

Review

# Acute ischemia-induced gap junctional uncoupling and arrhythmogenesis

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Received 7 October 2003; received in revised form 12 January 2004; accepted 29 January 2004

Time for primary review 28 days

## Abstract

Sudden cardiac death forms a major cause of mortality. Myocardial ischemia-induced ventricular fibrillation (VF) is frequently the underlying mechanism. Ventricular arrhythmias arise in two distinct phases during the first hour of ischemia. The first, the 1A phase, has been extensively studied, and few studies relate to the 1B phase. The latter is associated with intercellular electrical uncoupling, mediated by decreased conductance of gap junction channels.

Although the relation between gap junctional uncoupling and decreased conduction velocity appears clear under normoxic conditions, additional factors contribute to conduction slowing during ischemia, and VF occurs preferentially at moderate levels of uncoupling. A potential mechanism of arrhythmias depends on temporary electrotonic depression of intrinsically viable tissue by the large bulk of the ischemic zone. This causes conduction slowing and conduction block in the surviving layers, leading to arrhythmias. These arrhythmias then resolve with progression of uncoupling. It is unknown whether either accelerated uncoupling or maintenance of gap junctional communication is antiarrhythmic. Ischemic preconditioning postpones both gap junctional uncoupling and occurrence of VF. Given the burden of sudden death and the large number of casualties in the low-risk population, there is, even in the era of implantable cardiac defibrillators, need for further understanding the mechanism of ischemia-induced VF.

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*Keywords:* Ischemia; Gap junctions; Ventricular arrhythmia; Cellular uncoupling; Connexin; Arrhythmia mechanism

## 1. Introduction

Cardiovascular mortality remains the leading cause of death in the industrialized world [1,2]. Sudden death, defined as ‘death outside the hospital,’ ‘dead on arrival,’ or ‘dead in the emergency department,’ was responsible for almost 300,000 casualties in the United States alone in 1998 [3]. Ventricular arrhythmias, ventricular fibrillation (VF) in particular, are the cause of cardiac arrest in the majority of cases [1]. Acute myocardial ischemia is the major cause of sudden cardiac death [1,4]. Large-scale clinical trials with antiarrhythmic drugs have failed to reduce the incidence of sudden death and even caused an increased mortality in the treated groups [5–7]. Implantable defibrillators are efficient against

ventricular arrhythmias [8,9], but risk stratification of sudden death is only possible in the high-risk population, whereas most deaths occur in the far larger low-risk population [10]. Therefore, our understanding of the underlying mechanism of ischemia-induced ventricular arrhythmias appears to be incomplete. In this review, we will first discuss the mechanism of ventricular arrhythmias and gap junctional closure during acute myocardial ischemia, then look at the direct influence of gap junctional uncoupling on conduction velocity and the creation of heterogeneities. After that, we will review the data supporting the role of gap junctions in the genesis of ischemia-induced arrhythmias.

### 1.1. Electrophysiologic mechanism of ventricular fibrillation

Reentry is the underlying electrophysiological mechanism of ventricular fibrillation [4]. Reentry was first

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defined by Mines [11] in 1914 as a persisting electrical impulse that reactivates an area of previously activated myocardial tissue that is no longer refractory, resulting in a circus movement of activation. The length of such circle depends on its wavelength, defined by the mathematical product of refractory period and conduction velocity (plus an excitable gap when present) [12]. The requirements for reentrant activation in the intact heart are a region of unidirectional block and (regionally) slow-enough conduction velocity to allow the activation impulse to travel around the zone of block. It is facilitated by a short refractory period. The ultimate proof of reentry is its termination by interruption of the circle [11]. Our understanding of reentry has been extended by the introduction of different concepts of its mechanism such as single rotor reentry [13] and fibrillatory conduction [14,15] and the leading circle concept [16], but Mines' principles apply in all these forms of ventricular fibrillation.

For an arrhythmia to occur, both a suitable substrate (the preexisting circumstances that allow perpetuation of the arrhythmia) and a trigger (the event that sets off the arrhythmia within the substrate) need to be present [17]. Factors such as catecholamines,  $[K^+]_o$ , pH, and drugs might modulate both trigger and substrate. During acute myocardial ischemia, the conditions necessary for the initiation of reentry, both trigger and substrate, occur in concert and, indeed, ventricular fibrillation is often encountered [4].

In the course of ischemia, gap junctions close. This may result in electrophysiological effects. Its first results were noted as early as 1875 and 1879 by Engelmann [18], who stated that the cells live together but die singly, and by Burden-Sanderson and Page [19] and De Mello et al. [20], respectively, who described the phenomenon of 'healing over'—the reversal of (electrical) signs of myocardial injury following a stab wound of the heart. Gap junctional uncoupling may contribute to attaining conditions favorable for the initiation of reentrant activation and ventricular fibrillation by slowing of conduction (both directly and indirectly) and the creation of heterogeneities.

### 1.2. Closure of gap junctions in ischemia

Tissue impedance, the composite measure of resistance and reactance and an indirect measure for intercellular coupling, increases in a biphasic manner following coronary occlusion [21]. Immediately after interruption of coronary flow, a first rise of approximately 10–25% occurs, attributed to collapse of the vasculature [21]. Fleischhauer et al. [22] showed that the impedance of the perfusion fluids contributes little to total tissue impedance. However, collapse of the vasculature affects the relation between the intracellular and extracellular volume and might therefore affect whole tissue impedance. The second rise in impedance, after approximately 15 min, was attributed to closure of the gap junctions, and thus to decrease of intercellular conductance [21].

#### 1.2.1. Ischemia-related factors responsible for closure of gap junctions during ischemia

Although many factors change simultaneously during early ischemia, several individual factors that uncouple gap junctions have been identified. Increase of diastolic cytoplasmic  $[Ca^{2+}]_i$  is associated with gap junctional uncoupling [23,24]. Diastolic  $[Ca^{2+}]_i$  increases between 15 and 25 min of ischemia in rabbits [24] and closely precedes gap junctional uncoupling, which lasts between 10 min (rabbit) and 40 min (pig) [25–27] and results in conduction slowing and conduction block [24].

Ischemia-induced intracellular acidification also decreases gap junctional conductance [28]. Decreased intracellular pH also renders gap junctions more sensitive to increased  $[Ca^{2+}]_i$  [29].

Lysophosphoglycerides and arachidonic acid metabolites accumulate in the intercalated disks of ischemic cells after a few minutes of ischemia and decrease gap junctional conductance [30]. Catecholamines increase cAMP and  $[Ca^{2+}]_i$ , which in turn decrease gap junctional conductance [31].

Cx43 proteins dephosphorylate during ischemia [32,33] and transfer from the intercalated disks to intercellular pools [33]. The latter reports show a direct link between energy deprivation as the common pathway leading to membrane depolarization,  $Ca^{2+}$  overload, anaerobic glycolysis leading to decreased  $pH_i$ , and gap junctional uncoupling.

### 1.3. Gap junctions and conduction velocity

Conduction velocity partially depends on intercellular conductance, the combined resistance of the cytoplasm, and gap junctions.

The number of available gap junctions is much larger than needed for propagation of the action potential under normoxic conditions. Weingart and Maurer [34] showed action potential propagation within a pair of coupled cells at gap junctional conductance of  $>1.3$  nS, albeit with a considerable delay. Rudy and Quan [35] showed in a computer simulation that conduction velocity decreases discontinuously at high gap junctional resistance. Indeed, Jongasma and Wilders [36] confirmed that, under non-ischemic conditions, approximately 90% decrease of gap junctions is required to decrease conduction velocity with 50%. In another study, it was demonstrated that the safety for conduction is much higher when gap junctional coupling is reduced than when  $I_{Na}$  is decreased [37]. Thus, slow conduction without conduction block remains possible even at very low gap junctional conductance. These simulation studies corroborate studies by Gutstein et al. [38] in genetically engineered conditional Cx43  $-/-$  mice where a 90% reduction of Cx43 was associated with a decrease of approximately 50% in transversal and longitudinal conduction velocity. This is consistent with the studies of Morley et al. [39] and Thomas et al. [40] who showed little effect of heterozygous Cx43 reduction on

conduction velocity. However, others found that in Cx43 +/- mice, a 50% reduction of Cx43 protein corresponded to a decrease of conduction velocity of approximately 25% [41,42].

