496

A Novel Selective and Sensitive Electrochemical Sensor for Insulin Detection

Zulkarnain¹, Suprapto¹, Taslim Ersam², Fredy Kurniawan¹*

¹Instrumentation and Analytical Sciences Laboratory, Department of Chemistry, Institut Teknologi Sepuluh Nopember, Surabaya 60111, Indonesia ²Natural Product Chemistry and Synthesis Laboratory, Department of Chemistry, Institut Teknologi Sepuluh Nopember, Surabaya 60111, Indonesia

*Corresponding author, e-mail: fredy@chem.its.ac.id

Abstract

A novel selective and sensitive electrochemical sensor for insulin detection has been fabricated and investigated. The electrochemical sensor was made from a mixture of silica gel, chitosan and nickel hydroxide (Ni(OH)₂) nanoparticles, which was mounted on a silver wire and covered by a glass tube (silica gel/chitosan/Ni(OH)₂ nanoparticles paste electrode). The sensor was characterized using cyclic voltammetry (CV) and electrochemical impedance spectroscopy (EIS). The anodic and cathodic currents of the silica gel/chitosan paste electrode with Ni(OH)₂ are 580 μ A and -750 μ A, respectively. Without Ni(OH)₂, however, the currents are 150 μ A and -250 μ A, respectively. The sensitivity and limit of detection of the silica gel/chitosan/Ni(OH)₂ nanoparticle paste electrode for insulin detection are 5573 × 10⁵ pA/pMcm² and 0.25 pM, respectively. The sensor also shows good reproducibility of measurement for 35 days with an RSD of 0.29%. The fabricated electrodes also show good reproducibility, with an RSD of 1.39%.

Keywords: insulin, silica gel, chitosan, nickel hydroxide nanoparticles

Copyright © 2016 Institute of Advanced Engineering and Science. All rights reserved.

1. Introduction

The increase in the glucose concentration in the blood is influenced by the amount of carbohydrate that is consumed by a person. When the level of glucose in the blood increases, the pancreas produces insulin as a response to the amount of glucose to control the level of glucose in the blood [1]. Insulin is an important hormone that consists of polypeptide chains A and B, which are composed of 21 and 30 amino acids, respectively [2]. The inability of the pancreas to produce enough insulin to control glucose concentrations in the blood is called diabetes. According to the International Diabetes Federation (IDF), in 2014, there were 387 million cases of diabetes in the world, of which 9 million were in Indonesia. In 2012, the World Health Organization (WHO) estimated that approximately 1.5 million people died from diabetes. There are two types of diabetes. When the pancreas in the human body cannot produce insulin at all, it is called diabetes type 1. The second type is diabetes type 2, which occurs when the pancreas can still produce insulin but the amount is not enough to control glucose in the blood at a normal concentration [3-5]. Therefore, to obtain information related to the type of diabetes from the patient, it is important to have a method or instrument to measure the insulin in the blood directly before attempting medical treatment for the diabetes patient. The determination of insulin concentrations has been investigated intensively because it can be used as a tool to help treat diabetes patients.

The detection of a definite compound, e.g., insulin [6], glucose [7–9], dopamine [10], and urea [11], using a modified electrode with an electrochemical method has been reported. The electrochemical method has several advantages: the sample can be measured directly without complicated sample preparation, and the analysis process takes less than 5 minutes and does not use chemical reagents, so it is more effective and efficient [12]. Other methods such as high-performance liquid chromatography (HPLC) and immunoassay methods require complicated sample preparation, are time consuming, and require reagents such as HCl and dichloromethane [13] or polyethyleneimine and 3-amino propyltriethoxysilane [14] for detection, which are relatively expensive. Several modified electrodes for insulin determination have been

497

reported. A silica gel/carbon paste electrode was used to measure insulin in phosphate buffer. The sensitivity and limit of detection (LOD) of the electrode were 1511 pA/pMcm² and 36 pM, respectively [15]. Conversely, the sensitivity and LOD of a nickel nanoparticle (NiOx)/glassy carbon/guanine electrode were 803 pA/pMcm² and 22 pM, respectively [16]. Besides for insulin sensor, the carbon material was applied for gas sensor [17]. Salimi et al.'s electrode had a better LOD than Jaafariasl et al.'s electrode but worse sensitivity. In the present work, we fabricate a silica gel/chitosan/Ni(OH)₂ paste electrode to measure insulin in 0.1 M phosphate buffer (pH 7.4) that demonstrates better performance in terms of sensitivity and LOD than both Salimi et al.'s electrodes.

