

A tumour suppressor network relying on the polyamine–hypusine axis

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Tumour suppressor genes encode a broad class of molecules whose mutational attenuation contributes to malignant progression. In the canonical situation, the tumour suppressor is completely inactivated through a two-hit process involving a point mutation in one allele and chromosomal deletion of the other¹. Here, to identify tumour suppressor genes in lymphoma, we screen a short hairpin RNA library targeting genes deleted in human lymphomas. We functionally identify those genes whose suppression promotes tumorigenesis in a mouse lymphoma model. Of the nine tumour suppressors we identified, eight correspond to genes occurring in three physically linked ‘clusters’, suggesting that the common occurrence of large chromosomal deletions in human tumours reflects selective pressure to attenuate multiple genes. Among the new tumour suppressors are adenosylmethionine decarboxylase 1 (*AMD1*) and eukaryotic translation initiation factor 5A (*eIF5A*), two genes associated with hypusine, a unique amino acid produced as a product of polyamine metabolism through a highly conserved pathway². Through a secondary screen surveying the impact of all polyamine enzymes on tumorigenesis, we establish the polyamine–hypusine axis as a new tumour suppressor network regulating apoptosis. Unexpectedly, heterozygous deletions encompassing *AMD1* and *eIF5A* often occur together in human lymphomas and co-suppression of both genes promotes lymphomagenesis in mice. Thus, some tumour suppressor functions can be disabled through a two-step process targeting different genes acting in the same pathway.

Tumour suppressors may be disabled by point mutations, deletions and promoter methylation. Because mutations in one allele are often followed by deletion of the other¹, somatic deletions in human cancers often pinpoint tumour suppressor genes that function as ‘drivers’ of tumour evolution. However, such deletions often encompass other genes, termed ‘passengers’, whose disruption may have no functional consequence³. They also may include haploinsufficient tumour suppressors, whose partial loss of function contributes to cancer development in the absence of mutations targeting the second wild-type allele⁴.

B-cell non-Hodgkin’s lymphoma is a heterogeneous group of malignancies that frequently acquire large chromosomal deletions whose biological impact is poorly understood⁵. We previously showed that suppression of gene function using RNA interference can mimic tumour suppressor gene inactivation, and we have developed strategies to screen for driver tumour suppressors using mouse models as a biological filter^{6,7}. To do this for human B-cell non-Hodgkin’s lymphoma, we focused on genes present in deletions identified in several studies^{8–12} (Fig. 1a and Supplementary Tables 1 and 2). Although not comprehensive, this analysis identified 11 recurrent deletions (encompassing 3–103 genes) targeting a total of 323 recurrently deleted genes (Supplementary Table 3).

We next designed a short hairpin RNA (shRNA) library targeting the mouse orthologues of deleted genes using on-chip oligonucleotide synthesis and a bar-coding strategy such that shRNAs could be amplified from the mixtures in pools of a discrete size. The pools were individually screened for their ability to increase the lymphomagenic potential of pre-malignant haematopoietic stem and progenitor cells (HSPCs) from E μ -myc transgenic mice (Supplementary Fig. 1a), which express the *Myc* oncogene in the B-cell compartment and, although not producing a precise pathological match to human lymphoma, have been widely used to identify and characterize important human cancer genes¹³.

Using a modified syngeneic transplant model described previously¹⁴, we identified conditions such that a green fluorescent protein (GFP)-coupled p53-targeting shRNA efficiently accelerated lymphomagenesis when diluted at 1:200 with a neutral shRNA (Supplementary Fig. 1b). Although typically only about 20% of the HSPCs were transduced (as assessed by GFP fluorescence), the resulting lymphomas were GFP positive. By contrast, a control GFP-tagged shRNA targeting a neutral gene (luciferase) did not accelerate lymphomagenesis and most lymphomas that eventually arose were GFP negative (Supplementary Fig. 1c–e). Hence, GFP fluorescence provides confirmation that an shRNA confers a selective advantage during lymphomagenesis and provides a fluorescent marker to track and isolate lymphoma cells.

To determine whether lymphoma deletions are enriched for tumour suppressors specific to lymphoma, we tested in parallel equivalent-sized shRNA pools targeting genes deleted in breast cancer, acute myeloid leukaemia and hepatocellular carcinoma that show only modest overlap with lymphoma (Supplementary Fig. 2). Only 20% (3/15) of the control pools accelerated lymphomagenesis whereas more than half (8/14) of the lymphoma pools scored (Fig. 1b, c). Furthermore, the lymphoma pools scored more frequently than the breast cancer, acute myeloid leukaemia and hepatocellular carcinoma sets (Fig. 1d; $P = 0.0018$), implying that our approach enriched for shRNAs targeting tumour suppressors and that, despite some limitations, the E μ -myc model can accurately filter genetic interactions relevant to the human disease.

Sequencing of PCR-amplified shRNAs present in accelerated GFP-positive lymphomas was used to identify candidate ‘drivers’ of disease. Candidates were prioritized using an enrichment score based on (1) the number of shRNAs targeting the gene retrieved from the lymphomas, (2) the number of lymphomas carrying shRNAs targeting the gene and (3) the enrichment of each shRNA relative to its representation in the pool (Supplementary Table 4). Nine new tumour suppressors were subsequently validated (Supplementary Table 5), all of which (by their inclusion in the screen) were embedded within recurrent somatic deletions in human lymphoma (Supplementary Fig. 4) and for several of which there is additional evidence of mutation (*AMD1*, *SMPD2* and

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CSMD1) or promoter methylation (*KCTD11*) in other cancers. In all cases, several shRNAs per gene were re-assayed individually and knockdown was confirmed by immunoblotting or quantitative PCR with reverse transcription (Supplementary Figs 3 and 5).

