



A Review on Plants and Microorganisms Mediated Synthesis of Silver Nanoparticles, Role of Plants Metabolites and Applications

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Abstract: Silver nanoparticles are one of the most extensively studied nanomaterials due to their high stability and low chemical reactivity in comparison to other metals. They are commonly synthesized using toxic chemical reducing agents which reduce metal ions into uncharged nanoparticles. However, in the last few decades, several efforts were made to develop green synthesis methods to avoid the use of hazardous materials. The natural biomolecules found in plants such as proteins/enzymes, amino acids, polysaccharides, alkaloids, alcoholic compounds, and vitamins are responsible for the formation of silver nanoparticles. The green synthesis of silver nanoparticles is an eco-friendly approach, which should be further explored for the potential of different plants to synthesize nanoparticles. In the present review we describe the green synthesis of nanoparticles using plants, bacteria, and fungi and the role of plant metabolites in the synthesis process. Moreover, the present review also describes some applications of silver nanoparticles in different aspects such as antimicrobial, biomedicine, mosquito control, environment and wastewater treatment, agricultural, food safety, and food packaging.

Keywords: silver nanoparticles; green synthesis; plant metabolites

1. Introduction

Nanotechnology is one of the emergent cutting-edge technologies in a variety of different fields of science including biology, chemistry, and material science [1]. The continued utilization of nanotechnology for fabricating nano-scale products in research and development is growing [2,3]. Nanoparticles are small fragments which have a nano-scale dimension that ranges between 1–100 nm, with a very good thermal conductivity, catalytic reactivity, non-linear optical performance and chemical steadiness owing to their large surface area to volume ratio [4]. They are generally classified into different groups based on their sizes, shapes, and properties. The different groups include carbon-based nanoparticles, metal nanoparticles, ceramic nanoparticles, and polymeric nanoparticles and so many others [5]. The nanoparticles can be synthesized using several methods including chemical, physical, and biological methods. However, the chemical and physical methods are expensive, quite complicated, and potentially dangerous for the environment due to the toxic chemical compounds used as reducing agents [1,6].

In recent times, the biological method otherwise termed the green synthesis method has received increased attention due to the growing need to develop an environmentally benign technology in nanoparticle synthesis [7]. Due to its simple processes and cost effectiveness, both the research and industrial sectors are currently interested in green



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Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). production of silver nanoparticles for use in many applications in biomedicine, the environment, and industries [8]. Although there has been reported findings on the green synthesis of silver nanoparticles using several plants and microorganisms, the potential of plants and microorganisms as biological materials for the synthesis of silver nanoparticles is still yet to be fully explored.

The aim of this review was to examine the available published articles from 2001–2021 related to plant- and microorganism-mediated synthesis of silver nanoparticles, and the role of plant metabolites and applications. This review attempts to provide the general overview of various approaches of silver nanoparticles synthesis using green synthesis methods and the role of plant metabolites in the synthesis. The applications of silver nanoparticles in different aspects such as antimicrobes, biomedicine, mosquito control, environment controls, waste-water treatment, agricultural, food safety and food packaging will also be discussed. Lastly, the review provides direction and suggestions for future studies.

2. Methods of Silver Nanoparticles Synthesis

Silver nanoparticles (AgNPs) are synthesized using various physical, chemical, and biological techniques, which results in different shapes and sizes for use in numerous applications. These methods of synthesis are categorized into two main categories, namely "top-down" and "bottom-up" approaches. In the top-down approach, the size of silver metal in its bulk form is reduced mechanically to the nano-scale by using methods such as lithography, laser ablation, mechanical milling, etc. (Figure 1), whereas the bottom-up approach (self-assembly) involves the dissolution silver of salts in a solvent, reducing silver ions to their element with the use of a reducing agent, and then stabilizing the resulting neutral silver nanoparticles with stabilizing agents to prevent agglomeration [9,10]. A schematic diagram showing various top-down and bottom-up techniques for syntheses of silver nanoparticles is shown in Figure 1.

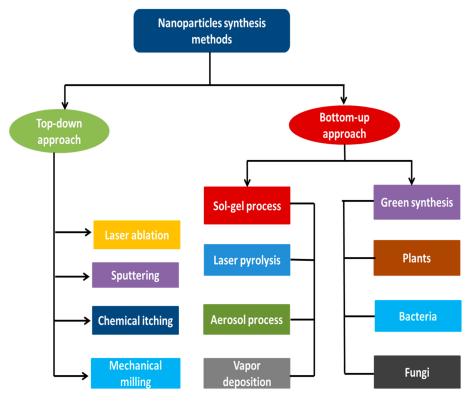


Figure 1. Different methods of synthesis of silver nanoparticles using the top-down and bottomup approaches.

2.1. Top-Down Approach

In this method of synthesis, a destructive approach is used to produce silver nanoparticles. It begins with breaking down a larger molecule which is decomposed into smaller units and then these units are converted into suitable nanoparticles. It is energy-demanding and requires extensive processing. The major advantage of this technique is the control of the size distribution and morphologies of nanoparticles [11]. Laser ablation, lithography, grinding/milling, spray pyrolysis, evaporation-condensation, and other decomposition processes are examples of this approach [12]. These approaches (laser ablation, lithography, grinding/milling, spray pyrolysis, evaporation-condensation) are also regarded as the physical synthesis methods. During these processes, the foundation material is placed in the middle of a furnace and vaporized into a carrier gas. The evaporation-condensation method has already been used to make carbon, lead, gold and silver nanoparticles [13]. Furthermore, a typical tube furnace takes several minutes of pre-heating time to achieve a constant working temperature, as well as electricity of several kilowatts. One of the limitations in this technique is the defects in the surface structure of the product, whereas physical properties of nanoparticles are dependent on the structure of the surface. The top-down approach was employed to synthesize colloidal carbon spherical particles with controlled sizes. The synthesis process was based on the continuous adsorption of chemical polyoxometalates on the interfacial surface of the carbon. Adsorption made the carbon black aggregates into relatively smaller spherical particles, with high dispersion capacity and narrow size distribution [14].

2.2. Bottom-Up Approach

The bottom-up technique is the reverse of the top-down method; it involves bringing together atoms and molecules to produce a diverse range of nanoparticles, therefore this approach is also called the building-up approach. Examples of this approach include sol-gel process, laser pyrolysis, aerosol process, chemical vapor deposition, and biological agents-assisted synthesis [15]. In this approach, nanoparticles can be synthesized using either chemical or biological methods by the self-assembly phenomenon of atoms to new nuclei which grows into nano-scaled particles [16]. The chemical reduction method is one of the methods used in this approach to synthesized silver nanoparticles. Various organic and inorganic agents such polyethylene glycol, sodium citrate, N,N-dimethylformamide, ascorbate, and sodium borohydride are used for the reduction of silver ions [17]. The green and biological bottom-up synthesis method of nanoparticles has drawn more attention in recent times by many researchers due to their less toxic effect and feasibility of the approach. These processes are also eco-friendly and cost-effective. In this approach nanoparticle synthesis is accomplished using biological systems such as using plant extracts such as bacteria, yeast, and fungi [18].

3. Syntheses of Silver Nanoparticles Using Plant Extracts

Synthesis of silver nanoparticles using plant materials is an important branch of biosynthesis processes. It has been known for a long time that plants have the potential to reduce metal ions both on their surface and in various organs and tissues remote from the ion penetration site [19]. The bioactive molecules found in plants include enzymes, proteins, amino acids, vitamins, polysaccharides, and organic acids such as citrates are potentially able to reduce metal ions [20].

In this regard, plant extracts are used for the bio-reduction of the metal ions to form their nanoparticles. The extract of various parts of plants such as leaves, flowers, seeds, barks, fruits, and roots have been applied for synthesis of silver nanoparticles and the plant extracts may serve as stabilizing and reducing agents in the synthesis process [21–23]. Several studies have been carried out on the synthesis of silver nanoparticles using plant extracts. Gardea et al., 2003 [24] was first to report the synthesis of metallic nanoparticles using alfalfa sprouts. The alfalfa roots have the ability to absorb silver (Ag) from agar medium and voyage them into shoots of plant in same oxidation state. In shoots, these

