A structural model of the active ribosome-bound membrane protein insertase YidC

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SUMMARY

The integration of most membrane proteins into the cytoplasmic membrane of bacteria occurs co-translationally. The universally conserved YidC protein mediates this process either individually as a membrane protein insertase, or in concert with the SecY complex (Dalbey et al. 2011; Kol et al. 2008). Here, we present a structural model of YidC based on evolutionary co-variation analysis (Hopf et al. 2012), lipid-versus-protein-exposure (Lai et al. 2013) and molecular dynamics simulations. The model suggests a distinctive arrangement of the conserved five transmembrane domains and a helical hairpin between transmembrane segment 2 (TM2) and TM3 on the cytoplasmic membrane surface. The model was used for docking into a cryo-electron microscopy reconstruction of a translating YidC-ribosome complex carrying the YidC substrate F_0c. This structure reveals how a single copy of YidC interacts with the ribosome at the ribosomal tunnel exit and identifies a site for membrane protein insertion at the YidC protein-lipid interface. Together, these data suggest a mechanism for the co-translational mode of YidC-mediated membrane protein insertion.

TEXT

At present, a mechanistic understanding of the function of YidC, as well as its mitochondrial and chloroplast counterparts Oxa1 and Alb3, respectively, is limited by a lack of structural information. High resolution structures are available only for the first periplasmic domain (P1) of *Escherichia coli* YidC (Fig. 1a) (Oliver and Paetzel 2008; Ravaud et al. 2008), however, this domain is poorly conserved, only present in Gram-negative bacteria and not essential for functionality (Jiang et al. 2003). Furthermore, the region(s) of YidC mediating the interaction with the ribosome have not been identified, and the oligomeric state of YidC during co-translational translocation remains controversial (Herrmann 2013; Kedrov et al. 2013; Kohler et al. 2009). Hence, we set out to determine a molecular model of ribosome-bound YidC during co-translational translocation of the substrate F_0c (van der Laan et al. 2004), an integral membrane subunit of the ATP synthase complex.

In order to build an initial structural model of YidC, we predicted contacts between pairs of residues based on covariation analysis (Hopf et al. 2012; Marks et al. 2011). For that purpose, we constructed a multiple sequence alignment of *E. coli* YidC excluding the nonconserved first transmembrane helix (TM1) and the P1 domain (Fig. 1a) and computed direct evolutionary couplings between pairs of YidC residues (Kamisetty et al. 2013). The resulting matrix of coupling strengths (Fig. 1b) contains several diagonal and anti-diagonal patterns of stronger coupling coefficients, which are indicative of parallel or anti-parallel helix-helix pairs, respectively. We computed probabilities for each possible helix-helix contact by aggregating the evidence of stronger coupling coefficients over the expected interaction patterns and calibrating the resulting raw scores on an independent dataset of helix-helix interactions to obtain accurate interaction probabilities. Seven helix-helix contacts attained probabilities above 57% (Fig. 1b-d) while all other possible contacts scored below 15%, demonstrating the specificity of the method (Fig. 1-figure supplement 1b).

We roughly positioned the five TM helices of *E. coli* YidC relative to each other using the predicted helix-helix contacts as constraints, and rotated them according to their predicted lipid or protein exposure (Lai et al. 2013) (Fig. 1c).

Next, we used MODELLER (Eswar et al. 2008) to create full length models based on the TM core, secondary structure prediction and the 50 residue-residue contacts with the highest coupling coefficients (39 excluding intrahelical contacts, indels and topology violations). In the resulting model (Fig. 1e & f), the conserved membrane integrated core of YidC forms a helical bundle arranged like the vertices of a pentagon, in the order 4-5-3-2-6 (clockwise) when viewed from the cytoplasm (Fig. 1f). Notably, all the predicted interactions between TM domains can be explained by monomeric YidC suggesting that dimer or oligomer formation may not be strictly required for YidC activity (see also below).

Outside the membrane region, strong helix-helix contacts were predicted within the cytoplasmic loop between TM2 and TM3, which can be explained the by formation of a helical hairpin (Fig. 1f). The base of this "helical paddle domain" (HPD) is structurally constrained by predicted contacts with TM3, its tip on the other hand is more mobile and appears to interact with lipid headgroups (see below).

While this manuscript was under review, two crystal structures were published of *Bacillus halodurans* YidC2 (BhYidC2, 34% sequence identity with *E. coli* YidC) (Kumazaki et al. 2014), providing us with a unique opportunity to directly assess the accuracy of our model. Overall, the root mean square deviation (RMSD) between the TM helices of our model and those of BhYidC2 is 7.5 Å (3WO6) and 7.3 Å (3WO7), which is within the resolution limits of our method. The global arrangement of TM helices is the same as in BhYidC2, yet, their tilt angle relative to the plane of the membrane is slightly different (Fig. 2). The tilt angle of the HPD also differs, as well as its side that faces the membrane (Video 1), which may be indicative of a high degree of flexibility of this domain, consistent with its high crystallographic B-factors (Kumazaki et al. 2014). Notably, the HPD is not essential for YidC function in *E. coli* since the deletion of the entire domain is possible without compromising cell viability (Jiang et al. 2003).

A qualitative difference between our model and BhYidC2 that may have more mechanistic importance is the relative position of TM3. In the structure of BhYidC2 a hydrophilic groove is formed on the cytoplasmic side of the TM bundle that has been proposed to form a binding site for YidC substrates (Kumazaki et al. 2014). Interestingly, the opening state of this groove differs

between the two crystal forms, *i.e.* it is more open in 3WO6 than in 3WO7 (Video 1), largely due to movement of the N-terminal half of TM3 (Fig. 2-figure supplement 1). In our model on the other hand, this hydrophilic groove is even more closed than in 3WO7 because we imposed covariation-based constraints between TM3 and TM5 (Pro^{425}-Pro^{499}) and between TM3 and TM6 (Cys^{423}-Gln^{528} & Phe^{433}-Thr^{524}) (Fig. 2 and Video 1). Strikingly, in BhYidC2 the distances between the Cβ atoms of these three pairs are outliers compared to other residue-residue pairs (20.5 Å/20.9 Å/14.9 Å *vs* an average of 8.2 Å, Fig. 2-figure supplement 2). Thus, given that (i) the position of TM3 differs in the two crystal forms, and (ii) that covariation analysis predicts with high accuracy a closer interaction of TM3 with TM6 and one contact with TM5, we conclude that movement of TM3 is a genuine feature of YidC. This movement and the accompanying dynamics of the hydrophilic groove may represent a crucial step in the functional cycle of the YidC insertase.

In summary, the overall structure of our YidC model agrees well with the BhYidC2 crystal structure, and a comparison of both structures reveals dynamic regions in YidC that may be of mechanistic importance. This further illustrates the power of covariation analysis not merely for structure prediction but also for obtaining dynamic insights (Hopf et al. 2012).

Next, in order to further characterize and validate our obtained YidC model, we assessed its stability and biochemical properties in the bacterial membrane by employing traditional molecular dynamics (MD) simulations. Overall, the model was found to be very stable during the simulation. While the five TM helices enable a rigid protein core, the polar loop regions tend to swim on the membrane surface (Fig. 3a). An analysis of inter-residue interactions within the TM region (Fig. 3b) provides a firm basis to the observed stability of YidC: hydrophobic residues on the exterior of the TM bundle stabilize interactions with the apolar lipid tails. The YidC core, in turn, is stabilized both via short and long-range interactions between the five helices. Residues towards the cytoplasmic side of the core are primarily polar or charged and, therefore, engaged in strong electrostatic or charge-dipole interactions. In contrast, residues on the periplasmic side are primarily aromatic and involved in stacking and other nonpolar dispersion interactions.

In order to verify the functional relevance of residues suggested by the MD simulations, we created alanine mutants and subjected them to an *in vivo* complementation assay. Some of the most stabilizing residues, T362 in TM2 and Y517 in TM6, both of which are located at the same height in the membrane, completely inactivated YidC when mutated to alanine (Fig. 3d & Fig. 3-figure supplement 1). Both mutants were stably expressed, indicating that the lack of complementation was not caused by instability of YidC (Fig. 3-figure supplement 2). Several residues close to this pair show intermediate activity levels (F433, M471 and F505), whereas residues further away do not show an effect (Fig. 3-figure supplement 1). Taken together, we provide a model for the overall arrangement of the conserved domains of YidC that is in good agreement with our covariation analysis, lipid exposure prediction, MD simulation, *in vivo* complementation analysis as well as the recent crystal structures.

