

## A novel antioxidant and antimicrobial compound produced by *Bacillus firmicutes*.

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### Abstract

In this study, antioxidant and antimicrobial compounds were produced from a dates seeds powder containing medium by *B. firmicutes*, a bacterial strain isolated from Al-Ain Alhara hot springs located in Gazan area in Saudi Arabia. The Total Antioxidant Capacity (TAC) of culture supernatant of *B. firmicutes* by the phosphomolybdenum was 2.5 mg/ml of culture medium, the total phenolic compounds is 5.3 mg/ml of culture medium and radical scavenging activity was done by DPPH. The greatest scavenging rate of antioxidants was 60%. In addition, its antibacterial activity was performed after treatment the culture medium with methanol or ethyl acetate. As antimicrobial ethyl acetate, extract is more effective than methanol. In both ethyl acetate and methanol extract the highest antibacterial activity was observed against *E. faecalis* with a maximum diameter inhibition zone of 25 mm, 15 mm respectively while, the lowest activity was detected against *P. aeruginosa* with inhibition diameter zone 14 mm and 10 mm respectively. Analysis by Thin Layer Chromatography (TLC) for ethyl acetate and methanol extracts revealed the presence one spot in the front in ethyl acetate while the spot of methanol come latter and maybe have another spot. The molecular mass of ethyl acetate extract was 304.53 m/z and methanol extract was 701.57 m/z as determined by MS.

**Keywords:** Antibacterial, Antioxidant, *Bacillus firmicutes*, Dates seed.

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### Introduction

The date seeds considered a waste product of many date processing plants producing pitted dates, date syrup and date confectionery. At present, seeds are used mainly for animal feeds in the cattle, sheep, and camel and poultry industries. Although, these date seed contain about 82% carbohydrates as reported by Besbes et al. [1] a little attention has been paid to investigate microbial conversion of date seeds to produce bioactive compounds such as antibiotics, mycotoxins, alkaloids, food grade pigments, plant growth factors, and antioxidant compounds [2]. Abou-Zeid et al. [3] was used date seed lipid and hydrolysate for production tetracycline.

Nowadays, to confront the growing antimicrobial resistances, modern medicine Focus on natural products for novel antibiotics and antimicrobials. The multifarious antimicrobial compounds produced by various *Bacillus* strains have the ability to address the multidrug resistance problems. *Bacillus* isolates are rather well known for the production of a vast array of structurally unrelated antimicrobial compounds, which include lipopeptides such as iturin, surfactin, fengycins,

bacteriocins, and bacteriocin such as inhibitory substances (BLIS) [4]. Moreover, these species produce other bioactive molecules such as catecholic siderophore bacillibactin [5]. Sequestration can reduce the occurrence of free radicals [6]. Free radicals or reactive oxygen species are known to cause oxidative damage to biological macromolecules, leading to a number of disorders including cancer, atherosclerosis, cardiovascular diseases, aging and inflammatory diseases [7]. Synthetic antioxidants that have been extensively used in industrial processing are being investigated for their toxic and carcinogenic effects [6,8]. Recently, the interest in finding natural antioxidant agents with low cytotoxicity has increased significantly [8]. Several studies have focused on plant compounds [9,10]. However, only a few reports have been conducted on the antioxidant power of microbial extracts [6]. Kumar et al. [11] reported that several bacteria were found to produce antioxidant compound as *Bacillus simplex*, *Bacillus natto*, *Bacillus cereus*, *Lactobacillus dextranicum*, *Micrococcus freudenreichii* and *Sarcina lutea*. Besides bacteria, some fungi, few Actinomycetes and yeasts were also found to produce the compounds with antioxidant activity. Dey

et al. [12] extracted antioxidant phenolic compounds from *Rhizopus oryzae* fungi cultured on wheat solid medium. Matsuo [13] observed that the oxidation of vegetable oil was inhibited by an addition of soybean-curd refuse fermented with *A. oryzae* or *Rhizopus oligosporus*. Lertcanawanichakul et al. [14] extracted A2 antioxidant and antimicrobial compounds from *Streptomyces lydicus*. *Streptomyces spp.* SRDP-H03 and BI244 exhibited antioxidant activity [15].

