



A Novel Bio-Sensor Based on DNA Strand Displacement

Xiaolong Shi, Zhiyu Wang, Chenyan Deng, Tao Song*, Linqiang Pan*, Zhihua Chen

Key Laboratory of Image Information Processing and Intelligent Control, School of Automation, Huazhong University of Science and Technology, Wuhan, Hubei, China

Abstract

DNA strand displacement technology performs well in sensing and programming DNA segments. In this work, we construct DNA molecular systems based on DNA strand displacement performing computation of logic gates. Specifically, a class of so-called “DNA neurons” are achieved, in which a “smart” way inspired by biological neurons encoding information is developed to encode and deliver information using DNA molecules. The “DNA neuron” is bistable, that is, it can sense DNA molecules as input signals, and release “negative” or “positive” signals DNA molecules. We design intelligent DNA molecular systems that are constructed by cascading some particularly organized “DNA neurons”, which could perform logic computation, including AND, OR, XOR logic gates, automatically. Both simulation results using visual DSD (DNA strand displacement) software and experimental results are obtained, which shows that the proposed systems can detect DNA signals with high sensitivity and accretion; moreover, the systems can process input signals automatically with complex nonlinear logic. The method proposed in this work may provide a new way to construct a sensitive molecular signal detection system with neurons spiking behavior in vitro, and can be used to develop intelligent molecular processing systems in vivo.

Citation: Shi X, Wang Z, Deng C, Song T, Pan L, et al. (2014) A Novel Bio-Sensor Based on DNA Strand Displacement. PLoS ONE 9(10): e108856. doi:10.1371/journal.pone.0108856

Editor: Sabato D’Auria, CNR, Italy

Received: May 27, 2014; **Accepted:** August 25, 2014; **Published:** October 10, 2014

Copyright: © 2014 Shi et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability: The authors confirm that all data underlying the findings are fully available without restriction. All relevant data are within the paper.

Funding: This work is supported by the National Science Foundations of China (Grant Nos. 61272071, 61033003, 61370105, and 61402187). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* Email: taosong@hust.edu.cn (TS); lqpan@hust.edu.cn (LP)

Introduction

Biomolecular computation refers to the study of exploiting biological macromolecules to implement relatively standard methods of computation, including molecular computing [1–3], membrane computing [4,5], storage media using bacteria rhodopsin [6,7] and biologically altered cells that do rudimentary operations within the paradigm of traditional computation, etc. In recent years, computing with programming DNA molecular has become a hot research topic and lot of work have contributed to this field, such as helical molecular programming [8], tabletop molecular communications with chemical signals [9], molecular computing methods to improve the accuracy of insertion site analysis in tumors [10], DNA self-assembly for computation [11,12], DNA strand displacement (DSD) technology [13,14].

DNA strand displacement technology has been proposed as an isothermal, in vitro DNA amplification technique in [13]. The technique is highly selective to the recognition of sequence [15], and has been used in detection of gene signals as a second-generation DNA probe system with the help of some DNA nanotechnology, see e.g. in [16–22]. DNA strand displacement is an isothermal and enzyme free technique, see e.g. [13,16], and it can be potentially applied as an intelligent molecular systems in vivo for DNA signal detection and processing. Some bio-molecular signal processing systems have been developed with using DNA strand displacement technology, such as

enzyme-free nucleic acid logic circuits [23], genetic programming and evolvable molecular machines [24], performing logic computation of Hopfield network auto-associative memory with DNA strands [25], kinetically controlled self-assembly of DNA oligomers [26].

In bio-molecular signal processing systems proposed in [23–26], single-stranded DNA molecules are generally used as input and output signals; all the strands needed for the computation are mixed together, and then the computation proceeds according to the design of DNA sequences without further intervention. The energy for this procedure is provided by the Watson-Crick complementary mechanism of DNA structures themselves, so whole system can run in basic wet labs. Note that all DNA strands can interact with each other automatically according to programmed logic with DNA strand displacement technology, that is, each DNA strand performs its own reactions independently with logically related strands respectively. The DNA strands cannot “cross talk” with other unrelated strands restricted by sequence design.

A molecular system can be seemed as a functioning unit with DNA strands representing particular partial biological functions, where information is encoded and stored in form of DNA strands, and each DNA strand performs its own reactions independently to achieve particular biological function. The output DNA strands in certain reaction can be taken as input

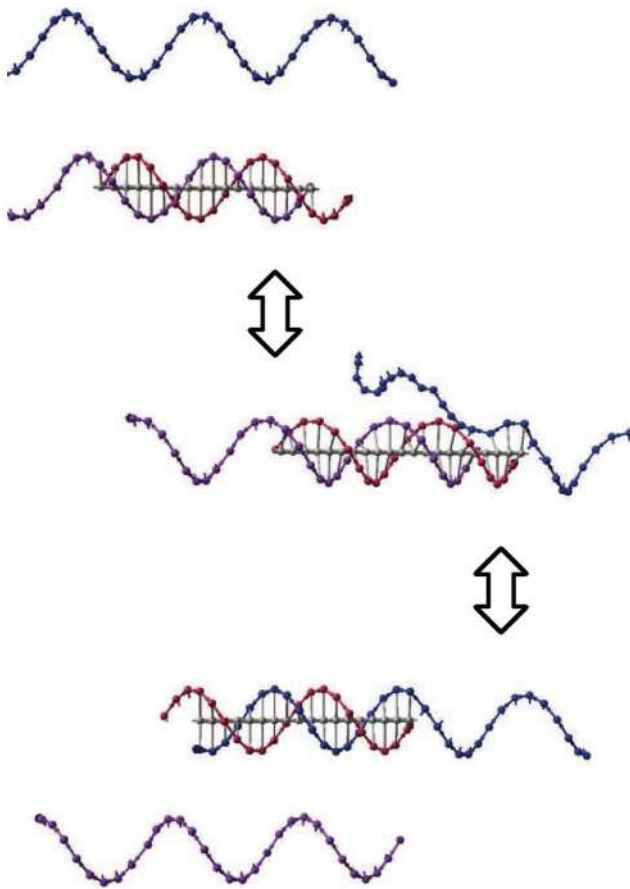


Figure 1. Thresholding reaction of a DNA neuron.
doi:10.1371/journal.pone.0108856.g001

of another DNA strand reactions, which in some sense can process information like functioning unit performing chain reactions.

Results

In this work, we present a novel method to achieve logic gate computation with DNA strand displacement technology. In general, group of “DNA neurons” are designed, and by cascading organization of the “DNA neurons” some intelligent molecular systems for logic gate computation are achieved.

In “DNA neurons”, we use a “smart” way that neurons encoding information by means of accumulation of spikes to encode information with “accumulation” of volumes of the input signal DNA molecules. Specifically, we use a amount of mol of pre-designed DNA strand as a basic information unit, and then encode information by different amounts of mols of basic DNA molecules. As well, some logically separated group of DNA strands, named “DNA neuron”, are designed for a particular reaction, which can perform its own function in a “separated region” (without interactions with other “DNA neurons”). The communication of “DNA neurons” with each

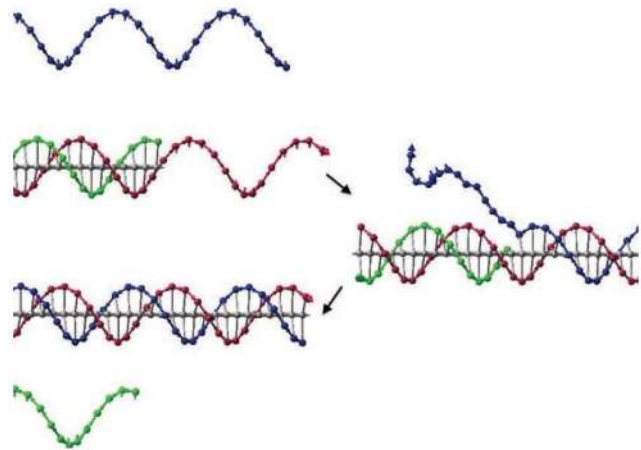


Figure 2. Irreversible DNA strand displacement.
doi:10.1371/journal.pone.0108856.g002

other is as follows: one “DNA neuron” performs its own reactions to generate particular output DNA strands, and the other “DNA neuron” starts its reaction only when it senses the generated output DNA strands.

With “DNA neurons”, we construct intelligent molecular systems for logic gates computation, including AND, OR, XOR gates. The systems perform logic computation with each “DNA neuron” detecting and processing DNA signals automatically until the computation result (which can be reported by “reporter DNA strand”) is obtained. To show the validity of our method, we firstly use visual DSD from [27] to do simulations of the computations in the systems for AND, OR, XOR logic gates, and then experiments are performed. Experimental results show that the systems with “DNA neurons” can work correctly and efficiently for performing logic gate computations, including AND, OR, and XOR logic gates.

Methods and Materials

Method

In general, “DNA neurons” can receive different combination of ssDNA strands as input spikes, and then release corresponding ssDNA strands as output signals. These output signals could be accepted by other DNA neurons as input spikes. This property makes it possible to cascade these DNA neurons systematically to build more complicated circuits. The XOR logic is such a creation that is constructed by organizing two AND logics in an artful way. The constructed XOR logic has an excellent attribute that the output is determined with given input spikes, which will be explained in details later.

