

A Randomized Clinical Trial of an Inactivated Avian Influenza A (H7N7) Vaccine

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Abstract

Background: Concern for a pandemic caused by a newly emerged avian influenza A virus has led to clinical trials with candidate vaccines as preparation for such an event. Most trials have involved vaccines for influenza A (H5N1), A (H7N7) or A (H9N2).

Objective: To evaluate dosage-related safety and immunogenicity of an inactivated influenza A (H7N7) vaccine in humans.

Design: One hundred twenty-five healthy young adults were randomized to receive two doses intramuscularly of placebo or 7.5, 15, 45 or 90 μg of HA of an inactivated subunit influenza A (H7N7) vaccine (25 per group), four weeks apart. Reactogenicity was evaluated closely for one week and for any adverse effect for six months after each dose. Serum hemagglutination-inhibiting and neutralizing antibody responses were determined four weeks after each dose and at six months.

Results: Reactogenicity evaluations indicated the vaccinations were well tolerated. Only one subject developed a ≥ 4 -fold serum hemagglutination-inhibition (HAI) antibody response and a final titer of $\geq 1:40$ four weeks after dose two and only five subjects developed a neutralizing antibody rise and a final titer of $\geq 1:40$ in tests performed at a central laboratory. Four of the five were given the 45 or 90 μg HA dosage. A more sensitive HAI assay at the study site revealed a dose-response with increasing HA dosage but only 36% in the 90 μg HA group developed a ≥ 4 -fold rise in antibody in this test and only one of these achieved a titer of $\geq 1:32$.

Conclusion: This inactivated subunit influenza A (H7N7) vaccine was safe but poorly immunogenic in humans.

Trials Registration: ClinicalTrials.gov NCT00546585

Citation: Couch RB, Patel SM, Wade-Bowers CL, Niño D (2012) A Randomized Clinical Trial of an Inactivated Avian Influenza A (H7N7) Vaccine. PLoS ONE 7(12): e49704. doi:10.1371/journal.pone.0049704

Editor: Eliane Namie Miyaji, Instituto Butantan, Brazil

Received: July 23, 2012; **Accepted:** October 4, 2012; **Published:** December 11, 2012

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Funding: Research performed by the authors and summarized in this report was supported by Public Health Service Contract NO1 AI 30039 from the National Institute of Allergy and Infectious Diseases. The funders participated in study design but had no role in data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist. The content of this publication does not necessarily reflect the views or policies of the Department of Health and Human Services, nor does mention of trade names, commercial products, or organizations imply endorsement by the U.S. Government.

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Introduction

The prevailing concept for the origin of new influenza A virus subtypes leading to pandemic influenza in humans is that a new virus with the ability to spread and cause illness contains an avian influenza virus hemagglutinin (HA) and/or neuraminidase glycoprotein (NA) acquired from an avian influenza virus [1–3]. This potential exists for the seventeen distinct HAs and nine NAs that have been described among influenza A viruses [1,4,5]. The level of concern for this happening was increased when an outbreak with avian influenza A (H5N1) occurred in humans in 1997 in Hong Kong that was from contacts with infected chickens and was reinforced with reappearance of human cases in other parts of the world in succeeding years [6–8]. Preparing candidate A/H5N1 vaccines for potential use in humans became an urgent need. Since then, an outbreak with avian influenza A (H7N7) occurred in humans in the Netherlands and cases of avian influenza A (H9N2)

have been recognized in humans [9–12]. Preparing prototype vaccines for these and perhaps other avian influenza A viruses is now an effort supported by public health authorities so as to be prepared with the knowledge of how to best proceed should one of these subtypes emerge as a pandemic among humans.

The present report is of a report of a clinical trial with an influenza A (H7N7) vaccine prepared by a manufacturer that used established methods for annual production of seasonal influenza vaccine. The objective was to evaluate dosage-related safety and immunogenicity. This vaccine proved to be safe but poorly immunogenic in humans. A second related report is of results comparing a number of the prototype inactivated vaccines containing avian HAs and NAs that have been evaluated in humans, including the A/H7N7 vaccine used in this report, in various in vitro tests in an effort to identify correlates that related to immunogenicity in humans other than the standard single radial immunodiffusion (SRID) assays of HA concentration.

