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Age-related Modulation of the Nitrogen Resorption Efficiency Response to Growth

2 Requirements and Soil Nitrogen Availability in a Temperate Pine Plantation

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ABSTRACT

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- Nitrogen (N) resorption is a key strategy for conserving N in forests, and is often affected by soil nutrient condition and N sink strength within the plant. However, our understanding of the age-related pattern of N resorption and how increasing N deposition will affect this pattern is limited. Here, we investigated N resorption along a chronosequence of stands ranging in age from 2 to 100 years old, and conducted a 4-year exogenous N input experiment in stands at age class 11, 20 and 45 in a Larix Principis-rupprechtii plantation in north China. We found a logarithmic increase in leaf NRE and green leaf N concentration, and a logarithmic decrease in senesced-leaf N concentration along the stand-age chronosequence. Leaf NRE was negatively correlated with plant-available N concentration. Stand-level N resorption was positively correlated with the annual N requirement for tree growth. N resorption contributed to 45%, 62% and 68% of the annual N supply in the 11-, 20- and 45-year-old stands, respectively. Our exogenous N input experiment showed that leaf NRE in the 11- and 20-year-old stands decreased 17 and 12% following a 50 kg N ha⁻¹ yr⁻¹ input. However, leaf NRE was not affected in the 45-year-old stand. The increases in leaf NRE and the contribution of N resorption to annual N supply along stand ages suggested that, with stand development, tree growth depends more on N resorption to supply its N need. Furthermore, the leaf NRE of mature stand was not decreased under exogenous N input, suggesting that mature stands can be stronger sinks for N deposition than young stands due to their higher capacity to retain the deposited N within plants via internal cycle. Ignoring age-related N use strategies can lead to a bias in N cycle models when evaluating forest net primary production under increasing global N deposition.
- 48 **Key words:** nitrogen resorption; green leaves; senesced leaves; nitrogen requirement; nitrogen deposition;
- stand age; plant-available nitrogen; annual stand biomass production.

INTRODUCTION

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Afforestation is one of the most important strategies for mitigating global climate change. In the past two decades, planted forest area has steadily increased globally by around 4.3 million hectares per year, and now makes up 6-7% of global forests. Most planted forests are distributed in temperate areas in Asia, Europe, and North America (Paquette and Messier 2009; FAO 2010). Forest plantations in temperate areas are often limited by nitrogen (N) availability (Vitousek and Howarth 1991; Magnani and others 2007), but also encounter rapid increases in anthropogenic N deposition (Galloway and others 2008). As planted stands develop, the balance between N supply and N requirement for tree growth often changes as well (Gholz and others 1985; Peri and others 2006). Without understanding how stand development and exogenous N input interact to affect temperate plantations' internal N cycling and demand, we could not evaluate the potential role of plantations in alleviating climate warming while supplying wood products. N resorption, the most important aspect of internal plant nutrient cycling (Killingbeck 1996), supplies about 36-76% of the annual N demand for forest growth (Bond-Lamberty and others 2006). Downregulation of N resorption efficiency (NRE) has often been observed in trees grown on N-rich soils or after N fertilization (Small 1972; Yuan and Chen 2015). Meanwhile, NRE can be affected by phloem transportation rates and loadings (Chapin and Moilanen 1991), and also the sink strength driving by demand such as producing N-rich reproductive structures (Tully and others 2013). Furthermore, rates of nutrient translocation may increase linearly with increases in the rates of tree growth (Nambiar and Fife 1991). As young stands begin to mature, tree growth rate tend to get its maximum and soil available N often decline (Gower and others 1996; Tang and others 2014). To adapt to the increasing severity of N limitation during stand development, old stands tend to be more economical in their N strategies than juvenile stands (Gholz and others 1985; Mediavilla and others 2014). To this end, N resorption, an

important process of internal N cycling and storage (Wang and others 2013), increased greatly after the canopy closed (at 8 years old) in a northern Florida pine forest (Gholz and others 1985). A study of several boreal stands also found that N resorption was significantly higher in older stands (Bond-Lamberty and others 2006). N resorption as an important component of plant physiological and metabolic processes (Wang and others 2013) is therefore expected to change with stand aging (Yuan and Chen 2010), although studies on the topic are scarce.

