



Advanced Routes of Biological and Bio-electrocatalytic Carbon Dioxide (CO₂) Mitigation Toward Carbon Neutrality

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Specialty section:

This article was submitted to
Carbon Capture, Storage, and
Utilization,
a section of the journal
Frontiers in Energy Research

Received: 24 February 2020

Accepted: 01 May 2020

Published: 12 June 2020

Citation:

Kondaveeti S, Abu-Reesh IM,
Mohanakrishna G, Bulut M and
Pant D (2020) Advanced Routes of
Biological and Bio-electrocatalytic
Carbon Dioxide (CO₂) Mitigation
Toward Carbon Neutrality.
Front. Energy Res. 8:94.
doi: 10.3389/fenrg.2020.00094

Changes in the environment due to multiple factors, such as combustion of fossil fuels, heating, transportation, deforestation, etc., have led to more greenhouse gases in the atmosphere, which eventually led to a rise in global temperatures. Carbon dioxide (CO₂) is the major factor for the rapid rise in global temperature. One of the most encouraging technological advances to address global warming is to transform CO₂ into value-added commodities that offer a win-win strategy. In this regard, intensive research has been pursued around the world for development of feasible systems in product recovery or product synthesis from CO₂-rich industrial emissions. We envision that the biological CO₂ reduction or conversion process can be beneficial for developing carbon-neutral technologies. The integration of CO₂-emitting industrial technologies with CO₂-converting biological systems can be helpful in achieving sustainable value-added products with no or minimal loss of energy and materials that are assuring for improved economics. The CO₂-converting bioprocesses can be directly integrated with the processes emitting a high amount of CO₂. This symbiotic integration can make the whole process carbon neutral. Herein, this review highlights an insight on research activities of biological CO₂ mitigation using photo catalysts (algae and photo bacteria), an anaerobic biocatalyst (bacteria), gas fermentation, and an enzymatic catalyst. Perspectives and challenges of these technologies are discussed.

Keywords: CO₂ capture and utilization, algae, bioenergy, greenhouse gases (GHG), gas fermentation, microbial electrosynthesis (MES), CO₂ reduction pathways

INTRODUCTION

Expansion of industries and world population are leading to an increase in the release of greenhouse gases, resulting in accelerated global warming (Kumar et al., 2016; Effendi and Ng, 2019; Youn et al., 2019). The higher levels of carbon dioxide (CO₂) emissions into the environment have become a major contributor to global warming (Figure 1A). Both reduction in CO₂ emission and capture of CO₂ are critical to reduce global warming (Benhelal et al., 2013). In this regard, carbon capture, sequestration, and utilization have proven to be efficient options for decreasing the atmospheric CO₂, which can be regarded as a waste-to-energy process (Liu et al., 2016). This energy

process can be either fuel generation or chemical and material production. Several industrial technologies have been developed and are operational, such as the alkaline adsorption process (CO₂ capture), mineralization process, landfill process (CO₂ sequestration), urea synthesis, methanol production, phenol carboxylation, epoxide carboxylation, and pyrrole carboxylation (CO₂ utilization) (Shi et al., 2015; Liu et al., 2016). Global research on biological CO₂ conversion for renewable and sustainable energy generation is increasing at a rapid pace in diversified research disciplines (Figure 1B). In addition, many bench laboratory-scale or pilot-scale technologies are also being developed to increase the performance of CO₂ capture, sequestration, and utilization toward value addition (Shi et al., 2015).

Capture and transformation of CO₂ to value-added products, such as biofuel, are of special interest, and several countries have been developing policies by providing subsidies and legislative incentives. For instance, the Renewable Energy Directive (RED)

under the European Union (EU) is moving toward a vision of 2020, which is targeting the generation of 10% of the energy required for the transportation sector that should be replaced with renewable biofuels (Chang et al., 2016; Sadhukhan et al., 2016). Likewise, in the United States, the Renewable Fuel Standard is targeted at increasing biofuel generation by 36% with a vision of 2022 (Rana et al., 2011). About 58% of biofuel generation is mainly from second- (e.g., organic waste, plant waste) and third-generation (e.g., algae biomass) biofuels.

Biological, chemical, photo-chemical, electrochemical, and enzymatic technologies have been developed for fixing and conversion of CO₂ to renewable energy generation (Benemann, 1997; Bond et al., 2001). However, a majority of the processes are linked with high energy input, which is making the process economically unfavorable. Among these methods, a biological system for CO₂ utilization using bacteria as a catalyst, which is of low cost and self-regenerative, is of particular interest and importance (Benemann, 1993; Jiang et al., 2019). Hence, the

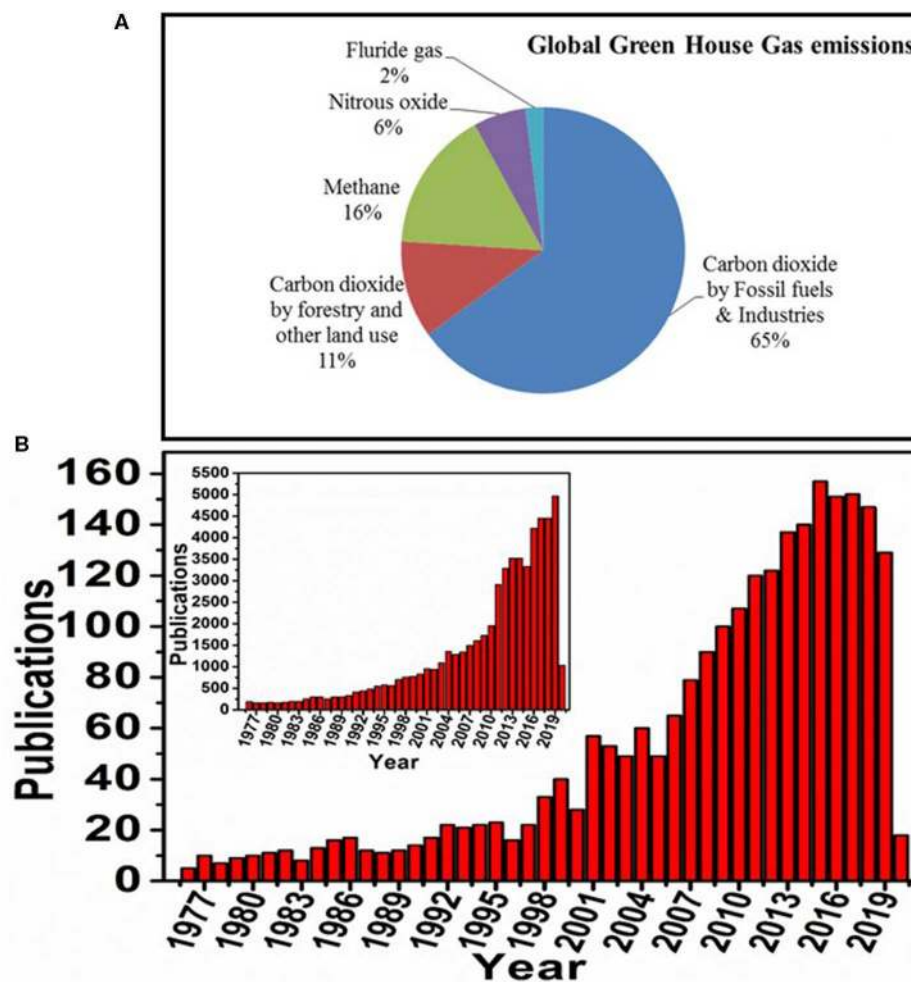
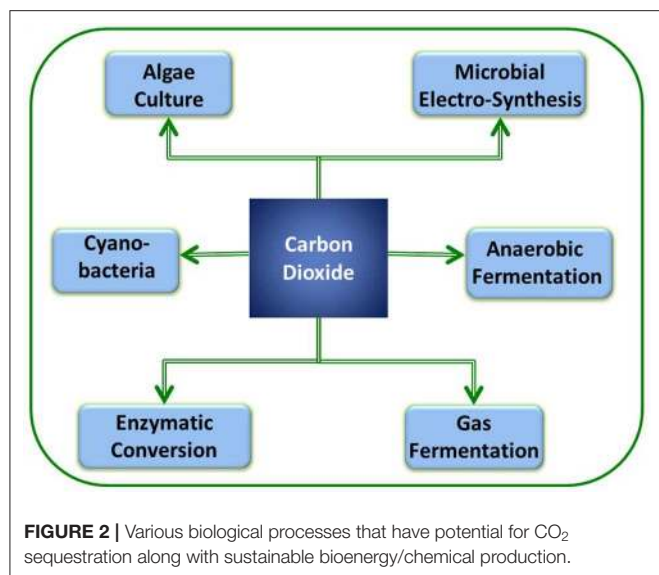


FIGURE 1 | (A) Graphical representation of global greenhouse gas emissions (source: <https://tinyurl.com/EPA-GHG-Data>), **(B)** number of articles published on biological conversion of CO₂ to value-added production (source: Scopus; key words: Biological carbon dioxide reduction or Biological carbon dioxide conversion, as on date: 21/04/2020). Inset figure exhibits the number of published articles on carbon dioxide reduction or carbon dioxide conversion irrespective of field.



potential of numerous biological CO₂ utilization methods to decrease the atmospheric CO₂ should be investigated further with an energy-neutral and biorefinery approach (Venkata Mohan et al., 2016; Hou et al., 2019). The petroleum refinery approach has been well-known for a long time for energy generation. However, the use of a CO₂ biorefinery approach is exciting and offers a sustainable and eco-friendly process (Packer, 2009; Venkata Mohan et al., 2016). Nonetheless, this area of biological CO₂ utilization research is obstructed with several questions regarding its efficiency, establishment cost, and fulfilling of current and future energy demands. Despite these criticisms, the research on utility of CO₂ should be increased toward a carbon-neutral footprint and energy neutrality. Therefore, the major objective of this review is to highlight the biological CO₂ utilization methods that can be implemented to generate bio-based products and elaborate on the biology of sequestration methods of different systems/microbes.

BIOLOGICAL MITIGATION OF CO₂

Photosynthetic CO₂ Reduction

Recently, several researchers have concentrated on sequestration of CO₂ using light energy to generate value-added products, including fuels, chemicals, and other materials. Most of the photosynthetic biological systems include oxygenic or anoxygenic photosynthesis routes (Hunt et al., 2010). Below, we discuss the CO₂ reduction using algae and photosynthetic bacteria (Figure 2). Algae cultivation for CO₂ conversion provides numerous advantages and liabilities that are documented in Table 1.

CO₂ Sequestration/Assimilation Using Algae

In the past decade, several studies have proven that CO₂ mitigation with algae is found to be a sustainable process with simultaneous generation of high calorific products, such as biodiesel, pigments, fatty acids, etc. (Chisti, 2007;

Demirbas, 2010) (Table 2). The potential utilization of algae is mainly attributed to its wide distribution, high biomass production, capability to adjust in adverse conditions, swift carbon uptake and utilization, and capability to generate value-added products (Figure 3). Both macro and micro algae have the capability to metabolize inorganic carbon by a photoautotrophic mechanism using carbonic anhydrase enzyme (CAE) (Clarens et al., 2010). The generated NADH₂ from the electron transport chain combines with the RuBisCo (Ribulose-1,5-bisphosphate carboxylase/oxygenase, provided by CAE) and helps in carbohydrate generation from CO₂ and provides the reducing power in the Calvin cycle for glucose synthesis (Shi et al., 2015).

Selection of suitable microalgae for CO₂ fixation

It is known that a higher concentration (generally greater than 5%) of CO₂ adversely impacts the growth and production rate in most microalgae (Clarens et al., 2010). Therefore, numerous research studies are being carried out to isolate a suitable algal strain for direct and efficient conversion of industrial CO₂ into products such as agar and alginate, etc. (Table 3). The criteria for selection is not only tolerance of CO₂, temperature, and toxic compounds, but also for attaining high growth rates and maximum cell densities (Kratz and Myers, 1955; Allen and Stanier, 1968).

Freshwater microalgae *Chlorella* HA-1 exhibited a maximum growth rate when it was evaluated at 5 and 10% of CO₂ (Watanabe et al., 1992). However, the growth was inhibited with further increases in CO₂ concentration up to 20%. This study proposed that *Chlorella* HA-1 can be used to treat industrial flue gas with a maximum CO₂ concentration of 10% (Watanabe et al., 1992). This strain exhibited a variation in response with a change in temperature and pH. *Chlorococcum littorale* culture isolated from a saline water pond was able to show an almost similar growth rate from 5 to 60% of CO₂, which is a much higher CO₂ concentration than in flue gas (20%). When the CO₂ concentration was boosted to 70%, a decrease in growth rate was identified. As this strain exhibits high growth at high CO₂ concentrations, it can be readily applied for industrial flue gases (Kodama et al., 1993). There were also several other strains of micro algae that have exhibited a high growth rate in adverse conditions, such as extreme pH and temperature (Kumar et al., 2011). The most industrially applicable algal product is lipids, which can be used as a source for biofuel productions (e.g., biodiesel) (Silva et al., 2014). Therefore, the assortment of strains for generation of high lipid generation and augmenting the culture conditions, such as light, temperature, and pH, are vital to enhance the productivity (Csavina et al., 2011; Georgianna and Mayfield, 2012). The unoptimized pH, during algae growth, can affect the distribution of carbon species and carbon availability. Also, at extreme pH, it can directly affect the metabolic activity of algae.

Flue gases from fossil fuel power plants and industries discharge elevated strengths of CO₂, varying from 10 to 20%, along with a biologically significant amount of NO_x and SO_x. The dosing of flue gases to algae ponds has shown to increase biomass yields by 3-fold and high energy generation (Hanagata et al.,

TABLE 1 | Diversified advantages and liabilities of algae based biofuels systems.

Advantages	Liabilities
Sustainable, renewable, and environmental friendly	Utilization of fossil fuel for process in small quantities
Low carbon to high carbon sources namely carbohydrates, lipids, and proteins	Indefinite availability of sustainable algae resources for biofuel generation
High value feedstock as a replacement to terrestrial biomass	Lack of origin and source certification for global monitoring and control of algae fuel
Non-toxic and non-edible	Regionally constrained market structures for biofuels
Diversification of fuel and energy generation	Production costs for operation of algae biofuel plant
Conservation of fossil fuel, High CO ₂ capture, energy conversion through photosynthesis	Economics of process need to be improved
High productivity and yields with rapid growth rates	Harmful algae blooms in global waters
Easily cultured, readily, and rapidly bioengineered	Utilization of arable land
Easy adaptability at various climatic conditions	Unclear utilization of type of waste materials
Can grow almost everywhere such as ponds, wastelands etc.,	May cause Neurotoxic problems
Restoration of contaminated areas and ponds with quick bioremediation with degradable sources	Limited expertise in optimization of industrial biofuel generation

TABLE 2 | Products from algae using CO₂ as a substrate and their applications.

Products	Applications	References
Agar	Food ingredient, brewing agents, biological/microbiological applications	Lee et al., 2017
Alginate (Kerogen)	Medical, paper, and cosmetic	Petersen et al., 2008
Antioxidants (Fucoxanthin)	Preservative in food, chemical, and cosmetic industries	Foo et al., 2017
Beta carotene and Carotenoids	Supplement in vitamin C and precursor for vitamin A	Fernando et al., 2016
Biobutanol, bioethanol, biodiesel, biomethane, biohydrogen,	Bioenergy and Biofuels	Xia et al., 2016
Biochar	Combustion and agriculture	Nautiyal et al., 2016
Biosorbents	Ion exchange materials that have the capacity in binding toxic heavy metals	Bulgariu and Bulgariu, 2016
Chemicals	Medical and industrial application	Trivedi et al., 2015
Cosmetics	Water binding material, antioxidants, and skin food	Wang et al., 2015
Digester residue	Compost and Vermi-compost	Vassilev and Vassileva, 2016
Feed	Animal and biological feedstock	Wang C. et al., 2015; Mark Ibekwe et al., 2017
Fertilizers	Rich in N,P and K minerals, as fertilizer	Rengasamy et al., 2016
Food	Juices, sauce, noodles, beverages, cheese, etc.,	Vigani et al., 2015
Preservatives	Food industry	Kerdudo et al., 2016; Lowe and LaLiberte, 2017
Pharmaceutical (Omega 3 and Omega 6)	Pharma industry	El Gamal, 2010; Samarakoon and Jeon, 2012
Pigments (phycocyanin, phycoerythrin)	Synthesized pigments can be helpful in textile industry	Venil et al., 2013; Holkar et al., 2016; Li et al., 2017

1992; Maeda et al., 1995). Fortunately, the benefits in injecting the flue gas to algae growth can be more helpful than the growth impacts attributed in the photorespiration process at high CO₂ concentrations. Recent studies have shown that direct submission of flue gases into algal ponds exhibited a 30% increase in biomass yields over using the equivalent concentration of CO₂ (Tsuzuki et al., 1990; Singh and Singh, 2014). This might be due to the existence of additional nutrients (oxides of nitrogen and sulfur) present in flue gas. This competence of CO₂ encapsulation by algae can vary according to the algae physiology, pond chemistry, and temperature. About 80 to 99% of CO₂ capture can be obtained under optimized conditions with gas residence times as short as 2 s (Keffer and Kleinheinz, 2002; Lee and Lee, 2003). For a 200 MW/h power plant, it has been noted that 3,600

acres of algae pond would be required to capture 80% of the plant's CO₂ emissions during the daylight hours, assuming a productivity rate of 20 g dry biomass/m²/d. Therefore, locating the algae ponds next to CO₂ point suppliers provides numerous potential low-cost and energy-saving advantages (Sayre, 2010) with a minimal carbon footprint.

