REVIEW

Addressing the Soy and Breast Cancer Relationship: Review, Commentary, and Workshop Proceedings

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The impact of soyfood intake on breast cancer risk has been investigated extensively. Much of this focus can be attributed to the soybean being a dietary source that is uniquely rich in isoflavones. The chemical structure of isoflavones is similar to that of estrogen, and isoflavones bind to both estrogen receptors (ERa and ERB) (although they preferentially bind to and activate ERB) and exert estrogen-like effects under some experimental conditions. Isoflavones also possess nonhormonal properties that are associated with the inhibition of cancer cell growth. Thus, there are several possible mechanisms by which soy may reduce the risk of breast cancer. However, the role of isoflavones in breast cancer has become controversial because, in contrast to the possible beneficial effects, some data from in vitro and animal studies suggest that isoflavones, especially genistein, the aglycone of the main soybean isoflavone genistin, may stimulate the growth of estrogen-sensitive tumors. Limited human data directly address the tumor-promoting effects of isoflavones and soy. Because the use of soyfoods and isoflavone supplements is increasing, it is important from a public health perspective to understand the impact of these products on breast cancer risk in women at high risk of the disease and on the survival of breast cancer patients. To this end, a workshop was held in November 2005 to review the existing literature and to make research recommendations. This paper summarizes the workshop findings and recommendations. The primary research recommendation is that the impact of isoflavones on breast tissue needs to be evaluated at the cellular level in women at high risk for breast cancer. [J Natl Cancer Inst 2006;98:1275-84]

The possibility that soyfoods reduce breast cancer risk first attracted widespread attention in 1990, when participants at a workshop sponsored by the National Cancer Institute concluded that there were several putative chemopreventive agents in soybeans and recommended funding research in this area (1). Among the various purported soybean chemopreventive agents, isoflavones have received the most attention; approximately 9000 papers have been published on these soybean constituents, about 20% of which involve cancer investigations. Although it is now recognized that the physiologic properties of isoflavones make them potentially applicable as chemopreventive agents for many types of cancer (2), most of the initial focus was on breast cancer (3). This potential role in breast cancer can be attributed to the historically low breast cancer incidence rates in Asia (4), where diets are rich in soyfoods; research demonstrating the potential for isoflavones—which have a similar chemical structure to the hormone estrogen-to exert antiestrogenic effects (5); and early epidemiologic (6) and rodent (7) studies showing associations between soy intake and reduced risk of breast and mammary cancer, respectively.

Despite the impressive amount of research conducted during the past 15 years, no clear consensus has emerged regarding the breast cancer preventive aspects of isoflavones. Although the limited epidemiologic data modestly support an inverse association between soy intake and breast cancer risk, many of the casecontrol and prospective studies have important limitations (8,9). Study limitations include the usual issues of sample size, dietary measurement error, and whether a study was specifically and appropriately designed to address the soy-breast cancer hypothesis. Comparison across studies is complicated by the variation in exposure measures used (e.g., intake of soy protein, soyfoods, or isoflavones; urinary or serum isoflavone levels) and the variation in amount and types of soy products consumed (9).

Rodent studies have shown that when isoflavones or soy protein are given before the administration of chemical carcinogens (10-13) or the implantation of cancer cells (14-16), mammary tumor development and/or growth is generally inhibited, although there are several exceptions (17-20). Furthermore, as discussed later, the timing of soy or isoflavone exposure relative to the implantation of cancer cells or the administration of carcinogens may be a critical factor in determining whether tumor development and growth is suppressed or enhanced in rodent models (1). In any event, there is little clinical evidence from largely short-term studies that soy or isoflavones favorably affect markers of breast cancer risk, including breast tissue density (21,22), serum estrogen concentrations (23,24), and breast cell proliferation (25). Some studies have found that, with high soy intake, estrogen metabolism is favorably altered (26) and menstrual cycle length increased (23); however, the impact of these changes on breast cancer risk is uncertain.

Considerable enthusiasm remains for the possibility that soyfood intake contributes to the low breast cancer rate in Asia but increasingly it appears that childhood and/or adolescence is the critical period of exposure. This hypothesis, which is supported by both epidemiologic (27-29) and animal (30,31) data, is

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See "Notes" following "References."

DOI: 10.1093/jnci/djj356 © 2006 The Author(s).

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consistent with the mounting evidence that early life events greatly impact breast cancer risk (32). However, it should be noted that although there is some evidence in rodents that in utero isoflavone exposure affects adult gene expression (33,34), there is no evidence that such exposure reduces mammary cancer risk (30,33–36). For example, in utero genistein exposure shifted the coat color of heterozygous viable yellow agouti offspring toward pseudoagouti (33) and the feeding of genistein and daidzein to Wistar-Kyoto rat dams during gestation caused the cardiac myocytes of their adult offspring to be shorter than in counterparts originating from mothers fed with a phytoestrogen-free caseinbased diet (34).

In addition to the potential protective effects, some data suggest that isoflavones could promote breast cancer. In vitro, genistein stimulates the growth of estrogen-sensitive mammary cancer cells (37), and in ovariectomized athymic mice, dietary genistein (37) and genistin (38) stimulate the growth of existing estrogensensitive mammary tumors. Consequently, in recent years, the estrogen-like effects of isoflavones have raised concerns that soyfoods are contraindicated for women at high risk of breast cancer and breast cancer patients with estrogen-sensitive tumors—approximately two-thirds of women with postmenopausal breast cancer are in this category (39). Numerous review articles and commentaries have been published on this topic (40–44).

Establishing the impact of soy intake on women at high risk for breast cancer and breast cancer patients is clearly an important public health goal. Many women at high risk underestimate their chance of getting breast cancer, and many at low risk overestimate it (45). Thus, it is possible that many women will either unnecessarily avoid soy or consume it when perhaps they should not. Moreover, many breast cancer patients consume soy and often list "anticancer effects" as a reason for doing so (46). Soyfoods, because of their purported health benefits, have become increasingly popular among non-Asians although partly because much of the soy protein added to traditional Western foods has a reduced isoflavone content, daily per capita isoflavone intake is quite low in the United States (47–49) and in Europe (50,51) typically less than 3 mg. It is however, much higher among health-conscious individuals and vegetarians (52,53).

To gain an understanding of the current state of knowledge regarding the safety of soy for breast cancer patients and for women at high risk for breast cancer and to identify research initiatives with the potential to resolve this controversy, a workshop was held on November 3, 2005. This meeting was organized by one of the authors (M. Messina) and was funded by the United Soybean Board. With one exception (W. McCaskill-Stevens), each of the 13 workshop participants received an honorarium for their participation, and all were given an opportunity to review the manuscript.

This review has three parts. First, a background section on isoflavones is provided. Next, the workshop proceedings are presented. Finally, an outline of the conclusions and the recommendations of the workshop are presented.

BACKGROUND

Isoflavones are a subclass of flavonoids that have a limited distribution in nature; among commonly consumed foods, they are found in dietary-relevant amounts only in the soybean (54). The three soybean isoflavone aglycones—genistein, daidzein, and

glycitein-are present in raw soybeans and in nonfermented soyfoods almost entirely as β -glycosides (genistin, daidzin, and glycitin), to which either an acetyl or malonyl group is attached (55) The biologically active form of isoflavones is the aglycone, but during digestion the glycoside is efficiently hydrolyzed such that the form in which isoflavones are ingested does not appear to markedly impact overall absorption and the resulting biologic effects (56). Genistein-genistin, daidzein-daidzin, and glyciteinglycitin make up approximately 50%, 40%, and 10%, respectively, of the total isoflavone content of the soybean. Each gram of soy protein in soybeans and in traditional Asian soyfoods contains approximately 3.5 mg of isoflavones (isoflavone weight throughout this paper refers to the aglycone weight) (55). However, processing reduces the isoflavone content of some soy protein products by as much as 80% (55). The daily isoflavone intake from traditional soyfoods of older Japanese adults ranges from 25 to 50 mg (57–60).

In response to the consumption of dietary amounts of isoflavones provided by soyfoods or extracts, postprandial isoflavone levels can reach the low micromolar range; however, at least 95% of the isoflavones in serum are conjugated and thought to be largely biologically inactive (56). Although isoflavones are extensively conjugated in both rats and humans, a higher percentage of both genistein and daidzein appear in the free or aglycone form in rats (61). Isoflavones have short half-lives (approximately 8 hours), and nearly all are excreted within 24 hours after ingestion (56). There is considerable interindividual variation in gut bacterial metabolism of genistein and daidzein (62-64), which leads to markedly different serum and urinary concentrations of the isoflavones and their metabolites in different individuals (62-64). This variation, coupled with differences in biologic activity among the isoflavonoids, has been offered as a possible explanation for the often inconsistent results from clinical trials (63). The varied chemical composition of the many soy products used in these trials further complicates interpretation of the literature (65).

