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Received 16 Jul 2012 | Accepted 11 Dec 2012 | Published 22 Jan 2013

DOI: 10.1038/ncomms2371

A novel pathway for the production of hydrogen sulfide from D-cysteine in mammalian cells

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In eukaryotes, hydrogen sulphide acts as a signalling molecule and cytoprotectant. Hydrogen sulphide is known to be produced from L-cysteine by cystathionine β -synthase, cystathionine γ -lyase and 3-mercaptopyruvate sulfurtransferase coupled with cysteine aminotransferase. Here we report an additional biosynthetic pathway for the production of hydrogen sulphide from D-cysteine involving 3-mercaptopyruvate sulfurtransferase and D-amino acid oxidase. Unlike the L-cysteine pathway, this D-cysteine-dependent pathway operates predominantly in the cerebellum and the kidney. Our study reveals that administration of D-cysteine protects primary cultures of cerebellar neurons from oxidative stress induced by hydrogen peroxide and attenuates ischaemia-reperfusion injury in the kidney more than L-cysteine. This study presents a novel pathway of hydrogen sulphide production and provides a new therapeutic approach to deliver hydrogen sulphide to specific tissues.

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Hydrogen sulphide (H_2S) modulates synaptic activity by enhancing the activity of *N*-methyl-*D*-aspartate receptors in neurons and by activating astrocytes that surround the synapse^{1,2}. It relaxes vascular smooth muscle by activating K^+ channels and regulates the release of insulin^{3–7}. In addition to its function as a signalling molecule, H_2S has a cytoprotective effect. It protects neurons from oxidative stress by recovering glutathione levels, scavenging ROS and suppressing intracellular Ca^{2+} concentrations^{8,9}. This finding led to the identification of H_2S -mediated cardio- and renal-protection from ischaemia-reperfusion injury^{10,11}.

Most studies about the production of H_2S have been concentrated on two pyridoxal 5'-phosphate-dependent enzymes, cystathionine β -synthase (CBS) and cystathionine γ -lyase (CSE)^{1,3,12,13}. Recently, 3-mercaptopyruvate sulfurtransferase (3MST)/cysteine aminotransferase (CAT) pathway has emerged as the third pathway. 3MST produces H_2S from 3-mercaptopyruvate (3MP), which is generated by CAT from *L*-cysteine and α -ketoglutarate (α -KG) in the presence of the cofactors thioredoxin and dihydrolipoic acid^{14–16}. *D*-Amino acid oxidase (DAO) can also produce achiral α -keto acids, including 3MP, from *D*-amino acids, some of which are found in mammalian tissues^{17–19}.

DAO is localized to peroxisomes, whereas 3MST is mainly found in mitochondria^{14,20}. Mitochondria and peroxisomes, which are essential cellular organelles in mammals, exchange various metabolites as well as enzymes via a specific form of vesicular trafficking, and are usually in close proximity to each other or have physical contact²¹. 3MST along with DAO can produce H_2S by the interaction of both organelles.

The present study identifies a novel pathway for H_2S production from *D*-cysteine and its cytoprotective role in the cerebellum and the kidney.

Results

H_2S production from *D*-cysteine. When we examined the production of H_2S from brain homogenates, we found that H_2S was produced from *D*-cysteine, a negative control for *L*-cysteine (Fig. 1a). Although H_2S is produced from *D*-cysteine in microorganisms and plants²², its production in mammals is not well understood. Therefore, we compared the catabolism of *D*-cysteine with that of *L*-cysteine. The production of H_2S from *D*-cysteine was optimal at pH 7.4, whereas production from *L*-cysteine was greatest at pH 8.4 (Fig. 1b). Next, we examined the PLP-dependent nature of the reaction because H_2S production from *L*-cysteine by 3MST/CAT and CBS depends on PLP^{1,14,15}. Our results revealed that hydroxylamine, which is an inhibitor of PLP-dependent enzymes, did not have an effect on H_2S production from *D*-cysteine except for at 0.5 mM (Fig. 1c). At 0.5 mM the production of H_2S from *D*-cysteine was increased probably due to the decrease in the competition for *D*-cysteine with PLP-dependent enzymes, which were greatly suppressed at 0.5 mM. However, at 5 mM, pH of the medium was shifted to acidic, which suppressed H_2S production from *D*-cysteine. A single freeze-thaw cycle differently affected enzymatic activity of the two pathways. Specifically, the H_2S production from *D*-cysteine was greatly increased, whereas *L*-cysteine catabolism was decreased (Fig. 1d). These observations suggest that the H_2S -producing pathway from *D*-cysteine is independent of PLP and distinct from the pathways involving *L*-cysteine.