Interestingly, we observed that YidC induces thinning of the lipid bilayer during the MD simulation. A significant thinning of 7-10 Å results from the hydrophobic mismatch between the TM helices and the membrane (Fig. 3e). The thinning is similar in the upper and lower leaflet, and the thinnest region is in proximity of TM3 and TM5. Since membrane inserting YidC substrates have been chemically cross-linked to both these helices (Klenner and Kuhn 2012; Klenner et al. 2008; Yu et al. 2008), we argue that thinning of this region in particular may be relevant for the molecular mechanism of YidC-dependent membrane insertion. In addition, the distribution of hydrophilic and hydrophobic residues within YidC revealed the presence of a hydrophilic environment on the cytoplasmic side of the YidC TM bundle (Fig. 3f), which continues into the mentioned hydrophobic cluster of aromatic residues towards the periplasmic side. It is tempting to speculate that this hydrophilic environment may receive the polar termini and loops of YidC substrates during the initiation of translocation, thus facilitating their transfer across the hydrophobic core of the (thinned) lipid bilayer (see below). Notably, essentially the same conclusions have been drawn on the basis of the BhYidC2 crystal structures and accompanying cross-linking studies (Kumazaki et al. 2014).

In order to provide a molecular model of YidC in its active state, we reconstituted purified full length YidC (extended with the C-terminus of *R. baltica* YidC (Seitl et

al. 2014)) with ribosome nascent chains (RNCs) exposing the first TM helix of F_Oc, and subjected the complex to cryo-EM and single particle analysis to a resolution of ~8 Å (Fig. 4a & b). In agreement with previous structural studies (Kohler et al. 2009; Seitl et al. 2014), YidC binds to the ribosomal exit site, however, the improved resolution now allows for a more detailed interpretation. Firstly, we were able to separate the weaker electron density of the detergent micelle from that of YidC (Fig. 4a). Secondly, the presence of elongated structural features (Fig. 4d-f) allowed us to dock our molecular model in a distinct orientation (cross correlation coefficient 0.865). Following placement of the YidC-core model, two prominent densities in the membrane region, one next to TM3 and one next to TM5, remained unaccounted for. These could be attributed to either TM1 of YidC or to the TM helix of the nascent chain (NC) F_Oc. Given that (i) YidC substrates are known to crosslink to TM3 (Klenner and Kuhn 2012; Klenner et al. 2008; Yu et al. 2008), and (ii) that the density neighboring TM3 is aligned with the ribosomal exit tunnel and (iii) that at the same relative position nascent chains have been observed inside the SecY channel (Frauenfeld et al. 2011) (Fig. 4-figure supplement 1), the most plausible assignment to the density near TM3 appeared to be the TM helix of F_Oc. To verify this, and to exclude that the density neighboring TM5 corresponds to the nascent chain, we reconstituted single cysteine mutants of YidC either in TM3 (M430C and P431C) or in TM5 (V500C and T503C) with RNCs of a single cysteine mutant of F_Oc(G23C), and exposed them to disulphide crosslinking. Upon exposure to the oxidator DTNB, only in the TM3 mutants a DTT-sensitive ~90 kDa product appeared that reacted with antibodies against the nascent chain (NC-tRNA~30 kDa, Fig. 4c) as well as YidC (~60 kDa, Fig. 4c). Thus, the adduct represented indeed the inserting F_Oc TM domain crosslinked to TM3 of YidC. RNCs lacking a cysteine in the nascent chain (Fig. 4-figure supplement 2) or YidC mutants with cysteines in TM5 did not yield any crosslinks (Fig. 4c). Hence, we conclude that the unaccounted electron density next to TM3 represents the TM of the nascent chain, and that the density neighbouring TM5 represents TM1 of YidC (Fig. 4d-f).

We attribute the remaining unaccounted electron density in the periplasmic region to the P1 domain; however, because it is substantially smaller than the crystal structure of P1, we did not include it in our molecular model. Flexibility

relative to the conserved membrane region of YidC is the most likely explanation for this finding. We did not observe density for the HPD, in agreement with its flexibility observed in both, the crystal structures of BhYidC2 and the MD simulations (Fig. 3c).

In order to validate our molecular model of co-translationally active YidC, we mutated residues that would be in direct contact with the ribosome (Fig. 5a & b) and analyzed their effect on functionality in the *in vivo* complementation test. Indeed, mutation of residues Y370A and Y377A (contacting ribosomal RNA helix 59) and D488K (contacting ribosomal protein uL23) severely interfere with YidC activity (Fig. 5c & Fig. 5-figure supplement 1) thereby emphasizing their functional significance. All these mutants were stably expressed, indicating that the lack of complementation was not caused by instability of YidC (Fig. 5-figure supplement 2). Given that YidC in general is known to be very tolerant to point mutations (Jiang et al. 2003), this provides further support for the overall correctness of our model of ribosome-bound YidC during membrane protein insertion.

Finally, it is notable that we observe only a single monomer of YidC bound to the active ribosome. This is in agreement with recent literature showing clearly that both YidC (Herrmann 2013; Kedrov et al. 2013; Seitl et al. 2014) and the SecY complex (Frauenfeld et al. 2011; Park et al. 2014; Park and Rapoport 2012; Taufik et al. 2013) can be fully active as monomers. However, the comparison of models for active YidC and active SecY (Fig. 5e & Fig. 4-figure supplement 1) reveals an important difference between the two proteins that has mechanistic implications. While SecY is known to translocate hydrophilic nascent chains through its central aqueous channel (Cannon et al. 2005; Driessen and Nouwen 2008; Rapoport 2007) and insert TM domains through a lateral gate (Gogala et al. 2014; Van den Berg et al. 2004), our model suggests that the YidC substrates are inserted at the protein-lipid interface. Two principal findings of our work suggest how YidC may facilitate this process: (i) it provides a hydrophilic environment within the membrane core for receiving the hydrophilic moieties (termini or loops) of a substrate, and (ii) it reduces the thickness of the lipid bilayer: initial interaction of the hydrophilic moieties of YidC substrates with the hydrophilic environment of YidC would allow for a partial insertion into the

membrane, while facilitating exposure of the hydrophobic TM domains to the hydrophobic core of the bilayer. The latter in turn may compensate for the energetic penalty of driving the hydrophilic moieties across the (already thinned) bilayer. Further biochemical and structural studies that capture the earlier stages of this translocation process are eagerly awaited to fully elucidate this mechanism.

METHODS

Covariation analysis

We constructed a multiple sequence alignment of YidC excluding the unconserved first transmembrane helix (TM1) and the periplasmic P1 domain. We searched for homologous sequences of *E. coli* YidC starting from the PFAM seed alignment of family PF02096 (Punta et al. 2012) and using the sensitive homology detection software HHblits (Remmert et al. 2012). First, 5 iterations of HHblits were run against the clustered Uniprot database with no filtering, to retrieve as many homologous sequences as possible. Then, we post-processed the alignment using HHfilter to generate a non-redundant alignment at 90% sequence identity. This resulted in an alignment containing 2366 sequences aligned across YidC helices TM2-TM6. Using this multiple sequence alignment, we computed direct evolutionary couplings between pairs of YidC residues using the method of Kamisetty et al (Kamisetty et al. 2013).

To compute probabilities for each possible helix-helix contact, we aggregated the evidence of stronger coupling coefficients over the expected interaction patterns for helix-helix contacts, taking into account the expected periodicity of ~3.5 residues per alpha helix turn. We built three non-redundant datasets of mainly-alpha proteins from the CATH database (Sillitoe et al. 2013). For each protein, we slid a square pattern (of size 17x17 residues = 289 cells) over the matrix of coupling strengths. For each pattern position, we used Bayes theorem to calculate the raw probability for a helix-helix interaction, given the 289 coupling strengths. The distributions of coupling strengths for interacting and non-interacting helix residues were fitted on dataset #1 (1118 proteins). We assigned different weights to the pattern cells, depending on their position within the pattern and the direction of the helix-helix interaction (parallel or antiparallel); these weights were optimized on dataset #2 (204 proteins). Finally, we calibrated the resulting raw scores on dataset #3 (85 proteins) to obtain accurate interaction probabilities. For cross-validation purposes, we also performed optimization on dataset #3 and calibration on dataset #2. Optimization on either dataset #2 or dataset #3 results in the same choice of weights for the pattern cells. The final posterior probabilities were obtained as the average of the values calibrated on datasets #2 and #3, weighted by dataset size. The calibration plots for datasets #2 and

#3 are shown in Fig. 1-figure supplement 1a. The histogram of final posterior probabilities obtained for YidC is shown in Fig. 1-figure supplement 1b, which illustrates the specificity of the helix-helix predictions.