The former observations are further supported by studies demonstrating conduction slowing (more prominent in transverse than in longitudinal conduction) upon administration of heptanol or palmitoleic acid [43–46]. Increase of gap junctional resistance with palmitoleic acid decreased conduction velocity to such an extent in cultured neonatal cells that activation fronts could propagate around the perimeter of a single cell [47]. Gap junctional conductance depends in a dynamic manner on transjunctional voltage [48,49] and effect that is larger when there are less gap junctions [49]. Most data on conduction slowing through gap junctional uncoupling relate to normoxic conditions. In the acutely ischemic intact heart, very slow conduction has not been demonstrated. Whether this is because gap junctional coupling remains sufficient for propagation and other factors are responsible for conduction slowing, or whether conduction block occurs before gap junctions uncouple is not exactly known.

#### 1.4. Unmasking of heterogeneities

Gap junctions allow intercellular exchange of ions and small molecules (with a molecular weight of up to 1000 Da [50]) and current, and cause equilibration of ionic concentrations [51,52] and energy-rich phosphates and propagation of the action potential. Their closure, by either pharmacological interventions or by pathological circumstances such as acute ischemia, might create an intercellular gradient in both nutrients and metabolites as in ions. Han and Moe became aware of this equilibrating effect. It was hypothesized that when two adjacent cells have a very different duration of the action potential, the current flowing from the cell with the longer action potential towards the cell with the shorter action potential during the plateau phase would allow the latter cell to depolarize and to generate a premature beat. Mendez et al. argued that this could not be the case between well-coupled cells because the current flowing from the cell with the longer action potential would prolong the short one and would itself shorten the longer action potential [53]. This would lead to an equilibration of action potential durations between adjacent cells. Thus, for a voltage gradient between cells to occur, partial gap junctional uncoupling is required. Indeed, closure of gap junctions causes gradients that are large enough to produce arrhythmias [51].

However, it appears that gap junctional conductance does not decrease to such an extent during ischemia, at least not while cells are excitable. In coupled cell pairs subjected to simulated ischemia, gap junctional coupling remained large enough to equilibrate action potential duration between the paired cells up to the moment of

inexcitability [54]. Moreover, the moment of 'ischemia'-induced rigor was exactly the same in two paired cells, whereas there was a large variation in a group of single myocytes [54]. Thus, gap junctional coupling remains sufficient to equilibrate action potential duration and moment of ischemia-induced rigor in cell pairs, suggesting that intercellular communication remains intact and prevents intercellular gradients on a cellular level. In support of the above, gap junctions were still permeable for sodium ions, on one hand, causing calcium overload via the  $\text{Na}^+/\text{Ca}^{2+}$  exchanger [52], and for luciferine yellow, on the other hand [55], at the moment rigor occurred. It remains to be determined what degree of uncoupling is required to cause physiologically significant heterogeneities in intact hearts.

In addition to the potential generalized effect of uncoupling on the unmasking of heterogeneities, gap junctional uncoupling is not equally distributed within the ischemic tissue. The increase in tissue impedance was significantly smaller in the ischemic border zone than in the central zone, although the time course of rise was identical [27]. Hence, ischemic and nonischemic myocardium interdigitate at the ischemic border [56] and ischemic tissue impedance increases, whereas nonischemic tissue impedance remains normal. Also, subepicardium and subendocardium are relatively unaffected by the ischemic burden through diffusion of oxygen and nutrients from surrounding tissues [57,58]. Irreversibly damaged cells will eventually die and reversibly challenged myocytes will dissociate from irreversibly damaged cells and survive ischemia (healing over) [19]. The ischemic myocardium that uncouples from the rest of the heart can therefore functionally no longer contribute to arrhythmogenesis.

## 2. Arrhythmogenesis during myocardial ischemia

### 2.1. Two phases of arrhythmias

Arrhythmias occur in two distinct phases during the first hour of coronary occlusion [4,59,60]. The first phase, called immediate ventricular arrhythmias by Kaplinsky et al. [59], later referred to as 1A [60] arrhythmias, lasts from 2 to 8 min of ischemia in the dog. After a relative arrhythmia-free interval, a second, delayed phase, now referred to as 1B [60], occurs from 15 to 45 min of coronary occlusion in dogs and pigs [25–27,59]. This phase has remained relatively poorly studied but appears to be more arrhythmogenic than the 1A phase [25,59]. The 1B phase of arrhythmias coincides with the increase of tissue impedance and therefore was thought to be causally related with gap junctional uncoupling [21]. Fig. 1, derived from the work of Smith et al. [25], shows the distribution of arrhythmic events in nine open-chested pigs. Arrhythmias occurred at the start of the rise of the tissue impedance. However, others have shown

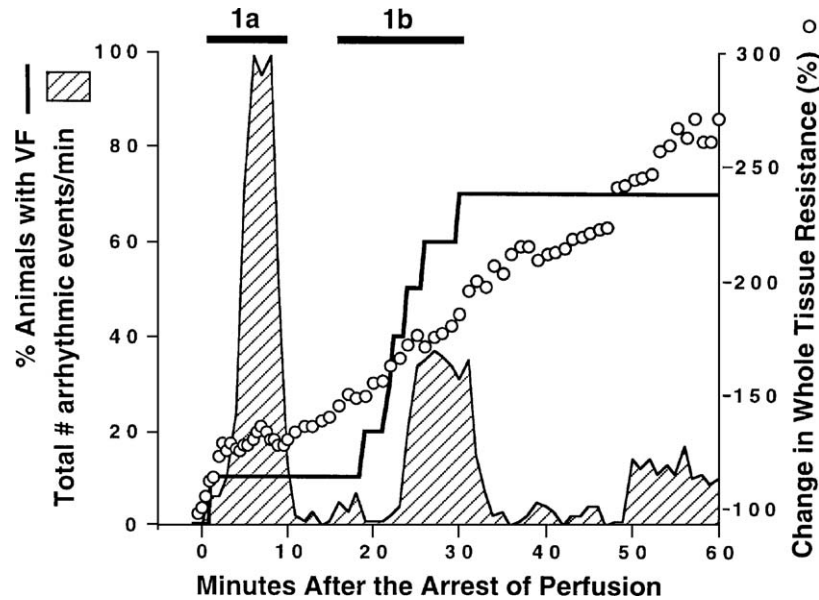


Fig. 1. Incidence of ventricular arrhythmias in regionally ischemic open-chested pig hearts (dashed area). Black line denotes percentage of animals with VF. The open circles indicate average rise in tissue impedance in all animals. The black line indicates the time course and number of animals with VF (reproduced with permission from Ref. [25]).

that the peak of 1B arrhythmias is of shorter duration than the duration of tissue impedance rise [26,59].

Both the ischemia-induced rise in tissue impedance and phase 1B ventricular arrhythmias can be successfully postponed after ischemic preconditioning [26]. Fig. 2 shows the distribution of PVCs during 4 h of regional ischemia in pigs without (upper panel) and with ischemic preconditioning (lower panel).

The two phases of arrhythmias have been described in dogs [59,61], pigs [25,26], sheep [62], and rats [63]. The bimodal distribution is less clear in cats [64] and rabbits [65], and may differ in individual animals [66]. It is unclear whether a similar distribution of ischemia-induced arrhythmias exists in man, as this is obviously extremely difficult to study.

## 2.2. Arrhythmogenic triggers

In case of ischemia-induced VF (both 1A and 1B), the trigger exists most often from a timely administrated or spontaneous premature ventricular complexes (PVCs) [4] that can be reentrant or nonreentrant in origin [67–69]. The mechanical stretch exerted by the viable myocardium on the rigid ischemic zone may result in PVCs arising preferentially from the ischemic border during the 1B phase [70]. Indeed, the number of PVCs was larger in working hearts than in isolated nonworking hearts, the triggers are initiated at the interface between the ischemic and the viable tissue, and premature beats occur preferentially following potentiated contractions in the viable myocardium [70]. However, a recent study has shown that gadolinium, a blocker of stretch-sensitive channels, did not abate the 1B phase of arrhythmias [71].

## 2.3. Arrhythmogenic substrates

### 2.3.1. Role of functional changes in gap junctional coupling

The mechanism by which gap junctional uncoupling causes conduction slowing and arrhythmias in the regionally ischemic heart has not completely been elucidated. Recently, a novel hypothesis was put forward involving the heterogeneity of the myocardium and the evolution of a surviving subepicardial and subendocardial layer [27]. Conduction slowing in the surviving tissue is caused by electrotonic interaction between the large mass of depolarized–dying–intramural cells and nonischemic subepicardial and subendocardial cells [72]. It has been established that a rim of subepicardial and subendocardial tissue survives ischemia and infarction [27,57] and that intramural sites become electrically inexcitable during prolonged ischemia, whereas subepicardial cells remain activated [25]. If the viable myocytes are electronically depressed by electrotonic interaction, slow conduction in the intrinsically viable layer would ensue, which would recover with progression of gap junctional uncoupling (decrease of electrotonic interaction), concomitant with the decrease of arrhythmias.