Isoflavones bind to both estrogen receptors (ER α and ER β) and exert some estrogen-like effects in vitro (66,67). However, in clinical studies, estrogen-like effects are often not observed (68–70). This discrepancy is not surprising because ER binding alone is a poor predictor of in vivo activity (71). ER-binding ligands often have very different and sometimes opposite physiologic effects, depending upon the manner in which the ligandreceptor complex interacts with coactivators and corepressors within the cell (72-74). Isoflavones have traditionally been considered weakly estrogenic, having 10⁻⁵-10⁻² less activity per mole than 17β-estradiol (75-77). However, in some in vitro systems, genistein and daidzein and their metabolites exert effects even greater than those of estradiol (78,79). Isoflavones are not unique in this regard; this phenomenon has been demonstrated for a number of other compounds, including resveratrol (79,80). It is difficult to accurately estimate the relative estrogenic activity of ER-binding ligands because it depends on many factors, including the dose and the type of tissue used in the study.

Although isoflavones can bind to both ER α and ER β , they preferentially bind to and activate ER β (81–83); for this reason, they are sometimes classified as selective estrogen receptor modulators (SERMs) (69,84,85). The selectivity of isoflavones may depend in part on the relative tissue distribution of the two ERs. However, isoflavones also possess a variety of nonhormonal properties; thus, classifying isoflavones as phyto-SERMs does not capture all of their properties (2,86,87). The preferential binding of isoflavones to ER β may have implications related to breast cancer risk; some data suggest that, when activated by certain ligands, ER β inhibits mammary cancer cell growth as well as the stimulatory effects of ER α (88). But the precise role of ER β activation in breast cancer is unclear (89).

In vitro, the isoflavone genistein inhibits the growth of most types of cancer cells, including both hormone-dependent and -independent breast cancer cells, through a variety of mechanisms (2,90,91). However, its effect on the growth of ER-positive (+) cells is biphasic (37,92–94). Genistein inhibits the growth of MCF-7 cells at higher (>10 µM) concentrations, whereas it stimulates growth at relatively low and physiologically relevant concentrations ($<1 \mu$ M). Current thinking is that growth stimulation and inhibition by isoflavonoids occur through estrogen-dependent (95) and -independent mechanisms, respectively (96). Genistein's estrogen-independent mechanisms include modulating genes that are related to the control of cell cycle and apoptosis, inhibiting the activation of nuclear factor-kB and Akt signaling pathways and inhibiting the activity of several enzymes and growth factors that control growth and differentiation (97-100). Furthermore, isoflavones have antioxidant activity (101) and may stimulate the immune system (102-104) and inhibit angiogenesis (105).

The clinical relevance of the in vitro data is a matter of considerable debate. A potentially important consideration is the extent to which the addition of physiologic levels of estrogen to culture medium affects the cancer cell growth-stimulatory effects of genistein. Some studies show that, in a high-estrogenic environment, genistein does not stimulate growth and can inhibit it (106), whereas others show a modest increase in growth with genistein (14,107–109). The hormonal milieu may also be an important factor determining the in vivo effects of isoflavones.

The in vitro growth-stimulatory effects of genistein were not fully appreciated (perhaps because they conflicted with the prevailing hypothesis) until dietary genistin was shown to stimulate the growth of existing estrogen-sensitive tumors in athymic ovariectomized mice (37). Even before this finding was published however, observations in humans suggested that soyfoods had the potential to exert estrogen-like effects on breast tissue. In a pilot study, premenopausal but not postmenopausal women who consumed isoflavone-rich isolated soy protein (which is ≥90% protein) had a two- to sixfold increase in nipple aspirate fluid volume compared with those who did not (110). Of greater concern was that epithelial hyperplasia was detected in seven of 24 postmenopausal women when they consumed soy. However, one important limitation of this study was the lack of a control group. Research published 3 years later by a different group showed that the consumption of textured vegetable (i.e., soy) protein for 2 weeks resulted in increased pS2 levels in breast biopsies taken from premenopausal women (111). The pS2 protein is expressed in response to estrogen (112); its activation in breast tissue in response to textured vegetable protein suggests that constituents of the intervention product are eliciting an ERmediated response (113). However, because of the short duration of this study and because breast cell proliferation was not increased, in contrast to the findings from a subset of this cohort that were published 1 year earlier (114), the authors of this study concluded that the long-term implications of these findings were unclear (111). Nevertheless, these latter two studies (110,111) suggested that soy has the potential to increase breast cancer risk and provided at least some of the impetus for further work in this area, the results of which form part of the discussion of the workshop proceedings.

WORKSHOP PROCEEDINGS

Epidemiologic Studies

Not surprisingly, concern over the effects of isoflavones on breast cancer risk is based in part on the role of estrogen in the etiology of the disease and on data suggesting that conventional hormone therapy increases risk of the disease (115). These data were briefly reviewed by W. McCaskill-Stevens (National Cancer Institute, Bethesda, MD), who noted that, in the Women's Health Initiative (WHI), risk of breast cancer was increased by 26% in response to the combination of estrogen plus progestin (116), whereas it was decreased by 23% in response to estrogen alone (117). In the Million Women Study, both treatments increased risk, but the risk associated with the use of combination hormones (odds ratio [OR] = 2.00, 95% confidence interval [CI] = 1.88 to 2.12) was higher than that of estrogen alone (OR = 1.30, 95% CI = 1.21 to 1.41) (P<.001) (118). The differing effects of estrogen and estrogen plus progestin on breast cancer risk, as highlighted by these studies and for which there are considerable supporting data (119,120), are noteworthy because isoflavones do not demonstrate progestin activity in vitro (121).

Both the WHI and the Million Women Study addressed the risk for generally healthy postmenopausal women to develop breast cancer, not on the impact of hormone therapy on the prognosis of breast cancer patients. In a comprehensive review, Creasman (122) recently concluded that there is relatively little evidence that conventional hormone therapy is contraindicated for breast cancer patients, although this issue remains highly controversial (123). In agreement with this conclusion, Durna et al. (124) found in a small study involving premenopausal breast cancer patients that hormone therapy use after diagnosis of breast cancer was not associated with increased breast cancer recurrence or mortality. However, there may be little chance of obtaining substantially more insight on this topic because the oncology community in general advises their ER+ breast cancer patients not to use postmenopausal hormones.

The impact of soy intake on recurrence or survival of breast cancer patients can be evaluated in an epidemiologic setting. In this regard, Anna Wu (University of Southern California, Los Angeles, CA) presented the experimental design for her ongoing investigation of the effects of lifestyle factors on breast cancer prognosis among Asian-Americans. A total of 1378 case patients are in the study, including 489 Chinese, 383 Japanese, and 506 Filipino women, all of whom reside in the Los Angeles area. Data are being collected about initial treatment and tumor characteristicstumor stage and size, lymph node status, extent of disease, histology, differentiation, grade, laterality, and estrogen-progesterone receptor status-as provided by the Los Angeles County Cancer Surveillance Program (member of the Surveillance End Results Program). Telephone interviews are being conducted 5 years after initial diagnosis of breast cancer to determine lifestyle characteristics, including body weight, physical activity, herbal and vitamin supplement use, and dietary pattern (main food groups include soy, tea, fruits and vegetables, red meat, white meat, fish, and alcohol). Postdiagnostic follow-up data also being collected include the number and type of breast surgeries and use of tamoxifen, raloxifene, aromatase inhibitors, herceptin, chemotherapy,

radiation, and other conventional treatments, as well as the use of alternative or complementary therapy. Data collection will be completed in 2008.

Relevant to this study are recently published findings from a prospective epidemiologic study conducted in Shanghai involving 1459 Chinese breast cancer patients (125). During the approximately 5-year follow-up period, 240 deaths occurred, but there was no association between the intake of soy protein or isoflavones before diagnosis of breast cancer and disease-free survival. The relationship between soy protein intake and breast cancer survival did not differ according to estrogen-progesterone receptor status, tumor stage, age at diagnosis, body mass index, waist-to-hip ratio, or menopausal status. Also, the results were not affected when the analysis was restricted to only women with ER+ tumors (63% of the total). One limitation of this study is that soy intake was determined only at baseline; however, when the analysis was restricted to only women who reported "no dietary change" during the follow-up period, the results were similar to findings involving all women.