YidC initial model building

The conserved TM helices of *E.coli* YidC were positioned according to the covariation based helix-helix contact prediction, and rotated based on their predicted lipid or protein exposure (Lai et al. 2013), resulting in a starting model of the conserved TM core of YidC. Additional information based on direct residue-residue interactions (covariance analysis) and secondary structure predictions by Jpred 3 (Cole et al. 2008) were used as structural restraints in MODELLER (Eswar et al. 2008). From a total of 10 output models that differed mainly in the relative orientation of the loop regions, the model that satisfied the imposed constraints best was used for further studies.

Molecular dynamics simulation

System preparation

All simulations were performed with the MD software NAMD 2.9 using the CHARMM36 force field for the proteins and lipids (Klauda et al. 2010). The TIP3P model is used to simulate water (Jorgensen et al. 1983). The YidC model was inserted into the membrane, solvated, and ionized using the Membrane Builder tools on CHARMM-GUI (Jo et al. 2008). The lipid composition is chosen to be 3 POPE to 1 POPG, as has been successfully used for modelling bacterial membranes in several past MD simulations (Ash et al. 2004; Mondal et al. 2013). An initial membrane surface of area 110 Å x 110 Å was constructed along the XY plane. The protein lipid-construct was solvated with 25 Å thick layers of water along the Cartesian Z directions, and ionized to charge neutralization using Monte Carlo sampling of Na^+ and Cl^- ions at 0.15 M concentration. The overall system size is 0.15 M. Prior to simulation the system was subjected to 10000 steps of conjugate gradient energy minimization, followed by 100 ps of thermalization and 25 ns of equilibration. During the first 10 ns of the equilibration stage, the protein was kept fixed, allowing the lipids, ions and water molecules to equilibrate. Subsequent 15 ns of equilibration included the protein as well. We then performed 500 ns of MD simulation at 300K.

315 The final 100 ns was repeated thrice to examine the statistical significance of the
316 result.
317 *Simulation parameters*
318 The systems were kept at constant temperature using Langevin dynamics for all non-
319 hydrogen atoms with a Langevin damping coefficient of $5ps^{-1}$. A constant pressure of
320 1atm was maintained using the Nose-Hoover Langevin piston with a period of 100fs
321 and damping timescale of 50fs. Simulations were performed with an integration time
322 step of 1fs where bonded interactions were computed every time step, short-range
323 non-bonded interactions every two time steps, and long range electrostatic
324 interactions every four time steps. A cutoff of 12 Å was used for van der Waals and
325 short-range electrostatic interactions: a switching function was started at 10 Å for van
326 der Waals interactions to ensure a smooth cutoff. The simulations were performed
327 under periodic boundary conditions, with full-system, long-range electrostatics
328 calculated by using the PME method with a grid point density of 1/ Å. The unit cell
329 was large enough so that adjacent copies of the system did not interact via short-range
330 interactions.
331 *Flexibility analysis*
332 The overall flexibility of the transmembrane helices relative to their average
333 configuration was compared. Positional variance of the helix residues was quantified
334 as a measure of their flexibility. Positional variance was computed by summing the
335 deviation of individual backbone atom position and dividing by the number of
336 backbone atoms in the loop. This measure is slightly different from the usual root
337 mean square fluctuation (RMSF) as contributions from overall displacements of the
338 helices and their motions relative to the rotation/translation and internal motions of
339 the protein are included to probe flexibility.
340 *Interaction energy, hydrogen bonds, and membrane thickness analysis*
341 To further understand the details of the structure and dynamics of the YidC model we
342 performed interaction energy, hydrogen bond, and membrane thinning analysis. These
343 analyses were carried out on the MD trajectory using standard tools available on
344 VMD. In particular, interaction energies were computed for each trajectory frame of
345 the final 100 ns simulation using the NAMD Energy plugin on VMD. The numbers
346 were then time averaged over the entire 100ns, locally averaged for every residue
347 over a cut-off distance of 10 Å, and plotted on the structure in Fig. 3b. Hydrogen
348 bonds are defined solely on the basis of geometric parameters (bond angle: 20°; bond-

length: 3.8 Å) between donors and acceptors. Thickness at a given point on the membrane surface was probed by finding the nearest lipid head group and measuring the minimum distance between the phosphate on that lipid head and one on the opposite leaflet.

Purification of ribosome nascent chain complexes (RNCs)

RNC constructs encoding residues 1-46 of F_Oc (preceded by an N-terminal His-tag and 3C rhinoprotease cleavage site, and followed by an HA-tag and TnaC stalling sequence) were cloned into a pBAD vector (Invitrogen) by standard molecular biology techniques, and expressed and purified as described in Bischoff *et al.*, (submitted). Briefly, *E.coli* KC6 Δ*smpB*Δ*ssrA* (Seidelt et al. 2009) carrying the plasmid for F_Oc was grown in LB with 100 µg/ml ampicilin at 37°C to an OD_{600} = 0.5 and expression was induced for 1h by adding 0.2 % arabinose. Cells were lysed and debris was removed by centrifugation for 20 min at 16.000 rpm in a SS34-rotor (Sorvall). The cleared lysate was spun overnight through a sucrose cushion at 45.000 rpm in a Ti45 rotor (Beckmann), the ribosomal pellet was resuspended for 1 h at 4°C and RNCs were purified in batch by affinity purification using Talon (Clontech). After washing the Talon beads with high salt buffer the RNCs were eluted and loaded onto a linear 10 % - 40 % sucrose gradient. The 70S peak was collected, RNCs were concentrated by pelleting, resuspended in an appropriate volume of RNC Buffer (20 mM HEPES pH 7.2, 100 mM KOAc, 6 mM $MgOAc_2$, 0.05% (w/v) dodecyl maltoside), flash frozen in liquid N_2 and stored at -80°C. The complete sequence of the nascent chain is:

MGHHHHHHHHDYDIPTTLEVLFQGPGTMENLNMDLLYMAAAVMMGLAAI
GAAIGIGILGGKFLEGAARQPDLIYPYDVPDYAGPNILHISVTSKWFNIDNKIV
DHRP

Purification of YidC

For purification and reconstitution studies, *E.coli* YidC extended with the C-terminus from *R. baltica* (Seitl et al. 2014) was re-cloned into pET-16 (Novagen) with an N-terminal His-tag followed by a 3C rhinovirus protease site. Expression and purification was performed essentially as described (Lotz et al. 2008). Briefly, *E.coli* C43(DE3) cells (Miroux and Walker 1996) harboring the YidC construct were grown at 37°C to an OD_{600} = 0.6 and expression was induced by adding 0.5 mM IPTG. YidC

was solubilized with Cymal-6 (Anatrace) and purified by affinity chromatography using TALON (Clontech). The N-terminal His-tag of the eluted protein was cleaved off with 3C protease during overnight dialysis at 4°C, followed by gel filtration chromatography (Superdex 200, GE Healthcare). Fractions of the monodisperse peak were pooled, concentrated to ~ 1 mg/ml in YidC Buffer (20 mM $NaPO_4$ pH 6.8, 100 mM KOAc, 10 % glycerol, 0.05 % Cymal-6) and directly used for further structural or biochemical assays.

Disulphide crosslinking

For disulphide crosslink analysis, $F_{OC}^{(G23C)}$-RNCs and single cysteine mutants of YidC were purified separately and reconstituted by incubating 100 pmol of RNCs with 500 pmol of freshly purified YidC for 30 min at 37°C. The endogenous cysteine in YidC at position 423 was replaced by serine. Disulphide crosslinking was induced by adding 1 mM 5,5'-dithiobis-(2-nitrobenzoicacid) (DTNB) for 10 min at 4°C and quenched by adding 20 mM N-Ethylmaleimide (NEM) for 20 min at 4°C. Crosslinked RNC-YidC complexes were separated from non-crosslinked YidC using a 10 % - 40 % linear sucrose gradient, and the 70S peak was harvested and analysed by SDS-PAGE followed by western blotting.