The DAO-3MST pathway produces H_2S from *D*-cysteine. We subsequently determined the subcellular localization of the activity to identify the specific pathway involved in the production of H_2S from *D*-cysteine. We concluded that ~70% of the

H_2S -producing activity was localized to the mitochondrial fraction, which includes peroxisomes as well as mitochondria (Fig. 1e). As mitochondrial 3MST produces H_2S from 3MP, the existence of an enzyme that produces 3MP from *D*-cysteine was predicted. It has been previously proposed in hepatocytes that DAO produces 3MP (Fig. 1f)¹⁹. Indeed, 3MP was produced from *D*-cysteine in the presence of the mitochondrial fraction, and the production was suppressed by the DAO inhibitor indole-2-carboxylate (I2CA) (Fig. 1g,h)²³. Although 3MP was also produced from *L*-cysteine with α -KG or *L*-cysteine alone to a lesser extent, the production was not suppressed by I2CA. In addition, H_2S production from *D*-cysteine was suppressed by I2CA, whereas that from 3MP was not (Fig. 1i). These observations suggest that DAO uses *D*-cysteine to produce 3MP, which is in turn metabolized to H_2S by 3MST.

We next utilized recombinant 3MST and DAO to confirm the production of H_2S via the 3MST/DAO pathway. The results with the recombinant proteins mimicked the endogenous results obtained with the mitochondrial fraction (Supplementary Fig. S1), confirming that the 3MST/DAO pathway produces H_2S from *D*-cysteine.

Tissue localization of DAO and 3MST. Subsequently, the tissue localization of DAO and 3MST was examined by western blot analysis. DAO mostly localized to the cerebellum and the kidney, and 3MST was also found in both tissues (Fig. 2a–d). We next examined whether the production of H_2S from *D*-cysteine correlated with the localization of DAO and 3MST. Indeed, H_2S was mainly produced from *D*-cysteine in the cerebellum and kidney homogenates (Fig. 2e,f). These observations suggest that H_2S is mainly produced by the 3MST/DAO pathway from *D*-cysteine in the cerebellum and the kidney.

The production of H_2S by isolated mitochondria and peroxisomes from *D*-cysteine was examined. The isolated cerebellar mitochondria and peroxisomes produced H_2S from *D*-cysteine. As mitochondria contain CAT, the isolated mitochondria produced H_2S from *L*-cysteine along with α -KG (Supplementary Fig. S2a,c). H_2S production from *D*-cysteine was greater than from *L*-cysteine along with α -KG in the presence of the isolated kidney mitochondria and peroxisomes (Supplementary Fig. S2b,c). H_2S production from *L*-cysteine alone was undetectable in the presence of isolated mitochondrial and peroxisomes of both tissues.

Developmental changes in H_2S production. We then examined the production of H_2S from *D*-cysteine and *L*-cysteine and the expression of 3MST and DAO in the cerebellum and the kidney during development. In the cerebellum, the levels of DAO increased from birth until the mice were 8 weeks old, while the 3MST levels were relatively high at birth and only slightly decreased after 8 weeks (Fig. 2g). H_2S production from *D*-cysteine increased over time, reaching the maximum production after 6 weeks, and then, it decreased, whereas H_2S production from *L*-cysteine was much less and did not significantly vary over time (Fig. 2i). Similar results were obtained in the kidney (Fig. 2h,j). These observations suggest that the altered production of H_2S from *D*-cysteine observed during development correlates well with the levels of DAO and 3MST, both in the cerebellum and the kidney.

Increase in bound sulfane sulphur by *D*-cysteine administration. Bound sulfane sulphur is the major physiologically relevant pool of H_2S capable of regulating enzyme activity and cell growth²⁴. As H_2S produced from *L*-cysteine is stored as bound sulfane sulphur¹⁴, H_2S produced from *D*-cysteine may also be