There are two kinds of DNA strand displacement strategies applied in this work: one is reversible strand displacement (shown in Figure 1), the other one is irreversible strand displacement (shown in Figure 2). For reversible strand displacement reaction, the input strand (blue strand in Figure 1) hybridizes with a gate complex (red and purple

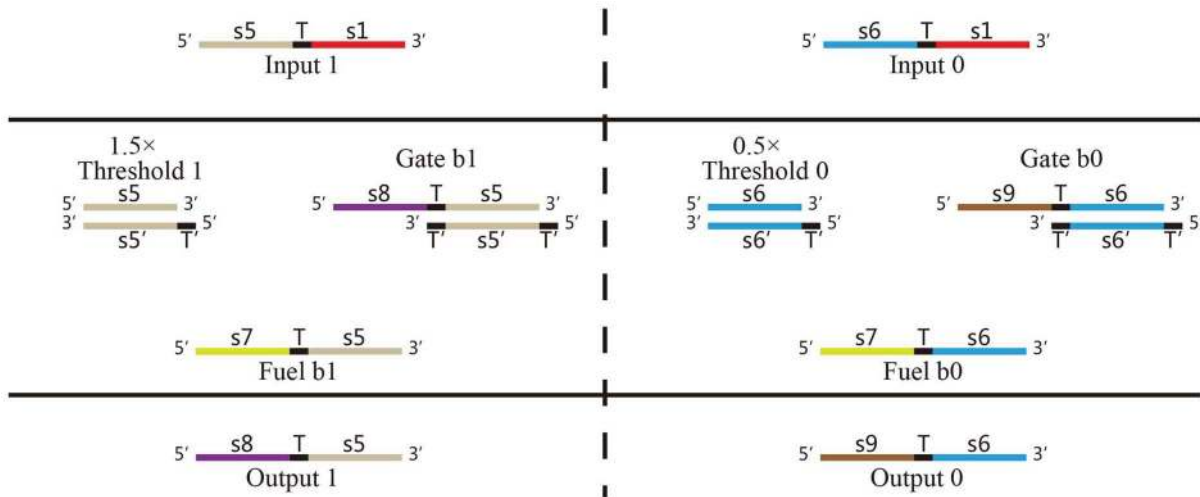


Figure 3. Abstract diagrams of implementation of a DNA neuron with AND logic.
doi:10.1371/journal.pone.0108856.g003

complex in Figure 1) through the exposed toehold, and then the branch migrates through recognition domain occurs and finally the previous strand (purple strand in Figure 1) will desquamate as an output signals of a DNA neuron. Note that there is the same uncovered toehold on the opposite side of the gate base strand (read strand in Figure 1), which makes this reaction reversible. In irreversible reaction, the input strand (blue strand in Figure 2) hybridizes with a threshold complex (red and green complex in Figure 2) via an extended toehold, and the original branch (green strand in Figure 2) bound to the threshold is replaced, and this reaction is not reversible since there is no uncovered toehold left.

Bistable DNA neurons with two stable output states are designed based on the two basic DNA strand displacement reactions above. It is shown how does the DNA neuron execute AND logic in Figure 3. The bistable DNA neuron can be divided in two parts. It is given in the left of Figure 3 the positive part which receives and releases the positive signal “1” and in the left of Figure 3 is the negative part corresponding with the negative neuron signal “0”. Each part consists of input, threshold, gate, fuel and output strands. The input strands is added in a form of accumulation of impulses, i.e., double amounts of input strand “1” added to represents the input of “11”; one time amount of input strand “0” and one time amount of input strand “1” represent

the input signal of “01”; and double amounts of input strand “0” represent the input signal of “00”. It is clear that rather than hybridizing with the gate, the input strand signals prefer to hybrid with the threshold since this reaction is irreversible. The threshold value in the positive part is 1.5, thus only when the input strand is “11”, an output signal strand in the positive part can be expected. The threshold value in the negative part, similarly, is set to be 0.5, thus output signal strand in the negative part can be generated only when the input signals are “10”, “01” and “00”. In this way, only if the input is “11”, the Output 1 strand could be detected; otherwise Output 0 strand will be detected. Inversely, if Output 1 strand is regarded as negative output signal 0, and Output 0 strand is regarded as output of positive signal 1, then AND logic can perform logic computation of OR logic gate.

In order to detect the strand representing positive output signal “1” and strand representing negative signal strand “0”, two kinds of fluorescent probes, namely reporter strands, are designed as shown in Figure 4. The reporter strands for output strand signals “1” and “0” (strand 8 and strand 9 in Figure 4) are labeled by fluorophores (HEX and FAM in Figure 4) and quenchers (IAbHQ and IAbFQ in Figure 4) occurring fluorescence quenching. When the output strand representing signal “1” or “0” releases from DNA neuron,

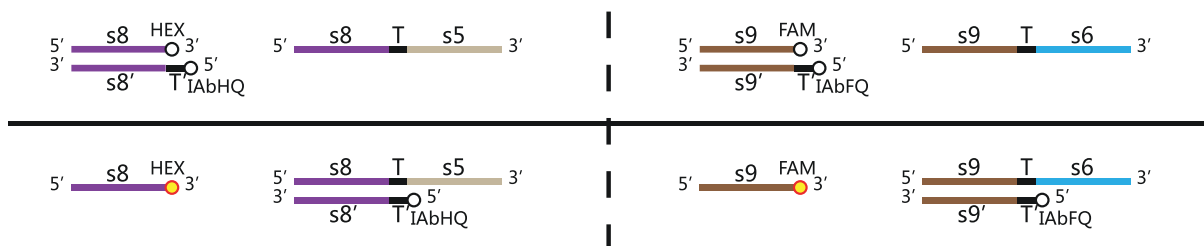


Figure 4. HEX and FAM fluorescent probes to detect the output spike “1/0” of DNA Neuron.
doi:10.1371/journal.pone.0108856.g004

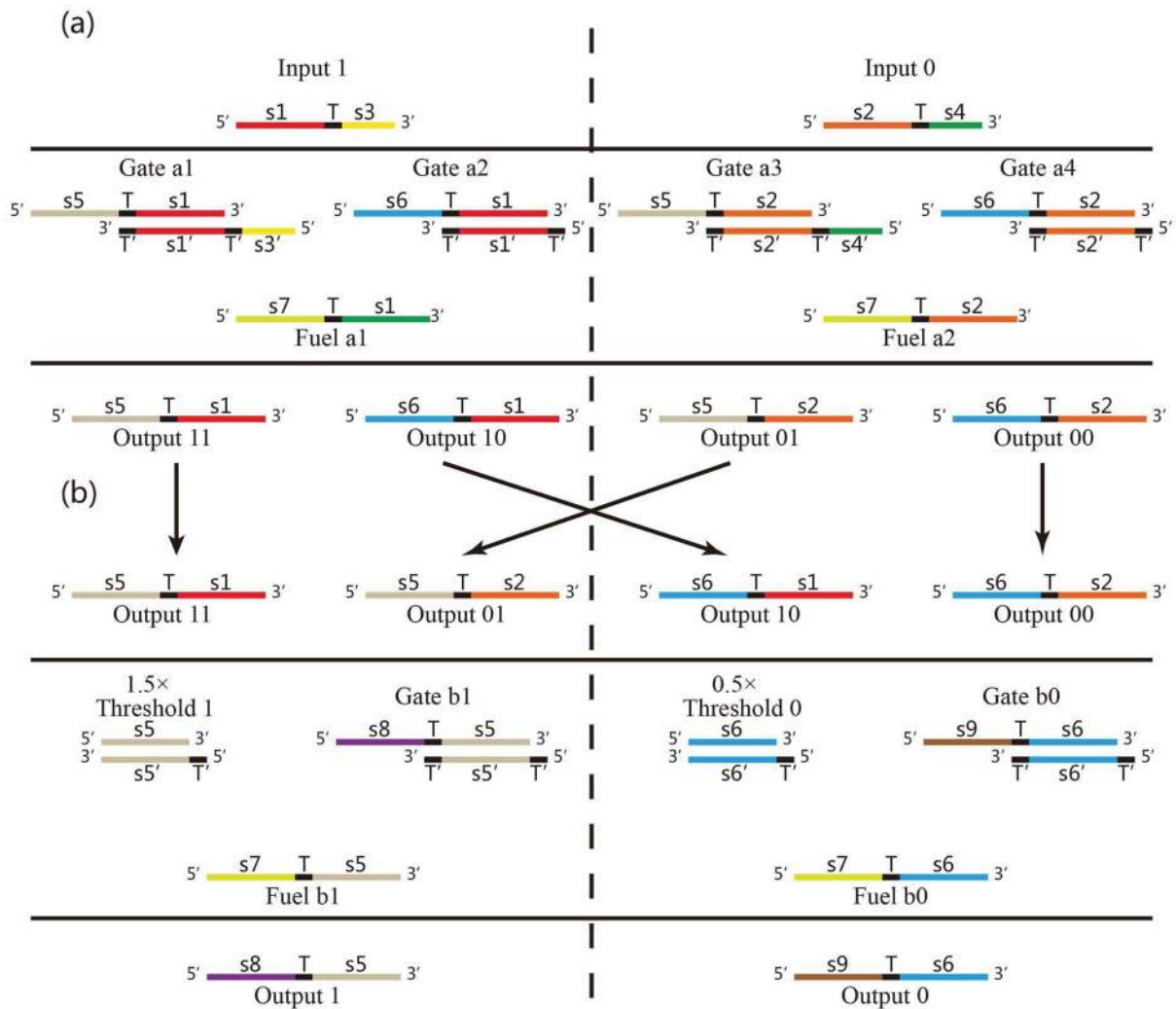


Figure 5. Abstract diagrams of two DNA neurons executing XOR logic.
doi:10.1371/journal.pone.0108856.g005

it will trigger the correspondence reporter and an irreversible DNA strand displacement take place. In this way, the two radicals will be separated at last and the fluorescence signal representing “1” or “0” can be monitored with a real-time PCR Machine.