Several observations support this hypothesis: (1) between two coupled cells, electrical depression of one cell can be transmitted via reduced gap junctional coupling to the other, provided that the mass of the depressant is large enough [72]; (2) VF could be induced with programmed stimulation between 14 and 53 min of ischemia [27]. Thereafter, the same induction protocol failed to induce VF [27]. Fig. 3 shows electrograms of VF inducibility in a typical isolated regionally ischemic pig heart. The number of PVCs to induce VF decreases from three during control to one at 32 min of ischemia, after which more PVC are required to



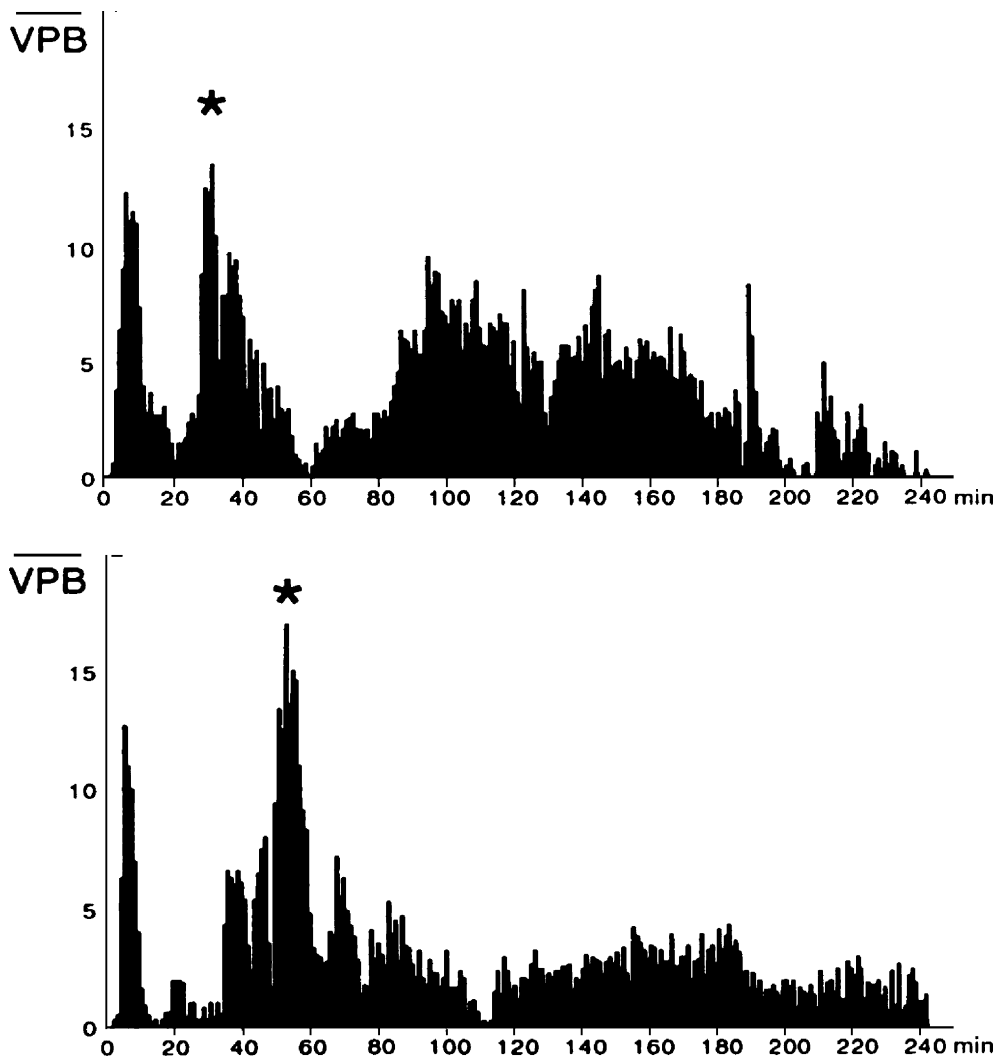


Fig. 2. Incidence of ventricular arrhythmias in regionally ischemic pig hearts without (upper panel) and with ischemic preconditioning (lower panel). The peak of arrhythmogenesis associated with the 1B phase of ischemia is postponed in the preconditioned group (reproduced with permission from Ref. [26]).

induce VF. The right panel shows the rise in tissue impedance in the same experiment. Fig. 4 shows overall data. Thus, by providing the necessary triggers for VF, these experiments indicate that the substrate for VF is evolving during the 1B phase, and that the optimum substrate is present when uncoupling is moderate; (3) spontaneous VF occurred at increase of tissue impedance of <math><50\%</math> in a study by Smith et al. [25]; (4) a decrease followed by an increase in

junctional conductance prevented conduction of ectopic activity to the nonischemic tissue [73].

An alternative hypothesis includes the occurrence of microreentry due to very slow conduction induced by gap junctional uncoupling [21,47]. Although in cultured myocytes exposed to palmitoleic acid very slow conduction was observed [47], microreentry is unlikely to be the underlying mechanism of 1B VF for the following reasons: (1) the reduction of

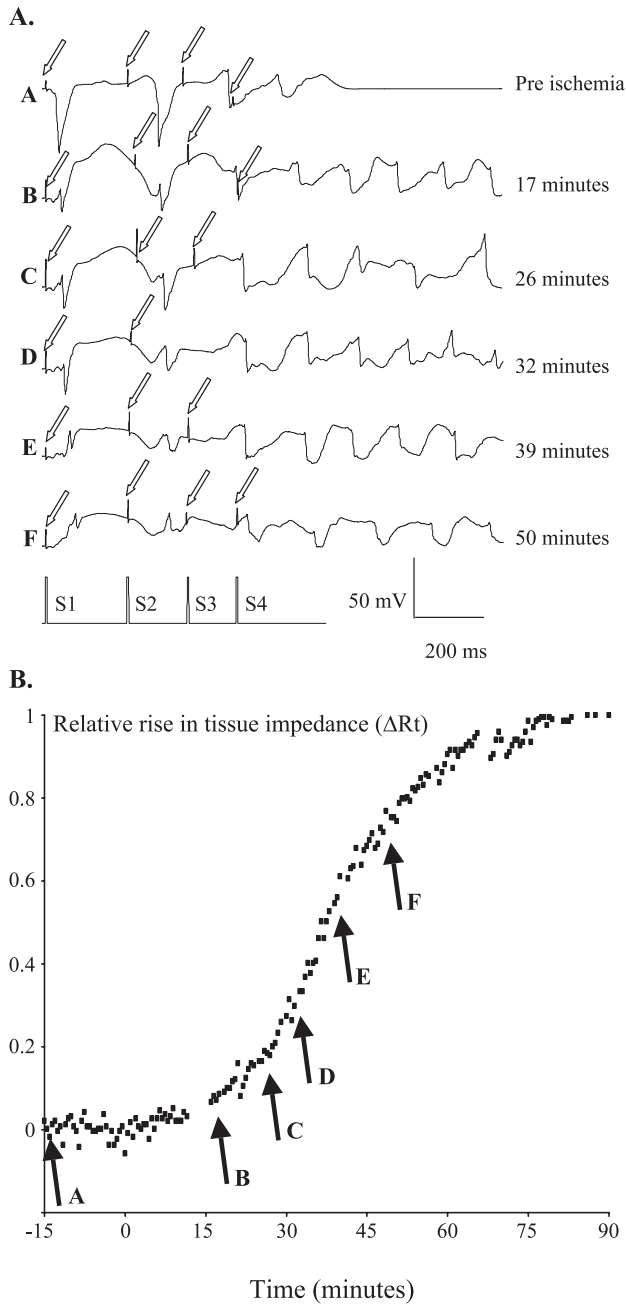


Fig. 3. Panel A: Unipolar electrograms of premature stimulation in a regionally ischemic pig heart. Arrows denote stimulation artifacts. Three PVC did not induce VF before ischemia. The number of PVC decreased from three to one at 32 min of ischemia, after which more stimuli are needed to induce VF. Panel B: Corresponding rise in tissue impedance in the same experiment as in panel A. Letters indicate when electrograms in panel A were recorded (reproduced with permission from Ref. [27]).

progressing gap junctional uncoupling [46]; (5) VF occurred or could be induced at a window of moderate uncoupling only [25,27,73]; (6) epicardial mapping of 1B ventricular arrhythmias showed reentrant circuits with wavelengths in the order of magnitude of centimeters [27] and VF could not be induced in isolated left ventricular preparations up to 9 g and macroreentry around slim lines of activation block was

observed [75]; and (7) Ruiz-Meana et al. [52] showed persistent dye coupling after ischemia-induced rigor, demonstrating open gap junctions.

