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A Preliminary Study of Smithport Plain Bottle Morphology in the Southern Caddo Area

Robert Z. Selden, Jr.

This study expands upon a previous analysis of the Clarence H. Webb collection, which resulted in the identification of two discrete shapes used in the manufacture of the base and body of Smithport Plain bottles. The sample includes the Smithport Plain bottles from the Webb collection, and four new bottles: two previously repatriated specimens in the Pohler Collection, and two from the Mitchell site (41BW4) to test whether those specimens align morphologically with the Belcher Mound or Smithport Landing specimens. Results indicate significant allometry and a significant difference in Smithport Plain body and base shapes for bottles produced at the Smithport Landing and Belcher Mound sites in northwest Louisiana. The Pohler and Mitchell specimens do not differ significantly from those found at Smithport Landing or Belcher Mound. Analysis of the aggregated sample indicates some significant relationships between bottle shape and size, bottle shape and type, and bottle shape and site, highlighting assemblage-level and type-specific variability. The test of morphological disparity by period indicates a gradual trend toward standardization, and the test of morphological integration indicates that Caddo bottles are significantly integrated, meaning that those discrete traits used to characterize their shape (rim, neck, body, and base) vary in a coordinated manner. The iterative development of this research design can lead to substantive theoretical gains that augment discussions of decorative components and motifs as well as ceramic technological attributes.

Defined as “a vessel with a spheroid or oval body, surmounted by a slender, cylindrical neck,” Caddo bottles were initially seen as a somewhat homogenous ceramic form (Harrington 1920:187); some with shapes and motifs so similar to be deemed the work of a single maker (Harrington 1920:188). In a more recent study, Caddo bottles were found to be more symmetrical than bowls and ollas (Selden, Jr. 2017); however, additional work is needed to identify whether—and to what extent—this holds true across a broader range of vessel shapes and types. Caddo vessel shapes are variable among groups and through time, reflecting stylistic, functional, and social change (Perttula 2010). Caddo potters elevated local ceramic production to high art, and “had no superiors short of the Pueblo country” (Swanton 1942:239), leading some analysts to posit that Caddo bottles rest at the apex of Native American ceramic technology (Gadus 2013). A division of the Caddo bottle category has been proposed for northeast Texas that segregates bottle forms into 27 shapes, each with distinct temporal and spatial distributions (Perttula 2015:Figure 2), and novel deployments of geographic information systems are aiding in the refinement of their probable geographic extents (McKinnon 2011).

This effort capitalizes on the quiddity of Caddo bottle shape for a small sample ($n = 8$) of Smithport Plain bottles previously posited to exhibit morphological differences (Selden Jr. 2018a; Suhm and Jelks 1962; Webb 1959). Three-dimensional (3D) meshes for the Webb Collection and four new samples from one site and one collection were used to test whether a significant difference in shape exists for Smithport Plain bottles by site, followed by a test for allometry. The Smithport Plain bottles were subsequently examined as part of the aggregated sample of Caddo bottles to demonstrate morphological variability, allometry, morphological disparity, and morphological integration among the types (Table 1 and Figure 1).

Taxonomic definitions for Caddo ceramics integrate semiotic and morphological attributes, and each type is characterized by a broad range of vessel shapes that often include bottles, bowls, carinated bowls, ollas, as well as other shapes (Suhm and Jelks 1962; Suhm et al. 1954). The Smithport Plain type was defined by Webb (1959) at the Smithport Landing site (16DS4) in northwest Louisiana and is believed to range in age from the Formative to Early Caddo periods (ca. AD 800–1200) (Webb 1963). All Caddo bottles used in this analysis fall

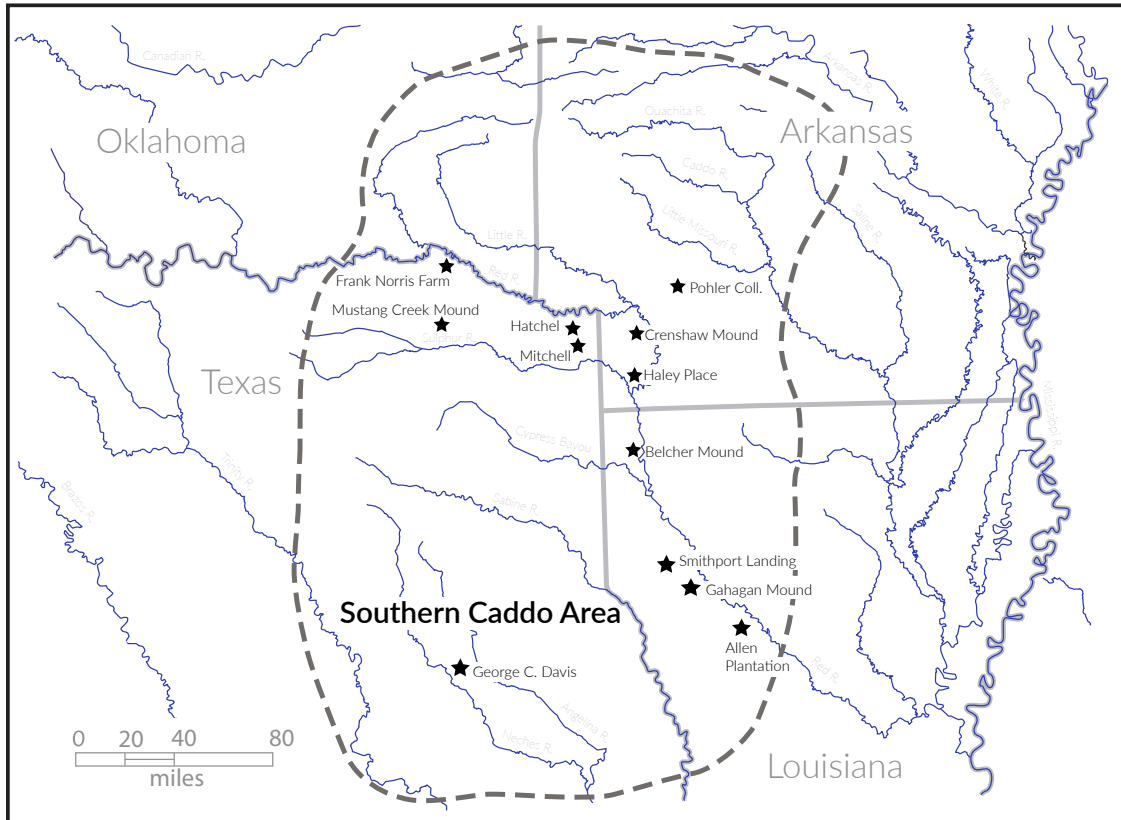


Figure 1. Locations of Allen Plantation, Hatchel, Belcher Mound, Crenshaw Mound, Frank Norris Farm, Gahagan Mound, George C. Davis, Haley Place, Mustang Creek Mound (also known as T. N. Cole), Paul Mitchell (Mitchell), specimens from the Pohler Collection (Clark County, Arkansas), and Smithport Landing.

Table 1. Caddo bottles used in this analysis.

| Specimen | Site Name | Trinomial | Context | Museum | Type |
|----------|---------------|-----------|-----------|--------|------------------|
| 256 | Belcher Mound | 16CD13 | Burial 5 | LSUMNH | Taylor Engraved |
| 267 | Belcher Mound | 16CD13 | Burial 5 | LSEM | Belcher Engraved |
| 269 | Belcher Mound | 16CD13 | Burial 5 | NSU | Belcher Engraved |
| 271 | Belcher Mound | 16CD13 | Burial 5 | LSUMNH | Taylor Engraved |
| 361 | Belcher Mound | 16CD13 | Burial 9 | NSU | Belcher Engraved |
| 363 | Belcher Mound | 16CD13 | Burial 10 | NSU | Belcher Engraved |
| 404 | Belcher Mound | 16CD13 | Burial 11 | NSU | Hickory Engraved |
| 405 | Belcher Mound | 16CD13 | Burial 11 | NSU | Smithport Plain |
| 430 | Belcher Mound | 16CD13 | Burial 12 | NSU | Smithport Plain |
| 775 | Belcher Mound | 16CD13 | Burial 15 | NSU | Belcher Engraved |
| 784 | Belcher Mound | 16CD13 | Burial 15 | LSUMNH | Keno Trailed |
| 787 | Belcher Mound | 16CD13 | Burial 15 | LSUMNH | Taylor Engraved |
| 788 | Belcher Mound | 16CD13 | Burial 15 | NSU | Belcher Engraved |
| 803 | Belcher Mound | 16CD13 | Burial 15 | LSUMNH | Belcher Engraved |
| 805 | Belcher Mound | 16CD13 | Burial 15 | NSU | Belcher Engraved |
| 845 | Belcher Mound | 16CD13 | Burial 17 | NSU | Belcher Engraved |
| 852 | Belcher Mound | 16CD13 | Burial 17 | NSU | Keno Trailed |
| 897 | Belcher Mound | 16CD13 | House 6 | NSU | Belcher Engraved |
| 997 | Belcher Mound | 16CD13 | Burial 24 | NSU | Belcher Engraved |
| 1054 | Belcher Mound | 16CD13 | Burial 26 | LSEM | Taylor Engraved |

Table 1. Caddo bottles used in this analysis. (Continued)

| Specimen | Site Name | Trinomial | Context | Museum | Type |
|-------------|-------------------------------|-------------|--------------|--------|------------------|
| 1073 | Belcher Mound | 16CD13 | House 6 | NSU | Belcher Engraved |
| 95 | Smithport Landing | 16DS4 | Burial 1 | NSU | Smithport Plain |
| 96 | Smithport Landing | 16DS4 | Burial 1 | NSU | Hickory Engraved |
| 152 | Smithport Landing | 16DS4 | Burial 10 | NSU | Smithport Plain |
| No # | Smithport Landing | 16DS4 | Unknown | NSU | Hickory Engraved |
| 142 | Allen Plantation | 16NA6 | Unknown | NSU | Hickory Engraved |
| 955 | Gahagan Mound | 16RR1 | Mound A | NSU | Hickory Engraved |
| 956 | Gahagan Mound | 16RR1 | Mound A | LSUMNH | Hickory Engraved |
| HFE1 | Haley Place | 3MI1 | Unknown | LSEM | Hickory Engraved |
| HFE2 | Haley Place | 3MI1 | Unknown | LSEM | Hickory Engraved |
| HFE3 | Haley Place | 3MI1 | Unknown | LSEM | Hickory Engraved |
| HFE4 | Haley Place | 3MI1 | Unknown | LSEM | Hickory Engraved |
| HFE5 | Haley Place | 3MI1 | Unknown | LSEM | Hickory Engraved |
| 55-1-8* | Crenshaw Mound | 3MI6 | Unknown | CNO | Hickory Engraved |
| 2002-01-18* | Unknown (Clark County, AR) | Pohler Coll | Unknown | CNO | Smithport Plain |
| 2002-01-20* | Unknown (Clark County, AR) | Pohler Coll | Unknown | CNO | Hickory Engraved |
| 2002-01-23* | Unknown (Clark County, AR) | Pohler Coll | Unknown | CNO | Hickory Engraved |
| 2002-01-27* | Unknown (Clark County, AR) | Pohler Coll | Unknown | CNO | Smithport Plain |
| FS7 | Hatchel | 41BW3 | Unknown | TARL | Hickory Engraved |
| 6-2-67 | Paul Mitchell | 41BW4 | | TARL | Smithport Plain |
| 6-2-78 | Paul Mitchell | 41BW4 | | TARL | Smithport Plain |
| 6-2-132 | Paul Mitchell | 41BW4 | Unknown | TARL | Hickory Engraved |
| 341-427 | Paul Mitchell | 41BW4 | Burial 9 | TARL | Hickory Engraved |
| 341-464 | Paul Mitchell | 41BW4 | Burial 21 | TARL | Hickory Engraved |
| 2015-1 | George C. Davis | 41CE19 | Burial F-154 | TARL | Hickory Engraved |
| 7 | Frank Norris Farm | 41RR2 | Unknown | TARL | Hickory Engraved |
| 2 | Mustang Creek Mound | 41RR3 | H. O. #568 | TARL | Hickory Engraved |