Complementation assay

For *in vivo* complementation studies, wildtype *E. coli* YidC was recloned into pTrc99a (Pharmacia), and mutants were created by standard molecular cloning techniques. *E.coli* FTL10 cells (Hatzixanthis et al. 2003) harboring pTrc99a plasmids encoding the YidC variants were grown overnight at 37°C in LB medium supplemented with 100 µg/ml ampiciline, 50 µg/ml kanamycin and 0.2% arabinose. YidC depletion was carried out by transferring the cells to LB medium supplemented with 100 µg/ml ampiciline, 50 µg/ml kanamycin and 0.2% glucose, followed by and additional incubation for 3h at 37°C. Cell suspensions of all constructs were adjusted to OD_{600} = 0.1 and either loaded onto SDS-PAGE gels for subsequent Western blotting, or further diluted to $OD_{600} = 10^{-5}$. Each dilution was spotted on LB agar plates supplemented 100 µg/ml ampiciline, 50 µg/ml kanamycin and either 0.2% arabinose or 0.2% glucose, and incubated overnight at 37°C.

Electron microscopy and image processing

For cryo-EM analysis, F_Oc-RNC:YidC complexes were reconstituted by incubating 10 pmol of RNCs with 100 pmol of freshly purified YidC for 30 min at 37°C in a final volume of 50 µl of RNC buffer. Samples were applied to carbon-coated holey grids according to standard methods (Wagenknecht et al. 1988). Micrographs were collected under low-dose conditions on a FEI TITAN KRIOS operating at 200 kV using a 4k x 4k TemCam-F416 CMOS camera and a final pixel size of 1.035Å on the object scale.

Image processing was done using the SPIDER software package (Shaikh et al. 2008). The defocus was determined using the TF ED command in SPIDER followed by automated particle picking using Signature (Chen and Grigorieff 2007). The machine-learning algorithm MAPPOS (Norousi et al. 2013) was used to subtract "false positive" particles from the data set and initial alignment was performed using an empty 70S ribosome as reference. The complete data set (876376 particles) was sorted using competitive projection matching in SPIDER followed by focused sorting for ligand density (Leidig et al. 2013), and refined to a final resolution of ~8.0 Å (Fourier shell correlation (FSC) cut-off 0.5). The final dataset consisted of 58960 particles showing electron density for P-site tRNA and ligand density at the tunnel exit.

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Author contributions

SW performed purifications, reconstitutions, cryo-EM image processing and model building, LB contributed to purifications and image processing, JA, SS and JS performed covariation analyses, AS and KS performed MD simulations and related analyses, OB performed cryo-EM data collection, EvdS designed experiments and supervised the project together with RB. All authors contributed to data interpretation and writing of the manuscript.

REFERENCES

1 Ash, W. L., Zlomislic, M. R., Oloo, E. O. & Tieleman, D. P. 2004. Computer simulations of membrane proteins. *Biochim Biophys Acta* **1666**, 158-189, doi:10.1016/j.bbamem.2004.04.012.

2 Cannon, K. S., Or, E., Clemons, W. M., Jr., Shibata, Y. & Rapoport, T. A. 2005. Disulfide bridge formation between SecY and a translocating polypeptide localizes the translocation pore to the center of SecY. *J Cell Biol* **169**, 219-225, doi:10.1083/jcb.200412019.

3 Chen, J. Z. & Grigorieff, N. 2007. SIGNATURE: a single-particle selection system for molecular electron microscopy. *J Struct Biol* **157**, 168-173, doi:10.1016/j.jsb.2006.06.001.

4 Cole, C., Barber, J. D. & Barton, G. J. 2008. The Jpred 3 secondary structure prediction server. *Nucleic Acids Res* **36**, W197-201, doi:10.1093/nar/gkn238.

5 Dalbey, R. E., Wang, P. & Kuhn, A. 2011. Assembly of bacterial inner membrane proteins. *Annu Rev Biochem* **80**, 161-187, doi:10.1146/annurev-biochem-060409-092524.

6 Driessen, A. J. & Nouwen, N. 2008. Protein translocation across the bacterial cytoplasmic membrane. *Annu Rev Biochem* **77**, 643-667, doi:10.1146/annurev.biochem.77.061606.160747.

7 Eswar, N., Eramian, D., Webb, B., Shen, M. Y. & Sali, A. 2008. Protein structure modeling with MODELLER. *Methods Mol Biol* **426**, 145-159, doi:10.1007/978-1-60327-058-8_8.

8 Frauenfeld, J. *et al.* 2011. Cryo-EM structure of the ribosome-SecYE complex in the membrane environment. *Nat Struct Mol Biol* **18**, 614-621, doi:10.1038/nsmb.2026.

9 Gogala, M. *et al.* 2014. Structures of the Sec61 complex engaged in nascent peptide translocation or membrane insertion. *Nature* **506**, 107-110, doi:10.1038/nature12950.

10 Hatzixanthis, K., Palmer, T. & Sargent, F. 2003. A subset of bacterial inner membrane proteins integrated by the twin-arginine translocase. *Mol Microbiol* **49**, 1377-1390.

11 Herrmann, J. M. 2013. The bacterial membrane insertase YidC is a functional monomer and binds ribosomes in a nascent chain-dependent manner. *J Mol Biol* **425**, 4071-4073, doi:10.1016/j.jmb.2013.08.003.

12 Hopf, T. A. *et al.* 2012. Three-dimensional structures of membrane proteins from genomic sequencing. *Cell* **149**, 1607-1621, doi:10.1016/j.cell.2012.04.012.

13 Jiang, F. *et al.* 2003. Defining the regions of Escherichia coli YidC that contribute to activity. *J Biol Chem* **278**, 48965-48972, doi:10.1074/jbc.M307362200.

14 Jo, S., Kim, T., Iyer, V. G. & Im, W. 2008. CHARMM-GUI: a web-based graphical user interface for CHARMM. *J Comput Chem* **29**, 1859-1865, doi:10.1002/jcc.20945.

15 Jorgensen, W. L., Chandrasekhar, J., Madura, J. D., Impey, R. W. & Klein, M. L. 1983. Comparison of Simple Potential Functions for Simulating Liquid

Water. *Journal of Chemical Physics* **79**, 926-935, doi:Doi 10.1063/1.445869.

16 Kamisetty, H., Ovchinnikov, S. & Baker, D. 2013. Assessing the utility of coevolution-based residue-residue contact predictions in a sequence- and structure-rich era. *Proc Natl Acad Sci U S A* **110**, 15674-15679, doi:10.1073/pnas.1314045110.

17 Kedrov, A. *et al.* 2013. Elucidating the native architecture of the YidC: ribosome complex. *J Mol Biol* **425**, 4112-4124, doi:10.1016/j.jmb.2013.07.042.

18 Klauda, J. B. *et al.* 2010. Update of the CHARMM all-atom additive force field for lipids: validation on six lipid types. *J Phys Chem B* **114**, 7830-7843, doi:10.1021/jp101759q.

19 Klenner, C. & Kuhn, A. 2012. Dynamic disulfide scanning of the membrane-inserting Pf3 coat protein reveals multiple YidC substrate contacts. *J Biol Chem* **287**, 3769-3776, doi:10.1074/jbc.M111.307223.

20 Klenner, C., Yuan, J., Dalbey, R. E. & Kuhn, A. 2008. The Pf3 coat protein contacts TM1 and TM3 of YidC during membrane biogenesis. *FEBS Lett* **582**, 3967-3972, doi:10.1016/j.febslet.2008.10.044.

21 Kohler, R. *et al.* 2009. YidC and Oxa1 form dimeric insertion pores on the translating ribosome. *Mol Cell* **34**, 344-353, doi:10.1016/j.molcel.2009.04.019.

22 Kol, S., Nouwen, N. & Driessen, A. J. 2008. Mechanisms of YidC-mediated insertion and assembly of multimeric membrane protein complexes. *J Biol Chem* **283**, 31269-31273, doi:10.1074/jbc.R800029200.

23 Kumazaki, K. *et al.* 2014. Structural basis of Sec-independent membrane protein insertion by YidC. *Nature*, doi:10.1038/nature13167.

24 Lai, J. S., Cheng, C. W., Lo, A., Sung, T. Y. & Hsu, W. L. 2013. Lipid exposure prediction enhances the inference of rotational angles of transmembrane helices. *BMC Bioinformatics* **14**, 304, doi:10.1186/1471-2105-14-304.