The XOR logic gate is constructed by cascading two DNA neurons executing AND logics in an artful way, which consists of two parts, XOR (a) and XOR (b), which is respectively illustrated in Figure 5 (a) and (b). In each part, it can generate two outputs: a positive output (Output signal strand “11” or “01”), and a negative output (Output signal strand “10” or “00”), the signal strands can be transferred to the DNA neuron in XOR (b) as input signals. It is noted that only input signal strands with doubled amount would generate negative output. Specifically, by receiving input signal “11”, XOR (a) will release one time amount of positive output (Output strand “11”) and one time amount of negative output (Output strand “10”), while by receiving the input signal “10”, it will release double amount of positive output (Output strand “11” and

“01”). The output signals of XOR (a) can be taken as input signal of XOR (b). If XOR (b) receives double amounts of positive input signals, it leads to release Output strand “1”; otherwise, it output negative signal by releasing Output strand “0”. With the explanation above, the system performing computation of XOR logic gate is constructed with DNA neurons. To make the computing process of DNA neurons for XOR logic clear, the following cases are discussed, where by $1 \times$ we denote the basic amount of mol as a basic information unit.

- We add $1 \times$ strand Input 0 and $1 \times$ strand Input 1 into the sample to represent the input signal being “01”, and then the Gate a1 and Gate a3 reaction can occur respectively. Eventually, the positive part will output positive signal ($1 \times$ strand Output strand “11”) and the negative part will output positive signal ($1 \times$ Output strand “01”). The outputs can be sensed as input signals of XOR (b). In XOR (b), if the concentration of the two positive input strands

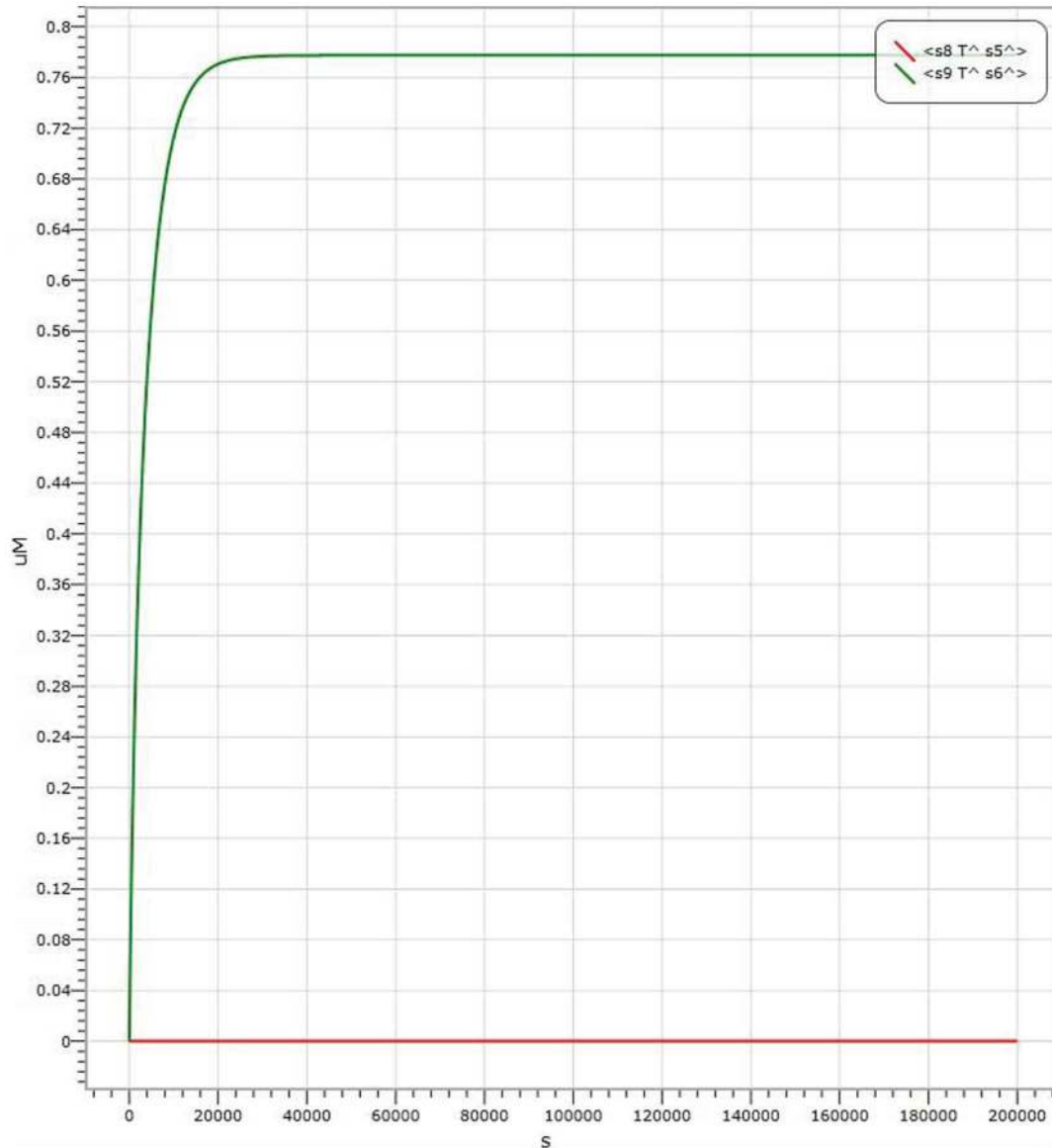


Figure 6. Visual DSD simulations of AND logic gate with input signal “00”.
doi:10.1371/journal.pone.0108856.g006

exceeds the threshold value $1.5\times$, and it outputs positive signal “1” consequently.

- We add $2\times$ strand Input 1 into the sample to represent the input signal being “11”. The Gate a1 and Gate a2 reactions occur successively in the positive part. Ultimately the positive part will output positive signal ($1\times$ Output strand “11”) and negative spike ($1\times$ Output strand “10”). The output strands will enter into XOR (b) as input signals, in which, the concentration of the negative signal will exceeds the threshold value $0.5\times$, and it outputs negative signal strand “0” consequently.
- As for input signal “00”, the XOR (a) release $1\times$ Output strand “01” and $1\times$ Output strand “00”. XOR (b) can sense

these strands as input signal “10” and release negative signal “0”.

The design of DNA neuron executing AND, OR, XOR logic gates with input based on amount variation is rational and has potential to be applied to construct circuit with more complexity.

Simulation

Based on the DSD mechanism analysis above, Visual DSD from [27] (a software package that could visualize the species and the reactions of the process of DNA strand displacement at the domain level then generate stochastic or deterministic simulations) is introduced as a tool of kinetics simulation to

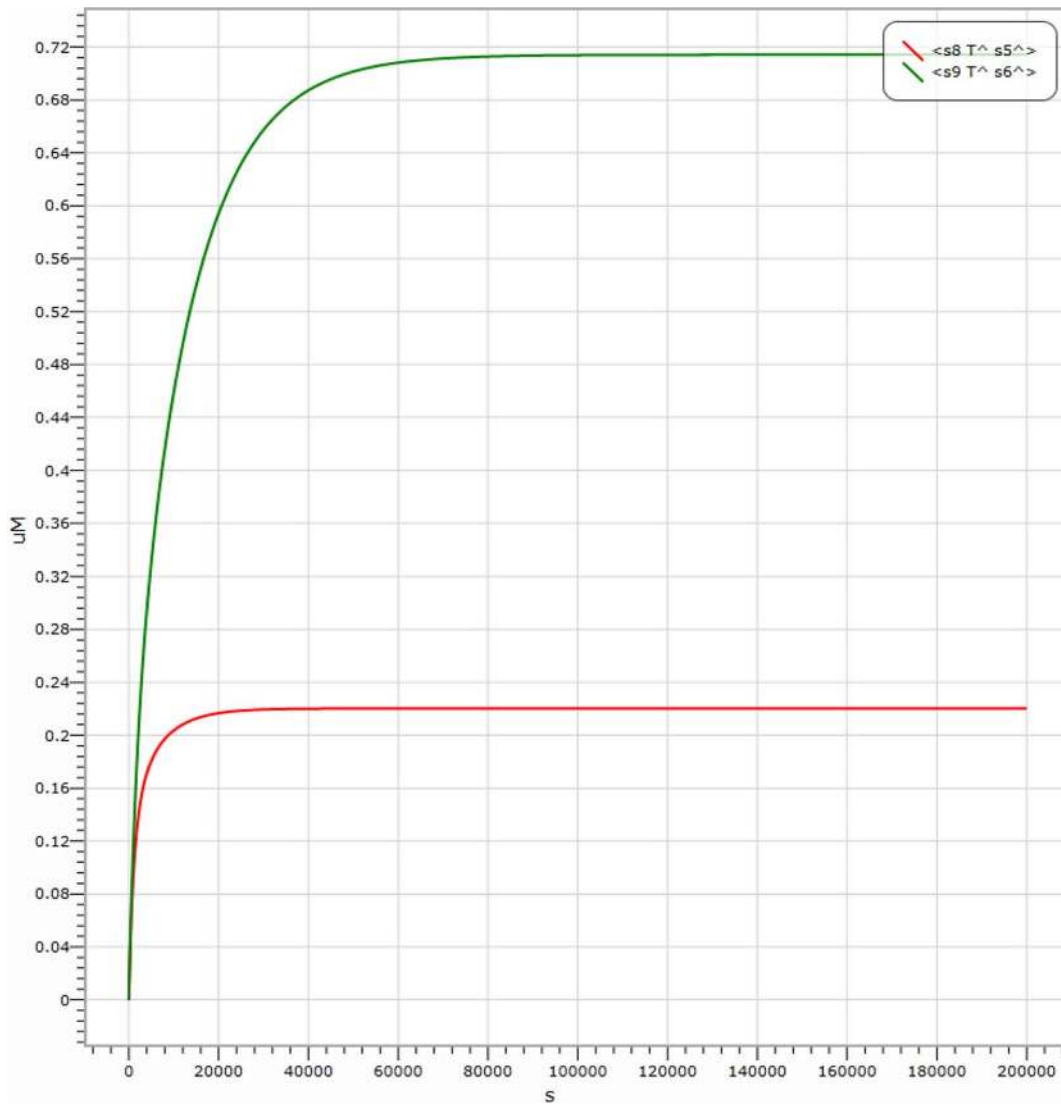


Figure 7. Visual DSD simulations of AND logic gate with input signal “01”.
doi:10.1371/journal.pone.0108856.g007

verify the feasibility of the bistable DNA neuron design. The models of AND logic and XOR logic gates are both programmed with Visual DSD, and for each model three types of input signals, “00”, “01” and “11” are tested with expected output. The strands used in the kinetics simulations are the same with the wet experiment that followed.

