

Review

# A Mini-Review on Functional Near-Infrared Spectroscopy (fNIRS): Where Do We Stand, and Where Should We Go?

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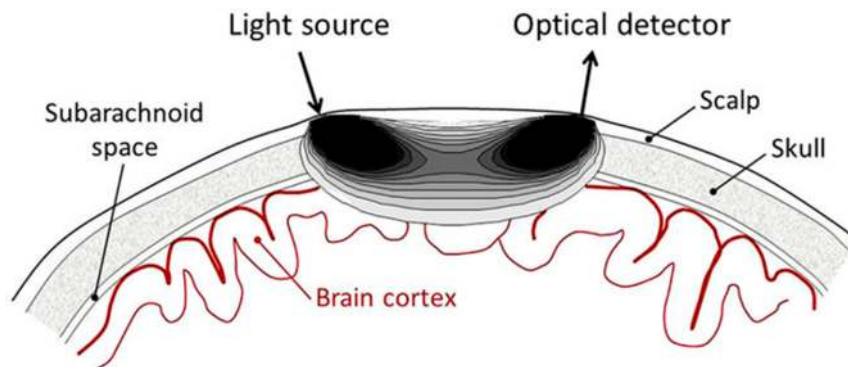
**Abstract:** This mini-review is aimed at briefly summarizing the present status of functional near-infrared spectroscopy (fNIRS) and predicting where the technique should go in the next decade. This mini-review quotes 33 articles on the different fNIRS basics and technical developments and 44 reviews on the fNIRS applications published in the last eight years. The huge number of review articles about a wide spectrum of topics in the field of cognitive and social sciences, functional neuroimaging research, and medicine testifies to the maturity achieved by this non-invasive optical vascular-based functional neuroimaging technique. Today, fNIRS has started to be utilized on healthy subjects while moving freely in different naturalistic settings. Further instrumental developments are expected to be done in the near future to fully satisfy this latter important aspect. In addition, fNIRS procedures, including correction methods for the strong extracranial interferences, need to be standardized before using fNIRS as a clinical tool in individual patients. New research avenues such as interactive neurosciences, cortical activation modulated by different type of sport performance, and cortical activation during neurofeedback training are highlighted.

**Keywords:** functional near-infrared spectroscopy; functional neuroimaging; optical imaging; cortical activation

## 1. Introduction

Biophotonics is a bridging discipline located at a critical juncture between fundamental advances in science/technology and biomedicine. Optical technologies have been playing an increasingly big role in the study of living organisms—the brain, in particular. Neurophotonics is an exploding research field that spans the intersection of light and neurons for fundamental discovery and clinical translation [1]. Neurophotonics employs a range of optical methodologies, from microscopies to spectroscopies, to achieve a multiscale understanding of the structure and function of normal and diseased brains as well as the nervous system. In particular, neurophotonics has employed photons to: (1) Interrogate the cellular processes of the nervous systems, (2) manipulate neurons to modulate function, and (3) detect diseases for clinical diagnosis and surgical guidance. This transdisciplinary field bridges the disciplines of optical physics, biochemistry, biomedical engineering, physiology, neuroscience, and neurosurgery. In 2005, Tanner et al. [2] published the first article including the term “neurophotonics” in the title. The results reported in that article were obtained by near-infrared (NIR) spectroscopy (NIRS). The discovery of this technique, now named medical NIRS, goes back to 1977 [3], when Frans Jöbsis, Professor of Physiology at Duke University (Durham, NC, USA), reported that the relatively high degree of transparency of brain tissue in the 650–900 nm NIR range (“optical window”), and the characteristic hemoglobin (Hb) absorption spectra in this wavelength region enable real-time non-invasive detection of Hb oxygenation using transillumination spectroscopy.

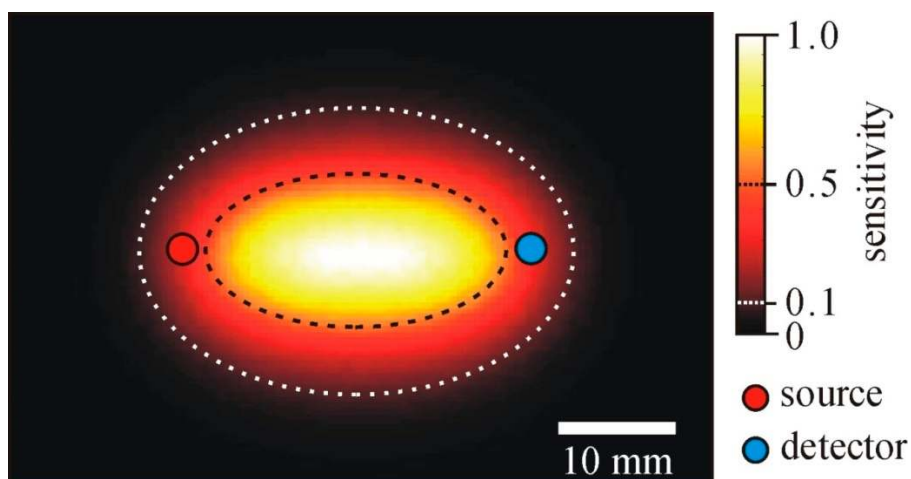
In medical NIRS measurements, the source (laser or light emitting diode) and detector probes are positioned over the scalp surface to detect the change in optical density caused by the hemodynamic changes mainly expected in the cortical grey matter [4]. Consequently, the light needs to pass through different extracranial and intracranial tissues (superficial layers, skull, cerebrospinal fluid, meninges, cortical grey matter) both before and after passing through the brain. At the end, the detected emerging NIR signal (as a result of the absorption and scattering phenomena) comes mainly from oxygenated Hb ( $O_2Hb$ ) and deoxygenated Hb (HHb) located in small vessels (<1 mm diameter). A schematic sketch representing the NIR light travelling through the different intracranial tissues is reported in Figure 1.



**Figure 1.** Schematic representation of the optical region of sensitivity (banana-shaped shaded area) in non-invasive near-infrared (NIR) studies of the human brain. The illumination and collection points (which are coupled to a light source and optical detector, respectively) are located on the scalp at a relative distance of the order of 3 cm. Light propagation is affected by the heterogeneity in the optical properties of tissues. Therefore, the sensitivity of the optical signal to the probed tissue is not spatially uniform (as indicated by the different grey levels within the region of sensitivity) and is maximal in the most superficial tissue layers (scalp and skull). Reproduced with permission from [5] © 2018 licensed under a Creative Commons Attribution (CC BY) license.

NIR photons propagate simultaneously in the entire illuminated volume of the head, and, due to multiple scattering, the photon paths have all possible shapes and lengths. The light intensity in the head cannot be non-invasively measured. Therefore, the light propagation in the head has been predicted by simulations using realistic head models and the Monte Carlo method [6]. The most meaningful way to characterize the variety of paths is to use the statistical quantities such as the mean total pathlength (typically 5–10 larger than the source–detector distance) and the partial pathlength—regions of the head, in particular. The partial pathlength in the brain of the adult subjects is small compared to the total pathlength (~10% of the total pathlength at a 3 cm separation).

It is important to obtain the sensitivity of the NIRS signal to the absorption change in the volume of sampled tissue, in particular in the cortical grey matter, with a particular source–detector pair. For this purpose, Sakakibara et al. [7], using Monte Carlo simulations and a five-layered head model, elaborated the spatial sensitivity profile on the surface of the grey matter (Figure 2). The source–detector pair detects the absorption change in the broad region in the grey matter. The sensitivity of the source–detector pair is the greatest at the measurement point, but the sensitivity decreases with an increase in the distance from the measurement point. The black and white lines in the figure indicate 50% and 10% with respect to the maximum sensitivity. The spatial distribution of the sensitivity of the probe arrangements depends on the positions of the measurement points and the direction of the spatial sensitivity profiles. Using the Monte Carlo method and the diffusion theory, several previous studies demonstrated that functional near-infrared spectroscopy (fNIRS) signals are more sensitive to the surface areas immediately under the optodes, i.e., the scalp, for review [5]. This limitation is less significant in young children, since, with thinner skull, the partial pathlength in the brain increases.



**Figure 2.** The spatial sensitivity profile on the grey matter surface. The black and white dotted lines indicate 50% and 10% with respect to the maximum sensitivity. This profile was obtained by a Monte Carlo simulation of a five-layered head model. Reproduced with permission from [7] © 2016 Springer.

