#### ORIGINAL RESEARCH

## A novel mutation panel for predicting etoposide resistance in small-cell lung cancer

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**Purpose:** Platinum-based chemotherapy, consisting of etoposide and cisplatin (EP), has been the cornerstone of therapy for extensive-stage small-cell lung cancer (ES-SCLC) for decades. Despite the marked initial sensitivity of SCLC to chemotherapy, EP regimens cannot avoid the emergence of drug resistance in clinical practice. With the rise of new chemotherapy regimens in recent years and the primary resistance or insensitivity of ES-SCLC to EP regimens, it is desirable to be able to identify patients with resistant or insensitive ES-SCLC.

**Methods:** The sequencing and drug sensitivity data of SCLC cell lines were provided by The Genomics of Drug Sensitivity in Cancer Project (GDSC). The data regarding sensitivity to etoposide of 54 SCLC cell lines were analyzed, and etoposide-sensitive cell lines and etoposide-resistant cell lines were differentiated according to the IC50 values defined by the GDSC. ROC curve analysis was performed on all mutations and combinations of mutations to select the optimal panel to predict resistance to etoposide.

**Results:** ROC analysis of etoposide resistance revealed that the most significant single gene mutation indicating resistance to etoposide was *CSMD3*, and the accuracy of predicting resistance to etoposide proved to be the highest when there was any mutation in *CSMD3/ PCLO/RYR1/EPB41L3*, area under the curve =0.804 (95% confidence interval: 0.679–0.930, P<0.001).

**Conclusion:** This study found that a panel with four genes (*CSMD3, EPB41L3, PCLO, and RYR1*) can accurately predict sensitivity to etoposide. These findings provide new insights into the overall treatment for patients with ES-SCLC that is resistant or insensitive to etoposide. **Keywords:** small-cell lung carcinoma, etoposide, EP regimens, IP regimens, gene mutation

#### Introduction

In recent years, humans have made significant progress in the early detection, early diagnosis, early treatment, and even prevention of cancer. However, lung cancer is the most commonly diagnosed cancer (11.6%) and the leading cause of cancer-related death (18.4%) worldwide.<sup>1</sup> Currently, there are approximately 2.1 million lung cancer patients worldwide.<sup>1</sup> Approximately 12–15% of new lung cancer patients are diagnosed with small-cell lung cancer (SCLC).<sup>2,3</sup> According to the latest National Comprehensive Cancer Network (NCCN) Guidelines, an estimated 29,654 new cases of SCLC occurred in the United States in 2017.<sup>4,5</sup> Studies have shown that the incidence of SCLC is attributable to cigarette smoking, and the smoking pack-years increases, so does the risk of SCLC. Ninety percent of patients with SCLC have been or are currently smokers, and smoking duration is positively associated with an increased risk of SCLC.<sup>6,7</sup> In addition, SCLC is characterized by a high growth fraction, a high degree of malignancy, and the early development of

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© 2019 Qiu et al. This work is published and licensed by Dove Medical Press Limited. The full terms of this license are available at https://www.dovepress.com/terms.php you hereby accept the Terms. Non-commercial uses of the work are permitted without any further permission from Dove Medical Press Limited, provided the work is properly attributed. For permission for commercial uses of this work, please see paragraphs 4.2 and 5 of our Terms (http://www.dovepress.com/terms.php). widespread metastases.<sup>8,9</sup> The 5-year survival rate in patients with SCLC is only 6.6%. Currently, SCLC is divided into limited-stage SCLC (LS-SCLC) and extensive-stage SCLC (ES-SCLC). Unfortunately, the 5-year survival rates are only 1.6% and 12.1% for patients with ES-SCLC (1/3) and ES-SCLC (2/3),<sup>8-11</sup> respectively.

At present, surgery is one of the main methods of cancer treatment, but it is rarely used in the treatment of patients with SCLC. It is only suitable for a small number of stage I patients with SCLC (2%-5%) who do not have mediastinal lymph node metastasis. In the past few decades, a platinum compound in combination with the topoisomerase-II inhibitor etoposide beyond 4 to 6 cycles of chemotherapy (EP) has become the cornerstone of treatment for patients with ES-SCLC for palliative care.<sup>11-13</sup> In recent years, the chemotherapy for ES-SCLC has mainly been irinotecan, cisplatin (IP) and EP regimens.<sup>14</sup> Despite the substantial initial sensitivity of SCLC to chemotherapy in the early stages of treatment, more than 90% of patients eventually develop clinical drug resistance and die as a result of relapse.<sup>8,9</sup> At present, there is a great deal of controversy about the therapeutic effect and safety tolerance of IP and EP in the treatment of ES-SCLC. In 2002, a randomized, multicenter, phase III trial (J9511) performed in Japan reported that patients with ES-SCLC who were treated with IP experienced a median survival of 12.8 months compared with 9.4 months for patients treated with EP (P=0.002). In addition, the 1-year survival rates were 58.4% vs 37.7% and the median progression-free survival (PFS) rates were 12.8 months vs 9.4 months in the IP and EP groups, respectively.<sup>15</sup> Furthermore, Hermes et al studied 220 patients with ES-SCLC, and the results showed that the median overall survival (OS) was slightly higher in those receiving IP than in those receiving EP (8.5 months vs 7.1 months, P=0.04).<sup>16</sup> However, it is surprising that there were no significant differences in the efficacy and survival of the IP and EP groups in 4 subsequent phase III trials.<sup>17-20</sup> In a cohort study from Korea, the median OS and median PFS of patients with ES-SCLC treated with IP were 10.9 months and 6.5 months, respectively, whereas the median OS and PFS in the EP arm were 10.3 months (P=0.120) and 5.8 months (P=0.115), respectively. Similarly, no significant differences were observed in the 1- and 2-year survival rates in the IP versus EP groups. In the subgroup analysis, males, patients <65 years old and patients with Eastern Cooperative Oncology Group performance status (ECOG PS)  $\leq 1$  were treated with IP or EP, and the two groups had significant therapeutic differences. In addition, there was a significant difference in the objective response rate (ORR) between the IP group and the EP group (62.4% vs 48.2%, *P*=0.006).<sup>21</sup>

Currently, 4 to 6 cycles EP is the standard therapy widely used for a majority of SCLC in the clinic, with an ORR of 50%-80%.<sup>22</sup> However, the median OS of patients with ES-SCLC is only 9 months, with only 2% of patients surviving after 5 years.<sup>14,23</sup> Although SCLC usually responds well to chemotherapy regimens in the early stages of treatment, subsequent clinical drug resistance and disease recurrence occur in more than 90% of patients.<sup>8,9</sup> This may be due to the existence of cancer stem cells that are relatively resistant to cytotoxic therapy. Chemotherapy cannot destroy residual tumor cells, leading to a high recurrence rate and a high drug resistance rate in SCLC.<sup>24</sup> Primary resistance or acquired resistance to chemotherapy is a major factor in the poor prognosis of patients with lung cancer.<sup>25–27</sup> In the drug sensitivity data from GDSC, we found that the IC50 of etoposide in the 54 SCLC cell lines ranged from 0.242 µM to 319  $\mu$ M, and the drug resistance cut-off value provided by the website was 16 µM. In total, 65% of patients have SCLC that is sensitive to etoposide, which is close to the response rate for etoposide.<sup>28</sup> Therefore, if we are able to select patients with ES-SLCL that is not sensitive to etoposide before treating them with standard chemotherapy, we could choose a different chemotherapy regimen to treat these patients, hopefully improving survival outcomes in those ES-SCLC patients. Survival time was significantly improved with the new chemotherapy compared with EP. However, there is currently no clinically relevant prediction factor and screening for appropriate means of insensitivity to etoposide.

To date, a growing number of studies have shown that the emergence of primary or acquired platinum and Topoisomerase Inhibitors resistance in EP is associated with certain gene expression changes or/and gene mutations.<sup>29</sup> Chiu et al<sup>30</sup> found that *FBXL7* is a biomarker of poor prognosis in patients with ovarian cancer. A high expression level of FBXL7 is positively associated with a low survival rate in ovarian cancer patients, and the FBXL7 mRNA level and ovarian cancer cell line paclitaxel (PTX) IC50 values were positively correlated, leading to the speculation that the upregulation of FBXL7 expression results in resistant ovarian cancer cell lines. In addition, Chiu et al<sup>31</sup> detected the transcriptional level of the shared gene in HCC38 (PTX-sensitive) and MDA-MB436 (PTX-resistant) TNBC cells posttreatment with paclitaxel. They found that the downregulation of miR-1180 may regulate OTUD7B, ultimately negatively regulating the NF-kB-Lin28 axis. This in turn triggers Let-7

microRNA-mediated caspase-3 downregulation, ultimately leading to resistance to PTX. Based on these findings, the sensitivity and drug resistance of tumor cells to chemotherapy can be predicted by gene expression levels. Thus, patients with ES-SLCL that is sensitive or insensitive to chemotherapy can be further distinguished. We hope that the sensitivity of ES-SCLC to etoposide can be predicted by gene mutation panels, allowing the selection of patients with ES-SCLC that is insensitive to etoposide before standard chemotherapy is administered and the development of personalized, precise chemotherapy to extend patients' OS and improve their quality of life (QOL).