### 2.3.2. Role of structural changes in gap junctional coupling

As a consequence of tissue anisotropy (more gap junctions along the fiber than perpendicular to fiber direction), conduction velocity and safety for conduction differ in longitudinal and transversal directions, and blockade of conduction in either transversal or longitudinal direction might have differential effects on arrhythmogenesis [46,76,77].

In Cx43 +/- mice, significantly more spontaneous and induced arrhythmias were observed during 1 h of regional ischemia [42]. Fig. 5 shows the incidence of arrhythmias in regionally ischemic wild-type versus Cx43 +/- mice. Both spontaneous and induced VT are more frequent in the Cx43 +/- animals. The hearts of these animals were morphologically normal and conduction velocity under normoxic circumstances was only marginally reduced; therefore, the increased arrhythmogenicity resulted from the interplay between acute ischemia and the genetic background of reduced gap junctional coupling [42]. Thomas et al. [40] showed in cultured Cx43 +/- cells that despite a 43% reduction in expressed level of Cx43 protein, no reduction of conduction velocity was observed.

The number of gap junctions decreases and lateralization occurs in failing hearts [78]. Therefore, load mismatches that cause either slow conduction, conduction block, or changed restitution of conduction velocity might arise. Derksen et al. [80] demonstrated that pathologic conduction curves, associated with high vulnerability of VF [79], occur in hearts with interstitial fibrosis and result from load mismatch. Interstitial fibrosis causes cellular uncoupling through insulation of myocardial fibers, and is associated with increased activation delay dependent on the type and amount [81]. Structural remodeling of gap junctions does not occur during the immediate phase of acute ischemia. It is conceivable, however, that arrhythmogenesis in the 1B phase of acute ischemia in hearts of patients with heart failure is more severe because of the preexisting morphological changes. Indeed, the progression of ischemia-induced changes is different in these hearts compared to normal hearts [82]. It has, moreover, been shown that gap junctional uncoupling decreases defibrillation success—a finding of particular importance for the growing population of patients with internal defibrillator [83].

## 2.4. Modulation of ischemia-induced arrhythmias

### 2.4.1. Preconditioning

Ischemic preconditioning delayed both gap junctional uncoupling and the occurrence of 1B VF in pigs [26] and was associated with decreased infarct size. Preconditioning may act through various pathways: (1) postponing the increase in  $[Ca^{2+}]_i$ ; and the closely associated rise in tissue

impedance [24]; (2) delayed gap junctional uncoupling in preconditioned hearts is likely related to diminished dephosphorylation and intracellular redistribution of Cx43 during prolonged ischemia [33,84]; (3) preconditioning is abol-

ished with glibenclamide, a blocker of the  $K_{ATP}$  channel [85]; (4) opening of the  $K_{ATP}$  channel postponed the second rise in  $[K^+]_o$  and the decreased catecholamine release from ischemic nerve endings in the isolated rabbit heart, but the decrease in ventricular arrhythmias did not reach statistical significance [86]. The role of closure of gap junctions as a preconditioning factor is unclear. Preconditioning preserved phosphorylation of Cx43, suggesting that the gap junctions remain opened [84,87]. Closure of gap junctions with heptanol in concentrations of 0.5 mM abolished the infarct size reduction by ischemic preconditioning [88], but at a concentration of 1 mM, heptanol was cardioprotective [89].

2.4.2. Reduction of wall stress

Arrhythmogenic triggers during the 1B phase are expected to decrease in occurrence when the wall tension on the ischemic border is lowered [70]. Indeed, unloading the heart with nitroprusside was effective in reducing sudden cardiac death [90].

2.4.3. Autonomic nervous system

Catecholamines, noradrenaline in particular, are released from the ischemic nerve endings with a time course similar to the onset of gap junctional uncoupling [86,91–93] and increase  $[Ca^{2+}]_i$  via a G-protein-dependent pathway. Gap junctional conductance increases upon catecholamine-induced increase in intracellular cAMP concentration [31,94]. Consequently, conduction velocity increased and  $dV/dt$  max did not change [94]. Parasympathetic stimulation has the opposite effect and decreases gap junctional coupling via a cGMP-dependent pathway [31]. Adrenergic blockade decreases ventricular arrhythmias, but in many studies, heart rate and blood pressure are altered as well [95,96] and the beneficiary effect appeared more prominent on phase-1A than on phase-1B arrhythmias [60,97]. Beta blockade during acute ischemia reduces mortality, partly because of reduction of ventricular rupture.

2.4.4. Other factors

Fatty acid metabolites accumulate in the proximity of intercalated disks and decrease gap junctional conductance, and may modulate arrhythmogenesis [30,98,99]. Preliminary studies with carbenoxolone, an ancient antiulcer drug that can be used as a specific blocker of gap junctions in cardiac muscles [100], showed that infusion in the ischemic zone just prior to coronary occlusion resulted in decreased transversal, but not longitudinal, conduction velocity com-

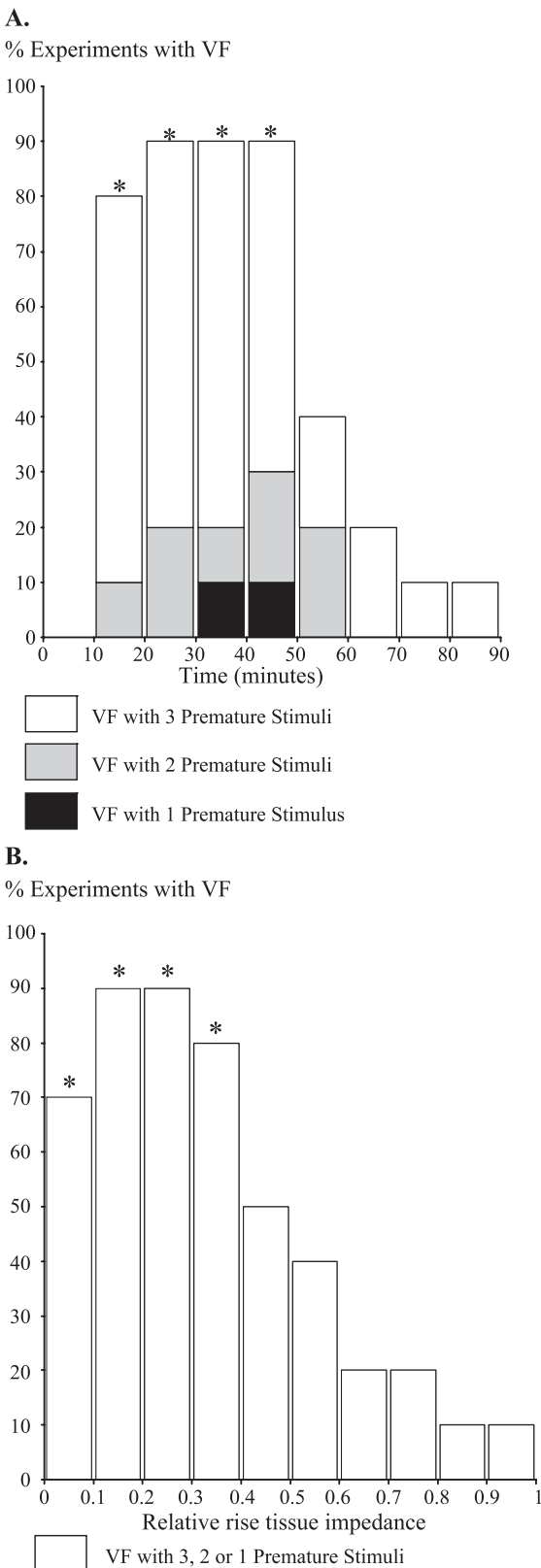


Fig. 4. Panel A: Percentage of VF inducibility in isolated pig hearts during 90 min of regional ischemia. VF is inducible with one (white bars), two (grey bars), or three premature stimuli (black bars) between 10 and 50 min of ischemia, after which the number of animals in which VF could be induced declined. Panel B: VF inducibility related to relative rise in tissue impedance. VF could be induced up to 40% of relative rise in tissue impedance; at higher degrees of uncoupling, inducibility declined (reproduced with permission from Ref. [27]).

