

# A STUDY ON CLAVULANIC ACID PRODUCTION BY *Streptomyces clavuligerus* IN BATCH, FED-BATCH AND CONTINUOUS PROCESSES

A. B. Neto<sup>\*</sup>, D. B. Hirata, L. C. M. Cassiano Filho, C. Bellão,  
A. C. Badino Júnior and C. O. Hokka

Federal University of São Carlos, Department of Chemical Engineering, P. O. Box, 676,  
Zip Code 13565-905, Rod Washington Luiz 235, São Carlos - SP, Brazil.  
E-mail: pabn@iris.ufscar.br<sup>\*</sup>,  
E-mail: alvarobn@hotmail.com

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**Abstract** - Clavulanic acid (CA) is a potent inhibitor of  $\beta$ -lactamases, enzymes that are responsible for the hydrolysis of  $\beta$ -lactam antibiotics. It is a secondary metabolite produced by the filamentous aerobic bacterium *Streptomyces clavuligerus* in submerged cultivations. In the present work clavulanic acid production in batch, fed-batch and continuous bioreactors was studied with the objective of increasing productivity. The operating conditions: temperature, aeration and agitation, were the same in all cases, 28° C, 0.5 vvm and 800 rpm, respectively. The CA concentration obtained in the fed-batch culture, 404 mg L<sup>-1</sup>, was ca twice the value obtained in the batch culture, 194 mg L<sup>-1</sup>, while 293 mg L<sup>-1</sup> was obtained in the continuous culture. The highest productivity was obtained in the continuous cultivation, 10.6 mg L<sup>-1</sup> h<sup>-1</sup>, as compared with 8.8 mg L<sup>-1</sup> h<sup>-1</sup> in the fed-batch process and 3.5 mg L<sup>-1</sup> h<sup>-1</sup> in the batch process, suggesting that continuous culture of *Streptomyces clavuligerus* is a promising strategy for clavulanic acid production.

**Keywords:** Clavulanic acid; Bioreactors; Enzymes.

## INTRODUCTION

Many pathogenic bacteria produce  $\beta$ -lactamases, enzymes able to catalyze the hydrolysis of molecules containing a  $\beta$ -lactam ring, such as those of penicillins and cephalosporins, thereby blocking their antibacterial property and giving rise to the problem of resistance to antibiotics in the treatment of infectious diseases. Brown et al. (1976), observed that the cell-free broth of *Streptomyces clavuligerus* NRRL 3585 (ATCC 27064) inhibited the growth of *Klebsiella aerogenes*, a  $\beta$ -lactamase-producing organism, when cultivated in agar containing benzylpenicillin. Although the chemical structure of the active component obtained from the filtered culture was some what similar to that of cephalosporins and the component had a low level of

antibacterial activity, the mode of action was significantly different. This new substance was designated clavulanic acid (CA) and is a potent inhibitor of  $\beta$ -lactamase.

Currently CA is used together with amoxycillin or ticarcillin, which are  $\beta$ -lactam antibiotics with high levels of antibacterial activity (Baggaley et al., 1997). In Brazil medicines, containing amoxycillin and potassium clavulanate are mainly produced by Glaxo-SmithKline Beecham Farmacêutica laboratories under the name of Clavulin ®.

Clavulanic acid can be produced industrially from cultivations of several species of *Streptomyces*, of which *Streptomyces clavuligerus* stands out (Butterworth, 1984). The author observes that there is no information regarding the industrial process, basing the article on patents and results in 1500 L