To this end, we analyzed the sequencing and drug sensitivity data for a SCLC cell line through the GDSC database to determine whether mutations can predict the primary resistance to etoposide and try to explain the potential underlying mechanism to provide first-line treatment recommendations for patients with ES-SCLC.

#### Methods

### Drug response, gene expression and mutation data

The natural logarithm half maximal inhibitory concentration (IC50) of all selected erlotinib-related cell lines were obtained from the GDSC (https://www.cancerrxgene.org/). Robust Multichip Average (RMA) normalized expression data from the Affymetrix Human Genome U219 array and gene mutation information found in cell lines by Illumina HiSeq 2000 whole-exome sequencing (WES) were downloaded from the GDSC.

#### Screening of mutated resistance genes

There were 54 SCLC cell lines in the GDSC with drug sensitivity data for etoposide. The GDSC site defined etoposide-resistant cell lines as those with IC50 values  $\geq 16 \ \mu M$  and etoposide-sensitive cell lines as those with IC50 values  $< 16 \ \mu M$ . ROC curve analysis was performed for all mutations, and the cell lines with areas under the curve (AUCs) > 0.5 were selected and randomly combined; then, resistance to etoposide was predicted by the combined mutation panels. The Youden Index values obtained by various combined ROC analyses were sorted to select the best combination.

#### Statistical analysis

The IC50 distribution for etoposide in various cell lines was obtained with the GDSC web tool. ROC analysis and mapping were performed with SPSS 21.0 (IBM SPSS Statistics, IBM Corporation); mutation and gene expression data were analyzed and mapped with the maftools<sup>32</sup> and limma packages<sup>33</sup> in R. In the differential analysis of the gene expression profiles, P<0.05 and FC>1.5 orFC<2/3 were considered to indicate significant differences. The survival analysis was with the log-rank test after the Kaplan-Meier analysis to investigate the predictive ability of a mutation panel with regard to survival. Gene Ontology (GO) annotation analysis and KEGG pathway enrichment analysis of the differentially expressed genes (DEGs) in this study were performed using the Database for Annotation, Visualization and Integrated Discovery (DAVID) (https://david.ncifcrf.gov/).

#### Results

The sensitivity of cancer cell lines to drugs is mainly expressed as the IC50 value, which refers to the concentration of drug that kills half of the tumor cells in vitro. Because the drug concentration is diluted to 1/10 or 1/100, we used lnIC50 values to distinguish between resistant or sensitive cell lines. Based on the GDCS 7.0 database (updated on March 20, 2018), there are 64 SCLC cell lines, but only 54 of them have etoposide susceptibility data (drug sensitivity data), WES mutation data and RNA Seq data.

Using the GDSC website tools, we obtained the IC50 distribution for etoposide by tissue type (Figure 1A). We found that most of the tumors are sensitive to etoposide, and the IC50 values of most cell SCLC lines indicate that they are sensitive to etoposide. By analyzing the IC50 values of the 54 SCLC cell lines shown in Figure 1B, we found that there are 35 cell lines that are sensitive to etoposide, accounting for 64.8% of the total, and their median and mean IC50 values were 2.06 µM (range: 0.242-15.2 µM) and 4.02±4.07 µM, respectively. In total, 19 strains were resistant to etoposide, accounting for 35.2% of the total, and their median and mean IC50 values were 50.0 µM (range: 16.4-319.0 µM) and 71.9  $\pm$ 71.8  $\mu$ M, respectively. The raw data for the IC50 values of all cell lines with regard to etoposide can be found in Table S1.

After sorting the IC50 values for etoposide, we found that in the mutation landscape of the 54 SCLC cell lines (Figure 2), the genes with the highest mutation frequencies were *TP53* (91%), *TTN* (78%) and *Rb1* (70%). Among them, *TP53* and *TTN* mutations were mainly missense mutations, while the *Rb1* mutations were mainly nonsense and splice mutations.

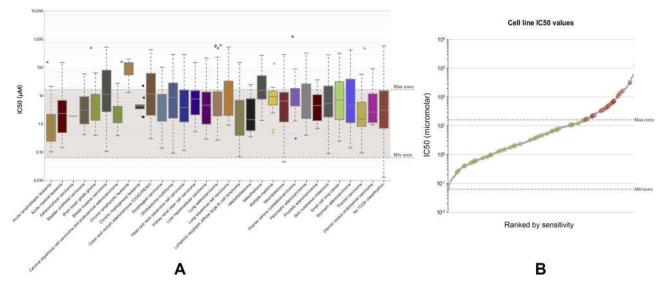


Figure I (A) IC50 distribution for etoposide by tissue type. (B) The scatter plot of IC50 distribution for etoposide of 54 SCLC cell lines. Abbreviation: IC50, half maximal inhibitory concentration.

We performed an ROC analysis of to predict etoposide resistance using all mutated genes (see Table S2). From the ROC curves, we found that the most significant single gene mutation associated with resistance to etoposide was *CSMD3*, with an AUC of 0.697 (P=0.016) (Table 1). By experimenting with different combinations, we found that when any mutations occurred in *CSMD3/PCLO/RYR1/EPB41L3*, the accuracy of predicting resistance to etoposide was the highest (AUC=0.804, 95% CI: 0.679–0.930, P<0.001) (Table 1). The ROC curve results of the panel composed of *CSMD3/PCLO/RYR1/EPB41L3* and the individual genes are shown in Figure 3A.

We performed a log-rank test with the Kaplan–Meier plots according to mutations and clinical follow-up data in 110 SCLCs published by George et al<sup>34</sup> In addition, we found a significantly lower average survival time in patients with CLC with any mutation in CSMD3/PCLO/RYR1/EPB41L3 than in those with no mutations in all four genes ( $35.6\pm5.3$  months vs 76.7±12.1 months, *P*=0.040) (Figure 3B). By analyzing significantly enriched KEGG pathways of DEGs, we found that there was a significant association between both CSMD3 and RYR1 mutations and MAPK signaling pathway (*P*=0.015 and *P*=0.023, respectively) (Table 2).

#### Discussion

EP has been the most common therapy for ES-SCLC for decades. As a standard treatment, it can inhibit tumor proliferation, relieve clinical symptoms, and achieve ideal results.<sup>13,34–37</sup> We found that 19 (35.2%) of the 54 SCLC cell lines were insensitive to etoposide according to

the data from the GDSC. Currently, the clinically accepted ORR of EP is 50–80%.<sup>23</sup> Based on the above findings, the majority of patients with SCLC do not receive survival benefits from EP, indicating that screening for patients with primary resistance to etoposide is necessary. Therefore, this study further analyzed the mutation, gene expression and etoposide sensitivity data of 54 ES-SCLC cell lines obtained from the GDSC. We identified four genes, namely, CSMD3, EPB41L3, PCLO, and RYR1; mutations in these genes predict resistance to etoposide. The predictive sensitivity this four-gene panel for resistance to etoposide is as high as 85%, with 77.8% accuracy when screening for patients with primary etoposide resistance. In addition, the ROC showed an AUC of 0.804 (95% CI 0.679-0.930), and the model was considered to have a high degree of confidence.

Recently, a small phase III trial performed in Japan compared the efficacy of IP and EP in patients with ES-SCLC<sup>15</sup>. The trial results showed a higher median OS (12.8 months vs 9.4 months), 1-year survival rate (58.4% vs 37.7%) and 2-year survival rate (19.5% vs 5.2%) after IP than after EP. In addition, Hermes et al<sup>16</sup> studied 220 patients with ES-SCLC, and the results showed a longer median OS resulting from the IP regimen compared with the EP regimen (8.5 months vs 7.1 months, P=0.04).

We analyzed the data and found that mutations in both *CSMD3* and *RYR1* can cause the activation of the downstream MAPK signaling pathway (Figure 4). In addition, Liu et al<sup>36</sup> found that etoposide activates the MAPK/ERK signaling pathway, inhibits p53 expression and enhances c-Myc expression

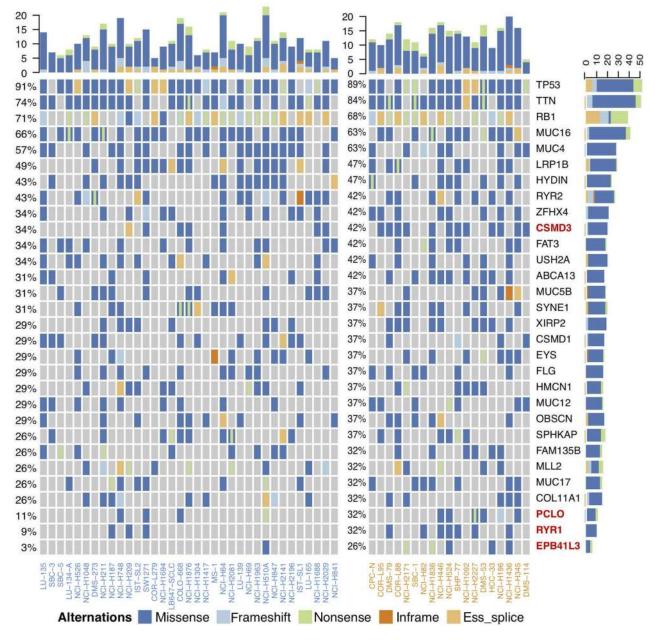


Figure 2 Mutation landscape of 54 SCLC cell lines. Abbreviation: SCLC, small-cell lung cancer.