2. Research Method

2.1. Chemicals and Materials

Silica gel G.60, K_2HPO_4 , and KH_2PO_4 were purchased from Merck and used without any purification. Insulin (100 IU/mL) was purchased from R. Lantus. Chitosan was prepared by Instrumentation and Analytical Sciences Laboratory, Institut Teknologi Sepuluh Nopember. Nickel hydroxide (Ni(OH)₂) nanoparticles were prepared according to the procedure developed by Budipramana et al. Briefly, nickel metal was electrolysed in sodium citrate solution at 100 °C. The potential of electrolysis was kept constant at 55 V via stirring for 30 minutes. The Ni(OH)₂ nanoparticle solution obtained was used in all experiments [18–20]. Solid paraffin, glass tubes (inner diameter, 0.2 cm; length, 5 cm), silver wire (outer diameter, 0.8 mm; length, 7 cm), and demineralized water were purchased from a local market.

2.2. Instrumentation

All electrochemical measurements were carried out using a potentiostat/galvanostat from eDAQ (potentiostat E161 and e-corder 410, which is equipped with e-chem software vs 2.0.1). A three-electrode cell system was used in all measurements unless otherwise described, with platinum as the counter electrode (CE), Ag/AgCl (KCl 3 M) as the reference electrode (RE), and a modified electrode as the working electrode (WE).

2.3. Fabrication of the Silica Gel/Chitosan Paste Electrode with Ni(OH)₂

The silica gel/chitosan/Ni(OH)₂ paste electrode was fabricated according to the following procedure. Silica gel and chitosan were mixed in a 4:6 weight ratio and thoroughly hand-mixed with an agate mortar and pestle to produce a homogeneous mixture. The mixture was heated to 65°C, and Ni(OH)₂ solution (2 mL/100 mg of the mixture) and paraffin (15 % from the mixture weight) were then added. The hand-mixing was continued until the mixture became slightly solid. The resulting mixture was inserted in the bottom of the tube glass (geometrical surface area: 0.5 cm²) and connected to silver wire from the top of the tube glass. The surface of the paste electrode was then polished using abrasion paper grade 2000. The silica gel/chitosan/Ni(OH)₂ paste electrode was used to measure insulin at concentrations of 0.0, 0.1, 0.3, 0.5, and 0.7 pM in 0.1 M phosphate buffer (pH 7.4) with cyclic voltammetry in the potential range of -1.0 to 1.0 V and a scan rate of 100 mV/s. The limit of detection (LOD) was calculated according to three times the standard deviation of the cyclic voltammetry response. The silica gel/chitosan/Ni(OH)₂ paste electrode was used to measure 1.0 μM glucose, 1.0 μM urea, 1.0 μM ascorbic acid, and 1.0 μM insulin. All of the foreign substances and insulin were dissolved in 0.1 M phosphate buffer at pH 7.4. Five silica gel/chitosan/Ni(OH)₂ paste electrodes were fabricated and used to measure 1.0 µM insulin. One of the silica gel/chitosan/Ni(OH), electrodes was used to measure 1.0 M insulin every day for 7 days and then on the 14th, 21st, 28th, and 35th days.

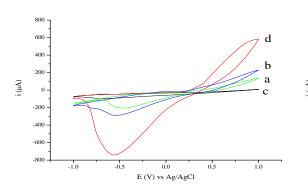
3. Results and Analysis

3.1. Electrochemical Characterization of the Silica Gel/Chitosan Paste Electrode

The preliminary experiment was carried out to observe the influence of Ni(OH)₂ on the silica gel/chitosan paste electrode in the response of 1.0 μ M insulin. The cyclic voltammograms of the silica gel/chitosan paste electrode with (c, d) and without (a, b) Ni(OH)₂ are shown in Figure 1. From the Figure 1 we know that the peak anodic current (i_{pa}) on the silica gel/chitosan paste electrode with Ni(OH)₂ (d) (580 μ A) and without Ni(OH)₂ (b) (150 μ A). The i_{pa} value

indicates the peak of oxidation to insulin at a peak anodic potential (E_{pa}) of 0.8 V. Conversely, there is also a reduction of insulin indicated by the value of the peak cathodic current (i_{pc}) with Ni(OH)₂ (d) (-750 µA) and without Ni(OH)₂ (-250 µA) that occurred at the peak cathodic potential (E_{pc}) of -0.5 V.