silver (Ag) atoms arranged themselves to produce silver nanoparticles (AgNPs). Ahmad and Sharma 2012 [6] also synthesized silver nanoparticles using Ananas comosus extract and characterized the synthesized silver nanoparticles using high resolution transmission electron microscopy (HRTEM), UV–Vis spectroscopy, energy dispersive X-ray spectroscopy (EDX), and selected area diffraction (SAD). The nanoparticles show a spherical shape with a diameter of 12 nm. In another study [25] silver nanoparticles were also synthesized using the leaf extract of Argemone mexicana that serve as reducing and capping agents. The properties of synthesized nanoparticles were analyzed using UV–Vis spectroscopy, X-ray diffraction (XRD), scanning electron microscopy (SEM), and Fourier transmission infrared spectrophotometer (FTIR). The average size of synthesized silver nanoparticles was 30 nm. Gavhane et al., 2012 [26] also synthesized silver by the reduction of aqueous AgNO₃ solution using the extract of neem and triphala. The properties of nanoparticles were analyzed using EDX, nanoparticles tracking analysis (NTA), and transmission electron microscope (TEM). NTA and TEM revealed that the spherical particle sizes were 43 nm and 59 nm. Velmuurugun et al., 2015 [27] also reported the synthesis of silver nanoparticles using peanut shell extract and their characteristics showed a spherical and oval shape with an average diameter up to 10–50 nm. Roy et al., 2014 [28] synthesized nanoparticles using the extract of Malus domestica with an average diameter of 20 nm. The nanoparticles were analyzed using UV–Vis spectroscopy, and their distinctive phases and morphology were confirmed using XRD and TEM, and FTIR was used to identify the biomolecules which are responsible for the reduction and stabilization of nanoparticles. *Polyalthia longifolia* leaf extract was also used as a reducing agent to synthesize silver nanoparticles [29]. Silver nanoparticles were also synthesized by Maqdoom et al., 2013 [30] using the fruit extract of Papaya and were characterized using absorption spectroscopy and FTIR. Spherical-shaped silver nanoparticles were synthesized using *Ocimum sanctum* leaf extract and characterized using UV–Vis spectroscopy, SEM, and XRD [31]. A leaf extract of Ceratonia siliqua was used to synthesize spherical silver nanoparticles with average particle sizes between 5–40 nm. UV–Vis spectroscopy, atomic absorption spectroscopy (AAS), XRD, SEM, and FTIR were used to characterize the nanoparticles. Jatropha curcas leaf extract was also used in syntheses involving the reduction of aqueous silver nitrate (AgNO₃) solution [21]. Silver nanoparticles were synthesized using the leaf extract of *Cassia auriculata*, and the leaf extract served as a reducing as well as a capping agent [32]. Shankar 2014 [33] synthesized silver nanoparticles using *Geranium* leaf extract. The silver nanoparticles synthesized were crystalline and stable with average size of size 40 nm. Stable and spherical shaped silver nanoparticles with average sizes between 10–50 nm were synthesized using the leaf extract of Ficus benghalensis. The characteristics of the synthesized nanoparticles were studied by FTIR, SEM, thermagravimetric analysis (TGA), and XRD [34]. Nakkala et al., 2014 [35] reported the syntheses of silver nanoparticles using Acorus calamus extract and evaluated its oxidation state, anti-cancer, and antibacterial effects. Another study by Kumar et al., 2014 [36] reported the syntheses of silver nanoparticles using the *Boerhaavia diffusa*. The synthesized nanoparticle showed an average size of 25 nm with a face-centered cubic geometry and spherical shape. The antimicrobial activity of synthesized silver nanoparticles was evaluated against Pseudomonas fluorescens, Aeromonas hydrophila, and Flavobacterium. Dwivedi et al., 2010 [37] also reported the syntheses of spherical shaped silver nanoparticles using the obnoxious weed Chenopodium album. The average nanoparticle sizes were between 10–30 nm as measured by TEM. In another study [38], Aldebasi et al. reported the synthesis of silver nanoparticles using *Ficus carica* leaf extract. Spherical shaped silver nanoparticles have been synthesized using the extract of Abutilon indicum and its antimicrobial activity was evaluated against Salmonella typhi, Escherichia coli, Salmonella aureus, and Bacillus subtilis microorganisms [39]. A similar study [40] by Awwad et al., reported on the syntheses of silver nanoparticles using the extract of Mulberry leaves. The 20 nm silver nanoparticles showed antimicrobial activity against *Staphylococcus aureus* and *Shigella* spp. Silver nanoparticles are synthesized by Awwad et al., 2012 [40] using the extract of Olea europaea and characterized by using SEM, XRD, and FTIR and yielded 15–40 nm particles

sizes. Silver nanoparticles were synthesized by the reduction of silver nitrate (AgNO₃) using the leaf extract of olive and the synthesized nanoparticles indicated effectiveness against drug-resistant bacteria isolates. The nanoparticles were characterized using UV–Vis spectroscopy, XRD, TGA, and SEM, and results showed that nanoparticles are mostly spherical with an average diameter between 20–25 nm [41]. *Alternanthera dentate* leaf extract was also used as a reducing and capping agent for synthesis of silver nanoparticles [42]. Studies by Murugan 2014 [43] reported that silver nanoparticles were synthesized using *Acacia leucophloea* extract with an average size that ranged between 38–72 nm. Another study by Arokiyaraj et al., 2014 [44] also reported that silver nanoparticles were synthesized using *Chrysanthemum indicum*, and nanoparticle sizes were 17–29 nm. Ashok et al., 2012 [45] and Kumar et al., 2013 [46] have synthesized silver nanoparticles using the leaf extract of *Parthenium hysterophorus*, *Premna herbacea*, and added to the aqueous solution of AgNO₃. The detail information on the plant and plant extracts used for the synthesis of silver nanoparticles are given in Table 1.

The use of plant extract in green synthesis has been well studied (presented in Table 1) and studies are still ongoing. Studies have demonstrated that the formation of metal nanoparticles using plant extracts can be completed in the metal salt solution at short duration of time at room temperature depending upon the nature of the plant extract. The main factors that can affect the formation of the nanoparticles are the concentration of the extract, temperature, metal salt, pH, and contact time [47]. The importance of using plants in the synthesis of nanoparticles is that plants are easy to obtain and all parts of plant (roots, latex, stem, seeds, and leaves) possess a large number of active agents that can be used in the reduction of Ag ion [48]. The active agents present in the plants which are responsible for the possible reduction of silver ion to silver nanoparticles includes terpenoids, polysaccharides, phenolics, alkaloids, flavones, amino acids, alcoholic compounds enzymes, and proteins [49,50]. Therefore, plant extracts in green synthesis were found to be ideal candidates for synthesizing silver nanoparticles due to their rapid growth, non-pathogenic and eco-friendly reaction conditions that occur in a single step using a highly economical protocol.

Plant Species	Part of Plant Used	Size (nm)	Shape	References
Nauclea latifolia	Fruit	10	Irregular	[51]
Citrus sinensis	Peels	10–70	Spherical	[52]
Cynara scolymus	Leaf extract	98.47 ± 2.04	Spherical	[53]
Alfalfa sprouts	Plant shoot	2–3	Icosahedral	[24]
Ananas comosus	Plant broth	12	Spherical	[6]
Argemone mexicana	Leaves extract	30	Spherical	[25]
Neem and Triphala	Leaves extract	43–59	Spherical	[26]
Peanut	Shell extract	10–50	Spherical/oval	[27]
Malus domestica	Leaf extract	20	Spherical	[28]
Polyalthia longifolia	Leaf extract	35–10	Spherical	[29]
Рарауа	Fruit extract	-	Spherical	[30]
Ocimum sanctum	Leaf extract	-	Spherical	[31]
Ceratonia siliqua	Leaf extract	5–40	-	[21]
Cassia auriculata	-	-	Spherical	[32]
Geranium spp.	Leaf extract	40	-	[33]
Ficus benghalensis	Leaf extract	10–50	Spherical	[34]

Table 1. Green synthesis of silver nanoparticles using plants.

Plant Species	Part of Plant Used	Size (nm)	Shape	References
Acorus calamus	Rhizome	31.83	Spherical	[35]
Boerhavia diffusa	-	25	-	[36]
Citrus limon	Peel	59	Spherical	[54]
Ananas comosus	Fruit	5–30	Spherical	[6]
Annona glabra	Leaf extract	10–100	Spherical	[55]

Table 1. Cont.

4. Synthesis of Silver Nanoparticles Using Bacteria

Microorganisms such as bacteria, are of great interest for nanoparticle synthesis, although this process is facing challenges, such as culture contamination, lengthy procedures and less control over nanoparticles size [12]. The bacteria are known to possess the extraordinary ability of reducing heavy metal ions and are therefore regarded as one of the best candidates for the synthesis of nanoparticles. Studies have reported that some bacteria species like Pseudomonas stutzeri and Pseudomonas aeruginosa have a developing ability to resort to specific defense mechanisms to overcome stresses like toxicity of heavy metal ions or metals and even survive and grow despite high metal ion concentrations [56]. Several studies have reported the synthesis of silver nanoparticles using bacteria (Table 2). Klaus et al. [57] reported the synthesis of silver nanoparticles with well-defined compositions and shapes using the *Pseudomonas stutzeri* strain. This study was one of the earliest studies that synthesized silver nanoparticles using microorganisms. Shivaji and Madhu 2011, [58] synthesized silver nanoparticles using culture supernatants of psychrophilic bacteria. Nanda and Sravanan 2009 [59] also studied the synthesis of silver nanoparticles using Staphylococcus aureus. A study by Kalimuthu et al., 2008 [60] also reported the synthesis of silver nanoparticles using the biomass of Bacillus licheniformis. It was reported that the size of the nanoparticles synthesis was between 40–50 nm, and these were stabilized using an enzyme nitrate. The summary of different bacteria strains used in synthesis of silver nanoparticles are shown in Table 2.

Table 2. Synthesis of silver nanoparticles using bacteria species.