Materials and Methods

The protocol for this trial and supporting CONSORT checklist are available as supporting information; see Checklist S1 and Protocol S1.

Subjects and Ethics Statement

Subjects were healthy male and nonpregnant females between the ages of 18 and 40 years. The study was reviewed and approved by the Institutional Review Board for Protection of Human Subjects in research at Baylor College of Medicine before commencing. The study was conducted in a clinic setting and all subjects gave written informed consent before any procedures were conducted. The Declaration of Helsinki principles were followed.

The H7 Vaccine

The tested vaccine is a monovalent inactivated influenza A (H7N7) vaccine produced by Sanofi Pasteur Inc. using their seasonal vaccine production methods. The virus used was a 6-2 reassortant generated in eggs. The virus donating the HA was A/Mallard/Netherlands/12/2000 (H7N3) and that donating the NA was A/Mallard/Netherlands/2/2000 (H10N7); both are low pathogenic avian influenza viruses. The six internal genes were donated by an influenza A (H1N1) strain, FDA-Resvir-12; the NP gene was from A/Johannesburg/82/96(H1N1) and the other five internal protein genes were from A/Puerto Rico/8/34 (H1N1) [13]. The vaccine virus was grown in embryonated eggs, inactivated with formalin, concentrated and purified and then detergent disrupted with Triton X-100 to produce a subunit virus antigen that was then further purified. It was formulated into single dose vials with 0.05% gelatin but no preservative as dosages of 7.5, 15, and 45 μ g per 0.5 ml.

Sample Size Determination

The sample size (25 per group), was selected by the data coordinating center of the Microbiology Division of the National Institutes of Allergy and Infectious Diseases to provide a robust initial safety database of 125 vaccinees receiving the antigen while providing some information concerning the dose-response immunogenicity in a timely fashion. Immunogenicity results were intended to be sufficient to establish a basis for narrowing the dose selection for further studies in infants, children, and elderly populations. A power calculation was not performed for this Phase I study but for an assumed serious adverse event frequency of 0.1% to 20%, the probability of detecting one or more events for 25 to 100 subjects ranged from 2.47% to 100%.

Design

The study was conducted in a randomized, double-blind fashion; randomization was conducted after enrollment at the data coordinating center of the sponsor. One hundred thirty-six subjects were screened and 125 were randomly assigned to groups of 25 each to receive 0.5 ml doses intramuscularly of placebo or 7.5 μ g, 15 μ g, or 45 μ g of vaccine HA on days 0 and 28. A fifth group of 25 received one ml of the 45 μ g per 0.5 ml dosage (total of 90 μ g) at each vaccination. Vaccine dosages were based on single radial immunodiffusion assays (SRID) for HA [14,15]. Vaccinations were done by persons who did not participate in any further work in the study. Individual vaccine dosages were transmitted to the study site from the coordinating center by code and the site pharmacist, with corroboration by the unblinded vaccination nurse, selected the vaccine dosage syringe to be used. The initial vaccination was on March 10, 2008 and the last on

May 19, 2008. The reactogenicity evaluations and antibody assays were performed by persons with no knowledge of the vaccine given. Reactogenicity evaluations consisted of daily recording by subjects of oral temperature, symptoms and any vaccine site changes on a scale of mild, moderate, severe for seven days after each vaccination, a phone check at one to three days and a clinic visit for evaluation at eight to twelve days after each vaccination. Additionally, any adverse effect (AE) up to 28 days after each vaccination and any severe adverse effects (SAE) up to day 208 was recorded and evaluated. Blood for antibody assays was obtained before each vaccination (0 and 28) and on days 56 and 208 after the initial vaccination.

Laboratory Tests

A central laboratory performed hemagglutination-inhibition (HAI) and neutralizing (neut) antibody assays on all serum specimens as described [16,17]. The starting dilution of serum for each assay was 1:10. Another laboratory experienced with avian influenza virus serology tested a sample of the specimens. The primary study site also did HAI tests on the day 0 and 56 serum samples using a starting dilution of 1:4 as described [18]. Planned analyses were for number of subjects developing a ≥ 4 fold increase, the number with a final titer $\geq 1:40$ and the geometric mean titer (GMT) for each group in the serum antibody tests.

