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AGROSTOLOGY

An Introduction to the Systematics of Grasses

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SECTION 1 - INTRODUCTION

1.01 - GRASSES: AN OVERVIEW

Nut-grass

Yellow-eyed-grass

Yellow-eyed-grass

"Of all things that live and grow upon this earth, grass is the most important." (Donald Culross Peattie. A Prairie Grove. 1938.)

The first word on the title page of this syllabus is an uncommon one. **Agrostology** [Greek, a kind of grass + body of knowledge] is the branch of systematic botany that deals with grasses, especially their identification, classification, and evolution. **Agriculture**, on the other hand, is the applied science that deals with cultivating land, and the raising and breeding of crops and livestock. **Agronomy** is the science of soil management and of crop production. Both terms are derived from the Greek root for fields, soils, and crops.

WHAT ARE GRASSES?

All true grasses belong to a single family of flowering plants, Gramineae or Poaceae. I use the phrase "true grasses" because there are many plants that have "grass" as part of their common name that are not, in fact, grasses. Many, but not all, have grass-like leaves that fool the uninitiated.

"GRASSES" THAT ARE NOT GRASSES

"GRASSES" THAT AR	E NOT GRASSES
Alkali-grass	Zigadenus elegans
Arrow-grass	Triglochin maritima
Bayonet-grass	Scirpus maritimus
Bear-grass	Xerophyllum tenax
Bear-grass	Nolina microcarpa
Bear-grass	Yucca filamentosa
Beavertail-grass	Calochortus coeruleus
Blue-eyed-grass	Sisyrinchium spp.
Cotton-grass	Eriophorum spp.
Eel-grass	Zostera marina
Fish-grass	Cabomba caroliniana
Gallow-grass	Cannabis sativa
Golden-eyed-grass	Sisyrinchium spp.
Goose-grass	Potentilla anserina
Grass	Cannabis sativa
Grass-of-Parnassus	Parnassia spp.
Grass-tree	Xanthorrhea spp.
Grass-wrack	Zostera marina
Indian basket-grass	Xerophyllum tenax
Iron-grass	Carex caryophyllea
Mat-grass	<i>Phyla nodiflora</i>
Merlin-grass	<i>Isoëtes</i> spp.

Milk-grass

Mondo-grass

Orange-grass Palm-grass Penny-grass Pepper-grass	Hypericum gentianoides Curculigo spp. Thlaspi spp. Lepidium spp.
Pigeon-grass	Verbena officinalis
Pineapple-grass	Astelia spp.
Pudding-grass	Mentha pulegium
Pudding-grass	Hedeoma pulegoides
Purple-eyed-grass	Sisyrinchium spp.
Ripple-grass	Plantago lanceolata
Saw-grass	Cladium jamaicense
Scorpion-grass	Myosotis arvensis
Sedge-grass	Carex pendula
Serpent-grass	Polygonum viviparum
Snake-grass	Equisetum arvense
Star-grass	Aletris farinosa
Surf-grass	Phyllospadix spp.
Tape-grass	Vallisneria spp.
Viper's-grass	Scorzonera hispanica
Whitlow-grass	Draba verna
Widgeon-grass	Ruppia maritima
Wire-grass	Juncus spp.

Cyperus esculentus

Sisyrinchium spp.

Xyris spp.

HOW BIG IS THE FAMILY?

Grasses, although they do not constitute the largest family of flowering plants, are economically the most important to us and ecologically they are the most dominant form of higher plants. Estimates of the size of the family vary, but ranges of 600-700 genera and about 10,000 species seem reasonable. The family ranks third in number of genera (behind the orchids and sunflowers) and fifth in number of species (after orchids, sunflowers, legumes, and members of the madder family).

NUMBER OF GENERA & SPECIES

Source	# Genera	# Species	
Linnaeus (1771) Trinius (1822) Hackel (1887) Bews (1929) Pilger (1954)	43 313 483 700	285 2457 3500 5871 8,000	
Prat (1960) Dahlgren (1985)	403 750	6250 10,000	

Valerianella locusta

Ophiopogon japonicum

Clayton & Renvoize (1986)	651	10,000
Tzvelev (1989)	898	10,300
Watson & Dallwitz (1992)	793	9,890
Takhtajan (1997)	850	11,000
Thorne (1999)	656	9,976

A GLOBAL SUMMARY BY SUBFAMILY

Subfamily	Genera	Species
Anomalochlooideae Pharoideae Bambusoideae Ehrhartoideae Poöideae Arundinoideae Danthonioideae Aristidoideae Centothecoideae Panicoideae Totals	2 (0.3%) 3 (0.5%) 65 (9.9%) 18 (2.7%) 154 (23.4%) 49 (7.4%) 19 (2.9%) 1 (0.1%) 10 (1.5%) 207 (31.5%) 658 (100%)	4 (0.1%) 12 (0.1%) 965 (9.7%) 150 (1.5%) 3275 (32.8%) 605 (6.1%) 275 (2.8%) 250 (2.5%) 25 (0.2%) 3290 (33.0%) 9976 (100%)

[Number of taxa from Thorne, 1999]

THE TWENTY LARGEST GRASS GENERA

North America	World-wide
Panicum (113)	Panicum (590)
Poa (96)	Poa (500)
Elymus (80)	Festuca (472)
Muhlenbergia (77)	Eragrostis (350)
Festuca (63)	Paspalum (330)
Eragrostis (60)	Stipa (300)
Bromus (58)	Aristida (290)
Paspalum (54)	Calamagrostis (230)
Aristida (51)	Digitaria (230)
Agrostis (48)	Agrostis (220)
Calamagrostis (47)	Elymus (221)
Stipa (43)	Muhlenbergia (160)
Sporobolus (35)	Sporobolus (160)
Digitaria (32)	Bromus (150)
Setaria (28)	Bambusa (150)
Bouteloua (25)	Axonopus (110)
Melica (24)	Setaria (110)
Andropogon (23)	Andropogon (100)
Glyceria (21)	Brachiaria (100)
Hordeum (20)	Isachne (100)

DISTRIBUTION

Grasses are the most cosmopolitan of all higher plants, occurring on all continents, including Antarctica. They are also the most frequently encountered vascular plants. There may well be more individual grass plants than there are all other vascular plants combined! They are found from the polar regions to the equator, from mountain tops to seashores. They occur in brackish and freshwater marshes, ponds, streams, rain forests, deserts, tundra, and arid slopes. About one-fourth of the

earth's plant cover is grasslands. They are dominant in the vast expanses of the world's prairies, steppes, pampas, paramos, and veldt. A major part of our agricultural lands is devoted to them. Grasses are with us in our cities, either as ornamentals or as weeds along sidewalks and in vacant lots. Grasses are never far away.

ECONOMIC IMPORTANCE

No other plant family, with the possible exception of the legumes and palms, can approach the grasses in direct economic importance to us. Major products include the cereal grains (wheat, rice, maize, barley, rye, sorghum, oats, and millets), hay, pasture, turf grasses, thatching material, timber, paper pulp, sugar (from sugar cane and sorghum), aromatic compounds (e. g., lemon grass and oil of vetiver), brooms, fishing poles, musical instruments, ornamentals, soil binders, starches, edible oils, alcohol, beverages, and food for most of the world's wild and domesticated animals.

We derive a major portion of our calories from cereals. Much of our agricultural land is devoted to the raising of cereals, especially wheat. Still more of the earth's surface is used for pastures for a variety of domesticated animals.

SELECTED REFERENCES

Arber, A. 1924. The Gramineae: a study of cereal, bamboo, and grass. Cambridge Univ. Press. New York, NY. 480 pp.

Barnard, C. (editor). 1964. Grasses & grasslands. Macmillan. London, England. 269 pp.

Bentham, G. 1881. Notes on Gramineae. J. Linn. Soc. 19: 14-134.

Bews, J. G. 1929. The world's grasses: their differentiation, distribution, economics, and ecology. Longmans, Green & Co. London, England. 408 pp.

Booth, W. E. 1964. Agrostology. Edwards Brothers. Ann Arbor, MI. 222 pp.

Cade, J. 1976. How to identify grasses. J. Agric. (Victoria) 74(11): 382-387.

Chapman, G. P. 1990. Reproductive versatility in the grasses. Cambridge Univ. Press. Cambridge, England. 296 pp.

Chapman, G. P. 1996. The biology of grasses. CAB International. Wallingford, England. 273 pp.

Chapman, G. P. (editor). 1992. Grass evolution and domestication. Cambridge Univ. Press. New York, NY. 352 pp.

Chapman, G. P. & W. E. Peat. 1992. An introduction to the grasses (including bamboos and cereals). CAB International. Wallingform, U. K. 111 pp.

Chase, A. & C. Niles. 1962. Index to grass species. Three volumes. G. K. Hall. Boston, MA.

Cheplick, G. P. (editor). 1998. Population biology of

- grasses. Cambridge Univ. Press. Cambridge, England. 399 pp.
- Clark, L. G. & R. W. Pohl. 1996. Agnes Chase's first book of grasses. Fourth edition. Smithsonian Inst. Press. Washington, D. C. 127 pp.
- Clayton, W. D. 1981. Evolution and distribution of grasses. Ann. Missouri Bot. Gard. 68: 5-14.
- Clayton, W. D. & S. A. Renvoize. 1986. Genera graminum: grasses of the world. Kew Bull. Addt. Series XIII. Royal Botanic Garden, Kew. London, England. 389 pp.
- Clifford, H. T. & L. Watson. 1977. Identifying grasses: data, methods, and illustrations. Univ. Queensland Press. St. Lucia, Australia. 146 pp.
- DeWet, J. M. J. 1981. Grasses and the culture history of man. Ann. Missouri Bot. Gard. 68: 87-104.
- Estes, J. R., R. J. Tyrl, & J. N. Brunken. 1982. Grasses and grasslands: systematics and ecology. Univ. Oklahoma Press. Norman. 312 pp.
- Gould, F. W. & R. B. Shaw. 1983. Grass systematics. Second edition. McGraw-Hill Book Co. New York, NY. 397pp.
- Hackel, E. 1887. Gramineae. <u>In</u>, Engler, A. & K. Prantl. Die natürlichen Pflanzenfamilien. W. Engelmann. Lepipiz, Germany. T. II, Abt. 2: 1-97.
- Hackel, E. 1890. The true grasses. Translated by F. Lamson-Scribner and E. A. Southworth from "Die natürlichen Pflanzenfamilien." Henry Holt & Co. New York, NY. 228 pp.
- Hahlenberg, M. H. & M. Schwartz. 1983. A book about grass: its beauty and uses. E. P. Dutton. New York, NY.
- Harrington, H. D. 1977. How to identify grasses and grasslike plants. Swallow Press. Chicago, IL. 142 pp.
- Hilu, K. W. 1985. Trends of variation and systematics of Poaceae. Taxon 34(1): 102-114.
- Hitchcock, A. S. 1914. A textbook of grasses of the United States, with special reference to the economic species of the United States. Macmillan & Co. New York, NY. 276 pp.
- Hitchcock, A. S. 1935. Grasses: what they are and where they live. Annual Report Board Regents Smithsonian Inst... 1934. Washington, D. C. Pp. 297-312 + 8 plts.
- Hitchcock, A. S. & A. Chase. 1931. Grass: old and new plant lore. Smithsonian Sci. Ser. 11: 201-250.
- Hubbard, C. E. 1984. Grasses: a guide to their structure, identification, uses, and distribution in the British Isles. Third edition. Revised by J. C. E. Hubbard. Penguin Books. Harmondsworth, England. 476 pp.
- Jacobs, S. W. L. & J. Everett (editors). 2000. Grasses: systematics and evolution. CSIRO. Collingwood, Australia. 406 pp.
- Judd, B. I. 1979. Handbook of tropical forage grasses.

- Garland Press. New York, NY. 116 pp.
- Kahlenberg, M. H. & M. Schwartz. 1983. A book about grass: its beauty and uses. E. P. Dutton. New York, NY. 112 pp.
- Knobel, E. 1980. Field guide to the grasses, sedges, and rushes of the United States. Second edition. Revised by M. E. Faust. Dover Publ. New York, NY. 83 pp.
- Pohl, R. W. 1978. How to know the grasses. Third edition. W. C. Brown. Dubuque, IA. 200 pp.
- Pohl, R. W. (editor). 1981. Evolution and systematics of the Gramineae. The 26th Systematics Symposium. Ann. Missouri Bot. Gard. 68: 1-104.
- Pohl, R. W. 1987. Man and the grasses: a history. $\underline{\text{In}}$, Soderstrom, T. R. et al. (editors). Pp. 355-358.
- Renvoize, S. A. & W. D. Clayton. 1992. Classification and evolution of the grasses. <u>In</u>, Chapman, G. P. Pp. 3-37.
- Roshevits, R. Y. 1937. Grasses. An introduction to the study of fodder and cereal grasses. Translated from the Russian. Publ. for the Smithsonian Inst. and the Natl. Science Found. by the Indian Natl. Sci. Doc. Centre. New Delhi. 635 pp.
- Skerman, P. J. & F. Riveros. 1990. Tropical grasses. Food and Agriculture Organization of the United Nations. Rome, Italy. 832 pp.
- Soderstrom, T. R. et al. (editors). 1987. Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. 472 pp.
- Stebbins, G. L. 1956. Taxonomy and the evolution of genera, with special references to the family Gramineae. Evol. 10: 235-245.
- Stebbins, G. L. 1956. Cytogenetics and evolution of grasses. Amer. J. Bot. 43: 890-906.
- Stebbins, G. L. 1972. The evolution of the grass family. $\underline{\text{In}}$, Younger & McKell. Pp. 1-17.
- Stebbins, G. L. 1981. Coevolution of grasses and herbivores. Ann. Missouri Bot. Gard. 68: 75-86.
- Stebbins, G. L. 1987. Grass systematics and evolution: past, present and future. <u>In</u>, Soderstrom et al. Pp. 359-367.
- Stefferud, A. (editor). 1948. Grass. The yearbook of agriculture 1948. U. S. Gov. Printing Office. Washington, D. C. 892 pp.
- Veldkamp, J. F. Grass literature. [A continuing series of computer disks with ASCII files that are available for purchase or in exchange for herbarium specimens]
- Watson, L. & M. J. Dallwitz. 1992. The grass genera of the world. CAB International. Wallingford, U. K. 1024 pp.
- Younger, V. B. & C. M. McKell (editors). 1972. The biology and utilization of grasses. Academic Press. New York, NY. 426 pp.

1.02 - VEGETATIVE STRUCTURE

"I believe a leaf of grass is no less than the journeywork of the stars." (Walt Whitman)

ROOTS

Most mature grasses have a root system that is **fibrous** -- finely divided and lacking a dominant one, as in a taproot system. It is also **adventitious**, in that the primary root system is short-lived and it is soon replaced by roots that are derived from some other node along the embryo axis, rather than developing from the branches of the primary root. The extent and penetration of the grass root system is variable. *Aristida pungens* of North Africa has roots more than 20 m long.

The cells that give rise to root hairs may be equal in length or alternately long and short. There is also a difference in the point of attachment and the angle of insertion of root hairs relative to the basal part of the cell.

The roots are of little direct economic importance to us, although they do retard erosion through the structuring of the soil. Some roots contain aromatic principles, as in oil of vetiver.

STEMS

The erect aerial stem of a grass plant is called a **culm**. The stem is generally soft and herbaceous in our temperate grasses. Some of the reeds and canes have much tougher culms. Typical bamboos appear to be quite woody, but plant anatomists will argue that what we are seeing is not actually woody tissue. A grass stem may be only a centimeter or so tall to as much as 40 m or so in some of the tropical bamboos. The stem is divided into **nodes** and **internodes**. The nodes are regions where leaves are attached; they are typically rather easy to located in grasses because they are swollen. The region between two successive nodes is the internode. It is typically hollow, but we do have several exceptions in some commonly encountered grasses. Some studies suggest that about half of the grasses may have solid internodes. These plants also have a specialized spikelet structure and tend to inhabit arid regions.

Branches of stems arise most commonly from buds at the base of a parent shoot. These basal branches are called **innovations**. In some crop plants, the innovations are called **tillers** or **suckers**. A secondary stem may elongate within a leaf sheath or it may break through it as it develops. We call the former situation **intravaginal branching** and the latter **extravaginal branching**. Intravaginal is the more common situation.

Grasses also commonly produce horizontal or **repent** stems. The two most frequently encountered types are the **rhizome** and the **stolon**. A rhizome is a horizontal stem at or below the surface of the ground. It bears reduced, scale-like leaves. Stolons, on the other hand, are horizontal stems running along the surface of the ground and they often bear ordinary foliage leaves.

Both serve as a means of vegetative reproduction. While these definitions sound precise, the distinction between the two is sometimes subtle. Bermuda grass (*Cynodon dactylon*) produces both rhizomes and stolons, depending upon environmental conditions.

Some grasses produce small onion-like **bulbs** (but without the odor), while others have **corms**, swollen hard stems surrounded by dry, papery, scale-like leaves, similar to the "bulb" of the gladiola.

The presence of rhizomes, stolons, bulbs, and corms is of taxonomic significance. All grass keys will, sooner or later, ask you whether these structures are present or absent. Make certain that you collect any underground parts of a grass plant when you are doing any serious collecting of grasses for identification or documentation.

Grass stems are very important as sources of building material, hay, forage, and packing material.

LEAVES

Grass leaves are **alternate** and **two-ranked**. They are alternate because only one leaf arises from a node. They are two-ranked because if the leaf borne at the first node comes off the left side of the culm, then the leaf at the second node will arise from the right side. Looking down on the stem and leaf system, the points of insertion or attachment are 180° opposite one another.

The leaves are typically composed of a **blade** or **lamina**, a **sheath**, and a **ligule**. The blade is usually linear -- most grass leaves, after all, do look like grass leaves! -- but it may be thread-like, needle-like, oval, or even arrowhead-shaped. In some of the tropical grasses, the leaf blades closely resemble those of some dicots. The venation is typically **parallel**, with all of the veins being more or less the same size or with one of them forming a more prominent midrib. Some tropical grasses have pinnate venation.

Some bamboos appear to have petioles, but I suspect they are best considered **pseudopetioles**. They appear to be nothing more than constrictions of the blade or sheath.

The grass sheath is usually interpreted as a flattened petiole. It is most often rounded, but in some grasses the sheath may be conspicuously flattened. Typically it is **open** -- the edges of the sheath come together and touch one another or they overlap slightly; but, they are not fused into a cylinder about the nodes. This is a useful character for separating most grasses from most sedges. But beware! Some very common grasses (orchard grass, onion grasses, and bromes) have **closed** sheaths, in which the two edges are joined. Wind action and careless use of a dissecting needle can convert closed sheaths to open ones.

The **ligule** is a membranous flap of tissue or a series of hairs (or both) at the junction of the blade and sheath. Its function may be to prevent water from entering the sheath or to hold the leaf tightly to the

culm. Not all grasses have liqules.

The first leaf of a culm branch or lateral shoot is the **prophyllum**. It is an unusual leaf, in that it lacks a blade. It protects the immature lateral stem axis and it provides mechanical support. The prophyllum has two prominent strands of vascular tissue running its length.

Grass leaves, especially those of the wheat or barley tribe (Triticeae) have ear- or claw-shaped appendages called **auricles**. These paired structures arise at the base of the blade in some grasses, but laterally at the sheath apex in others.

The leaf epidermis is an important source of taxonomic information. Typically the upper (adaxial) surface is different from the lower (abaxial) one. Both have cells arranged in columns over the vascular bundles (costal region) or in the zone between adjacent vascular bundles (intercostal zone). The cells are distinguished as long- or short-cells, depending upon their length-width ratio. Long-cells vary in wall thickness and appearance, being sinuous, papillate, or pitted. Those with sinuous walls are called ripple-wall cells.

Short cells occur singly or in pairs. There are two common types, **silica cells** and **cork cells**. The former have a silica-body in their lumen. The shape of this deposit determines the type of silica cell -- linear, rounded, irregular, saddle-shaped, dumbbell-shaped, cross-shaped, or double-axhead-shaped. Cork cells have cork in them.

Stomates are arranged in precise columns in the intercostal zone. Each is composed of two guard cells and two subsidiary cells.

An examination of the epidermis may also reveal **bulliform cells** -- large, colorless cells that are typically present in the intercostal zone of the adaxial surface. They are sometimes called mechanical cells, because they function in the rolling and unrolling (or

folding and unfolding) of the leaf blade.

The blade, when seen in cross-section (also referred to as a transverse-section) yields the following features:

mesophyll: thin-walled parenchyma and chlorenchyma cells

vascular bundles: composed to xylem and phloem tissue, surrounded by one or two bundle sheaths; the inner one (when present) is termed an **endodermis** or **mesostome sheath** by various authors

sclerenchyma fibers: typically present in clusters in the region between the epidermis and the outer bundle sheath

TEXTURES

cartilaginous - resembling cartilage; hard
 and tough, but flexible
chartaceous - with the texture of writing
 paper
coriaceous - with the texture of leather
crustaceous - with a brittle texture
hyaline - thin and translucent or transparent
indurate - hard or hardened
membranous - thin, soft, and flexible
pellucid - transparent, clear
scarious - thin, dry, membranous; not green

COLORS

cinereous - light gray; ash-colored ferruginous - rust-colored fuscous - brownish, dusky rufous - reddish-brown stramineous - pale yellow; straw-colored tawny - pale brown to dirty yellow

FEATURES OF THE SURFACES OF GRASSES*

Surface itself (exclusive of hairs, barbs, etc.)

glabrous - without hairs glaucous - with a whitish, waxy bloom **lustrous -** shining **papillae** - warty outgrowths of epidermal cells. pitted - with small depressions, pits, pin-holes, or cavities pruinose - with a waxy, powdery secretion on the surface
 punctate - dotted with pin-point impressions or translucent dots
 pustulose - with irregularly raised pimples reticulate - netted with regular, slightly elevated lines rugose - wrinkled **scurfy -** covered with minute scales **smooth** - not rough to the touch; not synonymous with glabrous **striated** - marked with longitudinal lines sulcate - furrowed with longitudinal channels tessellate - marked by square to oblong depressions tuberculate - with small projections; warty verrucose - another way of spelling tuberculate viscid - sticky

- Projections and depressions from surfaces, margins, and apices
 - Hairs branched or forked
 stellate with few- to several branched sessile or stalked hairs

malpighiaceous (dolabriform) - with forked hairs attached at middle

- Hairs simple, unbranched
 - Hooked or barbed antrorsely - with forward or upward directed barbs retrorsely - with backward or downward directed barbs uncinate - hooked, as in a fish hook
 - Not hooked nor barbed
 - Restricted to apex, base or margins
 ciliate with hairs along margins only
 fimbriate as in ciliate, but hairs coarser and longer
 comose with a tuft of hairs at apex or base
 - On surfaces
 - Curled, interwoven or entangled arachnoid - with slender, white, loosely tangled hairs floccose - with tufts of soft hairs that rub off easily lanate - with woolly or cottony hairs tomentose - with densely and softly matted hairs
 - Not curled. interwoven, nor entangled **bristle** a stiff slender hair-like appendage canescent - with a dense mat of grayish-white hairs echinate - with straight, ± large, prickle-like hairs **glabrate** - initially hairy, but becoming glabrous **glandular -** with swollen-tipped hairs; gland-bearing **hirsute -** with rough or coarse, ± erect hairs hoary - see canescent **hirsute** - with straight, ± stiff hairs hirtellous - minutely hirsute **hispid** - with long, rigid, bristly hairs **hispidulous** - minutely hispid microhairs - typically bicellular [rarely multicellular] hairs usually requiring magnification of a compound microscope macrohairs: typically one-celled hairs visible within the range of the ordinary dissecting microscope or good handlens; papillate - with pimple-like hairs papillose - see papillate **pilose -** with sparse, slender, soft hairs puberulent - minutely canescent pubescent - with short, soft, erect hairs; downy scabrous - with coarse, stiff, ascending hairs; rough sericeous - with long, fine, appressed hairs; silky setaceous - with bristly hairs **setose** - see setaceous strigose - with sharp, appressed, rigid, hairs that are often swollen at base velutinous - with dense, firm, straight hairs; velvety villous - with long, slender, soft (not matted) hairs; shaggy

*[Modified from Smith, J. P. 1977. Vascular plant families]

SELECTED REFERENCES

Amarasingehe, V. & L. Watson. 1988. Comparative ultrastructure of microhairs in grasses. Bot. J. Linn. Soc. 98: 303-319.

Arber, A. 1923. Leaves of the Gramineae. Bot. Gaz. 76: 374-388.

Brown, W. J. & C. Johnson. 1962. The fine structure of the grass guard cell. American J. Bot. 49: 110-115.

Burns, W. 1946. Corm and bulb formation in plants with special reference to the Gramineae. Trans. Proc. Bot. Soc. Edinburgh 34: 316-347.

Carolin, R. C. 1973. Structure of cells of mesophyll and parenchymatous bundle sheath of Gramineae. J. Linn. Soc. Bot. 66(4): 259-275.

Chase, A. 1923. Leaves of the Gramineae. Bot. Gaz. 119: 170-178.

Clark, L. G. & J. B. Fisher. 1987. Vegetative morphology of grasses: shoots and roots. $\underline{\text{In}}$, Soderstrom, T. R. et al. (editors). Pp. 37-45.

Collins, G. N. 1924. The prophyllum of grasses. Bot. Gaz. 78: 353, 354.

Ebinger, J. E. & J. L. Carlen. 1975. Culm morphology and grass systematics. Trans. Illinois Acad. 68: 87-

101.

Elias, R. P. 1976. A procedure for standardizing comparative leaf anatomy in the Poaceae. I. The leaf blade as viewed in transverse section. Bothalia 12: 65-109.

Elias, R. P. 1979. A procedure for standardizing comparative leaf anatomy in the Poaceae. II. The epidermis as seen in surface view. Bothalia 12: 641-671.

Evans, M. W. & J. E. Ely. 1935. The rhizomes of certain species of grass. J. American Soc. Agron. 27: 791-797.

Hilu, K. W. & J. L. Randall. 1984. Convenient method for studying grass leaf epidermis. Taxon 33(3): 413-415.

Johnson, Sister C. & W. V. Brown. 1973. Grass leaf ultrastructural variations. American J. Bot. 60: 727-735.

Langer, R. H. M. 1963. Tillering in herbage grasses. Herb. Abstr. 33: 141-148.

Liphschitz, N. & Y. Waisel. 1974. Existence of salt glands in various genera of the Gramineae. New Phytol. 73: 507-513.

Metcalfe, C. R. 1960. Anatomy of the monocotyledons. Vol. 1. Gramineae. Clarendon Press. Oxford, England. 731 pp.

Philipson, W. R. 1935. The development and morphology of the ligule in grasses. New Phylol. 34: 310-325.

Pohl, R. W. 1967. Controlled maceration of grass leaves in 40-80% nitric acid for preparation of epidermis for slides. Stain Tech. 42: 195-197.

Prat, H. 1922. L'epiderme des Gramineés. Ann. Sci. Nat. Bot., 10° Ser., Pp. 117-324.

1.03 - THE FLOWER, FRUIT, & SEED

FLOWERS

Most of us have never seen grass flowers and we are perhaps not even aware that grasses are flowering plants. The reasons are understandable. Grass flowers are small and hidden away from easy view by a system of reduced leaves (**bracts**). The brightly-colored sepals and petals that make the somewhat distantly-related lilies and orchids so attractive were lost through the gradual processes of evolutionary reduction. This is another way of saying that grass flowers do not strike most people as being terribly pretty. But, come closer!

All that remains of the grass perianth is two or three microscopic structures called **lodicules**. They are **hygoscopic**, swelling in the early morning and thereby forcing apart the bracts that enclose the flower. This process helps to facilitate wind pollination (**anemophily**). Not all grasses have lodicules.

The reproductive components of the grass flower that have been retained are modified for anemophily. The male part of the flower (**androecium**) is made up of stamens, each one consisting of a delicate, thread-like supporting stalk called a **filament** and a sac-like region of pollen-producing tissue, the **anther**. Most grasses have three stamens; some have two or one; a few have six; bamboo flowers may have hundreds of stamens!

The female portion of the flower (**gynoecium**) consists of a seed-producing **ovary**, two **styles** that are separate to their bases, and a terminal pair of feathery **stigmas** that trap airborne pollen.

Grass flowers vary in the presence or absence of reproductive parts. A **bisexual** or **perfect** flower is one that has both an androecium and a gynoecium. A **pistillate flower** has only the gynoecium; while the

staminate flower has only the complement of stamens. A **neuter** or **sterile** flower has no reproductive structures. All grass keys will require you to distinguish among perfect, staminate, pistillate, or neuter flowers. A friendly warning -- what appears so easily defined on this piece of paper is often very difficult to interpret under the dissecting microscope or handlens. Look at several flowers before reaching your decision. One of the more common causes of error arises when anthers develop early, shed their pollen, shrivel up, and fall from the plant. A quick glance can lead to the mistaken notion that the flower is pistillate. Look carefully for filaments as a clue to the presence of fallen anthers.

FRUIT, SEED, AND EMBRYO

The ovule has a single chamber (**locule**) and it is oneseeded. In the vast majority of grasses, it will mature into a fruit type known as the **caryopsis**, in which the seed coat is fused to the ovary wall, except at the funiculus. In a few grasses, the seed is more or less separate from the ovary wall, producing a fruit type called an **achene**. In some bamboos, the fruit is a large, fleshy, single-seeded **berry**.

The seed contains endosperm and the embryo itself. Endosperm results from the fusion of two polar nuclei and a sperm nucleus. It provides nourishment to the developing embryo and later to the young seedling. The endosperm is typically solid and starchy in most grasses; in a few it is in a liquid state.

The embryo consists of the embryo axis and its appendages. At the upper end is the shoot or **plumule**, enclosed in a protective sheath, the **coleoptile**. At the lower end of the embryo axis is the embryonic root or **radicle**, also covered by an enveloping cap, the **coleorhiza**.

The **scutellum** is the major lateral appendage of the embryo. It is embedded directly in the endosperm, where it enzymatically digests and absorbs the stored food material. This function appears to be unique in the flowering plants. The scutellum is often interpreted as a modified cotyledon. Some grasses have a second appendage, called an **epiblast**. Its origin is more controversial. It appears as a small outgrowth opposite the scutellum, at a node just above that of the coleorhiza. In some grasses, there is a distinct region, the **mesocotyl**, between the nodes where the scutellum and the coleoptile are inserted.

SELECTED REFERENCES

Anton, A. M. & M. E. Astegiano. 1973. Notas sobres la morphologia floral de gramineas. Kurtziana 7: 49-53.

Anton, A. M. & A. E. Cocucci. 1984. The grass megagametophyte and its possible phylogenetic implications. Plant Syst. Evol. 146(1-2): 117-121.

Barnard, C. 1956. Floral histogenesis in the monocotyledons. I. The Gramineae. Australian J. Bot. 5: 1-20.

Beetle, A. A. 1980. Vivipary, proliferation, and phyllody in grasses. J. Range Management 33(4): 256-261.

Brown, W. V. 1959. The epiblast and coleoptile of the grass embryo. Bull. Torrey Bot. Club 86: 13-16.

Butzin, F. 1965. Neue Untersuchungen über die Blüte der Gramineae. Druck d. Ernst-Reuter-Ges., Freie Univ. Berlin, Germany. 183 pp.

Celakovsky, L. J. 1897. Über die Homologien des Grasembryos. Bot. Ztg. 9: 141.

Clifford, H. T. 1961. Floral evolution in the family Gramineae. Evolution 15: 455-460.

Cocucci, A. E. & A. M. Anton. 1988. The grass flower: suggestions on its origin and evolution. Flora (Germany) 181(5-6): 353-362.

Davidse, G. 1987. Fruit dispersal in the Poaceae. <u>In,</u> Soderstrom, T. R. et al. (editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 143-155.

Guignard, J. & J. C. L. Maestre. 1970. L'embryon des graminées. Phytomorph. 20: 190-197.

Kinges, H. 1961. Merkmale des Gramineenembryos. Ein Beitrage zur Systematik der Graser. Bot. Jahr. 81(1-2): 50-93.

Negbi, M. 1984. The structure and function of the scutellum of the Gramineae. Bot. J. Linn. Soc. 88(3): 205-222.

Negbi, M. & D. Koller. 1962. Homologies in the grass embryo -- a re-evaluation. Phytomorph. 12(3): 289-296.

Norstog, K. 1972. The early development of the grass embryo and endosperm. Adv. Plant Morph. 1972: 113-125.

Page, J. S. 1978. A scanning electron microscope survey of grass pollen. Kew Bull. 32: 313-319.

Peart, M. H. 1979. Experiments on the biological significance of the morphology of seed dispersal units in grassès. J. Ecol. 67: 843-863.

Philipson, W. R. 1985. Is the grass gynoecium monocarpellary? Amer. J. Bot. 72(12): 1954-1961.

Reeder, J. R. 1977. The "germination flap" in certain Gramineae. Madrono 24: 123, 124.

Rost, T. L. & N. R. Lersten. 1973. A synopsis and selected bibliography of grass caryopsis anatomy and fine structure. Iowa State J. Res. 48(1): 47-87.

Roth, I. 1955. Zur morphologischen Deutung des Grasembryos und verwandter Embryotypen. Flora 142: 564-600.

Roth, I. 1961. Histogenesis and morphological interpretation of the grass embryo. $\underline{\text{In}}$, Recent advances in botany 1: 96-99.

Rowlee, W. W. 1898. The morphological significance of the iodicules of grasses. Bot. Gaz. 25: 199-302.