Bacteria Species	Size(nm)	Shape	References
Escherichia coli	1.2–62	Spherical/quasi-spherical	[61]
Pseudomonas stutzeri	200	Triangles and hexagons	[57]
Serratia nematodiphila	65–70	Spherical shape	[62]
Bacillus stearothermophilus	42–92	Spherical	[63]
Lactobacillus casei	25–50	Spherical	[64]
Nocardiopsis spp.	45 ± 0.15	Spherical	[65]
Streptomyces hygroscopicus	20-30	-	[66]
Staphylococcus aureus	160–180	Irregular	[59]
Rhodococcus spp.	5–50	Spherical	[67]
Marine Ochrobactrum spp.	38–85	Spherical	[68]
Escherichia coli	1–100	Spherical	[69]
Lactobacillus strains	15–500	Triangular/hexagonal	[70]
Bacillus methylotrophicus	10–30	Spherical	[71]
Vibrio alginolyticus	50-100	Crystalline/spherical	[62]
Fusarium semitectum	1–50	Ellipsoid/spherical	[72]

5. Synthesis of Silver Nanoparticles Using Fungi

Fungi are one of the reducing and stabilizing agents used in the biosynthesis of silver nanoparticles. The fungi mediated synthesis of silver nanoparticles has attracted more attention because of its ability to produce commercial quantities of silver nanoparticles with controlled size, morphology, and low toxicity of residues [73]. In the synthesis of silver nanoparticles using microorganisms, fungi strains are preferred over bacteria species due to metal accumulation properties and better tolerance of the fungi [74,75]. The mechanism of synthesis of nanoparticles using fungi could be intracellular or extracellular. However, in the case of intracellular synthesis, the precursor metal is added to the mycelial culture and is co-opted in the fungal biomass. Consequently, the extraction of the nanoparticles is required after the synthesis using chemical treatment, centrifugation and filtration are employed to disrupt the biomass and release the nanoparticles [76]. In extracellular synthesis methods, the metal precursor is added to the aqueous filtrate that is made up of only the fungal biomass biomolecules, thereafter resulting in the formation of free nanoparticles. This method does not require any protocols to release the nanoparticles from the cells and is the most widely used method [77]. Balaji et al., 2009 [78] synthesized silver nanoparticles by extracellular synthesis methods using *Cladosporium cladosporioides*. The synthesized silver nanoparticles showed average sizes that range between 10–100 nm. Spherical shaped silver nanoparticles with an average size of 1–20 nm were synthesized using Aspergillus terreus [79]. Mukherjee et al., 2001 [80] also reported that mono-dispersed silver nanoparticles were synthesized by utilizing the fungus Verticillium by the green approach. The summary of different fungal strains used in syntheses of silver nanoparticles is shown in Table 3.

Fungi Species	Size (nm)	Shape	References
Aspergillus niger	1–20	Polydispersed spherical	[45]
Alternaria alternata	32	Spherical	[81]
Penicillium fellutanum	5–25	Spherical	[82]
Fusarium semitectum	10–60	Crystlline/spherical	[83]
Schizophyllum commune	51–93	Spherical	[84]
Endophytic fungus	10–25	Hexagonaerel/spherical	[85]
Trichoderma viride	5–40	Spherical	[86]
Pestalotia spp.	12	Polydispersed/spherical	[87]
Penicillium citrinum	109	Uniform spherical	[88]
Fusarium acuminatum	13	Spherical	[89]
Aspergillus niger	1–20	Polydispersed/spherical	[45]
Fusarium oxysporum	5–13	Spherical	[90]
Guignardia mangiferae	5–30	Spherical	[91]
Duddingtonia flagrans	30-409	Spherical	[77]
Arthroderma fulvum	21	Spherical	[92]

Table 3. Synthesis of silver nanoparticles using fungal species.

6. Plants Secondary Metabolites and Their Role in Synthesis of Nanoparticles

The crude extracts of different plants are known to contain wide ranges of primary and secondary metabolite compounds, ranging from proteins to different low molecular weight compounds such as phenolic acid, flavonoids alkaloid, terpenoids, amino acids, alcoholic compounds, glutathiones, polysaccharides, antioxidants, organic acids (ascorbic, oxalic, malic, tartaric, protocatechuic acid), and quinones. It is generally known that these metabolites are involved in redox reaction processes [93]. They are responsible for the reduction of metal ions into metallic nanoparticles. Although the elements participating in the green synthesis of nanoparticles and the underlying mechanism of the bio-reduction of ions is yet to be fully understood, it has been hypothesized that the bio-reduction of silver first involves the trapping of silver ions on the surface of proteins in the plant extract by means of electrostatic interactions [94]. The silver ions are then reduced by proteins, which lead to transformation of their secondary structure and the formation of silver nuclei. The silver nuclei later grow by further reduction of silver ions and their accumulation at the nuclei [95]. The involvement and participation of the secondary metabolites (sugars, terpenoids, polyphenols, alkaloids, phenolic acids, and proteins) in the reduction of metal ions leading to nanoparticle formation and in supporting their subsequent stability has also been postulated [19]. Based on available literature, flavonoid was one of the most frequently reported or predicted compounds responsible for the synthesis of silver nanoparticles in the green synthesis approach. Different plant species and plant metabolites responsible for the synthesis of silver nanoparticles reported in the literature are summarized in Table 4.

Plant Species	Metabolites Identified	References
Acalypha indica	Quercetin	[96]
Nigella arvensis	Flavonoids, alkaloids	[97]
Lantana camara	Flavonoids	[98]
Mimusops elengi	Polyphenols	[99]
Zingiber officinale	Flavonoid, alkaloids	[100]
Solanum xanthocarpum	Alkaloids, phenolic, sugars	[101]
Trianthema decandra	Saponin	[102]
Aegle marmelos	Tannin	[103]
Anacardium occidentale	Proteins, polyols	[104]
Desmodium triflorum	Ascorbic acid	[105]
Decalepis hamiltonii	Polyols, phenols	[106]
Syzygium cumini	Polyphenols	[107]
Azadirachta indica	Flavonoids, terpenoids	[108]
Coleus aromaticus	Flavonoids	[109]
Hibiscus rosa-sinensis	Carboxylate ion groups	[110]
Helianthus annuus	Flavonoids, proteins,	[98]
Dioscorea bulbifera	Flavonoids, polyphenols	[111]
Glycyrrhiza glabra	Flavonoids and terpenoid	[112]
Achyranthes aspera	Polyols	[113]

Table 4. Plant metabolites responsible for synthesis of silver nanoparticles in different plant species.

7. Applications of Silver Nanoparticles

Silver nanoparticles have been found to have a wide range of applications as antimicrobials in biomedicine, in environment and wastewater treatment, mosquito control, agriculture, food safety and food packaging, and so many other applications. A schematic diagram showing some applications of silver nanoparticles is shown in Figure 2.

7.1. Applications of Silver Nanoparticles as Antimicrobial Agents

Microorganisms are powerful and known to develop a resistance to different antibiotics. However, due to recent increases in bacterial resistance to various antibiotics, alternative therapeutic agents that are nontoxic to human beings but toxic to microorganisms are urgently required. Therefore, the development of nanoparticle mediated antimicrobial agents is most warranted. Recently, several studies have been focused on nanoparticle-based therapeutic agents against different bacteria species [114]. According to Ghaed et al., 2015 [115] silver nanoparticles have a strong toxicity to broad ranges of both gram negative and gram positive bacteria. However, the antimicrobial activity of silver nanoparticles (AgNPs) can be changed with physical properties such as size, shape, mass, and composition (pH, ions, and macromolecules) of the nanoparticle [22]. The shape of the nanoparticle can play a vital role in their antibacterial activity. For example, smaller silver nanoparticles may have greater binding surfaces and show more bactericidal activity compared to larger nanoparticles [116,117]. It has also been reported that the silver nanoparticles that range between 1–10 nm attach to the surface of the cell membrane and interact with the outer membrane of the bacteria, which arrest and disturb its proper functions such as respiration and permeability [118]. The antibacterial property of silver nanoparticles against Staphylococcus aureus, Pseudomonas aeruginosa and Escherichia coli has been investigated [119]. Recently, plant-based synthesized silver nanoparticles using Lysiloma acapulcensis have shown high antimicrobial potency against E. coli, P. aeruginosa, S. aureus and C. albicans compared to chemically produced silver nanoparticles. Furthermore, the study demonstrated that biogenic silver nanoparticles maintain lower cytotoxicity than the silver nanoparticles produced chemically [61].

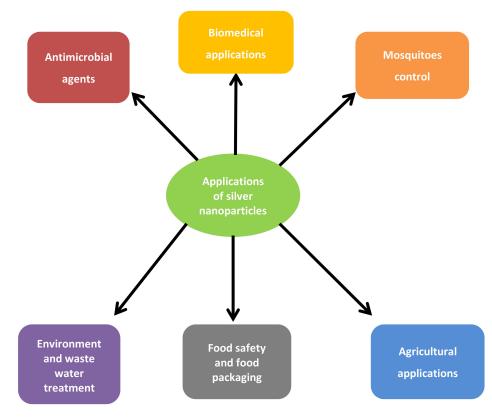


Figure 2. Applications of silver nanoparticles in different aspects.

7.2. Applications of Silver Nanoparticles in Biomedicine

The applications of silver nanoparticles in the field of biomedicine include drug delivery, cancer therapy, bio-imaging, dental technology, and many other applications. Recently, silver nanoparticles have attracted more attention in cancer therapeutics research because of their unique physical and chemical properties [120]. In comparison to available traditional anti-cancer therapies, metallic nanoparticles, especially silver nanoparticles, are considered as novel therapeutic agents or drug carriers in combination with drug candidates, and undesirable side-effects can be prevented by providing a targeted approach [121]. Previous studies have reported that silver nanoparticles exhibit good anti-cancer activities in different types of cancer, such as breast cancer, cervical, colon cancer, ovarian cancer, and lung cancer [122–126]. Furthermore, apart from anti-cancer therapy applications, nanoparticles have attracted more interest in the field of biomedicine because of their ability to deliver drugs in the optimum dosage range, often resulting in increased therapeutic efficiency of the drugs, weakened side effects and improved patient compliance [127]. Nanoparticles also have applications in cell bio-imaging and bio-sensing; however, the selection of the nanoparticles in question for achieving efficient contrast for cell imaging, biological application and photo thermal therapeutic applications is based on the optical properties of nanoparticles [128]. Silver nanoparticles also have applications in dentistry. It has been incorporated into some dental biomaterials for reducing biofilm formation due to its antibacterial activity. The silver nanoparticles joined into endodontic fillings showed a prolonged antibacterial impact against different strains of bacteria such as *Streptococcus milleri, Staphylococcus aureus* and *Enterococcus faecalis* [129]. The other applications of silver nanoparticles in biomedicine include wound dressings and catheters.