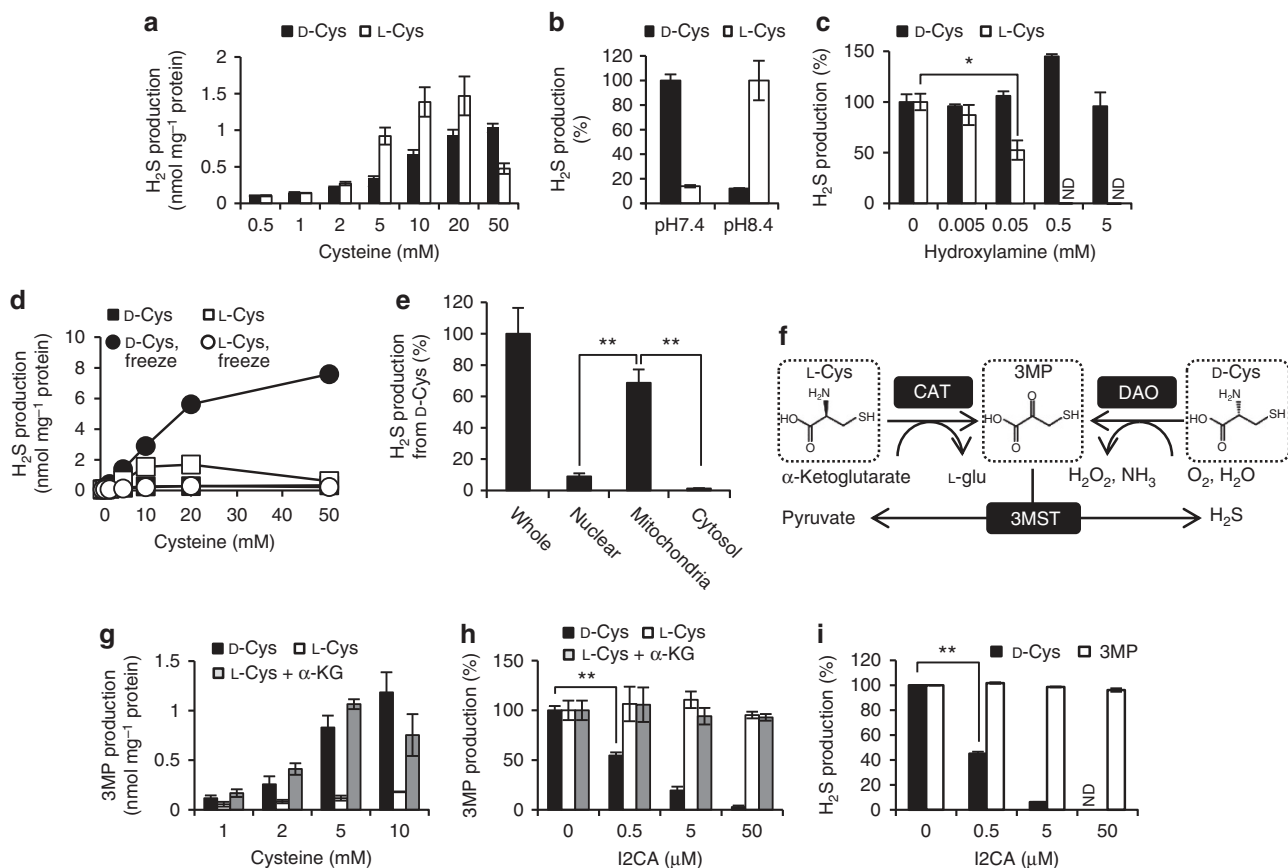


Figure 1 | H₂S-producing activity in the brain using D-cysteine and L-cysteine as substrates. **(a)** Production of H₂S from D-cysteine and L-cysteine in brain homogenate. **(b)** The optimal pH for H₂S-producing activities. Potassium phosphate buffer was used for pH 7.4, and Tris/HCl buffer for 8.4. **(c)** PLP dependency of H₂S-producing activity. **P* < 0.05. **(d)** The stability of H₂S-producing activities following a freeze-thaw cycle. Homogenates that were prepared from brains treated with or without a single freeze-thaw cycle were incubated with D-cysteine or L-cysteine. **(e)** Localization of H₂S-producing activities in the brain. Each fraction was incubated with 10 mM D-cysteine. The whole-brain activity (0.67 nmol mg⁻¹ protein) was taken as 100%. ***P* < 0.01. **(f)** Schematic representation of H₂S-producing pathways. **(g)** The 3MP-producing activities of the homogenates of mitochondrial fraction. Endogenous 3MP contents were subtracted. 3MP **(h)** and H₂S **(i)** production from D-cysteine was suppressed by indole-2-carboxylic acid (I2CA). The mitochondrial homogenates were incubated with 10 mM D-cysteine or L-cysteine or 2.5 mM α-ketoglutarate (α-KG). **P* < 0.05; ***P* < 0.01. All data are represented as the mean ± s.e.m. of at least three experiments.

stored. To examine this possibility, we orally administered D-cysteine to mice and measured the amount of bound sulfane sulphur in the cerebellum, cerebrum and kidney. The levels of bound sulfane sulphur in the cerebellum increased by 212% compared with the control 0.5 h after the administration of D-cysteine, and returned to the control level after 3 h; comparatively, L-cysteine did not significantly increase the levels of bound sulfane sulphur (Fig. 3a). Neither D-cysteine nor L-cysteine affected the cerebrum (Fig. 3b). D-Cysteine increased the levels of bound sulfane sulphur in the kidney by 236%, 218% and 196% at 0.5, 3 and 6 h after administration, respectively. L-cysteine increased the levels by 172% after 0.5 h, but they immediately declined thereafter (Fig. 3c). These observations suggest that D-cysteine more efficiently increased the levels of bound sulfane sulphur than L-cysteine in the cerebellum and the kidney.

D-cysteine protects neurons from oxidative stress. As H₂S protects neurons from oxidative stress^{8,9}, we examined whether D-cysteine protects cultures of cerebellar neurons from hydrogen peroxide (H₂O₂)-induced oxidative stress caused

using 4-3-(2-methoxy-4-nitrophenyl)-2-(4-nitrophenyl)-2H-5-tetrazolio-1,3-benzene disulfonate sodium salt (WST-8) assay, a tetrazolium salt, which is reduced by cellular activity to form intensely coloured formazans, and have often used to quantitate the number of living cells. Fifty micromolar H₂O₂ caused ~60% neuronal cell death 21 h after the application. The simultaneous application of D-cysteine with H₂O₂-protected neurons from H₂O₂-induced toxicity in a concentration-dependent manner (Fig. 3d). Both D- and L-cysteine at 1 mM increased the viability above 200% of a control. The oxidized form of D-cysteine, D-cystine, protected neurons greater than L-cystine, which is a major form in the extracellular fluid such as blood (Fig. 3e).