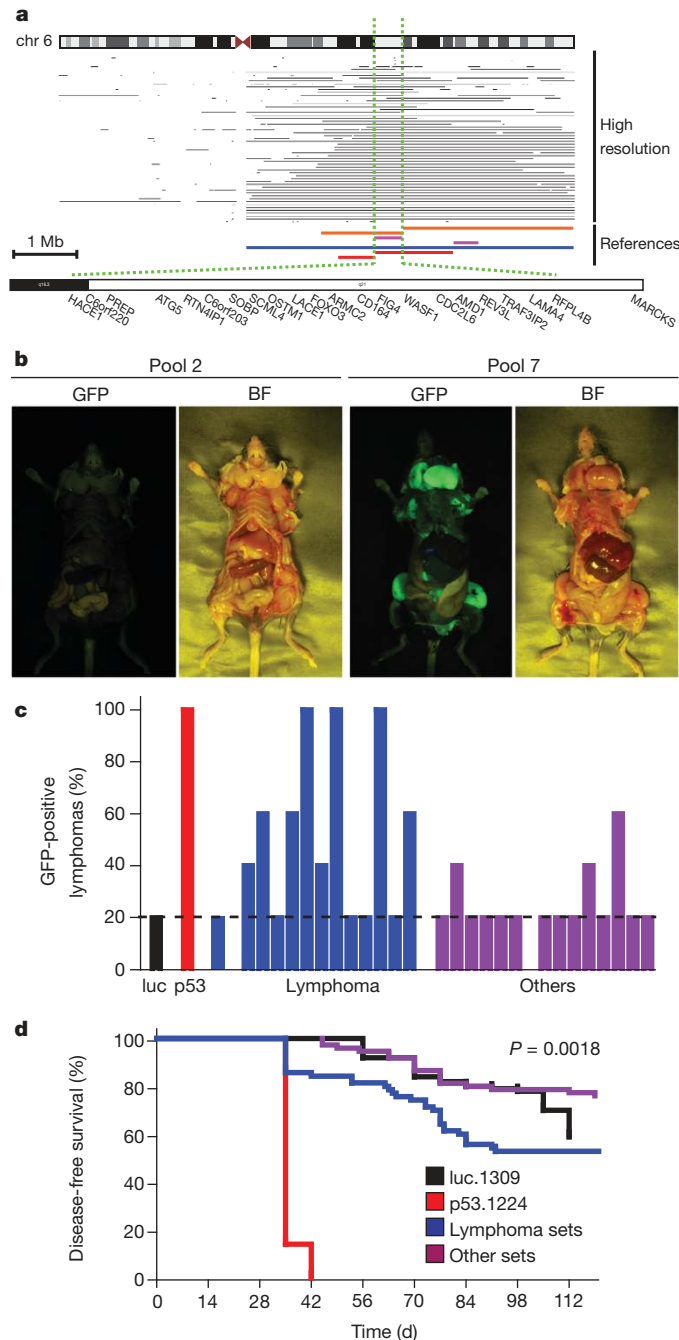


Figure 1 | An *in vivo* shRNA screen for tumour suppressors in lymphoma.

a, Screening interval for the 6q21 deletion. Top, high-resolution data from samples from patients. Bottom, 6q21 deletions as reported in the following references: orange⁸; violet⁹; blue¹⁰; red¹¹. The dotted green lines delimit the target interval. Representative genes are shown. **b**, Brightfield (BF) and GFP whole-body imaging of mice from lymphoma pools 2 and 7. **c**, GFP-positive lymphomas observed for mice transplanted with shRNA sets targeting genes deleted in lymphoma (blue) or in other cancers (violet). Luciferase (luc, black) and p53 (red) were used as negative and positive controls. The dotted line represents the threshold for sequencing lymphoma. Each bar represents a pool. **d**, Survival curves for mice transplanted with neutral control (luc.1309, black, $n = 100$), positive control (p53.1224, $n = 100$), lymphoma sets (blue, $n = 70$) and other sets (violet, $n = 75$).

Two pairs of tumour suppressors were connected by predicted functional relationships. *MED4* (13q14) and *Cyclin C* (6q16) are both components of the Mediator, a multi-unit complex that has been associated with transcription regulation¹⁵. Also, the candidate gene *AMD1* (6q21) controls the biochemical pathway leading to a unique post-translational modification of a second candidate, *eIF5A* (17p) (Fig. 2a–d and Supplementary Fig. 5a, b). Owing to physical linkage, both the human *AMD1* and *eIF5A* genes are frequently co-deleted with other genes scoring in our assay; indeed, *eIF5A* is adjacent to *TP53* on chromosome 17 (Supplementary Fig. 6). Additionally, as is seen in lymphoma patients with *TP53* mutations (ref. 16), underexpression of *eIF5A* protein was associated with a significantly reduced event-free survival ($P < 0.01$; Supplementary Fig. 7). Nonetheless, lymphomas triggered by suppression of *Amd1* or *Eif5a* arise independently of p53 loss, because shRNAs targeting each gene alone were sufficient to trigger lymphomas with aggressive features (Fig. 2e, f) and those lymphomas expressing *Eif5a* shRNAs retained intact p53 function as

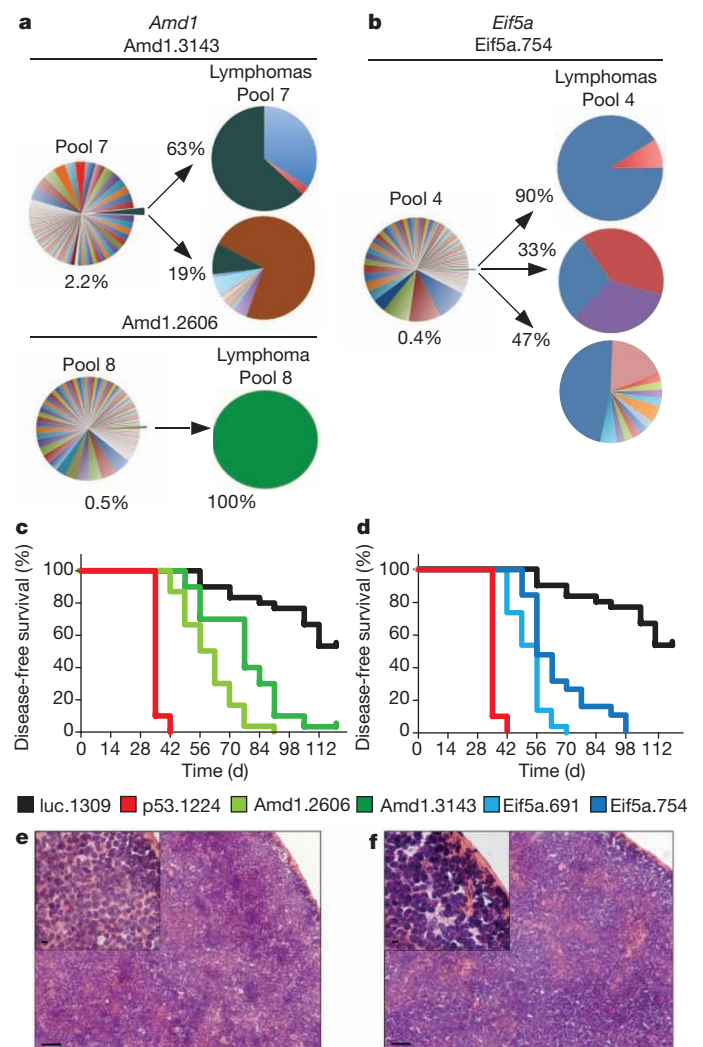


Figure 2 | Validation of *eIF5A* and *Amd1* as tumour suppressors in lymphoma.

The distribution of shRNAs targeting *Amd1* (**a**) or *Eif5a* (**b**) in lymphomas is compared with the corresponding pools. Percentages indicate the frequency of the shRNA in the pool and in the lymphomas. Survival curves for mice reconstituted with two shRNAs targeting *Amd1* (**c**, $n = 30$ for each shRNA, $P < 0.001$ for both shRNAs) or *Eif5a* (**d**, $n = 30$ each shRNA, $P < 0.001$ for both shRNAs). Controls are p53.1224 (red, $n = 30$, $P < 0.001$) and luc.1309 (black, $n = 30$). Haematoxylin and eosin sections of spleens from mice transplanted with shRNA targeting *Amd1* (**e**) or *Eif5a* (**f**). Disruption of tissue architecture is visible both in the spleen and lymph nodes. Scale bars, 50 μm ; insets, 5 μm .

assessed by several functional and molecular criteria (Supplementary Figs 8 and 9). These data support an emerging view that large chromosomal deletions can target several tumour suppressors whose attenuation contributes independently to tumorigenesis¹⁷.