Animal Studies

It was research demonstrating that dietary genistein (37) and genistin (38) stimulate the growth of estrogen-sensitive mammary tumors in rodents that first raised concern that isoflavones might be contraindicated for breast cancer patients. This research was conducted by William Helferich and colleagues at the University of Illinois, who used a model in which athymic BALB/c (nude) ovariectomized mice are subcutaneously injected with MCF-7 cells and implanted with estrogen pellets to stimulate estrogen-dependent tumor growth. Once tumors have reached a cross-sectional area of approximately 40 mm², the estrogen pellets are removed from all groups except the positive control. In mice that are fed the standard AIN-93G diet, tumors regress completely; however, diets containing either isoflavone-rich isolated soy protein (126) or isoflavone extracts (127) stimulate tumor growth in the mice. Furthermore, in this model, dietary genistein negates the ability of tamoxifen to inhibit tumor growth (128). In a recent publication from this group (129), dietary genistein stimulated estrogen-dependent tumor growth in athymic BALB/c ovariectomized mice implanted with silastic implants containing low levels of estradiol that produced plasma estradiol concentrations similar to those found in postmenopausal women. These data indicate that genistein can act as an ER agonist and can stimulate estrogen-dependent tumor growth in vivo.

A potentially crucial observation from Helferich and colleagues is that mice exposed to more processed soy products had faster tumor growth than mice exposed to less processed soy products even if the amount of genistein in both products was the same (127). A diet containing soy flour, which is minimally processed, did not promote tumor growth, although tumors did not regress to the extent that they did with the control diet lacking soy. The mechanism behind this processing effect is unclear, although two explanations have been proposed: one is that processing causes greater increases in serum levels of free genistein (130) and the other is that compounds removed during processing inhibit the tumor-stimulatory effects of isoflavones and/or directly inhibit mammary tumor growth (131). In contrast to genistein, daidzein only modestly stimulated the growth of MCF-7 cells in this mouse model. Moreover, tumor growth was not at all stimulated by equol, a bacterially derived metabolite

of daidzein that stimulates MCF-7 cell proliferation in vitro (132).

In a similar model to that used by Helferich's group (126,127), Lilian Thompson (University of Toronto) also observed tumorstimulatory effects of isoflavone-rich isolated soy protein although they were not as pronounced (133). According to Thompson, tumors initially regressed with dietary soy protein to the same extent as with the control diet, which did not contain soy, but after 10-12 weeks soy protein stimulated tumor regrowth. In contrast, flaxseed, a rich source of lignans, which like isoflavones are also diphenolic compounds classified as phytoestrogens, did not alter tumor growth from that in response to the control diet alone, i.e., tumor regression occurred to a similar extent. Furthermore, the addition of flaxseed to the isoflavone-rich isolated soy protein-containing diet caused tumor regression that was similar to the regression that occurred in response to the control diet. Thus, flaxseed inhibited the tumor-stimulatory effects of soy protein. In comparison to isoflavone-rich isolated soy protein alone, the addition of flaxseed caused a decrease in tumor cell proliferation and an increase in tumor cell apoptosis. Similar effects were noted when genistein and the enterolignans enterolactone and enterodiol were injected (10 mg/kg body weight), both alone and in a combination of all three (134). Tumors regressed initially in response to genistein alone and then stopped regressing after prolonged exposure, whereas the tumors continued to regress in response to the enterolignans and in response to the combination of enterolignans and genistein. As was observed for the combination of flaxseed and isoflavone-rich isolated soy protein, the addition of enterolignans markedly inhibited genistein-induced tumor cell proliferation compared with genistein alone. Finally, only the enterolignans increased apoptosis; there was no effect of the combination of genistein and the enterolignans on the percentage of cells that underwent apoptosis. Neither flaxseed nor the enterolignans inhibited the skeletal benefits of isoflavonerich isolated soy protein or genistein, respectively (135).

There is however considerable debate about the merits of using animal models of breast cancer to predict effects in humans. A specific criticism (136) of the athymic ovariectomized mouse model as used by the research groups of Helferich (126,127) and Thompson (133) is that, unlike pre- and postmenopausal women, these mice do not produce sufficient endogenous estrogen to promote or to even maintain tumors. Thus, this model is biased toward finding that even weakly estrogenic compounds stimulate the growth of existing estrogen-sensitive mammary tumors. However, mammary tumor stimulation has been noted in other rodent models in response to both dietary (18,20,129) and subcutaneously injected (137) genistein, including those in which estrogen levels are more reflective of the hormonal milieu of postmenopausal women (129).

Another criticism of the animal studies is the use of high oral doses of isoflavones. Studies using multiple treatment doses suggest that isoflavones at 200–500 ppm in the diet yield serum concentrations in rodents that are within the range observed in humans who consume soyfoods or use soy isoflavone supplements, whereas doses of approximately 1000 ppm result in excessive isoflavone concentrations (132,138,139). In addition, serum isoflavonoid molar ratios differ between rodents and humans because the rodent gut bacteria effectively convert daidzein to the metabolite equol, whereas only 30%–50% of humans carry bacteria with this metabolic capacity (61, 63). Furthermore, even in humans who are classified as equol producers, genistein is the

predominant isoflavone in the serum in response to the ingestion of soy or mixed isoflavones, whereas equol predominates in most other species, including both rodents and monkeys (61).

Angela Brodie (University of Maryland) described the different rodent models that are available for studying mammary cancer. These include aromatase-overexpressing transgenic mice, the BRCA1 conditional mutant mouse model, and the human ER+ MCF-7 aromatase (MCF-7C_a) cell xenograft model (140,141). In the first two models estrogen levels are regulated by the host. In the third model, however, estrogen-dependent tumor growth is regulated by the tumors, which produce their own estrogen. Thus, this third model may better reflect the hormonal environment of women than the first two. Furthermore, in the ER+ MCF-7 C_a model, mice are also often injected with androstendione to provide greater substrate for estrogen production, resulting in rapidly growing tumors that are sensitive to both antiestrogens and aromatase inhibitors. This model offers an opportunity to study the effect of soy and isoflavones on the growth of estrogen-sensitive mammary tumors.

Clinical Studies

As discussed previously, the hormonal milieu may affect the biologic activity of isoflavones. Therefore, it is important to have a clear understanding of the hormonal environment of both preand postmenopausal women in general and of normal and cancerous breast tissue in particular. Jürgen Geisler (Haukeland University Hospital, Norway) noted that in postmenopausal women, plasma concentrations of estrone, estradiol, and estrone sulfate are 60-80 pmol/L, 10-20 pmol/L, and 400-500 pmol/L, respectively (142). Breast tissue estrogen levels are largely determined by uptake from serum, by local production in tumor cells or in surrounding tissues, and by the metabolism of estrogens in breast and peripheral tissues. Free estrogens are taken up by breast tissue against a concentration gradient-estradiol, estrone, and estrone sulfate concentrations are 10–20 times, 2–10 times, and 10–20 times higher, respectively, in breast cancer tissue than in plasma (142). Furthermore, plasma estrogen levels do not predict tissue estrogen levels in postmenopausal breast cancer patients (142). Thus, it appears that, despite having lower serum estrogen levels, postmenopausal women have breast tissue estrogen concentrations that are similar to those of premenopausal women.

Clearly there is a need to determine the effect of soy consumption on markers of breast cancer risk in high-risk women and breast cancer patients. Unfortunately, few if any noninvasive or minimally invasive assays for markers of breast cancer risk have been identified. One that has been used extensively is breast tissue density. Dr Norman Boyd (Ontario Cancer Institute, Toronto, ON) noted that differences in the parenchymal pattern of the breast on mammography reflect differences in the amounts of stromal, epithelial, and fat tissue present in the breast (143). Stroma and epithelium are radiologically dense, whereas fat is lucent. Women who have extensive areas of mammographically dense breast tissue have a 4-6 times higher risk for breast cancer than women with little or no density (144). Furthermore, menopausal hormone (combined estrogen plus progestin) interventions, which are known to increase breast cancer risk, also increase breast tissue density (145). Nonetheless, the effects of hormone therapy on breast cancer risk do not appear to be mediated by effects on breast tissue density (146). These data suggest that if an intervention alters breast density, it does not necessarily follow that the intervention will alter breast cancer risk; conversely, interventions may alter risk of breast cancer without changing density.