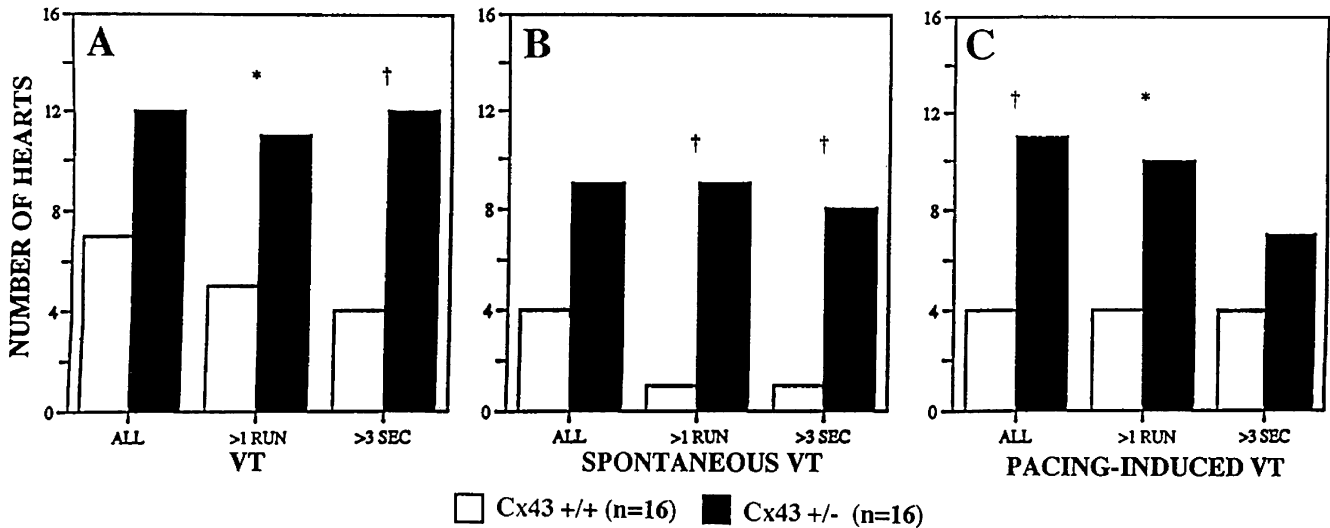


Fig. 5. Incidence of spontaneous and induced VT in wild-type (open bars) and Cx43 +/- mice (closed bars) subjected to regional ischemia. Panel A shows all occasions of VT, panel B and C show spontaneous and pacing induced VT respectively. More arrhythmias were observed in the Cx43 +/- group (reproduced with permission from Ref. [42]).

pared to untreated ischemic hearts, but no reduction in VF (J.R. de Groot, unpublished observation). An increased anisotropic ratio after partial gap junctional uncoupling has been reported previously [41,43,44].

It appears that commonly used antiarrhythmic drugs have little or no effect on gap junctional conductance [101].

### 3. Clinical relevance

#### 3.1. Lack of studies in humans

The natural time course of ischemia-induced arrhythmias as well as the presence of a 1B phase of arrhythmias and its potential contribution to mortality are unknown in man. It appeared that 30% of sudden cardiac death during the first 24 h of infarction occurred within the first 60 min [102]. This large percentage suggests that at least some of the victims succumb during the 1B phase of arrhythmias. Electrophysiological changes associated with the 1A phase in animals occur much more rapidly in patients undergoing thoracotomy subjected to regional ischemia of short duration [103]. It can be speculated therefore that because 1A changes occur more rapidly, the time window during which 1A arrhythmias can occur is restricted, and 1A arrhythmias occur less frequently in man than in the known animal models. This speculation might contrast with data of out-of-hospital cardiac resuscitation where sudden collapse without prior symptoms is frequently encountered [104]. The number of 1B events in the study by Waalewijn et al. might be underestimated by the patients who were able to seek help and experienced their cardiac arrest in the hospital. However, given the paucity of human data, these scenarios remain highly speculative.

#### 3.2. Can we affect the course of ischemia-induced arrhythmogenesis?

A moderate degree of gap junctional uncoupling is associated with ventricular arrhythmias [27], whereas more advanced uncoupling is antiarrhythmic. Pharmacologic uncoupling therefore may present as a novel target in antiarrhythmic therapy. Pharmacological uncoupling might be selective to the ischemic tissue by the preferential effect of uncoupling on already poorly coupled tissue: the same degree of uncoupling leads to a more pronounced decrease of conduction velocity in poorly coupled than in normally coupled tissue [36]. No clinical trials have, to our knowledge, tested the antiarrhythmic effect of uncoupling agents. Carbenoxolone, a saponin derivative, has been shown to selectively block gap junctions in cardiac tissue with no effect on the major transmembrane currents [100]. Its administration to isolated perfused healthy rabbit hearts did cause a small decrease in conduction velocity. In ischemic tissue, it does not decrease VF inducibility (J.R. de Groot, unpublished observation).

On the other hand, increased gap junctional coupling might be antiarrhythmic by maintaining conduction velocity during acute ischemia for a longer time. However, a large increase in gap junctional conductance could paradoxically cause conduction block through decreased safety for conduction because the current generated by a cell is insufficient to activate the many cells it is coupled to (current sink is too large). There are several agents that increase gap junctional coupling, including agents that increase intracellular cAMP [31,94]. Certain endogenous and synthetic peptides have been reported to decrease dispersion in refractoriness. Indeed, Dhein et al. [105] showed that AAP10 reduced dispersion of activation recovery intervals in a dose-dependent manner up to 10 nmol/l in regionally



ischemic rabbit hearts. It was shown that AAP10 increases gap junctional conductance [106] via a PKC-dependent mechanism [107] and prevents conduction slowing in hypoxic papillary muscles [108]. However, there was no significant reduction in VT or VF by AAP10 [105] nor by HP-5 in regionally ischemic rabbit hearts [109]. A recent study showed that another peptide, ZP123, an AAP analog with a longer plasma half-life compared to AAP10, increased gap junctional conductance with 69% [110]. ZP123 reversed conduction block and decreased the inducibility of reentrant monomorphic VT 1–4 h after coronary occlusion in open chest dogs [110]. The effects of ZP123 were restricted to ischemia, with no change in conduction velocity before ischemia. In summary, what these peptides have in common is that they reduce dispersion of action potential duration and maintain conduction velocity, but their use against acute ischemia-induced VF has not yet been shown convincingly.

### 3.3. Need of understanding VF in the internal cardiac defibrillator era

The absolute number of sudden deaths in the low-risk population is much larger than that in the high-risk population because the low-risk population is so much larger [10]. With this in mind, there is certainly a need for better understanding the mechanism of VF in normal hearts. The observation that ischemic and pharmacological preconditioning postpones both gap junctional uncoupling and the 1B VF is promising [26]. First, it creates a larger window during which the reversibly damaged myocardium can be salvaged. Second, there is more time to find medical assistance and to reach a hospital, ambulance, or defibrillator. However, the sequence of ischemia and reperfusion appears pivotal for ischemic preconditioning and, to the best of our knowledge, there are no drugs that precondition the heart once ischemia has already started.

From the hypothesis that arrhythmias are caused by a temporal electrotonic effect, mediated via residual gap junctional coupling, it follows that rapid uncoupling of the ischemic tissue would prove antiarrhythmic. This would narrow the arrhythmogenic time window. At this point, however, gap junctional uncoupling therapy is a pure theoretical speculation because there are no drugs that act preferentially within the ischemic zone without affecting the rest of the heart and the rest of the body. Moreover, experimental evidence that increased gap junctional uncoupling is antiarrhythmic is lacking. Once the unanswered questions about this issue are addressed, novel targets in medical therapy may contribute substantially to a decrease of sudden cardiac death in the population at large.

## 4. Conclusions

Gap junctions are essential for normal propagation of the activation impulse in the heart, and disruption of gap

junctional coupling results in discontinuous conduction and arrhythmias. Several factors that modulate gap junctional resistance and affect conduction velocity in vivo and in vitro have been identified.

Myocardial ischemia causes depletion of high-energy phosphates, decreased  $pH_i$ , and rise of  $[Ca^{2+}]_i$ . These factors are associated with dephosphorylation of gap junction proteins and the rise of tissue impedance, a hallmark of intercellular uncoupling. Gap junctional uncoupling causes conduction slowing and therefore provides circumstances leading to reentry, but its exact mechanisms during ischemia are unknown. VF occurs at the start of rise of tissue impedance [25].

Very slow conduction caused by gap junctional uncoupling, as has been demonstrated in vitro [34,46,47] and in silico [36,37], has never been shown in intact hearts, making microreentry an unlikely mechanism. Cx43 +/- mice have slower conduction velocities under normal circumstances and increased arrhythmogenesis during ischemia [42].