Challenges in using algae for CO₂ capturing/sequestration

On an industrial scale, algae are typically growing in photo bioreactors, open ponds, or raceway ponds (Wang, 2014). The use of open ponds can be simpler in architecture and operation. However, these are limited with a large surface area necessity, elevated cultivation cost, high chance of contamination, and low productivity. Photo bioreactors with closed systems have been

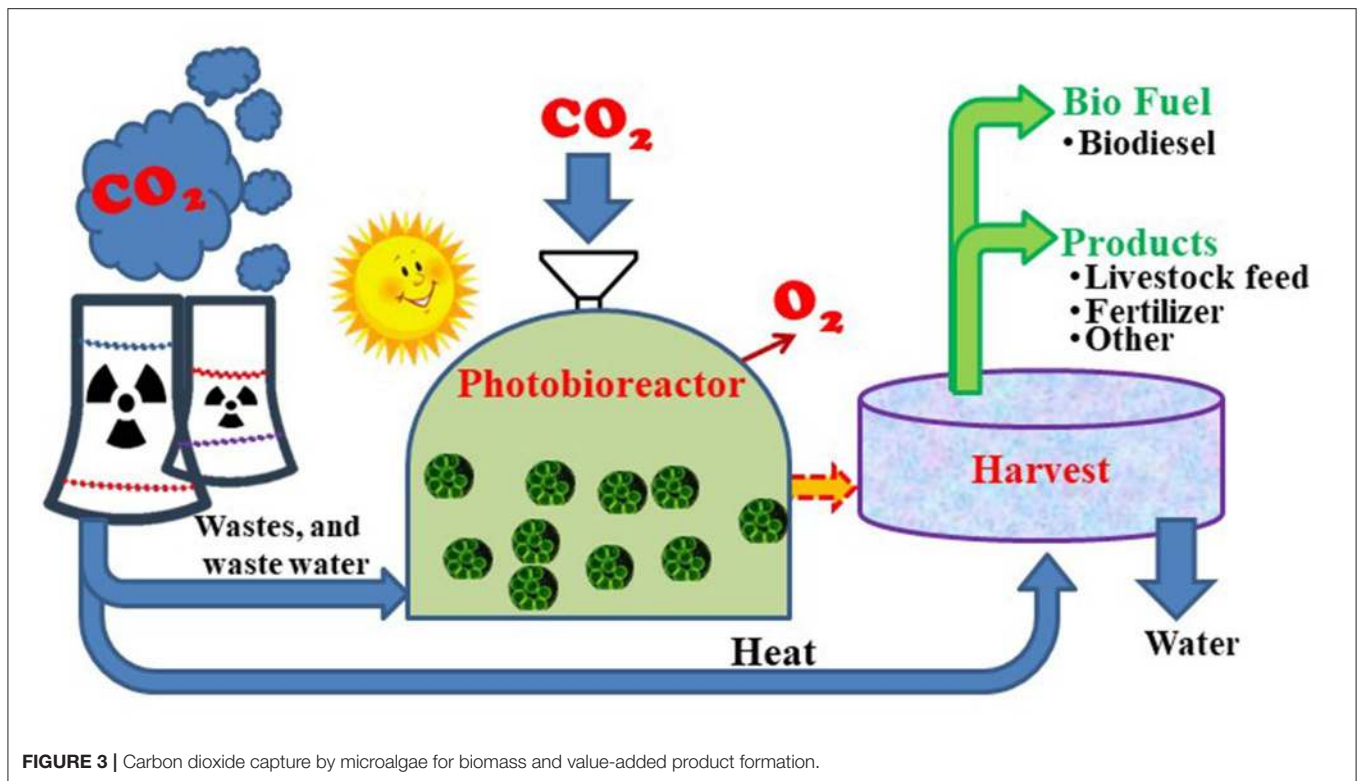


FIGURE 3 | Carbon dioxide capture by microalgae for biomass and value-added product formation.

proposed to overcome the above limitations (Li et al., 2014). A detailed review study published by Kumar et al. (2011) provided great aspects in the design of photo bioreactors and challenges noticed in industrial algae growth. Flue gas compositions are made up of different types of gases along with CO₂, which is viable for the growth of few algae. However, high temperatures, fluctuating flue gas composition, and the presence of sulfur and nitrogen can be toxic to some algal strains. Therefore, the selection of suitable algal processes and strains is needed. One of the other key aspects that play a crucial role in algae development is light intensity (Sorokin and Krauss, 1958). Insufficient light can lead to poor growth, and an increase in light intensity can also show adverse effects on the growth, known as photo inhibition (Xu et al., 2016). The mixing of cell cultures requires an extensive amount of energy input and constitutes a major portion of operational costs (Cloot, 1994). Stirring of cultures is important to reduce the mass transfer limitations occurring in photo bioreactors, which also favorably helps to eliminate oxygen accumulation. Several other advancements have been made to overcome the challenges that are discussed.

In general, the CO₂ uptake/capture with algae is calculated based on carbon concentration in influent and effluent (Anbalagan et al., 2017). However, it is difficult to consider that whole CO₂ uptake is carried out by algae and is utilized for its growth. But the carbon content in biomass can deliver a more precise value on the quantity of CO₂ consumption by algae. This scenario can only be useful if the provided culture media does not have an additional organic carbon source, other than inorganic carbon, such as bicarbonate or CO₂. Based on the following, CO₂

capture by algae can be calculated as

$$R_{CO_2} = P \cdot C_{CO_2} \frac{M_{CO_2}}{M_C}$$

where R_{CO_2} is the fixation rate of CO₂ concentration (g.l⁻¹ d⁻¹), P is the production of biomass concentration per day (g.l⁻¹ d⁻¹), C_{CO_2} is increase in carbon content of algae biomass from CO₂ utilization, M_{CO_2} is the molecular weight of CO₂, and M_C is the molecular mass of carbon (de Moraes and Costa, 2007a).

Tang et al. (2011) analyzed the carbon content and calculated the rate of CO₂ biofixation for different algae species that were grown in modified Blue-Green 11 (BG11) medium. In this study, they calculated the carbon capture rate by varying the CO₂ concentration from 0.03 to 50%. The carbon content in biomass was found to be around 50%, whereas the CO₂ fixation was in the range of 0.105 to 0.288 g.l⁻¹ d⁻¹.

Commercialization of microalgal technologies

To commercialize products from microalgal technologies, a designed biorefinery in a circular loop was found to be a sustainable option. A self-sustainable biorefinery (SSB) model of microalgae maximizes resource utilization along with zero waste discharge (Venkata Mohan et al., 2020). Utilization of defatted or de-oiled microalgal biomass (DMB) for other processes also helps to develop sustainable biorefineries. DMB can be used as a substrate for other biofuel generation, such as biomethane, biohydrogen, and bioethanol (Maurya et al., 2016). Thermochemical liquefaction, pyrolysis, and gasification are non-biological processes in which DMB can be used as substrate

TABLE 3 | Summary of diverse algal species for CO₂ fixation and conversion rates.

Type of algae	Conversion of CO ₂ (μmol/ g of CO ₂ /h)	References
<i>Nannochloris</i> sp.	57–569	Negoro et al., 1991
<i>Phaeodactylum tricornutum</i>	27–267	Negoro et al., 1991
<i>Chlorella</i> sp.	125–1,246	Sakai et al., 1995
<i>Synechocystis</i>	142–142	Kurano et al., 1995
<i>Chlorella vulgaris</i>	7–3,352	Scragg et al., 2002; Fan et al., 2008
<i>Scenedesmus</i> sp.	44–436	Jin et al., 2006; Yoo et al., 2010
<i>Spirulina</i> sp.	37–373	de Morais and Costa, 2007a,b
<i>Anabaena</i> sp.	137–1,373	González López et al., 2009
<i>Euglena gracilis</i>	36–362	Chae et al., 2006
<i>Microcystis</i> sp.	46–493	Jin et al., 2006
<i>Botryococcus braunii</i>	1,100	Murakami and Ikenouchi, 1997
<i>Synechocystis aquatilis</i>	1,500	Murakami and Ikenouchi, 1997
<i>Dunaliella tertiolecta</i>	272–313	Kishimoto et al., 1994; Sydney et al., 2010

to produce fuels and chemicals (Sarkar et al., 2015; Maurya et al., 2016). Compared to conventional methods, the microalgae industry is concerned about the two fundamental hurdles, which can be identified as low productivity and high costs (Maurya et al., 2016). Subjecting residual biomass or DMB (post lipid extraction) in various fields improves the economic feasibility and commercialization of algal bioprocesses (Singh et al., 2011; Rashid et al., 2013). The biomass of microalgae can be used as inoculum for new processes that reduce the maintenance of parent cultures. This helps in sustainable operation of microalgal bioreactors (Sarkar et al., 2015). Apart from utilizing DMB in fuel and chemical production, it can also be used as feed supplement and biofertilizer. DMB is rich in protein and essential amino acids that could be utilized as livestock and aquaculture feeds or as a dietary supplement. DMB has also been tested to replace fishmeal from wild fish caught in an ocean environment. Thus, it can mitigate ocean resource depletion (Zhang and Kendall, 2019). In wastewater treatment, DMB can be used as an active biosorbent to remove dyes/color and heavy metals (Maurya et al., 2016). During the extraction of lipids from algal biomass, the biomass used to rupture, which has a higher surface area than the raw biomass. This helps for the efficient functioning of DMB as an active biosorbent (Dong et al., 2016).

Bioenergy production with carbon capture and storage (CCS) plays a crucial role in CO₂ mitigation. Combination of bioenergy production and CCS help to keep CO₂ in geological reservoirs. Algae have high potential to store CO₂ as biomass, which is due to high photosynthetic efficiencies and high biomass yields (Moreira and Pires, 2016). Microalgal cultivation was designated as a negative emission technology (NET), which can reduce the impacts of ocean acidification and anthropogenic climate

change (Moreira and Pires, 2016). Since the scope of microalgae is versatile and possesses the expansive possibility of microalgae cultivation with a self-sustainability approach, it is designated as a blue-bioeconomy.

Autotrophic lipid/biodiesel generation using algae

World energy is mainly dependent on electricity and fuel segments. Still, both segments are needed to implement substantial carbon-neutral techniques to reach international legislated standards in regard to the CO₂ emissions target (International Energy Agency, 2019). Currently, 38% of the world's energy is generated in the form of electricity from fossil fuels, and still, these sectors are pursuing research in development of low-range carbon emission technologies, such as solar, wind, geothermal, hydroelectricity, etc. On the contrary, transportation fuels account for larger consumption (Dudley, 2018). Despite the significance of fuels, CO₂-neutral biodiesel, bioethanol, and biohydrogen are still being less used due to several concerns in biofuel production and environmental limitations. In this aspect, second-generation biomass, such as using lignocellulose and algae for fuels, have tremendous potential in a clean energy market with a value of US\$500 billion by 2050 (Schenk et al., 2008). Furthermore, based on several research studies, it is well-known that microalgae biofuel production systems exhibit high net energy balance, high water efficiency, and less land requirement in comparison to first-generation biofuels (plant biomass) (Benedetti et al., 2018).

In consideration of oil extracted from plant biomass vs. algal biomass, palm oil can generate a maximum of 4–5 vs. 30 ton/ha/yr, thus yielding a 5-fold higher product in comparison to a plant crop during similar posttreatment of biomass (Benedetti et al., 2018). In terms of area, algae are known to generate about 15–300 times more oil for biodiesel (Schenk et al., 2008). In contrast to crop plants' single harvesting process, algae allows multiple harvesting cycles in the same duration with high yield and greater light and CO₂ capture efficiencies.

Different oil concentrations are noted with various algae species, which is due to differences in the amount of protein/carbohydrates/fats. For instance, *S. maxima* has about 60–70% w/w proteins, *Porphyidium cruentum* generates around 57% proteins, and *Scenedesmus dimorphus* generates up to 40% w/w of lipids (Nigam and Singh, 2011; Venkata Mohan et al., 2011) (Table 4). In a few species of algae, such as *Botryococcus braunii* and *Chlorella protothecoides*, terpenoid hydrocarbons and lipids are noted, which can be further converted to short-chain hydrocarbons, similar to that of crude oil (Carlsson and Bowles, 2007). On the basis of prevalence of lipids or sugars, algae can be a wonderful substrate for biodiesel or bioalcohol, respectively (Kondaveeti et al., 2014a). Therefore, algae can be a great resource as a biofuel substrate under certain conditions. Several studies have employed different strategies for increasing lipid concentration of algae, such as temperature, CO₂ concentration, light strength/intensity, nutrient starvation, metal, and salinity stress. It is well-known that adequate light intensity helps in overproduction of microalgae lipids. This might be because a sufficient amount of light is beneficial for storing an excess of photo assimilates, which can be further transformed

TABLE 4 | Generation of biomass by pure microalgae cultures at specific CO₂ concentrations.

Algae type	Concentration of CO ₂ supply (%)	Generated biomass (g/l/d)	References
<i>Chlorella vulgaris</i>	0.03–6	0.21	Chinnasamy et al., 2009; Bhola et al., 2011
<i>Chlorococcum littorale</i>	N/A	9.2	Chen et al., 2009
<i>Scenedesmus obliquus</i>	5–15	1.14–2.3	de Morais and Costa, 2007c; Kaewkannetra et al., 2012
<i>Spirulina platensis</i>	10	2.91	Ramanan et al., 2010
<i>Nannochloropsis oculata</i>	2–15	0.37–0.48	Chiu et al., 2009
<i>Chlorella sp. KR-1</i>	10–70	0.6–0.7	Sung et al., 1999

into chemical energy (Zhu et al., 2016). The *Nannochloropsis* species of algae experienced the maximum accumulation of lipids (47% of dry weight) under light intensity of 51,800 lux (Zhu et al., 2016). In another study with *Scenedesmus abundans* by Mandotra et al. (2016) and Zhu et al. (2016), they noticed a 32.7% increase in lipid accumulation with an upsurge in light intensity from 3,000 to 6,000 lux. When light intensity of 5,000 and 3,000 lux were maintained, the lipid content was found to be 27 and 21%, respectively. It is recognized that higher lipid concentration is noted at maximum photosynthetic efficiency, which occurs at the light saturation point (Zhu et al., 2016). Hence, the extreme light intensity can cause photo inhibition, leading to damage of algal photo systems and, thus, reducing lipid accumulation. In the case of temperature, the optimal value for higher lipid generation achieved does vary from species to species. *C. vulgaris*, a microalgal species, achieved a maximum lipid concentration at 25°C. The decrease in external temperature led to an obvious decrease in lipids (Converti et al., 2009). Microalgae species *S. obliquus* had a lipid concentration of 18 and 40% of dry weight when the temperature was at 20 and 27.5°C, respectively (Xin et al., 2011). This suggests that an increase in temperature can increase total lipid content. However, this does not mean that all types of lipids experience an increase in concentration. A recent review by Zhu et al. (2016) provides detailed strategies for lipid enhancement.

Biodiesel is being developed as one of the most vital biofuels as basically all industrialized vehicles used for agriculture, transportation, and trade are diesel/fossil fuel-based. With an increase in algae biodiesel production, several studies had endeavored to calculate the cost of oilgae (algae oil) from large farms. Studies by Benemann and Oswald (1996) reported that the cost of oilgae would ranging from US\$39 to US\$69 per barrel from 4 Km² open ponds either by using pure CO₂ or flue gas from coal power stations with productivity in a range of 30 to 60 g/m²/day with a 50% algal lipid yield (Benemann and Oswald, 1996). In recent studies, it is noted that the production of oil from algae would cost around US\$66–153 per barrel (Nagarajan et al., 2013). Companies generating oilgae are not optimistic in terms of production cost. Seambiotic Ltd. calculated that the cost of dried algae would be US\$0.34 per kg with a productivity and total lipid content of 20 g/m²/d and 8~40%, respectively. Presuming the standard yield of lipid to be 24% without any additional cost, this would be equals to US\$1.42 per kg for lipid extractions, which is equal to US\$209 per barrel. With an increase in yield of lipids to 40%, the lipid extraction would cost around

US\$0.85 per kg, which is equal to US\$126 per barrel, which points to an increase in lipid concentration decreasing oil prices. Accounting for inflation in crude oil prices in 2017, the oil prices seem to be stabilizing around US\$51 per barrel (<https://www.eia.gov/todayinenergy/prices.php>, 2017). These estimates suggest that oilgae diesel production needs to be further improved to be economically viable and to compete with conventional fuels. However, the industrial production of biodiesel using algae is at early stages, and there is much scope for decrease in cost and increase in performance.

Reduction of CO₂ Using Photosynthetic Cyanobacteria

Most types of photosynthetic bacteria derive energy from ATP, which helps in the conversion of CO₂ to biomass and other products (Hatch and Slack, 1970). In comparison to eukaryotic plants and algae, photosynthetic bacteria have simpler photosystems due to the presence of pigments on their cytoplasm rather than specialized organelles (Pfennig, 1967). Photosynthetic bacteria are classified as five phyla: cyanobacteria, proteobacteria, chlorobi, choloflexi, and firmicutes (Pfennig, 1967). Cyanobacteria are capable of fixing atmospheric nitrogen and carbon. Similar to algae, they are distinct and broadly distributed and exist as biofilms or as suspended planktonic cells. Several scientists believe that cyanobacteria played a decisive role in atmospheric formation by decreasing CO₂ concentration and increasing oxygen (**Figure 1**) (Kasting and Siefert, 2002). In the present days also, cyanobacteria are found to be a key player by accounting for 20~30% of Earth's photosynthetic activity. Owing to their simpler nature, cyanobacteria are the most efficient in atmospheric carbon utilization over algae (Berman-Frank et al., 2003). However, cyanobacteria are still limited in biomass yield as compared to algae without any structural or genetic modifications.

In recent years, the production of ethanol using microbes has gained considerable attention. In general, the biomass from photosynthetic microbes is collected and subsequently converted to biofuel by solventogenesis. This process was found to be inefficient due to lower conversion efficiency (Lau et al., 2015). Thus, intense research is being carried out on direct conversion of CO₂ to fuel by photosynthetic bacteria. Although few cyanobacterial species are capable of generating ethanol in smaller concentrations as a product in biological fermentation, it is required to boost production rates as an economically viable process. Dexter and Fu (2009) made

an attempt to generate ethanol by introducing the genes of pyruvate decarboxylate (pdc) and alcohol dehydrogenase II (adh) from *Z. mobilis* into the chromosomes of *Synechocystis* sp. PCC6803, which achieved 550 mg/l of ethanol. Acetyl Co A-dependent 1-butanol production by *Synechococcus elongates* demonstrates its capability in generation of butanol through an autotrophic mechanism (Kusakabe et al., 2013). This species was metabolically engineered to generate isopropanol by using CO₂ and light to generate 26.5 mg/l of product following 9 days of cultivation (Kusakabe et al., 2013), whereas *Clostridium beijerinckii* generated 1.5 g/l through a heterotrophic mechanism (Chen and Hiu, 1986). These studies suggest that usage of genetically modified bacteria can be a viable advancement to increase product formation by employing CO₂ as an electron source rather than organics. Butanol is considered to be a better alternative to gasoline in contrast to ethanol as it exhibits higher energy density, less volatility, and low corrosivity (García et al., 2011; Zhang et al., 2018). Current advances in metabolic engineering of cyanobacteria for the generation of solvents from CO₂ were critically reviewed elsewhere (Quintana et al., 2011).

Cyanobacterial species, such as *Anabaena*, *Aphanocapsa*, *Calothrix*, *Microcystis*, *Nostoc*, and *Oscillatoria*, are equipped for the generation of hydrogen by using a photosynthetic autotrophic mechanism (Lambert and Smith, 1977; Lau et al., 2015). Filamentous cyanobacteria generate H₂ as a by-product during nitrogen fixation under N₂-limiting conditions. Additionally, cyanobacteria can generate H₂ by reversible activity of the hydrogenases enzyme. In general, these species are equipped to use bidirectional or reversible hydrogenase enzymes to oxidize oxygen to generate hydrogen. The enhancement in hydrogen generation can be achieved by blocking the pathways that use the hydrogenases for a reduction reaction (Tamagnini et al., 2002).

Cyanobacteria are also known to produce alkanes and alkenes, which have a desirable property for industrial application (Lau et al., 2015). Among these enzymes, acyl-acyl carrier protein reductase and aldehyde decarbonylases are the key enzymes noted for efficient conversion of fatty acids of metabolic intermediates to alkanes and alkenes (Schirmer et al., 2010). Additionally, due to similarity to algae metabolism, cyanobacteria are well-known for their effluent treatment of wastewaters that are rich in nitrogen and phosphorus contaminants.

Value-Added Products Using Algae and Cyanobacteria

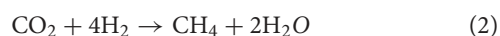
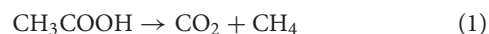
Apart from the biodiesel and fuels, algae are well-known for their usage in the food industry. The commercial products of algal powders are sold in the health and food industries. Algal products, such as unsaturated fatty acids and polysaccharides from microalgae and seaweed provide a dietary fiber and health benefits. The carotenoids that present in algae serve as antioxidants and vitamin A. In the textile industry, several algal products (pigments) are used as coloring agents. In aquatic fields, algae are used as feedstock in fish, shrimp, and other seafood that are grown in enclosed environments. Carotenoids, such as

astaxanthin, beta-carotene, bixin, and fucoxanthin, are used in the cosmetic and pharmaceutical industries (Hudek et al., 2014).