AMD1 encodes an enzyme controlling a rate-limiting step for the synthesis of spermidine, which is incorporated into eIF5A in the form of the non-standard amino acid hypusine¹⁸. Interestingly, eIF5A represents one of the two known hypusine-containing proteins (the other is eIF5A2, a family member not expressed in B cells¹⁹). The fact that both *Amd1* and *eIF5A* act as tumour suppressors suggests that the level of hypusinated eIF5A can be decreased by independent mechanisms, acting either on the total level of eIF5A or on the enzymes controlling hypusine synthesis. The latter mechanism was unexpected because high levels of polyamines are often linked to cancer, and as ornithine decarboxylase (ODC1), which regulates another key step in spermidine synthesis (Fig. 3a), is required for Myc-induced lymphoma development²⁰.

For clarification, we performed a secondary screen by designing shRNA pools (three or four shRNAs per gene) targeting each enzyme in the polyamine pathway and testing each for their tumour-promoting activity *in vivo* (Fig. 3b). The results of these experiments were decisive: although shRNAs targeting peripheral activities showed no effect, those targeting three of the four enzymes responsible for hypusine synthesis showed potent tumour-promoting effects. Hence, knockdown of *Srm* (spermidine synthase) and *Dhps* (deoxyhypusine synthase), which are both essential for the hypusine modification of eIF5A, promoted lymphomagenesis *in vivo* (Fig. 3b and Supplementary Fig. 10).

As predicted, lymphomas harbouring each of the tumour-promoting shRNA pools showed a reduction in eIF5A hypusination relative to controls as assessed by two-dimensional polyacrylamide gel electrophoresis (PAGE) followed by eIF5A immunoblotting (Fig. 3c, d and Supplementary Fig. 11). Furthermore, reintroduction of an shRNA-resistant wild-type *Eif5a* cDNA inhibited the outgrowth of lymphoma cells expressing *Eif5a* shRNAs, and a mutant (*Eif5a*^{K50A}) that could not be hypusinated²¹ had no effect (Fig. 3e and Supplementary Fig. 12). Finally, suppression of *Eif5a* attenuates spermidine-induced cell death in lymphomas driven by *Amd1* knockdown while not altering sensitivity to the cytotoxic drug adriamycin (Supplementary Fig. 13). Therefore, although *Amd1* and *Srm* may have additional cancer-relevant targets, our collective genetic and biochemical data provide strong evidence that the polyamine–hypusine axis is a potent tumour suppressor network in lymphomas.

To gain insights into the mechanism, we analysed pre-malignant B cells expressing each set of shRNAs. Although we observed no obvious effect on BrdU incorporation, S phase content and PCNA expression (data not shown), we noticed that suppression of *Amd1*, *Srm*, *Dhps* and *Eif5a* led to the reduction of apoptosis, as assessed by annexin V staining and by the absence of substantial caspase-3 cleavage (Fig. 3f and Supplementary Fig. 14). Conversely, enforced expression of a wild-type (but not mutant) eIF5A into a human line with 17p deletions also triggered apoptosis, as has been described in other contexts²² (Supplementary Fig. 15).

We next determined whether the expression of apoptotic regulators known to modulate Myc-induced lymphomagenesis (for example Bax, Bim, Puma, Bcl-2, Bcl-xL and Mcl-1) were altered in pre-malignant B cells expressing shRNAs targeting the hypusination biosynthetic pathway (Fig. 3g). Bax was the only protein consistently underexpressed (Fig. 3g) and was found reduced through a non-biased isobaric tag for relative and absolute quantitation (iTRAQ)-based proteomic analysis of pre-malignant B cells transduced with shRNAs targeting *Eif5a* and *Amd1* (Supplementary Fig. 16). Gene ontology analysis of the 431 differentially expressed proteins identified ‘apoptosis’ as the most significantly enriched functional category ($P = 1.9 \times 10^{-4}$; Supplementary Table 8). Precisely how Bax is controlled by *Eif5a* hypusination remains to be determined; nonetheless, Bax is a known eIF5A target²²

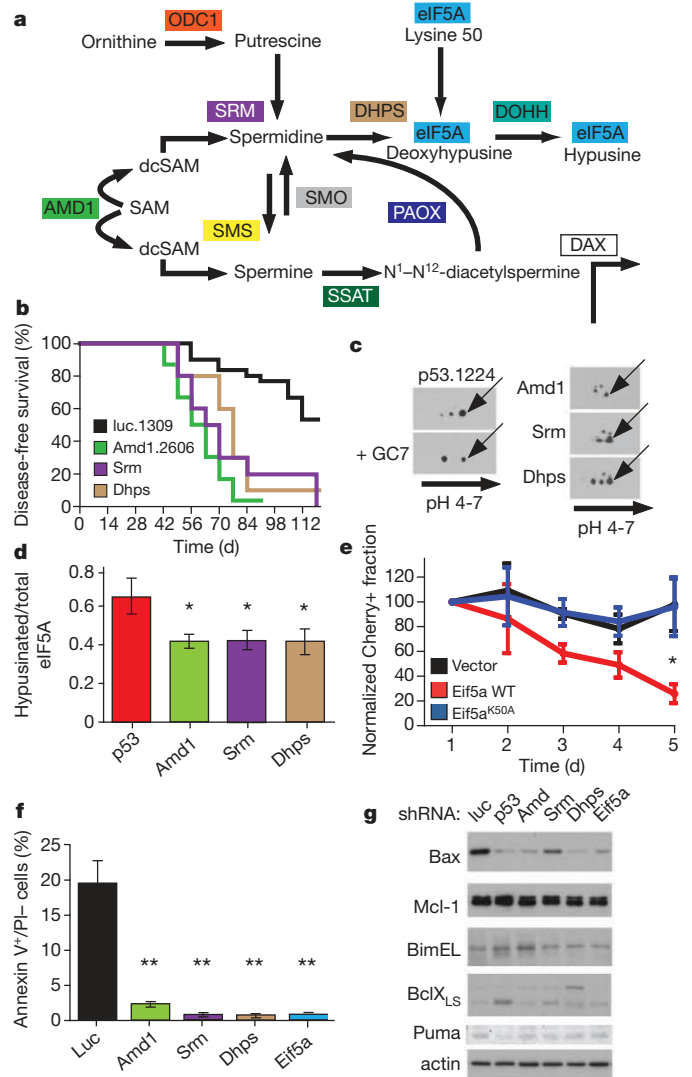


Figure 3 | Biosynthesis of hypusine as a tumour suppressor pathway.