 Table I Receiver operator characteristic curve analysis for four-gene panel and four genes separately to etoposide resistance status in

 small-cell lung cancer cell lines

Gene	Area under curve	95% confidence interval	Sensitivity	Specificity	Youden index	P-value
CSMD3	0.697	0.546–0.848	0.600	0.794	0.394	0.016
PCLO	0.591	0.429–0.754	0.300	0.882	0.182	0.267
RYR I	0.631	0.469–0.792	0.350	0.912	0.262	0.111
EPB41L3	0.610	0.447–0.774	0.250	0.971	0.221	0.179
Panel	0.804	0.679–0.930	0.850	0.706	0.556	<0.001

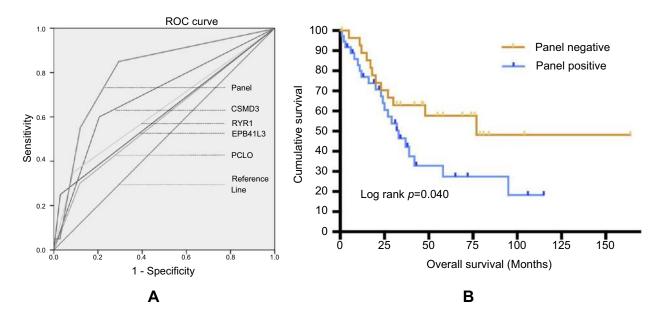


Figure 3 (A) ROC curve of the panel and four mutations; (B) Kaplan-Meier overall survival analyses for the four-gene panel in clincal trial of SCLC. Abbreviation: SCLC, small-cell lung cancer.

Mutation	Term	Count	P-value
CSMD3	hsa04142: Lysosome	8	0.002
	hsa04010: MAPK signaling pathway	10	0.015
	hsa05230: Central carbon metabolism in cancer	5	0.016
	hsa04610: Complement and coagulation cascades	5	0.021
	hsa01130: Biosynthesis of antibiotics	8	0.044
EPB41L3	hsa01200: Carbon metabolism	8	0.003
	hsa01130: Biosynthesis of antibiotics	11	0.004
	hsa01100: Metabolic pathways	33	0.010
	hsa00020: Citrate cycle (TCA cycle)	4	0.015
	hsa04730: Long-term depression	5	0.020
	hsa04130: SNARE interactions in vesicular transport	4	0.021
	hsa04720: Long-term potentiation	5	0.028
	hsa03022: Basal transcription factors	4	0.044
	hsa04726: Serotonergic synapse	6	0.045
PCLO	hsa04810: Regulation of actin cytoskeleton	11	<0.001
	hsa04151: PI3K-Akt signaling pathway	12	0.005
	hsa04510: Focal adhesion	9	0.005
	hsa04512: ECM-receptor interaction	6	0.005
	hsa03320: PPAR signaling pathway	5	0.011
	hsa05205: Proteoglycans in cancer	8	0.016
	hsa05160: Hepatitis C	6	0.031
	hsa05231: Choline metabolism in cancer	5	0.044
RYRI	hsa00500: Starch and sucrose metabolism	3	0.019
	hsa04010: MAPK signaling pathway	6	0.023
	hsa04960: Aldosterone-regulated sodium reabsorption	3	0.026
	hsa00280: Valine, leucine and isoleucine degradation	3	0.037
	hsa01130: Biosynthesis of antibiotics	5	0.048

Table 2 Significantly enriched KEGG pathways of	of DEGs
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Abbreviations: MAPK, mitogen activated kinase-like protein; TCA, tricarboxylic acid; SNARE, small NF90 (ILF3) associated RNA E; PI3K-Akt:phosphoinositide-3-kinase/ serine threonine kinase; ECM, extracellular matrix; PPAR, peroxisome proliferators-activated receptors.

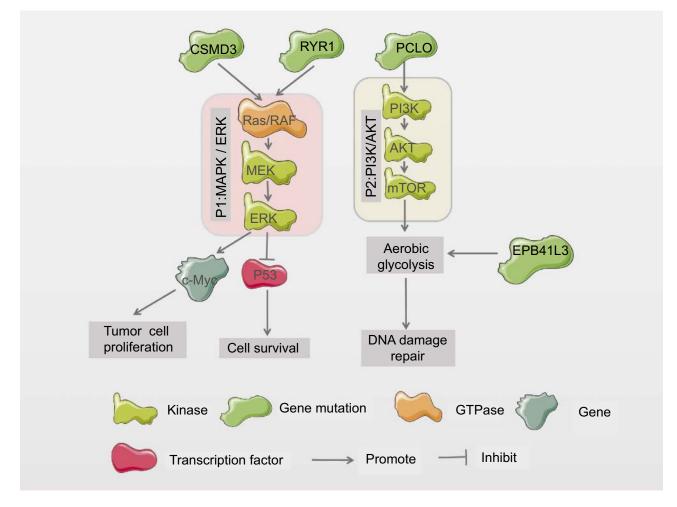


Figure 4 Potential mechanism of the four-gene panel to predict the resistance of etoposide in SCLC. Abbreviation: SCLC, small-cell lung cancer.

to decrease the sensitivity of gastric cancer cells to chemotherapy in. Therefore, we hypothesized that mutations in the CSMD3 and RYR1 genes may cause a significant resistance to etoposide in ES-SCLC via the downstream MAPK signaling pathway. It is well known that etoposide induces DNA double-strand breakage (DSB) and triggers the DNA damage response by activating the ataxia telangiectasia-mutated gene (ATM) DNA repair is a process of energy dissipation, and ATP-dependent chromatin remodeling complexes participate in DSB repair.<sup>37</sup> In aerobic conditions, tumor cells preferentially perform glycolysis rather than providing energy for cell growth through the more efficient oxidative phosphorylation pathway and are therefore characterized by high glucose uptake, glycolysis activity levels and lactic acid content in the metabolites. Glycolysis consumes more glucose but produces less ATP.<sup>38</sup> The PI3K/AKT signaling pathway promotes aerobic glycolysis by upregulating cell surface glucose transporters<sup>39</sup> and glycolytic enzymes in tumor cells.<sup>40,41</sup>

Surprisingly, we found that the mutation of the EPB41L3 gene caused increased activity of the glucose metabolism pathway in tumor cells. Therefore, we speculate that mutations in EPB41L3 may reduce sensitivity to etoposide through DNA repair in tumor cells. In addition, AKT is involved in the repair of DNA damage caused by genotoxicity, mainly by the action of DNA-dependent protein kinase (DNA-PK), the kinase ATM/ATM and nonhomologous end joining (NHEJ) to repair DSB.42 Makinoshima et al43 found that PI3K/AKT/mTOR signaling inhibitors can effectively inhibit the expression of GLUT1 on the cell membrane. They used RNAi to interfere with the expression of GLUT1, ultimately reducing the aerobic glycolysis process and cell proliferation rate. Furthermore, our results suggest that PCLO mutations cause activation of the PI3K-Akt pathway, so we hypothesized that PCLO mutations may enhance glucose metabolism by activating the PI3K/Akt pathway, thereby enhance the ability of the tumor cell to repair DNA.

n SCLC patients
t of etoposide in S
ive treatment o
s of alternativ
g clinical trial
Completed/ongoing
Table 3

- 00		_	-		
Drug name	Clincal phase	Comments	NCT No.	Treatment	Pathway/target
lrinotecan	3		NCT00168896	Carboplatin+Irinotecan	Topoisomerase I
	2		NCT01441349		
	2		NCT01441349	Carboplatin+Sunitinib+Irinotecan	
	2		NCT00695292		
	_		NCT00045604	Cisplatin+Irinotecan+Imatinib	
	_	c-kit positive	NCT00052494		
	2		NCT00248482		
	_		NCT00059761	Cisplatin+Irinotecan	
	2		NCT01441349		
	2		NCT01441349	Cisplatin+Simvastatin+Irinotecan	
	2		NCT00452634		
	2		NCT00546130	Cisplatin+Krestin+Irinotecan	
	2		NCT00118235	Cisplatin+Irinotecan+Bevacizumab	
Bevacizumab	2		NCT00118235	Cisplatin+Irinotecan+Bevacizumab	VEGF
Pemetrexed	2		NCT00051506	Carboplatin+Pemetrexed	TS, DHFR,GARFT
	2		NCT00494026		
	2		NCT00051506	Cisplatin+Pemetrexed	
	2		NCT00475657		
Dimethylkanthenone Acetic Acid (DMXAA)	2		NCT01057342	Carboplatin+Dimethylxanthenone Acetic Acid (DMXAA)+Paclitaxel	DT-diaphorase
Paclitaxel	2		NCT01057342	Carboplatin+Dimethylxanthenone Acetic Acid	Mitosis;Microtubule stabiliser
	2		NCT00454324	Carboplatin+Paclitaxel	
	_		NCT02069158	Carboplatin+Paclitaxel+PF-05212384	
PF-05212384	_		NCT02069158	Carboplatin+Paclitaxel+PF-05212384	ΡΙ3Κ/mTOR;ΡΙ3Κα, ΡΙ3Κγ,mTOR
Gemcitabine	2		NCT02722369	Carboplatin+Gemcitabine	DNA replication;Pyrimidine antimetabolite
Pegfilgrastim	2	Be able to receive growth	NCT01076504	Carboplatin+Pegfilgrastim+Amrubicin	Granulocyte colony-stimulating factor receptor;
					Neutrophil elastase
Amrubicin	2	Be able to receive growth	NCT01076504	Carboplatin+Pegfilgrastim+Amrubicin	Topoisomerase 2
Sunitinib	2		NCT00695292	Carboplatin+Sunitinib+Irinotecan	RTK signaling:PDGFR, KIT, VEGFR, FLT3, RET,
					CSFIR
					(Continued)