In 2012, the *Journal of Near Infrared Spectroscopy Special Issue on Medical Application* nicely summarized the most important aspects of the medical NIRS in 16 articles (mainly review articles) [8,9]. Brain/muscle oximetry and functional NIRS (fNIRS) represented the most established clinical and/or basic research areas. The first brain oximeter measuring cortical Hb saturation (in %) was built in 1989 by Hamamatsu Photonics K.K. (Japan). Today, more than 10 brain oximeters with Food and Drug Administration (FDA) and/or European Union (EU) approval are commercially available and utilized worldwide mainly in cardiac surgery and neonatal intensive care units [10–13]. The present mini-review does not cover the present and future applications of oximetry. Instead, it wants to focus exclusively on fNIRS applied to different medical fields.

In the last 20 years, there have been exponential developments in the field of neuroimaging. This field includes mainly magnetic resonance imaging (MRI) and molecular imaging, and most of the changes have occurred in the latter with advances in positron emission tomography (PET). Now, it is possible to image the brain glucose consumption as well many different chemicals like dopamine, serotonin, and acetylcholine. Non-invasive vascular-based neuroimaging techniques, such as functional MRI (fMRI) and fNIRS, map brain activity through hemodynamic-based signals and are invaluable diagnostic tools in several neurological disorders. Cerebral blood flow (CBF), adequate for brain activity and metabolic demand, is maintained through the processes of neurovascular coupling. More particularly, when a specific brain region is activated, CBF increases in a temporally and spatially coordinated manner tightly linked to changes in neural activity through a complex sequence of coordinated events involving neurons, glia, arteries/arterioles, and signaling molecules. fNIRS and fMRI rely on this coupling to infer changes in neural activity that are mirrored by the changes in the blood oxygenation in the region of the activated cortical area.

fNIRS, applying an array of sources/detectors over the scalp, maps (typical sampling rate 1–10 Hz) the concomitant increase in  $O_2Hb$  and the decrease in  $HHb$  only at level of cortical microcirculation blood vessels by means of the characteristic Hb absorption spectra in the NIR range; fMRI only maps the decrease in  $HHb$  in all brain regions with a spatial resolution ten times higher than fNIRS does. fNIRS also maps the total Hb (tHb) ( $tHb = O_2Hb + HHb$ ), though this is strictly related to cerebral blood volume. The hemodynamic signals are normally precisely related to the underlying neuronal activity through neurovascular coupling mechanisms that ensure the supply of glucose and oxygen to neurons [14] but also provide a heat sink to help cool the brain and removal of waste by-products [15,16]. In addition, the neurovascular coupling plays a key role in water dynamics inside the brain barrier [17]. As described recently in detail [18,19], the fNIRS signal includes six different components that can be classified according to their: (1) Source (cerebral versus

extra-cerebral); (2) stimulus/task relation (evoked versus non-evoked); and (3) physiological cause (neuronal versus systemic). The monitoring of the hemodynamic response due to neurovascular coupling is only one of these six components (i.e., the component neuronal/task-evoked/cerebral), while all the other components are the physiological noise that acts as confounders in fNIRS studies and must be removed by different methods [5,19]. While the strict relationship between CBF and neuronal activity forms a fundamental brain function, whether neurovascular coupling mechanisms are reliable across physiological and pathological conditions is still questionable; for instance, alterations of the brain vasculature compromise neurovascular coupling. The mechanisms that are involved in the neurovascular coupling are different in health and in diseases such as psychiatric disorders and stroke. In addition, neurovascular coupling mechanisms are probably affected by changing brain states like sleep, wakefulness, and attention. Though fMRI has been clinically utilized more extensively than fNIRS, in the last decade the functional activation of the human cerebral cortex has been successfully explored by fNIRS. The latest is also named: Optical topography, NIR imaging, diffuse optical imaging (DOI), or diffuse optical tomography (DOT). Unlike fMRI, fNIRS can be utilized on subjects while moving freely in naturalistic settings (such as face to face communications), in hyper-scanning studies, and in field studies on subjects practicing sports, playing a musical instrument, etc.

The present mini-review article is aimed at briefly summarizing the current status of fNIRS and at predicting where the technique should go in the next decade.

## 2. Where Do We Stand

In order to provide the readers with an update of the fNIRS methods, in Table 1 recent relevant references (33 articles published from 2012) are reported about several topics related to the fNIRS basics and technical developments. These articles were identified through the PubMed, Web of Science, and Scopus databases. The topics include: The basics of NIR photon migration, the state of the art of instrumentations/signal processing/statistical analysis, and the integration of fNIRS with other neuroimaging methods.

**Table 1.** Most relevant references about functional near-infrared spectroscopy (fNIRS): Basics and technical developments.

Topic	Year	1st Author [Ref]
Modeling near-infrared photon propagation in biological tissue	2012	Martelli [20]
	2016	Bigio [4]
	2018	Fantini [5]
History of fNIRS	2012	Ferrari [21]
State of the art of continuous-wave multispectral fNIRS instrumentation	2014	Scholkmann [18]
	2017	Yücel [22]
State of the art of continuous-wave hyperspectral fNIRS instrumentation	2016	Nsorati [23]
	2016	Pham [24]
	2018	Giannoni [25]
State of the art of time-domain fNIRS instrumentation	2014	Torricelli [26]
	2019	Yamada [27]
Clinical brain monitoring by time-domain fNIRS instrumentation	2019	Lange [28]
State of the art of diffuse optical imaging	2016	Hoshi [29]
	2017	Lee [30]
	2018	Fantini [5]
	2018	Zhao [31]
State of the art of wearable fNIRS	2018	Strangman [32]
	2018	Pinti [33]
State of the art of functional connectivity measurements	2018	Fantini [5]

Table 1. Cont.

Topic	Year	1st Author [Ref]
Factors influencing fNIRS data and recommendations	2010	Orihuela-Espina [34]
Caps for long term fNIRS measurements	2015	Kassab [35]
Selection of the optimum source–detector distance	2015	Brigadoi [36]
Mayer waves interference	2016	Yücel [37]
Multiple components of the fNIRS signal	2016	Tachtsidis [19]
Signal pre-processing procedures	2019	Pinti [38]
Anatomical guidance for fNIRS	2014	Tsuzuki [39]
	2015	Aasted [40]
Statistical analysis of fNIRS data	2014	Tak [41]
Pattern of hemodynamic response in newborn < 1 month	2018	de Roever [42]
Pattern of hemodynamic response in infants	2018	Issard [43]
Integration of fNIRS with:		
• Electroencephalography	2017	Chiarelli [44]
• Functional magnetic resonance imaging	2017	Scarapicchia [45]
• Transcranial magnetic stimulation	2019	Curtin [46]
Recent fNIRS general reviews including the advantages and limitations of fNIRS	2018	Fantini [5]
	2018	Pinti [47]
	2019	Quaresima [48]

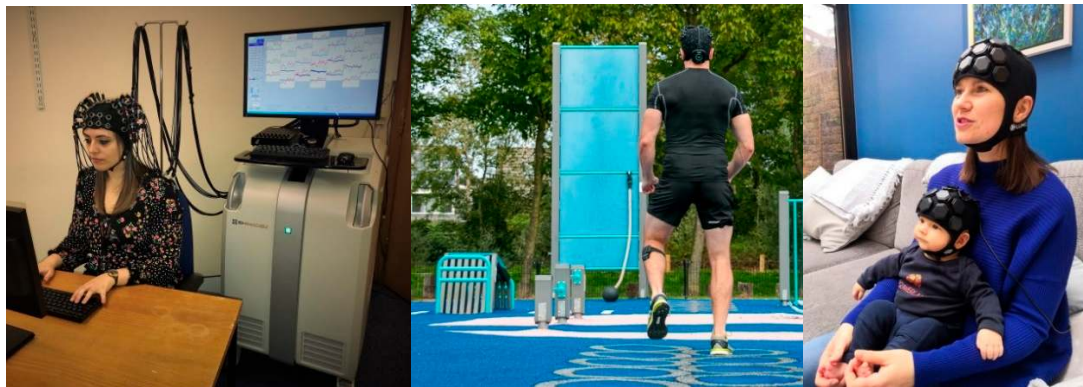
Ref: Reference number.