The bottle without a number (Webb Collection) is assumed to have come from the Smithport Landing site in fragments. The bottle was later reassembled, but a number was never assigned. * = repatriated to the Caddo Nation of Oklahoma. NSU = Northwestern State University (Williamson Museum); LSUMNS = Louisiana State University Museum of Natural Science; CNO = Caddo Nation of Oklahoma; TARL = Texas Archeological Research Laboratory; LSEM = Louisiana State Exhibit Museum.

under the Native American Graves Protection and Repatriation Act (NAGPRA), excepting those found in House 6 at the Belcher Mound site (see Table 1). The Caddo Nation of Oklahoma granted permission to scan the collections with the provision that any scan data used in the analysis must not include the texture (color) file. Full-resolution scan data were forwarded to the Caddo Nation of Oklahoma with the texture applied. This provides them with an accurate 3D record of each vessel, and a means of viewing a collection of bottles that is curated across numerous repositories.

Geometric Morphometrics in Archeology

Analyses of artifact shape are neither new or novel (Okumura and Araujo 2018), and it is not surprising that geometric morphometrics (GM) (*sensu* Corti (1993)) has captivated analysts of material culture due to the substantive contribution of morphology to lithic (Fox 2015; Thulman 2012; Wilczek et al. 2015) and ceramic typologies (Girulac 2006; Topi et al. 2017; Wilczek et al. 2014), additional categories of material culture (Chitwood 2014; Ros et al. 2014; Windhager et al. 2012), and

novel applications (Barceló 2010; Lenardi and Merwin 2010). The earliest study of artifacts was an analysis of irregular shapes by elliptic Fourier analysis (EFA) (Gero and Mazzullo 1984), and the adoption of the method by the archaeological community has grown to include an impressive array of applications (Figure 2).

EFA has been employed at an increasing rate in lithic and ceramic analyses (Cardillo et al. 2010; Costa 2010; Fox 2015; Ioviță 2009, 2010, 2011; Ioviță and McPherron 2011; Smith et al. 2014; Wilczek et al. 2014), where new approaches advance archeological applications. Creative research designs are also being developed to address challenges with incomplete specimens in the archeological record (Byrne et al. 2016; Rezek et al. 2011; Smith 2010; Smith and DeWitt 2016; Smith and Goebel 2018). These advancements have aided in the development of a useful suite of protocols applicable to wide-ranging research questions.

The recent fluorescence of landmark-based applications has been driven by advances in anthropology (Bookstein et al. 2004; Elewa 2010; Richtsmeier et al. 1992; Slice 2007) and a variety of other research domains (Adams et al. 2004, 2013; Bookstein 1982, 1991, 2016; Jensen 2003; MacLeod 2017; Marcus et al. 1996; Rohlf 1990, 1999; Rohlf and Marcus 1993; Rohlf and Slice 1990; Zelditch et al. 2004) that articulate with the rise of the Procrustes paradigm (Adams et al. 2013:8). Archaeological applications have included two-dimensional (2D) analyses of Clovis technology in North America (Buchanan and Collard 2010; Buchanan et al. 2011; Buchanan et al. 2015; Buchanan et al. 2013; Eren et al. 2015), Fishtail or Fell projectile points in South America (Castiñeira et al. 2011; Loponte et al. 2015), bifacial points from the Umbu Tradition in Brazil (Okumura and Araujo 2013, 2014, 2017), lanceolate points—*ayampitin*—from Argentina (Rivero and Heider 2017), the size and shape of projectile points from southern Patagonia (Charlin et al. 2014; Charlin and González-José 2012), bifacial tools from southern Poland (Serwatka 2015), Final Palaeolithic large tanged points (Serwatka and Riede 2016), Paleoindian point types from Florida (Thulman 2012) and the Southern High Plains (Buchanan et al. 2007), ceramics from Casas Grandes (Topi et al. 2017), flake morphology (Picin et al. 2014), and reduction effects (de Azevedo et al. 2014). All of these studies capitalize on the morphological variation that occurs in a single plane (Buchanan and Collard 2010; Velhagen and Roth 1997).

For research designs that incorporate questions associated with more complex geometry, 3D landmark-based approaches may be more appropriate. Examples from the literature include the development of novel tools and applications (Lycett et al. 2006) that cover a broad range of artifact categories including projectile points (Shott 2011; Shott and Trail 2010), bifaces (Archer and Braun 2010; Archer et al. 2015; Archer et al. 2016), percussive tools (Caruana et al. 2014), flake scars (Sholts et al. 2012), flake tools (Archer et al. 2017), handaxes (Lycett 2009; Lycett et al. 2010; Lycett and von Cramon-Taubadel 2013; Lycett et al. 2016; Wang et al. 2012), and Caddo ceramics (Selden Jr. 2017, 2018a; Selden Jr. et al. 2014). This study adduces the variation that occurs within a single plane (widest vessel profile) for a sample of Caddo bottles; however, 3D data were required to identify the widest profile. Additionally, a variety of landmark and semilandmark configurations are in development that provide for a more robust analysis of 3D morphology associated with specific elements of vessel morphology.

Methods

Bottles were scanned with a Creaform GoSCAN 50 at a 0.8 mm resolution or a Creaform GoSCAN20 at 0.5 mm resolution depending on their size. Scanner calibration was optimized prior to each scan, with positioning targets required for increased accuracy. Shutter speed was reconfigured in each instance; clipping planes were established to reduce the amount of superfluous data collected during each scan. Following data collection, resolution for the GoSCAN 50 meshes was increased to 0.5 mm, and meshes from both scanners were transferred to VX-model where the final mesh was rendered following application of the *clean mesh* function. This was used to remove isolated patches, self-intersections, spikes, small holes, singular vertices, creased edges, narrow triangles, outcropping triangles, narrow bridges, and non-manifold triangles prior to export as an ASCII stl file. The stl functions as a backup, and the ply was subsequently imported to Geomagic Design X (Dx).

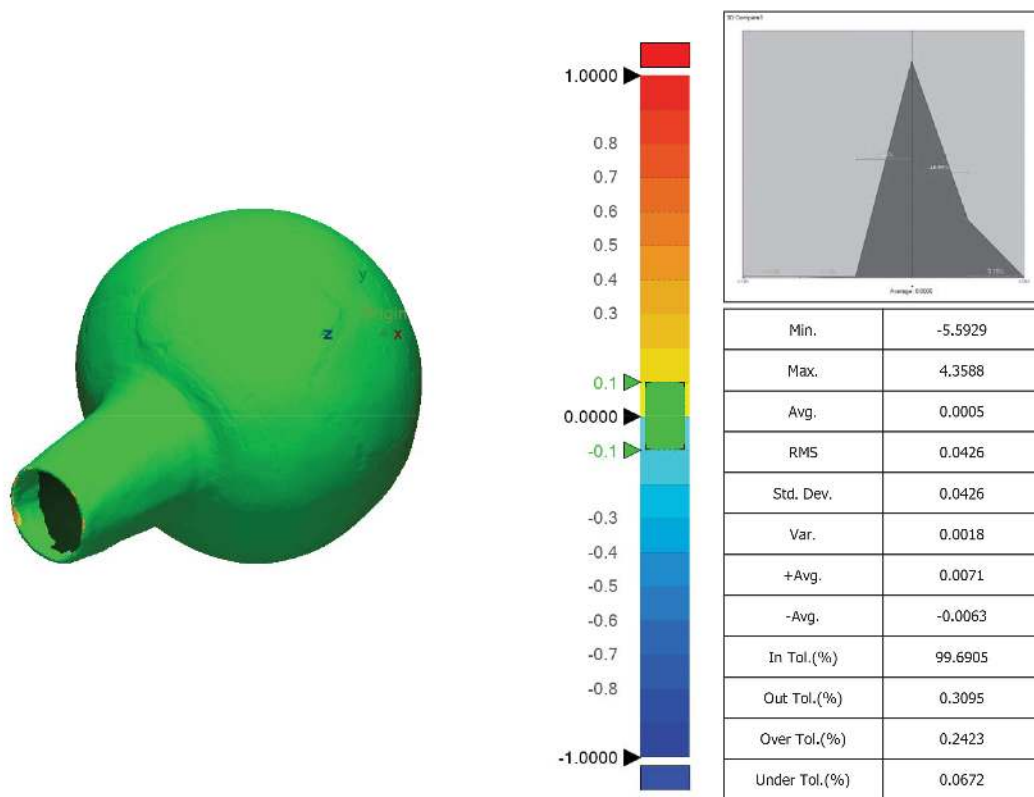
Prior to pursuing the mixed-method analysis employing data from two different scanners, two meshes of the same object—produced with the Creaform GoSCAN 50 and GoSCAN 20—were imported to a computer-aided inspection program (Geomagic Control X) in an effort to identify any

significant deviations that may exist between the meshes prior to the GM analysis (Figure 2). The tolerance level for the inspection was selected using the highest resolution of the GoSCAN 20 (0.1 mm). Small areas of the rim exhibited minor differences while the remainder of the vessel was at or below the arbitrary 0.1 mm tolerance; thus, results fell within an acceptable error range.

The histogram shown in Figure 2 illustrates the Gaussian distribution for the number of errors over the whole deviation. The graph is split into six segments: 1-Sigma at 31 percent from the average to the maximum deviation in each direction, 2-Sigma at 69 percent from the average to the maximum deviation

in each direction, and 3-Sigma at 93.3 percent from the average to the maximum deviation in each direction. The average (AVG) is the sum of all deviations divided by the number of all deviations, and the RMS is the square root of all squared deviations divided by the number of all deviations (sometimes referred to as the effective deviation). In tolerance (In Tol) and out tolerance (Out Tol) percentages indicate the percentage of deviations in or out of a given tolerance, and over tolerance (Over Tol) and under tolerance (Under Tol) percentages indicate the percentage of deviations over (positive direction) or under (negative direction) the tolerance range by the mesh normal of the reference mesh.

Result Data - 1 : 3D Compare1



| | | | | | |
|--------------|---------------|------------|--------|------|--------------|
| Product Name | 41BW4-341-464 | Department | CRHR | Date | Mar 11, 2018 |
| Part Name | 41BW4 341-464 | Inspector | Selden | Unit | mm |

Figure 2. Results of 3D compare for the GoSCAN 50 and GoSCAN 20 meshes of bottle 41BW4 341-464 indicating that 99.6905 percent of the vessel falls within the arbitrary 0.1 mm tolerance.