Anaerobic Fermentation of CO₂ for Value Addition

The increase in fossil fuel consumption is leading to a shortage of fuels and increase in energy prices. In order to reimburse energy consumption from the past few decades, research has been gaining attention on renewable energy sources with a prime focus on using inorganic carbon such as CO₂ as a substrate (Chang et al., 2016). The policy is to replace the fossil fuel with renewable energy sources and simultaneously decrease global carbon emissions. One of these renewable products includes CH₄ generation, which can be an assurance of chemical storage of energy (Villano et al., 2010). In several cases, anaerobic methane generation is productive and is frequently used in electricity generation and as a heat energy source (combustion process). Additionally, it can be used in the generation of biodiesel, methanol, and a few other hydrocarbons, such as ethanol and butanol (Ge et al., 2014; Zakaria and Kamarudin, 2016; Wang et al., 2017). In general, methane is generated in two ways: through biotic and abiotic pathways. Abiotic pathways include the catalytic conversion or thermal split of kerogen (Lawton and Rosenzweig, 2016). Biogenic methane generation is abundantly noticed from anaerobic digestion and from dairy farms. In fact, 20% of global natural gas is being produced from methanogens (Wang et al., 2017).

Methanogenic microbes are obligate anaerobes, which are under the Euryarchaeota phyla belonging to the Archaea domain. These obligate anaerobes have the capability of sequestering the CO₂ under optimized specific conditions (Zeikus, 1977; Strong et al., 2016). In gas fermentation, CO₂ sequestration is carried out by hydrogenotrophic methanogens with the help of gamma- and zeta-type carbonic anhydrase enzymes. The CO₂ can be digested with different forms of chemicals, such as bicarbonate, carboxylic acids, alcohols, and biogas with their respective enzymes. However, acetyl CoA is the central mediator molecule in several microbial processes for product synthesis. Methanogenic and acetogenic bacteria are frequently found in a pair when CO₂ is the reduction medium (Kuramochi et al., 2013; Shi et al., 2015). In conventional anaerobic digestion, the difference in operational conditions affects the behavior and fate of both acetogenic and hydrogenotrophic methanotrophs. Under optimized conditions, the acetogenic bacterium transforms the acetic acid to H₂ and CO₂, which is followed by the conversion to methane via acetate (Equation 1) or the indirect conversion of H₂ and CO₂ (Equation 2) by hydrogenotrophic methanogenic bacterial cells (Jiang et al., 2013; Bian et al., 2018).



For efficient and stable performance in the transformation of CO₂ and H₂ to CH₄ by hydrogenotrophic methanogens, a continuous supplement of molecular hydrogen is required. Based on several studies, the key parameters that control hydrogenotrophic methanogenic activity were identified

as pressure, redox conditions, temperature, headspace gas composition, hydraulic retention time, and pH (Rabaey et al., 2009; Lee et al., 2012). Scherer et al. (2000) analyzed the degradation of organics in municipal gray water under thermophilic and mesophilic environments using a lab-scale continuous mode of operation for biogas generation. The most probable number (MPN) technique illustrated that hydrogenotrophic methanogens (H₂-CO₂ utilizers) were in a range of 10⁸ to 10¹⁰/g total solids (TS), which seems to dominate the acetogenic methanotrophs by a factor of 10 to 10,000, due to short hydraulic retention time (HRT). Similarly, Ahring et al. (2001) noted that hydrogenotrophic methanogens exhibited higher specific methanogenic activity at 65°C. Although these studies prove that the presence of hydrogenotrophic methanogens can enhance methane formation, it is unclear whether enhanced methane formation is by direct or indirect conversion under practical conditions. Thus, future research in methane production should be encouraged toward the CO₂-utilizing metabolic pathways that control the symbiotic relationship among acetoclastic and hydrogenotrophic methanotrophs along with acidogenic bacteria.

Gas Fermentation of CO₂ to Products

The preceding advances in bio-energy generation have raised a feed vs. fuel debate. In this regard, gas fermentation looks to be a promising technology, in which gaseous substrates (CO₂) are converted to biochemicals. Also, many industrial processes generate large amounts of carbon gases, which are left unused. Thereby, utilization of CO₂ as a feedstock toward value-added products can shut down the feed vs. fuel debate. As aforementioned, conversion scenarios indicate that CO₂-rich waste gases can be a suitable substrate for gas fermentation. Several microbes have the ability to utilize CO₂ as a carbon source. Among these several microbes, acetogenic bacterium is mostly being studied and shows a capability for industrial applications. These bacterial species are equipped to convert CO₂ and CO by making use of the Wood-Ljungdahl pathway (WLP). Different acetogenic bacteria can naturally generate ethanol or butanol and some other goods of industrial interest, such as 2-oxobutyrate, hexanol, PHA, vitamin B12, polymers, and their precursors for other products.

In a recent study by Kim et al. (2016) acetate was noted as an end product in a multistage gas fermentation process by using steel mill off-gas as a feedstock. In the initial stage of fermentation, CO was transformed to CO₂ and H₂ by *Thermococcus onnurineus*. Later, the mixture of CO, CO₂, and H₂ were converted to acetate by *Thermoanaerobacter kivui* and resulted in a maximum generation rate of 0.33 mmol/l/h. When the initial step was absent, *T. kivui* was unable to generate acetate due to CO inhibition and lack of CO₂ as a substrate. Acetate formations in the multistep process also can be employed for the production of ethanol and other value-added products. Richter et al. (2013) operated a two-stage system for generation of ethanol with CO₂ as a feedstock by using *C. ljungdahlii*. The first stage of the reactor was employed for acetate; then the broth containing acetate was transferred to a second fermenter by lowering the pH for efficient conversion of acetate to ethanol. The production

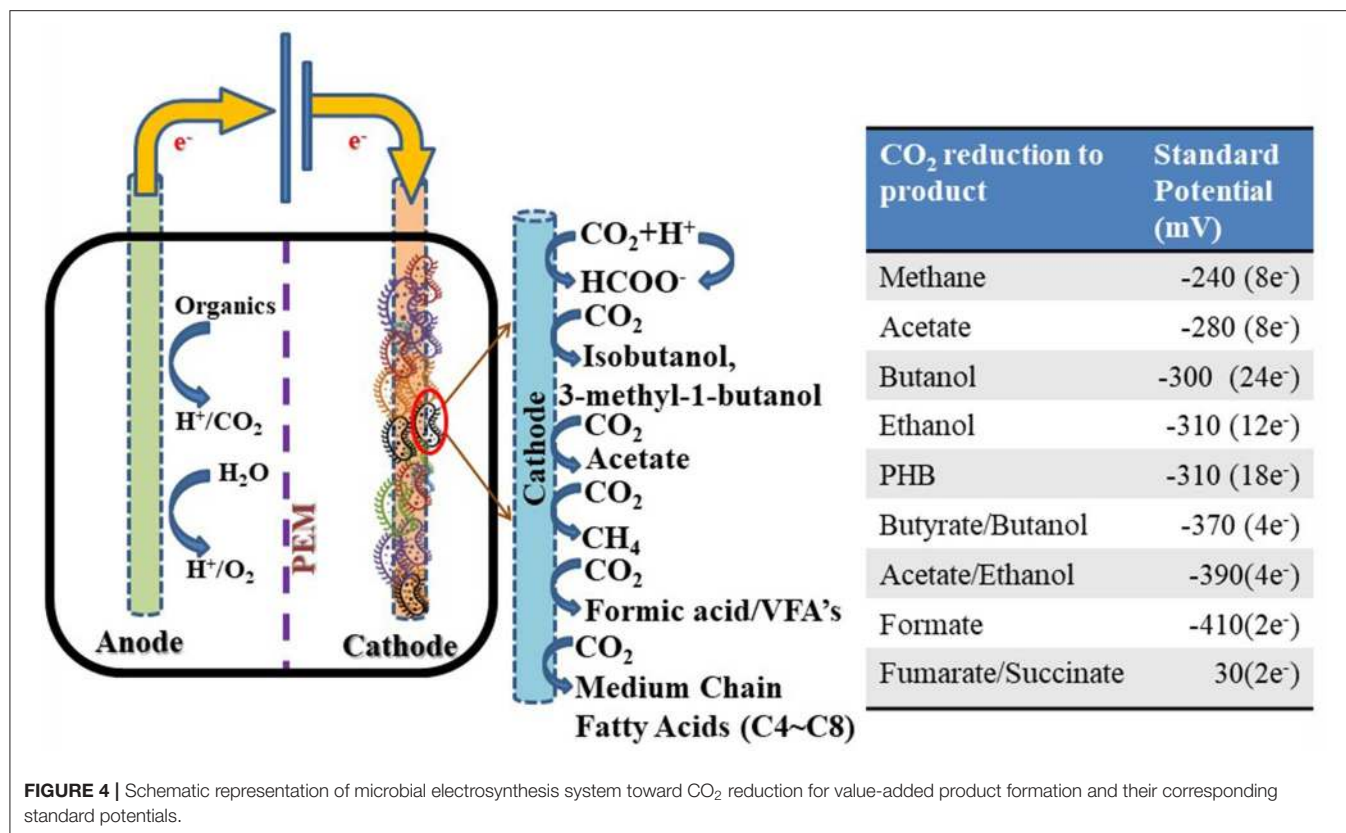
rate and maximum concentrations of ethanol were noted as 0.37 g/l/h and 450 mM, respectively. Different types of clostridium strains were tested in similar systems, and it was noted that *C. ljungdahlii* PETC was the most optimal producer for ethanol (Martin et al., 2016). Hu et al. (2016) developed a two-stage system for generation of longer chain fatty acids (C₁₆-C₁₈). This was carried out by *M. thermoacetica* in the first stage for acetate formation with syngas as a feedstock. The effluent broth was subjected to an aerobic reactor containing genetically modified *Yarrowia lipolytica*. The system could generate up to 18 g/l of C₁₆-C₁₈ lipids at a rate of 0.19 g/l/h. The system proves the conversion of carbon in syngas to complex substrates with a multistage system.

To generate value-added products by an integration approach along with biological processes, chemical processes are also feasible. Lanzatech, in association with Invista and SK innovation (<http://greenchemicalsblog.com/2014/06/05/invista-and-lanzatech-expand-collaboration/>; <http://www.invista.com/en/news/pr-invista-and-lanzatech-make-breakthrough-for-bio-derived-butadiene-production.html>), proposed production of 1, 3-butadiene in a two-step process, which is combination of biological and chemical steps. The recent review published by Liew et al. (2016) gives an excellent overview on the advances of synthetic biology on syngas-fermenting bacteria. However, we speculate that these technologies are limited by low concentration of substrate uptake, productivity, and selectivity in terms of product formation in comparison to current technologies.

Bioelectrochemical Conversion of CO₂ to Fuels and Chemicals

Bioelectrochemical systems (BES), such as microbial electrosynthesis (MES), microbial electrolysis cells (MECs), and microbial fuel cells (MFCs), are gaining importance in the field of renewable energy due to their promising approach for converting waste to energy (Kondaveeti et al., 2017; Mohanakrishna et al., 2020). In general, MFCs generate electricity from organics, whereas MES synthesizes value-added chemicals from CO₂ via electrogenic fermenting cathode microbes (biocathode) by using the cathode electrode as a sole electron source (Figure 4) (Logan, 2008). Nonetheless, MES is more beneficial in comparison to MFCs due to its capability in taking CO₂ and slashing the greenhouse gases. In addition, MES is interesting because of energy storage in the form of chemical production (e.g., hydrogen, alcohols, and C₁-C₆ medium chain fatty acids) for sustainability and decrease in dependency on non-renewable sources, such as fossil fuels (Rabaey et al., 2009).

The bioelectrochemical conversion of organics, such as acetate, in the anode of MFCs generates CO₂, protons, and electrons. The generated electrons move toward the cathode through an external circuit. The protons are diffused toward the cathode through a separator, typically a proton exchange membrane (PEM, e.g., Nafion-117) or cation exchange membrane (CEM). On the cathode, the diffused protons and electrons are combined together to form water by using O₂ as an electron acceptor (cathodic reduction reaction). The potential



disparity between the anode and cathode are captured as voltage. When an external electron acceptor, such as O₂, is absent, the diffused protons are converted to H₂ with the help of minimal external voltage (0.2 V), which was termed as MEC. The initial discovery for generation of hydrogen by modifying MFC to MEC was reported by the B.E. Logan group at Penn State University (Logan, 2008). MEC operations are more economic over MFC due to the ecologically beneficial production of valuable products, such as H₂. By the same process, the microbes at biocathodes can shuttle the e⁻ from the electrode for the reduction of CO₂ to value-added products. This can be achieved by using an anaerobic biotic anode with an electrogenic biocatalyst using organics or with an abiotic anode with an electrolysis process by applying additional potential. The bioprocess for value-added product formation was carried out by numerous metabolic pathways, such as the Calvin Benson cycle or the WLP (Rabaey and Rozendal, 2010; ElMekawy et al., 2016).

MES is capable of generating basic value-added products, such as H₂, acetate, and oxobutyrate (Kadier et al., 2020). These products can be directly used as fuels or can be used further to generate high carbon fuels or bioplastics. Initially, Nevin et al. (2010) stated reduction of CO₂ to acetate in small concentrations by operating MES at -0.6 V by using an acetogenic *Sporomusaovata* as cathodic biocatalyst (Nevin et al., 2010) and renewable energy (solar energy) as an external power source. This study also evidenced smaller amounts of oxobutyrate production (Table 5). Further, in 2011, acetate at

a production rate of 0.05 mM/d was achieved using mixed cultures (Nevin et al., 2011). In other studies using *S. ovata*, the production rates were 2.3-fold higher using a nickel nanoparticle-coated cathode over a plain graphite cathode (1.13 mM/d) (Zhang et al., 2013). Mohanakrishna et al. (2015) reported an acetate generation rate of 4.1 g/l by using enriched single chamber MES with VITO-activated carbon electrodes. Similarly, in other studies from the same group, they analyzed acetate formation by using bicarbonate and CO₂ as a substrate by varying the poised cathode potentials (-0.6 and -0.8 V vs. Ag/AgCl). Interestingly, in this study, they noted that MES operation with CO₂ at -0.8 V generated a higher acetate concentration of around 5.1 g/l. However, decreasing poised potentials (-0.6 V) during MES operation with bicarbonate exhibited maximum acetate concentrations of around 4.1 g/l (Mohanakrishna et al., 2016). Batlle-Vilanova et al. (2016) operated continuous mode MES for CO₂ reduction and noted an acetate production rate of 0.98 mM/d. Marshall et al. (2013) reported the highest acetate generation (17.25 mM/d) from CO₂ by using MES (-0.79 V) with a carbon bed cathode and enriched cultures from methanogenic bacteria and H₂. MES operation with a simultaneous production and extraction mechanism for acetate from CO₂ were operated by Gildemyn et al. (2015). In this study, the maximum concentration of acetate formation was noted as 13.5 g/l, which is found to be the highest to date, whereas the highest production rate of 0.78 g/l/h was registered by LaBelle and May (2017). This might be due to the ceasing

TABLE 5 | Bio electrochemical generation of solvents and biofuels from CO₂ under various operational and nutritional conditions.

Reactor	Substrate	Applied energy (voltage or current)	Operational condition	Dominant catalyst (microbes or enzymes)	Products	References
H-Type double chamber	CaCO ₃	+0.5 to -0.5 V (vs. Ag/AgCl)	Batch type; Anaerobic Fermentation	<i>Clostridium Sporogenes BE01</i>	Butanol, Ethanol, Fatty acids, Hydrogen	Gottumukkala et al., 2015
Double Chamber Fuel cell	Butyraldehyde + TRIS-HCl buffer	-0.6 V (vs. Ag/AgCl)	Batch type; Enzymatic Fuel cell	Alcohol dehydrogenase enzymes	Butanol	Schlager et al., 2015
H-Type double chamber	P2 electron carriers in medium + Glucose	-0.7 V (vs. Ag/AgCl)	Batch type; Anaerobic Fermentation	<i>Clostridium beijerinckii IB4</i>	Acetone, Butanol, Ethanol production and butanol as primary product	He et al., 2016
Two compartment Cell	CAB medium with electron carrier in buffer	-2.5 V poised at cathode	Batch type; Electrochemical cell	<i>Clostridium Acetobutylicum ATCC 4259</i>	Butanol and Acetone	Kim and Kim, 1988
H-Type double chamber	CO ₂ injection +DSMZ medium	-0.6 V (vs. Ag/AgCl)	Continuous mode operation	<i>Sporomusa, Geobacter, Clostridium, Morella</i>	Acetate, Formate, Butyrate, Propanol, Ethanol and 2-oxobutyrate	Lovley and Nevin, 2018
H-Type double chamber	CO ₂ (No organics in media)	-0.8 V (vs. SHE)	Batch type; Electrochemical cell	<i>Clostridium</i> species+ Carboxydrotrophic mixed culture	Ethanol, Butanol, acetate, butyrate	Puig et al., 2015
H-Type double chamber	Modified P 2 Medium + SMM medium	1.32 V as Applied voltage	Batch type; Electrochemical cell	<i>C. pasteurianum</i>	Butanol and by products as solvents and acids	Khosravanipour Mostafazadeh et al., 2016
Double chamber	CO ₂	-0.55 V (vs. NHE)	Batch type; Electrochemical cell	N/A	CH ₄	Van Eerten-Jansen et al., 2012

of acetate oxidation at the cathode by employing simultaneous production and extraction techniques. All these studies pointed out that, during long-term operation, the oxidation of acetate to methane is one of the major limitations. This can be controlled by employing a chemical treatment process, such as by using BESA (2-Bromoethanesulfonic acid) or employing pH shock treatment (Mohanakrishna et al., 2015, 2016; ElMekawy et al., 2016). The variation in acetate production might be due to differences in applied voltage, operational environment, reactor configuration, electrode type, biofilm activity, and type of bio-electrocatalyst at the cathode.

Compared to pure culture of *C. ljungdahlii*, enriched mixed culture results showed a higher acetate (1.3 mM/d) production rate at -1.1 V, which has hydrogen-generating potential (Bajracharya et al., 2015). This study also evidenced methane production and suggests an increase in CO₂ reduction by using mixed cultures by supplementing H₂. The supplementation of H₂ can be obtained by maintaining higher negative cathode potentials for the reduction of protons to H₂. However, generated H₂ can leak from the reactors, thereby restricting the CO₂-reduction process, which, in turn, decreases current and product formation. This complication can be overcome by using high gas absorption electrodes (e.g., platinum single crystal electrode) in order to decrease H₂ leakage (Will, 1965; Wang Q. et al., 2015). Several pure cultures, such as *Clostridia*, *Moorella*, and

Sporomusa, are proven for direct conversion of CO₂ by accepting electrons from a poised cathode electrode at -0.6 V (Nevin et al., 2010; Bajracharya et al., 2015); however, these are still limited for practical applications due to strict operational conditions.

In MES, acetate is one of the primary products noticed from the reduction of CO₂ (Mohanakrishna et al., 2016; Modestra and Mohan, 2017). However, its autotrophic production is not economically desirable due to lower yields and little commercial value. In this aspect, researchers have further explored the possibility of higher value-added product generation (Rabaey and Rozendal, 2010; Arends et al., 2017). In this facet, acetic and butyric acids were converted to bioalcohols using a poised biocathode at -0.65 V (Steinbusch et al., 2010; Sharma et al., 2013b). In this study, total alcohol production of 13.5 mM was achieved. Ethanol, butanol, propanol, and acetone were noticed as major alcohols along with propionic and caproic acids (Sharma et al., 2013a). Similarly, Kondaveeti et al. operated a BES and transformed the VFAs from anaerobic digester effluent to bioalcohols by varying applied voltage. Electron mediators, such as neutral red, helped for increasing product formation at higher redox potentials (Kondaveeti and Min, 2015). Ethanol and butanol are the major alcohols noticed at the applied voltage of 2 V (Cathode potential, -0.9 V). Likewise, Steinbusch et al. (2010) operated a microbial electrosynthesis system with various mediators [methyl viologen (MV), neutral red (NR),

anthraquinone-2,6-disulfonate (AQDS)] and noted a maximum yield of 13.5 mM of ethanol by using MV. In other studies pursued by Van Eerten-Jansen et al. (2013), a reduction in acetate to Caproate (6.8 mM) and butyrate (3 mM) were noticed in MES with cathode poised potential of -0.9 V. Recently, Vassilev et al. (2019) operated a dual cathode MES system for concurrent acetogenesis and solventogenesis followed by carbon chain elongation. This type of system can assist in operational cost reduction with a simultaneous increase in productivity due to the utilization of off-gas. All previous studies are found to be interesting in terms of product formation; however, these systems are still limited by selectivity of product formation and extraction of product using cheaper processes (del Pilar Anzola Rojas et al., 2018; Li et al., 2018; Vassilev et al., 2018). Most of the product extraction studies of MES include integrated membrane separation with an anion and cation exchange membrane or sequential operation. However, the operation of such processes can be expensive as they often need to be changed because of fouling during longer operation. The research in this regard needs to be pursued to increase the production rate and decrease operational costs for product extraction and selectivity of product formation.