Analyses

Reactogenicity. The number of subjects in each vaccine group reporting a specific solicited local or systemic adverse effect (AE) that was mild, moderate or severe for seven days after each vaccination and the number and description of unsolicited AEs for 28 days after each vaccination were totaled. Occurrence of severe AEs (SAE) was monitored for six months and individually evaluated in detail for any potential vaccine relationship.

Immunogenicity. The number of subjects in each vaccine group developing a ≥ 4 -fold serum antibody response and achieving a final titer of $\geq 1:40$ in HAI and neut assays was totaled 28 days after each vaccination and at 208 days. Also, the number (%) of subjects developing a ≥ 4 -fold and $\geq 1:32$ final titer 28 days after dose 2 (day 56) in the more sensitive HAI assay was totaled. GMT calculations were not done.

Statistical Analyses. Statistical analyses were conducted using SPSS version 17.0.

Registry Name, Number and Performance

Name. Safety, reactogenicity, and immunogenicity of inactivated influenza A/H7/N7 vaccine in healthy adults. No.: NCT00546585. Initiated March 10, 2008, completed February 10, 2009.

Performance. Study performance conformed to CONSORT guidelines and is reported conforming to Consort 10 guidelines (PLoS Medicine 2010, 7(3):1–7 e1000251).

Results

The Study

A flow diagram of the randomized trial is shown in Figure 1. One hundred thirty-six subjects were screened and 125 were enrolled. Eleven persons were excluded, one for over age 40, four for a blood pressure exceeding 140/90, four for a new or changed medication within the three month limit, one for having received a recent vaccination, one for a positive pregnancy test and one for an unstable medical condition.

