



Adaptive and Innate Immunity in Psoriasis and Other Inflammatory Disorders

Michael P. Schön*

Department of Dermatology, Venereology and Allergology, University Medical Center Göttingen, Göttingen, Germany

Over the past three decades, a considerable body of evidence has highlighted T cells as pivotal culprits in the pathogenesis of psoriasis. This includes the association of psoriasis with certain MHC (HLA) alleles, oligoclonal expansion of T cells in some cases, therapeutic response to T cell-directed immunomodulation, the onset of psoriasis following bone marrow transplantation, or induction of psoriasis-like inflammation by T cells in experimental animals. There is accumulating clinical and experimental evidence suggesting that both autoimmune and autoinflammatory mechanisms lie at the core of the disease. Indeed, some studies suggested antigenic functions of structural proteins, and complexes of self-DNA with cathelicidin (LL37) or melanocytic ADAMTSL5 have been proposed more recently as actual auto-antigens in some cases of psoriasis. These findings are accompanied by various immunoregulatory mechanisms, which we increasingly understand and which connect innate and adaptive immunity. Specific adaptive autoimmune responses, together with our current view of psoriasis as a systemic inflammatory disorder, raise the question of whether psoriasis may have connections to autoimmune or autoinflammatory disorders elsewhere in the body. While such associations have been suspected for many years, compelling mechanistic evidence in support of this notion is still scant. This review sets into context the current knowledge about innate and adaptive immunological processes in psoriasis and other autoimmune or autoinflammatory diseases.

Keywords: psoriasis, adaptive immunity, innate immunity, autoimmune disease, skin-immunology

SETTING THE STAGE: PSORIASIS AS AN IMMUNE-MEDIATED DISORDER

If I was to name diseases that in recent years have increased our understanding of both adaptive and innate immune mechanisms on the one hand and have contributed decisively to the development of modern biological therapies on the other, then psoriasis would certainly occupy one of the top ranks. Psoriasis is currently viewed as a systemic chronic inflammatory disease with an immunogenetic basis that can be triggered extrinsically or intrinsically (1, 2). Research into its pathophysiology has led to impressive therapeutic improvements (3, 4). The disease is based on close interactions between components of the adaptive and the innate branches of the immune system (3, 5-9) (**Figure 1**). Since it was shown in the late 1970s that psoriasis can be ameliorated by cyclosporin A (10), it can no longer be seriously denied that T lymphocytes play a central role in the pathogenesis of this disease. This view is substantiated by numerous subsequent observations

OPEN ACCESS

Edited by:

Kutty Selva Nandakumar, Southern Medical University, China

Reviewed by:

Marcus Schmitt-Egenolf, Umeå University, Sweden Antonio Costanzo, Sapienza University of Rome, Italy Ia Khmaladze, Oriflame Cosmetics AB, Sweden

*Correspondence:

Michael P. Schön michael.schoen@ med.uni-goettingen.de orcid.org/0000-0002-0698-4130

Specialty section:

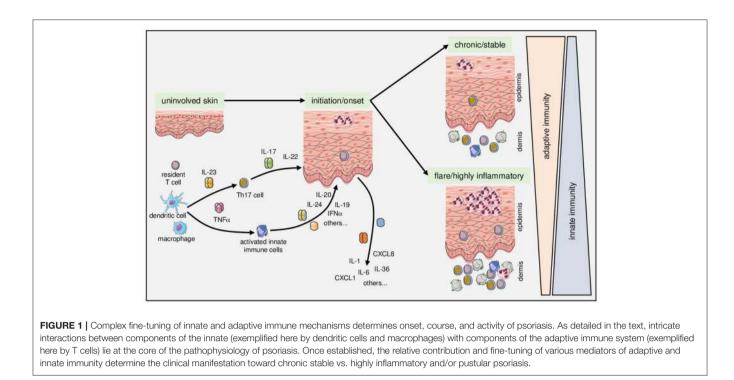
This article was submitted to Autoimmune and Autoinflammatory Disorders, a section of the journal Frontiers in Immunology

> **Received:** 31 May 2019 **Accepted:** 11 July 2019 **Published:** 26 July 2019

Citation:

Schön MP (2019) Adaptive and Innate Immunity in Psoriasis and Other Inflammatory Disorders. Front. Immunol. 10:1764. doi: 10.3389/fimmu.2019.01764

1



over the past four decades: psoriasis can be precipitated by bone marrow transplantation (11) and, similar to other autoinflammatory diseases, the disease is frequently associated with certain HLA expression patterns (7, 12-14). Drugs that specifically inhibit the function of T lymphocytes (such as CD2 blockade in the early days of biologics) can improve psoriasis (15). A therapeutic effect can also be achieved by interleukin (IL)-4, which pushes the cytokine milieu toward a Thelper (Th) cell 2-dominated immune response (16), probably through attenuation of Th17 function following diminished IL-23 production in antigen-presenting cells (17) and through induction of the transcription factor GATA3 (18, 19). IL-10 can also ameliorate psoriatic symptoms by modulating T cell functions (20). In addition, psoriasis-like skin inflammation in animal models can be initiated by certain CD4 + T cells (21–24), and T cells can induce psoriatic lesions in human skin xenografts (25, 26). Finally, the more recent discoveries that complexes of the antimicrobial peptide LL37 (a 37 amino acid C-terminal cleavage product of the antimicrobial peptide, cathelicidin) with own DNA or the melanocytic antigen ADAMTSL5 may function as autoantigens (27, 28), support the central role of T cells in the pathogenesis of psoriasis (29, 30).

AUTOIMMUNE PROCESSES IN PSORIASIS

The Plot Thickens: Actual Auto-Antigens in Psoriasis

Pathogenic T cells in psoriatic skin lesions facilitate hyperproliferation of keratinocytes, influx of neutrophilic granulocytes, as well as production of other inflammatory cytokines, chemokines and antimicrobial peptides. They feature

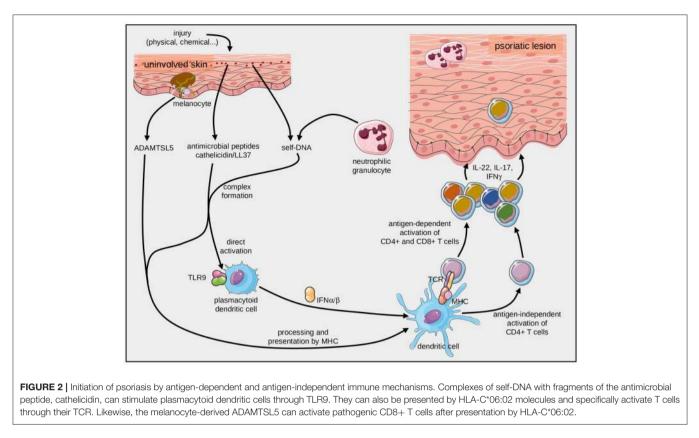
a Th17 signature, i.e., they express IL-17A, IL-22, and IFN- γ (3, 31, 32) (**Figure 2**). Dendritic cells maintain activation and differentiation of lesional Th17 cells primarily through secretion of IL-23 [reviewed in (8)].

In general, both HLA restriction and peptide specificity of a given T cell are determined by its T cell receptor (TCR) repertoire (33). Activation and clonal expansion of T cells occur upon antigenic stimulation. In the absence of foreign antigens, clonal T cell expansion is highly suggestive for autoimmunity in inflammatory diseases (34). Indeed, oligoclonal T cell expansion has been identified in psoriatic lesions in early well-designed studies (35–40) as well as in more recent investigations (41). It has been interpreted as an indicator for antigen-specific immune responses.

In psoriasis, oligoclonality of cutaneous T-cell populations is usually confined to lesional skin. This suggests that psoriasis is driven by locally presented antigens (35, 42–46). Likewise, the clonal TCRs arguably mark T cells which mediate the disease process. Several landmark publications during the past years lent support to this notion through identification of putative autoantigens in psoriasis.

Early concepts of autoimmune processes in psoriasis stemmed from the recognition of sequence homologies between keratinocyte structural proteins and streptococcal antigens (*S. pyogenes* in particular) (47–51). Mechanistic proof, however, is still lacking.

It was previously known that complexes of LL37 and self-DNA can activate dermal plasmacytoid dendritic cells (pDC) through toll-like receptor (TLR) signaling (52–54). These stimulate pDC then facilitate the psoriatic inflammatory cascade (52, 53, 55),



a mechanism that is alluded to in more detail below. The activation *via* innate immune mechanisms was extended later by the finding that complexes of self-DNA and LL37 can also induce adaptive antigen-specific immune responses. Indeed, LL37 can trigger profound TCR and MHC (*HLA-C*06:02*)-dependent T-cell responses (28). It remains to be confirmed, however, that the LL37-related candidate peptides can be derived from the parent protein by antigen processing within the antigen presenting cell and then be presented by HLA-class I-molecules.

A more recent strategy to identify potential targets of pathogenic T-cells in psoriasis was based on the generation of T-cell hybridomas expressing the paired Va3S1/VB13S1 TCR of clonal CD8⁺ psoriatic T cells of an HLA-C*06:02expressing psoriasis patient (27). This elegant approach identified melanocytes as target cells of the psoriatic immune response (27). A peptide derived from ADAMTS-like protein 5 (ADAMTSL5) by proteasomal cleavage and post-cleavage trimming induced the specific immune response. The auto-antigenic function of melanocytic ADAMTSL5 was then confirmed by mutation and knock-down experiments. Moreover, peripheral lymphocytes of the majority of psoriasis patients but not individuals without psoriasis responded to ADAMTSL5 with production of IL-17 or IFNy (27) (Figure 2). In contrast to LL37, which has been shown to activate both CD8⁺ cytotoxic T cells and CD4⁺ T helper cells, ADAMTSL5 appears to activate preferentially CD8⁺ T cells. Of note, both antigens are recognized by T cells when being presented by HLA-C*06:02, i.e., the most prominent psoriasis risk gene in the genome [located on PSORS1 (psoriasis susceptibility locus 1) on chromosome 6p21.3].

While the role of cellular adaptive immunity is becoming increasingly plausible, only recently autoantibodies, i.e., elements of humoral adaptive immunity, have been described in patients with psoriasis and psoriatic arthritis. Interestingly, these IgG are directed against (carbamylated/citrullinated) LL37 or ADAMTSL5 (56, 57). Since the serum concentrations of these antibodies were associated with the severity of psoriasis and since patients with psoriatic arthritis had higher serum levels, it is conceivable that a causal pathogenetic relationship and a contribution to systemic inflammation exist (56). It is also possible that the respective autoantibodies exert protective functions through scavenging autoantigens. However, their roles need to be clarified in future studies.

The Other Side: Antigen Presentation by HLA Molecules in Psoriasis

While most, if not all, autoimmune diseases are linked with certain HLA alleles (58–60), *HLA-C**06:02 is the predominant psoriasis risk gene (61–63). HLA class I molecules present short peptide antigens (8–10 amino acids) to $\alpha\beta$ TCRs of CD8+ T cells. Such antigenic peptides are usually derived within the antigen presenting cell from (intracellular) parent proteins by proteasomal cleavage and loaded onto HLA-class I molecules. The HLA/peptide complex is then transported to the cell membrane where it can be recognized by CD8+ T cells (64, 65). Thus, HLA-class I-restricted immune responses are usually directed against target cells which produce the antigenic peptide.

HLA-C*06:02-presented non-apeptides (9 amino acids long) possess anchor amino acids at residues 2 (arginine) and 9

(leucine, valine, and less frequently methionine and isoleucine), along with a putative anchor at residue 7 (arginine). HLA-C*06:02 features very negatively charged pockets and thus binds to distinct positively charged peptides. Given that between 1,000 and 3,000 different self-peptides have been detected on HLA-C*06:02 under experimental conditions, multiple cellular proteins should be, in principle, presented by this HLA molecule and recognizable by CD8+ T cells (66, 67).