The advantages and disadvantages of fNIRS have been widely reported in several recent review articles [5,47,48]. Unlike other neuroimaging modalities, fNIRS has a very high experimental flexibility. fNIRS is silent, tolerant to movement artefacts, and allows for long-time continuous measurements. fNIRS can be easily integrated with fMRI, PET, electroencephalography (EEG) or event related potentials. A detailed critical comparison between fNIRS and fMRI has been recently reported [5]. Among the disadvantages, it is noteworthy to mention: (1) fNIRS does not provide anatomical information, and (2) fNIRS measurements are restricted to the outer cortex and have a low spatial resolution (2–3 cm).

Roughly twenty multi-channel fNIRS systems, which utilize arrays of multiple NIR sources and detectors arranged over the scalp, are so far commercially available [47,48]. Figure 3 shows one stationary system and two mobile wireless systems.

Multi-channel fNIRS systems utilize different NIRS techniques: (1) The continuous wave (CW) multispectral and CW hyperspectral (broadband) techniques, both based on constant tissue illumination, measuring the light attenuation; (2) the frequency-domain (FD) method, based on intensity-modulated light, measuring both the attenuation and phase delay of emerging light; and (3) the time-domain (TD) technique, based on short pulses of light, measuring the shape of the pulse after propagation through tissues [18]. The CW hyperspectral technique allows for a more accurate separation of the chromophores than the CW multispectral technique that utilizes few wavelengths [25]. The O<sub>2</sub>Hb/HHb quantitation depends on the fNIRS adopted technology [5,18]. The most commonly used CW multispectral fNIRS instrumentation measures changes of O<sub>2</sub>Hb and HHb (with respect to an initial value arbitrarily set equal to zero) that are calculated using a modification of the Lambert–Beer law. Considering that the tissue optical pathlength is longer than the distance between the source and the detector (Figure 1), the O<sub>2</sub>Hb and HHb signal changes are expressed as  $\mu\text{molar}\cdot\text{cm}$  or  $\text{mmolar}\cdot\text{mm}$ . CW multispectral systems offer the advantages of being low-cost and easily transportable (Figure 3). fNIRS analysis methods permit the monitoring of real-time cortical hemodynamic changes. fNIRS data from multiple simultaneous measurement sites are displayed by fNIRS systems in the form of O<sub>2</sub>Hb/HHb map over a cortical area.





**Figure 3.** Continuous wave (CW) multispectral fNIRS instrumentations. Left panel: Stationary system in a conventional laboratory setting (LABNIRS, Shimadzu, Japan with three wavelengths and 52 channels) (year of release in USA 2015) (Photo courtesy of University College London, Department of Medical Physics and Biomedical Engineering). Central panel: Mobile wireless system in outdoor environment (Brite<sup>24</sup>, Artinis Medical Systems, The Netherlands with two wavelengths and 24 channels) (year of release 2018) (Photo courtesy of Artinis). Right panel: Mobile wireless system (LUMO, Gowerlabs Ltd., UK) (year of release 2019) (Photo courtesy of Gowerlabs). The first commercially available modular, wearable, high-density diffuse optical tomography (DOT) system consisting of a series of hexagonal sensor modules (‘tiles’), each of which provides two wavelength LED sources and four detectors. Channels are formed both within and across tiles with source–detector separations ranging from 10 to 40 mm. By connecting multiple tiles into the LUMO head cap, users can create lightweight, high-density fNIRS imaging arrays to cover any part of the cortex. We obtained permission from photographed subjects.

In 2014, the journal *Neuroimage* dedicated a Special Issue with 58 articles to celebrate the first 20 years of fNIRS research [49]. Thus far, fNIRS has lacked the combination of spatial resolution and wide field-of-view sufficient to map in detail distributed brain functions. The emergence of high-density DOT represents the last generation of multispectral CW fNIRS systems. Figure 3 depicts an example of a high-density DOT imaging system for children and adults. High-density DOT resolves the basic problem of the contribution from hemodynamic changes occurring in the scalp, skull, and other extra-cerebral tissue layers [5].

In order to provide the readers with an update on the fNIRS applications, in Table 2, 44 recent review articles (published from 2012) covering different applications are listed; the field of psychology/education is covered by 10 reviews, functional neuroimaging basic research by 13 reviews, and medicine by 18 reviews.

**Table 2.** Main reviews on the fNIRS applications in the fields of cognitive and social sciences, functional neuroimaging research, and medicine.

Field of Application	Topic	Year	N.	Subjects	1st Author [Ref]
Psychology/education	Cognition and food	2015	39	A	Val-Laillet [50]
	Cognition in infants	2015	171	C	Aslin [51]
	Development (typical and atypical)	2014	29	C	Vanderwert [52]
		2015	149	C	Wilcox [53]
	Development of mathematics/language skills in children	2018	7	C	Soltanlou [54]
	Emotion	2016	11	A	Bendall [55]
	Influence of exercise on cognition	2018	35	A	Herold [56]
	Interhemispheric organization	2014	32	A	Homae [57]
	Psychology general review	2012	106	A	Cutini [58]
	Social development during infancy	2018	29	C	McDonald [59]

Table 2. Cont.

Field of Application	Topic	Year	N.	Subjects	1st Author [Ref]
<i>Economics</i>	Neuroeconomic research	2014	15	A	Kopton [60]
<i>Linguistics</i>	Language and its development	2012	60	A C	Quaresima [61]
	Word and sentence processing	2012	9	C	Rossi [62]
<i>Neuroergonomics</i>	Neuroergonomics and fNIRS	2018	68	A	Curtin [63]
		2019	37	A	Zhu [64]
<i>Functional Neuroimaging Basic Research</i>	Brain computer interface	2015	33	A	Naseer [65]
	Driving research	2016	10	A	Liu [66]
		2019	13	A	Lohani [67]
	Hybrid fNIRS-EEG brain-computer interfaces	2017	11	A	Ahn [68]
		2018	43	A	Hong [69]
	Hyperscanning with multi-subject measurements	2013	7	A	Scholkmann [70]
		2018	15	A	Minagawa [71]
		2018	18	A	Wang [72]
	Postural and walking tasks	2017	57	A	Herold [73]
	Resting-state functional brain connectivity	2014	16	A	Niu [74]
	Walking	2017	31	A	Vitorio [75]
		2019	35	A	Pelicioni [76]
	Walking and balance tasks in older adults	2018	24	A	Stuart [77]
	<i>Medicine</i>	Attention deficit disorder	2018	11	C
Auditory cortex plasticity after cochlear implant		2018	7	A	Basura [79]
Autism spectrum disorder		2019	15	C	Liu [80]
		2019	30	C	Zhang [81]
Cognitive aging		2017	34	A	Agbangla [82]
Developmental age attention deficit/hyperactivity disorder		2019	13	C	Grazioli [83]
Eating disorders		2015	11	A	Val-Laillet [50]
Epilepsy		2016	23	A	Peng [84]
Gait disorders		2017	12	A	Gramigna [85]
Mild cognitive impairment		2017	8	A	Beishon [86]
Neurofeedback training		2018	127	A	Ehlis [87]
Pain assessment in infants		2017	9	C	Benoit [88]
Parkinson's disease and walking balance tasks		2018	5	A	Stuart [77]
Prolonged disorder of consciousness		2018	7	A	Rupawala [89]
Psychiatry	2014	168	A	Ehlis [90]	
Robot-assisted gait training	2019	2	A	Berger [91]	
Schizophrenic disorders	2017	17	A	Kumar [92]	
Stroke therapy/recovery/rehabilitation	2019	66	A	Yang [93]	

A: Adults; C: Children; EEG: Electroencephalography; N: Number of reviewed articles; Ref: Reference number.

The total number of the articles quoted by the 44 reviews is 1675. A detailed analysis of the very different fields of applications is beyond the aim of this mini-review. It is noteworthy to mention in the last five years, there has been an increasing number of clinical studies on psychiatric disorders and basic studies using the hyper-scanning approach. Hyper-scanning, which consists of the measurement of brain activity simultaneously on two or more people, has been adopted by fNIRS for investigating

inter-personal interactions in a natural context. fNIRS, more than any other neuroimaging modality, is suitable for investigating real social interactions by using the hyper-scanning approach.

Table 3 lists five recent video articles showing different applications; these videos very carefully illustrate different studies performed in a laboratory or outdoor area utilizing stationary or mobile/wireless instrumentations.

