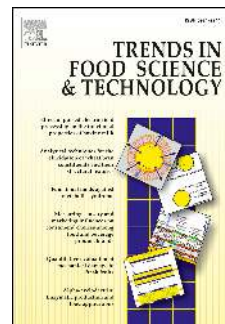


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1 **A review on the effects of light-emitting diode (LED) light on the nutrients of**
2 **sprouts and microgreens**

3

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13

14 **Abstract:**

15 **Background:** Sprouts and microgreens, which are tender, flavourful, rich in nutrients,
16 have a short growth cycle, and have been recognized as functional foods in the human
17 diet. Culturing under artificial light sources could regulate the growth, the
18 phytochemical compound content and antioxidant capacity of sprouts and microgreens.

19 **Scope and Approach:** In this review, the effects of light-emitting diode (LED) on
20 growth, phytochemical compound content and antioxidant capacity, as well as the
21 post-harvest quality of sprouts and microgreens were overviewed, and the underlying
22 mechanisms were discussed. The future applications and research, which aim to
23 improve the growth and nutritional quality of sprouts and microgreens, were also
24 investigated.

25 **Key Findings and Conclusions:** LED light can promote the accumulation of different
26 phytochemicals, such as phenolic compounds, vitamins, glucosinolates, chlorophyll
27 and carotenoids. Meanwhile, the antioxidant capacity could also be significantly
28 increased by growth under LED light, in particular UV-B light. The accumulation of
29 mineral elements (Ca^{2+} , Fe^{2+} , K^+) increased after light exposure. The effects of LED
30 light on the growth was species dependent. Therefore, growth under LED light is an
31 efficient and promising strategy for producing sprouts and microgreens with higher
32 nutritional values.

33

34 **Keywords:** sprouts; microgreens; light-emitting diode (LED); nutrition quality; light;
35 functional foods

36

37

38 1. Introduction

39 Vegetables are rich in phytonutrients and dietary fibers, which are indispensable
40 and beneficial for the human body. Recent researches have shown that regular intake
41 of vegetables is associated with a reduced risk of illnesses, such as cardiovascular
42 disease and cancer (Aune, et al., 2017; Moore & Thompson, 2015). Over the past two
43 decades, the interest in fresh, nutritional and organic vegetables has increased as living
44 standards have risen. In such cases, sprouts and microgreens are gaining increasing
45 popularity. According to the recently published papers, the definition of sprouts is “the
46 product obtained from the germination of seeds and their development in water or
47 another medium, harvested before the development of true leaves and which is intended
48 to be eaten whole, including the seed” (Di Gioia, Renna, & Santamaria, 2017). While,
49 microgreens are defined as “tender immature greens, produced from the seeds of
50 vegetables and herbs, having two fully developed cotyledon leaves with or without the
51 emergence of a rudimentary pair of first true leaves” (Xiao, Lester, Luo, & Wang,
52 2012). There is a large variety of sprouts and microgreens, among which legumes (e.g.
53 soybean, pea, cowpea, etc.) and *Brassica* species (e.g. radish, broccoli, red pak choi and
54 buckwheat, etc.) sprouts and microgreens are the most commonly consumed.

55 One common feature of sprouts and microgreens is that they are both young and
56 tender edible seedlings produced from the seeds of vegetables, herbs, or grains (The
57 production methods of sprouts and microgreens were summarised in Supplemental
58 methods and Figure 1) (Renna, Castellino, Leoni, Paradiso, & Santamaria, 2018).
59 During seed germination, a series of physiological and biochemical processes occur:
60 (1) the imbibed seeds germinate, the radicle and hypocotyl elongate and the cotyledon
61 expands; (2) the content of anti-nutritional factors decreases (Bora, 2014); (3)

62 macromolecules (such as polysaccharides and fats) are transformed into small
63 molecules (such as oligosaccharides and free amino acids), which increases their
64 digestibility (Márton, Mándoki, Csapókiss, & Csapó, 2010); and (4) the content of
65 bioactive phytochemicals and the antioxidant capacity increases (Di Gioia, Renna, &
66 Santamaria, 2017). The nutrients of sprouts and microgreens include, but are not
67 limited to, proteins, vitamins, phenolics, carotenoids, glucosinolate and minerals (Di
68 Gioia, Renna, & Santamaria, 2017; Ebert, 2012). It is worth noting that the content of
69 bioactive compounds in sprouts and microgreens are higher than those of their mature
70 counterparts (Kyriacou, et al., 2016).

71 Light is one of the most important environmental factors for plants, as it provides
72 not only the source of energy for photosynthesis but also the signal for a multitude of
73 physiological responses. Light quality (wavelength), light quantity (intensity),
74 direction and photoperiod (duration) are key components of light conditions (Ding, et
75 al., 2011; Kami, Lorrain, Hornitschek, & Fankhauser, 2010). The use of artificial light
76 sources (e.g. fluorescent lamps, halogen light, LED light and high-pressure sodium
77 lamps), which emit photons over a spectral range from 250 nm to 750 nm, make the
78 study of the effects of light on the nutrient quality of sprouts and microgreens more
79 convenient and the results more conclusive. Recently, an increased number of studies
80 have demonstrated the beneficial effects of LED light on plant growth and quality of
81 crops, including the accumulation of phytonutrients in sprouts and microgreens
82 (Ciska, Honke, & Kozłowska, 2008; Peng, Zou, Su, Fan, & Zhao, 2015;
83 Pérez-Balibrea, Moreno, & García-Viguera, 2008; Qian, et al., 2016; Samuolienė

84 Giedrè, et al., 2017). Compared with conventional light sources, LED light offers
85 cheap, cool and controllable sources of light that can selectively and quantitatively
86 provide different spectra. LEDs provide photons that can activate discrete
87 developmental pathways to change plant growth (e.g. leaf area, thickness, stem length)
88 and quality (e.g. metabolites) through photoreceptors include phytochrome and
89 cryptochrome (Folta & Carvalho, 2015; Folta & Childers, 2008; Heijde & Ulm, 2012;
90 Quail, 2002; Smith, 2000). Therefore, this provides us with a new opportunity to
91 manipulate the quality and quantity of vegetable products for markets and meet the
92 demands of retailers.

93 Here we have reviewed the most relevant progressive studies, from 2005 to date,
94 investigating the regulation of growth and nutrient quality of sprouts and microgreens
95 grown under light-controlled environments. The objective of this study was to 1)
96 summarise the effects of LED light (including other artificial lights) on growth,
97 phytochemical compound content and antioxidant capacity of sprouts and
98 microgreens; 2) discuss the underlying mechanisms; 3) assess the application
99 potential and prospect of LED light in the production of sprouts and microgreens.

100 **1.1 The artificial lighting systems in sprouts and microgreens production**

101 The main light sources in sprouts or microgreen production are artificial lights,
102 including high-pressure sodium lamps (HPSs), fluorescent lamps (FLs) and
103 light-emitting diodes (LEDs), among others (Bantis, et al., 2018). HPSs, with their
104 high electrical efficiencies, long operating life and a wide spectrum of light, are the
105 most common artificial light sources used for the culture of many plant species in

106 greenhouses (Islam, et al., 2012). FLs are widely used in growth chambers. When
107 used as additional light sources, FLs can provide sustained photosynthetic photon
108 fluence which is necessary for high productivity (Darko, Heydarizadeh, Schoefs, &
109 Sabzalian, 2014). However, HPSs and FLs both have serious limitations. For example,
110 HPSs require high voltage and emit intense radiant heat, in addition, they also contain
111 only 5% blue light, which is low compared to natural sunlight (18% blue light) (Islam,
112 et al., 2012). While FLs have limited photon output, less than ideal energy conversion
113 to light and a short effective lifespan (Rehman, et al., 2017). These features bring
114 several problems, in respect to the way the plants grow and develop, or overheating
115 due to infrared radiation (D'Souza, Yuk, Khoo, & Zhou, 2015). In contrast, LEDs
116 possess unique properties that are highly suitable for horticulture, including low
117 radiant heat emissions; high emissions of monochromatic light; photon efficiency;
118 long life expectancy, and flexibility, as well as the ability to be pulsatate (D'Souza, Yuk,
119 Khoo, & Zhou, 2015; Yeh & Chung, 2009). Therefore, LEDs have received
120 widespread attention and have been widely used in plant factory and other controlled
121 environment chambers.

122 **1.2 The LED light spectra and plant photoreceptors**

123 The range of wavelengths which modify the behavior of plants ranges from
124 ultraviolet (UV) light (~280 nm) to far-red light (700–750 nm) (Figure 2) (Kami, et
125 al., 2010; Rizzini, et al., 2011). Photoreceptors, which can sense a range of light
126 wavelengths, help plants to sense and utilize radiation from near UV (~350nm)
127 through the blue to red/far-red (~750nm) light (Möglich, Yang, Ayers, & Moffat,

128 2010). Blue (~450nm) and red (~650nm) light are strongly absorbed by plants, while
129 green (~530 nm) and far-red (~735 nm) light are reflected a large extent (Kami, et al.,
130 2010). Red/far-red light receptors phytochromes, UV-A/blue light receptors
131 phototropins, cryptochromes and ZEITLUPE family proteins, as well as UV-B
132 receptors UV RESPONSE LOCUS 8 (UVR8), form the three main systems that
133 respond to light signals in plants (Chen, Chory, & Fankhauser, 2004; Rizzini, et al.,
134 2011). In plants, there are two interconvertible forms of phytochromes: a red
135 light-absorbing Pr form and a far-red light-absorbing Pfr form (Chen, Chory, &
136 Fankhauser, 2004). Phytochromes have roles in seed germination, shade-avoidance
137 responses, seedling development and floral induction (Mazzella, Cerdán, Staneloni, &
138 Casal, 2001; Neff, Fankhauser, & Chory, 2000). In *Arabidopsis thaliana*,
139 phototropins are responsible for phototropism (Inoue, Kinoshita, Takemiya, Doi, &
140 Shimazaki, 2008), while cryptochromes are responsible for flowering, the entrainment
141 of the circadian clock, the regulation of stomatal opening and root development (Li &
142 Yang, 2007). UVR8 regulates a range of UV-B responses, including flavonoid
143 biosynthesis, hypocotyl growth inhibition and leaf cell expansion (Jenkins, 2014).
144 Although the specific green light photoreceptor remains to be identified in higher
145 plants, the effects of green light on plant growth should not be ignored. Increasing
146 evidences have shown the role of green light in photosynthesis and metabolism in
147 plants. For instance, addition of small portion of green light (~20%) has been shown
148 to enhance plant growth as green light penetrates deeper into the plant canopy and
149 enhances photosynthesis at lower leaf levels (Kim, Goins, Wheeler, & Sager, 2004).

150 Therefore, an intricate photosensory system may work in conjunction to control plant
151 physiological responses in response to spectral quality, intensity and duration (Neff,
152 Fankhauser, & Chory, 2000). The downstream signalling components that interact
153 with the photoreceptors remained to be gradually identified.

154 LEDs emit light with a narrow-band wavelength, which depends on the
155 properties of the semiconductor materials of the devices. With the improvement of
156 semiconductor technology, the peak emission wavelength of LED ranges from UV
157 (~250 nm) to infrared (~1000 nm) (Olle & Viršile, 2013). In other words, LED light
158 could emit both broad-band light (white light) and monochromatic light (e.g., UV,
159 blue, green, red and far-red light). When multiple LEDs are combined,
160 monochromatic lights with different intensities or a combination of light with
161 different spectral compositions could be emitted. Therefore, the accurate and flexible
162 light spectra control ability of LEDs allows it to provide optimal light wavelengths
163 that match the plant photoreceptors and photosynthetic pigments, and may contribute
164 to optimize plant growth and metabolism.

165 **1.3 Light spectra and intensity change photosynthesis**

166 Photosynthesis is a process to convert light energy (photons) to biomass in plant,
167 in which light intensity and spectra play fundamental roles. The highest
168 photosynthesis and productivity of plants could be achieved at appropriate light
169 intensity. When light intensity is lower than a certain compensation intensity,
170 photosynthesis will be exceeded by respiration, and the plants will become a net
171 consumer of oxygen. However, the photosynthesis and even plant growth could be

172 damaged with the increase of light intensity, due to the photoinhibition (Ooms, Dinh,
173 Sargent, & Sinton, 2016).

174 The most important light region for photosynthesis is 400 to 700 nm, known as
175 photosynthetically active radiation (PAR). Due to the emitting of narrow wave band
176 lights, the flexibility to control light spectral composition and the high PAR
177 conversion efficiency (Both, et al., 2017), LEDs are perceived as tailor-made light
178 sources and widely employed in analyzing photosynthesis under different light spectra.
179 It has long been known that blue and red light regions are most efficiently absorbed
180 by chlorophylls, the primary photosynthetic pigments, during the photosynthetic
181 processes (Chen & Blankenship, 2011). Therefore, red and blue lights are currently
182 two types of light spectra that have been most studied on plant photobiology . Red
183 light was reported to promote photosynthesis and vegetative growth by increasing the
184 content of chlorophyll, promoting the formation of photosynthetic apparatus and
185 probably by inducing stomatal opening (Zhu, Geng, Chakravorty, Guan, Chen, &
186 Assmann, 2019; Wang, Lu, Tong, & Yang, 2016). However, prolonged red light
187 illumination may result in 'red light syndrome', which is characterized by low
188 photosynthetic capacity, low maximum quantum yield of chlorophyll fluorescence
189 (F_v/F_m), low carbohydrate content accumulation and impaired growth (Miao, Chen,
190 Qu, Gao, & Hou, 2019)). Blue light, which is strongly absorbed by carotenoid
191 pigments (lutein and β -carotene), was reported to increase the chlorophyll content and
192 chlorophyll a/b ratio, promote stomatal opening, control the integrity of chloroplast
193 protein, and enhance F_v/F_m (Huché-Théliier, et al., 2016) . Meanwhile, many studies

194 showed that an optimized red: blue light ratio may be more beneficial for
195 photosynthesis. For instance, the net photosynthetic rate increased upon the decrease
196 of the red: blue ratio (Nanya, Ishigami, Hikosaka, & Goto, 2012), and the impaired
197 photosynthetic parameters and chloroplast development induced by red light could be
198 alleviated by adding blue light (Miao, Chen, Qu, Gao, & Hou, 2019).