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<sup>\*</sup>To whom correspondence should be addressed

fermentor at a pilot plant. Several types of bioreactor operation are being applied in the production of secondary metabolite; the fed-batch method is more frequently used due to its higher productivity and easy operation. In the literature several studies report on utilization of fed-batch cultivation to study the fermentation process in antibiotic production. For CA production, the effect of stirring conditions and oxygen transfer (Rosa, 2002) and dissolved oxygen concentration (Yegneswaran et al., 1991) are studied in fed-batch cultivations. Rosa (2002) utilized a medium containing glycerol and malt extract as the source of carbon and Samprosoy 90 NB as the source of nitrogen. Using an air flow rate of 0.5 vvm and 800 rpm the authors, obtained a productivity of 5 mg L<sup>-1</sup> h<sup>-1</sup>, much lower than the value obtained when working with an airflow rate of 0.5 vvm and 1000 rpm (11.5 mg L<sup>-1</sup> h<sup>-1</sup>).

The fed-batch processes may be used to maintain the microorganism in the stationary phase longer as well as to increase the total mass of cells. Mention should be made that the highest rate of production of secondary metabolites is found for the stationary phase. Therefore production is increased, keeping the cultivation in this phase longer.

There are few accounts in the literature of studies on fed-batch processes that address the increase in CA productivity. Mayer and Deckwer (1996) carried out fed-batch experiments for CA production from *Streptomyces clavuligerus* DSM 738. The authors used glycerol as the source of carbon and energy and soy flour or soy flour extract as the main source of nitrogen. In this work, the authors tried to keep the glycerol concentration between 1 and 4 g L<sup>-1</sup> by control of the feed flow of rate additional media as a strategy to obtain higher CA productivity. In that way, the authors obtained CA concentrations of 400 to 500 mg L<sup>-1</sup> in 90 h cultivations.

In a recent patent, Nabais and Fonseca (2000) proposed CA production by *Streptomyces clavuligerus* ATCC 27064 in fed-batch cultivation utilizing a bioreactor with a 8 L capacity and medium containing glycerol or glycerol plus dextrin as the source of carbon and soy flour as the sole source of nitrogen. Different flow rates for addition culture media with different amounts of glycerol or maltose and soy flour were adjusted in order to keep the protein concentration in the broth between 0.4 and 1.5 g L<sup>-1</sup> throughout the cultivation. The maximum CA concentration obtained was around 1.6 g L<sup>-1</sup>; thus higher concentrations could probably be obtained by minimizing the repression effect on the enzyme system related to CA production.

Repression by easily consumed sources of carbon is often described in the literature (Demain, 1971).

Continuous operation is more efficient than batch or fed-batch operation in terms of processing time since dead time, due to charge and discharge operations, bioreactor preparation and sterilization, is avoided. However productivity is often lowered by genetic variations liable to occur when operation continues over a long time. Also, the probability of contamination is much higher than in batch or fed-batch systems.

The only study on continuous cultivation of *Streptomyces clavuligerus* reported in the literature is the work of Kirk et al. (2000). The authors studied the metabolic flux of *Streptomyces clavuligerus* to understand the process of clavulanic acid production by this microorganism. For this purpose the authors conducted continuous cultivations with limited sources of carbon, nitrogen and phosphorus and obtained specific productivities of 0, 0.32 and 3.65 mg of CA per g of cells per hour, respectively.

Mention should be made that there are no accounts in the literature aiming at comparing the effects of bioreactor operating conditions on CA production. Therefore, the objective of the present work was to compare clavulanic acid productivity in batch, fed-batch and continuous fermentors by cultivating *Streptomyces clavuligerus* in a complex medium.

## MATERIALS AND METHODS

### Microorganism

Vegetative cells of *Streptomyces clavuligerus* ATCC 27064, stored in cryotubes (glycerol 10% v/v) at -70°C, were used throughout this work

#### Culture Media

In this work reactivation medium for reactivation of the microorganism from the cryotubes and production medium for the preparation of inoculum and fermentation steps were used.