**Table 3.** Recent fNIRS video articles.

Topic	Year	1st Author [Ref]	Device, Company, Country	Number of Channels
Brain development. Language processing study (rhyme judgment task) on primary school aged children.	2018	Jasińska [94]	LightNIRS, Shimadzu, Japan	47
Hyper-scanning. Parent–child dyads for analyzing brain-to-brain synchrony during a cooperative and a competitive computer task.	2019	Reindl [95]	ETG-4000, Hitachi, Japan	44
Motor cortex activation during different motor tasks (cycling, walking) on adults.	2014	Sukal-Moulton [96]	CW6, TechEn, Milford, MA, USA	24
Temporal cortex activation during a dance video game task revealed by fNIRS and fMRI on adults.	2015	Noah [97]	LABNIRS, Shimadzu, Japan	22
Wearable fNIRS. Real-world ecological prospective memory tasks on adults.	2015	Pinti [98]	WOT-100, NeU Corporation, Japan	16

fMRI = functional magnetic resonance imaging; Ref = reference number.

The top 10 cited articles on fNIRS [21,99–106], ranked according to their citations, account for between approximately 500 to over 1000 studies and (data from Scopus, Elsevier, Amsterdam, The Netherlands, June 2019) provide an insight into the historical developments and allows for the recognition of the important advances in the fNIRS field since 1993, the year of the first five fNIRS publications [21].

### 3. Where Should We Go?

The main question is: Might fNIRS improve people’s lives? Imagining the future of the fNIRS instrumentations and applications is quite difficult. Considering that a significant portion of the optical pathlength of the detected photons lies within the extra-cerebral tissue (Figure 1), the “vital” main requirement of all commercial fNIRS instrumentations (using different fNIRS methods) should be their capability to correct the skull/scalp blood flow/systemic effects. For this purpose, the ideal fNIRS instrumentation should be equipped with different source–detector distances that can provide the fNIRS data necessary for adopting the different strategies to disentangle the cerebral/extra-cerebral contributions of the NIRS signals. These strategies have been recently reviewed [5,19].

The sector of wearable health technology is gaining endless interest. The use of low-cost wearable monitoring devices or wearable biosensors that allow for the constant monitoring of physiological signals, such as fNIRS signals, is essential for the advancement of both the diagnosis and treatment of diseases, as well as for monitoring active life styles [32,107].

Since the first fNIRS studies in 1993, there has been a vast improvement in CW multispectral fNIRS systems. TD-fNIRS (the most quantitative methodology) is still not at its final stage; broadband or multi-wavelength laser sources and new detectors can be further miniaturized [108], and the signal to noise ratio can be consistently improved [109,110].



New developments in fNIRS technology will further allow for the monitoring, at least on newborns, of the cytochrome-c-oxidase redox state (CCO), which is also a metabolic marker of oxidative metabolism [111]. The hyperspectral CW NIR technique is currently used by some research groups to monitor in vivo human brain metabolism via measurements of the concentration changes in O<sub>2</sub>Hb, HHb and the oxidation state of CCO to achieve the quantification of cerebral metabolic activation in different situations from functional stimulation to response during oxygen-dependent conditions [23,24]. An increase of the CCO signal, typically corresponding to an increment in cerebral metabolism, was found, for instance, during the functional activation induced by simulated driving [23], the Stroop task [24] or working memory tasks [112]. Unlike the hemodynamic changes that are strongly affected by scalp blood flow changes, the changes in the CCO signal more specifically reflect the brain cortex activation. Therefore, the CCO measurement could represent an additional and more robust marker of cortical brain activation, thus allowing for the better identification of false positives and negatives [19].

The fNIRS integration with multimodal physiological monitoring and neuro-stimulation methodologies has already been demonstrated [46], but it needs to be better designed and defined. Very recently, Scholkmann et al. [113] introduced systemic-physiology-augmented functional near-infrared spectroscopy; SPA-fNIRS), which consists of a combination of fNIRS with physiological measurements. These measurements, obtained by a gas analyzer, a continuous noninvasive blood pressure monitor, and a skin conductance measuring device, can give an important integrative view because any brain stimulation could provoke systemic effects, which, in turn, could affect cerebral hemodynamics. Therefore, these effects should be investigated because the concept that cerebral hemodynamic changes are purely associated with brain activation is probably wrong, and it should be correctly revisited [113].

Considering that fNIRS has no age limitation, it is difficult to predict which would be the most useful clinical and basic science applications. Table 2 already includes some very useful clinical and basic science applications to be further investigated. The most important challenge is to improve patient care by translating the new technologies from basic science into clinical practice.

The FDA, industries and several research groups have become increasingly involved in efforts to develop international consensus standards that can facilitate the development of fNIRS devices with the potential to improve the related regulatory processes. Recommendations for conducting and reporting fNIRS findings should be also generated.

To extract and analyze the fNIRS information at single-subject level, novel methods should be conceived. Ideally, all clinical applications would require a single-subject analysis, even on-line in the case of, for example, the neurorehabilitation field. These new methods should be capable to identify the cortical circuitry and the brain function/dysfunction. For instance, several psychiatric and neurological symptoms are best explained by network-level changes rather than focal alterations. Given the broad range of related diseases and methodological variability, defining procedural clinical standards could be difficult. However, developing recommendations for patient and methodological challenges is highly desirable to move fNIRS into the clinical realm [114].

The multi-modal integrations of EEG-fNIRS seem to be promising in different fields [44]. For example, EEG-fNIRS can characterize the neurovascular coupling in the brain network dynamics induced by robot-assisted gait training [91]. In order to guide non-invasive brain stimulation protocols, a feedback of cortical activations patterns could be useful for the identifications of regions of hypo- or hyperactivity. Transcranial direct current stimulation (tDCS) is a non-invasive brain stimulation technique that involves the application of low intensity direct currents at the scalp for the modulation of central nervous system excitability [115]. tDCS is an increasingly important tool that is being used in a wide range of applications, including as a potential adjunct therapy for neurological/ psychiatric disorders. The integration of tDCS with EEG-fNIRS holds great promise for shedding light on the underlying neural mechanisms of stimulation effects [115].

A recent review article summarized the vast potential and bright future of all neuroimaging techniques [116]. Several advances in functional neuroimaging technologies offer promising opportunities to answer clinical questions and to address some of the most fundamental aspects of how the brain works. Local fluctuations in brain physiologic signals are highly correlated across brain regions organized within functional networks. Functional connectivity maps could also provide clear guidance for pre-surgical planning for the resection of brain tumors and epileptogenic lesions. In the future, such connectivity maps may allow clinicians to interrogate functionally perturbed networks controlling attention, memory, and other key cognitive domains. In addition, fNIRS will absolutely have a unique role in fields such interactive neurosciences [71], cortical activation in sport performance [117], and cortical activation during neurofeedback training [87].

Moreover, over the last 20 years, a complementary optical technique—NIR diffuse correlation spectroscopy (DCS)—has been developed for the continuous measurement of blood flow in tissue. Applications to the human brain cortex have been successfully demonstrated [118]. DCS uses the temporal fluctuations of diffusely-reflected light to quantify the motion of tissue scatterers (which are primarily red blood cells) and provides a non-invasive estimate of deep tissue microvascular blood flow. By combining oximetry and DCS flow measures, the tissue regional oxygen metabolic rate—a parameter closely linked to underlying physiology and pathological states—could finally be quantified [119,120]. Therefore, the combination of fNIRS with DCS could provide a very interesting tool for functional neuroimaging studies because it could give information about how surface/cortical blood flow changes affect the hemodynamic signals that are measured by fNIRS.

#### 4. Conclusions

fNIRS technology continues to evolve, and the nature of this approach provides distinct advantages when studying human cortical activation. Despite the current limitations that are largely isolated to a limited depth of penetration, a low spatial resolution, and strong extra-cranial interference, in our view, the feasibility and the success of applying fNIRS in some branches of medicine, neuroimaging basic research, and social sciences have been well documented. The development of fNIRS has strongly gained from the advances in microelectronics, computer technology, and optical engineering. With the advent of further miniaturization and integration such as integrated optics, wearable and even disposable fNIRS technology can be envisioned. The fNIRS systems that will emerge from these developments would further enlarge the number of fNIRS applications and make fNIRS findings more easily comparable with the other ones obtained by using other technologies. Before reaching the final goal, consisting of the use of fNIRS as a clinical tool in individual patients, fNIRS procedures need to be standardized.