Sargent, E. & A. Arber. 1915. The comparative morphology of the embryo and seedling in the Gramineae. Ann. Bot. 29: 161-222.

Schuster, J. 1910. Ueber die Morphologie der Grasblute. Flora (Germany) 100: 213-266.

Surkov, V. A. 1961. The ontogenesis and the morphological nature of flower parts in the Gramineae. Bot. Zh. 46: 1134-1143. [Russian, with English summary]

Theile, H. L., H. T. Clifford, & R. W. Rogers. 1996. Diversity in the grass pistil and its taxonomic significance. Australian Syst. Bot. 9: 902-912.

Van Tieghem, P. 1897. Morphologie de l'embryon et de la plantule chez les Graminees et les Cyperacees. Ann. Sci. Nat. Bot. Ser. 8, 3: 259-309.

Walker, E. R. 1906. On the structure of the pistils of some grasses. Nebraska Univ. Studies 6: 203-218.

1.04 - SPIKELET STRUCTURE

THE BASIC PLAN

Because of their small size, high degree of evolutionary reduction, and lack of easily observed features, grass flowers have not been used extensively as the basis for distinguishing genera and species within the family. Instead, the classification of grasses has been based heavily upon the structure of the bracts that enclose individual grass flowers and that subtend groupings of them.

Grass flowers, the minute stalks that support them, and the bract system associated with them make up the **spikelet**. Some spikelets, especially those containing a single flower, may be quite small. Others may be a few centimeters long and easily seen without magnification. The spikelet, although characteristic of Gramineae, is not its exclusive property. Sedges have spikelets, too. Because their spikelets are superficially similar, it is easy to confuse the two families. Refer back to Table 1 for a comparison of the features of the two families.

All grass spikelets are put together according to the same basic plan. The tiny flowers and bracts are attached either directly or indirectly to an unbranched central spikelet axis called a **rachilla**. At the base of the rachilla are two bracts that are empty or sterile, in that they do not flowers in their axils. Each of the two basal bracts is a **glume**. Careful inspection will show that one bract is inserted slightly below the other. The lower bract is the first glume; the one attached slightly above it is the second glume. The two bracts may be similar in length, width, shape, and texture or they may be significantly different from one another. While most grass spikelets have two glumes, a few have only one, and it appears that in a few species the glumes are completely suppressed.

In addition to glumes, a spikelet contains one or more **florets**, each inserted at its own point of attachment (**node**) on the rachilla. The etymology of floret would suggest to you that the term means "a small flower." Not so. A floret is not only an individual small flower, but two bracts that enclose it. A floret never has more than one flower in it. The number of florets in a spikelet is of great diagnostic importance. A spikelet with a single floret is said to be one-flowered; one with two florets is two-flowered; and so on.

The two bracts that enclose the flower are the **lemma** and the palea. The lemma is typically the more conspicuous bract -- larger and of firmer texture, its edges often partially obscuring the palea. The lemma typically has an odd number of nerves or veins of vascular tissue running its length. Occasionally it will appear veinless. The number of nerves on the lemma is of great importance in identifying an unknown grass. Counting their number can be a challenge. It is very easy to overlook submarginal veins, those that lie close to the edge of the lemma. In most instances, the nerves of the lemma will converge with one another towards its apex; but in some grasses, they remain parallel to one another. When viewed in cross-section, a lemma often appears to be a rounded bract. Sometimes it is conspicuously flattened or even Vshaped. It may also have a prominent rib (keel) running down its center, the term being derived from the structure found on the bottom of a ship or boat. The lemma is attached directly to the rachilla. Usually it has a flower in its axil, in which case it is a **fertile lemma**. If the flower is absent, then it is called a **sterile lemma**.

Unlike the glume or lemma, the palea is not the source of many taxonomic features. It tends to be a delicate, membranous, two-nerved bract. It may be as long as the lemma, but it is usually somewhat shorter. The palea is not inserted directly on the rachilla. You will have to take my word for it, because it is all but impossible to see this level of detail under the dissecting microscope. The palea subtends the flower itself and it is attached to the tiny flower stalk.

The apex of a glume or lemma may bear a short, sharp point called a **mucro**. These bracts may also have a more elongate, substantial, hair-like projection known as an **awn**. It may be a few millimeters to several centimeters long. Awns may be straight, bent (**geniculate**), or twisted. Some, as in oats, function in the self-planting of the seed-like fruits. Some awns are terminal, while others arise from the back of a glume or lemma at about their midpoints. Others come from at or near the base of the bract. While glumes and/or lemmas are commonly awned, it is unusual in temperate grasses to find an awned palea.

The hardened base of a lemma or of a floret is its **callus**. In some instances, the callus is a combination of lemma and rachilla tissue. It may be rounded or sharp-pointed, as in the needle grasses. The callus may lack hairs or it may be clothed in a conspicuous tuft of hairs.

Spikelet parts are homologous with the stems and leaves of a grass plant. The rachilla is the homolog of a stem, the glumes and lemmas are homologous with ordinary foliage leaves, and the palea with the specialized first leaf of a side branch, the prophyllum.

COMPRESSION

Spikelets are either round in cross-section (**terete**) or they are flattened (**compressed**). Terete spikelets are relatively uncommon, but they occur in such common plants as the Indian rice-grass. Compressed spikelets come in two models. If the bracts are flattened as though pressure were brought to bear from the sides of the bracts, then the spikelet is **laterally compressed**. If the spikelet is flattened as though pressure were brought to bear from the backs of the bracts, then it is **dorsally compressed**.

Perhaps this distinction between dorsal and lateral compres-sion may be made clearer by drawing on two familiar animals that are flattened. Turtles show dorsal compres-sion, while fish are laterally compressed.

DISARTICULATION

At maturity, most spikelets will break apart at predeter-mined points of separation. The process is

called **disarticulation** and it occurs in various ways:

- below the first glume, so that the entire spikelet falls from the plant;
- above the glumes and between the florets so that empty glumes are all that remains behind;
- florets may fall separately or in clusters, sometimes with a prominent segment of the rachilla remaining attached.
- between the first and second glume (an unusual situation); or
- above glumes, but lemmas persisting (an unusual situation).

It takes some practice to determine disarticulation. You can force it to occur with a dissection needle, but not necessarily where it would under natural conditions. I recommend that you always observe older inflorescences -- ones that may otherwise look uninviting -- to find bare pedicels or empty glumes.

There is a tendency -- and it is nothing more than that -- for spikelets that are laterally compressed to disarticulate above the glumes and for those that are dorsally compressed to disarticulate below the glumes.

SEXUALITY

An individual floret or grass plant may be:

- bisexual (perfect or hermaphroditic), if it has both stamens and carpels;
- staminate (male), if it has only male florets;
- pistillate (female), if it has only female florets;
- sterile (neuter or barren), if it lacks either functional carpels or stamens (or, especially in older literature, if a floret were staminate).

There is another level of complexity. In many grasses, the lower florets of a spikelet are typically bisexual, with the upper florets progressively smaller and sterile. Another com-mon situation is the spikelet of panicoid grasses, in which the upper floret is bisexual and the lower is sterile. A less common possibility is the several-flowered spikelet that has both upper and lower florets that are sterile, while those in the middle are fertile.

Grass species, depending on the distribution of their stamens and carpels, may be described as:

- ⇒ bisexual;
- monoecious, if an individual plant produces both staminate and pistillate spikelets;
- dioecious, if an individual plant produces either staminate or pistillate spikelets;
- various combinations of perfect and unisexual spikelets, on the same or different plants of a species.

VARIATION ON A THEME

To summarize, a typical grass spikelet consists of two

glumes and one or more florets attached to a rachilla. It is laterally or dorsally compressed, or less often terete. The spikelet disarticulates above or below the glumes. The lemmas and/or the glumes may be awned.

While this is the basic plan, the spikelet is subject to a fascinating series of modifications. One of the most important of these is the reduction and loss of spikelet parts. One or both glumes may be missing. In spikelets with more than one floret, the upper one(s) are often smaller than the lower one(s) and they may be sterile. Sometimes the uppermost floret is well-developed and fertile, while the one or two florets below it are reduced. In a few grasses, the middle florets are best developed, while those above and below are reduced or sterile. The palea may be reduced or even absent, as in the bent grasses. This variation can be frustrating at first, but proper interpretation can be more easily assured if you take the time now to learn the basic positional relationships of the spikelet parts.

SPIKELET FORMULAE

A convenient system for summarizing the number of flower parts characteristic of various plant families is called a **floral formula**. In this system of notation, each of the four floral series is given an abbreviation, such as K (for calyx), C (for corolla), A (for androecium), and G (for gynoecium). Exponents or superscripts indicate the number of sepals, petals, stamens, and carpels. I developed the following little system for describing spikelets.

G = glume

 G^0 = glumes absent

G¹ = one glume present

G² = two similar glumes present

 G^{1+1} = two dissimilar glumes

F = fertile floret

 $F^{2-x} = 2$ to several fertile florets

F^a = awned floret

L = sterile lemma (of a sterile floret)

 $L^{x} = 3$ to several sterile lemmas

SOME SPIKELET MODELS

Agrostis $G^2 + F$ Andropogon $G^2 + L^1 + F^a$ Avena $G^2 + F^{2-3}$ Panicum $G^{1+1} + L + F$ Paspalum $G^1 + L + F$ Phalaris $G^2 + L^{1-2} + F$

Poa $G^2 + F^{2-x}$ Uniola $G^2 + L^x + F^x + L^x$ Zizaniopsis $G^0 + F$

SELECTED REFERENCES

Allred, K. W. & J. T. Columbus. 1988. The grass spikelet formula: an aid in teaching and identification. J. Range Management 41(4): 350, 351.

Chevalier, G. 1949. Observations sur les squamules de Graminees. Ann. Sc. Nat. Bot. 11 serie, Pp. 123-129.

Clayton, W. D. 1990. The spikelet. <u>In</u>, Chapman, G. P. (editor). Reproductive versatility in the grasses. Cambridge Univ. Press. Cambridge, England. Pp. 32-51.

Janchen, E. 1938. Der morphologische wert der Gramineen-vorspelze. Oster Bot. Ztschur. 87: 51-61.

McClure, F. A. 1972. The agrostological term, anthoecium. Taxon 21(1): 153, 154.

Nozeran, R. 1955. L'epillet des graminées. <u>In</u>, Contribution à l'étude de quelques structures florales. Ann. Sc. Nat. Bot. 11 serie. 16: 1-224.

Philipson, W. R. 1934. The morphology of the lemma in the grasses. New Phytol. 33: 359-371.

Pilger, R. 1939. Zur Morphologie des Aehrchens der Gramineen. Bot. Jahrb. Syst. 69: 401-418.

Pilger, R. 1948. Additamenta agrostologica. 2. Der Vorspelze der Gramineen. Bot. Jahr. 74: 199-265.

Piper, C. V. 1906. The terminology of the parts of the grass spikelet. Science 23: 789, 790.

Pissarek, H. -P. 1971. Untersuchungen uber Bau and Funktion der Gramineen-Lodiculae. Beitr. Zur Biol. Pflanz. 217: 313-370.

Thomasson, J. R. 1985. Miocene fossil grasses: possible adaptation in reproductive bracts (lemma and palea). Ann. Missouri Bot. Gard. 72: 843-851.

Tran, T.-T.-H. 1965. Les glumelles inferieures aristees de quelques Graminées: anatomie morphologie. Bull. Jard. Bot. Bruxelles. 35(3): 219-284.

Thomasson, J. R. 1976. Clearing, cuticle removal, and staining for the fertile bracts, lemmas, and palea of grass enthoecia. Stain Technol. 53: 233-236.

von Mohl, H. 1845. Ueber die Bedeutung der untern Blumenspelze der Graser. Bot. Ztg. 3: 3-33.

Wycherley, P. R. 1954. Vegetative proliferation of floral spikelets in British grasses. Ann. Bot. 18: 119-127.

1.05 - INFLORESCENCE TYPES

Grass stems, whether they are the primary culm or a lateral branch, may emerge from their sheaths and bear one to several hundred spikelets. This flowering portion of the grass plant is its **inflorescence**. A stem may bear only one inflorescence or it may have several of them. If it emerges from the uppermost sheath of a primary stem, it is a **terminal** inflorescence. If it arises from a lower sheath, it is an **axillary** or **lateral** inflorescence.

At first, it may be difficult for you to determine just how much of what you are looking at makes up a single inflorescence. A good rule of thumb is that there are never well-developed foliage leaves within an inflorescence. Whether terminal or axillary, the uppermost or outermost spikelet delimits the top of an inflorescence. The lowest typical foliage leaf marks its base.

The upper portion of the culm that supports the entire inflorescence is the **peduncle**, while the stalk that supports an individual spikelet is its **pedicel**. This terminology is not consistent with usage in other plant families. A pedicel is usually the stalk that supports a single flower. The true pedicel of a grass flower is, of course, within the spikelet. This error in terminology goes back about two hundred years to a time when attempts were made by Linnaeus and others to interpret the spikelet as a flower. The interpretation is incorrect, but unfortunately, the term persists.

If there is a clearly defined axis within the inflorescence, it is called a **rachis**. Note that the rachis is the axis of an entire inflorescence of spikelets, while the rachilla is the central axis of an individual spikelet. The rachis may be delicate, wiry, or even thickened with spikelets partially embedded in its tissue.

The exact arrangement of spikelets determines the inflorescence type. You will find this terminology frustrating because it has not been standardized and authors of keys and descriptions vary shamefully in their usage. No scheme is without its problems, but I have found the following one useful.

SIMPLE INFLORESCENCES

In the **spike**, the spikelets are inserted directly on an unbranched rachis. Pedicels are, for all practical purposes, absent. The number of spikelets attached at a given node on the rachis is variable. One, two, three, and a cluster of several spikelets per node are common. Many grasses have this inflorescence type.

In the **raceme**, spikelets are borne on well-developed pedicels arising from an unbranched rachis. Typically spikelets occur in pairs or trios at a given node, infrequently only one spikelet per node, as in the semaphore grasses. It is much less common than the spike. The distinction between the raceme and spike is arbitrary, the degree of pedicel development marking the difference. I use 1 mm as the dividing line.

The **rame** is a specialized modification of the raceme in which pedicellate and sessile spikelets occur

together in pairs or trios. The pedicels are of equal or unequal length. The rame is typical of the barley and bluestem tribes (Triticeae and Andropogoneae). Few authors recognize the rame as distinct from the raceme.

The **panicle** is probably the most common inflorescence type in the family. Here the spikelets are borne on pedicels that are themselves secondary or tertiary branches of a much-branched system. This means that the spikelets are not attached directly to a central axis as they are in the spike, raceme, or rame. Panicles may be large, open, and very conspicuously branched or it may be so contracted and dense that they appear to be some sort of spike.

An extreme form of the panicle is the **solitary spikelet**, in which the peduncle bears a single spikelet, as in the poverty oats. In such instances, we believe that the solitary spikelet is the result of evolutionary reduction of a more typical muchbranched panicle with multiple spikelets.

In the spike, raceme, or rame, the spikelets may be more or less evenly attached on opposite sides of the rachis so that the inflorescence is **balanced**, or they may be obviously attached on just one side of the rachis, so that the inflorescence is **one-sided**. If the spikelets are tightly packed along one side, as in the teeth of a comb, the inflorescence is said to be **pectinate**. Examples may be seen in grama grass (*Bouteloua*) and in toothache grass (*Ctenium*).

COMPOUND INFLORESCENCES

In many grasses, we see inflorescences composed of unbranched or sparingly branched arms. If we look at any particular branch, it bears a spike of spikelets, a raceme of spikelets, or a rame of spikelets. We may refer to them as compound spikes, compound racemes, and compound rames, respectively. In older literature, these are also considered to be panicles because they are branched inflorescences. In these compound inflorescences, the branches may be clustered at the apex of a peduncle (digitate) or they may be attached at various points along a rachis and be racemose.

SELECTED REFERENCES

Allred, K. W. 1982. Describing the grass inflorescence. J. Range Management 35(5): 672-675.

Butzin, F. 1977. Evolution of inflorescences in the panicoid grasses with bristles. Willdenowia 8(1): 67-80.

Càmara-Hernàndez, J. & G. H. Rua. 1991. The synflorescence of Poaceae. Beitr. Zur Biol. Der Pflanzen 66: 297-311.

Gram, K. 1961. The inflorescence of the grasses. Bot. Tid. 56: 293-313.

Vegetti, A. C. & A. M. Anton. 1995. Some trends in Poaceae inflorescences. Flora 190: 225-228.

Vegetti, A. C. & A. M. Anton. 2000. The grass inflorescence. In, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 29-31.

Vegetti, A. C. & F. Weberling. 1996. The structure of the paracladial zone in Poaceae. Taxon 45(3): 453-

Wilhelm, W. W. & G. S. McMaster. 1996. Spikelet and floret naming scheme for grasses with a spike inflorescence. Crop Sci. 36(4): 1071-1073.

1.06 - POLLINATION & REPRODUCTION

"... grasses break almost all of the rules that many other groups of animals and plants observe....

INTRODUCTION

Pollination is the transfer of pollen from an anther to the surface of a stigma. The term is clearly not synonymous with fertilization, which involves the union of egg and sperm nuclei. In most flowering plants, the two structures are in different flowers and we speak of **cross-pollination**. It offers the selective advantage of yielding new genetic combinations from two different plants. On the other hand, crosspollination requires a pollinator - insects, birds, water, wind, moths, etc.

In many instances, the stigma of a particular flower is receptive to its own pollen grains or to other flowers on the same plant and **self-pollination** occurs. No pollinator is required. This mechanism obviously is not found in dioecious species, but it may occur in monoecious species, such as maize. Most plants have developed mechanisms that prevent or retard selfpollination.

WIND POLLINATION

Grasses are heavily adapted for wind pollination and cross-fertilization. The syndrome of adaptation that we find in the family includes:

- lodicules sensitive to weather;
- elongation of stamen filament;
- ℧ new orientation of anthers;
- quick shedding of pollen;
- pollen released with high temperatures and lowered humidity; pollen that is light, abundant, easily dispersed;
- light pollen has lowered terminal velocities;
- lower probability of entrapment; ℧
- large, easily exserted stigma;
- inflorescences well-elevated above vegetative plant parts; and
- lack of nectaries.

INSECT POLLINATION

Not all grasses are wind pollinated (anemophilous). A number of visits by insects have been recorded in the literature. They appear to be looking for food, in the form of pollen grains or various sweet liquids made by the *Claviceps* fungi. Grasses are commonly visited when they are in flower. Insects move from one flower to another and thereby transfer pollen.

Examples may be found in our temperate grasses, but it is perhaps more common in tropical rain forests where there is little, if any, wind to effect wind pollination.

INCOMPATIBILITY

I have already mentioned that most plants have developed mechanisms that favor cross-pollination, or to put it differently they are self-sterile or self-incompatible. Several possibilities come easily to mind: pollen is produced at a time when the stigmatic surface of that same flower is not receptive; stamens may have short filaments and the styles of the same flower may be very long, thereby physically making self-pollination more difficult. Less obvious are those that may involve genetic incompatibility.

The incompatibility mechanism found in Gramineae is unique. It is referred to as the "SZ incompatibility system." Here are the basic elements:

- Compatibility or incompatibility rests on the interaction of two genes, S and Z.
- Both S and Z occur in multi-allelic series, S1, S2, S3, etc.
- In a diploid grass, a haploid pollen grain would contain one S allele and one Z allele. The diploid stigma and style tissue would have nuclei with two of each of them.
- Incompatibility is determined by whether the pollen grain and stigma/style share alleles. If they share none, they are compatible. If they share one, they are compatible. If they share both, they are incompatible.
- Prevents fertilization between genotypes identical in incompatibility alleles.
- Greater the allelic differences -> greater chance that pollen will function.

Stigma/Style	Pollen Grain	Outcome
\$1\$2Z1Z2 \$1\$2Z1Z2 \$1\$2Z1Z2 \$1\$2Z1Z2 \$1\$2Z1Z2	S1Z1 S1Z2 S2Z1 S2Z2	Fails Fails Fails Fails
S1S2Z1Z2 S1S2Z1Z2 S1S2Z1Z2 S1S2Z1Z2	S1Z1 S1Z3 S2Z1 S2Z3	Fails OK Fails OK

S1S2Z1Z2	S1Z1	Fails
S1S2Z1Z2	S1Z3	OK
S1S2Z1Z2	S3Z1	OK
S1S2Z1Z2	S2Z3	OK
\$1\$2Z1Z2	S3Z3	OK
\$1\$2Z1Z2	S3Z4	OK
\$1\$2Z1Z2	S4Z3	OK
\$1\$2Z1Z2	S4Z4	OK

[After Chapman & Peat, 1992]

BREEDING SYSTEMS

Grasses may be predominantly or exclusively outcrossing or cross-fertilized (**allogamous**) or they are predominantly self-fertile (**autogamous**) or they may be outcrossing for some period of time and then switch to selfing at another stage. The taxonomic consequences of crossing and selfing can be both amazing and very frustrating, especially to those who adhere to the classical version of the biological species concept.

DISTRIBUTION OF REPRODUCTIVE STRUCTURES

The terms **fertile** and **sterile** (**barren**, **neuter**, or **neutral**) have different meanings, depending on the author. In the broad sense, a flower, spikelet, or grass plant is fertile if it bears stamens and/or carpels. In H & C, they are fertile only if they bear functional carpels.

Similarly, a flower, spikelet, or grass plant is sterile if it lacks functional carpels, even if the stamens are func-tional! In other words, fertility and sterility are defined by the presence or absence of female reproductive structures.

A flower, a spikelet, or a grass plant that has both stamens and carpels is **perfect** or **bisexual**. If it has either stamens or carpels, but not both, it is **imperfect** or **unisexual**, and a flower, spikelet, or plant is either male (**staminate**) or female (**pistillate**).

A plant that bears both staminate and pistillate flowers or spikelets is **monoecious**. If male and female flowers or spikelets occur on separate plants, then we have the **dioecious** condition. These terms do not apply to flowers or spikelets – only to plants or species.

Plants that bear both perfect and imperfect flowers or spikelets are said to be **polygamous**. Four flavors are recognized:

- ♂ + bisexual on same plant = andromonoecious
- → + bisexual on different plants = androgynoecious
- ♀ + bisexual on same plant = gynomonoecious
- ♀ + bisexual on different plants = gynodioecious

TYPES OF REPRODUCTION

"Grasses, like old men, have found a substitute for sex."

[Richard W. Pohl, a personal revelation]

There are basically two kinds of reproduction -- **sexual** and **asexual**. Sexual reproduction involves the union of egg and sperm nuclei. Grasses have developed a series of mechanisms that favor outcrossing, so that the gametes that unite come from different plants. These include:

- imperfect flowers and spikelets;
- self-sterility;
- chasmagamous flowers (ones that are open pollinated); and
- protandrous flowers, in which the stamens shed their pollen before the stigmas are receptive.

On the other hand, mechanisms that favor inbreeding or selfing include:

- perfect flowers or spikelets;
- self-fertility; and
- cleistogamous flowers, those that are closed and self-pollinated.

Apomixis is the general term used for all types of asexual reproduction, where there is no union of egg and sperm. The simplest form of apomixis is vegetative reproduction by means of rhizomes, bulbs, corms, bulblets or bulbils (vegetative proliferations that replace flowers), and fragmentation of stems. New plants arise vegetatively because these various structures contain buds that will produce new stems, roots, leaves, and spikelets if suitable moisture and nutrients are available. Many of our most successful weedy grasses have exploited asexual reproduction. Each of those chopped up rhizome segments or bulbs is fully capable of yielding a new, independent plant that is a genetic carbon copy of its mother.

Grasses can be much more subtle about their asexual reproduction. In **agamospermy**, seeds are produced, but they are not the product of the union of egg and sperm. In **adventitious embryony**, the embryo develops directly from the diploid tissue of the nucellus or ovule integment. The gametophytic generation has been completely bypassed.

In **gametophytic apomixis**, alternation of generation occurs, but the gametophytes arise without meiosis having occurred. Five versions are recognized:

- apospory, in which a diploid embryo sac is formed directly from a cell of the nucellus or of the inner integument;
- diplospory, in which a diploid embryo sac is formed from a cell of the archegonium
- parthenogenesis, in which the embryo forms from a diploid egg cell;
- apogamy, in which the embryo forms from some cell in the embryo sac other than the egg cell; and
- pseudogamy, in which pollination is required to stimulate seed set.

KEY TO STRATEGIES

- Reproduction by means of the union of egg and sperm nuclei (sexual reproduction) ->
- Reproduction by means of vegetative tissue only or by means of seeds formed without the union of egg and sperm (asexual reproduction or apomixis)

-> 3

- 2. Plants self- or closed pollinated -> Cleistogamy
- Plants cross- or open-pollinated ->Chasmogamy
- Reproduction by means of rhizomes, tillers, stolons, bulbs, corms, or aerial stem fragments (node + bud) -> Vegetative reproduction
- Reproduction by means of modified spikelet parts or by means of seeds formed without union of egg and sperm ->
- 4. Palea and/or lemma modified into bulblets ->Vivipary (proliferation)
- Neither palea nor lemma so modified; fertile seed formed through partial use of sexual life cycle (agamospermy or apomixis s. s.) ->
- Embryo sac not formed; embryo developing from somatic cells -> Adventitious embryony
- 5. Embryo sac formed ->
- Embryony sac derived from somatic cell of the ovary -> Apospory
- 6. Embryo sac derived from a megaspore mother cell-> Diplospory

SOME EXAMPLES

Inbreeding: Aira, Avena, Bouteloua, Brachypodium, Briza, Bromus, Chloris, Cortaderia, Danthonia, Deschampsia, Digitaria, Echinochloa, Eleusine, Elymus, Eragrostis, Festuca, Glyceria, Hordeum, Hyparrhenia, Koeleria, Lagurus, Lolium, Oryza, Panicum, Paspalum, Phalaris, Phleum, Poa, Polypogon, Puccinellia, Rottboellia, Secale, Setaria, Sieglingia, Sorghum, Sporobolus, Stipa, Trisetum, Triticum, Vulpia, Zizania.

Cleistogamy: Agrostis, Amphicarpum, Arctagrostis, Aristida, Avena, Bothriochloa, Bouteloua, Briza, Bromus, Calamagrostis, Chasmanthium, Chloris, Cottea, Dactyloctenium, Danthonia, Deschampsia, Dichanthelium, Dichanthium, Digitaria, Echinochloa, Eleusine, Enneapogon, Eragrostis, Erianthus, Festuca, Gymnopogon, Helictotrichon, Hemarthria, Hordeum, Leersia, Leptochloa, Melica, Microchloa, Microstegium, Muhlenbergia, Oryza, Oryzopsis, Panicum, Pappophorum, Paspalum, Phippsia, Piptochaetium, Poa, Schizachyrium, Secale, Setaria, Sieglingia, Sorghum, Spartina, Sporobolus, Stipa, Tridens, Triplasis, Trisetum, Uniola, Vulpia, Willkommia.

Apomixis: Anthephora, Bothriochloa, Brachiaria, Calamagrostis, Cenchrus, Chloris, Coix, Cortaderia, Dichanthelium, Elymus, Eragrostis, Eriochloa, Heteropogon, Hierochloe, Hilaria, Hyparrhenia, Nardus, Panicum, Paspalum, Pennisetum, Poa, Saccharum, Setaria, Sorghum, Themeda, Tripsacum, Urochloa.

Hidden Cleistogenes: Amphicarpum, Andropogon, Aristida, Chloris, Cottea, Danthonia, Dichanthelium, Digitaria, Diplachne, Enneapogon, Leersia, Muhlenbergia, Pappophorum, Paspalum, Pennisetum, Phippsia, Sieglingia, Stipa, Triplasis.

Subterranean Cleistogenes: *Amphicarpum, Chloris, Paspalum*

Only Unisexual Florets: Allolepis, Arrhenatherum,

Buchloe, Cathestecum, Coix, Cortaderia, Distichlis, Gynerium, Heteropogon, Luziola, Monanthochloe, Olyra, Opizia, Pharus, Poa, Scleropogon, Tripsacum, Zea, Zizania, Zizaniopsis.

Monoecious: Buchloe, Coix, Distichlis, Luziola, Olyra, Pharus, Scleropogon, Tripsacum, Zea, Zizania, Zizaniopsis.

Dioecious: Allolepis, Buchloe, Cortaderia, Distichlis, Gynerium, Monanthochloe, Opizia, Poa, Scleropogon.

*After Watson, L. 1990. <u>In</u>, Chapman, G. P. Reproductive versatility in the grasses. Cambridge Univ. Press. Pp. 14, 15.

SELECTED REFERENCES

POLLINATION

Adams, D. E., W. E. Perkins, & J. R. Estes. 1981. Pollination systems in *Paspalum dilatatum* Poir. (Poaceae): an example of insect pollination in a temperate grass. American J. Bot. 68: 389-394.

Karr, J. R. 1976. An association between a grass (*Paspalum virgatum*) and moths. Biotropica 8(4): 284, 285.

Sendulsky, T. 1993. First report of ballistochory [ballistic ejection of grains] in the Poaceae. Ann. Missouri Bot. Gard. 80(2): 518-521.

Soderstrom, T. R. & C. E. Calderón. 1971. Insect pollination in tropical rain forest grasses. Biotropica 3: 1-16.

REPRODUCTION

Berg, A. R. 1972. Grass reproduction. $\underline{\text{In}}$, Younger & McKell. Pp. 335-347.

Brown, W. V. & W. H. P. Emery. 1958. Apomixis in the Gramineae. American J. Bot. 45: 253-263.

Campbell, C. S. et al. 1983. Cleistogamy in grasses. Ann. Rev. Ecol. Syst. 14: 411-441.

Chapman, G. P. 1990. Reproductive versatility in grasses. Cambridge Univ. Press. Cambridge, England. 296 pp.

Chapman, G. P. 1992. Apomixis and evolution. <u>In</u>, Grass evolution and domestication. Cambridge Univ. Press. Cambridge, England. Pp. 138-155.

Connor, H. E. 1979. Breeding systems in grasses: a survey. New Zealand J. Bot. 17: 547-574.

Connor, H. E. 1981. Evolution of reproductive systems in the Gramineae. Ann. Missouri Bot. Gard. 68: 48-74.

Connor, H. E. 1987. Reproductive biology in the grasses. <u>In</u>, Soderstrom, T. R. et al. Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 117-132.

Fryxell, P. A. 1957. Modes of reproduction in higher plants. Bot. Rev. 23: 135-233.

Knox, R. B. 1967. Apomixis: seasonal and population differences in a grass. Science 157: 6-15.

Lundquist, A. 1968. The mode of origin of self-fertility in grasses. Hereditas 59: 413-426.

Marshall, D. R. & A. H. D. Brown. 1981. The evolution of apomixis. Heredity 47: 1-15.

Quarin, C. 1992. The nature of apomixis and its origin in panicoid grasses. Apomixis Newsletter 5: 8-15.

Quinn, J. A. 2000. Adaptive plasticity in reproduction and reproductive systems of grasses. <u>In</u>, Jacobs, S. W.

L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 281-286.

Reddy, P. S. 1977. Evolution of apomictic mechanisms in Gramineae – a concept. Phytomorph. 27: 45-50.

Stebbins, G. L. 1941. Apomixis in the angiosperms. Bot. Rev. 7: 507-542.

1.07 - GENOMES, HYBRIDS, & POLYPLOIDS

GENOMES

A **genome** is a complete set of chromosomes. In a diploid, it is either of the two sets of chromosomes derived from the parents. In a polyploid, a genome is any of the sets derived from its ancestors. Genomes are usually designated by a single capital letter, as in the B genome of wheat. The number of chromosomes in a genome is often designated by a small letter \mathbf{x} , as in $\mathbf{x} = \mathbf{7}$.

In a **euploid**, the individual, cell, or nucleus contains an exact multiple of x. If x = 10, then 10, 20, 30, 40, etc. would constitute a euploid series. In **aneuploids**, we have chromosome numbers that are not exact multiples of the base chromosome number. Individual chromosomes have been added or lost. Several kinds of aneuploids are recognized. In the **nullisomic**, both members of a chromosome pair are missing. In a **monosomic**, only one member is missing. In a **trisomic**, one member of a pair is in triplicate. In a **tetrasomic**, both members of the chromosome pair are duplicated.

We use the small letter \mathbf{n} to designate the number of chromosomes found in a sex cell or gamete. The number of chromosomes found in a vegetative cell is shown as $2\mathbf{n}$, as in $2\mathbf{n} = 10$. These two conditions are also called the gametic and somatic chromosome numbers, respectively.

A convention has been adopted in plant genetics that tells the chromosome number and ploidy level. According to this scheme the "genetic formula" for bread wheat is 2n = 6x = 42. This means that the vegetative cells contain 42 chromosomes (the 2n indicating a somatic number, rather than a gametic one), and that the plant is a hexploid (6x). If 6x = 42, then x = 7.

POLYPLOIDY

An individual, a cell, or a nucleus that contains one and only one complete sets of chromosomes is **haploid**. If it contains two complete sets, it is a **diploid**. An individual, a cell, or a nucleus that contains three or more complete sets of chromosomes is a **polyploid**. The combination of the prefixes tri-, tetra-, penta-, hexa-, octa-, etc., and the suffix -ploid, indicates the particular number of complete sets. A hexaploid has six sets.