HLA- $C^*06:02$, and other psoriasis-related HLA types such as HLA- $C^*07:01$, HLA- $C^*07:02$, and HLA- B^*27 utilize identical anchor residues and present partially overlapping peptide residues (66, 67). Moreover, a negatively charged binding pocket is shared with another risk allele, HLA- $C^*12:03$ (68, 69), resulting in similar functional domains and peptidebinding characteristics (67, 70). Thus, several HLA-class I types implicated in psoriasis appear to share similar peptide-binding properties. It is, therefore, conceivable that they can substitute for each other in conferring psoriasis risk. However, HLA- $C^*06:02$ is the prototype allele within this spectrum and is associated with the highest risk for psoriasis.

Supporting Acts: Indispensable Players in the Ensemble of Psoriasis Immunology

Autoantigen presentation alone does not suffice to induce the psoriatic cascade in genetically predisposed individuals. Rather, costimulatory effects of various gene products orchestrate the activation of the actual autoimmune response. Such risk gene variants modulate inflammatory signaling pathways (e.g., the IL-23 pathway), peptide epitope processing and/or Th/c17 differentiation (a selection of important factors is summarized in **Table 1**).

These genetic variations create costimulatory signals which modulate innate and adaptive immune mechanisms and shape the proinflammatory environment. In sum and in conjunction with the appropriate HLA molecules and autoantigens, they may eventually exceed the thresholds for activation and maintenance of pathogenic autoimmune and autoinflammatory responses in psoriasis (29, 71). Likewise, regulatory mechanisms involving programmed death (PD)-1 signals have emerged recently as modulators of chronic inflammation in psoriasis (72). However, the complex interactions of various players are by no means fully understood. Therefore, they are listed here only as a whole.

The autoantigens described so far cannot fully explain the genesis of psoriasis. To give just one example of the latter notion: Psoriatic lesions can also occur in vitiligo foci that do not contain melanocytes (73, 74). Alterations of resident cell types such as vascular endothelial cells or the cutaneous nervous system are also involved in the disease process (75–77). Further research is certainly needed here.

SHADES OF GRAY: CROSSTALK BETWEEN ADAPTIVE AND INNATE IMMUNITY IN PSORIASIS

In addition to the antigen-specific facilitation of inflammation in psoriasis, there are several strong connections to components of the innate immune system. The crosstalk between the innate and adaptive branches of the immune system in psoriasis is complex and can only be highlighted by a few selected examples. Its fine-tuning arguably determines the actual clinical correlate within the spectrum of the disease. Indeed, there is accumulating circumstantial evidence that in patients with stable and mild disease, mechanisms of adaptive immunity are more likely to be in the foreground, while innate mechanisms seem to be more important in patients with active severe disease, systemic involvement and comorbid conditions (78) (**Figure 1**). The impact on systemic comorbid diseases has been interpreted, at least in part, as a systemic "*spillover*" of innate inflammatory processes in severe psoriasis (78). Of course, such factors are not specific for psoriasis, but appear to account for a general inflammatory state in patients with severe psoriasis.

Patients with severe psoriasis have increased levels of inflammatory cytokines, CRP, fibrinogen, $\alpha 2$ macroglobulin or PAI-1 (plasminogen activator inhibitor-1) in the blood (79–81), they show transcriptomic, proteomic and metabolomic abnormalities (82) and there are connections with chronic stress (83) and biophysical properties of the skin (84).

The serum levels of inflammatory cytokines have been proposed as parameters for disease severity (85). Such general inflammatory markers are accompanied by increased numbers of Th1, Th17, and Th22 cells in patients with severe psoriasis (86, 87), which provides a direct link with autoimmune (adaptive) processes. Moreover, there is an increasing number of modulating factors, such as autoimmune reactivity to ribonucleoprotein A1 (HNRNPA1) (88), which impact on the course and severity of psoriasis.

One of the perhaps most vivid recent examples of how individual mediators influence the spectrum of psoriasis by shifting innate or adaptive immune processes comes from research on the interplay between IL-17- and IL-36-driven inflammation (89). The three IL-36 isoforms (IL-36 α , β , and γ) belong to the IL-1 family and are upregulated in psoriatic skin (90, 91). They bind to the IL-36 receptor (IL-36R), thereby inducing transcription of several inflammatory mediators through NF-κβ activation. IL-36Ra (IL-1F9), an antiinflammatory natural IL36R antagonist, is encoded by the IL36RN gene and is abundantly present in the skin of patients with psoriasis vulgaris, which may constitute part of the "checks and balances" that control the psoriatic inflammation (90, 92). Function-abrogating mutations in the IL36RN gene may result in unrestrained inflammatory effects of IL-36. Absence of IL-36Ra then leads to excessive neutrophil accumulation as observed in some cases of familial generalized pustular psoriasis (92-94). Palmoplantar pustular psoriasis, however, seems to be related to CARD14 variants rather than IL36RN mutations (95, 96).

While most cases of pustular psoriasis occur without such mutations (97), IL-36-related processes appear to contribute decisively to the actual clinical manifestation of specific psoriatic phenotypes: It has recently been shown elegantly that the skin of patients with psoriasis vulgaris differs significantly from that of patients with pustular psoriasis—in a sense, opposite ends of the spectrum of psoriasis: while in both forms numerous genes are expressed abnormally, these differentially expressed

TABLE 1 | Genetic factors implicated in psoriasis.

Pathogenic function	Psoriasis-associated gene locus
IL-23/IL-17A signaling	IL23R, interleukin-23 receptor
	IL12B, interleukin-12 subunit p40, also part of IL-23
	IL12RB, interleukin-12 receptor subunit beta 1, also termed IL-12RB1
	IL23A, interleukin-23 subunit alpha, p19
	IL23R, interleukin-23 receptor, a Janus kinase-2 associated type I cytokine receptor that activates STAT3 upon ligand binding
	TYK2, tyrosine-protein kinase 2, a Janus kinase family member, facilitates type I and II cytokine receptor and type I and III interferon signaling pathways, involved in innate and adaptive immune processes
	STAT3, signal transducer and activator of transcription 3, a central transcription factor in inflammatory processes
	STAT5A/B, signal transducer and activator of transcription 5A/B
	SOCS1, suppressor of cytokine signaling 1, a member of the STAT-induced STAT inhibitor family, negative regulator of cytokine signaling
	ETS1, a member of the E26 transformation-specific transcription factors, a negative regulator of Th17 cells
	TRAF3IP2, tumor necrosis factor receptor-associated factor 3-interacting protein-2, central role in response to inflammatory signals
	KLF4, Krüppel-like factor 4, influences NF-κB-mediated inflammatory pathways
	IF3, eukaryotic translation initiation factor 3
Effector T-cell function and differentiation	ETS1, E26 transformation-specific transcription factor 1, a negative regulator of Th17 cells
	RUNX3, runt-related transcription factor 3, a runt domain-containing transcription factor involved in regulation of many cellular processes
	TNFRSF9, tumor necrosis factor receptor superfamily member 9, CD137, costimulator of T cells
	MBD2, methyl-CpG-binding domain protein 2, binds to methylated DNA and regulates transcription from methylated gene promote
	IRF4, interferon regulatory factor 4
Type I interferon and cytokine	ELMO1, engulfment and cell motility protein 1
signaling	TYK2, non-receptor tyrosine-protein kinase, a Janus kinase family member (see above)
	SOCS1, suppressor of cytokine signaling 1, a member of the STAT-induced STAT inhibitor family, negative regulator of cytokine signaling
	IFIH1/MDA5, Interferon-induced helicase C domain-containing protein 1/melanoma differentiation-associated protein 5, a CARD (caspase activation and recruitment domain) protein involved in IL-1 and IL-18 processing and in regulation of inflammation
	RNF114, ring finger protein 114, a ubiquitin ligase
	IRF4, interferon regulatory factor 4
	RIG1/DDX58, retinoic acid inducible gene I encoded by the DDX58 (DExD/H-Box Helicase 58) gene; contains a RNA helicase motif and a caspase recruitment domain (CARD), involved in regulation of immune responses
	IFNLR1/IL28RA, interferon lambda receptor 1, forms complex with IL10RB and interacts with IL-28A, IL-28B, and IL-29, involved in immune regulation
	IFNGR2, interferon gamma receptor 2, non-ligand-binding beta chain of the IFN γ chain
Regulation of NF-κB-associated inflammatory signaling pathways	TNFAIP3, TNFα induced protein 3, a ubigitin-editing enzyme that inhibits NF-κB activation and TNF-mediated apoptosis; involved in the cytokine-mediated inflammatory responses
	TNIP1, TNFAIP3 interacting protein 1, a regulator of NF-κB activation
	TYK2, non-receptor tyrosine-protein kinase, a Janus kinase family member (see above)
	REL, reticuloendotheliosis oncogene, a NF-kB subunit (c-Rel) involved in apoptosis, inflammation and immune responses, SNPs are also associated with ulcerative colitis and rheumatoid arthritis
	NFkBIA, NF- κ B inhibitor α , interacts with NF- κ B/c-Rel involved in inflammatory responses.
	CARD14, caspase recruitment domain family member 14, a scaffold protein involved in cell adhesion, signal transduction and cell polarity, involved in NF-κB activation.
	CARM1, coactivator associated arginine methyltransferase 1, catalyzes methylation of histones and other chromatin-associated proteins, involved in regulation of gene expression
	UBE2L3, ubiquitin conjugating enzyme E2 L3, an E2 ubiquitin-conjugating enzyme, participates in ubiquitination of the p105 NF-κB precursor
	FBXL19, F-box and leucine rich repeat protein 19, an E3 ubiquitin ligase, binds to interleukin 1 receptor-like 1 and regulates its ubiquitination, associated with pulmonary inflammation and psoriasis
Antigen processing (N-terminal trimming)	ERAP1, endoplasmic reticulum aminopeptidase 1, involved in trimming of HLA class I-binding precursors enabling them to be presented on MHC class I molecules

Most genetic associations have direct connections with immune functions implicated in the pathophysiology of psoriasis.

Adaptive and Innate Immunity in Psoriasis

genes overlap only to a relatively small extent. In psoriasis vulgaris, genes involved in adaptive (T-cell-associated) immune processes predominate, whereas in pustular psoriasis processes of innate immunity (mainly neutrophil-associated) they are dysregulated. Interestingly, IL-36 seems to play an important role for the accumulation of neutrophilic granulocytes and a pustular phenotype of psoriasis (89). The balance between IL-36 and IL-17 seems to contribute—at least partially—to clinical symptoms of psoriasis vulgaris vs. psoriasis pustulosa. If this interpretation of the data is correct, then this would constitute a mechanism that regulates the fine-tuning between innate and adaptive immune processes.

THE IL-23/IL-17 PATHWAY CONNECTS INNATE AND ADAPTIVE IMMUNITY IN PSORIASIS

The notion of psoriasis featuring elements of both antigenspecific autoimmunity and non-specific autoinflammation needs to be considered a bit more closely, in particular downstream of innate and/or adaptive activation processes. As of today, interfering with IL-17A or IL-23 are the most efficient treatment modalities against psoriasis (98). Indeed, the IL-23/IL-17 axis seems to be particularly well-suited to exemplify the intricate crosstalk between adaptive and innate immunity in psoriasis.