In AND logic gate, shown in Figure 4, the amount of Threshold 1 is $1.5 \times$; that of Threshold is $0.5 \times$; that of Gate b1 and that of Gate b0 are both $1 \times$, while Threshold of Fuel b1 and Fuel b0 are both $2 \times$. The results of kinetics simulations are shown in Figures 6, 7 and 8, where the red curve denotes the amount of molecule P15 ($\langle s8 T^{\wedge} s5^{\wedge} \rangle$) representing the output of signal “1” and the green curve denotes the amount of molecule P20

($\langle s9 T^{\wedge} s6^{\wedge} \rangle$) representing the output signal of “0”. It is shown in Figure 6 the simulation result with the input signal of

“00” ($2 \times$ Input strand 0, representing by molecule labelled with P7); the simulation result with the input signal of “01” is indicated in Figure 7, where the output is $1 \times$ Input strand 0 and $1 \times$ Input 1 strand representing by molecule labelled with P1); the simulation result with the input signal of “11” is shown in Figure 8. It is worth to point out that in Figures 6, 7 and 8, there are large margins to distinguish the right result from the wrong. It means the molecular system for performing computation of AND logic gate is with high confidence level.

The simulation results of XOR logic gate are shown in Figures 9, 10 and 11. In the figures, red curves represent the amount of molecules representing positive signal “1” and green curves represent the amount of molecules representing negative signal “0”. The detailed amounts of involved DNA molecules go as shown in Table 1.

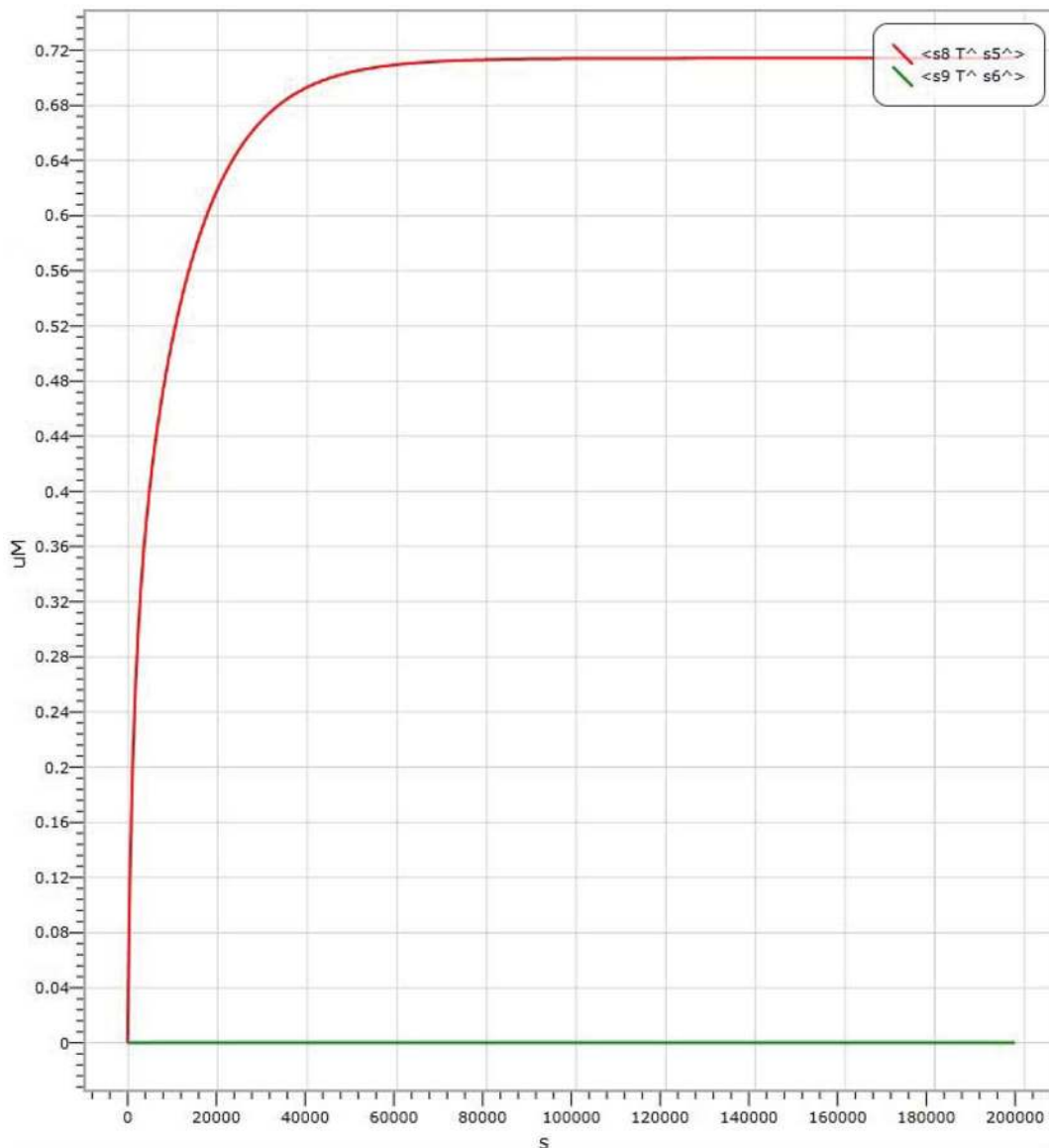


Figure 8. Visual DSD simulations of AND logic gate with input signal “11”.
doi:10.1371/journal.pone.0108856.g008

Experiments

Materials

DNA Oligodeoxynucleotide Strands. All the DNA strands used in the experiments are purchased and from Sangon Biotech (Shanghai, China) Co., Ltd. Most of them are with PAGE purification expect fluorescent ones. The DNA strands, purified by HPLC, are 3'-labeled respectively with fluorophore 6-FAM (FITC), HEX and 5'-labeled with other corresponding quenchers. Fluorescence spectra of two fluorophores are shown in Figure 12 (Fluorescence SpectraViewer from Thermo Fisher Scientific Inc). It confirms little interference between two sets of wavelengths of excitation (dotted curves) and emission (solid curves).

Reagents and Equipment. All the mixtures are dissolved with ultrapure water. $1 \times$ TAE/Mg²⁺ buffer consists of 40 mM Tris (pH 7.6), 2 mM EDTA, 20 mM acetic acid and 35 mM magnesium acetate. The whole reactions are occurred in the real-time PCR from Xi'An TianLong Science and Technology Co., Ltd.

Oligonucleotide Sequences. We design 26 different DNA strands with reusable domains for both AND logic and XOR logic of “DNA neurons”. Sequences of the strands from P1 to P26 and reusable domains from s1 to s10 are listed in Tables 2 and 3. Component samples of “DNA neurons” constructed from T1 to T16 are shown in Table 4 and compositions of fluorescent probes highlight in Table 5.