199 On the other hand, other lights with wavelengths outside the red and blue light regions
200 were also reported to play considerable roles in photosynthesis, which was once
201 neglected. For example, green light was once ignored because it is absorbed weakly by
202 the chlorophylls. The literature recently showed that green light could penetrate the leaf
203 further than blue and red light, increasing carbon fixation and maybe yield (Terashima,
204 Fujita, Inoue, Chow, & Oguchi, 2009; Terashima, Fujita, Inoue, Chow, & Oguchi,
205 2009; Smith, Mcausland, & Murchie, 2017). Moreover, green light could also reverse
206 UV-B and blue light-mediated stomatal opening (Smith, Mcausland, & Murchie,
207 2017). Similarly, far-red light has long been considered as photosynthetically
208 inefficient radiation, due to its poor ability in driving photosynthetic reaction.
209 However, far-red light was now reported to be more active in enhancing photochemical
210 efficiency than commonly believed, because of the synergistic effect between far-red
211 light and light with shorter wavelengths (Zhen & van Iersel, 2017; Zhen, Haidekker, &
212 van Iersel, 2019). UV light was traditionally considered harmful for photosynthesis,
213 while the recent studies have changed this stereotype. Although, high UV radiation
214 could impair the biosynthesis of carotenoids and damage the photosystem II, a
215 moderate UV dose might have positive effects on photosynthesis by inducing the
216 biosynthesis of flavonoids with photoprotective activity (Johnson & Day, 2010;
217 Verdagner, Jansen, Llorens, Morales, & Neugart, 2017; Guidi, et al., 2016).

218 Understanding the physiological responses induced by other lights (not only red and
219 blue light) will expand our knowledge about how different portions of the light
220 spectrum independently or cooperatively regulate plant morphogenesis, which can lead
221 to the optimization of crop growth.**2. The effects of LED light on the growth of**
222 **sprouts and microgreens**

223 Photomorphogenic process, which is a default developmental process for the
224 light-grown seedlings, is triggered by light in sprouts and microgreens during their
225 development from seeds to edible vegetable products. Increasing numbers of studies
226 have revealed the vital role of light signals on the growth of sprouts and microgreens
227 (Table S1). Hypocotyls are one of the main edible parts of sprouts and microgreens,
228 and the growth of hypocotyls could be affected by artificial lights. For instance, FLs
229 light exposure was reported has negative effects on hypocotyl elongation. Compared
230 with the dark conditions, illuminating with FLs light significantly reduced hypocotyl
231 length of tartary buckwheat sprouts and soybean sprouts by 40% and 16%,
232 respectively (Peng, Zou, Su, Fan & Zhao, 2015; Yuan, et al., 2015a). Similarly, FLs
233 light illuminated bean sprouts, such as dolichos and cowpea sprouts, have been
234 reported to have decreased radical length when compared with their counterparts
235 grown in the dark (Martín-Cabrejas, et al., 2008). However, FLs light could
236 significantly increase the diameter of soybean sprout hypocotyls by 12%, compared
237 with the sprouts grown in the dark (Yuan, et al., 2015a). The results also showed that
238 sprouts and microgreens grown in the dark show etiolation phenotype, with white and
239 long hypocotyls. When grown under light, the shorten hypocotyl of light-grown

240 sprouts and microgreens may be due to the inactivation of COP1 (CONSTITUTIVE
241 PHOTOMORPHOGENIC 1), a repressor for photomorphogenesis, and the
242 stabilization of HY5 (LONG HYPOCOTYL 5) and HYH (LONG HYPOCOTYL 5
243 HOMOLOG), transcription factors that promote photomorphogenesis (Deng, Caspar,
244 & Quail, 1991). Furthermore, plant hormones, such as gibberellins and ethylene, may
245 also contribute in mediating light-regulated hypocotyl elongation (Alabadí, et al.,
246 2008; Yu & Huang, 2017). Compared with red and blue combined LED light, LED
247 blue light alone could significantly increase hypocotyl length of buckwheat sprouts
248 (Lee, et al., 2014). Similarly, supplementation of LED far-red light on the basis of
249 LED red + blue light could significantly increase the hypocotyl length in kohlrabi
250 microgreens and mustard microgreens (Gerovac, Craver, Boldt, & Lopez, 2016). UV
251 light has been reported to suppress the growth of seedlings (Huché-Thélier, et al.,
252 2016), while Brazaitytė et al. (2015a) reported that supplemental LED UV-A (402 nm)
253 irradiation significantly increased the hypocotyl length of basil, beet and pak choi
254 microgreens. The results of Wu et al. (2007) showed that LED blue light and red light
255 both significantly increased stem length of pea microgreens, when compared with
256 LED white light. The above conclusions, which are in contradiction with previous
257 research results, may be caused by the lower light intensity under LED blue and red
258 light. The study on *Brassicaceae* microgreens showed that the hypocotyl length of
259 tatsoi, red pak choi and mustard microgreens grown under customized LED light was
260 significantly shorter as compared with those of HPSs, even though the light intensity
261 of the two treatments was the same ($150 \mu\text{mol m}^{-2} \text{s}^{-1}$) (Vaštakaitė & Viršilė, 2015).

262 The undesirable hypocotyl length of LED-grown *Brassicaceae* microgreens could be
263 attributed to the high percentage of blue light of customized LED lamps (~16%), as it
264 was reported that hypocotyl elongation could be prevented by the adding of blue light
265 (Darko, Heydarizadeh, Schoefs, & Sabzalian, 2014). The effects of light quality on
266 the leaf area in sprouts and microgreens differ between species. For example, LED red
267 light was reported to have positive effects on leaf area of pea microgreens (Wu, et al.,
268 2007), while having negative effects on leaf area of mustard and tatsoi microgreens
269 (Brazaitytė, et al., 2016). LED light also has positive effects on leaf area. For instance,
270 LED green light was reported to increase leaf area of mizuna microgreens, and the
271 same is true for LED UV-A light in basil and pak choi microgreens (Brazaitytė, et al.,
272 2015b; Gerovac, Craver, Boldt, & Lopez, 2016). As for the effects of light intensity, it
273 was showed that high light intensity could decrease the leaf area in both kohlrabi and
274 mustard microgreens (Gerovac, Craver, Boldt, & Lopez, 2016).

275 Fresh weight is one of the most important growth qualities of sprouts and
276 microgreens. The effects of light on fresh weight varies depend upon the light spectra
277 applied and varied among sprouts and microgreen species. For example, FLs light was
278 reported to increase the fresh weight of mucuna sprouts, while decrease the fresh
279 weight of cowpea sprouts (Martín-Cabrejas, et al., 2008). Similarly, LED UV-A light
280 was shown to increase the fresh weight of basil microgreens, but decrease the fresh
281 weight of beet microgreens (Brazaitytė, et al., 2015a). Both red and blue light are
282 effective for enhancing plant growth because they are more efficiently absorbed by
283 photosynthetic pigments than other regions of the light spectrum. Therefore, it is not

284 surprising that the adding of LED red and/or blue light could increase fresh weight of
285 sprouts and microgreens (Kopsell, Sams, Barickman, & Morrow, 2014; Lee, et al.,
286 2014; Wu, et al., 2007). However, when compared with the dark, LED red and blue
287 light showed inhibitory effects on fresh weight of common buckwheat sprouts (Nam,
288 Lim, & Eom, 2018). Furthermore, light exposure might significantly decrease the
289 yield of soybean sprouts. For example, a study on soybean sprouts with 1-7 days
290 showed that the yield and dry weight of fluorescent light-grown sprouts were lower
291 than that of dark-grown sprouts. The above-mentioned observations may result from
292 the higher protein and lipid consumption and lower water absorption of light-grown
293 sprouts. (Chen & Chang, 2015). The researches on the light intensity showed that
294 LED light with higher light intensity could significantly increase the fresh weight of
295 mustard microgreens (Gerovac, et al., 2016). The details of effects of LED light
296 illumination on the growth of sprouts and microgreens are listed in the Table S1.

297 **3. The effects of LED light on the nutrients content of sprouts and microgreens**

298 The metabolite levels of sprouts and microgreens are mostly determined by the
299 culturing conditions, especially when produced under artificial light and
300 controlled-environment growing conditions. Here, we summarised the changes in
301 main phytochemical compounds content and the antioxidant capacity of sprouts and
302 microgreens grown under LED light, we also reviewed the changes of gene
303 expression levels in response to the different LED light.

304 **3.1 Phenolic compounds**

305 Phenolic compounds, which are ubiquitous in plants, comprise an extremely rich

306 group of secondary metabolites. They confer the colour, taste, and aroma of fruits and
307 vegetables, and are an integral part of the human diet (Shahidi & Ambigaipalan,
308 2015). Numerous studies have highlighted the antioxidant properties and potential
309 health-improving effects of a phenolic-rich diet, such as antioxidant,
310 anti-inflammatory, anti-adhesive and antibacterial effects (Balasundram, Sundram, &
311 Samman, 2006; Ferreira, Martins, & Barros, 2017; Rice-Evans, Miller, & Paganga,
312 1997). The content of phenolic compounds is an important quality index of sprouts
313 and microgreens, and the accumulation of phenolic phytochemicals can be stimulated
314 by cultivation under different LEDs (Table 1). Compared to the dark, light exposure
315 increased the total phenolic content of soybean, Chinese kale, and pea sprouts, as well
316 as sprouts of *Brassica oleracea* varieties (Kim, et al., 2006; Liu, et al., 2016; Qian, et
317 al., 2016; Vale, Cidade, Pinto, & Oliveira, 2014). However, when grown in the
318 growth chamber at controlled light cycle (16 h/8 h, light/dark), the total phenolic
319 content decreased in galega kale, penca cabbage and broccoli sprouts, while increased
320 in red cabbage sprouts (Vale, Cidade, Pinto, & Oliveira, 2014). The studies on the
321 light quality showed that total phenolic content of Chinese kale sprouts and common
322 buckwheat sprouts was significantly decreased under LED red light, while
323 significantly increased under LED blue light, as compared with white light (Nam,
324 Kim, & Eom, 2018; Qian, et al., 2016). In a study of pea sprouts, LED light of 585
325 nm was observed to decrease total phenolic content (Liu, et al., 2016). Several studies
326 on tatsoi and basil microgreens reported an increase in total phenolic content under
327 LED red light or supplemental LED red light (Brazaitytė, et al., 2016b; Samuolienė,

328 et al., 2016).

329 Flavonoids represent one of the largest classes of phenolic compounds in fruits
330 and vegetables (Zoratti, Karppinen, Escobar, Häggman, & Jaakola, 2014). In sprouts
331 and microgreens, isoflavone, anthocyanin, rutin and quercetin are the most common
332 phenolic compounds. Similar to the promoting effects on total phenolic content, light
333 exposure significantly increased the content of the above-mentioned phenolic
334 compounds (Table 1). For instance, light exposure was reported to boost isoflavone
335 content in soybean and chickpea sprouts (Aisyah, Gruppen, Madzora, & Vincken,
336 2013; Gao, Yao, Zhu, & Ren, 2015). In addition, LED blue light showed promoting
337 effects on total flavonoid content of common buckwheat sprouts, while LED red light
338 showed the opposite effect (Nam, Kim & Eom, 2018). It is reported that secondary
339 metabolites in plants are a response to the environmental stresses (Ramakrishna &
340 Ravishankar, 2011). This has led to a series of studies on the effects of UV light on
341 secondary metabolites in sprouts and microgreens. For example, UV-B lamps has
342 been reported to increase total flavonoid content in broccoli sprouts by 92%, when
343 compared with white light (Mewis, et al., 2012). UV-B light at 313 nm was reported
344 to increase isoflavone content in soybean sprouts (Jiao, Yang, & Gu, 2016; Jiao, Yang,
345 & Gu, 2017), while UV-B at 310 nm was reported to decrease isoflavone content in
346 red clover sprouts (Grażyna, et al., 2018). In soybean sprouts, there were consistent
347 strong positive correlations between the isoflavone content and radiation intensity of
348 UV-B (from 0, 5, 10 and 20 $\mu\text{W cm}^{-2}$) (Ma, Wang, Yang, & Gu, 2018). Besides, nitric
349 oxide signaling was reported involved in UV-B-induced isoflavone biosynthesis by

350 regulating the gene expression of key enzymes related (i.e. phenylalanine ammonia
351 lyase, PAL and chalcone synthase, CHS) (Jiao, Yang, Zhou, & Gu, 2016). Therefore,
352 the influence of UV-B on isoflavone accumulation is dependent on the species, the
353 UV-B wavelength as well as the radiation intensity (UV dose). In a study of tartary
354 buckwheat sprouts, LED blue light followed by UV-C light enhanced the
355 accumulation of bioactive compounds, while the opposite combination (UV-C
356 followed by LED blue light) showed weaker effects (Ji, Wen, Zhou, & Ying, 2016).
357 Thus, the content of phenolic compounds was not only dependent on the light quality
358 but also the way they are combined.