The reactivation medium was composed of the following (g L<sup>-1</sup>): glycerol, 10; bacto peptone, 10; malt extract, 10; yeast extract, 1.0; K<sub>2</sub>HPO<sub>4</sub>, 2.5; MgSO<sub>4</sub>·7H<sub>2</sub>O, 0.75; MnCl<sub>2</sub>·4H<sub>2</sub>O, 0.001; FeSO<sub>4</sub>·7H<sub>2</sub>O, 0.001 and ZnSO<sub>4</sub>·7H<sub>2</sub>O, 0.001

The production media were composed of the following (g L<sup>-1</sup>): glycerol, 15; Samprosoy 90NB (soybean hydrolyzed protein from Bunge Alimentos S.A., Esteio RS, Brazil), 10; malt extract, 10; yeast extract, 1.0; K<sub>2</sub>HPO<sub>4</sub>, 2.5; MgSO<sub>4</sub>·7H<sub>2</sub>O, 0.75;

$\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$ , 0.001;  $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ , 0.001 and  $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ , 0.001.

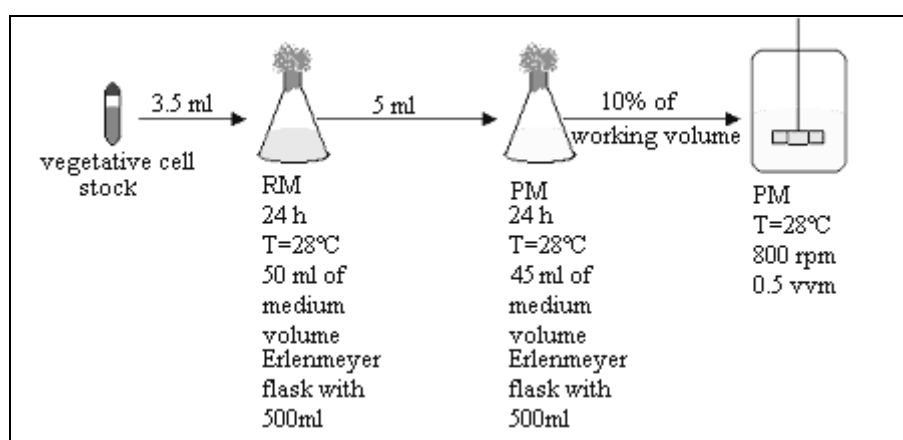
The pH value was maintained at 6.8 in the reactivation and preparation of inoculum with a 100 mM of MOPS buffer (3-[N-Morpholino]-propanesulfonic acid).

In the bioreactor runs,  $0.1 \text{ g L}^{-1}$  of silicone antifoam was added.

### Fermentation Equipment and Cultivation Procedure

Cell suspension from cryotubes (3.5 mL), with a cell concentration of  $\text{ca } 5 \text{ g L}^{-1}$ , were added to 50 mL reactivation medium in a 500 mL Erlenmeyer flask

and incubated in a rotary shaker (New Brunswick Sci.) at  $28^\circ\text{C}$  and 250 rpm for 24 h. Erlenmeyer flasks of 500 mL with 45 mL of production medium were inoculated with 5 mL of the cultivation above and incubated in a rotary shaker at  $28^\circ\text{C}$  and 250 rpm for 24 h. The entire content of the flasks was transferred to the fermentor containing the production medium, making up 10% of the inoculum volume. All cultivations were conducted at  $28^\circ\text{C}$  and the pH was automatically maintained at  $6.8 (\pm 0.1)$  by adding 4 M HCl or 1 M NaOH solution. Stirrer speed was kept constant at 800 rpm and the specific air flow rate, at 0.5 vvm. The procedure described above is illustrated in Figure 1.



**Figure 1:** Schematic flowsheet of the experimental procedure

In batch cultivations, the initial volume was 4 L. Samples of about 20 mL were taken approximately every three hours during the first 24 hours and every four hours afterwards. Some of the samples were used for determination of rheological parameters, i.e., consistency index (K) and behavior index (n) from the power law model. The remainder of the samples were centrifuged at  $3720 \text{ g}$  and  $5^\circ\text{C}$  for 20 min and the supernatants were utilized to determine clavulanic acid and glycerol concentrations.