Several investigators have examined the impact of either soyfoods or isoflavones on breast tissue density in intervention (21, 22,147-149) and epidemiologic (150-154) studies. Gertraud Maskarinec (Cancer Research Center of Hawaii, Honolulu, HI) discussed three intervention studies: a 1-year study that examined the impact of isoflavone supplements derived from red clover on mammographic density in postmenopausal women (21), a 1-year study in which premenopausal women were given 100 mg/d soybean isoflavones in supplement form (147,148), and a 2-year study in which premenopausal women consumed two servings of soyfoods per day that provided approximately 50 mg of isoflavones (22,149). Maskarinec concluded that these studies showed that there is no effect of 1-2 years of soy or isoflavone consumption on breast density in premenopausal women. No published studies have examined the impact of soy isoflavones on density in postmenopausal women; however, red clover isoflavones, which lead to blood isoflavone concentrations similar to those achieved with the ingestion of soyfoods (155), had no effect (21). Thus, these studies show evidence of neither harm nor benefit on breast cancer density, in contrast to the effects of hormone therapy, which increases breast tissue density (156).

Jeffrey A. Tice (University of California San Francisco, San Francisco, CA) also discussed the effects of soy on breast tissue density. Tice and his colleagues carried out a doubleblinded study in which 47 postmenopausal women at high risk (defined by Gail risk \geq 1.67% and mammographic breast density \geq 50%) for breast cancer were randomly assigned to either a daily dose of 25 g casein or 25 g isoflavone-rich isolated soy protein that provided approximately 50 mg of isoflavones. At 6 months, there were no differences between the groups in the change in breast density timed to the menstrual cycle; there were also no differences in circulating levels of insulin-like growth factor 1 (IGF-1), insulin-like growth factor binding protein-3 (IGFBP-3), or the IGF-1 : IGFBP-3 ratio. Thus, these results are consistent with the lack of effect of isoflavones on breast density in the studies reviewed by Maskarinec.

In the not too distant future, more information about the impact of soy on breast tissue density will be available. Lee-Jane Lu (The University of Texas Medical Branch, Galveston, TX) presented the experimental designs for her two ongoing doubleblinded, randomized, placebo-controlled, parallel group studies. Both trials are 2 years in duration and involve healthy premenopausal women aged 30-42 years not using contraceptive medications. In one study, women will consume daily either 40 g soy protein without isoflavones or 40 g casein, and in the other, women will take a placebo or 130 mg/d isoflavones as a supplement. Serum hormones, bone density, and breast density will be measured at baseline and yearly during the intervention period. There will be approximately 100 women per group in each study. Thus, this research, both because of the duration and size, may provide the most definitive data to date on the effect of both soy protein and isoflavones on breast tissue density in premenopausal women.

Direct histologic analysis of breast tissue—short of monitoring for tumor development—provides the optimal approach for determining cancer risk. Melanie Palomares (City of Hope Comprehensive Cancer Center, Duarte, CA), presented the results of her pilot randomized controlled trial in which postmenopausal breast cancer survivors were given either a placebo or an isoflavone supplement (100 mg/d) for 1 year. To qualify, women had to have a history of unilateral stage I–II infiltrating ductal or infiltrating lobular carcinoma or ductal carcinoma in situ and not to have used estrogen-modulating therapy, including SERMs, aromatase inhibitors, hormone therapy, or hormonally active herbal supplements within 3 months of enrollment. Also, women were excluded if their baseline diet included more than three servings of soyfoods per day (average 10 mg/d of isoflavones).

Normal breast tissue from the contralateral breast was sampled using ultrasound-guided core biopsy at baseline and at 6 and 12 months. At none of the time points were there statistically significant differences in cell proliferation (Ki67 index), histology (hyperplasia with or without atypia), or ER expression between the two groups. However, because of the small sample size (n = 23) of this study the findings should be interpreted cautiously. Interestingly, the baseline Ki67 indices were higher and the incidence of hyperplasia in these women was greater than what has been observed for healthy individuals in other studies, supporting the observation that breast cancer patients are at an increased risk of developing contralateral breast cancer (157).

Finally, Carol Fabian (University of Kansas Medical School, Kansas City, KS) reviewed her research demonstrating the use of random periareolar fine-needle aspiration (RPFNA) for obtaining breast tissue to study the effects of different interventions on breast cancer risk (158). The advantages of this approach include the capacity to assess precancerous changes at the tissue level, the availability of tissue for other response biomarkers (e.g., Ki67) and those predictive of response (e.g., ER expression), and minimal discomfort on the part of the subject. Disadvantages include interpretation and sampling variance; approximately 25% of a placebo-treated group will show improvement (40% will show overall categoric change) when a high-risk cohort member exhibiting hyperplasia +/- atypia is treated for 6 months. In one 6-month study using RPFNA among women on a stable dose of hormone therapy, letrozole reduced cell proliferation (Ki67 index) by two-thirds but did not affect breast tissue density, thus emphasizing the importance of analyzing tissue to assess risk. Dr Fabian presented the experimental design for an ongoing study in which RPFNA will be used to investigate the effects of the plant lignan secoisolariciresinol diglycoside on breast cancer risk in premenopausal women at high risk for breast cancer.

It is clear from the above discussion that biomarkers of breast cancer risk are limited and often their association with causality is not well understood. The clinical studies presented as part of this workshop suggest that biomarkers measured in the target tissue (e.g., breast tissue hormone concentrations or epithelial cell proliferation), rather than surrogate measures (e.g., breast density or serum hormone concentrations), may be more appropriate for evaluating the impact of an intervention on risk of breast cancer. The limitations of these existing biomarkers highlight the critical need to develop biomarkers definitively linked to breast cancer as an outcome.

WORKSHOP CONCLUSIONS AND RESEARCH RECOMMENDATIONS

Neither the existing animal nor human data allow definitive conclusions to be drawn about the effect of soyfoods or isoflavones on breast cancer risk in high-risk women and on the survival of breast cancer patients. There is an important public health imperative to determine the safety of soyfoods in both groups of women. Definitively establishing that soyfoods do not adversely affect the survival of breast cancer patients may not be possible. To do so will likely require conducting a long-term intervention trial in which tumor recurrence or survival are endpoints. However, conducting such studies may be prohibitively expensive and raise ethical concerns. Assessing the potential impact of soyfoods on breast cancer risk in high-risk women is possible by examining cancer risk markers (e.g., cell proliferation, apoptosis) using breast tissue samples obtained via RPFNA or ultrasoundguided biopsies. Such research is urgently needed and should be designed to determine both safety and efficacy. Careful consideration should be given to the types of soy products used for such interventions; emphasis should be placed on using products that allow findings to be extrapolated to as broad a range of soy products as possible.