The present study addressed partial characterization of new antioxidant compound along with an evaluation of its antibacterial produced from *B. firmicutes*.

## Materials and Methods

### Materials

Maddina aqgwa dates pits powder was obtained from local market in Maddina. Other chemicals used were obtained in analytical grade. Microorganism was isolated from two thermal hot springs located in Gazan area in Saudi Arabia.

### Media

**Mineral dates seed powder medium (MDPM) (g/l):** NH<sub>4</sub>Cl, 1; K<sub>2</sub>HPO<sub>4</sub>, 0.5; KH<sub>2</sub>PO<sub>4</sub>, 1; MgSO<sub>4</sub>.7H<sub>2</sub>O, 0.2; Dates seed powder, 5. pH was adjusted to pH 7.0-20 g/L agar was added in solid medium (MDPMA).

**Maintenance medium (g/l):** Meat extract, 10, peptone, 10; NaCl, 5, dates seed powder 10, agar, 20. pH adjusted at 7. The slants were inoculated with bacterial strain and incubated at 37°C for 24 h. The slants were maintained at 0-4°C in refrigerators.

**Inoculum medium (g/l):** *Bacillus*

**Basal medium:** Unless stated otherwise the bacterium will be grown in shake flasks in a basal medium of the following composition: (g/l) 2.5 KH<sub>2</sub>PO<sub>4</sub>; 2.5 Na<sub>2</sub>HPO<sub>4</sub>; 1 NaCl; 2 NaNO<sub>3</sub>; 2 yeast extract; 0.05 MgSO<sub>4</sub>.7H<sub>2</sub>O and 0.075 CaCl<sub>2</sub>. The basal medium will be supplemented with 10 g dates seed powder. The pH of the medium will be adjusted to pH 7.0. The basal medium was sterilized by autoclaving at 121°C for 15 min. Erlenmeyer flasks (250 ml) containing 50 ml of basal medium were inoculated at 10% (v/v) level and incubated at 37°C on a rotary shaker (150 rpm) for 48 h. Samples were removed at intervals for the determination of growth of cells and antioxidant concentration.

**Inoculum medium nutrient broth medium:** Peptone-5 g/L meat extract-1 g/L yeast extract-2 g/L sodium chloride-5 g/L, pH-7.0 ± 0.2 nutrient agar for maintenance bacteria, the same nutrient broth in addition to 20 g/L agar.

**Müller-Hinton agar:** It is typically contains: 2.0 g/L beef extract, 17.5 g/L casein hydrolysate, 1.5 g/L starch, 17.0 g/L agar dissolved in 1 litre of distilled water. pH was adjusted to neutral at 25°C [16]. Müller-Hinton broth the same medium without agar.

### Microorganisms

The tested microorganisms included the following:

Gram-positive bacteria; *Enterococcus faecalis* ATTC (292121); *Staphylococcus aureus* ATTC (29213); Gram-negative bacteria; *Escherichia coli* ATCC (25922); *Klebsiella pneumoniae* ATTC (70060); *Proteus mirabilis* ATTC (14153); *Pseudomonas aeruginosa* (27853); These strains were obtained from King Abdul-Aziz University Hospital.

### Methods

#### Isolation of *Bacillus firmicutes*

Water samples were collected from two thermal hot springs located in Gazan area in Saudi Arabia, these are Al-Ain Alhara (or Al-Khawaba) located at a distance of about 50 km southeast of Gazan city.

Samples were collected in 500 ml sterile thermal glass containers, which keep the temperature of the water samples constant. 5 ml from each water sample was inoculated to 500 ml Erlenmeyer flask containing 50 ml (MDPM) medium.

The inoculated flasks will be incubated at 37°C with shaking at 180 rpm for 48 h millilitre of cultured flask were striking on petri dish containing (MDPMA) media, then incubated at 37°C in a bench top incubator for 24 to 48 h. For the purpose of getting pure culture, distinctive colonies were picked up and transferred to a fresh (MDPMA) agar plates, then incubated at 37°C in a bench top incubator for 24 and 48 h till pure culture were obtaining.