a, Schematic of the polyamine–hypusine pathway. Enzymes and compounds are indicated as follows: ODC1, ornithine decarboxylase; SRM, spermidine synthase; SMS, spermine synthase; DHPS, deoxyhypusine synthase; DOHH, deoxyhypusine hydrolase; SMO(X), spermine oxidase; PAOX, polyamine oxidase; SSAT, spermidine–spermine acetyltransferase; DAX, diamine transporter. SAM, S-adenosylmethionine; dc-SAM, decarboxylated S-adenosylmethionine. The SSAT–PAOX axis can also convert spermidine to putrescine (not shown). **b**, Survival curves for mice ($n = 10$ per pool) transduced with the following shRNAs or shRNA pools: *Srm* (violet, $P < 0.001$), *Dhps* (blue, $P < 0.001$), *Amd1* (*Amd1.2606*, green, $P < 0.001$) and control (*luc.1309*, black). **c**, Two-dimensional PAGE followed by eIF5A immunoblotting of lymphomas driven by the indicated shRNAs. The p53.1224 lymphomas were treated with $10 \mu\text{M}$ N¹-guanyl-1,7-diaminoheptane (GC-7) in short-term culture conditions. Arrows indicate the hypusinated form. **d**, Quantification of hypusinated/total eIF5A ratio for the indicated shRNAs ($n = 3$ per group, $*P < 0.05$). Error bars, s.d. **e**, Lymphomas generated by transduction of *Eif5a* shRNAs were retrovirally transduced with control vector (black), a complementary DNA encoding wild-type *Eif5a* (red) or a mutant complementary DNA that cannot be hypusinated because of the substitution of the modified lysine (K50A, blue). Cherry percentages were monitored for 5 days and normalized to the Cherry fraction at day 1 (100). Error bars, s.d. ($n = 4$ for each time point, $*P < 0.05$). **f**, Annexin V⁺/PI⁻ fractions of GFP-positive B cells 3 weeks after adoptive transplant of Eµ-myc HSPCs transduced with the indicated shRNAs or shRNA pools ($n = 3$ per each shRNA; $**P < 0.01$). Error bars, s.d. **g**, Western blot analysis for expression of the indicated proteins in Eµ-myc pre-malignant B cells transduced with the indicated shRNAs and sorted 21 days after transplant. Cells from three mice were pooled for each shRNA.

and is an established suppressor of lymphomagenesis in the E μ -myc model²³.

Next, we returned to our analysis of human lymphoma to search for somatic mutations or evidence of an epistatic relationship between *eIF5A* and *AMD1*. Sequencing efforts so far have only identified two non-synonymous mutations in *AMD1* (ref. 24) and none in *eIF5A*, raising the possibility that these tumour suppressors are haploinsufficient. Accordingly, we never observed complete suppression of either *eIF5A* or *Amd1* in lymphomas driven by their corresponding shRNAs (Supplementary Fig. 5a, b), and primary B cells with near-complete suppression had poor viability (Supplementary Fig. 17). Surprisingly, analysis of publicly available comparative genomic hybridization data of diffuse large B-cell lymphomas²⁵ showed a significant co-association between the deletion of *AMD1* (6q21) and *DHPS* (19p13) (Supplementary Fig. 18) and *AMD1* and *eIF5A* (17p13) (Fig. 4a;

$P < 0.0001$), with the 17p and 6q co-deleted samples enriched in the ABC subtype. Although other genes in the deleted regions probably also contribute to tumour phenotypes, the co-deletion of two genes in the same pathway was unexpected.

Because *AMD1* and *eIF5A* are each essential genes^{26,27}, we reasoned that attenuation of two haploinsufficient tumour suppressors might selectively target the anti-tumour activities of polyamine biogenesis while leaving essential functions intact. As an initial test, we co-transduced HSPCs with shRNAs targeting *Eif5a* and *Amd1*, tagged with either GFP or Cherry fluorescent markers, which allowed us to track cells transduced with one or both shRNAs (Fig. 4b). Strikingly, the co-knockdown of *Eif5a* and *Amd1* accelerated lymphomagenesis over both individual shRNAs ($P < 0.0001$; Fig. 4c), and the resulting lymphomas were invariably GFP–Cherry double positive (Fig. 4e and Supplementary Fig. 19). High-resolution isoelectric point (pI) separation followed by *eIF5A* immunoblotting indicated that co-depletion of *Amd1* and *Eif5a* led to lower levels of hypusinated *eIF5A* than single knockdown of either gene, providing a biochemical basis for the accelerated lymphomagenesis driven by co-suppression of both genes (Supplementary Fig. 20).

Using a mouse lymphoma model to distinguish driver from passenger lesions, we functionally validated nine tumour suppressors of B-cell non-Hodgkin's lymphoma. Two of these, *AMD1* and *eIF5A*, are involved in the biosynthesis of hypusine. This highly conserved pathway involves two enzymes (DHPS, DOHH) that specifically target the same lysine on—and apparently only on—*eIF5A*². Although its action is not established²⁸, studies in model organisms suggest that hypusinated *eIF5A* controls proliferation by regulating translation initiation or elongation^{29,30}. Paradoxically, the *eIF5A* family member *eIF5A2* can function as an oncogene in a murine hepatocellular carcinoma model⁶ but not in the system described here (data not shown), highlighting the contextual action of pathway components that, in pre-malignant B cells, act primarily to promote apoptosis. The fact that an offshoot pathway of polyamine biogenesis suppresses tumorigenesis may partly explain the failure of strategies to inhibit the polyamine pathway in the clinic³¹.

Our study also shows a previously unanticipated mechanism for disabling tumour suppressor networks, which are typically thought to be inactivated through mutation and/or loss of two alleles of the same gene. Here the ‘two hits’ arise through inactivating a single allele of two genes in the same network, whose combined effect in this case presumably reduces *eIF5A* function below the threshold needed to restrict cancer development while retaining sufficient polyamines for essential roles in gene regulation and membrane stability³¹. Although the prevalence of this mode of tumour suppressor inactivation remains to be determined, such cooperation between haploinsufficient tumour suppressors provides one strategy for minimizing the anti-proliferative output of complex networks that also control essential metabolic processes. As such interactions are impossible to identify from genomic approaches alone, our results further highlight the importance of integrating genomic analysis with functional studies to decode the complexity of cancer genomes.