25 Leidig, C. *et al.* 2013. Structural characterization of a eukaryotic chaperone--the ribosome-associated complex. *Nat Struct Mol Biol* **20**, 23-28, doi:10.1038/nsmb.2447.

26 Lotz, M., Haase, W., Kuhlbrandt, W. & Collinson, I. 2008. Projection structure of yidC: a conserved mediator of membrane protein assembly. *J Mol Biol* **375**, 901-907, doi:10.1016/j.jmb.2007.10.089.

27 Marks, D. S. *et al.* 2011. Protein 3D structure computed from evolutionary sequence variation. *PLoS One* **6**, e28766, doi:10.1371/journal.pone.0028766.

28 Miroux, B. & Walker, J. E. 1996. Over-production of proteins in Escherichia coli: mutant hosts that allow synthesis of some membrane proteins and globular proteins at high levels. *J Mol Biol* **260**, 289-298, doi:10.1006/jmbi.1996.0399.

29 Mondal, S., Khelashvili, G., Shi, L. & Weinstein, H. 2013. The cost of living in the membrane: a case study of hydrophobic mismatch for the multi-segment protein LeuT. *Chem Phys Lipids* **169**, 27-38, doi:10.1016/j.chemphyslip.2013.01.006.

30 Norousi, R. *et al.* 2013. Automatic post-picking using MAPPOS improves particle image detection from cryo-EM micrographs. *J Struct Biol* **182**, 59-66, doi:10.1016/j.jsb.2013.02.008.

31 Oliver, D. C. & Paetzel, M. 2008. Crystal structure of the major periplasmic domain of the bacterial membrane protein assembly facilitator YidC. *J Biol Chem* **283**, 5208-5216, doi:10.1074/jbc.M708936200.
32 Park, E. *et al.* 2014. Structure of the SecY channel during initiation of protein translocation. *Nature* **506**, 102-106, doi:10.1038/nature12720.
33 Park, E. & Rapoport, T. A. 2012. Bacterial protein translocation requires only one copy of the SecY complex in vivo. *J Cell Biol* **198**, 881-893, doi:10.1083/jcb.201205140.
34 Punta, M. *et al.* 2012. The Pfam protein families database. *Nucleic Acids Res* **40**, D290-301, doi:10.1093/nar/gkr1065.
35 Rapoport, T. A. 2007. Protein translocation across the eukaryotic endoplasmic reticulum and bacterial plasma membranes. *Nature* **450**, 663-669, doi:10.1038/nature06384.
36 Ravaud, S., Stjepanovic, G., Wild, K. & Sinning, I. 2008. The crystal structure of the periplasmic domain of the Escherichia coli membrane protein insertase YidC contains a substrate binding cleft. *J Biol Chem* **283**, 9350-9358, doi:10.1074/jbc.M710493200.
37 Remmert, M., Biegert, A., Hauser, A. & Soding, J. 2012. HHblits: lightning-fast iterative protein sequence searching by HMM-HMM alignment. *Nat Methods* **9**, 173-175, doi:10.1038/nmeth.1818.
38 Seidelt, B. *et al.* 2009. Structural insight into nascent polypeptide chain-mediated translational stalling. *Science* **326**, 1412-1415, doi:10.1126/science.1177662.
39 Seitl, I., Wickles, S., Beckmann, R., Kuhn, A. & Kiefer, D. 2014. The C-terminal regions of YidC from Rhodopirellula baltica and Oceanicaulis alexandrii bind to ribosomes and partially substitute for SRP receptor function in Escherichia coli. *Mol Microbiol* **91**, 408-421, doi:10.1111/mmi.12465.
40 Shaikh, T. R. *et al.* 2008. SPIDER image processing for single-particle reconstruction of biological macromolecules from electron micrographs. *Nat Protoc* **3**, 1941-1974, doi:10.1038/nprot.2008.156.
41 Sillitoe, I. *et al.* 2013. New functional families (FunFams) in CATH to improve the mapping of conserved functional sites to 3D structures. *Nucleic Acids Res* **41**, D490-498, doi:10.1093/nar/gks1211.
42 Taufik, I., Kedrov, A., Exterkate, M. & Driessen, A. J. 2013. Monitoring the activity of single translocons. *J Mol Biol* **425**, 4145-4153, doi:10.1016/j.jmb.2013.08.012.
43 Van den Berg, B. *et al.* 2004. X-ray structure of a protein-conducting channel. *Nature* **427**, 36-44, doi:10.1038/nature02218.
44 van der Laan, M., Bechtluft, P., Kol, S., Nouwen, N. & Driessen, A. J. 2004. F1F0 ATP synthase subunit c is a substrate of the novel YidC pathway for membrane protein biogenesis. *J Cell Biol* **165**, 213-222, doi:10.1083/jcb.200402100.
45 Wagenknecht, T., Grassucci, R. & Frank, J. 1988. Electron microscopy and computer image averaging of ice-embedded large ribosomal subunits from Escherichia coli. *J Mol Biol* **199**, 137-147.
46 Yu, Z., Koningstein, G., Pop, A. & Luirink, J. 2008. The conserved third transmembrane segment of YidC contacts nascent Escherichia coli inner

membrane proteins. *J Biol Chem* **283**, 34635-34642, doi:10.1074/jbc.M804344200.

FIGURE LEGENDS

Figure 1

Evolutionary covariation based structural model of *E. coli* YidC

a: Membrane topology of YidC, with helix colouring as in all subsequent Figures.

b: Matrix of coupling strengths between pairs of YidC residues based on an alignment of 2366 non-redundant sequences. Helix-helix pairs with posterior probabilities higher than 57% are outlined in boxes; the 50 residue-residue pairs with highest coupling coefficients are indicated with red crosses.

c: Overall arrangement of TM helices viewed from the cytoplasm based on the prediction of helix-helix pairs (black lines) and exposure to lipid (yellow) or protein (green). The first residue of each helix is indicated with an asterisk.

d: Linear representation of YidC with the seven most probable helix-helix pairs indicated by arches, with thicknesses approximating posterior probabilities.

e & f: Side view and cytoplasmic view, respectively, of the *E. coli* YidC model based on covariation analysis, with predicted residue-residue pairs indicated by yellow pseudobonds.

Figure 1-figure supplement 1:

a: Calibration plots for the prediction of helix-helix interactions.

Calibration plots for dataset #2 (left), dataset #3 (middle) and combined datasets #2 and #3 (right). The empirical fraction of true positives is plotted depending on the uncalibrated probability (raw score) obtained from our method. Points correspond to empirical averages over bins of 60 predictions (ordered by increasing uncalibrated probability). Lines correspond to maximum likelihood fits of the calibration plots using a transformed Bernoulli distribution with 4 parameters.

b: Histogram of posterior probabilities for helix-helix interactions.

Distribution of predicted calibrated posterior probabilities for YidC (TM2 – TM6) which contains 7 predicted helices, thus 21 possible helix-helix contacts. The histogram of predicted probabilities shows the specificity of the predictions: there is a large gap between 15% and 55% probability and most possible contacts have probability < 15%.

Figure2:

Covariation-based model *vs* homology model

Comparison of the *E. coli* YidC covariation-based model (a & b) to a homology model of *E. coli* YidC based on the crystal structure of BhYidC2 (3WO6) (c & d). Predicted residue-residue pairs are indicated by yellow pseudobonds. Note that extracellular helix 1 (white) was not present in our multiple sequence alignment and is thus not included in the model.

Figure 2-figure supplement 1:

Local deviations among YidC structures

a: Smoothed Cα distances between the two BhYidC2 crystal forms (3WO6 *vs* 3WO7, red), between our model of *E. coli* YidC and 3WO6 (green) and between our model and 3WO7 (blue). b: Overall root mean square deviations (RMSD) between (the TM helices of) our model of *E. coli* YidC and the two BhYidC2 crystal forms.

Figure 2-figure supplement 2:

Top 50 scoring residue-residue pairs in covariation analysis

Table showing the 50 residue-residue pairs with the highest covariation scores, and the distances between the Cβ atoms in the final model of the 39 pairs that were used as constraints for model building. For comparison, the corresponding distances in 3WO6 are also given. The eleven residue-residue pairs that were excluded for model building are in italics, with the reason for their exclusion indicated on the right.