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Most forests growth is generally thought to be stimulated by exogenous N input (Thomas and others 2010), which results in increases in annual net primary production (Bown and others 2010; Vicca and others 2012). However, stands of different ages differ in their growth responses to N input. For example, exogenous N input stimulated the growth of 1-year-old Cryptomeria japonica seedlings in Japan (Nakaji and others 2001). By contrast, N input stimulated the growth of 18-year-old Pinus sylvestris stands in a boreal forest in Sweden for the first 7 years, but reduced it after that (Högberg and others 2006). Such age-based responses may occur because the growth rates of dominant tree species, plant community composition, and N requirements change with stand development (Lehtonen and others 2004; Tang and others 2014). In addition to vegetation growth and composition, many studies have found that stand age also regulates the responses of microbial community composition and activity to N deposition (Allison and others 2010; Ma and others 2013), which could further influence N cycling in the soil. We expect that the N needed for growth in mature forests depends largely on the internal N cycle, such as greater N withdrawal from senescing leaves and reduced dependence on external nutrient availability. Thus, the effects of exogenous N input on N resorption should vary with forest age, we expect older forests to have a higher NRE and be less responsive to exogenous N inputs, yet few data with which to test this hypothesis are available.

Nitrogen resorption and plant-available N in soil are both vital in whether the supply of N can satisfy the demand for forest growth. However, to our knowledge, no attempts have been made to assess the dynamics between N resorption and N demand during stand development under increasing N deposition. Here, we investigate NRE in larch (*Larix principis-rupprechtii*) plantations of five ages (2–100 years) at Saihanba National Forest Park in China, the largest plantation (~94,700 ha) in East Asia. We also simulated N deposition by adding N to Larch plantations of three different ages. We measured N concentration in both green and senesced leaves of *Larix Principis-rupprechtii* from the various stand-age classes. We assessed annual production in stand biomass, N required for tree growth, and soil N availability in the forest under ambient and simulated N deposition treatments. We hypothesized that 1) N resorption and requirements vary during stand development; 2) the faster growing stands have a higher N requirement and therefore lead to a greater N resorption; and 3) N deposition alleviates N limitation and therefore reduces NRE.

MATERIALS AND METHODS

Site Description and Experimental Design

The study site was located at the Saihanba Ecological Station (42°25′N, 117°15′E, 1505 m a.s.l) of Peking University in Saihanba National Forest Park, Hebei Province, China. The topography of the study site is relatively flat, and the soil is predominantly sandy. The mean annual temperature is -1.4°C (-21.8°C in January and 16.2°C in July), and the mean annual precipitation is 450 mm. The site is frost-free for 81 d each year (Ma and others 2014). Snowfall normally begins in November, and snowmelt occurs in early April. In winter, the snow depth is typically < 30 cm. The ambient nitrogen deposition is 13 kg ha⁻¹ yr⁻¹.

Five stands, aged 2, 11, 20, 45, and 100 years, were selected in August 2009. The dominant tree

species was *Larix principis-rupprechtii*, with the 45-year-old stand thinned in 1989. The distance between any two stands was less than 2 km, and all stands have similar climate and the soils are all classified as sandy soil. In 2010, the 11-, 20-, and 45-year-old stands were selected for a long-term exogenous N input experiment. Details of the three stands' properties are listed in Table 1. In each of these three forests, the experimental area consisted of nine 20 × 20-m² plots, with wide buffer zones (> 10 m) between them. In each stand, nine plots were randomly assigned as control (no N added, N0), low N input (20 kg N ha⁻¹ year⁻¹, N20), or high N input (50 kg N ha⁻¹ year⁻¹, N50), with three replicates for each N input level. Urea was applied to the soil surface six times yearly from early May to early October using backpack sprayers. The amount of water added to the soil through this N application was equivalent to 0.0625 mm of rainfall, and the same amount of water was applied to the control plots. Comprehensive investigations of soil and plant nutrient status were only conducted in N0 and N50, and the results for N20 are therefore not presented in this study.

Plant and Soil Sampling, and Chemical Analysis

To examine leaf NRE, we sampled green and senesced needles from the five age classes in mid-August and early October, respectively. Appendix table S1 lists the sampling years for each stand age and exogenous N input treatment. Leaves were collected from upper and lower crown positions. Senescing needles were collected by hand while still attached to the tree, with the collection time determined by their color and whether they were ready to flush to the ground, according to Killingbeck and others (1990). We collected 100 needles from each sample tree in mid-August and early October to estimate the leaf mass lost between the green and senescent stages. Needle samples were placed in plastic bags, transported to the laboratory, and oven-dried at 60°C to a constant mass. Leaf litter mass loss was determined by dividing the mass of

100 needles sampled in October by the mass of 100 needles sampled in August.

We collected litterfall twice per year from 2010 onward, using two traps installed in each plot (for a total of 18 litter traps in each stand). The traps measured $1 \times 1 \text{ m}^2$ and were constructed of mesh and metal frames mounted on 0.4 and 0.8 m high polyvinyl chloride rods in the 11-year-old and 20- and 45-year-old stands. Samples from the two traps were composited to generate one sample per plot, dried to constant mass at 60° C, and weighed. Almost no woody or other fraction was found in the traps in our study. Thus the amount of litterfall was effectively equal to the foliage mass.