The concept of residual coupling between ischemic and nonischemic tissue, whereby the ischemic inexcitable cells electrotonically depress the intrinsically viable tissue, may form an arrhythmogenic mechanism for gap junction uncoupling related to arrhythmias [72]. Indeed, VF inducibility was restricted to the first 40% of tissue impedance rise [27] and gap junctional coupling remains intact longer than excitability [54]. These experimental findings are supported by simulations that provide a mechanism explaining why arrhythmias only occur at modest but not complete uncoupling [73]. However, the evidence for temporary electrotonic interaction between viable and irreversibly damaged myocardium remains circumstantial, and the change in gap junctional conductance between the ischemic subepicardium and the midmyocardium is unknown.

Thus, although many studies have improved our understanding of the role of gap junctional uncoupling in the occurrence of arrhythmias in the regionally ischemic heart, the picture is yet far from complete. The main question that needs to be answered is: Can we locally and specifically modulate gap junctional coupling, either by reducing or by increasing it, and thereby suppress or even prevent lethal arrhythmias during the 1B phase of myocardial ischemia?

## Acknowledgements

This work was supported by the Netherlands Heart Foundation grant 2000T020 and EU projects IST-1999-13047 (MicroTrans) and Esprit IV 33485 (MicroCard).

## References

- [1] Zipes DP, Wellens HJJ. Sudden cardiac death. *Circulation* 1998;98: 2334–51.
- [2] Lenfant C. Task force on research in epidemiology and prevention of cardiovascular diseases. *Circulation* 1994;90:2609–17.

- [3] Zheng Z-J, Croft JB, Giles WR, Mensah GA. Sudden cardiac death in the United States, 1989 to 1998. *Circulation* 2001;104:2158–63.
- [4] Janse MJ, Wit AL. Electrophysiological mechanisms of ventricular arrhythmias resulting from myocardial ischemia and infarction. *Physiol Rev* 1989;69:1049–169.
- [5] The Cardiac Arrhythmia Suppression Trial II Investigators. Effect of the antiarrhythmic agent moricizine on survival after myocardial infarction. *N Engl J Med* 1992;327:227–33.
- [6] The Cardiac Arrhythmia Suppression Trial (CAST) Investigators. Preliminary report: effect of encainide and flecainide on mortality in a randomized trial of arrhythmia suppression after myocardial infarction. *N Engl J Med* 1989;321:406–12.
- [7] Waldo AL, Camm AJ, deRuyter H, et al. On behalf of the SWORD investigators. Effect of D-sotalol on mortality in patients with left ventricular dysfunction after recent and remote myocardial infarction. *Lancet* 1996;348:7–12.
- [8] Moss AJ, Zareba W, Hall WJ, et al. For the Multicenter Automatic Defibrillator Implantation Trial II Investigators. Prophylactic implantation of a defibrillator in patients with myocardial infarction and reduced ejection fraction. *N Engl J Med* 2002;346:877–83.
- [9] The Antiarrhythmic Versus Implantable Defibrillators (AVID) Investigators. A comparison of antiarrhythmic-drug therapy with implantable defibrillators in patients resuscitated from near-fatal ventricular arrhythmias. *N Engl J Med* 1997;337:1576–83.
- [10] Huikuri HV, Castellanos A, Myerburg RJ. Sudden death due to cardiac arrhythmias. *N Engl J Med* 2001;345:1473–82.
- [11] Mines GR. On circulating excitations in heart muscle and their possible relation to tachycardia and fibrillation. *Trans R Soc Can* 1914; IV:43–52.
- [12] Han J, Moe GK. Nonuniform recovery of excitability in ventricular muscle. *Circ Res* 1964;14:44–60.
- [13] Winfree AT. Electrical turbulence in three-dimensional heart muscle. *Science* 1994;266:1003–6.
- [14] Chen J, Mandapati R, Berenfeld O, Skanes AC, Gray RA, Jalife J. Dynamics of wavelets and their role in atrial fibrillation in the isolated sheep heart. *Cardiovasc Res* 2000;48:220–32.
- [15] Jalife J. Ventricular fibrillation: mechanisms of initiation and maintenance. *Annu Rev Physiol* 2000;62:25–50.
- [16] Allesie MA, Bonke FIM, Schopman FJG. Circus movement in rabbit atrial muscle as a mechanism of tachycardia: III. The “Leading Circle” concept: a new model of circus movement in cardiac tissue without the involvement of an anatomical obstacle. *Circ Res* 1977; 41:9–18.
- [17] Coumel P. The management of clinical arrhythmias. An overview on invasive versus non-invasive electrophysiology. *Eur Heart J* 1987;8: 92–9.
- [18] Engelmann TW. Ueber die Leitung der Erregung im Herzmuskel. *Pflügers Arch* 1875;11:465–80.
- [19] Burden-Sanderson J, Page FJM. On the time-relations of the excitatory process in the ventricle of the heart of the frog. *J Physiol* 1879;2: 384–429.
- [20] De Mello WC, Motta G, Chapeau M. A study on the healing-over of myocardial cells of toads. *Circ Res* 1969;24:475–87.
- [21] Kléber AG, Riegger CB, Janse MJ. Electrical uncoupling and increase of extracellular resistance after induction of ischemia in isolated, arterially perfused rabbit papillary muscle. *Circ Res* 1987;61: 271–9.
- [22] Fleischhauer J, Lehmann L, Kléber AG. Electrical resistances of interstitial microvascular space as determinants of the extracellular electrical field and velocity of propagation in ventricular myocardium. *Circulation* 1995;92:587–94.
- [23] De Mello WC. Effect of intracellular injection of calcium and strontium on cell communication in the heart. *J Physiol* 1975; 250:231–45.
- [24] Dekker LRC, Fiolet JWT, VanBavel E, et al. Intracellular Ca<sup>2+</sup>, intercellular electrical coupling, and mechanical activity in ischemic rabbit papillary muscle. Effects of preconditioning and metabolic blockade. *Circ Res* 1996;79:237–46.
- [25] Smith WT, Fleet WF, Johnson TA, Engle CL, Cascio WE. The 1b phase of ventricular arrhythmias in ischemic in situ porcine heart is related to changes in cell-to-cell electrical coupling. *Circulation* 1995; 92:3051–60.
- [26] Cinca J, Warren M, Carreño A, et al. Changes in myocardial electrical impedance induced by coronary artery occlusion in pigs with and without preconditioning. Correlation with local ST segment potential and ventricular arrhythmias. *Circulation* 1997;96:3079–86.
- [27] De Groot JR, Wilms-Schopman FJG, Opthof T, Remme CA, Coronel R. Late ventricular arrhythmias during acute regional ischemia in the isolated blood perfused pig heart. Role of electrical cellular coupling. *Cardiovasc Res* 2001;50:362–72.
- [28] Yan G-X, Kléber AG. Changes in extracellular and intracellular pH in ischemic rabbit papillary muscle. *Circ Res* 1992;71:460–70.
- [29] Kléber AG. The potential role of Ca<sup>2+</sup> for electrical cell-to-cell uncoupling and conduction block in myocardial tissue. *Basic Res Cardiol* 1992;87:131–43.
- [30] Wu J, McHowat J, Saffitz J, Yamada KA, Corr PB. Inhibition of gap junctional conductance by long-chain acylcarnitines and their preferential accumulation in junctional sarcolemma during hypoxia. *Circ Res* 1993;72:879–89.
- [31] Burt JM, Spray DC. Inotropic agents modulate gap junctional conductance between cardiac myocytes. *Am J Physiol* 1988;254: H1206–10.
- [32] Huang X-D, Sandusky GE, Zipes DP. Heterogeneous loss of connexin43 protein in ischemic dog hearts. *J Cardiovasc Electrophysiol* 1999;10:79–91.
- [33] Beardslee MA, Lerner DL, Tadros PN, et al. Dephosphorylation and intracellular redistribution of ventricular connexin43 during electrical uncoupling induced by ischemia. *Circ Res* 2000;87:656–62.
- [34] Weingart R, Maurer P. Action potential transfer in cell pairs isolated from adult rat and guinea pig ventricles. *Circ Res* 1988;63:72–80.
- [35] Rudy Y, Quan WL. A model study of the effects of the discrete cellular structure on electrical propagation in cardiac tissue. *Circ Res* 1987;61:815–23.
- [36] Jongsma HJ, Wilders R. Gap junctions in cardiovascular disease. *Circ Res* 2000;86:1193–97.
- [37] Shaw RM, Rudy Y. Ionic mechanisms of propagation in cardiac tissue. Roles of sodium and L-type calcium currents during reduced excitability and decreased gap junction coupling. *Circ Res* 1997;81: 727–41.
- [38] Gutstein DE, Morley GE, Tamadonn H, et al. Conduction slowing and sudden arrhythmic death in mice with cardiac-restricted inactivation of connexin43. *Circ Res* 2001;88:333–9.
- [39] Morley GE, Vaidya D, Samie FH, Lo C, Delmar M, Jalife J. Characterization of conduction in the ventricles of normal and heterozygous Cx43 knockout mice using optical mapping. *J Cardiovasc Electrophysiol* 1999;10:1361–75.
- [40] Thomas SP, Kucera JP, Bircher-Lehmann L, Rudy Y, Saffitz JE, Kléber AG. Impulse propagation in synthetic strands of neonatal cardiac myocytes with genetically reduced levels of connexin43. *Circ Res* 2003;92:1209–16.
- [41] Eloff BC, Lerner DL, Yamada KA, Schuessler RB, Saffitz JE, Rosenbaum DS. High resolution optical mapping reveals conduction slowing in connexin43 deficient mice. *Cardiovasc Res* 2001;51:681–90.
- [42] Lerner DL, Yamada KA, Schuessler RB, Saffitz JE. Accelerated onset and increased incidence of ventricular arrhythmias induced by ischemia in Cx43-deficient mice. *Circulation* 2000;101:547–52.
- [43] Delmar M, Michaels DC, Johnson T, Jalife J. Effects of increasing intercellular resistance on transverse and longitudinal propagation in sheep epicardial muscle. *Circ Res* 1987;60:780–5.
- [44] Dhein S, Krüsemann K, Schaefer T. Effects of the gap junction uncoupler palmitoleic acid on the activation and repolarization wavefronts in isolated rabbit hearts. *Br J Pharmacol* 1999;128:1375–84.
- [45] Jalife J, Sicouri S, Delmar M, Michaels DC. Electrical uncoupling