7.3. Applications of Nanoparticles in Environment and Waste Water Treatment

Metallic nanoparticles are one of the most attractive and cost-effective nanomaterials that have wide ranges of applications in the environment. The applications include environmental protection, water remediation, monitoring, purification, and treatment, and agriculture waste water treatment [3]. Clean and quality drinking water quality is one of the priorities of any nation. Plant extract-mediated green synthesized silver and gold nanoparticles can be employed for water treatment. For example, water and wastewater detoxification can be achieved by adsorption, photo-catalytic degradation, and nano-filtration techniques using nanoparticles as well [130]. Several studies have described the process of pesticides removal in water using gold and silver nanoparticles such as chlorprifos malathion and atrazine [131]. Moreover, the extraction of pesticides and other chemicals can be achieved by their adsorption onto nanoparticles which retain them on their surface, interacting for long periods until the complex precipitates. Therefore, these nanoparticles represent a suitable, convenient, and cost effective means of removing pesticides for either drinking water or irrigation projects. Water pollution with pathogenic microorganisms such as bacteria represent a high risk for water-borne diseases. The antimicrobial properties of metallic nanoparticles have been reported to be effective in this type of water purification [132].

7.4. Applications of Nanoparticles in the Control of Mosquitoes

The prevention and control of mosquitoes are important measures to control the spread of mosquito-borne diseases. The most commonly used method for the control of mosquitoes is chemical control. However, the persistent and repeated use of these synthetic chemical agents have caused the mosquito vector to become resistant, thus the need for the alternative new insecticides as novel biological tools for the control of mosquitoes [133]. Recently silver nanoparticles are emerging as one of the fastest growing materials due to their unique physical, chemical, and biological properties, small size and high specific surface area. Studies have indicated that synthesized silver nanoparticle using plants extracts of different plant species has shown to have tremendous mosquitocidal activity against various species of mosquitoes [133]. Moreover, nano-encapsulated pesticides and herbicides have shown enhanced properties in terms of solubility, specificity, permeability, and stability, as the nanostructure protects the active substance from early degradation and provides pest control for longer periods.

7.5. Applications of Nanoparticles in Agriculture

Nanotechnology in agriculture can provide wide ranges of applications for sustainable development through the development of nano-fertilizers, nano-pesticides, and nano-herbicides. Silver nanoparticle-based fertilizers have been developed in order to control nutrient release with plant uptake. This system helps to preserve or maintain soil fertility by reducing nutrient loss, contamination of ground water and soil, and chemical reactions between water, soil, and microbes that change them into unusable or dangerous chemicals for plants [134,135]. The control of pathogens such as bacteria and fungi responsible

for causing disease in plants can be achieved by the spraying of nanoparticle solutions directly on seeds, grains, or foliage to prevent the invasion of plant pathogens. The nanopesticides and nano-herbicides have shown enhanced properties in terms of solubility, stability, specificity, and permeability, as the nano-structure can protect and enhance the active substances from early degradation, and they provide active pest control ingredients for longer periods of time [3].

7.6. Applications of Nanoparticles in Food Safety and Food Packaging

The microbial contamination and spoilage of foods are major problems in food industries, considering the implication to public health due to food-borne diseases [136]. In order to minimize these problems, there is a need to develop active food packages with antimicrobial properties using active biocidal substances which may prevent food spoilage, microbial contamination, and increase the quality of the product. The materials used for packaging in food industries were organic acids, enzymes, and polymers (biodegradable and non-degradable). Recently, nanoparticles of metals or metallic oxides have been introduced with greater advantages compared with organic and inorganic acids, as they are resistant to the most severe processing conditions [136], such as exposure to high temperatures [137]. Therefore, the application of nanoparticles in the food packaging industry may offer potential solutions for the challenge presented by short shelf-life products, improving their quality and keeping them free of microbial adhesion. Metallic nanoparticles such as silver, magnesium oxide, copper oxide, zinc oxide, cadmium selenite/tellurite, titanium, and gold have been used in food industries due to their antimicrobial activity [138].

8. Conclusions

The green synthesis of silver nanoparticles offers a potentially ecofriendly, non-toxic, and cost-effective approach for the synthesis of nanoparticles. Different plant extracts and microorganisms can be used for the synthesis of silver nanoparticles. In the present review, the synthesis methods, applications, and role of plant metabolites in the synthesis of silver nanoparticles were highlighted. It is understood that different types of natural compounds present in plant extracts can act as reducing and stabilizing agents in the synthesis of silver nanoparticles. Plant-mediated silver nanoparticles are also stable due to the presence of natural capping agents such as proteins, which prevent the particles from aggregation. Furthermore, silver nanoparticles generated by green synthesis have potential applications, especially as antimicrobial agents of certain microorganisms for which their efficacy has been scientifically proven, in biomedicine as therapeutics agents, in mosquito control, in environment and wastewater treatment, in agriculture, in food safety, and in food packaging. Therefore, the green synthesis of silver nanoparticles using plant extracts has several advantages such as eco-friendliness, biocompatibility and cost-effectiveness. It is concluded that due to these unique properties, silver nanoparticles will have a key role in many of the nanotechnology-based processes.

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References

- 1. Pirtarighat, S.; Ghannadnia, M.; Baghshahi, S. Green synthesis of silver nanoparticles using the plant extract of Salvia spinosa grown in vitro and their antibacterial activity assessment. *J. Nanostruct. Chem.* **2019**, *9*, 1–9. [CrossRef]
- Albrecht, M.A.; Evans, C.W.; Raston, C.L. Green chemistry and the health implications of nanoparticles. *Green Chem.* 2006, 8, 417–432. [CrossRef]
- Castillo-Henríquez, L.; Alfaro-Aguilar, K.; Ugalde-Álvarez, J.; Vega-Fernández, L.; Montes de Oca-Vásquez, G.; Vega-Baudrit, J.R. Green Synthesis of Gold and Silver Nanoparticles from Plant Extracts and Their Possible Applications as Antimicrobial Agents in the Agricultural Area. *Nanomaterials* 2020, 10, 1763. [CrossRef]
- Agarwal, H.; Kumar, S.V.; Rajeshkumar, S. Resource-Efficient Technologies Review article A review on green synthesis of zinc oxide nanoparticles—An eco-friendly approach. *Resour. Technol.* 2017, *3*, 406–413.
- 5. Khan, I.; Saeed, K.; Khan, I. Nanoparticles: Properties, applications and toxicities. Arab. J. Chem. 2019, 12, 908–931. [CrossRef]
- Ahmad, N.; Sharma, S. Green Synthesis of Silver Nanoparticles Using Extracts of Ananas comosus. *Green Sustain. Chem.* 2012, 2, 141–147. [CrossRef]
- Sesuvium, L.; Nabikhan, A.; Kandasamy, K.; Raj, A.; Alikunhi, N.M. Synthesis of antimicrobial silver nanoparticles by callus and leaf extracts from saltmarsh plant, *Sesuvium portulacastrum* L. Asmathunisha. *Colloids Surf. B Biointerfaces* 2010, 79, 488–493.
- 8. McNamara, K.; Tofail, S.A.M. Nanoparticles in biomedical applications. *Adv. Phys.* 2017, *2*, 54–88. [CrossRef]
- 9. Tolaymat, T.M.; El Badawy, A.M.; Genaidy, A.; Scheckel, K.G.; Luxton, T.P.; Suidan, M. An evidence-based environmental perspective of manufactured silver nanoparticle in syntheses and applications: A systematic review and critical appraisal of peer-reviewed scientific papers. *Sci. Total Environ.* **2010**, *408*, 999–1006. [CrossRef] [PubMed]
- Leela, A.; Vivekanandan, M. Tapping the unexploited plant resources for the synthesis of silver nanoparticles. *Afr. J. Biotechnol.* 2008, 7, 3162–3165.
- 11. Wang, Y.; Xia, Y. Bottom-up and top-down approaches to the synthesis of monodispersed spherical colloids of low melting-point metals. *Nano Lett.* **2014**, *4*, 2047–2050. [CrossRef]
- 12. Rafique, M.; Sadaf, I.; Rafique, M.S.; Tahir, M.B. A review on green synthesis of silver nanoparticles and their applications. *Artif. Cells Nanomed. Biotechnol.* **2017**, 45, 1272–1291. [CrossRef]
- 13. Hurst, S.J.; Lytton-Jean, A.K.; Mirkin, C.A. Maximizing DNA loading on a range of gold nanoparticle sizes. *Anal. Chem.* **2006**, 78, 8313–8318. [CrossRef]
- 14. Garrigue, P.; Delville, M.H.; Labrugère, C.; Cloutet, E.; Kulesza, P.J.; Morand, J.P.; Kuhn, A. Top–down approach for the preparation of colloidal carbon nanoparticles. *Chem. Mater.* **2004**, *16*, 2984–2986. [CrossRef]
- 15. Iravani, S. Green synthesis of metal nanoparticles using plants. Green Chem. 2011, 13, 2638. [CrossRef]
- 16. Daniel, M.C.; Astruc, D. Gold nanoparticles: Assembly, supramolecular chemistry, quantum-size-related properties, and applications toward biology, catalysis, and nanotechnology. *Chem. Rev.* **2004**, *104*, 293–346. [CrossRef] [PubMed]
- 17. Singh, A.; Kaur, K. Biological and Physical Applications of Silver Nanoparticles with Emerging Trends of Green Synthesis. In *Synthesis of Silver Nanoparticles*; IntechOpen Limited: London, UK, 2018; pp. 15–20.
- 18. Parveen, K.; Banse, V.; Ledwani, L. Green synthesis of nanoparticles: Their advantages and disadvantages. *AIP Conf. Proc.* 2016, 1724, 020048.
- 19. Makarov, V.V.; Love, A.J.; Sinitsyna, O.V.; Makarova, S.S.; Yaminsky, I.V.; Taliansky, M.E.; Kalinina, N.O. "Green" nanotechnologies: Synthesis of metal nanoparticles using plants. *Acta Nat.* **2014**, *6*, 35–44. [CrossRef]
- 20. Mohammadlou, M.; Maghsoudi, H.; Jafarizadeh-Malmiri, H.J.I.F.R.J. A review on green silver nanoparticles based on plants: Synthesis, potential applications and eco-friendly approach. *Int. Food Res. J.* **2016**, *23*, 446–463.
- 21. Bar, H.; Bhui, D.K.; Sahoo, G.P.; Sarkar, P.; Pyne, S.; Misra, A. Green synthesis of silver nanoparticles using seed extract of Jatropha curcas. *Colloids Surf. A Physicochem. Eng. Asp.* 2009, 348, 212–216. [CrossRef]
- 22. Marambio-Jones, C.; Hoek, E.M. A review of the antibacterial effects of silver nanomaterials and potential implications for human health and the environment. *J. Nanopart Res.* **2010**, *12*, 1531–1551. [CrossRef]
- 23. Velayutham, K.; Rahuman, A.A.; Rajakumar, G.; Mohana, S.; Elango, G.; Kamaraj, C.; Marimuthu, S.; Iyappan, M.; Siva, C. Larvicidal activity of green synthesized silver nanoparticles using bark aqueous extract of Ficus racemosa against Culex quinquefasciatus and Culex gelidus. *Asian Pac. J. Trop. Med.* **2013**, *6*, 95–101. [CrossRef]
- 24. Gardea-Torresdey, J.L.; Gomez, E.; Peralta-Videa, J.R.; Parsons, J.G.; Troiani, H.; Jose-Yacaman, M. Alfalfa sprouts: A natural source for the synthesis of silver nanoparticles. *Langmuir* **2003**, *19*, 1357–1361. [CrossRef]