In the past few years, research on generation of methane from CO₂ using bioelectrochemical techniques has increased significantly. It has been noted that the optimized electrode potential (direct electron donor) for direct conversion of CO₂ to methane was found to be around -0.6 to -0.7 V vs. Ag/AgCl (Cheng et al., 2009; Villano et al., 2010; ElMekawy et al., 2016; Bajracharya et al., 2017). Villano et al. (2010) reported that, using -1.0 V of applied external potential, 96% of the obtained current was captured in the form of methane. Based on current and obtained abiotic hydrogen, it was noted that CH₄ was produced directly from electrons derived from a cathode electrode rather than using the hydrogen gas. Alternatively, they also have suggested that some amount of methane generation is by hydrogenophilic methanogens by consuming H₂ from abiotic water reduction. The performances of biocathodes were further tested by inoculating with methanogenic culture. These cultures were greatly able to reduce the inorganic carbon to methane (0.055 mmol/d for mg/VSS) with 80% electron capture efficiency (Villano et al., 2010). The standard theoretical potential for the reduction of CO₂ to CH₄ is -0.44 V vs. Ag/AgCl. The required energy for this cathodic reaction can be achieved from the bioanode (organic oxidation) process. However, in practical operation, these can be greatly limited with over-potentials. These over-potentials can be reduced by implementing biocathode systems. In another study, Villano et al. (2013) demonstrated a double chamber MEC for methane generation with anode potential of -0.397 V vs. Ag/AgCl by using acetate as an energy source. Simultaneously, a cathode was constantly supplemented along with CO₂ and N₂ for pH correction and carbonate supply. It was reported that a 94% reduction in acetate at the anode chamber with 91% of coulombic efficiency, which proves that the generated current was mainly from methane production. Yasin et al. operated an enriched activated sludge system to generate methane from CO₂. In this system, in the presence of CO₂ and H₂, they noticed a 70-fold

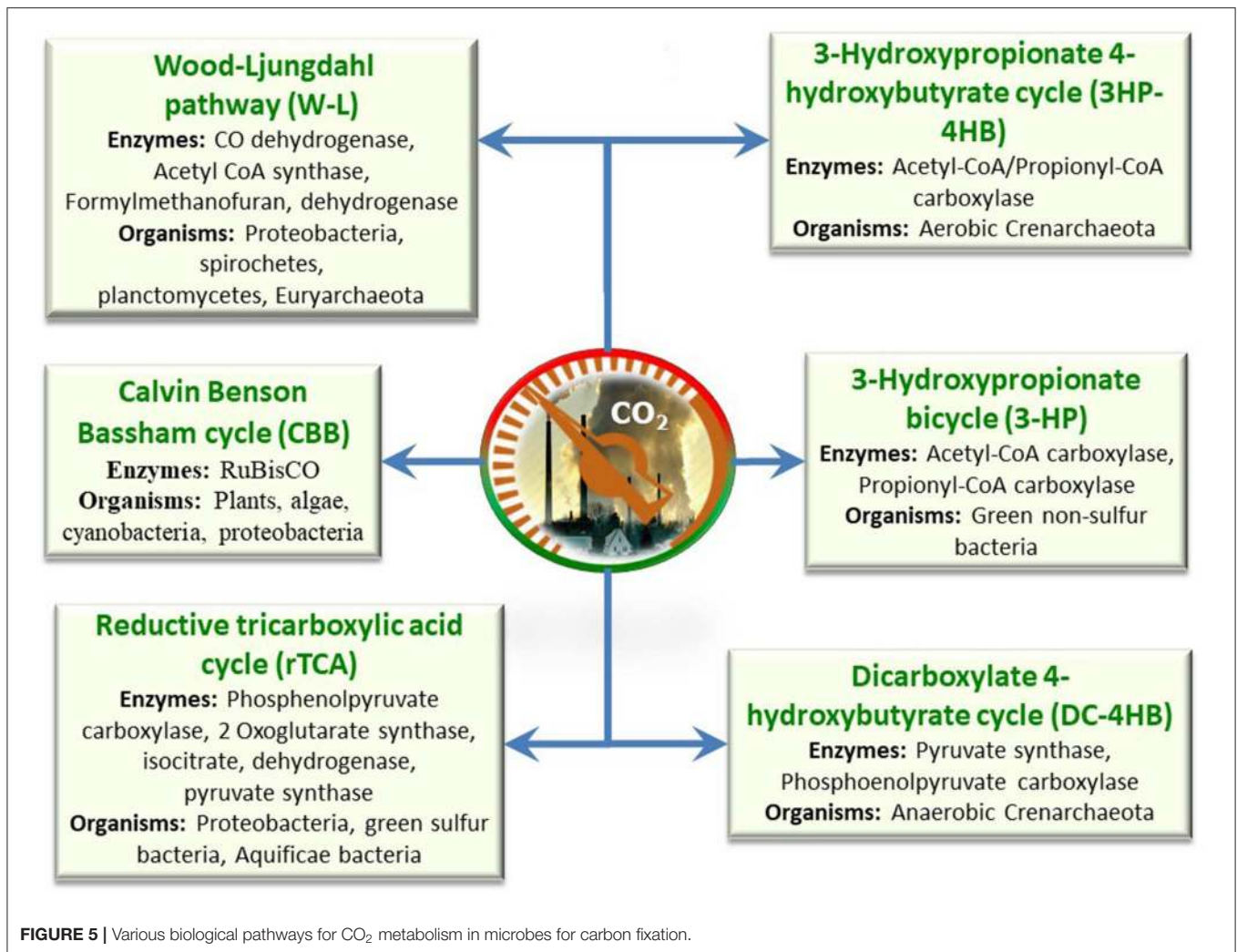
increase in methane generation after 72 h of operation. The complete utilization of CO₂ was noticed after 96 h of incubation under supplementation of CO₂ and H₂ as carbon and electron sources, respectively (Mohd Yasin et al., 2015). Based on a recent study by Xenia et al., the microbial electrosynthesis cells were proven to be economical and sustainable with CO₂ as a substrate for product formation, such as methanol, ethanol, and formate (Christodoulou et al., 2017).

OTHER CO₂-UTILIZING BACTERIA

β -Proteobacteria

Proteobacteria are Gram-negative bacteria, mainly composed of lipopolysaccharides in their outer membrane. These are found to be a geologically, environmentally, and evolutionarily important group of microorganisms. The members of this bacterial phylum exhibit an enormous metabolic diversity. Most of these bacteria have an industrial, medical, and agricultural significance (Sezenna, 2011). The photosynthetic proteobacteria are known as purple bacteria, concerning their reddish pigments. One of the most common and well-known photosynthetic β -proteobacteria is *Ralstonia eutropha*. This bacterium can grow on variable carbon environments, such as heterotrophic, autotrophic, and mixotrophic. In the absence of a heterotrophic carbon environment as an electron source, this bacterium has proven to utilize hydrogen as an energy source for fixing CO₂ via the Calvin-Bassham cycle (CBB) (Figure 5) (Jajesniak et al., 2014).

R. eutropha is a convenient photosynthetic bacterium to handle because of its aerobic H₂ oxidizing capabilities over obligate anaerobes to generate chemicals and other value-added products. This fascinating interest in *R. eutropha* has led to generation of bioplastics (PHAs/poly hydroxyalkanoates) by storing carbon in cytoplasm. PHAs are generally composed of PHB (poly 3-hydroxybutyrate) and PHBV (poly 3-hydroxybutyrate-co-3-hydroxyvalerate) (Luengo et al., 2003; Jajesniak et al., 2014). These bioplastics are gaining high importance due to credentials such as biodegradability over conventional plastics and their green synthesis with CO₂ sink. These polymers with desired length can be pursued with genetic modifications (Reinecke and Steinbüchel, 2009). Few studies are pursued in genetic modification of *R. eutropha* for the generation of products with variable properties (mechanical strength) and production yield. Voss et al. modified *R. eutropha* and generated a cyanophycin, a protein-like polymer, with an increased production quantity (Voss and Steinbüchel, 2006). A few other studies have synthesized several useful compounds, such as ferulic acid (precursor to vanillin biotransformation) and 2 methyl citric acid, by genetic modification (Overhage et al., 2002; Ewering et al., 2006). The limitation in these studies is use of organic carbon as an energy source rather than inorganic CO₂. Beller et al. engineered *R. eutropha* and generated methyl ketones of diesel range with productivity in a range of 50–180 mg/l by using CO₂ and H₂ as energy and electron sources, respectively (Müller et al., 2013). In a previous study, Li et al. (2012) described an integrated approach for production of isobutanol and 3-methyl-1-butanol by using a microbial electrochemical process. In this system, initially, CO₂ was converted to formate



and tailed by isobutanol and 3-methyl-1-butanol generation. This system achieved a final concentration of 850 and 570 mg/l of isobutanol and 3-methyl-1-butanol, respectively.

Idenolla species is another type of photosynthetic bacteria similar to *R. eutropha* in generation of polymers by using CO₂. An additional superiority of *Idenolla* over *R. eutropha* is the generation of products using carbon monoxide (CO) as an energy source. These bacterial species have an additional advantage by synthesizing products from exhaust gas (Tanaka et al., 2011).

Clostridia

Most clostridia are anaerobic and Gram-positive bacteria. They are found to have great importance in (i) human and animal health and physiology, (ii) anaerobic mortification of simple and complex carbohydrates, (iii) the carbon cycle, and (iv) bioremediation/degradation of complex organic chemicals (Kondaveeti et al., 2014b). Many strains of clostridia are capable of autotrophic fixation of CO₂ or CO by using H₂ as an electron donor (Willey et al., 2014). Clostridia also can use simple chain organic carbon molecules, such as formate or methanol, as an energy source. They can achieve this by the WLP pathway in

which two molecules of CO₂ are scaled down to generate one molecule of Acetyl-CoA with CO or H₂ being used as a reducing proportionate (Tracy et al., 2012).

Additionally, *Clostridium* species provide many fascinating characteristics for biotechnological utilization: (i) ability to use simple as well as complex substrates such as H₂, CO₂, and CO; (ii) equipped with diversified pathways for generation of value-added products; and (iii) tolerant to toxic metabolites and substrates. However, as they are strictly anaerobic, the cultivation of *Clostridium* is very difficult (Willey et al., 2014). Normal atmospheric and facultative conditions are lethal to *Clostridium* species. Early work with *Clostridium* was mostly carried out for the production of acetic acid and other related products. The synthetic biology/genetic modification of *Clostridium* are being pursued at a slow pace due to their difficulty in lack of genetic tools as found in *R. eutropha*. However, the repertoires of genetic tools are applicable for *Clostridium* with development in plasmid DNA and chromosomal manipulation technologies (Tracy et al., 2012). The uses of mobile group II introns are generally targeted for gene disruption and genetic engineering of *Clostridium*. Cooksley et al. (2012) demonstrated a

Clostron application through genetic engineering in *Clostridium acetobutylicum* for acetone-butanol-ethanol (ABE) fermentation and for expression of cellulosome. Leang et al. (2012) employed double crossover homologous recombination technology and an improved electroporation protocol to the process in a proof-of-concept gene deletion study on the species of *C. ljundahlii*. Likewise, recent studies by the D.R. Lovley group demonstrated introduction of foreign genes and deletion of certain genes in *C. ljundahlii* to enhance the production of biocommodities. Here, deletion of the ethanol-production pathway led to an increase in acetate production. The homolog recombination method described in their study can be applicable for the introduction of desired metabolic genes or reporters into the chromosomes of *C. ljundahlii* (Leang et al., 2012; Ueki et al., 2014).

Archaea

Archaea are generally found in abnormal ecological niches, such as high and low temperatures, acidic and high-saline environments, and anaerobic atmospheres. The hydrogenotrophic methanogens of Archaea use H₂ as an energy carrier and CO₂ as a carbon source for CH₄ generation. Furthermore, several strains of Archaea belonging to the haloarchael group are equipped with a metabolic pathway for accumulation of PHA from CO₂. As most of the Archaea are thermophilic in nature, this becomes an excellent source of the carbonic anhydrases (CA) enzyme, which is thermo stable and convenient for industrial CO₂ capture and a platform for gas fermentation (Willey et al., 2014). The recognition of genetic tools for Archaea might enhance the product yield in the gas fermentation process. Recently, genetic reengineering of Archaea for valuable product formation has been rising. Keller et al. (2013) reported a heterologous expression of five genes from a carbon fixation pathway in *Metallosphaera sedula* into hyper thermophilic *Pyrococcus furiosus*, which grows on organic carbon at 100°C. Engineered *P. furiosus* is capable of incorporating CO₂ for generation of 3-hydroxypropionate-4-hydroxybutyrate (3BHP) at an optimum condition of *M. sedula*; i.e., at 70°C. This temperature-dependent strategy for generation of valuable product formations is established in the work of Basen et al. (2012).

KEY ROLE OF ENZYMES IN MICROBIAL CELLS FOR CONVERSION OF CO₂

Adaptation of CO₂ to value-added products in the microbial kingdom has been found with six different pathways, such as WLP, Calvin Benson Bassham cycle, reductive tricarboxylic acid cycle (reductive/reverse TCA cycle), dicarboxylate 4-hydroxybutyrate cycle, 3-hydroxypropionate bicycle, and 3-hydroxypropionate 4-hydroxybutyrate cycle. These pathways are not found in every species, but specific pathways are found in specific organisms (Figure 5). Among these, the first three routes were found to be dominant for CO₂ conversion, and these are discussed in detail.

Calvin Cycle for the Phototrophic Reduction of CO₂

The Calvin cycle is one of the important biochemical cycles in photosynthetic organisms, which are capable of incorporating CO₂ into carbon metabolism. Thus, this metabolic function is dominant for CO₂ reduction in nature. In general, three key enzymes and three vital steps of the CO₂ process are noticed in the Calvin cycle (Venkata Mohan et al., 2016). The steps are known for CO₂ fixation, CO₂ reduction, and regeneration of CO₂. In the first step, the 1-5, bisphosphate ribulose bis-phosphate carboxylase (RuBisCo) catalyzes the reaction between CO₂ and 1-5, bisphosphate ribulose, and this process generates 3-phosphoglycerate (Shi et al., 2015). Later, 3-phosphoglycerate is further converted to 1,3-diphosphoglycerate by accepting inorganic phosphate (Pi) from ATP and further reduced to 3-phosphate glyceraldehyde with the help of the phosphoglyceraldehyde dehydrogenase enzyme. In the final step, phosphate glyceraldehyde is further transformed to 5-phosphate ribulose via a series of enzymatic reactions following further with its activation by phosphoribulokinase in ATP-dependent condensation to synthesize 1-5, bisphosphate ribulose for CO₂ collection for further cycles (Willey et al., 2014). One sixth of 3-phosphate glyceraldehyde is transformed as sugars, fatty acids, amino acids, etc.

Reverse TCA (Tricarboxylic Acid Cycle) Cycle

The reverse TCA (rTCA) cycle is known as the reductive citric acid cycle (reverse Krebs cycle), which basically runs in the reverse pathway. In this cycle, CO₂ and water are converted to carbon compounds (Lehninger et al., 2005). The reductive TCA cycle was first identified in anaerobic *Chloorbium limicola*, a green photoautotrophic sulfur-reducing bacteria. It was also identified in another thermophilic bacterium that uses hydrogen and sulfur as an energy source. The reverse TCA cycle contains four steps of carboxylation, in which, initially, succinyl-CoA is reductively carboxylated with CO₂ by α -Ketoglutarate synthase/2-oxoglutarate synthase (ΔG_0 , +19 kJ/mol; first step) to generate α -Ketoglutarate/2-oxoglutarate by utilizing two equivalents of reduced ferredoxin. Next, α -Ketoglutarate/2-oxoglutarate along with CO₂ is converted to isocitrate (ΔG_0 , +8 kJ/mol; second step) by using isocitrate dehydrogenase and NADPH. Further, isocitrate is converted to citrate by a cleavage mechanism, followed by oxaloacetate and acetate-CoA by ATP citrate lyase. The terminal compounds are carboxylated with CO₂, which requires a pyruvate synthase enzyme (ΔG_0 , +19 kJ/mol). The synthesized pyruvate is activated by pyruvate kinase to yield phosphoenolpyruvate (PEP), which is further converted to bicarbonate by a carboxylation mechanism (ΔG_0 , -24 kJ/mol). As an outcome, oxaloacetate is generated and finally converted to succinyl-CoA by a series of enzymes (Venkata Mohan et al., 2016).

Reverse Acetyl-CoA Cycle

The reverse acetyl-CoA pathway is predominantly found in strict acetogenic (Eubacteria) and methanogenic (Euryarchaeota)

bacteria. This route was suggested by Ljungdahl and Wood (2013), and it was named after them (Garrett and Grisham, 2008). Unlike the Calvin and reductive TCA cycles, the reductive acetyl-CoA is a non-cyclic route. In this route, initially, CO₂ is converted to formate by using NADH-dependent formate dehydrogenase (F_{ate}DH) (ΔG_0 +22 kJ/mol). Later, formate is apprehended by tetrahydrofolate and reduced into a methyl group by generating methyl-H4 folate. Methyltransferase transfers the methyl group of methyl-H4 folate to the cobalt center of hetero dimeric corrinoid iron sulfur protein, thus generating methylated corrinoid protein. Further, CO dehydrogenase (CODH) functions to convert CO₂ into CO via acetyl CoA synthase accepting the methyl group from CH₃-Co^{III} and converts CO, CoASH, and methyl groups to acetyl CoA (Mohanakrishna et al., 2015; Shi et al., 2015). As noted in the above enzymatic reaction, the autotrophic mechanism is by either CO₂ or bicarbonate as the carbon source. This mainly depends on the specific enzyme in a specific CO₂ fixation/conversion reaction. The attribute difference is noted in carbon source, and a fast-reliable conversion of CO₂ to bicarbonate is required.

SYNTHETIC BIOLOGY IN THE CONVERSION OF CO₂

Direct use of microbes in generation of value-added products is typically hindered by several characteristics/properties, such as low product yield, expensive cultivation condition, and inadequate growth rate. In this aspect, synthetic biology has gained importance due to its interest in product formation by altering biochemical pathways or by introduction of heterologous pathways into microbes for favorable features (Ferry et al., 2012). With consideration of recent advances in metabolic and protein engineering, the synthetic biology proves to be an excellent tool for boosting up biological process and to improve economic viability. In this section, *E. coli* is given special attention as it is the most often used microbe in the field of synthetic biology (Sawitzke et al., 2007).