Video 1:

Conformational states of YidC

Animation showing conformational differences in YidC starting from BhYidC2 crystal form 1 (3WO6), towards crystal form 2 (3WO7) and ending with our covariation based YidC model. Views are from within the membrane (left) and from the cytoplasm (right). Note the movement of the HPD and the closing of the hydrophilic groove between TM3 (orange) and TM5 (green).

Figure 3:

Molecular dynamics simulation of the YidC model

a: Side view (left) and cytoplasmic view (right) of the stable YidC model after a 500 ns MD simulation in a lipid bilayer composed of 3:1 POPE:POPG.

b: Ribbon representation of the stable model according to inter-helix energy (in kcal/mol), blue: -7.5 to -1; white: -1 to -0.002; red: ≥ -0.00.2. Residues that inactivate YidC upon mutagenesis are indicated by spheres.

c: Ribbon representation of the stable model according to flexibility (in $Å^2$), blue: 0.04 to 0.09; white: 0.09-1; red: ≥1.0.

d: *In vivo* complementation assay of YidC mutants T362A (TM2) and Y517A (TM6).

e: Thickness of the cytoplasmic and periplasmic leaflet of the simulated bilayer after 500 ns, highlighting the membrane thinning effect in the vicinity of YidC. The membrane surface is defined by positions of polar head groups in the lipids, and thickness at a given point on the surface is taken to be the shortest distance between the head groups from opposite leaflets. The thickness values are averaged over the MD trajectory and presented as a contour plot on the membrane surface with a color-scale from red, indicating thicker region representing bulk bilayer lipids, to blue showing thinned regions close to YidC suggesting hydrophobic mismatch.

f: Distribution of hydrophobic (red) and hydrophilic residues (blue) in YidC at various heights of the membrane, highlighting the hydrophilic environment in the center of YidC on the cytoplasmic side.

Figure 3-figure supplement 1:

Complementation of MD-based mutants

In vivo complementation assay of YidC mutants identified as structurally important by MD simulations. Positions in YidC that were also identified by covariation analyses are indicated in the right column.

Figure 3-figure supplement 2:

Expression of MD-based mutants

712 Western blot of whole FTL10 cells grown on arabinose or glucose, showing the
713 stable expression of inactive YidC mutants that were identifed by MD
714 simulations.
715
716 Figure 4:
717 **Cryo-EM structure of RNC bound YidC and structural model of the active**
718 **state**
719 **a:** Side view of the ~8 Å resolution cryo-EM based electron density of the
720 RNC:YidC complex, with the small subunit depicted in yellow, the large subunit
721 in grey, P-site tRNA and nascent chain in green, YidC in red and the detergent
722 micelle in blue.
723 **b:** As in **a**, but sliced through the ribosomal exit tunnel.
724 **c:** Validation of the active state model by disulphide crosslinking. RNCs carrying
725 the mutant $F_Oc^{(G23C)}$ were reconstituted with the indicated single cysteine YidC
726 mutants, oxidized, applied to a linear sucrose gradient and harvested from the
727 70S peak before SDS-PAGE and western blotting. Immunodetection was
728 performed with antibodies raised against the HA-tag (located in the nascent
729 chain inside the ribosomal exit tunnel) and anti-YidC antibodies. YidC, nascent
730 chain-tRNA (NC-tRNA) and the expected crosslink product (NC-tRNA x YidC) are
731 indicated.
732 **d-f:** Structural model of YidC during membrane protein insertion, viewed from
733 two sides within the membrane (d & e) and from the cytoplasm (f). The
734 detergent micelle was removed for clarity, the TM helix of F_Oc is depicted in
735 magenta, and the disulphide crosslink between YidC and F_Oc with -SS-.
736
737 Figure 4-figure supplement 1:
738 **Comparison of the active states of YidC and SecY**
739 Left: Molecular model of YidC during co-translational translocation of F_Oc, and
740 the contour of active SecY. Middle: Composite model of active YidC with F_Oc
741 replaced by the hydrophilic part of nascent FtsQ as found in active SecY. Right:
742 Molecular model of SecY during co-translational translocation of FtsQ. For
743 clarity, the N-terminal signal anchor of FtsQ was omitted.
744

Figure 4-figure supplement 2:

Negative control for RNC-YidC crosslinking

Crosslinking was performed with a cysteine-less F_0c RNC as described in the legend to Figure 3c. A poorly reproducible unknown product is indicated with an asterisk.

Figure 5:

Contacts between active YidC and the ribosome

a & b: Close-up views from within the membrane region highlighting the contact between H59 of the ribosome and the 2/3 loop of YidC (a) and ribosomal protein uL23 and the 4/5 loop of YidC (b). Residues that inactivate YidC upon mutagenesis or deletion are indicated by magenta spheres.

c: *In vivo* complementation assay of YidC point mutants D488A, D488K, deletion mutant Δ487-489 and the double mutants Y370A/Y377A and Y370F/Y377F.

d: Periplasmic view of the active ribosome-bound YidC model, with the YidC contour outlined in red. The polypeptide exit tunnel is indicated with an asterisk.

e: Cartoon based comparison of active SecY (left) and active YidC (right) during membrane insertion of FtsQ and F_0c, respectively. The ribosome is depicted in grey, the aqueous channel in SecY as well as the hydrophilic environment within YidC are shaded blue, hydrophobic TM domains of the substrates are depicted magenta, hydrophilic parts in green and the P1 domain by a dashed oval.

Figure 5-figure suppement 1:

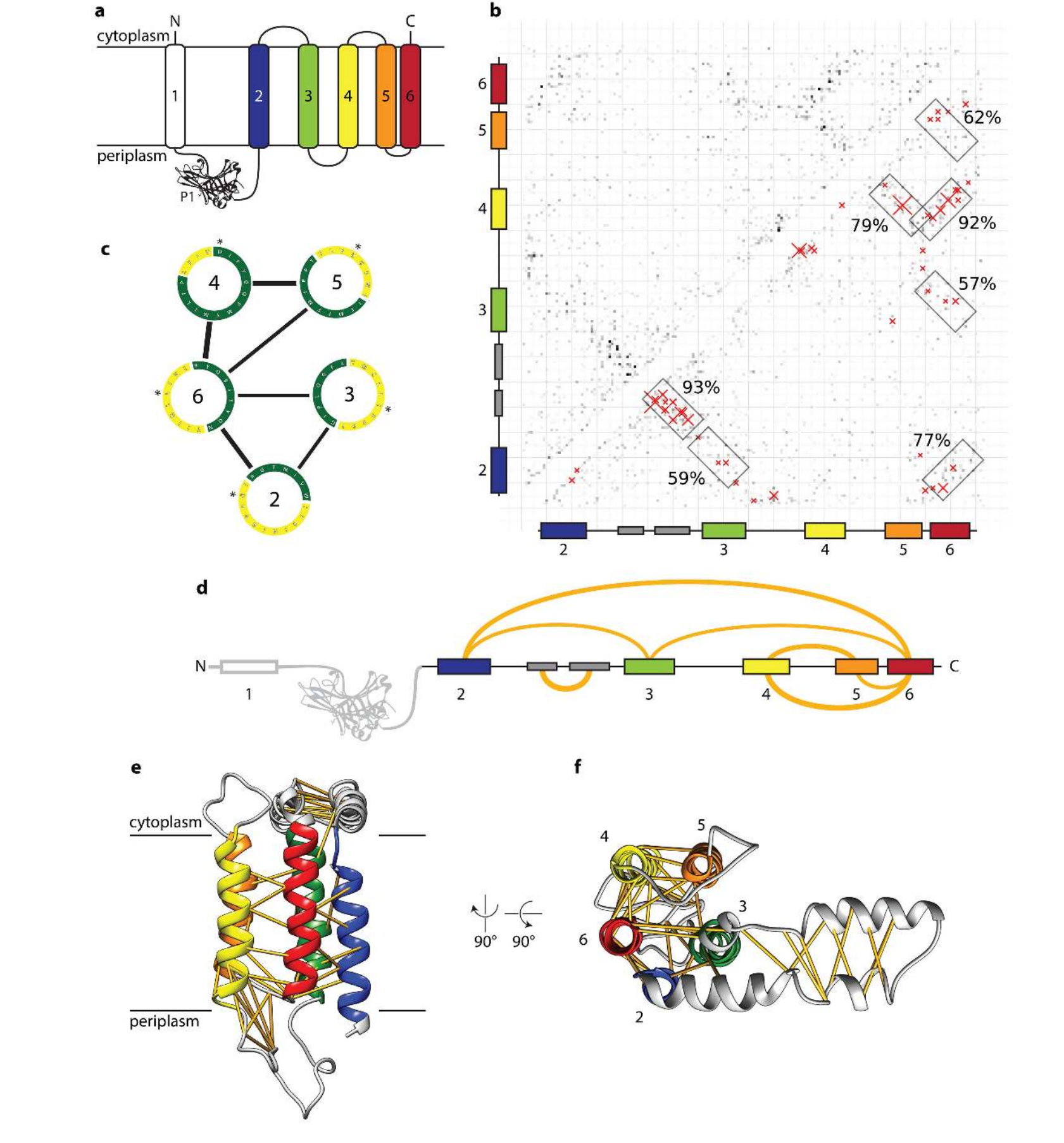
Complementation of ribosome interaction mutants

In vivo complementation assay of YidC mutants involved in ribosome binding.