In the fed-batch cultivation, the initial working volume was 2.5 L. Additional medium was added after 24 hours of cultivation at a constant flow rate ( $0.08 \text{ L h}^{-1}$ ). Samples were taken approximately every three hours during the first 24 hours and every six hours afterwards. The same analyses as those made in the batch cultivations were carried out. The additional medium had the same composition as that of the production medium except for the glycerol concentration used, which was  $10 \text{ g L}^{-1}$ .

The working volume in the continuous cultivation reactor was 3 L. The continuous culture was

operated at six different dilution rates (D) between  $0.036$  and  $0.18 \text{ h}^{-1}$ , corresponding to a range of residence time from 5.5 to 27.8 h.

### Analytical Methods

Clavulanic acid concentration was determined by the HPLC method using a reversed-phase column (C-18  $\mu$ -bondapak, Waters Co.) and prederivatization with imidazole solution, as described by Foulstone and Reading (1982). The eluent was composed of 100 mM  $\text{KH}_2\text{PO}_4$  at pH 3.2 with a buffer-to-methanol ratio of 94:6 and the flow rate was  $2.5 \text{ mL min}^{-1}$ .

Glycerol concentration was determined by HPLC. NaOH (1 mM) solution was used as the mobile phase. The equipment was operated at  $34^\circ\text{C}$  with a flow rate of  $1 \text{ mL min}^{-1}$ . A Shodex 802 column (Lonpak, division of Millipore) was utilized.

The rheological parameters (K and n) were determined using a cylindrical concentric rheometer, model DVIII+ from Brookfield Engineering Inc.

The cell concentrations were determined by a correlation of rheological parameter and cell concentration obtained by assuming a cell-to-substrate yield coefficient. The correlation, eq. 1, was described by Rosa (2002).

$$C_x = 3.76K^{0.333}n^{-0.079} \quad (1)$$

Where  $K$  is the consistency index,  $n$  is the flow behavior index and  $C_x$  is the cell concentration

The values for volumetric productivity of the clavulanic acid for batch and fed-batch cultivations were calculated with eq. 2 below. The values for volumetric productivity of clavulanic acid in continuous cultivation were calculated with eq. 3.

$$P_p = \frac{C_p}{t} \quad (2)$$

$$P_p = C_p D \quad (3)$$

Where  $P_p$  is the volumetric productivity of clavulanic acid ( $\text{mg L}^{-1} \text{h}^{-1}$ ),  $t$  is the cultivation time (h),  $C_p$  is the clavulanic acid concentration ( $\text{mg L}^{-1}$ ) and  $D$  is the dilution rate ( $\text{h}^{-1}$ ).