Genetic Engineering for CO₂ Conversion

Despite being heterotrophic in nature, the Gram-negative *E. coli* gained its place at the table of genetic engineering due to its simple cultivation, low cost, easy transformation, fermentation, and high production for new technologies, which include using CO₂ as a carbon source (Willey et al., 2014). Zhuang et al. utilized different fermentation conditions for heterologous expression of RuBisCo and phosphoribulokinase (PRK) in *E. coli* (Zhuang and Li, 2013). In this study, they noticed a 67-mg CO₂/mole arabinose/l/h as the CO₂ fixation rate by a 15% decrease in CO₂ reduction during the fermentation process. Heterologous expression of carbonic anhydrase (CA) in *E. coli* is extensively reported. CA expression of *Methanobacterium thermoautotrophicum* in periplasm of *E. coli* resulted in a whole cell biocatalyst for CO₂ hydration. These systems successfully hydrated the CO₂ with a catalytic efficiency and productivity similar to free enzymes (Patel et al., 2013). The smaller difference observed in productivity might be due to mass transport

limitations. In the presence of CaCl₂ and CaCO₃, a 50~70% increase in carbon precipitation is noticed in comparison to operation without calcium (carbonate and chloride) components (Jo et al., 2013). Despite using free enzymes, whole cell utilization provides additional advantages such as (i) an increase in stability (100% in mortality activity after 24 h of use) and (ii) by passing the protein purification (whole cells are isolated and recycled). Similarly, as *M. thermoautotrophicum*, the periplasmic expression of CA is noticed in *Neisseria gonorrhoeae* (Jo et al., 2013). These advances noted in *E. coli* are not confined in the expression of individual enzymes. Bonacci et al. (2012) studied the expression of efficient carboxysomes from *Halothiobacillus neapolitanus* and encoded 10 genes in *E. coli* and noted that these bacteria are capable of carbon fixation. Further, they noticed an *in vitro* CO₂ fixation mechanism by using expressed carboxysomes with a sucrose gradient centrifugation as a purification step.

Synthetic biology of *E. coli* has provided a possibility of encoding photo bacterial genetics for the generation of value-added products. For instance, brown algae are well-known feedstock for the generation of carbon-neutral biofuel. However, from the point of economic industrial viability, these are limited by their inability to utilize alginate. Wargacki et al. (2012) reported the DNA fragments in *Vibrio splendidus* that are responsible for secretion of enzymes that are capable of utilization of alginate. This enzyme expression in *E. coli* resulted in a bacterial platform that is capable of metabolizing alginate with a yield of 0.281 weight ethanol/dry weight of microalgae that is equivalent to 80% of the theoretical basis of total polysaccharide composition. Along with several bacterial species, *Saccharomyces cerevisiae* also gained attention in conversion of CO₂ to value-added products. However, it has been limited with low productivity as compared to *E. coli*.

FUTURE PROSPECTS

The concepts of green and biological energy and/or chemical generation have been settling firmly in academia and industrial aspects, especially in utilizing CO₂ (Kuk et al., 2019). These processes guide the development of environmentally friendly products by optimized design concepts. The existing carbon capture technologies require huge land areas, large volumes, high energy, and expensive catalysts, and they also consume chemicals (ElMekawy et al., 2016). Therefore, based on reviewing the biological processes here, we suggest that biological systems align perfectly with green chemistry platforms for sustainable carbon capture and sequestration. The importance of biological processes for chemical generation using CO₂ as a carbon source is increasing rapidly (Venkata Mohan et al., 2016). From an economic point of view, all the biological processes for chemical/fuel production cannot be an industrially viable process. However, in consideration of carbon-neutral green energy generation, challenges, and their improvements, the biological adaptation of CO₂ utilization seems to be promising. The integration of a biological CO₂-mitigation process with conventional wastewater treatment technologies can minimize the economics related to waste remediation (Sadhukhan et al.,

2016). Concurrently, process integration can be beneficial in generating sustainable energy or fuels with a minimum carbon footprint. Yet the conversion of low concentrated waste organics into fuels or chemical commodities is required to achieve selective product formation with use of biocatalytic technology by economic evaluation, mathematical modeling, optimization, and integration of energy-efficient processes (ElMekawy et al., 2016; Sadhukhan et al., 2016). In this regard, the MES technology has proven to be a potential approach for conversion of CO₂ to chemicals and fuels.

Moreover, the scaling up of MES is an advancement to MFC that produces high-value products. Likewise, it has the capability to become an alternative to fossil fuels and also to eliminate the food vs. fuel debate. However, there are few constraints that should be solved, for instance, the low electron intake by bacteria at the cathode and low productivity, stability, selectivity,

etc. The synergistic effects of the substrate, electrode materials, and bacterial cell should be studied and well-understood for up-scaling. The conventional anaerobic fermentation system necessitates ~3 kg of glucose for production of 1 kg of butanol, which seems to be non-optimal in spite of existing large-scale processes (He et al., 2016). In this regard, alcohol generation in MES using organic-rich wastewater seems to be a promising choice (Rabaey and Rozendal, 2010). More specifically, influence of current and electrical potential on the microbial metabolism and microbial strains related to electro fermentation or MES are required to consider for industrial application (Mohanakrishna et al., 2016). Microbial carbon capture is likely to be a standalone process that could be competitive to conventional technologies. Therefore, coupling of biological CO₂ processes with chemical processes, such as gasification and water gas shift reaction, could be beneficial. Complex biomass sources, such as

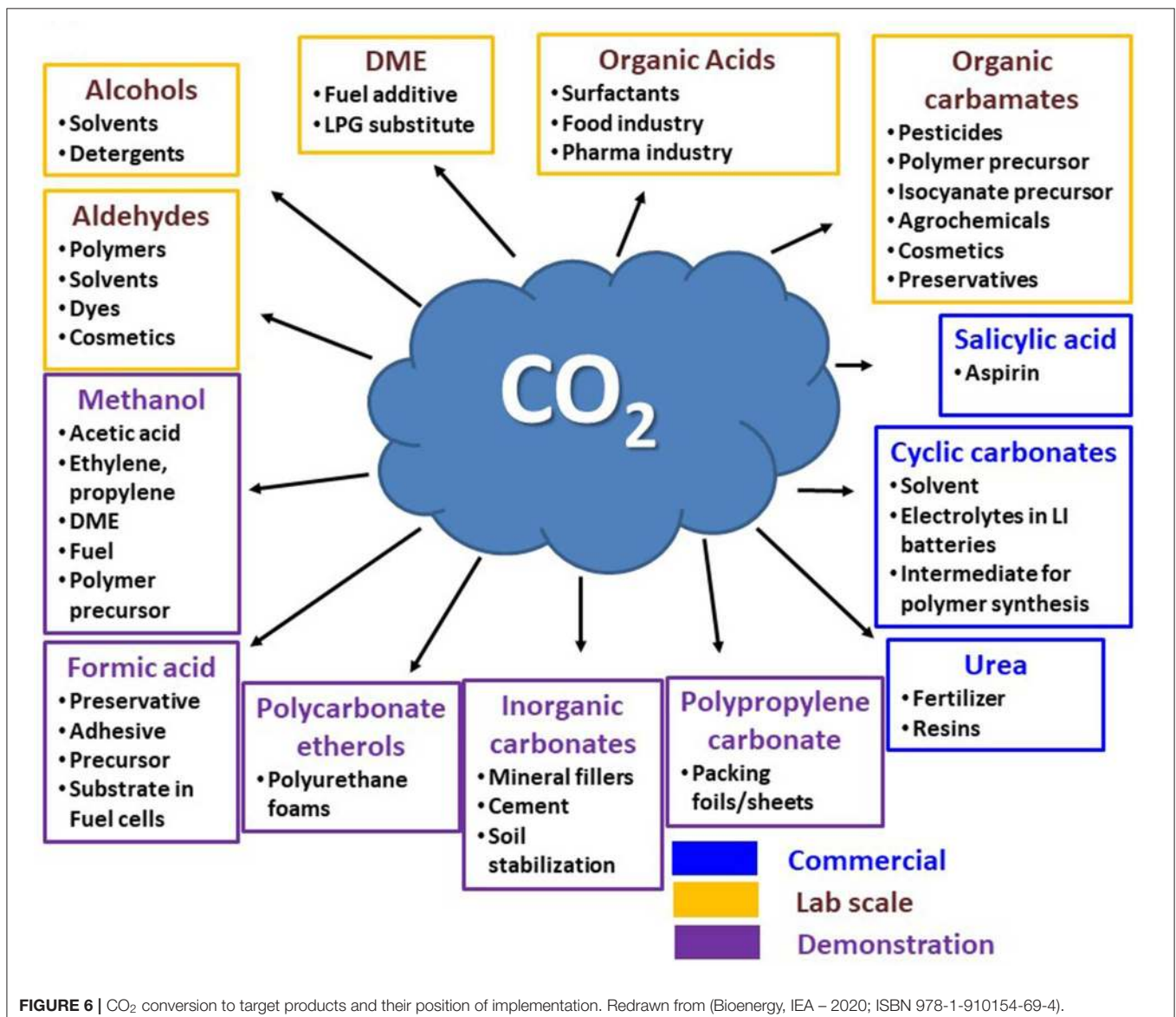


FIGURE 6 | CO₂ conversion to target products and their position of implementation. Redrawn from (Bioenergy, IEA – 2020; ISBN 978-1-910154-69-4).

cellulosic, wood, and straw materials, cannot be easily degraded or fermented. A good alternative is biomass gasification, which converts carbonaceous compounds into syngas through partial oxidation. H₂ and CO₂ in syngas also can be utilized by microbes for product formation. The conversion of syngas to products with a higher productivity rate can be achieved with bioelectrochemical and photobiological processes (Jiang et al., 2013). Further research would be required in order to enhance biofuel generation with low energy utilization and a minimal amount of toxic by-product generation (Rabaey and Rozendal, 2010; Mohanakrishna et al., 2015; ElMekawy et al., 2016; Venkata Mohan et al., 2016).

In terms of biotechnology of CO₂ utilization, algae and cyanobacteria have gained importance due to enhanced biomass generation for a feedstock and transesterification process. However, β -proteobacteria, clostridia, archaea, and other bacterial species can overtake the photosynthetic microbes due to rigorous research and efforts (Wang et al., 2012). Nonetheless, in utilization of various microbes for successful bioprocess development, one needs to pay attention to vital necessities, such as (i) suitable and appropriate strain selection for higher lipid generation (e.g., cell growth, autotrophic carbon fixation rate, product type and yield), (ii) supplementation minerals/chemical in CO₂ fixation, (iii) operation parameters (gas concentration, composition, etc.), (iv) type of bioprocess operation (batch or continuous), and (v) reactor configuration (CSTR, fixed bed reactor, membrane reactor) (Li et al., 2014). A few decisive factors in biological processes and engineering of carbon capture and utilization include (i) safety (especially during the H₂ utilization as electron donor/source), (ii) type of product selectivity and formation (concerning the purification and extraction economy), (iii) optimized operational conditions, and (iv) an increase in productivity by amplifying the cell density, etc.

In overview, the earlier described bioelectrochemical reaction pathways using CO₂ as the feedstock are based on H₂ as a reaction companion, whereas algae can transform CO₂ into value-added products, such as omega fatty acids, proteins, and amino acids and can simultaneously reduce pollutants. Likewise, other technologies, such as enzymes (e.g., formate dehydrogenase) can utilize/transform CO₂ by providing necessary energy. **Figure 6** provides an overview of different CO₂-based product formation pathways and their current status of employment. Apart from urea, the leading CO₂-based product formations from other industrial processes involve the production of cyclic carbonates and salicylic acids, which are being produced at about 0.1 million tons/year (technology readiness level (TRL): 9) (Bazzanella and Ausfelder, 2017). The poly (propylene) carbonate and polycarbonate etherols are extensively produced polymers in industries using CO₂ as a building block. Novomer incorporation from the United States and Covestro (former Bayer Material Science) are dynamic in this area with large-scale operations (TRL7-9). DNV (Det Norske Veritas) from Norway has constructed a pilot-scale operation plant for formic acid generation with a capability of 1 kg/day via electrochemical CO₂ reduction. Similarly, Mantra Energy Alternative Limited from Vancouver, British Columbia, Canada, is constructing a plant for 100 kg/day formic acid

generation (TRL7) by electro-reduction of CO₂ (Bazzanella and Ausfelder, 2017). Mineral carbonation, also known as carbon mineralization, is already being used in smaller and semi-commercial plants for various purposes, such as treatment of industrial waste, polluted soil, and generation of cement-like building materials (TRL7-9). There are several other processes being researched at lab scale with significantly lower TRL. Among these, one approach is noteworthy to point out because of its huge potential in the direct synthesis of dimethyl ether (DME) from CO₂. This method allows a CO₂ reduction of 0.125 t CO₂ /t DME, which is known to be higher in comparison to the state-of-the-art process with an intermediation of methanol equivalent to a 30% reduction. There is more production pathways research that involves direct production of sodium acrylate from ethylene and CO₂ or electrocatalytic conversion of CO₂ to ethylene (Alexis Bazzanella, 2017).

One of the crucial parameters to be considered for commercialization of CO₂ reduction via biological routes is the CO₂ that is stoichiometrically converted per ton of product. Based on earlier studies, acetic acid, ethanol, succinic acid, and caproic acid have stoichiometric production of 1.47, 1.91, 1.49, and 2.28 t CO₂/t, respectively. These chemicals (acetic acid, ethanol, succinic acid, and caproic acid) have a projected market value of 16.4 (Acumen Research Consulting, 2019), 54.63 (Coherent Market Insights, 2017), 0.237 (Markets Markets, 2019), and 0.27 (Market Study Report, 2019) billion dollars by 2026. Moreover, these products, such as acetate, can be used as building blocks for higher valued product generation, such as ethyl acetate, with a projected market value of 95 billion dollars by 2024 (Global Market Insights, 2018). Also, the combination of ethanol and other compounds (e.g., volatile fatty acids) can be used for a carbon chain elongation reaction. On the basis of these projected market values, the mitigation of CO₂ to product formation can be beneficial. However, these biological approaches need a further life cycle assessment (LCA) in order to compare benchmarks with the state-of-the-art production, considering energy aspects, etc.

In order to implement innovative biological ideas on a commercial scale and to reach the visionary target, consideration of the following aspects can be beneficial: (i) high-performing conversion technology on the level of CO₂ upstream: The technology should ideally cope with given impurities and/or CO₂ gas concentrations of their client's CO₂ waste streams at minimal pretreatment costs. CO₂ conversion process: convert CO₂ in an efficient way (total CO₂ conversion and selectivity toward product of interest) at an appreciable productivity (space-time yield) toward product in order to minimize investment and operational costs and obtain a techno-economic feasible route. Downstream: obtain high product titers and high product selectivity in order to reduce further downstream processing steps or costs. (ii) Local value chains fit for given CO₂-to-product pathway via biological route. (iii) Legislation for the end product in case of high-volume application (ethanol, fuels) (Carr, 2005).

Indeed, some of the major challenges of CO₂ reduction are the availability of renewable energy to sufficiently supply the

potential biological CO₂ conversion strategies (via green H₂ production), location, and transport of product. In this regard, low potential photovoltaic energy prices in the Middle East, Southern Europe, and North Africa would enable the production of liquid CO₂ fuels that can be easily transported to Europe, for instance. This enables the creation of new value chains and business cases of interest. As energy prices are high today and scales are small, we need high-value product routes or high volume that is supported by proper legislation. In the future, at low energy prices and larger scale, the challenge would be the energy supply in the most ambitious scenario, in which the full potential for given chemical/fuels is valorized via the CO₂ route (Tahvonen et al., 1994; Illing et al., 2018).

CONCLUSIONS

This report has overviewed and discussed potential applications of biological processes for CO₂ fixation with generation of value-added products. Despite achieving some progress, there is still a need for finding suitable processes or microbial strains for CO₂ mitigation. For instance, the productivity of microbial CO₂ sequestration should be more than 3-fold higher than traditional CO₂ technologies, which seems to be a daunting goal with current methods. Thus, the combination of industrial processes with microbial CO₂ mitigation can be sustainable and economically viable for reduction of CO₂. The metabolism of CO₂ conversion by algae and photosynthetic processes can be equally viable as

chemical and electrochemical technologies. Furthermore, market analysis and commercialization need to be further discussed by considering the real field application. Bioelectrochemical CO₂ conversion and photo biological processes for value-added product synthesis seems to be an economical and sustainable approach toward future technological development and commercialization. However, integration of suitable CO₂-converting processes with CO₂-generating processes helps for the development of a CO₂-based bio-refinery that may lead to carbon-neutral industrialization.

AUTHOR CONTRIBUTIONS

SK: inception of review concept, major write-up part of the review. IA-R: editing of review and supervision and Co-Lead PI of the project. GM: involved in writing two sections of the manuscript, editing of review and supervision. MB: involved in writing future prospective of the CO₂ transformation/reduction section of the review. DP: editing of review and supervision. All authors contributed to the article and approved the submitted version.

FUNDING

This publication was made possible by NPRP grant # 6-289-2-125 from the Qatar national research fund (a member of Qatar Foundation). The statements made herein are solely the responsibility of the authors.

REFERENCES

- Acumen Research and Consulting (2019). *Acetic Acid Market Value to Reach US 16.4 Billion by 2026*. Available online at: <https://www.acumenresearchandconsulting.com/acetic-acid-market> (accessed February 15, 2020).
- Ahring, B. K., Ibrahim, A. A., and Mladenovska, Z. (2001). Effect of temperature increase from 55 to 65°C on performance and microbial population dynamics of an anaerobic reactor treating cattle manure. *Water Res.* 35, 2446–2452. doi: 10.1016/S0043-1354(00)00526-1
- Alexis Bazzanella, D. K. (2017). *Technologies for Sustainability and Climate Protection – Chemical Processes and Use of CO₂*. Available online at: https://dechema.de/en/energyandclimate_CO2_Buch_engl.pdf (accessed February 15, 2020).
- Allen, M. M., and Stanier, R. Y. (1968). Selective isolation of blue-green algae from water and soil. *Microbiology* 51, 203–209. doi: 10.1099/00221287-51-2-203
- Anbalagan, A., Toledo-Cervantes, A., Posadas, E., Rojo, E. M., Lebrero, R., González-Sánchez, A., et al. (2017). Continuous photosynthetic abatement of CO₂ and volatile organic compounds from exhaust gas coupled to wastewater treatment: evaluation of tubular algal-bacterial photobioreactor. *J. CO₂ Util.* 21, 353–359. doi: 10.1016/j.jcou.2017.07.016
- Arends, J. B. A., Patil, S. A., Roume, H., and Rabaey, K. (2017). Continuous long-term electricity-driven bioproduction of carboxylates and isopropanol from CO₂ with a mixed microbial community. *J. CO₂ Util.* 20, 141–149. doi: 10.1016/j.jcou.2017.04.014
- Bajracharya, S., Ter Heijne, A., Dominguez Benetton, X., Vanbroekhoven, K., Buisman, C. J. N., Strik, D. P. B. T. B., et al. (2015). Carbon dioxide reduction by mixed and pure cultures in microbial electrosynthesis using an assembly of graphite felt and stainless steel as a cathode. *Bioresour. Technol.* 195, 14–24. doi: 10.1016/j.biortech.2015.05.081
- Bajracharya, S., Yuliasni, R., Vanbroekhoven, K., Buisman, C. J. N., Strik, D. P. B. T. B., and Pant, D. (2017). Long-term operation of microbial electrosynthesis cell reducing CO₂ to multi-carbon chemicals with a mixed culture avoiding methanogenesis. *Bioelectrochemistry* 113, 26–34. doi: 10.1016/j.bioelechem.2016.09.001
- Basen, M., Sun, J., and Adams, M. W. W. (2012). Engineering a hyperthermophilic archaeon for temperature-dependent product formation. *MBio* 3, e00053–e00012. doi: 10.1128/mBio.00053-12
- Battle-Vilanova, P., Puig, S., Gonzalez-Olmos, R., Balaguer, M. D., and Colprim, J. (2016). Continuous acetate production through microbial electrosynthesis from CO₂ with microbial mixed culture. *J. Chem. Technol. Biotechnol.* 91, 921–927. doi: 10.1002/jctb.4657
- Bazzanella, A., and Ausfelder, F. (2017). *Low Carbon Energy and Feedstock for the European Chemical Industry*. Frankfurt am Main: DECHEMA; Gesellschaft für Chemische Technik und Biotechnologie eV.
- Benedetti, M., Vecchi, V., Barera, S., and Dall'Osto, L. (2018). Biomass from microalgae: the potential of domestication towards sustainable biofactories. *Microb. Cell Fact.* 17:173. doi: 10.1186/s12934-018-1019-3
- Benemann, J. R. (1993). Utilization of carbon dioxide from fossil fuel-burning power plants with biological systems. *Energy Convers. Manage.* 34, 999–1004. doi: 10.1016/0196-8904(93)90047-E
- Benemann, J. R. (1997). CO₂ mitigation with microalgae systems. *Energy Convers. Manage.* 38(Suppl.), S475–S479. doi: 10.1016/S0196-8904(96)00313-5
- Benemann, J. R., and Oswald, W. J. (1996). *Systems and Economic Analysis of Microalga Ponds for Conversion of CO₂ to Biomass*. Final report. Berkeley, CA: California Univ., Dept. of Civil Engineering. doi: 10.2172/493389
- Benhelal, E., Zahedi, G., Shamsaei, E., and Bahadori, A. (2013). Global strategies and potentials to curb CO₂ emissions in cement industry. *J. Clean. Prod.* 51, 142–161. doi: 10.1016/j.jclepro.2012.10.049
- Berman-Frank, I., Lundgren, P., and Falkowski, P. (2003). Nitrogen fixation and photosynthetic oxygen evolution in cyanobacteria.