A polyploid in which all of the chromosome sets are

derived from the same genetic lineage, so that a chromosome in one set is capable of pairing with its corresponding number in another set, is an **autoploid** or **autopolyploid**. If, on the other hand, we are dealing with a polyploid in which the chromosome sets are derived from different species, subspecies, or varieties, then we have an **allopolyploid**. Here the chromosome sets are different enough that pairing of chromosomes is impaired -- slightly to completely. In a **segmental allopolyploid**, the polyploid appears to be of both auto- and allopolyploid, origin. A series of interrelated polyploids, often demonstrating morphological similarities to one another constitute a **polyploid complex**.

HYBRIDIZATION

C. D. Darlington (1937) defined a hybrid as "a zygote produced by the union of dissimilar gametes." While theoretically acceptable, this definition is too broad for general use. Essentially all sexually reproducing plants and animals would be hybrids. Your classes are filled with hybrid students. We will use the term in a more restricted sense—the offspring of interbreeding (crossing) between two or more taxa. The process itself is called hybridization. There are hundreds (probably thousands) of crosses between grass species (interspecific hybrids) and a long list of crosses between grass genera (intergeneric hybrids). Here is a long, but incomplete list of the latter.

Aegilops X Elymus Aegilops X Secale Aegilops X Triticum Agropyron X Elymus Agropyron X Hordeum Agropyron X Secale Agropyron X Triticum Agrostis X Calamagrostis Agrostis X Polypogon Ammophila X Calamagrostis Arctophila X Dupontia Arrhenatherum X Avena Avena X Arrhenatherum Bothriochloa X Dichanthium Bromus X Festuca Calamagrostis X Agrostis Chloris X Cynodon Colpodium X Phippsia Cynodon X Chloris Danthonia X Sieglingia Dichanthium X Bothriochloa Dupontia X Arctophila

Elymus X Aegilops Elymus X Hordeum Erianthus X Saccharum Festuca X Bromus Festuca X Lolium Festuca X Vulpia Hordeum X Elymus Hordeum X Secale Hordeum X Triticum Imperata X Saccharum Koeleria X Trisetum Leptochloa X Oryza Lolium X Festuca Miscanthus X Saccharum Oryza X Leptochloa Oryza X Pennisetum Oryza X Sorghum Oryza X Triticum Oryzopsis X Stipa Pennisetum X Oryza Pennisetum X Zea Phippsia X Colpodium Phippsia X Puccinellia Polypogon X Agrostis Puccinellia X Phippsia Saccharum X Erianthus Saccharum X Imperata Saccharum X Miscanthus Saccharum X Sorghum Saccharum X Žea Secale X Aegilops Secale X Elymus Secale X Triticum Sieglingia X Danthonia Sorghum X Oryza Sorghum X Saccharum Sphenopholis X Trisetum Stipa X Oryzopsis Tripsacum X Zea Trisetum X Koeleria Trisetum X Sphenipholis Triticum X Aegilops Triticum X Elymus Triticum X Secale Vulpia X Festuca Zea X Saccharum Zea X Tripsacum

NAMED INTERGENERIC HYBRIDS

Sometimes the offspring of crossing between plants in closely related genera are stable enough to warrant naming, even though they are partially to completely sterile. The following intergeneric hybrids have been named. Not all of them are legal according to the ICBN. It is drawn primarily after Clayton & Renvoize (1986) and Watson & Dallwitz (1992).

Aegilosecale Aegilops x Secale Aegilops x Triticum Agrostis x Calamagrostis Aegilotriticum Agrocalamagrostis Agroelymus Agropyron x Elymus Agropyron x Hordeum Agrohordeum Agrostis x Polypogon Agropogon Agrositanion Agropyron x Śitanion Agrotrisecale Agropyron x Triticum x Secale Agropyron x Triticum Agrotriticum Ammophila x Calamagrostis Ammocalamagrostis Arctophila x Dupontia Arctodupontia Bromofestuca Bromus x Festuca Calammophila Calamagrostis x Ammophila Cynodon x Chloris Cynochloris

Danthosieglingia Danthonia x Sieglingia Dupontia x Poa Dupoa Dupontopoa Dupontia x Poa Elyhordeum Elymus x Hordeum Elyleymus Élymus x Leymus Elymostachys Elymus x Psathyrostachys Elymus x Elytrigia Elymotrigia[°] Elymotriticum Elymus x Triticum Elymus x Sitanion Elysitanion Euchlaezea Euchlaena x Zea Festulolium Festuca x Lolium Festulpia Festuca x Vulpia Hordeum x Secale Hordale Leymopyron Leymus x Agropyron Oryza x Triticum Puccinellia x Phippsia Oryzticúm Pucciphippsia Sitanion x Hordeum Sitordeum Stipa x Oryzopsis Stiporyzopsis Trisetobromus Trisetum x Bromus Trisetum x Koeleria Trisetokoeleria Triticale Triticum x Secale Triticum x Secale Triticosecale Triticum x Hordeum Tritordeum

SELECTED REFERENCES

Anderson, E. & G. L. Stebbins. 1954. Hybridization as an evolutionary stimulus. Evolution 8: 378-388.

Calzada, J.-P. V., C. F. Crane, & D. M. Stelly. 1996. Apomixis: the asexual revolution. Science 274: 1322, 1323.

Darlington, C. D. 1937. What is a hybrid? J. Heredity 28: 308.

De Wet, J. M. J. 1987. Hybridization and polyploidy in the Poaceae. <u>In</u>, Soderstrom, T. R. et al. (editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 188-194.

Hackel, E. 1906. Uber Kleistogamie bei den Grasern. Osterreichische Bot. Zeitschrift 56: 81-88; 143-154; 180-186.

Harlan, J. R. and J. M. J. de Wet. 1963. The compilospecies concept. Evolution 17:497-501.

Hermsen, J. G. T. 1984. Nature, evolution, and breeding of polyploids. Iowa State J. Res. 58(4): 411-420.

Heslop-Harrison, J. 1979. The pollen-stigma interaction in grasses: a brief review. New Zealand J. Bot. 17(4): 537-546.

Hunziker, H. H. & G. L. Stebbins. 1987. Chromosomal evolution in the Gramineae. <u>In, Soderstrom, T. R. et al. (editors)</u>. Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 179-187.

Johnson, B. L. 1972. Polyploidy as a factor in the evolution and distribution of grasses. <u>In</u>, Younger & McKell. Pp. 19-35.

Jenkins, T. J. 1933. Intergeneric and interspecific hybrids in herbage grasses. Initial crosses. J. Genet. 28: 205-264.

Jenkins, T. J. 1954. Interspecific and intergeneric hybrids in herbage grasses. IV- VIII. J. Genet. 52: 239-331.

Jenkins, T. J. 1955. Interspecific and intergeneric hybrids in herbage grasses. IX-XVIII. J. Genet. 53: 81-130; 380-486.

Kellogg, E. A. 1990. Variation and species limits in agamospermous grasses. Syst. Bot. 15(1): 112-123.

Kimber, W. M. 1947. Cytology and genetics of forage grasses. Bot. Rev. 13: 319-421.

Knobloch, I. W. 1963. The extent of hybridization in the Gramineae. Darwinia 12: 624-628.

Knobloch, I. W. 1968. A check list of crosses in the Gramineae: Dept. of Botany & Plant Pathology. Michigan State Univ. East Lansing.

Löve, A. 1951. Taxonomical evaluation of polyploids. Caryologia 3(3): 263-284.

Morgan, W. G. 1976. A technique for the production of polyploids in grasses. Euphytica 25(2): 443-446.

Myers, W. M. 1947. Cytology and genetics of forage grasses. Bot. Rev. 13: 319-421.

Niklas, K. J. 1985. The aerodynamics of wind pollination. Bot. Rev. 51: 328-386.

Niklas, K. J. 1985. Wind pollination -- a study in controlled chaos. American Sci. 73: 462-470.

Peart, M. H. 1981. Further experiments on the biological significance of the morphology of seed-dispersal units in grasses. J. Ecol. 69: 425-436.

Peart, M. H. 1984. The effects of morphology, orientation and position of grass diaspores on seedling survival. J. Ecol. 72: 437-453.

Rabinowitz, D. & J. K. Rapp. 1981. Dispersal abilities of seven sparse and common grasses from a Missouri prairie. American J. Bot. 68: 616-624.

Smith, D. C. 1956. Progress in grass breeding. Adv. Agron. 8: 127-162.

Stebbins, G. L. 1947. Types of polyploids: their classification and significance. Adv. Genet. 1: 403-429.

Stebbins, G. L. 1949. The evolutionary significance of natural and artificial polyploids in the family Gramineae. Proc. 8th Inter. Congr. Genet., 1948. Pp. 461-485.

Stebbins, G. L. 1952. Species hybrids in grasses. Proc. Inter. Grassland Congr. 6: 247-253.

Stebbins, G. L. 1956. Taxonomy and the evolution of genera, with special reference to the family Gramineae. Evol. 10: 235-245.

Stebbins, G. L. 1975. The role of polyploid complexes in the evolution of North American grasslands. Taxon 24: 67-80.

Stebbins, G. L. 1985. Polyploidy, hybridization and the invasion of new habitats. Ann. Missouri Bot. Gard 72: 824-382.

Stebbins, G. L. & R. M. Love. 1941. A cytological study of California forage grasses. American J. Bot. 28: 371-382.

SECTION 2 – SYSTEMATICS OF THE GRASSES

2.01 - GRASSES & THEIR RELATIVES

The grass family belongs to a major subgroup of flowering plants called Liliopsida (Monocotyledoneae of earlier systems). Informally called "monocots," these plants have a single cotyledon, scattered vascular bundles in the stem, a circular pattern of vascular tissue in the roots, parallel venation in the leaf blade, and floral parts in 3's or multiples of 3. All of these features are usually seen in most grasses, except for the characters of the flower. They are highly reduced and are not easily interpreted. More about them later.

The closest relatives of the grasses belong to little-known tropical and subtropical plant families, such as Flagellariaceae, Centrolepidaceae, and Restionaceae. None of them is represented in the flora of North America. There are, however, two other families whose plants resemble grasses, at least superficially. The rushes (Juncaceae) and sedges (Cyperaceae) occur widely in North America. Rushes are rather easily distinguished from grasses by their 3-parted calyx and corolla and by their fruit, a many-seeded capsule. Sedges require closer inspection. Table 1 presents a comparison of typical members of the two families.

TECHNICAL FAMILY DESCRIPTIONS

GRAMINEAE (THE GRASS FAMILY)

Annual or perennial herbs, sometimes + woody in the canes, reeds, and ornamental bamboos. Roots generally fibrous and adventitious at maturity; rhizomes and stolons frequent, some with bulbs. Stems generally round, sometimes flattened, erect to prostrate; nodes swollen, solid; internodes generally hollow. Leaves alternate, 2-ranked, simple, generally elongate and differentiated into a blade with parallel veins, a sheath (typically open with its edges meeting or overlapping slightly) that encircles the stem, and ligule (a membranous flap or series of hairs at inner apex of sheath). Inflorescence complex, consisting generally of numerous basic units, the spikelets, which are themselves tiny spikes. Spikelets round or flattened (dorsally or laterally) in cross-section; generally consisting of 2 sterile, overlapping, basal bracts (the first and second glume) and one or more florets (flowers and subtending bracts, the palea and lemma), these 2-ranked on an internal axis (the rachilla); glumes equal or unequal in size and shape, awned or awnless; 0 to many-nerved; lemmas similar to glumes in appearance and texture or quite dissimilar, awned or awnless, 0 to many-nerved; palea generally thin, transparent, awnless, 2-nerved, and \pm enclosed by the lemma; floret base sometimes

forming a sharp-pointed and/or hairy callus; breaking apart at maturity either below or above the glumes. Spikelets borne in secondary inflorescences (spikes, spike-like panicles, panicles, etc.), rarely solitary. Flowers generally bisexual, minute, wind-pollinated, the perianth reduced to 2 or 3 microscopic structures (lodicules); stamens [1] 3 [6], anthers generally comparatively large; stigmas generally 2, typically dissected and feather-like; ovary 1-chambered. Fruit a caryopsis (or grain), with the fruit wall + completely fused with the seed coat of the single seed inside. 651 genera; + 10,000 species; cosmopolitan, probably the most frequently encountered flowering plants, found in a wide variety of habitats and on all continents, including Antarctica.

CYPERACEAE (THE SEDGE FAMILY)

Perennial [rarely annual] herbs of wet and marshy sites. Plants often with creeping rhizomes. Stems generally with solid internodes and often 3-sided. Leaves from basal tufts or cauline and 3-ranked; sheaths usually closed; ligule generally absent. Flowers minute, bisexual or unisexual (the species generally monoecious), spirally or distichously arranged in tiny spikes. Each flower is subtended by a small bract (often called a glume). The spike of reduced flowers and subtending bracts form the spikelet, which are themselves arranged in panicles, umbels, or spike-like inflorescences. Perianth of bristles, hairs, scales, or absent. Stamens 3 [rarely 1 or 6]. Carpels 2 or 3, united, 1-ovuled, with as many style-branches as carpels; ovary superior. Fruit an achene or nutlet, lenticular or 3-sided, sometimes enclosed in a membranous sac (perigynium). 90 genera; 4000 species; widespread, particularly in the cool temperate and subarctic regions. Of little economic importance; a few are edible and some are grown as ornamentals.

JUNCACEAE (THE RUSH FAMILY)

Perennial or annual herbs, from erect or horizontal rhizomes. Leaves generally basal, linear, sheathing at base, sheaths generally open; blades sometimes absent. Flowers small, green, actinomorphic, bisexual or unisexual (species diocecious); in heads, panicles, or corymbs. Tepals 6 (in two sets of 3), sepaloid. Stamens 6 [rarely 3]. Carpels 3, united, unilocular; placentation axillary or parietal; style 1; stigmas 3, brush-like; ovary superior. Fruit a capsule. 9 genera; 400 species; largely cool temperate and subarctic damp and wet sites. Of no direct economic importance; a few are grown as ornamentals.

CENTROLEPIDACEAE

Annual or perennial grass-like, rush-like, or moss-like herbs. Leaves linear, bristle-like, cauline or in basal rosettes. Flowers tiny, unisexual, in cymose false inflorescences (pseudanthia) of 1 or 2 male flowers and 2-many female flowers. Pseudanthia terminal, spikelike, and subtended by 2-several glume-like bracts. Male flowers reduced to 1 stamen. Female flowers reduced to 1 carpel, the ovary unilocular. Fruit dehiscent or indehiscent, 1-seeded. 5 genera; 30 species; found primarily in New Zealand, Australia, and the southern tip of South America. None occur in North America.

RESTIONACEAE

Perennial, rhizomatous herbs. Stem internodes solid or hollow. Leaves reduced to open sheaths, blades and ligules generally absent. Flowers small, regular, unisexual (species dioecious). Perianth absent or scale-like. Stamens 3 [rarely 1 or 4]. Carpels 1 or 3, united, the ovary superior. Fruit an achene, nut, or capsule. 38 genera; 400 species; found primary in the southern hemisphere [1 sp. in Viet Nam], especially well-represented in the Cape region of South Africa where 180 ssp. are endemic! Of little economic importance; a few are used for thatch and to make brooms. None occur in North America.

FLAGELLARIACEAE

Glabrous vines. Leaves spirally arranged, blades parallel-veined and terminating in a tendril; ligule absent. Flowers small, regular, bisexual, 3-merous, wind-pollinated, in terminal bracteate panicles. Tepals 3 + 3. Stamens 6. Carpels 3, united; ovary trilocular, superior. Fruit a drupe. 1 genus; 4 species; found primarily in the Old World tropics and islands of the South Pacific. Of little economic importance; one species is used in basket-making in Asia. None occur in North America.

JOINVILLEACEAE

Erect, perennial, rhizomatous herbs. Stem internodes hollow. Leaves grass-like, spirally arranged; ligulate and auriculate. Flowers bisexual, 3-merous, in terminal bracteate panicles. Tepals 3 + 3. Stamens 6. Carpels 3, united; ovary 3-locular, superior. Fruit a drupe. 1 genus; 2 species; restricted to Malaysia and the islands of the Pacifc. The family was only recently separated out of Flagellariaceae. Of no economic importance.

A COMPARISON OF GRASSES AND SEDGES

Feature	Grasses	Sedges		
Stems:				
Shape in x-section Internodes	Round [flat]* Hollow [solid]	Triangular [round] Solid		
Leaves:				
# of Ranks Sheaths Ligule	2 Present	3 Open [closed]Closed Absent		
Spikelets:				
Bract insertion Bracts per flower	Distichous 2	Distichous, spiral 1		
Flowers:				
Perianth Stamen number Anthers Stigmas Carpels Pollen shed as	2 [3] lodicules 3 [6, 1, many] Monads	0-6 bristles 1-3 VersatileBasifixed 23 2 [3]3 [2] Pseudomonads		
Fruit and Seed:				
Fruit type Embryo position	Caryopsis Lateral	Achene Central		
Chromosomes:				
Centromeres	Monocentric	Diffuse		

^{*} Character states in brackets indicate the less typical situation.

SELECTED REFERENCES

- Bessey, C. E. 1915. The phylogeny of the grasses. Rept. Michigan Acad. Sci. 19: 239-245.
- Bruhl, J. J. 1995. Sedge genera of the world: relationships and a new classification of the Cyperaceae. Australian Syst. Bot. 8: 125-305.
- Bruhl, J. J., L. Watson, & M. J. Dallwitz. 1992. Genera of Cyperaceae: interactive identification and information retrieval. Taxon 41: 225-234.
- Chanda, S. 1966. On the pollen morphology of the Centrolepidaceae, Restionaceae and Flagellariaceae, with special reference to taxonomy. Grana Palynol. 6: 355-415.
- Clayton, W. D. 1981. Evolution and distribution of grasses. Ann. Missouri Bot. Gard. 68: 5-14.
- Clifford, H. T. 1970. Monocotyledon classification with special reference to the origin of the grasses (Poaceae). Bot. J. Linn. Soc., Suppl. 1: 35-34.
- Crepet, W. L. & G. D. Feldman. 1991. The earliest remains of grasses in the fossil record. American J. Bot. 78(7): 1010-1014.
- Cutler, D. F. 1966. Anatomy and taxonomy of the Restionaceae. Jodrell Lab. Notes 4: 1-25.
- Cutler, D. F. & H. K. Airy Shaw. 1965. Anarthriaceae and Ecdeiocolaceae: two new monocotyledonous families separated from the Restionaceae. Kew Bull. 19: 489-499.
- Daghlian, C. P. 1981. A review of the fossil record of the monocotyledons. Bot. Rev. 47(4): 517-555.
- Doyle, J. J. et al. 1992. Chloroplast DNA inversions and the origin of the grass family (Poaceae). Proc. Natl. Acad. Sci., U. S. A. 89: 7722-7726.
- Gardener, J. S. 1886. Fossil grasses. Proc. Geol. Assoc. London 9: 433-454.
- Gonzalez-Elizondo, M. S. & P. A. Peterson. 1997. A classification of and key to the supraspecific taxa in *Eleocharis* (Cyperaceae). Taxon 46(3): 433-449.
- Hamann, U. 1975. Neue Untersuchungenen zur Embryologie und Systematik der Centrolepidaceae. Bot. Jahrb. Syst. 96: 154-191.
- Hurd, E. G., S. Goodrich, & N. L. Shaw. 1994. Field guide to Intermountain rushes. Gen. Tech. Rep. INT-306. U. S. Dept. Of Agric. Forest Service. Intermountain Res. Sta. Ogden, UT. 56 pp.
- Kircher, P. 1986. Untersuchungen zur Bluten- und Infloreszenmorphologie, Embryologie und Systematik der Restionaceae im Vergeich mit Gramineen und verwandten Familien. J. Cramer. Berlin. 219 pp.
- Koyama, T. 1961. Classification of the family Cyperaceae. Pt. 1. J. Fac. Sci. Univ. Tokyo, Sect. 3, Bot. 8: 37-148.
- Koyama, T. 1962. Classification of the family Cyperaceae. Pt. 2. J. Fac. Sci. Univ. Tokyo, Sect. 3, Bot. 8: 149-278.

- Lee, D. W., et al. 1975. Serological evidence on the distinctness of the monocotyledonous families Flagellariaceae, Hanguanaceae and Joinvilleaceae. J. Linn. Soc. Bot. 70: 77-81.
- Linder, H. P. 1987. The evolutionary history of the Poales/Restionales -- a hypothesis. Kew Bull. 42(2): 297-318.
- Linder, H. P. & I. K. Ferguson. 1985. On the pollen morphology and phylogeny of the Restionales and Poales. Grana 24: 65-76.
- Linder, H. P. & P. J. Rudall. 1993. The megagametophyte in *Anarthria* (Anarthriaceae, Poales) and its implication for the phylogeny of the Poales. American J. Bot. 80: 1455-1464.
- Newell, T. K. 1969. A study of the genus *Joinvillea* (Flagellariaceae). J. Arnold Arbor. 50: 527-555.
- Sharma, M. L. 1979. Some considerations on the phylogeny and chromosomal evolution in grasses. Cytologia 44: 679-685.
- Smithson, 1957. The comparative anatomy of the Flagellariaceae. Kew Bull. 1956: 491-501.
- Thomasson, J. R. 1980. Paleoagrostology: an historical review. Iowa State J. Res. 54(3): 301-317.
- Tomlinson, P. B. & A. C. Smith. 1970. Joinvilleaceae, a new family of monocotyledons. Taxon 19: 887-889.
- Torrey, J. 1836. Monograph of North American Cyperaceae. Ann. Lyceum Nat. Hist. New York 3: 239-448.
- Tucker, G. C. 1987. The genera of Cyperaceae in the southeastern United States. J. Arnold Arbor. 68: 361-445.
- Ziegenspeck, H. 1939. Die phylogenie der Glumiflorae. Bot. Arch. 39: 177-205.

2.02 - SYSTEMS OF CLASSIFICATION [Historical Survey]

"The Grasses are dreadfully difficult and systematically a chaos of imperfect descriptions, erroneous identifications, confused synonymy and imbecile attempts. We have upwards of a century of collections and not an attempt at a classification. Each Botanist in his own country has worked at his sweet will in ignorance of his predecessors' and contemporaries' work, with imperfect materials and often no books -- 'Hinc illae lachrymae.' (Sir Joseph Dalton Hooker)

The purpose of this section is to outline the major attempts in the last four hundred years to classify grasses. While there have been important conceptual advances since Burbidge made his comment of thirty years ago, it still has a ring of truth about it.

Jacob Theodore (Tabernaemontanus) (1590) wrote thirty-five chapters on grasses, including rushes, arrow-grasses, plantains, horsetails, and some caryophylls. His classification was based upon general plant form and a plant's habitat.

Caspari Bauhin (1623) recognized twenty-four groups based upon form of the grass plant. His was an unnatural assemblages of plants, lumping together grasses and cattails.

Joseph Pitton de Tournefort (1694) was the first to present an index to technical and common names. His Class 15 included grasses, chenopods, nettles, smartweeds, sedges, etc. Section 3 was devoted to grasses and it contained 9 "genera" (1-7, cereals; 9, reed grasses; 8, others).

John Ray (1703) urged simplification in classification and the use of easier features. He also encouraged the use of dichotomous keys.

Pier Antonio Micheli (1728) developed a system based upon the completeness of flower and spikelet. He found the "lost petals" of the grass flower (lodicules).

Carolus Linnaeus (1753) recognized thirty-eight genera of grasses (one of them actually a sedge!). They were put into six of his artificial groups.

Robert Brown (1814) was one of England's most influential botanists. He was the first to distinguish the two great groups of grasses: Panicoideae and Pooideae.

Palisot de Beauvois (1812) recognized two "families," divided into tribes and then into sections. He recognized 210 genera based upon glume arrangement and disarticulation. His name appears frequently as the author of numerous genera and species of grasses.

Carl Bernard von Trinius (1822) prepared a summary of grass classification and synonymy, in which he listed 2457 species, along with their author

and place of publication.

George Bentham (1881) was one of Victorian England's most famous botanists. He often collaborated with Joseph Dalton Hooker. Bentham's system was used by A. S. Hitchcock and others until mid-20th century.

Series A: Paniceae

Tribe: Paniceae Tribe: Maydeae Tribe: Oryzeae Tribe: Tristegineae Tribe: Andropogonea

Tribe: Andropogoneae

Series B: Poaceae
Tribe: Phalarideae
Tribe: Agrostideae
Tribe: Aveneae
Tribe: Chlorideae
Tribe: Festuceae

Tribe: Hordeae Tribe: Bambuseae

Eduard Hackel (1887) developed a system much like that of Bentham. He recognized 13 tribes and 313 genera.

George Valentine Nash (1909) wrote major portions of the treatment of Gramineae for the "North American Flora." He recognized 13 tribes, but did not place them in subfamilies.

Tribe: Maydeae Tribe: Andropogoneae

Tribe: Zoysieae
Tribe: Tristegineae
Tribe: Paniceae
Tribe: Oryzeae
Tribe: Phalarideae
Tribe: Agrostideae
Tribe: Aveneae
Tribe: Festuceae
Tribe: Chlorideae

John Bews was the author of "The World's Grasses: Their Differentiation, Distribution, and Ecology." Although quite dated, it remains a useful reference. In it, he recognized two subfamilies, 15 tribes, 481 genera, and 5871 species.

Subfamily: Pooideae

Tribe: Bambuseae Tribe: Phareae Tribe: Festuceae Tribe: Aveneae Tribe: Chlorideae Tribe: Hordeae Tribe: Agrostideae Tribe: Zoysieae Tribe: Phalarideae Tribe: Arundinelleae

Tribe: Oryzeae
Subfamily: Panicoideae

Tribe: Melinideae Tribe: Paniceae Tribe: Andropogoneae Tribe: Maydeae

Nikolai Pavlovic Avdulov was a famous Russian botanist. His 1931 cytological examination of Gramineae is considered by many the foundation of the "new agrostology." In it, he listed two series and 13 tribes.

Series A

Tribe: Maydeae Tribe: Andropogoneae Tribe: Zoysieae Tribe: Tristagineae Tribe: Paniceae Tribe: Oryzeae

Series B

Tribe: Phalarideae Tribe: Agrostideae Tribe: Aveneae Tribe: Chlorideae Tribe: Festuceae Tribe: Hordeae Tribe: Bambuseae

Romain Roschevicza, another Soviet botanist, in 1937 recognized two subfamilies (Poatae and Sacchariferae), 5 series, and 28 tribes.

Subfamily: Poatae

Series: Bambusiformes Tribe: Bambuseae Tribe: Phareae Series: Phragmitiformes Tribe: Centotheceae Tribe: Arundineae Tribe: Oryzeae Tribe: Stipeae Tribe: Brachypodieae Tribe: Unioleae Series: Festuciformes Tribe: Festuceae Tribe: Nardeae Tribe: Aveneae Tribe: Phalarideae Tribe: Agrostideae
Subfamily: Sacchariferae

Series: Eragrostiformes Tribe: Eragrosteae Tribe: Pappophoreae Tribe: Chlorideae Tribe: Sporoboleae Series: Paniciformes Tribe: Paniceae

Tribe: Melinideae Tribe: Zoysieae Tribe: Andropogoneae Tribe: Maydeae

Albert Spear Hitchcock, Curator of Grasses at the Smithsonian Institution, was the most influential American agrostologist of the early 20th century. The system he used in "The Manual" (1935, 1951) has two subfamilies and 14 tribes and it closely follows the thinking of George Bentham. This is the scheme that I learned when I took agrostology.

Subfamily: Festucoideae

Tribe: Bambuseae Tribe: Festuceae Tribe: Hordeae Tribe: Aveneae Tribe: Agrostideae

Tribe: Zoysieae Tribe: Chlorideae Tribe: Phalarideae Tribe: Oryzeae Tribe: Zizanieae **Subfamily: Panicoideae**

Tribe: Melinideae

Tribe: Paniceae

Tribe: Andropogoneae Tribe: Tripsaceae

Robert Pilger and Eva Potzal, two eminent German botanists, prepared portions of a world wide treatment of grasses. They recognized nine subfamilies, 34 tribes, and 555 genera. Although the individual generic treatments are still useful, their system is not widely accepted.

Alan Beetle, an American botanist, was one of the first to support more than two subfamilies for our North American material. He recognized:

Subfamily: Bambusoideae Subfamily: Pharoideae Subfamily: Festucoideae Subfamily: Panicoideae

Tsugo Tateoka is one of Japan's most influential botanists. His system, published in 1957, recognized five subfamilies:

Subfamily: Pharoideae Subfamily: Pooideae Subfamily: Eragrostoideae Subfamily: Panicoideae Subfamily: Arundinoideae

Henri Prat (1960), in his world-wide survey of grasses, recognized six subfamilies, 26 tribes, 403 genera, and 6250 species.

G. Ledyard Stebbins and Beecher Crampton were both professors at the University of California at Davis. In 1961, they published a provisional scheme for North American grasses. It turned out to be very influential.

Subfamily: Bambusoideae

Tribe: Arundinarieae Subfamily: Oryzoideae Tribe: Phareae

Tribe: Olyreae Tribe: Ehrharteae Tribe: Oryzeae Tribe: Zizanieae

Subfamily: Arundinoideae

Tribe: Arundineae Tribe: Danthonieae Tribe: Unioleae Tribe: Aristideae

Subfamily: Festucoideae Tribe: Ampelodesmeae

Tribe: Stipeae Tribe: Brachyelytreae Tribe: Nardeae Tribe: Monermeae Tribe: Meliceae Tribe: Diarrheneae Tribe: Festuceae Tribe: Hordeae Tribe: Aveneae

Subfamily: Eragrostoideae

Tribe: Aeluropideae Tribe: Spartineae

Tribe: Pappophoreae Tribe: Eragrosteae Tribe: Chlorideae Tribe: Zoysieae **Subfamily: Panicoideae** Tribe: Paniceae Tribe: Andropogoneae

Henri Jacques-Felix, in his 1962 survey of tropical African grasses, recognized twelve subfamilies and 35 tribes. His names employ the endings of his native French.

Subfamily: Olyroide
Subfamily: Bambusoide
Subfamily: Streptogynoide
Subfamily: Stipoide
Subfamily: Oryzoide
Subfamily: Ehrhartoide
Subfamily: Zizanioide
Subfamily: Centothecoide
Subfamily: Arundinoide
Subfamily: Festucoide
Subfamily: Chloridoide
Subfamily: Panicoide

Frank Gould, in the 1968 edition of his textbook "Grass Systematics," recognized six subfamilies, 23 tribes, 122 genera, and 1083 native species in the United States. Gould, a faculty member at Texas A & M University, was one of the most productive agrostologists in this country.

Subfamily: Festucoideae Subfamily: Panicoideae Subfamily: Eragrostoideae Subfamily: Bambusoideae Subfamily: Oryzoideae Subfamily: Arundinoideae

Charles Edward Hubbard, of the Royal Botanic Garden at Kew, was the dean of the British agrostologists, with an encyclopedic knowledge of the world's grasses. He did not follow the increasingly popular view of recognizing four to six subfamilies by listing only two (Festucoideae and Panicoideae) and 27 tribes.

W. Derek Clayton and **Stephen Andrew Renvoize** are the authors of one of the two major world-wide surveys to be published in the past few years. The descriptions of genera are relatively brief. The taxa are arranged systematically and keys to tribes and genera are provided.

Subfamily: Bambusoideae

Tribe: Bambuseae Tribe: Anomochloeae Tribe: Streptochaeteae Tribe: Olyreae

Tribe: Olyreae Tribe: Parianeae Tribe: Phareae

Tribe: Phaenospermateae

Tribe: Streptogyneae Tribe: Oryzeae Tribe: Phyllorachideae Tribe: Ehrharteae Tribe: Diarrheneae Tribe: Brachyelytreae

Subfamily: Pooldeae

Tribe: Nardeae Tribe: Lygeae Tribe: Stipeae Tribe: Poeae Tribe: Hainardieae Tribe: Meliceae Tribe: Aveneae Tribe: Bromeae Tribe: Triticeae

Subfamily: Centothecoideae

Tribe: Centotheceae **Subfamily: Arundinoideae**

Tribe: Arundineae
Tribe: Thysanolaeneae
Tribe: Micraireae
Tribe: Aristideae

Subfamily: Chloridoideae

Tribe: Pappophoreae Tribe: Orcuttieae Tribe: Eragrostideae Tribe: Leptureae

Tribe: Cynodonteae **Subfamily: Panicoideae**Tribe: Paniceae

Tribe: Paniceae Tribe: Isachneae Tribe: Hubbardieae Tribe: Eriachneae

Tribe: Steyermarkochloeae Tribe: Arundinelleae Tribe: Andropogoneae

N. N. Tzvelev is Russia's most prominent agrostologist. In 1989, he published a comprehensive review of the family in which he recognized only two subfamilies and 28 tribes. Note that he is the only recent author who goes back to the once common two subfamily view.