1200



900 600 300 -300 -300 -120 -1.0 -1.0 -0.5 -0.0 -0.5 -0.0 -0.5 -0.0 -0.5 -0.0 -0.5 -0.0 -0.5 -0.0 -0.5 -0.0 -0.5 -0.0 -0.5 -0.0 -0.5 -0.0 -0.5 -0.0 -0.5 -0.0 -0.5 -0.0 -0.5 -0.0 -0.5 -0.

Figure 1. The Cyclic Voltammogram of Phosphate Buffer, pH 7.4, solution (a, c) and 1.0 μM insulin in phosphate buffer, pH 7.4, solution (b, d) for silica gel/chitosan (a, b) and silica gel/chitosan/Ni(OH)₂ paste electrodes (c, d). Scan rate, 100 mV/s

Figure 2. The Linear sweep voltammogram of insulin using silica gel/chitosan/Ni(OH)₂ nanoparticles paste electrode, (a) Anodic sweep, (b) Cathodic sweep, and (c) Anodic and Cathodic sweep of blank solutions

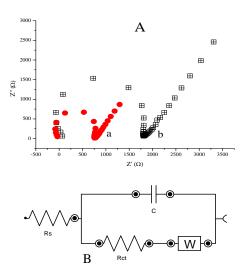


Figure 3. (A) Nyquist plot of silica gel/chitosan paste electrodes with (a) and without $Ni(OH)_2$ (b) in 1.0 μ M insulin solution in 0.1 M phosphate buffer at pH 7.4. (B) EIS equivalent circuit used to fit EIS diagrams.

The presence of Ni(OH)₂ on a silica gel/chitosan paste electrode can be used as a good insulin sensor because Ni(OH)₂ has good electrocatalytic properties and can oxidize and reduce insulin on the electrode surface. The i_{pa} and i_{pc} values obtained upon blank measurement with the silica gel/chitosan paste electrode with (c) are much smaller than the i_{pa} and i_{pc} values obtained without Ni(OH)₂. This indicates that the silica gel/chitosan paste electrode with Ni(OH)₂ is more sensitive than that without Ni(OH)₂ because the Ni(OH)₂ has high reactivity properties so fast electron transfer occurs and is able to interact with insulin detection even at a low concentration. A possible reaction between Ni(OH)₂ and insulin (amino acid) is the following: [21].

 $Ni(OH)_2 \leftrightarrow NiOOH + e^{-} + H^{+}$ (1)

NiOOH + Insulin \rightarrow Ni(OH)₂ + Insulin_(red)

Figure 2 shows the Linear Sweep Voltammetry of the insulin with (a) for anodic sweep, (b) for cathodic sweep in phosphate buffer pH 7.4 using silica gel/chitosan/Ni(OH)₂ nanoparticles paste electrode. Anodic sweep does not have any peak of oxidation of insulin. Increasing of current in the end of the voltammogram is probably due to the oxidation of Ni(OH)₂. Conversely, the cathodic sweep show a reduction peak of insulin. This is indicated that the insulin only can be reduced by this electrode while in the other works reported oxidation of the insulin [16, 21]. The most probably reduction reaction is carboxylic acid group change to aldehyde group.

The silica gel/chitosan paste electrodes with and without $Ni(OH)_2$ were also characterized by electrochemical impedance spectroscopy (EIS) in 0.1 M phosphate buffer at pH 7.4. Figure 3 (A) shows the Nyquist plot of the silica gel/chitosan paste electrode with (a) and without $Ni(OH)_2$ (b), and Figure 3 (B) shows the equivalent circuit. R_{ct} , R_s C, and W are the charge transfer resistance, solution resistance, capacitance, and Warburg impedance, respectively.