Alignment and Reference Geometry

Following transfer to Dx, each mesh was subjected to an additional quality check to eliminate non-manifold poly-vertices, folded poly-faces, dangling poly-faces, small clusters, small poly-faces, non-manifold poly-faces, crossing poly-faces, and small tunnels. Due to the paucity of homologous landmarks on cultural artifacts (Lycett 2009), reference geometry was constructed around each vessel in a manner that yielded a replicable configuration of nine landmarks, and 46 equidistant semilandmarks along the widest vessel profile, with notable similarities to previous landmark configurations used by Girrulat (2006:Figure 4), Selden Jr. et al. (2014:Figure 5), Selden, Jr. (2018a:Figure 3), and Topi et al. (2017:Figure 4), all of which largely follow Birkhoff (1933).

The first component of reference geometry added, and the principal assumption, was a reference vector. A sampling ratio of 100 percent was used to apply the reference vector on a revolving axis, after which a reference point was added by projecting it atop the mesh surface at the location where the reference vector exits the base of the vessel. A reference plane was inserted using the *pick multiple points* function, by adding a series of 10 points around the circumference of the bottle's base. Each element of reference geometry (vector, point, and plane) was then used in an interactive 3-2-1 alignment where the vessel was aligned to a global origin, orienting it in 3D space where it sat upright atop a planar surface (assumed to be the intent of the maker). Following alignment, the reference plane and point were deleted.

The widest profile is defined as the location on a mesh that lies farthest from that point where the reference vector exits the vessel base while oriented atop the planar surface. To identify that location, a mesh sketch was generated with the planar method using the plane at the base of the vessel to identify and sketch the widest vessel circumference. By using the plane located at the base of the vessel for the sketch, the point at which the reference vector exits the mesh remains linked to the remainder of the reference geometry. A circle was then sketched using the vector as the center, extending outward until the whole of the vessel fit within. Using the mesh sketch, a cylinder (surface) was extruded around the vessel. The accuracy analyzer in Dx was then used to identify the point on the vessel with the lowest deviation from the extruded surface, and a plane (MPlane) was inserted

coplanar to the vector and oriented to the widest point, bisecting the vessel along the widest profile.

Using the MPlane as the basis for a second mesh sketch, a spline with 15 interpolation points was sketched on one rim. Above that sketch, a horizontal line was added where both the spline and horizontal line determine the horizontal tangent of the rim. A vertical line was subsequently added that bisected the rim at the location of the tangent. This operation was repeated for the opposing rim. The addition of this added step was necessary because surface scanners are unable to collect data from the interior of the bottles, so the spline needed to be cut in a replicable location. Since the Smithport Plain bottles exhibit slightly inverted-to-vertical rims, the preceding step was extended to include an additional measure. A line was drawn between each rim tangent, then a second from the intersection of the line and reference vector to a point 10 mm down the vector, where a horizontal line (parallel with the rim peaks) was inserted to intersect with both external walls of the bottle (Selden, Jr. 2018a:Figure 3). It is at this intersection that the final mesh sketch was cut to discriminate between the neck and rim. While this step admittedly appears odd in the context of a comparison of bottle shapes that all exhibit direct rims, it is of considerable import for inter-type comparisons where other bottle types exhibit differing rim morphologies (i.e., everted, etc.) (Selden, Jr. 2018b).

Using the MPlane as the basis for a third sketch, a spline was populated for the entirety of the silhouetted profile. That spline was split at the location of the horizontal tangent on each rim, and the remaining sections that continued into the bottle interior were deleted. The second split was added at the intersection of the spline and reference vector (center of base). Four additional splits were subsequently added at the juncture of the base/body and body/neck on each side of the vessel at the points of highest curvature. The point of highest curvature used to split the spline was identified using the *curvature* function in Dx, and does not represent an arbitrary location.

Landmarks and Semilandmarks

A total of nine landmarks and 46 semilandmarks segregated each bottle into four discrete components corresponding with the rim, neck, body, and base (Table 2 and Figure 3). Landmarks and semilandmarks were populated along the spline, and

numbering always began on that side of the profile determined to include the widest point. Divisions between each component articulate with those of the spline splits, where landmarks were placed at each point in Table 2, with a series of equidistant semilandmarks between them.

While sliding semilandmarks were an early consideration of this research design, the decision to use equidistant semilandmarks rather than sliding

semilandmarks was based upon results from an earlier iteration of the Webb collection analysis (Selden, Jr. 2018a:Figure 3). In the study of the Webb collection, the first landmark and sliding semilandmark configuration did not split the spline between the neck and rim, and when mean shapes were generated for each type, an anomaly, from the everted rims of Belcher Engraved bottles in that case, was added to the otherwise direct or tapered necks of the Hickory

Table 2. Landmarks used in this analysis.

| Landmark | Location | Definition |
|----------|------------|---|
| Point01 | Rim peak | Horizontal tangent of rim curvature on widest side of vessel |
| Point06 | Rim/Neck | Point of highest curvature (everted rim) or intersection of horizontal line 10 mm below rim tangents (direct rim) |
| Point15 | Neck/Body | Point of highest curvature |
| Point24 | Body/Base | Point of highest curvature |
| Point28 | CenterBase | Intersection of vector and external surface of the 3D mesh |
| Point32 | Body/Base | Point of highest curvature |
| Point41 | Neck/Body | Point of highest curvature |
| Point50 | Rim/Neck | Point of highest curvature (everted rim) or intersection of horizontal line 10 mm below rim tangents (direct rim) |
| Point55 | Rim peak | Horizontal tangent of rim curvature |

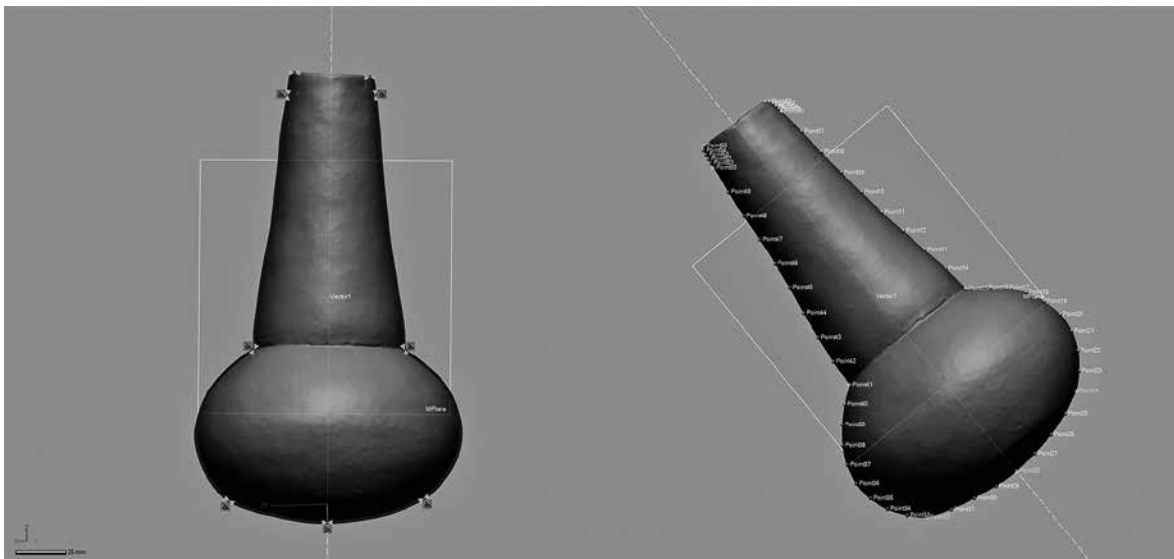


Figure 3. Spline splits for discrete components (rim, neck, body, and base) used in the GM analysis (left) segregated by landmarks (blue), with equidistant semilandmarks (white) populated between (right).

Engraved and Smithport Plain bottles. Given that the use of sliding semilandmarks could potentially influence the results of this analysis by introducing a morphological attribute to specimens where one does not exist, they were abandoned.

Analysis

Landmarks and equidistant semilandmarks were exported as x, y, and z coordinate data from Dx. Those data were aligned to a global coordinate system (Kendall 1981, 1984; Slice 2001), achieved through generalized Procrustes superimposition (Rohlf and Slice 1990) performed in R 3.5.0 (R Development Core Team 2018) using the *geomorph* library v.3.0.6 (Adams et al. 2017; Adams and Otárola-Castillo 2013). Procrustes superimposition translates, scales, and rotates the coordinate data to allow for comparisons among objects (Gower 1975; Rohlf and Slice 1990). The *geomorph* package uses a partial Procrustes superimposition that projects the aligned specimens into tangent space subsequent to alignment in preparation for the use of multivariate methods that assume linear space (Rohlf 1999; Slice 2001).

Principal components analysis (Jolliffe 2002) was used as an exploratory means of visualizing shape variation among the bottles. The shape changes described by each principal axis are commonly visualized using thin-plate spline warping of a reference 3D mesh (Klingenberg 2013; Sherratt et al. 2014). A residual randomization permutation procedure (RRPP; $n=1000$ permutations) was used for all Procrustes ANOVAs (Adams and Collyer 2015; Collyer and Adams 2018), which has higher statistical power and a greater ability to identify patterns in the data should they be present (Anderson and Ter Braak 2003). To assess whether shape differs by size (allometry) and site, Procrustes ANOVAs (Goodall 1991) were run that also enlist effect-sizes (z-scores) computed as standard deviates of the generated sampling distributions (Collyer et al. 2015). For the aggregated sample, a Procrustes ANOVA was run to assess whether shape changes with size, and the assumption of allometric slope homogeneity was tested with the *procD.allometry* function using the PredLine option (Adams and Nistri 2010). Should this test not be significant, then allometric slopes are similar—if not identical—across time and types.

A Procrustes ANOVA and pairwise test was used to identify sites where bottle shapes and types differ.

The pairwise test is conceptually similar to trajectory analysis (Adams and Collyer 2007, 2009; Collyer and Adams 2007, 2013) in that pairwise statistics are vector lengths between vectors, but differs in that a factorial model is not explicitly needed to contrast vectors between point factor levels nested within group factor levels (Adams et al. 2017). Procrustes variance was used to discriminate between groups and to compare the amount of shape variation (morphological disparity) across communities (Zelditch et al. 2004), which is estimated as the Procrustes variance using residuals of linear model fit (Adams et al. 2017).

Morphological integration was assessed for the aggregated sample of whole vessels. Integration between pairs of traits was tested using a two-block partial least-squares (2B-PLS) analysis to evaluate relationships for two blocks of variables collected from the same specimens (Bookstein et al. 2003; Rohlf and Corti 2000; Wold 1966), using shape coordinates in all blocks of variables (Bastir and Rosas 2006; Bookstein et al. 2003; Gunz and Harvati 2007). To assess whether the different modules (RIM_{neck} , $NECK_{body}$, and $BODY_{base}$ in particular) are integrated, a two-sample test using effect sizes calculated as standard deviates in sampling distributions from the 2B-PLS analyses were used to determine the significance and strength of integration between the modules (Adams and Collyer 2016).