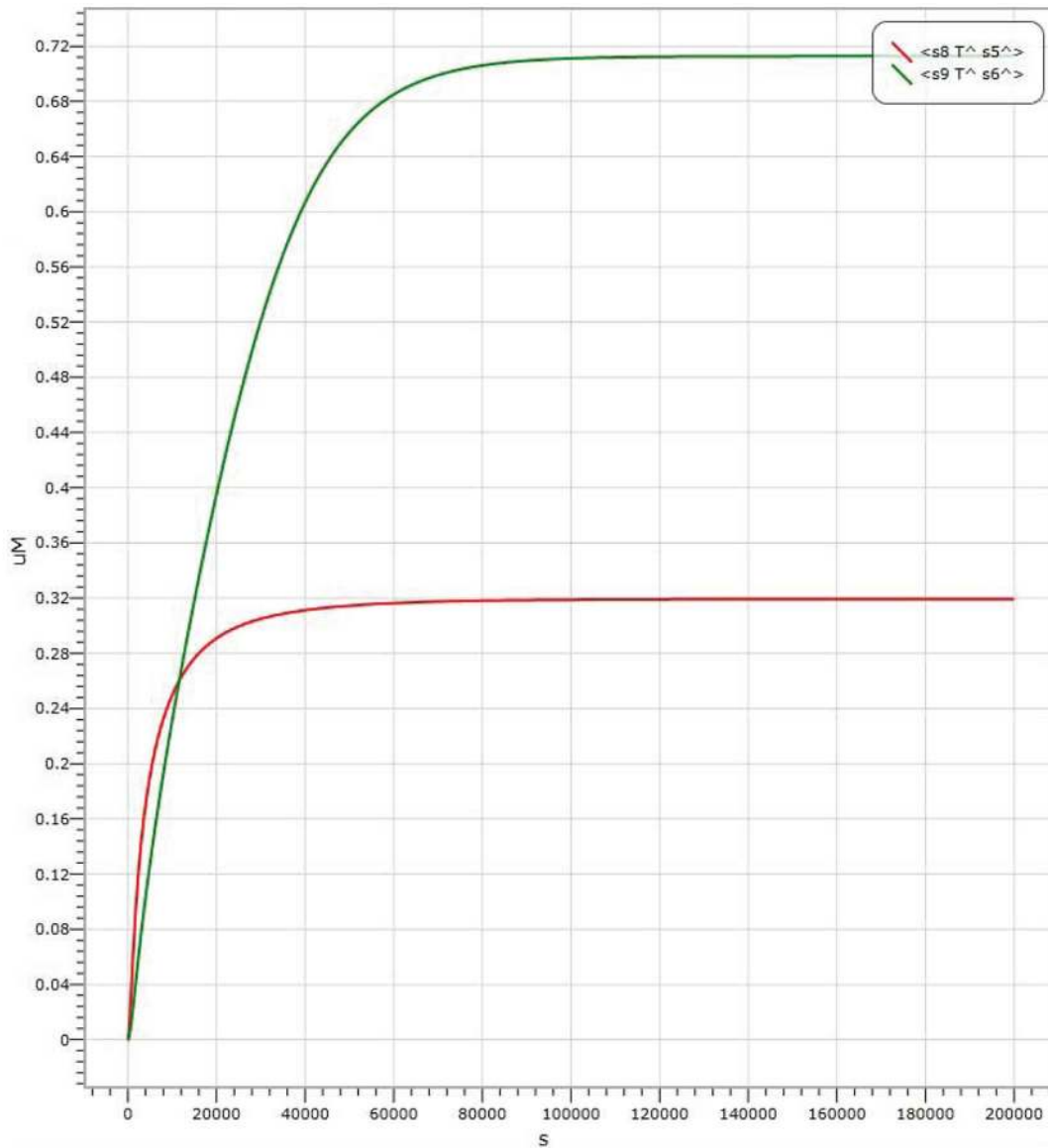


Figure 9. Visual DSD simulations of XOR logic gate with input signal “00”.
doi:10.1371/journal.pone.0108856.g009

Experimental Process

The standard concentration, $1 \times$ DNA strands, is 20 nM.

- To form a double strand, two specified single DNA strands, named strand 1 and strand 2, are added into 20 μ l of solution which contains 4 μ M strand a, 4 μ M strand b, 0.5 \times TAE/Mg²⁺+buffer. There are ten different solutions included double strands that are formed by strand a and strand b. They are corresponding respectively to P2 and P3 for T2, P4 and P5 for T3, P8 and P9 for T6, P10 and P11 for T7, P13 and P14 for T9, P15 and P16 for T10, P18 and P19 for T12, P20 and P21 for T13, P23 and P24 for T15, P25 and P26 for T16. All the solutions are incubated at 95 °C for 3 minutes and then cooled down to 4 °C for 16 hours.
- Mix with $1 \times$ P15, $1 \times$ P20, $1 \times$ T15, $1 \times$ T16 and 0.5 \times TAE/Mg²⁺+ buffer into the standard solution. Incubate at 25 °C for more than 8 hours.
- 100 μ l mixture of a DNA neuron AND logic includes $1 \times$ T10, $1 \times$ T13, 1.5 \times T9, 0.5 \times T12, 2 \times P17, 2 \times P22, $1 \times$ T15, $1 \times$ T16 and 0.5 \times TAE/Mg²⁺+ buffer. Each of the three solutions has the different concentration of input strands. The mixture of the input signal “11” contains 2 \times P1; the mixture of the input “01/10” contains $1 \times$ P1, $1 \times$ P7; the mixture of the input “00” contains 2 \times P7. All the mixtures incubate in the qRT-PCR at 25 °C for 8 hours.
- A DNA neuron for XOR logic gate is 100 μ l mixture of XOR (a) and XOR (b). XOR (a) solution includes 1.5 \times T2, 1.5 \times T3, 1.5 \times T6, 1.5 \times T7, 5 \times P6, 5 \times P12 and 0.5 \times

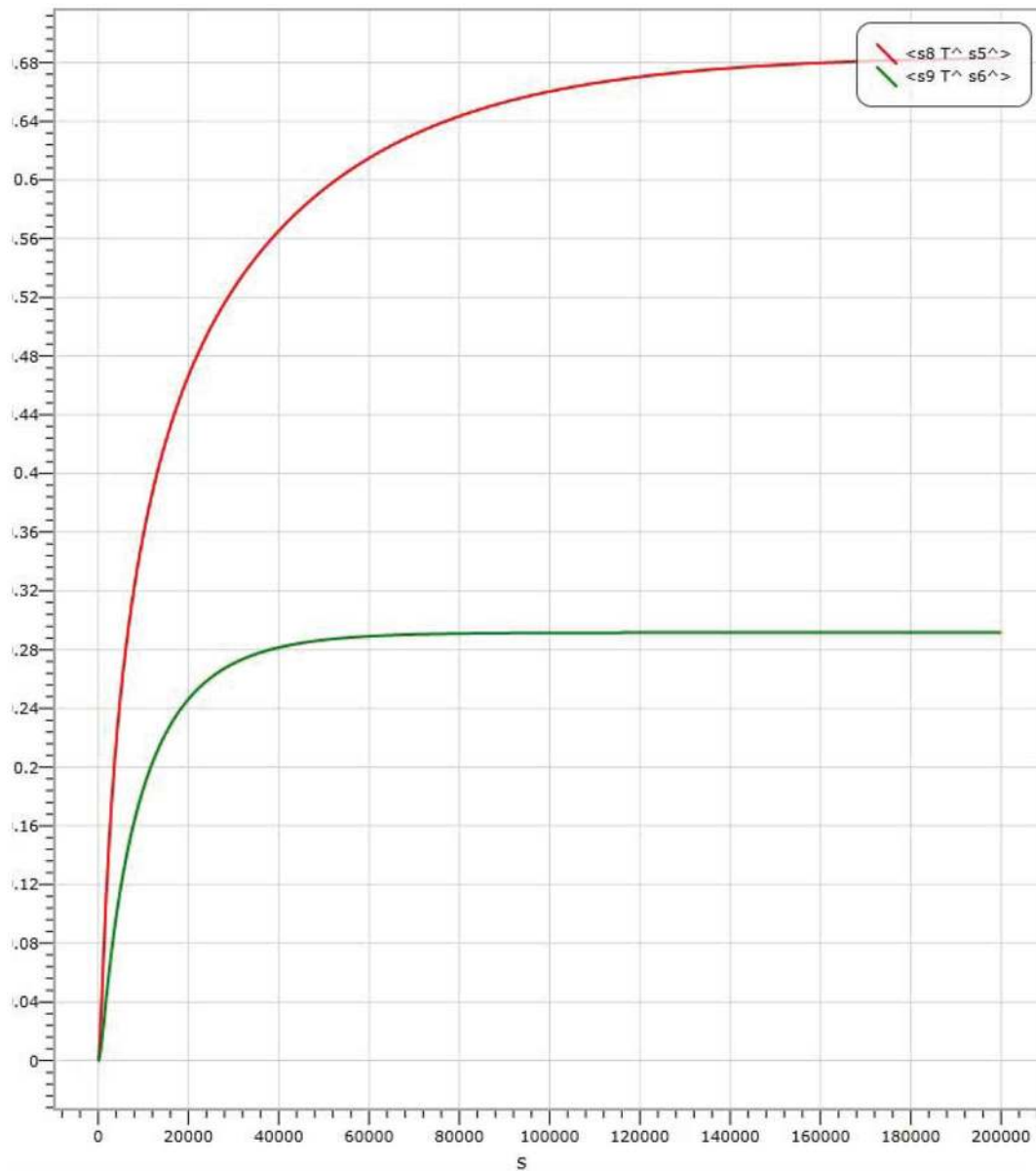


Figure 10. Visual DSD simulations of XOR logic gate with input signal “01”.
doi:10.1371/journal.pone.0108856.g010

TAE/Mg²⁺+buffer. XOR (b) includes 1 × T10, 1 × T13, 1.5 × T9, 0.5 × T12, 2 × P17, 2 × P22, 1 × T15, 1 × T16 and 0.5 × TAE/Mg²⁺+buffer. Then put two solutions XOR (a) and XOR (b) together as XOR logic mixture.

Experimental Results

Designed for the experiment, the solutions containing specific DNA strands deploy “DNA neurons”. Two fluorescent probes T15 and T16 compose reporter “1” and reporter “0” to detect the output signal of “DNA neurons”. The standard solution is for measures of fluorescent detection of two channels. P2 and P4

represent input signals for “DNA neurons”; P1 corresponds to positive signal “1” and P7 represents negative signal “0” of AND logic gate shown in Figure 4.