The R_{ct} values for the silica gel/chitosan paste electrode with and without Ni(OH)₂ are 767 Ω and 1400 Ω , respectively. This indicates that the silica gel/chitosan paste electrode with Ni(OH)₂ is approximately two times more conductive than that without Ni(OH)₂. Higher conductivity means better electron transfer during the oxidation and reduction reactions on the electrode surface, which in turn can increase the sensitivity and limit of detection of the electrode [22, 23].

The calculated C values of the silica gel/chitosan paste electrode with and without $Ni(OH)_2$ are 1.91 nF and 1.29 nF, respectively. This indicates that the ability of the silica gel/chitosan paste electrode with $Ni(OH)_2$ to store electrical energy is greater than that of the electrode without $Ni(OH)_2$. The W values of the silica gel/chitosan paste electrodes with and without $Ni(OH)_2$ are 187 μ Mho and 66.4 μ Mho, respectively. This indicates that the mass transport occurring at the silica gel/chitosan paste electrode with $Ni(OH)_2$ is twice that without $Ni(OH)_2$. When the mass transport increases, conductivity also increases and can increase the electrode sensitivity [21, 24]. A higher W value means better electrical potential on the electrode surface, and this can increase the electric induction that is proportional to the current flowing in the electrode [25].

3.2. Calibration Curve of Insulin by Silica Gel/Chitosan/Ni(OH)₂ Paste Electrode

The silica gel/chitosan/Ni(OH)₂ paste electrode was studied to measure insulin at different concentrations. Figure 4 shows (A) the cyclic voltammograms of insulin at various concentrations and (B) a calibration curve that is obtained using an i_{pc} value of -0.5 V. The silica gel/chitosan/Ni(OH)₂ paste electrode was studied to measure insulin at different concentrations. Figure 4 shows (A) the cyclic voltammograms of insulin at various concentrations and (B) a calibration curve that is obtained using an i_{pc} value of -0.5 V. The silica gel/chitosan/Ni(OH)₂ paste electrode was studied to measure insulin at different concentrations. Figure 4 shows (A) the cyclic voltammograms of insulin at various concentrations and (B) a calibration curve that is obtained using an i_{pc} value of -0.5 V.

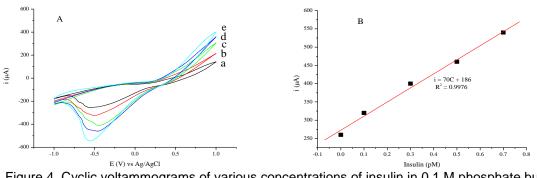


Figure 4. Cyclic voltammograms of various concentrations of insulin in 0.1 M phosphate buffer solution at pH 7.4 (A): 0 pM (a), 0.1 pM (b), 0.3 pM (c), 0.5 pM (d), 0.7 pM (e). Calibration curve of insulin (B)

499

(2)

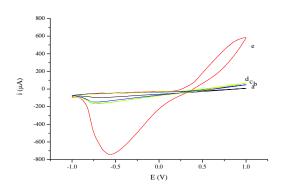
The correlation coefficient (R^2), sensitivity and limit of detection (LOD) of the calibration curve are 0.9976, 5573 × 10⁵ pA/pM cm² and 0.25 pM, respectively. The performance of several electrodes for insulin detection is shown at Table 1.

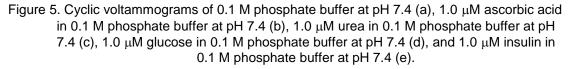
Table 1. Performance Comparison for Several Electrodes				
Researcher	Electrode	LOD (pM)	Sensitivity (pA/pMcm)	pН
Salimi et al, 2008	NiOx/glassy carbon/ guanine	22	803	7.4
Jafariasl et al, 2011	Silica gel/carbon	36	1511	2.0
Rafiee et al, 2013	NiOx/carbon nanotube	6100	1800×10 ³	7.4
This work, 2016	Silica gel/chitosan /Ni(OH)₂	0.25	5573×10⁵	7.4

Table 1 shows that the silica gel/chitosan/Ni(OH)₂ paste electrode has a higher sensitivity and LOD compared to the other working electrode. Accordingly, Ni(OH)₂ has good electrocatalytic properties against insulin. Thus, the silica gel/chitosan/Ni(OH)₂ paste electrode can be promoted as an alternative method for insulin determination.