359 The flavonoids is biosynthesized through a branched phenylpropanoid pathway,
360 and most of the flavonoid synthesizing enzymes have been found. Recently, the
361 transcriptional levels of flavonoid biosynthetic genes were investigated in sprouts and
362 microgreens. For example, it was shown that the time duration and amount of light
363 strongly affected the phenylpropanoids content in tartary buckwheat sprouts, the three
364 anthocyanin compounds (namely cyanidin 3-*O* glucoside, cyanidin 3-*O*-rutinoside
365 and delphindin-3-*O*-coumarylglucoside) in 4-day-old Hokkai T10 buckwheat sprouts
366 grown under light/dark condition was nearly 4-fold more than those grown in the dark
367 (Li, et al., 2012). The transcriptional results of flavonoid biosynthetic genes also
368 showed that *FtFLS2*, *FtF3'H1*, *FtF3'H2* and *FtANS* were up-regulated by light. In
369 addition, the transcription factor, *FtMYB*-like gene, was markedly induced in
370 light-treated buckwheat sprouts, indicating that *FtMYB*-like gene was possibly
371 involved in the light-regulated flavonoid biosynthesis (Li, et al., 2012). In another

372 study, it was shown that rutin was the main phenolic compounds in Hokkai T8
373 buckwheat sprouts, and the maximum rutin content was observed at 4 day after LED
374 exposure under blue light, as compared to the LEDA white and red light. Meanwhile,
375 the cyanidin 3-O-rutinoside content under LED blue light was 2.8- and 10.6-fold
376 higher than that under LED white and red light, respectively. The analysis of gene
377 expression showed that the transcript levels of *FtC4H*, *FtCHI*, *FtFLS-2*, and *FtANS*
378 was higher in sprouts grown under LED blue light (Thwe, et al., 2014). Studies on
379 UV irradiation showed that UV-B and UV-A could increase the anthocyanin content
380 of radish sprouts and soybean sprouts, respectively (Su, et al., 2016; Su, et al., 2017).
381 The anthocyanin accumulation was significantly decreased by shading treatment in
382 soybean sprouts, which was attributed to the down-regulation of anthocyanin
383 biosynthesis genes (*GmDFR*, *GmANS* and *GmUFGT*) (Su, et al., 2017). Those results
384 strongly suggested that the accumulation of anthocyanin was closely related to light
385 exposure. On one hand, light is a necessary condition for the biosynthesis of
386 anthocyanins. On the other hand, different light qualities exhibit different effects on
387 anthocyanin biosynthesis, and short-wavelength light, such as blue light and UV light,
388 might be more effective to increase anthocyanin content (Qian, et al., 2016; Seo, Arasu,
389 Kim, Park, & Kim, 2015; Thwe, et al., 2014). The details of effects of LED light on
390 the phenolic compounds content in sprouts and microgreens are summarized in the
391 Table 1.

392 In addition to light spectra, light intensity and light duration, pulsed light was
393 reported has profound effects on the accumulation of phytochemicals in microgreen.

394 Vaštakaitė, et al. (2017) reported that adding monochromatic (455, 470, 505, 590 and
395 627nm) LEDs with frequencies at 2, 256, and 1024 Hz had most positive effects on
396 total phenolic compounds in mustard microgreens grown under HPS lamps. On the
397 contrary, adding the monochromatic LEDs at 32 Hz significantly decreased the total
398 phenolic content in mustard microgreens. However, the monochromatic LED
399 frequencies at 32 Hz were the most suitable for the accumulation of anthocyanins in
400 red pak choi and tatsoi microgreens.

401 **3.2 Vitamins**

402 Vitamins are a class of organic compounds that cannot be synthesized by the
403 human body and can only be absorbed from the diet. They are essential for human
404 development and health, and the intake of vitamins help to reduce the risk of diseases,
405 such as cardiovascular disease and coronary heart disease (Ashor, Lara, Mathers, &
406 Siervo, 2014; Stampfer, et al., 1993). Vitamin C (ascorbic acid) is a dietary nutrient
407 required as a co-factor for many enzymes, and is an especially effective antioxidant
408 owing to its high electron-donating power (Jacob & Sotoudeh, 2002). Therefore,
409 vitamin C is one of the most important vitamins in the human diet. Artificial light has
410 been reported to significantly increase vitamin C content of sprouts and microgreens
411 (Table S2). For example, compared to growth in the dark, white light and UV + red
412 LED light significantly increased vitamin C content of broccoli sprouts and soybean
413 sprouts by 87% and 79%, respectively (Pérez-Balibrea, Moreno, & García-Viguera,
414 2008; Xu, Dong, & Zhu, 2005). In addition, LED red light was reported to play a
415 positive role in promoting vitamin C content of microgreens (Brazaitytė,

416 Jankauskienė, & Novičkovas, 2013; Brazaitytė, et al., 2016). Moreover, it was shown
417 that supplementary LED light, on the basis of HPS light, could also promote the
418 content of vitamin C of tatsoi microgreens (Vaštakaitė & Viršilė, 2015).

419 Vitamin E, the major lipid-soluble component in the cell antioxidant defense
420 system, has long been considered as a cytoprotective factor with roles in anti-ageing,
421 arthritis, cataracts, Type 2 diabetic disease and kidney diseases (Jiang, 2014; Rizvi, et
422 al., 2014). Alpha-tocopherols, the predominant form of vitamin E, was reported to be
423 regulated by artificial light (LED and HPS lamps) in microgreens. As shown in Table
424 S2, using the combination of LEDs or HPSs as the primary light source, supplemental
425 green, blue and red LED light could promote the increase in vitamin E content of
426 microgreens (Brazaitytė, et al., 2016; Samuolienė, et al., 2014; Samuolienė, et al.,
427 2017). However, LED red light was reported to decrease the content of vitamin E in
428 *Perilla frutescens* (L.) microgreens and red pak choi microgreens (Brazaitytė,
429 Jankauskienė, & Novičkovas, 2013; Brazaitytė, et al., 2016). Therefore, the effects of
430 LED light on vitamin E content depend on the light spectra and intensity and vary
431 among microgreen species (Table S2).

432 **3.3 Photosynthetic pigment contents**

433 Following exposure to light, sprouts and microgreens undergo
434 photomorphogenesis and inevitably synthesize photosynthetic pigments, such as
435 chlorophyll and carotenoids. Chlorophylls are the most obvious and widespread
436 pigments of plants, as they are required for photosynthesis. In addition, chlorophylls
437 have been shown to play important roles in maintain human health, in view of their

438 potent anti-inflammatory property and anti-oxidant activity (Lee, Nishizawa, Shimizu,
439 & Saeki, 2017; Subramoniam, et al., 2012). Several preliminary studies revealed the
440 effects of artificial light on chlorophyll and carotenoid content of sprouts and
441 microgreens (Table S3). It was reported that 20% LED blue light with 80% LED red
442 light treatment could significantly increase the content of total chlorophyll,
443 chlorophyll a and chlorophyll b of broccoli microgreens (Kopsell, Sams, Barickman,
444 & Morrow, 2014). Moreover, UV-A was also reported to increase total chlorophyll
445 content in broccoli sprouts (Moreira-Rodríguez, Nair, Benavides, Cisneros-Zevallos,
446 & Jacobo-Velázquez, 2017b). However, a recent study has revealed that LED blue
447 light could decrease the total chlorophyll content of common buckwheat sprouts, as
448 compared with the white light (Nam, Lim, & Eom 2018). Similarly, supplemental
449 LED blue light significantly decreased the chlorophyll b content of broccoli
450 microgreens (Kopsell, & Sams, 2013). Additionally, the total chlorophyll content of
451 microgreens decreases when exposed to high-intensity light. For example, total
452 chlorophyll content of mustard microgreens decreased by 27% under fluorescent light
453 of $463 \mu\text{mol m}^{-2} \text{s}^{-1}$, in comparison to $275 \mu\text{mol m}^{-2} \text{s}^{-1}$ exposure (Kopsell,
454 Pantanizopoulos, Sams, & Kopsell, 2012).

455 Carotenoids (α -carotene, β -carotene, violaxanthin, neoxanthin and lutein, etc.)
456 play fundamental roles in photosynthetic organisms. They act as accessory
457 light-harvesting pigments, but they also perform photoprotective role by quenching
458 triplet state chlorophyll molecules and scavenging singlet oxygen and other toxic
459 oxygen species formed within the chloroplast (Young, 1991). Dietary carotenoids are

460 thought to play an important role in maintaining good health and preventing human
461 diseases, such as cancers, diabetes and eye disease (Johnson, 2002; Sluijs, et al.,
462 2015). Generally, LED light is beneficial for the accumulation of carotenoids in
463 sprouts and microgreens (Table S3). For example, the supplementation of LED yellow
464 light (595 nm) could increase total carotenoids content of tatsoi microgreens by 16%
465 (Brazaitytė, et al., 2015b). Similarly, supplemental blue, red and green LED light was
466 reported to increase the content of α -carotene and β -carotene of microgreens
467 (Brazaitytė, et al., 2015b; Brazaitytė, et al., 2016; Kopsell, Sams, Barickman, &
468 Morrow, 2014; Samuolienė, et al., 2017). However, supplemental LED orange light
469 (622 nm) decreased β -carotene content (Brazaitytė, et al., 2015b). Studies on
470 *Brassicaceae* microgreens and beet microgreens showed that supplemental green,
471 yellow and blue LED light could promote violaxanthin content, while supplemental
472 LED orange light showed the opposite effect (Brazaitytė, et al., 2015b; Samuolienė, et
473 al., 2017). It was also reported that supplemental LED green light increased
474 neoxanthin content of red pak choi microgreens by 5%, while supplemental LED
475 yellow light lead to a decrease in neoxanthin content by 69% (Brazaitytė et al., 2015b).
476 A study on the effects of high light intensity reported that high light intensity could
477 decrease neoxanthin content of mustard microgreens (Kopsell, Pantanizopoulos, Sams,
478 & Kopsell, 2012). It was reported that lutein and β - carotene are two main carotenoid
479 compositions in tartary buckwheat sprouts, and white florescent light exposure (16 h/8
480 h light/dark cycles) could significantly increase their content, even if the light
481 intensity was very low ($35 \mu\text{mol m}^{-2} \text{s}^{-1}$) (Tuan, et al., 2013a). The total carotenoid

482 content reached the maximum level at 9 or 6 days after sowing (DAS) in light-grown
483 T8 or T10 tartary buckwheat sprouts, respectively. The transcription levels of
484 carotenoid biosynthetic genes were analyzed during the sprouts' developmental stages,
485 showing that the expression of *FtPSY*, *FtPDS*, *FtZDS*, *FtLCYB* and *FtLCYE* genes
486 increased drastically from 3 DAS to 9 DAS and then decreased at 12 DAS in T10
487 tartary buckwheat sprouts, both in the dark and under light. And the higher expression
488 levels of carotenoid biosynthetic genes and carotenoid content was observed in the
489 light-grown T10 tartary buckwheat sprouts. While, in T8 tartary buckwheat sprouts,
490 the expression of the above-mentioned genes decreased from 3 DAS to 9 or 6 DAS
491 then increased at 12 DAS, whether in the presence of light or not. Although no
492 difference was observed in the transcript levels of carotenoid biosynthetic-related
493 genes between light and dark conditions, the carotenoid contents in the light-grown
494 T8 tartary buckwheat sprouts were significantly higher. The discrepancy between
495 gene expression and carotenoid content in T8 and T10 tartary buckwheat sprouts
496 suggested that other isoforms of carotenoid biosynthetic genes may participate in
497 controlling the flux into carotenoid biosynthesis. In addition, the accumulation of
498 carotenoid was potentially controlled by additional determinants, such as the
499 post-transcriptional and metabolic mechanisms (Tuan, et al., 2013a). At the same year,
500 it was also reported that the total carotenoids content under LED white light (1282.63
501 $\mu\text{g g}^{-1}$ dry weight) was much higher than that under LED blue (858.29 $\mu\text{g g}^{-1}$ dry
502 weight) and LED red (908.64 $\mu\text{g g}^{-1}$ dry weight) light in T8 tartary buckwheat sprouts
503 at 10 DAS. The accumulation of carotenoids was induced under white light from 2

504 DAS to 10 DAS, while it increased from 2 to 6 DAS (red light) or 8 DAS (blue light)
505 then decreased at 10 DAS. At 8 DAS, the transcription levels of *FtPSY*, *FtLCYB*,
506 *FtLCYE*, *FtCHXB*, *FtCHXE*, and *FtZEP* were higher in sprouts grown under white
507 light than in those grown under blue and red lights, which might lead to the rich
508 amount of carotenoids accumulation under white light (Tuan, et al., 2013b). LED red
509 light, whether used alone or as a supplementary light, was reported to increase lutein
510 content of tatsoi and basil microgreens, but decreased lutein content of mustard
511 microgreens (Brazaitytė, et al., 2016; Samuolienė, et al., 2017). Therefore, the effects
512 of LED red light on carotenoid accumulation might depend on both the species and
513 the light conditions of this processing (Table S3).