Fluorescence data of DNA neurons execute AND logic are shown in Figures 13, 14 and 15 with input signals “00”, “01” and “11”, respectively. As it is shown in Figure 13, the relative intensity of FAM signal that represents negative output signal “0” increases gradually from about 10% up to about 70%, while the HEX signal that represents positive output signal “1” keeps at about 10% to 20% in the sample with input signal “00”. It means the output signal is “0”. In Figure 15, the plot of input signal “11” displays that HEX signal intensity of positive signal “1” increases gradually from about 10% up to about 70%, while FAM signal intensity keeps below 20%. In

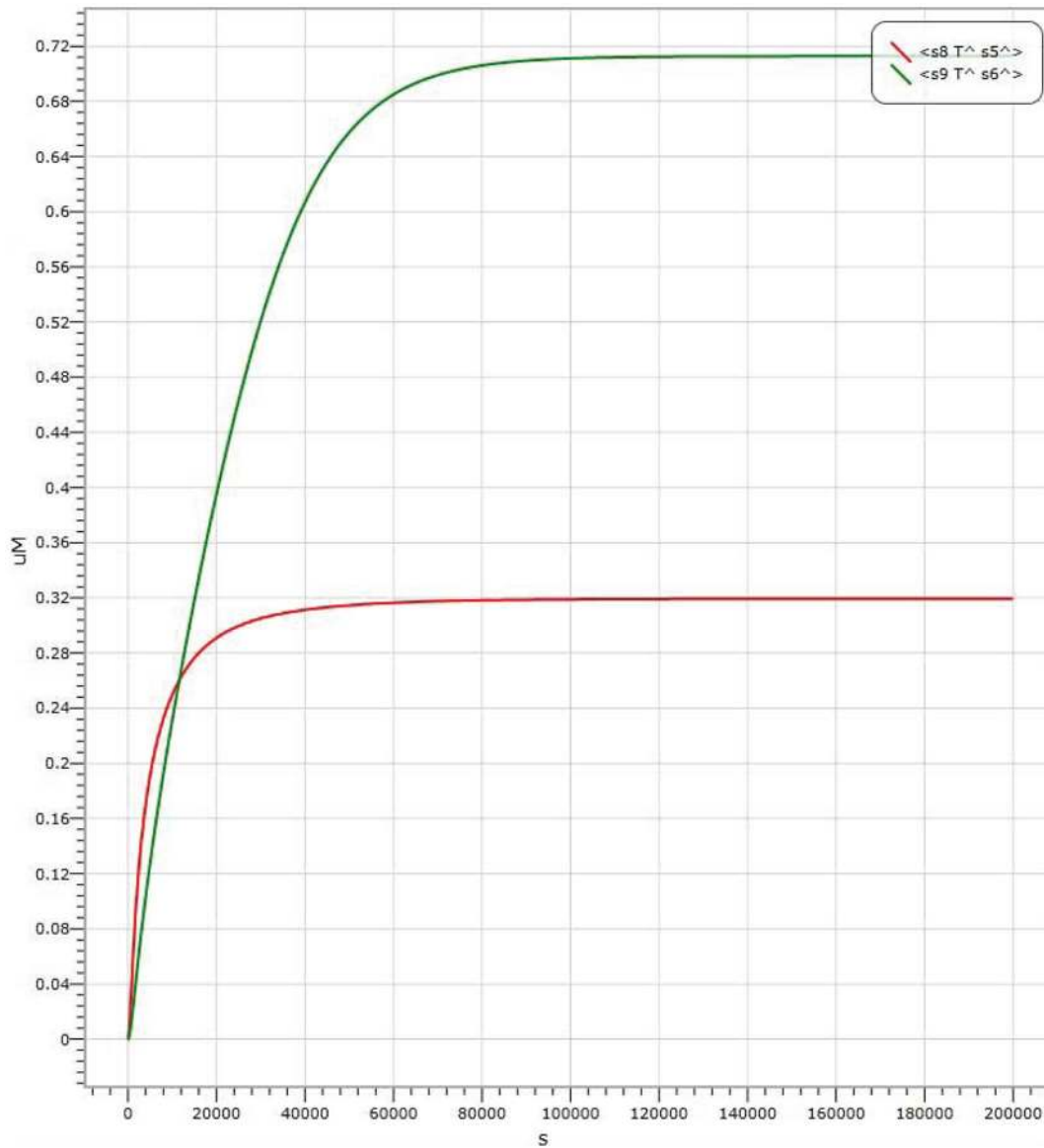


Figure 11. Visual DSD simulations of XOR logic gate with input signal “11”.
doi:10.1371/journal.pone.0108856.g011

Figure 14, the situation is illustrated in the sample with input “01/10”. The intensities of both HEX and FAM signals has the parallel trace and the similar growth during the reaction, while the FAM signal growth two times faster than HEX

signal. It is shown that all the correct strands can be detected from the AND logic gate.

In the solution of XOR logic indicated in Figure 5, the calculation in XOR (b) is the same with the AND logic gate. The

Table 1. The detailed amounts of involved DNA molecules in XOR logic gate.

| Name | Gate a1 | Gate a2 | Gate a3 | Gate a4 | Fuel a1 | Fuel a2 |
|--------|-------------|---------|-------------|---------|---------|---------|
| Amount | 1.5 × | 1.5 × | 1.5 × | 1.5 × | 5 × | 5 × |
| Name | Threshold 1 | Gate b1 | Threshold 0 | Gate b0 | Fuel b1 | Fuel b0 |
| Amount | 1.8 × | 1 × | 0.9 × | 1 × | 2 × | 2 × |

doi:10.1371/journal.pone.0108856.t001

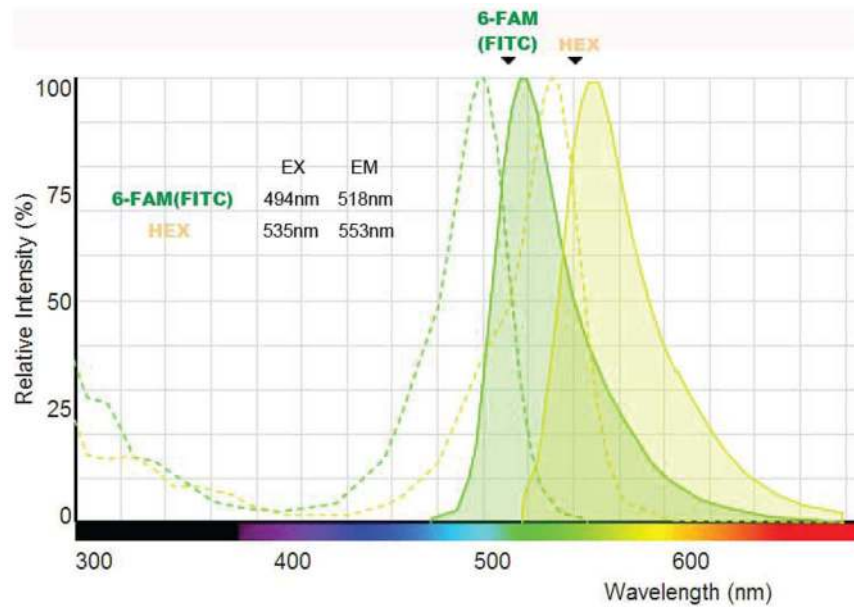


Figure 12. Fluorescence spectra of 6-FAM(FITC) and HEX.
doi:10.1371/journal.pone.0108856.g012

Table 2. All Oligonucleotides for “DNA neurons”.

| ID | Domains | Sequence (5'-3') |
|-----|---------------|-------------------------------------|
| P1 | s1+T+s3 | CACCCTAAAATCTCATCTCACATAACA |
| P2 | s5+T+s1 | CATCCATTCCACTCATCTCACCTAAAATCTCA |
| P3 | s3'+T'+s1'+T' | TGTTATGTGAGATGAGATTTTAGGGTGAGA |
| P4 | s6+T+s1 | CACCACAAAATTCATCTCACCTAAAATCTCA |
| P5 | T'+s1'+T' | AGATGAGATTTTAGGGTGAGA |
| P6 | s7+T+s1 | CAACATATCAATTCATCTCACCTAAAATCTCA |
| P7 | s2+T+s4 | CATAACACAATCACATCTCAAAAACAAA |
| P8 | s5+T+s2 | CATCCATTCCACTCATCTCATAACACAATCACA |
| P9 | s4'+T'+s2'+T' | TTGTTTTGAGATGTGATTGTGTTATGAGA |
| P10 | s6+T+s2 | CACCACAAAATTCATCTCATAACACAATCACA |
| P11 | T'+s2'+T' | AGATGTGATTGTGTTATGAGA |
| P12 | s7+T+s2 | CAACATATCAATTCATCTCATAACACAATCACA |
| P13 | s5 | CATCCATTCCACTCA |
| P14 | T'+s5' | AGATGAGTGAATGGATG |
| P15 | s8+T+s5 | CACCATCAAAATACATCTCATCCATTCCACTCA |
| P16 | T'+s5'+T' | AGATGAGTGAATGGATGAGA |
| P17 | s7+T+s5 | CAACATATCAATTCATCTCATCCATTCCACTCA |
| P18 | s6 | CACCACAAAATTCATA |
| P19 | T'+s6' | AGATGAAGTTTGGTGGTG |
| P20 | s9+T+s6 | CACTAACATACAACATCTCACCCACAAAATTCATA |
| P21 | T'+s6'+T' | AGATGAAGTTTGGTGGTGAGA |
| P22 | s7+T+s6 | CAACATATCAATTCATCTCACCCACAAAATTCATA |
| P23 | PC+T'+s8' | TGAGATGTTATTTGATGGTG |
| P24 | s8 | CACCATCAAAATAACA |
| P25 | PC+T'+s9' | TGAGATGTTGATGTTAGTG |
| P26 | s9 | CACTAACATAACAACA |

doi:10.1371/journal.pone.0108856.t002

Table 3. Domains of “DNA neurons”.