D-cysteine protects the kidney from ischaemia-reperfusion injury. We also examined the effect of D-cysteine on ischaemia-reperfusion injury in the kidney. The staining with 2, 3, 5-triphenyltetrazolium chloride (TTC), which is another tetrazolium salt, showed that the cortex was stained in mice orally administered D-cysteine, while it was not stained 24 h after ischaemia-reperfusion in control mice, suggesting that D-cysteine protects

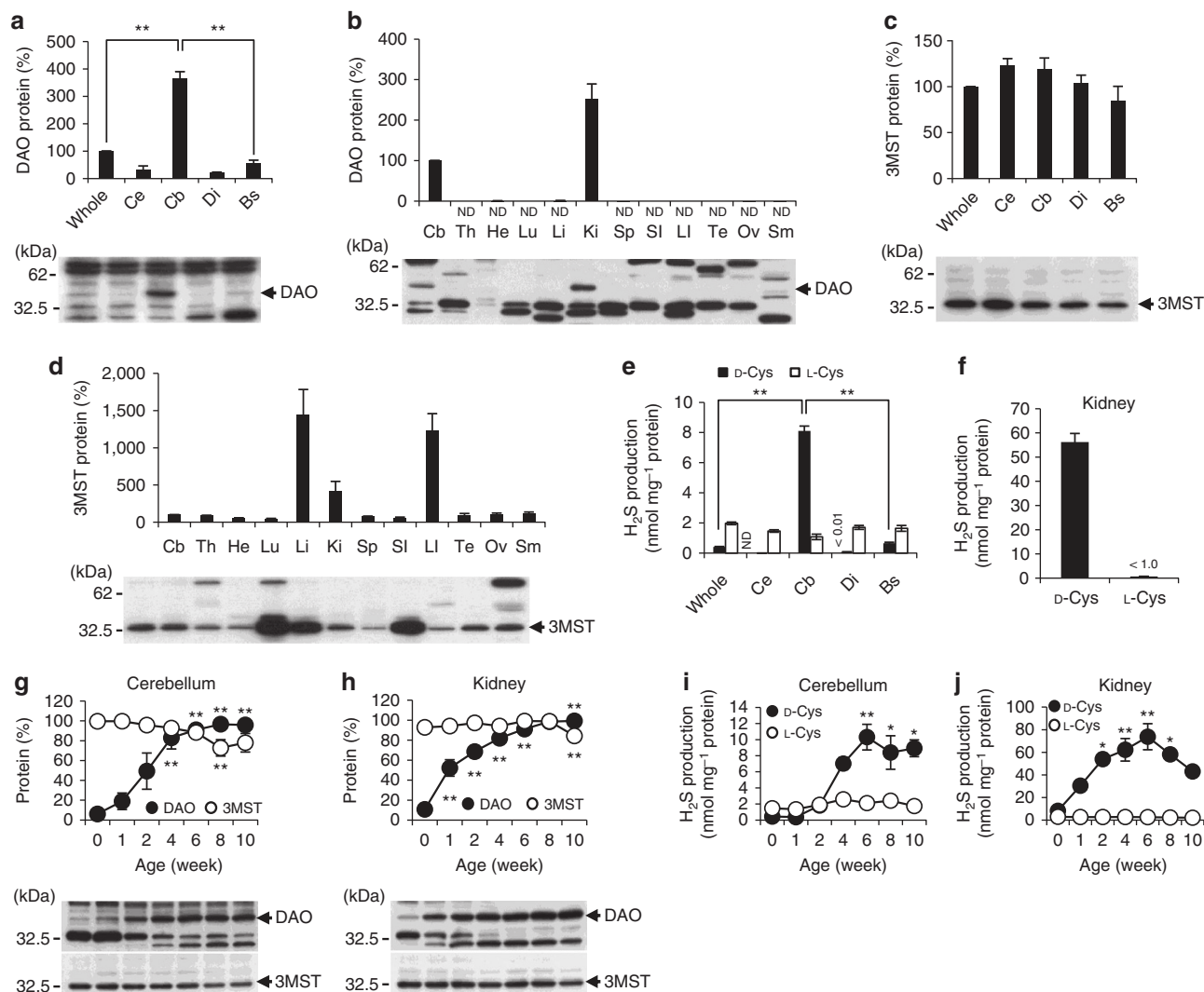


Figure 2 | Tissue localization of DAO and 3MST and developmental changes in H₂S production. (**a–d**) Tissue localization of DAO and 3MST. Western blot analysis was performed with antibodies against DAO (**a,b**) and 3MST (**c,d**) for the brain (**a,c**) and peripheral tissues (**b,d**). Relative amounts analysed by NIH Image J are shown in the upper column. ** $P < 0.01$. (**e,f**) H₂S-producing activities from D-cysteine and L-cysteine in the brain (**e**) and kidney (**f**). Homogenates were incubated with 10 mM D-cysteine or L-cysteine. ** $P < 0.01$. (**g,h**) The developmental changes in the levels of DAO and 3MST. Western blot analysis was performed and analysed by using NIH image J. (**i,j**) H₂S production from D-cysteine and L-cysteine. Homogenates were incubated with 10 mM D-cysteine or L-cysteine. (**g–j**) * $P < 0.05$; ** $P < 0.01$; versus 0 week of each group. All data are expressed as the mean \pm s.e.m. of three experiments. Bs, brain stem; Ce, cerebrum; Cb, cerebellum; Di, diencephalon; He, heart; Ki, kidney; Li, liver; LI, large intestine; Lu, lung; Ov, ovary; Sp, spleen; SI, small intestine; Sm, skeletal muscle.

the renal cortex (Fig. 3f). TTC staining in the renal cortex in mice administered D-cysteine was greater than that in mice administered L-cysteine, suggesting that D-cysteine protects the renal cortex more efficiently than L-cysteine does.