Subfamily: Bambusoideae

Tribe: Arundineae Tribe: Bambuseae Tribe: Dendrocalameae

Subfamily: Pooideae

Tribe: Brachypodieae
Tribe: Triticeae
Tribe: Bromeae
Tribe: Poeae
Tribe: Phleeae
Tribe: Meliceae
Tribe: Diarrheneae
Tribe: Brachyelytreae
Tribe: Ampelodesmeae

Tribe: Stipeae
Tribe: Nardeae
Tribe: Oryzeae
Tribe: Ehrharteae
Tribe: Centosteceae
Tribe: Arundineae
Tribe: Thysanolaeneae
Tribe: Aristideae
Tribe: Cynodonteae

Tribe: Paniceae Tribe: Andropogoneae

Leslie Watson and **Michael J. Dallwitz** in 1992 published "The Grass Genera of the World," a compendium that provides very detailed descriptions. The genera are arranged alphabetically and there are no keys. Their system of classification appears below:

Subfamily: Stipoideae

Tribe: Nardeae Tribe: Ampelodesmeae

Tribe: Stipeae
Tribe: Brachyelytreae
Subfamily: Pooideae
Tribe: Triticeae

Tribe: Brachypodieae Tribe: Bromeae Tribe: Aveneae Tribe: Poeae Tribe: Meliceae

Subfamily: Bambusoideae

Tribe: Oryzeae Tribe: Diarrheneae Tribe: Phareae Tribe: Bambuseae

Subfamily: Centothecoideae Tribe: Centotheceae

Subfamily: Arundinoideae Tribe: Arundineae Tribe: Danthonieae

Tribe: Aristideae **Subfamily: Chloridoideae** Tribe: Pappophoreae

Tribe: Orcuttieae Main Chloridoid Assemblage

Subfamily: Panicoideae

Tribe: Paniceae Tribe: Andropogoneae Tribe: Maydeae

Grass Phylogeny Working Group. The GPWG is composed of eight individuals located here in the United States, and in South Africa and Switzerland. The system is based on a cladistic analysis of six molecular sequence data sites, chloroplast restriction site data, and more traditional morphological data.

Subfamily: Bambusoideae Tribe: Bambuseae Tribe: Olyreae

Subfamily: Anomochlooideae Tribe: Streptochaeteae **Subfamily: Pharioideae** Tribe: Phareae

Subfamily: Ehrhartoideae Tribe: Oryzeae Tribe: Ehrharteae

Subfamily: Centothecoideae

Tribe: Centotheceae Tribe: Thysanolaeneae

Subfamily: Pooideae

Tribe: Triticeae Tribe: Brachypodieae Tribe: Broméae Tribe: Poeae

Tribe: Ampelodesmeae

Tribe: Meliceae Tribe: Stipeae Tribe: Nardeae Tribe: Brachyelytreae Tribe: Diarrheneae **Subfamily: Arundinoideae** Tribe: Arundineae

Subfamily: Danthonioideae Tribe: Danthonieae

Subfamily: Aristidoideae Tribe: Aristideae

Subfamily: Chloridoideae Tribe: Pappophoreae

Tribe: Orcuttieae Tribe: Cynodonteae Tribe: Eragrostideae Tribe: Leptureae Subfamily: Panicoideae

Tribe: Paniceae Tribe: Andropogoneae

Flora of North America. 'The Manual of Grasses' will

soon be replaced by two volumes in the 'Flora of North America' series. One volume has been published; the second is to appear in 2006. Mary Barkworth, an agrostologist at Utah State Univ., is the senior editor. The unpublished scheme that will be used appears below.

Subfamily: Bambusoideae

Tribe: Bambuseae Tribe: Phareae **Subfamily: Oryzoideae** Tribe: Oryzeae

Subfamily: Pooideae Tribe: Ehrharteae

Tribe: Diarrheneae Tribe: Brachyelytreae Tribe: Nardeae Tribe: Stipeae

Tribe: Poeae Tribe: Hainardieae Tribe: Meliceae Tribe: Aveneae Tribe: Bromeae Tribe: Brachypodieae Tribe: Triticeae

Subfamily: Centothecoideae Tribe: Centotheceae

Subfamily: Arundinoideae

Tribe: Arundineae Tribe: Danthonieae

Tribe: Aristideae Subfamily: Chloridoideae Tribe: Pappophoreae

Tribe: Orcuttieae Tribe: Cynodonteae

Subfamily: Panicoideae

Tribe: Paniceae

Tribe: Andropogoneae

THE SYSTEM-BUILDERS

1880 C. O. Hartz (Germany)

George Bentham (England) 1881

Eduard Hackel (Germany) 1887

1929 John Bews (England)

1931 1934

Nikolai P. Avdulov (Russia) Charles E. Hubbard (England)

1936 Henri Prat (France)

Romain Roshevitz (Russia) 1946

Robert Pilger (Germany) Alan Bettle (United States) 1954

1955

Tsugo Tateoka (Japan) 1957

G. Ledyard Stebbins & Beecher Crampton 1961

(United States) W. Derek Clayton & Stephen A. Renvoize 1986

(England)

1988 Leslie Watson & Michael J. Dallwitz (Australia)

Nikolai Tzvelev (Russia) 1989

Grass Phylogeny Working Group 2001

SELECTED REFERENCES

Avdulov, N. P. 1931. Karyo-systematische Untersuchungen der Familie Gramineen. Bull. Appl. Bot. Genet. Plant Breed. Suppl. 44: 1-428. [In Russian. German Zusammenfassung, pp. 353-425].

Bauhin, C. 1623. Pinax theatri botanici.... Impensis Joannis Regis. Basileae. 522 pp.

- Beauvois, A. M. F. J. Palisot de. 1812. Essai d'une nouvelle agrostographie ou nouveaux genres des Graminées. Fain. Paris, France. 182 pp.
- Beetle, A. A. 1955. The four subfamilies of the Gramineae. Bull. Torrey Bot. Club 82: 196, 197.
- Bentham, G. 1881. Notes on Gramineae. J. Linn. Soc. Bot. 19: 14-134.
- Bews, J. G. 1929. The world's grasses: their differentiation, distribution, economics, and ecology. Longmans, Green & Co., Ltd. London, England. 408 pp.
- Bowden, W. M. 1960. An experimental taxonomist examines the classification of grasses. Rev. Canadian Biol. 19: 279-292.
- Bowden, B. N. 1965. Modern grass taxonomy. Outlook Agric. 4: 243-253.
- Brown, R. 1814. A voyage to Terra Australia. Two volumes. G. & W. Nicol. London, England.
- Butzin, F. 1969. Zur Klassifizierung der Spelzenformen bei den Gramineen mit besonderer Berucksichtigung der begrannten Spelzen. Willdenowia 5: 445-470.
- Butzin, F. 1973. Die Namen der Supragenerischen Einheiten der Gramineae (Poaceae). Willdenowia 7: 113-168.
- Campbell, C. S. & E. A. Kellogg. 1987. Sister group relationships of the Poaceae. In, Soderstrom, T. R. et al. Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 217-224.
- Clark, L. G. & E. J. Judziewicz. 1996. The grass subfamilies Anomochlooideae and Pharoideae (Poaceae). Taxon 45(4): 641-645.
- Clark, L. G., W. Zhang, & J. F. Wendel. 1995. A phylogeny of the grass family (Poaceae) based upon ndhF sequence data. Syst. Bot. 20(4): 436-460.
- Clayton, W. D. 1981. Early sources of tribal names in Gramineae. Kew Bull. 36: 483-485.
- Clayton, W. D. & S. A. Renvoize. 1986. Genera graminum: grasses of the world. Kew Bull. Addt. Series XIII. Royal Botanic Garden, Kew. London, England. 389 pp.
- Clayton, W. D. & S. A. Renvoize. 1992. A system of classification for the grasses. <u>In</u>, Chapman, G. P. (Editor). Grass evolution and domestication. Cambridge Univ. Press. Cambridge, England. Pp. 338-353.
- Clifford, H. T. 1965. The classification of the Poaceae: a statistical study. Queensland Pap. (Botany) 4(15): 243-253.
- Clifford, H. T. & D. W. Goodall. 1967. A numerical contribution to the classification of the Poaceae. Australian J. Bot. 15(3): 499-519.
- Clifford, H. T., W. T. Williams, & G. W. Lance. 1969. A further numerical contribution to the classification of the Poaceae. Australian J. Bot. 17: 119-131.
- Duvall, M. R. & B. R. Morton. 1996. Molecular phylogenetics of Poaceae: an expanded analysis of

- rbcL sequence data. Mol. Phylogenet. Evol. 5: 352-358.
- Gould, F. W. 1968. Grass systematics. McGraw-Hill Book Co. New York, NY. P. 97.
- Grass Phylogeny Working Group. 2001. Phylogeny and subfamilial classification of the grasses (Poaceae). Ann. Missouri Bot. Gard. 88(3): 373-457.
- Hackel, E. 1887. Gramineae. In, Engler, A. & K. Prantl. Die Natürlichen Pflanzenfamilien.
- Hamby, R. K. & E. A. Zimmer. 1988. Ribosomal RNA sequences for inferring phylogeny within the grass family (Poaceae). Pl. Systm. Evol. 34: 393-400.
- Harz, C. O. 1880. Beitrage zur Systematik der Gramineen. Linnaea 43: 1-30.
- Hayek, A. 1929. Zur Systematik der Gramineen. Oster. Bot. Zeit. 74: 249-255.
- Herrera, A. Y. 1994. Perspectiva historica de la classification de la familia Poaceae. Phytologia 77(1): 8-22.
- Hilu, K. W. & K. Wright. 1982. Systematics of Gramineae: a cluster analysis study. Taxon 31: 9-36.
- Hitchcock, A. S. 1951. Manual of the grasses of the United States. Second edition, revised by A. Chase. Misc. Publ. 200. U. S. Dept. of Agric. Washington, D. C. 1051 pp.
- Hubbard, C. E. 1973. Gramineae. <u>In</u>, Hutchinson, J. The families of flowering plants. Oxford Univ. Press. XXX
- Jacques-Felix, H. 1962. Les Graminées (Poaceae) d'Afrique tropicale. I. Généralités, classification description des genres. Inst. Rech. Agron. Trop. et Cult. Viv. Bull. Sci. No. 8.
- Kellogg, E. A. & C. S. Campbell. 1987. Phylogenetic analyses of the Gramineae. In, Soderstrom, T. R. et al. Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 310-322.
- Kunth, C. S. 1829-1835. Revision des Graminées. Luetiae Parisiorum. Paris, France.
- Kunth, C. S. 1883. Agrostographia. Synoptica sive Enumeratio Graminearum omnium Hucusque Cognitarium adjectes characteribus, differentiis et synonymie.
- Linnaeus, C. 1753. Species plantarum. Facsimile edition. Ray Society. London, England. 560 pp.
- Mehra, K. L. & E. Anderson. 1959. Character association analysis: its use in grass taxonomy. Ninth Intern. Bot. Congr. 2: 258, 259.
- Micheli, P. A. 1728. Nova plantarum genera juxta Tournefortii methodum disposita.... Bernardi Paperinii. Florence. 108 pp.
- Nash, G. V. 1909. Poaceae. North American Flora 17(1): 77, 78.
- Parodi, L. R. 1961. La taxonomia de las gramineas Argentinas a la luz de las investigaciones mas recientes. Recent Advances in Botany 1: 125-130.

Pilger, R. 1954. Das System der Gramineen unter Ausschluss der Bambusoideae. Bot. Jahrb. 76: 281-384.

Prat, H. 1936. La systématique des Graminées. Ann. Sci. Nat. Bot., Ser. 10, 18: 165-258.

Prat, H. 1960. Revue d'agrostologie vers une classification naturelle des Graminées. Bull. Soc. Bot. France 107: 32-79.

Raspail, F. V. 1824. Essai d'une classification generale des graminees fondee sur l'etude physiologique des characters de cette famille. Annales des Sciences Naturelles 4: 423-451; 5: 287-311; 5: 433-460.

Ray, J. 1703. Methodus plantarum emendata ... accedit Graminum, Juncorum et Cyperorum specialis eddem auctore. Smith & Walford. London, England. 202 pp.

Roshevits, R. Y. 1937. Zlaki. Vvedenie v Izuchenie Kormovykh i Khlebnykn Zlakov.

Roshevits, R. Y. 1946. Systematics of the grasses in relation to their evolution. Collection of Scientific Works. Komarov Inst. Acad. Sci. U. S. S. R. Pp. 25-40.

Scheuchzer, J. 1719. Agrostographia sive graminum, juncorum, cyperorum, cyperoidum, iisque affinium historia. Bodmerianis. Zurich. 512 pp.

Soderstrom, T. R. 1981. The grass subfamily Centothecoideae. Taxon 30: 614, 615.

Soreng, R. J. & J. I. Davis. 1995. Phylogenetic structure of the Poaceae: results from morphology, molecules, and total evidence. American J. Bot. 82(Suppl.): 163 (abstract).

Stebbins, G. L. & B. Crampton. 1961. A suggested revision of the grass genera of temperate North America. Recent Advances in Botany 1: 133-145.

Tateoka, T. 1957. Miscellaneous Papers on the Phylogeny of Poaceae. X. Proposition of a new phylogenetic system of Poaceae. J. Japanese Bot. 32: 275-287.

Tournefort, J. P. de. 1694. Elémens de botanique:, ou méthode pour connoitre les plantes. Three volumes. De L'Imprimerie Royale. Paris, France.

Trinius, C. B. 1822. Clavis agrostographiae antiquioris. Coburg.

Trinius, C. B. 1828. Species graminum. Iconibus et descriptionibus illustravit. Vol. 1. Imp. Acad. Imp. Scient. Petropoli. 680. Pars. 2. Icones.

Tzvelev, N. N. 1989. The system of grasses (Poaceae) and their evolution. Bot. Rev. 55(3): 141-203.

Vasey, G. 1883. The grasses of the United States: being a synopsis of the tribes and genera, with descriptions of the genera, and a list of the species. Special Report No. 63. U. S. Dept. of Agriculture. Washington, D. C. 47 pp.

Watson, L. & M. J. Dallwitz. 1992. The grass genera of the world. CAB International. Wallingford, U. K. 1024 pp.

Watson, L., H. T. Clifford, & M. J. Dallwitz. 1985. The classification of the Poaceae: subfamilies and supertribes. Australian J. Bot. 33: 433-484.

2.03 - THE BASIS OF MODERN SYSTEMS

The great agrostologists of the past were limited by the instruments and the technology of their period, just as we are today. For the most part, botanists had to rely upon the features that were seen under the light microscope. At first, these devices were relatively crude. You can still see Linnaeus's microscope if you visit his home in Sweden. Workers of the 19th century and the early part of the 20th based their systems of classification primarily on features that could be seen with increasingly better instruments, especially the dissecting microscope. They focused their attention on the morphological details of stems, leaves, the inflorescence, and the spikelet.

By the time A. S. Hitchcock had completed the first edition of "The Manual" in 1935, botanists in various countries around the world had begun using new techniques to understand the relationships in the family and to suggest new systems of subfamilies and tribes. These more modern systems of classification relied heavily upon a series of what we might call **microcharacters** -- anatomical, cytological, genetic, and chemical traits of grasses. The result is a system of subfamilies, tribes, and genera that is strikingly different from the ones produced by the great figures of the 19th and early 20th centuries. General reviews of these new features upon which the modern classification of the family rests are found in Auquier (1963), Avdulov (1931), Clifford (1969), and Gould & Shaw (1983).

A CHRONOLOGY

- 1892 Embryo structure
- 1926 Chromosome complements
- 1931 Chromosome karotypes
- 1932 Leaf epidermis
- 1953 Root hair development
- 1957 Persistent nucleoli
- Shoot apex and meristem activity
- 1957 1957 Embryo structure
- 1958 Germination responses to IPC
- 1958 Leaf anatomy
- Nodal pulvini and internode structure 1959
- 1960 Culm anatomy
- Seedling leaf 1961
- Antigen/antibody reactions 1961
- 1962 Starch grains
- 1968 Starch versus fructosan accumulation
- 1969 Underground seedling organs
- 1974 Photosynthetic pathways/leaf anatomy
- Flavonoid patterns Stomatal insertion 1976
- 1978
- Pollen antigens 1983
- 1987 Photosynthetic pathways
- 1991 Chloroplast DNA analysis
- 2000 Phytochrome B

UNDERGROUND SEEDLING **FEATURES**

Hoshikawa (1969) recognized six patterns of underground organs of the grass seedling based upon:

- the presence or absence of transitory node roots (TNR);
- whether or not the mesocotyl elongates, and
- the presence or absence of mesocotyl roots (MR).

He also discovered seven patterns of seedling establishment based upon features of the TNR, MR, and crown node roots (CNR). He then used the six patterns of organs and the seven of establishment to characterize various subfamilies and tribes.

FIRST LEAF

Kuwabara (1960, 1961) investigated the shape, position, and length/width ratio of the first leaf of the grass seedling. He found that in festucoids the leaf blade was linear, perpendicular, and had a L/W ratio = 10.3-106. In panicoids, the blade was oval to lanceolate, horizontal to ascendent, and L/W = 1.3-12. In eragrostoids, the blade was heterogeneous and L/W = 15.5-25.9.

ROOT HAIRS

Row & Reeder 1957) and Reeder & von Maltzalen (1953) found two kinds of root hairs in grasses:

Type A, the hairs were made up of alternating long and short cells and the hairs themselves arose from apical end at a 45 angle. Typical of pooid grasses.

Type B, the hairs were composed of cells of equal length and arose from the midpoint at 90° angles. Found more commonly in chloridoid and panicoid taxa.

LEAF IN TRANSVERSE SECTION

Based upon the studies of Avdulov (1931), Prat (1936), and Brown (1958), we have been able to recognize six different patterns: festucoid, bambusoid, arundinoid, panicoid, aristidoid, and chloridoid. The characters used to distinguish the six types include those of the vascular bundles, the endodermis, the mestome sheath, the kind and location of plastids, and the pattern of chlorenchyma cells. In leaves of grasses that employ the C_4 photosynthetic pathway, there are specialized cells around the vascular bundles.

CULM IN TRANSVERSE SECTION

Brown, Harris, & Graham (1959) found that while almost all of the poold grasses that they examined had hollow internodes, that 49-100% of the chloridoid and panicoids had \pm solid internodes. They concluded that there was a positive correlation between the latter condition and hot, arid habitats.

CULM NODE AND SHEATH PULVINI

Brown et al. (1959) investigated the presence of leaf sheath pulvini and its correlation with hollow vs. solid culm internodes. Pulvini are meristematic swellings at the base of a leaf sheath. They are sometimes called "motor organs" because they make it possible for a stem that has been trampled or blown down to right itself. They discovered that pooid grasses have pulvini, while most panicoid and chloridoids do not. Therefore, pulvini are correlated with hollow internodes.

LEAF EPIDERMIS

Prat (1932, 1936) found that the leaf epidermis was a rich source of useful characters. Based upon the appearance of long cells, short cells, silica bodies, cork cells, stomata, microhairs, macrohairs, prickle-hairs, and papillae, it is possible to recognize four groups: bambusoids, pooids, chloridoids, and panicoids.

Tateoka et al. (1959) and Johnston & Watson (1976) have carried out extensive investigations of bicellular microhairs on grass leaf epidermises. Their findings are summarized as follows:

width of hair/length of hair panicoids: 0.694 eragrostoids: 2.265

 ratio of upper cell to lower cell length panicoids: 1.731 eragrostoids: 2.681

 angle between axis of hair and shortest line panicoids: 2.014 eragrostoids: 0.890

cell wall thickness in upper and lower cells
 Type A: about same thickness
(eragrostoids)

Type B: intermediate
Type C: upper thinner than lower

(panicoids)

TUNICA LAYERS IN SHOOT APEX

At the apex of a grass shoot is a one- or two-layered tunica layer that covers the main body of cells. Brown et al. (1957) found that grasses always appear to have a two-layered tunica; while Barnard (1964) found that chloridoid and panicoid grasses have only a 1-layered tunica. This is the sort of major controversy that will cause you to toss and turn all night long.

LODICULES

Lodicules are small green or whitish flaps of tissue at the base of the grass flower. Most grasses have two; bamboos have three. Lodicules are usually interpreted as perianth remnants. Studies by Stebbins (1956), Jirásek & Jozífová (1968), and Guédès & Dupuy (1976) studied variation in lodicule structure at the subfamily and tribal levels. Their findings suggest the following patterns:

- Panicoids: short, truncate, thick, and heavily vasculated;
- Pooids: elongate, pointed, thick base and

membranous above with little or no vasculature;

- Bambusoids: similar to festucoids, but heavily vasculated;
- Chloridoids: similar to panicoids, but little vasculature

LODICULE MICROHAIRS

Tateoka (1967) found that microhairs on lodicules are frequent in bambusoids, rare in most subfamilies, and absent in festucoids.

EMBRYO STRUCTURE

The grass embryo has been studied intensively for over a century, beginning with Bruns (1892), and continuing with Van Tieghem (1897), Yakovlev (1950), Reeder (1957, 1962), and Kinges (1961). John Reeder's 1957 paper is especially important. He examined four characters:

- whether the vascular trace to the scutellum and coleoptile diverged at the same point [F] or were separated by an internode [P];
- whether the epiblast was present [+] or absent [];
- whether the scutellum was free from the coleorhiza [P] or fused with it [F]; and
- whether the embryonic leaf margins over-lapped [P] or merely met [F].

Based upon combinations of these character states, Reeder recognized seven embryo types:

STARCH GRAINS

For over a century, agrostologists have been studying the starch grains in the grass endosperm, hoping that the variation that they had discovered had some systematic significance. Tateoka (1962) investigated almost 800 taxa and found appreciable variation even within a single species. Although he could not find correlations that held with any subfamilies, he did recognize four different types of starch grains:

Type 1: Triticum-type. Simple grains, broadly elliptic, rounded

Type 2: Panicum-type. Simple grains, angular

Type 3: Miscanthus-type. Simple or of 2-4 granules

Type 4: Festuca-Eragrostis type. Compound grains only.

BASE CHROMOSOME NUMBER

The research carried out by Avdulov (1931) and Carnahan & Hill (1961) provided the breadth of chromosome numbers needed to survey variation in grasses. The range, by the way, is impressive: 2n = 4 to 220. In addition to the numbers reported below, base chromosome numbers of 4, 6, 8, 11, 13, 17, 19, and 23 have been cited.

 $\begin{array}{lll} \text{Bambusoideae} & x = 12 \\ \text{Pooideae} & x = 7 \\ \text{Arundinoideae} & x = 6 \text{ or } 12 \\ \text{Ehrhartoideae} & x = 12 \\ \text{Chloridoideae} & x = 12 \\ \text{Chloridoideae} & x = 9 \text{ or } 10 \\ \text{Panicoideae} & x = 5, 9, \text{ or } 10 \end{array}$

PERSISTENT NUCLEOLI

When the nuclei of somatic cells divide, the nucleolus typically disappears before metaphase. The nucleoli are then reconstituted in the nuclei of daughter cells. Frew & Bowen (1927) and Brown & Emory (1957) found that in certain grasses the nucleolus persists after metaphase. Their studies showed that pooid grasses do not have persistent nucleoli, whereas they do persist in chloridoid and panicoids.

EFFECT OF IPC

Al-Aish & Brown (1958) studied the effect of IPC (isopropyl-n-phenyl carbamate) on grass seed germination. Festucoid seeds are very sensitive to IPC and did not germinate in its presence. Panicoid seeds are able to tolerate it; all seeds germinated.

LOW OXYGEN TENSION

The same authors also investigated the ability of grass seeds to germinate in low oxygen atmospheres. The panicoids tested did germinate; the pooids did not do well. Rice seeds were most successful.

CARBOHYDRATE STORAGE

Grasses are either sacchifers (storing carbohydrates only in the form of starch or sucrose) or laevulifers (storing them not only as starch or sucrose, but also as fructose polymers).

Pooids: laevulifers Chloridoids: sacchifers Panicoids: sacchifers

HEAT PRODUCTION OF CARYOPSES

Pooids: weakly exothermic Chloridoids: strongly exothermic Panicoids: strongly exothermic

PHOTOSYNTHETIC PATHWAYS

Grasses are just like all other higher plants in having a two-phase photosynthetic process. The **light**

reaction is a photochemical process (photophosphorylation) that occurs in the chloroplasts and chlorenchyma cells of the mesophyll. Adenosine diphosphate (ADP) is converted to adenosine triphosphate (ATP). NADP is also reduced to NADPH. During the **dark reaction**, CO_2 enters through the stomates and combines with ribulose diphosphate to form a 6-carbon intermediary molecule. It quickly divides into two 3-carbon units, PGA (3-phosphoglyceric acid). These processes occur within the chlorenchyma cells in the mesophyll of the leaf.

About thirty years ago, another version of the dark reaction was discovered that involved 4-carbon units. This alternate pathway is variously known as the Hatch-Slack pathway (after its discoverers) or the kranz-type pathway (from the German word for "ring," an anatomical reference to the arrangement of some specialized leaf cells when seen in cross-section. This second photosynthetic pathway is found in many grasses. Not only are the intermediary compounds different, but the C_4 pathway takes place not only within the chlorenchyma cells of the mesophyll, but also in parenchyma sheath cells (often called kranz cells). Three subtypes of this pathway, based upon the decarboxy-lating enzymes found in the kranz cells, have been recognized. Smith & Brown (1973), Brown (1975, 1977), and Waller & Lewis (1979) investigated the systematic significance of the two pathways and found the following:

Bambusoideae C3 Ehrhartoideae C3 Pooideae C3

Arundinoideae C3 (mostly) and C4 C4 (mostly) and C3 Panicoideae C3 and C4

IMMUNOLOGY AND SEROLOGY

Almost a century ago, two workers in Germany and in England discovered that the combination of antigens and antibodies yielded a visible precipitate and that this phenomenon could be used to investigate how closely related to organisms might be by testing how similar their proteins are to one another. Fairbrothers & Johnson (1961) employed this technique in the grasses and they discovered, for instance, that the Festuceae of George Bentham and later workers was a hodgepodge of unrelated taxa. Their work supported carving out a number of genera and placing them in an entirely different subfamily -- the Chloridoideae. Similar studies of amino acids in various grasses have also been carried out.

Another kind of protein interaction occurs when grass pollen encounters the tissues lining the nasal passages in humans. The taxonomic significance of allergic reactions to grasses has been explored by Watson (1983), and Watson & Knox (1976). Correspondence with Dr. Watson revealed that he and I were both amazed to see that we were allergic to some tribes of grasses, but not to others.

FLAVONOIDS

Flavonoids are secondary metabolites in plants. More specifically, they are kinds of phenolic compounds -- a loose assemblage of chemicals based upon a phenol nucleus. A number of the more unusual plant

pigments are flavonoids. Harborne & Williams (1976, 1987) have studied these pigments in grasses.

DNA & RNA STUDIES

When I was a graduate student, chromosomes were the Messiah. Yes, chromosomes had been invented that long ago! Their number, morphology, and behavior during meiosis would provide an objective index for determining whether two plants or two populations of plants belonged to a single species. What a disappointment it was to discover that grasses were not that simple.

More recent techniques focus on genetic information at an even more fundamental level. If chromosomes do not tell us what we need to know, then certainly examining the sequence of base pairs in the genetic material itself ought to do the trick. The earliest attempts to examine nucleic acid sequences involved extracting the total DNA of one plant and comparing it with the total DNA from another. The degree to which the two DNA samples annealed or reassociated was taken as a measure of how closely related the two entities were.

This methodology worked well in microorganisms, but less so in higher plants. Long stretches of repeated DNA sequences and the fact that DNA occurred in chromosomes, mitochondria, and plastids made analysis of these life forms more difficult. Today investigators use only specific sections of DNA that are separated by enzymes (restriction endonucleases) into well defined restriction fragments. These fragments can be separated from one another by gel electrophoresis. These studies rest on the assumption that examination of comparable restriction site fragments of DNA will yield more useful data. One of the more elegant papers on this subject is the work of Jones & Flavell (1982) on rye.

SELECTED REFERENCES

Al-Aish, M. & W. V. Brown. 1958. Grass germination responses to isopropyl-n-phenyl carbonate and classification. American J. Bot. 45: 16-23.

Amarasingehe, V. & L. Watson. 1988. Comparative ultrastructure of microhairs in grasses. Bot. J. Linnean Soc. 98: 303-319.

Augustine, R. 1959. Grass pollen allergens. I: paper chromatography and membrane diffusion studies. Immunology 2: 1-18.

Augustine, R. 1959. Grass pollen allergens. II: antigen-antibody precipitation patterns in gel -- their interpretation as a serological problem and in relation to skin reactivity. Immunology 2: 148-169.

Augustine, R. 1959. Grass pollen allergens. III: their differentation from the other pollen antigens by immuno-electrophretic studies in relation to skin reactivity, enzymic digestions heat and pH stabilities. Immunology 2: 230-251.

Augustine, R. 1963. Antigens and other allergens of grass pollens. Allergologia Immunopathologia 1: 3-24.

Auquier, P. 1963. Critères anciens et modernes dans

la systematique des Gramineés. Nat. Monsana 18: 1-63.

Avdulov, N. P. 1931. Karyo-systematische Untersuchungen der Familie Gramineen. Bull. Appl. Bot. Genet. Plant Breed. Suppl. 44: 1-428. (Zusammenfassung, pp. 353-425).

Baum, B. R. 1987. Numerical taxonomic analysis of the Poaceae. <u>In</u>, Soderstrom, T. R. et al. Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 334-342.

Bonnett, G. D. et al. 1997. Structural diversity of fructan in relation to the taxonomy of the Poaceae. New Phytol. 136(1): 11-17.

Brown, W. V. 1958. Leaf anatomy in grass systematics. Bot. Gaz. 119: 170-178.

Brown, W. V. 1959. Grass leaf anatomy: its use in systematics. [Abstract]. Proc. 9th Intern. Bot. Congr. 2:50.

Brown, W. V. 1960. A cytological difference between the Eupanicoideae and the Chloridoideae. Southw. Nat. 5: 7-11.

Brown, W. V. 1965. The grass embryo: a rebuttal. Phytomorph. 15: 274-284.

Brown, W. V. 1974. Another cytological difference among the Kranz subfamilies of the Gramineae. Bull. Torrey Bot. Club 101: 120-124.

Brown, W. V. 1975. Variations in anatomy, associations, and origins of Kranz tissue. American J. Bot. 62: 395-402.

Brown, W. V. 1977. The kranz syndrome and its subtypes in grass systematics. Mem. Torrey Bot. Club 23(3): 1-97.

Brown, W. V. 1975. Variations in anatomy, associations, and origins of kranz tissue. American J. Bot. 62: 395-402.

Brown, W. V. & B. N. Smith. 1972. Grass evolution, the kranz syndrome, 13C/12C ratios, and continental drift. Nature 239: 345, 346.

Brown, W. V. & W. H. P. Emery. 1957. Persistent nucleoli and grass systematics. Amer. J. Bot. 44: 585-590.

Brown, W. V., C. Heimsch, & W. H. P. Emery. 1957. The organization of the grass shoot apex and systematics. American J. Bot. 44: 590-595.

Brown, W. V., G. A. Pratt, & H. M. Mobley. 1959. Grass morphology and systematics. II. The nodal pulvinus. Southwest. Nat. 4: 126-130.

Brown, W. V., W. F. Harris, & J. D. Graham. 1959. Grass morphology and systematics. I. The internode. Southwest. Nat. 4: 126-130.

Bruns, E. 1892. Der Grasembryo. Flora 76: 1-33.

Carolin, R. C., W. I. Jacobs, & M. Vesk. 1973. The structure of the cells of the mesophyll and parenchymatous bundle sheath of Gramineae. Bot. J. Linnean Soc. 66(4): 259-275.

- Chandra, N. 1963. Some ovule characters in the systematics of Gramineae. Current Science 32(6): 277-279.
- Cheadle, V. I. 1955. The taxonomic use of specialization of vessels in the metaxylem of Gramineae, Cyperaceae, Juncaceae, and Restionaceae. J. Arnold Arbor. 36: 141-157.
- Cheadle, V. I. 1960. Vessels in grasses: kinds, occurrences, taxonomic implications. J. South Afr. Biol. Soc. 1: 27-37.
- Clifford, H. T. 1969. Attribute correlations in the Poaceae (grasses). Bot. J. Linnean Soc. 62: 59-67.
- Cugnac, A. de 1931. Les glucides des Graminees. Ann. Sci. Nat., Bot series 10, 13: 1-29.
- Dengler, N. G. et al. 1994. Quantitative leaf anatomy of C_3 and C_4 grasses (Poaceae): bundle sheath and mesophyll surface area relationships. Ann. Bot. 73: 241-255.
- Dengler, N. G., R. E. Dengler, & P. W. Hattersley. 1985. Differing ontogenetic origins of PCR ("kranz") sheaths in leaf blades of C4 grasses (Poaceae). American J. Bot. 72: 284-302.
- De Wet, J. M. J. 1960. Culm anatomy in relation to taxonomy. Bothalia 7: 311-316.
- Downton, J., J. Berry, & E. B. Tregunna. 1969. Photosynthesis: temperate and tropical characteristics within a single grass genus. Science 163: 78, 79.
- Doyle, J. J. et al. 1992. Chloroplast DNA inversions and the origin of the grass family (Poaceae). Proc. Natl. Acad. Sci. (USA) 89: 7722-7726.
- Ebinger, J. E. & J. L. Carlen. 1975. Culm morphology and grass systematics. Trans. Illinois Acad. Sci. 68: 87-101.
- Esen, A. & K. Hilu. 1989. Immunological affinities among subfamilies of the Poaceae. American J. Bot. 76: 196-203.
- Evans, G. 1926. Chromosome complements in grasses. Nature 118: 841.
- Fairbrothers, D. E. & M. A. Johnson. 1961. The precipitation reaction as an indicator of relationships in some grasses. Recent Adv. Bot. 1: 116-120.
- Felsenstein, J. 1988. Phylogenies from molecular sequences: inferences and reliability. Ann. Rev. Genetics 22: 521-565.
- Feinberg, J. G. 1960. Immunological inadequacy of randomly selected grass pollen extracts in specific hay fever therapy. Int. Archs. Allergy 16: 1-16.
- Frew, P. & R. H. Bowen. 1929. Nucleolar behavior in the mitosis of plant cells. Quart. J. Microscop. Sci. 73: 197-214.
- Gould, F. W. & R. B. Shaw. 1983. Grass classification. In, Grass systematics. Texas A & M Univ. Press. College Station. Pp. 93-105.
- Grob, A. 1896. Beitrage zur Anatomie der Epidermis der Gramineenblatter. Biblio. Bot. 36. 123 pp. + 10 plates.