#### Microbial methods

**Phylogenetic analysis:** Analysis of genes sequences of bacteria was done by phylogenetic method using reference sequences homology from the NCBI database using MEGA 4 programme (Tamura et al.). Phylogenetic trees were constructed by distance matrix-based cluster algorithms *viz.* un-weighted pair group method with averages un-weighted pair group method using arithmetic averages (UPGMA), neighbour-joining, maximum likelihood, and maximum-parsimony analysis as described elsewhere [17]. The trees were rooted using *Escherichia coli* strain K 12 MG1655 (accession no. U00096) as out-group; the stability of trees obtained from above cluster analysis was assessed by using BOOTSTRAP program in sets of 1000 re-samplings (MEGA 4) [18].

**Antimicrobial activity:** For the experiments, a loopful of the bacterial working stocks were enriched on a tube containing 9 ml of Nutrient broth then incubated at 37°C for 24 h.

Antimicrobial activity was done according to the protocol described by Hajlaoui et al. [19]. Agar diffusion well-variant: according to Smânia et al. [20].

The bacterial inoculum was uniformly spread using sterile cotton swab on a sterile petri dish contain Müller-Hinton agar. 40 µL of each the supernatant extract (Methanol-ethyl acetate) were added to each of the wells (7 mm diameter holes cut in

the agar gel). The systems were incubated for 24 h at 37°C ± 1°C, under aerobic conditions. After incubation, confluent bacterial growth was observed. Inhibition of the bacterial growth was measured in mm. Tests were performed in duplicate.

**Determination of minimum inhibitory concentration (MIC):** The MIC for different strains was determined by using a microtiter plate dilution method in appropriate broth using 96-well plates according to the Clinical and Laboratory Standard Institute (CLSI) guidelines [21]. Using 0.5 McFarland standards, CFU/mL of each indicator bacteria was prepared and subjected to varying concentration of the methanol and ethyl acetate 0.4-100.0 µg/mL. The microtiter plates were incubated at incubator under shaking condition. The lowest concentration that inhibited growth of indicator strains was recorded.

**Chemicals analyses of the extract**

**Preparation of the methanol and ethyl acetate extracts:** *B. firmicutes* were cultured for 48 h then centrifuged and 200 ml of the supernatant were concentrated to 20 ml. Then 10 ml was treated with methanol and another 10 ml was treated with ethyl acetate. Both fractions were concentrated to 2 ml then it used for antibacterial and TLC analyses.

**Determination of moisture:** The method reported the A.O.A.C [22] for moisture determination. Sample (5 g) was weighed in watch glass the sample was dried at 120 ± 2°C to constant weight.

**Determination of ash:** The method reported in the A.O.A.C [22] was applied. Sample (2 g) was weighed into porcelain crucible and in muffle furnace at 550°C until constant weight.

**Antioxidant assays**

**Test for phenolics [23]:** Neutral ferric chloride was added to each fraction. Appearance of bluish green colour indicated presence of phenolics.

**Test for flavonoids:** Dilute ammonia (5 ml) was added to a portion of sample solution in water. Concentrated sulphuric acid (1 ml) was added. A yellow colouration that disappear standing indicated the presence of flavonoids.

**Condensed tannins:** Condensed tannins were estimated using the method of Sun et al. [24] with some modifications.

**Total antioxidant capacity (TAC):** Phosphomolybdenum assay. The total antioxidant capacity (TAC) of culture filtrate was spectrophotometrically determined by the phosphomolybdenum assay using the method described by Prieto et al. [25]. The antioxidant capacity of each sample was expressed as Ascorbic Acid (AA) equivalent using the following linear equation established using ascorbic acid (Figure 1) as standard:  $(A=0.0037C+0.0343; R^2=0.991)$  where A is the absorbance at 695 nm and C the concentration as ascorbic acid equivalent (µg/ml) [26]. The values are presented as the means of triplicate analysis.