Results

The mean consensus configuration and Procrustes residuals were calculated using a generalized Procrustes analysis (GPA) (Figure 4). This initial view of the data demonstrates the degree of variability in Caddo bottles that occurs across the sample. As an exploratory measure, GM methods—to include GPA—aid in clarifying shape differences, and in the production of novel *a posteriori* hypotheses (Mitteroecker and Gunz 2009).

Principal components analysis (PCA) was conducted on scaled, translated, and rotated landmarks and semilandmarks, and demonstrates that the first two PC's account for 68 (PC1) and 27 (PC2) percent of the variation in bottle shape (Table 3 and Figure 6). Together, PC1 and PC2 account for 95 percent of shape variation, with all remaining PCs representing three or fewer percent of the variation (see Table 3). The first two PCs are plotted in Figure 5, where warp grids represent the shape changes along PC1 and PC2. This plot indicates that shape

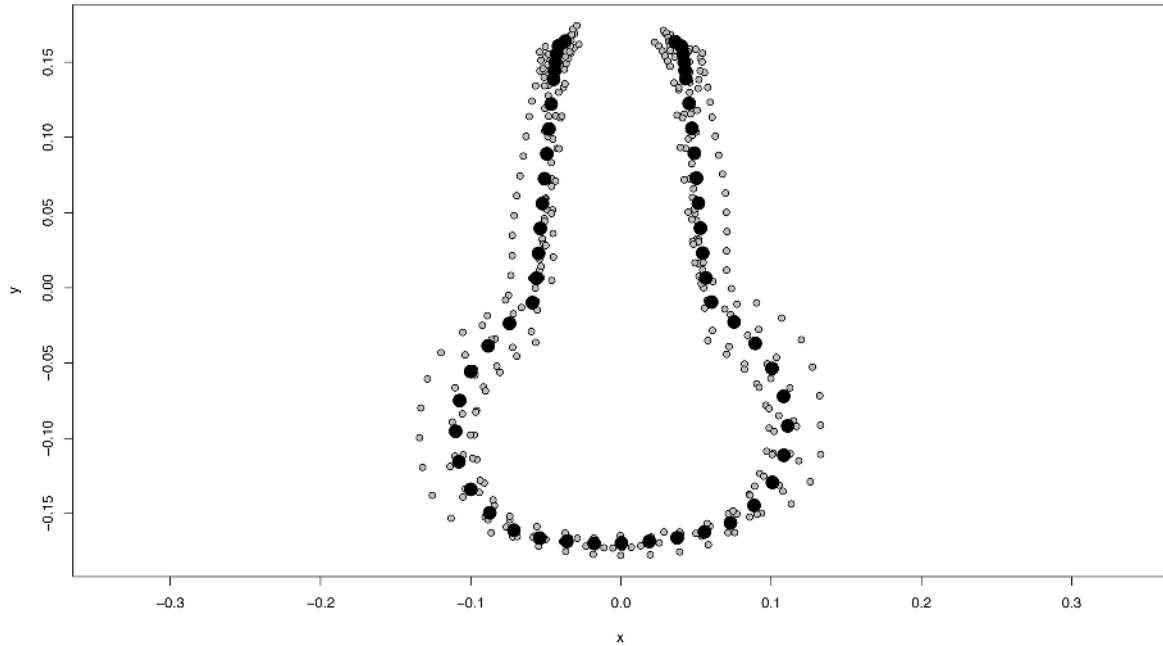


Figure 4. Results of generalized Procrustes analysis for Smithport Plain whole bottles. Mean consensus configuration shown in black; samples in gray.

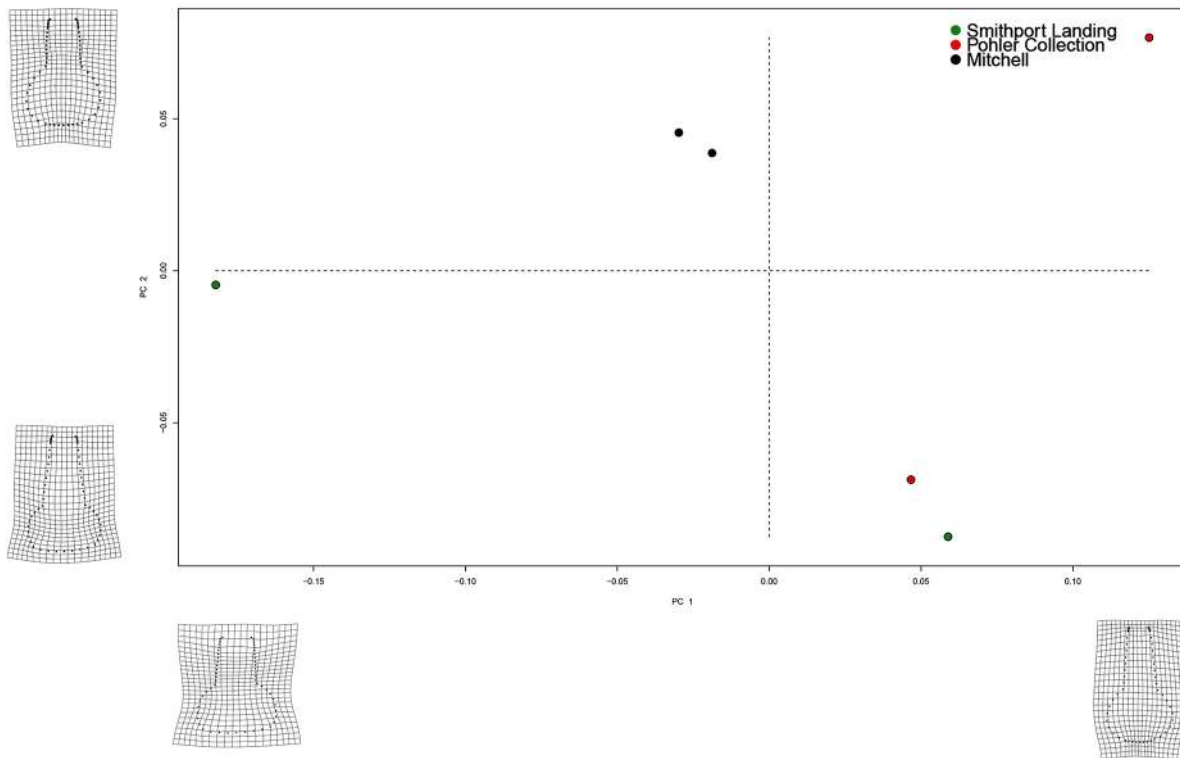


Figure 5. Results of PCA summarizing shape variation in the whole bottle sample.

Table 3. Results of PCA.

| | SD | PV | CP |
|-----|-------|-------|-------|
| PC1 | 0.106 | 0.683 | 0.683 |
| PC2 | 0.066 | 0.268 | 0.951 |
| PC3 | 0.023 | 0.033 | 0.985 |
| PC4 | 0.014 | 0.013 | 0.997 |
| PC5 | 0.007 | 0.003 | 1.000 |

SD = standard deviation; PV = proportion of variance; CVE = cumulative proportion.

changes associated with PC1 articulate most readily with base and body shape, and shape changes associated with PC2 articulate with base, body, and neck shape.

A Procrustes ANOVA was used to test for significant allometry. Results of the ANOVA indicate significant allometry in the sample (RRPP = 1000, $Rsq = 0.59427$, $Pr(>F) = 0.0075$), indicating that Smithport Plain bottle shapes change with size. A Procrustes ANOVA was used to test for a significant difference in bottle shape by site, and results indicate that there is not a significant difference in bottle shape by site (RRPP = 1000, $Rsq = 0.40907$, $Pr(>F) = 0.537$).

Bottle base and body morphology

Two Smithport Plain bottles, specimen numbers 405 and 430, from Burials 11 and 12 at the Belcher Mound site are missing the upper portions of the neck and rim, and therefore could not be included in the analysis of whole vessels. Using a subset of the same constellation of landmarks and equidistant semilandmarks from the analysis of whole vessels (landmarks/semilandmarks 15-41), these two samples were added for an analysis of bottle base and body morphology. The mean consensus configuration and Procrustes residuals were calculated using a GPA for the base and body sample (Figure 6).

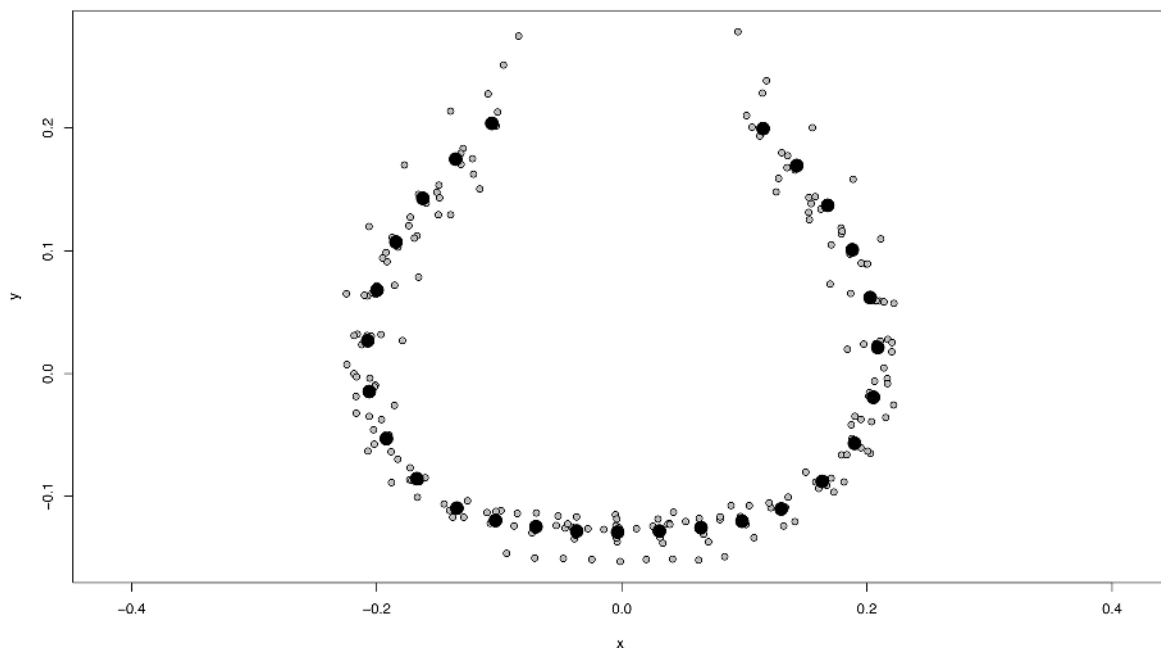


Figure 6. Results of generalized Procrustes analysis for Smithport Plain base and body sample. Mean consensus configuration shown in black; samples in gray.

PCA was conducted on scaled, translated, and rotated landmarks and semilandmarks, and demonstrates that the first two PCs account for 83 (PC1) and 15 (PC2) percent of the variation in bottle base and body shape (Table 4 and Figure 8). Together, PC1 and PC2 account for 98 percent of the variation for base and body shape, with each remaining PC representing less than two percent of the variation (see Table 4). The first two PCs are plotted in Figure 7, where warp grids represent the shape

changes along PC1 and PC2. The plot indicates that shape changes associated with PC1 articulate most readily with a tall, pear-shaped body and narrow base at the maximum, and a shorter, globular body and wide base at the minimum. For PC2, the maximum values articulate with a pear-shaped body that is widest near a broad base, and a shorter, globular body with a narrow and rounded base at the minimum.