- Res. Microbiol.* 154, 157–164. doi: 10.1016/S0923-2508(03)00029-9
- Bhola, V., Desikan, R., Santosh, S. K., Subburamu, K., Sanniyasi, E., and Bux, F. (2011). Effects of parameters affecting biomass yield and thermal behaviour of *Chlorella vulgaris*. *J. Biosci. Bioeng.* 111, 377–382. doi: 10.1016/j.jbiotec.2010.11.006
- Bian, B., Alqahtani, M. F., Katuri, K. P., Liu, D., Bajracharya, S., Lai, Z., et al. (2018). Porous nickel hollow fiber cathodes coated with CNTs for efficient microbial electrosynthesis of acetate from CO₂ using *Sporomusa ovata*. *J. Mater. Chem. A* 6, 17201–17211. doi: 10.1039/C8TA05322G
- Bonacci, W., Teng, P. K., Afonso, B., Niederholtmeyer, H., Grob, P., Silver, P. A., et al. (2012). Modularity of a carbon-fixing protein organelle. *Proc. Natl. Acad. Sci. U.S.A.* 109, 478–483. doi: 10.1073/pnas.1108557109
- Bond, G. M., Stringer, J., Brandvold, D. K., Simsek, F. A., Medina, M-G., and Egeland, G. (2001). Development of integrated system for biomimetic CO₂ sequestration using the enzyme carbonic anhydrase. *Energy Fuels* 15, 309–316. doi: 10.1021/ef000246p
- Bulgariu, D., and Bulgariu, L. (2016). Potential use of alkaline treated algae waste biomass as sustainable biosorbent for clean recovery of cadmium(II) from aqueous media: batch and column studies. *J. Clean. Prod.* 112, 4525–4533. doi: 10.1016/j.jclepro.2015.05.124
- Carlsson, A. S., and Bowles, D. J. (2007). *Micro- and Macro-algae: Utility for Industrial Applications: Outputs from the EPOBIO Project*. Speen; Newbury, UK: CPL Press.
- Carr, M. (2005). Energy bill boosts industrial biotechnology; sweeping new legislation looks to bioproducts and biofuels to cut peak oil-dependency. *Ind. Biotechnol.* 1, 142–143. doi: 10.1089/ind.2005.1.142
- Chae, S. R., Hwang, E. J., and Shin, H. S. (2006). Single cell protein production of *Euglena gracilis* and carbon dioxide fixation in an innovative photo-bioreactor. *Bioresour. Technol.* 97, 322–329. doi: 10.1016/j.biortech.2005.02.037
- Chang, X., Wang, T., and Gong, J. (2016). CO₂ photo-reduction: insights into CO₂ activation and reaction on surfaces of photocatalysts. *Energy Environ. Sci.* 9, 2177–2196. doi: 10.1039/C6EE00383D
- Chen, J-S., and Hiu, S. F. (1986). Acetone-butanol-isopropanol production by *Clostridium beijerinckii* (synonym, *clostridium butylicum*). *Biotechnol. Lett.* 8, 371–376. doi: 10.1007/BF01040869
- Chen, W., Zhang, C., Song, L., Sommerfeld, M., and Hu, Q. (2009). A high throughput Nile red method for quantitative measurement of neutral lipids in microalgae. *J. Microbiol. Methods* 77, 41–47. doi: 10.1016/j.mimet.2009.01.001
- Cheng, S., Xing, D., Call, D. F., and Logan, B. E. (2009). Direct biological conversion of electrical current into methane by electromethanogenesis. *Environ. Sci. Technol.* 43, 3953–3958. doi: 10.1021/es803531g
- Chinnasamy, S., Ramakrishnan, B., Bhatnagar, A., and Das, K. C. (2009). Biomass production potential of a wastewater alga *Chlorella vulgaris* ARC 1 under elevated levels of CO₂ and temperature. *Int. J. Mol. Sci.* 10, 518–532. doi: 10.3390/ijms10020518
- Chisti, Y. (2007). Biodiesel from microalgae. *Biotechnol. Adv.* 25, 294–306. doi: 10.1016/j.biotechadv.2007.02.001
- Chiu, S-Y., Kao, C-Y., Tsai, M-T., Ong, S-C., Chen, C-H., and Lin, C-S. (2009). Lipid accumulation and CO₂ utilization of *Nannochloropsis oculata* in response to CO₂ aeration. *Bioresour. Technol.* 100, 833–838. doi: 10.1016/j.biortech.2008.06.061
- Christodoulou, X., Okoroafor, T., Parry, S., and Velasquez-Orta, S. B. (2017). The use of carbon dioxide in microbial electrosynthesis: advancements, sustainability and economic feasibility. *J. CO₂ Util.* 18(Suppl. C), 390–399. doi: 10.1016/j.jcou.2017.01.027
- Clarens, A. F., Resurreccion, E. P., White, M. A., and Colosi, L. M. (2010). Environmental life cycle comparison of algae to other bioenergy feedstocks. *Environ. Sci. Technol.* 44, 1813–1819. doi: 10.1021/es902838n
- Clout, A. (1994). Effect of light intensity variations on the rate of photosynthesis of algae: a dynamical approach. *Math. Comput. Model.* 19, 23–33. doi: 10.1016/0895-7177(94)90038-8
- Coherent Market Insights (2017). *Global Ethanol Market to Surpass US 54.63 Billion by 2026, Attributed to Increasing Production of Renewable Fuels*. Available online at: <https://www.coherentmarketinsights.com/press-release/global-ethanol-market-to-surpass-us-531-billion-by-2025-fuelled-by-increased-production-of-renewable-fuels-314>. (accessed February 15, 2020).
- Converti, A., Casazza, A. A., Ortiz, E. Y., Perego, P., and Del Borghi, M. (2009). Effect of temperature and nitrogen concentration on the growth and lipid content of *Nannochloropsis oculata* and *Chlorella vulgaris* for biodiesel production. *Chem. Eng. Process. Process Intensific.* 48, 1146–1151. doi: 10.1016/j.cep.2009.03.006
- Cooksley, C. M., Zhang, Y., Wang, H., Redl, S., Winzer, K., and Minton, N. P. (2012). Targeted mutagenesis of the *Clostridium acetobutylicum* acetone–butanol–ethanol fermentation pathway. *Metab. Eng.* 14, 630–641. doi: 10.1016/j.ymben.2012.09.001
- Csavina, J. L., Stuart, B. J., Guy Riefler, R., and Vis, M. L. (2011). Growth optimization of algae for biodiesel production. *J. Appl. Microbiol.* 111, 312–318. doi: 10.1111/j.1365-2672.2011.05064.x
- de Morais, M. G., and Costa, J. A. V. (2007a). Biofixation of carbon dioxide by *Spirulina* sp. and *Scenedesmus obliquus* cultivated in a three-stage serial tubular photobioreactor. *J. Biotechnol.* 129, 439–445. doi: 10.1016/j.jbiotec.2007.01.009
- de Morais, M. G., and Costa, J. A. V. (2007b). Carbon dioxide fixation by *Chlorella kessleri*, *C. vulgaris*, *Scenedesmus obliquus* and *Spirulina* sp. cultivated in flasks and vertical tubular photobioreactors. *Biotechnol. Lett.* 29, 1349–1352. doi: 10.1007/s10529-007-9394-6
- de Morais, M. G., and Costa, J. A. V. (2007c). Isolation and selection of microalgae from coal fired thermoelectric power plant for biofixation of carbon dioxide. *Energy Convers. Manage.* 48, 2169–2173. doi: 10.1016/j.enconman.2006.12.011
- del Pilar Anzola Rojas, M., Zaiat, M., Gonzalez, E. R., De Wever, H., and Pant, D. (2018). Effect of the electric supply interruption on a microbial electrosynthesis system converting inorganic carbon into acetate. *Bioresour. Technol.* 266, 203–210. doi: 10.1016/j.biortech.2018.06.074
- Demirbas, A. (2010). Use of algae as biofuel sources. *Energy Convers. Manage.* 51, 2738–2749. doi: 10.1016/j.enconman.2010.06.010
- Dexter, J., and Fu, P. (2009). Metabolic engineering of cyanobacteria for ethanol production. *Energy Environ. Sci.* 2, 857–864. doi: 10.1039/b811937f
- Dong, T., Knoshaug, E. P., Pienkos, P. T., and Laurens, L. M. L. (2016). Lipid recovery from wet oleaginous microbial biomass for biofuel production: a critical review. *Appl. Energy* 177, 879–895. doi: 10.1016/j.apenergy.2016.06.002
- Dudley, B. (2018). *BP Statistical Review of World Energy*. London: BP Statistical Review.
- Effendi, S. S. W., and Ng, I. S. (2019). The prospective and potential of carbonic anhydrase for carbon dioxide sequestration: a critical review. *Process Biochem.* 87, 55–65. doi: 10.1016/j.procbio.2019.08.018
- El Gamal, A. A. (2010). Biological importance of marine algae. *Saudi Pharm. J.* 18, 1–25. doi: 10.1016/j.jsps.2009.12.001
- ElMekawy, A., Hegab, H. M., Mohanakrishna, G., Elbaz, A. F., Bulut, M., and Pant, D. (2016). Technological advances in CO₂ conversion electro-biorefinery: a step toward commercialization. *Bioresour. Technol.* 215, 357–370. doi: 10.1016/j.biortech.2016.03.023
- Ewering, C., Heuser, F., Benölken, J. K., Brämer, C. O., and Steinbüchel, A. (2006). Metabolic engineering of strains of *Ralstonia eutropha* and *Pseudomonas putida* for biotechnological production of 2-methylcitric acid. *Metab. Eng.* 8, 587–602. doi: 10.1016/j.ymben.2006.05.007
- Fan, L-H., Zhang, Y-T., Zhang, L., and Chen, H-L. (2008). Evaluation of a membrane-sparged helical tubular photobioreactor for carbon dioxide biofixation by *Chlorella vulgaris*. *J. Membr. Sci.* 325, 336–345. doi: 10.1016/j.memsci.2008.07.044
- Fernando, I. P. S., Nah, J-W., and Jeon, Y-J. (2016). Potential anti-inflammatory natural products from marine algae. *Environ. Toxicol. Pharmacol.* 48, 22–30. doi: 10.1016/j.etap.2016.09.023
- Ferry, M. S., Hasty, J., and Cookson, N. A. (2012). Synthetic biology approaches to biofuel production. *Biofuels* 3, 9–12. doi: 10.4155/bfs.11.151
- Foo, S. C., Yusoff, F. M., Ismail, M., Basri, M., Yau, S. K., Khong, N. M. H., et al. (2017). Antioxidant capacities of fucoxanthin-producing algae as influenced by their carotenoid and phenolic contents. *J. Biotechnol.* 241, 175–183. doi: 10.1016/j.jbiotec.2016.11.026
- García, V., Pääkilä, J., Ojamo, H., Muurinen, E., and Keiski, R. L. (2011). Challenges in biobutanol production: how to improve the efficiency? *Renew. Sustain. Energy Rev.* 15, 964–980. doi: 10.1016/j.rser.2010.11.008
- Garrett, R. H., and Grisham, C. M. (2008). *Biochemistry*. Boston, MA: Cengage Learning.

- Ge, X., Yang, L., Sheets, J. P., Yu, Z., and Li, Y. (2014). Biological conversion of methane to liquid fuels: status and opportunities. *Biotechnol. Adv.* 32, 1460–1475. doi: 10.1016/j.biotechadv.2014.09.004
- Georgianna, D. R., and Mayfield, S. P. (2012). Exploiting diversity and synthetic biology for the production of algal biofuels. *Nature* 488, 329–335. doi: 10.1038/nature11479
- Gildemyn, S., Verbeeck, K., Slabbinck, R., Andersen, S. J., PrévotEAU, A., and Rabaey, K. (2015). Integrated production, extraction, and concentration of acetic acid from CO₂ through microbial electrosynthesis. *Environ. Sci. Technol. Lett.* 2, 325–328. doi: 10.1021/acs.estlett.5b00212
- Global Market Insights (2018). *Ethyl Acetate Market size, Industry Analysis Report, Regional Outlook, Application Development Potential, Price Trends, Competitive Market Share and Forecast, 2019 - 2025*. Available online at: <https://www.gminsights.com/industry-analysis/ethyl-acetate-market> (accessed February 15, 2020).
- González López, C. V., Ación Fernández, F. G., Fernández Sevilla, J. M., Sánchez Fernández, J. F., Cerón García, M. C., and Molina Grima, E. (2009). Utilization of the cyanobacteria *Anabaena* sp. ATCC 33047 in CO₂ removal processes. *Bioresour. Technol.* 100, 5904–5910. doi: 10.1016/j.biortech.2009.04.070
- Gottumukkala, L. D., Sukumaran, R. K., Mohan, S. V., Valappil, S. K., Sarkar, O., and Pandey, A. (2015). Rice straw hydrolysate to fuel and volatile fatty acid conversion by *Clostridium sporogenes* BE01: bio-electrochemical analysis of the electron transport mediators involved. *Green Chem.* 17, 3047–3058. doi: 10.1039/C5GC00310E
- Hanagata, N., Takeuchi, T., Fukujyu, Y., Barnes, D. J., and Karube, I. (1992). Tolerance of microalgae to high CO₂ and high temperature. *Phytochemistry* 31, 3345–3348. doi: 10.1016/0031-9422(92)83682-O
- Hatch, M., and Slack, C. (1970). Photosynthetic CO₂-fixation pathways. *Annu. Rev. Plant Physiol.* 21, 141–162. doi: 10.1146/annurev.pp.21.060170.01041
- He, A. Y., Yin, C. Y., Xu, H., Kong, X. P., Xue, J. W., Zhu, J., et al. (2016). Enhanced butanol production in a microbial electrolysis cell by *Clostridium beijerinckii* IB4. *Bioprocess Biosyst. Eng.* 39, 245–254. doi: 10.1007/s00449-015-1508-2
- Holkar, C. R., Jadhav, A. J., Pinjari, D. V., Mahamuni, N. M., and Pandit, A. B. (2016). A critical review on textile wastewater treatments: possible approaches. *J. Environ. Manage.* 182, 351–366. doi: 10.1016/j.jenvman.2016.07.090
- Hou, X., Huang, L., Zhou, P., Tian, F., Tao, Y., and Li Puma, G. (2019). Electrosynthesis of acetate from inorganic carbon (HCO₃⁻) with simultaneous hydrogen production and Cd(II) removal in multifunctional microbial electrosynthesis systems (MES). *J. Hazard Mater.* 371, 463–473. doi: 10.1016/j.jhazmat.2019.03.028
- Hu, P., Chakraborty, S., Kumar, A., Woolston, B., Liu, H., Emerson, D., et al. (2016). Integrated bioprocess for conversion of gaseous substrates to liquids. *Proc. Natl. Acad. Sci. U.S.A.* 113, 3773–3778. doi: 10.1073/pnas.1516867113
- Hudek, K., Davis, L. C., Ibbini, J., and Erickson, L. (2014). “Commercial products from algae,” in *Algal Biorefineries: Volume 1: Cultivation of Cells and Products*, eds R. Bajpai, A. Prokop, and M. Zappi (Dordrecht: Springer Netherlands), 275–295. doi: 10.1007/978-94-007-7494-0_11
- Hunt, A. J., Sin, E. H. K., Marriott, R., and Clark, J. H. (2010). Generation, capture, and utilization of industrial carbon dioxide. *ChemSusChem* 3, 306–322. doi: 10.1002/cssc.200900169
- Illing, L., Natelson, R., Resch, M., Rowe, I., and Babson, D. (2018). *Rewiring the Carbon Economy: Engineered Carbon Reduction Listening Day Summary Report*. USDOE Office of Energy Efficiency and Renewable Energy (EERE), Washington, DC. doi: 10.2172/1419624
- International Energy Agency (2019). *CO₂ Status Report*. Paris, France: IEA (International Energy Agency).
- Jajesniak, P., Ali, H. E., and Wong, T. S. (2014). Carbon dioxide capture and utilization using biological systems: opportunities and challenges. *J. Bioprocess. Biotech.* 4:1. doi: 10.4172/2155-9821.1000155
- Jiang, Y., May, H. D., Lu, L., Liang, P., Huang, X., and Ren, Z. J. (2019). Carbon dioxide and organic waste valorization by microbial electrosynthesis and electro-fermentation. *Water Res.* 149, 42–55. doi: 10.1016/j.watres.2018.10.092
- Jiang, Y., Su, M., Zhang, Y., Zhan, G., Tao, Y., and Li, D. (2013). Bioelectrochemical systems for simultaneously production of methane and acetate from carbon dioxide at relatively high rate. *Int. J. Hydrogen Energy* 38, 3497–3502. doi: 10.1016/j.ijhydene.2012.12.107
- Jin, H.-F., Lim, B.-R., and Lee, K. (2006). Influence of nitrate feeding on carbon dioxide fixation by microalgae. *J. Environ. Sci. Health A Toxicol. Hazard Substain. Environ. Eng.* 41, 2813–2824. doi: 10.1080/10934520600967928
- Jo, B. H., Kim, I. G., Seo, J. H., Kang, D. G., and Cha, H. J. (2013). Engineered *Escherichia coli* with periplasmic carbonic anhydrase as a biocatalyst for CO₂ sequestration. *Appl. Environ. Microbiol.* 79, 6697–6705. doi: 10.1128/AEM.02400-13
- Kadier, A., Jain, P., Lai, B., Kalil, M. S., Kondaveeti, S., Alabbosh, K. F. S., et al. (2020). Biorefinery perspectives of microbial electrolysis cells (MECs) for hydrogen and valuable chemicals production through wastewater treatment. *Biofuel Res. J.* 7, 1128–1142. doi: 10.18331/BRJ2020.7.1.5
- Kaewkannetra, P., Enmak, P., and Chiu, T. (2012). The effect of CO₂ and salinity on the cultivation of *Scenedesmus obliquus* for biodiesel production. *Biotechnol. Bioprocess Eng.* 17, 591–597. doi: 10.1007/s12257-011-0533-5
- Kasting, J. F., and Siefert, J. L. (2002). Life and the evolution of earth's atmosphere. *Science* 296, 1066–1068. doi: 10.1126/science.1071184
- Keffer, J. E., and Kleinheinz, G. T. (2002). Use of *Chlorella vulgaris* for CO₂ mitigation in a photobioreactor. *J. Ind. Microbiol. Biotechnol.* 29, 275–280. doi: 10.1038/sj.jim.7000313
- Keller, M. W., Schut, G. J., Lipscomb, G. L., Menon, A. L., Iwuchukwu, I. J., Leuko, T. T., et al. (2013). Exploiting microbial hyperthermophilicity to produce an industrial chemical, using hydrogen and carbon dioxide. *Proc. Natl. Acad. Sci. U.S.A.* 110, 5840–5845. doi: 10.1073/pnas.1222607110
- Kerdudo, A., Burger, P., Merck, F., Dingas, A., Rolland, Y., Michel, T., et al. (2016). Development of a natural ingredient – natural preservative: a case study. *Comptes Rendus Chimie* 19, 1077–1089. doi: 10.1016/j.crci.2016.06.004
- Khosravanipour Mostafazadeh, A., Drogui, P., Brar, S. K., Tyagi, R. D., Le Bihan, Y., Buelna, G., et al. (2016). Enhancement of biobutanol production by electromicrobial glucose conversion in a dual chamber fermentation cell using *C. pasteurianum*. *Energy Convers. Manage.* 130, 165–175. doi: 10.1016/j.enconman.2016.10.050
- Kim, T. S., and Kim, B. H. (1988). Electron flow shift in *Clostridium acetobutylicum* fermentation by electrochemically introduced reducing equivalent. *Biotechnol. Lett.* 10, 123–128. doi: 10.1007/BF01024638
- Kim, T. W., Bae, S. S., Lee, J. W., Lee, S.-M., Lee, J.-H., Lee, H. S., et al. (2016). A biological process effective for the conversion of CO-containing industrial waste gas to acetate. *Bioresour. Technol.* 211, 792–796. doi: 10.1016/j.biortech.2016.04.038
- Kishimoto, M., Okakura, T., Nagashima, H., Minowa, T., Yokoyama, S.-Y., and Yamaberi, K. (1994). CO₂ fixation and oil production using micro-algae. *J. Ferment. Bioeng.* 78, 479–482. doi: 10.1016/0922-338X(94)90052-3
- Kodama, M., Ikemoto, H., and Miyachi, S. (1993). A new species of highly CO₂ tolerant fast growing marine microalga suitable for high-density culture. *J. Mar. Biotechnol.* 1, 21–25.
- Kondaveeti, S., Choi, K. S., Kakarla, R., and Min, B. (2014a). Microalgae *Scenedesmus obliquus* as renewable biomass feedstock for electricity generation in microbial fuel cells (MFCs). *Front. Environ. Sci. Eng.* 8, 784–791. doi: 10.1007/s11783-013-0590-4
- Kondaveeti, S., Kakarla, R., Kim, H. S., Kim, B.-G., and Min, B. (2017). The performance and long-term stability of low-cost separators in single-chamber bottle-type microbial fuel cells. *Environ. Technol.* 39, 288–297. doi: 10.1080/09593330.2017.1299223
- Kondaveeti, S., Lee, S.-H., Park, H.-D., and Min, B. (2014b). Bacterial communities in a bioelectrochemical denitrification system: the effects of supplemental electron acceptors. *Water Res.* 51, 25–36. doi: 10.1016/j.watres.2013.12.023
- Kondaveeti, S., and Min, B. (2015). Bioelectrochemical reduction of volatile fatty acids in anaerobic digestion effluent for the production of biofuels. *Water Res.* 87, 137–144. doi: 10.1016/j.watres.2015.09.011
- Kratz, W. A., and Myers, J. (1955). Nutrition and growth of several blue-green algae. *Am. J. Bot.* 42, 282–287. doi: 10.1002/j.1537-2197.1955.tb11120.x
- Kuk, S. K., Ham, Y., Gopinath, K., Boonmongkolras, P., Lee, Y., Lee, Y. W., et al. (2019). CO₂ reduction: continuous 3D titanium nitride nanoshell structure for solar-driven unbiased biocatalytic CO₂ reduction. *Adv. Energy Mater.* 9:1970097. doi: 10.1002/aenm.201970097
- Kumar, B., Brian, J. P., Atla, V., Kumari, S., Bertram, K. A., White, R. T., et al. (2016). New trends in the development of heterogeneous catalysts for electrochemical CO₂ reduction. *Catal. Today* 270, 19–30. doi: 10.1016/j.cattod.2016.02.006