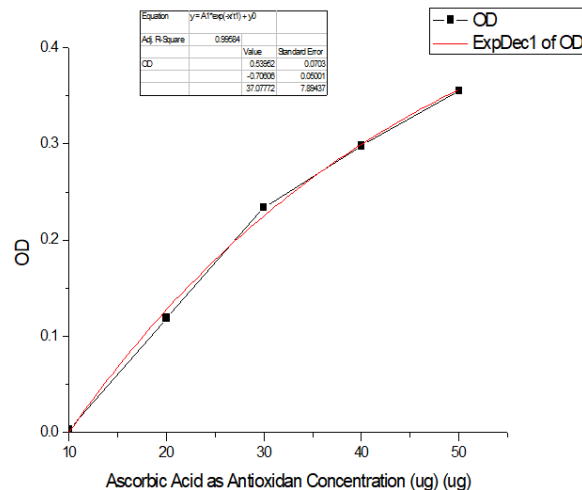


Figure 1. Stander curve for total antioxidant capacity.

**DPPH scavenging activity assay:** DPPH radical scavenging activity was measured using the method described with slight modifications [27].

1 mL of sample was added to 2 mL of DPPH (0.15 mM). 1 mL of 100% methanol was added to 2 mL of DPPH as blank control. The mixture was allowed to stand for 30 min in the dark at room temperature. After 30 min, absorbance was measured at 517 nm. All samples were tested in triplicate. The results were expressed in percentage of radical scavenging activity, calculated using the following formula:

$$\% \text{ radical scavenging activity} = (A \text{ Blank} - A \text{ Sample}) \times 100\%$$

**A Blank:** A Blank refers to the absorbance of the blank control whereas A Sample is the absorbance of the tested samples.

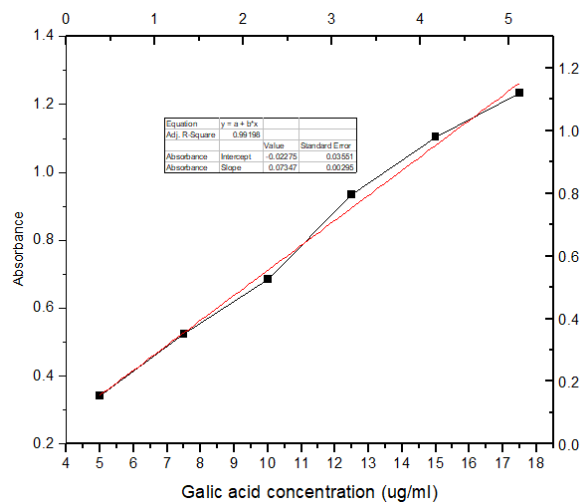


Figure 2. Calibration curve of standard Gallic acid for determination of total phenolics.

**Total phenolic content analysis:** Total phenolic content of the supernatant extract of *B. firmicutes* was determined with Folin-Ciocalteu reagent according to the method [28] with slight modification, using Gallic Acid as a standard. A 0.1 mL of extract was placed into a 10 mL measuring flask and 0.5 mL of

Folin-Ciocalteu reagent was added to the extract. The measuring flask was allowed to stand at room temperature for 5 min. Then, 1.5 mL of 20% (w/v) Na<sub>2</sub>CO<sub>3</sub> was added to the mixture. The mixture was adjusted until the volume reaches 10 mL with distilled water. After 30 min at room temperature, absorbance was measured at 765 nm versus a blank by using a spectrophotometer (Shimadzu UV 1800, Japan). Total phenol value was expressed as mg Gallic Acid 9 (Figure 2) Equivalent/g.

### Chromatographic analysis

**Thin layer chromatography (TLC):** (A) Identification of antioxidant for different cultured time. TLC was used to assess the purity and number of compounds in the supernatant extract of *B. firmicutes* for different days (1-7 d) was analyzed on pre-coated 5 × 10 cm, 0.25 mm thickness silica gel 60 F254 aluminium plates; (B) Both methanol and ethyl acetate were analyzed on pre-coated 5 × 10 cm, 0.25 mm thickness silica gel 60 F254 aluminium plates.

1. The extracts were spotted on TLC plate by using a fine glass capillary tube.
2. The mobile: Chloroform: Methanol (10:1).
3. The developed TLC was visualized under UV lamp.

