				
82	GTAFEGKS	44-51	1ISA (A)	Superoxide dismutase				
83	GTAFEGKS	44-51	1ISA (B)	Superoxide dismutase				
84	GKGGIGKS	9–16	1G20 (E)	Nitrogenase iron protein				
85	GIIAPGKS	539-546	1E8G (A)	Vanillyl-alchohol oxidase				
86	GVRFPGKS	313-320	1BAG	α -Amylase (<i>B.subtilis</i>)				
87	GGSCTGKS	434-441	1JAE	α-Amylase (yellow mealworm)				
88	GRSGRGKS	138–145	1HJC (A)	RUNT-related transcription factor-1				
89	GPLYLGKS	187–194	1CMX (A)	Ubiquitin				
90	GQAMPGKS	105–112	1DDZ (A)	Carbonic anhydrase				
91	GVVQPGKS	510-517	1DEE (B)	IgM heavy chain				
92	GENGIGKS	42-49	1DYW (A)	Cylophilin 3				
93	GAAAGKS	893-900	1EJ6 (A)	Reovirus core protein $\lambda 2$				
94	GLVSPGKS	951-958	1HQM (C)	Bacterial RNA polymerase β subunit				
95	GDVSPGKS	457-464	1QHB (A)	Vanadium bromo-peroxidase				
96	GGNGAGKS	34-41	1QHL (A)	MukB N-terminal domain				

^aMc, Methylococcus capsulatus; Mt, Methylosinus trichosporum

	Position No.								
	1	2	3	4	5	6	7	8	
Set I [45] Mean (ϕ, ψ) R.m.s. (ϕ, ψ)	(159,166) (18,11)	(-67,162) (10,10)	(-63,141) (8,11)	(79,12) (14,15)	(-87,-11) (17,14)	(104,20) (16,11)	(-60,-46) (7,10)	(-66,-40) (5,5)	
Set II [5] Mean (ϕ, ψ) R.m.s. (ϕ, ψ)	(158,171) (11,7)	(-66,157) (7,8)	(-60,-36) (13,7)	(-82,6) (19,15)	(-86,-5) (20,14)	(73,29) (12,13)	(-63,-32) (5,9)	(-66,-43) (4,8)	
<i>Set III [2]</i> Mean (φ,ψ) R.m.s.(φ,ψ)	(-138,176) (35,25)	(-138,154) (2,6)	(-106,114) (17,8)	(-127,125) (8,4)	(41,54) (1,3)	(84,–11) (3,5)	(-115,149) (10,13)	(-104,134) (7,7)	
<i>Set IV</i> [2] Mean (φ,ψ) R.m.s.(φ,ψ)	(-63,-45) (4,4)	(-62,-44) (9,7)	(-65,-38) (1,3)	(-62,-21) (3,9)	(-97,7) (7,8)	(82,16) (2,2)	(-99,163) (12,19)	(-114,141) (13,16)	
<i>Set V</i> [2] Mean (φ,ψ) R.m.s.(φ,ψ)	(-52,-47) (8,2)	(-67,-45) (4,6)	(-61,-28) (5,7)	(-76,-44) (7,5)	(-73,-36) (8,2)	(-71,-34) (4,12)	(-44,-38) (17,3)	(-63,-49) (1,1)	
<i>Set VI [2]</i> Mean (φ,ψ) R.m.s.(φ,ψ)	(-141,138) (8,2)	(-65,151) (11,3)	(-62,147) (6,14)	(89,-2) (3,32)	(-95,144) (21,29)	(-70,85) (23,18)	(-78,-40) (26,9)	(-59,-52) (11,4)	

The corresponding values do not have any meaning for set VII, which has structurally dissimilar conformations. The number of examples in each set is given in parentheses along with the set number in the first column.