- Singh, A.; Jain, D.; Upadhyay, M.K.; Khandelwal, N.; Verma, H.N. Green synthesis of silver nanoparticles using Argemone mexicana leaf extract and evaluation of their antimicrobial activities. *Dig. J. Nanomater. Biosci.* 2010, 5, 483–489.
- Gavhane, A.J.; Padmanabhan, P.; Kamble, S.P.; Jangle, S.N. Synthesis of silver nanoparticles using extract of neem leaf and triphala and evaluation of their antimicrobial activities. *Int. J. Pharm. Bio. Sci.* 2012, *3*, 88–100.
- Velmurugan, P.; Sivakumar, S.; Young-Chae, S.; Seong-Ho, J.; Pyoung-In, Y.; Jeong-Min, S.; Sung-Chul, H. Synthesis and characterization comparison of peanut shell extract silver nanoparticles with commercial silver nanoparticles and their antifungal activity. J. Ind. Eng. Chem. 2015, 31, 51–54. [CrossRef]
- 28. Roy, K.; Sarkar, C.K.; Ghosh, C.K. Green synthesis of silver nanoparticles using fruit extract of Malus domestica and study of its antimicrobial activity. *Dig. J. Nanomater. Biostruct.* **2014**, *9*, 1137–1147.
- 29. Kaviya, S.; Santhanalakshmi, J.; Viswanathan, B. Green synthesis of silver nanoparticles using *Polyalthia longifolia* leaf extract along with D-sorbitol: Study of antibacterial activity. *J. Nanotechnol.* **2011**, 2011, 1–5. [CrossRef] [PubMed]
- Maqdoom, F.; Sabeen, H.; Zarina, S. Papaya fruit extract: A potent source for synthesis of bionanoparticle. *J. Environ. Res. Dev.* 2013, 7, 15–18.
- Rout, Y.; Behera, S.; Ojha, A.K.; Nayak, P.L. Green synthesis of silver nanoparticles using Ocimum sanctum (Tulashi) and study of their antibacterial and antifungal activities. J. Microbiol. Antimicrob. 2012, 4, 103–109. [CrossRef]
- 32. Udayasoorian, C.; Kumar, K.V.; Jayabalakrishnan, M. Extracellular synthesis of silver nanoparticles using leaf extract of Cassia auriculata. *Dig. J. Nanomater. Biostruct.* **2011**, *6*, 279–283.
- Shankar, S.; Jaiswal, L.; Aparna, R.S.L.; Prasad, R.G.S.V. Synthesis, characterization, in vitro biocompatibility, and antimicrobial activity of gold, silver and gold silver alloy nanoparticles prepared from *Lansium domesticum* fruit peel extract. *Mater. Lett.* 2014, 137, 75–78. [CrossRef]
- 34. Saware, K.; Sawle, B.; Salimath, B.; Jayanthi, K.; Abbaraju, V. Biosynthesis and characterization of silver nanoparticles using Ficus benghalensis leaf extract. *Int. J. Res. Eng. Technol.* **2014**, *3*, 868–874.
- 35. Nakkala, J.R.; Mata, R.; Gupta, A.K.; Sadras, S.R. Biological activities of green silver nanoparticles synthesized with Acorous calamus rhizome extract. *Eur. J. Med. Chem.* **2014**, *85*, 784–794. [CrossRef]
- Kumar, P.V.; Pammi SV, N.; Kollu, P.; Satyanarayana, K.V.V.; Shameem, U. Green synthesis and characterization of silver nanoparticles using Boerhaavia diffusa plant extract and their anti bacterial activity. Ind. Crop. Prod. 2014, 52, 562–566. [CrossRef]
- 37. Dwivedi, A.D.; Gopal, K. Biosynthesis of silver and gold nanoparticles using *Chenopodium album* leaf extract. *Physicochem. Eng. Asp.* **2010**, 369, 27–33. [CrossRef]
- 38. Aldebasi, Y.H.; Aly, S.M.; Khateef, R.; Khadri, H. Noble silver nanoparticles (AgNPs) synthesis and characterization of fig Ficus carica (fig) lea extract and its antimicrobial effect against clinical isolates from corneal ulcer. *Afr. J. Biotechnol.* **2015**, *13*, 275–281.
- 39. Ashokkumar, S.; Ravi, S.; Kathiravan, V.; Velmurugan, S. Synthesis of silver nanoparticles using A. indicum leaf extract and their antibacterial activity. *Spectrochim. Acta A Mol. Biomol. Spectrosc.* **2015**, *134*, 34–39. [CrossRef]
- Awwad, A.M.; Salem, N.M. Green synthesis of silver nanoparticles by mulberry leaves extract. *Nanosci. Nanotechnol.* 2012, 125–128. [CrossRef]
- Khalil, M.M.; Ismail, E.H.; El-Baghdady, K.Z.; Mohamed, D. Green synthesis of silver nanoparticles using olive leaf extract and its antibacterial activity. Arab. J. Chem. 2014, 7, 1131–1139. [CrossRef]
- 42. Kumar, B.; Smita, K.; Cumbal, L.; Debut, A.; Pathak, R.N. Sonochemical Synthesis of Silver Nanoparticles Using Starch: A Comparison. *Bioinorg. Chem. Appl.* 2014, 2014, 784268. [CrossRef]
- 43. Murugan, K.; Senthilkumar, B.; Senbagam, D.; Al-Sohaibani, S. Biosynthesis of silver nanoparticles using Acacia leucophloea extract and their antibacterial activity. *Int. J. Nanomed.* **2014**, *9*, 243–244.
- 44. Arokiyaraj, S.; Arasu, M.V.; Vincent, S.; Prakash, N.U.; Choi, S.H.; Oh, Y.K. Rapid green synthesis of silver nanoparticles from Chrysanthemum indicum L and its antibacterial and cytotoxic effects: An in vitro study. *Int. J. Nanomed.* **2014**, *9*, 379. [CrossRef]
- 45. Sagar, G.; Ashok, B. Green synthesis of silver nanoparticles using Aspergillus niger and its efficacy against human pathogens. *Eur. J. Exp. Biol.* **2012**, *2*, 654–658.
- 46. Kumar, P.; Selvi, S.S.; Govindaraju, M. Seaweed-mediated biosynthesis of silver nanoparticles using Gracilaria corticata for its antifungal activity against *Candida* spp. *Appl. Nanosci.* **2013**, *3*, 495–500. [CrossRef]
- 47. Mittal, A.K.; Chisti, Y.; Banerjee, U.C. Chisti Synthesis of metallic nanoparticles using plant extracts. *Biotechnol. Adv.* 2013, 31, 346–356. [CrossRef]
- Kharissova, O.V.; Dias, H.R.; Kharisov, B.I.; Pérez, B.O.; Pérez, V.M.J. The greener synthesis of nanoparticles. *Trends Biotechnol.* 2013, *31*, 240–248. [CrossRef] [PubMed]
- 49. Sharma, V.K.; Yngard, R.A.; Lin, Y. Silver nanoparticles: Green synthesis and their antimicrobial activities. *Adv. Colloid Interface Sci.* 2009, 145, 83–96. [CrossRef]
- 50. Bindhu, M.R.; Umadevi, M. Synthesis of monodispersed silver nanoparticles using Hibiscus cannabinus leaf extract and its antimicrobial activity. *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* 2013, 101, 184–190. [CrossRef]
- 51. Ayodele, M.; Chikodiri, V.; Adebayo-Tayo, B.C. Green synthesis and cream formulations of silver nanoparticles of Nauclea latifolia (*African peach*) fruit extracts and evaluation of antimicrobial and antioxidant activities. *Sustain. Chem. Pharm.* **2020**, 15, 100197.
- Carmalita, M.; Niluxsshun, D.; Masilamani, K.; Mathiventhan, U. Green Synthesis of Silver Nanoparticles from the Extracts of Fruit Peel of *Citrus tangerina*, *Citrus sinensis*, and *Citrus limon* for Antibacterial Activities. *Bioinorg. Chem. Appl.* 2021, 2021, 6695734.