**Identification by using MS/MS:** MS experiments were carried out using a Thermo LCmsms, model LTQ XL Instrument and with an accelerating voltage of 16 kV. Hypersil GOLD™ C18 selectivity LC columns.

## Results and Discussion

Date products such as syrup, jam, vinegar and sweets are all important and used by local consumers. Date seeds considered a waste product of many date processing plants producing pitted dates, date syrup and date confectionery. At present, seeds are used mainly for animal feeds in the cattle, sheep and camel and poultry industries. With world production of dates reaching 6.9 million tons in 2004, from this approximately 863 thousand tons of date seeds are produced [29].

### Dates seed ago (pits) chemical composition

Results for chemical composition of date ago seed pits are shown in Table 1. The ash, moisture, protein, fat, and fiber contents (dry weight basis) were 0.9, 4.8, 5.47, 6.1, 9.0 and 22%, respectively. Accordingly, total carbohydrate content was 57.2%. Our results fall within the range of date pits values previously reported by Attalla et al. [30], Hamada et al. [31] and Al-Farsi et al [32] who researched the functional properties of date seeds, their reported composition was 3.1-7.1% moisture, 2.3-6.4% protein, 5.0-13.2 fat, 0.9-1.8% ash and 22.5-80.2% dietary fiber.

**Table 1.** Chemical composition of date pits/100 g powder.

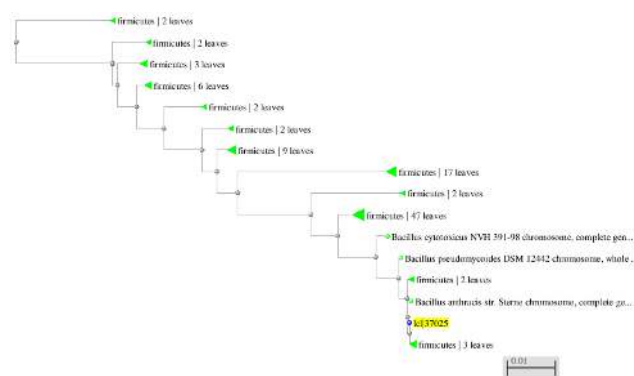
Ingredients	DP/100 g
Ash (g)	0.9

Moisture (g)	4.8
Protein (g)	6.1
Total fat (g)	9
Fiber (g)	22
Carbohydrates (g)	57.2

### Identification of Bacillus strain

Molecular characterization of these strains was done by DNA isolation (CTAB method) and 16S rDNA analysis. Further these amplified 16S rDNA sequences of the bacterial strains was blasted using online tool (MEGA 4). The taxonomical identification was done by the phylogenetic tree construction and the comparison of this bacterial strain sequences with other homologous bacterial sequences.

The strain was gram positive, aerobic, spore forming short rod *Bacillus*. The organism was able to grow between 30-60°C and Several biochemical tests as Catalase reaction positive, oxidase reaction positive, nitrate oxidase positive, gram staining and 3% KOH negative, idols negative and maltose negative in addition to phylogenetic tree based on different species of *Bacillus* was constructed using neighbour joining method (Figure 3), indicated that strain was very closely related with *Bacillus firmicutes* strain. Therefore it was named *B. firmicutes* strain.



**Figure 3.** Identification of Bacillus by using phylogenetic tree method indicated that strain was very closely related with *Bacillus firmicutes* strain.

### Phytochemical investigation

The present study carried out on the supernatant of the culture medium after incubation for 2 d revealed the presence of medicinally active constituents (Table 2). It was observed from the preliminary phytochemical investigation that phenolics, flavonoids and tannin were present in the supernatant. Deschamps et al. [33] reported that bacterial strains identified as *Klebsiella pneumonia* and *Corynebacterium sp.* were produced Gallic acid.

**Table 2.** Phytochemical constituents of culture ago pits medium.

Test	Supernatant
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Tannins	+
Sugars	+
Phenolic	+
Flavonoids	+

**Total antioxidant capacity (TAC)**

The total antioxidant capacity (TAC) of culture supernatant of *B. firmicutes* was determined by the phosphomolybdenum assay using the method described by Prieto et al. [25]. As shown in Figure 1 that the total (TAC) compounds in the culture supernatant is 2.5 mg/ml of culture medium. The same results were detected by Janardhan et al. [34] who detected potent total antioxidant activity produced by isolated actinomycetes.