These observations were confirmed by Periodic-Acid-Schiff staining. The structure of the renal cortex was disintegrated after ischaemia-reperfusion (Fig. 3g). In contrast, it was well preserved in mice administered D-cysteine. The structure of glomeruli showed a significant difference from that of a control. In mice administered L-cysteine, glomeruli were shrunk and a wide space was observed between glomerulus and the surrounding capsule (Fig. 3g). These observations suggest that D-cysteine increases the levels of bound sulfane sulphur to protect the kidney from the ischaemia-reperfusion injury more efficiently than L-cysteine (Fig. 3c,g). Serum creatinine levels, which are increased by renal injury, were measured after the ischaemia-reperfusion of unilateral kidney. The ischaemia-reperfusion significantly increased

serum creatinine, and D-cysteine decreased the level of serum creatinine more efficiently than L-cysteine but the difference was not statistically significant (Supplementary Fig. S3).

Sources of D-cysteine. There are at least two possible sources of D-cysteine, namely, racemase-induced chiral change of L-cysteine and absorption from food. As cysteine is structurally similar to serine with an OH replaced by an SH, and because aspartate racemase is homologous to CAT, which has an affinity for both aspartate and cysteine^{25–27}, it is possible that serine racemase or aspartate racemase changes L-cysteine to D-cysteine. However, we determined that neither recombinant racemase was capable of this conversion (Supplementary Fig. S4).

We also examined the racemase activity in the brain. The cerebellar homogenates produced a substance from L-cysteine detected at a retention time of 6.98 min after 30 min, and the peak