References

- Messina M, Barnes S. The role of soy products in reducing risk of cancer. J Natl Cancer Inst 1991;83:541–6.
- (2) Sarkar FH, Li Y. Soy isoflavones and cancer prevention. Cancer Invest 2003;21:744–57.
- (3) Messina MJ, Persky V, Setchell KD, Barnes S. Soy intake and cancer risk: a review of the in vitro and in vivo data. Nutr Cancer 1994;21:113–31.
- (4) Pisani P, Bray F, Parkin DM. Estimates of the world-wide prevalence of cancer for 25 sites in the adult population. Int J Cancer 2002;97:72–81.
- (5) Folman Y, Pope GS. The interaction in the immature mouse of potent oestrogens with coumestrol, genistein and other utero-vaginotrophic compounds of low potency. J Endocrinol 1966;34:215–25.
- (6) Lee HP, Gourley L, Duffy SW, Esteve J, Lee J, Day NE. Dietary effects on breast-cancer risk in Singapore. Lancet 1991;337:1197–2000.
- (7) Barnes S, Grubbs C, Setchell KD, Carlson J. Soybeans inhibit mammary tumors in models of breast cancer. Prog Clin Biol Res 1990;347:239–53.
- (8) Yan L, Spitznagel E. A meta-analysis of soyfoods and risk of breast cancer in women. Int J Cancer Prev 2005;1:281–93.
- (9) Trock BJ, Hilakivi-Clarke L, Clarke R. Meta-analysis of soy intake and breast cancer risk. J Natl Cancer Inst 2006;98:459–71.
- (10) Messina MJ, Loprinzi CL. Soy for breast cancer survivors: a critical review of the literature. J Nutr 2001;131:3095S–108S.
- (11) Magee PJ, Rowland IR. Phyto-oestrogens, their mechanism of action: current evidence for a role in breast and prostate cancer. Br J Nutr 2004; 91:513–31.
- (12) Yan L, Li D, Yee JA. Dietary supplementation with isolated soy protein reduces metastasis of mammary carcinoma cells in mice. Clin Exp Metastasis 2002;19:535–40.
- (13) Constantinou AI, Lantvit D, Hawthorne M, Xu X, van Breemen RB, Pezzuto JM. Chemopreventive effects of soy protein and purified soy isoflavones on DMBA-induced mammary tumors in female Sprague-Dawley rats. Nutr Cancer 2001;41:75–81.
- (14) Shao ZM, Wu J, Shen ZZ, Barsky SH. Genistein exerts multiple suppressive effects on human breast carcinoma cells. Cancer Res 1998;58: 4851–7.
- (15) Zhou JR, Yu L, Mai Z, Blackburn GL. Combined inhibition of estrogendependent human breast carcinoma by soy and tea bioactive components in mice. Int J Cancer 2004;108:8–14.
- (16) Gallo D, Ferlini C, Fabrizi M, Prislei S, Scambia G. Lack of stimulatory activity of a phytoestrogen-containing soy extract on the growth of breast cancer tumors in mice. Carcinogenesis 2006;27:1404–9.
- (17) Cohen LA, Zhao Z, Pittman B, Scimeca JA. Effect of intact and isoflavone-depleted soy protein on NMU-induced rat mammary tumorigenesis. Carcinogenesis 2000;21:929–35.
- (18) Day JK, Besch-Williford C, McMann TR, Hufford MG, Lubahn DB, MacDonald RS. Dietary genistein increased DMBA-induced mammary adenocarcinoma in wild-type, but not ER alpha KO, mice. Nutr Cancer 2001;39:226–32.

- (19) Thomsen AR, Mortensen A, Breinholt VM, Lindecrona RH, Penalvo JL, Sorensen IK. Influence of Prevastein(R), an isoflavone-rich soy product, on mammary gland development and tumorigenesis in Tg.NK (MMTV/ c-neu) mice. Nutr Cancer 2005;52:176–88.
- (20) Allred CD, Allred KF, Ju YH, Clausen LM, Doerge DR, Schantz SL, et al. Dietary genistein results in larger MNU-induced, estrogen-dependent mammary tumors following ovariectomy of Sprague-Dawley rats. Carcinogenesis 2004;25:211–8.
- (21) Atkinson C, Warren RM, Sala E, Dowsett M, Dunning AM, Healey CS, et al. Red-clover-derived isoflavones and mammographic breast density: a double-blind, randomized, placebo-controlled trial. Breast Cancer Res 2004;6:R170–9.
- (22) Maskarinec G, Takata Y, Franke AA, Williams AE, Murphy SP. A 2-year soy intervention in premenopausal women does not change mammographic densities. J Nutr 2004;134:3089–94.
- (23) Kurzer MS. Hormonal effects of soy in premenopausal women and men. J Nutr 2002;132:5708–38.
- (24) Maskarinec G, Franke AA, Williams AE, Hebshi S, Oshiro C, Murphy S, et al. Effects of a 2-year randomized soy intervention on sex hormone levels in premenopausal women. Cancer Epidemiol Biomarkers Prev 2004;13:1736–44.
- (25) Palomares MR, Hopper L, Goldstein L, Lehman CD, Storer BE, Gralow JR. Effect of soy isoflavones on breast proliferation in postmenopausal breast cancer survivors. Breast Cancer Res Treat 2004;88(Suppl 1):4002.
- (26) Brown BD, Thomas W, Hutchins A, Martini MC, Slavin JL. Types of dietary fat and soy minimally affect hormones and biomarkers associated with breast cancer risk in premenopausal women. Nutr Cancer 2002; 43:22–30.
- (27) Shu XO, Jin F, Dai Q, Wen W, Potter JD, Kushi LH, et al. Soyfood intake during adolescence and subsequent risk of breast cancer among Chinese women. Cancer Epidemiol Biomarkers Prev 2001;10:483–8.
- (28) Wu AH, Wan P, Hankin J, Tseng CC, Yu MC, Pike MC. Adolescent and adult soy intake and risk of breast cancer in Asian-Americans. Carcinogenesis 2002;23:1491–6.
- (29) Korde L, Fears T, Wu A, West D, Pike M, Hoover R, et al. Adolescent and childhood soy intake and breast cancer risk in Asian-American women. Breast Cancer Res Treat 2005;88(Suppl 1):S149.
- (30) Lamartiniere CA, Zhao YX, Fritz WA. Genistein: mammary cancer chemoprevention, in vivo mechanisms of action, potential for toxicity and bioavailability in rats. J Womens Cancer 2000;2:11–9.
- (31) Hilakivi-Clarke L, Onojafe I, Raygada M, Cho E, Skaar T, Russo I, et al. Prepubertal exposure to zearalenone or genistein reduces mammary tumorigenesis. Br J Cancer 1999;80:1682–8.
- (32) Russo J, Mailo D, Hu YF, Balogh G, Sheriff F, Russo IH. Breast differentiation and its implication in cancer prevention. Clin Cancer Res 2005;11:931s–6s.
- (33) Dolinoy DC, Weidman JR, Waterland RA, Jirtle RL. Maternal genistein alters coat color and protects Avy mouse offspring from obesity by modifying the fetal epigenome. Environ Health Perspect 2006;114:567–72.
- (34) Souzeau E, Belanger S, Picard S, Deschepper CF. Dietary isoflavones during pregnancy and lactation provide cardioprotection to offspring rats in adulthood. Am J Physiol Heart Circ Physiol 2005;289:H715–21.
- (35) Hilakivi-Clarke L, Cho E, Clarke R. Maternal genistein exposure mimics the effects of estrogen on mammary gland development in female mouse offspring. Oncol Rep 1998;5:609–16.
- (36) Hilakivi-Clarke L, Cho E, Onojafe I, Raygada M, Clarke R. Maternal exposure to genistein during pregnancy increases carcinogen-induced mammary tumorigenesis in female rat offspring. Oncol Rep 1999;6:1089–95.
- (37) Hsieh CY, Santell RC, Haslam SZ, Helferich WG. Estrogenic effects of genistein on the growth of estrogen receptor-positive human breast cancer (MCF-7) cells in vitro and in vivo. Cancer Res 1998;58:3833–8.
- (38) Allred CD, Ju YH, Allred KF, Chang J, Helferich WG. Dietary genistin stimulates growth of estrogen-dependent breast cancer tumors similar to that observed with genistein. Carcinogenesis 2001;22:1667–73.
- (39) Hulka BS. Epidemiology of susceptibility to breast cancer. Prog Clin Biol Res 1996;395:159–74.
- (40) Meng L, Maskarinec G, Wilkens L. Ethnic differences and factors related to breast cancer survival in Hawaii. Int J Epidemiol 1997;26:1151–8.
- (41) Yonemoto RH. Breast cancer in Japan and United States: epidemiology, hormone receptors, pathology, and survival. Arch Surg 1980;115:1056–62.