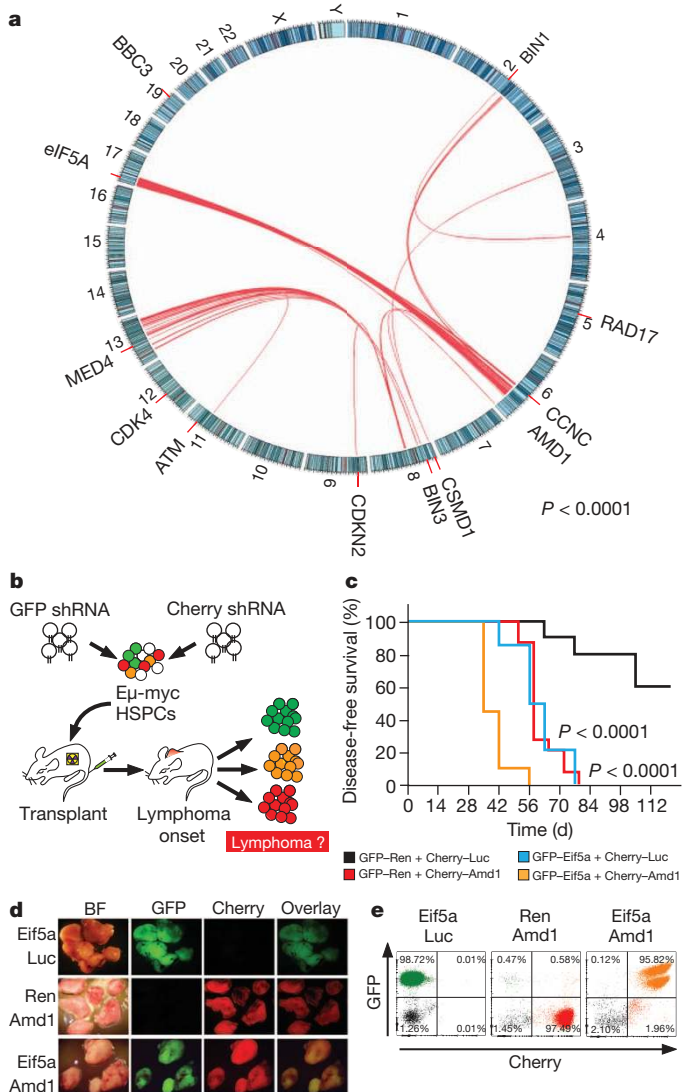


Figure 4 | Loss of *eIF5A* and *AMD1* cooperate in lymphoma progression. **a**, Deletions of *eIF5A* (17p) and *AMD1* (6q21) are significantly associated in human lymphoma. Red lines identify significant pairwise co-deletion frequency for the indicated genomic regions. Representative tumour suppressors are shown. **b**, Outline of the two-colour *in vivo* cooperation assay. **c**, Survival curves for the following shRNA combinations: GFP–Ren + Cherry–Luc ($n = 10$, black); GFP–*Eif5a* + Cherry–Luc ($n = 15$, blue); GFP–Ren + Cherry–*Amd1* ($n = 15$, red); GFP–*Eif5a* + Cherry–*Amd1* ($n = 20$, orange). P values refer to Mantel–Cox tests of the *Eif5a*–*Amd1* knockdown versus *Eif5a* + Luc or Ren + *Amd1*. Bright-field (BF), GFP and Cherry imaging (**d**) and flow cytometry (**e**) for the indicated shRNA combinations. The first shRNA was tagged with GFP, the second with Cherry.

METHODS SUMMARY

Oncogenomics-driven shRNA screen. Commonly deleted intervals were defined on the basis of oncogenomics data that were retrieved from publicly available data sets (see Methods). A mir30-based shRNA library was designed to target the mouse orthologues of genes residing in the commonly deleted intervals. The library was subdivided into 14 pools of about 100 shRNAs each and cloned in the LMS vector.

Lymphoma adoptive transplant model. E μ -myc HSPCs derived from embryonic day (E)12.5–E14.5 fetal livers were retrovirally transduced with the shRNA pools and used to reconstitute sublethally irradiated (600 rad) syngeneic C57BL/6 females. Mice were monitored weekly for development of palpable lymph nodes. All lymphomas were analysed for GFP content and B-cell immunophenotype. GFP-positive lymphomas arising in pools that significantly promoted lymphomagenesis over neutral shRNA were deep-sequenced to establish shRNA representation.

Full Methods and any associated references are available in the online version of the paper at www.nature.com/nature.

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- Knudson, A. G. Cancer genetics. *Am. J. Med. Genet.* **111**, 96–102 (2002).
- Park, M. H. The post-translational synthesis of a polyamine-derived amino acid, hypusine, in the eukaryotic translation initiation factor 5A (eIF5A). *J. Biochem.* **139**, 161–169 (2006).
- Bignell, G. R. *et al.* Signatures of mutation and selection in the cancer genome. *Nature* **463**, 893–898 (2010).
- Berger, A. H. & Pandolfi, P. P. Haplo-insufficiency: a driving force in cancer. *J. Pathol.* **223**, 137–146 (2011).
- Lenz, G. & Staudt, L. M. Aggressive lymphomas. *N. Engl. J. Med.* **362**, 1417–1429 (2010).
- Zender, L. *et al.* An oncogenomics-based *in vivo* RNAi screen identifies tumor suppressors in liver cancer. *Cell* **135**, 852–864 (2008).
- Bric, A. *et al.* Functional identification of tumor-suppressor genes through an *in vivo* RNA interference screen in a mouse lymphoma model. *Cancer Cell* **16**, 324–335 (2009).
- Salaverria, I. *et al.* Specific secondary genetic alterations in mantle cell lymphoma provide prognostic information independent of the gene expression-based proliferation signature. *J. Clin. Oncol.* **25**, 1216–1222 (2007).
- Rubio-Moscardo, F. *et al.* Mantle-cell lymphoma genotypes identified with CGH to BAC microarrays define a leukemic subgroup of disease and predict patient outcome. *Blood* **105**, 4445–4454 (2005).
- Chen, W. *et al.* Array comparative genomic hybridization reveals genomic copy number changes associated with outcome in diffuse large B-cell lymphomas. *Blood* **107**, 2477–2485 (2006).
- Bea, S. *et al.* Diffuse large B-cell lymphoma subgroups have distinct genetic profiles that influence tumor biology and improve gene-expression-based survival prediction. *Blood* **106**, 3183–3190 (2005).
- Mestre-Escorihuela, C. *et al.* Homozygous deletions localize novel tumor suppressor genes in B-cell lymphomas. *Blood* **109**, 271–280 (2007).
- Hanahan, D., Wagner, E. F. & Palmiter, R. D. The origins of oncomice: a history of the first transgenic mice genetically engineered to develop cancer. *Genes Dev.* **21**, 2258–2270 (2007).
- Schmitt, C. A. *et al.* Dissecting p53 tumor suppressor functions *in vivo*. *Cancer Cell* **1**, 289–298 (2002).
- Kornberg, R. D. Mediator and the mechanism of transcriptional activation. *Trends Biochem. Sci.* **30**, 235–239 (2005).
- Young, K. H. *et al.* Mutations in the DNA-binding codons of TP53, which are associated with decreased expression of TRAILreceptor-2, predict for poor survival in diffuse large B-cell lymphoma. *Blood* **110**, 4396–4405 (2007).
- Xue, W. *et al.* A cluster of cooperating tumor-suppressor gene candidates in chromosomal deletions. *Proc. Natl Acad. Sci. USA* **109**, 8212–8217 (2012).
- Cooper, H. L., Park, M. H. & Folk, J. E. Posttranslational formation of hypusine in a single major protein occurs generally in growing cells and is associated with activation of lymphocyte growth. *Cell* **29**, 791–797 (1982).
- Jenkins, Z. A., Haag, P. G. & Johansson, H. E. Human eIF5A2 on chromosome 3q25-q27 is a phylogenetically conserved vertebrate variant of eukaryotic translation initiation factor 5A with tissue-specific expression. *Genomics* **71**, 101–109 (2001).
- Nilsson, J. A. *et al.* Targeting ornithine decarboxylase in Myc-induced lymphomagenesis prevents tumor formation. *Cancer Cell* **7**, 433–444 (2005).
- Cano, V. S. *et al.* Mutational analyses of human eIF5A-1 – identification of amino acid residues critical for eIF5A activity and hypusine modification. *FEBS J.* **275**, 44–58 (2008).
- Sun, Z., Cheng, Z., Taylor, C. A., McConkey, B. J. & Thompson, J. E. Apoptosis induction by eIF5A1 involves activation of the intrinsic mitochondrial pathway. *J. Cell. Physiol.* **223**, 798–809 (2010).
- Eischen, C. M., Roussel, M. F., Korsmeyer, S. J. & Cleveland, J. L. Bax loss impairs Myc-induced apoptosis and circumvents the selection of p53 mutations during Myc-mediated lymphomagenesis. *Mol. Cell. Biol.* **21**, 7653–7662 (2001).
- Hudson, T. J. *et al.* International network of cancer genome projects. *Nature* **464**, 993–998 (2010).
- Lenz, G. *et al.* Molecular subtypes of diffuse large B-cell lymphoma arise by distinct genetic pathways. *Proc. Natl Acad. Sci. USA* **105**, 13520–13525 (2008).
- Parreiras-e-Silva, L. T. *et al.* Evidences of a role for eukaryotic translation initiation factor 5A (eIF5A) in mouse embryogenesis and cell differentiation. *J. Cell. Physiol.* **225**, 500–505 (2010).
- Nishimura, K. *et al.* Essential role of S-adenosylmethionine decarboxylase in mouse embryonic development. *Genes Cells* **7**, 41–47 (2002).
- Park, M. H., Nishimura, K., Zanelli, C. F. & Valentini, S. R. Functional significance of eIF5A and its hypusine modification in eukaryotes. *Amino Acids* **38**, 491–500 (2009).
- Henderson, A. & Hershey, J. W. Eukaryotic translation initiation factor (eIF) 5A stimulates protein synthesis in *Saccharomyces cerevisiae*. *Proc. Natl Acad. Sci. USA* **108**, 6415–6419 (2011).
- Saini, P., Eyley, D. E., Green, R. & Dever, T. E. Hypusine-containing protein eIF5A promotes translation elongation. *Nature* **459**, 118–121 (2009).
- Casero, R. A. & Marton, L. J. Targeting polyamine metabolism and function in cancer and other hyperproliferative diseases. *Nature Rev. Drug Discov.* **6**, 373–390 (2007).

