

Routine recovery experiments were done by adding known amounts of neomycin to tissue homogenates. After centrifugation for 2 min at $12,000 \times g$ the supernatant fraction² was assayed by a disc plate microbiological assay^{3,4}. For liver, kidney and other body tissues recoveries were complete (table). Recovery from homogenates of inner ear tissues, however, was low and variable, $39 \pm 22\%$ (range, 15–88%). To investigate the cause of the low yields buffer was 'homogenized' without added tissues in glass-glass homogenizers and transferred to polypropylene tubes. Recovery of neomycin from this buffer was similar as with the ear tissues, $42 \pm 21\%$ (range, 21–55%). As only the homogenization but not the subsequent assays were carried out in glass tubes it seems that preparation of tissue

Recovery of neomycin from homogenates

Tissue	Technique	Neomycin recovered (%)
Kidney	Polytron	125 ± 27
Liver	Polytron	91 ± 13
Inner ear	Glass/glass	39 ± 22
None, buffer only	Glass/glass	42 ± 21

Tissue homogenates were prepared in 0.2 M sodium phosphate, pH 8 and standard amounts of neomycin ($5\text{--}20 \mu\text{g}/100 \mu\text{l}$ homogenate) were added. Inner ear tissues are a pool of stria vascularis, spiral ligament and organ of Corti. Numbers are means \pm SD.

homogenates in glass homogenizers can lead to sufficient abrasion of glass to lower the yields of antibiotics. It is important to note that loss of drug during tissue preparation is detected only in recovery studies with the original homogenate because glass particles are removed by centrifugation. Drug standards generated with the supernatant fraction of ear tissue homogenates showed recoveries of 112 and 122%.

A number of variables may influence recovery such as the amount of tissue in relation to the surface of the glass homogenizer; the concentration of the drug; the force with which homogenization is carried out; the use of glass or teflon pestles in connection with a glass tube; the affinity of the aminoglycosides for glass. From these considerations, it seems indicated to avoid glass homogenizers and to conduct recovery experiments by adding the aminoglycoside prior to the homogenization of tissues.

- 1 Acknowledgment. Supported by research grant NS-13792 and Program Project grant NS-05785 from the National Institutes of Health.
- 2 D.C. Grove and W.A. Randall, Assay methods of antibiotics: A laboratory manual. Medical Encyclopedia, Inc., New York 1955.
- 3 W.W. Davis, Appl. Microbiol. 22, 659 (1971).
- 4 W.W. Davis, Appl. Microbiol. 22, 666 (1971).

CORRIGENDUM

S.-C. Chen: A simple synthesis of (E)-3-formylbut-2-enenitrile, and its use as a precursor of isotope-labelled zeatin and (\pm)dihydrozeatin, *Experientia* 37, 543 (1981). On page

544, line 19 from the bottom as well as in the figure (right hand side between formula 4 and 6) it should correctly read $-\text{CoCl}_2$ instead of $-\text{COCl}_2$.

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