- (42) Morrison AS, Lowe CR, MacMahon B, Ravnihar B, Yuasa S. Some international differences in treatment and survival in breast cancer. Int J Cancer 1976;18:269–73.
- (43) Ohsumi S, Sakamoto G, Takashima S, Koyama H, Shin E, Suemasu K, et al. Long-term results of breast-conserving treatment for early-stage breast cancer in Japanese women from multicenter investigation. Jpn J Clin Oncol 2003;33:61–7.
- (44) Kanemori M, Prygrocki M. Results of breast conservation therapy from a single-institution community hospital in Hawaii with a predominantly Japanese population. Int J Radiat Oncol Biol Phys 2005;62:193–7.
- (45) Haas JS, Kaplan CP, Des Jarlais G, Gildengoin V, Perez-Stable EJ, Kerlikowske K. Perceived risk of breast cancer among women at average and increased risk. J Womens Health (Larchmt) 2005;14:845–51.
- (46) Fang CY, Tseng M, Daly MB. Correlates of soy food consumption in women at increased risk for breast cancer. J Am Diet Assoc 2005;105: 1552–8.
- (47) Horn-Ross PL, John EM, Canchola AJ, Stewart SL, Lee MM. Phytoestrogen intake and endometrial cancer risk. J Natl Cancer Inst 2003; 95:1158–64.
- (48) Goodman-Gruen D, Kritz-Silverstein D. Usual dietary isoflavone intake is associated with cardiovascular disease risk factors in postmenopausal women. J Nutr 2001;131:1202–6.
- (49) 2004Q-0151: Qualified Health Claim (QHC): Soy Protein and Cancer. Available at: http://www.fda.gov/ohrms/dockets/dockets/04q0151/ 04q0151.htm. [Last accessed: December 10, 2005.]
- (50) van Erp-Baart MA, Brants HA, Kiely M, Mulligan A, Turrini A, Sermoneta C, et al. Isoflavone intake in four different European countries: the VENUS approach. Br J Nutr 2003;89(Suppl 1):S25–30.
- (51) van der Schouw YT, Kreijkamp-Kaspers S, Peeters PH, Keinan-Boker L, Rimm EB, Grobbee DE. Prospective study on usual dietary phytoestrogen intake and cardiovascular disease risk in Western women. Circulation 2005;111:465–71.
- (52) Kirk P, Patterson RE, Lampe J. Development of a soy food frequency questionnaire to estimate isoflavone consumption in US adults. J Am Diet Assoc 1999;99:558–63.
- (53) Frankenfeld CL, Patterson RE, Kalhorn TF, Skor HE, Howald WN, Lampe JW. Validation of a soy food frequency questionnaire with plasma concentrations of isoflavones in US adults. J Am Diet Assoc 2002;102: 1407–13.
- (54) Franke AA, Custer LJ, Wang W, Shi CY. HPLC analysis of isoflavonoids and other phenolic agents from foods and from human fluids. Proc Soc Exp Biol Med 1998;217:263–73.
- (55) Murphy PA, Song T, Buseman G, Barua K, Beecher GR, Trainer D, et al. Isoflavones in retail and institutional soy foods. J Agric Food Chem 1999;47:2697–704.
- (56) Rowland I, Faughnan M, Hoey L, Wahala K, Williamson G, Cassidy A. Bioavailability of phyto-oestrogens. Br J Nutr 2003;89(Suppl 1): S45–58.
- (57) Nagata C, Inaba S, Kawakami N, Kakizoe T, Shimizu H. Inverse association of soy product intake with serum androgen and estrogen concentrations in Japanese men. Nutr Cancer 2000;36:14–8.
- (58) Nagata C, Kabuto M, Kurisu Y, Shimizu H. Decreased serum estradiol concentration associated with high dietary intake of soy products in premenopausal Japanese women. Nutr Cancer 1997;29:228–33.
- (59) Nagata C, Takatsuka N, Kawakami N, Shimizu H. A prospective cohort study of soy product intake and stomach cancer death. Br J Cancer 2002;87:31–6.
- (60) Somekawa Y, Chiguchi M, Ishibashi T, Aso T. Soy intake related to menopausal symptoms, serum lipids, and bone mineral density in postmenopausal Japanese women. Obstet Gynecol 2001;97:109–15.
- (61) Gu L, House SE, Prior RL, Fang N, Ronis MJ, Clarkson TB, et al. Metabolic phenotype of isoflavones differ among female rats, pigs, monkeys, and women. J Nutr 2006;136:1215–21.
- (62) Wiseman H, Casey K, Bowey EA, Duffy R, Davies M, Rowland IR, et al. Influence of 10 wk of soy consumption on plasma concentrations and excretion of isoflavonoids and on gut microflora metabolism in healthy adults. Am J Clin Nutr 2004;80:692–9.
- (63) Setchell KD, Brown NM, Lydeking-Olsen E. The clinical importance of the metabolite equol—a clue to the effectiveness of soy and its isoflavones. J Nutr 2002;132:3577–84.

- (65) Erdman JW Jr, Badger TM, Lampe JW, Setchell KD, Messina M. Not all soy products are created equal: caution needed in interpretation of research results. J Nutr 2004;134:1229S–33S.
- (66) Kuiper GG, Carlsson B, Grandien K, Enmark E, Haggblad J, Nilsson S, et al. Comparison of the ligand binding specificity and transcript tissue distribution of estrogen receptors alpha and beta. Endocrinology 1997;138:863–70.
- (67) Kuiper GG, Lemmen JG, Carlsson B, Corton JC, Safe SH, van der Saag PT, et al. Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor beta. Endocrinology 1998;139:4252–63.
- (68) Teede HJ, Dalais FS, McGrath BP. Dietary soy containing phytoestrogens does not have detectable estrogenic effects on hepatic protein synthesis in postmenopausal women. Am J Clin Nutr 2004;79:396–401.
- (69) Yildiz MF, Kumru S, Godekmerdan A, Kutlu S. Effects of raloxifene, hormone therapy, and soy isoflavone on serum high-sensitive Creactive protein in postmenopausal women. Int J Gynaecol Obstet 2005; 90:128–33.
- (70) D'Anna R, Baviera G, Corrado F, Cancellieri F, Crisafulli A, Squadrito F. The effect of the phytoestrogen genistein and hormone replacement therapy on homocysteine and C-reactive protein level in postmenopausal women. Acta Obstet Gynecol Scand 2005;84:474–7.
- (71) Pike AC, Brzozowski AM, Hubbard RE, Bonn T, Thorsell AG, Engstrom O, et al. Structure of the ligand-binding domain of oestrogen receptor beta in the presence of a partial agonist and a full antagonist. EMBO J 1999;18:4608–18.
- (72) Naciff JM, Jump ML, Torontali SM, Carr GJ, Tiesman JP, Overmann GJ, et al. Gene expression profile induced by 17alpha-ethynyl estradiol, bisphenol A, and genistein in the developing female reproductive system of the rat. Toxicol Sci 2002;68:184–99.
- (73) Pearce V, Nawaz Z, Xiao W, Wiedenfeld D, Boyle N, Smith D. 4-Ethoxymethylphenol: a novel phytoestrogen that acts as an agonist for human estrogen receptors. J Steroid Biochem Mol Biol 2003;84:431–9.
- (74) Dey M, Lyttle CR, Pickar JH. Recent insights into the varying activity of estrogens. Maturitas 2000;34(Suppl 2):S25–33.
- (75) Markiewicz L, Garey J, Adlercreutz H, Gurpide E. In vitro bioassays of non-steroidal phytoestrogens. J Steroid Biochem Mol Biol 1993; 45:399–405.
- (76) Mayr U, Butsch A, Schneider S. Validation of two in vitro test systems for estrogenic activities with zearalenone, phytoestrogens and cereal extracts. Toxicology 1992;74:135–49.
- (77) Nagel SC, vom Saal FS, Welshons WV. The effective free fraction of estradiol and xenoestrogens in human serum measured by whole cell uptake assays: physiology of delivery modifies estrogenic activity. Proc Soc Exp Biol Med 1998;217:300–9.
- (78) Totta P, Acconcia F, Virgili F, Cassidy A, Weinberg PD, Rimbach G, et al. Daidzein-sulfate metabolites affect transcriptional and antiproliferative activities of estrogen receptor-{beta} in cultured human cancer cells. J Nutr 2005;135:2687–93.
- (79) Freyberger A, Schmuck G. Screening for estrogenicity and anti-estrogenicity: a critical evaluation of an MVLN cell-based transactivation assay. Toxicol Lett 2005;155:1–13.
- (80) Gehm BD, McAndrews JM, Chien PY, Jameson JL. Resveratrol, a polyphenolic compound found in grapes and wine, is an agonist for the estrogen receptor. Proc Natl Acad Sci U S A 1997;94:14138–43.
- (81) An J, Tzagarakis-Foster C, Scharschmidt TC, Lomri N, Leitman DC. Estrogen receptor beta-selective transcriptional activity and recruitment of coregulators by phytoestrogens. J Biol Chem 2001;276:17808–14.
- (82) Margeat E, Bourdoncle A, Margueron R, Poujol N, Cavailles V, Royer C. Ligands differentially modulate the protein interactions of the human estrogen receptors alpha and beta. J Mol Biol 2003;326:77–92.
- (83) Kostelac D, Rechkemmer G, Briviba K. Phytoestrogens modulate binding response of estrogen receptors alpha and beta to the estrogen response element. J Agric Food Chem 2003;51:7632–5.
- (84) Brzezinski A, Debi A. Phytoestrogens: the "natural" selective estrogen receptor modulators? Eur J Obstet Gynecol Reprod Biol 1999;85:47–51.
- (85) Diel P, Geis RB, Caldarelli A, Schmidt S, Leschowsky UL, Voss A, et al. The differential ability of the phytoestrogen genistein and of estradiol to induce uterine weight and proliferation in the rat is associated

with a substance specific modulation of uterine gene expression. Mol Cell Endocrinol 2004;221:21–32.