Healthy human skin contains only a few IL-17-producing T cells (99), a population of $CD4^+$ T cells distinct from the "classical" Th1 and Th2 cells. They were eponymously named for their production of IL-17 (100). In psoriasis (31, 101, 102), palmoplantar pustulosis (103) and other inflammatory disorders (104, 105), Th17 lymphocytes are vastly expanded and are thought to contribute decisively to the pathogenesis of these conditions (Figure 3). The resulting imbalance between Th17 and regulatory T cells (Treg) favors inflammation (106). The IL-17 production of T lymphocytes is further stimulated by activated keratinocytes, thus creating a positive feedback loop (107). Th17 cells are controlled by regulatory T cells through IL-10 (108). In psoriatic skin, IL-17A is considered the most relevant of the six known isoforms (102). IL-17A is not only secreted by CD4+ Th17 cells, but also by CD8+ T cells (109) and certain cells of the innate immune system including neutrophilic granulocytes (110-112), thus further highlighting the tight connection of innate and adaptive immunity in psoriasis. The presentation of IL-17 by neutrophil extracellular traps (NETs), which are generated upon activation of neutrophils in a clearly defined manner (113) and are prominently present in both pustular and plaque-type psoriasis (114), may also play a role (115).

In this context it should be mentioned that so-called tissue resident memory cells ($T_{\rm rm}$ cells) in psoriatic skin remain in the long term even after resolution of the lesions, which contribute as mediators of the local adaptive immune response to renewed exacerbations. Although the role of these cells is not yet fully understood, there is growing evidence of their pathogenic role in psoriasis and other chronic inflammatory diseases (116–118). $T_{\rm rm}$ cells in psoriatic lesions are CD8⁺ but lack CD49a

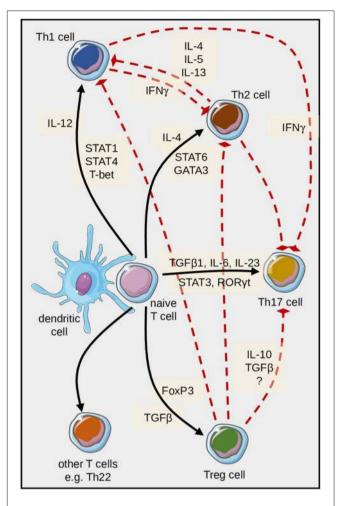


FIGURE 3 Differentiation of pathogenic T cells in psoriasis is embedded in a complex regulatory network. Naïve T cells can differentiate into several directions; this is mainly determined by the cytokines and transcription factors depicted here. In addition, various regulatory feedback mechanisms exist, some of which are schematically highlighted here with particular reference to Th17 cell differentiation and function.

(α 1 integrin) expression; they predominantly generate IL-17 responses that promote local inflammation (119).

In addition to Th17 cells, T cells which produce both IL-17 and IFN γ (termed Th17/Th1-T cells) and IL-22-producing T cells can also be detected in psoriatic skin (31). Naive T cells express several cytokine receptors including the IL-23 receptor. DC-derived TGF β 1, IL-1 β , IL-6, and IL-23 facilitate priming and proliferation of Th17 cells (120–122), while IL-12 assumes these functions for Th1 cells, and IL-6 and TNF α contribute to the programming of Th22 cells.

The balance of Th17 cells and Th1 cells appears to be critical for the pathogenesis of psoriasis (123) and other related conditions (124). There are several exogenous factors such as ultraviolet light or vitamin D3 (125, 126), or other cytokines like IL-9 (127) that can modulate Th17-dependent inflammation. Of note, IL-17A can also be produced independent of IL-23, e.g., by

 $\gamma\delta$ T lymphocytes or invariant natural killer (iNKT) cells (128–130). However, it is not clear yet whether this alternative pathway impacts on the accrual and course of inflammatory disorders or potential undesired effects of either IL-23 or IL-17 inhibition.

In any case, the IL-23/IL-17 axis in psoriasis clearly illuminates the close interaction of the innate immune system (represented by IL-23-producing myeloid cells) with cells of the adaptive immune system (in this case Th17- and IL17-expressing CD8⁺ T-cells). Psoriasis could again serve as a "model disease" to clarify such relationships.

CONTRIBUTION OF RESIDENT SKIN CELLS TO IMMUNOLOGICAL PROCESSES IN PSORIASIS

Multiple genetic and environmental factors influence the immunopathology of psoriasis (131). The mechanisms leading to the first occurrence of psoriasis in predisposed individuals are only partly known. Infections with streptococci, medications such as lithium, antimalarials, or ß-blockers, or physical or

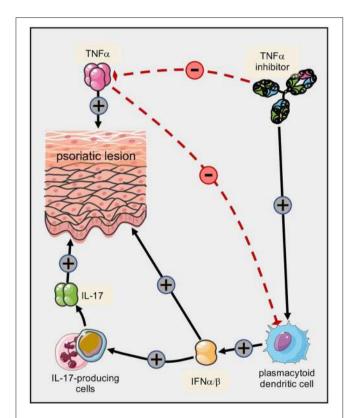


FIGURE 4 | Paradoxical psoriasis triggered by TNF inhibitors in predisposed individuals. Several cytokines contribute to the pathogenesis of psoriatic skin lesions, with TNF α and IL-17A playing prominent roles. However, TNF α also exerts an inhibitory effect on plasmacytoid dendritic cells. Upon therapeutic inhibition of TNF α , this inhibitory effect is abrogated and the resulting shift toward increased production of type I interferons fuels the secretion of IL-17. It is conceivable that additional mechanisms contribute to the shift of cytokines ultimately resulting in "paradoxical" psoriatic lesions.

chemical stress may trigger the disease. Minimal trauma can induce rapid immigration and activation of immune cells including T-cells and neutrophils (132), the so-called Köbner phenomenon (133, 134). Feedback loops between adaptive immune cells (T cells), innate immune cells (neutrophilic granulocytes, macrophages, dendritic cells), and resident skin cells (keratinocytes, endothelial cells) result in an amplification and chronification of the inflammatory response. Aspects of systemic inflammation in patients with severe psoriasis are thought to contribute to comorbid diseases (135).

Hyperproliferative keratinocytes in psoriatic plaques produce large amounts of antimicrobial peptides and proteins (AMP). These positively charged peptides, which have been termed alarmins, have strong proinflammatory properties. Most studies have addressed cathelicidin and its fragment, LL37, which is highly expressed psoriatic skin (136, 137). The positively charged LL37 can associate with negatively charged nucleic acids (DNA and RNA), thus forming immunostimulatory complexes. The free DNA required for such complexes probably comes from neutrophils (which form NETs) and damaged resident skin cells (e.g., traumatized keratinocytes). Plasmacytoid DC (pDC) and myeloid dendritic cells (DC) take up these complexes. Subsequently, RNA motifs stimulate toll-like receptors (TLR) 7 and 8, and DNA triggers TLR9 signaling (52, 138). Cytokines such as TNF, IL-23, and IL-12 are produced by TLR7/8stimulated myeloid DC, while pDC make type I-interferons (IFNa), all of which fuel the psoriatic inflammation (131). A prominent role in psoriasis and other autoimmune diseases has been attributed to the so-called 6-sulfo LacNAc (slan) DC (139).

Several other skin-derived alarmins such as S100 proteins are inflammatory AMPs also implicated in the pathogenesis of psoriasis. Indeed, IL-17A induces the production of S100A7 (psoriasin) and S100A15 (koebnerisin) by keratinocytes (140, 141). Likewise, myeloid cells and keratinocytes produce the calgranulins, S100A8 and S100A9 [also termed myeloid-related protein (Mrp) 8 and Mrp14], both of which induce T-cell mediated autoimmune reactions and inflammatory changes in keratinocytes (142, 143). Similar to LL37, human ß-defensin (HBD) 2 and HBD4 bind DNA, trigger TLR9 and stimulate pDC (144). Innate immune sensing is also facilitated by IL-26 bound to self-DNA (145).

Activated DC in turn can program the differentiation of naive T into pathogenic T cells [reviewed in (8)]. Neutrophilic granulocytes, too, release AMP, inflammatory cytokines, proteases, free oxygen radicals, and NETs, all of which have been implicated in the inflammatory cascade in psoriasis (8, 114).

NOT ALONE: RELATIONS AND SIMILARITIES OF PSORIASIS WITH OTHER AUTOIMMUNE AND AUTOINFLAMMATORY DISORDERS

The highlights outlined so far show that both adaptive and innate immune processes contribute to psoriasis. Their balance and finetuning seem to determine the development of certain clinical forms of the disease, but also organ-specific manifestations. On the one hand, the outlined long-term systemic inflammatory processes probably contribute to the pathogenesis of important metabolic, cardiovascular, and mental concomitant diseases. In these areas, the evidence of a causal relationship is becoming increasingly clear and numerous publications prove this. A more detailed overview can be found elsewhere in this thematic focus. On the other hand, the contoured adaptive and innate immune mechanisms are not specific for psoriasis. Rather, many of them have been found—in varying degrees and weightings—in a whole range of other autoimmune and autoinflammatory diseases. In any case, although this interplay of different components of the immune system is certainly not yet fully understood, parallels with other chronic inflammatory and autoimmune diseases emerge that underpin our current view of psoriasis as a systemic disease.

Indeed, the prevalence of several autoimmune and/or autoinflammatory diseases including rheumatoid arthritis, celiac disease, Crohn's disease, multiple sclerosis, systemic lupus erythematosus, vitiligo, Sjögren's syndrome, alopecia areata, or autoimmune thyroiditis appears to be increased in patients with psoriasis compared to that in healthy controls (146, 147). Several other and more uncommon associations have also been reported (148). Such associations have been attributed to certain genetic and immunological similarities and "overlaps" (146, 149). Three such disease complexes associated with psoriasis, i.e., rheumatoid arthritis, Crohn's disease and systemic lupus erythematosus, will be briefly discussed as examples.

Psoriasis susceptibility 1 candidate gene 1 (*PSORS1C1*), a gene thought to be involved in IL-17 and IL-1β regulation, is increased in immune cells from patients with rheumatoid arthritis (150). Moreover, aberrant expression of runt-related transcription factor 1 (*RUNX1*) has been implicated in defective regulation of sodium-hydrogen antiporter 3 regulator 1 (*SLC9A3R1*) and N-acetyltransferase 9 (*NAT9*) in both psoriasis and rheumatoid arthritis (151–153). Polymorphisms of the IL-23R gene have also been implicated in both diseases, which further underscores the general relevance of the IL-23/IL-17 axis (154). TNFαinduced protein 3 (*TNFAIP3*), which negatively regulates NF-κB signaling, is another gene thought to be involved in rheumatoid arthritis and psoriasis alike, but also in Crohn's disease, celiac disease, and systemic lupus erythematosus (155, 156).

Similar functional imbalances between Th17 and regulatory T cells (Tregs) as well as similar central cytokines including TNF α , IL-23, and IL-17A, but also IL-1 β , IL-6, IL-17F, and IL-21 contribute to both diseases (131, 157–159). Such striking parallels result in the response of both disorders to the same therapies.

Similar to rheumatoid arthritis, Crohn's disease is significantly more prevalent in patients with psoriasis compared to healthy controls and *vice versa* (160–164). Moreover, considerable genetic overlap exists between both diseases as exemplified by seven mutual susceptibility loci (165). Genes involved in the same way include some relevant for the IL-23/I-17 axis such as *IL23R*, *IL12B*, and *TYK2* (166–169).

One of the first immunological parallels found between psoriasis and Crohn's disease was the central pathogenic role of TNF α (170–172). Hence, TNF α inhibitors ameliorate both

disorders (172-175). Paradoxical induction of psoriasis in patients treated with TNFa inhibitors has been attributed to shifts within the balance of $TNF\alpha$ and type I interferons (IFNa) with impact on plasmacytoid dendritic cells (176-181) (Figure 4). The pathophysiology of such paradoxical reactions in other immunomodulating settings is less clear (182, 183). The composition of the inflammatory infiltrate (T cells, macrophages, dendritic cells and neutrophilic granulocytes) as well as inflammatory mediators (IFNy, IL-12, IL-6, IL-17) are conspicuously similar in psoriasis and Crohn's disease (131, 184). A dysregulated balance between Th17 cells and CD4+CD25high Foxp3+ Tregs is thought to lie at the core of both diseases (101, 157, 185-187). In addition, there may even be IL-17 producing Tregs in lesions of both Crohn's disease and psoriasis (188, 189), suggesting differentiation of Tregs toward a proinflammatory phenotype. However, a putative protective role of IL-17 in Crohn's disease (190) may explain, at least in part, the worsening of gut inflammation in some cases upon inhibition of IL-17A (191).