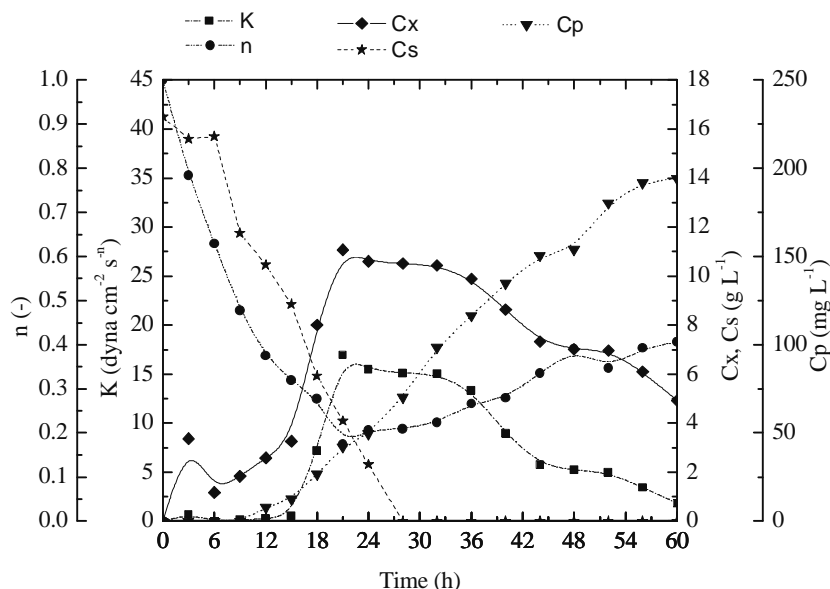
## RESULTS AND DISCUSSION

The time courses of the batch, fed-batch and continuous cultivations are presented in Figures 2, 3 and 4, respectively.

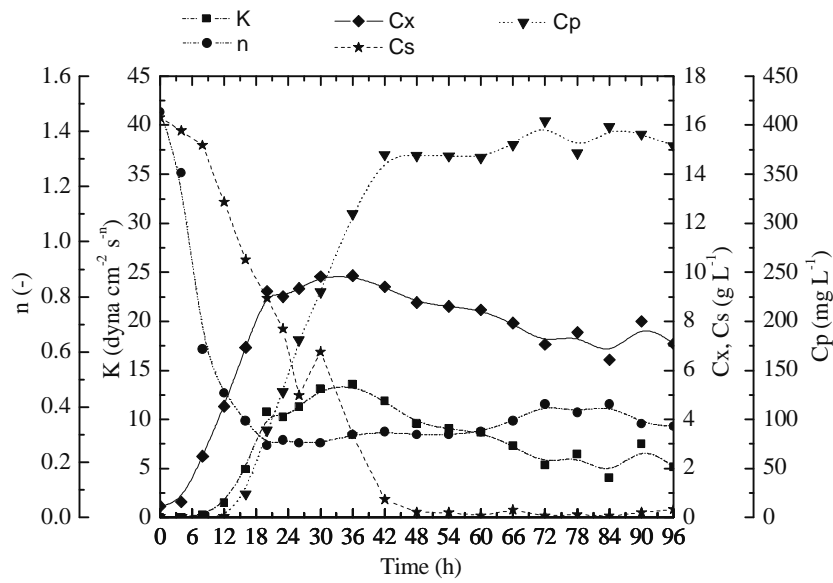
In the batch cultivation production of clavulanic acid started at between 12 and 18 h of cultivation, still during the growth phase; however most of the production occurred later, after the glycerol had been consumed (ca 27 h). The production rate remained almost constant up to 60 h, ca 32 h after glycerol consumption had been completed. Also cell growth occurred in approximately 27 h. This fact indicates that production of clavulanic acid is not associated with microorganism growth, in agreement with the results obtained by Baptista Neto et al. (2000).

The same behavior was observed in the fed-batch cultivation; however the results show that the rate of clavulanic acid production decreased after 42 hours of cultivation. This fact indicates that a product of the metabolism of the microorganism may be acting as a repressor or inhibitor in the metabolic pathway of CA production.

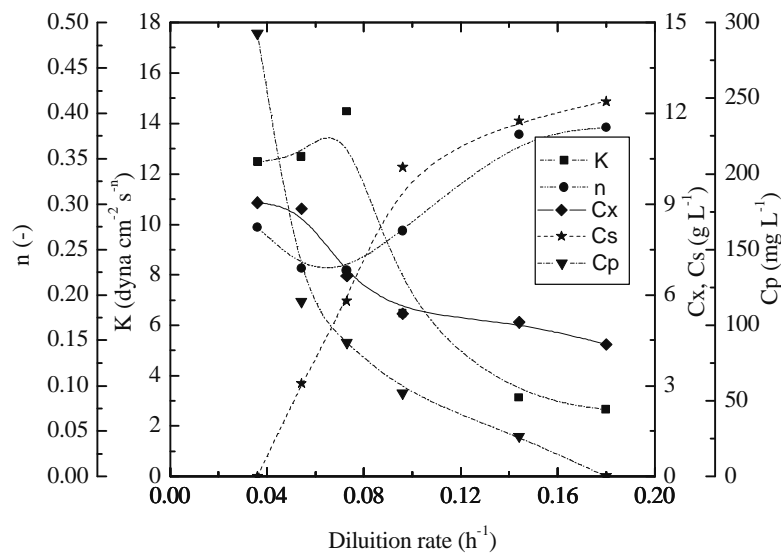
In the three operation modes studied, production started whenever the glycerol concentration reached values below ca  $11.0 \text{ g L}^{-1}$ , in agreement with the results obtained by Baptista Neto et al. (2000).