**Determination of total phenolic compounds**

Phenolic compounds were responsible for the antioxidant activity and many biological activities such as antibacterial, anticancer, anti-allergic, anti-diabetes, anti-aging, anti-inflammatory and anti-HIV activities the most important classes of natural products [35]. The produced total phenolic compounds the culture supernatant of the isolated, selected *B. firmicutes* were determined and expressed as Gallic acid equivalent. The highest amount of phenolic compounds is 5.3 mg/ml of culture medium as shown in Figure 2. The same results were reported by Tebbal et al. [36] who found phenolic compounds in the isolated cyanobacteria. Martins et al. [37] have reported that the use of methanol for extraction of the total phenolic content has manifested a higher amount yield in *Oscillatoria agardhii* and *Anabaena sphaerica*.

**DPPH radical scavenging activity**

Free radical scavenging is one of the known mechanisms by which antioxidants inhibit cellular damage. DPPH is one of the compounds that have a proton free radical with a characteristic absorption, which decreases significantly on exposure to proton radical scavengers. Furthermore, it is well accepted that the DPPH free radical scavenging by antioxidants is due to their hydrogen-donating ability. The present findings showed that antioxidants were found in the culture supernatant of *B. firmicutes* had a noticeable DPPH free radical scavenging activity Figure 4. It was observed that the DPPH scavenging activity was increased in a dose dependent (0-100 µg/mL) manner. The greatest scavenging rate of antioxidants was 60%. The results were less than that done by Liang et al. [38] who showed that scavenging rate of EPS isolated from *P. mucilaginosus* was 80%, the 72% observed for *Serratia ureilytica* [39] and 77% for *Paenibacillus sp.* [40]. The half-maximal effective concentration (EC50) of antioxidant 57 µg/mL, which was lower than that of the standard antioxidant ascorbic acid (60 µg /mL); antioxidants found in the culture supernatant of *B. firmicutes* were a potent and natural antioxidant that could be used as an alternative to synthetic antioxidants.

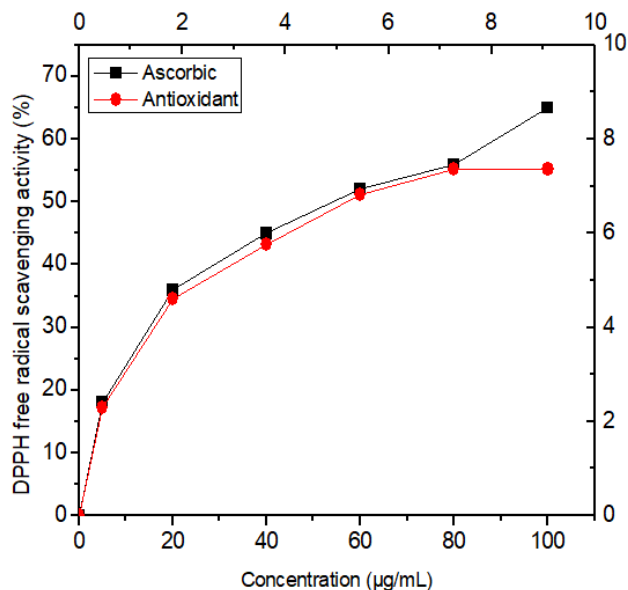


Figure 4. DPPH free radical scavenging ability *B. firmicutes*.

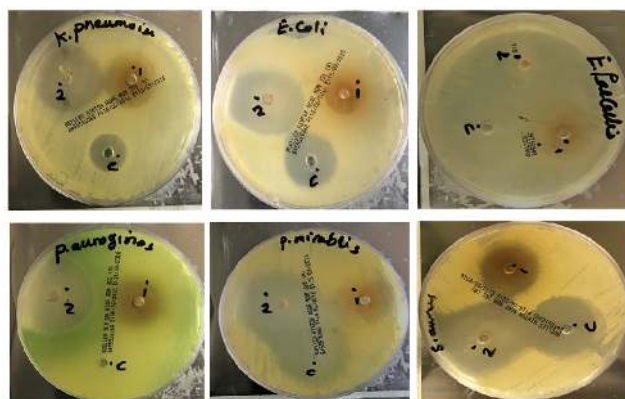


Figure 5. Inhibition zone produced by *B. firmicutes*. 1 methanol extract, 2 ethyl acetate extract, Augmentin antibiotic.