514 **3.4 Other nutritional compounds**

515 In addition to the bioactive substances mentioned above, sprouts and
516 microgreens also contain other nutrients and bioactive compounds (Table S4).
517 Glucosinolates are an important group of secondary plant metabolites. They are
518 sulfonated thioglycosides with a variable aglycone side chain, based on the structure
519 of which glucosinolates are divided into three classes: aliphatic, indolyl and aromatic
520 glucosinolates (Mewis, et al., 2012). Apart from the well-known role in plant defence
521 response to insects and pathogens, glucosinolates also have health benefits in lowering
522 the risk of myocardial infarction, cancer and coronary heart disease (Ma, et al., 2018;
523 Traka, 2016). The biosynthesis of glucosinolates can be triggered by a series of biotic
524 and abiotic factors (Kissen, et al., 2016). Therefore, UV irradiation was used as an
525 effective strategy to accumulate high levels of glucosinolates in sprouts and

526 microgreens. It was reported that UV-A, UV-B light doses and harvesting time
527 differentially tailor glucosinolate profiles in broccoli sprouts, and UV-B could
528 significantly increase the content of glucosinolate in broccoli sprouts
529 (Moreira-Rodríguez, Nair, Benavides, Cisneros-Zevallos, & Jacobo-Velázquez, 2017a;
530 2017b) (Table S4). Light exposure has positive effect on glucosinolates accumulation
531 in sprouts. For instance, compared with the dark, exposure to fluorescent light was
532 reported to significantly increased the total glucosinolate content of broccoli sprouts
533 by 35% (Pérez-Balibrea, Moreno, & García-Viguera, 2008). However, white light was
534 reported to significantly decrease total glucosinolates content of white mustard
535 sprouts by 8% (Ciska, Honke, & Kozłowska, 2008). The effects of different light
536 qualities (white LED, red and blue lights) on glucosinolates content in Chinese kale
537 sprouts were investigated, the results showed that the total glucosinolates content in
538 the shoot under white and red LED light exhibited no striking change, but was
539 significantly decreased under LED blue light, when compared to those grown in the
540 dark (Qian, et al., 2016). However, Kopsell and Sams (2013) reported that
541 supplemental LED blue light could significantly increase the glucosinolate content of
542 broccoli microgreens. Combining red and blue LED light was also reported to
543 increase the glucosinolate content of broccoli microgreens (Kopsell, Sams, Barickman,
544 & Morrow, 2014). Recently, different cultivars of rapeseed sprouts are studied to
545 identify the effects of blue/red light ratios on glucosinolate accumulation. The results
546 showed that the glucosinolate content varied between different cultivars, but neither
547 high blue ratio (31.7% blue light/ 66.3% red light) nor low blue ratio (14.8%

548 blue/81.3% red light) could significantly alter the glucosinolate content. Therefore,
549 the glucosinolates content of rapeseed sprouts might be optimized when treated with
550 appropriate light treatment (Groenbaek, Tybirk, & Kristensen, 2018).

551 The total protein content of cowpea sprouts, jack bean sprouts and red cabbage
552 sprouts were significantly increased under white light, as compared to those grown in
553 the dark. However, the total protein content of mucuna sprouts and penca cabbage
554 sprouts was significantly decreased under white light (Martín-Cabrejas, et al., 2008;
555 Vale, et al., 2015). Therefore, the influence of white light on total protein content is
556 species- and cultivars-dependent. In a study of mustard sprouts, 9 essential or semi
557 essential amino acids and 12 nonessential amino acids were detected, and the contents
558 of glycine, arginine, and isoleucine were up-regulated by light conditions, reached the
559 maximum after 9 or 12 days. As for the total amino acids, the highest content was
560 observed in dark-grown mustard sprouts (Li, et al., 2013). Among those detected
561 amino acids, γ -aminobutyric acid (GABA) is a non-protein amino acid and has been
562 considered a bioactive plant component. Glutamate decarboxylase (GAD) is the
563 rate-limiting enzyme for GABA accumulation in GABA shunt. The expression of
564 GAD-encoding genes showed higher expression level in the dark than under the light
565 conditions (16 h/8 h, light/dark). However, GABA content in light-grown mustard
566 sprouts was significantly higher than that of dark-grown sprouts. The discrepancy
567 between the *GAD* expression level and GABA content of light-grown mustard sprouts
568 may resulted from the activity of GABA transaminase and diamine oxidase (DAO),
569 which catalyze the degradation of GABA and GABA polyamine degradation pathway,

570 respectively (Shelp, 1997). This study provided a foundation for understanding the
571 effects of light on GABA accumulation in sprouts, nevertheless, further studies are
572 necessary to identify more regulators and genes that are associated with these
573 compounds' biosynthesis.

574 Dietary fiber is reported to reduce the risk of colorectal adenoma and colorectal
575 cancer (Aune, et al., 2011; Ben, et al., 2014), with vegetables being one of the main
576 sources of dietary fiber. Vale, et al. (2015) reported that exposure to light determined a
577 lower content of total dietary fiber in the four studied *Brassica oleracea* sprouts (red
578 cabbage, broccoli, Galega kale and Penca cabbage). Similarly, Martín-Cabrejas, et al.
579 (2008) reported that the soluble dietary fiber content was significantly decreased in
580 cowpea sprouts and dolichos sprouts under 12 h/d and 24 h/d illumination, while
581 significantly increased in jack bean sprouts and soybean sprouts under 24 h/d
582 illumination. It is well-documented that high dietary nitrate consumption is associated
583 with an increased risk of gastrointestinal cancer (Muscaritoli, Amabile, & Molfino,
584 2016). Approximately 80% of human dietary nitrates comes from vegetables, so ways
585 to reduce the nitrate content in vegetables has become a primary concern. It was
586 reported that LED red light could reduce nitrate content of *Perilla frutescens* (L.)
587 microgreens and radish microgreens, while increases were seen in basil microgreens.
588 It is also reported that LED green light decreases nitrate content of radish microgreens,
589 while increases nitrate content of basil microgreens (Samuolienė, et al., 2014).
590 Mineral elements are also important nutrient of sprouts and microgreens and light
591 could also manipulate the mineral elements content in sprouts and microgreens. For

592 example, it was reported that, compared with the HPS treatment, compound LED
593 light could significantly increase the content of minerals, such as Ca^{2+} , K^+ , Na^+ and P^+ ,
594 in *Brassicaceae* microgreens (Vaštakaitė & Viršilė, 2015).

595 **3.5 Antioxidant capacity**

596 Antioxidants are compounds that, in low concentration, can prevent
597 biomolecules (e.g. proteins, nucleic acids, polyunsaturated lipids, and sugars) from
598 undergoing oxidative damage through free radical-mediated reactions (Bendary,
599 Francis, Ali, Sarwat, & El Hady, 2013). They contribute to oxidative stress-related
600 diseases, e.g. neurodegenerative diseases, cancer and heart disease (Apak, Özyürek,
601 Güçlü, & Çapanoğlu, 2016). Various methods are used to measure the antioxidant
602 activity of dietary antioxidants. The most commonly used methods for detecting
603 antioxidant capacity of sprouts and microgreens include
604 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay, ferric ion reducing antioxidant power
605 (FRAP) assay, 2,2'-azino-bis-3-ethylbenzthiazoline-6-sulphonic acid (ABTS) assay
606 and oxygen radical absorbance capacity (ORAC) assay (Table S5).

607 There are many studies show that sprouts and microgreens are good sources of
608 dietary antioxidants. Generally, the antioxidant capacity of sprouts and microgreens
609 reflects the composition and content of their bioactive compounds, such as phenolics,
610 vitamins, chlorophyll, carotenoids and glucosinolates (Keum, Jeong, & Kong, 2004;
611 Podsędek, 2007; Williamson, Faulkner, & Plumb, 1998). The structure-activity
612 relationships (SAR) among those bioactive compounds hence provide us with a
613 preliminary insight into their antioxidant capacity. Many studies showed that the
614 antioxidant activity of sprouts and microgreens was positively correlated with
615 flavonoids compounds content and was closely depended on the light conditions

616 (Table S5). For example, in the Chinese kale sprouts, the antioxidant capacity was
617 significantly increased in sprouts under light exposure, and the highest antioxidant
618 capacity was observed in sprouts grown under LED blue light, which was in
619 accordance with the variation tendency of anthocyanin content (Qian, et al., 2016). In
620 a study of soybean sprouts, the antioxidant activities of two varieties of soybean
621 sprouts, yellow soybean sprouts (YSS, grown in the dark) and green soybean sprouts
622 (GSS, grown under light), with different germination days were studied using DPPH,
623 FRAP and ORAC analysis methods. The results showed that the GSS had lower
624 DPPH radical scavenging capacity but higher ORAC value than YSS on day 7. It was
625 reported that ORAC could measure both lipophilic and hydrophilic antioxidants,
626 which is considered the most relevant to human biology among the antioxidant assay
627 methods (e.g. DPPH, ORAC and FRAP) (Prior, Xianli, & Karen, 2005). Thus, it was
628 speculated that ORAC value is more fundamentally representative of the antioxidant
629 status of soybean sprouts than DPPH. And it is better to produce GSS from the
630 viewpoint of antioxidant capacity because ORAC values were higher than those of
631 YSS. (Chen & Chang, 2015). In another study, light illumination (12 h light and 12 h
632 dark) significantly enhanced the DPPH radical scavenging activity and the FRAP
633 value of polysaccharides from soybean sprouts, which indicating that light
634 illumination during germination has a significant effect on structural and functional
635 properties of polysaccharides derived from soybean sprouts (Yuan, et al., 2015b). In
636 addition, the activity of antioxidant enzymes could also be significantly increased by
637 UV-B (Jiao, Yang, Zhou, & Gu, 2016). The correlation analysis of phenolic
638 compounds with antioxidant capacity showed that the antioxidant activities were
639 correlated significantly and positively with phenolic content of lentil sprouts and pea
640 sprouts (Liu, et al., 2016; Świeca, Gawlik-Dziki, Kowalczyk, & Złotek, 2012).

641 However, studies have also shown that light has no effect or even negative effect on
642 the antioxidant capacity of sprouts and microgreens (Brazaitytė, Jankauskienė, &
643 Novičkovas, 2013; Chen & Chang, 2015; Świeca, Gawlik-Dziki, Kowalczyk, &
644 Złotek, 2012; Vale, Cidade, Pinto, & Oliveira, 2014).

645 **3.6 Postharvest quality**

646 Sprouts and microgreens are delicate and have a short shelf life due to high
647 moisture content and rapid deterioration, which limits their commercial use. In
648 addition, the nutritional quality of sprouts decreases during postharvest storage.
649 Studies have shown that light treatments after harvesting of fruits and vegetables can
650 play a role in delaying ageing, reducing nutrient loss and extending shelf life (Büchert,
651 Gómez Lobato, Villarreal, Civello, & Martínez, 2011; Jin, Yao, Xu, Wang, & Zheng,
652 2015). However, little information is available describing the postharvest physiology
653 of sprouts and microgreens, and the effects of light during this process have not yet
654 been well quantified. Recently, several studies have shown that postharvest light
655 treatment could affect the shelf life and the bioactive compounds content of the
656 post-harvested sprouts and microgreens. For example, it was reported that
657 continuous fluorescent light exposure during storage increased the content of ascorbic
658 acid while having no effect on α -tocopherol, total phenolic concentrations or DPPH
659 radical scavenging capacity of radish microgreens (Xiao, et al., 2014). Although
660 UV-LEDs with wavelengths around 240-370 nm are commercially available, the
661 application of UV-LEDs is restricted due to their lower output optical power and lack
662 of complete monochromaticity (Bui & Hauser, 2015; Li, Dvořák, Nesterenko,

663 Nuchtavorn, & Macka, 2018). Nevertheless, the use of UV florescent lamps have
664 shown that postharvest UV-B radiation could further boost glucosinolates levels and
665 extend the shelf life of broccoli microgreens (Lu, et al., 2018). Further studies are
666 needed to investigate the effects of light exposure on the shelf life and nutrient
667 contents of sprouts and microgreens.

668 **4. Conclusions**

669 LED, an innovative artificial light source for plants, is a promising lighting for
670 improving the nutrient quality in sprouts and microgreens, both as a supplemental and
671 a sole-source illumination. In these studies, it has been preliminarily established that a
672 series of structural genes related to the biosynthesis of phytochemical compounds,
673 such as flavonoids and carotenoids, are regulated in response to LED light in sprouts
674 and microgreens. However, the expression patterns of the biosynthetic related genes
675 cannot always explain their accumulation levels. Therefore, further work is necessary
676 to thoroughly delineate the underlying mechanisms. Sprouts and microgreens are
677 young seedlings that are sensitive to light and are a good model system for studying
678 the metabolism. We would suggest the following topics that are worth considering for
679 further exploration:

680 1) for the more efficient use of LED light and the improvement of the nutritional
681 and health value of sprouts and microgreen vegetables, specific LED light recipes (the
682 properly designed LED lighting system combining different spectral composition and
683 light intensities) should be identified to increase the yield and maximize the content of
684 phytochemical compounds to promote health or prevents diseases;

685 2) the integrated approaches of transcriptomic, proteomic and metabolomic
686 analysis are needed to reveal the dynamics of the metabolism under LED light;

687 3) the role of photoreceptors (e.g. phytochromes) and light signal transduction
688 pathways in regulating phytochemical compounds metabolism under LED light
689 remained to be clarified;

690 4) further analysis should be carried out to identify the target genes and markers
691 associated with secondary metabolites biosynthesis for future molecular breeding.

692 The studies on the regulatory effect of LED light spectra on the growth and
693 metabolism of sprouts and microgreens will provide an increasingly mature system in
694 which to provide adequate quantity and quality of vegetables, as well as to explore the
695 basic responses of plant seedlings to light signals. Therefore, the application of LED
696 light in vegetable production has enormous potential health benefits, and provides
697 possibilities for feeding the expanding population sustainably and efficiently in the
698 future.