- 53. Erdogan, O.; Abbak, M.; Demirbolat, M.; Id, F.B. Green synthesis of silver nanoparticles via Cynara scolymus leaf extracts: The characterization, anticancer potential with photodynamic therapy in MCF7 cells. *PLoS ONE* **2019**, *14*, e0216496.
- Alkhulaifi, M.M.; Alshehri, J.H.; Alwehaibi, M.A.; Awad, M.A.; Al-Enazi, N.M.; Aldosari, N.S.; Hatamleh, A.A.; Raouf, N.A. Green synthesis of silver nanoparticles using Citrus limon peels and evaluation of their antibacterial and cytotoxic properties. *Saudi J. Biol. Sci.* 2020, 27, 3434–3441. [CrossRef] [PubMed]
- Amarasinghe, L.D.; Wickramarachchi, P.A.S.R.; Aberathna, A.A.A.U.; Sithara, W.S.; De Silva, C.R. Comparative study on larvicidal activity of green synthesized silver nanoparticles and Annona glabra (Annonaceae) aqueous extract to control Aedes aegypti and Aedes albopictus (Diptera: Culicidae). *Heliyon* 2020, 6, e04322. [CrossRef]
- 56. Iravani, S. Bacteria in Nanoparticle Synthesis: Current Status and Future Prospects. Int. Sch. Res. Not. 2014, 2014, 1–18. [CrossRef]
- Klaus, T.; Joerger, R.; Olsson, E.; Granqvist, C.G. Silver-based crystalline nanoparticles, microbially fabricated Tanja. Proc. Natl. Acad. Sci. USA 1999, 96, 13611–13614. [CrossRef]
- Shivaji, S.; Madhu, S.; Singh, S. Extracellular synthesis of antibacterial silver nanoparticles using psychrophilic bacteria. *Process Biochem.* 2011, 46, 1800–1807. [CrossRef]
- Nanda, A.; Saravanan, M. Biosynthesis of silver nanoparticles from Staphylococcus aureus and its antimicrobial activity against MRSA and MRSE. *Nanomed. Nanotechnol. Biol. Med.* 2009, 5, 452–456. [CrossRef]
- 60. Kalimuthu, K.; Babu, R.S.; Venkataraman, D.; Bilal, M.; Gurunathan, S. Biosynthesis of silver nanocrystals by Bacillus licheniformis. *Colloids Surf. B Biointerfaces* **2008**, *65*, 150–153. [CrossRef]
- Garibo, D.; Borbón-Nuñez, H.A.; de León, J.N.D.; Mendoza, E.G.; Estrada, I.; Toledano-Magaña, Y.; Tiznado, H.; Marroquin, M.O.; Ramos, A.G.S.; Blanco, A.; et al. Green synthesis of silver nanoparticles using Lysiloma acapulcensis exhibit high—Antimicrobial activity. Sci. Rep. 2020, 10, 12805. [CrossRef]
- 62. Malarkodi, C.; Rajeshkumar, S.; Paulkumar, K.; Vanaja, M.; Jobitha, G.D.G.; Annadurai, G. Bactericidal activity of bio mediated silver nano-particles synthesized by Serratia nematodiphila. *Drug Invent. Today* **2013**, *5*, 119–125. [CrossRef]
- Hallol, M.M.A.M.A. Studies on Bacterial Synthesis of Silver Nanoparticles Using Gamma Radiation and Their Activity against Some Pathogenic Microbes. Master's Thesis, Cairo University, Cairo, Egypt, 2013.
- Korbekandi, H.; Iravani, S.; Abbasi, S. Optimization of biological syn- thesis of silver nanoparticles using *Lactobacillus casei* subsp. casei. *J. Chem. Technol. Biotechnol.* 2012, 87, 932–937. [CrossRef]
- 65. Manivasagan, P.; Venkatesan, J.; Senthilkumar, K.; Sivakumar, K.; Kim, S.K. Biosynthesis, antimicrobial and cytotoxic effect of silver using a novel *Nocardiopsis* sp. MBRC-1. *BioMed. Res. Int.* **2013**, 2013, 287638. [CrossRef] [PubMed]
- 66. Sadhasivam, S.; Shanmugam, P.; Yun, K. Biosynthesis of silver nanoparticles by Streptomyces hygroscopicus and antimicrobial activity against medically important pathogenic microorganisms. *Colloids Surf. B* **2010**, *81*, 358–362. [CrossRef] [PubMed]
- 67. Otari, S.V.; Patil, R.M.; Ghosh, S.J.; Thorat, N.D.; Pawar, S.H. Intracellular synthesis of silver nanoparticle by actinobacteria and its antimicrobial activity. *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* **2015**, *136*, 1175–1180. [CrossRef]
- Thomas, R.; Janardhanan, A.; Varghese, R.T.; Soniya, E.V.; Mathew, J.; Radhakrishnan, E.K. Antibacterial properties of silver nanoparticles synthesized by marine *Ochrobactrum* sp. *Braz. J. Microbiol.* 2014, 45, 1221–1227. [CrossRef] [PubMed]
- 69. Ghorbani, H.R. Biosynthesis of silver nanoparticles by Escherichia coli. Asian J. Chem. 2013, 25, 1247–1249.
- 70. Nair, B.; Pradeep, T. Coalescence of nanoclusters and formation of submicron crystallites assisted by Lactobacillus strains. *Crys. Growth Des.* **2002**, *2*, 293–298. [CrossRef]
- 71. Wang, C.; Kim, Y.J.; Singh, P.; Mathiyalagan, R.; Jin, Y.; Yang, D.C. Green synthesis of silver nanoparticles by Bacillus methylotrophicus, and their antimicrobial activity. *Artif. Cells Nanomed. Biotechnol.* **2015**, *15*, 10–11.
- 72. Shelar, G.B.; Chavan, A.M. Myco-synthesis of silver nanoparticles from Trichoderma harzianum and its impact on germination status of oil seed. *Biolife* **2015**, *3*, 109–113.
- Gade, A.K.; Bonde, P.P.; Ingle, A.P.; Marcato, P.D.; Duran, N.; Rai, M.K. Exploitation of Aspergillus niger for synthesis of silver nanoparticles. J. Biobased Mater. Bioenergy 2008, 2, 243–247. [CrossRef]
- 74. Ahmad, S.; Munir, S.; Ullah, A.; Khan, B. Green nanotechnology: A review on green synthesis of silver nanoparticles—An ecofriendly approach. *Int. J. Nanomed.* **2019**, *4*, 5087–5107. [CrossRef] [PubMed]
- Sastry, M.; Ahmad, A.; Mukherjee, P.; Senapati, S. Extracellular biosynthesis of silver nanoparticles using the fungus, Fusarium oxysporum. *Colloids Surf. B* 2003, 28, 313–318.
- Rajput, S.; Werezuk, R.; Lange, R.M.; Mcdermott, M.T. Fungal isolate optimized for biogenesis of silver nanoparticles with enhanced colloidal stability. *Langmuir* 2016, 32, 8688–8697. [CrossRef] [PubMed]
- Silva LP, C.; Oliveira, J.P.; Keijok, W.J.; da Silva, A.R.; Aguiar, A.R.; Guimarães, M.C.C. Extracellular biosynthesis of silver nanoparticles using the cell-free filtrate of nematophagus fungus Duddingtonia flagans. *Int. J. Nanomed.* 2017, 12, 6373. [CrossRef] [PubMed]
- Balaji, D.S.; Basavaraja, S.; Deshpande, R.; Mahesh, D.B.; Prabhakar, B.K.; Venkataraman, A. Extracellular biosynthesis of functionalized silver nanoparticles by strains of Cladosporium cladosporioides fungus. *Colloids Surf. B Biointerfaces* 2009, 68, 88–92. [CrossRef] [PubMed]
- Li, X.; Xu, H.; Chen, Z.S.; Chen, G. Biosynthesis of nanoparticles by microorganisms and their applications. J. Nanomater. 2011, 2011, 270974. [CrossRef]