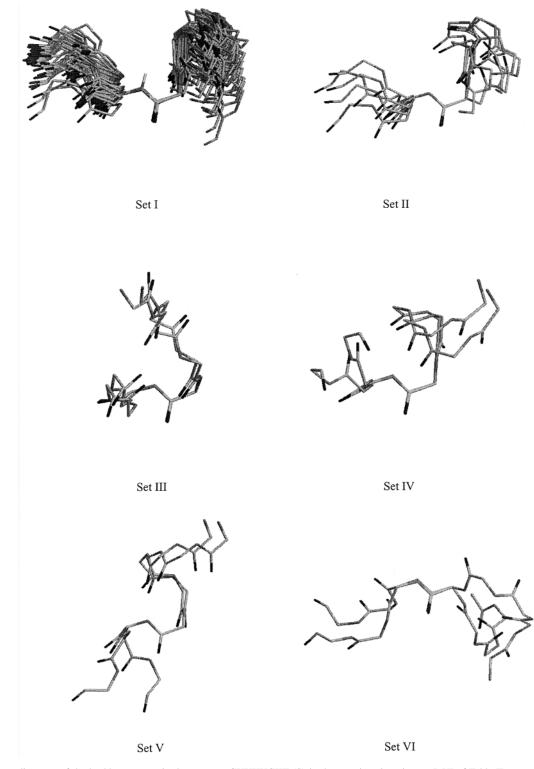
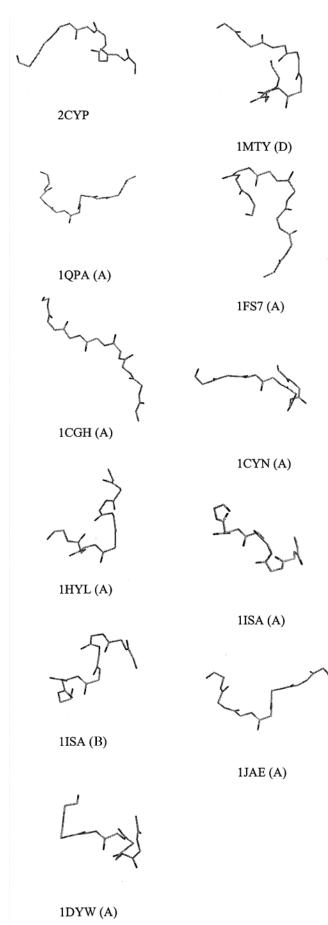


Fig. 1. Wire-frame diagrams of the backbone atoms in the segment GXXXXGKT (S) in the proteins given in sets I–VI of Table II.

groups in Table I which had only one entry, the choice was unambiguous; (2) for those groups having more than one entry, one with the best resolution shown in bold face in Table I had been picked up as the representative of the group/sub-group. These collectively gave 107 examples which were regrouped solely on the basis of similarity of the Ramachandran angles (ϕ , ψ). Of the new sets thus

obtained, 53 (out of a total of 107) entries constituted the major set. Another set had five entries, while seven others had two entries each. The last set comprised 35 entries, without any structural similarity among them. In any particular set, proteins with high overall sequence homology could be found, although the sequences of the variable region were different. These are as follows: (1AYL, 10EN);



(1A4R, 1MH1); (1DPF, 1TX4); (1CIP, 1AS0), (1AGP, 1CTQ, 1RVD, 421P, 821P); pairs [(2CYP, 1CCG); (1MHY (D), 1MTY (D)], as well as [1DT0 (A), 1ISA (B)]. Since the structures in such cases are expected to be similar, the entries that had the best resolution were retained. These were 1AYL, 1MH1, 1TX4, 1CIP, 1CTQ, 2CYP, 1MTY (D) and 1ISA (B). The final grouping thus obtained is given in Table II, which has 45 proteins in set I, five in set II, two each in sets III, IV, V and VI and the remaining 38 in set VII. The mean and r.m.s. (ϕ , ψ) values of sets I–VI are given in Table III and these are small enough to warrant structural similarity among the members. The r.m.s. has no relevance for the last set (set VII).

For easy comprehension of the structural grouping, the line diagrams of the backbone of GXXXXGKT (S) segments of the proteins in sets I–VI, drawn with the peptide unit spanning residues 5-(X) and 6-(G) as the common internal frame of reference, are shown in Figure 1. The sickle-like folding with overlap of the atoms of the backbone is seen with members of the set I (Figure 1). Nearly the same structure of segment 5–8 is found in set II, but that of segment 1–4 is different (Table III). Set VII is comprised of structures of differing conformations indicating the flexibility of Walker sequences to acquire random folding. Out of the 38 examples in this set, only those which have resolution of 1.8 Å or better are shown in Figure 2.