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#### References

1. Hutchinson, M.R.; Stoddart, P.R.; Mahadevan-Jansen, A. Challenges and opportunities in neurophotonics discussed at the International Conference on Biophotonics. *Neurophotonics* **2018**, *5*, 040402.
2. Tanner, K.; D'Amico, E.; Kaczmarowski, A.; Kukreti, S.; Malpeli, J.; Mantulin, W.W.; Gratton, E. Spectrally resolved neurophotonics: A case report of hemodynamics and vascular components in the mammalian brain. *J. Biomed. Opt.* **2005**, *10*, 064009. [[CrossRef](#)]
3. Jöbsis, F.F. Noninvasive, infrared monitoring of cerebral and myocardial oxygen sufficiency and circulatory parameters. *Science* **1977**, *198*, 1264–1267. [[CrossRef](#)]
4. Bigio, I.J.; Fantini, S. *Quantitative Biomedical Optics Theory, Methods, and Applications*, 1st ed.; Cambridge University Press: Cambridge, UK, 2016.

5. Fantini, S.; Frederick, B.; Sassaroli, A. Perspective: Prospects of non-invasive sensing of the human brain with diffuse optical imaging. *APL Photonics* **2018**, *3*, 110901. [[CrossRef](#)]
6. Wang, L.; Jacques, S.L.; Zheng, L. MCML—Monte Carlo modeling of light transport in multi-layered tissues. *Comput. Methods Programs Biomed.* **1995**, *47*, 131–146. [[CrossRef](#)]
7. Sakakibara, Y.; Kurihara, K.; Okada, E. Evaluation of improvement of diffuse optical imaging of brain function by high-density probe arrangements and imaging algorithms. *Opt. Rev.* **2016**, *23*, 346–353. [[CrossRef](#)]
8. Ferrari, M.; Norris, K.H.; Sowa, M.G. Medical near infrared spectroscopy 35 years after the discovery. *J. Near Infrared Spectrosc.* **2012**, *20*, vii–ix. [[CrossRef](#)]
9. Ferrari, M.; Quaresima, V. Review: Near infrared brain and muscle oximetry: From the discovery to current applications. *J. Near Infrared Spectrosc.* **2012**, *20*, 1–14. [[CrossRef](#)]
10. Garvey, A.A.; Dempsey, E.M. Applications of near infrared spectroscopy in the neonate. *Curr. Opin. Pediatr.* **2018**, *30*, 209–215. [[CrossRef](#)]
11. La Cour, A.; Greisen, G.; Hyttel-Sorensen, S. In vivo validation of cerebral near-infrared spectroscopy: A review. *Neurophotonics* **2018**, *5*, 040901. [[CrossRef](#)]
12. Serraino, G.F.; Murphy, G.J. Effects of cerebral near-infrared spectroscopy on the outcome of patients undergoing cardiac surgery: A systematic review of randomised trials. *BMJ Open.* **2017**, *7*, e016613. [[CrossRef](#)]
13. Yu, Y.; Zhang, K.; Zhang, L.; Zong, H.; Meng, L.; Han, R. Cerebral near-infrared spectroscopy (NIRS) for perioperative monitoring of brain oxygenation in children and adults. *Cochrane Database Syst. Rev.* **2018**, *1*, CD010947. [[CrossRef](#)]
14. Lecrux, C.; Bourourou, M.; Hamel, E. How reliable is cerebral blood flow to map changes in neuronal activity? *Auton. Neurosci.* **2019**, *217*, 71–79. [[CrossRef](#)]
15. Shetty, P.K.; Galeffi, F.; Turner, D.A. Cellular links between neuronal activity and energy homeostasis. *Front. Pharmacol.* **2012**, *3*, 43. [[CrossRef](#)]
16. Tsao, A.; Galwaduge, P.T.; Kim, S.H.; Shaik, M.; Hillman, E.M.C. Measuring the thermodynamic effects of neurovascular coupling in the awake, behaving mouse brain. In *Biomedical Optics 2016, OSA Technical Digest*; Optical Society of America: Washington, DC, USA, 2016.
17. Nakada, T.; Kwee, I.L. Fluid dynamics inside the brain barrier: Current concept of interstitial flow, glymphatic flow, and cerebrospinal fluid circulation in the brain. *Neuroscientist* **2019**, *25*, 155–166. [[CrossRef](#)]
18. Scholkmann, F.; Kleiser, S.; Metz, A.J.; Zimmermann, R.; Pavia, J.M.; Wolf, U.; Wolf, M. A review on continuous wave functional near-infrared spectroscopy and imaging instrumentation and methodology. *Neuroimage* **2014**, *85*, 6–27. [[CrossRef](#)]
19. Tachtsidis, I.; Scholkmann, F. False positives and false negatives in functional near-infrared spectroscopy: Issues, challenges, and the way forward. *Neurophotonics* **2016**, *3*, 031405. [[CrossRef](#)]
20. Martelli, F. An ABC of near infrared photon migration in tissues: The diffusive regime of propagation. *J. Near Infrared Spectrosc.* **2012**, *20*, 29–42. [[CrossRef](#)]
21. Ferrari, M.; Quaresima, V. A brief review on the history of human functional near-infrared spectroscopy (fNIRS) development and fields of application. *Neuroimage* **2012**, *63*, 921–935. [[CrossRef](#)]
22. Yücel, M.A.; Selb, J.J.; Huppert, T.J.; Franceschini, M.A.; Boas, D.A. Functional near infrared spectroscopy: Enabling routine functional brain imaging. *Curr. Opin. Biomed. Eng.* **2017**, *4*, 78–86. [[CrossRef](#)]
23. Nosrati, R.; Vesely, K.; Schweizer, T.A.; Toronov, V. Event-related changes of the prefrontal cortex oxygen delivery and metabolism during driving measured by hyperspectral fNIRS. *Biomed. Opt. Express* **2016**, *7*, 1323–1335. [[CrossRef](#)]
24. Phan, P.; Highton, D.; Lai, J.; Smith, M.; Elwell, C.; Tachtsidis, I. Multi-channel multi-distance broadband near-infrared spectroscopy system to measure the spatial response of cellular oxygen metabolism and tissue oxygenation. *Biomed. Opt. Express* **2016**, *7*, 4424–4440. [[CrossRef](#)]
25. Giannoni, L.; Lange, F.; Tachtsidis, I. Hyperspectral imaging solutions for brain tissue metabolic and hemodynamic monitoring: Past, current and future developments. *J. Opt.* **2018**, *20*, 044009. [[CrossRef](#)]
26. Torricelli, A.; Contini, D.; Pifferi, A.; Caffini, M.; Re, R.; Zucchelli, L.; Spinelli, L. Time domain functional NIRS imaging for human brain mapping. *Neuroimage* **2014**, *85*, 28–50. [[CrossRef](#)]
27. Yamada, Y.; Suzuki, H.; Yamashita, Y. Time-domain near-infrared spectroscopy and imaging: A review. *Appl. Sci.* **2019**, *9*, 1127. [[CrossRef](#)]