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# Table 3 (Continued)

Drug name	Clincal phase	Comments	NCT No.	Treatment	Pathway/target
Topotecan	2 3		NCT00316186 NCT00043927 NCT00028925	Carboplatin+Topotecan Cisplatin+Topotecan Carboplatin+Topotecan+G-CSF	DNA topoisomerases
Belotecan	3		NCT00826644	Cisplatin+Belotecan	HDAC
Imatinib	2   	c-kit positive	NCT00248482 NCT00045604 NCT00052494	Cisplatin+Irinotecan+Imatinib	RTK signaling:ABL, KIT, PDGFR
Simvastatin	2 2		NCT01441349 NCT00452634	Cisplatin+Simvastatin+Irinotecan	HMG-CoA Reductase
Krestin Sagopilone	2 2 2		NCT01441349 NCT00546130 NCT00359359	Carboplatin+Irinotecan+Simvastatin Cisplatin+Krestin+Irinotecan Cisplatin+Sagopilone	Apoptosis;p21(WAF/Cip1) Microtubule stabiliser
Notes: TS, Thymidylate Synthetase: DHFR, Dihydrofolate Redu Methylglutaryl Coenzyme A Reductase; RTK, Receptor Tyrosine Ki FLT3, Fms Related Tyrosine Kinase; RET, Ret Proto-Oncogene; C granulocyte colony stimulating factor; SCLC,small-cell lung cancer.	DHFR, Dihydrof RTK, Receptor ET, Ret Proto-O SCLC,small-cell	olate Reductase; GARFT, Formylgycii Tyrosine Kinase; PDGFR, Platelet-Deri Incogene; CSFIR, Colony Stimulating I lung cancer.	namide Ribotide Ami ved Growth Factor Ri Factor I Receptor; H	Notes: T5, Thymidylate Synthetase; DHFR, Dihydrofolate Reductase; GARFT, Formylgycinamide Ribotide Amidotransferase; PI3K/mTOR, Phosphoinosmde-3-Kinase/The Mammalian Target of Rapamycin; HMG-CoA, Hydroxy Methylglutaryl Coenzyme A Reductase; RTK, Receptor Tyrosine Kinase; PDGFR, Platelet-Derived Growth Factor Receptor; KIT, KIT proto-oncogene, Receptor Tyrosine Kinase; VEGFR, Vascular Endothelial Growth Factor Receptor; FLT3, Fms Related Tyrosine Kinase; RET, Ret Proto-Oncogene; CSF1R, Colony Stimulating Factor I Receptor; HDAC, Histone Deacetylase; ABL, Abl Tyrosine Kinase; p21(WAF/Cip1), Cyclin Dependent Kinase Inhibitor; G-CSF granulocyte colony stimulating factor; SCLC,small-cell lung cancer.	Notes: TS, Thymidylate Synthetase; DHFR, Dihydrofolate Reductase; GARFT, Formylgycinamide Ribotide Amidotransferase; P13K/mTOR, Phosphoinosmde-3-Kinase/The Mammalian Target of Rapamycin; HMG-CoA, Hydroxy Methylglutaryl Coenzyme A Reductase; RTK, Receptor Tyrosine Kinase; PDGFR, Platelet-Derived Growth Factor Receptor; FLT3, Fms Related Tyrosine Kinase; RET, Ret Proto-Oncogene; CSF1R, Colony Stimulating Factor 1 Receptor; HDAC, Histone Deacetylase; ABL, Abl Tyrosine Kinase; p21(WAF/Cip1), Cyclin Dependent Kinase Inhibitor; G-CSF, granulocyte colony stimulating factor; SCLC,small-cell lung cancer.

Identifying outpatients with ES-SCLC that is not sensitive to etoposide and treating them with another combination therapy are important steps in improving the survival of patients with SCLC. Screening for the sensitivity to etoposide in patients with SCLC who are receiving chemotherapy for the first time allows clinicians to use a different combination chemotherapy regimen (Table 3) in these patients to avoid treatment failure due to primary resistance to etoposide. Currently, alternative treatment options that are commonly used in clinical practice include IP protocols, platinum-based drugs plus paclitaxel, and IP plus sunitinib. A phase II clinical trial (NCT00454324) on the use of a platinum-based compound plus paclitaxel in patients with ES-SCLC has shown good efficacy.<sup>44</sup> In a phase II clinical trial (NCT00695292),45 sunitinib combined with IP for patients with ES-SCLC showed potential clinical efficacy and safety, with an ORR of 59%, a oneyear survival rate of 54% and a median PFS of 7.6 months. In recent years, combinations of various chemotherapy regimens have been shown to provide excellent survival advantages in patients with ES-SCLC. It may be possible to classify patients by adding inclusion criteria and then use a more specific new chemotherapy regimen as a clinical treatment to achieve individualized and precise treatment of ES-SCLC patients, overcoming the treatment bottleneck for patients with ES-SCLC that is resistant to EP and ultimately prolonging their survival time and improving their OOL.

There were some limitations in this study. First, the most suitable alternative drug at present is irinotecan. GDSC does not provide data regarding the sensitivity to irinotecan, and the sensitivity of etoposide-resistant ES-SCLC to irinotecan is still unclear. Second, currently, there are no suitable large-sample clinical datasets that directly support our conclusions, and relevant clinical research needs to be further conducted to verify our hypothesis; moreover, we have initialed a clinical trial(NCT03162705) and hope this onging clincal trial could provide more direct evidence onni. Third, the accuracy of the model prediction is inadequate, and it may be necessary to expand the model to optimize it.

#### Conclusion

In conclusion, we analyzed the mutation and gene expression data from the GDSC of 54 ES-SCLC cell lines with regard to etoposide susceptibility and found that the panel including *CSMD3*, *EPB41L3*, *PCLO*, and *RYR1* can likely predict the sensitivity of ES-SCLC to etoposide and, therefore, the clinical survival of patients with SCLC.

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#### Disclosure

The authors report no conflicts of interest in this work.

#### References

- Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* 2018;68:394–424. doi:10.3322/caac.21492
- Haddadin S, Perry MC. History of small-cell lung cancer. Clin Lung Cancer. 2011;12(2):87–93. doi:10.1016/j.cllc.2011.03.002
- Sher T, Dy GK, Adjei AA. Small cell lung cancer. *Mayo Clin Proc*. 2008;83:355–367. doi:10.4065/83.3.355
- 4. Howlader N, Noone AM, Krapcho M, et al. SEER cancer statistics review, 1975–2014, National Cancer Institute. Bethesda, MD, based on November 2016 SEER data submission, posted to the SEER website, April 28, 2017. Available from: https://seer.cancer.gov/csr/1975\_2014/. Accessed April 28, 2017.
- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2018. CA Cancer J Clin. 2018;68:7–30. doi:10.3322/caac.21442
- Torre LA, Siegel RL, Jemal A. Lung cancer statistics. Adv Exp Med Biol. 2016;893:1–19. doi:10.1007/978-3-319-24223-1\_1
- Conen K, Hagmann R, Hess V, et al. Incidence and predictors of Bone metastases (BM) and Skeletal-related events (SREs) in Small cell lung cancer (SCLC): a Swiss patient cohort. *J Cancer*. 2016;7:2110–2116. doi:10.7150/jca.16211
- Schiller JH, Adak S, Cella D, et al. Topotecan versus observation after cisplatin plus etoposide in extensive-stage small-cell lung cancer: E7593–a phase III trial of the eastern cooperative oncology group. J Clin Oncol. 2001;19:2114–2122. doi:10.1200/JCO.200 1.19.8.2114
- Jett JR, Schild SE, Kesler KA, et al. Treatment of small-cell lung cancer: diagnosis and management of lung cancer, 3rd ed: American college of chest physicians evidence-based clinical practice guidelines. *Chest.* 2013;143(5Suppl):e4008–e419S. doi:10.1378/chest.12-2363
- SEER cancer statistics review. 1975–2009. Available from: https:// seer.cancer.gov/csr/1975\_2009\_pops09/, based on November 2011 SEER data submission, posted to the SEER website, April 30, 2012.. Accessed April 30, 2012.
- Stinchcombe TE, Gore EM. Limited-stage small-cell lung cancer: current chemoradiotherapy treatment paradigms. *Oncologist*. 2010;15:187–195. doi:10.1634/theoncologist.2009-0298
- Morabito A, Carillio G, Daniele G, et al. Treatment of small-cell lung cancer. *Crit Rev Oncol Hematol.* 2014;9:257–270. doi:10.1016/j. critrevonc.2014.03.003
- Früh M, De Ruysscher D, Popat S, et al. Small-cell lung cancer (SCLC): ESMO clinical practice guidelines for diagnosis, treatment and follow-up. *Ann Oncol.* 2013;24:vi99–vi105. doi:10.1093/annonc/ mdt178
- Ogino H, Hanibuchi M, Kakiuchi S, et al. Analysis of the prognostic factors of extensive disease small-cell lung cancer patients intokushima university hospital. J Med Invest. 2016;63:286–293.
- Noda K, Nishiwaki Y, Kawahara M, et al. Irinotecan plus cisplatin compared with etoposide plus cisplatin for extensive small-cell lung cancer. N Engl J Med. 2002;346:85–91. doi:10.1056/NEJMoa003034