- Mukherjee, P.; Ahmad, A.; Mandal, D.; Senapati, S.; Sainkar, S.R.; Khan, M.I.; Parishcha, R.; Ajaykumar, P.; Alam, M.; Kumar, R. Fungus-mediated synthesis of silver nanoparticles and their immobilization in the mycelial matrix: A novel biological approach to nanoparticle synthesis. *Nano Lett.* 2001, 1, 515–519. [CrossRef]
- 81. Gajbhiye, M.; Kesharwani, J.; Ingle, A.; Gade, A.; Rai, M. Fungus-mediated synthesis of silver nanoparticles and their activity against pathogenic fungi in combination with fluconazole. *Nanomed. Nanotechnol. Biol. Med.* **2009**, *5*, 382–386. [CrossRef]
- 82. Kathiresan, K.; Manivannan, S.; Nabeel, M.A.; Dhivya, B. Studies on silver nanoparticles synthesized by a marine fungus, Penicillium fellutanum isolated from coastal mangrove sediment. *Colloids Surf. B Biointerfaces* **2009**, *71*, 133–137. [CrossRef]
- 83. Basavaraja, S.; Balaji, S.D.; Lagashetty, A.; Rajasab, A.H.; Venkataraman, A. Extracellular biosynthesis of silver nanoparticles using the fungus Fusarium semitectum. *Mater. Res. Bull.* **2008**, *43*, 1164–1170. [CrossRef]
- Arun, G.; Eyini, M.; Gunasekaran, P. Green synthesis of silver nanoparticles using the mushroom fungus Schizophyllum commune and its biomedical applications. *Biotechnol. Bioprocess Eng.* 2014, 19, 1083–1090. [CrossRef]
- Verma, V.C.; Kharwar, R.N.; Gange, A.C. Biosynthesis of antimicrobial silver nanoparticles by the endophytic fungus Aspergillus clavatus. *Nanomedicine* 2010, 5, 33–40. [CrossRef] [PubMed]
- Fayaz, A.M.; Balaji, K.; Girilal, M.; Yadav, R.; Kalaichelvan, P.T.; Venketesan, R. Biogenic synthesis of silver nanoparticles and their synergistic effect with antibiotics: A study against gram-positive and gram-negative bacteria. *Nanomed. Nanotechnol. Biol. Med.* 2010, *6*, 103–109. [CrossRef] [PubMed]
- 87. Raheman, F.; Deshmukh, S.; Ingle, A.; Gade, A.; Rai, M. Silver nanoparticles: Novel antimicrobial agent synthesized from an endophytic fungus Pestalotia sp. isolated from leaves of *Syzygium cumini* (L.). *Nano Biomed. Eng.* **2011**, *3*, 174–178. [CrossRef]
- 88. Honary, S.; Barabadi, H.; Gharaei-Fathabad, E.; Naghibi, F. Green synthesis of silver nanoparticles induced by the fungus Penicillium citrinum. *Trop J. Pharm. Res.* **2013**, *12*, 7–11. [CrossRef]
- 89. Ingle, A.; Gade, A.; Pierrat, S.; Sonnichsen, C.; Rai, M. Mycosynthesis of silver nanoparticles using the fungus Fusarium acuminatum and its activity against some human pathogenic bacteria. *Curr. Nanosci.* **2008**, *4*, 141–144. [CrossRef]
- Husseiny, S.M.; Salah, T.A.; Anter, H.A. Biosynthesis of size controlled silver nanoparticles by Fusarium oxysporum, their antibacterial and antitumoral activities. *Beni Suef. Univ. J. Basic Appl. Sci.* 2015, 2, 225–231.
- 91. Balakumaran, M.D.; Ramachandran, R.; Kalaicheilvan, P.T. Exploitation of endophytic fungus, Guignardia mangiferae for extracellular synthesis of silver nanoparticles and their in vitro biological activities. *Microbiol. Res.* 2015, 178, 9–17. [CrossRef]
- 92. Xue, B.; He, D.; Gao, S.; Wang, D.; Yokoyama, K.; Wang, L. Biosynthesis of silver nanoparticles by the fungus Arthroderma fulvum and its antifungal activity against genera of Candida, Aspergillus and Fusarium. *Int. J. Nanomed.* **2016**, *11*, 1899–1906.
- Aromal, S.A.; Philip, D. Green synthesis of gold nanoparticles using Trigonella foenum-graecum and its size dependent catalytic activity. Spectrochim. Acta Part A Mol. Biomol. Spectrosc. 2012, 97, 1–5. [CrossRef] [PubMed]
- 94. Marslin, G.; Siram, K.; Id, Q.M.; Selvakesavan, R.K.; Kruszka, D.; Kachlicki, P.; Franklin, G. Secondary Metabolites in the Green Synthesis of Metalic Nanoparticles. *Materials* **2018**, *11*, 940. [CrossRef] [PubMed]
- 95. Li, S.; Shen, Y.; Xie, A.; Yu, X.; Qiu, L.; Zhang, L.; Zhang, Q. Green synthesis of silver nanoparticles using *Capsicum annuum* L. extract. *Green Chem.* **2007**, *9*, 852–858. [CrossRef]
- Krishnaraj, C.; Jagan, E.G.; Rajasekar, S.; Selvakumar, P.; Kalaichelvan, P.T.; Mohan, N. Synthesis of silver nanoparticles using Acalypha indica leaf extracts and its antibacterial activity against water borne pathogens. *Colloids Surf. B Biointerfaces* 2010, 76, 50–56. [CrossRef] [PubMed]
- Chahardoli, A.; Karimi, N.; Fattahi, A. Biosynthesis, characterization, antimicrobial and cytotoxic effects of silver nanoparticles using Nigella arvensis seed extract. *Iran. J. Pharm. Res.* 2017, *16*, 1167–1175. [PubMed]
- Ajitha, B.; Ashok Kumar Reddy, Y.; Shameer, S.; Rajesh, K.M.; Suneetha, Y.; Sreedhara Reddy, P. Lantana camara leaf extract mediated silver nanoparticles: Antibacterial, green catalyst. J. Photochem. Photobiol. B 2015, 149, 84–92. [CrossRef]
- Kiran Kumar, H.A.; Mandal, B.K.; Mohan Kumar, K.; Maddinedi, S.B.; Sai Kumar, T.; Madhiyazhagan, P.; Ghosh, A.R. Antimicrobial and antioxidant activities of Mimusops elengi seed extract mediated isotropic silver nanoparticles. *Spectrochim. Acta A Mol. Biomol. Spectrosc.* 2014, 130, 13–18. [CrossRef]
- Velmurugan, P.; Anbalagan, K.; Manosathyadevan, M.; Lee, K.J.; Cho, M.; Lee, S.M.; Park, J.H.; Oh, S.G.; Bang, K.S.; Oh, B.T. Green synthesis of silver and gold nanoparticles using Zingiber officinale root extract and antibacterial activity of silver nanoparticles against food pathogens. *Bioprocess Biosyst. Eng.* 2014, 37, 1935–1943. [CrossRef]
- 101. Sengottaiyan, A.; Mythili, R.; Selvankumar, T.; Aravinthan, A.; Kamala-Kannan, S.; Manoharan, K.; Thiyagarajan, P.; Govarthanan, M.; Kim, J.-H. Green synthesis of silver nanoparticles using *Solanum indicum* L. and their antibacterial, splenocyte cytotoxic potentials. *Res. Chem. Int.* 2016, 42, 3095–3103. [CrossRef]
- Geethalakshmi, R.; Sarada, D.V.L. Characterization and antimicrobial activity of gold and silver nanoparticles synthesized using saponin isolated from *Trianthema decandra* L. Ind. Crop. Prod. 2013, 75, 107–115.
- Jagajjanani Rao, K.; Paria, S. Green synthesis of silver nanoparticles from aqueous Aegle marmelos leaf extract. *Mater. Res. Bull.* 2013, 48, 628–634. [CrossRef]
- Mukunthan, K.S.; Balaji, S. Cashew apple juice (*Anacardium occidentale* L.) speeds up the synthesis of silver nanoparticles. *Int. J. Green Nanotechnol.* 2012, 4, 71–79. [CrossRef]
- 105. Ahmad, N.; Sharma, S.; Singh, V.N.; Shamsi, S.F.; Fatma, A.; Mehta, B.R. Biosynthesis of Silver Nanoparticles from Desmodium triflorum: A Novel Approach TowardsWeed Utilization. *Biotechnol. Biotechnol. Res. Int.* **2011**, *4*, 454090. [CrossRef] [PubMed]