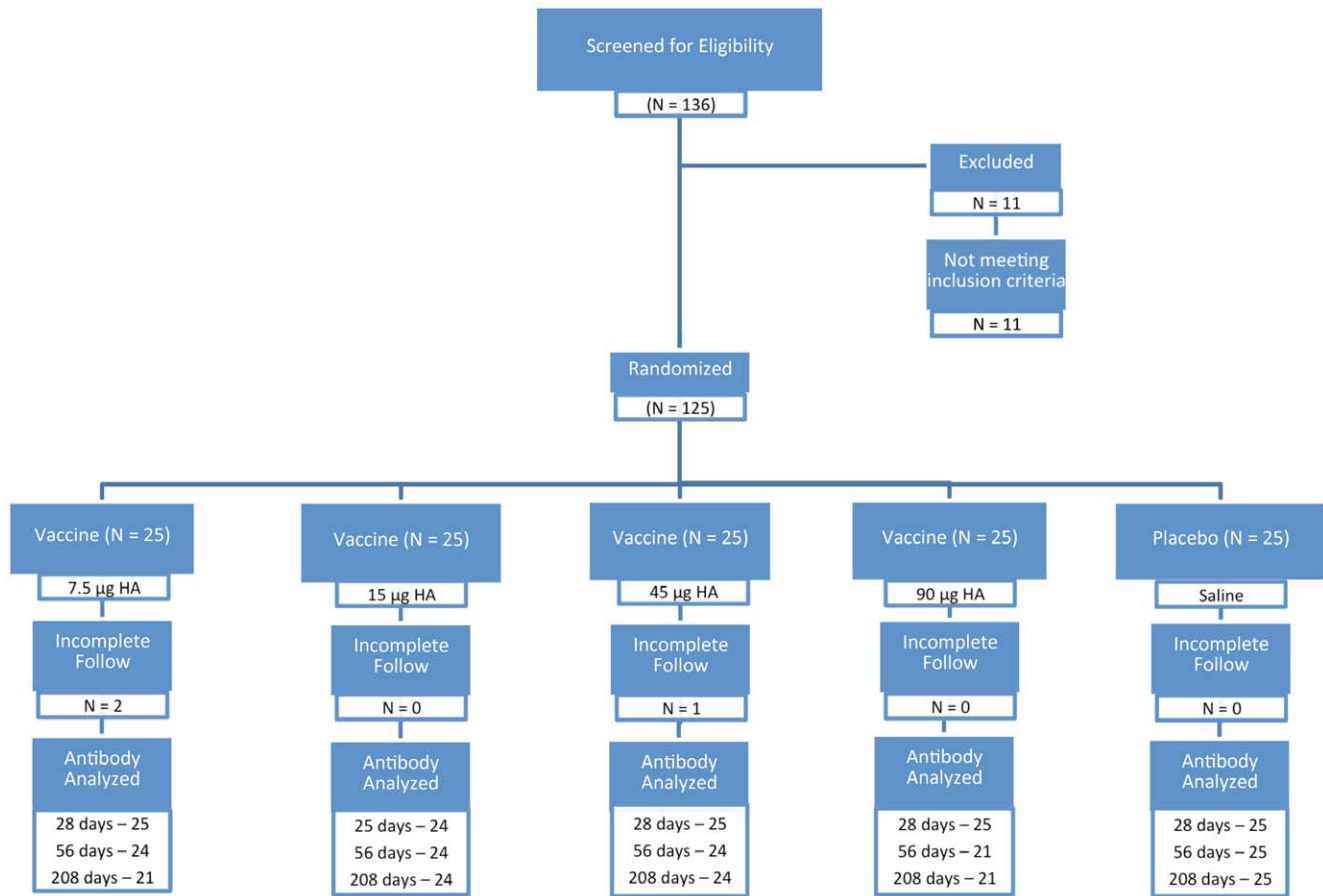


Figure 1. Flow diagram of the randomized clinical trial of an inactivated subunit influenza A (H7N7) vaccine.
doi:10.1371/journal.pone.0049704.g001

One hundred twenty-five subjects were randomized; complete followup for 208 days was available on 122 (97.6%). Of the three who withdrew, one thought dose 1 had caused an acute respiratory illness and two withdrew for travel. Four others were not given dose 2 because of an SAE (see reactogenicity), and one of these had started a new medicine (exclusion criterion). The number of subjects in each vaccination group tested for antibody at each specified time is shown in the figure.

Demographics of the 125 enrolled subjects are shown in Table 1. The median age was 26.4 years and the range was 18 to 40 years. The male and female percent of participants was 49 and 51%, respectively, and most subjects were white. There were no significant differences among the different dosage groups by gender, race, or age (Table 1).

Reactogenicity

Solicited adverse events in the seven days following each vaccination are summarized in Table 2. After dose one, fever ($\geq 37.7^{\circ}\text{C}$), was reported by one subject given the 7.5 μg dosage and two given the 90 μg dosage. After dose two, fever was reported by one subject given the 7.5 μg dosage and one given 45 μg . The most commonly reported systemic symptoms in the week postvaccination were headache and malaise; the most commonly reported local symptoms were redness and tenderness. There was an increased frequency of moderate AE among those given the higher dosages but there were no severe AEs. There were no severe unsolicited reactions; mild or moderate unsolicited

AEs thought to be vaccine related were transient and were injection site bruise – 2, site pruritus – 1, dizzy – 2, vomiting – 1, throat irritation – 1, anxiety – 1, and a new herpes labialis lesion – 1. Overall, the vaccinations were well tolerated.

Four subjects were designated as an SAE during the study period. Two persons were hospitalized, one for moderately severe back pain considered related to a car accident two years earlier but the hospitalization was extended because of depression; both illnesses resolved without sequelae. The other hospitalization was for stress-related severe depression that improved during hospitalization and was considered resolved on discharge. A 40-year old female was noted to have eyelid edema when reporting for dose two and reported a new physician-prescribed medication; she also reported experiencing eyelid edema in the past. Follow-up evaluations led to a diagnosis of mixed connective tissue disease and special tests on her prevaccination and earlier available sera revealed evidence of a preexisting autoimmune disorder. The illness was considered moderately-severe. None of these three subjects were given the second vaccination. Both the site investigators and the independent safety monitoring committee considered these SAEs as not associated with vaccination. The fourth SAE was a mild transient decrease in hearing by the left ear with onset nine days after the initial vaccination that did not interfere with normal activities. A specific cause other than vaccination was not identified and treatment provided by ENT consultation did not lead to improvement. Hearing returned to normal in 84 days; the second vaccination was not done. Both the site investigators and the independent safety monitoring commit-

Table 1. Demographics by Study Group.