Increased expression of IL-6 has been demonstrated in psoriatic plaques and inflamed intestinal mucosa alike (192, 193). IL-6 signaling induces STAT3 phosphorylation, which leads to relative resistance of effector T cells toward Tregs (194, 195).

The association of psoriasis and systemic lupus erythematosus is uncommon and controversially discussed (196, 197). However, dysfunctional interaction of RUNX1 with its binding site due to nucleotide polymorphisms links psoriasis not only with rheumatoid arthritis but also with systemic lupus erythematosus (153, 198, 199). RUNX1 binding on chromosome 2 is defective in some patients with SLE, while RUNX1 binding on chromosome 17 seems to be altered in some psoriasis patients.

TNF receptor-associated factor 3 Interacting Protein 2 (*TRAF3IP2*) has been described as a genetic susceptibility locus for psoriasis and appears to facilitate IL-17 signaling in both psoriasis and systemic lupus erythematosus (200–205). On the cellular level, psoriasis and systemic lupus erythematosus share impaired Treg functions (157, 192, 206, 207), thus suggesting that similar genetic and immune alterations govern pathological immune reactions in both psoriasis and systemic lupus erythematosus.

In summary, psoriasis shows elements of both autoimmune and autoinflammatory mechanisms, whose fine-tuning determines the actual clinical symptoms within the broad spectrum of the disease. Given that psoriasis is a systemic disease that shares conspicuous genetic and immunological similarities with other autoimmune and autoinflammatory disorders, it may serve as a model disorder for research into general mechanisms of such complex immunological regulations.

AUTHOR CONTRIBUTIONS

The author confirms being the sole contributor of this work and has approved it for publication.

REFERENCES

- Boehncke WH, Sterry W. Psoriasis-a systemic inflammatory disorder: clinic, pathogenesis and therapeutic perspectives. J Dtsch Dermatol Ges. (2009) 7:946–52. doi: 10.1111/j.1610-0387.2009.07083.x
- Schäkel K, Schön MP, Ghoreschi K. Pathogenesis of psoriasis. Hautarzt. (2016) 67:422–31. doi: 10.1007/s00105-016-3800-8
- Boehncke WH, Schön MP. Psoriasis. Lancet. (2015) 386:983–94. doi: 10.1016/S0140-6736(14)61909-7
- Volc S, Ghoreschi K. Pathophysiological basis of systemic treatments in psoriasis. J Dtsch Dermatol Ges. (2016) 14:557–72. doi: 10.1111/ddg.13050
- Schön MP, Boehncke WH. Psoriasis. N Engl J Med. (2005) 352:1899–912. doi: 10.1056/NEJMra041320
- Nestle FO, Kaplan DH, Barker J. Psoriasis. N Engl J Med. (2009) 361:496–509. doi: 10.1056/NEJMra0804595
- Harden JL, Krueger JG, Bowcock AM. The immunogenetics of psoriasis: a comprehensive review. J Autoimmun. (2015) 64:66–73. doi: 10.1016/j.jaut.2015.07.008
- Schön MP, Erpenbeck L. The Interleukin-23/Interleukin-17 axis links adaptive and innate immunity in psoriasis. *Front Immunol.* (2018) 9:1323. doi: 10.3389/fimmu.2018.01323
- Prinz JC. Human leukocyte antigen-class I alleles and the autoreactive T cell response in psoriasis pathogenesis. *Front Immunol.* (2018) 9:954. doi: 10.3389/fimmu.2018.00954
- Mueller W, Herrmann B. Cyclosporin A for psoriasis. N Engl J Med. (1979) 301:555. doi: 10.1056/NEJM197909063011015
- Gardembas-Pain M, Ifrah N, Foussard C, Boasson M, Saint Andre JP, Verret JL. Psoriasis after allogeneic bone marrow transplantation. *Arch Dermatol.* (1990) 126:1523. doi: 10.1001/archderm.1990.01670350139033
- Burden AD, Javed S, Bailey M, Hodgins M, Connor M, Tillman D. Genetics of psoriasis: paternal inheritance and a locus on chromosome 6p. J Invest Dermatol. (1998) 110:958–60. doi: 10.1046/j.1523-1747.1998.00213.x
- Trembath RC, Clough RL, Rosbotham JL, Jones AB, Camp RD, Frodsham A, et al. Identification of a major susceptibility locus on chromosome 6p and evidence for further disease loci revealed by a two stage genome-wide search in psoriasis. *Hum Mol Genet.* (1997) 6:813–20. doi: 10.1093/hmg/6.5.813
- 14. Chen L, Tsai TF. HLA-Cw6 and psoriasis. Br J Dermatol. (2018) 178:854–62. doi: 10.1111/bjd.16083
- Ellis CN, Krueger GG, Group. ACS. Treatment of chronic plaque psoriasis by selective targeting of memory effector T lymphocytes. *N Engl J Med.* (2001) 345:248–55. doi: 10.1056/NEJM200107263450403
- Ghoreschi K, Thomas P, Breit S, Dugas M, Mailhammer R, van Eden W, et al. Interleukin-4 therapy of psoriasis induces Th2 responses and improves human autoimmune disease. *Nat Med.* (2003) 9:40–6. doi: 10.1038/nm804
- Guenova E, Skabytska Y, Hoetzenecker W, Weindl G, Sauer K, Tham M, et al. IL-4 abrogates T(H)17 cell-mediated inflammation by selective silencing of IL-23 in antigen-presenting cells. *Proc Natl Acad Sci USA*. (2015) 112:2163–8. doi: 10.1073/pnas.1416922112
- Onderdijk AJ, Baerveldt EM, Kurek D, Kant M, Florencia EF, Debets R, et al. IL-4 downregulates IL-1β and IL-6 and induces GATA3 in psoriatic epidermal cells: route of action of a Th2 cytokine. J Immunol. (2015) 195:1744–52. doi: 10.4049/jimmunol.1401740
- Rácz E, Kurek D, Kant M, Baerveldt EM, Florencia E, Mourits S, et al. GATA3 expression is decreased in psoriasis and during epidermal regeneration; induction by narrow-band UVB and IL-4. *PLoS ONE*. (2011) 6:e19806. doi: 10.1371/journal.pone.0019806
- Asadullah K, Sterry W, Stephanek K, Jasulaitis D, Leupold M, Audring H, et al. IL-10 is a key cytokine in psoriasis. Proof of principle by IL-10 therapy: a new therapeutic approach. *J Clin Invest.* (1998) 101:783–94. doi: 10.1172/JCI1476
- Breban M, Hammer RE, Richardson JA, Taurog JD. Transfer of the inflammatory disease of HLA-B27 transgenic rats by bone marrow engraftment. J Exp Med. (1993) 178:1607–16. doi: 10.1084/jem.178.5.1607
- Hammer RE, Maika SD, Richardson JA, Tang JP, Taurog JD. Spontaneous inflammatory disease in transgenic rats expressing HLA-B27 and human beta 2m: an animal model of HLA-B27-associated human disorders. *Cell.* (1990) 63:1099–112. doi: 10.1016/0092-8674(90)90512-D

- Schön MP, Detmar M, Parker CM. Murine psoriasis-like disorder induced by naive CD4+ T cells. Nat Med. (1997) 3:183–8. doi: 10.1038/nm0297-183
- Hong K, Chu A, Lúdvíksson BR, Berg EL, Ehrhardt RO. IL-12, independently of IFN-gamma, plays a crucial role in the pathogenesis of a murine psoriasislike skin disorder. *J Immunol.* (1999) 162:7480–91.
- Boehncke WH, Dressel D, Zollner TM, Kaufmann R. Pulling the trigger on psoriasis. *Nature*. (1996) 379:777. doi: 10.1038/379777a0
- Wrone-Smith T, Nickoloff BJ. Dermal injection of immunocytes induces psoriasis. J Clin Invest. (1996) 98:1878–87. doi: 10.1172/JCI118989
- Arakawa A, Siewert K, Stöhr J, Besgen P, Kim SM, Rühl G, et al. Melanocyte antigen triggers autoimmunity in human psoriasis. J Exp Med. (2015) 212:2203–12. doi: 10.1084/jem.20151093
- Lande R, Botti E, Jandus C, Dojcinovic D, Fanelli G, Conrad C, et al. The antimicrobial peptide LL37 is a T-cell autoantigen in psoriasis. *Nat Commun.* (2014) 5:6595. doi: 10.1038/ncomms6621
- 29. Prinz JC. Autoimmune aspects of psoriasis: heritability and autoantigens. *Autoimmun Rev.* (2017) 16:970–9. doi: 10.1016/j.autrev.2017.07.011
- Prinz JC. Melanocytes: target cells of an HLA-C*06:02-restricted autoimmune response in psoriasis. J Invest Dermatol. (2017) 137:2053–8. doi: 10.1016/j.jid.2017.05.023
- Lowes MA, Kikuchi T, Fuentes-Duculan J, Cardinale I, Zaba LC, Haider AS, et al. Psoriasis vulgaris lesions contain discrete populations of Th1 and Th17 T cells. *J Invest Dermatol.* (2008) 128:1207–11. doi: 10.1038/sj.jid.5701213
- Perera GK, Di Meglio P, Nestle FO. Psoriasis. Annu Rev Pathol. (2012) 7(385-422). doi: 10.1146/annurev-pathol-011811-132448
- Davis MM, Bjorkman PJ. T-cell antigen receptor genes and T-cell recognition. *Nature*. (1988) 334:395–402. doi: 10.1038/334395a0
- Woodsworth DJ, Castellarin M, Holt RA. Sequence analysis of T-cell repertoires in health and disease. *Genome Med.* (2013) 5:98. doi: 10.1186/gm502
- Menssen A, Trommler P, Vollmer S, Schendel D, Albert E, Gürtler L, et al. Evidence for an antigen-specific cellular immune response in skin lesions of patients with psoriasis vulgaris. *J Immunol.* (1995) 155:4078–83.
- Bour H, Puisieux I, Even J, Kourilsky P, Favrot M, Musette P, et al. T-cell repertoire analysis in chronic plaque psoriasis suggests an antigen-specific immune response. *Hum Immunol.* (1999) 60:665–76. doi: 10.1016/S0198-8859(99)00027-0
- Prinz JC, Vollmer S, Boehncke WH, Menssen A, Laisney I, Trommler P. Selection of conserved TCR VDJ rearrangements in chronic psoriatic plaques indicates a common antigen in psoriasis vulgaris. *Eur J Immunol.* (1999) 29:3360–8.
- Costello PJ, Winchester RJ, Curran SA, Peterson KS, Kane DJ, Bresnihan B, et al. Psoriatic arthritis joint fluids are characterized by CD8 and CD4 T cell clonal expansions appear antigen driven. *J Immunol.* (2001) 166:2878–86. doi: 10.4049/jimmunol.166.4.2878
- Lin WJ, Norris DA, Achziger M, Kotzin BL, Tomkinson B. Oligoclonal expansion of intraepidermal T cells in psoriasis skin lesions. J Invest Dermatol. (2001) 117:1546–53. doi: 10.1046/j.0022-202x.2001.01548.x
- Borgato L, Puccetti A, Beri R, Codella O, Frigo A, Simeoni S, et al. The T cell receptor repertoire in psoriatic synovitis is restricted and T lymphocytes expressing the same TCR are present in joint and skin lesions. *J Rheumatol.* (2002) 29:1914–9.
- Matos TR, O'Malley JT, Lowry EL, Hamm D, Kirsch IR, Robins HS, et al. Clinically resolved psoriatic lesions contain psoriasis-specific IL-17-producing αβ T cell clones. *J Clin Invest.* (2017) 127:4031–41. doi: 10.1172/JCI93396
- Conrad C, Boyman O, Tonel G, Tun-Kyi A, Laggner U, de Fougerolles A, et al. Alpha1beta1 integrin is crucial for accumulation of epidermal T cells and the development of psoriasis. *Nat Med.* (2007) 13:836–42. doi: 10.1038/nm1605
- Di Meglio P, Villanova F, Navarini AA, Mylonas A, Tosi I, Nestle FO, et al. Targeting CD8(+) T cells prevents psoriasis development. J Allergy Clin Immunol. (2016) 138:274–6. doi: 10.1016/j.jaci.2015.10.046
- Chang JC, Smith LR, Froning KJ, Schwabe BJ, Laxer JA, Caralli LL, et al. CD8+ T cells in psoriatic lesions preferentially use T-cell receptor V beta 3 and/or V beta 13.1 genes. *Proc Natl Acad Sci USA*. (1994) 91:9282–6. doi: 10.1073/pnas.91.20.9282