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706

707 **References**

- 708 Aisyah, S., Gruppen, H., Madzora, B., & Vincken, J.-P. (2013). Modulation of isoflavonoid
709 composition of *Rhizopus oryzae* elicited soybean (*Glycine max*) seedlings by light and
710 wounding. *Journal of Agricultural and Food Chemistry*, 61, 8657-8667.
- 711 Alabadí, D., Gallego-Bartolomé, J., Orlando, L., García-Cárcel, L., Rubio, V., Martínez, C., et al.
712 (2008). Gibberellins modulate light signaling pathways to prevent Arabidopsis seedling
713 de-etiolation in darkness. *The Plant Journal*, 53, 324-335.
- 714 Apak, R. a., Özyürek, M., Güçlü, K., & Çapanoğlu, E. (2016). Antioxidant activity/capacity
715 measurement. 3. Reactive oxygen and nitrogen species (ROS/RNS) scavenging assays,
716 oxidative stress biomarkers, and chromatographic/chemometric assays. *Journal of*
717 *Agricultural and Food Chemistry*, 64, 1046–1070.
- 718 Ashor, A. W., Lara, J., Mathers, J. C., & Siervo, M. (2014). Effect of vitamin C on endothelial
719 function in health and disease: A systematic review and meta-analysis of randomised
720 controlled trials. *Atherosclerosis*, 235, 9–20.
- 721 Aune, D., Chan, D. S. M., Lau, R., Vieira, R., Greenwood, D. C., Kampman, E., et al. (2011).
722 Dietary fibre, whole grains, and risk of colorectal cancer: systematic review and
723 dose-response meta-analysis of prospective studies. *BMJ*, 343, d6617.
- 724 Aune, D., Giovannucci, E., Boffetta, P., Fadnes, L. T., Keum, N., Norat, T., et al. (2017). Fruit and
725 vegetable intake and the risk of cardiovascular disease, total cancer and all-cause
726 mortality—a systematic review and dose-response meta-analysis of prospective studies.
727 *International Journal of Epidemiology*, 46, 1029–1056.
- 728 Balasundram, N., Sundram, K., & Samman, S. (2006). Phenolic compounds in plants and

- 729 agri-industrial by-products: Antioxidant activity, occurrence, and potential uses. *Food*
730 *chemistry*, 99, 191–203.
- 731 Bantis, F., Smirnakou, S., Ouzounis, T., Koukounaras, A., Ntagkas, N., & Radoglou, K. (2018).
732 Current status and recent achievements in the field of horticulture with the use of
733 light-emitting diodes (LEDs). *Scientia Horticulturae*, 235, 437–451.
- 734 Barber, J., & Andersson, B. (1992). Too much of a good thing: light can be bad for photosynthesis.
735 *Trends in biochemical sciences*, 17, 61–66.
- 736 Ben, Q., Sun, Y., Chai, R., Qian, A., Xu, B., & Yuan, Y. (2014). Dietary fiber intake reduces risk
737 for colorectal adenoma: A meta-analysis. *Gastroenterology*, 146, 689–699.
- 738 Bendary, E., Francis, R., Ali, H., Sarwat, M., & El Hady, S. (2013). Antioxidant and structure–
739 activity relationships (SARs) of some phenolic and anilines compounds. *Annals of*
740 *Agricultural Sciences*, 58, 173–181.
- 741 Bora, P. (2014). Anti-nutritional factors in foods and their effects. *Journal of Academia and*
742 *Industrial Research*, 3, 285–290.
- 743 Both, A. J., Bugbee, B., Kubota, C., Lopez, R. G., Mitchell, C., Runkle, E. S., et al. (2017).
744 Proposed product label for electric lamps used in the plant sciences. *HortTechnology*, 27,
745 544–549.
- 746 Brazaitytė, A., Jankauskienė, J., & Novičkovas, A. (2013). The Effects of
747 supplementary short-term red LEDs lighting on nutritional quality of *Perilla frutescens* L.
748 microgreens. *Rural development*, 2013, 54–58.
- 749 Brazaitytė, A., Sakalauskienė, S., Samuolienė, G., Jankauskienė, J., Viršilė, A., Novičkovas, A., et
750 al. (2015b). The effects of LED illumination spectra and intensity on carotenoid content in
Brassicaceae microgreens. *Food chemistry*, 173, 600–606.

- 751 Brazaitytė, A., Sakalauskienė, S., Viršilė, A., Julė, J., Samuoliene, G., Sirtautas, R., et al. (2016).
752 The effect of short-term red lighting on *Brassicaceae* microgreens grown indoors. *Acta*
753 *Horticulturae*, *1123*, 177–184.
- 754 Brazaitytė, A., Viršilė, A., Jankauskienė, J., Sakalauskienė, S., Samuolienė, G., Sirtautas, R., et al.
755 (2015a). Effect of supplemental UV-A irradiation in solid-state lighting on the growth and
756 phytochemical content of microgreens. *International Agrophysics*, *29*, 13–22.
- 757 Büchert, A. M., Gómez Lobato, M. E., Villarreal, N. M., Civello, P. M., & Martínez, G. A. (2011).
758 Effect of visible light treatments on postharvest senescence of broccoli (*Brassica oleracea*
759 L.). *Journal of Agricultural and Food Chemistry*, *91*, 355–361.
- 760 Bui, D. A., & Hauser, P. C. (2015). Analytical devices based on light-emitting diodes—a review of
761 the state-of-the-art. *Analytica chimica acta*, *853*, 46–58.
- 762 Chen, M., & Blankenship, R. E. (2011). Expanding the solar spectrum used by photosynthesis.
763 *Trends in Plant Science*, *16*, 427–431.
- 764 Chen, M., Chory, J., & Fankhauser, C. (2004). Light signal transduction in higher plants. *Annual*
765 *Review of Genetics*, *38*, 87–117.
- 766 Chen, Y., & Chang, S. K. C. (2015). Macronutrients, phytochemicals, and antioxidant activity of
767 soybean sprout germinated with or without light exposure. *Journal of Food Science*, *80*,
768 1391–1398.
- 769 Ciska, E., Honke, J., & Kozłowska, H. (2008). Effect of light conditions on the contents of
770 glucosinolates in germinating seeds of white mustard, red radish, white radish, and rapeseed.
771 *Journal of Agricultural and Food Chemistry*, *56*, 9087–9093.
- 772 Darko, E., Heydarizadeh, P., Schoefs, B., & Sabzalian, M. R. (2014). Photosynthesis under

- 773 artificial light: the shift in primary and secondary metabolism. *Philosophical Transactions of*
774 *The Royal Society B Biological Sciences*, 369, 20130243.
- 775 Deng, X., Caspar, T., & Quail, P. H. (1991). cop1: a regulatory locus involved in light-controlled
776 development and gene expression in Arabidopsis. *Genes development*, 5, 1172–1182.
- 777 Di Gioia, F., Renna, M., & Santamaria, P. (2017). Sprouts, microgreens and “baby Leaf”
778 vegetables. In *Minimally Processed Refrigerated Fruits and Vegetables*; Springer: Boston,
779 MA, USA, pp. 403–432.
- 780 Ding, Z., Galván-Ampudia, C. S., Demarsy, E., Łangowski, Ł., Kleine-Vehn, J., Fan, Y., et al.
781 (2011). Light-mediated polarization of the PIN3 auxin transporter for the phototropic
782 response in Arabidopsis. *Nature Cell Biology*, 13, 447–452.
- 783 D'Souza, C., Yuk, H. G., Khoo, G. H., & Zhou, W. (2015). Application of light-emitting diodes in
784 food production, postharvest preservation, and microbiological food safety. *Comprehensive*
785 *Reviews in Food Science and Food Safety*, 14, 719–740.
- 786 Ebert, A. (2012). Sprouts, microgreens, and edible flowers: the potential for high value specialty
787 produce in Asia. In *High Value Vegetables in Southeast Asia: Production, Supply and*
788 *Demand, Proceedings SEAVEG 2012* (Chiang Mai: Asian Vegetable Research and
789 Development Center), 216–227.
- 790 Ferreira, I. C. F. R., Martins, N., & Barros, L. (2017). Phenolic Compounds and its bioavailability:
791 In vitro bioactive compounds or health promoters? *Advances in Food and Nutrition Research*,
792 82, 1–44.
- 793 Folta, K. M., & Carvalho, S. D. (2015). Photoreceptors and control of horticultural plant traits.
794 *HortScience*, 50, 1274–1280.

- 795 Folta, K. M., & Childers, K. S. (2008). Light as a growth regulator: controlling plant biology with
796 narrow-bandwidth solid-state lighting systems. *HortScience*, *43*, 1957–1964.
- 797 Gao, Y., Yao, Y., Zhu, Y., & Ren, G. (2015). Isoflavone content and composition in chickpea
798 (*Cicer arietinum* L.) sprouts germinated under different conditions. *Journal of Agricultural
799 and Food Chemistry*, *63*, 2701–2707.
- 800 Gerovac, J. R., Craver, J. K., Boldt, J. K., & Lopez, R. G. (2016). Light intensity and quality from
801 sole-source light-emitting diodes impact growth, morphology, and nutrient content of
802 *Brassica* microgreens. *HortScience*, *51*, 497–503.
- 803 Goins, G. D., Yorio, N. C., Sanwo, M. M., & Brown, C. S. (1997). Photomorphogenesis,
804 photosynthesis, and seed yield of wheat plant grown under red light-emitting diodes (LED)
805 with and without supplemental blue lighting. *Journal of Experimental Botany*, *48*, 1407–
806 1413.
- 807 Grażyna, B., Ilona, G.-C., Ewelina, B., Joanna, G., Radosław, M., Dorota, Ż., et al. (2018).
808 Evaluation of estrogenic activity of red clover (*Trifolium pratense* L.) sprouts cultivated
809 under different conditions by content of isoflavones, calorimetric study and molecular
810 modelling. *Food chemistry*, *245*, 324–336.
- 811 Groenbaek, M., Tybirk, E., & Kristensen, H. L. (2018). Glucosinolate and carotenoid content of
812 white-and yellow-flowering rapeseed grown for human consumption as sprouts and seedlings
813 under light emitting diodes. *European Food Research and Technology*, *244*, 1121–1131.
- 814 Guidi, L., Brunetti, C., Fini, A., Agati, G., Ferrini, F., Gori, A., & Tattini, M. (2016). UV radiation
815 promotes flavonoid biosynthesis, while negatively affecting the biosynthesis and the
816 de-epoxidation of xanthophylls: Consequence for photoprotection? *Environmental and*

- 817 *Experimental Botany*, 127, 14–25.
- 818 Heijde, M., & Ulm, R. (2012). UV-B photoreceptor-mediated signalling in plants. *Trends in Plant*
819 *Science*, 17, 230–237.
- 820 Huché-Théliér, L., Crespel, L., Le Gourrierc, J., Morel, P., Sakr, S., & Leduc, N. (2016). Light
821 signaling and plant responses to blue and UV radiations—Perspectives for applications in
822 horticulture. *Environmental and Experimental Botany*, 121, 22–38.
- 823 Inoue, S., Kinoshita, T., Takemiya, A., Doi, M., & Shimazaki, K. (2008). Leaf positioning of
824 *Arabidopsis* in response to blue light. *Molecular plant*, 1, 15–26.
- 825 Islam, M. A., Kuwar, G., Clarke, J. L., Blystad, D. R., Gislerød, H. R., Olsen, J. E., et al. (2012).
826 Artificial light from light emitting diodes (LEDs) with a high portion of blue light results in
827 shorter poinsettias compared to high pressure sodium (HPS) lamps. *Scientia Horticulturae*,
828 147, 136–143.
- 829 Jacob, R. A., & Sotoudeh, G. (2002). Vitamin C function and status in chronic disease. *Nutrition*
830 *in Clinical Care*, 5, 66–74.
- 831 Jenkins, G. I. (2014). The UV-B photoreceptor UVR8: from structure to physiology. *The Plant*
832 *Cell*, 26, 21–37.
- 833 Ji, H. B., Wen, T., Zhou, X. L., & Ying, W. (2016). Combined effects of blue and ultraviolet lights
834 on the accumulation of flavonoids in tartary buckwheat sprouts. *Polish Journal of Food and*
835 *Nutrition Sciences*, 66, 93–98.
- 836 Jiang, Q. (2014). Natural forms of vitamin E: metabolism, antioxidant, and anti-inflammatory
837 activities and their role in disease prevention and therapy. *Free Radical Biology and Medicine*,
838 72, 76–90.

- 839 Jiao, C., Yang, R., & Gu, Z. (2016). Cyclic ADP-ribose and IP3 mediate abscisic acid-induced
840 isoflavone accumulation in soybean sprouts. *Biochemical and Biophysical Research
841 Communications*, 479, 530–536.
- 842 Jiao, C., Yang, R., & Gu, Z. (2017). Cyclic ADP-ribose mediates nitric oxide-guanosine
843 3',5'-cyclic monophosphate-induced isoflavone accumulation in soybean sprouts under
844 UV-B radiation. *Canadian Journal of Plant Science*, 98, 47–53.
- 845 Jiao, C., Yang, R., Zhou, Y., & Gu, Z. (2016). Nitric oxide mediates isoflavone accumulation and
846 the antioxidant system enhancement in soybean sprouts. *Food chemistry*, 204, 373–380.
- 847 Jin, P., Yao, D., Xu, F., Wang, H., & Zheng, Y. (2015). Effect of light on quality and bioactive
848 compounds in postharvest broccoli florets. *Food chemistry*, 172, 705–709.
- 849 Johnson, E. J. (2002). The role of carotenoids in human health. *Nutrition in Clinical Care*, 5, 56–
850 65.
- 851 Johnson, G. A., & Day, T. A. (2010). Enhancement of photosynthesis in *Sorghum bicolor* by
852 ultraviolet radiation. *Physiologia Plantarum*, 116, 554–562.
- 853 Kami, C., Lorrain, S., Hornitschek, P., & Fankhauser, C. (2010). Light-regulated plant growth and
854 development. In *Current Topics in Developmental Biology* . Ed. Timmermans, M. C. P.
855 (Academic Press), 29–66.
- 856 Keum, Y. S., Jeong, W. S., & Kong, A. N. T. (2004). Chemoprevention by isothiocyanates and
857 their underlying molecular signaling mechanisms. *Mutation research*, 555, 191–202.
- 858 Khattak, A. B., Zeb, A., & Bibi, N. (2008). Impact of germination time and type of illumination on
859 carotenoid content, protein solubility and in-vitro protein digestibility of chickpea (*Cicer
860 arietinum* L) sprouts. *Food Chemistry*, 109, 797–801.