We sampled branches in the upper and lower crown positions simultaneously with the senesced leaves. To determine the stem N concentration, three trees were randomly selected in each plot, and the sapwood and heartwood of each were sampled at a height of 1.3 m using an increment borer. The three samples from each plot were combined and oven-dried to constant mass at 60°C.

To estimate root N concentration, four soil cores (0-40 cm) were obtained using a 10 cm-diameter metal auger at randomly selected positions in each plot at the end of the growing season. Soil cores were divided into three depths (0–10, 10–20, and 20–40 cm). Roots were separated by hand from each sample and oven-dried to constant mass at 60°C. Root N concentration was the average of the four samples.

All plant samples were ground in a Wiley mill (2 mm mesh). The C and N concentrations in plant samples were determined using an elemental analyzer (2400 II CHNS/O Elemental Analyzer, Perkin-Elmer, Waltham, MA, USA).

Soil samples were collected the month before N additions in the growing seasons of 2011 and 2013. Soil was randomly sampled from three points in each plot using a corer (internal diameter 4 cm) to a depth of 20 cm, stored in an icebox, and transported to the laboratory immediately after collection. The three samples from each plot were composited and roots were hand-sorted and soil passed through a 2-mm sieve.

We performed a KCl (0.5 M) on soil subsamples, and the extractant was analyzed for NH_4^+ -N and NO_3^- -N using the Type AA3 Continuous Flow Analytical System (BranLubbe, Germany).

Nitrogen Resorption Calculations

The NRE at leaf level was calculated as follows (Aerts 1996; Killingbeck 1996):

$$NRE = \left[\frac{N_{green} - N_{senesced}}{N_{green}}\right] \times 100\%$$
 (1)

where N_{green} and $N_{senesced}$ are the nitrogen concentrations (mass of N per unit dry mass) in mature green and senesced leaves, respectively.

Nitrogen resorption at the stand level was determined by the difference in the amount of N between green and senesced leaves in a plot. The green leaf mass in a plot was assessed by dividing the litterfall mass by the rate of leaf mass loss.

Annual Stand Biomass Production

Annual stand biomass production was used to represent the growth potential of trees in a plot (Chapin and others 1990). Annual stand biomass production in each plot was determined in the 11-, 20-, and 45-year-old stands from 2010 to 2013. To examine the increment in tree height and diameter, 15–20 trees were randomly selected in each plot. We installed metal bands at breast height (1.3 m) on each tree in the spring of 2010 to measure the diameter at breast height (DBH) of each tree. The change of the window length between two measurements was the yearly circumference growth, and was recorded at the beginning (early may) and end (late October) of the growing season. At the same time, tree height was measured using a hypsometer. The height and DBH of trees in each plot were calculated as the means of 15–20 trees.

We estimated the biomass of branches, stems, and roots using an allometric equation relating each

biomass component to the DBH and tree height, respectively. This allometric equation was established using forest inventory data from our study area (personal communication with Chao Yue), and is as follows:

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$$Ln(biomass) = a \times Ln(D^2H) + b$$
 (2)

where D and H are the mean DBH and tree height, respectively. Table 2 summarizes the values of a and b for the biomass of branches, stems, and roots in all study stands.

Measuring the Annual Biomass Production and the N Required for Tree Growth

Annual biomass production was defined as the sum of the annual increase in foliage, branch, stem, and root biomasses. The annual biomass production in a plot was calculated as:

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$$B = B_f + (B_b - B_b') + (B_s - B_s') + (B_r - B_r')$$
 (3)

where B (Mg ha⁻¹) is the annual biomass production in a plot, B_f (Mg ha⁻¹) is the foliage biomass in a plot in August, B_b ', B_s ', and B_r ' (Mg ha⁻¹) are the branch, stem, and root biomasses in a plot at the beginning of the local growing season, respectively, and B_b , B_s , and B_r (Mg ha⁻¹) are the branch, stem, and root biomasses in a plot at the end of the local growing season, respectively.

The annual N requirement was defined as the amount of N taken up by vegetation per square meter per year (g m⁻² yr⁻¹), which is difficult to determine directly (Chapin and others 1986). Therefore, we used total N content in annual accumulated biomass as a surrogate of the annual N requirement for tree growth (Gholz and others 1985). Thus, the annual N requirement for tree growth was defined as the annual N accumulation in leaves, branches, stems, and roots in a plot (Berendse and Aerts 1987; Tomaszewski and others 2003). The annual N requirement in a plot was calculated as:

$$Nr = \sum (B_i \times c_i) \tag{4}$$

where Nr (g m $^{-2}$ yr $^{-1}$) is the annual N requirement for the growth of leaves, branches, stems, and roots in a plot, and B $_i$ (Mg ha $^{-1}$) and c $_i$ (mg kg $^{-1}$) are the annual production in stand biomass and average N concentration of foliage, branches, stems, and roots in a plot, respectively.