28. Lange, F.; Tachtsidis, I. Clinical brain monitoring with time domain NIRS: A review and future perspectives. *Appl. Sci.* **2019**, *9*, 1612. [[CrossRef](#)]
29. Hoshi, Y.; Yamada, Y. Overview of diffuse optical tomography and its clinical applications. *J. Biomed. Opt.* **2016**, *21*, 091312. [[CrossRef](#)]
30. Lee, C.W.; Cooper, R.J.; Austin, T. Diffuse optical tomography to investigate the newborn brain. *Pediatr. Res.* **2017**, *82*, 376–386. [[CrossRef](#)]
31. Zhao, H.; Cooper, R.J. Review of recent progress toward a fiberless, whole-scalp diffuse optical tomography system. *Neurophotonics* **2018**, *5*, 011012. [[CrossRef](#)]
32. Strangman, G.E.; Ivkovic, V.; Zhang, Q. Wearable brain imaging with multimodal physiological monitoring. *J. Appl. Physiol.* **2018**, *124*, 564–572. [[CrossRef](#)]
33. Pinti, P.; Aichelburg, C.; Gilbert, S.; Hamilton, A.; Hirsch, J.; Burgess, P.; Tachtsidis, I. A review on the use of wearable functional near-infrared spectroscopy in naturalistic environments. *Jpn. Psychol. Res.* **2018**, *60*, 347–373. [[CrossRef](#)]
34. Orihuela-Espina, F.; Leff, D.R.; James, D.R.C.; Darzi, A.W.; Yang, G.Z. Quality control and assurance in functional near infrared spectroscopy (fNIRS) experimentation. *Phys. Med. Biol.* **2010**, *55*, 3701–3724. [[CrossRef](#)]
35. Kassab, A.; Le Lan, J.; Vannasing, P.; Sawan, M. Functional near-infrared spectroscopy caps for brain activity monitoring: A review. *Appl. Opt.* **2015**, *54*, 576–586. [[CrossRef](#)]
36. Brigadoi, S.; Cooper, R. How short is short? Optimum source–detector distance for short-separation channels in functional near-infrared spectroscopy. *Neurophotonics* **2015**, *2*, 025005. [[CrossRef](#)]
37. Yücel, M.A.; Selb, J.; Aasted, C.M.; Lin, P.Y.; Borsook, D.; Becerra, L.; Boas, D.A. Mayer waves reduce the accuracy of estimated hemodynamic response functions in functional near-infrared spectroscopy. *Biomed. Opt. Express* **2016**, *7*, 3078–3088. [[CrossRef](#)]
38. Pinti, P.; Scholkmann, F.; Hamilton, A.; Burgess, P.; Tachtsidis, I. Current status and issues regarding pre-processing of fNIRS neuroimaging data: An investigation of diverse signal filtering methods within a general linear model framework. *Front. Hum. Neurosci.* **2019**, *12*, 505. [[CrossRef](#)]
39. Tsuzuki, D.; Dan, I. Spatial registration for functional near-infrared spectroscopy: From channel position on the scalp to cortical location in individual and group analyses. *Neuroimage* **2014**, *85*, 92–103. [[CrossRef](#)]
40. Aasted, C.M.; Yücel, M.A.; Cooper, R.J.; Dobb, J.; Tsuzuki, D.; Becerra, L.; Petkov, M.P.; Borsook, D.; Dan, I.; Boas, D.A. Anatomical guidance for functional near-infrared spectroscopy: AtlasViewer tutorial. *Neurophotonics* **2015**, *2*, 020801. [[CrossRef](#)]
41. Tak, S.; Ye, J.C. Statistical analysis of fNIRS data: A comprehensive review. *Neuroimage* **2014**, *85*, 72–91. [[CrossRef](#)]
42. De Roeve, I.; Bale, G.; Mitra, S.; Meek, J.; Robertson, N.J.; Tachtsidis, I. Investigation of the pattern of the hemodynamic response as measured by functional near-infrared spectroscopy (fNIRS) studies in newborns, less than a month old: A systematic review. *Front. Hum. Neurosci.* **2018**, *12*, 371. [[CrossRef](#)]
43. Issard, C.; Gervain, J. Variability of the hemodynamic response in infants: Influence of experimental design and stimulus complexity. *Dev. Cognit. Neurosci.* **2018**, *33*, 182–193. [[CrossRef](#)]
44. Chiarelli, A.M.; Zappasodi, F.; Di Pompeo, F.; Merla, A. Simultaneous functional near-infrared spectroscopy and electroencephalography for monitoring of human brain activity and oxygenation: A review. *Neurophotonics* **2017**, *4*, 041411. [[CrossRef](#)]
45. Scarapicchia, V.; Brown, C.; Mayo, C.; Gawryluk, J.R. Functional magnetic resonance imaging and functional near-infrared spectroscopy: Insights from combined recording studies. *Front. Hum. Neurosci.* **2017**, *11*, 419. [[CrossRef](#)]
46. Curtin, A.; Tong, S.; Sun, J.; Wang, J.; Onaral, B.; Ayaz, H. A systematic review of integrated functional near-infrared spectroscopy (fNIRS) and transcranial magnetic stimulation (TMS) studies. *Front. Hum. Neurosci.* **2019**, *13*, 84. [[CrossRef](#)]
47. Pinti, P.; Tachtsidis, I.; Hamilton, A.; Hirsch, J.; Aichelburg, C.; Gilbert, S.; Burgess, P.W. The present and future use of functional near-infrared spectroscopy (fNIRS) for cognitive neuroscience. *Ann. N. Y. Acad. Sci.* **2018**. [[CrossRef](#)]
48. Quaresima, V.; Ferrari, M. Functional near-infrared spectroscopy (fNIRS) for assessing cerebral cortex function during human behavior in natural/social situations: A concise review. *Organ. Res. Methods* **2019**, *22*, 46–68. [[CrossRef](#)]

49. Boas, D.A.; Elwell, C.E.; Ferrari, M.; Taga, G. Twenty years of functional near-infrared spectroscopy: Introduction for the special issue. *Neuroimage* **2014**, *85*, 1–5. [[CrossRef](#)]
50. Val-Laillet, D.; Aarts, E.; Weber, B.; Ferrari, M.; Quaresima, V.; Stoeckel, L.E.; Alonso-Alonso, M.; Audette, M.; Malbert, C.H.; Stice, E. Neuroimaging and neuromodulation approaches to study eating behavior and prevent and treat eating disorders and obesity. *Neuroimage Clin.* **2015**, *8*, 1–31. [[CrossRef](#)]
51. Aslin, R.N.; Shukla, M.; Emberson, L.L. Hemodynamic correlates of cognition in human infants. *Annu. Rev. Psychol.* **2015**, *66*, 349–379. [[CrossRef](#)]
52. Vanderwert, R.E.; Nelson, C.A. The use of near-infrared spectroscopy in the study of typical and atypical development. *Neuroimage* **2014**, *85*, 264–271. [[CrossRef](#)]
53. Wilcox, T.; Biondi, M. fNIRS in the developmental sciences. *Wiley Interdiscip. Rev. Cognit. Sci.* **2015**, *6*, 263–283. [[CrossRef](#)]
54. Soltanlou, M.; Sitnikova, M.A.; Nuerk, H.C.; Dresler, T. Applications of functional near-infrared spectroscopy (fNIRS) in studying cognitive development: The case of mathematics and language. *Front. Psychol.* **2018**, *9*, 277. [[CrossRef](#)]
55. Bendall, R.C.; Eachus, P.; Thompson, C. A brief review of research using near-infrared spectroscopy to measure activation of the prefrontal cortex during emotional processing: The importance of experimental design. *Front. Hum. Neurosci.* **2016**, *10*, 529. [[CrossRef](#)]
56. Herold, F.; Wiegel, P.; Scholkmann, F.; Müller, N.G. Applications of functional near-infrared spectroscopy (fNIRS) neuroimaging in exercise–cognition science: A systematic, methodology-focused review. *J. Clin. Med.* **2018**, *7*, 466. [[CrossRef](#)]
57. Homae, F. A brain of two halves: Insights into interhemispheric organization provided by near-infrared spectroscopy. *Neuroimage* **2014**, *85*, 354–362. [[CrossRef](#)]
58. Cutini, S.; Basso Moro, S.; Bisconti, S. Functional near infrared optical imaging in cognitive neuroscience: An introductory review. *J. Near Infrared Spectrosc.* **2012**, *20*, 75–92. [[CrossRef](#)]
59. McDonald, N.M.; Perdue, K.L. The infant brain in the social world: Moving toward interactive social neuroscience with functional near-infrared spectroscopy. *Neurosci. Biobehav. Rev.* **2018**, *87*, 38–49. [[CrossRef](#)]
60. Kopton, I.M.; Kenning, P. Near-infrared spectroscopy (NIRS) as a new tool for neuroeconomic research. *Front. Hum. Neurosci.* **2014**, *8*, 549. [[CrossRef](#)]
61. Quaresima, V.; Bisconti, S.; Ferrari, M. A brief review on the use of functional near-infrared spectroscopy (fNIRS) for language imaging studies in human newborns and adults. *Brain Lang.* **2012**, *121*, 79–89. [[CrossRef](#)]
62. Rossi, S.; Telkemeyer, S.; Wartenburger, I.; Obrig, H. Shedding light on words and sentences: Near-infrared spectroscopy in language research. *Brain Lang.* **2012**, *121*, 152–163. [[CrossRef](#)]
63. Curtin, A.; Ayaz, H. The age of neuroergonomics: Towards ubiquitous and continuous measurement of brain function with fNIRS. *Jpn. Psychol. Res.* **2018**, *60*, 374–386. [[CrossRef](#)]
64. Zhu, Y.; Rodriguez-Paras, C.; Rhee, J.; Mehta, R.K. Methodological approaches and recommendations for functional near-infrared spectroscopy applications in HF/E research. *Hum. Factors* **2019**. [[CrossRef](#)]
65. Naseer, N.; Hong, K.S. fNIRS-based brain-computer interfaces: A review. *Front. Hum. Neurosci.* **2015**, *9*, 3. [[CrossRef](#)]
66. Liu, T.; Pelowski, M.; Pang, C.; Zhou, Y.; Cai, J. Near-infrared spectroscopy as a tool for driving research. *Ergonomics* **2016**, *59*, 368–379. [[CrossRef](#)]
67. Lohani, M.; Payne, B.R.; Strayer, D.L. A review of psychophysiological measures to assess cognitive states in real-world driving. *Front. Hum. Neurosci.* **2019**, *13*, 57. [[CrossRef](#)]
68. Ahn, S.; Jun, S.C. Multi-modal integration of EEG-fNIRS for brain-computer interfaces—Current limitations and future directions. *Front. Hum. Neurosci.* **2017**, *11*, 503. [[CrossRef](#)]
69. Hong, K.S.; Khan, M.J.; Hong, M.J. Feature extraction and classification methods for hybrid fNIRS-EEG brain-computer interfaces. *Front. Hum. Neurosci.* **2018**, *12*, 246. [[CrossRef](#)]
70. Scholkmann, F.; Holper, L.; Wolf, U.; Wolf, M. A new methodical approach in neuroscience: Assessing inter-personal brain coupling using functional near-infrared imaging (fNIRI) hyperscanning. *Front. Hum. Neurosci.* **2013**, *7*, 813. [[CrossRef](#)]
71. Minagawa, Y.; Xu, M.; Morimoto, S. Toward interactive social neuroscience: Neuroimaging real-world interactions in various populations. *Jpn. Psychol. Res.* **2018**, *60*, 374–386. [[CrossRef](#)]