Supplementary Information is linked to the online version of the paper at www.nature.com/nature.

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Author Contributions C.S., C.M. and S.W.L. designed the study. C.S. and C.M. performed the *in vivo* screen and analysed data. C.S., L.L. and J.P. performed eIF5A hypusination assays and analysed data. C.R. and D.P. performed the iTRAQ analysis. C.S., J.R., S.Y. and A.K. analysed the oncogenomics data. S.W.L. supervised the project. C.S., C.M. and S.W.L. wrote the paper. C.S., C.M. and I.A. performed experiments on human cell lines. J.T. analysed tissue microarrays. All authors reviewed the manuscript.

Author Information The cell line data are deposited in the National Center for Biotechnology Information Gene Expression Omnibus database under accession number GSE36967. Reprints and permissions information is available at www.nature.com/reprints. The authors declare no competing financial interests. Readers are welcome to comment on the online version of this article at www.nature.com/nature. Correspondence and requests for materials should be addressed to S.W.L. (lowes@mskcc.org).

METHODS

Oncogenomics data. To generate the lymphoma screening interval, deletion data were retrieved from the references and clustered if reported by several sources. Overlapping intervals (Supplementary Table 2) were used to generate the gene list (Supplementary Table 3). Murine orthologues were obtained from Biomart and were used to generate the shRNA library. Control libraries consist of shRNA pools targeting genes deleted in other cancers including the following: hepatocellular carcinoma (eight pools⁶); breast cancer (three pools, targeting genes frequently deleted (>5%) and included in focal deletions (<5 Mb)); acute myeloid leukaemia (four pools, targeting genes included in focal deletions at 7q^{32,33}). SNP profiles of human lymphoma cell lines used in this study are deposited in the Gene Expression Omnibus database under accession number GSE36967.

Vectors and library construction. To generate the shRNA library, 101-base-pair oligonucleotides (including the mir30-shRNA precursor, XhoI–EcoRI sites and a pool-specific barcode) were synthesized on a 55k customized oligonucleotide array (Agilent Technologies). Pool-specific PCR products were individually cloned in the LMS vector³⁴. The LMS–Cherry vector was constructed from the LMS vector by standard cloning techniques. Viruses were produced by transient transfection of Phoenix ecotropic packaging cell line. shRNA sequences are reported in Supplementary Table 6.

Tumour sequencing. DNA was extracted from lymphoma cell pellets using a modified Laird protocol. Briefly, lymphoma single-cell suspensions were prepared by passing lymph nodes through a 100 μM mesh. Laird buffer (0.2% SDS, 5 mM EDTA, 200 mM NaCl, 100 mM Tris, pH 8.5) supplemented with 5 $\mu\text{g ml}^{-1}$ Proteinase K (Roche) was added to lymphoma pellets that were incubated overnight in an Eppendorf Thermomixer at 55 °C at 6.5g agitation. After centrifugation, DNA was extracted by adding a volume of isopropanol. DNA pellets were washed in 70% ethanol, air dried and resuspended in TE buffer. shRNA representation was determined by both mass Sanger sequencing (30–100 colonies per tumour) and by Solexa bar-coded deep-sequencing. For the former method, shRNAs were amplified by PCR with vector-specific primers (MSCV.Bgl2, 5'-TCCTTCTCTAGGCGCCGGAATTAG-3' mir30.EcoRI 5'-CTAAAGTAGC CCCTGAATTCCGAGGCAGTAGGCA-3'). The PCR products were digested and cloned in the LMS vector. shRNAs were identified by sequence alignment using the Blat algorithm³⁵. For the latter method, a PCR-based barcoding strategy was designed. Briefly, shRNA sequences were amplified from the genomic tumour DNA with primers annealing to the shRNA loop (forward primer) and the miR30 3'-region (reverse primer). The primers also contained the P5 and P7 adaptor sequences required for sequencing on the Illumina GAII platform. Furthermore, the forward primer carried an eight-nucleotide barcode sequence between the loop- and the P5-sequence. After amplification, PCR products from individual tumour samples amplified with specific barcodes were purified, quantified and pooled. Approximately 2×10^6 50-base-pair reads were acquired for each sample. By reading 50 nucleotides into the amplicon starting from the shRNA guide strand, we were able to deconvolute the different tumour samples according to the sample-specific barcode. The specific sequences were subsequently identified by comparing the sequences with the original library sequences using the BLAT algorithm, and the relative distribution was calculated. A gene enrichment score was calculated by adding the percentages of enrichment of all shRNAs in each tumour and multiplying it by the number of lymphomas and the number of different shRNAs targeting the same gene.

Haematopoietic stem and progenitor cell isolation and *in vivo* adoptive transfer. $\text{E}\mu\text{-myc}$ HSPCs were prepared from E13.5 fetal livers as described³⁶ and retrovirally transduced with shRNA pools or individual shRNA. After short-term *in vitro* expansion, 6×10^5 HSPCs were transplanted by tail-vein injection into sublethally irradiated (600 rad) 6- to 8-week-old syngenic C57/BL6 females. Recipient mice were administered ciprofloxacin (125 mg l^{-1} , Sigma–Aldrich; supplemented with sucrose 20 g l^{-1}) in the drinking water for 1 week before and 2 weeks after transplantation.