Statistical Analysis

We performed a two-way ANOVA to evaluate the effects of stand age and exogenous N input on all the variables. Significant effects were determined at P < 0.05 unless otherwise stated. The relationships between variables were analyzed using exponential or linear models. Analyses were conducted using SPSS 16.0 software (SPSS Inc., Chicago, IL, USA). Data were expressed as mean values \pm S.E. (standard error).

RESULTS

Leaf N Concentration and NRE

Green leaf N concentration logarithmically increased with increasing stand age (Figure 1A). During stands' initial development (2–23 years old), green leaf N concentration increased sharply with increasing stand age from 1.53% to 2.51%. After canopy closure (20–100 years old), green leaf N concentration increased slowly, ranging from 2.16% to 2.87%.

In contrast to green leaf N concentration, senesced-leaf N concentration logarithmically decreased with increasing stand age (Figure 1B). A sharp decrease (from 1.65% to 0.60%) in senesced-leaf N concentration occurred from the initial development to canopy closure stages. After canopy closure, the senesced-leaf N concentration remained relatively low, ranging from 0.40% to 0.99%.

Like green leaf N concentration, NRE logarithmically increased with increasing stand age (Figure 1C).

In 2–23 years old trees, the NRE increased sharply with increasing stand age from 8% to 76%. After age

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The annual biomass production was significantly age-related (Figure 2A), and was largest in the 20-year-old stand, followed by the 45-year-old stand, and lowest in the 11-year-old stand. The biomass production of foliage, branches, stems, and roots displayed the same pattern among stand ages (Figure 2A). However, biomass allocation to these tissues varied among ages (Figure 2A), with 6.9%, 34.2%, 17.1%, and 41.9% in the 11-year-old stand, 12.9%, 48.3%, 13.5%, and 25.3% in the 20-year-old stand and 11.6%, 45.1%, 8.6%, and 34.6% in the 45-year-old stand to roots, stems, branches and leaves, respectively (Figure 2A). Exogenous N input significantly increased annual biomass production (P = 0.037), and did not alter the biomass allocation among tissues (Figure 2A). Different from green leaves, the N concentrations in roots, branches, and stems decreased with stand age (Table 3). The exogenous N input significantly increased N concentration in green leaves, but had no impact on the N concentrations of branches, stems and roots (Table 3). The annual N requirement, estimated by summing N content in newly accumulated biomass in different organs, displayed the same pattern as the annual increase in stand biomass, with the highest value in the 20-year-old stand, followed by the 45-year-old stand, and the lowest value in the 11-year-old stand (Figure 2B and Table 3). The majority of annual N supply was used for leaf growth, accounting for 71.6, 78.3 and 84.2% of the total annual N requirement in 11-, 20- and 45-year-old stands, respectively (Figure 2B). The ratios of annual N requirement for stems and branches to the total both declined with stand age: 9.5, 8.7 and 5.4% for branches and 15.1, 4.9 and 4.5% for stems in 11-, 20- and 45-year-old stands, respectively (Figure 2B).

However, the ratios of annual N requirement for roots to the total were higher in the 20- (8.2%) and

45-year-old (5.9%) stands than in the 11-year-old stands (3.8%) (Figure 2B). The exogenous N input did not alter the annual N requirement for leaves, branches and stems, but increased the annual N requirement for roots, although the increase was significant only in the 45-year-old stands (Table 3).

N Resorption at Leaf and Stand-level

Younger stands tended to have higher N concentrations in senesced leaves (P < 0.001, Figure 3A). The exogenous N input increased N concentration in senesced leaves (P < 0.001), but the degree of the increase differed among stands. Among the three age classes, the greatest increase in senesced leaf N concentration occurred in the 11- year-old stands (Figure 3A).

The NRE at leaf level ranged from 34.6 to 77.8%, which was increased significantly from the 11- to 45-year-old stands (P < 0.001, Figure 3B). The exogenous N input decreased NRE in 11- and 20-year-old stands, while not altered NRE in 45-year-old stand.

N resorption at stand-level also differed among the three stands (P < 0.001, Figure 3C). The highest N resorption was 8.3 ± 0.74 g m⁻² in the 20-year-old stand and the lowest was 1.8 ± 0.46 g m⁻² in the 11-year-old stand. However the exogenous N input did not altered the N resorption at stand level (Figure 3C).