- Diluvio L, Vollmer S, Besgen P, Ellwart JW, Chimenti S, Prinz JC. Identical TCR beta-chain rearrangements in streptococcal angina and skin lesions of patients with psoriasis vulgaris. *J Immunol.* (2006) 176:7104–11. doi: 10.4049/jimmunol.176.11.7104
- Kim SM, Bhonsle L, Besgen P, Nickel J, Backes A, Held K, et al. Analysis of the paired TCR alpha- and beta-chains of single human T cells. *PLoS ONE*. (2012) 7:e7338. doi: 10.1371/journal.pone.0037338
- Valdimarsson H, Thorleifsdottir RH, Sigurdardottir SL, Gudjonsson JE, Johnston A. Psoriasis–as an autoimmune disease caused by molecular mimicry. *Trends Immunol.* (2009) 30:494–501. doi: 10.1016/j.it.2009.07.008
- 48. Thorleifsdottir RH, Sigurdardottir SL, Sigurgeirsson B, Olafsson JH, Sigurdsson MI, Petersen H, et al. Improvement of psoriasis after tonsillectomy is associated with a decrease in the frequency of circulating T cells that recognize streptococcal determinants and homologous skin determinants. *J Immunol.* (2012) 188:5160–5. doi: 10.4049/jimmunol.1102834
- 49. Sigmundsdottir H, Sigurgeirsson B, Troye-Blomberg M, Good MF, Valdimarsson H, Jonsdottir I. Circulating T cells of patients with active psoriasis respond to streptococcal M-peptides sharing sequences with human epidermal keratins. *Scand J Immunol.* (1997) 45:688–97. doi: 10.1046/j.1365-3083.1997.d01-438.x
- Besgen P, Trommler P, Vollmer S, Prinz JC. Ezrin, maspin, peroxiredoxin 2, heat shock protein 27: potential targets of a streptococcal-induced auto- immune response in psoriasis. *J Immunol.* (2010) 184:5392–402. doi: 10.4049/jimmunol.0903520
- Gudmundsdottir AS, Sigmundsdottir H, Sigurgeirsson B, Good MF, Valdimarsson H, Jonsdottir I. Is an epitope on keratin 17 a major target for autoreactive T lymphocytes in psoriasis? *Clin Exp Immunol.* (1999) 117:580–6. doi: 10.1046/j.1365-2249.1999.01013.x
- Lande R, Gregorio J, Facchinetti V, Chatterjee B, Wang YH, Homey B, et al. Plasmacytoid dendritic cells sense self-DNA coupled with antimicrobial peptide. *Nature*. (2007) 449:564–9. doi: 10.1038/nature06116
- Nestle FO, Conrad C, Tun-Kyi A, Homey B, Gombert M, Boyman O, et al. Plasmacytoid predendritic cells initiate psoriasis through interferon-alpha production. J Exp Med. (2005) 202:135–43. doi: 10.1084/jem.20050500
- Kahlenberg JM, Kaplan MJ. Little peptide, big effects: the role of LL-37 in inflammation and autoimmune disease. *J Immunol.* (2013) 191:4895–901. doi: 10.4049/jimmunol.1302005
- Lande R, Ganguly D, Facchinetti V, Frasca L, Conrad C, Gregorio J, et al. Neutrophils activate plasmacytoid dendritic cells by releasing self-DNApeptide complexes in systemic lupus erythematosus. *Sci Transl Med.* (2011) 3:73ra19. doi: 10.1126/scitranslmed.3001180
- Frasca L, Palazzo R, Chimenti MS, Alivernini S, Tolusso B, Bui L, et al. Anti-LL37 antibodies are present in Psoriatic Arthritis (PsA) patients: new biomarkers in PsA. *Front Immunol.* (2018) 9:1936. doi: 10.3389/fimmu.2018.01936
- Yuan Y, Qiu J, Lin ZT, Li W, Haley C, Mui UN, et al. Identification of novel autoantibodies associated with psoriatic arthritis. *Arthritis Rheumatol.* (2019) 71:941–951. doi: 10.1002/art.40830
- De Bakker PI, Raychaudhuri S. Interrogating the major histocompatibility complex with high-throughput genomics. *Hum Mol Genet*. (2012) 21:R29– 36. doi: 10.1093/hmg/dds384
- Fernando MM, Stevens CR, Walsh EC, De Jager PL, Goyette P, Plenge RM, et al. Defining the role of the MHC in autoimmunity: a review and pooled analysis. *PLoS Genet.* (2008) 4:e1000024. doi: 10.1371/journal.pgen.1000024
- Horton R, Wilming L, Rand V, Lovering RC, Bruford EA, Khodiyar VK, et al. Gene map of the extended human MHC. *Nat Rev Genet.* (2004) 5:889–99. doi: 10.1038/nrg1489
- Nair RP, Stuart PE, Nistor I, Hiremagalore R, Chia NV, Jenisch S, et al. Sequence and haplotype analysis supports HLA-C as the psoriasis susceptibility 1 gene. *Am J Hum Genet.* (2006) 78:827–51. doi: 10.1086/503821
- Tiilikainen A, Lassus A, Karvonen J, Vartiainen P, Julin M. Psoriasis and HLA-Cw6. Br J Dermatol. (1980) 102:179–84. doi: 10.1111/j.1365-2133.1980.tb05690.x
- Zhou F, Cao H, Zuo X, Zhang T, Zhang X, Liu X, et al. Deep sequencing of the MHC region in the Chinese population contributes to studies of complex disease. *Nat Genet.* (2016) 48:740–6. doi: 10.1038/ng.3576

- Blum JS, Wearsch PA, Cresswell P. Pathways of antigen processing. Annu Rev Immunol. (2013) 31:443–73. doi: 10.1146/annurev-immunol-032712-095910
- Neefjes J, Jongsma ML, Paul P, Bakke O. Towards a systems understanding of MHC class I and MHC class II antigen presentation. *Nat Rev Immunol.* (2011) 11:823–36. doi: 10.1038/nri3084
- 66. Di Marco M, Schuster H, Backert L, Ghosh M, Rammensee HG, Stevanovic S. Unveiling the peptide motifs of HLA-C and HLA-G from naturally presented peptides and generation of binding prediction matrices. *J Immunol.* (2017) 199:2639–2651. doi: 10.4049/jimmunol.1700938
- Mobbs JI, Illing PT, Dudek NL, Brooks AG, Baker DG, Purcell AW, et al. The molecular basis for peptide repertoire selection in the human leucocyte antigen (HLA) C*06:02 molecule. J Biol Chem. (2017) 292:17203– 15. doi: 10.1074/jbc.M117.806976
- Okada Y, Han B, Tsoi LC, Stuart PE, Ellinghaus E, Tejasvi T, et al. Fine mapping major histocompatibility complex associations in psoriasis and its clinical subtypes. *Am J Hum Genet.* (2014) 95:162–72. doi: 10.1016/j.ajhg.2014.07.002
- Fitzgerald O, Haroon M, Giles JT, Winchester R. Concepts of pathogenesis in psoriatic arthritis: genotype determines clinical phenotype. *Arthritis Res Ther.* (2015) 17:115. doi: 10.1186/s13075-015-0640-3
- Helms C, Saccone NL, Cao L, Daw JA, Cao K, Hsu TM, et al. Localization of PSORS1 to a haplotype block harboring HLA-C and distinct from corneodesmosin and HCR. *Hum Genet.* (2005) 118:466–76. doi: 10.1007/s00439-005-0048-2
- Indhumathi S, Rajappa M, Chandrashekar L, Ananthanarayanan PH, Thappa DM, Negi VS. Polymorphisms in T helper 1 proinflammatory cytokine genes and the risk of psoriasis in a South Indian Tamil cohort. J Dtsch Dermatol Ges. (2017) 15:208–11. doi: 10.1111/ddg.13043
- Geller S, Pulitzer M, Horwitz SM, Moskowitz AJ, Myskowski PL. Mycosis fungoides, Psoriasis and Anti-PD-1 - A New Aspect of Known Associations. *J Dtsch Dermatol Ges.* (2019) 17:186–8. doi: 10.1111/ddg.13715_g
- Yazdanpanah MJ, Banihashemi M, Pezeshkpoor F, Moradifar M, Feli S, Esmaeili H. Evaluation between Association of Psoriasis and Vitiligo. J Cutan Med Surg. (2015) 19:140–3. doi: 10.2310/7750.2014.14074
- Furue K, Ito T, Tsuji G, Kadono T, Nakahara T, Furue M. Autoimmunity and autoimmune co-morbidities in psoriasis. *Immunology*. (2018) 154:21–7. doi: 10.1111/imm.12891
- Zibert JR, Wallbrecht K, Schön M, Mir LM, Jacobsen GK, Trochon-Joseph V, et al. Halting angiogenesis by non-viral somatic gene therapy alleviates psoriasis and murine psoriasiform skin lesions. *J Clin Invest.* (2011) 121:410–21. doi: 10.1172/JCI41295
- Raychaudhuri SP, Sanyal M, Weltman H, Kundu-Raychaudhuri S. K252a, a high-affinity nerve growth factor receptor blocker, improves psoriasis: an *in vivo* study using the severe combined immunodeficient mouse-human skin model. *J Invest Dermatol.* (2004) 122:812–9. doi: 10.1111/j.0022-202X.2003.12602.x
- Raychaudhuri SK, Raychaudhuri SP. NGF and its receptor system: a new dimension in the pathogenesis of psoriasis and psoriatic arthritis. *Ann N Y Acad Sci.* (2009) 1173:470–7. doi: 10.1111/j.1749-6632.2009.04652.x
- Christophers E, van de Kerkhof PCM. Severity, heterogeneity and systemic inflammation in psoriasis. J Eur Acad Dermatol Venereol. (2019) 33:643–7. doi: 10.1111/jdv.15339
- Rocha-Pereira P, Santos-Silva A, Rebelo I, Figueiredo A, Quintanilha A, Teixeira F. The inflammatory response in mild and in severe psoriasis. Br J Dermatol. (2004) 150:917–28. doi: 10.1111/j.1365-2133.2004.05984.x
- Siegel D, Devaraj S, Mitra A, Raychaudhuri SP, Raychaudhuri SK, Jialal I. Inflammation, atherosclerosis, and psoriasis. *Clin Rev Allergy Immunol.* (2013) 44:194–204. doi: 10.1007/s12016-012-8308-0
- Gisondi P, Malerba M, Malara G, Puglisi Guerra A, Sala R, Radaeli A, et al. C-reactive protein and markers for thrombophilia in patients with chronic plaque psoriasis. *Int J Immunopathol Pharmacol.* (2010) 23:1195– 202. doi: 10.1177/039463201002300423
- Jiang S, Hinchliffe TE, Wu T. Biomarkers of An autoimmune skin disease-psoriasis. *Genomics Proteomics Bioinformatics*. (2015) 13:224–33. doi: 10.1016/j.gpb.2015.04.002
- Breuer K, Göldner FM, Jäger B, Werfel T, Schmid-Ott G. Relationship between chronic stress and CRP levels in women with psoriasis. J Dtsch Dermatol Ges. (2016) 14:528–30. doi: 10.1111/ddg.12850