72. Wang, M.Y.; Luan, P.; Zhang, J.; Xiang, Y.T.; Niu, H.; Yuan, Z. Concurrent mapping of brain activation from multiple subjects during social interaction by hyperscanning: A mini-review. *Quant. Imaging Med. Surg.* **2018**, *8*, 819–837. [[CrossRef](#)]
73. Herold, F.; Wiegel, P.; Scholkmann, F.; Thiers, A.; Hamacher, D.; Schega, L. Functional near-infrared spectroscopy in movement science: A systematic review on cortical activity in postural and walking tasks. *Neurophotonics* **2017**, *4*, 041403. [[CrossRef](#)]
74. Niu, H.; He, Y. Resting-state functional brain connectivity: Lessons from functional near-infrared spectroscopy. *Neuroscientist* **2014**, *20*, 173–188. [[CrossRef](#)]
75. Vitorio, R.; Stuart, S.; Rochester, L.; Alcock, L.; Pantall, A. Fnirs response during walking - Artefact or cortical activity? A systematic review. *Neurosci. Biobehav. Rev.* **2017**, *83*, 160–172. [[CrossRef](#)]
76. Pelicioni, P.H.S.; Tijmsa, M.; Lord, S.R.; Menant, J. Prefrontal cortical activation measured by fNIRS during walking: Effects of age, disease and secondary task. *Peer J.* **2019**, *7*, e6833. [[CrossRef](#)]
77. Stuart, S.; Vitorio, R.; Morris, R.; Martini, D.N.; Fino, P.C.; Mancini, M. Cortical activity during walking and balance tasks in older adults and in people with Parkinson's disease: A structured review. *Maturitas* **2018**, *113*, 53–72. [[CrossRef](#)]
78. Mauri, M.; Nobile, M.; Bellina, M.; Crippa, A.; Brambilla, P. Light up ADHD: I. Cortical hemodynamic responses measured by functional near infrared spectroscopy (fNIRS). *J. Affect. Disord.* **2018**, *234*, 358–364. [[CrossRef](#)]
79. Basura, G.J.; Hu, X.S.; Juan, J.S.; Tessier, A.M.; Kovelman, I. Human central auditory plasticity: A review of functional near-infrared spectroscopy (fNIRS) to measure cochlear implant performance and tinnitus perception. *Laryngoscope Investig. Otolaryngol.* **2018**, *3*, 463–472. [[CrossRef](#)]
80. Liu, T.; Liu, X.; Yi, L.; Zhu, C.; Markey, P.S.; Pelowski, M. Assessing autism at its social and developmental roots: A review of autism spectrum disorder studies using functional near-infrared spectroscopy. *Neuroimage* **2019**, *185*, 955–967. [[CrossRef](#)]
81. Zhang, F.; Roeyers, H. Exploring brain functions in autism spectrum disorder: A systematic review on functional near-infrared spectroscopy (fNIRS) studies. *Int. J. Psychophysiol.* **2019**, *137*, 41–53. [[CrossRef](#)]
82. Agbangla, N.F.; Audiffren, M.; Albinet, C.T. Use of near-infrared spectroscopy in the investigation of brain activation during cognitive aging: A systematic review of an emerging area of research. *Ageing Res. Rev.* **2017**, *38*, 52–66. [[CrossRef](#)]
83. Grazioli, S.; Mauri, M.; Crippa, A.; Maggioni, E.; Molteni, M.; Brambilla, P.; Nobile, M. Light up ADHD: II. Neuropharmacological effects measured by near infrared spectroscopy: Is there a biomarker? *J. Affect. Disord.* **2019**, *244*, 100–106. [[CrossRef](#)]
84. Peng, K.; Pouliot, P.; Lesage, F.; Nguyen, D.K. Multichannel continuous electroencephalography-functional near-infrared spectroscopy recording of focal seizures and interictal epileptiform discharges in human epilepsy: A review. *Neurophotonics* **2016**, *3*, 031402. [[CrossRef](#)]
85. Gramigna, V.; Pellegrino, G.; Cerasa, A.; Cutini, S.; Vasta, R.; Olivadese, G.; Martino, I.; Quattrone, A. Near-infrared spectroscopy in gait disorders: Is it time to begin? *Neurorehabil. Neural Repair* **2017**, *31*, 402–412. [[CrossRef](#)]
86. Beishon, L.; Haunton, V.J.; Panerai, R.B.; Robinson, T.G. Cerebral hemodynamics in mild cognitive impairment: A systematic review. *J. Alzheimers Dis.* **2017**, *59*, 369–385. [[CrossRef](#)]
87. Ehlis, A.; Barth, B.; Hudak, J.; Storchak, H.; Weber, L.; Kimmig, A.S.; Kreifelts, B.; Dresler, T.; Fallgatter, A.J. Near-infrared spectroscopy as a new tool for neurofeedback training: Applications in psychiatry and methodological considerations. *Jpn. Psychol. Res.* **2018**, *60*, 225–241. [[CrossRef](#)]
88. Benoit, B.; Martin-Misener, R.; Newman, A.; Latimer, M.; Campbell-Yeo, M. Neurophysiological assessment of acute pain in infants: A scoping review of research methods. *Acta Paediatr.* **2017**, *106*, 1053–1066. [[CrossRef](#)]
89. Rupawala, M.; Dehghani, H.; Lucas, S.J.E.; Tino, P.; Cruse, D. Shining a light on awareness: A review of functional near-infrared spectroscopy for prolonged disorders of consciousness. *Front. Neurol.* **2018**, *9*, 350. [[CrossRef](#)]
90. Ehlis, A.C.; Schneider, S.; Dresler, T.; Fallgatter, A.J. Application of functional near-infrared spectroscopy in psychiatry. *Neuroimage* **2014**, *85*, 478–488. [[CrossRef](#)]
91. Berger, A.; Horst, F.; Müller, S.; Steinberg, F.; Doppelmayr, M. Current state and future prospects of EEG and fNIRS in robot-assisted gait rehabilitation: A brief review. *Front. Hum. Neurosci.* **2019**, *13*, 172. [[CrossRef](#)]