The Effects of Plant-Available N Concentration and N Growth Requirement on N Resorption Plant-available N concentration was different among the three age classes (P = 0.007), with the value was higher in the 11-year-old stand than in the 20- and 45-year-old stands. The exogenous N input significantly increased the plant-available N concentration (P = 0.022, Figure 4A). Leaf NRE decreased with increasing plant-available N concentration, and exogenous N input did not alter this relationship (Figure 4B).

The contribution of N resorption to annual N supply, as indicated by the ratio of N resorption at stand-level to the annual N requirement for stand growth, significantly increased with increasing stand age (P < 0.001), with mean values of $45 \pm 4.7\%$ for the 11-year-old stand, $62 \pm 2.5\%$ for the 20-year-old stand, and $68 \pm 2.1\%$ for the 45-year-old stand (Figure 5A). The exogenous N input decreased the contribution of N resorption to annual N supply (P = 0.020), Figure 5A). The correlation between N requirement and N resorption at stand level was also analyzed with linearly regression. To avoid autocorrelation, the analysis was conducted using the total annual N requirement for branches, stems, and roots but not leaves. The results showed that N resorption at stand level increased with increasing annual N requirement regardless of with $(R^2 = 0.54)$, $(R^2 = 0.54)$, $(R^2 = 0.64)$, $(R^2 =$

DISCUSSION

We found that along the stand-age chronosequence from 2 to 100 years old, leaf NRE increased logarithmically from 8% to 82% (Figure 1C). Leaf N is an important component of proteins, which are abundant in chloroplasts, and minor amounts are found in cytosolic proteins, chlorophyll, and amino acids (Estiarte and Peñuelas 2015). During leaf senescence, proteins are hydrolyzed into amino acids, which subsequently retranslocated to woody tissues (Chapin and Kedrowski 1983). 62% of the N is removed from leaves by this process (Vergutz and others 2012). Killingbeck (1996) suggested that any N concentration in senesced leaves $< 7 \text{ mg g}^{-1}$ can be considered "complete resorption," and concentrations $> 10 \text{ mg g}^{-1}$ "incomplete resorption". In our study, the N concentration of senesced needles in young stands ranged from 0.74% to 1.65%. After canopy closure, N in senesced needles decreased to 0.40% to 0.99%

(Figure 1B), suggesting that N resorption shifted from incomplete to complete, and indicating that the N strategy of the larch plantation gradually improved with stand aging. This change in N resorption may be driven by changes in soil nutrient conditions and forest growth rates during stand development.

The high soil N availability could lead to low efficiency of N use and luxury N consumption of plant (Vitousek 1982; Yuan & Chen 2015). Numerous studies found that leaf NRE is generally lower in N-rich conditions than in N-poor conditions (Small 1972; Kobe and others 2005; Vergutz and others 2012; Yuan and Chen 2015). At our study sites, N availability in the soil decreased with stand age (Figure 4C). The decline in plant-available N could be due to increased N use for aboveground productivity with stand development and, simultaneously, soil N availability limited by the return of N from litterfall due to relatively low litterfall decomposition (Polglase and others 1992; Farley and Kelly 2004). In addition, N mineralization and nitrification are strongly controlled by litter decomposition, which in turn is controlled by litter N concentration (Melillo and others 1982; Manzoni and others 2008). At our study site, mature stands have a relatively low N concentration in senesced leaves, ranging from 0.40% to 1.0% after canopy closure (Figure 1B). The low N concentration in senesced leaves, coupled with the high C:N ratio of leaf detritus (Table 1), could slow the litter decomposition rate in mature forest, feeding back on N mineralization and therefore soil nutrient availability (Aerts 1997). The decrease in plant-available N during stand development therefore led to an increase in leaf NRE with stands aging (Figure 4B).

Temperate forests are facing increasing N deposition, which generally increases plant-available N in the soils. A recent meta-analysis based on a global dataset found that leaf NRE declined in response to N fertilization (Yuan and Chen 2015). However, it is largely unknown whether leaf N resorption in stands of different ages will respond differently to exogenous N input. Our experiments found that exogenous N input increased N concentration in both green leaves and senesced leaves, however the responses changed

with stand development (Table 3, Figure 3A). Compared to older stands, younger stands tend to have a lower increase in green leaf N concentration, but a greater increase in senesced leaf N concentration under exogenous N input (Table 3, Figure 3A). Exogenous N input was therefore significantly decreased leaf NRE in young stands, but such reduction diminished in mature stands (Figure 3C). Our study suggested that increase in plant-available N could be the main reason leading to the decline in leaf NRE under exogenous N input (Figure 4B). However, the degree of the responses was regulated by the physiological stage of the stands (Figure 3B). Increase in N deposition could decease the efficiency of young stands to recycle N within plants, whereas mature stands can be a stronger sink for N deposition, with better capacity to retain the deposited N within plants via internal cycle.