- Schmuth M, Blunder S, Dubrac S, Gruber R, Moosbrugger-Martinz V. Epidermal barrier in hereditary ichthyoses, atopic dermatitis, and psoriasis. *J Dtsch Dermatol Ges.* (2015) 13:1119–23. doi: 10.1111/ddg.12827
- Jafri K, Bartels CM, Shin D, Gelfand JM, Ogdie A. Incidence and management of cardiovascular risk factors in psoriatic arthritis and rheumatoid arthritis: a population-based study. *Arthritis Care Res.* (2017) 69:51–7. doi: 10.1002/acr.23094
- Kagami S, Rizzo HL, Lee JJ, Koguchi Y, Blauvelt A. Circulating Th17, Th22, and Th1 cells are increased in psoriasis. J Invest Dermatol. (2010) 130:1373–83. doi: 10.1038/jid.2009.399
- Owczarczyk-Saczonek A, Placek W. Interleukin-17 as a factor linking the pathogenesis of psoriasis with metabolic disorders. *Int J Dermatol.* (2017) 56:260–8. doi: 10.1111/ijd.13420
- Guarneri C, Aguennouz M, Guarneri F, Polito F, Benvenga S, Cannavò SP. Autoimmunity to heterogeneous nuclear ribonucleoprotein A1 in psoriatic patients and correlation with disease severity. J Dtsch Dermatol Ges. (2018) 16:1103–7. doi: 10.1111/ddg.13631
- Johnston A, Xing X, Wolterink L, Barnes DH, Yin Z, Reingold L, et al. IL-1 and IL-36 are dominant cytokines in generalized pustular psoriasis. *J Allergy Clin Immunol.* (2017) 140:109–20. doi: 10.1016/j.jaci.2016.08.056
- Johnston A, Xing X, Guzman AM, Riblett M, Loyd CM, Ward NL, et al. IL-1F5, -F6, -F8, -F9: a novel IL-1 family signaling system that is active in psoriasis and promotes keratinocyte antimicrobial peptide expression. J Immunol. (2011) 186:2613–22. doi: 10.4049/jimmunol.1003162
- Muhr P, Zeitvogel J, Heitland I, Werfel T, Wittmann M. Expression of interleukin (IL)-1 family members upon stimulation with IL-17 differs in keratinocytes derived from patients with psoriasis and healthy donors. *Br J Dermatol.* (2011) 165:189–93. doi: 10.1111/j.1365-2133.2011.10302.x
- Lowes MA, Russell CB, Martin DA, Towne JE, Krueger JG. The IL-23/ T17 pathogenic axis in psoriasis is amplified by keratinocyte responses. *Trends Immunol.* (2013) 34:174–81. doi: 10.1016/j.it.2012.11.005
- Marrakchi S, Guigue P, Renshaw BR, Puel A, Pei XY, Fraitag S, et al. Interleukin-36-receptor antagonist deficiency and generalized pustular psoriasis. N Engl J Med. (2011) 365:620–8. doi: 10.1056/NEJMoa1013068
- Ayala-Fontanez N, Soler DC, McCormick TS. Current knowledge on psoriasis and autoimmune diseases. *Psoriasis Targets Ther.* (2016) 6:7–32. doi: 10.2147/PTT.S64950
- 95. Mössner R, Frambach Y, Wilsmann-Theis D, Löhr S, Jacobi A, Weyergraf A, et al. Palmoplantar pustular psoriasis is associated with missense variants in CARD14, but not with loss-of-function mutations in IL36RN in European patients. J Invest Dermatol. (2015) 135:2538–41. doi: 10.1038/jid.2015.186
- Sugiura K, Muto M, Akiyama M. CARD14 c.526G>C (p.Asp176His) is a significant risk factor for generalized pustular psoriasis with psoriasis vulgaris in the Japanese cohort. J Invest Dermatol. (2014) 134:1755–7. doi: 10.1038/jid.2014.46
- Wilsmann-Theis D, Schnell LM, Ralser-Isselstein V, Bieber T, Schön MP, Hüffmeier U, et al. Successful treatment with interleukin-17A antagonists of generalized pustular psoriasis in patients without IL36RN mutations. J Dermatol. (2018) 45:850–4. doi: 10.1111/1346-8138.14318
- Girolomoni G, Strohal R, Puig L, Bachelez H, Barker J, Boehncke WH, et al. The role of IL-23 and the IL-23/TH 17 immune axis in the pathogenesis and treatment of psoriasis. *J Eur Acad Dermatol Venereol.* (2017) 31:1616–26. doi: 10.1111/jdv.14433
- Poot J, Thewissen M, Booi D, Nieuwhof C. Characterization of skin-resident T cells using a crawl-out method and flow cytometry. *Exp Dermatol.* (2013) 22:554–5. doi: 10.1111/exd.12191
- 100. Steinman L. A brief history of T(H)17, the first major revision in the T(H)1/T(H)2 hypothesis of T cell-mediated tissue damage. *Nat Med.* (2007) 13:139–45. doi: 10.1038/nm1551
- 101. Di Cesare A, Di Meglio P, Nestle FO. The IL-23/Th17 axis in the immunopathogenesis of psoriasis. J Invest Dermatol. (2009) 129:1339–50. doi: 10.1038/jid.2009.59
- Lynde CW, Poulin Y, Vender R, Bourcier M, Khalil S. Interleukin 17A: toward a new understanding of psoriasis pathogenesis. J Am Acad Dermatol. (2014) 71:141–50. doi: 10.1016/j.jaad.2013.12.036
- 103. Murakami M, Hagforsen E, Morhenn V, Ishida-Yamamoto A, Iizuka H. Patients with palmoplantar pustulosis have increased IL-17 and IL-22

levels both in the lesion and serum. *Exp Dermatol.* (2011) 20:845-7. doi: 10.1111/j.1600-0625.2011.01325.x

- Neurath MF. IL-23: a master regulator in Crohn disease. Nat Med. (2007) 13:26–8. doi: 10.1038/nm0107-26
- 105. Kebir H, Kreymborg K, Ifergan I, Dodelet-Devillers A, Cayrol R, Bernard M, et al. Human TH17 lymphocytes promote blood-brain barrier disruption and central nervous system inflammation. *Nat Med.* (2007) 13:1173–5. doi: 10.1038/nm1651
- Chen X, Oppenheim JJ. Th17 cells and Tregs: unlikely allies. J Leukoc Biol. (2014) 95:723–31. doi: 10.1189/jlb.1213633
- 107. Muhr P, Renne J, Schaefer V, Werfel T, Wittmann M. Primary human keratinocytes efficiently induce IL-1-dependent IL-17 in CCR6+ T cells. *Exp Dermatol.* (2010) 19:1105–7. doi: 10.1111/j.1600-0625.2010.01134.x
- Huber S, Gagliani N, Esplugues E, O'Connor WJ, Huber FJ, Chaudhry A, et al. Th17 cells express interleukin-10 receptor and are controlled by Foxp3? and Foxp3+ regulatory CD4+ T cells in an interleukin-10-dependent manner. *Immunity.* (2011) 34:554–65. doi: 10.1016/j.immuni.2011. 01.020
- 109. Kryczek I, Bruce AT, Gudjonsson JE, Johnston A, Aphale A, Vatan L, et al. Induction of IL-17+ T cell trafficking and development by IFN-gamma: mechanism and pathological relevance in psoriasis. J Immunol. (2008) 181:4733–41. doi: 10.4049/jimmunol.181.7.4733
- Keijsers RR, Joosten I, van Erp PE, Koenen HJ, van de Kerkhof PC. Cellular sources of IL-17 in psoriasis: a paradigm shift? *Exp Dermatol.* (2014) 23:799– 803. doi: 10.1111/exd.12487
- 111. Dyring-Andersen B, Honoré TV, Madelung A, Bzorek M, Simonsen S, Clemmensen SN, et al. Interleukin (IL)-17A and IL-22-producing neutrophils in psoriatic skin. Br J Dermatol. (2017) 177:e321-e2. doi: 10.1111/bjd.15533
- 112. Schön MP. The plot thickens while the scope broadens: a holistic view on IL-17 in psoriasis and other inflammatory disorders. *Exp Dermatol.* (2017) 23:804–6. doi: 10.1111/exd.12541
- 113. Neubert E, Meyer D, Rocca F, Günay G, Kwaczala-Tessmann A, Grandke J, et al. Chromatin swelling drives neutrophil extracellular trap release. *Nat Commun.* (2018) 9:3767. doi: 10.1038/s41467-018-06263-5
- 114. Schön MP, Broekaert SM, Erpenbeck L. Sexy again: the renaissance of neutrophils in psoriasis. *Exp Dermatol.* (2017) 26:305–11. doi: 10.1111/exd.13067
- 115. Lin AM, Rubin CJ, Khandpur R, Wang JY, Riblett M, Yalavarthi S, et al. Mast cells and neutrophils release IL-17 through extracellular trap formation in psoriasis. J Immunol. (2011) 187:490–500. doi: 10.4049/jimmunol. 1100123
- Chemin K, Gerstner C, Malmström V. Effector functions of CD4⁺ T cells at the site of local autoimmune inflammation-lessons from rheumatoid arthritis. *Front Immunol.* (2019) 10:353. doi: 10.3389/fimmu.2019.00353
- 117. Wu H, Liao W, Li Q, Long H, Yin H, Zhao M, et al. Pathogenic role of tissueresident memory T cells in autoimmune diseases. *Autoimmun Rev.* (2018) 17:906–911. doi: 10.1016/j.autrev.2018.03.014
- Pan Y, Kupper TS. Metabolic reprogramming and longevity of tissue-resident memory T cells. Front Immunol. (2018) 9:1347. doi: 10.3389/fimmu.2018.01347
- 119. Cheuk S, Schlums H, Gallais Sérézal I, Martini E, Chiang SC, Marquardt N, et al. CD49a expression defines tissue-resident CD8⁺ T cells poised for cytotoxic function in human skin. *Immunity*. (2017) 46:287–300. doi: 10.1016/j.immuni.2017.01.009
- 120. McGeachy MJ, Chen Y, Tato CM, Laurence A, Joyce-Shaikh B, Blumenschein WM, et al. The interleukin 23 receptor is essential for the terminal differentiation of interleukin 17-producing effector T helper cells *in vivo*. *Nat Immunol.* (2009) 10:314–24. doi: 10.1038/ni.1698
- 121. Bettelli E, Korn T, Kuchroo VK. Th17: the third member of the effector T cell trilogy. *Curr Opin Immunol.* (2007) 19:652–7. doi: 10.1016/j.coi.2007.07.020
- 122. Stritesky GL, Yeh N, Kaplan MH. IL-23 promotes maintenance but not commitment to the Th17 lineage. J Immunol. (2008) 181:5948–55. doi: 10.4049/jimmunol.181.9.5948
- Ghoreschi K, Weigert C, Röcken M. Immunopathogenesis and role of T cells in psoriasis. *Clin Dermatol.* (2007) 25:574–80. doi: 10.1016/j.clindermatol.2007.08.012