92. Kumar, V.; Shivakumar, V.; Chhabra, H.; Bose, A.; Venkatasubramanian, G.; Gangadhar, B.N. Functional near infra-red spectroscopy (fNIRS) in schizophrenia: A review. *Asian J. Psychiatr.* **2017**, *27*, 18–31. [[CrossRef](#)]
93. Yang, M.; Yang, Z.; Yuan, T.; Feng, W.; Wang, P. A systemic review of functional near-infrared spectroscopy for stroke: Current application and future directions. *Front. Neurol.* **2019**, *10*, 58. [[CrossRef](#)]
94. Jasińska, K.K.; Guei, S. Neuroimaging field methods using functional near infrared spectroscopy (NIRS) neuroimaging to study global child development: Rural sub-saharan africa. *J. Vis. Exp.* **2018**, *132*, e57165. [[CrossRef](#)]
95. Reindl, V.; Konrad, K.; Gerloff, C.; Kruppa, J.A.; Bell, L.; Scharke, W. Conducting hyperscanning experiments with functional near-infrared spectroscopy. *J. Vis. Exp.* **2019**, *143*, e58807. [[CrossRef](#)]
96. Sukal-Moulton, T.; de Campos, A.C.; Stanley, C.J.; Damiano, D.L. Functional near infrared spectroscopy of the sensory and motor brain regions with simultaneous kinematic and EMG monitoring during motor tasks. *J. Vis. Exp.* **2014**, *94*, e52391. [[CrossRef](#)]
97. Noah, J.A.; Ono, Y.; Nomoto, Y.; Shimada, S.; Tachibana, A.; Zhang, X.; Bronner, S.; Hirsch, J. Fmri validation of fNIRS measurements during a naturalistic task. *J. Vis. Exp.* **2015**, *100*, e52116. [[CrossRef](#)]
98. Pinti, P.; Aichelburg, C.; Lind, F.; Power, S.; Swingler, E.; Merla, A.; Hamilton, A.; Gilbert, S.; Burgess, P.; Tachtsidis, I. Using fiberless, wearable fNIRS to monitor brain activity in real-world cognitive tasks. *J. Vis. Exp.* **2015**, *106*, e53336. [[CrossRef](#)]
99. Villringer, A.; Chance, B. Non-invasive optical spectroscopy and imaging of human brain function. *Trends Neurosci.* **1997**, *20*, 435–442. [[CrossRef](#)]
100. Villringer, A.; Planck, J.; Hock, C.; Schleinkofer, L.; Dirnagl, U. Near infrared spectroscopy (NIRS): A new tool to study hemodynamic changes during activation of brain function in human adults. *Neurosci. Lett.* **1993**, *154*, 101–104. [[CrossRef](#)]
101. Strangman, G.; Culver, J.P.; Thompson, J.H.; Boas, D.A. A quantitative comparison of simultaneous BOLD fMRI and NIRS recordings during functional brain activation. *Neuroimage* **2002**, *17*, 719–731. [[CrossRef](#)]
102. Obrig, H.; Villringer, A. Beyond the visible—Imaging the human brain with light. *J. Cereb. Blood Flow Metab.* **2003**, *23*, 1–18. [[CrossRef](#)]
103. Maki, A.; Yamashita, Y.; Ito, Y.; Watanabe, E.; Mayanagi, Y.; Koizumi, H. Spatial and temporal analysis of human motor activity using noninvasive NIR topography. *Med. Phys.* **1995**, *22*, 1997–2005. [[CrossRef](#)]
104. Strangman, G.; Boas, D.A.; Sutton, J.P. Non-invasive neuroimaging using near-infrared light. *Biol. Psychiatry* **2002**, *52*, 679–693. [[CrossRef](#)]
105. Boas, D.A.; Dale, A.M.; Franceschini, M.A. Diffuse optical imaging of brain activation: Approaches to optimizing image sensitivity, resolution, and accuracy. *Neuroimage* **2004**, *23*, S275–S288. [[CrossRef](#)]
106. Hoshi, Y.; Kobayashi, N.; Tamura, M. Interpretation of near-infrared spectroscopy signals: A study with a newly developed perfused rat brain model. *J. Appl. Physiol.* **2001**, *90*, 1657–1662. [[CrossRef](#)]
107. Ward, J.; Pinti, P. Wearables and the brain. *IEEE Pervasive Comput.* **2019**, *18*, 94–100. [[CrossRef](#)]
108. Buttafava, M.; Martinenghi, E.; Tamborini, D.; Contini, D.; Mora, A.D.; Renna, M.; Torricelli, A.; Pifferi, A.; Zappa, F.; Tosi, A. A compact two-wavelength time-domain NIRS system based on SiPM and pulsed diode lasers. *IEEE Photonics J.* **2017**, *9*, 7792611. [[CrossRef](#)]
109. Di Sieno, L.; Dalla Mora, A.; Torricelli, A.; Spinelli, L.; Re, R.; Pifferi, A.; Contini, D. A versatile setup for time-resolved functional near infrared spectroscopy based on fast-gated single-photon avalanche diode and on four-wave mixing laser. *Appl. Sci.* **2019**, *9*, 2366. [[CrossRef](#)]
110. Lange, F.; Dunne, F.; Hale, L.; Tachtsidis, I. Maestros: A multiwavelength time-domain NIRS system to monitor changes in oxygenation and oxidation state of cytochrome-c-oxidase. *IEEE J. Sel. Top. Quant. Electron.* **2019**, *25*, 1–12. [[CrossRef](#)]
111. Bale, G.; Elwell, C.E.; Tachtsidis, I. From Jöbsis to the present day: A review of clinical near-infrared spectroscopy measurements of cerebral cytochrome-c-oxidase. *J. Biomed. Opt.* **2016**, *21*, 091307. [[CrossRef](#)]
112. De Roeve, I.; Bale, G.; Cooper, R.J.; Tachtsidis, I. Functional NIRS measurement of cytochrome-c-oxidase demonstrates a more brain-specific marker of frontal lobe activation compared to the haemoglobins. *Adv. Exp. Med. Biol.* **2017**, *977*, 141–147.
113. Scholkmann, F.; Hafner, T.; Metz, A.J.; Wolf, M.; Wolf, U. Effect of short-term colored-light exposure on cerebral hemodynamics and oxygenation, and systemic physiological activity. *Neurophotonics* **2017**, *4*, 045005. [[CrossRef](#)]

114. Beisteiner, R.; Pernet, C.; Stippich, C. Can we standardize clinical functional neuroimaging procedures? *Front. Neurol.* **2019**, *9*, 1153. [[CrossRef](#)]
115. Woods, A.J.; Bikson, M.; Chelette, K.; Dmochowski, J.; Dutta, A.; Esmaeilpour, Z.; Gebodh, N.; Nitsche, M.A.; Stagg, C. Transcranial direct current stimulation integration with magnetic resonance imaging, magnetic resonance spectroscopy, near infrared spectroscopy imaging and electroencephalography. In *Practical Guide to Transcranial Direct Current Stimulation*, 1st ed.; Knotkova, H., Nitsche, M., Bikson, M., Woods, A., Eds.; Springer International Publisher: Cham, Switzerland, 2019; pp. 293–345.
116. Wintermark, M.; Colen, R.; Whitlow, C.T.; Zaharchuk, G. The vast potential and bright future of neuroimaging. *Br. J. Radiol.* **2018**, *91*, 20170505. [[CrossRef](#)]
117. Perrey, S.; Besson, P. Studying brain activity in sports performance: Contributions and issues. *Prog. Brain Res.* **2018**, *240*, 247–267.
118. Durduran, T.; Yodh, A.G. Diffuse correlation spectroscopy for non-invasive, micro-vascular cerebral blood flow measurement. *Neuroimage* **2014**, *15*, 51–63. [[CrossRef](#)]
119. Roche-Labarbe, N.; Carp, S.A.; Surova, A.; Patel, M.; Boas, D.A.; Grant, P.E.; Franceschini, M.A. Noninvasive optical measures of CBV, StO<sub>2</sub>, CBF index, and rCMRO<sub>2</sub> in human premature neonates' brains in the first six weeks of life. *Hum. Brain Mapp.* **2010**, *31*, 341–352. [[CrossRef](#)]
120. Andresen, B.; De Carli, A.; Fumagalli, M.; Giovannella, M.; Durduran, T.; Michael Weigel, U.; Contini, D.; Spinelli, L.; Torricelli, A.; Greisen, G. Cerebral oxygenation and blood flow in normal term infants at rest measured by a hybrid near-infrared device (BabyLux). *Pediatr. Res.* **2019**. [[CrossRef](#)]



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