| Name | Sequence | Length |
|------|------------------|--------|
| T | TCT | 3 |
| PC | TG | 2 |
| s1 | CACCCCTAAAATCTCA | 15 |
| s2 | CATAACACAATCACA | 15 |
| s3 | CACATAACA | 9 |
| s4 | CAAAACAAA | 9 |
| s5 | CATCCATTCCACTCA | 15 |
| s6 | CACCACCAAATTCA | 15 |
| s7 | CAACATATCAATTCA | 15 |
| s8 | CACCATCAAATAACA | 15 |
| s9 | CACTAACATACAACA | 15 |

doi:10.1371/journal.pone.0108856.t003

Table 4. Components samples.

| Sample ID | Formation | Function |
|-----------|-----------|-------------|
| T1 | P1 | Input 1 |
| T2 | P2 P3 | Gate a1 |
| T3 | P4 P5 | Gate a2 |
| T4 | P6 | Fuel a1 |
| T5 | P7 | Input 0 |
| T6 | P8 P9 | Gate a3 |
| T7 | P10 P11 | Gate a4 |
| T8 | P12 | Fuel a2 |
| T9 | P13 P14 | Threshold 1 |
| T10 | P15 P16 | Gate b1 |
| T11 | P17 | Fuel b1 |
| T12 | P18 P19 | Threshold 0 |
| T13 | P20 P21 | Gate b0 |
| T14 | P22 | Fuel b0 |
| T15 | P23 P24 | Reporter 1 |
| T16 | P25 P26 | Reporter 0 |

doi:10.1371/journal.pone.0108856.t004

Table 5. Probes with Fluorophores.

| Strand | Probe | Sequences (5'-3') |
|--------|------------|------------------------------|
| P23 | Reporter 1 | TGAGATGTTATTGATGGTG/3HEX/ |
| P24 | Reporter 1 | /5IAbHQ/CACCATCAAATAACA |
| P25 | Reporter 0 | TGAGATGTTGTATGTTAGTG/36-FAM/ |
| P26 | Reporter 0 | /5IAbFQ/CACTAACATACAACA |

doi:10.1371/journal.pone.0108856.t005

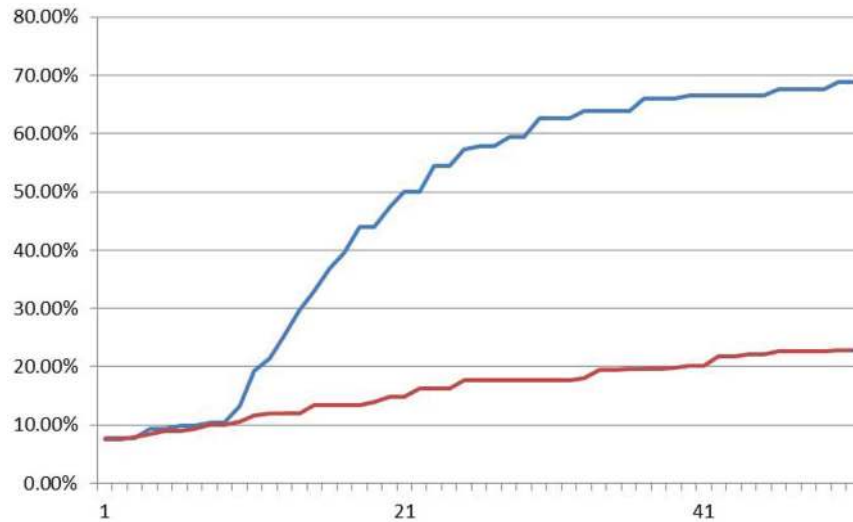


Figure 13. Fluorescence data of DNA Neurons execute AND logic with input “00”. The X-axis is cycling time of Real-time PCR, the time span of each cycle is 10 minutes, temperature of each cycle keeps in 24–25°C. The Y-axis is relative intensity of HEX (red curve) and FAM (blue curve). doi:10.1371/journal.pone.0108856.g013

differences are the changes of the input signals and the addition of XOR (a) including Gate a1, a2, a3, a4, Fuel a1 and a2. By mixing XOR (a) and XOR (b), the molecular system are cascaded as XOR logic gate.

Experimental results by fluorescence data of XOR logic gate are shown in Figures 16, 17 and 18 with input signals “00”, “01/10” and “11” respectively. It demonstrates in Figure 16 that fluorescent signal intensity of FAM (representing output signal “0”) increases gradually from zero to about 80% and that of HEX (representing output signal “1”) keeps below 40% in the sample with input signal “00”. A similar case happens in the sample with input signal “11” shown in Figure 18. The fluorescent signal intensity of FAM (representing output signal

“0”) keeps below 40%, but signal intensity HEX (representing output signal “1”) increases gradually from zero to about 80%. In the sample with input signal “01/10”, it performs that intensity of HEX increases gradually from about zero up to above 60% and that of FAM keeps below 30% see Figure 17. All the plots clarify that it takes out correct strands through the XOR logic.

Author Contributions

Conceived and designed the experiments: XS TS CD. Performed the experiments: XS TS. Analyzed the data: ZC LP. Contributed reagents/materials/analysis tools: ZC LP ZW. Wrote the paper: XS TS.

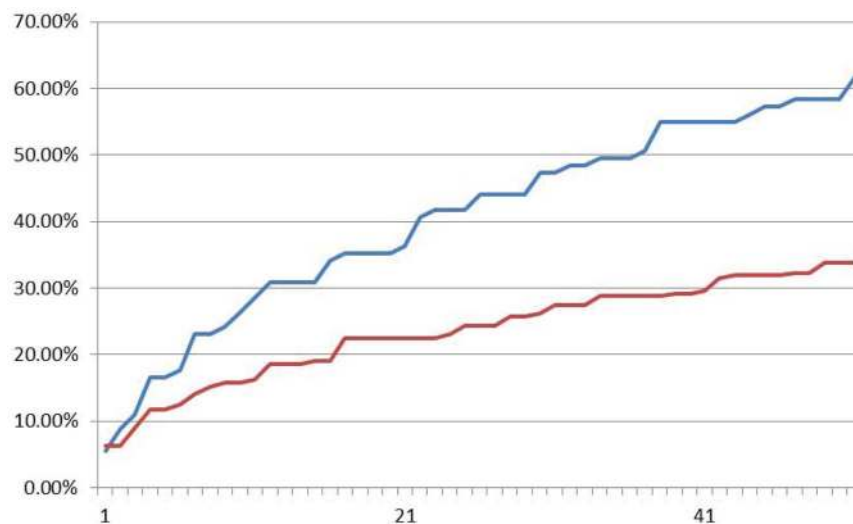


Figure 14. Fluorescence data of DNA Neurons execute AND logic with input “01”. The X-axis is cycling time of Real-time PCR, the time span of each cycle is 10 minutes, temperature of each cycle keeps in 24–25°C. The Y-axis is relative intensity of HEX (red curve) and FAM (blue curve). doi:10.1371/journal.pone.0108856.g014

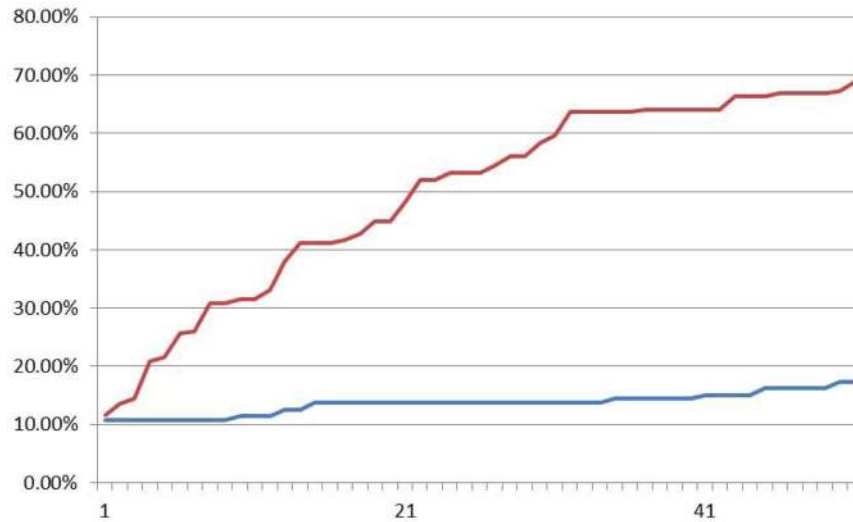


Figure 15. Fluorescence data of DNA Neurons execute AND logic with input "11". The X-axis is cycling time of Real-time PCR, the time span of each cycle is 10 minutes, temperature of each cycle keeps in 24–25°C. The Y-axis is relative intensity of HEX (red curve) and FAM (blue curve). doi:10.1371/journal.pone.0108856.g015