	ALL (No. = 125)	7.5 mcg (No. = 25)	15 mcg (No. = 25)	45 mcg (No. = 25)	90 mcg (No. = 25)	Placebo (No. = 25)
Gender – No. (%)¹						
Male	61 (49)	10 (40)	11 (44)	14 (56)	13 (52)	13 (52)
Female	64 (51)	15 (60)	14 (56)	11 (44)	12 (48)	12 (48)
Race – No. (%)²						
American Indian/ Alaskan Native	0	0	0	0	0	0
Asian	14 (11)	4 (16)	2 (8)	2 (8)	4 (16)	2 (8)
Hawaiian/Pacific Islander	0	0	0	0	0	0
Black/African American	10 (8)	2 (8)	0	5 (20)	1 (4)	2 (8)
White	94 (75)	19 (76)	23 (92)	15 (60)	18 (72)	19 (76)
Multi-Racial	6 (5)	0	0	2 (8)	2 (8)	2 (8)
Other/Unknown	1 (1)	0	0	1 (4)	0	0
Age in Years³						
Mean (STD)	28.1 (5.0)	29.1 (5.5)	29.6 (5.5)	27.8 (4.9)	28.2 (4.8)	25.7 (3.7)
Median	26.4	28.8	28.4	26.1	28.0	25.2
Min,Max	(18, 40)	(22, 40)	(18, 40)	(21, 38)	(19, 38)	(20, 38)

¹Male vs. nonmale frequency by dosage, chi-square 1.73, p = .785.

²White vs. nonwhite frequency by dosage, chi-square 7.04, p = .134.

³Age vs. dosage, Anova, p > .05.

doi:10.1371/journal.pone.0049704.t001

tee designated the AE as vaccination associated; the independent monitoring committee required an SAE designation because of “significant disability.”

Immunogenicity

Serum antibody response measurements at the central laboratory 28 days after each vaccination are shown in Table 3. As noted, very few significant responses were seen but most were after the 45 and 90 µg dosages. None of the subjects retained a 4-fold response and titer $\geq 1:40$ in HAI tests of sera collected at 208 days

and only one subject (15 µg HA dosage) retained an increase and final $\geq 1:40$ titer in neut tests (data not shown). The infrequent responses reported by the central laboratory were confirmed by the laboratory with experience in serology for avian influenza viruses (data not shown). Results of testing for HAI antibody at the primary test site are shown in Table 4. The greater sensitivity of the assay revealed a dose response in frequency of significant increases in titer (logistic regression, p = .001) although the frequency for the highest dosage was only 36% and only two of the subjects achieved a titer of 1:32.

Table 2. Solicited Adverse Events (AE) after Influenza A (H7N7) Vaccinations.

Adverse Events	Dosage ¹ (µg HA)	No. (%) with \geq One AE in Category							
		Dose 1				Dose 2			
		No. ²	Mild	Mod	Severe	No. ²	Mild	Mod	Severe
Systemic	0	25	5 (20)	3 (12)	0	22	5 (23)	1 (5)	0
	7.5	25	10 (40)	1 (4)	0	24	1 (4)	1 (4)	0
	15	25	9 (36)	2 (8)	0	24	5 (21)	1 (4)	0
	45	25	5 (20)	2 (8)	0	23	3 (13)	0 (0)	0
	90	25	4 (16)	6 (24)	0	25	4 (16)	1 (4)	0
Local	0	25	13 (52)	0 (0)	0	22	12 (55)	0 (0)	0
	7.5	25	15 (60)	0 (0)	0	24	14 (58)	0 (0)	0
	15	25	11 (44)	2 (8)	0	24	17 (71)	1 (4)	0
	45	25	14 (56)	1 (4)	0	23	15 (65)	3 (13)	0
	90	25	14 (56)	3 (12)	0	25	13 (52)	3 (12)	0

¹As determined in single radial immunodiffusion assays.