- Kumar, K., Dasgupta, C. N., Nayak, B., Lindblad, P., and Das, D. (2011). Development of suitable photobioreactors for CO₂ sequestration addressing global warming using green algae and cyanobacteria. *Bioresour. Technol.* 102, 4945–4953. doi: 10.1016/j.biortech.2011.01.054
- Kuramochi, Y., Fu, Q., Kobayashi, H., Ikarashi, M., Wakayama, T., Kawaguchi, H., et al. (2013). Electromethanogenic CO₂ conversion by subsurface-reservoir microorganisms. *Energy Procedia* 37, 7014–7020. doi: 10.1016/j.egypro.2013.06.636
- Kurano, N., Ikemoto, H., Miyashita, H., Hasegawa, T., Hata, H., and Miyachi, S. (1995). Fixation and utilization of carbon dioxide by microalgal photosynthesis. *Energy Convers. Manage.* 36, 689–692. doi: 10.1016/0196-8904(95)00099-Y
- Kusakabe, T., Tatsuke, T., Tsuruno, K., Hirokawa, Y., Atsumi, S., Liao, J. C., et al. (2013). Engineering a synthetic pathway in cyanobacteria for isopropanol production directly from carbon dioxide and light. *Metab. Eng.* 20, 101–108. doi: 10.1016/j.ymben.2013.09.007
- LaBelle, E. V., and May, H. D. (2017). Energy efficiency and productivity enhancement of microbial electrosynthesis of acetate. *Front. Microbiol.* 8:756. doi: 10.3389/fmicb.2017.00756
- Lambert, G. R., and Smith, G. D. (1977). Hydrogen formation by marine blue-green algae. *FEBS Lett.* 83, 159–162. doi: 10.1016/0014-5793(77)80664-9
- Lau, N.-S., Matsui, M., and Abdullah, A. A.-A. (2015). Cyanobacteria: photoautotrophic microbial factories for the sustainable synthesis of industrial products. *Biomed. Res. Int.* 2015:754934. doi: 10.1155/2015/754934
- Lawton, T. J., and Rosenzweig, A. C. (2016). Biocatalysts for methane conversion: big progress on breaking a small substrate. *Curr. Opin. Chem. Biol.* 35, 142–149. doi: 10.1016/j.cbpa.2016.10.001
- Leang, C., Ueki, T., Nevin, K. P., and Lovley, D. R. (2012). A genetic system for *Clostridium ljungdahlii*: a chassis for autotrophic production of biocommodities and a model homoacetogen. *Appl. Environ. Microbiol.* 79, 1102–1109. doi: 10.1128/AEM.02891-12
- Lee, J. C., Kim, J. H., Chang, W. S., and Pak, D. (2012). Biological conversion of CO₂ to CH₄ using hydrogenotrophic methanogen in a fixed bed reactor. *J. Chem. Technol. Biotechnol.* 87, 844–847. doi: 10.1002/jctb.3787
- Lee, J.-S., and Lee, J.-P. (2003). Review of advances in biological CO₂ mitigation technology. *Biotechnol. Bioprocess Eng.* 8:354. doi: 10.1007/BF02949279
- Lee, W.-K., Lim, Y.-Y., Leow, A. T.-C., Namasivayam, P., Ong Abdullah, J., and Ho, C.-L. (2017). Biosynthesis of agar in red seaweeds: a review. *Carbohydr. Polym.* 164, 23–30. doi: 10.1016/j.carbpol.2017.01.078
- Lehninger, A. L., Nelson, D. L., and Cox, M. M. (2005). *Lehninger Principles of Biochemistry*. New York, NY: W. H. Freeman.
- Li, H., Oppenorth, P. H., Wernick, D. G., Rogers, S., Wu, T.-Y., Higashide, W., et al. (2012). Integrated electromicrobial conversion of CO₂ to higher alcohols. *Science* 335:1596. doi: 10.1126/science.1217643
- Li, J., He, J., and Huang, Y. (2017). Role of alginate in antibacterial finishing of textiles. *Int. J. Biol. Macromol.* 94, 466–473. doi: 10.1016/j.ijbiomac.2016.10.054
- Li, K., An, X., Park, K. H., Khraishah, M., and Tang, J. (2014). A critical review of CO₂ photoconversion: catalysts and reactors. *Catal. Today* 224, 3–12. doi: 10.1016/j.cattod.2013.12.006
- Li, X., Angelidaki, I., and Zhang, Y. (2018). Salinity-gradient energy driven microbial electrosynthesis of value-added chemicals from CO₂ reduction. *Water Res.* 142, 396–404. doi: 10.1016/j.watres.2018.06.013
- Liew, F., Martin, M. E., Tappel, R. C., Heijstra, B. D., Mihalcea, C., and Köpke, M. (2016). Gas fermentation—a flexible platform for commercial scale production of low-carbon-fuels and chemicals from waste and renewable feedstocks. *Front. Microbiol.* 7:694. doi: 10.3389/fmicb.2016.00694
- Liu, M., Pang, Y., Zhang, B., De Luna, P., Voznyy, O., Xu, J., et al. (2016). Enhanced electrocatalytic CO₂ reduction via field-induced reagent concentration. *Nature* 537, 382–386. doi: 10.1038/nature19060
- Ljungdahl, L., and Wood, H. G. (1965). Incorporation of C¹⁴ From Carbon Dioxide into Sugar Phosphates, Carboxylic Acids, and Amino Acids by *Clostridium thermoaceticum*. *J. Bacteriol.* 89, 1055–1064.
- Logan, B. E. (2008). *Microbial Fuel Cells*. Hoboken, NJ: John Wiley and sons, Inc.
- Lovley, D. R., and Nevin, K. (2018). *Microbial Production of Multi-carbon Chemicals and Fuels from Water and Carbon Dioxide Using Electric Current*. U.S. Patent, 9856449. Available online at: <https://tinyurl.com/yahnawoc>
- Lowe, R. L., and LaLiberte, G. D. (2017). “Chapter 11 - benthic stream algae: distribution and structure,” in *Methods in Stream Ecology*, 3rd Edn, Vol. 1. (Boston, MA: Academic Press), 193–221. doi: 10.1016/B978-0-12-416558-8.00011-1
- Luengo, J. M., Garcia, B., Sandoval, A., Naharro, G., and Olivera, E. A. R. (2003). Bioplastics from microorganisms. *Curr. Opin. Microbiol.* 6, 251–260. doi: 10.1016/S1369-5274(03)00040-7
- Maeda, K., Owada, M., Kimura, N., Omata, K., and Karube, I. (1995). CO₂ fixation from the flue gas on coal-fired thermal power plant by microalgae. *Energy Convers. Manage.* 36, 717–720. doi: 10.1016/0196-8904(95)00105-M
- Mandotra, S., Kumar, P., Suseela, M., Nayaka, S., and Ramteke, P. (2016). Evaluation of fatty acid profile and biodiesel properties of microalga *Scenedesmus abundans* under the influence of phosphorus, pH and light intensities. *Bioresour. Technol.* 201, 222–229. doi: 10.1016/j.biortech.2015.11.042
- Mark Ibekwe, A., Murinda, S. E., Murry, M. A., Schwartz, G., and Lundquist, T. (2017). Microbial community structures in high rate algae ponds for bioconversion of agricultural wastes from livestock industry for feed production. *Sci. Total Environ.* 580, 1185–1196. doi: 10.1016/j.scitotenv.2016.12.076
- Market Study Report (2019). *Capric Acid Market Size is Expected to Exhibit 270 million USD by 2024*. Available online at: <https://www.marketwatch.com/press-release/capric-acid-market-size-is-expected-to-exhibit-270-million-usd-by-2024-2019-09-05> (accessed February 15, 2020).
- Markets and Markets (2019). *Succinic Acid Market worth \$237.8 million by 2027*. Available online at: <https://www.marketsandmarkets.com/PressReleases/succinic-acid.asp> (accessed February 15, 2020).
- Marshall, C. W., LaBelle, E. V., and May, H. D. (2013). Production of fuels and chemicals from waste by microbiomes. *Curr. Opin. Biotechnol.* 24, 391–397. doi: 10.1016/j.copbio.2013.03.016
- Martin, M. E., Richter, H., Saha, S., and Angenent, L. T. (2016). Traits of selected *Clostridium* strains for syngas fermentation to ethanol. *Biotechnol. Bioeng.* 113, 531–539. doi: 10.1002/bit.25827
- Maurya, R., Paliwal, C., Ghosh, T., Pancha, I., Chokshi, K., Mitra, M., et al. (2016). Applications of de-oiled microalgal biomass towards development of sustainable biorefinery. *Bioresour. Technol.* 214, 787–796. doi: 10.1016/j.biortech.2016.04.115
- Modestra, J. A., and Mohan, S. V. (2017). Microbial electrosynthesis of carboxylic acids through CO₂ reduction with selectively enriched biocatalyst: microbial dynamics. *J. CO₂ Util.* 20, 190–199. doi: 10.1016/j.jcou.2017.05.011
- Mohanakrishna, G., Abu Reesh, I. M., Vanbroekhoven, K., and Pant, D. (2020). Microbial electrosynthesis feasibility evaluation at high bicarbonate concentrations with enriched homoacetogenic biocathode. *Sci. Total Environ.* 715:137003. doi: 10.1016/j.scitotenv.2020.137003
- Mohanakrishna, G., Seelam, J. S., Vanbroekhoven, K., and Pant, D. (2015). An enriched electroactive homoacetogenic biocathode for the microbial electrosynthesis of acetate through carbon dioxide reduction. *Faraday Discuss.* 183, 445–462. doi: 10.1039/C5FD00041F
- Mohanakrishna, G., Vanbroekhoven, K., and Pant, D. (2016). Imperative role of applied potential and inorganic carbon source on acetate production through microbial electrosynthesis. *J. CO₂ Util.* 15(Suppl. C), 57–64. doi: 10.1016/j.jcou.2016.03.003
- Mohd Yasin, N. H., Maeda, T., Hu, A., Yu, C.-P., and Wood, T. K. (2015). CO₂ sequestration by methanogens in activated sludge for methane production. *Appl. Energy* 142, 426–434. doi: 10.1016/j.apenergy.2014.12.069
- Moreira, D., and Pires, J. C. M. (2016). Atmospheric CO₂ capture by algae: negative carbon dioxide emission path. *Bioresour. Technol.* 215, 371–379. doi: 10.1016/j.biortech.2016.03.060
- Müller, J., MacEachran, D., Burd, H., Sathitsuksanoh, N., Bi, C., Yeh, Y.-C., et al. (2013). Engineering of *Ralstonia eutropha* H16 for autotrophic and heterotrophic production of methyl ketones. *Appl. Environ. Microbiol.* 79, 4433–4439. doi: 10.1128/AEM.00973-13
- Murakami, M., and Ikenouchi, M. (1997). The biological CO₂ fixation and utilization project by rite (2) — screening and breeding of microalgae with high capability in fixing CO₂ —. *Energy Convers. Manage.* 38(Suppl.), S493–S497. doi: 10.1016/S0196-8904(96)00316-0
- Nagarajan, S., Chou, S. K., Cao, S., Wu, C., and Zhou, Z. (2013). An updated comprehensive techno-economic analysis of algae biodiesel.