- 861 Kim, E. H., Kim, S. H., Chung, J. I., Chi, H. Y., Kim, J. A., & Chung, I. M. (2006). Analysis of
862 phenolic compounds and isoflavones in soybean seeds (*Glycine max* (L.) Merrill) and sprouts
863 grown under different conditions. *European Food Research and Technology*, 222, 201–208.
- 864 Kim, H.H., Goins, G. D., Wheeler, R. M., & Sager, J. C. (2004). Green-light supplementation for
865 enhanced lettuce growth under red-and blue-light-emitting diodes. *HortScience*, 39, 1617–
866 1622.
- 867 Kissen, R., Eberl, F., Winge, P., Uleberg, E., Martinussen, I., & Bones, A. M. (2016). Effect of
868 growth temperature on glucosinolate profiles in *Arabidopsis thaliana* accessions.
869 *Phytochemistry*, 130, 106–118.
- 870 Kopsell, D. A., & Sams, C. (2013). Increases in shoot tissue pigments, glucosinolates, and mineral
871 elements in sprouting broccoli after exposure to short-duration blue light from light emitting
872 diodes. *Journal of the American Society for Horticultural Science*, 138, 31–37.
- 873 Kopsell, D. A., Pantanizopoulos, N. I., Sams, C. E., & Kopsell, D. E. (2012). Shoot tissue pigment
874 levels increase in ‘Florida Broadleaf’ mustard (*Brassica juncea* L.) microgreens following
875 high light treatment. *Scientia Horticulturae*, 140, 96–99.
- 876 Kopsell, D. A., Sams, C., Barickman, T., & Morrow, R. (2014). Sprouting broccoli accumulate
877 higher concentrations of nutritionally important metabolites under narrow-band light-emitting
878 diode lighting. *Journal of the American Society for Horticultural Science*, 139, 469–477.
- 879 Kyriacou, M. C., Roupael, Y., Di Gioia, F., Kyratzis, A., Serio, F., Renna, M., et al. (2016).
880 Micro-scale vegetable production and the rise of microgreens. *Trends in Food Science and*
881 *Technology*, 57, 103–115.
- 882 Lee, D., Nishizawa, M., Shimizu, Y., & Saeki, H. (2017). Anti-inflammatory effects of dulse

- 883 (*Palmaria palmata*) resulting from the simultaneous water-extraction of phycobiliproteins
884 and chlorophyll a. *Food Research International*, *100*, 514–521.
- 885 Lee, S.W., Seo, J. M., Lee, M. K., Chun, J. H., Antonisamy, P., Arasu, M. V., et al. (2014).
886 Influence of different LED lamps on the production of phenolic compounds in common and
887 Tartary buckwheat sprouts. *Industrial Crops and Products*, *54*, 320–326.
- 888 Li, Q., & Yang, H. (2007). Cryptochrome signaling in plants. *Photochemistry and Photobiology*,
889 83, 94–101.
- 890 Li, X., Kim, Y. B., Uddin, M. R., Lee, S., Kim, S. J., & Park, S. U. (2013). Influence of light on
891 the free amino acid content and γ -aminobutyric acid synthesis in *Brassica juncea* seedlings.
892 *Jornal of Agricultural and Food Chemistry*, *61*, 8624–8631.
- 893 Li, X., Thwe, A. A., Park, N. I., Suzuki, T., Kim, S. J., & Park, S. U. (2012). Accumulation of
894 phenylpropanoids and correlated gene expression during the development of tartary
895 buckwheat sprouts. *Jornal of Agricultural and Food Chemistry*, *60*, 5629–5635.
- 896 Li, Y., Dvořák, M., Nesterenko, P. N., Nuchtavorn, N., & Macka, M. (2018). High power deep
897 UV-LEDs for analytical optical instrumentation. *Sensors Actuators B: Chemical*, *255*, 1238–
898 1243.
- 899 Liu, H., Chen, Y., Hu, T., Zhang, S., Zhang, Y., Zhao, T., et al. (2016). The influence of
900 light-emitting diodes on the phenolic compounds and antioxidant activities in pea sprouts.
901 *Journal of Functional Foods*, *25*, 459–465.
- 902 Lobiuc, A., Vasilache, V., Pintilie, O., Stoleru, T., Burducea, M., Oroian, M., et al. (2017). Blue
903 and red led illumination improves growth and bioactive compounds contents in acyanic and
904 cyanic *Ocimum Basilicum* L. microgreens. *Molecules*, *22*, 2111.

- 905 Lu, Y., Dong, W., Alcazar, J., Yang, T., Luo, Y., Wang, Q., et al. (2018). Effect of preharvest CaCl₂
906 spray and postharvest UV-B radiation on storage quality of broccoli microgreens, a richer
907 source of glucosinolates. *Journal of Food Composition and Analysis*, *67*, 55–62.
- 908 Ma, L., Liu, G., Zong, G., Hu, F. B., Rimm, E. B., Willett, W. C., et al. (2018). Intake of
909 glucosinolates and risk of coronary heart disease in three large prospective cohorts of US
910 men and women. *Clinical Epidemiology*, *10*, 749–762.
- 911 Ma, M., Wang, P., Yang, R., & Gu, Z. (2018). Effects of UV-B radiation on the isoflavone
912 accumulation and physiological-biochemical changes of soybean during germination:
913 Physiological-biochemical change of germinated soybean induced by UV-B. *Food chemistry*,
914 *250*, 259–267.
- 915 Martín-Cabrejas, M. A., Díaz, M. F., Aguilera, Y., Benítez, V., Mollá, E., & Esteban, R. M. (2008).
916 Influence of germination on the soluble carbohydrates and dietary fibre fractions in
917 non-conventional legumes. *Food chemistry*, *107*, 1045–1052.
- 918 Márton, M., Mándoki, Z., Csapókiss, Z., & Csapó, J. (2010). The role of sprouts in human
919 nutrition. A review. *Gerontologist*, *31*, 318–324.
- 920 Mazzella, M. A., Cerdán, P. D., Staneloni, R. J., & Casal, J. J. (2001). Hierarchical coupling of
921 phytochromes and cryptochromes reconciles stability and light modulation of *Arabidopsis*
922 development. *Development*, *128*, 2291–2299.
- 923 Mewis, I., Schreiner, M., Nguyen, C. N., Krumbein, A., Ulrichs, C., Lohse, M., et al. (2012).
924 UV-B irradiation changes specifically the secondary metabolite profile in broccoli sprouts:
925 Induced signaling overlaps with defense response to biotic stressors. *Plant Cell Physiology*,
926 *53*, 1546–1560.

- 927 Miao, Y., Chen, Q., Qu, M., Gao, L., & Hou, L. (2019). Blue light alleviates 'red light syndrome'
928 by regulating chloroplast ultrastructure, photosynthetic traits and nutrient accumulation in
929 cucumber plants. *Scientia Horticulturae*, 257, 108680.
- 930 Möglich, A., Yang, X., Ayers, R. A., & Moffat, K. (2010). Structure and function of plant
931 photoreceptors. *Annual Review of Plant Biology*, 61, 21–47.
- 932 Moore, L. V., & Thompson, F. E. (2015). Adults meeting fruit and vegetable intake
933 recommendations-United States, 2013. *MMWR. Morbidity and Mortality Weekly Report*, 64,
934 709–713.
- 935 Moreira-Rodríguez, M., Nair, V., Benavides, J., Cisneros-Zevallos, L., & Jacobo-Velázquez, D. A.
936 (2017a). UVA, UVB light doses and harvesting time differentially tailor glucosinolate and
937 phenolic profiles in broccoli sprouts. *Molecules*, 22, 1065.
- 938 Moreira-Rodríguez, M., Nair, V., Benavides, J., Cisneros-Zevallos, L., & Jacobo-Velázquez, D. A.
939 (2017b). UVA, UVB light, and methyl jasmonate, alone or combined, redirect the
940 biosynthesis of glucosinolates, phenolics, carotenoids, and chlorophylls in broccoli sprouts.
941 *International journal of molecular sciences*, 18, 2330.
- 942 Muscaritoli, M., Amabile, M. I., & Molino, A. (2016). Foods and their components promoting
943 gastrointestinal cancer. *Current Opinion in Clinical Nutrition and Metabolic Care*, 19, 377–
944 381.
- 945 Nam, T. G., Kim, D. O., & Eom, S. H. (2018). Effects of light sources on major flavonoids and
946 antioxidant activity in common buckwheat sprouts. *Food Science and Biotechnology*, 27,
947 169–176.
- 948 Nam, T. G., Lim, Y. J., & Eom, S. H. (2018). Flavonoid accumulation in common buckwheat

- 949 (*Fagopyrum esculentum*) sprout tissues in response to light. *Horticulture Environment and*
950 *Biotechnology*, 59, 19–27.
- 951 Nanya, K., Ishigami, Y., Hikosaka, S., & Goto, E. (2012). Effects of blue and red light on stem
952 elongation and flowering of tomato seedlings. *Acta Horticulturae*, 956, 261–266.
- 953 Neff, M. M., Fankhauser, C., & Chory, J. (2000). Light: an indicator of time and place. *Genes Dev.*,
954 *14*, 257–271.
- 955 Olle, M., & Viršile, A. (2013). The effects of light-emitting diode lighting on greenhouse plant
956 growth and quality. *Agricultural and Food Science*, 22, 223–234.
- 957 Ooms, M. D., Dinh, C. T., Sargent, E. H., Sinton, D. (2016). Photon management for augmented
958 photosynthesis. *Nature communications*, 7, 12699.
- 959 Peng, L., Zou, L., Su, Y., Fan, Y., & Zhao, G. (2015). Effects of light on growth, levels of
960 anthocyanin, concentration of metabolites in *Fagopyrum tataricum* sprout cultures.
961 *International Journal of Food Science and Technology*, 50, 1382–1389.
- 962 Pérez-Balibrea, S., Moreno, D., & García-Viguera, C. (2008). Influence of light on
963 health-promoting phytochemicals of broccoli sprouts. *Journal of the Science of Food and*
964 *Agriculture*, 88, 904–910.
- 965 Phommalth, S., Jeong, Y. S., Kim, Y. H., Dhakal, K. H., & Hwang, Y. H. (2008). Effects of light
966 treatment on isoflavone content of germinated soybean seeds. *Jornal of Agricultural and*
967 *Food Chemistry*, 56, 10123–10128.
- 968 Podsędek, A. (2007). Natural antioxidants and antioxidant capacity of *Brassica* vegetables: A
969 review. *LWT-Food Science and Technology*, 40, 1–11.
- 970 Prior, R. L., Xianli, W., & Karen, S. (2005). Standardized methods for the determination of

- 971 antioxidant capacity and phenolics in foods and dietary supplements. *Journal of Agricultural*
972 *and Food Chemistry*, 53, 4290–4302.
- 973 Qian, H., Liu, T., Deng, M., Miao, H., Cai, C., Shen, W., et al. (2016). Effects of light quality on
974 main health-promoting compounds and antioxidant capacity of Chinese kale sprouts. *Food*
975 *Chemistry*, 196, 1232–1238.
- 976 Quail, P. H. (2002). Phytochrome photosensory signalling networks. *Nature reviews Molecular*
977 *Cell Biology*, 3, 85–93.
- 978 Ramakrishna, A., & Ravishankar, G. A. (2011). Influence of abiotic stress signals on secondary
979 metabolites in plants. *Plant Signaling and Behavior*, 6, 1720-1731.
- 980 Rehman, M., Ullah, S., Bao, Y., Wang, B., Peng, D., & Liu, L. (2017). Light-emitting diodes:
981 whether an efficient source of light for indoor plants? *Environmental Science and Pollution*
982 *Research*, 24, 24743–24752.
- 983 Renna, M., Castellino, M., Leoni, B., Paradiso, V. M., & Santamaria, P. (2018). Microgreens
984 production with low potassium content for patients with impaired kidney function. *Nutrients*,
985 10, 675.
- 986 Rice-Evans, C., Miller, N., & Paganga, G. (1997). Antioxidant properties of phenolic compounds.
987 *Trends in Food Science and Technology*, 2, 152–159.
- 988 Rizvi, S., Raza, S. T., Ahmed, F., Ahmad, A., Abbas, S., & Mahdi, F. (2014). The role of vitamin E
989 in human health and some diseases. *Sultan Qaboos University Medical Journal*, 14, e157–
990 e165.
- 991 Rizzini, L., Favory, J. J., Cloix, C., Faggionato, D., O'Hara, A., Kaiserli, E., et al. (2011).
992 Perception of UV-B by the Arabidopsis UVR8 protein. *Science*, 332, 103–106.