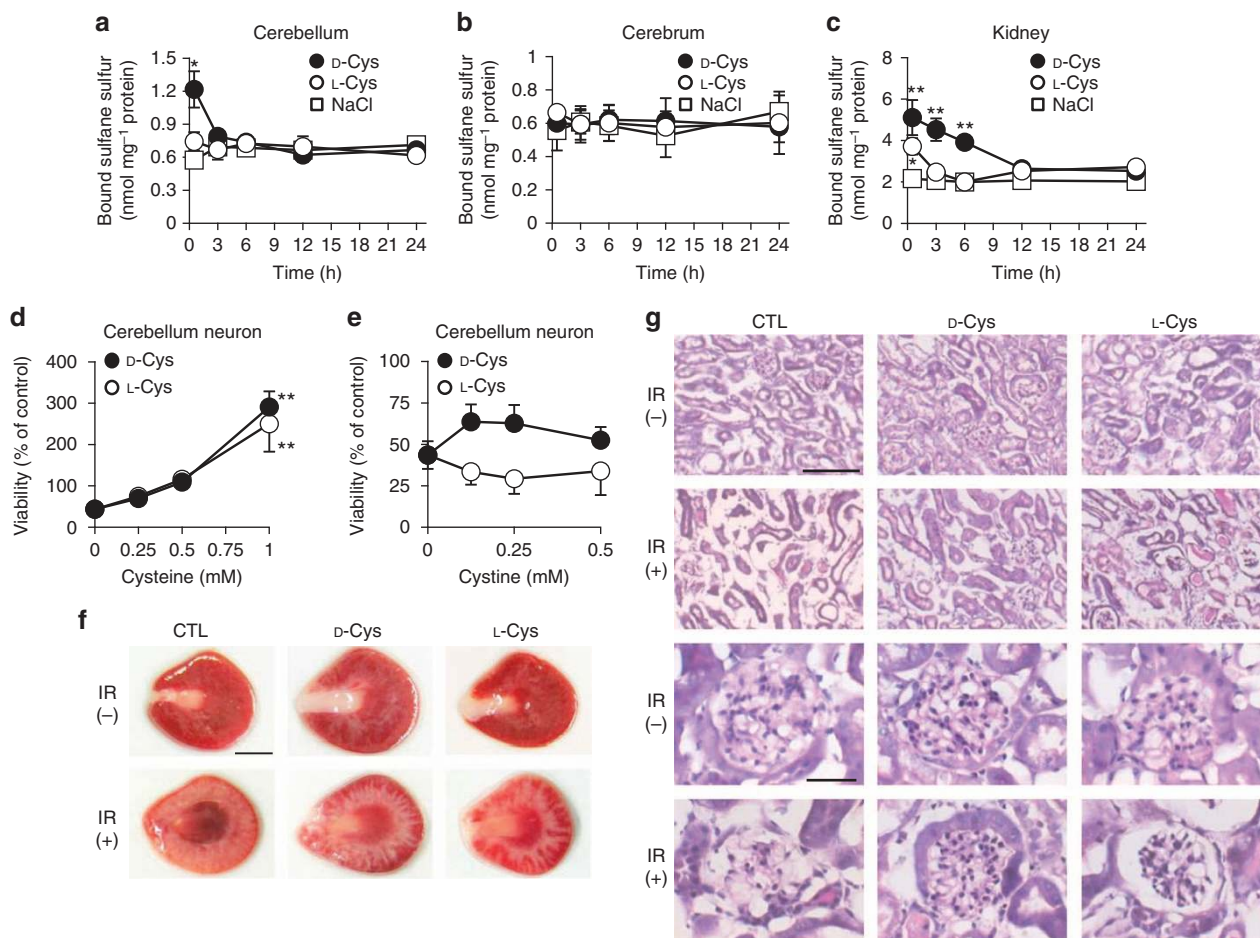


Figure 3 | D-Cysteine increases bound sulfane sulphur and protects cerebellar neurons and kidney. **(a–c)** Concentrations of bound sulfane sulphur after administration of D- or L-cysteine. Tissues were obtained from mice at the times indicated after the administration of D- or L-cysteine. * $P < 0.05$; ** $P < 0.01$. **(d, e)** The protection of cerebellar neurons from H₂O₂-induced oxidative stress by D- or L-cysteine **(d)** and D- or L-cysteine **(e)**. The viability of cultured neurons was assessed by WST-8 assay. **(f, g)** Protection of the kidney from ischaemia-reperfusion injury by D- or L-cysteine. Kidney slices were stained with TTC **(f)**, scale bar, 3 mm or Periodic-Acid-Schiff (PAS) **(g)** 24 h after the ischaemia-reperfusion (IR) injury with or without the oral administration of D- or L-cysteine. The structure of the renal cortex **(g)**, upper two columns, scale bar, 400 μ m and that of glomerulus on a magnified scale **(g)**, lower two columns, scale bar, 40 μ m were shown. All data are represented as the mean \pm s.e.m. of at least three experiments.

was shifted to 6.93 min after 60-min incubation (Supplementary Fig. S5b,c). However, considering the difference in the retention time, we determined that neither peak was identical to D-cysteine (6.34 min), (Supplementary Fig. S5e).

Discussion

L-Amino acids are nonenzymatically racemized by heat and alkaline conditions applied during food processing, and 21–44% of L-cysteine is changed to D-cysteine by alkaline treatment^{28,29}. D-cysteine may thus be mostly provided from food. Although D-cysteine was not detected in tissues after the oral administration, the levels of bound sulfane sulphur were increased to protect the kidney from ischaemia-reperfusion injury (Fig. 3). These observations with the previous finding that D-cysteine is easily absorbed through the gastrointestinal tract and enters the blood stream³⁰ suggest that D-cysteine may be immediately metabolized to produce bound sulfane sulphur in tissues.

Both D-cysteine and L-cysteine protect cerebellar neurons from oxidative stress, and further increase cell viability at the higher concentrations (Fig. 3d). Our previous study similarly showed that H₂S specifically increased cell viability of neuronal cells³¹.

D-Cysteine protected cerebellar neurons more efficiently than L-cysteine (Fig. 3e). It is probably because the transport activity for D-cysteine is greater than that for L-cysteine when compared at the same concentrations³².

D-cysteine protects the renal cortex more efficiently than L-cysteine (Fig. 3f,g). As H₂S-producing enzymes, CBS, CSE and 3MST along with DAO are localized to the renal cortex^{33–36}, the administration of D-cysteine or L-cysteine protects the renal cortex but not the medulla (Fig. 3f,g). D-cysteine, which increases the levels of bound sulfane sulphur more greatly than L-cysteine, more efficiently protects the renal cortex than L-cysteine does (Fig. 3f,g). The observation also confirms the interaction between mitochondrial 3MST and peroxisomal DAO *in vivo*.

L-Cysteine is an excitotoxin comparable in potency to other excitatory amino acids³⁷. In contrast, the subcutaneous administration of D-cysteine does not cause excitotoxic damage to the brain³⁸. Therefore, systemic application of D-cysteine may be less toxic than that of L-cysteine. The administration of D-cysteine may provide a new therapeutic approach to protect specific cells from oxidative stress or ischaemia-reperfusion injury through its conversion to H₂S via a novel pathway with 3MST and DAO.