- (86) Constantinou A, Huberman E. Genistein as an inducer of tumor cell differentiation: possible mechanisms of action. Proc Soc Exp Biol Med 1995;208:109–15.
- (87) Barnes S. Soy isoflavones-phytoestrogens and what else? J Nutr 2004; 134:1225S-8S.
- (88) Strom A, Hartman J, Foster JS, Kietz S, Wimalasena J, Gustafsson JA. Estrogen receptor {beta} inhibits 17{beta}-estradiol-stimulated proliferation of the breast cancer cell line T47D. Proc Natl Acad Sci U S A 2004;101:1566–71.
- (89) Park BW, Kim KS, Heo MK, Ko SS, Hong SW, Yang WI, et al. Expression of estrogen receptor-beta in normal mammary and tumor tissues: is it protective in breast carcinogenesis? Breast Cancer Res Treat 2003;80:79–85.
- (90) Peterson G, Barnes S. Genistein inhibition of the growth of human breast cancer cells: independence from estrogen receptors and the multi-drug resistance gene. Biochem Biophys Res Commun 1991;179:661–7.
- (91) Dampier K, Hudson EA, Howells LM, Manson MM, Walker RA, Gescher A. Differences between human breast cell lines in susceptibility towards growth inhibition by genistein. Br J Cancer 2001;85:618–24.
- (92) Le Bail JC, Champavier Y, Chulia AJ, Habrioux G. Effects of phytoestrogens on aromatase, 3beta and 17beta-hydroxysteroid dehydrogenase activities and human breast cancer cells. Life Sci 2000;66:1281–91.
- (93) Fioravanti L, Cappelletti V, Miodini P, Ronchi E, Brivio M, Di Fronzo G. Genistein in the control of breast cancer cell growth: insights into the mechanism of action in vitro. Cancer Lett 1998;130:143–52.
- (94) Power KA, Thompson LU. Ligand-induced regulation of ERalpha and ERbeta is indicative of human breast cancer cell proliferation. Breast Cancer Res Treat 2003;81:209–21.
- (95) Chen YM, Ho SC, Lam SS, Ho SS, Woo JL. Beneficial effect of soy isoflavones on bone mineral content was modified by years since menopause, body weight, and calcium intake: a double-blind, randomized, controlled trial. Menopause 2004;11:246–54.
- (96) Chen WF, Huang MH, Tzang CH, Yang M, Wong MS. Inhibitory actions of genistein in human breast cancer (MCF-7) cells. Biochim Biophys Acta 2003;1638:187–96.
- (97) Akiyama T, Ogawara H. Use and specificity of genistein as inhibitor of protein-tyrosine kinases. Methods Enzymol 1991;201:362–70.
- (98) Akiyama T, Ishida J, Nakagawa S, Ogawara H, Watanabe S, Itoh N, et al. Genistein, a specific inhibitor of tyrosine-specific protein kinases. J Biol Chem 1987;262:5592–5.
- (99) Constantinou A, Kiguchi K, Huberman E. Induction of differentiation and DNA strand breakage in human HL-60 and K-562 leukemia cells by genistein. Cancer Res 1990;50:2618–24.
- (100) Kim H, Peterson TG, Barnes S. Mechanisms of action of the soy isoflavone genistein: emerging role for its effects via transforming growth factor beta signaling pathways. Am J Clin Nutr 1998;68:1418S–25S.
- (101) Ruiz-Larrea MB, Mohan AR, Paganga G, Miller NJ, Bolwell GP, Rice-Evans CA. Antioxidant activity of phytoestrogenic isoflavones. Free Radic Res 1997;26:63–70.
- (102) Wang W, Higuchi CM, Zhang R. Individual and combinatory effects of soy isoflavones on the in vitro potentiation of lymphocyte activation. Nutr Cancer 1997;29:29–34.
- (103) Zhang R, Li Y, Wang W. Enhancement of immune function in mice fed high doses of soy daidzein. Nutr Cancer 1997;29:24–8.
- (104) Zhang Y, Song TT, Cunnick JE, Murphy PA, Hendrich S. Daidzein and genistein glucuronides in vitro are weakly estrogenic and activate human natural killer cells at nutritionally relevant concentrations. J Nutr 1999;129:399–405.
- (105) Su SJ, Yeh TM, Chuang WJ, Ho CL, Chang KL, Cheng HL, et al. The novel targets for anti-angiogenesis of genistein on human cancer cells. Biochem Pharmacol 2005;69:307–18.
- (106) Han D, Tachibana H, Yamada K. Inhibition of environmental estrogeninduced proliferation of human breast carcinoma MCF-7 cells by flavonoids. In Vitro Cell Dev Biol Anim 2001;37:275–82.
- (107) Wang C, Kurzer MS. Effects of phytoestrogens on DNA synthesis in MCF-7 cells in the presence of estradiol or growth factors. Nutr Cancer 1998;31:90–100.
- (108) Zava DT, Duwe G. Estrogenic and antiproliferative properties of genistein and other flavonoids in human breast cancer cells in vitro. Nutr Cancer 1997;27:31–40.

- (109) Miodini P, Fioravanti L, Di Fronzo G, Cappelletti V. The two phytooestrogens genistein and quercetin exert different effects on oestrogen receptor function. Br J Cancer 1999;80:1150–5.
- (110) Petrakis NL, Barnes S, King EB, Lowenstein J, Wiencke J, Lee MM, et al. Stimulatory influence of soy protein isolate on breast secretion in preand postmenopausal women. Cancer Epidemiol Biomarkers Prev 1996; 5:785–94.
- (111) Hargreaves DF, Potten CS, Harding C, Shaw LE, Morton MS, Roberts SA, et al. Two-week dietary soy supplementation has an estrogenic effect on normal premenopausal breast. J Clin Endocrinol Metab 1999;84:4017–24.
- (112) Thompson AM, Elton RA, Hawkins RA, Chetty U, Steel CM. PS2 mRNA expression adds prognostic information to node status for 6-year survival in breast cancer. Br J Cancer 1998;77:492–6.
- (113) Kim J, Petz LN, Ziegler YS, Wood JR, Potthoff SJ, Nardulli AM. Regulation of the estrogen-responsive pS2 gene in MCF-7 human breast cancer cells. J Steroid Biochem Mol Biol 2000;74:157–68.
- (114) McMichael-Phillips DF, Harding C, Morton M, Roberts SA, Howell A, Potten CS, et al. Effects of soy-protein supplementation on epithelial proliferation in the histologically normal human breast. Am J Clin Nutr 1998;68:14318–5S.
- (115) Wren BG. Do female sex hormones initiate breast cancer? A review of the evidence. Climacteric 2004;7:120–8.
- (116) Writing Group for the Women's Health Initiative Investigators. Risks and benefits of estrogen plus progestin in healthy postmenopausal women: principal results From the Women's Health Initiative randomized controlled trial. JAMA 2002;288:321–33.
- (117) Anderson GL, Limacher M, Assaf AR, Bassford T, Beresford SA, Black H, et al. Effects of conjugated equine estrogen in postmenopausal women with hysterectomy: the Women's Health Initiative randomized controlled trial. JAMA 2004;291:1701–12.
- (118) Beral V. Breast cancer and hormone-replacement therapy in the Million Women Study. Lancet 2003;362:419–27.
- (119) Colditz GA. Estrogen, estrogen plus progestin therapy, and risk of breast cancer. Clin Cancer Res 2005;11:909s–17s.
- (120) Shah NR, Borenstein J, Dubois RW. Postmenopausal hormone therapy and breast cancer: a systematic review and meta-analysis. Menopause 2005;12:668–78.
- (121) Zava DT, Dollbaum CM, Blen M. Estrogen and progestin bioactivity of foods, herbs, and spices. Proc Soc Exp Biol Med 1998;217:369–78.
- (122) Creasman WT. Hormone replacement therapy after cancers. Curr Opin Oncol 2005;17:493–9.
- (123) Colditz GA. Menopausal hormone therapy after breast cancer. Breast Cancer Res 2005;7:168–70.
- (124) Durna EM, Wren BG, Heller GZ, Leader LR, Sjoblom P, Eden JA. Hormone replacement therapy after a diagnosis of breast cancer: cancer recurrence and mortality. Med J Aust 2002;177:347–51.
- (125) Boyapati SM, Shu XO, Ruan ZX, Dai Q, Cai Q, Gao YT, et al. Soyfood intake and breast cancer survival: a followup of the Shanghai Breast Cancer Study. Breast Cancer Res Treat 2005;92:11–7.
- (126) Allred CD, Allred KF, Ju YH, Virant SM, Helferich WG. Soy diets containing varying amounts of genistein stimulate growth of estrogen-dependent (mcf-7) tumors in a dose-dependent manner. Cancer Res 2001;61: 5045–50.
- (127) Allred CD, Allred KF, Ju YH, Goeppinger TS, Doerge DR, Helferich WG. Soy processing influences growth of estrogen-dependent breast cancer tumors. Carcinogenesis 2004;25:1649–57.
- (128) Ju YH, Doerge DR, Allred KF, Allred CD, Helferich WG. Dietary genistein negates the inhibitory effect of tamoxifen on growth of estrogendependent human breast cancer (MCF-7) cells implanted in athymic mice. Cancer Res 2002;62:2474–7.
- (129) Ju YH, Allred KF, Allred CD, Helferich WG. Genistein stimulates growth of human breast cancer cells in a novel, postmenopausal animal model, with low plasma estradiol concentrations. Carcinogenesis 2006;27: 1292–9.
- (130) Allred CD, Twaddle NC, Allred KF, Goeppinger TS, Churchwell MI, Ju YH, et al. Soy processing affects metabolism and disposition of dietary isoflavones in ovariectomized BALB/c mice. J Agric Food Chem 2005;53:8542–50.
- (131) Ju YH, Clausen LM, Allred KF, Almada AL, Helferich WG. Betasitosterol, beta-sitosterol glucoside, and a mixture of beta-sitosterol and