- 16. Hermes A, Bergman B, Bremnes R, et al. Irinotecan plus carboplatin versus oral etoposide plus carboplatin in extensive small-cell lung cancer: a randomized phase III trial. J Clin Oncol. 2008;26:4261– 4267. doi:10.1200/JCO.2007.15.7545
- Hanna N, Bunn PA Jr, Langer C, et al. Randomized phase III trial comparing irinotecan/cisplatin with etoposide/cisplatin in patients with previously untreated extensive-stage. J Clin Oncol. 2006;24:2038–2043. doi:10.1200/JCO.2005.04.8595
- Lara PN Jr, Natale R, Crowley J, et al. Phase III trial of irinotecan/ cisplatin compared with etoposide/cisplatin in extensive-stage small-cell lung cancer: clinical and pharmacogenomic results from SWOG S0124. *J Clin Oncol.* 2009;27:2530–2535. doi:10.1200/JCO.2008.20.1061
- Schmittel A, Sebastian M, Fischer von Weikersthal L, et al. A German multicenter, randomized phase III trial comparing irinotecan-carboplatin. *Ann Oncol.* 2011;22:1798–1804. doi:10.1093/ annonc/mdq652
- 20. Zatloukal P, Cardenal F, Szczesna A, et al. A multicenter international randomized phase III study comparing cisplatin in combination with irinotecan or etoposide in previously untreated small-cell lung cancer patients with extensive disease. *Ann Oncol.* 2010;21(9):1810– 1816. doi:10.1093/annonc/mdq036
- 21. Kim DW, Kim HG, Kim JH, et al. Randomized phase III trial of irinotecan plus cisplatin versus etoposide plus cisplatin in chemotherapy-naïve Korean patients with extensive-disease small-cell lung cancer. *Cancer Res Treat.* 2019;51:119–127. doi:10.4143/crt.2018.019
- 22. Rossi A, Di Maio M, Chiodini P, et al. Carboplatin- or cisplatin-based chemotherapy in first-line treatment of small-cell lung cancer: the COCIS meta-analysis of individual patient data. J Clin Oncol. 2012;30:1692–1698. doi:10.1200/JCO.2011.40.4905
- Chute JP, Chen T, Feigal E, et al. Twenty years of phase III trials for patients with extensive-stage small-cell lung cancer: perceptible progress. *J Clin Oncol.* 1999;17:1794–1801. doi:10.1200/JCO.1999.17.6.1794
- 24. Goldie JH, Coldman AJ. A mathematic model for relating the drug sensitivity of tumors to their spontaneous mutation rate. *Cancer Treat Rep.* 1979;63(11–12):1727–1733.
- 25. van Meerbeeck JP, Fennell DA, de Ruysscher DK. Small-cell lung cancer. *Lancet*. 2011;378(9804):1741–1755. doi:10.1016/S0140-6736 (11)60984-7
- 26. Dingemans AM, Witlox MA, Stallaert RA, et al. Expression of DNA topoisomerase IIalpha and topoisomerase IIbeta genes predicts survival and response to chemotherapy in patients with small-cell lung cancer. *Clin Cancer Res.* 1999;5:2048–2058.
- Viktorsson K, De Petris L, Lewensohn R. The role of p53 in treatment responses of lung cancer. *Biochem Biophys Res Commun.* 2005;331:868–880. doi:10.1016/j.bbrc.2005.03.192
- Yang W, Soares J, Greninger P, et al. Genomics of drug sensitivity in cancer (GDSC): a resource for therapeutic biomarker discovery in cancer cells. *Nucleic Acids Res.* 2013;41:D955–61. doi:10.1093/nar/gks1111
- Bonanno L, Favaretto A, Rugge M, et al. Role of genotyping in nonsmall-cell lung cancer treatment: current status. *Drugs*. 2011;71:2231–2246. doi:10.2165/11597700-000000000-00000
- 30. Chiu HW, Chang JS, Lin HY, et al. FBXL7 upregulation predicts a poor prognosis and associates with a possible mechanism for paclitaxel resistance in ovarian cancer. *J Clin Med.* 2018;7(10):330. doi:10.3390/jcm7100330

- 31. Chiu HW, Lin HY, Tseng IJ, et al. OTUD7B upregulation predicts a poor response to paclitaxel in patients with triple-negative breast cancer. Oncotarget. 2017;9:553–565. doi:10.18632/ oncotarget.23074
- 32. Mayakonda A, Lin DC, Assenov Y, et al. Maftools: efficient and comprehensive analysis of somatic variants in cancer. *Genome Res.* 2018;28(11):1747–1756. doi:10.1101/gr.239244.118
- 33. Ritchie ME, Phipson B, Wu D, et al. limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Res.* 2015;43:e47. doi:10.1093/nar/gkv007
- 34. George J, Lim JS, Jang SJ, et al. Comprehensive genomic profiles of small-cell lung cancer. *Nature*. 2015;524(7563):47–53. doi:10.1038/ nature14664
- 35. Liu SQ, Yu JP, Yu HG, et al. Activation of Akt and ERK signalling pathways induced by etoposide confer chemoresistance in gastric cancer cells. *Dig Liver Dis.* 2006;38:310–318. doi:10.1016/j. dld.2006.01.012
- Bakkenist CJ, Kastan MB. DNA damage activates ATM through intermolecular autophosphorylation and dimer dissociation. *Nature*. 2003;421(6922):499–506. doi:10.1038/nature01368
- Lans H, Marteijn JA, Vermeulen W. ATP-dependent chromatin remodeling in the DNA-damage response. *Epigenetics Chromatin*. 2012;5:4. doi:10.1186/1756-8935-5-4
- 38. Warburg O. On the origin of cancer cells. *Science*. 1956;123 (3191):309–314.
- DeBerardinis RJ, Lum JJ, Hatzivassiliou G, et al. The biology of cancer: metabolic reprogramming fuels cell growth and proliferation. *Cell Metab.* 2008;7:11–20. doi:10.1016/j.cmet.2007.10.002
- 40. Majewski N, Nogueira V, Bhaskar P, et al. Hexokinase-mitochondria interaction mediated by Akt is required to inhibit apoptosis in the presence or absence of Bax and Bak. *Mol Cell*. 2004;16:819–830. doi:10.1016/j.molcel.2004.11.014
- Buzzai M, Bauer DE, Jones RG, et al. The glucose dependence of Akttransformed cells can be reversed by pharmacologic activation of fatty acid beta-oxidation. *Oncogene*. 2005;24:4165–4173. doi:10.1038/sj. onc.1208622
- 42. Xu N, Lao Y, Zhang Y, et al. Akt: a double-edged sword in cell proliferation and genome stability. J Oncol. 2012;2012:951724. doi:10.1155/2012/951724
- 43. Makinoshima H, Takita M, Saruwatari K, et al. Signaling through the Phosphatidylinositol 3-kinase (PI3K)/Mammalian target of Rapamycin(mTOR) axis is responsible for aerobic glycolysis mediated by glucose transporter in Epidermal growth factor receptor(EGFR) -mutated lung adenocarcinoma. J Biol Chem. 2015;290:17495–17504. doi:10.1074/jbc.M115.660498
- 44. Grilley-Olson JE, Keedy VL, Sandler A, et al. A randomized phase II study of carboplatin with weekly or every-3-week nanoparticle albumin-bound paclitaxel (abraxane) in patients with extensive-stage small-cell lung cancer. *Oncologist.* 2015;20:105–106. doi:10.1634/ theoncologist.2014-0327
- 45. Spigel DR, Greco FA, Rubin MS, et al. Phase II study of maintenance sunitinib following irinotecan and carboplatin as first-line treatment for patients with extensive-stage small-cell lung cancer. *Lung Cancer.* 2012;77:359–364. doi:10.1016/j.lungcan.20 12.03.009

#### Supplementary materials

Cell line	IC50 (μM)	AUC
LU-135	0.242	0.262
SBC-3	0.276	0.292
SBC-5	0.406	0.344
LU-134-A	0.407	0.363
NCI-H526	0.515	0.393
NCI-H1048	0.563	0.405
DMS-273	0.595	0.42
NCI-H211	0.618	0.423
NCI-H187	0.758	0.458
NCI-H748	0.838	0.475
NCI-H209	0.97	0.495
IST-SL2	0.978	0.496
SW1271	1.29	0.537
COR-L279	1.39	0.555
NCI-H1694	1.52	0.566
LB647-SCLC	1.77	0.585
COLO-668	2.01	0.61
NCI-H1876	2.06	0.614
NCI-HI304	2.34	0.629
NCI-HI4I7	3.26	0.669
MS-I	3.62	0.709
NCI-H64	3.93	0.742
NCI-H2081	4.28	0.715
LU-139	4.7	0.71
NCI-H69	5.35	0.74
NCI-H1963	6.37	0.795
NCI-H510A	6.78	0.795
NCI-H847	7.38	0.827
NCI-H2141	7.39	0.797
NCI-H2196	8.08	0.798
IST-SLI	10.5	0.83
LU-165	10.9	0.821
NCI-H1688	11	0.825
NCI-H2029	12.3	0.867
NCI-H841	15.2	0.871
CPC-N	16.4	0.865
COR-L95	17.5	0.86
DMS-79	21.4	0.877
COR-L88	22	0.876
NCI-H2171	23.8	0.933
SBC-1	33.3	0.935
NCI-H82	36	0.942
NCI-H1836	41.1	0.928
NCI-H446	45.6	0.936
NCI-H524	50	0.965
SHP-77	57.7	0.97
NCI-H1092	65.2	0.96
NCI-H2227	69.3	0.949
DMS-53	71.3	0.955
	I (Cor	ntinued)