Nutrient resportion is a key strategy that plants conserve previously acquired nutrients (Zhang and others 2015). The N demands of living tissues constitute sinks for resorbed N, as observed in early studies (Chapin and Kedrowski 1983; Chapin and Moilanen 1991; Tully and others 2013). The potential of forests to grow living tissues varies with stand ages. We expected that stands with higher growth potential require more N to support biomass production, therefore should have a higher NRE. This hypothesis was supported by a study in radiate pine saplings in South Australia. The study found that periods of high N resorption coincided with periods of high shoot production, indicating that growth demand of N greatly determined nutrient resorption (Nambiar and Fife 1987, 1991).

In the current study, we found that the annual increase in stand biomass and its allocation in tissues shifted across different stand ages (Figure 2A), in line with many other studies on various tree species (Gower and others 1996; Ryan and others 2004; He and others 2012; Taylor and others 2014). Due to changes in the annual accumulated biomass and N concentrations in plant organs, the annual N requirement showed age-related variation, ranging from 3.8 ± 0.46 to 13.4 ± 1.19 g m⁻² yr⁻¹ (Figure 2B). Middle-aged

stands had the highest N requirements, consistent with the results of Bond-Lamberty and others (2006) for a boreal black spruce stand. We observed a strong positive relationship between the stand-level N resorption and the annual N requirement for stand growth (Figure 5B), suggesting that the shift in annual N requirement could the main factor responsible for the increase in stand-level N resorption along the age chronosequence.

A recent global analysis suggested that leaf N resportion contributes to 31% of annual N plant demand (Cleveland and others 2013). In our sites, N derived from senescing leaves can supply 45–68% of the N required for stand growth (Figure 3A). Both leaf NRE and the contribution of recycled N to annual N requirement increased with increasing stand age (Figure 3B and Figure 5A). Those evidences indicated that mature stands are more efficient in recycling N than are younger ones; mature stands are therefore have a greater capacity to supply growth required N via internal N cycling in plants. Our exogenous N input experiments further found that N input decreased the contribution of recycled N to the total annual N requirement by an average of 6.8%, and stand ages did not alter the responses (Figure 5A). With the continuing increase in N deposition, we expect that the N sources for stand growth would be shifted. Plants will depend more on root uptake of newly deposited N, but less on recycled N via resorption.

In the current study, we use a chronosequence approach to investigate how stand age and its interaction with N deposition affect N resorption at leaf level and stand level. Our studied stands along the age chronosequence have similar climate, topography and soil type, we therefore expected that the difference among the stands is predominately driven by the difference in stand ages. Still the findings should be interpreted alongside the limitations of the space-for-time approach (Johnson and Miyanishi 2008). Considering factors such as soil nutrient condition and stand successional trajectory could be changed due to continue receiving N deposition, the chronosequential response of a stand could be different

from our prediction using the space-for-time approach. Long-term time series studies are needed to improve our understanding on this topic.

CONCLUSIONS

We found a logarithmic shift in the green- and senesced-leaf N concentration and leaf NRE of larch along a stand-age chronosequence, indicating that NRE can change with stand development. In models, NRE is often set to a constant value of 50% (Aerts 1996; Van Heerwaarden and others 2003; Vergutz and others 2012). As a key process in biogeochemical models, neglecting age-related changes in N resorption can lead to bias when evaluating N-derived changes in forest net primary production in N cycle models, especially with increasing global N deposition. Thus, further experimental and modeling studies are needed to accurately quantify the age-related pattern of NRE in other tree species and to better address the implications for N-derived changes in forest growth.

Our data also suggested that the age-related pattern of stand-level N resorption was controlled by the changes in soil N availability and annual N requirement for forest growth during stand development. In addition, the ratios of stand-level N resorption to the annual N requirement for stand growth increased with stand age. These results not only imply that the growth potential of trees can influence their NRE with stand development, but also show that mature stands are probably stronger sinks for N from atmospheric deposition. Ignoring these results could lead to a bias in N-cycling models when evaluating N-derived changes in forest net primary production under increasing global N deposition.