- and impulse propagation in isolated sheep Purkinje fibers. *Am J Physiol* 1989;257:H179–89.
- [46] Rohr S, Kucera JP, Fast VG, Kléber AG. Paradoxical improvement of impulse conduction in cardiac tissue by partial cellular uncoupling. *Science* 1997;275:841–4.
- [47] Rohr S, Kucera JP, Kléber AG. Slow conduction in cardiac tissue: I. Effects of a reduction of excitability versus a reduction of electrical coupling on microconduction. *Circ Res* 1998;83:781–94.
- [48] Vogel R, Weingart R. Mathematical model of vertebrate gap junctions derived from electrical measurements on homotypic and heterotypic channels. *J Physiol* 1998;510:177–89.
- [49] Henriquez AP, Vogel R, Muller-Borer BJ, Henriquez CS, Weingart R, Cascio WE. Influence of dynamic gap junction resistance on impulse propagation in ventricular myocardium: a computer simulation study. *Biophys J* 2001;81:2112–21.
- [50] Kumar NM, Gilula N.B. The gap junction communication channel. *Cell* 1996;84:381–88.
- [51] Lesh MD, Pring M, Spear JF. Cellular uncoupling can unmask dispersion of action potential duration in ventricular myocardium. A computer model study. *Circ Res* 1989;65:1426–40.
- [52] Ruiz-Meana M, Garcia-Dorado D, Hofstaetter B, Piper HM, Soler-Soler J. Propagation of cardiomyocyte hypercontracture by passage of Na<sup>+</sup> through gap junctions. *Circ Res* 1999;85:280–7.
- [53] Mendez C, Mueller WJ, Meridith J, Moe GK. Interaction of transmembrane potentials in canine Purkinje fibers and at Purkinje fiber–muscle junctions. *Circ Res* 1969;24:361–72.
- [54] De Groot JR, Schumacher CA, Verkerk AO, Baartscheer A, Fiolet JWT, Coronel R. Intrinsic heterogeneity in repolarization is increased in isolated failing rabbit cardiomyocytes during simulated ischemia. *Cardiovasc Res* 2003;59:705–14.
- [55] Ruiz-Meana M, Garcia-Dorado D, Lane S, et al. Persistence of gap junction communication during myocardial ischemia. *Am J Physiol* 2001;280:H2563–71.
- [56] Janse MJ, Cinca J, Morena H, et al. The “Border Zone” in myocardial ischemia. An electrophysiological, metabolic and histochemical correlation in the pig heart. *Circ Res* 1979;44:576–88.
- [57] Wilensky RL, Trantum-Jensen J, Coronel R, Wilde AAM, Fiolet JWT, Janse MJ. The subendocardial border zone during acute ischemia of the rabbit heart: an electrophysiologic, metabolic, and morphologic correlative study. *Circulation* 1986;74:1137–46.
- [58] Fujiwara H, Ashraf M, Sato S, Millard RW. Transmural cellular damage and blood flow distribution in early ischemia in pig hearts. *Circ Res* 1982;51:683–93.
- [59] Kaplinsky E, Ogawa S, Balke W, Dreifus LS. Two periods of early ventricular arrhythmia in the canine acute myocardial infarction model. *Circulation* 1979;60:397–403.
- [60] Menken U, Wiegand V, Bucher P, Meesman W. Prophylaxis of ventricular fibrillation after acute experimental coronary occlusion by chronic beta-adrenoceptor blockade with atenolol. *Cardiovasc Res* 1979;13:588–94.
- [61] Russell DC, Lawrie JS, Riemersma RA, Oliver MF. Mechanisms of phase 1a and 1b early ventricular arrhythmias during acute myocardial ischemia in the dog. *Am J Cardiol* 1984;53:307–12.
- [62] Euler DE, Spear JF, Moore EN. Effect of coronary occlusion on arrhythmias and conduction in the ovine heart. *Am J Physiol* 1983;245:H82–9.
- [63] Parratt JR. Inhibitors of the slow calcium current and early ventricular arrhythmias. In: Parratt JR, editor. *Early Arrhythmias Resulting from Myocardial Ischemia. Mechanism and Prevention by Drugs*. London: Macmillan; 1982. p. 329–46.
- [64] Penkoske PA, Sobel BE, Corr PB. Disparate electrophysiological alterations accompanying dysrhythmia due to coronary occlusion and reperfusion in the cat. *Circulation* 1978;58:1023–35.
- [65] Bril A, Forest M-C, Gout B. Ischemia and reperfusion induced arrhythmias in rabbits with chronic heart failure. *Am J Physiol* 1991;261:H301–7.
- [66] Curtis MJ. Characterisation, utilisation and clinical relevance of isolated perfused heart models of ischemia-induced ventricular fibrillation. *Cardiovasc Res* 1998;39:194–215.
- [67] Janse MJ, Van Capelle FJL, Morsink H, et al. Flow of “injury” current and patterns of excitation during early ventricular arrhythmias in acute regional myocardial ischemia in isolated porcine and canine hearts. Evidence for two different arrhythmogenic mechanisms. *Circ Res* 1980;47:151–65.
- [68] Coronel R, Wilms-Schopman FJG, Opthof T, Van Capelle FJL, Janse MJ. Injury current and gradients of diastolic stimulation threshold, TQ potential, and extracellular potassium concentration during acute regional ischemia in the isolated perfused pig heart. *Circ Res* 1991;68:1241–9.
- [69] Pogwizd SM, Corr PB. Reentrant and nonreentrant mechanisms contribute to arrhythmogenesis during early myocardial ischemia: results using three-dimensional mapping. *Circ Res* 1987;61:352–71.
- [70] Coronel R, Wilms-Schopman FJG, De Groot JR. Origin of ischemia-induced phase 1B ventricular arrhythmias in pig hearts. *J Am Coll Cardiol* 2002;39:166–76.
- [71] Barrabes JA, Garcia-Dorado D, Agulló L, et al. Blockade of stretch-activated channels with intracoronary Gd<sup>3+</sup> does not reduce the incidence of phase 1b ventricular arrhythmias during acute coronary occlusion in swine. *Eur Heart J* 2003;24:510 [Abstract].
- [72] Tan RC, Joyner RW. Electrotonic influences on action potentials from isolated ventricular cells. *Circ Res* 1990;67:1071–81.
- [73] Pollard AE, Cascio WE, Fast VG, Knisley SB. Modulation of triggered activity by uncoupling in the ischemic border. A model study with phase 1b-like conditions. *Cardiovasc Res* 2002;56:381–92.
- [74] Wilders R, Wagner MB, Golod DA, et al. Effects of anisotropy on the development of cardiac arrhythmias associated with focal activity. *Pflugers Arch Eur J Physiol* 2000;441:301–12.
- [75] De Groot JR, Wilms-Schopman FJG, Janse MJ, Coronel R. Reentry around slim lines of functional activation block constitutes the mechanisms of 1B ventricular arrhythmias. *PACE* 2000;23:585 [Abstract].
- [76] Spach MS, Miller III WT, Geselowitz DB, Barr RC, Kootsey JM, Johnson EA. The discontinuous nature of propagation in normal canine cardiac muscle. Evidence for recurrent discontinuities of intracellular resistance that affect membrane currents. *Circ Res* 1981;48:39–54.
- [77] Spach MS, Miller III WT, Dolber PC, Kootsey JM, Sommer JR, Mosher CE. The functional role of structural complexities in the propagation of depolarization in the atrium of the dog. Cardiac conduction disturbances due to discontinuities of effective axial resistivity. *Circ Res* 1982;50:175–91.
- [78] Peters NS, Green CR, Poole-Wilson PA, Severs NJ. Reduced content of connexin43 gap junctions in ventricular myocardium from hypertrophied and ischemic human hearts. *Circulation* 1993;88:864–75.
- [79] Saumarez RC, Camm AJ, Panagos A, et al. Ventricular fibrillation in hypertrophic cardiomyopathy is associated with increased fractionation of paced right ventricular electrograms. *Circulation* 1992;86:467–74.
- [80] Derksen R, Van Rijen HVM, Wilders R, et al. Tissue discontinuities affect conduction velocity restitution. A mechanism by which structural barriers may promote wave break. *Circulation* 2003;108:882–8.
- [81] Kawara T, Derksen R, De Groot JR, et al. Activation delay after premature stimulation in chronically diseased human myocardium relates to the architecture of interstitial fibrosis. *Circulation* 2001;104:3069–75.
- [82] Dekker LRC, Rademaker H, Vermeulen JT, et al. Cellular uncoupling during ischemia in hypertrophied and failing rabbit ventricular myocardium. Effects of preconditioning. *Circulation* 1998;97:1724–30.
- [83] Sims JJ, Schoff KL, Loeb JM, Wiegert NA. Regional gap junction inhibition increases defibrillation thresholds. *Am J Physiol* 2003;285:H10–6.
- [84] Jain SK, Schuessler RB, Saffitz JE. Mechanisms of delayed electrical uncoupling induced by ischemic preconditioning. *Circ Res* 2003;92:1138–44.
- [85] Duncker DJ, Verdouw PD. Role of K<sub>ATP</sub> channels in ischemic pre-