**Figure 2:** Biomass concentration ( $C_x$ ), glycerol concentration ( $C_s$ ), consistency index ( $K$ ), flow behavior index ( $n$ ) and clavulanic acid concentration ( $C_p$ ) in the batch cultivation overtime



**Figure 3:** Biomass concentration (Cx), glycerol concentration (Cs), consistency index (K), flow behavior index (n) and clavulanic acid concentration (Cp) in the fed-batch cultivation overtime



**Figure 4:** Variation in biomass concentration (Cx), glycerol concentration (Cs), consistency index (K), flow behavior index (n) and clavulanic acid concentration (Cp) with dilution rate in the continuous cultivation

**Table 1: Maximum clavulanic acid concentration ( $C_{p,max}$ ) and maximum volumetric productivity of clavulanic acid ( $P_{p,max}$ ) obtained in the batch, fed-batch and continuous cultivations.**

Experiment	$C_{p,max}$ (g L <sup>-1</sup> )	$P_{p,max}$ (g L <sup>-1</sup> h <sup>-1</sup> )
batch cultivation	194	3.5
fed-batch cultivation	404	8.8
continuous cultivation	293	10.6

A comparison of the maximum value of concentration of clavulanic acid ( $C_{p_{max}}$ ) and the maximum value of volumetric productivity of clavulanic acid ( $P_{p_{max}}$ ) obtained in the three modes of operation is in Table 1.

The results in Table 1 show that the highest maximum concentration of clavulanic acid was obtained in the fed-batch cultivation and the highest volumetric productivity of clavulanic acid was obtained in the continuous cultivation.

The highest maximum concentration of clavulanic acid obtained in the fed-batch cultivation is a typical result; since this mode of operation favors production of secondary metabolites such as CA, which occurs mostly during the stationary phase. The glycerol feed increased the stationary phase, characterized by high cell concentration and an increased and steady rate of secondary metabolite production.

In the fed-batch cultivation, a decrease in the rate of CA production was observed after 42 hours. At this time the cell concentration was still high and the glycerol concentration was lower than the value that causes repression or inhibition of CA production. This indicates that, in this process, the decrease in production rate is associated with the biosynthesis of metabolic byproducts that repress or inhibit the metabolic pathway to clavulanic acid.

This repression or inhibition was not observed in the continuous process. In this process the continuous feed of fresh medium and the continuous removal of spent medium allowed for a lower concentration of the CA pathway repressor or inhibitor.

Based on the results obtained above, studies on the process of CA production in a system with cell recycle equipment allowing for a high cell concentration, are underway in our laboratories.

## CONCLUSIONS

The results obtained in this work showed that clavulanic acid is partially associated with growth; however most of the production phase occurs with no association with the growth phase. The highest CA productivity was obtained in the continuous cultivation, while the fed-batch culture attained the highest CA concentration. Batch cultivation was shown to be the least efficient mode of operation since both CA productivity and maximum CA concentration were the lowest in this mode of operation. In conclusion, it can be said that the continuous culture of *Streptomyces*

*clavuligerus* is a very promising strategy for production of clavulanic acid.