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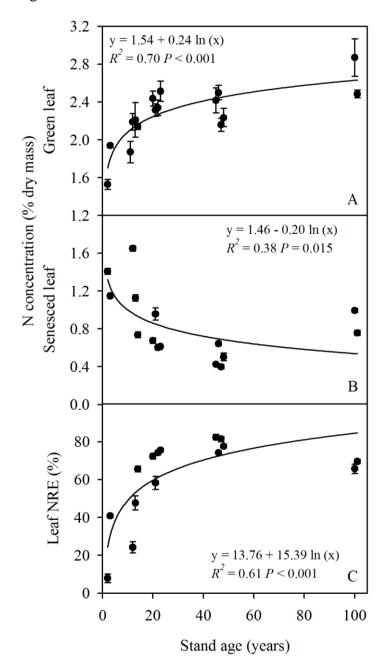
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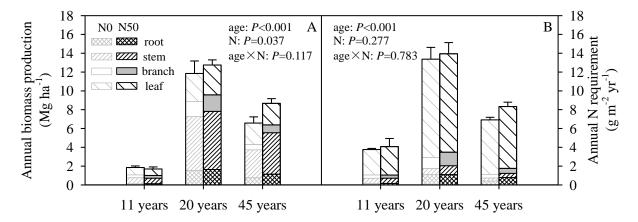
502	Figure legends
503	
504	Figure 1 Variations in green (A) and senesced (B) leaf N concentrations and leaf NRE (C) along a
505	stand-age chronosequence in a Larix plantation in north China. The regression equations represent green-
506	and senesced-leaf N concentrations and leaf NRE $vs.$ stand age. The vertical bars represent the mean \pm 1
507	SE.
508	
509	Figure 2 Annual stand biomass productions (A), N requirement for stand growth (B) in different stand-age
510	classes. Annual stand biomass production and N requirement for stand growth include the contributions of
511	leaves, branches, stems, and roots in each plot. The error bars represent the standard deviations of the
512	annual stand biomass increase or the N requirement for stand growth.
513	
514	Figure 3 The effects of stand age and exogenous N input on senesced-leaf N concentrations (A), leaf NRE
515	(B) and the amount of stand-level N resorption (C). Vertical bars represent the mean \pm 1 SE.
516	
517	Figure 4 The effects of stand age and exogenous N input on the plant-available N concentration (A) and
518	the relationship between plant-available N concentration (0-20-cm soil depth) and leaf NRE under two N
519	input levels (B). Data in figure B are the mean values from the three stands in 2011 and 2013. The circles
520	triangle and squares symbols represent the stands aged 11-, 20- and 45-year-old, respectively. Black and
521	gray lines are regressions of the plant-available N and NRE under N0 and N50 treatments, respectively.
522	

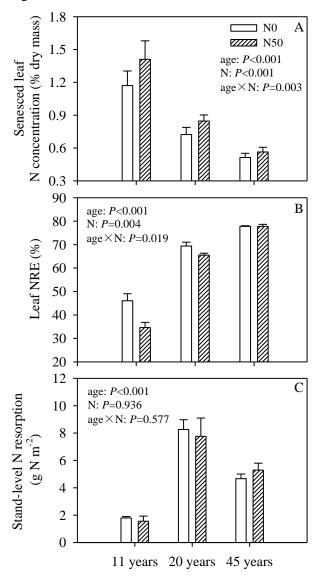
Figure 5 The effects of stand age and exogenous N input on the ratio of stand-level N resorption to annual

N requirement for stand growth (A), and the relationship between the total annual N requirement for branches, stems, and roots and the amount of stand-level N resorption for stands aged 11 (circles), 20 (triangles), and 45 (squares) years (B). The data are mean values. Black and gray lines are regressions of the amount of stand-level N resorption and the annual N requirement for tree growth under N0 and N50 treatments, respectively.

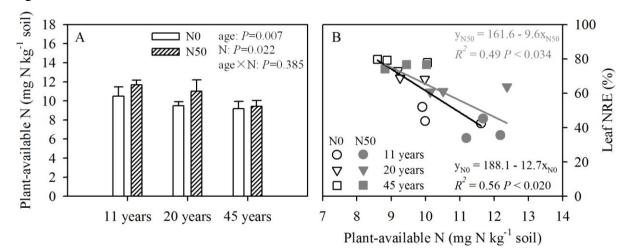


535 Figure 2





542 Figure 4



544 Figure 5

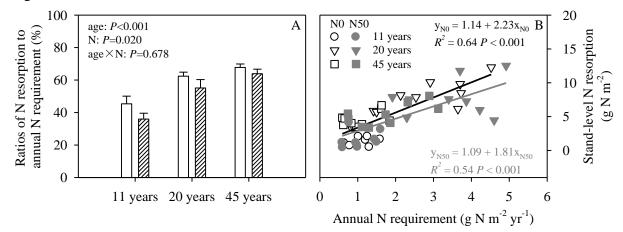


Table 1 Forest structure and litter (organic horizon) and soil (0–10-cm depth) properties of the three stand ages of *Larix principis-rupprechtii* plantations in 2013.

