# Adiponectin Dysregulation and Insulin Resistance in Type 1 Diabetes

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**Context:** Type 1 diabetes (T1D) is associated with insulin resistance despite elevated levels of the insulin-sensitizing protein adiponectin. Whether the expected positive correlation between adiponectin and insulin sensitivity is preserved in a T1D population is unknown.

**Objective:** We measured the correlation between total and high-molecular-weight (HMW) adiponectin and insulin sensitivity in T1D patients and nondiabetic controls and identified determinants of adiponectin levels in patients with T1D.

**Design and Participants:** Fasting total and HMW adiponectin were measured in 86 subjects from the Coronary Artery Calcification in T1D (CACTI) cohort (39T1D, 47 nondiabetic; age  $45 \pm 8$  yr; 55% female). The association of adiponectin levels with insulin sensitivity was analyzed.

Setting: The study was conducted at an academic research institute.

**Methods:** Fasting total and HMW adiponectin were measured by RIA and ELISA, respectively. Insulin sensitivity was measured by a hyperinsulinemic-euglycemic clamp. Multivariate linear regression was used to identify determinants of adiponectin levels.

**Results:** Adiponectin levels positively correlated with insulin sensitivity in both subject groups (total adiponectin, r = 0.33 P < 0.05 for T1D, r = 0.29 P < 0.05 controls), but insulin sensitivity was lower in T1D subjects at any given level of total or HMW adiponectin. Adiponectin levels were independently associated with age, gender, and trunk fat, but these variables did not account for increased adiponectin in patients with T1D.

**Conclusion:** Adiponectin levels are positively correlated with insulin sensitivity in T1D patients. However, T1D patients have decreased insulin sensitivity compared with controls at every level of adiponectin, suggesting an important adaptive change of adiponectin set point. (*J Clin Endocrinol Metab* 97: E642–E647, 2012)

Type 1 diabetes (T1D) is primarily a disease of insulin deficiency but is also characterized by insulin resistance (1, 2). In nonautoimmune, insulin-resistant states including obesity and type 2 diabetes, circulating levels of the fat-derived protein adiponectin are decreased compared with controls (3, 4), and insulin resistance correlates closely to decreased adiponectin (5). In contrast, T1D (6) and other autoimmune disorders (7, 8) are associated with increased adiponectin. The relationship between circulating adiponectin levels and insulin action in insulin-resistant patients with T1D remains poorly understood.

Adiponectin, an adipocyte-secreted protein, is an important determinant of whole-body insulin sensitivity (9) and has protective cardiovascular effects (10). Adiponec-

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Abbreviations: BMI, Body mass index; CACTI, Coronary Artery Calcification in T1D; CV, coefficient of variation; DM, diabetes mellitus; FFM, fat free mass; GIR, glucose infusion rate; HEC, hyperinsulinemic-euglycemic clamp; HMW, high molecular weight; T1D, type 1 diabetes.

tin is found in the circulation in three distinct multimeric forms: low molecular weight, medium molecular weight, and high molecular weight (HMW) (11). HMW adiponectin, the presumed active form, is most closely associated with insulin sensitivity (12). To our knowledge, the association of HMW adiponectin to insulin sensitivity in T1D has not been described.

In the present study, we examined the relationship between adiponectin and insulin sensitivity in subjects with and without T1D to determine whether the expected relationship between these measures is altered in patients with T1D. Furthermore, we sought to identify determinants of adiponectin in subjects with T1D and also nondiabetes mellitus (non-DM) controls.

#### **Materials and Methods**

#### Study population

The present analysis includes subjects from the Coronary Artery Calcification in Type 1 Diabetes (CACTI) hyperinsulinemiceuglycemic clamp substudy (2). This substudy cohort consisted of subjects recruited from the previously described CACTI study (13) who completed a hyperinsulinemic-euglycemic clamp. Subjects were adults with T1D or nondiabetic controls and had no history of cardiovascular disease. Subjects in the substudy were representative of the full CACTI cohort (2). One subject was excluded from the present analysis due to elevated creatinine. We present data for the remaining 86 subjects (39 T1D and 47 nondiabetic) in the CACTI clamp substudy. All participants provided informed consent, and the study was approved by the Colorado Combined Institutional Review Board.

## **Body composition**

Dual x-ray absorptiometry scans were performed for body composition (percentage body fat, percentage trunk fat) and determination of fat free mass. Abdominal computed tomography scans for calculation of abdominal visceral and sc fat areas were performed within 1 yr of the clamp study. Anthropometric measures included height, weight, and waist circumference.

#### Continuous glucose monitoring

All subjects with T1D underwent masked continuous glucose monitoring (MiniMed Gold System; Medtronic, Minneapolis, MN) for the 3 d before the clamp. Continuous glucose monitoring measures included overall glycemic control, hypoglycemia, hyperglycemia, and glycemic variability as previously described (14).