**Antimicrobial activity**

In the present study, the antibacterial activity of the extracted (methanolic, ethyl acetate) of *B. firmicutes* against pathogenic gram-negative and positive bacteria strains responsible for nosocomial infection such as *E. coli*, *K. pneumonia*, *Acinetobacter baumannii*, *Ps. aeruginosa* and *Sta. aureus* has been investigated by the using of agar-well diffusion method, based on the measure of the formed diameter of the inhibition zone. Both extracts exhibited a potent antibacterial activity against the tested bacteria, however the diameter of the inhibition zone depends mainly on the type of the used solvent for the extraction, the ethyl acetate is more effective than methanol against pathogenic bacteria.

As shown in Figure 5 and Table 3, highest antibacterial activity in ethyl acetate extract was observed against *E. faecalis* with a maximum diameter inhibition zone 25 mm while; the lowest activity was detected against *P. aeruginosa* diameter inhibition zone 14 mm. While, the highest antibacterial activity in methanol extract was observed against *K. pneumoniae* and *E.*

*faecalis* with a maximum diameter inhibition zone 15 mm and, the lowest activity was detected against *P. aeruginosa* diameter inhibition zone 10 mm. On the other hand, the methanol extract did not exhibit antimicrobial activity against *E. coli*. Several lipopeptide biosurfactants obtained from *Bacillus amyloliquefaciens* M1 [41], *Bacillus mojavensis* A21 [42] and *Bacillus methylotrophicus* [43] showed strong antibacterial activity.

**Table 3.** Antimicrobial activity spectrum of *B. firmicutes*.

Bacteria name	G	Control/m	Methanol	Ethyl acetate
<i>K. pneumoniae</i>	G-	10	15	17
<i>E. coli</i>	G-	14	9	19
<i>P. mirabilis</i>	G-	16	12	19
<i>P. aeruginosa</i>	G-	0	10	14
<i>E. faecalis</i>	G+	14	15	25
<i>S. aureus</i>	G+	14	14	18

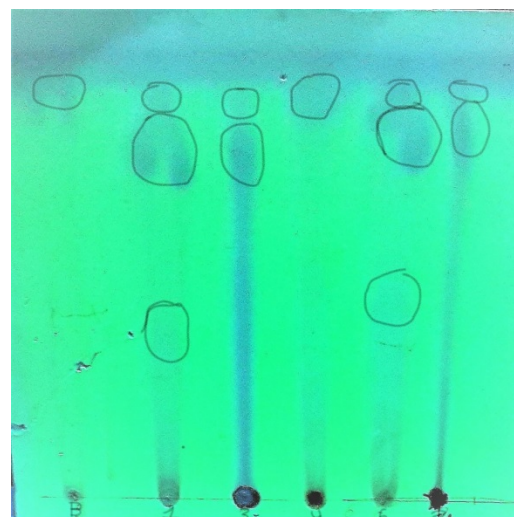
### Minimum inhibitory concentration (MIC)

In ethyl acetate extract *E. faecalis*, *E. coli* and *P. mirabilis* were the most sensitive bacteria with MIC values of 15, 20 and 21 mg/mL respectively while *P. aeruginosa* 30 mg/ml. The methanol has less efficient than ethyl acetate extract. In case of ethyl acetate extract both *K. pneumoniae* and *E. faecalis* were the most sensitive bacteria with MIC values of 35 and 40 mg /ml respectively. Many studies have underlined the importance of bacterial antibiotics in the food industry. Indeed, both nisin and pediocin PA-1 produced by lactic acid bacteria have been approved as food additives in many countries [44]. Our results are in agreement with the results of [45] that showed that new antibiotics and antioxidant S07-2 compound produced by *Bacillus subtilis* B38.