Lymphoma monitoring and analysis. Mice were examined weekly by lymph node palpation. Disease state was defined by the presence of palpable lymph nodes or paralysis. Statistical evaluation of survival curves was performed using the log-rank (Mantel–Cox) test. To assess cell immunophenotype and GFP content, single-cell suspensions were obtained by passing lymph nodes through a 100 μM mesh, treated with ACK buffer (0.15 M NH_4Cl , 10 mM KHCO_3 , 0.1 mM EDTA) for 5 min and resuspended in PBS. Pe-Cy5-B220 and Pe-Cy5-IgM (Biolegend) were used for B-cell immunophenotyping. Samples were acquired using a LSR-II Flow Cytometer System (BD Biosciences). All animal experiments were performed according to federal, National Institutes of Health and local Institutional Animal Care and Use Committee guidelines. All mouse experiments were approved by the Cold Spring Harbor Laboratory Institutional Animal Care and Use Committee.

Apoptosis assays. Spleens of asymptomatic mice were dissected 3 weeks after transplant in B6/SJL mice (CD45.1⁺), and cells were treated with ACK buffer as described above. Half a million cells were washed in PBS, and incubated for 15 min at room temperature in 100 μl Annexin V Buffer (10 mM Hepes, 140 mM NaCl, 25 mM CaCl_2 , pH 7.4) with 2 $\mu\text{g ml}^{-1}$ propidium iodide (Sigma–Aldrich) and 5 μl Pacific Blue Annexin V (Biolegend). For cleaved caspase-3 immunoblotting, spleen single-cell suspensions were first depleted of host cells using MagnaBind Streptavidin Beads (Pierce) coupled to a biotin anti-mouse CD45.2 antibody (Biolegend) according to the manufacturer's protocol. The unbound fraction was then positively selected for B cells using a Dynabeads Mouse Pan B cell kit (Invitrogen). Correct immunophenotype was confirmed by flow cytometry for B220 expression as described above. The neutral control sample (luc) was obtained by pooling spleens from three mice.

Western blot. Lymphoma pellets were lysed in modified RIPA buffer (20 mM Tris pH 7.5, 100 mM NaCl, 1 mM EDTA, 1 mM EGTA, 0.1% NP-40, 0.1% SDS) and protein concentration was determined using the DC Protein Assay (Biorad). Twenty micrograms per sample were subjected to SDS–PAGE and transferred to Immobilon P-membrane (Millipore). The following antibodies were used: anti-eIF5A (ab32407, Abcam; 1:2,000); anti-Med4 (ab75791, Abcam; 1:1,000); anti-Bin3 (ab67032, Abcam; 1:1,000); anti-cleaved caspase-3 (number 9664, Cell Signalling, 1:500); anti-p19Arf (Upstate, 1:500); anti-Puma (number 7467, Cell Signalling 1:1,000); HRP-anti-actin (A3854, Sigma; 1:20,000). For Cmsd1, a rabbit polyclonal serum was developed by Prime Biotech. Blots were visualized using ECL (Amersham).

Two-dimensional PAGE. Lymphomas were washed in PBS and then lysed in urea lysis buffer (8 M urea, 2% Chaps, 10 mM DTT, 0.5% IPG pH 4–7 buffer (Amersham), trace of bromophenol blue) at room temperature for 20 min. Membranes were removed by centrifugation, and 75 μg of extract was loaded on Immobiline Dry Strips (pH 4–7, 13 cm) (GE Healthcare) for isoelectric focusing with an IPGphor (Amersham Pharmacia Biotech). Strips were rehydrated in sample for 11–14 h, and run at 150 V for 2 h, 500 V for 1 h, 1,000 V for 1 h and 8,000 V for 4 h. Isoelectric focusing strips were then equilibrated for 15 min in modified SDS sample buffer (50 mM Tris pH 6.8, 6 M urea, 30% glycerol, 2% SDS, 1% DTT and trace bromophenol blue), separated in the second dimension by 15% SDS–PAGE, transferred to PVDF membranes and subjected to eIF5A and β -actin immunoblotting as an internal control. GC-7 was purchased from Biosearch. Quantification of hypusinated eIF5A was performed by densitometry (Image J). Error bars, s.d.

OFFGEL fractionation. Mouse embryo fibroblasts (p53^{-/-}) were infected with the indicated combination of shRNAs. Flow cytometry analysis indicated that GFP + Cherry + content defining the double-infected fraction was greater than 90% in all conditions. After 3 days, cells were lysed as described above and 1 mg protein was precipitated using one volume of TCA. Protein pellets were washed three times with acetic acid and resuspended in the manufacturer's buffer (Agilent). Samples were separated on a 3100 OFFGEL Fractionator (Agilent) using high-resolution pH 4–7 OFFGEL Strips according to the manufacturer's instructions. After fractionation was completed, fractions 4–17 were separated on a 12% SDS–PAGE, transferred to PVDF membranes and blotted for eIF5A.

eIF5A hypusination quantification. Images from films were acquired with CanoScan Lide 110 in TIFF format (600 dots per inch, black and white mode). Image processing and quantification were performed with ImageJ 1.43. After noise was removed with the Despeckle option (Process/Noise/Despeckle), images were inverted (edit/invert). For one-dimensional SDS–PAGE, the measure area was set to rectangular selection so that it would include the entire length of the band. Background was quantified by averaging 10 random rectangular selections across the image in areas where no signal was present and subtracted from the signal for both eIF5A and actin. For the hypusination profile, total eIF5A was quantified by one-dimensional SDS–PAGE as described. After two-dimensional SDS–PAGE and eIF5A immunoblotting, images were acquired and elliptical selection was used to quantify each spot. The ratio of hypusinated eIF5A to total was calculated by dividing the densitometry value of the hypusinated form by the sum of the values for all the forms.

Quantitative PCR. RNA was extracted from tumours or cells using Rneasy Kit (Qiagen). RNA (2 μg) was used for first strand synthesis (Taqman RT Reagents Kit, Applied Biosystem). Quantitative PCRs were performed in triplicate on a iCycler iQ5 (Biorad) with SYBR Green PCR MasterMix (Applied Biosystem). Beta-actin was used as normalization control. Error bars, s.d. Quantification was based on a standard curve obtained by serial dilution of the indicated control RT reaction. Primer sequences are reported in Supplementary Table 7.

Spermidine competition assay. Lymphomas (sh.Amd) were plated in B-cell media (45% Iscove's modified Eagle medium, 45% Dulbecco's minimal essential medium, 10% fetal bovine serum supplemented with 100 U ml^{-1} penicillin and streptomycin and 55 μM 2-mercaptoethanol) and partly transduced with the

indicated retroviral constructs. Cells were then treated with the indicated concentration of spermidine (Sigma-Aldrich) in modified B cell media supplemented with 1 mM aminoguanidine (Sigma-Aldrich) and dialysed bovine serum (Gibco). After treatment for 24 h, cells were diluted with fresh media and allowed to recover for 72 h. Cherry-positive fractions were assessed using a LSR-II Flow Cytometer System (BD Biosciences). The adriamycin competition assay was performed in μ -myc Arf^{-/-} lymphomas using GFP-tagged shRNA vectors as previously described.