Table SI (Continued)

Cell line	IC50 (μM)	AUC
HCC-33	73.8	0.964
NCI-H196	108	0.971
NCI-H1436	133	0.968
NCI-H345	162	0.978
DMS-114	319	0.984

 $\label{eq:abbreviations: AUC, area under the curve; IC50, half maximal inhibitory concentration; SCLC, small cell lung cancer.$ 

Test result variable(s)	Area	Standard error <sup>a</sup>	Asymptotic significance	Asymptotic 95% con	fidence interval
				Lower bound	Upper bound
CSMD3	0.697	0.077	0.016	0.546	0.848
USP34	0.685	0.099	0.053	0.49	0.879
MYO18B	0.679	0.096	0.061	0.491	0.867
ABCA13	0.673	0.093	0.07	0.491	0.855
DNAH2	0.673	0.099	0.07	0.479	0.866
LAMA5	0.661	0.099	0.092	0.468	0.854
SCN4A	0.655	0.101	0.105	0.457	0.853
ARAP2	0.643	0.101	0.134	0.446	0.84
CNTRL	0.643	0.101	0.134	0.446	0.84
ENSG00000250423	0.643	0.101	0.134	0.446	0.84
RYRI	0.631	0.082	0.111	0.469	0.792
EYS	0.631	0.096	0.17	0.443	0.818
HSPG2	0.631	0.1	0.17	0.435	0.827
NLRP5	0.631	0.1	0.17	0.435	0.827
UNCI3C	0.631	0.1	0.17	0.435	0.827
DDX12	0.619	0.1	0.212	0.424	0.814
XIRP2	0.619	0.096	0.212	0.432	0.806
EPB41L3	0.61	0.083	0.179	0.447	0.774
COL3AI	0.607	0.099	0.261	0.413	0.802
NIPBL	0.607	0.099	0.261	0.413	0.802
NLRP3	0.607	0.099	0.261	0.413	0.802
POLQ	0.607	0.099	0.261	0.413	0.802
GRM5	0.601	0.101	0.289	0.404	0.798
PKDILI	0.601	0.097	0.289	0.411	0.792
REG3G	0.601	0.101	0.289	0.404	0.792
AHNAK	0.595	0.099	0.318	0.404	0.789
PCLO		0.083	0.267	0.402	0.754
	0.591	0.085	0.349	0.393	0.785
AC027369_8 BRIP1	0.589	0.1	0.349	0.393	0.785
		0.1			
COL6A3	0.589	0.1	0.349	0.393	0.785
ERBB4	0.589		0.349	0.393	0.785
FAM135B	0.589	0.097	0.349	0.399	0.779
FBNI	0.589	0.1	0.349	0.393	0.785
FREMI	0.589	0.1	0.349	0.393	0.785
HFMI	0.589	0.1	0.349	0.393	0.785
KDR	0.589	0.1	0.349	0.393	0.785
MYHI	0.589	0.1	0.349	0.393	0.785
NDST4	0.589	0.1	0.349	0.393	0.785
PPP1R9A	0.589	0.1	0.349	0.393	0.785
SMARCA4	0.589	0.1	0.349	0.393	0.785
THSD7B	0.589	0.1	0.349	0.393	0.785
UBQLN3	0.589	0.1	0.349	0.393	0.785
NAV3	0.583	0.098	0.382	0.391	0.776
ADAMTS16	0.577	0.099	0.417	0.383	0.772
AKAP13	0.577	0.099	0.417	0.383	0.772
ALPK2	0.577	0.099	0.417	0.383	0.772
COLI4AI	0.577	0.099	0.417	0.383	0.772
DPPIO	0.577	0.099	0.417	0.383	0.772
EML5	0.577	0.099	0.417	0.383	0.772
1	1		I a semi-	1	

0.417

 Table S2 ROC curve of all genes (mutation frequency >10%)

(Continued)

KIAA I 109

0.577

0.099

0.772

0.383

Test result variable(s)	Area	ea Standard error <sup>a</sup>	Asymptotic significance	Asymptotic 95% confidence interval		
				Lower bound	Upper bound	
LYST	0.577	0.099	0.417	0.383	0.772	
MYH13	0.577	0.099	0.417	0.383	0.772	
MYH7	0.577	0.099	0.417	0.383	0.772	
PDGFRA	0.577	0.099	0.417	0.383	0.772	
ZEBI	0.577	0.099	0.417	0.383	0.772	
.RRK2	0.571	0.098	0.454	0.38	0.763	
CAN	0.565	0.099	0.492	0.372	0.759	
DAMTSLI	0.565	0.099	0.492	0.372	0.759	
DCY8	0.565	0.099	0.492	0.372	0.759	
ALMS I	0.565	0.099	0.492	0.372	0.759	
NKSIB	0.565	0.099	0.492	0.372	0.759	
NTNAP4	0.565	0.099	0.492	0.372	0.759	
RASI	0.565	0.099	0.492	0.372	0.759	
AMAT	0.565	0.099	0.492	0.372	0.759	
MORCI	0.565	0.099	0.492	0.372	0.759	
AUC16	0.565	0.092	0.492	0.385	0.746	
AUC5B	0.565	0.097	0.492	0.376	0.755	
TPRB	0.565	0.099	0.492	0.372	0.759	
SIGLEC I 0	0.565	0.099	0.492	0.372	0.759	
TAB2	0.565	0.099	0.492	0.372	0.759	
YNEI	0.565	0.097	0.492	0.376	0.755	
JBR4	0.565	0.099	0.492	0.372	0.759	
DNAH8	0.56	0.097	0.533	0.368	0.751	
RELN	0.56	0.097	0.533	0.368	0.751	
<b>FP53</b>	0.56	0.089	0.533	0.385	0.734	
WDR72	0.56	0.099	0.533	0.365	0.754	
ZNF831	0.56	0.099	0.533	0.365	0.754	
DAMTS12	0.554	0.098	0.574	0.361	0.746	
DGB	0.554	0.098	0.574	0.361	0.746	
BN2	0.554	0.098	0.574	0.361	0.746	
GPR I I 2	0.554	0.098	0.574	0.361	0.746	
TGAD	0.554	0.098	0.574	0.361	0.746	
KALRN	0.554	0.098	0.574	0.361	0.746	
KIF2B	0.554	0.098	0.574	0.361	0.746	
YKHD I LI	0.554	0.098	0.574	0.361	0.746	
G	0.554	0.098	0.574	0.361	0.746	
VDR87	0.554	0.098	0.574	0.361	0.746	
NKRDII	0.548	0.099	0.618	0.354	0.741	
INTN5	0.548	0.099	0.618	0.354	0.741	
COLIZAI	0.548	0.097	0.618	0.357	0.738	
COLI 7A I	0.548	0.099	0.618	0.354	0.741	
PS I	0.548	0.099	0.618	0.354	0.741	
DAPKI	0.548	0.099	0.618	0.354	0.741	
NAH6	0.548	0.099	0.618	0.354	0.741	
CGBP	0.548	0.097	0.618	0.357	0.738	
5L13	0.548	0.099	0.618	0.354	0.741	
SRIN2B	0.548	0.099	0.618	0.354	0.741	
IECWI	0.548	0.099	0.618	0.354	0.741	
HYDIN	0.548	0.095	0.618	0.361	0.735	