Differing structures with same Walker sequence

Structural differences between segments having the same Walker sequence are also perceivable from the foregoing data. There are nine such examples in the present data set. The PDB codes along with the Ramachandran angles at the eight positions of Walker sequence of these nine pairs are given in Table IV. Large differences in Ramachandran angles are observed at four different locations within the segment (shown in bold face in the table). These are as follows: (i) 1VOM–2MYS (A), (ii) 1F5N (A)–1DG3 (A), (iii) 1FP6 (A)–1G20 (E) and (iv) 1EFT–1EFU (A) all at locations 3/4; (v) 1MMO (D)–1MMO (E) at locations 4/5; (vi) 1ISA (A)–1ISA (B), (vii) 1D9X (A)–1D9Z (A) and (viii) 1BMF (D)–1BMF (E) at locations 5/6; and (ix) 1G3I (A)–1G3I (S) at locations 6/7. These large changes arise owing to a flip of the peptide unit spanning the two residues.

There are more than two examples with differing conformations for two of the sequences. The first example consists of 1EFT, 1EFU (A) and 1ETU, which have the same sequence, GHVDHGKT (iv in Table IV), but the peptide unit between residues 3 and 4 in 1EFT and 1EFU (A) is flipped. The conformation of the third member of this group, 1ETU, does not match in entirety with the other two examples. A close examination reveals that the conformations of 1EFT and 1ETU differ only in the segment 18–20. The second example is of 1BMF (D, E), 1MAB (B) and 1E1R (E) having the same sequence GGAGVGKT. The peptide unit between locations 5 and 6 in 1BMF (E) and 1BMF (D) is flipped, as also is the one between locations 6 and 7 in 1BMF (D) and 1MAB (B) (viii in

Fig. 2. Wire-frame diagrams of the backbone atoms in the segment GXXXXGKT (S) in the proteins belonging to set VII of Table II. Only those examples occurring in protein structures which have a resolution of 1.8 Å or better are shown.

Table IV Ramachandran angles (ϕ, ψ) (°) in the segment GXXXXGKT for those examples with same sequence for XXXX but with different conformations

No.	Sequence, PDB code (chain) and segment locations	d (ϕ,ψ) at position							
		1	2	3	4	5	6	7	8
(i)	GESGAGKT, 1VOM, 179–186	(150,155)	(-73,171)	(-59,131)	(86,4)	(-67,-26)	(117,18)	(-61,-47)	(-67,-37)
	GESGAGKT, 2MYS(A), 179–186	(178,174)	(-72,166)	(-35,-42)	(-101,2)	(-61,-25)	(82,57)	(-82,-47)	(-44,-48)
(ii)	GLYRTGKS, 1F5N(A), 45–52	(155,173)	(-53,147)	(-63,157)	(60,34)	(-111,13)	(79,28)	(-66,-52)	(-60,-41)
	GLYRTGKS, 1DG3(A), 45–52	(149,178)	(-75,161)	(-60,-42)	(-54,-22)	(-71,-14)	(65,31)	(-67,-17)	(-60,-32)
(iii)	GKGGIGKS, 1FP6(A), 9–16	(155,176)	(-64,163)	(-73,131)	(74,13)	(-79,-24)	(130,20)	(-75,-25)	(-76,-35)
	GKGGIGKS, 1G20(E), 9–16	(137,156)	(-53,138)	(-80,-35)	(-62,10)	(-97,-49)	(134,39)	(-61,-46)	(-70,-32)
(iv)	GHVDHGKT, 1EFT, 18–25	(147,165)	(-65,164)	(-66,133)	(75,18)	(-91,-20)	(125,31)	(-67,-46)	(-73,-34)
	GHVDHGKT, 1EFU(A), 18–25	(175,176)	(-63,141)	(-45,-41)	(-102,24)	(-101,4)	(84,40)	(-68,-44)	(-67,-47)
	GHVDHGKT, 1ETU, 18–25	(-126, -166)	(-120,-41)	(143,82)	(133,-27)	(-47,-37)	(119,10)	(-49,-47)	(-57,-46)
(v)	GLRSDGKT, 1MMO(D), 487–494	(-80,42)	(-125,158)	(-79,161)	(-62,137)	(125,-29)	(74,23)	(-135,-59)	(-69,136)
	GLRSDGKT, 1MMO(E) ^a , 487–494	(-84,50)	(-129,148)	(-70,170)	(-70,-1)	(-86,-2)	(65,27)	(-145,-53)	(-80,138)
(vi)	GTAFEGKT, 1ISA(A), 44–51	(81,8)	(-112,173)	(-70,-13)	(-74,-23)	(-69,134)	(92,-12)	(-87,162)	(-79,166)
	GTAFEGKT, 1ISA(B), 44–51	(83,9)	(-107,174)	(-67,-19)	(-68,-21)	(-64,-25)	(-100,31)	(-126,159)	(-76,161)
(vii)	GATGTGKT, 1D9X(A), 39–46	(-122,133)	(-75,149)	(-68,161)	(92,-34)	(-73,173)	(-92,68)	(-52,-48)	(-70,-48)
	GATGTGKT, 1D9Z(A), 39–46	(-110,129)	(-68, -169)	(-68,156)	(48,26)	(-99,2)	(76,84)	(-97,-53)	(-41,-41)
(viii)	GGAGVGKT, 1BMF(E), 156–163	(93,153)	(-125,-107)	(-86,159)	(78,-37)	(-86,133)	(-48,30)	(-41,-58)	(-69,22)
	GGAGVGKT, 1BMF(D), 156–163	(-174,145)	(-75,176)	(-65,127)	(68,41)	(-112,-8)	(128,5)	(- 52,-60)	(-64,-43)
	GGAGVGKT, 1MAB(B), 156–163	(-153,160)	(-76,116)	(-100,105)	(69,89)	(-160,-25)	(179, -158)	(88, -106)	(-45,-52)
	GGAGVGKT, 1E1R(E), 156–163	(-170,-87)	(111, -176)	(-74,-71)	(66,17)	(-131,23)	(79,17)	(-51,-54)	(-63,-35)
(ix)	GPTGVGKT, 1G3I(A), 57–64	(-161,179)	(-82, -172)	(-80,136)	(43,65)	(-124,8)	(126,-23)	(-34,-51)	(-80,-40)
	GPTGVGKT, 1G3I(S), 57–64	(-174,158)	(-47, -165)	(-118,92)	(141,–19)	(73,-4)	(82,98)	(-120,-23)	(-89,-33)