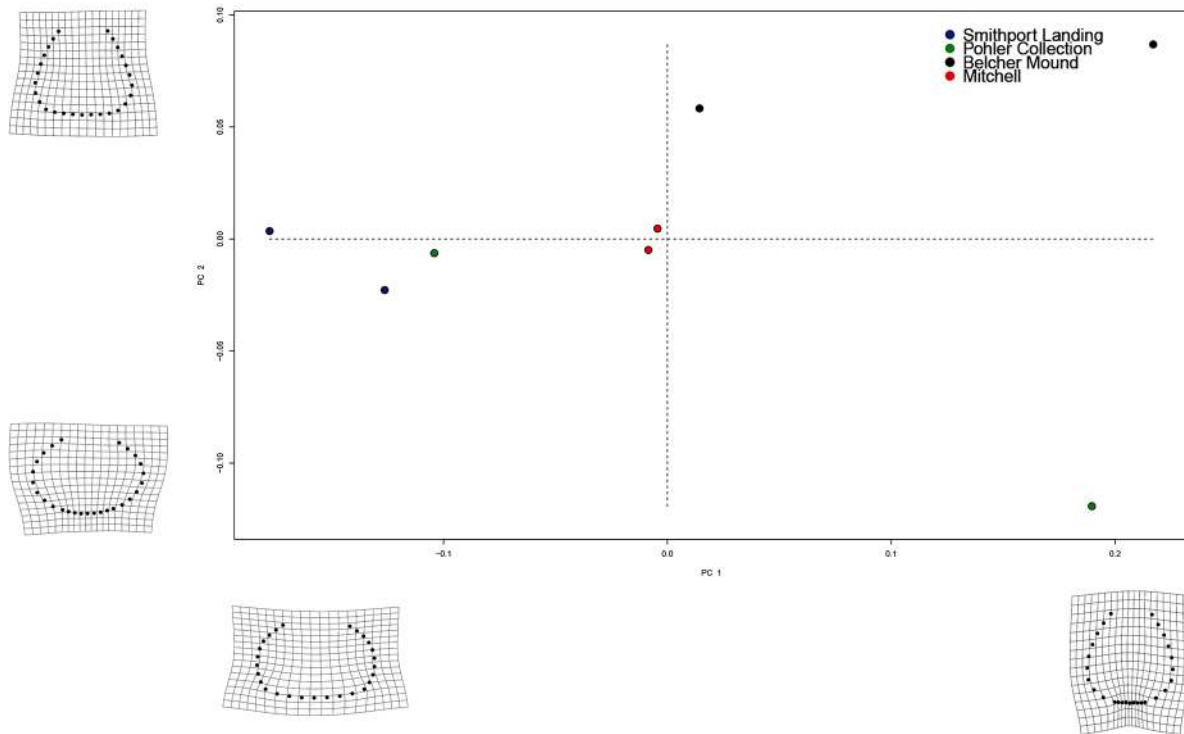


Figure 7. Results of PCA summarizing shape variation in the base and body sample.

Table 4. Results of PCA.

| | SD | PV | CP |
|-----|-------|-------|-------|
| PC1 | 0.142 | 0.826 | 0.826 |
| PC2 | 0.061 | 0.150 | 0.976 |
| PC3 | 0.020 | 0.017 | 0.993 |
| PC4 | 0.009 | 0.003 | 0.996 |
| PC5 | 0.006 | 0.002 | 0.998 |
| PC6 | 0.005 | 0.001 | 0.999 |
| PC7 | 0.004 | 0.001 | 1.000 |

SD = standard deviation; PV = proportion of variance; CVE = cumulative proportion.

A Procrustes ANOVA was used to test for significant allometry. Results of the ANOVA indicate that allometry is not significant in the base and body sample (RRPP = 1000, Rsq = 0.17667, Pr(>F) = 0.275). A second Procrustes ANOVA was used to test for a significant difference in bottle shape by site. The advanced Procrustes ANOVA and pairwise test demonstrates a significant difference between Smithport Plain bottles from the Smithport Landing and Belcher Mound sites (Table 5).

Synthesis with aggregated sample

The Smithport Plain bottles were then added to the aggregated sample (see Table 1), omitting the two previously mentioned incomplete specimens (405 and 430). The aggregated sample of whole vessels (n=45) consists of five Caddo bottle types from 12 sites curated across five repositories in three states, with iterative analytical improvements achieved as new samples are added. The mean consensus configuration and Procrustes residuals were calculated using a GPA for the aggregated sample (Figure 8).

PCA was conducted on scaled, translated, and rotated landmarks and semilandmarks, and demonstrates that the first two PCs account for 59 (PC1) and 20 (PC2) percent of the variation in bottle shape (Table 6 and Figure 9). Together, PC1 and PC2 account for 79 percent of the variation in

bottle shape, with each remaining PC representing ≤ 10 percent of the variation. The first two PCs are plotted in Figure 9, where warp grids represent the shape changes along PC1 and PC2. The plot indicates that shape changes associated with PC1 articulate with relative differences in base, body, neck, and rim shapes. Differences include a sharp or diffuse angle at the base and body juncture, flat versus slightly convex geometry, and a difference in relative width. Body differences range from globular to sub-globular, necks from everted to tapering, and rims from everted to slightly inverted. Shape changes associated with PC2 articulate with differences in relative base width, and a slightly carinated to globular body shape. In addition to a difference in relative width, the bottle necks also range from direct to slightly tapering, with rims that are vertical to slightly everted.

A Procrustes ANOVA was used to test for allometry, and significant allometry was identified in this sample (RRPP = 1000, Rsq = 0.18337, Pr(>F) = 0.001). Plots of predicted allometric trajectories for period (Formative-Early and Late-Historic Caddo) and type factors are presented in Figure 10. The null hypothesis of parallel slopes is rejected by the homogeneity of slopes test for group allometries, as the allometric trajectories differ significantly by period (RRPP = 1000, Rsq = 0.03711, Pr(>F) = 0.010). Allometric trajectories also differ significantly by type (RRPP = 1000, Rsq = 0.10182,

Table 5. Results of advanced Procrustes ANOVA and pairwise test (RRPP = 1000) of Smithport Plain bottle shape by site.

| | Belcher Mound | Mitchell | Pohler Coll | Smithport Landing |
|-------------------|--------------------------------|--------------------------|-------------------------|-------------------|
| Belcher Mound | 0 0 1.000 | | | |
| Mitchell | 0.143 0.040 0.431 | 0 0 1.000 | | |
| Pohler Coll | 0.155 0.172 0.362 | 0.086 -0.816 0.785 | 0 0 1.000 | |
| Smithport Landing | 0.280 2.010 0.032 | 0.147 0.105 0.393 | 0.203 0.945 0.183 | 0 0 1.000 |

Least-squares means distance matrix (top), effect sizes (middle), and P-values (bottom--significant results in bold).

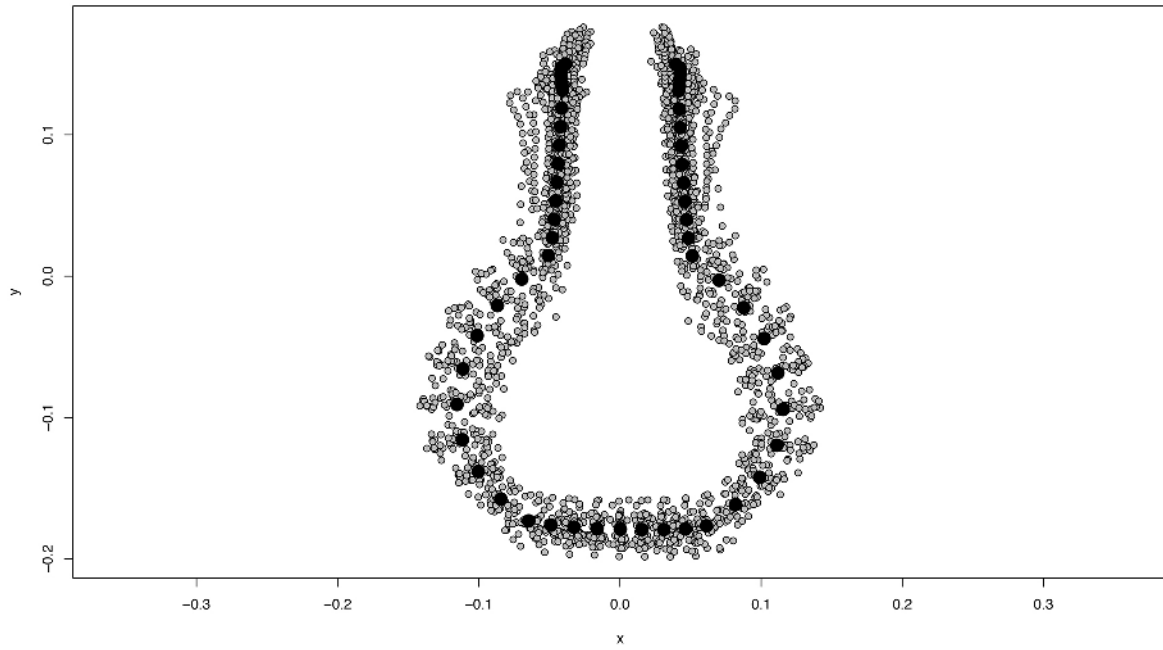


Figure 8. Results of generalized Procrustes analysis for the aggregated sample. Mean consensus configuration shown in black; samples in gray.