Test result variable(s)	Area	Standard error <sup>a</sup>	Asymptotic significance	Asymptotic 95% confidence interval		
				Lower bound	Upper bound	
GSF3	0.548	0.099	0.618	0.354	0.741	
KIAA I 409	0.548	0.099	0.618	0.354	0.741	
LING02	0.548	0.099	0.618	0.354	0.741	
LRRIQ I	0.548	0.099	0.618	0.354	0.741	
MADD	0.548	0.099	0.618	0.354	0.741	
MCF2	0.548	0.099	0.618	0.354	0.741	
PLXNA4	0.548	0.099	0.618	0.354	0.741	
RYR2	0.548	0.095	0.618	0.361	0.735	
SORCS3	0.548	0.099	0.618	0.354	0.741	
JNC80	0.548	0.097	0.618	0.357	0.738	
NDR17	0.548	0.099	0.618	0.354	0.741	
CUBN	0.542	0.098	0.662	0.351	0.733	
DSCAMLI	0.542	0.098	0.662	0.351	0.733	
ENSG00000121031	0.542	0.098	0.662	0.351	0.733	
ENSG00000188219	0.542	0.098	0.662	0.351	0.733	
FAT3	0.542	0.096	0.662	0.353	0.73	
AMA2	0.542	0.098	0.662	0.351	0.733	
SYNE2	0.542	0.098	0.662	0.351	0.733	
TAFIL	0.542	0.098	0.662	0.351	0.733	
INN	0.542	0.098	0.662	0.351	0.733	
ZNF99	0.542	0.098	0.662	0.351	0.733	
ACSM2B	0.536	0.098	0.708	0.344	0.727	
ASPM	0.536	0.098	0.708	0.344	0.727	
ATPIOD	0.536	0.098	0.708	0.344	0.727	
BCLAFI	0.536	0.098	0.708	0.344	0.727	
CI2orf35	0.536	0.098	0.708	0.344	0.727	
26	0.536	0.098	0.708	0.344	0.727	
CACNATH	0.536	0.098	0.708	0.344	0.727	
CDH19	0.536	0.098	0.708	0.344	0.727	
COLI9AI	0.536	0.098	0.708	0.344	0.727	
COL24A I CREBBP	0.536	0.098 0.098	0.708 0.708	0.344 0.344	0.727 0.727	
	0.536					
DCHS2	0.536	0.098	0.708	0.344	0.727	
DNAH17	0.536	0.098	0.708	0.344	0.727	
DOCK7	0.536	0.098	0.708	0.344	0.727	
EP400	0.536	0.098	0.708	0.344	0.727	
GF2R	0.536	0.098	0.708	0.344	0.727	
TBPI	0.536	0.098	0.708	0.344	0.727	
MUCI7	0.536	0.097	0.708	0.346	0.725	
MYHTT	0.536	0.098	0.708	0.344	0.727	
NOTCHI	0.536	0.098	0.708	0.344	0.727	
DTOF	0.536	0.098	0.708	0.344	0.727	
PIK3CG	0.536	0.098	0.708	0.344	0.727	
POMI2ILI2	0.536	0.098	0.708	0.344	0.727	
POTEC	0.536	0.098	0.708	0.344	0.727	
POTEG	0.536	0.098	0.708	0.344	0.727	
PTEN	0.536	0.098	0.708	0.344	0.727	
ROBO4	0.536	0.098	0.708	0.344	0.727	
SCNIA	0.536	0.098	0.708	0.344	0.727	

Test result variable(s)	Area	Standard error <sup>a</sup>	Asymptotic significance	Asymptotic 95% confidence interval	
				Lower bound	Upper bound
SLC5A10	0.536	0.098	0.708	0.344	0.727
SLIT3	0.536	0.098	0.708	0.344	0.727
SRCAP	0.536	0.098	0.708	0.344	0.727
RHDE	0.536	0.098	0.708	0.344	0.727
TN	0.536	0.093	0.708	0.354	0.718
WA3B	0.536	0.098	0.708	0.344	0.727
VBSCR I 7	0.536	0.098	0.708	0.344	0.727
VNK3	0.536	0.098	0.708	0.344	0.727
NF208	0.536	0.098	0.708	0.344	0.727
INF804B	0.536	0.098	0.708	0.344	0.727
SCAN20	0.536	0.098	0.708	0.344	0.727
OCKII	0.53	0.098	0.755	0.338	0.722
KHDI	0.53	0.097	0.755	0.34	0.72
ΡΤΑΙ	0.53	0.097	0.755	0.34	0.72
FHX4	0.53	0.096	0.755	0.342	0.718
INF536	0.53	0.097	0.755	0.34	0.72
BCA12	0.524	0.097	0.803	0.334	0.714
BCBI	0.524	0.097	0.803	0.334	0.714
C007731.1	0.524	0.097	0.803	0.334	0.714
NKRD30B	0.524	0.097	0.803	0.334	0.714
20orf26	0.524	0.097	0.803	0.334	0.714
7orf58	0.524	0.097	0.803	0.334	0.714
CACNAIC	0.524	0.097	0.803	0.334	0.714
DMD	0.524	0.097	0.803	0.334	0.714
DPP6	0.524	0.097	0.803	0.334	0.714
LG2	0.524	0.097	0.803	0.334	0.714
RM I	0.524	0.097	0.803	0.334	0.714
IMCNI	0.524	0.096	0.803	0.335	0.712
NAGEC I	0.524	0.097	0.803	0.334	0.714
1DN I	0.524	0.097	0.803	0.334	0.714
IGAM	0.524	0.097	0.803	0.334	0.714
1KI67	0.524	0.097	0.803	0.334	0.714
AUC12	0.524	0.096	0.803	0.335	0.712
1UC2	0.524	0.097	0.803	0.334	0.714
IID2	0.524	0.097	0.803	0.334	0.714
DR8K I	0.524	0.097	0.803	0.334	0.714
APPA	0.524	0.097	0.803	0.334	0.714
TPN I 3	0.524	0.097	0.803	0.334	0.714
AMD9	0.524	0.097	0.803	0.334	0.714
I	0.524	0.097	0.803	0.334	0.714
РНКАР	0.524	0.096	0.803	0.335	0.712
PO	0.524	0.097	0.803	0.334	0.714
ISP32	0.524	0.097	0.803	0.334	0.714
CAN	0.524	0.097	0.803	0.334	0.714
VRN	0.524	0.097	0.803	0.334	0.714
EB2	0.524	0.097	0.803	0.334	0.714
 INF479	0.524	0.097	0.803	0.334	0.714
NAHTI	0.518	0.096	0.851	0.329	0.707
NAH14	0.518	0.096	0.851	0.329	0.707

Test result variable(s)	Area	Standard error <sup>a</sup>	Asymptotic significance	Asymptotic 95% confidence interval	
				Lower bound	Upper bound
GABRA5	0.518	0.097	0.851	0.328	0.708
VPS13B	0.518	0.096	0.851	0.329	0.707
ABCCII	0.512	0.096	0.901	0.323	0.7
CCDC141	0.512	0.096	0.901	0.323	0.7
CDH10	0.512	0.096	0.901	0.323	0.7
CDH8	0.512	0.096	0.901	0.323	0.7
CEP350	0.512	0.096	0.901	0.323	0.7
COLI I A2	0.512	0.096	0.901	0.323	0.7
CRBI	0.512	0.096	0.901	0.323	0.7
DOCK2	0.512	0.096	0.901	0.323	0.7
LAMA3	0.512	0.096	0.901	0.323	0.7
POTEH	0.512	0.096	0.901	0.323	0.7
PXDNL	0.512	0.096	0.901	0.323	0.7
SAMD9L	0.512	0.096	0.901	0.323	0.7
SPAG I 7	0.512	0.096	0.901	0.323	0.7
TPTE	0.512	0.096	0.901	0.323	0.7
CACNATE	0.506	0.096	0.95	0.318	0.694
FAM5B	0.506	0.096	0.95	0.318	0.694
FAT4	0.506	0.096	0.95	0.318	0.693
HRNR	0.506	0.096	0.95	0.318	0.693
MDGA2	0.506	0.096	0.95	0.318	0.694
MYCBP2	0.506	0.096	0.95	0.318	0.694
NBPF10	0.506	0.096	0.95	0.318	0.693
ORIOJI	0.506	0.096	0.95	0.318	0.694
TNXB	0.506	0.096	0.95	0.318	0.693
TRPAI	0.506	0.096	0.95	0.318	0.694
ZICI	0.506	0.096	0.95	0.318	0.694
ABCA9	0.5	0.095		0.313	0.687
DNAH3	0.5	0.095	1	0.313	0.687
FAM75D4	0.5	0.095		0.313	0.687
FMN2	0.5	0.095		0.313	0.687
KIAA0947	0.5	0.095		0.313	0.687
MTUS2	0.5	0.095		0.313	0.687
MYH4	0.5	0.095		0.313	0.687
NEB	0.5	0.095		0.313	0.687
OR14KI	0.5	0.095		0.313	0.687
SLC8A3	0.5	0.095		0.313	0.687
TEPI	0.5	0.095		0.313	0.687
THSD7A	0.5	0.095		0.313	0.687
USH2A	0.5	0.095		0.313	0.687
C15orf2	0.494	0.095	0.95	0.308	0.68
CDH20	0.494	0.095	0.95	0.308	0.68
COLIIAI	0.494	0.095	0.95	0.308	0.68
COL5A2	0.494	0.095	0.95	0.308	0.68
DNAH9	0.494	0.095	0.95	0.308	0.68
FSTL5	0.494	0.095	0.95	0.308	0.68
GRIPI	0.494	0.095	0.95	0.308	0.68
KIF2 I A	0.494	0.095	0.95	0.308	0.68
MYO7A	0.494	0.095	0.95	0.308	0.68