^aThe entry 1MMO (E) has been taken in the place of 1MTY (D) of Table II since both of these are structurally and sequentially highly homologous.

Table V. Distribution of the different types secondary structures flanking
Walker sequence GXXXXGKT (S) in the examples given in Tables I and II

Secondary structure flanking Walker sequence		Examples in Table I	Examples in Table II
Preceding	Following		
α	α	62	9
α	β	19	4
α	X	91	9
β	α	418	60
β	β	21	8
β	X	4	3
X	α	9	0
Х	β	23	3
X	X	2	1

 $\alpha = \alpha$ -Helix; $\beta = \beta$ -strand; X = neither α nor β .

Table IV, shown as overlapping boxes). The fourth entry, 1EIR (E), has an altogether different conformation.

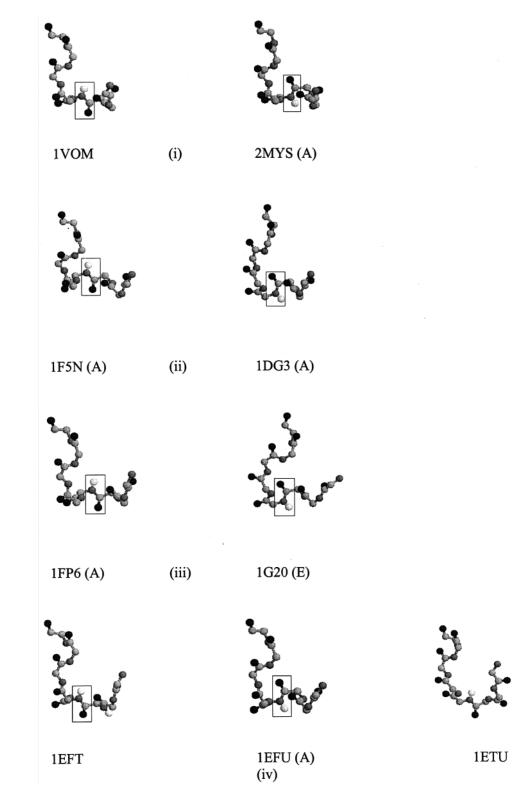
The last entry in Table IV corresponds to the sequence GPTGVGKT occurring in the two chains A and S of the protein 1G3I and the conformations are different. In this case the peptide unit between locations 6 and 7 show a rotation of $\approx 90^{\circ}$ about the virtual C^{α} – C^{α} bond, instead of a flip, as is found in the other examples.