Figure 5-figure supplement 2:

Expression of ribosome interaction mutants

Western blot of whole FTL10 cells grown on arabinose or glucose, showing the stable expression of inactive YidC mutants that interact with the ribosome.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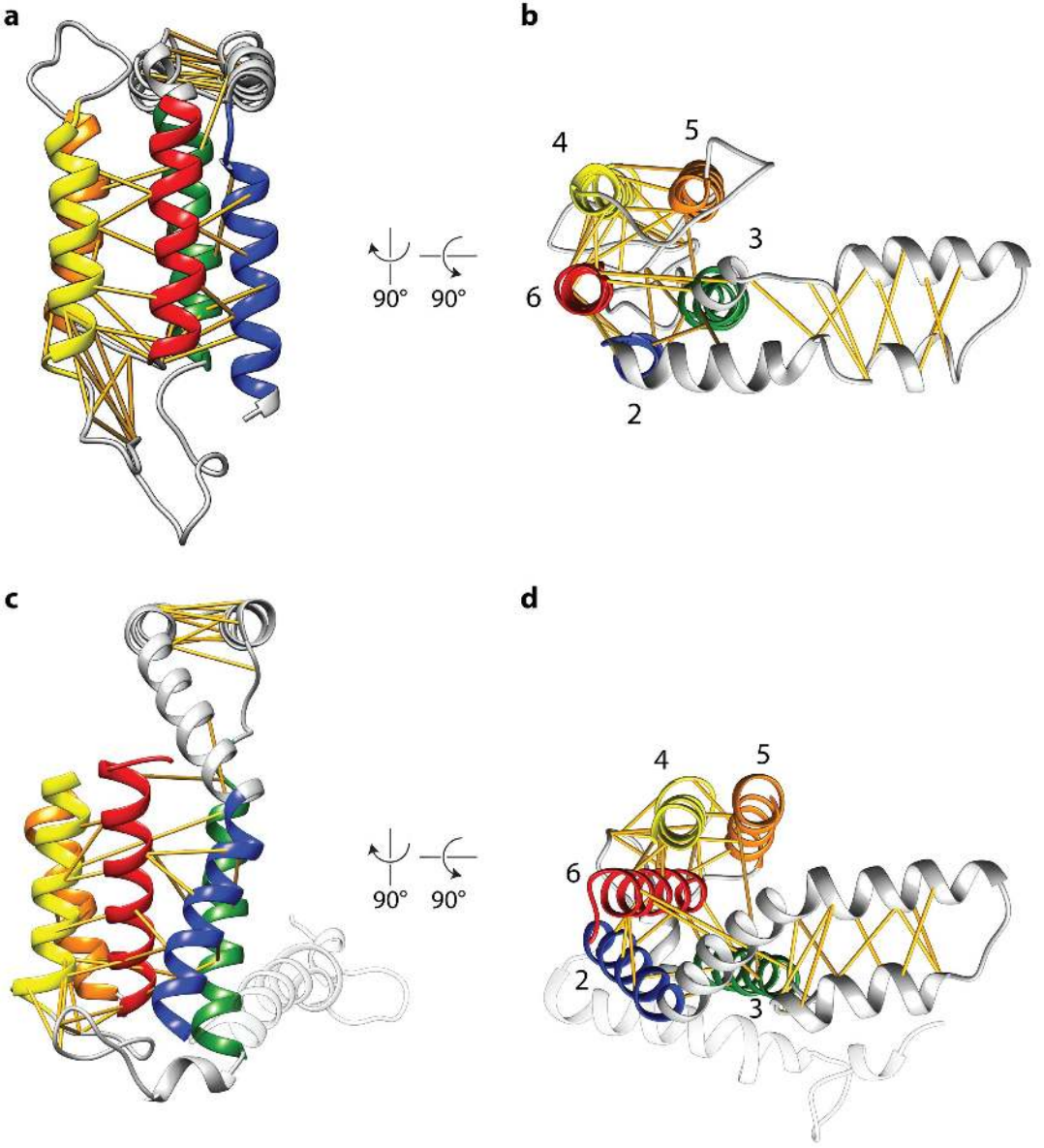