#### Hyperinsulinemic-euglycemic clamp (HEC) visit

A detailed description of the HEC protocol used has been published previously (2). A three-stage HEC was performed using the method of DeFronzo *et al.* (15). Briefly, a primed continuous infusion of insulin was administered at a rate of 4 mU/  $m^2 \cdot min$  for 1.5 h, 8 mU/m<sup>2</sup>  $\cdot min$  for 1.5 h, and then 40 mU/  $m^2 \cdot min$  for the final 1.5 h. An infusion of 20% dextrose was titrated throughout the procedure to maintain blood glucose approximately 90 mg/dl. Arterialized blood was sampled every 5 min for bedside determination of glucose concentration (Analox, Lunenberg, MA). Arterialized blood samples for hormone and substrate measurements were taken at baseline and during the last 10 min of each stage. A steady state was achieved during the last 30 min of the high insulin infusion stage and mean glucose infusion rate [GIR; milligram per kilogram fat free mass (FFM) per minute] during this time was used as the measure of insulin sensitivity.

#### Measurements

Fasting plasma samples for adiponectin measurements were obtained at the time of the clamp visit. Total adiponectin levels were measured by RIA (LINCO Research, Inc., St. Charles, MO) with an intraassay precision of 1.78–3.59% coefficient of variation (CV), and interassay precision of 6.90–9.25% CV. The HMW adiponectin was measured by ELISA (Millipore, Billerica, MA); intraassay precision was 2.5–4.7% CV and interassay precision was 5.8–6.9% CV.

#### **Statistical analyses**

Student's *t* tests were used to compare the means of continuous variables between the T1D and non-DM groups. Factors correlated with total and HMW adiponectin were identified using Pearson correlation coefficients. Linear regression models were run to identify independent predictors of adiponectin levels for both total and HMW adiponectin. The independent association of adiponectin levels with insulin sensitivity was explored using forward selection multivariate regression analysis. SAS version 9.2 (SAS Institute, Inc., Cary, NC) was used for all statistical analyses. A P < 0.05 was considered statistically significant.

# Results

Characteristics of the study participants are presented in Table 1, stratified by diabetes status and gender. T1D and non-DM groups were similar for gender distribution, age, blood pressure, body mass index (BMI), and adiposity (waist circumference, percentage body fat, percentage trunk fat, and visceral fat area). Expected diabetes-group differences were observed in glycosylated hemoglobin, fasting insulin, and fasting glucose. Lipid panels showed lower total cholesterol, triglycerides, and low-density lipoprotein cholesterol, and higher low-density lipoprotein cholesterol in the T1D group compared with the non-DM group. As previously reported (2), subjects with T1D were significantly more insulin resistant than subjects without diabetes, requiring lower GIR of  $5.4 \pm 3.7$  mg/kg FFM per minute in men and  $6.2 \pm 3.4$  mg/kg FFM per minute in women with T1D, compared with  $10.0 \pm 5.2$  mg/kg FFM per minute and  $15.6 \pm 5.0$  mg/kg FFM per minute in their non-DM counterparts. T1D patients had higher mean total and HMW adiponectin (microliters per milliliter) than nondiabetic controls  $(12.3 \pm 5.8 vs. 9.6 \pm 5.5, P = 0.03;$ and  $6.5 \pm 4.6 vs. 4.4 \pm 3.5$ , P = 0.02, respectively) despite having lower insulin sensitivity. Stratified by gender, both