- Guédès, M. & P. Dupuy. 1976. Comparative morphology of lodicules in grasses. Bot. J. Linnean Soc. 73: 317-331.
- Hamby, R. F. & E. A. Zimmer. 1988. Ribosomal RNA sequences for inferring phylogeny within the grass family (Poaceae). Plant Syst. Evol. 160: 29-37.
- Harberd, D. J. 1972. A note on the relevance of the mesocotyl in the systematics of the Gramineae. Ann. Bot. 36: 599-603.
- Harborne, J. B. & C. A. Williams. 1976. Flavonoid patterns in leaves of the Gramineae. Biochem. Syst. Ecol. 4(4): 267-280.
- Harborne, J. B. & C. A. Williams. 1987. Flavonoid patterns of grasses. <u>In</u>, Soderstrom, T. R. et al. (editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 107-116.
- Hattersley, P. W. 1984. Characterization of C4 type leaf anatomy in grasses (Poaceae). Mesophyll: bundle sheath area ratios. Ann. Bot. 53: 163-179.
- Hattersley, P. W. 1987. Variations in photosynthetic pathway. <u>In</u>, Soderstrom, T. R. et al. (editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 49-64.
- Hattersley, P. W. & L. Watson. 1975. Anatomical parameters for predicting photosynthetic pathways of grass leaves: the "maximum lateral cell count" and the "maximum cell distant count." Phytomorphology 25: 325-333.
- Hilu, K. W. 1987. Chloroplast DNA in the systematics and evolution of the Poaceae. <u>In</u>, Soderstrom, T. R. et al. (Editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 65-72.
- Hilu, K. W. 2000. Contributions of prolamin size diversity and structure to the systematics of the Poaceae. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 241-247.
- Hilu, K. W. & J. L. Johnson. 1991. Chloroplast DNA reassociation and grass phylogeny. Plant Syst. Evol. 176(1-2): 21.
- Hock-Hin, Y. & L. Watson. 1987. Taxonomic patterns in protein amino acid profiles of grass leaves and caryopses. <u>In</u>, Soderstrom, T. R. et al. (Editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 88-96.
- Hoshikawa, K. 1969. Underground organs of the seedlings and the systematics of Gramineae. Bot. Gaz. 130(3): 192-203.
- Hsiao, C. et al. 1994. Phylogenetic relationships of 10 grass species: an assessment of phylogenetic utility of the internal transcribed spacer region in nuclear ribosomal DNA in monocots. Genome 37: 112-120.
- Hunter, A. W. 1934. A karyosystematic investigation in the Gramineae. Canadian J. Research 11: 213-241.
- Janchen, E. 1938. Der morphologischen Wert der Gramineen-Vorspelze. Osterr. Bot. Z. 87: 51-61.
- Jirásek, V. & M. Jozífová. 1968. Morphology of

- lodicules, their variability and importance in the taxonomy of the Poaceae family. Bot. Soc. Arg. Bot. 12: 324-349.
- Johnston, C. R. & L. Watson. 1976. Microhairs: a universal characteristic of non-festucoid grass genera? Phytomorphology 26(3): 297-301.
- Johnston, C. & W. V. Brown. Grass leaf ultrastructure variations. American J. Bot. 60: 727-735.
- Jones, J. D. G. & R. B. Flavell. 1982. The structure, amount, and chromosomal localization of defined repeated DNA sequences in species of the genus *Secale*. Chromosoma 86: 613-641.
- Kabuye, C. H. S. & D. Wood. 1969. A first record of multicellular glandular hairs in the Gramineae. Bot. J. Linnean Soc. 62: 69, 70.
- Kahler, A. L. & S. C. Price. 1987. Isozymes in population genetics, systematics, and evolution of grasses. <u>In</u>, Soderstrom, T. R. et al. (Editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 97-106.
- Kemp, P. R., G. L. Cunningham, & H. P. Adams. 1983. Specialization of mesophyll morphology in relation to C_4 photosynthesis in the Poaceae. American J. Bot. 70(3): 349-354.
- Kennedy, P. B. 1899. The structure of the caryopsis of grasses with reference to their morphology and classification. Bull. No. 19. Div. of Agrostology. U. S. Dept. Agric. 44 pp.
- Kinges, H. 1961. Merkmale des Gramineen embryos. Bot. Jahrb. 81: 50-93.
- Kuwabara, Y. 1960. The first seedling leaf in grass systematics. J. Japanese Bot. 35(5): 139-145.
- Kuwabara, Y. 1961. On the shape and the direction of leaves of grass seedlings. J. Japanese Bot. 36: 368-373.
- Kuwabara, Y. 1962. Caryopsis and systematics in Japanese grasses. J. Japanese Bot. 37: 207-217.
- Kuwabara, Y. 1968. Systematic and autecological studies on grasses, especially on the taxonomic characteristics of grass seedlings. Bol. Soc. Argent. Bot. 12: 316-323.
- Lewis, R. F. & N. V. Rothwell. 1964. Implications of nucleolar differences in the root epidermis among several grass species. American J. Bot. 51: 1107-1113.
- Liang, H. & K. W. Hilu. 1996. Application of the matK gene sequences to grass systematics. Canadian J. Bot. 74(1): 125-134.
- Liphschitz, N. 1974. Existence of salt glands in various genera of Gramineae. New Phytol. 73(3): 507-513.
- Lommasson, R. C. 1961. Grass leaf venation. Recent Adv. Bot. 1: 108-111.
- Maffei, M. 1996. Chemotaxonomic significance of leaf wax alkanes in the Gramineae. Biochem. Syst. Ecol. 24(1): 53-64.
- Mlada, J. 1974. The histological structure of the grass

- embryo and its significance for the taxonomy of the family Poaceae. Acta Univ. Carol. Biol. 2/3: 51-156.
- Myers, W. M. 1947. Cytology and genetics of forage grasses. Bot. Rev. 13: 318-421.
- Nadot, S., R. Bajon, & B. Lejeune. 1994. The chloroplast rps4 as a tool for the study of Poaceae phylogeny. Plant Syst. Evol. 191: 27-38.
- Neyra, C. A. & J. Dobereiner. 1977. Nitrogen fixation in grasses. Adv. Agron. 29: 1-38.
- Ono, H. & T. Tateoka. 1953. Karyotaxonomy in Poaceae. I. Chromosomes and taxonomic relations in some Japanese grasses. Bot. Mag. (Tokyo) 66: 18-27.
- Page, J. S. 1978. A scanning electron microscope survey of grass pollen. Kew Bull. 32(2): 313-319.
- Palmer, P. G. 1976. Grass cuticles: a new paleoecological tool for East African lake sediments. Canadian J. Bot. 54: 1725-1733.
- Piperno, D. R. & D. M. Pearsall. 1998. The silica bodies of tropical American grasses: morphology, taxonomy, and implications for grass systematics and fossil phytolith identification. Smithsonian Contr. Bot. No. 85. Smithsonian Inst. Press. Washington, D. C. 40 pp.
- Prat, H. 1932. L'épiderme des Graminées; étude anatomique et systématique. Ann. Sci. Nat. Bot., Ser. 10, 14: 117-324.
- Prat, H. & C. Vignal. 1968. Utilisation des particularities de l'epiderme pour l'identification et al recherche des affinites des Gramineés. Bol. Soc. Argent. Bot. 12: 155-160.
- Prendergast, H. D. V., P. W. Hattersley, & N. E. Stone. 1987. New structural/biochemical associations in leaf blades of C^4 grasses (Poaceae). Australian J. Plant Physiol. 14: 403-420.
- Reeder, J. R. 1957. The embryo in grass systematics. American J. Bot. 44: 756-769.
- Reeder, J. R. 1962. The bambusoid embryo: a reappraisal. American J. Bot. 49: 639-641.
- Reeder, J. R. & K. von Maltzalen. 1953. Taxonomic significance of root-hair development in the Gramineae. Proc. Nat. Acad. Sci. (U. S. A.) 39: 593-598.
- Rosengurtt, B. et al. 1971. Lipids in central endosperm of Gramineae in relation to taxonomy. Adansonia, Ser. 2, 11(2): 383-391.
- Rothwell, N. V. 1964. Nucleolar size differences in the grass root epidermis. American J. Bot. 51: 172-179.
- Row, H. C. & J. R. Reeder. 1957. Root hair development as evidence of relationships among genera of Gramineae. American J. Bot. 44: 596-601.
- Seberg, O. 1988. Genome analysis, phylogeny, and classification. Plant Syst. Evol. 166: 159-171.
- Singh, D. & M. B. E. Godward. 1960. Cytological studies in the Gramineae. Heredity 15: 193-197.
- Sinha, N. R. & E. A. Kellogg. 1996. Parallelism and

- diversity in multiple origins of C⁴ photosynthesis in the grass family. American J. Bot. 83(11): 1458-1470.
- Smith, B. N. 1973. Kranz syndrome in Gramineae as indicated by carbon isotopic ratios. American J. Bot. 60(6): 505-513.
- Smith, B. N. & W. V. Brown. 1973. The Kranz syndrome in the Gramineae as indicated by carbon isotype ratios. American J. Bot. 60: 505-513.
- Smith, D. 1968. Classification of several native North American grasses as starch or fructosan accumulators in relation to taxonomy. J. Brit. Grassland Soc. 23: 306-309.
- Smith, P. 1969. Serological relationships and taxonomy in certain tribes of the Gramineae. Ann. Bot. 33: 591-613.
- Sparrow, A. H. & A. F. Nauman. 1974. Evolutionary changes in genome and chromosome sizes and in DNA content in grasses. Brookhaven Symp. Biol. 23: 367-389.
- Stebbins, G. L. 1987. Grass systematics and evolution: past, present and future. <u>In</u>, Soderstrom, T. R. et al. Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 359-367.
- Taira, H. 1968. Amino acid pattern of grass seed and systematics. Proc. Japanese Soc. Plant Tax. 2: 14-17.
- Takeda, T. & M. Fukuyama. 1971. Studies on the photosynthesis of the Gramineae. Part 1. Differences in photosynthesis among subfamilies and their relations with the systematics of the Gramineae. Proc. Crop Sci. Jap. 40(1): 12-20.
- Tateoka, T. 1960. Cytology in grass systematics: a critical review. Nucleus 3: 81-110.
- Tateoka, T. 1962. Starch grains of endosperm in grass systematics. Bot. Mag. Tokyo 75: 377-383.
- Tateoka, T. 1967. Notes on some grasses. XIX. Systematic significance of microhairs of lodicule epidermis. Bot. Mag. (Tokyo) 80: 394-403.
- Tateoka, T. 1969. Notes on some grasses. Part 20. Systematic significance of the vascular bundle sheath in the mesocotyl. Bot. Mag. (Tokyo) 82(976): 387-391.
- Tateoka, T. 1969. Root anatomy in grass systematics. Bull. Nat. Sci. Mus. (Tokyo) 12(3): 643-651.
- Tateoka, T., S. Inoue, & S. Kawano. 1959. Notes on some grasses. IX. Systematic significance of bicellular microhairs of the leaf epidermis. Bot. Gaz. 121: 80-91.
- Terrell, E. E. 1971. Survey of occurrences of liquid or soft endosperm in grass genera. Bull. Torrey Bot. Club 98: 264-268.
- Tulloch, A. P. & L. L. Hoffman. 1977. Composition of epicuticular waxes of some grasses. Canad. J. Bot. 55(8): 853-857.
- Van Tieghem, P. 1897. Morphologie de l'embryon et de la plantule chez les Graminées et les Cyperacées. VIII. Ann. Sci. Nat. Bot. 3: 259-309.

- Waller, S. S. & J. K. Lewis. 1979. Occurrence of C_3 and C_4 photosynthetic pathways in North American grasses. J. Range Management 32: 12-28.
- Watson, L. 1972. Smuts on grasses: some general implications of the incidence of Ustilaginales on the genera of Gramineae. Q. Rev. Biol. 47: 46-62.
- Watson, L. 1983. Taxonomic patterns in grass pollen antigens and allergens. Proc. Sydney Allergen Group 3: 37-47.
- Watson, L. & C. R. Johnston. 1978. Taxonomic variation and stomatal insertion among grass leaves. Australian J. Bot. 26: 235-238.
- Watson, L. & E. M. Bell. 1975. A surface-structural survey of some taxonomically diverse grass pollens. Australian J. Bot. 23: 981-990.
- Watson, L. & R. B. Knox. 1976. Pollen wall antigens and allergens: taxonomically-ordered variation among grasses. Ann. Bot. 40: 399-408.
- Wodehouse, R. P. 1955. Antigenic analysis by gel diffusion. II. Grass pollen. Int. Archs. Allergy 6: 65-70.
- Wright, G. L. T. & H. T. Clifford. 1965. The relationship of intermediate grass pollen skin reactions to taxonomic groups of grasses. Med. J. Australia 2: 74, 75.
- Yakovlev, M. S. 1950. Structure of embryo and endosperm in grasses as a systematic feature. Morf. Anat. Trudy Bot. Akad. Nauk. S. S. S. R. (Series 7) 1: 121-218.
- Yeoh, H. H., N. E. Stone, & L. Watson. 1982. Taxonomic variation in the subunit amino acid compositions of RuBP carboxylates from grasses. Phytochemistry 21: 71-80.

SECTION 3 - SURVEY OF SUBFAMILIES, TRIBES, & GENERA

3.01 - SUBFAMILY BAMBUSOIDEAE

TECHNICAL DESCRIPTION

Habit: Mostly perennial; culms woody or herbaceous; leaf blades often pseudopetiolate

Root Hairs: Unequal, arising from the middle of the cell

Leaf Epidermis: Bicellular microhairs; short cells paired or in rows; silica-cells cross-, saddle-, or dumbbell-shaped; stomata with dome-shaped subsidiary cells

Leaf in Cross-section: Double vascular bundle sheath, the inner with thicker cell walls; large fusoid cells and arm cells present; non-kranz anatomy

Inflorescence: Typically paniculate; often subtended by a spathe

Spikelets: Bisexual or unisexual; 1- to manyflowered; glumes 2-5; awned or awnless, disarticulation above the glumes and between the florets.

Flowers: Lodicules [1] 3 [10]; stamens 3, 6, or more; gynoecium with 1, 2, 3 or more stigmas, tricarpellate

Fruit: Caryopsis, nut, berry, or utricle

Embryo Formula: F + P P

Cytology: x = 10, 11, or 12

Photosynthetic Pathway: C₃

Distribution: Asia, Africa, the Americas, and the islands of the Pacific and Indian oceans; abundant in the tropics; absent from Europe; mostly in forests, woodlands, and wet places.

SYSTEMATICS

Unfortunately, the delimitation of our first subfamily remains controversial. One hint that there is a problem is that these grasses may be treated as one, two or three distinct subfamilies. The most liberal interpretation places the woody and herbaceous bamboos in Bambusoideae, the oryzoid grasses (rice and its relatives) in Ehrhartoideae, and the pharoid grasses (only one species of which is in North

America) in Pharoideae. Note how various authors have disposed of our North American bamboos:

Bentham (1881): Bambuseae
Roshevits (1937): Bambuseae, Phareae
Stebbins & Crampton (1961): Bambuseae
Potztal (1964): Bambuseae, Arundinariae
Tzvelev (1989): Bambuseae, Arundinariae
Clayton & Renvoize (1992): Bambuseae, Olyreae,
Phareae, Oryzeae, Ehrharteae, Diarrheneae,
Brachyelytreae
Watson & Dallwitz (1998): Bambuseae, Oryzeae,
Phareae, Diarrheneae

Grass Phylogeny Working Wroup (2001): Bambuseae, Olyreae

A common error is to assume that any really large grass with what appears to be woody stems must be a bamboo. Look also in Arundinoideae for large bamboo-like grasses, such as the giant reed grass (*Arundo donax*).

TRIBE: BAMBUSEAE

Woody perennials from extensive rhizomes. Culms 2-10+ m tall, freely-branched; internodes hollow to solid. Leaves of axillary branches pseudopetiolate, while those of the culm itself typically without blades or those that fall early. Inflorescence usually a large, many-flowered panicle; sometimes reduced to 1 spikelet. Spikelets typically large, several-flowered, bisexual, and disarticulating above the glumes and between the florets. Glumes 2 (the first often reduced), shorter than the lemmas. Lemmas similar to glumes, 5- to many-nerved, usually awnless. Palea often 2-keeled. Lodicules typically 3; stamens usually 6; stigmas usually 3. [= Arundinarieae in older literature].

Bamboos, the tree grasses, are variable in habit. They range from a few centimeters to more than 40 m tall. The culms may be solid or have hollow internodes. Larger bamboos are almost 25 cm in diameter with internodes of up to 1.5 m. Not all bamboos are woody; plants in two of the three tribes represented in the flora of North America are herbaceous. One bamboo is a vine 30 m long.

Rhizome development is extensive. Two types of rhizome systems are recognized. The **pachymorph** rhizome is short and thick, with the lateral buds producing only rhizomes. New culms arise from the apex of the rhizome system. Bamboos of this sort typically have a "clump" growth form. The **leptomorph** rhizome system is long and slender, with many nodes capable of producing a new culm and adventitious roots. This is the rhizome system

characteristic of the "running bamboos."

Leaves on the main culm axis are typically bladeless or with blades that fall early. The leaves of lateral branches appear to be petiolate, but they are probably best considered **pseudopetiolate**. Sheaths and scale leaves have played an important role in the traditional taxonomy of the group.

The inflorescence of many bamboos is a panicle or is reduced to a single spikelet, subtended by a few to several bracts, this combination known as a **pseudospikelet**. It may be simple or compound.

The flowers are the most monocot-like found in the family. They are composed of three lodicules, 3 or 6 stamens (rarely as many as 120!), and three united carpels. Blooming is erratic. There seems to be three basic patterns. Some bamboos flower gregariously, in regular cycles during which they set tremendous quantities of seeds, and then die within a year or so. Other bamboos are characterized by irregular flowering, during which vegetative growth of the plant is stunted; but, the plants do not die. Still other species flower annually.

It now appears that most of the woody bamboos are probably wind pollinated, but that at least some of the herbaceous bambusoids are insect pollinated. Some bear their spikelets beneath the litter of the forest floor; others produce underground fruits, as in the peanut.

The economic uses of bamboos are almost endless -- "No growing things on earth have so many and so varied uses as ... bamboos" (Soderstrom, 1979). They include ornamentals building materials for houses, furniture, ships, aqueducts, carts, umbrella frames, bird cages, tiger cages, chop sticks, musical instruments, springs for carts, food, fibers, cordage, oars, masts, baskets, mats, spear shafts, bows, arrows, knives, ladders, rafts, pails, churns, curtains, tiles for roofs, beehives, fans, fishing poles, medicine ... and several hundred more!

ARUNDINARIA. Cane, canebrake, giant cane, switch cane. Rhizomatous perennial. Culms woody, 2-8 m tall. Leaves pseudopetiolate, the blades disarticulating from sheaths. Inflorescence spicate to paniculate; pseudospikelets absent. Spikelets bisexual, large, several-flowered, laterally compressed, disarticulating above the glumes and between the florets. A gigantea is the only truly woody grass native to N. America and the only commonly occurring native bamboo here. The plants often form dense colonies. The species occurs from Ohio and Illinois into the Southeast. Three subspecies are now recognized.

PHYLLOSTACHYS. Fishpole bamboo, madake, timber bamboo, black bamboo, moso bamboo. Shrubby to arborescent perennials. Culms woody, 3-20+ m tall. Leaves pseudopetiolate. Inflorescence spicatepaniculate, often subtended by a spathe. Spikelets large (to 8 cm!), several- to many-flowered, bisexual, disarticulating above the glumes and between the florets. A genus of about 50 eastern Asian species. Of considerable economic importance as a source of handsome ornamentals, fishing rods, walking sticks, furniture, and edible young shoots.

BAMBUSA. Giant bamboo, hedge bamboo, timber bamboo, Oldham's bamboo. Shrubby to arborescent, rhizomatous perennials. Culms 2-35+ m tall. Leaves

pseudopetiolate. Inflorescence in panicles or fascicles; pseudospikelets present. Spikelets terete to laterally compressed, large (to 8 cm!), several-flowered, bisexual, disarticulating above the glumes and between the florets. A large genus of about 120 species native to tropical and subtropical areas of the Old and New World. Of considerable economic importance as a source of rods, poles, scaffolding for construction, furniture, fibers for weaving, paper pulp, and edible shoots.

PSEUDOSASA. Metake, arrow bamboo. Shrubby, rhizomatous perennial. Culms woody, 2-5 m tall. Leaves pseudopetiolate with auriculate setae. Inflorescence a terminal panicle of bisexual spikelets. A genus of 8 species native to eastern Asia. *P. japonica* is commonly cultivated.

TRIBE: OLYREAE

Caespitose perennials. Culms woody, to 3 m. Leaves broad, with asymmetric blades and cordate to sagittate bases. Inflorescence paniculate, the spikeletbearing branches persisting. Spikelets unisexual (species monoecious), several-flowered, disarticulating above the glumes and between the florets. A tribe of about 18 genera. *Olyra latifolia* occurs in Florida.

SELECTED REFERENCES

Calderón, C. E. & T. R. Soderstrom. 1980. The genera of Bambusoideae (Poaceae) of the American continent: key and comments. Smithsonian Contr. Bot. 44: 1-27.

Camus, A. 1935. Classification des bambusées. Arch. Mus. d'Hist. Natur. 12: 601-603.

Camus, E.-G. 1913. Les bambusées: monographie, biologie, culture, principaux usages. Two vols. P. Lechevalier. Paris. 215 pp. + 101 plates.

Chapman, G. P. 1997. The bamboos. Linnean Soc. Symposium Series No. 19. Academic Press. San Diego, CA. 370 pp.

Clark, L. G. 1997. Bamboos: the centerpiece of the grass family. <u>In</u>, Chapman, G. P. (editor). The bamboos. Academic Press. San Diego, CA. Pp. 237-248.

Clark, L. G. 1998. An overview of bamboo systematics today. Monogr. Syst. Bot. Missouri Bot. Gard. 68: 391-396.

Farrelly, D. 1984. The book of bamboo. Sierra Club Books. San Francisco, CA. 332 pp.

Galloway, B. T. 1957. Bamboos: their culture and uses in the United States. Bull No. 1329. U. S. Dept. Agric. Washington, D. C. 000 pp.

Ghopal, B. H. & H. Y. Mohan Ram. 1985. Systematic significance of mature embryo of bamboo. Plant Syst. Evol. 148: 239-246.

Ghorai, A. & A. Sharma. 1980. Bambuseae -- a review. Feddes Repert. 91(5-6): 281-289.

- Grosser, D. & W. Liese. 1973. Present status and problems of bamboo classification. J. Arnold Arbor. 54: 293-308.
- Hanke, D. E. 1990. Seeding the bamboo revolution. Nature 334: 291, 292.
- Haun, J. R., T. F. Clark, & G. A. White. 1966. Fiber and papermaking characteristics of bamboo. Tech. Bull. 1361. U. S. Dept. Agric. Washington, D. C. 19 pp.
- Hidalgo, O. 1974. Bambú: su cultivo y applicaciones en fabricacíon de papel, construccíon, arquitectura, ingeniería, artesanía. Estudios Técnicos Colombianos. Cali. Colombia. 318 pp.
- Holttum, R. E. 1956. The classification of bamboos. Phytomorph. 6: 73-90.
- Janzen, D. H. 1976. Why bamboos wait so long to flower. Ann. Rev. Ecol. Syst. 7: 347-391.
- John, C. K. et al. 1993. On the "monocarpic" flowering of bamboos. Current Sci. 65: 665, 666.
- Judziewicz, E. J. 1987. Taxonomy and morphology of the tribe Phareae. Ph. D. dissertation. Univ. Wisconsin. Madison. 529 pp.
- Judziewicz, E. J. et al. 1999. American bamboos. Smithsonian Inst. Press. Washington, D. C. 392 pp.
- Judziewicz, E. J. et al. 2000. Catalogue of New World grasses (Poaceae): I. Subfamilies Anomochlooideae, Bambusoideae, Ehrhartoideae, and Pharoideae. Contr. U. S. Natl. Herbarium 39: 1-128.
- Kellogg, E. A. & L. Watson. 1993. Phylogenetic studies of a large data set. I. Bambusoideae, Andropogoneae, and Pooideae (Gramineae). Bot. Rev. 59(4): 273-343.
- Keng, P. C. 1982 -->. A revision of the genera of bamboos from the world. J. Bamboo Research. Pt. 1, 1(1): 1-19. Pt. 2, 1(2): 31-46. Pt. 3, 2(1): 11-27. Pt. 4, 2(2): 1-17. Pt. 5, 3(1): 22-42. Pt. 6, 3(2): 1-22. [In Chinese with brief English summary on last page of each part].
- Lawson, A. H. 1968. Bamboo: a gardener's guide to their cultivation in temperate climates. Taplinger Publ. Co. New York, NY. 192 pp.
- Mahadevan, V., S. S. Negi, & O. N. Agarwala. 1962. Flowers to rats. Nature 192(480): 470, 471.
- Marden, L. 1980. Bamboo, the giant grass. Natl. Geogr. 158(4): 502-529.
- McClintock, D. 1967. The flowering of bamboos. J. Royal Hort. Soc. 92(12): 520-525.
- McClure, F. A. 1935. Bamboo -- a taxonomic problem and an economic opportunity. Sci. Monthly 51: 193-204.
- McClure, F. A. 1943. Bamboo as panda food. J. Mammalogy 24: 267, 268.
- McClure, F. A. 1953. Bamboo as a building material. Foreign Agric. (U. S. Dept. of Agric.). 52 pp.
- McClure, F. A. 1956. Bamboo in the economy of Oriental peoples. Econ. Bot. 10(4): 335-361.

- McClure, F. A. 1957-1963. Typification of the genera of the Bambusoideae. Taxon 6: 199-210; 8: 208, 209; 9: 194; 11: 141; 12: 127.
- McClure, F. A. 1961. Toward a fuller description of the Bambusoideae (Gramineae). Kew Bull. 15(2): 321-324.
- McClure, F. A. 1963. A new feature in bamboo rhizome anatomy. Rhodora 65: 134-136.
- McClure, F. A. 1966. A glossary of the bamboos. Taxon 15: 220-235.
- McClure, F. A. 1966. The bamboos: a fresh perspective. Harvard Univ. Press. Cambridge, MA. 347 pp.
- McClure, F. A. 1973. Genera of bamboos native to the New World (Gramineae: Bambusoideae). Smithsonian Contr. Bot. 9: 1-148.
- Metcalfe, C. R. 1956. Some thoughts on the structure of bamboo leaves. Bot. Mag. (Tokyo) 69: 391-400.
- Munro, W. 1868. A monograph of the Bambusaceae, including descriptions of all the species. Trans. Linnean Soc. London 26: 1-157.
- Nadgauda, R. S. et al. 1990. Precocious flowering and seeding behaviour in tissue cultured bamboos. Nature 334: 335, 336.
- Nadgauda, R. S. et al. 1993. Why wait so long for the bamboos to flower? BIC-India Bull. 3: 14-20.
- Ohrnberger, D. & J. Goerrings. 1984. The bamboos of the world: a preliminary study of the names and distribution of the herbaceous and woody bamboos (Bambusoideae Nees v. Esenbeck) -- documented in lists and maps. J. Amer. Bamboo Soc. 5: 1-46.
- Piper, J. M. 1992. Bamboo and rattan: traditional uses and beliefs. Oxford Univ. Press. New York, NY. 88 pp.
- Pohl, R. W. 1982. On the flowering of bamboos in Central America. Brenesia 19/20: 465-475.
- Pohl, R. W. 1991. Blooming history of the Costa Rican bamboos. Rev. Biol. Trop. 39: 111-124.
- Pope, G. G. 1989. Bamboo and human evolution. Nat. Hist. 98(10): 49-57.
- Rashford, J. H. 1995. The past and present uses of bamboo in Jamaica. Econ. Bot. 49(4): 395-405.
- Recht, C. & M. E. Wetterwald. 1992. Bamboos. Timber Press. Portland, OR. 128 pp.
- Reeder, J. R. 1962. The bambusoid embryo: a reappraisal. Amer. J. Bot. 49(6): 639-641.
- Renvoize, S. A. 1985. A survey of leaf-blade anatomy in grasses. V. The bamboo allies. Kew Bull. 40: 509-535.
- Restrepo, E. A. et al. 1990. Tropical bamboo. Rizzoli International. New York, NY. 175 pp.
- Shou-liang, C. & C. Liang-chi. 1988. Chinese bamboos. Science Press. Beijing, China. 120 pp.
- Simmons, J. C. & J. Brandenburg. 1986. The most

useful plant in the world. Audubon 88(1): 58-69.

Sineath, H. H. et al. 1953. Industrial raw materials of plant origin. 5. A survey of the bamboos. Bull. Georgia Inst. Tech., Engr. Exp. Sta. 18: 1-230.

Soderstrom, T. R. 1981. Some evolutionary trends in the Bambusoideae (Poaceae). Ann. Missouri Bot. Gard. 68: 15-47.

Soderstrom, T. R. 1984. In quest of the pygmy bamboos. Bull. Fairchild Trop. Gard. 39(3): 6-15.

Soderstrom, T. R. 1985. Bamboo systematics: yesterday, today and tomorrow. J. American Bamboo Soc. 6: 4-16.

Soderstrom, T. R. & C. E. Calderón. 1974. Primitive forest grasses and evolution of the Bambusoideae. Biotropica 6: 141-153.

Soderstrom, T. R. & C. E. Calderón. 1979. A commentary on the bamboos (Poaceae: Bambusoideae). Biotropica 11(3): 161-172.

Soderstrom, T. R. & C. E. Calderón. 1979. Distribution and environment of the Bambusoideae. <u>In</u>, Numata, M. (Editor). Ecology of grasslands and bamboolands in the world. Junk. The Hague, Netherlands. Pp. 223-236.

Soderstrom, T. R. & C. E. Calderón. 1980. In search of the primitive bamboos. Nat. Geogr. Soc. Res. Reports 12: 647-654.

Soderstrom, T. R. & R. P. Ellis. 1987. The position of bamboo genera and allies in a system of grass classification. <u>In</u>, Soderstrom, T. R. et al. (Editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 225-238.

Soderstrom, T. R. & S. M. Young. 1983. A guide to collecting bamboos. Ann. Missouri Bot. Gard. 70: 128-136.

Soderstrom, T. R. et al. 1988. Distributional patterns of neotropical bamboos. <u>In</u>, Heyer, W. R. & P. E. Vanzolini (editors). Proceedings of a workshop on neotropical distributional patterns. Academia Brasileira de Ciencias. Rio de Janeiro. Pp. 121-157.

Takama, S. 1983. The world of bamboo. Heian Int. Publ. San Francisco, CA. 236 pp.

Tewari, D. N. 1993. A monograph on bamboo. International Book Distr. Dehra Dun, India. 498 pp.

Von Shibata, K. 1990. Beitrag zur Wachstumgeschichte der Bambusgewachse. J. Coll. Sci. Imperial Univ. Tokyo 13: 427-496.

Wen, T.-H. 1986. Some ideas about the origin of bamboos. J. Amer. Bamboo Soc. 6: 104-111.

Wu, M. C. -Y. 1958. The anatomical study of bamboo leaves. Qtrly. J. Taiwan Mus. 11: 349-370.

Wu, M. C.-Y. 1962. The classification of Bambuseae based upon leaf anatomy. Bot. Bull. Acad. Sinica, n. s., 3(1): 83-108.

Young, R. A. 1945-1946. Bamboos for American horticulture. Nat. Hort. Mag. 24: 171-196, 274-291; 25: 40-64, 352-365.

Young, R. A. 1954. Flavor qualities of some edible oriental bamboos. Econ. Bot. 8(4): 377-386.

Young, R. A. & J. R. Haun. 1961. Bamboo in the United States: description, culture, and utilization. Handbook No. 193. U. S. Dept. Agric. Washington, D. C. 74 pp.

Zhang, W. 1996. Phylogeny and classification of the bamboos (Poaceae: Bambusoideae) based on molecular and morpho-logical data. Ph. D. dissertation. Iowa State Univ. Ames.

Zhang, W. & L. G. Clark. 2000. Phylogeny and classification of the Bambusoideae (Poaceae). <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 35-42.

Zhang, W., L. G. Clark, & J. F. Wendel. 1995. nhdF gene sequences and phylogeny of subfamily Bambusoideae (Poaceae). American J. Bot. 82 (Suppl.): 173 (abstract).

Zuloaga, F. O. et al. 1993. Endemic herbaceous bamboo genera of Cuba (Poaceae: Bambusoideae: Olyreae). Ann. Missouri Bot. Gard. 80(4): 846-861.

ARUNDINARIA

Gilly, C. L. 1943. A preliminary investigation of the North American canes (*Arundinaria*). Bull. Torrey Bot. Club 70(3): 297-309.

Hughes, R. H. 1951. Observation of cane (*Arundinaria*) flowers, seed, and seedlings in the North Carolina coastal plan. Bull. Torrey Bot. Club 78: 113-121.

Marsh, D. L. 1977. The taxonomy and ecology of cane, *Arundinaria Gigantea* (Walter) Muhlenberg. Ph. D. dissertation. Univ. Arkansas. Little Rock.