- 993 Ryo, M., Keiko, O. K., Kazuhiro, F., Eiji, G., & Kenji, K. (2004). Photosynthetic characteristics of
994 rice leaves grown under red light with or without supplemental blue light. *Plant Cell*
995 *Physiology*, *45*, 1870–1874.
- 996 Samuolienė, G., Brazaitytė, A., Duchovskis, P., Viršilė, A., Sirtautas, R., Jankauskienė, J., et al.
997 (2014). Light-emitting diodes: a tool for growth and metabolism handle. In *Light Sources*
998 *2014, Proceedings of the 14th International Symposium on the Science and Technology of*
999 *Lighting*, Como, Italy, LP74.
- 1000 Samuolienė, G., Brazaitytė, A., Viršilė, A., Jankauskienė, J., Sakalauskienė, S., & Duchovskis, P.
1001 (2016). Red light-dose or wavelength-dependent photoresponse of antioxidants in herb
1002 microgreens. *PloS one*, *11*, e0163405.
- 1003 Samuolienė, G., Viršilė, A., Brazaitytė, A., Jankauskienė, J., Sakalauskienė, S., Vaštakaitė, V., et al.
1004 (2017). Blue light dosage affects carotenoids and tocopherols in microgreens. *Food*
1005 *Chemistry*, *228*, 50–56.
- 1006 Seo, J. M., Arasu, M. V., Kim, Y. B., Park, S. U., & Kim, S.J. (2015). Phenylalanine and LED
1007 lights enhance phenolic compound production in Tartary buckwheat sprouts. *Food Chemistry*,
1008 *177*, 204–213.
- 1009 Shahidi, F., & Ambigaipalan, P. (2015). Phenolics and polyphenolics in foods, beverages and
1010 spices: Antioxidant activity and health effects—A review. *Journal of Functional Foods*, *18*,
1011 820–897.
- 1012 Shelp, B. J. (1997). The metabolism and functions of γ -aminobutyric acid. *Plant Physiology*, *115*,
1013 1–5.
- 1014 Sluijs, I., Cadier, E., Beulens, J. W. J., van der A, D. L., Spijkerman, A. M. W., & van der Schouw,

- 1015 Y. T. (2015). Dietary intake of carotenoids and risk of type 2 diabetes. *Nutrition, Metabolism*
1016 *and Cardiovascular Diseases, 25*, 376–381.
- 1017 Smith, H. (2000). Phytochromes and light signal perception by plants—an emerging synthesis.
1018 *Nature, 407*, 585–591.
- 1019 Smith, H. L., Mcausland, L., & Murchie, E. H. (2017). Don't ignore the green light: exploring
1020 diverse roles in plant processes. *Journal of Experimental Botany, 68*, 2099–2110.
- 1021 Sood, S., Tyagi, A. K., & Tripathy, B. C. (2004). Inhibition of photosystem I and photosystem II in
1022 wheat seedlings with their root-shoot transition zones exposed to red light. *Photosynthesis*
1023 *Research, 81*, 31–40.
- 1024 Stampfer, M. J., Hennekens, C. H., Manson, J. E., Colditz, G. A., Rosner, B., & Willett, W. C.
1025 (1993). Vitamin E consumption and the risk of coronary disease in women. *New England*
1026 *Journal of Medicine, 328*, 1444–1449.
- 1027 Su, N., Lu, Y., Wu, Q., Liu, Y., Xia, Y., Xia, K., et al. (2016). UV-B-induced anthocyanin
1028 accumulation in hypocotyls of radish sprouts continues in the dark after irradiation. *Journal*
1029 *of the Science of Food and Agriculture, 96*, 886–892.
- 1030 Su, N., Wu, Q., Qi, N., Liu, Y., Li, N., & Cui, J. (2017). Effect of partial shading treatments on
1031 anthocyanin synthesis in the hypocotyls of soybean sprouts under UV-A irradiation. *Jornal*
1032 *of Plant Growth Regulation, 36*, 50–59.
- 1033 Subramoniam, A., Asha, V. V., Nair, S. A., Sasidharan, S. P., Sureshkumar, P. K., Rajendran, K. N.,
1034 et al. (2012). Chlorophyll revisited: Anti-inflammatory activities of chlorophyll a and
1035 inhibition of expression of TNF- α gene by the same. *Inflammation, 35*, 959–966.
- 1036 Świeca, M., Gawlik-Dziki, U., Kowalczyk, D., & Złotek, U. (2012). Impact of germination time

- 1037 and type of illumination on the antioxidant compounds and antioxidant capacity of *Lens*
1038 *culinaris* sprouts. *Scientia Horticulturae*, 140, 87–95.
- 1039 Terashima I, Fujita T, Inoue T, Chow WS, & Oguchi R. (2009) Green light drives leaf
1040 photosynthesis more efficiently than red light in strong white light: revisiting the enigmatic
1041 question of why leaves are green, *Plant and Cell Physiology*, 50, 684–697.
- 1042 Terashima,I.,Fujita,T.,Inoue,T.,Chow,W.S., & Oguchi,R. (2009). Green light drives leaf
1043 photosynthesis more efficiently than red light in strong white light: revisiting the enigmatic
1044 question of why leaves are green. *Plant Cell Physiology*, 50, 684–697.
- 1045 Thwe, A. A., Kim, Y. B., Li, X., Seo, J. M., Kim, S. J., Suzuki, T., et al. (2014). Effects of
1046 light-emitting diodes on expression of phenylpropanoid biosynthetic genes and accumulation
1047 of phenylpropanoids in *Fagopyrum tataricum* sprouts. *Jornal of Agricultural and Food*
1048 *Chemistry*, 62, 4839–4845.
- 1049 Traka, M. H. (2016). Health Benefits of Glucosinolates. In: *Kopriva (Ed.), Glucosinolates,*
1050 *Advances in Botanical Research. Vol.80. Elsevier, Netherlands. 247–279*
- 1051 Tsurunaga, Y., Takahashi, T., Katsube, T., Kudo, A., Kuramitsu, O., Ishiwata, M., et al. (2013).
1052 Effects of UV-B irradiation on the levels of anthocyanin, rutin and radical scavenging activity
1053 of buckwheat sprouts. *Food chemistry*, 141, 552–556.
- 1054 Tuan, P. A., Thwe, A. A., Kim, J. K., Kim, Y. B., Lee, S., & Park, S. U. (2013a). Molecular
1055 characterisation and the light-dark regulation of carotenoid biosynthesis in sprouts of tartary
1056 buckwheat (*Fagopyrum tataricum* Gaertn.). *Food Chemistry*, 141, 3803–3812.
- 1057 Tuan, P. A., Thwe, A. A., Kim, Y. B., Kim, J. K., Kim, S. J., Lee, S., et al. (2013b). Effects of
1058 white, blue, and red light-emitting diodes on carotenoid biosynthetic gene expression levels

- 1059 and carotenoid accumulation in sprouts of tartary buckwheat (*Fagopyrum tataricum* Gaertn.).
1060 *Journal of Agricultural and Food Chemistry*, 61, 12356–12361.
- 1061 Vale, A. P., Cidade, H., Pinto, M., & Oliveira, M. B. P. P. (2014). Effect of sprouting and light
1062 cycle on antioxidant activity of *Brassica oleracea* varieties. *Food chemistry*, 165, 379–387.
- 1063 Vale, A. P., Santos, J., Brito, N. V., Peixoto, V., Carvalho, R., Rosa, E., et al. (2015). Light
1064 influence in the nutritional composition of *Brassica oleracea* sprouts. *Food Chemistry*, 178,
1065 292–300.
- 1066 Vaštakaitė, V. & Viršilė, A. (2015). Light-emitting diodes (LEDs) for higher nutritional quality of
1067 *Brassicaceae* microgreens. *Food Science*, 1, 111–117.
- 1068 Vaštakaitė, V., Viršilė, A., Brazaitytė, A. r., Samuolienė, G., Jankauskienė, J., Novičkovas, A., et
1069 al. (2017). Pulsed light-emitting diodes for a higher phytochemical level in microgreens.
1070 *Jornal of Agricultural and Food Chemistry*, 65, 6529–6534.
- 1071 Verdaguer, D., Jansen, M. A. K., Llorens, L., Morales, L. O., & Neugart, S. (2017). UV-A
1072 radiation effects on higher plants: Exploring the known unknown. *Plant Science*, 255, 72–81.
- 1073 Wang, J., Lu, W., Tong, Y., & Yang, Q. (2016). Leaf morphology, photosynthetic performance,
1074 chlorophyll fluorescence, stomatal development of lettuce (*Lactuca sativa* L.) exposed to
1075 different ratios of red light to blue light. *Frontiers in Plant Science*, 7, 250.
- 1076 Williamson, G., Faulkner, K., & Plumb, G. W. (1998). Glucosinolates and phenolics as
1077 antioxidants from plant foods. *European Journal of Cancer Prevention*, 7, 17–21.
- 1078 Wu, M., Hou, C., Jiang, C., Wang, Y., Wang, C., Chen, H., et al. (2007). A novel approach of LED
1079 light radiation improves the antioxidant activity of pea seedlings. *Food Chemistry*, 101,
1080 1753–1758.

- 1081 Wu, Q., Su, N., Zhang, X., Liu, Y., Cui, J., & Liang, Y. (2016). Hydrogen peroxide, nitric oxide
1082 and UV RESISTANCE LOCUS8 interact to mediate UV-B-induced anthocyanin biosynthesis
1083 in radish sprouts. *Scientific Reports*, *6*, 29164.
- 1084 Xiao, Z., Lester, G. E., Luo, Y., & Wang, Q. (2012). Assessment of vitamin and carotenoid
1085 concentrations of emerging food products: Edible microgreens. *Journal of Agricultural and*
1086 *Food Chemistry*, *60*, 7644–7651.
- 1087 Xiao, Z., Lester, G. E., Luo, Y., Xie, Z. K., Yu, L. L., & Wang, Q. (2014). Effect of light exposure
1088 on sensorial quality, concentrations of bioactive compounds and antioxidant capacity of
1089 radish microgreens during low temperature storage. *Food Chemistry*, *151*, 472–479.
- 1090 Xu, M. J., Dong, J. F., & Zhu, M. Y. (2005). Effects of germination conditions on ascorbic acid
1091 level and yield of soybean sprouts. *Journal of the Science of Food and Agriculture*, *85*, 943–
1092 947.
- 1093 Yan, X., Lee, K., Cheng, S., & Alvarado, J. C. (2019). Tunable LED emitter with continuous
1094 spectrum. In: Google Patents.
- 1095 Yeh, N., & Chung, J. P. (2009). High-brightness LEDs—Energy efficient lighting sources and
1096 their potential in indoor plant cultivation. *Renewable and Sustainable Energy Reviews*, *13*,
1097 2175–2180.
- 1098 Young, A. J. (1991). The photoprotective role of carotenoids in higher plants. *Physiologia*
1099 *Plantarum*, *83*, 702–708.
- 1100 Yu, Y., & Huang, R. (2017). Integration of ethylene and light signaling affects hypocotyl growth in
1101 *Arabidopsis*. *Frontiers in Plant Science*, *8*, 57.
- 1102 Yuan, M., Jia, X., Ding, C., Zeng, H., Du, L., Yuan, S., et al. (2015a). Effect of fluorescence light

- 1103 on phenolic compounds and antioxidant activities of soybeans (*Glycine max* L. Merrill)
1104 during germination. *Food Science and Biotechnology*, 24, 1859–1865.
- 1105 Yuan, M., Jia, X., Yang, Y., Ding, C., Du, L., Yuan, S., et al. (2015b). Effect of light on structural
1106 properties and antioxidant activities of polysaccharides from soybean sprouts. *Process*
1107 *Biochemistry*, 50, 1152-1157.
- 1108 Zhen, S., & van Iersel, M. W. (2017). Far-red light is needed for efficient photochemistry and
1109 photosynthesis. *Journal of Plant Physiology*, 209, 115–122.
- 1110 Zhen, S., Haidekker, M., & van Iersel, M. W. (2019). Far-red light enhances photochemical
1111 efficiency in a wavelength-dependent manner. *Physiologia plantarum*, 167, 21–33.
- 1112 Zhu, M., Geng, S., Chakravorty, D., Guan, Q., Chen, S., & Assmann, S. M. (2019). Metabolomics
1113 of red-light-induced stomatal opening in *Arabidopsis thaliana*: Coupling with abscisic acid
1114 and jasmonic acid metabolism. *The Plant Journal*, doi:10.1111/tpj.14594
- 1115 Zoratti, L., Karppinen, K., Escobar, A. L., Häggman, H., & Jaakola, L. (2014). Light-controlled
1116 flavonoid biosynthesis in fruits. *Frontiers in Plant Science*, 5, 534.

1117 **Figure captions:**

1118 **Figure 1.** The current production methods of sprouts and microgreens.

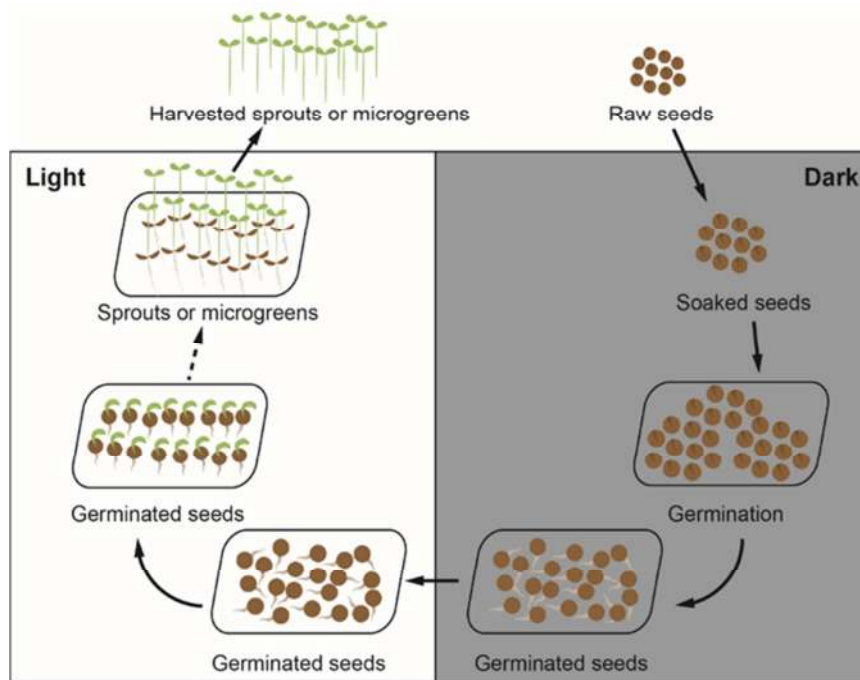
1119 **Figure 2.** The effective spectra for plants and plant photoreceptors. UVR8, UV

1120 RESPONSE LOCUS 8.

1121

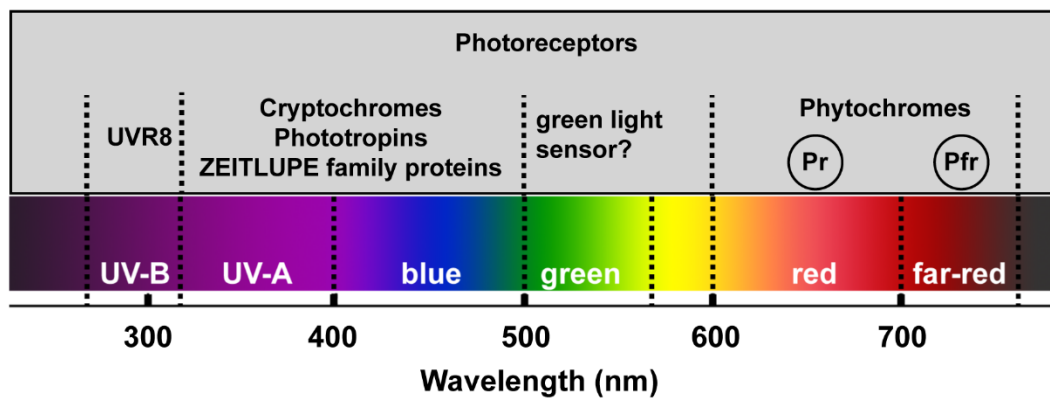
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1122 **Figure 1**



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1124 **Figure 2**



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Table 1. The effects of LED light on the phenolic compounds content of sprouts and microgreens.