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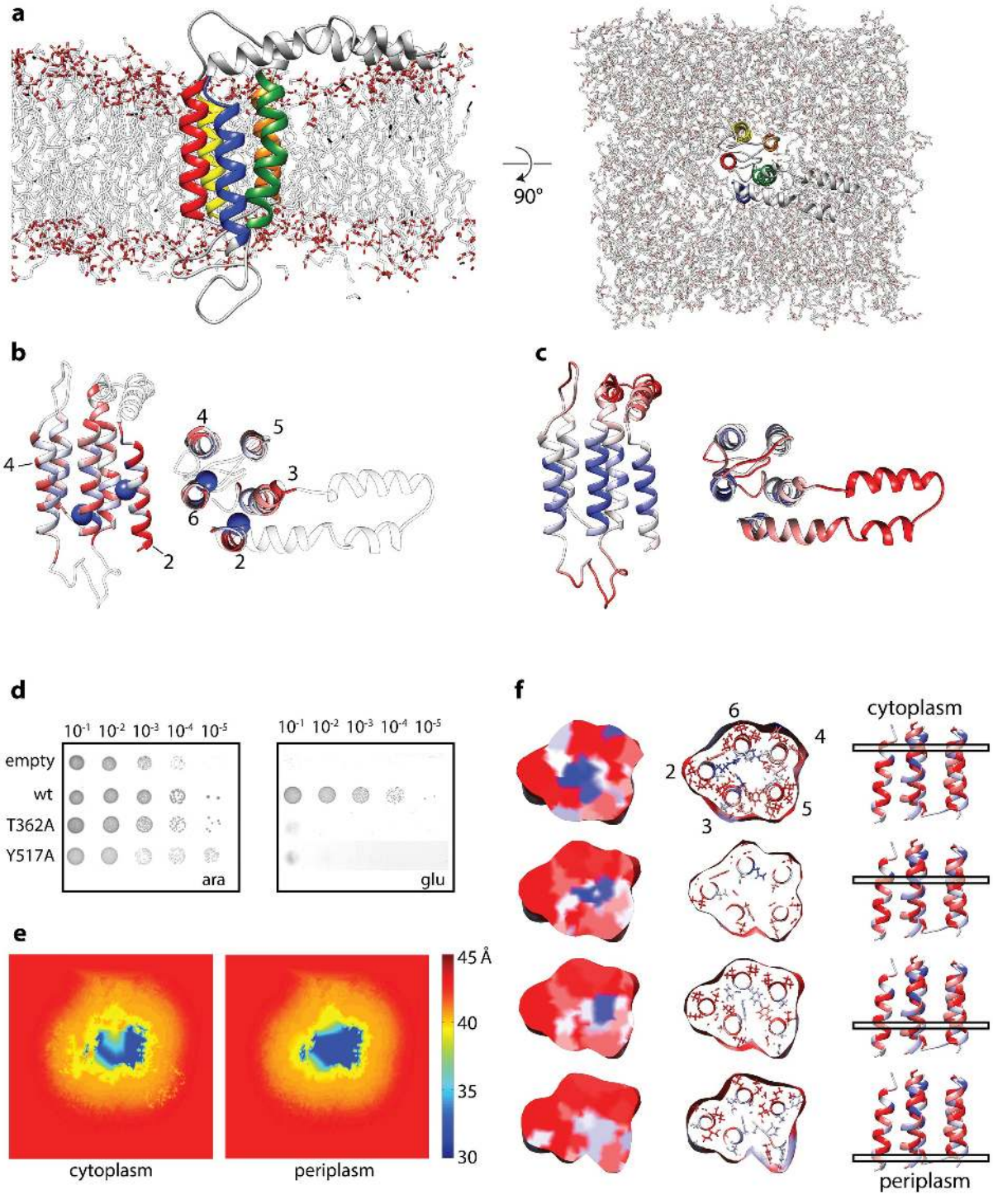
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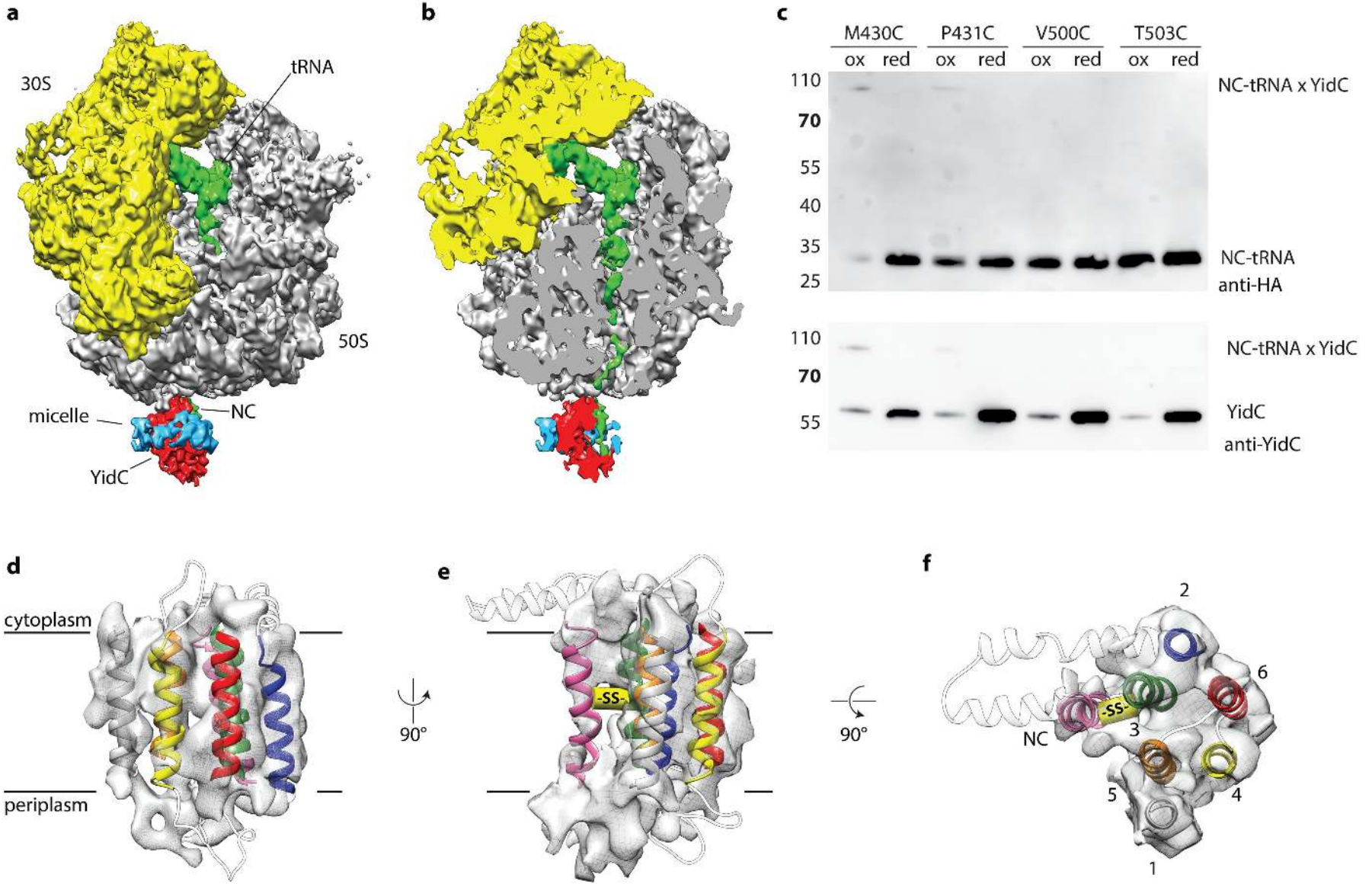
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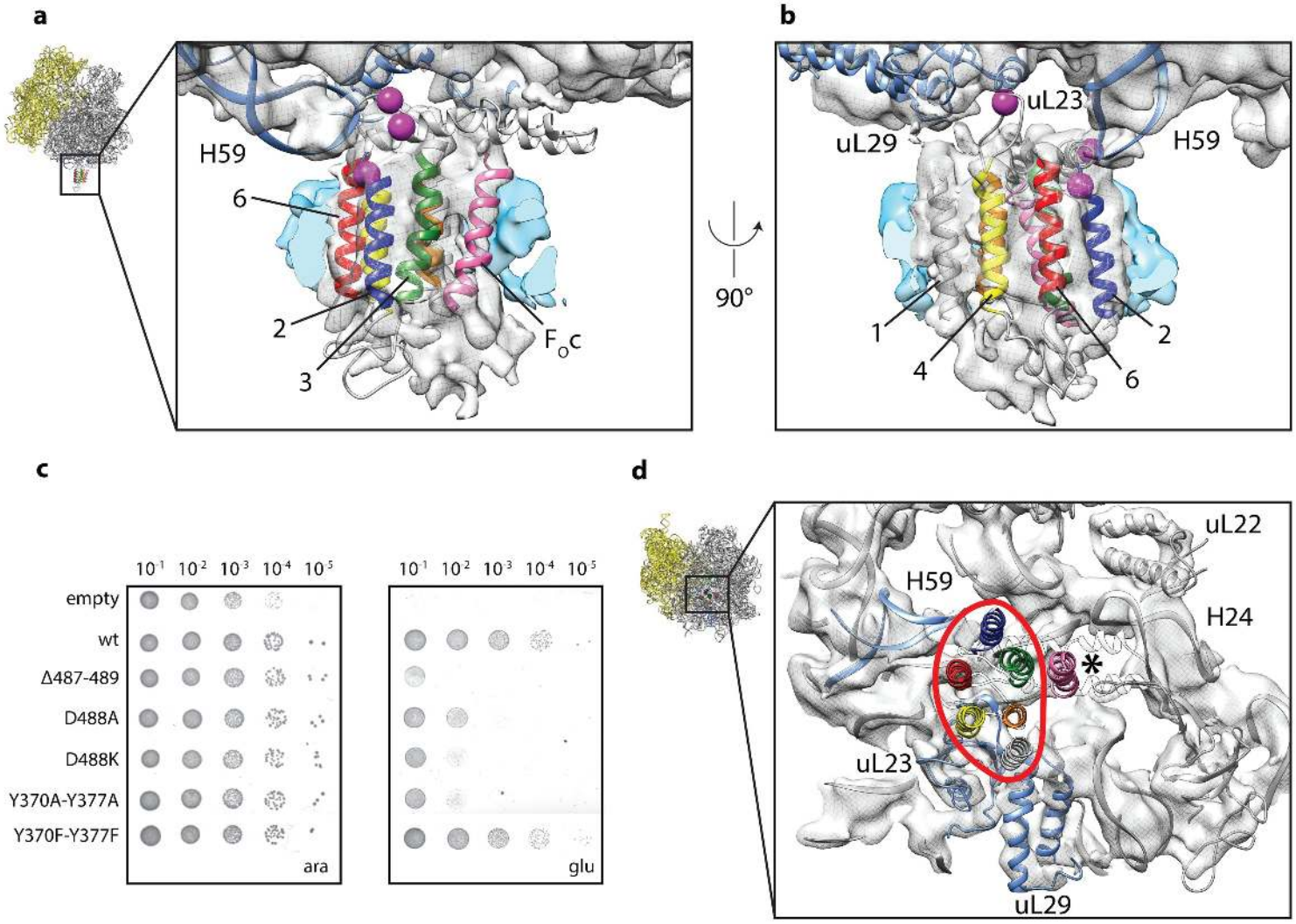
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