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## NOMENCLATURE

$C_p$	clavulanic acid concentration	( $\text{mg L}^{-1}$ )
$C_{p_{max}}$	maximum value of clavulanic acid concentration	( $\text{mg L}^{-1}$ )
$C_s$	glycerol concentration	( $\text{g L}^{-1}$ )
$C_x$	cell concentration	( $\text{g L}^{-1}$ )
$D$	dilution rate	( $\text{h}^{-1}$ )
$K$	consistency index	( $\text{dyna cm}^{-2} \text{s}^{-n}$ )
$n$	behavior index	
$P_p$	volumetric productivity of clavulanic acid	( $\text{mg L}^{-1} \text{h}^{-1}$ )
$P_{p_{max}}$	maximum volumetric productivity of clavulanic acid	( $\text{mg L}^{-1} \text{h}^{-1}$ )
$t$	cultivation time	(h)

## REFERENCES

- Baggaley, K.H., Brown, A.G. and Schofield, C.J., Chemistry and Biosynthesis of Clavulanic Acid and Another Clavans. Natural Product Report, vol. 57, pp. 309-333, 1997.
- Baptista-Neto, A., Gouveia, E.R., Badino-Jr, A.C. and Hokka, C.O., Phenomenological Model of the Clavulanic Acid Production Process Utilizing *Streptomyces clavuligerus*, Brazilian Journal of Chemical Engineering, vol. 17, n. 04-07, pp.809-818, 2000.
- Battaglia-Hirata, D., Produção de Ácido Clavulânico por *Streptomyces clavuligerus* e Extração em Sistema de Duas Fases Aquosas, São Carlos-SP, University Federal of São Carlos, (Master's thesis), 2003.
- Brown, A.G., Butterworth, D., Cole, M., Hanscomb, G., Hood, J.D., Reading, C. and Roinson, G.N., Naturally Occurring  $\beta$ -lactamase Inhibitors with Antibacterial Activity. The Journal of Antibiotics, vol.29, n. 6, pp. 668-669,1976.

- Butterworth, D., Clavulanic acid: Properties Biosynthesis, and Fermentation. In: Vandamme, E.J. Biotechnology of Industrial Antibiotics, New York, Marcel Dekker, vol.22, pp.225-235, 1984.
- Demain, A.L., Overproduction of Microbial Metabolites and Enzymes due to Alteration of Regulation, Advances in Biotechnological Engineering, vol. 1, 113 p., (1971)
- Foulstone, M. and Reading, C., Assay of Amoxillin and Clavulanic Acid, the Components of Augmentin, in Biological Fluids With HPLC, Ant. Ag. Chem., vol.22, pp.753-762, 1982.
- Kirk S, Avignone-Rossa C.A. and Bushell M.E., Growth Limiting Substrate Affects Antibiotic Production and Associated Metabolic Fluxes in *Streptomyces clavuligerus*, Biotechnology Letters; vol. 22; n. 22, pp. 1803-1809; 2000.
- Mayer, A.F. and Deckwer, W.-D. Simultaneous Production and Decomposition of Clavulanic Acid During *Streptomyces clavuligerus* Cultivations. Appl. Microbiol. Biotechnol., vol.45, pp. 41-46, 1996.
- Nabais, A.M.A. and Fonseca, M.M.R. Improved Process for Clavulanic Acid Production, Patent Number WO0005397, 2000.
- Rosa, J.C., Influência das Condições de Transferência de O<sub>2</sub> na Produção de Ácido Clavulânico por *Streptomyces clavuligerus*, São Carlos-SP, University Federal of São Carlos, (Master's thesis), 2002.
- Yegneswaran, P.K., Gray, M.R. and Thompson, B.G., Experimental Simulation of Dissolved Oxygen Fluctuations in Large Fermentors: Effect on *Streptomyces clavuligerus*, Biotech. Bio., vol.38, pp. 1203-1209, 1991.