- Boniface K, Blom B, Liu YJ, de Waal Malefyt R. From interleukin-23 to Thelper 17 cells: human T-helper cell differentiation revisited. *Immunol Rev.* (2008) 226:132–46. doi: 10.1111/j.1600-065X.2008.00714.x
- 125. Singh TP, Schön MP, Wallbrecht K, Michaelis K, Rinner B, Mayer G, et al. 8-methoxypsoralen plus ultraviolet A therapy acts via inhibition of the IL-23/Th17 axis and induction of Foxp3+ regulatory T cells involving CTLA4 signaling in a psoriasis-like skin disorder. *J Immunol.* (2010) 184:7257–67. doi: 10.4049/jimmunol.0903719
- 126. Drozdenko G, Heine G, M. W. Oral vitamin D increases the frequencies of CD38+ human B cells and ameliorates IL-17-producing T cells. *Exp Dermatol.* (2014) 23:107–12. doi: 10.1111/exd.12300
- 127. Singh TP, Schön MP, Wallbrecht K, Gruber-Wackernagel A, Wang XJ, Wolf P. Involvement of IL-9 in Th17-associated inflammation and angio- genesis of psoriasis. *PLoS ONE*. (2013) 8:e51752. doi: 10.1371/journal.pone.0051752
- Cua DJ, Tato CM. Innate IL-17-producing cells: the sentinels of the immune system. Nat Rev Immunol. (2010) 10:479–89. doi: 10.1038/nri2800
- 129. Lee JS, Tato CM, Joyce-Shaikh B, Gulen MF, Cayatte C, Chen Y, et al. Interleukin-23-independent IL-17 production regulates intestinal epithelial permeability. *Immunity.* (2015) 43:727–38. doi: 10.1016/j.immuni.2015.09.003
- 130. Yoshiga Y, Goto D, Segawa S, Ohnishi Y, Matsumoto I, Ito S, et al. Invariant NKT cells produce IL-17 through IL-23-dependent and -independent pathways with potential modulation of Th17 response in collagen-induced arthritis. *Int J Mol Med.* (2008) 22:369–74. doi: 10.3892/ijmm_00000032
- Lowes MA, Suarez-Farinas M, Krueger JG. Immunology of psoriasis. Annu Rev Immunol. (2014) 32:227–55. doi: 10.1146/annurev-immunol-032713-120225
- 132. Keijsers RR, Hendriks AGM, van Erp PE, van Cranenbroek B, van de Kerkhof PCM, Koenen HJ, et al. *In vivo* induction of cutaneous inflammation results in the accumulation of extracellular trap-forming neutrophils expressing RORγt and IL-17. *J Invest Dermatol.* (2014) 134:1276–84. doi: 10.1038/jid.2013.526
- 133. Wu CY, Lin FL. Hand-foot-and-mouth-disease-induced Koebner phenomenon in psoriasis. J Dtsch Dermatol Ges. (2019) 17:549–51. doi: 10.1111/ddg.13824
- Elsner P, Darr-Foit S, Schliemann S. Occupational koebnerization of psoriasis caused by glyphosate. J Dtsch Dermatol Ges. (2018) 16:70–1. doi: 10.1111/ddg.13393
- 135. Wohlrab J, Fiedler G, Gerdes S, Nast A, Philipp S, Radtke MA, et al. Recommendations for detection of individual risk for comorbidities in patients with psoriasis. *Arch Dermatol Res.* (2013) 305:91–8. doi: 10.1007/s00403-013-1318-9
- 136. Frohm M, Agerberth B, Ahangari G, Ståhle-Bäckdahl M, Lidén S, Wigzell H, et al. The expression of the gene coding for the antibacterial peptide LL-37 is induced in human keratinocytes during inflammatory disorders. *J Biol Chem.* (1997) 272:15258–63. doi: 10.1074/jbc.272.24.15258
- 137. Morizane S, Yamasaki K, Mühleisen B, Kotol PF, Murakami M, Aoyama Y, et al. Cathelicidin antimicrobial peptide LL-37 in psoriasis enables keratinocyte reactivity against TLR9 ligands. J Invest Dermatol. (2012) 132:135–43. doi: 10.1038/jid.2011.259
- 138. Ganguly D, Chamilos G, Lande R, Gregorio J, Meller S, Facchinetti V, et al. Self-RNA-antimicrobial peptide complexes activate human dendritic cells through TLR7 and TLR8. J Exp Med. (2009) 206:1983–94. doi: 10.1084/jem.20090480
- Döbel T, Schäkel K. The role of human 6-sulfo LacNAc dendritic cells (slanDCs) in autoimmunity and tumor diseases. *J Dtsch Dermatol Ges.* (2014) 12:874–80. doi: 10.1111/ddg.12439_suppl
- 140. Hegyi Z, Zwicker S, Bureik D, Peric M, Koglin S, Batycka-Baran A, et al. Vitamin D analog calcipotriol suppresses the Th17 cytokineinduced proinflammatory S100 "alarmins" psoriasin (S100A7) and koebnerisin (S100A15) in psoriasis. *J Invest Dermatol.* (2012) 132:1416–24. doi: 10.1038/jid.2011.486
- 141. Batycka-Baran A, Maj J, Wolf R, Szepietowski JC. The new insight into the role of antimicrobial proteins-alarmins in the immunopathogenesis of psoriasis. *J Immunol Res.* (2014) 2014:628289. doi: 10.1155/2014/ 628289
- 142. Lee Y, Jang S, Min JK, Lee K, Sohn KC, Lim JS, et al. S100A8 and S100A9 are messengers in the crosstalk between epidermis and dermis modulating

a psoriatic milieu in human skin. Biochem Biophys Res Commun. (2012) 423:647–53. doi: 10.1016/j.bbrc.2012.05.162

- 143. Loser K, Vogl T, Voskort M, Lueken A, Kupas V, Nacken W, et al. The Tolllike receptor 4 ligands Mrp8 and Mrp14 are crucial in the development of autoreactive CD8+ T cells. *Nat Med.* (2010) 16:713–7. doi: 10.1038/nm.2150
- 144. Tewary P, de la Rosa G, Sharma N, Rodriguez LG, Tarasov SG, Howard OM, et al. β-Defensin 2 and 3 promote the uptake of self or CpG DNA, enhance IFN-α production by human plasmacytoid dendritic cells, promote inflammation. J Immunol. (2013) 191:865–74. doi: 10.4049/jimmunol.1201648
- 145. Meller S, Di Domizio J, Voo KS, Friedrich HC, Chamilos G, Ganguly D, et al. T(H)17 cells promote microbial killing and innate immune sensing of DNA via interleukin 26. *Nat Immunol.* (2015) 16:970–9. doi: 10.1038/ni.3211
- 146. Wu JJ, Nguyen TU, Poon KY, Herrinton LJ. The association of psoriasis with autoimmune diseases. J Am Acad Dermatol. (2012) 67:924–30. doi: 10.1016/j.jaad.2012.04.039
- 147. Hsu LN, Armstrong AW. Psoriasis and autoimmune disorders: a review of the literature. J Am Acad Dermatol. (2012) 67:1076–9. doi: 10.1016/j.jaad.2012.01.029
- 148. Claus S, Ziemer M, Simon JC, Treudler R. Coincidence of annular pustular psoriasis, pemphigus foliaceus, and leukocytoclastic vasculitis associated with chronic cholecystitis. J Dtsch Dermatol Ges. (2016) 14:830–1. doi: 10.1111/ddg.12619
- 149. Makredes M, Robinson DJ, Bala M, Kimball AB. The burden of autoimmune disease: a comparison of prevalence ratios in patients with psoriatic arthritis and psoriasis. J Am Acad Dermatol. (2009) 61:405–10. doi: 10.1016/j.jaad.2009.02.015
- 150. Sun H, Xia Y, Wang L, Wang Y, Chang X. PSORS1C1 may be involved in rheumatoid arthritis. *Immunol Lett.* (2013) 153:9–14. doi: 10.1016/j.imlet.2013.06.001
- Alarcon-Riquelme ME. Role of RUNX in autoimmune diseases linking rheumatoid arthritis, psoriasis and lupus. *Arthritis Res Ther.* 204;6:169–73. doi: 10.1186/ar1203
- Okuda T, Nishimura M, Nakao M, Fujita Y. RUNX1/AML1: a central player in hematopoiesis. *Int J Hematol.* (2001) 74:252–7. doi: 10.1007/BF02982057
- 153. Helms C, Cao L, Krueger JG, Wijsman EM, Chamian F, Gordon D, et al. A putative RUNX1 binding site variant between SLC9A3R1 and NAT9 is associated with susceptibility to psoriasis. *Nat Genet.* (2003) 35:349–56. doi: 10.1038/ng1268
- 154. Gregersen PK, Olsson LM. Recent advances in the genetics of autoimmune disease. Annu Rev Immunol. (2009) 27:363–91. doi: 10.1146/annurev.immunol.021908.132653
- 155. Vereecke L, Beyaert R, van Loo G. The ubiquitin-editing enzyme A20 (TNFAIP3) is a central regulator of immunopathology. *Trends Immunol.* (2009) 30:383–91. doi: 10.1016/j.it.2009.05.007
- 156. Wang F, Zhang X, Xia P, Zhang L, Zhang Z. Upregulation of tumor necrosis factor alpha-induced protein 3 interacting protein 1 mRNA in psoriasis vulgaris. J Dtsch Dermatol Ges. (2015) 13:159–61. doi: 10.1111/ddg.12462
- 157. Sugiyama H, Gyulai R, Toichi E, Garaczi E, Shimada S, Stevens SR, et al. Dysfunctional blood and target tissue CD4+CD25high regulatory T cells in psoriasis: mechanism underlying unrestrained pathogenic effector T cell proliferation. *J Immunol.* (2005) 174:164–73. doi: 10.4049/jimmunol.174.1.164
- 158. Hueber W, Patel DD, Dryja T, Wright AM, Koroleva I, Bruin G, et al. Effects of AIN457, a fully human antibody to interleukin-17A, on psoriasis, rheumatoid arthritis, and uveitis. *Sci Transl Med.* (2010) 2:52ra72. doi: 10.1126/scitranslmed.3001107
- McInnes IB, Schett G. The pathogenesis of rheumatoid arthritis. N Engl J Med. (2011) 365:2205–19. doi: 10.1056/NEJMra1004965
- 160. Yates VM, Watkinson G, Kelman A. Further evidence for an associa- tion between psoriasis, Crohn's disease and ulcerative colitis. *Br J Dermatol.* (1982) 106:323–30. doi: 10.1111/j.1365-2133.1982.tb01731.x
- 161. Bernstein CN, Wajda A, Blanchard JF. The clustering of other chronic inflammatory diseases in inflammatory bowel disease: a population- based study. *Gastroenterology*. (2005) 129:827–36. doi: 10.1053/j.gastro.2005.06.021
- 162. Weng X, Liu L, Barcellos LF, Allison JE, Herrinton LJ. Clustering of inflammatory bowel disease with immune mediated diseases among

members of a northern california-managed care organization. *Am J Gastroenterol.* (2007) 102:1429–35. doi: 10.1111/j.1572-0241.2007.01215.x