²Number subjects evaluated.

doi:10.1371/journal.pone.0049704.t002

Table 3. Serum Antibody Responses to Influenza A (H7N7) Vaccinations¹.

Dosage (μg HA) ²	Post Dose 1					Post Dose 2				
	No. ³	No. (%) ≥ 4 -Fold Increase		No. (%) $\geq 1:40$		No. ³	No. (%) ≥ 4 -Fold Increase		No. (%) $\geq 1:40$	
		HAI ⁴	Neut ⁴	HAI ⁴	Neut ⁴		HAI ⁴	Neut ⁴	HAI ⁴	Neut ⁴
0	25	0	0	0	0	25	0	0	0	0
7.5	25	0	0	0	0	24	0	0	0	0
15	24	0	1 (4)	0	1 (4)	24	0	1 (4)	0	1 (4)
45	25	0	0	0	0	24	0	2 (8)	0	2 (8)
90	25	0	0	0	0	21	1 (5)	2 (10)	1 (5)	2 (10)

¹As determined at central laboratory; increase defined as ≥ 4 -fold or < 10 to 40.

²As determined in single radial immunodiffusion assays.

³Number of subjects in evaluation.

⁴Hemagglutination-inhibition (HAI) and neutralizing (neut) antibody assays at central laboratory.

doi:10.1371/journal.pone.0049704.t003

Discussion

The major finding of this study is that an inactivated influenza A (H7N7) subunit vaccine prepared using manufacturing methods approved for seasonal influenza vaccines was poorly immunogenic in healthy young adults despite their having been given two doses a month apart of dosages up to 90 μg of the HA as determined in SRID assays. The neut assay at the central laboratory appeared to be slightly more sensitive than the HAI assay whereas the HAI assay appeared more sensitive than the neut assay at a laboratory with considerable experience with avian influenza virus serology (data not shown). Nevertheless, the serologic findings at the central laboratory were essentially confirmed. The more sensitive HAI assay performed at the study site provided a more complete assessment of immunogenicity. That test indicated that immunogenic HA was present in the vaccine although the immunogenicity was still considerably lower than what was expected for the dosages administered. The major limitation of these findings is that the sample size was not selected to produce definitive immunogenicity data on any of the dosages tested. However, the desired guidance for any further testing was obtained.

Influenza A (H7N7) is proposed as a priority for vaccine development for potential use in humans along with H5N1 and H9N2 because of human infections that have been detected with these viruses [3,19]. The case for H7N7 was emphasized when an

outbreak with H7N7 viruses occurred in 2003 in the Netherlands among persons exposed to poultry; one infected person died and evidence for human-to-human transmission was provided [9,10]. Also, in an outbreak of H7N3 infections in poultry in Canada in 2004, transmission to humans was detected [20,21].

Considerable effort has been expended on vaccine development with H5N1 viruses for potential use in humans [22]. Influenza A/H7 vaccines have been developed and shown to be immunogenic and to induce protection in animals [13,23–25]. Thus, the H7 HA appears fully immunogenic in vaccinations of animals and should also be fully immunogenic for humans. Other clinical trials in humans with H7 vaccines have been reported. Low immunogenicity was reported in a clinical trial using a split virus vaccine prepared in tissue culture with A/H7N1 plasmids derived from a chicken virus [26]. Two vaccinations three weeks apart of 12 and 24 μg dosages of the HA with and without alum were each given to 15 healthy adults (total 60). The 12 and 24 μg dosages without alum induced HAI and/or neut antibody responses in 21 and 23% of subjects, respectively and 50 and 62% respectively with adjuvant. A clinical trial with an H7N3 live attenuated virus using the A/Ann Arbor (H2N2) cold-adapted Master Donor Strain given to 17 subjects showed minimal virus shedding but serum HAI and neut antibody responses in 43% and 48%, respectively [27].