Marsh, D. L. 1978. The distribution of cane, *Arundinaria gigantea* (Walt.) Muhl. (Poaceae); Bambusoideae). Proc. Arkansas Acad. Sci. 31: 72-76.

McClure, F. A. 1973. *Arundinaria*. <u>In</u>, Genera of bamboos native to the New World (Gramineae: Bambusoideae). Smithsonian Contr. Bot. 9: 21-40.

Platt, S. G. & C. G. Brantley, 1997. Canebrakes: an ecological and historical perspective. Castanea 62(1): 8-21.

PHYLLOSTACHYS

Hodkinson, T. R. et al. 2000. A comparison of ITS nuclear rDNA sequence data and AFLP markers for phylogenetic studies in *Phyllostachys* (Bambusoideae, Poaceae). J. Plant Res. 113(1111): 259-269.

McClure, F. A. 1957. Bamboos of the genus *Phyllostachys* under cultivation in the United States. Agric. Handbook No. 114. U. S. Dept. of Agric. Washington, D. C. 69 pp.

Renzoize, S. 1995. From fishing poles and ski sticks to vegetables and paper: the bamboo genus *Phyllostachys*. Curtis's Bot. Mag. 12(1): 8-15.

Wang, C.-P. et al. 1980. A taxonomical study of *Phyllostachys*. Acta Phytotax. Sinica 18: 15-19; 168-193.

3.02 - SUBFAMILY PHAROIDEAE

TECHNICAL DESCRIPTION

Habit: Rhizomatous perennial herbs; leaves distichous, pinnately-veined, pseudopetiolate, twisted, the blade resupinate; culms typically solid

Root Hairs:

Leaf Epidermis: Bicellular microhairs and papillae absent; silica bodies dumbbell-shaped; bulliform cells absent or poorly developed

Leaf in Cross-section: Fusoid cells prominent and well-developed; arm cells weakly to moderately well-developed

Inflorescence: Open, terminal panicle

Spikelets: Unisexual, 1-flowered, the male and female spikelets mostly paired on short branchlets; terete (?) or laterally compressed (?); female spikelets covered with uncinate microhairs

Flowers: Lodicules 0 or 3 (3); stamens 6; stigmas

Fruit: Caryopsis
Embryo Formula:

Cytology: X= 12

Photosynthetic Pathway: C₃

Distribution: Pantropical; in New World from Florida southward to Argentina and Uruguay.

SYSTEMATICS

This subfamily was first defined in 1955 by Alan Beetle. It was formally described and published by Clark & Judziewicz in 1996. The subfamily consists of a single tribe of three genera and eleven species. The pharoid grasses are known in North America from a single species collected in Florida.

TRIBE: PHAREAE

Herbaceous perennials. Culms decumbent and rooting at the nodes, the internodes solid. Leaves with broad blades and a twisted, petiole-like constriction above the sheath. Inflorescence a panicle. Spikelets unisexual (species monoecious), 1-flowered, the pistillate sessile and the staminate pedicellate. Glumes 2, shorter than floret, 5- or 7-nerved. Lemmas 5- to 7-nerved, membranous (σ) or indurate (φ). Palea narrow, 2-keeled, 2-nerved.

A tribe of 3 genera. *Pharus lappulaceus* occurs in Florida, based on historic collections. It has probably been extirpated there.

SELECTED REFERENCES

Judziewicz, E. J. 1987. Taxonomy and morphology of the tribe Phareae. Ph. D. dissertation. Univ. Wisconsin. Madison. 529 pp.

Judziewicz, E. J. et al. 2000. Catalogue of New World grasses (Poaceae): I. Subfamilies Anomochlooideae, Bambusoideae, Ehrhartoideae, and Pharoideae. Contr. U. S. Natl. Herbarium 39: 1-128.

3.03 - SUBFAMILY EHRHARTOIDEAE

TECHNICAL DESCRIPTION

Habit: Herbaceous annuals and perennials

Root Hairs: No studies have been made.

Leaf Epidermis: Bicellular microhairs present; silica cells often broader than long, silica bodies mostly dumbbell-shaped; long cells with rows of papillae, as in the bambusoids

Leaf Anatomy: Arms cells and fusoid cells present, as in the bambusoids; vascular bundles with double sheath, the inner with thicker cell walls; non-kranz

Spikelets: Spikelets 1- or 3-flowered, perfect or unisexual, laterally compressed or terete; glumes reduced or absent; lemma 5- to several-nerved; palea 2- to 3+-nerved

Flowers: Stamens 6, 3, or rarely 1; stigmas 1 or 2

Embryo: F + PF or F + PP

Cytology: x = 12 for most species; x = 15 in

Zizania

Photosynthetic Pathway: C₃

Distribution: Marshy and aquatic sites of the

tropics and subtropics.

SYSTEMATICS

Hitchcock and Chase placed the members of this subfamily in three different tribes of their Festucoideae. Until the mid-1990's, the core of this group was rice and its relatives and the subfamily was called Oryzoideae. More recent studies suggest the appropriateness of including Ehrharteae here, which requires a name change.

The ehrhartoid, oryzoid, pharoid, and bambusoid grasses share many features, to the point where many agrostologists combine them into a single subfamily. I have followed that practice for a number of years. However, a number of recent studies, particularly at the molecular level, recognize them as distinct. For nomenclatural reasons that we need not explore, the transfer of *Ehrharta* into the Oryzoideae (rice subfamily) requires that its name be changed to Ehrhartoideae.

TRIBE: EHRHARTEAE

A small tribe of 4 genera, none of them with native to North America. Our only representative is *Ehrharta*.

EHRHARTA. Veldt grass. Annual or perennial herbs. Culms (ours) herbaceous, to 1 m tall. Inflorescence racemose to paniculate. Spikelets laterally compressed or terete, 3-flowered (the lower 2

reduced to sterile lemmas), bisexual, disarticulation above the glumes, but not between the florets. Glumes 2, 5-nerved, shorter or much longer than florets; sterile lemmas often transversely wrinkled; fertile lemma 5- to 7-nerved; palea 2-keeled. Lodicules 2; stamens 3, 4, or 6; stigmas 2. A genus of 27 species native to the Old World. *E. calycina*, *E. erecta*, and *E. longiflora* are weedy in California.

TRIBE: ORYZEAE

Annual or perennial herbs. Inflorescence a panicle. Spikelets laterally compressed, bisexual or unisexual, disarticulating above the glumes. Glumes reduced or absent, represented by a cup-like pedicel apex, their place seemingly occupied by sterile lemmas; fertile floret one. A tribe of about 13 genera.

ORYZA. Rice. Annual or perennial herbs. Inflorescence and open to contracted panicle. Spikelets laterally compressed, appearing 1-flowered, bisexual, disarticulating above the glumes. Glumes reduced to a 2-lobed cupule; lemmas 5-nerved, awned or awnless; palea 2-nerved. Lodicules 2; stamens 6; stigmas 2. A genus of about 25 species native to moist, wet, shady regions of the Old and New World; none native to North American. Rice, *O. sativa*, is one of the principal cereal crops that we consume. It is widely cultivated in flooded fields or on dry land. Another species, *O. rufipogon*, an Old World weedy species, has recently been found in the U. S.

THE SPECIES OF ORYZA

Canama

Nativo

Species	Ge	nome Native
Diploids	[2n	= 2x = 24]
O. australiensis O. barthii O. brachyantha O. eichingeri O. glaberrima	EE AA FF CC AA	Australia West Africa West & central Africa East & central Africa West Africa
O. granulata O. longisminata O. meyeriana O. nivara O. officinalis	AA AA CC	South & Southeast Asia Africa Southeast Asia, China Asia, China, Australia Asia, China, New Guinea
O. punctata O. rufipogon O. sativa O. schlechteri	BB AA AA	Africa Asia and China Asia New Guinea

Tetraploids [2n = 4x = 48]

O. alta O. eichingeri O. grandiglumis O. latifolia O. longiglumis	CCDD BBCC CCDD CCDD	C. & S. America East & central Africa South America C. & S. America New Guinea
O. minuta	BBCC	Southeast Asia

O. punctata
O. ridleyi

BBCC

Africa Southeast Asia

[After Simmonds, 1976]

LEERSIA. Cutgrass, rice cutgrass, white grass. Mostly rhizomatous perennials. Inflorescence a panicle. Spikelets laterally compressed, bisexual, 1-flowered, disarticulating below the glumes. Glumes 0; lemmas 5-nerved, usually awnless. Lodicules 2; stamens 1-6; stigmas 2. A genus of about 18 species, mostly of moist to wooded sites in the Old and New World; 5 occur in North America. *Leersia oryzoides* is our only California species.

ZIZANIA. Wild-rice. Tall, reed-like annuals or perennials. Culms herbaceous, 1-3 m tall. Inflorescence a conspicuous, terminal panicle. Spikelets 1-flowered, unisexual; female spikelets on stiff, erect branches; male spikelets on lower spreading branches. Glumes 0; pistillate lemma 3-nerved, awned; staminate lemma 5-nerved, awntipped; pistillate palea 2-nerved; staminate palea 3-nerved. Stamens 6. A genus of 3 species native to North America and Eurasia. Z. aquatica, the wild rice of commerce, is native to the eastern half of the United States. A second species, Z. texana, occurs only in Texas. The third species (Z. latifolia) is cultivated for its edible shoots.

ZIZANIOPSIS. Marsh millet, southern wildrice. Coarse, rhizomatous perennials. Inflorescence a conspicuous, terminal panicle. Spikelets 1-flowered, unisexual (male and female spikelets on the same branches), disarticulation below the glumes. Staminate spikelet: glumes 0; lemma 5-nerved, awnless; palea 3-nerved; stamens 6. Pistillate spikelet: glumes 0; lemma 7-nerved, acuminate to spikelet: glumes 0; lemma 7-nerved, acuminate to awned; palea 3-nerved; stigmas 2. A genus of 3-5 species native to North and South America. Z. miliacea, the only species found in North America, grows in wet freshwater and brackish sites from Maryland through the southern states to Texas.

LUZIOLA. Southern water grass. Low growing perennials in ponds, marshy sites, and along stream banks. Inflorescences unisexual (σ terminal and the φ axillary). Stamens 6-16. Fruit an achene. As treated here, the genus includes *Hydrochloa*. Three species are found in North America, mostly in the Southeast.

SELECTED REFERENCES

Duvall, M. R. et al. 1993. Phylogeny of North American oryzoid grasses as construed from maps of plastid DNA restriction sites. American J. Bot. 80(1): 83-88.

Jacques-Félix, H. 1955. Les tribus de la serie Oryzoide. J. Agric. Trop. & Bot. Appl. 2: 600-619.

Judziewicz, E. J. et al. 2000. Catalogue of New World grasses (Poaceae): I. Subfamilies Anomochlooideae, Bambusoideae, Ehrhartoideae, and Pharoideae. Contr. U. S. Natl. Herbarium 39: 1-128.

Tateoka, T. 1963. Notes on some grasses. XIII. Relationship between Oryzeae and Ehrharteae, with special reference to leaf anatomy and histology. Bot. Gaz. 124(4): 264-270.

Terrell, E. E. and H. Robinson. 1974. Luziolinae, a new subtribe of oryzoid grasses. Bull. Torrey Bot. Club 101(5): 235-245.

Vegetti, A. C. 2000. Typology of synflorescence in Oryzeae (Poaceae). Phyton (Austria) 40(1): 71-88.

Weatherwax, P. 1929. The morphology of the spikelets of six genera of Oryzeae. Amer. J. Bot. 16: 547-555.

ORYZA

Gopalakrishnan, R. & S. Sampath. 1966. The American species of Oryza. Oryza 3(1): 35-40.

McIntyre, C. L. et al. 1992. Relationships between *Oryza* species (Poaceae) based on 5S DNA sequences. Plant Syst. Evol. 183: 249-264.

Michaud, V. 1944. Morphology of the rice spikelet. Bull. Torrey Bot. Club 71(6): 624-626.

Nanda, J. S. & S. D. Sharma (editors). 2003. Monograph on genus *Oryza*. Science Publ. Enfield, NH. 400 pp.

Ng, N. Q. et al. 1981. Morphological studies of Asian rice. Bot. J. Linn. Soc. 61: 303-313.

Sampath, S. 1963. The genus *Oryza*: its taxonomy and species relationships. Oryza 1: 2-29.

Sasaki, T. & G. Moore (editors). 1997. Oryza: from molecule to plant. Kluwer Acad. Publ. Dordrecht. The Netherlands. 254 pp.

Sharma, S. D. and S. V. S. Shastry. 1964. Neglected characters in taxonomy of genus *Oryza* L. Curr. Sci. 33: 316,317.

Sokolova, I. I. 1969. The systematics of the genus *Oryza*. Tr. Prikl. Bot. Genet. Selek. Vses. Inst. Rastenievod 41(2): 117-147.

Shuguo, F. et al. 2000. History and situation on the research on the classification of *Oryza* L. J. Wuhan Bot. Res. 18(4): 329-337.

Tateoka, T. 1963. Taxonomic studies of *Oryza*, III. Key to species and their enumeration. Bot. Mag. Tokyo 76: 165-173.

Tateoka, T. 1964. Notes on some grasses. XVI. Embryo structure of the genus *Oryza* in relation to the systematics. American J. Bot. 51: 539-543.

Terrell, E. E., P. M. Peterson, & W. P. Wergin. 2001. Epidermal features and spikelet micromorphology in *Oryza* and related genera (Poaceae: Oryzeae). Smithsonian Contr. Bot. No. 91. 50 pp.

Vandiver, V. V. et al. 1992. Discovery of *Oryza rufipogon* (Poaceae: Oryzeae), new to the United States, with its implications. Sida 15(1): 105-109.

Vaughn, D. A. 1989. The genus *Oryza* L.: current status of taxonomy. IRRI Res. Paper Ser. 138. 21 pp.

Vaughn, D. A. 1994. The wild relatives of rice: a genetic resources handbook. International Rice Research Inst. Manila, The Philippines. 137 pp.

LEERSIA

Pyrah, G. L. 1969. Taxonomic and distributional studies in *Leersia*. Iowa State J. Sci. 44(2): 215-270.

ZIZANIA

Aiken, S. G. et al. 1988. Wild rice in Canada. Research Branch. Agric. Canada. Publ. No. 1830. 130 pp.

Beaty, H. E. 1975. Texas wildrice. Texas Hort. 2: 9-11.

Barber, P. H. et al. 1991. Phylogenetic relationships of annual and perennial wild rice. Probing by direct DNA sequencing. Theor. Appl. Genet. 81: 693-702.

de Wet, J. M. J. & E. A. Oelke. 1978. Domestication of American wild rice (*Zizania aquatica* L., Gramineae). J. Agric. Tradit. Bot. Appl. 25: 67-84.

Dore, W. G. 1969. Wild rice. Publ. No. 1393. Research Branch. Canad. Dept. Agric. 84 pp.

Emery, W. H. P. 1977. Current status of Texas wild rice (*Zizania texana* Hitchc.) Southw. Nat. 51: 393, 394.

Fassett, N. C. 1924. A study of the genus *Zizania*. Rhodora 26: 153-160.

Hayes, P. M. et al. 1989. The domestication of American wildrice (*Zizania palustris*, Poaceae). Econ. Bot. 43(2): 203-214.

Horne, F. & A. Kahn. 1997. Phylogeny of North American wild rice, a theory. Southwest Nat. 42(4): 423-434.

Terrell, E. E. et al. 1978. Observations on *Zizania texana* (Texas wildrice), an endangered species. Bull. Torrey Bot. Club 105: 50-57.

Terrell, E. E. & W. P. Wergin. 1981. Epidermal features and silica deposition in lemmas and awns of *Zizania* (Gramineae). American J. Bot. 68(5): 697-707.

Terrell, E. E. & S. W. T. Batra. 1984. Insects collect pollen of eastern wild rice, *Zizania aquatica* (Poaceae). Castanea 49: 31-34.

Terrell, E. E. et al. 1997. Taxonomy of North American

species of Zizania (Poaceae). Sida 17(3): 533-549.

Walsh, R. 1998. Wild and wilder. Nat. Hist. 107(7): 82-85.

Warwick, S. I. & S. G. Aiken. 1986. Electrophoretic evidence for the recognition of two species in annual wild rice (*Zizania*, Poaceae). Syst. Bot. 11(3): 464-473.

ZIZANIOPSIS

Holmes, W. C. & D. T. Stalling. 1990. Studies on the repro-ductive strategy of *Zizaniopsis miliacea* (Michx.) Doell & Asch. (Gramineae: tribe Oryzeae). Castanea 55: 113-121.

LUZIOLA

Swallen, J. R. 1965. The grass genus *Luziola*. Ann. Missouri Bot. Gard. 52(3): 472-475.

TRIBE EHRHARTEAE

Tateoka, T. 1960. Notes on some grasses. XIII. Relationships between Oryzeae and Ehrharteae with special reference to leaf anatomy and histology. Bot. Gaz. 124: 264-270.

Tateoka, T. 1967. Lodicules of the tribes Ehrharteae and Aristideae (Gramineae). Bull. Natl. Sci. Mus. (Tokyo) 10: 443-453.

EHRHARTA

Brey, C. 1996. What? Another *Ehrharta*? CalEPPC Newsletter 4(2): 4, 5.

Gibbs Russell, G. E. & R. P. Ellis. 1987. Species groups in the genus *Ehrharta* (Poaceae) in southern Africa. Bothalia 17: 51-65.

Howell, J. T. 1957. Records of *Ehrharta calycina* in California. Leaflts. West. Bot. 8(5): 144.

Smith, J. P., Jr. 1993. *Ehrharta*. <u>In</u>, Hickman, J. C. The Jepson manual: higher plants of California. Univ. California Press. Berkeley. P. 1253.

3.04 - SUBFAMILY CENTOTHECOIDEAE

TECHNICAL DESCRIPTION

Habit: Perennial herbs

Root Hairs: No studies have been made.

Leaf Epidermis: Bicellular microhairs rodlike to linear; silica cells mostly dumbbell-shaped, confined to the costal region

Leaf Anatomy: Vascular bundles with a double sheath, the outer with large, thin-walled parenchyma cells; arm cells present in some species; mesophyll with large intercellular spaces; bulliform cells large, occupying a major portion of the blade in transverse section.

Spikelets: Spikelets 2- to many-flowered, with reduction above or below the fertile florets; perfect or unisexual

Flowers: Lodicules 2 or 0, many-nerved; stamens 2 or 3; stigmas 2

Embryo: P + P P

Cytology: x = 12

Photosynthetic Pathway: C₃

Distribution: Mostly grasses of shaded, warm woodlands and tropical forests of the Americas, Africa, and Asia

arica, ariu Asia

SYSTEMATICS

The subfamily consists of about 13 genera and only 30 or so species. It is represented in North America by a single tribe containing a single genus, *Chasmanthium*. The subfamily is clearly bambusoid in its affinities and it is merged with Bambusoideae in some modern treat-ments. Others place the genera in Arundinoideae.

In recent literature, the subfamily and tribe names were sometimes incorrectly spelled Centostecoideae.

TRIBE: CENTOTHECEAE

The characters are those of the single genus below.

CHASMANTHIUM. Wild oats, spangle grass, broadleaved uniola. Broad-leaved perennials, often rhizomatous. Culms herbaceous, to 1.5 m tall. Leaf blades broad, flat. Inflorescence an open to contracted panicle. Spikelets 2- to many-flowered (the lower 1-6 sterile), bisexual, laterally compressed, disarticulating above the glumes and between the florets. Glumes 2, 3- to 7-nerved; lemmas 5- to many-nerved, awnless; palea 2-keeled, winged. Lodicules 2; stamens 1 [3]; stigmas 2, red. A genus of 5 or 6 species, of the southeastern and southwestern United States and

adjacent Mexico. H & C treated it as part of *Uniola* and placed the genus in their tribe Festuceae. *C. latifolium* is probably the most conspicuous species; it is increasingly popular as an ornamental. Other species may be confused with plants of the genus *Diarrhena*.

CHASMANTHIUM AND UNIOLA

Feature	Chasmanthium	Uniola
Habitat	Mesic forests Coa	stal sand dunes
Stamen Number	One	Three
Spikelet Color	Greenish	Straw-colored
Disarticulation	Above glumes	Below glumes
Embryo Type	P + P P	P - P F
Leaf Anatomy	Bambusoid	Chloridoid
Chromosomes	x = 12, 24	x = 20

SELECTED REFERENCES

Soderstrom, T. R. 1981. The grass subfamily Centostecoideae. Taxon 30(3): 614-616.

Tenorio, E. 1976. The subfamily Centostecoideae (Gramineae). Ph.D. dissertation. University of Maryland.

CHASMANTHIUM

Clark, L. G. 1990. A new combination in *Chasmanthium* (Poaceae). Ann. Missouri Bot. Gard. 77: 601.

Yates, H. O. 1966. Revision of grasses traditionally referred to *Uniola*, II. *Chasmanthium*. Southw. Nat. 11(4): 415-455.

Yates, H. O. & R. B. Channell. 1973. Nomenclatural history and taxonomic status of the Linnaean genus *Uniola* (Gramineae). J. Tennessee Acad. Sci. 48(1): 12-15.

3.05 - SUBFAMILY POÖIDEAE

TECHNICAL DESCRIPTION

Habit: Annual or perennial herbs

Root Hairs: Equal

Leaf Epidermis: Relatively simple; bicellular microhairs absent (except in some Stipeae); silica cells round, elliptical, solitary or paired with cork cells; stomata low, dome-shaped or with parallel-sided subsidiary cells

Leaf Anatomy: Vascular bundle sheath usually double; chlorenchyma irregular; non-kranz anatomy

Inflorescence: Typically a panicle, rarely a spike or raceme

Spikelets: Spikelets 1- to many-flowered, laterally compressed or terete; florets 1 to many, the upper usually reduced or aborted; disarticulation usually above the glumes and between the florets; lemmas 5- to many-nerved; palea typically 2-keeled

Flowers: Lodicules 2; stamens usually 3; stigmas

Embryo: F + F F

Cytology: x = 7, often relatively large (x = 8, 9 or 10 in Meliceae; x = 7 and 11 in Stipeae); no persistent nucleoli

Photosynthetic Pathway: C₃

Distribution: Herbaceous grasses, primarily of the cool and temperate regions of the world or of the alpine areas of the tropics and subtropics.

SYSTEMATICS

This subfamily, called Festucoideae by Hitchcock and Chase, was recognized by all of the classical workers, but in a much broader sense than we see it now defined. The other genera included by H & C now reside in every other subfamily, except Panicoideae.

FATE OF H & C'S SUBFAMILY?

I. Tribes moved to other subfamilies:

Bambuseae →	Bambusoideae
Chlorideae →	Chloridoideae
Oryzeae →	Oryzoideae
Zizanieae →	Oryzoideae
Zoysieae →	Chloridoideae

II. Portions of tribes moved:

Agrostideae →	Arundinoideae
Agrostideae →	Chloridoideae
Agrostideae →	Aristidoideae

Aveneae →ArundinoideaeAveneae →DanthoniodeaeFestuceae →ArundinoideaeFestuceae →Chloridoideae

III. Tribes remain more or less intact:

Agrostideae = Aveneae: Alopecurinae Aveneae = Aveneae: Aveninae Hordeae = Triticeae Phalarideae = Aveneae: Phalarinae

IV. New tribes carved out of old ones:

Aristideae out of Agrostideae
Bromeae out of Festuceae
Danthonieae out of Aveneae
Hainardieae out of Hordeae
Meliceae out of Festuceae
Nardeae out of Hordeae
Stipeae out of Agrostideae

TRIBE: DIARRHENEAE

The tribe consists of the single genus described below.

DIARRHENA. Perennial herbs. Inflorescence a few-flowered panicle, its branches often drooping. Spikelets 3- to 5-flowered, bisexual, disarticulating above the glumes and between the florets. Glumes 2, the first 1-nerved, the second 3-nerved; lemmas 3-nerved, awnless; palea 2-nerved. Lodicules 2; stamens [1] 2 or 3; stigmas 2; fruit a shiny, turgid, beaked achene. The tribe consists of the 5 Old World and New World species of this genus. In North America, we see *Diarrhena americana*, a grass of rich or moist woods in the central and eastern states.

TRIBE: BRACHYELYTREAE

This tribe consists of the single genus described below.

BRACHYELYTRUM. Perennial, rhizomatous herbs. Culms herbaceous, to 1 m; internodes solid. Inflorescence a few-flowered panicle. Spikelets terete to dorsally compressed, 1-flowered, disarticulating above the glumes. First glume minute to absent; second glume short, awned or awnless. Lemma 5-nerved, awned. Lodicules 2; stamens 3; stigmas 2. This is the only genus in the tribe. It consists of 2 species, one in Japan and Korea; *B. erectum* is native to moist or rocky woods in eastern North America.

TRIBE: POEAE

Annual or perennial herbs. Inflorescence a panicle, rarely a raceme or spike. Spikelets 2- to many- [1-] flowered, laterally compressed, bisexual [rarely unisexual]. One or both glumes shorter than the lemmas; lemmas 5- to many-nerved, awned or

awnless, its apex variable.

Festuca Sect. Vulpia

FESTUCA. Fescue, fescue grass. Caespitose, rhizomatous, or stoloniferous perennial herbs. Culms to 2 m tall, internodes hollow or soild. Leaves mostly basal; sheaths open. Inflorescence an open to contracted panicle. Spikelets 2- to several-flowered, laterally compressed, bisexual, disarticulating above the glumes and between the florets. Glumes 2, unequal, 1- to 3-nerved, shorter than the florets; lemmas similar to glumes in texture or much firmer, typically [3-] 5- to 7-nerved, awned from an entire to minutely bifid apex or awnless; palea relatively long, apically notched. Lodicules 2; stamens 3; stigmas 2; flowers open-pollinated.

A genus of about 360+ species of temperate and mountainous regions worldwide. Of economic importance as a source of fodder, lawn grasses, and major weedy species. Common species include *F. ovina* (sheep fescue), *F. arundinacea* (tall or alta fescue), *F. idahoensis* (Idaho fescue), *F. pratensis* (meadow fescue), and *F. rubra* (red fescue).

The annual fescues (*Festuca* sec. *Vulpia* in H & C) are now most often treated as belonging to the segregate genus *Vulpia*. The table below presents the name changes in the North American species. You will note that several species have disappeared into synonymy.

ANNUAL FESTUCA TO VULPIA

Combination in Vulpia

F. arida → V. microstachys var. m. F. confusa → V. microstachys var. confusa V. bromoides F. dertonensis → V. microstachys var. ciliata F. eastwoodiae → F. grayi → V. microstachys var. ciliata V. myuros var. hirsuta F. megalura → V. microstachys var. m. F. microstachys → F. myuros → V. myuros var. m. F. octoflora var. glauca → V. octoflora var. g. F. octoflora var. hirtella → V. octoflora var. h.

F. octoflora var. glauca → V. octoflora var. g.
F. octoflora var. hirtella → V. octoflora var. h.

F. octoflora var. octoflora → V. octoflora var. o.
F. octoflora var. tenella → V. octoflora var. t.
F. pacifica var. p. → V. microstachys var. pauciflora
F. pacifica var. simulans → V. m. var. pauciflora
F. reflexa → V. microstachys var. pauciflora
F. sciurea → V. microstachys var. confusa

V. microstachys var. confusa

VULPIA. Annual fescue. Tufted annuals [rarely perennial] herbs. Culms to 9 dm tall, the internodes solid or hollow. Inflorescence a contracted or open spike-like panicle. Spikelets 2- to many-flowered, laterally compressed, bisexual, disarticulating above the glumes and between the florets. Glumes 2, very unequal, 1- to 3-nerved, shorter than the florets; lemmas tapered, firmer than the glumes, 3- to 5-nerved (often inconspicuously so), awned or acuminate; palea relatively long, apically notched. Lodicules 2; anthers 1 [3]; stigmas 2; flowers cleistogamous.

A genus of about 23 species widespread in the temperate regions of the world where they are often weedy. Included in *Festuca* by H & C. Common species include *V. octoflora* (sixweeks fescue), *V. myuros*, and

V. bromoides. Some agrostologists, myself included, prefer to merge this genus with *Festuca*, but this is not a popular view.

COMPARISON OF FESTUCA AND VULPIA

Festuca	Vulpia
Perennial [annual]	Annual [perennial]
Stamens 3 [1]	1 [3]
Pollination open [closed]	closed [open]

LOLIUM. Rye grass. Caespitose annuals or perennial from rhizomes or stolons. Culms herbaceous, to 1+ m tall, the internodes hollow. Inflorescence a single spike (spikelets attached edgewise to rachis). Spikelets several-flowered, laterally compressed, bisexual, disarticulating above the glume and between the florets. Glume 1 (lower missing), except in the uppermost spikelets; lemmas 5- to 9-nerved, awned or awnless; palea relatively long, often ciliate. Lodicules 2, stamens 3; stigmas 2.

A genus of 8-10 Old World species, especially African and Eurasian. Of economic significance because of fodder, lawn grasses, and common weeds. Formerly placed in Triticeae because of its inflorescence type; *Lolium* now treated as a close relative of *Festuca* because of its interfertility. Recent research suggests it should be merged with that genus. *Lolium perenne* is an important pasture grass; *L. temulentum* (darnel) is commonly toxic because of a fungal infection.

PUCCINELLIA. Alkali grass. Annuals or perennials herbs. Culms herbaceous, to 1 m tall. Leaves sheaths open [rarely closed]. Inflorescence an open or congested panicle. Spikelets small, several-flowered, laterally compressed, bisexual, disarticulating above the glumes and between the florets. Glumes 2, very unequal, shorter than the florets, first 1-nerved and the second 3-nerved; lemmas with 5 weak or strong, parallel nerves; palea equalling or exceeding the lemma. Lodicules 2; stamens 3; stigmas 2.

A genus of about 80 species native to the north temperate zone of the Old and New World, especially North America; often of wet or marshy, especially alkaline sites. A few species listed in H & C have been transferred to *Glyceria* and *Torreyochloa* by some workers. *P. airoides* (Nuttall's alkali grass) is an important forage grass in some areas; otherwise, the genus is of little direct economic importance.

TORREYOCHLOA. Stoloniferous or caespitose perennials. Culms herbaceous, to 5 dm tall. Leaf sheaths open. Inflorescence a panicle. Spikelets 3- to 7-flowered, bisexual, laterally compressed, and disarticulating above the glumes and between the florets. Glumes 2, very unequal, 1- or 3-nerved; lemmas 5- to 7-nerved, awnless; palea relatively long. Lodicules 2; stamens 3; stigmas 2.

A genus of 4 species native to northern Asia and North America, often found in wet meadows and aquatic sites. The genus is doubtfully distinct from *Glyceria*.

POA. Blue grass, mutton grass. Annuals or perennials, many rhizomatous. Culms herbaceous, to 1.5m tall, the internodes hollow. Leaf blade with a bow-shaped tip. Inflorescence an open to congested panicle, sometimes reduced to a raceme. Spikelets small, [1-]

2- to 10-flowered, laterally compressed, bisexual or infrequently unisexual (species dioecious), and disarticulating above the glumes and between the florets. Glumes 2, \pm equal, 1- or 3-nerved; lemmas typically 5-nerved, keeled, awnless, its base glabrous or with a web of fine, cottony hairs; palea 2-nerved. Lodicules 2; stamens 3; stigmas 2.

A large and notoriously difficult cosmopolitan genus of about 500 species, found typically in grasslands and meadows, but in a variety of other habitats as well; many are weedy. Recent treatments have often resulted in reducing the number of taxa, certainly a step in the right direction. *P. pratensis* (Kentucky bluegrass) may well be the most important perennial pasture grass in North America; *P. fendleriana* (mutton grass) is a very important pasture grass in the Rocky Mountain region; *P. compressa* (Canada bluegrass) is often used as a lawngrass.

BRIZA. Quaking grass, rattlesnake grass. Tufted annuals or perennials. Culms herbaceous, ours to 0.5 m tall. Inflorescence an open panicles, the pedicels slender and often drooping. Spikelets several-flowered (florets crowded and spreading at right angles to the rachilla), bisexual, laterally compressed, and disarticulating above the glumes and between the florets. Glumes 2, ± equal, thin and papery, rounded, 3- to 15-nerved; lemmas similar to glumes in texture, 7- to 15-nerved, awnless; palea 2-nerved. Lodicules 2, stamens 3; stigmas 2.

A genus of about 16-20 species native to Europe, Mexico, and principally South America. Three European species are adventive in North America. They come in three sizes, *B. maxima*, *B. media*, and *B. minor*. The first species is often found as a roadside weed and it is gathered up for dried arrangements.

DACTYLIS. Orchard grass. Tall, densely caespitose perennials. Culms herbaceous, to 2 m tall. Leaf sheaths keeled and closed. Inflorescence a panicle, with secund spikelets clumped at ends of panicle branches. Spikelets 2- to 5-flowered, bisexual, laterally compressed, and disarticulating above the glumes and between the florets. Glumes 2, shorter than or equaling the florets, 1- or 3-nerved; lemmas 5-nerved, awned or awnless; palea 2-nerved and 2-keeled. Lodicules 2; stamens 3; stigmas 2.

A monotypic genus (or 3-5 species if you are a splitter), native to temperate Eurasia. *D. glomerata* is used as a pasture and fodder grass. It is also a major weed.