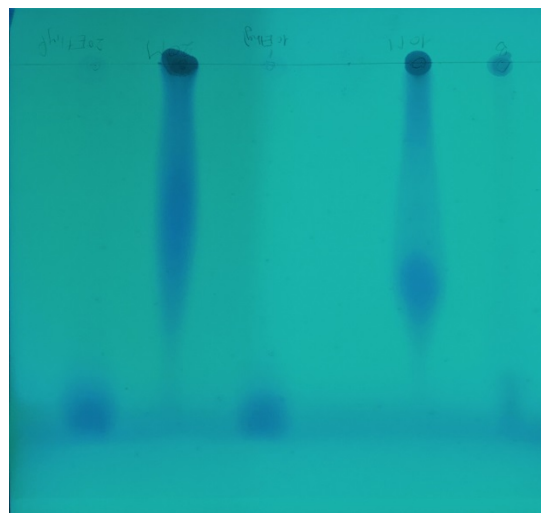
### Thin layer chromatography

**TLC for detection the antioxidant at different time:** 50 ml of culture medium were taken every day for 7 d. The culture medium was centrifuged to separate the bacterial cells. The supernatants were evaporated to about 1.5 ml. 40  $\mu$ l for every day was applied on TLC (silica gel 60 F254). The developed TLC was visualized under UV lamp. UV-active organic compounds appeared as purple spots on TLC as seen Figure 6. As shown in Figure 6, in the front, first spot was found in all days even in the blank although it increases in the color in 4<sup>th</sup> day. New spot detected from 1<sup>st</sup> day, trace spot was detected in 1<sup>st</sup> and 6<sup>th</sup> day.

**Detection of Antioxidant in methanol and ethyl acetate extracts:** As shown in Figure 7 both ethyl acetate and methanol at two concentrations (10-20  $\mu$ L) were applied in TLC plate. The developed TLC was visualized under UV lamp. The front spot was found in ethyl acetate while methanol extract was found later from the front.



**Figure 6.** Thin-layer chromatography TLC analysis of antioxidant products for different fermentation time, B: one spot; 1 d: 3 spots; 3 d: 2 spots; 4 d: 1 spot; 6 d: 3 spots; 7 d: 2 spots.

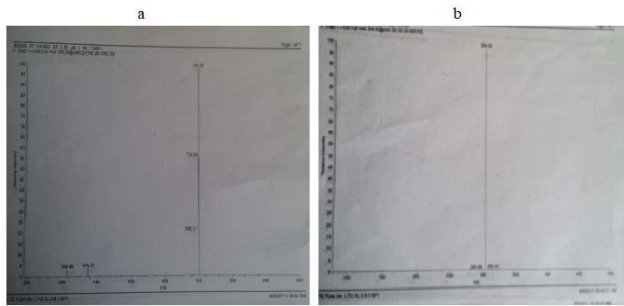


**Figure 7.** TLC of ethyl acetate and methanol extract of *B. firmicutes* supernatant.

### MS analysis

The molecular mass of both extracts methanol and ethyl acetate compound were determined LCMS (Figures 8a and 8b). The mass spectrum confirmed the purity of methanol sample showed one major peak at  $m/z$  701. Expansion of the chromatogram also showed minor species at  $m/z$  325.

On the other hand, ethyl acetate sample showed one major peak at  $m/z$  304 and minor one at  $m/z$  284. The same results were observed by Hagelin et al. [46] who produced cyclic peptide antibiotics by *B. subtilis* species generally exhibit molecular masses 41000 Da, ranging from 1447.7 to 1519.8 Da in the case of the maltacine complex, from 800 to 1500 Da in the case of lipopeptides [47].



**Figure 8.** Mass spectrum profile of the methanol and ethyl acetate extracts. a) Methanol extract; b) Ethyl acetate.

## Conclusion

Both ethyl acetate and methanol extract possess highest antibacterial activity against *E. faecalis* with a maximum diameter inhibition zone of 25 mm, 15 mm respectively while, the lowest activity was detected against *P. aeruginosa* with diameter inhibition zone 14 mm and 10 mm respectively.

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