The ball and stick diagrams of these nine examples with a flipped peptide unit shown within a box are given in Figure 3. The overlap of the polypeptide backbones appears good. The examples of pairs i–vi correspond to the flip occurring at the middle peptide unit of the well-known $4\rightarrow1$ hydrogenbonded β -turns of types I and II (Venkatachalam, 1968; Gunasekaran *et al.*, 1998). However, the flip of the peptide unit observable in pairs vii–ix does not correspond to the β -

turn flip as the values of (ϕ, ψ) are far different from those characteristic of β -turn ranges. Further, the $4\rightarrow 1$ hydrogen bond is also absent. Notwithstanding the flip, the same overall backbone structure is retained.

The examples of nucleotide-binding proteins are arranged in Figure 3 with the nucleotide bound forms on the left and the free forms on the right. These are as follows: myosin ATPase [1VOM-2MYS (A)], guanylate binding protein [1F5N (A)-1DG3(A)], nitrogenase, [1FP6(A)-1G20(E)] elongation factor Tu [1EFT-1EFU(A), uvrB protein (A)-DNA helicase [1D9Z(A)-1D9X(A)], F₁-ATPase [1BMF(D)-1BMF(E) and 1MAB(B)] and the HSLUV protease chaperone complex [1G3I(A)–1G3I(S)]. Wherever the nucleotide is bound the N-H of the flipped peptide unit projects inwards of the loop. In the case of F₁-ATPase (Abrahams et al., 1994), this N–H forms a hydrogen bond with P=O of the β -phosphate. It appears that the presence or absence of the nucleotide makes the difference between the two structural forms. The residues of Walker sequence in such proteins not only bind to the nucleotide phosphates but also show consequent localized structural changes. This feature has important implications in the biochemical events that occur at this site.

In the case of proteins with oxygen-related reactions, the difference appears to be present in the polypeptides as isolated. The two proteins of methane monooxygenase, showing a flip at position 4/5, are derived from two organisms. The Walker sequence is present only in the Fe-form of superoxide dismutase and the two identical subunits of this enzyme protein exhibit this flip of a peptide unit. It is possible that the O=O group may act as the P=O in nucleotide phosphate in protein–substrate interactions. No relationship has so far been found between the peptide flips in Walker sequences and the activities of these proteins.



The secondary structures flanking the Walker sequence

The foregoing analysis indicated that the variable region is unlikely to determine the conformation of the Walker sequence A found in many nucleotide-binding proteins. The characteristic loop structure of the Walker sequence in these proteins is known to be preceded by a β -strand and followed by an α helix (see, for an example, Abrahams *et al.*, 1994). It was therefore of interest to examine the occurrence of the flanking secondary structure of Walker sequences in proteins listed in Tables I and II. For this purpose, segments of eight residues on either side of Walker sequences were examined for the presence of secondary structures ($\alpha = \alpha$ -helix; $\beta = \beta$ -strand; X = neither α nor β ; W = Walker sequence A). All nine possible combinations do occur and their distribution is given in Table V. The majority of the examples fall into the category of β -W- α . This structural motif is present in all cases in the sets 1, 2 and 6 and some in set 7 of Table II. Interestingly, each of these proteins can bind to nucleotides leading to