CYNOSURUS. Dogtail. Caespitose annuals or perennials. Culms herbaceous, to 1 m tall. Inflorescence a head-like or cylindric, spike-like panicle of two very heteromorphic fertile and sterile spikelets. Fertile spikelets sessile, 1- to 5-flowered, bisexual, laterally compressed, and dis-articulating above the glumes and between the florets. Glumes 2, ± equal; lemmas 5-nerved, awned; palea relatively long; 2-nerved. Lodicules 2; stamens 3; stigmas 2. Sterile spikelets pedicellate, its awns and lemmas rigid, lanceolate, 1-nerved, and ± concealing the fertile ones.

A genus of 4-8 species native to Eurasia and Africa. Two species are adventive in North America. *C. echinatus* is an annual weed of disturbed, open ground. *C. cristatus*, also weedy, has been used as a pasture grass.

LAMARCKIA. Goldentop. Caespitose annual. Culms herbaceous, to 2 dm tall. Leaf sheaths keeled and closed for 2/3 their length. Inflorescence a contracted panicle of drooping fascicles of two very heteromorphic fertile and sterile spikelets. Terminal spikelets in each fascicle bisexual (with a single fertile floret and a stipitate rudimentary one), laterally compressed, and disarticulating below the glumes. Glumes 2, ± equal, 1-nerved; lemmas papery, 2-lobed, awned; palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2. Staminate or sterile spikelets with 3- to 6-flowered, the lemmas awnless.

A monotypic Mediterranean and Asian genus. *L. aurea* is adventive in Texas, Arizona, and California. It is attractive enough to be an ornamental, but I have never seen it offered in nurseries.

TRIBE: BROMEAE

This tribe, traditionally included in Poeae (= Festuceae of older systems), is separated from it on the basis of microcharacters, such as starch grain type and features of the gynoecium. The ovary is hairy, with a con-spicuous apical appendage. Our only representative in North America is the genus below.

BROMUS. Brome grass, rescue grass, chess, ripgut. Annuals or rhizomatous or stoloniferous perennials. Culms herbaceous, to 2 m tall, the internodes hollow [rarely solid]. Leaf sheaths closed. Inflorescence an open to contracted panicle, sometimes reduced to a raceme or even 1- to a few spikelets. Spikelets several-flowered, laterally compressed, bisexual, disarticulating above the glumes and between the florets. Glumes 2, typically very unequal, 1- to 5-nerved, usually awnless; lemmas 5- to several-nerved, usually awned from a bifid apex; palea 2-nerved, often adnate to the caryopsis. Lodicules 2; stamens 1-3; stigmas 2; ovary hairy, with an apical appendage and lateral styles.

A large and complex genus of about 150 species native to a variety of habitats in temperate regions of the Old World and New World, and in cooler, mountainous regions of the tropics and subtropics. It is of economic significance as a source of fodder, hay, and major weedy species. Five sections are recognized. They are sometimes treated as separate genera.

The Sections of Bromus

Ceratochloa. Native annuals, biennials, and perennials of the Pacific and mountain states; spikelets large, distinctly laterally compressed; glumes and lemmas keeled. *B. carinatus* (California brome), *B. marginatus*, and *B. unioloides* (= *B. catharticus* in H & C) (rescue grass).

Bromopsis. Native perennials of woodlands and grass-lands; panicles mostly open, lemmas rounded. *B. inermis* (smooth brome).

Bromium. Annual Mediterranean weeds of the grainfields; glumes and lemmas comparatively broad. *B. mollis* (soft chess), *B. brizaeformis* (rattlesnake chess), *B. secalinus* (chess), *B. commutatus* (hairy chess), and *B. japonicus* (Japanese brome).

Bromus (= **Eubromus** in H & C). Annual

Mediterranean weeds of disturbed sites; glumes and lemmas narrow, long-awned; callus sharp; the ripgut grasses are mechanically injurious to cattle. $B.\ rubens$ (foxtail chess), $B.\ tectorum$ (downy chess), and $B.\ diandrus$ (= $B.\ rigidus$ in H & C) (ripgut).

Neobromus. Introduced South American annual; lemmas lanceolate, deeply bifid, the awn twisted and geniculate. *B. berterianus* (Chilean chess) occurs in Oregon, California, and the Southwest.

EVOLUTION IN BROMUS*

	Bromposis	Ceratochloa	Neobromus
14	LL <i>B. ciliatus</i>	$A_1A_1 \times B_2B_2$	$B_3B_3 \times B_4B_4$
	B. anomalus	•	▼
		•	•
		•	•
28	LLLL	$A_1A_1B_2B_2$	$B_3B_3B_4B_4$
		▼	•
		▼	•
		•	•
42	LLLLLL	$A_1A_1B_2B_2B_2B_2$ <i>B.</i> catharticus	A ₂ A ₂ B ₃ B ₃ B ₄ B ₄ B. berterianus
	•	•	▼
	•	▼	▼
	•	•	•
56	•	A ₁ A ₁ B ₂ B ₂ B ₂ B ₂ LL <i>B. carinatus</i>	•
		B. marginatus	▼
		•	▼
		•	▼
		▼	•
84		$A_1A_1A_2A_2B_2B_2B_2B_2B_3B_3$ B_4B_4 B. arizonicus	•
		B. arizonicus	*After Stebbins (1981)

TRIBE: AVENEAE

This group is conceptually difficult. Hitchcock and Chase recognized a relatively large assemblage of grasses characterized by a paniculate inflorescence and spikelets with elongate glumes, at least the second one being as long as the lowest lemma. All of the florets were typically fertile, except for the uppermost. The lemmas were usually awned from the base, back, or from a bifid apex. The genera seemed especially arbitrary. Other workers have argued that the group is nothing more than a segment of a larger reduction series involving *Phalaris* and its relatives with reduced florets below the single fertile one above and *Agrostis* and its relatives with a single fertile floret.

I have tended to accept the more recent views of the group and to recognize a single large tribe (Aveneae) that included the Aveneae, Phalarideae, and Agrostideae of Hitchcock and Chase. However, recent cluster analysis studies suggest that there is some justification for the view that there are three tribes, but not as described by H & C. The summary below is a compromise, recognizing the three major "lumps" at the subtribal level.

SUBTRIBE: AVENINAE

The subtribe contains most of the genera of H & C's Aveneae. Typical features include open to contracted panicles of spikelets with 2-several florets whose glumes are as long as or longer than the first floret, often all of them. The lemmas usually have a twisted, geniculate awn arising from the back or apex. The rachilla is often prolonged beyond the upper floret as a slender stalk, sometimes bearing a rudimentary floret

AVENA. Oats, wild oats. Caespitose annuals. Culms herbaceous, to 1 m tall. Inflorescence an open panicle, the branches capillary and pendulous, sometimes reduced to a raceme or even a solitary spikelet in depauperate material. Spikelets large, typically 2- or 3-flowered, bisexual, laterally compressed, disarticulating above the glumes and between the florets (except in cultivated oats). Glumes 2, \pm equal, 3- to 11-nerved, as long as or longer than the florets, awnless; lemmas rounded, 5- to 7-nerved, with a stout, geniculate awn (reduced or absent in cultivated oats); palea 2-nerved. Lodicules 2; stamens 3; stigmas 2.

A genus of 15-30 species native to the Old World. *A. fatua* var. *sativa* is cultivated oats; *A. fatua* var. *fatua* is one of several important weeds called wild oats.

SUMMARY OF AVENA SPECIES

Ploidy: Species Comment

Diploids [2n = 2x = 14]

A. strigosa (sand oat) Fodder plant; now weedy A. brevis (short oats) Grown in s. Europe for fodder

Tetraploids [2n = 4x = 28]

A. barbata (slender oat)

Weedy

A. abyssinica (Abyssinian oat) Wild/cultivated forms

Hexaploids [2n = 6x = 42]

A. byzantina (red oat)
A. fatua (wild oats)
A. nuda (naked oat)
A. sativa (cultivated oat)
A. sterilis (animated oat)

Eaten by livestock and us
Pernicious weed
Grain crop in China
Widely cultivated
Fodder and ornamental

ARRHENATHERUM. Tall oat grass. Caespitose perennials. Culms herbaceous, to 2 m tall; corms sometimes present. Inflorescence a narrow panicle. Spikelets typically 2-flowered (upper perfect, the lower larger and staminate), laterally compressed, bisexual, disarticulating above the glumes (2 florets falling together); rachilla extended as bristle beyond uppermost floret. Glumes 2, very unequal, the first 1-nerved and the second 3-nerved, awnless; lemmas 5-to 9-nerved, awnless or with a dorsal awn; palea relatively long. Lodicules 2; stamens 3; stigmas 2.

A genus of 4-6 species native to Europe and the Mediterranean. *A. elatius* (tall oat grass) is planted for pasture and has escaped in many areas of North America.

HELICTOTRICHON. Spike-oat, alpine-oat. Caespitose perennials. Culms herbaceous, to 1.5 m tall. Inflorescence a contracted panicle. Spikelets large, 2-to 7-flowered, bisexual, laterally compressed, disarticulating above the glumes and between the florets. Glumes 2, equal to or shorter than the florets, 1- to 5-nerved; lemmas 5- to 7-nerved, apex toothed, with a stout, twisted, geniculate dorsal awn; palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2.

A genus of about 90 species native to the Old and New World. Two are native to North America (*H. hookeri* and *H. mortonianum*); one is introduced from Europe (*H. pubescens*).

AIRA. Silver hair grass. Delicate, caespitose annuals. Culms herbaceous, to 0.25 m tall. Inflorescence an open or panicle. Spikelets 2-flowered, laterally compressed, bisexual, disarticulating above the glumes and between the florets. Glumes 2, \pm equal, as long as or longer than the florets, 1- or 3-nerved, lemmas 5-nerved, a geniculate, hair-like awn arising from below the middle, apex with 2 slender teeth or setae; palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2.

A genus of 8-10 species native to the northern and southern temperate regions of the Old World. *A. caryophyllea*, *A. elegans*, and *A. praecox* are adventive in North America.

DESCHAMPSIA. Hair grass. Mostly caespitose perennials; a few are annuals. Culms herbaceous, to 1 m tall. Inflorescence an open or congested panicle, often with capillary branches. Spikelets small, 2-flowered, laterally compressed, bisexual, disarticulating above the glumes and between the florets, the rachilla hairy and prolonged above the upper floret (sometimes with a rudiment at its apex). Glumes 2, ± equal, the first 1-nerved and the second 3-nerved, as long as or longer than the lower floret; lemmas 5- to 7-nerved (often obscurely so), its apex several toothed or cleft, and with a geniculate awn from or below the middle; palea relatively long. Lodicules 2; stamens 3; stigmas 2.

A genus of about 40 species native to the temperate and cooler portions of the Old and New World. *D. danthonioides*, the only annual species in North America, is common in the West; *D. caespitosa* (tufted hair grass) occurs in mountain meadows in the same region.

TRISETUM. Trisetum. Mostly caespitose perennials. Culms herbaceous, to 1.5 m. Inflorescence a contracted panicle. Spikelets 2-flowered [rarely 3- or 4-flowered], laterally compressed, bisexual, disarticulation above or below the glumes, or not disarticulating, the rachilla prolonged above the uppermost floret. Glumes 2, equal or unequal, the first 1- to 3-nerved and the second 1- to 5-nerved; lemmas 5- to 7-nerved, its apex bifid, the teeth awned, with a straight or bent awn from the base of the cleft; palea relatively long, with two apical setae. Lodicules 2; stamens 3; stigmas 2.

A genus of about 75-85 species of the temperate and cooler regions of the Old and New World. Of economic importance as a source of pasture and fodder grasses. *T. spicatum* (spike trisetum) occurs in the Rocky Mountains and in the West.

SPHENOPHOLIS. Wedgescale. Caespitose annuals and perennials. Culms herbaceous, to 1+ m tall. Inflorescence a contracted panicle. Spikelets 2- or 3-flowered, laterally compressed, bisexual, disarticulating below the glumes. Glumes 2, dimorphic (first narrow and 1- [3-] nerved and the second broad and 3- to 5-nerved); lemmas faintly 5-nerved, awnless or less often awned; palea relatively long. Lodicules 2; stamens 3; stigmas 2.

A genus of 4 or 5 species native to North America and the West Indies. *S. obtusata* (prairie wedgescale) is widespread in North America.

KOELERIA. June grass. Caespitose annuals or perennials. Culms herbaceous, to 1+ m tall. Inflorescence a spike-like panicle. Spikelets 2- to 4-flowered, laterally compressed, bisexual, disarticulating above the glumes and between the florets, rachilla extended as a bristle beyond the uppermost floret. Glumes 2, ± equal, as long as or shorter than the florets, dimorphic (first acute, 1-nerved and the second broader, longer, obscurely 3-to 5-nerved); lemmas 3- to 5-nerved, awnless or awned from a bifid apex; palea relatively long, apically notched, 2-nerved. Lodicules 2; stamens 3; stigmas 2.

A genus of about 60 species native to the temperate regions of the Old and New World. The annual species have been treated as the genus *Lophochloa*. *K. macrantha* (= *K. cristata* in H & C) is native to prairies and wooded areas over much of North America. *K. phleoides* is a small, introduced annual.

HOLCUS. Velvet grass. Caespitose perennials. Culms herbaceous, weak, succulent, to 1 m tall. Leaves velvety-pubescent, especially the sheaths. Inflorescence a contracted panicle. Spikelets 2-flowered (lower perfect, the upper staminate or neuter), laterally compressed, bisexual, rachilla prolonged above the uppermost fertile floret, disarticulating below the glumes. Glumes 2, as long as or longer than the florets, the first 1-nerved and the second 3-nerved; lemmas faintly 3- to 5-nerved, the upper one with a short, hooked awn from near its apex; palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2. A genus of 6-8 species native

to Eurasia and Africa. *H. lanatus* and *H. mollis* are adventive in North America.

BECKMANNIA. Slough grass. Annuals with thick culms. Inflorescence a panicle of appressed or ascending spikes. Spikelets 1- or 2-flowered, ± orbicular, laterally com-pressed, sessile, disarticulating below the glumes, the rachilla often prolonged. Glumes 2, equal, broad, inflated, 3-nerved, keeled, and apiculate at apex. Lemma ± equal to glumes, narrow, 5-nerved, tapering to a slender tip. Palea narrow, shorter than lemma. Lodicules 2; stamens 3; stigmas 2.

A genus of two species native to Eurasia and North America. *Beckmannia syzigachne*, American slough grass, is native to marhes and wet sites over much of the northern and western U. S. It is sometimes frequent enough to use for hay or forage.

SUBTRIBE: PHALARIDINAE

As here defined, the subtribe is equivalent to the Phalarideae of H & C. The group is defined by its spikelets with a single fertile floret with 1 or 2 staminate or sterile florets below it, these sometimes reduced to inconspicuous, scale-like rudiments.

ANTHOXANTHUM. Sweet vernal grass. Annuals or perennials, pleasantly fragrant because of coumarin. Culms herbaceous, to 1 m tall. Inflorescence a contracted to spike-like panicle. Spikelets with a single fertile floret subtended by two sterile ones, laterally compressed, bisexual, disarticulating above the glumes (the three florets falling as a group). Glumes 2, very unequal, the first 1-nerved, the second 3-nerved, longer than the florets; sterile lemmas 3-nerved, hairy, awned from a notched apex; fertile lemma 3- to 5-nerved, glabrous except at apex, awnless; palea 1- or 3-nerved. Lodicules 0; stamens 2; stigmas 2.

A small genus of about 4 species native to the Old World. Two species, *A. odoratum* (sweet vernal grass) and *A. aristatum* are adventive in North America. The genus is doubtfully distinct from *Hierochloë*.

HIEROCHLOË. Holy grass, sweet grass. Caespitose or rhizomatous perennials, pleasantly fragrant because of coumarin. Culms herbaceous, to 1+ m tall. Inflorescence an open to contracted panicle. Spikelets with a single fertile floret subtended by two staminate ones, laterally compressed, bisexual, disarticulating above the glumes (the three florets falling as a group). Glumes 2, equal, the first 1- to 5-nerved and the second 3- to 5-nerved, as long as or slightly shorter than the florets; lemmas of staminate florets awnless or with a short awn from a notched apex, 5-nerved; lemma of fertile floret 3- to 5-nerved, awnless; palea 1- or 3-nerved. Lodicules 2; stamens 2 (bisexual florets) or 3 (staminate florets).

A genus of 20-30 species native to the temperate and cooler regions of both hemispheres. Three species are found in North America. *H. alpina* occurs in from the New England states across Montana; *H. odorata* is found through much of the northern and western states; and *H. occidentale* coours along the Pacific Coast. Schouten & Veldkamp (1985) proposed merging this genus with *Anthoxanthum*, but their work seems to have gone largely unnoticed.

PHALARIS. Canary grass, reed canary grass.

Caespitose annuals or perennials, often from rhizomes. Culms herbaceous, to 2 m tall. Inflorescence typically a spike-like panicle. Spikelets with a single bisexual floret subtended by two reduced (sometimes very tiny!) scale-like florets, laterally compressed, disarticulating above or below the glumes or not disarticulating. Glumes 2, ± equal, as long as or longer than the florets, usually with a winged dorsal keel; sterile lemmas reduced to small scales; fertile lemma 5-nerved, awnless; palea relatively long, 1- or 2-nerved. Lodicules 2; stamens 3; stigmas 2.

A genus of 15 or 16 native to Eurasia, Africa, and the New World. Five are native to North America; four are adventive. *Ph. arundinacea* (reed canary grass) is an important pasture grass and weed; *Ph. canariensis* is one of the standard ingredients in commercial birdseed mixtures.

SUBTRIBE: ALOPECURINAE

This subtribe contains some, but by no means all, of the genera of H & C's Agrostideae. The plants are primarily of the temperate and Arctic regions. The inflorescence is typically a panicle. The spikelets are 1-flowered, small, and mostly laterally compressed. The glumes tend to be longer than the floret. The lemmas are thin, 3- to 5-nerved, and rarely awned.

AMMOPHILA. Beach grass, European beach grass. Coarse, rhizomatous perennials. Culms herbaceous, to 1.5 m tall. Inflorescence a dense, spike-like panicle. Spikelets large, 1-flowered, laterally compressed, bisexual, rachilla prolonged as a hairy bristle beyond the floret, disarticulat-ing above the glumes. Glumes 2, ± equal, the first 1-nerved and the second 1- to 3-nerved, as long as or longer than the floret; lemma 5-nerved, awnless, with long hairs at the base; palea relatively long, 4- to several-nerved. Lodicules 2; stamens 3; stigmas 2.

A genus of 2-4 species native to North America and Europe. *A. breviligulata* (American beach grass) is native to the East Coast and around the Great Lakes; *A. champlainensis* is native to the Northeast; *A. arenaria* (European beach grass) was introduced to stabilize sand dunes along the Pacific Coast.

CALAMAGROSTIS. Reed grass, bluejoint. Caespitose, rhizomatous, or stoloniferous perennials, often of wet and marshy sites. Culms herbaceous, sometimes reed-like, to 2 m tall. Inflorescence a panicle, typically contracted. Spikelets 1-flowered, laterally compressed, bisexual, disarticulating above the glumes, rachilla often extended beyond the single floret as a slender bristle. Glumes 2, ± equal, longer than the floret, the first 1-nerved and the second 3-nerved; lemma 3- or 5-nerved, its apex notched or toothed, bearing a slender, geniculate dorsal awn; callus long-hairy; palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2.

A genus of over 230 species native to the temperate and cooler regions of the Old and Nedw World. A number of species reported for North America, all but one of them native. *C. canadensis* (bluejoint), which occurs over much of the northern part of the continent, is an important forage grass. *C. purpurascens* (purple reedgrass) is found over much of the western United States; *C. nutkaensis* (Pacific reedgrass) is native along the Pacific coast from Alaska to California; *C. inexpansa* (northern

reedgrass) ranges from Greenland to California.

AGROSTIS. Bent grass, bent, redtop, tickle grass. Annuals and caespitose, rhizomatous, or stoloniferous perennials. Culms herbaceous, to 1 m tall. Inflorescence an open to contracted panicle. Spikelets small (sometimes distressingly so), 1-flowered, laterally compressed, bisexual, disarticulating above the glumes. Glumes 2, \pm equal, the first typically 1-nerved and the second 1- [3-] nerved, as long as or longer than the floret; lemma 3- or 5-nerved, glabrous or hairy at base, awned dorsally from or below the middle or awnless; palea typically small or even absent. Lodicules 2; stamens 3; stigmas 2.

A complex genus of over 200 species native to the temperate and cooler regions of both hemispheres. Several of our North American representatives are of considerable economic importance as forage grasses, lawn grasses, and as the substrate on which some humans use sticks to knock little white balls into small holes in the ground. A number of them are also weedy. A. gigantea (= A. alba in H & C) is redtop; A. hiemalis is winter bentgrass, and A. palustris is the creeping bent used at golf courses.

CINNA. Woodreed. Caespitose or rhizomatous perennials, often of wet, shady sites. Culms herbaceous, to 1.5 m tall. Inflorescence an open or congested panicle. Spikelets 1-flowered, laterally compressed, bisexual, disarticulating below the glumes, the rachilla prolonged above the floret as a stub or bristle. Glumes 2, equal or the first shorter, the lower 1-nerved and the upper 1- or 3-nerved, lemmas 3- to 5- nerved, the mid-nerve extended as a short, straight awn; palea about as long as the lemma, 1-nerved. Lodicules 2; stamen 1 or 2 [3]; stigmas 2.

A genus of 3-4 species native to the temperate regions of both hemispheres. *C. latifolia* (drooping woodreed) is widespread in moist, shaded areas; *C. arundinacea* (stout woodreed) is found in moist woods in the eastern and central portions of North America; *C. bolanderi* is endemic to California.

PHLEUM. Timothy. Caespitose annuals and perennials. Inflorescence an ovoid to cylindrical, spikelike panicle. Culms herbaceous, to 1.5 m tall, the bases sometimes tuberous. Spikelets 1-flowered, strongly laterally compressed, bisexual, disarticulating above or below the glumes. Glumes 2, ± equal, longer than the floret, abruptly narrowed to an awn or mucro, 3-nerved; lemma 5- to 7-nerved, blunt, awnless.

A genus of 10-15 species native to the temperate regions of Eurasia and North America. *Ph. alpinum* is native in mountainous areas; *Ph. pratense* (timothy) is perhaps the leading hay grass of the eastern states.

POLYPOGON. Rabbitfoot grass. Annuals or perennials. Culms herbaceous, often weak and decumbent and rooting at nodes, to 1+ m tall. Inflorescence a soft, dense, contracted panicle (hence the common name). Spikelets small, 1-flowered, ± laterally compressed, bisexual, disarticulating below the glumes. Glumes 2, ± equal, longer than the floret, 1-nerved, bearing a long awn from an entire or notched apex; lemma 5-nerved, its apex typically toothed, awned or awnless; palea relatively long, 2-nerved. Lodicules 2; stamens 1 or 3; stigmas 2.

A genus of 10-20 species native mostly to the

temperate regions of the Old World. Several Eurasian species have been introduced; *P. monospeliensis* (rabbitfoot grass) is a common weed in wet areas across North America. *P. elongatus* is native to Arizona and California.

Lagurus. Hare's tail. Plants annual. Inflorescence a dense, pale, ovoid to oblong capitate panicle. Sheaths and blades pubescent. Spikelets 1-flowered. Glumes thin, narrow, villous, 1-nerved, tapering to a plumose awn-point. Lemma shorter than glumes, thin, glabrous, bearing an awn form its back, its apex awn-tipped. Palea thin, narrow, the two keels ending in minute awns.

One species, *L. ovatus*, native to the Mediterranean. Cultivated as an ornamental, popular in dried arrangements. Sparingly escaped in California and elsewhere.

ALOPECURUS. Meadow foxtail. Annuals and caespitose or rhizomatous perennials. Culms herbaceous, to 1 m tall. Inflorescence a dense, spikelike panicle. Spikelets 1-flowered, laterally compressed, bisexual, disarticulating below the glumes. Glumes 2, their lower edges united, ciliatekeeled, equal, 3-nerved, equaling the floret; lemma 5-nerved, its lower margins joined, awned from below the middle; palea absent or much-reduced. Lodicules 0; stamens 3; stigmas 2.

A genus of 25-35 species native to the northern temperate regions of both hemispheres. Of economic significance as a source of fodder, pasture grasses, and weeds. *A. carolinianus*, a native annual, is widely distributed. *A. pratensis* (meadow foxtail), a European pasture grass, is grown in the northern states.

GASTRIDIUM. Nit grass. Caespitose annuals. Culms herbaceous, to 5 dm tall. Inflorescence a dense, spike-like panicle. Spikelets 1-flowered, laterally compressed, bisexual, disarticulating below the glumes, the rachilla prolonged above the floret as a bristle. Glumes 2, unequal, longer than the floret, 1-nerved; lemma 5-nerved, awnless or awned from below the toothed apex; palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2.

A genus of only two species, both native to western Europe and the Mediterranean. *G. ventricosum* (nit grass) is adventive along the Pacific coast and scattered locations in Texas and the East.

TRIBE: TRITICEAE

"Abandon hope all ye who enter here." (Dante Alighieri, writing on an another distressing topic)

Annuals or perennials. Leaf blades usually auriculate. Inflorescence a balanced spike or rame, its axis continuous or breaking apart at maturity. Spikelets commonly 1-3 per node, sometimes as many as 6. Lemmas 5- to 9-nerved, awned from the tip or awnless. Caryopsis free from or adhering to the anthoecium. Hybridization rampant; polyploidy common. X = 7. The tribe is more or less equivalent to the Hordeae of H & C.

I am tempted to offer Dante's admonition about abandoning all hope to those who enter here. Generic delimitations within the tribe are exceedingly difficult, often appearing quite arbitrary, and they remain

controversial. To give you some flavor of the difference of opinion as to how to treat this group, some workers recommend recognizing a single genus, *Triticum*. Others, including myself, believe that our North American material can be accommodated in about six genera. The most liberal disposition is that we ought to give generic recognition to each distinct genome or combination of genomes, which yields Avogadro's number of segregate (and to my way of thinking, useless) genera.

EVOLVING VIEWS OF TRITICEAE

Bentham (1882). Agropyron, Elymus, Hordeum, Hystrix, Secale, Triticum.

Nevski (1933). Aegilops, Agropyron, Brachypodium, Critesion, Elymus, Elytrigia, Eremopyrum, Hordeum, Psathyrostachys, Secale, Sitanion, Taeniatherum, Triticum,

Hitchcock (1951). Aegilops, Agropyron, Elymus, Hordeum, Hystrix, Lolium, Monerma, Nardus, Parapholis, Scribneria², Secale, Sitanion, Triticum.

Tzvelev (1976). Aegilops, Agropyron, Elymus, Elytrigia, Eremopyrum, Hordeum, Hystrix, Leymus, Psathyrostachys, Secale, Taeniatherum, Triticum.

Baum (1982): Aegilops, Agropyron, Elymus, Elytrigia, Eremopyrum, Hordeum, Hystrix, Leymus, Psathyrostachys, Secale, Taeniatherum, Triticum.

Estes & Tyrl (1982): Agropyron, Elymus, Hordeum, Secale, Triticum.

Gould (1983): Agropyron, Elymus, Hordeum, Hystrix, Secale, Sitanion, Taeniatherum, Triticum.

Löve (1984): Aegilops, Aegilopodes, Agropyron, Critesion, Cylindropyrum, Elymus, Elytrigia, Eremopoa, Eremopyrum, Hordeum, Hystrix, Leymus, Lophopyrum, Pascopyrum, Psathyrostachys, Pseudoroegneria. Roegneria, Secale, Sitanion, Taeniatherum, Thinopyrum, Triticum.

Barkworth (1983, 1985): Aegilops, Agropyron, Critesion, Elymus, Elytrigia, Eremopyrum, Hordeum, Leymus, Pascopyrum, Psathyrostachys, Pseudoroegneria, Secale, Taeniatherum, Thinopyrum, Triticum.

Clayton & Renvoize (1986): Aegilops, Agropyron, Brachypodium, Crithopsis, Elymus, Eremopyrum, Hordeum, Hystrix, Leymus, Psathyrostachys, Secale, Sitanion, Taeniatherum, Triticum.

Watson & Dallwitz (1992): Aegilops, Agropyron, Elymus, Elytrigia, Eremopyrum, Hordeum, Hystrix, Leymus, Lophopyrum, Pascopyrum, Psathyrostachys, Pseudoroegneria, Secale, Sitanion, Taeniatherum, Thinopyrum, Triticum.

Czerepanov (1995): Aegilops, Agropyron, Elymus, Elytrigia, Eremopyrum, Hordeum, Hystrix, Leymus, Psathyrostachys, Secale, Taeniatherum, Triticum.

GENOME-BASED GENERA

Askell Löve and others have argued that the genera of Triticeae should be based on genomes. Here is how that would look.

Genome	Genus
A	Crithodium
AB	Gigachilon
ABD	Triticum
B	Sitopsis
BU	Aegilemma
C	Orrhopygium
CD	Cylindropyrum
D	Patropyrum
DM	Gastropyrum
DMU	Aegilonearum
E	Lophopyrum
EJS	Elytrigia
F	Eremopyrum
G	Festucopsis
H	Critesion
HJNS	Pascopyrum
HS	Elymus
HT	Hordelymus
I	Hordeum
J	Thinopyrum
JN	Leymus
K	Crithopsis
L	Chennapyrum
M	Comopyrum
MU	Aegilops
N	Psathyrostachys
O	Henrardia
P	Agropyron
Q	Heteranthelium
R	Secale
S	Pseudoroegneria
T	Taeniatherum
U	Kiharapyrum
V	Dasypyrum
Z	Amblyopyrum

INTERGENERIC HYBRID

Given the promiscuous nature of the grasses in this group, you will not be surprised to learn that there are many intergeneric hybrids, as seen below.

X Aeailosecale X Aegilotriticum X Agroelymus X Agrohordeum X Agrositanion X Ağrotrisecale X Agrotriticum X Elyhordeum X Elýmordeum X Elymopyrum X Elyleymus X Elysitanion X Élytesion X Leytesion X Pseudelymus X Sitordeum X Triticosecale X Tritordeum

FATE OF H & C's GENERA

The genera recognized by Hitchcock & Chase (1951) appear in the left column. Their current disposition as per various recent monographs is shown on the right. You will note that the concept of some genera, such as Secale and Triticum, has changed little. Some genera, such as Hystrix, have disappeared. In still other cases, H & C recognized a large genus, such as Elymus, and recent workers also recognize an entity called Elymus, but it is more narrowly defined because one or more species have been transferred to segregate genera.

In a few instances, genera are now seen as members of tribes other than Triticeae. They

Aegilops	Aegilops Cylindropyrum Triticum
Agropyron	Agropyron Elymus Eremopyrum Pascopyrum Pseudoroegneria Roegneria Thinopyrum
Elymus	Elymus Elytrigia Leymus Lophopyrum Psathyrostachys Taeniatherum
Hordeum	Hordeum Critesion
Hystrix	Elymus
Lolium	Lolium (Poeae)
Monerma	Hainardia (Hainardieae)
Nardus Parapholis	<i>Nardus</i> (Nardeae) <i>Parapholis</i> (Hainardieae)
Scribneria	Scribneria (Hainardieae)
Secale	Secale
Sitanion	Sitanion Elymus
Triticum	Triticum

AGROPYRON. Crested wheatgrass. Caespitose or rhizomatous perennials. Culms herbaceous, to 1 m tall. Inflorescence a balanced spike, the rachis continuous or shattering at maturity, spikelets ± pectinate, 1 per node, inserted flatwise to axis. Spikelets 3- to 8-flowered, laterally compressed, bisexual, disarticulating above the glumes and between the florets. Glumes 2, ± equal, shorter than the florets, broad or narrow, awned; lemmas 5- or 7-nerved, awned or awnless; palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2.

As defined here, the genus consists of about 15 species native to the Mediterranean area and Asia. Our North American representatives are *A. cristatum* (crested wheatgrass) and *A. desertorum* (desert

wheatgrass). I follow Stebbins and Gould in placing the other species listed in H & C in the expanded version of Elymus. The increasingly popular disposition of these taxa is to place them in a series of segregate genera.

ELYMUS. Wild rye. Caespitose or rhizomatous perennials. Culms herbaceous, to 2 m tall. Inflorescence a balanced spike [compound in some species], the rachis continuous. Spikelets typically [1] 2 or 3 [6] per node; [2-] 3-7 [9-] flowered, laterally compressed, bisexual, disarticulating above the glumes and between the florets. Glumes 2, equal or unequal, 3- to 7-nerved, broad to quite narrow, almost awn-like in some; lemmas 5- to 7-nerved, tapering to an awn [rarely awnless]; palea relatively long, 2-nerved. Lodicules 2; stamens 3, stigmas 2.

A genus of about 150 species widespread in the temperate and cooler regions of both hemispheres. As treated here, the genus includes most of the taxa assigned to *Agropyron* in H & C. Some species are important forage grasses. *E. virginicus* (Virginia wildrye), *E. canadensis* (Canada wildrye), *E. glaucus* (blue wildrye), *E. cinereus*.

Common species transferred from *Agropyron* include *E. smithii* (western wheatgrass), *E. repens* (quack grass), and *E. spicatus* (bluebunch wheatgrass). The genus also includes all of our North American species of *Sitanion* (squirreltail grasses) and *Hystrix* (bottlebrush grasses) that appear in H & C.