	T1D (n = 39)		Non-DM (n = 47)	
	Man		Man	(II – 47) Women
	(n - 18)	(n - 21)	(n - 20)	(n - 27)
	(11 - 16)	(11 - 21)	(11 - 20)	(11 - 27)
Age (yr)	$47 \pm 10$	$44 \pm 9$	$47 \pm 6$	45 ± 8
Diabetes duration (yr)	$23 \pm 8$	$22 \pm 8$	N/A	N/A
Insulin dose (units/kg • d)	$0.57 \pm 0.13$	$0.57 \pm 0.19$	N/A	N/A
HbA1c (%)	$7.5 \pm 0.9^{a}$	$7.5 \pm 0.9^{a}$	$5.4 \pm 0.3$	$5.5 \pm 0.3$
Fasting insulin ( $\mu$ U/ml)	28.0 ± 16.0 <sup>a</sup>	$36.2 \pm 35.3^{a}$	$10.4 \pm 5.0^{b}$	$7.4 \pm 2.2$
Fasting glucose (mg/dl)	122.7 ± 54.5	$109.2 \pm 19.5^{a}$	99.1 $\pm$ 8.7 <sup>b</sup>	92.9 ± 6.1
Systolic blood pressure (mm Hg)	116 ± 11	$111 \pm 10$	120 ± 9 <sup>b</sup>	109 ± 11
Diastolic blood pressure (mm Hg)	$79 \pm 6^{b}$	73 ± 8	$80 \pm 7^{b}$	73 ± 7
BMI (kg/m <sup>2</sup> )	$28.3 \pm 4.2$	25.8 ± 4.3	27.2 ± 3.6	$25.2 \pm 4.3$
Waist circumference (cm)	$94.7 \pm 9.3^{\circ}$	82.4 ± 11.7	95.6 ± 10.6 <sup>c</sup>	79.5 ± 8.6
Body fat (%)	$24.3 \pm 6.2^{\circ}$	$32.5 \pm 6.7$	$24.2 \pm 3.2^{\circ}$	$33.6 \pm 6.6$
Trunk fat (%)	$25.7 \pm 7.5$	$30.4 \pm 8.6$	$27.4 \pm 4.4^{b}$	31.2 ± 7.6
Visceral fat area (cm <sup>3</sup> )	56.5 (32.9–73.0)	37.5 (25.0-41.4)	60.1 (45.2–87.9) <sup>c</sup>	35.0 (25.0-44.7)
Total cholesterol (mg/dl)	$143 \pm 32^{d}$	$135 \pm 33^{d}$	171 ± 25	171 ± 32
Triglycerides (mg/dl)	$70 \pm 23^{d}$	69 ± 42 <sup>d</sup>	126 ± 73	94 ± 38
LDL cholesterol (ma/dl)	$68 \pm 24^{a}$	$66 \pm 25^{a}$	101 ± 25	93 ± 27
HDL cholesterol (mg/dl)	$61 \pm 31^{d}$	$56 \pm 13$	$45 \pm 9^{\circ}$	$60 \pm 15$
Serum creatinine (mg/dl)	$1.1 \pm 0.2^{b}$	$0.9 \pm 0.1^{a}$	$1.2 \pm 0.2^{b}$	$1.0 \pm 0.1$
Albumin excretion rate (mg/min)	6.0 (4.0-9.4)	3.9 (3.1-6.0)	4.0 (3.4-5.7)	3.3 (3.0-5.2)
GIR (ma/ka FFM per min)	$54 + 37^{d}$	$62 + 34^{a}$	$10.0 + 5.2^{\circ}$	156 + 50
Total adiponectin ( $\mu$ g/ml)	$11.3 \pm 5.6^{d}$	$13.4 \pm 6.1$	$7.3 \pm 4.4^{b}$	$11.9 \pm 6.0$
Total adiponectin	$10.2 + 1.8^{d}$	12.2 + 1.6	$6.2 + 1.8^{b}$	10.4 + 1.7
(geometric mean µg/ml)	10.2 = 1.0	12.2 = 1.0	0.2 = 1.0	10.1 = 1.7
HMW adiponectin (µg/ml)	46+29 <sup>b, d</sup>	75+47	2 7 + 1 4 <sup>c</sup>	58+40
HMW adiponectin	40 + 24	59 + 22	$2.7 \pm 1.1$ $2.2 \pm 2.0^{b}$	44+23
(geometric mean, $\mu$ g/ml)	1.0 = 2.4	5.5 - 2.2	2.2 - 2.0	= 2.5

#### **TABLE 1.** Characteristics of study participants by diabetes status and gender

Data are presented as mean ± sD, percentage; median (interquartile range). HbA1c, Glycosylated hemoglobin; LDL, low-density lipoprotein; HDL, low-density lipoprotein.

<sup>a</sup> P < 0.001 for comparison by diabetes group within sex.

<sup>b</sup> P < 0.05 for comparison by sex within diabetes group.

 $^{c}P < 0.001$  for comparison by sex within diabetes group.

 $^{d}$  P < 0.05 for comparison by diabetes group within sex.

total and HMW adiponectin levels were higher in men with T1D compared with non-DM control men (11.3  $\pm$ 5.6 vs. 7.3  $\pm$  4.4 and 4.6  $\pm$  2.9 vs. 2.7  $\pm$  1.4 µg/ml, respectively, P < 0.05 for both), although values were not significantly different in the women. The magnitude of the differences observed in total adiponectin was larger than that in HMW adiponectin, consistent with increases in all adiponectin multimers in the group with T1D. However, most (62%) of the increase in total adiponectin was attributed to an increase in the HMW adiponectin isoform.

To identify possible determinants of total and HMW adiponectin levels, associations between these measures and variables thought likely to influence adiponectin levels were measured within each group. Total adiponectin levels were positively associated with older age (r = 0.37, P < 0.05) and greater diabetes duration (r = 0.42, P < 0.05) and negatively associated with daily weight-adjusted insulin dose (r = -0.50, P < 0.05) in subjects with T1D. Expected negative correlations were found between total adiponectin and adiposity measures including BMI,

percentage trunk fat, and visceral fat area. The associations with HMW adiponectin were similar to those observed with total adiponectin. No associations were observed between adiponectin and renal function measures including serum creatinine and albumin excretion rate.