- conditioning and cardioprotection. *Cardiovasc Drug Ther* 2000; 14:7–16.
- [86] Remme CA, Schumacher CA, De Jong JWJ, et al.  $K_{ATP}$  channel opening during ischemia: effects on myocardial noradrenaline release and ventricular arrhythmias. *J Cardiovasc Pharmacol* 2001;38: 406–16.
- [87] Schulz R, Gres P, Skyschally A, et al. Ischemic preconditioning preserves connexin43 phosphorylation during sustained ischemia in pig hearts in vivo. *FASEB J* 2003;17:1355–7.
- [88] Li G, Whittaker P, Yao M, Kloner RA, Przyklenk K. The gap junction uncoupler heptanol abrogates infarct size reduction with preconditioning in mouse hearts. *Cardiovasc Pathol* 2002;11:158–65.
- [89] Saltman AE, Aksehirli TO, Valiunas V, et al. Gap junction uncoupling protects the heart against ischemia. *J Thorac Cardiovasc Surg* 2002;124:371–6.
- [90] Durrer JD, Lie KI, Van Capelle FJL, Durrer D. Effect of sodium nitroprusside on mortality in acute myocardial infarction. *N Engl J Med* 1982;306:1121–8.
- [91] Schömig A, Dart AM, Dietz R, Mayer E, Kübler W. Release of endogenous catecholamines in the ischemic myocardium of the rat: Part A. Locally mediated release. *Circ Res* 1984;55:689–701.
- [92] Wilde AAM, Peters RJG, Janse MJ. Catecholamine release and potassium accumulation in the isolated globally ischemic rabbit heart. *J Mol Cell Cardiol* 1988;20:887–96.
- [93] Lameris TW, De Zeeuw S, Alberts G, et al. Time course and mechanism of myocardial catecholamine release during transient ischemia in vivo. *Circulation* 2000;101:2645–50.
- [94] Darrow BJ, Fast VG, Kléber AG, Beyer EC, Saffitz J. Functional and structural assessment of intercellular communication. Increased conduction velocity and enhanced connexin expression in dibuteryl cAMP-treated cultured cardiac myocytes. *Circ Res* 1996;79:174–83.
- [95] Bergey JL, Wendt RL, Nocella K, McCallum JD. Acute coronary artery occlusion–reperfusion arrhythmias in pigs: antiarrhythmic and antifibrillatory evaluation of verapamil, nifedipine, prenylamine and propranolol. *Eur J Pharmacol* 1984;97:95–103.
- [96] Kjekshus JK. Importance of heart rate in determining beta-blocker efficacy in acute and long-term acute myocardial infarction intervention trials. *Am J Cardiol* 1986;57:43F–9F.
- [97] Puddu PE, Jouve R, Langlet F, et al. Prevention of postischemic ventricular fibrillation by long term beta adrenoceptor blockade with acebutolol in the anaesthetized dog. *Cardiovasc Res* 1986;20:721–6.
- [98] DaTorre SD, Creer MH, Pogwizd SM, Corr PB. Amphipathic lipid metabolites and their relation to arrhythmogenesis in the ischemic heart. *J Mol Cell Cardiol* 1991;23:11–22.
- [99] Yamada KA, McHowat J, Yan G-X, et al. Cellular uncoupling induced by accumulation of long-chain acylcarnitine during ischemia. *Circ Res* 1994;74:83–95.
- [100] De Groot JR, Veenstra T, Verkerk AO, et al. Conduction slowing by the specific gap junctional uncoupler carbenoxolone. *Cardiovasc Res* 2003;60:288–97.
- [101] Daleau P. Effects of antiarrhythmic agents on junctional resistance of guinea pig ventricular cell pairs. *J Pharmacol Exp Ther* 1998;284: 1174–9.
- [102] Löwel H, Lewis M, Hörmann A. Prognostische bedeutung der Prähospitalphase beim akuten myokardinfarkt. Ergebnisse des Augsburger Herzinfarktregisters 1985–1988. *Deutsch Med Wochenschr* 1991;116:729–33.
- [103] Taggart P, Sutton PMI, Opthof T, et al. Inhomogeneous transmural conduction during early ischaemia in patients with coronary artery disease. *J Mol Cell Cardiol* 2000;32:621–30.
- [104] Waalewijn RA, De Vos R, Koster RW. Out-of-hospital cardiac arrests in Amsterdam and its surrounding areas: results from the Amsterdam resuscitation study (ARREST) in Utstein style. *Resuscitation* 1998;38:157–67.
- [105] Dhein S, Manicone N, Müller A, et al. A new synthetic antiarrhythmic peptide reduces dispersion of epicardial activation recovery intervals and diminishes alterations of epicardial activation patterns induced by regional ischemia. *Naunyn-Schmiedeberg's Arch Pharmacol* 1994;350:174–84.
- [106] Müller A, Gottwald M, Tudyka T, Linke W, Klaus W, Dhein S. Increase in gap junction conductance by an antiarrhythmic peptide. *Eur J Pharmacol* 1997;327:65–72.
- [107] Weng S, Lauen M, Schaefer T, Polontchouk L, Grover R, Dhein S. Pharmacological modification of gap junction coupling by an antiarrhythmic peptide via protein kinase C activation. *FASEB J* 2002;16:1114–6.
- [108] Müller A, Schaefer T, Linke W, et al. Actions of the antiarrhythmic peptide AAP10 on intercellular coupling. *Naunyn-Schmiedeberg's Arch Pharmacol* 1997;356:76–82.
- [109] Kjolbye AL, Holstein-Rathlou NH, Petersen JS. Anti-arrhythmic peptide *N*-3-(4-hydroxyphenyl)propionyl Pro-Hyp-Gly-Ala-Gly-OH reduces dispersion of action potential duration during ischemia/reperfusion in rabbit hearts. *J Cardiovasc Pharmacol* 2002;40: 770–9.
- [110] Xing D, Kjolbye AL, Nielsen MS, et al. ZP123 increases gap junctional conductance and prevents reentrant ventricular tachycardia during myocardial ischemia in open chest dogs. *J Cardiovasc Electrophysiol* 2003;14:510–20.