The present report and the two summarized clinical trials, in combination with the good immunogenicity in animal immunizations, indicate that a satisfactory H7 vaccine can be made for humans. Because of the potential need for an H7N7 vaccine for humans, developing a vaccine for potential use in humans seems indicated. To aid in this goal, we have directed research toward gaining some understanding of why this H7N7 vaccine of high HA dosage was so poorly immunogenic in humans. These studies are reported in a companion report on in vitro correlates of immunogenicity of avian influenza virus vaccines in humans.

Supporting Information

Checklist S1 Consort checklist.

(DOC)

Protocol S1 Study protocol.

(PDF)

Table 4. Serum Antibody Responses to H7N7 Vaccinations¹.

Dosage/ μg HA ²	No. ³	HAI Antibody Post Dose 2	
		No. (%) ≥ 4 Fold Increase ⁴	No. (%) $\geq 1:32$
0	25	0	0
7.5	21	2 (9.5)	0
15	21	2 (9.5)	1 (5)
45	24	6 (25)	0
90	22	8 (36)	1 (5)

¹Hemagglutination-inhibition antibody assays at primary study 'site.

²As determined in single radial immunodiffusion assays.

³Number of subjects in evaluation.

⁴Increasing dosage induced increased % increase; logistic regression, $p = .001$.

doi:10.1371/journal.pone.0049704.t004

Acknowledgments

Statistical analyses: Robert L. Atmar, M.D.
 Manuscript preparation: I. Darlene Kirk, CCRP.
 Data summaries: EMMES Corporation, Heather Hill.
 Study design and vaccine contribution: National Institute of Allergy and Infectious Diseases, Roland Levandowski.