Methods

Determination of H₂S-producing activity. All animal procedures were approved by the National Institute of Neuroscience Animal Care Use Committee. Whole-brain extracts were prepared from male C57BL/6N mice (Clea Japan, Tokyo, Japan) with nine volumes of ice-cold buffer A (100 mM potassium phosphate buffer (pH 7.4), 1 mM dithiothreitol (DTT), and a complete protease inhibitor cocktail (Roche Diagnostics, Mannheim, Germany)) by using a Potter-type glass homogenizer with a Teflon pestle or a Polytron-type homogenizer. To examine the stability of the enzymes following a single freeze-thaw cycle, the whole brain was immediately frozen at -80°C after excision and thawed on ice. The extracts of the frozen-and-thawed tissue were prepared as described above. Enzymatic production of H₂S was measured by following our previously described method¹⁰.

Subcellular fractionation. Whole-mouse brains were homogenized with three volumes of the fractionation buffer (5 mM MOPS (pH 7.4), 0.32 M sucrose, 1 mM EDTA, 0.1% ethanol and protease inhibitor cocktail) by using a Potter-type glass homogenizer with a Teflon pestle (1,200 r.p.m., three strokes) and centrifuged at 1,000 g for 10 min. The pellet was rehomogenized as described above and centrifuged twice at 600 g for 10 min. The final pellet was recovered as a nuclear fraction that was resuspended in fractionation buffer at 10% of the original volume. The post-nuclear supernatant was centrifuged at 16,500 g for 30 min to obtain the cytosolic and mitochondrial fractions. The mitochondrial pellets were resuspended in a volume of fractionation buffer equivalent to the supernatant.

Determination of 3MP. The mitochondrial pellets were resuspended in 10% of the original volume with 100 mM potassium phosphate buffer (pH 7.4) and a complete protease inhibitor cocktail. For the enzyme reactions, 11 μl of D-cysteine was added to 0.1 ml of the mitochondrial fraction to the final concentrations indicated in the figure legend and the reaction mixture was incubated at 37°C for 50 min. The thiol group of 3MP is highly reactive and can form dimers in multiple combinations through interactions with itself and other thiols³⁹. To block the reactivity, an equal volume of 0.1 M N-ethylmaleimide, which is a thiol-alkylating agent, was added to the mixture and incubated at 37°C for 10 min. After centrifugation at 12,000 g for 10 min, the supernatant was centrifuged through Amicon Ultra centrifugal filters (Millipore, Bedford, MA) with a cutoff of 3 kDa. The alkylated 3MP in the filtrate was determined by the pre-labelling high-performance liquid chromatography assay using 1,2-diamino-4,5-dimethoxybenzene⁴⁰.

Western blot analysis. The protein samples (10 μg) were fractionated by SDS-PAGE on a 12 or 15% polyacrylamide gel (DRC, Tokyo, Japan) and transferred to a polyvinylidene difluoride membrane (Millipore). The membrane was blocked by using PBS-T (137 mM NaCl, 10 mM Na₂HPO₄, 2.7 mM KCl, 1.8 mM KH₂PO₄ and 0.1% Tween 20) containing 2% skim milk (Wako, Osaka, Japan) overnight at 4°C and incubated with either anti-MPST (1:3,000; Atlas Antibodies AB, Stockholm, Sweden) or anti-DAO⁴¹ (BioResource room, The Institute for Enzyme Research, The University of Tokushima) at 4°C for 4 h. After additional 2-h incubation with the secondary anti-rabbit antibodies conjugated with horseradish peroxidase diluted at 1:20,000 (GE Healthcare), the binding of the antibodies was visualized using the Millipore Immobilon Western Chemiluminescent HRP substrate (Millipore) or Western Lightning ECL Pro (PerkinElmer, Waltham, MA).

Cell culture. Primary cerebellar neurons were prepared from embryonic day 16 imprinting control region (ICR) mice (Clea Japan). The cerebellum was digested with 0.25% trypsin and 0.1% DNase I in Ca²⁺/Mg²⁺-free PBS for 10 min at 37°C . After pipetting cells were seeded on poly-D-lysine-coated 96-well microtiter dishes at 100,000 cells per 100 μl in each well. The cells were cultured in Neurobasal medium (Life technologies, Carlsbad, CA) supplemented with B27, 0.5 mM L-glutamine, 100 U ml⁻¹ penicillin and 100 μg ml⁻¹ streptomycin at 37°C in 10% CO₂. For the toxicity study, 5-day-old cells were exposed to 50 μM H₂O₂, and then cysteine or cystine was added to the concentrations indicated in the figure legend. WST-8 (2-(2-methoxy-4-nitrophenyl)-3-(4-nitrophenyl)-5-(2,4-disulphophenyl)-2H-tetrazolium, monosodium salt) viability assay was performed with the cell-counting kit-8 (Dojindo, Kumamoto, Japan) 21 h after the application of H₂O₂. Ten microliters of WST-8 was added to each well, cells were incubated at 37°C for 2 h and absorption values at 450 nm were determined using a microplate reader. Values of samples were normalized by subtracting values of direct reduction of WST-8 by the chemicals.