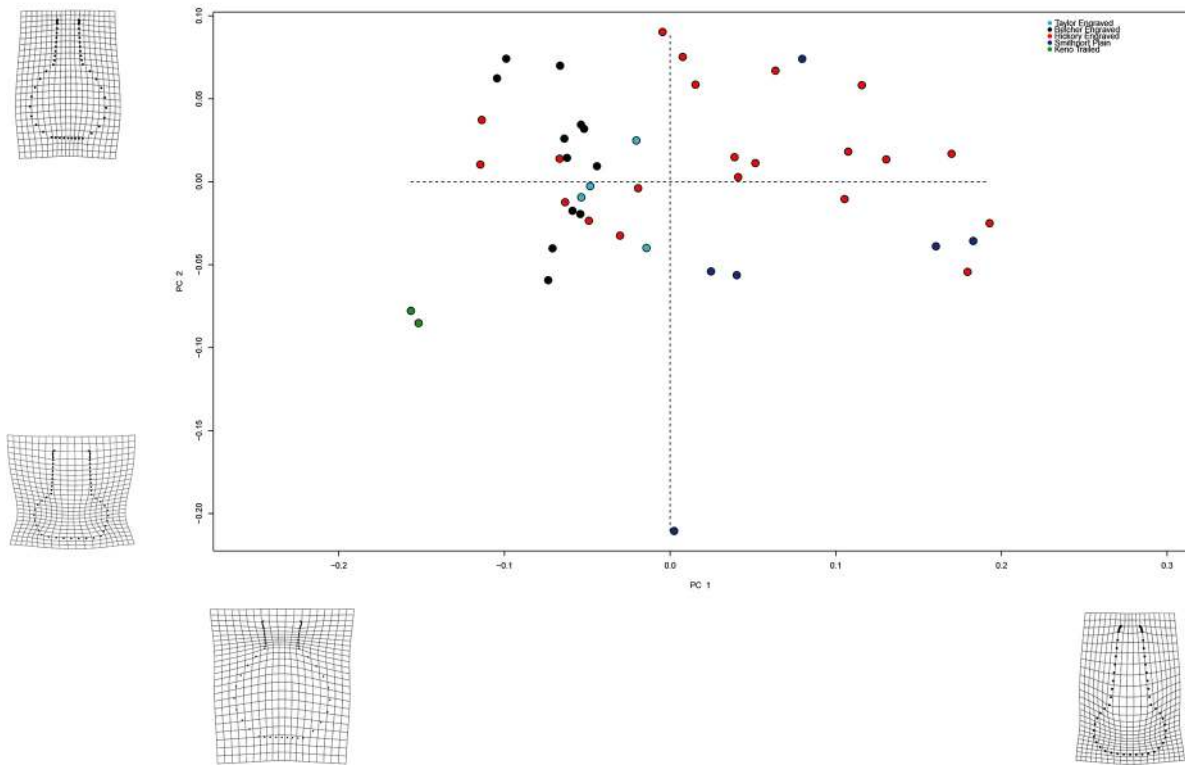


Figure 9. Results of PCA summarizing shape variation in the aggregated sample.

Table 6. Results of PCA.

| | SD | PV | CP |
|------|-------|-------|-------|
| PC1 | 0.093 | 0.592 | 0.592 |
| PC2 | 0.055 | 0.205 | 0.797 |
| PC3 | 0.038 | 0.100 | 0.897 |
| PC4 | 0.026 | 0.045 | 0.942 |
| PC5 | 0.018 | 0.022 | 0.964 |
| PC6 | 0.013 | 0.011 | 0.975 |
| PC7 | 0.010 | 0.007 | 0.982 |
| PC8 | 0.008 | 0.004 | 0.986 |
| PC9 | 0.007 | 0.004 | 0.990 |
| PC10 | 0.006 | 0.003 | 0.993 |

SD = standard deviation; PV = proportion of variance; CVE = cumulative proportion.

Pr(>F) = 0.001) (see Figure 10), and the size of Formative-Early bottles, specifically those of the Hickory Engraved type, extends beyond the range of the Late-Historic and Smithport Plain types.

A second Procrustes ANOVA was used to test for a difference in bottle shape by site, which is (RRPP = 1000, Rsq = 0.52213, Pr(>F) = 0.001). An advanced Procrustes ANOVA and pairwise comparison was used to identify those sites where bottle assemblages differ, and whether that difference is in magnitude, direction, or both (Table 7). Those sites with bottle samples found to differ significantly include Belcher Mound compared to Gahagan Mound, Smithport Landing, Haley Place, Mitchell, and the Pohler Collection. In addition, Smithport Landing also differs significantly compared to Haley Place, and Mitchell.

A third Procrustes ANOVA was used to test for a difference in bottle shape by type, which is significant (RRPP = 1000, Rsq = 0.3907, Pr(>F) = 0.001). An advanced Procrustes ANOVA and pairwise comparison was used to identify which bottle types differ and whether that difference is in magnitude, direction, or both (Table 8). Those bottle types found to differ significantly include Keno Trailed compared to Belcher Engraved, Hickory Engraved, Smithport Plain, and Taylor Engraved; also Belcher Engraved compared to Hickory Engraved, and Smithport Plain; and finally, Smithport Plain compared to Taylor Engraved.

A test of morphological disparity indicates that Hickory Engraved and Smithport Plain bottles display a greater range of shape variation among individual bottles relative to other groups, and differ significantly from the Belcher Engraved, Keno Trailed, and Taylor Engraved bottles (Table 9). This indicates that the Formative-Early Caddo bottles may encompass a greater range of morphological variability than the Late-Historic Caddo bottles in the sample; an assertion that was later confirmed in a subsequent test of morphological disparity by period (Table 10).

The 2B-PLS analyses, each enlisting 1000 random permutations, was used to test for morphological integration between combinations of bottle components (rim, neck, body, and base). The results indicate significant integration between the rim and neck ($r_{\text{PLS}} = 0.969$, P-value = 0.001), rim and body ($r_{\text{PLS}} = 0.942$, P-value = 0.001), rim and base ($r_{\text{PLS}} = 0.663$, P-value = 0.001), neck and body ($r_{\text{PLS}} = 0.962$, P-value = 0.001), neck and base ($r_{\text{PLS}} = 0.869$, P-value = 0.001), and the body and base ($r_{\text{PLS}} = 0.859$, P-value = 0.001) for bottles in the sample (Figure 11). A pairwise test of morphological integration was used to identify combinations of traits that covary. Results indicate that for this sample of Caddo bottles, the RIM_{neck} and RIM_{base} , RIM_{body} and RIM_{base} , and RIM_{base} and $\text{NECK}_{\text{body}}$ are significantly integrated (Table 11).

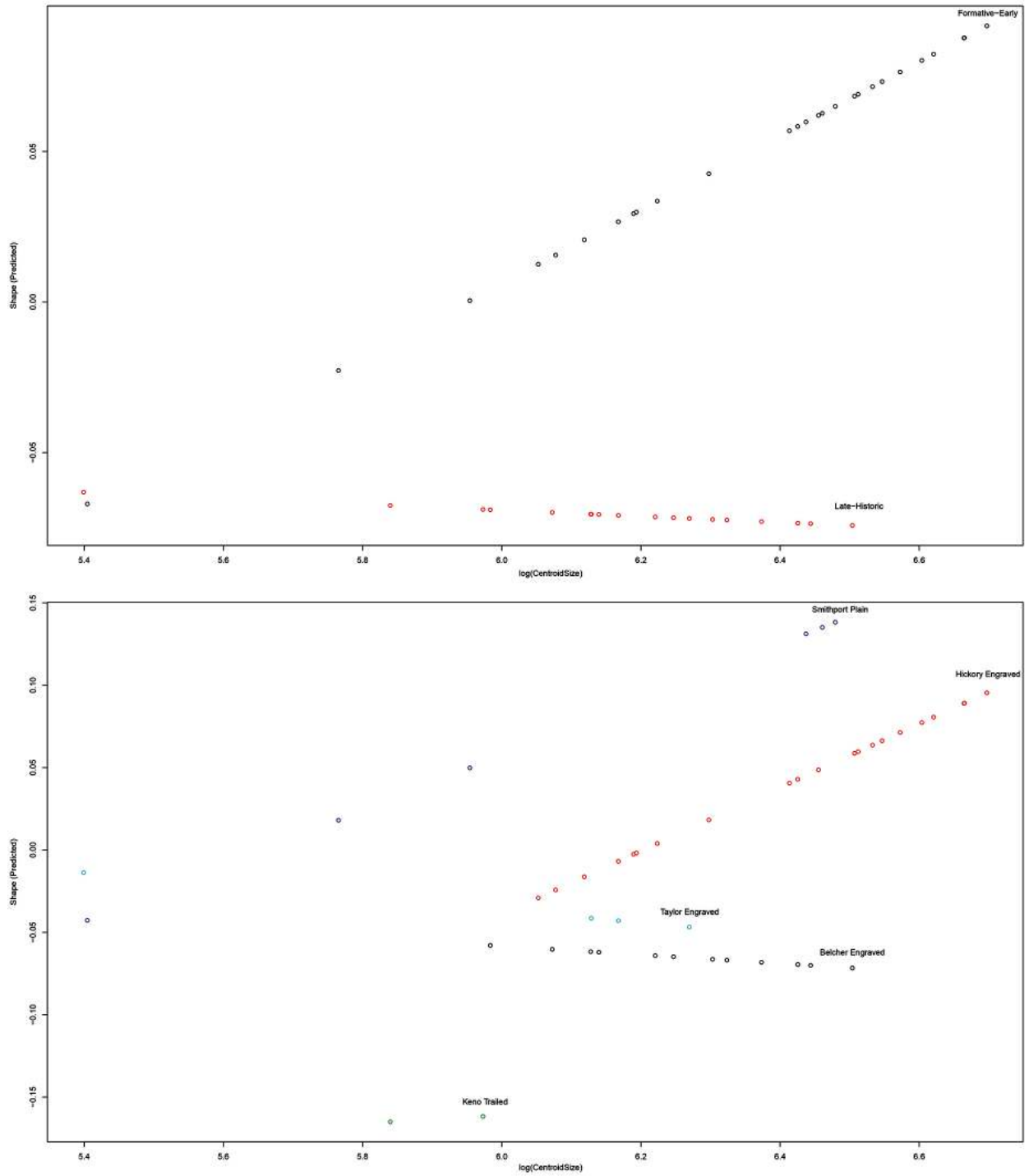


Figure 10. Predicted values of Caddo bottle shape from temporal (top) and type (bottom) regressions versus log(CentroidSize).

Table 7. Least-squares mean distance matrix (top), effect sizes (middle), and P-values (bottom) for advanced Procrustes ANOVA and pairwise test (RRPP = 1000) of bottle shape by site.