The correlation between GIR and both total adiponectin (r = 0.33, P < 0.05 for patients with T1D; r = 0.29, P < 0.05 for controls) and HMW adiponectin (r = 0.47, P < 0.05 for patients with T1D; r = 0.28, P = 0.055 for controls) was similar for the two groups (Fig. 1). However, at any given level of total or HMW adiponectin, patients with T1D had decreased insulin sensitivity (lower GIR) compared with nondiabetic controls (P < 0.0001 for the difference in GIR by diabetes status when adjusted for either total or HMW adiponectin). In contrast, insulinstimulated free fatty acid suppression was not associated with either total or HMW adiponectin (data not shown).

The relationships between total and HMW adiponectin and glycemic control were explored in subjects with T1D using data obtained during continuous glucose monitor-



**FIG. 1.** Relationship between total (A) and HMW (B) adiponectin and GIR in subjects with T1D and non-DM controls. Scatterplot and predicted regression lines show the relationship between adiponectin and GIR. A presents total adiponectin and B presents HMW adiponectin. Data for subjects with T1D is presented by *open circles* and *gray lines*. Data for non-DM control subjects is presented by *star symbols* and *black lines*. P < 0.0001 indicates the difference in GIR by diabetes status when adjusted for either total or HMW adiponectin.

ing. Neither total nor HMW adiponectin levels were correlated with any included measure of glycemic control or glycemic variability.

In the forward stepwise regression analyses, T1D status, older age, female gender, and percentage trunk fat were independently associated with both total and HMW adiponectin. However, T1D status remained a strong independent positive determinant of both adiponectin measures and was associated with 2.6  $\mu$ g/ml higher mean total adiponectin (R<sup>2</sup> = 0.21, P = 0.03) and 1.6  $\mu$ g/ml higher mean HMW adiponectin levels (R<sup>2</sup> = 0.41, P = 0.02).

# Discussion

We measured the correlation between total and HMW adiponectin and insulin sensitivity in subjects with T1D

and non-DM controls. Consistent with previous reports (6, 16), mean total and HMW adiponectin were higher in patients with T1D compared with non-DM controls. We found that group differences in adiponectin levels are largely but not entirely due to increased HMW adiponectin in T1D patients. As observed in populations without T1D, total and HMW adiponectin levels were positively associated with insulin sensitivity within each group. Thus, among patients with T1D, higher adiponectin was associated with higher insulin sensitivity. However, despite elevated mean adiponectin levels, patients with T1D had decreased insulin sensitivity compared with nondiabetic controls. In fact, insulin sensitivity was lower for subjects with T1D compared with non-DM controls at any given level of total or HMW adiponectin, suggesting an increased set point or dysregulation of adiponectin function in the subjects with T1D. Our findings are consistent with a relative adiponectin resistance among subjects with T1D and support the hypothesis that factors unrelated to adiponectin contribute to decreased insulin sensitivity in this population.

Although the expected correlations between adiponectin and age, gender, and adiposity were found, these variables did not explain the observed increased levels of adi-

ponectin in the T1D population. Adiponectin has previously been reported to be inversely associated with renal function in patients with T1D (17). However, we did not observe this association in our cohort of patients with normal renal function, and renal function did not explain the observed increased levels of adiponectin in the T1D group. Further mechanistic studies are required to elucidate the causative factors of increased total and HMW adiponectin in subjects with T1D.

Recent studies have reported a positive association between adiponectin and cardiovascular as well as all-cause mortality in patients with T1D (18, 19) unexplained by renal dysfunction, catabolism, inflammation, or preexisting cardiovascular disease. Our finding of an expected positive association between adiponectin and insulin sensitivity despite decreased insulin sensitivity in patients with T1D at any given level of adiponectin suggests a complex association of total and HMW adiponectin with insulin sensitivity and health outcomes in people with T1D.

Our findings are limited by a relatively small sample size, although the CACTI study of 86 subjects is large for a clamp study, especially given the complexity of such studies in subjects with T1D. In addition, our data are cross-sectional and we are not able to determine causation based on our study design.

In summary, we find that both total and HMW adiponectin are increased in patients with T1D compared with nondiabetic controls, and this observation is not explained by group differences in age, gender, or fat distribution. Among patients with T1D, adiponectin levels are positively correlated to insulin sensitivity, similar to the relationship among adults without diabetes. However, insulin sensitivity is lower for patients with T1D at any given level of total or HMW adiponectin suggesting a change in total and HMW adiponectin set point in these patients.

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