Administration of cysteine. Male ICR mice (Clea Japan) with a body weight of 30–40 g were housed with *ad libitum* access to food and water. The animals were divided into three groups: D-cysteine, L-cysteine and control. The D- and L-cysteine (8 mmol kg⁻¹ body weight) were orally administered by gastric intubation. The cysteine solution was adjusted to pH 2.0–2.5 with 5 M NaOH. The control mice received the same amount of NaCl. At the times indicated in the figure legend after oral administration, the mice were transcardially perfused with saline. The tissues were immediately removed, frozen with liquid nitrogen and then stored at -80°C until the bound sulfane sulphur was measured.

Determination of bound sulfane sulphur. To determine the amount of bound sulfane sulphur, the frozen tissues were thawed on ice and homogenized with ice-cold lysis buffer B (10 mM potassium phosphate buffer (pH 7.4), 1% TritonX-100 and the protease inhibitor cocktail) with 10 mM of hydroxylamine and sodium benzoate (each), which suppress the activities of PLP-dependent enzymes and DAO that are involved in enzymatic H₂S production. The lysates were centrifuged at 12,000 g for 10 min, and the supernatant was recovered. For measurement of the H₂S released from the bound sulfane sulphur, 0.1 ml of sample (2.5 mg protein per ml) and 0.1 ml of 15 mM DTT in 0.2 M Tris/HCl (pH 9.0) were placed in a 15-ml centrifugation tube, sealed with Parafilm and then incubated at 37°C for 50 min. After adding 0.4 ml of 1 M sodium citrate buffer (pH 6.0), the mixtures were incubated at 37°C for 10 min with shaking at 125 rpm on a rotary shaker NR-3 (Taitec) to facilitate the release of H₂S gas from the aqueous phase. H₂S concentration was determined according to the method described for the determination of the H₂S produced by the enzymes. A reaction mixture without the samples was used as a control to account for the release of H₂S from DTT.

Renal ischaemia-reperfusion. Male ICR mice were anesthetized with an intraperitoneal injection of sodium pentobarbital (65 mg kg⁻¹ body weight). Body temperature was maintained with a heating blanket. Male ICR mice with a body weight of 30–40 g were divided into three groups: D-cysteine, L-cysteine and a control. The D- and L-cysteine (8 mmol kg⁻¹ body weight) were orally administered 30 min before ischaemia. Unilateral renal blood flow was interrupted for 30 min by clamping a nontraumatic vascular clamp over renal pedicles. The control mice were subjected to the same surgical procedures without the oral administration.

Evaluation of ischaemia-reperfusion injury. Twenty-four h after reperfusion, the kidney was sliced coronally into 2-mm-thick sections by a tissue slicer. To visualize the area of the ischaemia-reperfusion injury, the slices were incubated with 2% TTC in saline at room temperature for 30 min. For histological analysis with a light-microscopy, mice were perfused with 4% (w/v) paraformaldehyde transcardially. The kidney was removed, post-fixed in paraformaldehyde at 4°C overnight and immersed in 5, 15 and 30% (w/v) sucrose in order at 4°C . The kidney was then submerged into an O.C.T. compound and frozen. The kidney sections with 4 μm thickness were cut by a cryostat at -20°C . The sections were stained with Periodic-Acid-Schiff to highlight glomerular capillary loops and tubular epithelium. Stained sections were observed with an Axiophoto microscope (Carl Zeiss, Jena, Germany) equipped with a cooled CCD digital camera (Penguin 600CL; Pixera, Los Gatos, CA).

Statistics. All statistical analyses of the data were performed using Excel 2007 with the add-in software Statcel2 (OMS, Saitama, Japan). Differences between three or more groups were analysed with one-way analysis of variance. *Post hoc* multiple comparisons were made using the Bonferroni test.

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Acknowledgements

We thank Dr David Schubert for a critical reading of the manuscript. This work was supported by a grant from the National Institute of Neuroscience, a KAKENHI (23659089) Grant-in-Aid for Challenging Exploratory Research to H.K., a KAKENHI (23700434) Grant-in-Aid for Young Scientists (B), a Health Labour Sciences Research Grant from the Ministry of Health Labour and Welfare to N.S., and a KAKENHI (22590258) Grant-in-Aid for Scientific Research (C) to Y.K.

Author contributions

N.S., S.K., M.I., M.T. and Y.K. provided the experimental data. Y.O. provided suggestions for experiments. K.F. provided an antibody against DAO. N.N. provided an antibody against 3MST. N.S. and H.K. planned the experiments.

Additional information

Supplementary Information accompanies this paper at <http://www.nature.com/naturecommunications>

Competing financial interests: The authors declare no competing financial interests.

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How to cite this article: Shibuya, N. *et al.* A novel pathway for the production of hydrogen sulfide from D-cysteine in mammalian cells. *Nat. Commun.* **4**:1366 doi: 10.1038/ncomms2371 (2013).