- Bioresour. Technol.* 145, 150–156. doi: 10.1016/j.biortech.2012.11.108
- Nautiyal, P., Subramanian, K. A., and Dastidar, M. G. (2016). Adsorptive removal of dye using biochar derived from residual algae after *in-situ* transesterification: alternate use of waste of biodiesel industry. *J. Environ. Manage.* 182, 187–197. doi: 10.1016/j.jenvman.2016.07.063
- Negoro, M., Shioji, N., Miyamoto, K., and Micira, Y. (1991). Growth of microalgae in high CO₂ gas and effects of SOX and NOX. *Appl. Biochem. Biotechnol.* 28, 877–886. doi: 10.1007/BF02922657
- Nevin, K. P., Hensley, S. A., Franks, A. E., Summers, Z. M., Ou, J., Woodard, T. L., et al. (2011). Electrosynthesis of organic compounds from carbon dioxide is catalyzed by a diversity of acetogenic microorganisms. *Appl. Environ. Microbiol.* 77, 2882–2886. doi: 10.1128/AEM.02642-10
- Nevin, K. P., Woodard, T. L., Franks, A. E., Summers, Z. M., and Lovley, D. R. (2010). Microbial electrosynthesis: feeding microbes electricity to convert carbon dioxide and water to multicarbon extracellular organic compounds. *mBio* 1:e00103-10. doi: 10.1128/mBio.00103-10
- Nigam, P. S., and Singh, A. (2011). Production of liquid biofuels from renewable resources. *Prog. Energy Combust. Sci.* 37, 52–68. doi: 10.1016/j.pecs.2010.01.003
- Overhage, J., Steinbüchel, A., and Priefert, H. (2002). Biotransformation of eugenol to ferulic acid by a recombinant strain of *Ralstonia eutropha* H16. *Appl. Environ. Microbiol.* 68, 4315–4321. doi: 10.1128/AEM.68.9.4315-4321.2002
- Packer, M. (2009). Algal capture of carbon dioxide; biomass generation as a tool for greenhouse gas mitigation with reference to New Zealand energy strategy and policy. *Energy Policy* 37, 3428–3437. doi: 10.1016/j.enpol.2008.12.025
- Patel, T. N., Park, A-H. A., and Banta, S. (2013). Periplasmic expression of carbonic anhydrase in *Escherichia coli*: a new biocatalyst for CO₂ hydration. *Biotechnol. Bioeng.* 110, 1865–1873. doi: 10.1002/bit.24863
- Petersen, H. I., Rosenberg, P., and Nytoft, H. P. (2008). Oxygen groups in coals and alginite-rich kerogen revisited. *Int. J. Coal Geol.* 74, 93–113. doi: 10.1016/j.coal.2007.11.007
- Pfennig, N. (1967). Photosynthetic bacteria. *Annu. Rev. Microbiol.* 21, 285–324. doi: 10.1146/annurev.mi.21.100167.001441
- Puig, S., Batlle-Vilanova, R.G. P., Dolers Balaguer, M., and Colprim, J. (2015). Microbial electrosynthesis of C₂-C₄ platform chemicals from carbon dioxide. *ACS Sustain. Chem. Eng.* 6, 8485–8493.
- Quintana, N., Van der Kooy, F., Van de Rhee, M. D., Voshol, G. P., and Verpoorte, R. (2011). Renewable energy from cyanobacteria: energy production optimization by metabolic pathway engineering. *Appl. Microbiol. Biotechnol.* 91, 471–490. doi: 10.1007/s00253-011-3394-0
- Rabaey, K., Angenent, L., Schroder, U., and Keller, J. (2009). *Bioelectrochemical Systems*. London: IWA Publishing.
- Rabaey, K., and Rozendal, R. A. (2010). Microbial electrosynthesis — revisiting the electrical route for microbial production. *Nat. Rev. Microbiol.* 8, 706–716. doi: 10.1038/nrmicro2422
- Ramanan, R., Kannan, K., Deshkar, A., Yadav, R., and Chakrabarti, T. (2010). Enhanced algal CO₂ sequestration through calcite deposition by *Chlorella* sp. and *Spirulina platensis* in a mini-raceway pond. *Bioresour. Technol.* 101, 2616–2622. doi: 10.1016/j.biortech.2009.10.061
- Rana, D., Kim, Y., Matsuura, T., and Arafat, H. A. (2011). Development of antifouling thin-film-composite membranes for seawater desalination. *J. Membr. Sci.* 367, 110–118. doi: 10.1016/j.memsci.2010.10.050
- Rashid, N., Rehman, M. S. U., and Han, J-I. (2013). Recycling and reuse of spent microalgal biomass for sustainable biofuels. *Biochem. Eng. J.* 75, 101–107. doi: 10.1016/j.bej.2013.04.001
- Reinecke, F., and Steinbüchel, A. (2009). *Ralstonia eutropha* strain H16 as model organism for PHA metabolism and for biotechnological production of technically interesting biopolymers. *J. Mol. Microbiol. Biotechnol.* 16, 91–108. doi: 10.1159/000142897
- Rengasamy, K. R. R., Kulkarni, M. G., Papenfus, H. B., and Van Staden, J. (2016). Quantification of plant growth biostimulants, phloroglucinol and eckol, in four commercial seaweed liquid fertilizers and some by-products. *Algal Res.* 20, 57–60. doi: 10.1016/j.algal.2016.09.017
- Richter, H., Martin, M. E., and Angenent, L. T. (2013). A two-stage continuous fermentation system for conversion of syngas into ethanol. *Energies* 6, 3987–4000. doi: 10.3390/en6083987
- Sadhukhan, J., Lloyd, J. R., Scott, K., Premier, G. C., Yu, E. H., Curtis, T., et al. (2016). A critical review of integration analysis of microbial electrosynthesis (MES) systems with waste biorefineries for the production of biofuel and chemical from reuse of CO₂. *Renew. Sustain. Energy Rev.* 56, 116–132. doi: 10.1016/j.rser.2015.11.015
- Sakai, N., Sakamoto, Y., Kishimoto, N., Chihara, M., and Karube, I. (1995). *Chlorella* strains from hot springs tolerant to high temperature and high CO₂. *Energy Convers. Manage.* 36, 693–696. doi: 10.1016/0196-8904(95)00100-R
- Samarakoon, K., and Jeon, Y-J. (2012). Bio-functionalities of proteins derived from marine algae — a review. *Food Res. Int.* 48, 948–960. doi: 10.1016/j.foodres.2012.03.013
- Sarkar, O., Agarwal, M., Naresh Kumar, A., and Venkata Mohan, S. (2015). Retrofitting heterotrophically cultivated algae biomass as pyrolytic feedstock for biogas, bio-char and bio-oil production encompassing biorefinery. *Bioresour. Technol.* 178, 132–138. doi: 10.1016/j.biortech.2014.09.070
- Sawitzke, J. A., Thomason, L. C., Costantino, N., Bubunenko, M., Datta, S., and Court, D. L. (2007). Recombining: *in vivo* genetic engineering in *E. coli*, *S. enterica*, and Beyond. *Methods Enzymol.* 421, 171–199. doi: 10.1016/S0076-6879(06)21015-2
- Sayre, R. (2010). Microalgae: the potential for carbon capture. *Bioscience* 60, 722–727. doi: 10.1525/bio.2010.60.9.9
- Schenk, P. M., Thomas-Hall, S. R., Stephens, E., Marx, U. C., Mussgnug, J. H., Posten, C., et al. (2008). Second generation biofuels: high-efficiency microalgae for biodiesel production. *Bioenergy Res.* 1, 20–43. doi: 10.1007/s12155-008-9008-8
- Scherer, P. A., Vollmer, G-R., Fakhouri, T., and Martensen, S. (2000). Development of a methanogenic process to degrade exhaustively the organic fraction of municipal “grey waste” under thermophilic and hyperthermophilic conditions. *Water Sci. Technol.* 41, 83–91. doi: 10.2166/wst.2000.0059
- Schirmer, A., Rude, M. A., Li, X., Popova, E., and Del Cardayre, S. B. (2010). Microbial biosynthesis of alkanes. *Science* 329, 559–562. doi: 10.1126/science.1187936
- Schlager, S., Neugebauer, H., Haberbauer, M., Hinterberger, G., and Sariciftci, N. S. (2015). Direct electrochemical addressing of immobilized alcohol dehydrogenase for the heterogeneous bioelectrocatalytic reduction of butyraldehyde to butanol. *ChemCatChem* 7, 967–971. doi: 10.1002/cctc.201402932
- Scragg, A. H., Illman, A. M., Carden, A., and Shales, S. W. (2002). Growth of microalgae with increased calorific values in a tubular bioreactor. *Biomass Bioenergy* 23, 67–73. doi: 10.1016/S0961-9534(02)00028-4
- Sezenna, M. L. (2011). *Proteobacteria: Phylogeny, Metabolic Diversity and Ecological Effects*. New York, NY: Nova Science.
- Sharma, M., Aryal, N., Sarma, P. M., Vanbroekhoven, K., Lal, B., Benetton, X. D., et al. (2013a). Bioelectrocatalyzed reduction of acetic and butyric acids via direct electron transfer using a mixed culture of sulfate-reducers drives electrosynthesis of alcohols and acetone. *Chem. Commun.* 49, 6495–6497. doi: 10.1039/c3cc42570c
- Sharma, M., Jain, P., Varanasi, J. L., Lal, B., Rodríguez, J., Lema, J. M., et al. (2013b). Enhanced performance of sulfate reducing bacteria based biocathode using stainless steel mesh on activated carbon fabric electrode. *Bioresour. Technol.* 150, 172–180. doi: 10.1016/j.biortech.2013.09.069
- Shi, J., Jiang, Y., Jiang, Z., Wang, X., Wang, X., Zhang, S., et al. (2015). Enzymatic conversion of carbon dioxide. *Chem. Soc. Rev.* 44, 5981–6000. doi: 10.1039/C5CS00182J
- Silva, C., Soliman, E., Cameron, G., Fabiano, L. A., Seider, W. D., Dunlop, E. H., et al. (2014). Commercial-scale biodiesel production from algae. *Ind. Eng. Chem. Res.* 53, 5311–5324. doi: 10.1021/ie403273b
- Singh, A., Nigam, P. S., and Murphy, J. D. (2011). Mechanism and challenges in commercialisation of algal biofuels. *Bioresour. Technol.* 102, 26–34. doi: 10.1016/j.biortech.2010.06.057
- Singh, S. P., and Singh, P. (2014). Effect of CO₂ concentration on algal growth: a review. *Renew. Sustain. Energy Rev.* 38, 172–179. doi: 10.1016/j.rser.2014.05.043
- Sorokin, C., and Krauss, R. W. (1958). The effects of light intensity on the growth rates of green algae. *Plant Physiol.* 33, 109–113. doi: 10.1104/pp.33.2.109
- Steinbusch, K. J. J., Hamelers, H. V. M., Schaap, J. D., Kampman, C., and Buisman, C. J. N. (2010). Bioelectrochemical ethanol production through mediated acetate reduction by mixed cultures. *Environ. Sci. Technol.* 44, 513–517. doi: 10.1021/es902371e
- Strong, P. J., Kalyuzhnaya, M., Silverman, J., and Clarke, W. P. (2016). A methanotroph-based biorefinery: potential scenarios for generating multiple

- products from a single fermentation. *Bioresour. Technol.* 215, 314–323. doi: 10.1016/j.biortech.2016.04.099
- Sung, K. D., Lee, J. S., Shin, C. S., and Park, S. C. (1999). Isolation of a new highly CO₂ tolerant fresh water microalga *Chlorella* SP. KR-1. *Renew. Energy* 16, 1019–1022. doi: 10.1016/S0960-1481(98)00362-0
- Sydney, E. B., Sturm, W., de Carvalho, J. C., Thomaz-Soccol, V., Larroche, C., Pandey, A., et al. (2010). Potential carbon dioxide fixation by industrially important microalgae. *Bioresour. Technol.* 101, 5892–5896. doi: 10.1016/j.biortech.2010.02.088
- Tahvonen, O., Von Storch, H., and Storch, J.-S. (1994). Economic efficiency of CO₂ reduction programs. *Clim. Res.* 4, 127–141. doi: 10.3354/cr004127
- Tamagnini, P., Axelsson, R., Lindberg, P., Oxelfelt, F., Wünschiers, R., and Lindblad, P. (2002). Hydrogenases and hydrogen metabolism of cyanobacteria. *Microbiol. Mol. Biol. Rev.* 66, 1–20. doi: 10.1128/MMBR.66.1.1-20.2002
- Tanaka, K., Miyawaki, K., Yamaguchi, A., Khosravi-Darani, K., and Matsusaki, H. (2011). Cell growth and P(3HB) accumulation from CO₂ of a carbon monoxide-tolerant hydrogen-oxidizing bacterium, *Ideonella* sp. O-1. *Appl. Microbiol. Biotechnol.* 92, 1161–1169. doi: 10.1007/s00253-011-3420-2
- Tang, D., Han, W., Li, P., Miao, X., and Zhong, J. (2011). CO₂ biofixation and fatty acid composition of *Scenedesmus obliquus* and *Chlorella pyrenoidosa* in response to different CO₂ levels. *Bioresour. Technol.* 102, 3071–3076. doi: 10.1016/j.biortech.2010.10.047
- Tracy, B. P., Jones, S. W., Fast, A. G., Indurthi, D. C., and Papoutsakis, E. T. (2012). Clostridia: the importance of their exceptional substrate and metabolite diversity for biofuel and biorefinery applications. *Curr. Opin. Biotechnol.* 23, 364–381. doi: 10.1016/j.copbio.2011.10.008
- Trivedi, J., Aila, M., Bangwal, D. P., Kaul, S., and Garg, M. O. (2015). Algae based biorefinery—how to make sense? *Renew. Sustain. Energy Rev.* 47, 295–307. doi: 10.1016/j.rser.2015.03.052
- Tsuzuki, M., Ohnuma, E., Sato, N., Takaku, T., and Kawaguchi, A. (1990). Effects of CO₂ concentration during growth on fatty acid composition in microalgae. *Plant Physiol.* 93, 851–856. doi: 10.1104/pp.93.3.851
- Ueki, T., Nevin, K. P., Woodard, T. L., and Lovley, D. R. (2014). Converting carbon dioxide to butyrate with an engineered strain of *Clostridium ljungdahlii*. *mBio* 5:e01636-14. doi: 10.1128/mBio.01636-14
- Van Eerten-Jansen, M. C. A. A., Heijne, A. T., Buisman, C. J. N., and Hamelers, H. V. M. (2012). Microbial electrolysis cells for production of methane from CO₂: long-term performance and perspectives. *Int. J. Energy Res.* 36, 809–819. doi: 10.1002/er.1954
- Van Eerten-Jansen, M. C. A. A., Ter Heijne, A., Grootsholten, T. I. M., Steinbusch, K. J. J., Sleutels, T. H. J. A., Hamelers, H. V. M., et al. (2013). Bioelectrochemical production of caproate and caprylate from acetate by mixed cultures. *ACS Sustain. Chem. Eng.* 1, 513–518. doi: 10.1021/sc300168z
- Vassilev, I., Hernandez, P. A., Battle-Vilanova, P., Freguia, S., Krömer, J. O., Keller, J., et al. (2018). Microbial electrosynthesis of isobutyric, butyric, caproic acids, and corresponding alcohols from carbon dioxide. *ACS Sustain. Chem. Eng.* 6, 8485–8493. doi: 10.1021/acssuschemeng.8b00739
- Vassilev, I., Kracke, F., Freguia, S., Keller, J., Krömer, J. O., Ledezma, P., et al. (2019). Microbial electrosynthesis system with dual biocathode arrangement for simultaneous acetogenesis, solventogenesis and carbon chain elongation. *Chem. Commun.* 55, 4351–4354. doi: 10.1039/C9CC00208A
- Vassilev, S. V., and Vassileva, C. G. (2016). Composition, properties and challenges of algae biomass for biofuel application: an overview. *Fuel* 181, 1–33. doi: 10.1016/j.fuel.2016.04.106
- Venil, C. K., Zakaria, Z. A., and Ahmad, W. A. (2013). Bacterial pigments and their applications. *Process Biochem.* 48, 1065–1079. doi: 10.1016/j.procbio.2013.06.006
- Venkata Mohan, S., Hemalatha, M., Chakraborty, D., Chatterjee, S., Ranadheer, P., and Kona, R. (2020). Algal biorefinery models with self-sustainable closed loop approach: trends and prospective for blue-bioeconomy. *Bioresour. Technol.* 295:122128. doi: 10.1016/j.biortech.2019.122128
- Venkata Mohan, S., Modestra, J. A., Amulya, K., Butti, S. K., and Velvizhi, G. (2016). A circular bioeconomy with biobased products from CO₂ sequestration. *Trends Biotechnol.* 34, 506–519. doi: 10.1016/j.tibtech.2016.02.012
- Venkata Mohan, S., Prathima Devi, M., Mohanakrishna, G., Amarnath, N., Lenin Babu, M., and Sarma, P. N. (2011). Potential of mixed microalgae to harness biodiesel from ecological water-bodies with simultaneous treatment. *Bioresour. Technol.* 102, 1109–1117. doi: 10.1016/j.biortech.2010.08.103
- Vigani, M., Parisi, C., Rodríguez-Cerezo, E., Barbosa, M. J., Sijtsma, L., Ploeg, M., et al. (2015). Food and feed products from micro-algae: market opportunities and challenges for the EU. *Trends Food Sci. Technol.* 42, 81–92. doi: 10.1016/j.tifs.2014.12.004
- Villano, M., Aulenta, F., Ciucci, C., Ferri, T., Giuliano, A., and Majone, M. (2010). Bioelectrochemical reduction of CO₂ to CH₄ via direct and indirect extracellular electron transfer by a hydrogenophilic methanogenic culture. *Bioresour. Technol.* 101, 3085–3090. doi: 10.1016/j.biortech.2009.12.077
- Villano, M., Scardala, S., Aulenta, F., and Majone, M. (2013). Carbon and nitrogen removal and enhanced methane production in a microbial electrolysis cell. *Bioresour. Technol.* 130, 366–371. doi: 10.1016/j.biortech.2012.11.080
- Voss, I., and Steinbüchel, A. (2006). Application of a KDPG-aldolase gene-dependent addition system for enhanced production of cyanophycin in *Ralstonia eutropha* strain H16. *Metab. Eng.* 8, 66–78. doi: 10.1016/j.ymben.2005.09.003
- Wang, B., Albarracín-Suazo, S., Pagán-Torres, Y., and Nikolla, E. (2017). Advances in methane conversion processes. *Catal. Today* 285, 147–158. doi: 10.1016/j.cattod.2017.01.023
- Wang, B., Wang, J., Zhang, W., and Meldrum, D. R. (2012). Application of synthetic biology in cyanobacteria and algae. *Front. Microbiol.* 3:344. doi: 10.3389/fmicb.2012.00344
- Wang, C., Scott Buchanan, J., Kliever, W. R., and Qian, K. (2015). Water-washing to reduce metals in oils extracted from nanochloropsis algae for potential FCC feedstock. *Fuel* 155, 63–67. doi: 10.1016/j.fuel.2015.03.074
- Wang, H.-M. D., Chen, C.-C., Huynh, P., and Chang, J.-S. (2015). Exploring the potential of using algae in cosmetics. *Bioresour. Technol.* 184, 355–362. doi: 10.1016/j.biortech.2014.12.001
- Wang, Q., Dong, H., Yu, H., and Yu, H. (2015). Enhanced performance of gas diffusion electrode for electrochemical reduction of carbon dioxide to formate by adding polytetrafluoroethylene into catalyst layer. *J. Power Sources* 279, 1–5. doi: 10.1016/j.jpowsour.2014.12.118
- Wang, W.-N. (2014). Comparison of CO₂ photoreduction systems: a review. *Aerosol Air Qual. Res.* 14, 533–549. doi: 10.4209/aaqr.2013.09.0283
- Wargacki, A. J., Leonard, E., Win, M. N., Regitsky, D. D., Santos, C. N. S., Kim, P. B., et al. (2012). An engineered microbial platform for direct biofuel production from brown macroalgae. *Science* 335, 308–313. doi: 10.1126/science.1214547
- Watanabe, Y., Ohmura, N., and Saiki, H. (1992). Isolation and determination of cultural characteristics of microalgae which functions under CO₂ enriched atmosphere. *Energy Convers. Manage.* 33, 545–552. doi: 10.1016/0196-8904(92)90054-Z
- Will, F. G. (1965). Hydrogen adsorption on platinum single crystal electrodes I. Isotherms and heats of adsorption. *J. Electrochem. Soc.* 112, 451–455. doi: 10.1149/1.2423567
- Wiley, J. M., Sherwood, L. M., and Woolverton, C. J. (2014). *Prescott's Microbiology*. New York, NY: McGraw-Hill Education.
- Xia, A., Jacob, A., Tabassum, M. R., Herrmann, C., and Murphy, J. D. (2016). Production of hydrogen, ethanol and volatile fatty acids through co-fermentation of macro- and micro-algae. *Bioresour. Technol.* 205, 118–125. doi: 10.1016/j.biortech.2016.01.025
- Xin, L., Hong-Ying, H., and Yu-Ping, Z. (2011). Growth and lipid accumulation properties of a freshwater microalga *Scenedesmus* sp. under different cultivation temperature. *Bioresour. Technol.* 102, 3098–3102. doi: 10.1016/j.biortech.2010.10.055
- Xu, Y., Ibrahim, I. M., and Harvey, P. J. (2016). The influence of photoperiod and light intensity on the growth and photosynthesis of *Dunaliella salina* (chlorophyta) CCAP 19/30. *Plant Physiol. Biochem.* 106, 305–315. doi: 10.1016/j.plaphy.2016.05.021
- Yoo, C., Jun, S.-Y., Lee, J.-Y., Ahn, C.-Y., and Oh, H.-M. (2010). Selection of microalgae for lipid production under high levels carbon dioxide. *Bioresour. Technol.* 101(Suppl. 1), S71–S74. doi: 10.1016/j.biortech.2009.03.030
- Youn, M. H., Park, K. T., Lee, Y. H., Kang, S.-P., Lee, S. M., Kim, S. S., et al. (2019). Carbon dioxide sequestration process for the cement industry. *J. CO₂ Util.* 34, 325–334. doi: 10.1016/j.jcou.2019.07.023

- Zakaria, Z., and Kamarudin, S. K. (2016). Direct conversion technologies of methane to methanol: an overview. *Renew. Sustain. Energy Rev.* 65, 250–261. doi: 10.1016/j.rser.2016.05.082
- Zeikus, J. (1977). The biology of methanogenic bacteria. *Bacteriol. Rev.* 41, 514–541. doi: 10.1128/MMBR.41.2.514-541.1977
- Zhang, L., Singh, R. D. S., Guo, Z., Li, J., Chen, F., et al. (2018). An artificial synthetic pathway for acetoin, 2,3-butanediol, and 2-butanol production from ethanol using cell free multi-enzyme catalysis. *Green Chem.* 20, 230–242. doi: 10.1039/C7GC02898A
- Zhang, T., Nie, H., Bain, T. S., Lu, H., Cui, M., Snoeyenbos-West, O. L., et al. (2013). Improved cathode materials for microbial electrosynthesis. *Energy Environ. Sci.* 6, 217–224. doi: 10.1039/C2EE23350A
- Zhang, Y., and Kendall, A. (2019). Consequential analysis of algal biofuels: benefits to ocean resources. *J. Clean. Prod.* 231, 35–42. doi: 10.1016/j.jclepro.2019.05.057
- Zhu, L. D., Li, Z. H., and Hiltunen, E. (2016). Strategies for lipid production improvement in microalgae as a biodiesel feedstock. *Biomed. Res. Int.* 2016:8792548. doi: 10.1155/2016/8792548
- Zhuang, Z.-Y., and Li, S.-Y. (2013). Rubisco-based engineered *Escherichia coli* for *in situ* carbon dioxide recycling. *Bioresour. Technol.* 150, 79–88. doi: 10.1016/j.biortech.2013.09.116

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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