- 163. Lee FI, Bellary SV, Francis C. Increased occurrence of psoriasis in patients with Crohn's disease and their relatives. *Am J Gastroenterol.* (1990) 85:962–3.
- 164. Scarpa R, Manguso F, D'Arienzo A, D'Armiento FP, Astarita C, Mazzacca G, et al. Microscopic inflammatory changes in colon of patients with both active psoriasis and psoriatic arthritis without bowel symptoms. *J Rheumatol.* (2000) 27:1241–6.
- 165. Ellinghaus D, Ellinghaus E, Nair RP, Stuart PE, Esko T, Metspalu A, et al. Combined analysis of genome-wide association studies for Crohn disease and psoriasis identifies seven shared susceptibility loci. Am J Hum Genet. (2012) 90:636–47. doi: 10.1016/j.ajhg.2012.02.020
- 166. Ferguson LR, Han DY, Fraser AG, Huebner C, Lam WJ, Morgan AR. IL23R and IL12B SNPs and haplotypes strongly associate with Crohn's disease risk in a New Zealand population. *Gastroenterol Res Pract.* (2010) 2010:539461. doi: 10.1155/2010/539461
- 167. Cargill M, Schrodi SJ, Chang M, Garcia VE, Brandon R, Callis KP, et al. A large-scale genetic association study confirms IL12B and leads to the identification of IL23R as psoriasis-risk genes. *Am J Hum Genet.* (2007) 80:273–90. doi: 10.1086/511051
- 168. Nair RP, Ruether A, Stuart PE, Jenisch S, Tejasvi T, Hiremagalore R, et al. Polymorphisms of the IL12B and IL23R genes are associated with psoriasis. *J Invest Dermatol.* (2008) 128:1653–61. doi: 10.1038/sj.jid.5701255
- 169. Genetic Analysis of Psoriasis Consortium & the Wellcome Trust Case Control Consortium 2, Strange A, Capon F, Spencer CC, Knight J, Weale ME, et al. A genome-wide association study identifies new psoriasis susceptibility loci and an interaction between HLA-C and ERAP1. *Nat Genet.* (2010) 42:985–90. doi: 10.1038/ng.694
- Braegger CP, Nicholls S, Murch SH, Stephens S, MacDonald TT. Tumour necrosis factor alpha in stool as a marker of intestinal inflammation. *Lancet.* (1992) 339:89–91. doi: 10.1016/0140-6736(92)90999-J
- 171. Murch SH, Braegger CP, Walker-Smith JA, MacDonald TT. Location of tumour necrosis factor alpha by immunohistochemistry in chronic inflammatory bowel disease. *Gut.* (1993) 34:1705–9. doi: 10.1136/gut.34.12.1705
- 172. Oh CJ, Das KM, Gottlieb AB. Treatment with anti-tumor necrosis factor alpha (TNF-alpha) monoclonal antibody dramatically decreases the clinical activity of psoriasis lesions. J Am Acad Dermatol. (2000) 42(5 pt 1):829–30. doi: 10.1067/mjd.2000.105948
- 173. Chaudhari U, Romano P, Mulcahy LD, Dooley LT, Baker DG, Gottlieb AB. Efficacy and safety of infliximab monotherapy for plaque-type psoriasis: a randomised trial. *Lancet.* (2001) 357:1842–7. doi: 10.1016/S0140-6736(00)04954-0
- 174. Plevy SE, Landers CJ, Prehn J, Carramanzana NM, Deem RL, Shealy D, et al. A role for TNF-alpha and mucosal T helper-1 cytokines in the pathogenesis of Crohn's disease. *J Immunol.* (1997) 159:6276–82.
- 175. Targan SR, Hanauer SB, van Deventer SJ, Mayer L, Present DH, Braakman T, et al. A short-term study of chimeric monoclonal antibody cA2 to tumor necrosis factor alpha for Crohn's disease. Crohn's Disease cA2 Study Group. N Engl J Med. (1997) 337:1029–35. doi: 10.1056/NEJM199710093371502
- Amschler K, Broekaert SMC, Mohr J, Schön MP, Mössner R. Persistent neutrophilic scarring alopecia triggered by anti-TNF blockade for Crohn's disease. *Eur J Dermatol.* (2018) 28:403–5. doi: 10.1684/ejd.2018.3270
- 177. Angelucci E, Cocco A, Viscido A, Vernia P, Caprilli R. Another paradox in Crohn's disease: new onset of psoriasis in a patient receiving tumor necrosis factor-alpha antagonist. *Inflamm Bowel Dis.* (2007) 13:1059–61. doi: 10.1002/ibd.20137
- 178. Cottone M, Sapienza C, Macaluso FS, Cannizzaro M. Psoriasis and Inflammatory Bowel Disease. *Dig Dis.* (2019) 10:1–7. doi: 10.1159/000500116
- 179. Fiorino G, Allez M, Malesci A, Danese S. Review article: anti TNF- alpha induced psoriasis in patients with inflammatory bowel disease. *Aliment Pharmacol Ther.* (2009) 29:921–7. doi: 10.1111/j.1365-2036.2009.03955.x
- 180. Mylonas A, Conrad C. Psoriasis: classical vs. paradoxical. The Yin-Yang of TNF and type I interferon. *Front Immunol.* (2018) 9:2746. doi: 10.3389/fimmu.2018.02746
- Lindhaus C, Tittelbach J, Elsner P. Cutaneous side effects of TNF-alpha inhibitors. J Dtsch Dermatol Ges. (2017) 15:281–8. doi: 10.1111/ddg.13200

- 182. Fujimoto N, Matsuo S, Satoh T. Psoriasis vulgaris in a patient with pemphigus vulgaris on corticosteroid therapy. J Dtsch Dermatol Ges. (2018) 16:606–8. doi: 10.1111/ddg.13505
- Amschler K, Meyersburg D, Kitze B, Schön MP, Mössner R. Onset of psoriasis upon interferon beta treatment in a multiple sclerosis patient. *Eur J Dermatol.* (2016) 26:211–2. doi: 10.1684/ejd.2015.2602
- Baumgart DC, Sandborn WJ. Crohn's disease. Lancet. (2012) 380:1590–605. doi: 10.1016/S0140-6736(12)60026-9
- 185. Brand S. Crohn's disease: Th1, Th17 or both? The change of a paradigm: new immunological and genetic insights implicate Th17 cells in the pathogenesis of Crohn's disease. *Gut.* (2009) 58:1152–67. doi: 10.1136/gut.2008.163667
- 186. Eastaff-Leung N, Mabarrack N, Barbour A, Cummins A, Barry S. Foxp3+ regulatory T cells, Th17 effector cells, and cytokine environment in inflammatory bowel disease. J Clin Immunol. (2010) 30:80–9. doi: 10.1007/s10875-009-9345-1
- Galvez J. Role of Th17 cells in the pathogenesis of human IBD. ISRN Inflamm. (2014) 2014:928461. doi: 10.1155/2014/928461
- Hovhannisyan Z, Treatman J, Littman DR, Mayer L. Characterization of interleukin-17-producing regulatory T cells in inflamed intestinal mucosa from patients with inflammatory bowel diseases. *Gastroenterology*. (2011) 140:957–65. doi: 10.1053/j.gastro.2010.12.002
- 189. Bovenschen HJ, van de Kerkhof PC, van Erp PE, Woestenenk R, Joosten I, Koenen HJ. Foxp3+ regulatory T cells of psoriasis patients easily differentiate into IL-17A-producing cells and are found in lesional skin. *J Invest Dermatol.* (2011) 131:1853–60. doi: 10.1038/jid.2011.139
- 190. O'Connor WJ, Kamanaka M, Booth CJ, Town T, Nakae S, Iwakura Y, et al. A protective function for interleukin 17A in T cell-mediated intestinal inflammation. *Nat Immunol.* (2009) 10:603–9. doi: 10.1038/ni.1736
- 191. Hohenberger M, Cardwell LA, Oussedik E, Feldman SR. Interleukin-17 inhibition: role in psoriasis and inflammatory bowel disease. J Dermatolog Treat. (2018) 29:13–8. doi: 10.1080/09546634.2017.1329511
- Goodman WA, Levine AD, Massari JV, Sugiyama H, McCormick TS, Cooper KD. IL-6 signaling in psoriasis prevents immune suppression by regulatory T cells. *J Immunol.* (2009) 183:3170–6. doi: 10.4049/jimmunol.0803721
- 193. Atreya R, Mudter J, Finotto S, Müllberg J, Jostock T, Wirtz S, et al. Blockade of interleukin 6 trans signaling suppresses T-cell resistance against apoptosis in chronic intestinal inflammation: evidence in crohn disease and experimental colitis *in vivo*. *Nat Med.* (2000) 6:583–8. doi: 10.1038/75068
- 194. Goodman WA, Young AB, McCormick TS, Cooper KD, Levine AD. Stat3 phosphorylation mediates resistance of primary human T cells to regulatory T cell suppression. *J Immunol.* (2011) 186:3336–45. doi: 10.4049/jimmunol.1001455
- 195. Sano S, Chan KS, DiGiovanni J. Impact of Stat3 activation upon skin biology: a dichotomy of its role between homeostasis and diseases. J Dermatol Sci. (2008) 50:1–14. doi: 10.1016/j.jdermsci.2007.05.016
- 196. Janjumratsang P, Phainupong D, Chanjanakijskul S, Roongphibulsopit P. Positive direct immunofluorescence and autoantibody profiles in psoriasis patients. *J Dermatol.* (2008) 35:508–13. doi: 10.1111/j.1346-8138.2008.00512.x
- 197. Dubois EL, Wierzchowiecki M, Cox MB, Weiner JM. Duration and death in systemic lupus erythematosus. An analysis of 249 cases. JAMA. (1979) 227:1399–402. doi: 10.1001/jama.227.12.1399
- 198. Tokuhiro S, Yamada R, Chang X, Suzuki A, Kochi Y, Sawada T, et al. An intronic SNP in a RUNX1 binding site of SLC22A4, encoding an organic cation transporter, is associated with rheumatoid arthritis. *Nat Genet.* (2003) 35:341–8. doi: 10.1038/ng1267
- 199. Prokunina L, Castillejo-López C, Oberg F, Gunnarsson I, Berg L, Magnusson V, et al. A regulatory polymorphism in PDCD1 is associated with susceptibility to systemic lupus erythematosus in humans. *Nat Genet.* (2002) 32:666–9. doi: 10.1038/ng1020
- 200. Perricone C, Ciccacci C, Ceccarelli F, Di Fusco D, Spinelli FR, Cipriano E, et al. TRAF3IP2 gene and systemic lupus erythematosus: association with disease susceptibility and pericarditis development. *Immunogenetics*. (2013) 65:703–9. doi: 10.1007/s00251-013-0717-6
- 201. Hüffmeier U, Uebe S, Ekici AB, Bowes J, Giardina E, Korendowych E, et al. Common variants at TRAF3IP2 are associated with susceptibility to psoriatic arthritis and psoriasis. *Nat Genet.* (2010) 42:996–9. doi: 10.1038/ng.688

- Wu L, Wang C, Boisson B, Misra S, Rayman P, Finke JH, et al. The differential regulation of human ACT1 isoforms by Hsp90 in IL-17 signaling. *J Immunol.* (2014) 193:1590–9. doi: 10.4049/jimmunol.1400715
- 203. Chen XQ, Yu YC, Deng HH, Sun JZ, Dai Z, Wu YW, et al. Plasma IL-17A is increased in new-onset SLE patients and associated with disease activity. J *Clin Immunol.* (2010) 30:221–5. doi: 10.1007/s10875-009-9365-x
- 204. Wen Z, Xu L, Xu W, Yin Z, Gao X, Xiong S. Interleukin-17 expression positively correlates with disease severity of lupus nephritis by increasing anti-double-stranded DNA antibody production in a lupus model induced by activated lymphocyte derived DNA. *PLoS ONE.* (2013) 8:e58161. doi: 10.1371/journal.pone.0058161
- 205. Yu B, Guan M, Peng Y, Shao Y, Zhang C, Yue X, et al. Copy number variations of interleukin-17F, interleukin-21, and interleukin-22 are associated with systemic lupus erythematosus. *Arthritis Rheum*. (2011) 63:3487–92. doi: 10.1002/art.30595
- 206. Barreto M, Ferreira RC, Lourenço L, Moraes-Fontes MF, Santos E, Alves M, et al. Low frequency of CD4+CD25+ Treg in SLE patients: a heritabletrait

associated with CTLA4 and TGFbeta gene variants. *BMC Immunol.* (2009) 10:5. doi: 10.1186/1471-2172-10-5

207. Franz B, Fritzsching B, Riehl A, Oberle N, Klemke CD, Sykora J, et al. Low number of regulatory T cells in skin lesions of patients with cutaneous lupus erythematosus. *Arthritis Rheum.* (2007) 56:1910–20. doi: 10.1002/art. 22699

Conflict of Interest Statement: The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Schön. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.