beta-sitosterol glucoside modulate the growth of estrogen-responsive breast cancer cells in vitro and in ovariectomized athymic mice. J Nutr 2004;134:1145–51.

- (132) Ju YH, Fultz J, Allred KF, Doerge DR, Helferich WG. Effects of dietary daidzein and its metabolite, equol, at physiological concentrations on the growth of estrogen-dependent human breast cancer (MCF-7) tumors implanted in ovariectomized athymic mice. Carcinogenesis 2006;27: 856–63.
- (133) Saarinen NM, Power K, Chen J, Thompson LU. Flaxseed attenuates the tumor growth stimulating effect of soy protein in ovariectomized athymic mice with MCF-7 human breast cancer xenografts. Int J Cancer 2006;119:925–31.
- (134) Power KA, Saarinen NM, Chen JM, Thompson LU. Mammalian lignans enterolactone and enterodiol, alone and in combination with the isoflavone genistein, do not promote the growth of MCF-7 xenografts in ovariectomized athymic nude mice. Int J Cancer 2006;118:1316–20.
- (135) Power KA, Ward WE, Chen JM, Saarinen NM, Thompson LU. Genistein alone and in combination with the mammalian lignans enterolactone and enterodiol induce estrogenic effects on bone and uterus in a postmenopausal breast cancer mouse model. Bone 2006;39:117–24.
- (136) Clarkson TB, Appt SE, Wood CE, Cline JM. Lessons to be learned from animal studies on hormones and the breast. Maturitas 2004;49:79–89.
- (137) Kijkuokool P, Parhar IS, Malaivijitnond S. Genistein enhances Nnitrosomethylurea-induced rat mammary tumorigenesis. Cancer Lett 2005: Dec 6.
- (138) Fritz WA, Coward L, Wang J, Lamartiniere CA. Dietary genistein: perinatal mammary cancer prevention, bioavailability and toxicity testing in the rat. Carcinogenesis 1998;19:2151–8.
- (139) Lamartiniere CA, Wang J, Smith-Johnson M, Eltoum IE. Daidzein: bioavailability, potential for reproductive toxicity, and breast cancer chemoprevention in female rats. Toxicol Sci 2002;65:228–38.
- (140) Brodie A, Jelovac D, Long BJ. Predictions from a preclinical model: studies of aromatase inhibitors and antiestrogens. Clin Cancer Res 2003; 9:4558–98.
- (141) Brodie A, Jelovac D, Macedo L, Sabnis G, Tilghman S, Goloubeva O. Therapeutic observations in MCF-7 aromatase xenografts. Clin Cancer Res 2005;11:884s–8s.
- (142) Geisler J. Breast cancer tissue estrogens and their manipulation with aromatase inhibitors and inactivators. J Steroid Biochem Mol Biol 2003;86:245–53.
- (143) Boyd NF, Martin LJ, Stone J, Greenberg C, Minkin S, Yaffe MJ. Mammographic densities as a marker of human breast cancer risk and their use in chemoprevention. Curr Oncol Rep 2001;3:314–21.
- (144) Boyd NF, Rommens JM, Vogt K, Lee V, Hopper JL, Yaffe MJ, et al. Mammographic breast density as an intermediate phenotype for breast cancer. Lancet Oncol 2005;6:798–808.
- (145) Greendale GA, Reboussin BA, Slone S, Wasilauskas C, Pike MC, Ursin G. Postmenopausal hormone therapy and change in mammographic density. J Natl Cancer Inst 2003;95:30–7.
- (146) Boyd NF, Martin LJ, Li Q, Sun L, Chiarelli AM, Hislop G, et al. Mammographic density as a surrogate marker for the effects of hormone therapy on risk of breast cancer. Cancer Epidemiol Biomarkers Prev 2006;15:961–6.
- (147) Maskarinec G, Williams AE, Inouye JS, Stanczyk FZ, Franke AA. A randomized isoflavone intervention among premenopausal women. Cancer Epidemiol Biomarkers Prev 2002;11:195–201.
- (148) Maskarinec G, Williams AE, Carlin L. Mammographic densities in a oneyear isoflavone intervention. Eur J Cancer Prev 2003;12:165–9.
- (149) Maskarinec G, Robbins C, Riola B, Kane-Sample L, Franke AA, Murphy S. Three measures show high compliance in a soy intervention among premenopausal women. J Am Diet Assoc 2003;103:861–6.
- (150) Maskarinec G, Meng L. An investigation of soy intake and mammographic characteristics in Hawaii. Breast Cancer Res 2001;3:134–41.
- (151) Jakes RW, Duffy SW, Ng FC, Gao F, Ng EH, Seow A, et al. Mammographic parenchymal patterns and self-reported soy intake in Singapore Chinese women. Cancer Epidemiol Biomarkers Prev 2002;11: 608–13.
- (152) Frankenfeld CL, McTiernan A, Aiello EJ, Thomas WK, LaCroix K, Schramm J, et al. Mammographic density in relation to daidzeinmetabolizing phenotypes in overweight, postmenopausal women. Cancer Epidemiol Biomarkers Prev 2004;13:1156–62.

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- (153) Nagel G, Mack U, von Fournier D, Linseisen J. Dietary phytoestrogen intake and mammographic density—results of a pilot study. Eur J Med Res 2005;10:389–94.
- (154) Maskarinec G, Pagano I, Lurie G, Wilkens LR, Kolonel LN. Mammographic density and breast cancer risk: the multiethnic cohort study. Am J Epidemiol 2005;162:743–52.
- (155) Beck V, Rohr U, Jungbauer A. Phytoestrogens derived from red clover: an alternative to estrogen replacement therapy? J Steroid Biochem Mol Biol 2005;94:499–518.
- (156) Warren R. Hormones and mammographic breast density. Maturitas 2004;49:67–78.
- (157) MertensWC,HilbertV,Makari-JudsonG.Contralateralbreastcancer: factors associated with stage and size at presentation. Breast J 2004;10:304–12.

(158) Fabian CJ, Kimler BF, Mayo MS, Khan SA. Breast-tissue sampling for risk assessment and prevention. Endocr Relat Cancer 2005;12:185–213.

Notes

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Funding to pay the Open Access publication charges for this article was provided by the Soy Nutrition Institute (St Louis, MO), which is funded by industry members, and by the United Soybean Board, a US farmer–operated organization.

Manuscript received April 7, 2006; revised June 23, 2006; accepted July 19, 2006.