3.3. Interference Study

The response of the silica gel/chitosan/Ni(OH)₂ paste electrode was studied against glucose, urea, and ascorbic acid. Figure 5 shows voltammograms for (a) ascorbic acid, (b) urea, (c) glucose, and (d) insulin.





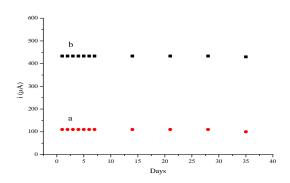


Figure 6. The anodic current (a) and cathodic current (b) of silica gel/chitosan/(NiOH)₂ paste electrode for the determination of 1.0 μ M insulin in 0.1 M phosphate buffer at pH 7.4 for 35 days.

The i_{pa} and i_{pc} values of insulin on the silica gel/chitosan/Ni(OH)₂ paste electrode are 590 μ A and -710 μ A, respectively. These are higher than the i_{pa} and i_{pc} values of glucose, urea, or ascorbic acid. This indicates that the electrode has good selectivity to insulin.

3.4. Stability of Electrode

The stability of the silica gel/chitosan/Ni(OH) $_2$ paste electrode was evaluated for 35 days. The results obtained can be seen in Figure 6.

Five silica gel/chitosan/Ni(OH)₂ paste electrodes were used to measure insulin. The results show that the electrode has a relative standard deviation (RSD) of 1.39%, which means that the electrodes can be fabricated with good precision. The anodic and cathodic currents of insulin on the surface of the silica gel/chitosan/Ni(OH)₂ paste electrode remain constant for 35 days at 110 μ A and -430 μ A, respectively. This indicates that the silica gel/chitosan/Ni(OH)₂ paste electrode has a good reproducibility of measurement (RSD 0.29 %).

4. Conclusions

The silica gel/chitosan/Ni(OH)₂ paste electrode has shown good performance for insulin determination. The sensitivity and LOD of the electrode are $5573 \times 10^5 \text{ pA/pM cm}^2$) (R² = 0.9976) and 0.25 pM, respectively. This electrode also shows good selectivity for insulin detection; the signals of the insulin are relatively high in comparison with those of glucose, ascorbate acid, and urea. The reproducibility of the five electrodes is relatively good, with an RSD of 1.39 %. The stability of the electrode for 35 days is also relatively good, with an RSD of 0.29 %.

Acknowledgements

We acknowledge the Indonesian Government, especially the Minister of Research, Technology, and Higher Education, for their financial support.

References

- Scheuing N, Thon A, Konrad K et al. Carbohydrate intake and insulin requirement in children, adolescents and young adults with cystic fibrosis-related diabetes: A multicenter comparison to type 1 diabetes. *Clinical Nutrition*. 2015; 34(4): 732–738.
- [2] Nasrallah, Nasrallah, L Raymond Reynolds. Insulin Degludec, the New Generation Basal Insulin or Just another Basal Insulin? *Clinical Medicine Insights: Endocrinology and Diabetes*. 2012: 31.
- [3] Tama BA, S RF, Hermansyah H. An Early Detection Method of Type-2 Diabetes Mellitus in Public Hospital. *TELKOMNIKA (Telecommunication Computing Electronics and Control)*. 2013; 9(2):287-294.
- [4] Gough SCL. A review of human and analogue insulin trials. *Diabetes Research and Clinical Practice*. 2007; 77(1): 1-15.
- [5] Joshi S, Joshi P. A review of insulin and insulin regimens in type 2 diabetes. *South African Family Practice*. 2009.
- [6] Arvinte A, Westermann AC, Sesay AM, Virtanen V. Electrocatalytic oxidation and determination of insulin at CNT-nickel–cobalt oxide modified electrode. Sensors and Actuators B: Chemical. 2010; 150(2): 756–763.
- [7] El-Ads EH, Galal A, Atta NF. Electrochemistry of glucose at gold nanoparticles modified graphite/SrPdO3 electrode-Towards a novel non-enzymatic glucose sensor. *Journal of Electroanalytical Chemistry*. 2015; 749: 42–52.
- [8] Kurniawan F, Tsakova V, Mirsky VM. Gold nanoparticles in nonenzymatic electrochemical detection of sugars. *Electroanalysis.* 2006; 18(19-20): 1937–1942.
- [9] Kurniawan F, Fitriyana F. Polyaniline-Invertase-Gold Nanoparticles Modified Gold Electrode for Sucrose Detection. *Indonesian Journal of Chemistry*. 2015; 15(3): 226–233.
- [10] Kurniawan F, Tsakova V, Mirsky VM. Analytical Applications of Electrodes Modified by Gold Nanoparticles: Dopamine Detection. *Journal of Nanoscience and Nanotechnology*. 2009; 9(4): 2407– 2412.
- [11] Ahmad R, Tripathy N, Hahn Y-B. Highly stable urea sensor based on ZnO nanorods directly grown on Ag/glass electrodes. *Sensors and Actuators B: Chemical.* 2014; 194: 290–295.
- [12] Businova P, Prasek J, Chomoucka J et al. Voltammetric Sensor for Direct Insulin Detection. *Procedia Engineering*. 2012; 47: 1235–1238.