Stand age	Location	Community		Litter and soil physiochemical properties (0-10 cm)			
(years)	(latitude, longitude)	Density (Trees ha ⁻¹)	DBH (cm)	Height (m)	Litter C:N	Soil C:N	Soil pH (soil:water = 1:2.5)
11	42°23.3′N, 117°14.0′E	2640(157) ^b	4.1(0.1) ^c	3.6(0.1) ^c	65.6(2.7) ^c	10.3(0.2) ^b	6.2(0.2) ^b
20	42°23.6′N, 117°14.1′E	3060(132) ^a	$10.5(0.1)^{b}$	$8.6(0.1)^{b}$	79.8(3.1) ^b	11.7(0.3) ^a	$6.5(0.0)^a$
45	42°23.9′N, 117°14.8′E	870(48) ^c	20.7(0.2) ^a	16.5(0.2) ^a	102.2(8.0) ^a	13.2(0.9) ^a	$6.3(0.2)^{ab}$

Values in parentheses are SDs of the mean (n = 3). Different superscript letters within the same column indicate significant differences between stands (one-way

ANOVA, post hoc LSD test, P < 0.05). DBH = diameter at breast height.

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Items	a	b	R^2	P
11-year-old stand				
Stems	0.5360	4.7647	0.89	< 0.01
Branches	0.3481	5.2004	0.52	< 0.01
Roots	0.3896	4.0268	0.62	< 0.01
20- and 45-year-old stands				
Stems	0.8882	3.6574	0.99	< 0.01
Branches	0.7051	3.8495	0.90	< 0.01
Roots	0.8725	2.4581	0.95	< 0.01

The models were developed from Larix principis-rupprechtii plantations in Saihanba National Forest.

The equation is: $Ln(biomass) = a \times Ln(D^2H) + b$, where D is the mean DBH, and H is the mean tree height.

Table 3 N concentrations and annual N requirement for newly accumulated biomass in different organs.

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Stand age	N treatment		N	concentration	(%)	A	annual N requir	ement (g m ⁻² yı	r ⁻¹)
(years)		Green leaves	Branches	Stems	Roots	Leaves	Branches	Stems	Roots
11	N0	2.18(0.10)	1.14(0.01)	0.90(0.03)	1.14(0.05)	2.70(0.11)	0.36(0.01)	0.57(0.04)	0.14(0.01)
	N50	2.15(0.05)	1.12(0.04)	0.93(0.02)	1.34(0.08)	3.02(0.88)	0.33(0.07)	0.55(0.13)	0.16(0.04)
20	N0	2.39(0.03)	0.76(0.06)	0.11(0.01)	0.72(0.09)	10.47(0.98)	1.16(0.12)	0.65(0.07)	1.09(0.14)
	N50	2.48(0.02)	0.84(0.05)	0.16(0.02)	0.67(0.03)	10.47(1.31)	1.43(0.14)	0.96(0.10)	1.09(0.04)
45	N0	2.30(0.06)	0.62(0.02)	0.11(0.02)	0.53(0.02)	5.82(0.32)	0.37(0.02)	0.31(0.07)	0.41(0.02)
	N50	2.54(0.08)	0.60(0.05)	0.10(0.01)	0.69(0.05)	6.57(0.48)	0.53(0.07)	0.45(0.07)	0.78(0.08)
Source (P-values)									
	age	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001
	N	0.014	0.669	0.272	0.058	0.590	0.081	0.065	0.043
	$age \times N$	0.024	0.467	0.585	0.112	0.892	0.265	0.22	0.036

Data are the means of 3 years from 2011 to 2013. P-values were results of the LSD test which followed the Two-way ANOVA on the effects of stand age and exogenous N input on the concentrations of branches, stems and roots and N requirement of foliage, branches, stems and roots.

Table S1 The arrangement of exogenous N input treatments and leaf sampling for each stand of the five age classes.

Stand age N input treatment		treatment	Sampling year			
(years)	N0	N50	2010	2011	2012	2013
2	\checkmark			$\sqrt{}$	$\sqrt{}$	
11	\checkmark	$\sqrt{}$		$\sqrt{}$	\checkmark	$\sqrt{}$
20	\checkmark	$\sqrt{}$	$\sqrt{}$	$\sqrt{}$	$\sqrt{}$	$\sqrt{}$
45	\checkmark	\checkmark	\checkmark	$\sqrt{}$	$\sqrt{}$	$\sqrt{}$
100	$\sqrt{}$			\checkmark	\checkmark	

[&]quot; $\sqrt{}$ " indicates that the exogenous N input treatment was carried out in this stand, or that green and senesced

leaves were sampled in this year.

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