Phytochemicals	Quantification	Sprouts/microgreens	germination time	Light conditions			References
				Light quality & light sources	Light intensity or UV-irradiance	Photoperiod illumination (h) or time	
Total Phenolic	+62%	Soybean sprouts (<i>Glycine max</i> L.)	6 days	Dark (Con ^a), Green and yellow (colored cellophane tapes and an acryl film)	NA ^b	96 h	(Kim, et al., 2006)
	-6%	Penca cabbage sprouts (<i>B. oleracea</i> L. var. <i>costata</i> DC)	12 days	Dark (Con), White (LNA ^c)	NA	16 h light/8 h dark photoperiod	(Vale, Cidade, Pinto, & Oliveira, 2014)
	-17%	Broccoli sprouts (<i>B. oleracea</i> L. var. <i>italic</i> Plenck)	7 days				
	+30%	Red cabbage sprouts (<i>B. oleracea</i> L. var. <i>capitata</i> f. <i>rubra</i>)	12 days				
	-17%	Galega kale sprouts (<i>B. oleracea</i> L. var. <i>acephala</i> DC)	7 days				
	+34% under white light, +69% under blue light	Chinese kale sprouts (<i>Brassica oleracea</i>)	7 days	Dark (Con), White (440-660 nm), Red (660 nm), Blue (470 nm) (LED lamps)	30 μ mol m ⁻² s ⁻¹	16 h light/8 h dark photoperiod	(Qian, et al., 2016)
	NQ ^d (significantly improved by red, blue and	Pea sprouts (<i>Pisum sativum</i> L.)	6 days	Dark (Con), Red (635 nm), Blue (460 nm), Yellow (585 nm),	30 μ mol m ⁻² s ⁻¹	12 h light/12 h dark photoperiod	(Liu, et al., 2016)

white light)					white (LED lamps for red, blue and yellow light, fluorescent lamps for white light)				
+10% under blue light, -17% under red light	Common sprouts (<i>Fagopyrum esculentum</i> Möench)	buckwheat	7 days		White (Con), Blue (460 nm), Red (625 nm), (LED lamps for blue and red light, fluorescent lamps for white light)	35 $\mu\text{ mol m}^{-2} \text{ s}^{-1}$	16 h light/8 h dark photoperiod		(Nam, Kim, & Eom, 2018)
+40%	Lentil sprouts (<i>Lens culinaris</i>)		8 days		Dark, White (LNA)	NQ	24 h/day		(Świeca, Gawlik-Dziki, Kowalczyk, & Złotek, 2012)
-34%	Red pak microgreens (<i>Brassica rapa</i> var. <i>rosularis</i>)	choi	10 days		Combination of LEDs (combination of 447 nm, 638 nm, 665 nm and 731 nm) (Con), Red (638 nm) (LED lamps)	300 $\mu\text{ mol m}^{-2} \text{ s}^{-1}$	16 h light/8 h dark photoperiod		(Brazaitytė, et al., 2016)
+13%	Tatsoi microgreens (<i>Brassica rapa</i> var. <i>rosularis</i>)		10 days		HPS light, HPS supplemental red light (638 nm) (HPS lamps, LED lamps)	HPS: 300 $\mu\text{ mol m}^{-2} \text{ s}^{-1}$, HPS 210 $\mu\text{ mol m}^{-2} \text{ s}^{-1}$ + LED 90 $\mu\text{ mol m}^{-2} \text{ s}^{-1}$	16 h light/8 h dark photoperiod		(Samuolienė, et al., 2016)
+58%	basil microgreens (<i>Ocimum basilicum</i> L.)		13 days		Dark (Con), White (fluorescent lamps)	120 $\mu\text{ mol m}^{-2} \text{ s}^{-1}$	12 h light/12 h dark		(Yuan et al., 2015)
NQ (significantly increased under white light)	Soybean sprouts (<i>Glycine max</i> L. Merrill)		3-7 days						

Total Flavonoid	+56% UV-C, +51% UV-A, +24% under blue light	under	Tartary sprouts (<i>Fagopyrum esculentum</i>)	buckwheat	3 days	Dark (Con), UV-C (254 nm), UV-A (365 nm), blue (Fluorescent lamps for blue light & UV lamps for UV light)	UV light: 70.32 W m ⁻² , blue light: 66.93 W m ⁻²	12 h light/12 h dark	(Ji, Wen, Zhou & Ying, 2016)
	NQ (significantly improved by all the light quality treatment)		Pea sprouts (<i>Pisum sativum</i> L.)		6 days	Dark (Con), Red (635 nm), Blue (460 nm), Yellow (585 nm), white (LED lamps for red, blue and yellow light, fluorescent lamps for white light)	30 μ mol m ⁻² s ⁻¹	12 h light/12 h dark photoperiod	(Liu, et al., 2016)
	+12% blue light, -13% under red light	under	Common sprouts (<i>Fagopyrum esculentum</i> Moench)	buckwheat	7 days	White (Con), Blue (460 nm), Red (625 nm), (LED lamps for blue and red light, fluorescent lamps for white light)	35 μ mol m ⁻² s ⁻¹	16 h light/8 h dark photoperiod	(Nam, Kim, & Eom, 2018)
	+92%		Broccoli sprouts (<i>B. oleracea</i> L.)		13 days	White (Con), UV-B (UV lamps)	0.6 kJ m ⁻² d ⁻¹	240 min	(Mewis, et al., 2012)
	+55%		Lentil sprouts (<i>Lens culinaris</i>)		8 days	Dark (Con), White (LNA)	NA	24 h/day	(Świeca, Gawlik-Dziki, Kowalczyk, & Złotek, 2012)

Isoflavone	+176%	Soybean sprouts (<i>Glycine max</i> L. cv Yunhe)	4 days	Dark (Con), UV-B (313 nm) (UV-B light bulb)	4 $\mu\text{W cm}^{-2}$	NA	(Jiao, Yang, & Gu, 2016; 2017)
	+148%	Soybean sprouts (<i>Glycine max</i> L.)	6 days	Dark (Con), Green and yellow (colored cellophane tapes and an acryl film)	NA	96 h	(Kim, et al., 2006)
	+58%	Soybean sprouts (<i>Glycine max</i> L.)	4 days	Dark, UV-B (UV-B lamps)	10 $\mu\text{W cm}^{-2}$	6h light/18 h dark	(Ma, et al., 2018)
	-49%	Red clover sprouts (<i>Trifolium pratense</i> L.)	10 days	White (Con), UV-B (340 nm) (LNA)	NA	24 h/day	(Grażyna, et al., 2018)
	+20%	Soybean sprouts (<i>Glycine max</i> L. cv Aga3)	7 days	Dark (Con), White (greenhouse lamps)	8 $\mu\text{mol m}^{-2} \text{s}^{-1}$	12 h light/12 h dark photoperiod	(Phommalth, Jeong, Kim, Dhakal, & Hwang, 2008)
Anthocyanin	+103%	Buckwheat sprouts (<i>Fagopyrum esculentum</i> Moench.)	NA	White (Con), UV-B (> 300 nm) (fluorescent lamps)	white light: 110 $\mu\text{mol m}^{-2} \text{s}^{-1}$ and 12 W m^{-2} , UV-B: 10 $\mu\text{mol m}^{-2} \text{s}^{-1}$ and 890 W m^{-2}	24 h	(Tsurunaga, et al., 2013)
	+465% under blue light, +454% under white light	Tartary buckwheat sprouts (<i>Fagopyrum sp.</i>)	11 days	Dark (Con), Blue (430 nm), White (380 nm) (LED lamps)	blue light: 177 $\mu\text{mol m}^{-2} \text{s}^{-1}$, white light: 198 $\mu\text{mol m}^{-2} \text{s}^{-1}$	16 h light/8 h dark photoperiod	(Seo, Arasu, Kim, Park, & Kim, 2015)
	NQ (significantly improved by all the light treatment,	Chinese kale sprouts (<i>Brassica oleracea</i>)	7 days	Dark (Con), White (440-660 nm), Red (660 nm), Blue (470 nm)	30 $\mu\text{mol m}^{-2} \text{s}^{-1}$	16 h light/8 h dark photoperiod	(Qian, et al., 2016)

	especially blue light)				(LED lamps)				
	+610%	Tartary buckwheat sprouts (<i>Fagopyrum tataricum</i>)	10 days	Dark (Con), White light (LNA)	4000 LX	16 h light/8 h dark photoperiod			(Peng, Zou, Su, Fan, & Zhao, 2015)
	+65%	Tartary buckwheat sprouts (<i>Fagopyrum tataricum</i> , cv Hokkai T8)	4 days	White (380 nm) (Con), Blue (470 nm) (LED lamps)	50 $\mu\text{ mol m}^{-2} \text{ s}^{-1}$	16 h light/8 h dark photoperiod			(Thwe, et al., 2014)
	NQ (significantly increased under UV-B)	Radish sprouts (<i>Raphanus sativus</i> L.)	4 days	Dark UV-B (UV lamps)	5.5 W m^{-2}	12 h light/12 h dark photoperiod			(Su, et al., 2016)
	NQ (significantly increased under UV-A)	Soybean sprouts (<i>Glycine max</i> L. 'Dongnong 690')	4 days	Dark UV-A (UV lamps)	5.5 W m^{-2}	36 h			(Su, et al., 2017)
	NQ (significantly increased under UV-B)	Radish sprouts (<i>Raphanus sativus</i> L.)	3 days	White (Con), UV-B (LED lamps for white light, UV lamps for UV-B)	white light: 50 $\mu\text{ mol m}^{-2} \text{ s}^{-1}$, UV-B 10 W m^{-2}	36 h			(Wu, et al., 2016)
	+25%	<i>Perilla frutescens</i> (L.) microgreens	NA	White (Con), Red (638 nm) (LED lamps)	300 $\mu\text{ mol m}^{-2} \text{ s}^{-1}$	18 h light/6 h dark photoperiod			(Brazaitytė, et al., 2013)
	+56%	Basil microgreens (<i>Ocimum basilicum</i> L.)	15 days	White (Con), Red:blue light ratio = 1:2 (LED lamps)	120 $\mu\text{ mol m}^{-2} \text{ s}^{-1}$	12 h light/12 h dark photoperiod			(Lobiuc, et al., 2017)
Rutin	+43%	Buckwheat sprouts (<i>Fagopyrum esculentum</i> Moench.)	NA	White (Con), UV-B (> 300 nm) (fluorescent lamps)	white light: 110 $\mu\text{ mol m}^{-2} \text{ s}^{-1}$ and 12 W m^{-2} , UV-B: 10 $\mu\text{ mol m}^{-2} \text{ s}^{-1}$ and 890	24 h			(Tsurunaga, et al., 2013)

	+66%	Soybean sprouts (<i>Glycine max</i> L. Merrill)	6 days	Dark (Con), White (fluorescent lamp)	$100 \mu \text{ mol m}^{-2} \text{ s}^{-1}$	12 h light/12 h dark photoperiod	(Yuan, et al., 2015)
	+8%	common buckwheat sprouts (<i>Fagopyrum esculentum</i>)	12 days	Red+blue (Con), Red (LED lamps)	red+blue light : $9.19 \mu \text{ mol m}^{-2} \text{ s}^{-1}$, red light: $4.75 \mu \text{ mol m}^{-2} \text{ s}^{-1}$	NA	(Lee, et al., 2014)
	+5%	Tartary buckwheat sprouts (<i>Fagopyrum tataricum</i> , cv Hokkai T8)	4 days	White (380 nm) (Con), Blue (470 nm) (LED lamps)	$50 \mu \text{ mol m}^{-2} \text{ s}^{-1}$	16 h light/8 h dark photoperiod	(Thwe, et al., 2014)
Quercetin	+19%	Soybean sprouts (<i>Glycine max</i> L. Merrill)	6 days	Dark (Con), White (fluorescent lamp)	$100 \mu \text{ mol m}^{-2} \text{ s}^{-1}$	16 h light/8 h dark photoperiod	(Yuan, et al., 2015)
	-31%	Lentil sprouts (<i>Lens culinaris</i>)	8 days	Dark (Con), White (LNA)	NQ	24 h/day	(Świeca, Gawlik-Dziki, Kowalczyk, & Złotek, 2012)
Luteolin	+138%	Lentil sprouts (<i>Lens culinaris</i>)	8 days	Dark (Con), White (LNA)	NQ	24 h/day	(Świeca, Gawlik-Dziki, Kowalczyk, & Złotek, 2012)

^a Con: control group;

^b NA: not available information;

^c LNA: light source not available;

^d NQ: not quantitative.

Highlights

1. Sprouts/microgreens are functional foods with multiple health benefits.
2. Sprouts/microgreens respond to spectra by altering the morphology and metabolism.
3. LEDs provide new opportunities to manipulate the quality of produce for markets.
4. An optimized LED light recipe unique for each plant species can be programmed.