- Rashmi, V.; Sanjay, K.R. Green synthesis, characterisation and bioactivity of plant-mediated silver nanoparticles using Decalepis hamiltonii root extract. *IET Nanobiotechnol.* 2017, 11, 247–254. [CrossRef] [PubMed]
- Kumar, V.; Yadav, S.C.; Yadav, S.K. Syzygium cumini leaf and seed extract mediated biosynthesis of silver nanoparticles and their characterization. J. Chem. Technol. J. Chem. Technol. Biotechnol. 2010, 85, 1301–1309. [CrossRef]
- Ahmed, S.; Ahmad, M.; Swami, B.L.; Ikram, S.; Saifullah. Green synthesis of silver nanoparticles using Azadirachta indica aqueous leaf extract. J. Radiat. Res. Appl. Sci. 2016, 9, 1–7. [CrossRef]
- 109. Vanaja, M.; Annadurai, G. Coleus aromaticus leaf extract mediated synthesis of silver nanoparticles and its bactericidal activity. *Appl. Nanosci.* **2013**, *3*, 217–223. [CrossRef]
- Nayak, D.; Ashe, S.; Rauta, P.R.; Nayak, B. Biosynthesis, characterisation and antimicrobial activity of silver nanoparticles using Hibiscus rosa-sinensis petals extracts. *IET Nanobiotechnol.* 2015, *9*, 288–293. [CrossRef] [PubMed]
- Ghosh, S.; Patil, S.; Ahire, M.; Kitture, R.; Kale, S.; Pardesi, K.; Cameotra, S.S.; Bellare, J.; Dhavale, D.D.; Jabgunde, A. Synthesis of silver nanoparticles using Dioscorea bulbifera tuber extract and evaluation of its synergistic potential in combination with antimicrobial agents. *Int. J. Nanomed.* 2012, 7, 483–496.
- 112. Dinesh, S.; Karthikeyan, S.; Arumugam, P. Biosynthesis of silver nanoparticles from Glycyrrhiza glabra root extract. *Arch. Appl. Sci. Res.* **2012**, *4*, 178–187.
- 113. Elumalai, D.; Kaleena, P.K.; Ashok, K.; Suresh, A.; Hemavathi, M. Green synthesis of silver nanoparticle using Achyranthes aspera and its larvicidal activity against three major mosquito vectors. *Eng. Agric. Environ. Food* **2016**, *9*, 1–8. [CrossRef]
- Beyth, N.; Houri-Haddad, Y.; Domb, A.; Khan, W.; Hazan, R. Alternative antimicrobial approach: Nano-antimicrobial materials. *Evid.-Based Complement Altern. Med.* 2015, 11, 11–22. [CrossRef] [PubMed]
- 115. Ghaedi, M.; Yousefinejad, M.; Safarpoor, M.; Khafri, H.Z.; Purkait, M.K. Rosmarinus officinalis leaf extract mediated green synthesis of silver nanoparticles and investigation of its antimicrobial properties. J. Ind. Eng. Chem. 2015, 31, 167–172. [CrossRef]
- Monteiro, D.R.; Gorup, L.F.; Takamiya, A.S.; Ruvollo-Filho, A.C.; de Camargo, E.R.; BDMonteiro, D.R.; Gorup, L.F. The growing importance of materials that prevent microbial adhesion: Antimicrobial effect of medical devices containing silver. *Int. J. Antimicrob. Agents* 2009, 34, 103–110. [CrossRef] [PubMed]
- 117. Kvitek, L.; Panacek, A.; Soukupova, J.; Kolar, M.; Vecerova, R.; Prucek, R.; Holecova, M.; Zbořil, R. Effect of surfactants and polymers on stability and antibacterial activity of silver nanoparticles (NPs). *J. Phys. Chem.* **2008**, *112*, 5825–5834. [CrossRef]
- Morones, J.R.; Elechiguerra, J.L.; Camacho, A.; Holt, K.; Kouri, J.B.; Ramírez, J.T.; Yacaman, M.J. The bactericidal effect of silver nanoparticles. *Nanotechnology* 2005, 16, 2346–2353. [CrossRef]
- 119. Rai, M.; Yadav, A.; Gade, A. Silver nanoparticles as a new generation of antimicrobials. Biotechnol. Adv. 2009, 27, 76–83. [CrossRef]
- 120. Shi, J.; Kantoff, P.W.; Wooster, R.; Farokhzad, O.C. Cancer nanomedicine: Progress, challenges and opportunities. *Nat. Rev. Cancer* **2017**, *17*, 20–37. [CrossRef]
- 121. Da Silva, P.B.; Machado, R.T.; Pironi, A.M.; Alves, R.C.; De Araújo, P.R.; Dragalzew, A.C.; Dalberto, I.; Chorilli, M. Recent Advances in the Use of Metallic Nanoparticles with Antitumoral Action-Review. *Curr. Med. Chem.* 2019, 26, 2108–2146. [CrossRef]
- Gurunathan, S.; Park, J.H.; Han, J.W.; Kim, J.H. Comparative assessment of the apoptotic potential of silver nanoparticles synthesized by Bacillus tequilensis and Calocybe indica in MDA-MB-231 human breast cancer cells: Targeting p53 for anticancer therapy. *Int. J. Nanomed.* 2015, 10, 4203–4222. [CrossRef]
- Al-Sheddi, E.S.; Farshori, N.N.; Al-Oqail, M.M.; Al-Massarani, S.M.; Saquib, Q.; Wahab, R. Anticancer Potential of Green Synthesized Silver Nanoparticles Using Extract of Nepeta deflersiana against Human Cervical Cancer Cells (HeLA). *Bioinorg. Chem. Appl.* 2018, 3, 939–945. [CrossRef]
- 124. Gurunathan, S.; Qasim, M.; Park, C.; Yoo, H.; Kim, J.H.; Hong, K. Cytotoxic Potential and Molecular Pathway Analysis of Silver Nanoparticles in Human Colon Cancer Cells HCT116. *Int. J. Mol. Sci.* **2018**, *19*, 2269. [CrossRef]
- 125. Yuan, Y.G.; Peng, Q.L.; Gurunathan, S. Silver nanoparticles enhance the apoptotic potential of gemcitabine in human ovarian cancer cells: Combination therapy for effective cancer treatment. *Int. J. Nanomed.* **2017**, *12*, 6487–6502. [CrossRef]
- Fard, N.N.; Noorbazargan, H.; Mirzaie, A.; Hedayati Ch, M.; Moghimiyan, Z.; Rahimi, A. Biogenic synthesis of AgNPs using Artemisia oliveriana extract and their biological activities for an effective treatment of lung cancer. *Artif. Cells Nanomed. Biotechnol.* 2018, 46, S1047–S1058. [CrossRef] [PubMed]
- 127. Alexis, F.; Pridgen, E.; Molnar, L.K.; Farokhzad, O.C. Factors affecting the clearance and biodistribution of polymeric nanoparticles. *Mol. Pharm.* **2018**, *5*, 505–515. [CrossRef] [PubMed]
- 128. Jain, K.; Mehra, N.K.; Jain, N.K. Potentials and emerging trends in nanopharmacology. *Curr. Opin. Pharmacol.* **2014**, *15*, 97–106. [CrossRef]
- 129. Lahiri, D.; Nag, M.; Sheikh, H.I.; Sarkar, T.; Edinur, H.A.; Pati, S.; Ray, R.R. Microbiologically-Synthesized Nanoparticles and Their Role in Silencing the Biofilm Signaling Cascade. *Front. Microbiol.* **2021**, *12*, 180. [CrossRef]
- 130. Sheng, Z.; Liu, Y. Effects of silver nanoparticles on wastewater biofilms. Water Res. 2011, 45, 6039–6050. [CrossRef] [PubMed]
- 131. Saifuddin, N.; Nian, C.Y.; Zhan, L.W.; Ning, K.X. Chitosan-silver Nanoparticles Composite as Point-of-use DrinkingWater Filtration System for Household to Remove Pesticides in Water. *Asian J. Biochem.* **2011**, *6*, 142–159. [CrossRef]
- Morsi, R.E.; Alsabagh, A.M.; Nasr, S.A.; Zaki, M.M. Multifunctional nanocomposites of chitosan, silver nanoparticles, copper nanoparticles and carbon nanotubes for water treatment: Antimicrobial characteristics. *Int. J. Biol. Macromol.* 2017, 97, 264–269. [CrossRef] [PubMed]

- 133. Balakrishnan, S.; Srinivasan, M. Biosynthesis of silver nanoparticles from mangrove plant (*Avicennia marina*) extract and their potential mosquito larvicidal property. *J. Parasit. Dis.* **2016**, *40*, 991–996. [CrossRef] [PubMed]
- Martínez-Fernández, D.; Barroso, D.; Komárek, M. Root water transport of *Helianthus annuus* L. under iron oxide nanoparticle exposure. *Environ. Sci. Pollut. Res.* 2016, 23, 1732–1741. [CrossRef] [PubMed]
- Panpatte, D.G.; Jhala, Y.K.; Shelat, H.N.; Vyas, R.V. The Next Generation Technology for Sustainable Agriculture. In *Microbial Inoculants in Sustainable Agricultural Productivity*; Springer: New Delhi, India, 2016; Volume 2, pp. 289–300.
- 136. Carbone, M.; Donia, D.T.; Sabbatella, G.; Antiochia, R. Silver nanoparticles in polymeric matrices for fresh food packaging. *J. King Saud Univ. Sci.* 2016, *28*, 273–279. [CrossRef]
- 137. Emamifar, A.; Kadivar, M.; Shahedi, M.; Solimanian-Zad, S. Effect of nanocomposite packaging containing Ag and ZnO on reducing pasteurization temperature of orange juice. *J. Food Process. Preserv.* **2012**, *34*, 104–112. [CrossRef]
- 138. AbdelRahim, K.; Mahmoud, S.Y.; Ali, A.M.; Almaary, K.S.; Mustafa AE ZM, A.; Husseiny, S.M. Extracellular biosynthesis of silver nanoparticles using Rhizopus stolonifer. *Saudi J. Biol. Sci.* **2017**, *24*, 208–216. [CrossRef]