- [13] Yilmaz B, Kadioglu Y. Determination of insulin in rabbit plasma using high-performance liquid chromatography. Kafkas Univ Vet Fak Derg. 2012; 18(1): 31–36.
- [14] Suri CR, Jain PK, Mishra GC. Development of piezoelectric crystal based microgravimetric immunoassay for determination of insulin concentration. 1995.
- [15] Jaafariasl M, Shams E, Amini MK. Silica gel modified carbon paste electrode for electrochemical detection of insulin. *Electrochimica Acta*. 2011; 56(11): 4390–4395.
- [16] Salimi A, Noorbakhash A, Sharifi E, Semnani A. Highly sensitive sensor for picomolar detection of insulin at physiological pH, using GC electrode modified with guanine and electrodeposited nickel oxide nanoparticles. *Biosensors and Bioelectronics*. 2008; 24(4): 792–798.
- [17] Gunawan B, Rivai M, Juwono H. Characterization of Polymeric Chemiresistors for Gas Sensor. *TELKOMNIKA (Telecommunication Computing Electronics and Control).* 2012; 10(2): 275.
- [18] Zulkarnain Z, Kurniawan F, Ersam T, Suprapto S. Influence of Ni(OH)2 Nanoparticles on Insulin Sensor Sensitivity. ARPN Journal of Engineering and Applied Sciences. 2016.
- [19] Budipramana Y, Kurniawan F, Ersam T, Suprapto S. Influence of CTAB and Sonication on Nickel Hydroxide Nanoparticles Synthesis by Electrolysis at High Voltage. Proceedings of Second International Conference on Electrical Systems, Technology and Information 2015 (ICESTI 2015).
- [20] Budipramana Y, Ersam T, Kurniawan F. Synthesis nickel hydroxide by electrolysis at high voltage. *Journal of Engineering & Applied Sciences*. 2014.
- [21] Rafiee B, Fakhari AR. Electrocatalytic oxidation and determination of insulin at nickel oxide nanoparticles-multiwalled carbon nanotube modified screen printed electrode. *Biosensors and Bioelectronics*. 2013; 46: 130–135.
- [22] Amini N, Gholivand MB, Shamsipur M. Electrocatalytic determination of traces of insulin using a novel silica nanoparticles-Nafion modified glassy carbon electrode. *Journal of Electroanalytical Chemistry*. 2014; 714-715: 70–75.
- [23] Kurniawan F, Madurani KA. Electrochemical and optical microscopy study of red pepper seed oil corrosion inhibition by self-assembled monolayers (SAM) on 304 SS. Progress in Organic Coatings. 2015; 88: 256–262.
- [24] Serafín V, Agüí L, Yáñez-Sedeño P, Pingarrón JM. Electrochemical immunosensor for the determination of insulin-like growth factor-1 using electrodes modified with carbon nanotubes– poly(pyrrole propionic acid) hybrids. *Biosensors and Bioelectronics*. 2014; 52: 98–104.
- [25] Sanghavi BJ, Kalambate PK, Karna SP, Srivastava AK. Voltammetric determination of sumatriptan based on a graphene/gold nanoparticles/Nafion composite modified glassy carbon electrode. *Talanta*. 2014; 120: 1–9.