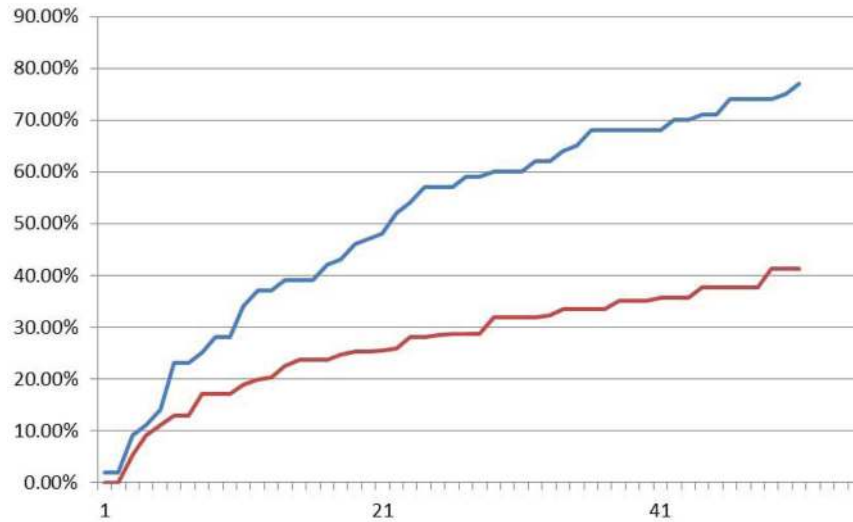


Figure 16. Fluorescence data of DNA Neurons execute XOR logic with input "00". The X-axis is cycling time of Real-time PCR, the time span of each cycle is 10 minutes, temperature of each cycle keeps in 24–25°C. The Y-axis is relative intensity of HEX (red curve) and FAM (blue curve). doi:10.1371/journal.pone.0108856.g016

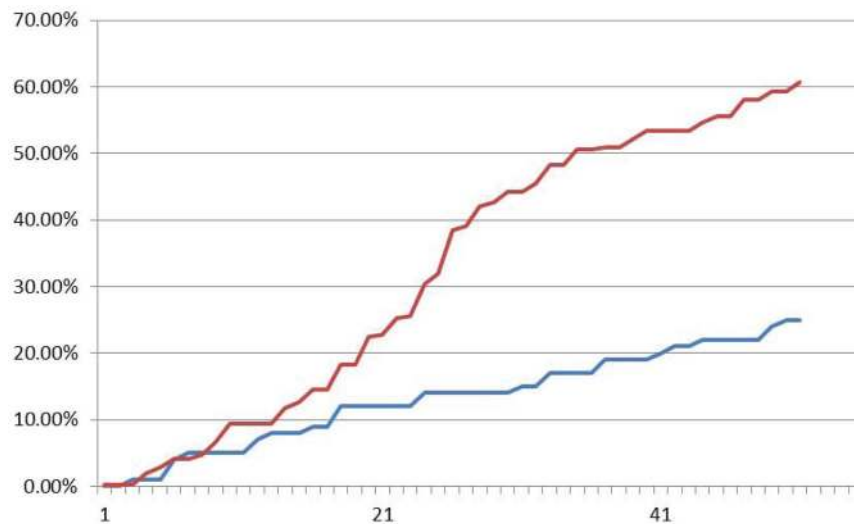


Figure 17. Fluorescence data of DNA Neurons execute XOR logic with input "01". The X-axis is cycling time of Real-time PCR, the time span of each cycle is 10 minutes, temperature of each cycle keeps in 24–25°C. The Y-axis is relative intensity of HEX (red curve) and FAM (blue curve). doi:10.1371/journal.pone.0108856.g017

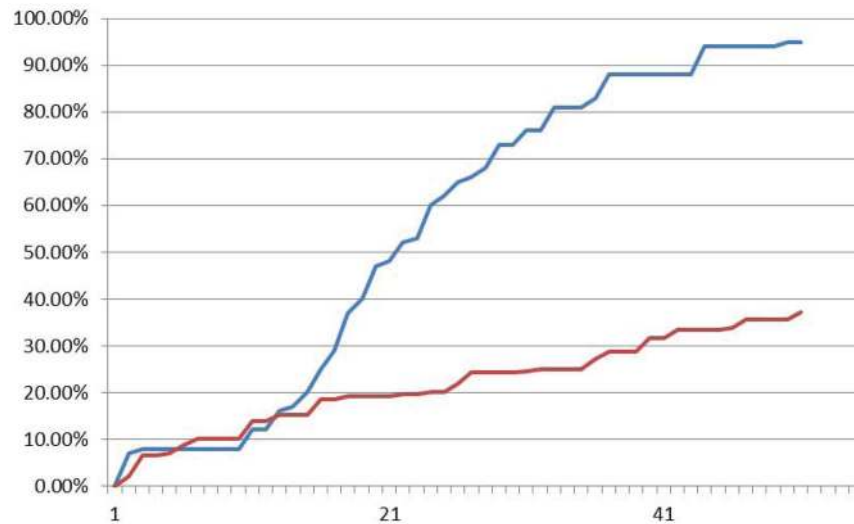


Figure 18. Fluorescence data of DNA Neurons execute XOR logic with input "11". The X-axis is cycling time of Real-time PCR, the time span of each cycle is 10 minutes, temperature of each cycle keeps in 24–25°C. The Y-axis is relative intensity of HEX (red curve) and FAM (blue curve). doi:10.1371/journal.pone.0108856.g018

References

- Adleman LM (1994) Molecular computation of solutions to combinatorial problems. *Science-AAAS-Weekly Paper Edition* 266: 1021–1023.
- Rothemund PW, Papadakis N, Winfree E (2004) Algorithmic self-assembly of DNA sierpinski triangles. *PLoS biology* 2: e424.
- Benenson Y, Paz-Elizur T, Adar R, Keinan E, Livneh Z, et al. (2001) Programmable and autonomous computing machine made of biomolecules. *Nature* 414: 430–434.
- Păun G (2002) *Membrane computing: an introduction*. Springer.
- Paun G, Rozenberg G, Salomaa A (2010) *The Oxford handbook of membrane computing*. Oxford University Press, Inc.
- Hampp N (2000) Bacteriorhodopsin as a photochromic retinal protein for optical memories. *Chemical Reviews* 100: 1755–1776.
- Stuart JA, Marcy DL, Wise KJ, Birge RR (2002) Volumetric optical memory based on bacteriorhodopsin. *Synthetic metals* 127: 3–15.
- Rowan AE, Nolte RJ (1998) Helical molecular programming. *Angewandte Chemie International Edition* 37: 63–68.
- Farsad N, Guo W, Eckford AW (2013) Tabletop molecular communication: text messages through chemical signals. *PLoS one* 8: e82935.
- Brett BT, Berquam-Vrieze KE, Nannapaneni K, Huang J, Scheetz TE, et al. (2011) Novel molecular and computational methods improve the accuracy of insertion site analysis in sleeping beauty-induced tumors. *PLoS one* 6: e24668.
- Winfree E, Liu F, Wenzler LA, Seeman NC (1998) Design and self-assembly of two-dimensional DNA crystals. *Nature* 394: 539–544.
- de Vries SJ, Zacharias M (2012) Attract-em: A new method for the computational assembly of large molecular machines using cryo-em maps. *PLoS one* 7: e49733.
- Walker GT, Fraiser MS, Schram JL, Little MC, Nadeau JG, et al. (1992) Strand displacement amplification: an isothermal, in vitro DNA amplification technique. *Nucleic Acids Research* 20: 1691–1696.
- Zhang DY, Seelig G (2011) Dynamic DNA nanotechnology using strand-displacement reactions. *Nature chemistry* 3: 103–113.

15. Nielsen PE, Egholm M, Berg RH, Buchardt O (1991) Sequence-selective recognition of DNA by strand displacement with a thymine-substituted polyamide. *Science* 254: 1497–1500.
16. Little MC, Andrews J, Moore R, Bustos S, Jones L, et al. (1999) Strand displacement amplification and homogeneous real-time detection incorporated in a second-generation DNA probe system, bdpobecet. *Clinical chemistry* 45: 777–784.
17. LaBean TH, Li H (2007) Constructing novel materials with DNA. *Nano Today* 2: 26–35.
18. Samano EC, Pilo-Pais M, Goldberg S, Vogen BN, Finkelstein G, et al. (2011) Self-assembling DNA templates for programmed artificial biomineralization. *Soft Matter* 7: 3240–3245.
19. Pilo-Pais M, Goldberg S, Samano E, LaBean T, Finkelstein G (2011) Connecting the nanodots: programmable nanofabrication of fused metal shapes on DNA templates. *Nano letters* 11: 3489–3492.
20. Shi X, Li X, Zhang Z, Xu J (2005) Improve capability of DNA automaton: DNA automaton with three internal states and tape head move in two directions. In: *Advances in Intelligent Computing*, Springer. pp. 71–79.
21. Xiaolong S, Linqiang P, Jin X, et al. (2006) General DNA automaton model with r/w tape. In: *Computational Intelligence and Bioinformatics*, Springer. pp. 258–266.
22. Wang Y, Hu P, Shi X, Cui G (2012) DNA self-assembly for graph vertex 3-coloring problem. *Journal of Computational and Theoretical Nanoscience* 9: 2086–2092.
23. Seelig G, Soloveichik D, Zhang DY, Winfree E (2006) Enzyme-free nucleic acid logic circuits. *science* 314: 1585–1588.
24. Poli R, McPhee NF, Rowe JE (2004) Exact schema theory and markov chain models for genetic programming and variable-length genetic algorithms with homologous crossover. *Genetic Programming and Evolvable Machines* 5: 31–70.
25. Qian L, Winfree E (2011) Scaling up digital circuit computation with DNA strand displacement cascades. *Science* 332: 1196–1201.
26. Lubrich D, Green SJ, Turberfield AJ (2009) Kinetically controlled self-assembly of DNA oligomers. *Journal of the American Chemical Society* 131: 2422–2423.
27. Phillips A, Cardelli L (2009) A programming language for composable DNA circuits. *Journal of the Royal Society Interface* 6: S419–S436.