| | Allen | Belcher | Crenshaw | FNF | Gahagan | GCD | Haley | Hatchel | MCM | Mitchell | Pohler | Smithport |
|-----------|--------|--------------|----------|--------|--------------|--------|--------------|---------|--------|--------------|--------|-----------|
| Allen | 0 | | | | | | | | | | | |
| | 0 | | | | | | | | | | | |
| | 1.000 | | | | | | | | | | | |
| Belcher | 0.181 | 0 | | | | | | | | | | |
| | 1.478 | 0 | | | | | | | | | | |
| | 0.118 | 1.000 | | | | | | | | | | |
| Crenshaw | 0.161 | 0.132 | 0 | | | | | | | | | |
| | 0.059 | 0.333 | 0 | | | | | | | | | |
| | 0.433 | 0.319 | 1.000 | | | | | | | | | |
| FNF | 0.070 | 0.204 | 0.147 | 0 | | | | | | | | |
| | -1.289 | 1.870 | -0.117 | 0 | | | | | | | | |
| | 0.930 | 0.075 | 0.488 | 1.000 | | | | | | | | |
| Gahagan | 0.096 | 0.251 | 0.218 | 0.092 | 0 | | | | | | | |
| | -0.672 | 4.902 | 1.382 | -0.754 | 0 | | | | | | | |
| | 0.706 | 0.001 | 0.101 | 0.756 | 1.000 | | | | | | | |
| GCD | 0.150 | 0.127 | 0.057 | 0.143 | 0.203 | 0 | | | | | | |
| | -0.060 | 0.278 | -1.470 | -0.197 | 1.211 | 0 | | | | | | |
| | 0.455 | 0.328 | 0.954 | 0.513 | 0.138 | 1.000 | | | | | | |
| Haley | 0.134 | 0.092 | 0.066 | 0.134 | 0.202 | 0.068 | 0 | | | | | |
| | 0.313 | 1.598 | -1.095 | 0.190 | 2.739 | -1.072 | 0 | | | | | |
| | 0.303 | 0.078 | 0.894 | 0.337 | 0.013 | 0.899 | 1.000 | | | | | |
| Hatchel | 0.117 | 0.202 | 0.121 | 0.067 | 0.129 | 0.121 | 0.127 | 0 | | | | |
| | -0.531 | 1.830 | -0.507 | -1.307 | -0.081 | -0.496 | 0.106 | 0 | | | | |
| | 0.652 | 0.069 | 0.648 | 0.915 | 0.457 | 0.632 | 0.378 | 1.000 | | | | |
| MCM | 0.116 | 0.139 | 0.151 | 0.135 | 0.142 | 0.122 | 0.124 | 0.140 | 0 | | | |
| | -0.593 | 0.509 | -0.124 | -0.321 | 0.110 | -0.521 | 0.033 | -0.246 | 0 | | | |
| | 0.693 | 0.279 | 0.511 | 0.559 | 0.400 | 0.665 | 0.382 | 0.546 | 1.000 | | | |
| Mitchell | 0.131 | 0.064 | 0.096 | 0.146 | 0.201 | 0.087 | 0.040 | 0.148 | 0.107 | 0 | | |
| | 0.188 | 0.376 | -0.503 | 0.473 | 2.942 | -0.630 | -1.029 | 0.527 | -0.273 | 0 | | |
| | 0.368 | 0.308 | 0.624 | 0.268 | 0.007 | 0.697 | 0.849 | 0.264 | 0.531 | 1.000 | | |
| Pohler | 0.115 | 0.092 | 0.088 | 0.124 | 0.175 | 0.070 | 0.048 | 0.124 | 0.079 | 0.040 | 0 | |
| | -0.196 | 1.178 | -0.722 | -0.034 | 1.969 | -1.020 | -0.874 | -0.034 | -0.866 | -1.186 | 0 | |
| | 0.493 | 0.124 | 0.742 | 0.433 | 0.046 | 0.869 | 0.813 | 0.440 | 0.809 | 0.924 | 1.000 | |
| Smithport | 0.075 | 0.200 | 0.195 | 0.094 | 0.105 | 0.177 | 0.153 | 0.144 | 0.133 | 0.147 | 0.136 | 0 |
| | -0.942 | 5.374 | 1.298 | -0.634 | 0.241 | 1.000 | 2.435 | 0.348 | 0.160 | 2.315 | 1.668 | 0 |
| | 0.849 | 0.001 | 0.138 | 0.683 | 0.354 | 0.163 | 0.026 | 0.309 | 0.370 | 0.027 | 0.072 | 1.000 |

Significant results in bold; FNF = Frank Norris Farm, GCD = George C. Davis, MCM = Mustang Creek Mound.

Table 8. Least-squares mean distance matrix (top), effect sizes (middle), and P-values (bottom) for advanced Procrustes ANOVA and pairwise test (RRPP = 1000) of bottle shape by type

| | Belcher Eng | HE | Keno Tr | Smithport Pl | Taylor Eng |
|------------------|--------------------------------|--------------------------------|--------------------------------|--------------------------------|-----------------|
| Belcher Engraved | 0 0 1.000 | | | | |
| Hickory Engraved | 0.114 4.499 0.002 | 0 0 1.000 | | | |
| Keno Trailed | 0.171 2.449 0.023 | 0.226 4.131 0.001 | 0 0 1.000 | | |
| Smithport Plain | 0.168 4.812 0.002 | 0.085 1.589 0.071 | 0.251 4.133 0.001 | 0 0 1.000 | |
| Taylor Engraved | 0.046 -0.738 0.745 | 0.085 0.932 0.184 | 0.176 1.931 0.049 | 0.130 2.021 0.034 | 0 0 1.000 |

Significant results in bold.

Table 9. Pairwise absolute differences between variances (top) and P-values (bottom) for the test of morphological disparity by type.

| | Belcher Eng | HE | Keno Tr | Smithport Pl | Taylor Eng |
|------------------|-----------------------|----------------|----------------|----------------|------------|
| Belcher Engraved | 0 1.000 | | | | |
| Hickory Engraved | 0.009 0.004 | 0 1.000 | | | |
| Keno Trailed | 0.001 0.906 | 0.010 0.082 | 0 1.000 | | |
| Smithport Plain | 0.011 0.015 | 0.002 0.698 | 0.012 0.093 | 0 1.000 | |
| Taylor Engraved | 0.001 0.827 | 0.008 0.088 | 0.002 0.760 | 0.009 0.080 | 0 1.000 |

Significant results in bold, RRPP = 1000.

Table 10. Pairwise absolute differences between variances (top) and P-values (bottom) for the test of morphological disparity by time period.

| | Formative-Early | Late-Historic |
|-----------------|-----------------------|---------------|
| Formative-Early | 0 1.000 | |
| Late-Historic | 0.007 0.007 | 0 1.000 |

Significant results in bold, RRPP = 1000.

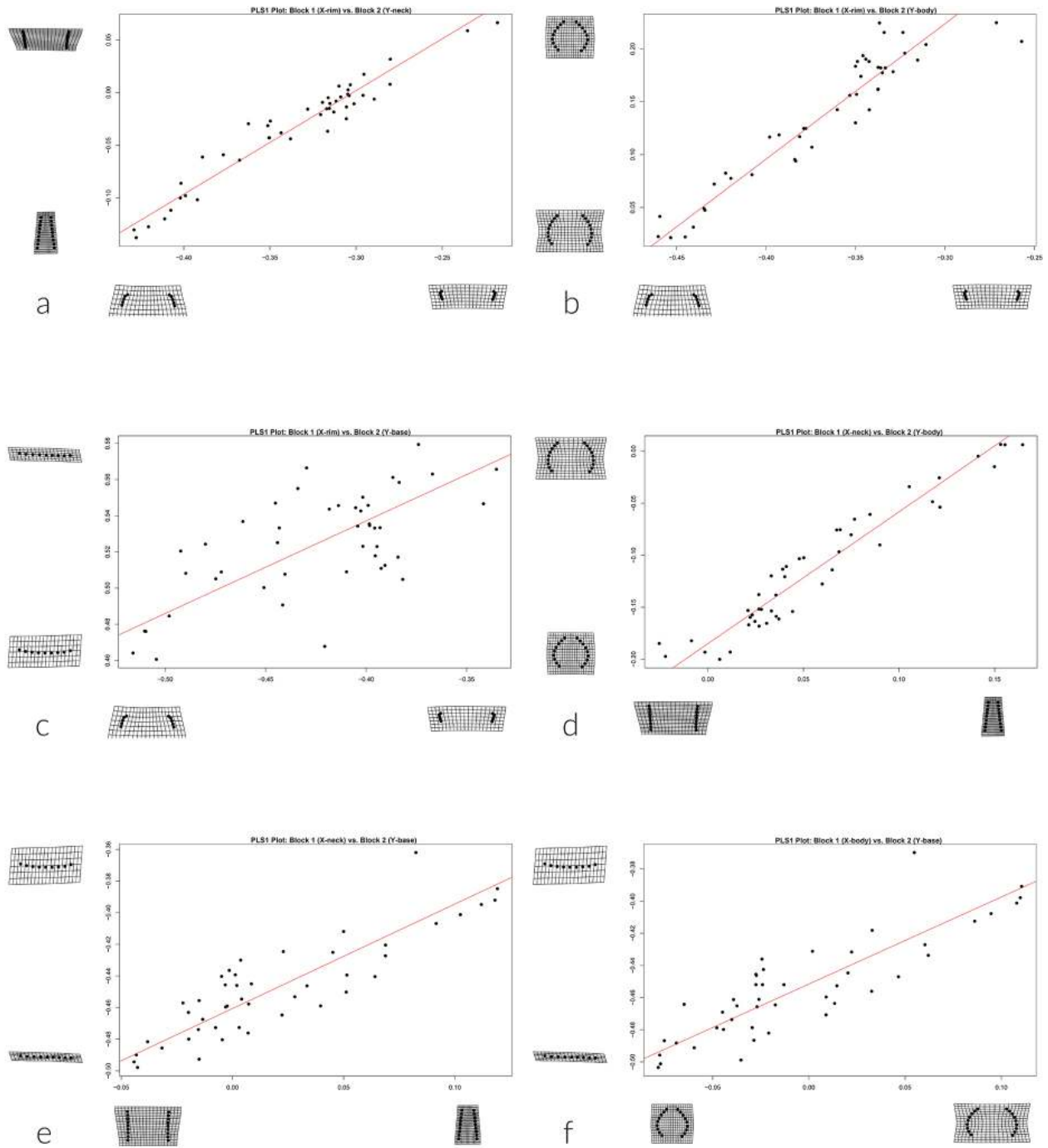


Figure 11. Results of 2B-PLS analyses for pairs of morphological components; (a) rim and neck, (b) rim and body, (c) rim and base, (d) neck and body, (e) neck and base, and (f) body and base.

Table 11. Comparison of morphological integration between modules.

| Z | RIM _{neck} | RIM _{body} | RIM _{base} | NECK _{body} | NECK _{base} | BODY _{base} | P | RIM _{neck} | RIM _{body} | RIM _{base} | NECK _{body} | NECK _{base} | BODY _{base} |
|----------------------|---------------------|---------------------|---------------------|----------------------|----------------------|----------------------|----------------------|---------------------|---------------------|---------------------|----------------------|----------------------|----------------------|
| RIM _{neck} | 0 | | | | | | RIM _{neck} | 1.000 | | | | | |
| RIM _{body} | 0.451 | 0 | | | | | RIM _{body} | 0.326 | 1.000 | | | | |
| RIM _{base} | 2.727 | 2.401 | 0 | | | | RIM _{base} | 0.003 | 0.008 | 1.000 | | | |
| NECK _{body} | 0.493 | 0.048 | 2.331 | 0 | | | NECK _{body} | 0.311 | 0.481 | 0.010 | 1.000 | | |
| NECK _{base} | 1.120 | 0.726 | 1.566 | 0.674 | 0 | | NECK _{base} | 0.131 | 0.234 | 0.059 | 0.250 | 1.000 | |
| BODY _{base} | 1.424 | 1.030 | 1.362 | 0.972 | 0.258 | 0 | BODY _{base} | 0.077 | 0.152 | 0.087 | 0.165 | 0.398 | 1.000 |

Matrix of pairwise differences in PLS effect sizes (left), and their associated significance levels (right).

Discussion and Conclusion

This repository-based analysis of a curated and majority-NAGPRA collection of intact or reconstructed Caddo bottles resulted in an improved characterization of Caddo bottle shapes, while highlighting similarities, differences, and a general trend toward standardization for the aggregated sample. Specifically, it resulted in a test of Smithport Plain bottle shapes confirming that discrete morphological characteristics (body and base) differ significantly between the Belcher Mound and Smithport Landing sites, supporting the morphological assertion initially posited by Webb (1959). The test included an analysis of Smithport Plain bottles from the Pohler Collection and Mitchell site, which do not differ significantly in shape from those recovered at the Smithport Landing or Belcher Mound sites. Analysis of the aggregate sample indicates allometric trajectories that are not homogenous for Formative-Early and Late-Historic Caddo types, a significant difference in bottle shape by site and type, significant morphological disparity between

the Formative-Early and Late-Historic Caddo types, and significant morphological integration of pairs and suites of bottle components.