Deletion association study. High-resolution lymphoma comparative genomic hybridization data were obtained from the Gene Expression Omnibus database (accession number GSE11318). Normalized comparative genomic hybridization profiles were segmented using the circular binary segmentation algorithm implemented in DNACopy (www.bioconductor.org). Copy number gains and losses were determined by identifying individual segments with a mean \log_2 ratio greater than 0.2 and lower than -0.2, respectively. To identify gene pairs with significant co-deletion frequency for each gene pair in a particular gene set, we computed the *P* value for the observed co-deletion frequency under the assumption of independent deletion (Fisher's exact test). When evaluating the significance of particular gene pairs, and to account for possible confounding factors such as genomic distance and deletion size, we obtained a null distribution by computing the *P* values for random pairs of genes residing on the same chromosomes of the tested genes; if these resided on the same chromosome, we used random pairs with a similar genomic distance as the tested genes. Significant gene pairs were determined as those having a *P* value less than the 95% quantile of the null distribution.

iTRAQ. Pelleted cells were lysed mechanically with 18- to 25-gauge needles in 300 μ l lysis buffer (10 mM HEPES pH 8, 0.5 M EDTA, 0.5% NP-40) supplemented with phosphatase inhibitors and protease inhibitor cocktail (P2850, P5726, P8340; Sigma-Aldrich). Protein concentration was measured using BCA Protein Assay. An aliquot of 100 μ g of each sample was further solubilized by adding ProteaseMax (Promega) to 0.1%. Samples were reduced with tris-(2-carboxyethyl) phosphine (5 mM) and alkylated with methyl methanethiosulphonate (10 mM). Each sample was then precipitated using a methanol/chloroform extraction. After reconstitution in 50 mM triethylammonium bicarbonate (TEAB), 0.1% ProteaseMax, each sample was digested with trypsin (1:50) overnight at 37 °C. The tryptic peptides were reduced to a final volume of 20 μ l in a speed-vac, and 30 μ l of 500 mM TEAB solution were added to each solution. iTRAQ labelling of peptides was according to the previous report³⁷. After labelling, each solution was acidified by the addition of 3 μ l trifluoroacetic acid and combined. Mixed samples were reduced to approximately 40 μ l. Combined samples were then analysed by two-dimensional liquid chromatography MudPIT³⁸, using a two-dimensional Vented Column Setup with a Proxeon nano-flow high-performance liquid pump³⁹. An equivalent of about 50 μ g of sample was loaded onto a tri-phasic, fused silica capillary column: 250 μ m internal diameter packed with 3 cm of 5 μ m Aqua C₁₈, followed by 3 cm of 5 μ m Luna SCX and 2 cm of 5 μ m Aqua C₁₈. For each salt step, peptides were eluted on an analytical column of a 100 μ m internal diameter capillary with a 5 μ m pulled tip and packed with 15 cm of 3 μ m Aqua C₁₈ on line with an LTQ Orbitrap Velos (Thermo Electron). Data were acquired in profile mode using the following parameters: for full-scan Fourier transform mass spectrometry, resolution = 60,000, *m/z* = 380–1,700 and the 10 most intense ions were fragmented with higher-collision dissociation at a normalized collision energy of 40% and an activation time of 0.1. Minimum threshold signal was at 5,000 and isolation width at 1.2. Dynamic exclusion settings were repeat count 1, repeat duration of 30, exclusion list size 500, exclusion duration 60 and exclusion mass width 10 p.p.m.

For data analysis, peaklist files were generated by Distiller (Matrix Science). Protein identification and quantification was performed on Mascot 2.3 (ref. 40) against the International Protein Index mouse database (version 3.86; 58,667 sequences; 26,399,545 residues). Methylthiolation of cysteine, amino (N)-terminal and lysine iTRAQ modifications were set as fixed modifications, methionine oxidation and deamidation as variables. Trypsin was used as cleavage enzyme and one missed cleavage allowed. Mass tolerance was set at 20 p.p.m. for intact peptide mass and 0.5 Da for fragment ions. Results were re-scored with Mascot percolator to give an overall 0.8% false discovery rate for protein identification. Protein-level iTRAQ ratios were calculated as intensity weighted, using only peptides with expectation values less than 0.05. Global ratio normalization was performed using intensity summation, with no outlier rejection. To assign fold-change significance, protein ratios in each iTRAQ channel were fitted to a three-parameter Burr distribution (a member of the log-normal distribution family). Protein ratio fold-changes greater or less than 2 s.d. were assigned as significant for up- or downregulation respectively.

Tissue microarrays. Tissue microarrays were constructed as previously published^{41,42}, using a fully automated Beecher Instrument, ATA-27. The study cohort comprised diffuse large B-cell lymphomas (173 cases) consecutively ascertained at the Memorial Sloan-Kettering Cancer Center between 1989 and 2008. All biopsies were evaluated at Memorial Sloan-Kettering Cancer Center, and the histological diagnosis was based on haematoxylin and eosin staining. Use of tissue blocks was approved by the Institutional Review Board and Human Biospecimen Utilization Committees. Anti-eIF5A (rabbit monoclonal antibody, EP57Y, ABCAM catalogue number ab32407) was used at a 1:2,000 dilution for 30 min after heat-induced epitope retrieval with citrate buffer for 30 min, polymer from envision kit from Dako for 30 min, then 3,3'-diaminobenzidine for 5 min. TP53 was performed on an automated platform according to the manufacturer's instructions (Ventana). For eIF5A and TP53, tumour cells were scored as follows: 0, negative; 1, less than 50% of tumour cells positive; 2, greater than 50% of tumour cells positive. Array chromosomal comparative genomic hybridization was performed in collaboration with J. Houldsworth at Cancer Genetics.

32. Curtiss, N. P. *et al.* Isolation and analysis of candidate myeloid tumor suppressor genes from a commonly deleted segment of 7q22. *Genomics* **85**, 600–607 (2005).
33. Kratz, C. P. *et al.* Candidate gene isolation and comparative analysis of a commonly deleted segment of 7q22 implicated in myeloid malignancies. *Genomics* **77**, 171–180 (2001).
34. Dickens, R. A. *et al.* Probing tumor phenotypes using stable and regulated synthetic microRNA precursors. *Nature Genet.* **37**, 1289–1295 (2005).
35. Kent, W. J. BLAT—the BLAST-like alignment tool. *Genome Res.* **12**, 656–664 (2002).
36. McCurrach, M. E. & Lowe, S. W. Methods for studying pro- and antiapoptotic genes in nonimmortal cells. *Methods Cell Biol.* **66**, 197–227 (2001).
37. Ross, P. L. *et al.* Multiplexed protein quantitation in *Saccharomyces cerevisiae* using amine-reactive isobaric tagging reagents. *Mol. Cell. Proteom.* **3**, 1154–1169 (2004).
38. Washburn, M. P. Utilisation of proteomics datasets generated via multidimensional protein identification technology (MudPIT). *Brief. Funct. Genom. Proteom.* **3**, 280–286 (2004).
39. Taylor, P. *et al.* Automated 2D peptide separation on a 1D nano-LC-MS system. *J. Proteome Res.* **8**, 1610–1616 (2009).
40. Perkins, D. N., Pappin, D. J., Creasy, D. M. & Cottrell, J. S. Probability-based protein identification by searching sequence databases using mass spectrometry data. *Electrophoresis* **20**, 3551–3567 (1999).
41. Hedvat, C. V. *et al.* Application of tissue microarray technology to the study of non-Hodgkin's and Hodgkin's lymphoma. *Hum. Pathol.* **33**, 968–974 (2002).
42. Hemann, M. T. *et al.* Evasion of the p53 tumour surveillance network by tumour-derived MYC mutants. *Nature* **436**, 807–811 (2005).