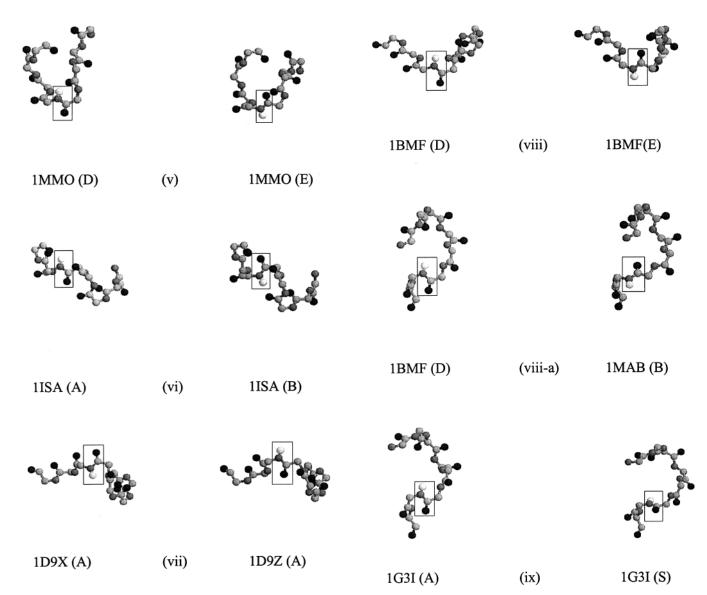


Fig. 3. Ball and stick diagrams of the backbone atoms in the segment GXXXXGKT (S) in the examples given in Table II. The flip of the peptide unit can be seen in the box, shown at the stated positions; 3 and 4 for i–iv; 4 and 5 for v; 5 and 6 for vi–viii; 6 and 7 for viii-a and ix. Shown as a white ball in this peptide unit, the hydrogen atom has been geometrically fixed.

hydrolysis of the terminal phosphate to provide energy for accompanying reactions (e.g. ATPases) in a large number of cases and in some cases transfer the phosphate to acceptors (kinases). This is true of the examples of proteins in the miscellaneous set 7. Hence it appears that the structural motif β -W- α , but not W alone, is the determining factor for nucleotide binding. The examples in sets 3, 4 and 5 of Table II, although small in number (only two each), show distinctive motifs of X-W- β , α -W- β and α -W- α , respectively.

Discussion

The noteworthy observation in this study is that the Walker sequence is present in many proteins and is not limited to those that bind and/or use nucleotides in their actions. Because of the belief that it provides the loop for phosphate binding, the so-called P-loop, this sequence was looked for only in such proteins and was invariably found. A search in the PDB files for its general occurrence, undertaken in this study, revealed its broad distribution (Tables I and II). The diversity of these proteins is truly amazing. These include peroxidases (of cytochrome, lignin), proteases (cathepsin, collagenase, serine protease), enzymes (methane monooxygenase, superoxide dismutase, α -amylase, glutamate dehydrogenase, carbonic anhydrase, Taq polymerase, etc.), binding proteins (lectin, trypsin inhibitor) and miscellaneous proteins (α -toxin, cyclophilin B, enterotoxin). An examination of the structures of cytochrome peroxidase and superoxide dismutase indicated that these sequences are present at some distance from the active metal centers. It is to be ascertained whether Walker sequences in these proteins are utilized in their actions or their presence is incidental. It becomes obvious that the Walker sequence is more widely distributed and presence of the P-loop seems to be restricted to the nucleotide binding proteins.

The second observation in this study is the sharing of a common loop structure in proteins of the major group which use and bind nucleotide phosphates. These include kinases, phosphatases, ATPases, heat shock proteins, transfer/transport ATPases, permeases, myosin motor domain and elongation factor. The variable quartet (XXXX) has little influence on the bend as seen from the minor variation of overlap in this region (Figure 1). Indeed, the variable quartet is so highly random in sequence that it gives no clue on the looping. Of these, G (13.3%), A (11.9%), S (9.8%), V (8.4%) and T (5.9%) occur more commonly than other amino acids, but no sequence can be identified with a set or a sub-set of proteins. Thus the formation of the β -turn loop seems to depend less on this sequence and more on the polypeptide chains on either side of the P loop, characteristically a β -sheet at the N-terminus and an α -helix at the C-terminus. The absence of the classical $4\rightarrow 1$ hydrogen bond in these loop structures appears to provide more room to surround and manipulate the phosphate chain of nucleotides for exchanging terminal phosphate.

Finally, the minor, local differences in the structures with the same Walker sequence, in our opinion, are of importance as they offer possibilities of participation in the functions of these proteins. These relate to the flip of peptide units in four positions (3-4, 4-5, 5-6, 6-7 in Table IV) in these sequences. The large differences in Ramachandran angles indeed brings to light these structural variants. Three examples are noted in the pairs that show these flips: the same enzyme protein from two different organisms (methyl monooxygenase), the two subunits of a homodimer protein (Fe-superoxide dismutase) and the binding of nucleotide to one of the two subunits (F₁-ATPase, β -subunit). The last example is a case with possible interaction of the substrate and the backbone structure of the enzyme active site and offers interesting mechanistic possibilities. Details of this have been reported elsewhere (Ramasarma and Ramakrishnan, 2002).

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