In the aggregated sample, significant assemblage-level differences in bottle shape exist between Belcher Mound compared with Gahagan Mound and Smithport Landing, Gahagan Mound compared with Haley Place, Mitchell, Pohler Collection, Haley Place and Smithport Landing, and Mitchell compared with those from Smithport Landing (see Table 7 and Figure 12). The results imply that bottle shapes employed by Formative-Early Caddo potters differ from those produced by Late-Caddo potters; an assertion echoed by the analyses of allometric trajectories (see Figure 10) and morphological disparity (see Table 10). While only a small sample has been examined thus far, the results of morphological disparity by period highlight a gradual trend toward standardization, where bottles produced in the Late-Historic Caddo periods occupy a more restricted range of morphospace than those manufactured in the Formative-Early Caddo periods. This dynamic assertion is subject to change as more bottles are added to the analysis.

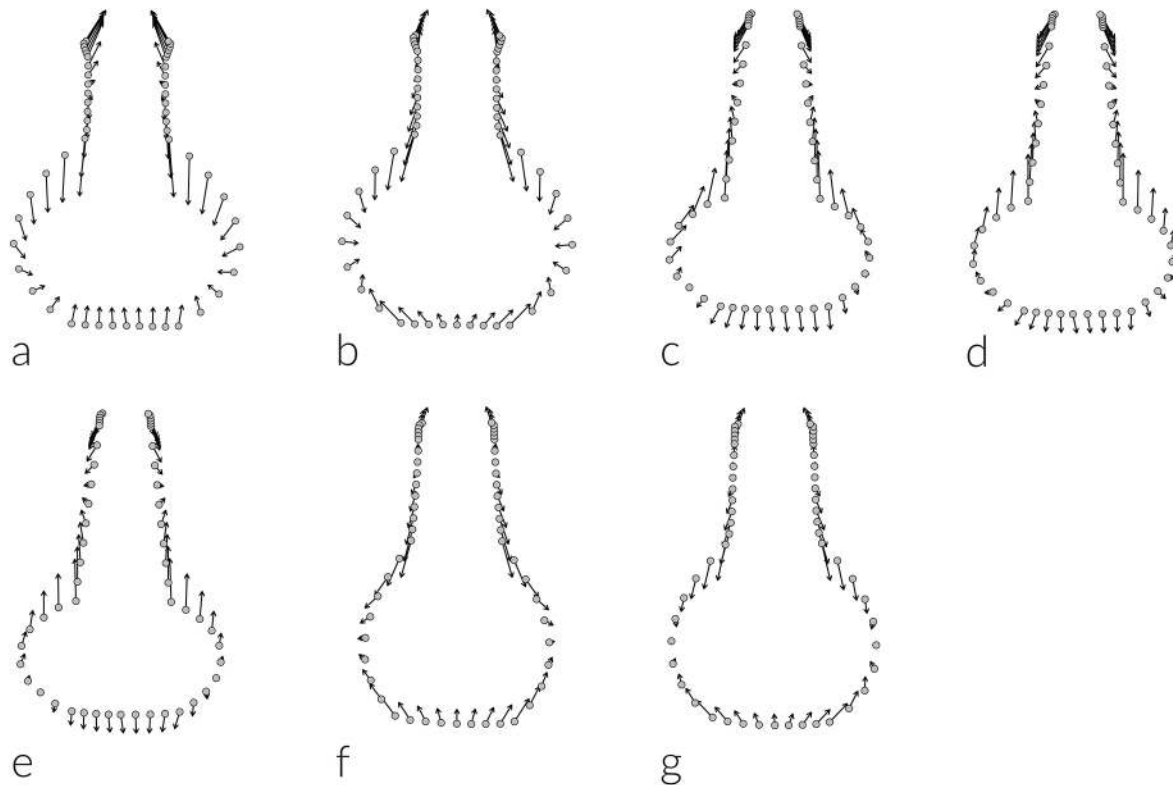


Figure 12. Comparison of mean bottle shapes by site for those sites found to differ significantly; (a) Belcher Mound (gray) and Gahagan Mound, (b) Belcher Mound (gray) and Smithport Landing, (c) Gahagan Mound (gray) and Haley Place, (d) Gahagan Mound (gray) and Mitchell, (e) Gahagan Mound (gray) and Pohler, (f) Haley Place (gray) and Smithport Landing, and (g) Mitchell (gray) and Smithport Landing.

Significant type-specific differences in bottle shape occur between Belcher Engraved compared with Hickory Engraved, Keno Trailed, and Smithport Plain; Keno Trailed compared with Smithport Plain and Taylor Engraved; Hickory Engraved compared with Keno Trailed; and Smithport Plain compared with Taylor Engraved (see Table 8 and Figure 13). The test of morphological disparity indicated that Hickory Engraved and Smithport Plain bottles occupy a significantly greater range of morphospace than the Belcher Engraved bottles (see Table 9). Elsewhere it may be the case that dimensional attributes are inappropriate for use in studies of standardization and diversity (Rice 1991); however, the morphological disparity results suggest a high degree of utility in clarifying questions of standardization and diversity through the employment of morphological traits associated with Caddo bottles. This can, in turn, provide evidence for varying degrees of tolerance in production, where a higher tolerance yields a greater range of variation in shape that decreases as shapes become more standardized (Eerkins and Bettinger

2001). In this sample, the tolerance for diversity in Caddo bottle shape is higher in the Formative-Early Caddo period, and becomes more restricted through time. Results specify that the tolerance for variation in Caddo bottle shapes is greater in the case of Smithport Plain and Hickory Engraved than it is for Belcher Engraved.

Results from the test of morphological integration indicate that Caddo bottles are significantly integrated (see Figure 11), lending some support to the hypothesis that Caddo potters were adhering to a template of vessel shape associated with specific decorative motifs (Early 2012). However, the suites of attributes were not found to covary in a predicted manner, as it is the RIM_{neck} and RIM_{base} , RIM_{body} and RIM_{base} , and RIM_{base} and $NECK_{body}$ that exhibit significant integration (see Table 11). An important component of the expanded research program will be type-specific tests of morphological integration following an increase in sample size.

The significant difference in the production of Smithport Plain body and base shapes at the

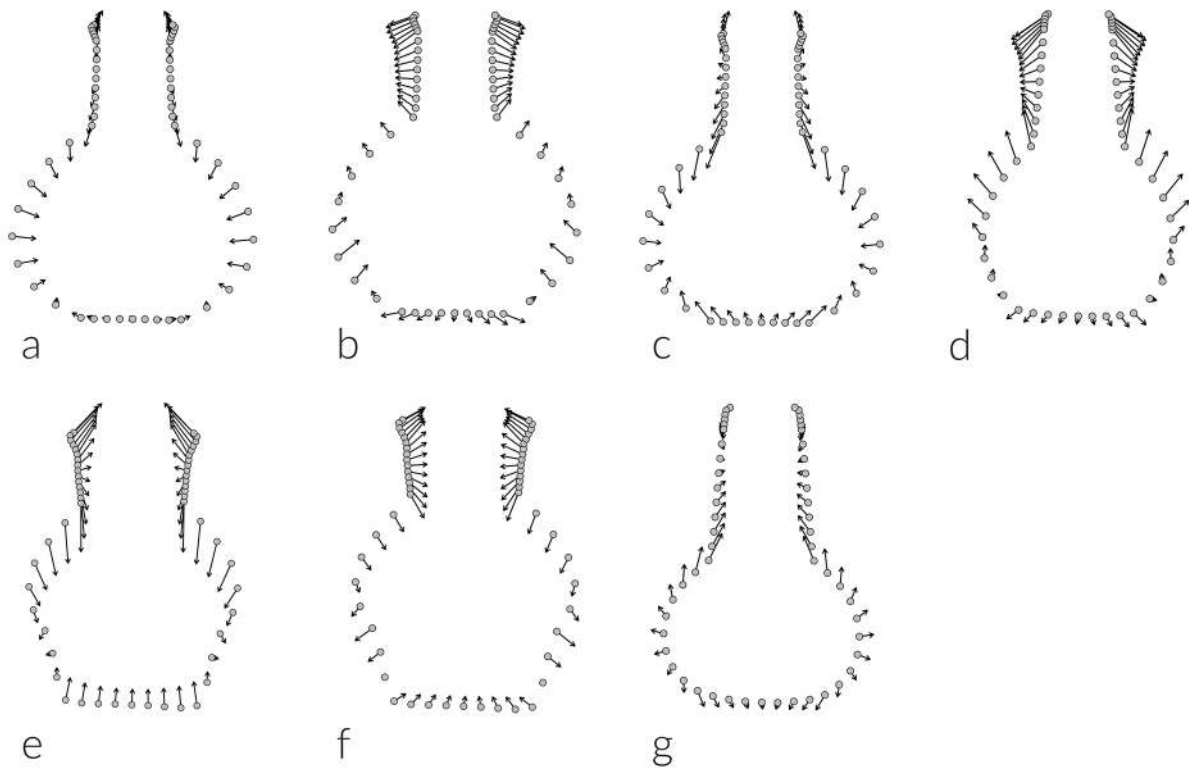


Figure 13. Comparison of mean bottle shapes by type for those types found to differ significantly; (a) Belcher Engraved (gray) and Hickory Engraved, (b) Belcher Engraved (gray) and Keno Trailed, (c) Belcher Engraved (gray) and Smithport Plain, (d) Hickory Engraved (gray) and Keno Trailed, (e) Keno Trailed (gray) and Smithport Plain, (f) Keno Trailed (gray) and Taylor Engraved, and (g) Smithport Plain (gray) and Taylor Engraved.

Smithport Landing and Belcher Mound sites was posited in the initial analysis (Selden, Jr. 2018a) where the differences in shape were seen as a possible north-south transition for the combined sample of Hickory Engraved and Smithport Plain types. In comparing these results with a recent analysis of the Hickory Engraved sample (Selden, Jr. 2018b), it is evident that while these two Formative-Early Caddo types exhibit similar morphological differences over geographic space, the differences are dynamic and will be further clarified by the continued and iterative expansion of type-specific analyses.

The contribution of GM methods to questions of Caddo ceramic morphology holds substantial promise. Those results presented here provide a succinct preview of a rigorous and systematic research design that capitalizes on the variability of Caddo ceramic shapes through an analysis of type-specific (Smithport Plain) morphology that is followed by an analysis of the aggregated sample of Caddo bottles. Iterative improvements to this research program will continue as new specimens are made available and incorporated. That progression will include the addition of Caddo bottles from the Bison B site in northwest Louisiana (Woodall 1969) curated at Southern Methodist University, and an expansion of the Belcher and Taylor Engraved samples. This will test whether similarity in Late-Historic Caddo bottle shape is a local, regional, or area-wide trend. Also considered will be the continued analyses of morphological disparity between different temporal periods to test whether significant morphological disparity and allometry between the temporal periods varies elsewhere; and the continued use of morphological integration to identify which of those morphological traits associated with Caddo bottle production might be said to covary.

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