References

1. Webster RG, Bean WJ, Gorman OW, Chambers TM, Kawaoka Y (1992) Evolution and ecology of influenza A viruses. *Microbiol Rev* 56: 152–79.
2. Taubenberger JK, Reid AH, Lourens RM, Wang R, Jin G, et al. (2005) Characterization of the 1918 influenza virus polymerase genes. *Nature* 437: 889–93.
3. Yen HL, Webster RG (2009) Pandemic influenza as a current threat. *Curr Top Microbiol Immunol* 333: 3–24.
4. Fouchier RA, Munster V, Wallensten A, Bestebroer TM, Herfst S, et al. (2005) Characterization of a novel influenza A virus hemagglutinin subtype (H16) obtained from black-headed gulls. *J Virol* 79: 2814–2822.
5. Tong S, Li Y, Rivaller P, Conrardy C, Castillo DA, Chen LM, et al. (2012) A distinct lineage of influenza A virus from bats. *Proc Natl Acad Sci U S A* 109: 4269–74.
6. Yuen KY, Chan PKS, Peiris M, Tsang DNC, Que TL, Shortridge KF, et al. (1998) Clinical features and rapid viral diagnosis of human disease associated with avian influenza A H5N1 virus. *Lancet* 351: 467–471.
7. Hien TT, Liem NT, Dung NT, San LT, Mai PP, van Vinh Chau N, et al. (2004) Avian influenza A (H5N1) in 10 patients in Vietnam. *N Engl J Med* 350: 1179–88.
8. Writing Committee of the Second World Health Organization Consultation on Clinical Aspects of Human Infection with Avian Influenza A(H5N1) Virus. (2008) Update on avian influenza A(H5N1) virus infection in humans. *N Engl J Med* 358: 261–273.
9. Fouchier RA, Schneberger PM, Rozendaal FW, Brockman JM, Kemink SA, et al. (2004) Avian influenza A virus (H7N7) associated with human conjunctivitis and a fatal case of acute respiratory distress syndrome. *Proc Natl Acad Sci USA* 101: 1356–1361.
10. Koopmans M, Wilbrink B, Conyn M, Natrop G, van der Nat H, et al. (2004) Transmission of H7N7 avian influenza A virus to human beings during a large outbreak in commercial poultry farms in the Netherlands. *Lancet* 363: 587–593.
11. Peiris M, Yuen KY, Leung CW, Chan KH, Lai RWM, et al. (1999) Human infection with influenza H9N2. *Lancet* 354: 916–917.
12. Butt KM, Smith GJD, Chen H, Zhang IJ, Leung YHC, et al. (2005) Human infection with an avian H9N2 influenza A virus in Hong Kong in 2003. *J Clin Microbiol* 43: 5760–5767.
13. Jadhao SJ, Achenbach J, Swayne DE, Donis R, Cox N, et al. (2008) Development of Eurasian H7N7/PR high growth reassortant virus for clinical evaluation as an inactivated pandemic influenza vaccine. *Vaccine* 26: 1742–1750.
14. Wood JM, Schild GC, Newman RW, Seagroatt V (1977) An improved single-radial-immunodiffusion technique for the assay of influenza haemagglutinin antigen: application for potency determination of inactivated whole virus and subunit vaccines. *J Biol Stand* 5: 237–247.
15. Williams MS (1993) Single-radial-immunodiffusion as an in vitro potency assay for human inactivated viral vaccines. *Vet Microbiol* 37: 253–262.
16. Stephenson I, Wood JM, Nicholson KG, Charlett A, Zambon MC (2004) Detection of anti-H5 responses in human sera by HI using horse erythrocytes following MF59-adjuvanted influenza A/Duck/Singapore/97 vaccine. *Virus Res* 103: 91–95.
17. Rowe T, Abernathy RA, Hu-Primmer J, Thompson WW, Lu X, et al. (1999) Detection of antibody to avian influenza A (H5N1) virus in human serum by using a combination of serologic assays. *J Clin Microbiol* 37: 937–943.
18. Couch RB, Winokur P, Brady R, Belshe R, Chen WH, et al. (2007) Safety and immunogenicity of a high dosage trivalent influenza vaccine among elderly subjects. *Vaccine* 25: 7656–7663.
19. Belsler JA, Bridges CB, Katz JM, Tumpey TM (2009) Past, present, and possible future human infection with influenza virus A subtype H7. *Emerg Infect Dis* 15: 859–865.
20. Hirst M, Astell CR, Griffith M, Coughlin SM, Moksa M, et al. (2004) Novel avian influenza H7N3 strain outbreak, British Columbia. *Emerg Infect Dis* 10: 2192–95.
21. Tweed SA, Skowronski DM, David ST, Larder DA, Petri M, et al. (2004) Human illness from avian influenza H7N3, British Columbia. *Emerg Infect Dis* 10: 2196–2199.
22. Keitel WA, Atmar RL (2009) Vaccines or pandemic influenza: summary of recent clinical trials. In: Compans RW, Orenstein WA, editors. *Vaccines for pandemic influenza*. Berlin Heidelberg: Springer-Verlag; pp. 431–447.
23. Pappas C, Matsuoka Y, Swayne DE, Donis RO (2007) Development and evaluation of an influenza virus subtype H7N2 vaccine candidate for pandemic preparedness. *Clin Vaccine Immunol* 14: 1425–32.
24. Itoh Y, Ozaki H, Ishigaki H, Sakoda Y, Nagata T, et al. (2010) Subcutaneous inoculation of a whole virus particle vaccine prepared from a non-pathogenic virus library induces protective immunity against H7N7 highly pathogenic avian influenza virus in cynomolgus macaques. *Vaccine* 28: 780–789.
25. Min JY, Vogel L, Matsuoka Y, Lu B, Swayne D, et al. (2010) A live attenuated H7N7 candidate vaccine virus induces neutralizing antibody that confers protection from challenge in mice, ferrets, and monkeys. *J Virol* 84: 11950–11960.
26. Cox RJ, Madhun AS, Hauge S, Sjurson H, Major D, et al. (2009) A phase I clinical trial of a PER.C6® cell grown influenza H7 virus vaccine. *Vaccine* 27: 1889–1897.
27. Talaat KR, Karron RA, Callahan KA, Luke CJ, DiLorenzo SC, et al. (2009) A live attenuated H7N3 influenza virus vaccine is well-tolerated and immunogenic in a phase I trial in healthy adults. *Vaccine* 27: 3744–3753.

Author Contributions

Conceived and designed the experiments: RBC. Performed the experiments: RBC SMP CLW DN. Analyzed the data: RBC. Wrote the paper: RBC.