Test result variable(s)	Area	Standard error <sup>a</sup>	Asymptotic significance	Asymptotic 95% confidence interval	
				Lower bound	Upper bound
MYPN	0.494	0.095	0.95	0.308	0.68
NALCN	0.494	0.095	0.95	0.308	0.68
РНКВ	0.494	0.095	0.95	0.308	0.68
PRUNE2	0.494	0.095	0.95	0.308	0.68
SCN7A	0.494	0.095	0.95	0.308	0.68
SPEG	0.494	0.095	0.95	0.308	0.68
TFAP2D	0.494	0.095	0.95	0.308	0.68
ZFPM2	0.494	0.095	0.95	0.308	0.68
ZNF142	0.494	0.095	0.95	0.308	0.68
AHNAK2	0.488	0.095	0.901	0.303	0.673
DNAH7	0.488	0.095	0.901	0.303	0.673
HCNI	0.488	0.095	0.901	0.303	0.673
CDH15	0.488	0.095	0.901	0.303	0.673
ZNF729	0.488	0.095	0.901	0.303	0.673
3SN	0.482	0.094	0.851	0.298	0.666
CENPF	0.482	0.094	0.851	0.298	0.666
LSTN2	0.482	0.094	0.851	0.298	0.666
ELNC	0.482	0.094	0.851	0.298	0.666
HEATR I	0.482	0.094	0.851	0.298	0.666
(IAA   239	0.482	0.094	0.851	0.298	0.666
CT	0.482	0.094	0.851	0.298	0.666
PHN3	0.482	0.094	0.851	0.298	0.666
MLL2	0.482	0.094	0.851	0.297	0.667
DDZ2	0.482	0.094	0.851	0.298	0.666
DR5T2	0.482	0.094	0.851	0.298	0.666
DR6Y1	0.482	0.094	0.851	0.298	0.666
CDHIIX	0.482	0.094	0.851	0.298	0.666
CDHB7	0.482	0.094	0.851	0.298	0.666
PKD1L2	0.482	0.094	0.851	0.298	0.666
PLCH I	0.482	0.094	0.851	0.298	0.666
TPRD	0.482	0.094	0.851	0.298	0.666
RGPD3	0.482	0.094	0.851	0.298	0.666
SELP	0.482	0.094	0.851	0.298	0.666
SYTL2	0.482	0.094	0.851	0.298	0.666
TKTL2	0.482	0.094	0.851	0.298	0.666
Ϋ́R	0.482	0.094	0.851	0.298	0.666
JTP20	0.482	0.094	0.851	0.298	0.666
/WF	0.482	0.094	0.851	0.298	0.666
POB	0.476	0.094	0.803	0.293	0.66
CNTNAP5	0.476	0.094	0.803	0.293	0.66
P300	0.476	0.094	0.803	0.293	0.66
IEATR7B2	0.476	0.094	0.803	0.293	0.66
ROSI	0.476	0.094	0.803	0.293	0.66
ZIM2	0.476	0.094	0.803	0.293	0.66
BCA8	0.47	0.093	0.755	0.288	0.652
BCC12	0.47	0.093	0.755	0.288	0.652
CSM5	0.47	0.093	0.755	0.288	0.652
DAM2	0.47	0.093	0.755	0.288	0.652
ANKRD55	0.47	0.093	0.755	0.288	0.652

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#### Table S2 (Continued)

Test result variable(s)	Area	Standard error <sup>a</sup>	Asymptotic significance	Asymptotic 95% confidence interval		
				Lower bound	Upper bound	
ATP1A2	0.47	0.093	0.755	0.288	0.652	
CIOorfII2	0.47	0.093	0.755	0.288	0.652	
CI 2orf5 I	0.47	0.093	0.755	0.288	0.652	
CMYA5	0.47	0.093	0.755	0.288	0.652	
SMDI	0.47	0.094	0.755	0.286	0.654	
CYPIIBI	0.47	0.093	0.755	0.288	0.652	
OCHSI	0.47	0.093	0.755	0.288	0.652	
SEL	0.47	0.093	0.755	0.288	0.652	
YSF	0.47	0.093	0.755	0.288	0.652	
AT I	0.47	0.093	0.755	0.288	0.652	
ERC2	0.47	0.093	0.755	0.288	0.652	
CNUI	0.47	0.093	0.755	0.288	0.652	
RPIB	0.47	0.095	0.755	0.284	0.656	
ISH4	0.47	0.093	0.755	0.288	0.652	
IYH I 5	0.47	0.093	0.755	0.288	0.652	
IYH2	0.47	0.093	0.755	0.288	0.652	
IYO9A	0.47	0.093	0.755	0.288	0.652	
LRP4	0.47	0.093	0.755	0.288	0.652	
BSCN	0.47	0.094	0.755	0.286	0.654	
RDM9	0.47	0.093	0.755	0.288	0.652	
TPRU	0.47	0.093	0.755	0.288	0.652	
ZT2	0.47	0.093	0.755	0.288	0.652	
NR	0.47	0.093	0.755	0.288	0.652	
RPM2	0.47	0.093	0.755	0.288	0.652	
TRN	0.47	0.093	0.755	0.288	0.652	
NF462	0.47	0.093	0.755	0.288	0.652	
NF534	0.47	0.093	0.755	0.288	0.652	
NK2	0.464	0.093	0.708	0.282	0.646	
OL22AI	0.464	0.093	0.708	0.282	0.646	
ST	0.464	0.093	0.708	0.282	0.646	
RIN2A	0.464	0.092	0.708	0.285	0.644	
YR3	0.464	0.093	0.708	0.282	0.646	
COIBI	0.464	0.092	0.708	0.285	0.644	
BCB5	0.458	0.092	0.662	0.279	0.638	
A13	0.458	0.092	0.662	0.279	0.638	
5orf42	0.458	0.092	0.662	0.279	0.638	
D163	0.458	0.092	0.662	0.279	0.638	
сс	0.458	0.092	0.662	0.279	0.638	
YO7B	0.458	0.092	0.662	0.279	0.638	
LRP12	0.458	0.092	0.662	0.279	0.638	
DZI	0.458	0.092	0.662	0.279	0.638	
DZ3	0.458	0.092	0.662	0.279	0.638	
R8H3	0.458	0.092	0.662	0.279	0.638	
DE4DIP	0.458	0.092	0.662	0.279	0.638	
IMS2	0.458	0.092	0.662	0.279	0.638	
100	0.450			0.070		

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Test result variable(s)	Area	Standard error <sup>a</sup>	Asymptotic significance	Asymptotic 95% confidence interval	
				Lower bound	Upper bound
Clorf173	0.452	0.092	0.618	0.272	0.633
DOCK4	0.452	0.09	0.618	0.275	0.629
GPR98	0.452	0.092	0.618	0.272	0.633
KIAA I 549	0.452	0.09	0.618	0.275	0.629
MACFI	0.452	0.092	0.618	0.272	0.633
CDH18	0.446	0.091	0.574	0.269	0.624
CTNNA2	0.446	0.091	0.574	0.269	0.624
NAH5	0.446	0.091	0.574	0.269	0.624
AM5C	0.446	0.091	0.574	0.269	0.624
RRAP	0.446	0.091	0.574	0.269	0.624
RWD3	0.44	0.089	0.533	0.266	0.615
CACHD I	0.44	0.089	0.533	0.266	0.615
DH7	0.44	0.089	0.533	0.266	0.615
DSCAM	0.44	0.089	0.533	0.266	0.615
RP2	0.44	0.091	0.533	0.262	0.619
AUC19	0.44	0.091	0.533	0.262	0.619
DRIIHI2	0.44	0.089	0.533	0.266	0.615
DR52R1	0.44	0.089	0.533	0.266	0.615
IGLEC8	0.44	0.089	0.533	0.266	0.615
MEM132D	0.44	0.091	0.533	0.262	0.619
NUC4	0.435	0.094	0.492	0.25	0.619
IMI	0.429	0.088	0.454	0.257	0.6
CARDII	0.429	0.088	0.454	0.257	0.6
COL5A3	0.429	0.088	0.454	0.257	0.6
SMD2	0.429	0.088	0.454	0.257	0.6
YA4	0.429	0.088	0.454	0.257	0.6
REM3	0.429	0.088	0.454	0.257	0.6
KIAA0240	0.429	0.088	0.454	0.257	0.6
(IAA   2   I	0.429	0.088	0.454	0.257	0.6
AMC3	0.429	0.088	0.454	0.257	0.6
PA	0.429	0.088	0.454	0.257	0.6
RFN5	0.429	0.088	0.454	0.257	0.6
VAV2	0.429	0.088	0.454	0.257	0.6
VCAM2	0.429	0.088	0.454	0.257	0.6
DK I	0.429	0.088	0.454	0.257	0.6
ETD2	0.429	0.088	0.454	0.257	0.6
HROOM3	0.429	0.088	0.454	0.257	0.6
PTB	0.429	0.088	0.454	0.257	0.6
NKRD30A	0.423	0.089	0.417	0.249	0.596
DTOG	0.423	0.089	0.417	0.249	0.596
APPA2	0.423	0.089	0.417	0.249	0.596
10orf71	0.417	0.086	0.382	0.247	0.586
OL6A6	0.417	0.086	0.382	0.247	0.586
LG	0.417	0.09	0.382	0.241	0.592
SCB	0.417	0.086	0.382	0.247	0.586
CNX	0.417	0.086	0.382	0.247	0.586
KDH	0.417	0.086	0.382	0.247	0.586
BODIL	0.405	0.085	0.318	0.238	0.571
RRC7	0.405	0.085	0.318	0.238	0.571

Test result variable(s)	Area	Standard error <sup>a</sup>	Asymptotic significance	Asymptotic 95% confidence interval	
				Lower bound	Upper bound
RPILI	0.405	0.085	0.318	0.238	0.571
ADAMTS20	0.399	0.086	0.289	0.23	0.568
MLL3	0.393	0.084	0.261	0.229	0.557
DNAH10	0.369	0.081	0.17	0.21	0.528
RBI	0.369	0.096	0.17	0.182	0.557

Note:  $\ensuremath{^a\!Under}$  the nonparametric assumption.

Abbreviation: ROC, receiver operating characteristic.

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