TAENIATHERUM. Medusa head. Annuals. Culms herbaceous, to 6 dm tall. Inflorescence a bristly, balanced spike, spikelets paired, the rachis continuous. Spikelets 2-flowered (the lower bisexual and the upper rudimentary), dorsally compressed, disarticulating above the glumes. Glumes 2, subulate, ndurate, 1- or 3-nerved, joined at base, tapering to a stiff awn; lemma narrow, 5-nerved, with a long, flattened awn; palea about as long as lemma, 2-nerved. Lodicules 2; stamens 3; stigmas 2.

A genus of 2-3 species, doubtfully distinct from *Elymus*, native to Eurasia. *T. caput-medusae* [= *Elymus c.-m.* in H & C] is a serious weed in several of the western states, including California.

HORDEUM. Barley, foxtail barley, little barley, squirreltail. Annuals or caespitose perennials. Culms herbaceous, to 1+ m tall. Inflorescence a dense, balanced, often bristly spike, continuous or shattering at maturity. Spikelets typically 3 per node, the central sessile and fertile, the laterals often pedicellate and sterile (all three fertile in H. vulgare). Central spikelet 1-flowered, the rachilla extended above it and bearing a rudiment, laterally or dorsally compressed, bisexual, disarticulating below the glumes or not disarticulating in cultivars. Glumes 2, narrow, rigid, 1-nerved, subulate or awned; lemma dorsally flattened, 5-nerved (often difficult to distinguish), acuminate to an awn or awn-point; palea about as long as lemma, often adnate to caryopsis. Lateral spikelets often reduced to awn-like glumes. Lodicules 2; stamens 3; stigmas 2.

A genus of 25-40 species native to the temperate regions of both hemispheres; a number of them are weedy. A dozen or so species are found in North America, most of them native. H. jubatum (foxtail barley) is native in the West; H. brachyantherum (meadow barley) provides good forage; H. pusillum (little barley) is a native annual found over much of

North America; *H. vulgare* is cultivated barley, whose grains are eaten and sprouted to make malt for the brewing and distilling industries.

SUMMARY OF NORTH AMERICAN HORDEUM

Ploidy Level: Taxon Nativity (Duration)

2n = 2x = 14

H. bulbosum	Mediterranean (P)
H. californicum	Western United States (P)
H. euclaston	South America (A)	
H. intercedens	Western United States (A)
H. marinum ssp. marin	<i>num</i> Mediterranean (A)
H. marinum ssp. gusso		
H. murinum ssp. glauci)
H. pusillum	North America (A)
H. vulgare ssp. vulgare	Cultivated (A))

2n = 4x = 28

2n = 6x = 42

H. arizonicum Southwest United States (A/P) H. murinum ssp. leporinum Mediterranean (A)

[After R. von Bothmer in Shewry, P. R. (1992)]

AEGILOPS. Goat grass. Caespitose or rhizomatous annuals. Culms herbaceous, to 8 dm tall. Inflorescence a single, balanced spike, the spikelets solitary at each node. Spikelets turgid or cylindrical, placed flatwise and fitting into the rachis, 2- to 5-flowered, rounded to \pm laterally compressed, bisexual, disarticulating above or below the glumes, or not disarticulating. Glumes 2, several-nerved, 1- or 3-awned; lemmas usually 5- to 7-nerved, awnless, mucronate, or 1- or 2-awned; palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2.

The genus is completely interfertile with *Triticum* and the two genera are often merged. A genus of 20+ species native to the Mediterranean and Asia. *Ae. cylindrica* (jointed goatgrass) is a widespread weed in the central and southern states; *Ae. ovata* and *Ae. triuncialis* are common weeds in California.

TRITICUM. Wheat, bread wheat. Caespitose annuals. Culms herbaceous, the internodes hollow or solid, to 1 m tall. Inflorescence a thick, balanced spike, the spikelets solitary at each node and attached flatwise to the rachis. Spikelets 2- to 5-flowered, laterally compressed or rounded, bisexual, disarticulating above or below the glumes, or not disarticulating in cultivars. Glumes 2, \pm equal, shorter than the florets, 5- to 11-nerved, mucronate or with 1 or more awns at its apex; lemmas keeled or rounded, glabrous or pubescent, several-nerved, awnless or with 1 or 3 awns.

A genus of about 8 species native to Eurasia. *T. aestivum* (bread wheat) and *T. durum* (durum or macaroni wheat) are the principal economic species.

SUMMARY OF WHEAT SPECIES

Species (Common Name) Genome(s)

Diploids [2n = 2x = 14]

T. boeoticum (wild einkorn wheat)
AA
T. monococcum (einkorn wheat)
AA

Tetraploids [2n = 4x = 28]

Hexaploids [2n = 6x = 42]

T. spelta (spelt wheat)	AABBDD
T. macha (macha wheat)	AABBDD
T. vavilovii (Vavilov's wheat)	AABBDD
T. compactum (club wheat)	AABBDD
T. sphaerococcùm (shot wheat)	AABBDD
T. aestivum (bread`or common wheat)	AABBDD

EVOLUTION OF MODERN HEXAPLOID WHEATS

Phase I: Diploid to Tetraploid

Triticum boeoticum		Aegilops speltoides
(Wild einkorn wheat)		(Goat grass)
[2n = 2x = 14]	Χ	[2n = 2x = 14]
[Genome: AA]		[Genome: BB]
	∇	
	∇	

Sterile F₁ Hybrid [2n = 2x = 14] [Genomes: AB]

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Unreduced Gametes ("Chromosome Doubling")

 ∇ ∇

Triticum dicoccoides (Wild emmer wheat) [2n = 4x = 28] [Genomes: AABB]

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Domestication

 ∇ ∇

Triticum dicoccum (Cultivated emmer wheat) [2n = 4x = 28] [Genomes: AABB]

Phase II: Tetraploid to Hexaploid

Triticum dicoccum (Cultivated emmer wheat) (Goat grass) [2n = 4x = 28] [2n = 2x = 14] [Genomes: AABB] X [Genomes: DD]

Sterile Hybrid [2n = 3x = 21] [Genomes: ABD]

 ∇ ∇

Unreduced Gametes ("Chromosome Doubling")

 ∇ ∇

Triticum aestivum
(Bread wheat)
[2n = 6x = 42]
[Genomes: AABBDD]

 ∇ ∇

Domestication

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Hulled/Free-threshing cultivars

 $\nabla \nabla$ ∇

Recent domestication

 $\begin{array}{c} \nabla \\ \nabla \end{array}$

Genetic engineering

SECALE. Rye. Annuals, sometimes perennials. Culms herbaceous, to 1+ m tall. Inflorescence a balanced spike, the spikelets solitary at the node, and placed flatwise to the rachis. Spikelets typically 2-flowered, laterally compressed, bisexual, disarticulating below the glumes or not disarticulating in cultivars. Glumes 2, narrow, 1-nerved, subulate; lemmas sharply keeled, ciliate on the keels, 5-nerved, tapering to a long awn.

A genus of 5 species native to Eurasia. *S. cereale* (rye) escapes from cultivation.

X TRITICOSECALE. Triticale. The hybrid between wheat and rye, combining desirable features from both of these important crop plants. It is the source of triticale berries that you see in the health food markets.

TRIBE BRACHYPODIEAE

The characters of the tribe are those of the genus below.

BRACHYPODIUM. False brome, purple-brome. Mostly perennial herbs. Inflorescence a series of linear racemes on stiffly erect branches. Spikelets 1 per node, ± sessile, divergent. Spikelets 2- to 20-flowered, ± terete to slightly laterally compressed, disarticulating above the glumes. Glumes 2, 5- to 9-nerved; lemmas herbaceous to thickened at maturity, 7- to 9-nerved, extending into an awn. Lodicules 2; stamens 3; stigmas 2, white.

A genus of 16 species native to the temperate regions of Europe and Asia, especially of woodlands and open grasslands. Two are native to Mexico. *B. distachyon* is a weedy European introduction of scattered sites in the U. S. A.

TRIBE: MELICEAE

A small tribe of grasses often associated with wet sites. Leaf sheaths typically closed. Inflorescence a panicle or raceme. Spikelets several-flowered; disarticulation above or below the glumes. H & C included these genera in their Festuceae. See Section 4 for a key to genera.

MELICA. Onion grass, melic. Perennials. Culms herbaceous, to 1 m tall, bases often swollen into bulb-like corms. Leaf sheaths closed. Inflorescence an open to contracted panicle. Spikelets several-flowered, rounded to laterally compressed, bisexual, the upper florets sterile, over-lapping, and forming a knob-like cluster, disarticulating above the glumes and between the florets. Glumes 2, 1- to 7-nerved, the margins scarious; lemmas thin, 5- to several-nerved, scarious-margined, awnless; palea present, 2-nerved. Lodicules 2; stamens 3; stigmas 2.

A genus of about 60-80 species native to the temperate regions of both hemispheres; about 20 of them are found in North America. *M. mutica* (two-flowered melic) is native to much of the eastern United States. *M. nitens* and *M. mutica* occur in the eastern states; *M. subulata* (Alaska onion grass) occurs from Alaska to California; *M. porteri* (Porter's melic) is found in the Southwest.

GLYCERIA. Manna grass. Rhizomatous, stoloniferous, or caespitose perennials of wet sites. Culms herbaceous, often decumbent and rooting at the nodes, to 1+ m tall. Inflorescence an open to contracted panicle, sometimes reduced to a raceme. Spikelets fragile, linear, awnless, laterally compressed or almost terete, several-flowered, bisexual, disarticulating above the glumes and between the florets. Glumes 2, unequal, 1- [3-] nerved, shorter than the florets; lemmas broad, with [5] 7 [9] conspicuous, parallel nerves, the apex acute, obtuse, or truncate; palea broad, slightly longer than lemma, 2-nerved. Lodicules 2; stamens 2 or 3; stigmas 2.

A genus of about 35-40 species native to the temperate regions of the Old and New World; about 15 of them are found in North America. Some species of the genus have been treated under *Puccinellia* and *Torreyochloa* by recent workers. Several species are important sources of food for humans and waterfowl.

G. striata (fowl manna grass) is probably the most widespread representative of the genus in North America, ranging from Newfoundland to Mexico; G. borealis (northern manna grass) occurs from Newfoundland to Washington and south to California; G. septentrionalis (eastern manna grass) is found from Canada to Florida, west to Texas.

PLEUROPOGON. Semaphore grass. Caespitose perennials or annuals. Culms herbaceous, to 1+ m. Leaf sheaths closed. Inflorescence a single raceme, rarely a panicle. Spikelets large, linear, several-flowered, laterally compressed, bisexual, disarticulating above the glumes and between the florets. Glumes 2, unequal, shorter than the florets, the first 1-nerved and the second 3-nerved; lemmas 7-nerved, apex entire or bifid, awned; palea 2-nerved, 2-keeled, keels winged on lower portion, awned in 2 species. Lodicules 2; stamens 3; stigmas 2.

A genus of 5 species endemic from Washington to California; three of them known only from relatively few sites in California. A sixth species (*P. sabinii*) is circumpolar. An alternative treatment is to place the American species in the genus *Lophochlaena*, with *P. sabinii* being the only taxon in *Pleuropogon*.

TRIBE: STIPEAE

Plants of this tribe were placed in the Agrostideae of H & C. They differ in details of spikelet morphology and chromosome number. This group is sometimes placed in the subfamily Arundinoideae; sometimes it is treated as the distinct subfamily Stipoideae. See Section 4 for a key to the North American genera of Stipeae.

STIPA. Needle grass, porcupine grass. Caespitose perennials. Culms herbaceous or woody, to 2 m tall. Leaves often in a basal clump; cleistogamous spikelets often present within sheaths. Inflorescence an open to contracted panicle. Spikelets 1-flowered, terete to laterally compressed, bisexual, disarticulating above the glumes. Glumes 2, 1- to 6-nerved, longer than the floret; lemmas long, narrow, firm to indurate at maturity, often tightly wrapped around palea and caryopsis, terminating in a conspicuous awn; lemma base and rachilla forming a sharp-pointed callus, this usually clothed in stiff hairs; palea relatively long, 2-nerved. Lodicules 3 [2]; stamens 3; stigmas 2-4.

A large genus of about 300 species native to temperate and tropical regions of the Old and New World. Of economic significance as a source of pasture grasses, fibers for cordage and mats, and weeds. The sharp-pointed florets can cause mechanical injury to domesticated and wild animals. S. spartea (porcupine grass) occurs in the prairies; S. pulchra (purple needle grass) is found in the California Coast Ranges; S. avenacea (blackseed needle grass) is native to the eastern and southeastern states. S. hymenoides [= Oryzopsis h. in H & C] (Indian ricegrass) is an important forage grass in the drier regions of the West. S. robusta (sleepy grass) is toxic, especially to horses.

Barkworth (1993) has, for the most part, elevated the sections of *Stipa* (or their equivalent) to the generic level. Our North American material would then fall into the genera *Achnatherum*, *Hesperostipa*, and *Nassella*. If we accept this disposition, which I do not, the new combinations would be as follows:

STIPA RENT ASUNDER

Stipa (Traditional) Stipa (Sensu Barkworth)

Stipa arida → Achnatherum aridum Stipa x bloomeri → Achnatherum X bloomeri Stipa brachychaeta → Achnatherum brachychaetum Stipa californica → Achnatherum occidentale var. c. Nassella cernua Stipa cernua → Stipa comata → Hesperostipa comata Stipa coronata → Achnatherum coronatum Hesperostipa curtiseta Stipa curtiseta → Stipa curvifolia → Achnatherum curvifolium Stipa diegoensis → Achnatherum diegoense Stipa eminens → Achnatherum eminens Stipa formicarum → Nassella formicarum Stipa hymenoides → Achnatherum hymenoides Stipa x latiglumis → Achnatherum X latiglume Stipa lemmonii → Achnatherum lemmonii Stipa lepida → Nassella lepida Achnatherum lettermanii Stipa lettermanii → Nassella leucotricha Stipa leucotricha → Stipa lobata → Achnatherum lobatum Stipa neesiana → Nassella neesiana Stipa nelsonii → Achnatherum nelsonii Stipa neomexicana → Hesperostipa neomexicana Achnatherum nevadense Stipa nevadensis → Stipa occidentalis → Achnatherum occidentale Stipa parishii → Achnatherum parishii Achnatherum pinetorum Stipa pinetorum → Stipa porteri → Ptilagrostis mongholica ssp. porteri Stipa pulchra → Nassella pulchra Achnatherum richardsonii Stipa richardsonii → Stipa robusta → Achnatherum robustum Stipa scribneri → Achnatherum scribneri Stipa spartea → Hesperostipa spartea Stipa speciosa → Achnatherum speciosum Stipa stillmanii → Achnatherum stillmanii Stipa tenuissima → Nassella tenuissima Stipa thruberiana → Achnatherum thurberianum Stipa viridula → Nassella viridula Stipa webberi → Achnatherum webberi

ORYZOPSIS. Ricegrass. Caespitose perennials. Culms herbaceous, the internodes solid or hollow, to 1+ m tall. Inflorescence an open or contracted panicle, the branches sometimes capillary. Spikelets 1-flowered, terete to dorsally compressed, bisexual, disarticulating above the glumes. Glumes 2, ± equal, as long or longer than the floret, 3- to 7-nerved, broad, acuminate to obtuse; lemma 3- to 5-nerved, firm to indurate, with a deciduous awn; callus short and blunt; palea relatively long, similar to lemma in texture and marginally or completely covered by it. Lodicules 2 or 3; stamens 3; stigmas 3.

A genus of 20-35 species native to temperate and subtropical regions of both hemispheres. Plants of this genus hybridize freely with *Stipa*. As treated here, *Oryzopsis* includes *Piptatherum*, the latter with an incurved callus, 3 lodicules, marginally covered palea, and free styles. The genus occurs widely in the United States, with species often being locally important as forage. *O. miliacea* (smilo grass) is a weedy

Mediterranean grass established on both coasts; *O. micrantha* (little ricegrass) is native in the West from Canada to California; *O. hymenoides* (Indian ricegrass) has been transferred to *Stipa*.

PIPTOCHAETIUM. Pinyon ricegrass, blackseed needle grass. Caespitose perennials. Culms herbaceous, stiffly erect, to 1+ m tall. Leaves basal, the blades filiform, usually involute. Inflorescence an open panicle. Spikelets 1-flowered, turgid, ± terete, bisexual, disarticulating above the glumes. Glumes 2, ± equal, somewhat longer than the floret, 3- to 7-nerved; lemma 5-nerved, dark-pigmented, with a stout, twisted, geniculate awn; palea 2-nerved, 2-keeled, with a narrow sulcus between the keels. Lodicules 2 or 3; stamens 3; stigmas 2.

A genus of 20-30 species native to the New World, mostly in South America. *P. fimbriatum* (pinyon ricegrass) is an important forage grass in the Southwest; *P. avenaceum* [= Stipa a. in H & C] (blackseed needle grass) is native to the wooded areas of the eastern U. S. A few other adventive species have been reported in California.

TRIBE: HAINARDIEAE

Grasses of the sea coasts. With their spicate inflorescences, they are similar to *Aegilops*. The spikelets are 1-flowered, disarticulating along with a segment of the rachis. The two genera below were assigned to Hordeae in H & C. See Section 4 for a key to the North American taxa.

HAINARDIA. Thintail. Caespitose annuals. Culms herbaceous, the internodes solid, to 5 dm tall. Inflorescence a single spike, spikelets solitary at each node, \pm embedded in notches in a thickened, cylindrical rachis. Spikelets 1-flowered, dorsally compressed, bisexual, disarticulating below the glume, with a rachis segment attached. Glume 1, the first absent and the second firm to indurate, acute, 3- to several-nerved, longer than the floret, closing over the cavity in the rachis; lemma 3-nerved, thin, awnless; palea relatively long, hyaline, 2-nerved. Lodicules 2; stamens 1-3; stigmas 2. A monotypic genus native to the Mediterranean area. *H. cylindrica* [= *Monerma c.* in H & C] is an introduction in California salt marshes.

PARAPHOLIS. Sickle grass. Caespitose annuals. Culms erect to \pm spreading, the internodes hollow, to 2 dm tall. Inflorescence a single spike, curved or straight, the spikelets solitary at each node, \pm embedded in a thickened rachis, disarticulating below the glumes with a rachis segment attached. Glumes 2, \pm equal, longer than the floret, 3- or 5-nerved, leathery, their attachment displaced so that they appear side-by-side in front; lemma hyaline, 1-nerved; awnless; palea relatively long, tightly clasped by lemma, 2-nerved. Lodicules 2; stamens 3; stigmas 2

A genus of 4-6 species native to maritime soils and salt marshes of the Old World; commonly occupying the same habitats as adventives. *P. incurva* is weedy on mud flats and in salt marshes of the Atlantic, Gulf, and Pacific coasts. A second species (*P. strigosa*) is known only from around Humboldt Bay. Its spikes are not curved, as in the other species. It was discovered a few years ago by Thomas Worley, who was an HSU student at the time.

SCRIBNERIA. Low, tufted annual. Inflorescence a slender spike; spikelets 1 per node, inserted flatwise against the rachis. Spikelets 1-flowered, disarticulating above the glumes; rachilla extended as a tiny, hairy bristle. Glumes 2, ± equal, awnless; first 2-nerved and the second 4-nerved. Lemma membranous, minutely bidentate, the midnerve extended as a short, straight awn. Palea about as long as lemma. Stamen 1; stigmas 2. 2n = 36. A monotypic genus. *S. bolanderi* is native from British Columbia to California. H & C placed the genus in their Hordeae.

SELECTED REFERENCES

Catalán, P., E. A. Kellogg, & R. G. Olmstead. 1997. Phylo-geny of Poaceae subfamily Pooideae based on chloroplast *ndh*F gene sequences. Mol. Phylo. Evol. 8: 150-166.

Chandra, N. 1962. Morphological studies in the Gramineae. I. Vascular anatomy of the spikelet in the Pooideae. Proc. Nat. Inst. Sci. India 28B: 545-562.

Hartley, W. 1973. Studies on the origin, evolution, and distribution of the Gramineae. V. The subfamily Festucoideae. Australian J. Bot. 21: 201-234.

Hsiao, C. et al. 1995. Molecular phylogeny of the Pooideae (Poaceae) based on nuclear rDNA (ITS) sequences. Theor. Appl. Genetics 90: 389-398.

Macfarlane, T. D. 1987. Poaceae subfamily Pooideae. In, Soderstrom, T. R. et al. (editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 265-276.

Macfarlane, T. D. & L. Watson. 1980. The circumscription of Poaceae subfamily Pooideae, with notes on some controversial genera. Taxon 29(6): 645-666.

Macfarlane, T. D. & L. Watson. 1982. The classification of the Poaceae subfamily Pooideae. Taxon 31(2): 178-203.

Soreng, R. J. & J. I. Davis. 2000. Phylogenetic structure in Poaceae subfamily Pooideae as inferred from molecular and morphological characters: misclassification versus reticulation. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Colling-wood, Australia. Pp. 61-74.

Soreng, R. J. et al. 1990. A phylogenetic analysis of chloroplast DNA restriction site variation in Poaceae subfam. Pooideae. Plant Syst. Evol. 172: 83-97.

Tucker, G. C. 1996. The genera of Pooideae (Gramineae) in the southeastern United States. Harvard Papers in Bot. 9: 11-90.

TRIBE DIARRHENEAE

DIARRHENA

Brandenburg, D. M., J. R. Estes, & S. L. Collins. 1991. A revision of *Diarrhena* (Poaceae) in the United States. Bull. Torrey Bot. Club 118(2): 128-136.

Schwab, C. A. 1971. Floral structure and embryology of *Diarrhena* (Gramineae). Ph. D. dissertation. Iowa State Univ. Ames. 121 pp.

Tateoka, T. 1957. Notes on grasses. III. Affinities of the genus *Brylkinia*. Systematic position of the genus *Diarrhena*. Bot. Mag. Tokyo 70: 8-12.

TRIBE BRACHYELYTREAE

BRACHYELYTRUM

Babel, W. K. 1943. The variations of *Brachypodium erectum*. Rhodora 45: 260-262.

Campbell, C. S., P. E. Garwood, & L. P. Specht. 1986. Bambusoid affinities of the north temperate genus *Brachyelytrum* (Gramineae). Bull. Torrey Bot. Club 113(2): 135-141.

Stephenson, S. N. 1971. The biosystematics and ecology of the genus *Brachyelytrum* (Gramineae) in Michigan. Michigan Bot. 10: 19-33.

Tateoka, T. 1957. Notes on some grasses. IV. Systematic position of the genus *Brachyelytrum*. J. Japanese Bot. 32: 111-114.

TRIBE POEAE

Bowden, W. M. 1960. Chromosome numbers and taxonomic notes on northern grasses. II. Tribe Festuceae. Canadian J. Bot. 38: 117-131.

Bulinska-Radomska, Z. & R. N. Lester. 1988. Intergeneric relationships of *Lolium, Festuca*, and *Vulpia* (Poaceae) and their phylogeny. Plant Syst. Evol. 159(3-4): 217-227.

Decker, H. F. 1964. An anatomic-systematic study of the classical tribe Festuceae (Gramineae). American J. Bot. 51: 453-463.

Lehvaslaiho, H. et al. 1987. Chloroplast DNA variation in the grass tribe Festuceae. Theor. Appl. Genet. 74: 298-302.

Lehvaslaiho, H., A. Saura, & J. Lokki. 1987. Chloroplast DNA variation in the grass tribe Festuceae. Theor. Appl. Genet. 74: 298-302.

Prat, H. 1935. Contribution a l'etude systematique et histologique des Festucees. Bull. Soc. Bot. France 82: 498-506.

Soreng, R. J. 1998. An infrageneric classification for *Poa* in North America, and other notes on sections, species, and sub-species of *Poa*, *Puccinellia*, and *Dissantherlium* (Poaceae). Novon 8(2): 187-202.

Tateoka, T. 1960. Notes on grasses. X. Some thoughts on the Festuceae: Festucineae with some special reference to their morphology. Canadian J. Bot. 38: 951-967.

Tateoka, T. 1962. Notes on grasses. XIV. Enumeration of the genera of the Festuceae. Bot. Mag. Tokyo 75: 336-343.

ARCTAGROSTIS

- Aiken, S. G. & L. P. Lefkovitch. 1990. *Arctagrostis* (Poaceae: tribe Pooideae) in North America and Greenland. Canadian J. Bot. 68: 2422-2432
- Mitchell, W. W. 1992. Cytogeographical races of *Arctagrostis* (Poaceae) in Alaska. Canadian J. Bot. 70: 80-83.

CUTANDIA

Stace, C. A. 1978. Notes on *Cutandia* and related genera. Bot. J. Linnean Soc. 76: 350-352.

CYNOSURUS

- Arber, A. 1928. Studies in the Gramineae. IV. 1. The sterile spikelets of *Cynosurus* and *Lamarckia*. Ann. Bot. 42: 173-182.
- Jirasek, V. & J. Chertek. 1964. Zur Frage des taxonomischen Wertes der Gattung *Cynosurus* L. Novit. Bot. Inst. Bot. Univ. Carol. Prag. 1964: 23-27.

DACTYLIS

- Borrill, M. 1961. The pattern of morphological variation within diploid and tetraploid *Dactylis*. J. Linnean Soc. Bot. 56: 441-452.
- Borrill, M. & R. Linder. 1971. Diploid-tetraploid sympatry in *Dactylis* (Gramineae). New Phytol. 70(6): 1111-1124.
- Jones, K. 1962. Chromosomal status, gene exchange and evolution in *Dactylis*. Genetica 32: 272-295.
- Stebbins, G. L. & D. Zohary. 1959. Cytogenetic and evolutionary studies in the genus *Dactylis*. I. Morphology, distribution, and inter-relationships of the diploid species. Univ. California Publ. Bot. 31: 1-40.
- Zohary, D. & U. Nur. 1959. Natural triploids in the orchard grass, *Dactylis glomerata* L., polyploid complex and their significance for gene flow from diploid to tetraploid levels. Evolution 13: 311-317.

FESTUCA

- Aiken, S. G. & S. J. Darbyshire. 1990. Fescue grasses of Canada. Agriculture Canada. Ottawa. 113 pp.
- Aiken, S. G., S. J. Darbyshire, & L. P. Lefkovitvh. 1985. Restricted taxonomic value of leaf sections in Canadian narrow-leaved *Festuca* (Poaceae). Canadian J. Bot. 63: 995-1007.
- Ainscough, M. M. et al. 1986. Natural hybrids between *Festuca* and species of *Vulpia* section *Vulpia*. Watsonia 16(2): 143-151.
- Alekseev, E. B. 1985. Genus *Festuca* L. in Mexico et America Centrali. Nov. Syst. Plant. Vasc. 21: 25-58.
- Auquier, P. 1970. Les concepts de Dumortier dans le genre *Festuca* L. (Poaceae). Bull. Jard. Nat. Belg. 40: 393-404.
- Bulinska-Radomska, Z. & R. N. Lester. 1986. Species relation-ships in *Festuca* (sect. *Ovinae*, Poaceae). Plant Syst. Evol. 154(3-4): 175-182.
- Burg, W. J. van der & G. Vierbergen. 1979.

- Distinguishing *Festuca rubra* and *F. ovina*. Seed Sci. Tech. 7(4): 569.
- Charmet, G., C. Ravel, & F. Balfourier. 1997. Phylogenetic analysis in the *Festuca-Lolium* complex using molecular markers and ITS rDNA. Theoret. Appl. Genet. 94: 1038-1046.
- Darbyshire, S. J. 1993. Realignment of *Festuca* subgenus *Schedonorus* with the genus *Lolium* (Poaceae). Novon 3(3): 239-243.
- Darbyshire, S. J. & S. I. Warwick. 1992. Phylogeny of North American *Festuca* (Poaceae) and related genera using chloroplast DNA restrictive site variation. Canadian J. Bot. 70(12): 2415-2429.
- Dubé. M. & P. Morisset. 1996. La variation des caractères épidermiques foliares chez le *Festuca rubra* sensu lato (Poaceae) dan l'est du Canada. Canadian J. Bot. 74(9): 1425-1438.
- Frederiksen, S. 1982. *Festuca brachyphylla, Festuca saxi-montana*, and related species in North America. Nordic J. Bot. 2: 525-536.
- Gaut, B. S. et al. 2000. Phylogenetic relationships and genetic diversity among members of the *Festuca-Lolium* complex (Poaceae) based on ITS sequence data. Plant Syst. Evol. 224(1-2): 33-53.
- Harms, V. L. 1985. A reconsideration of the nomenclature and taxonomy of the *Festuca altaica* complex (Poaceae) in North America. Madroño 32(1): 1-10.
- Hertzsch, W. 1961. Gattungskreuzungen Zwischen den Gattungen Festuca und Lolium C. Die Fl -- Bastarde, ihr Verhalten und ihr Aussehen. Z. Pfalzen 45: 345-360.
- Hill, H. D. 1965. Karyology of species of *Bromus, Festuca*, and *Arrhenatherum*. Bull. Torrey Bot. Club 92: 192-197.
- Jauhar, P. P. 1975. Chromosome relationships between *Lolium* and *Festuca* (Gramineae). Chromosoma 52(2): 103-121.
- Pavlick, L. E. 1985. A new taxonomic survey of the *Festuca rubra* complex in northwestern North America, with emphasis on British Columbia. Phytologia 57(1): 1-17.
- Pavlick, L. E. 1983. Notes on the taxonomy and nomenclature of Festuca occidentalis and F. idahoensis. Canadian J. Bot. 61(1): 337-344.
- Pavlick, L. E. & J. Looman. 1984. Taxonomy and nomenclature of rough fescues, *Festuca altaica, F. campestris (F. scabrella* var. *major)*, and *F. hallii* in Canada and the adjacent part of the United States. Canadian J. Bot. 62: 1739-1749.
- Piper, C. V. 1906. North American species of *Festuca*. Contr. U. S. Natl. Herb. 10(1): 1-49.
- Piper, C. V. 1913. Supplementary notes on American species of *Festuca*. Contr. U. S. Natl. Herb. 16(5): 197-199.
- Reveal, J. L., E. E. Terrell, & H. Scholz. 1991. Proposal to reject *Festuca elatior* L. with comments on the typification of *F. pratensis* and *F. arundinacea*

(Poaceae). Taxon 40(1): 135-137.

Saint-Yves, A. 1925. Contribution a l'étude des *Festuca* (subgen. *EuFestuca*) de l'Amerique du Nord et du Mexique. Candollea 2: 229-316.

Soreng, R. J. & E. E. Terrell. 1997. Taxonomic notes on *Schedonorus*, a segregate genus from *Festuca* or *Lolium*, with a new nothogenus X *Schedololium*, and new combinations.

Stace, C. A. et al. 1992. The distinction between the *Festuca ovina* L. and *Festuca rubra* L. aggregates in the British Isles. Watsonia 19: 107-112.

Tenant, F. 1973. Genetic structure of *Festuca microstachys* pop-ulations. Ph. D. dissertation. Univ. California, Davis.

Terrell, E. E. 1967. Meadow fescue: *Festuca elatior* L. or *F. pratensis* Hudson? Brittonia 19: 129-132.

Terrell, E. E. 1968. Notes on *Festuca arundinacea* and *Festuca pratensis* in the U. S. A. Taxonomy and distribution. Rhodora 70(784): 564-568.

Torrecilla, P. & P. Catalá. 2002. Phylogeny of broadleaved and fine-leaved *Festuca* lineages (Poaceae) based on nuclear ITS sequences. Syst. Bot. 27(2): 241-251.

Xu, W.-W. & D. D. Sleper. 1994. Phylogeny of tall fescues and related species using RFLPs. Theor. Appl. Genet. 88: 685-690.

LOLIUM

Bennett, S. J. 1997. A phenetic analysis and lateral key of the genus *Lolium* (Gramineae). Genet. Resources Crop Evol. 44(1): 63-72.

Bulinska-Radomska, Z. 1985. Relationships between five *Lolium* species (Poaceae). Plant Syst. Evol. 148: 169-176.

Bulinska-Radomska, Z. 1988. Intergeneric relationships of *Lolium*, *Festuca*, and *Vulpia* (Poaceae) and their phylogeny. Plant Syst. Evol. 159(3-4): 217-227.

Bulinska-Radomska, Z. & R. N. Lester. 1988. Intergeneric relationships of *Lolium*, *Festuca*, and *Vulpia* (Poaceae) and their phylogeny. Plant Syst. Evol. 159(3-4): 217-227.

Charmet, G. et al. 1996. Taxonomic relationships and interspecific hybridization in the genus *Lolium* (grasses). Genet. Res. Crop Evol. 43(4): 319-327.

Charmet, G. & F. Balfourier. 1994. Isozyme variation and species relationships in the genus *Lolium* L. (ryegrasses, Graminaceae) [sic]. Theoret. Appl. Genetics 87(6): 641-649.

Essad, S. 1954. Contribution a la systématique du genu *Lolium*. Ann. de L'Amélior des Plantes 4: 325-351.

Howlett, B. J. et al. 1981. Immunofluorescent localization of two water-soluble glycoproteins including the major allergen from pollen of rye grass (*Lolium perenne*). Histochem. J. 13: 461-480.

Jauhar, P. P. 1975. Chromosome relationships

between *Lolium* and *Festuca* (Gramineae). Chromosoma 52(2): 103-121.

Jenkin, T. J. 1959. The rye grasses (*Lolium L.*). Handb. Pflanzenzuchtung 4: 435-452.

Johnson, P. & D. G. Marsh. 1965. The isolation and characterization of allergens from the pollen of rye grass (*Lolium pereene*). J. Europ. Polymer 1: 63-77.

Lester, R. N. 1985. Relationships between five species of *Lolium* (Poaceae). Plant Syst. Evol. 148: 169-175.

Loos, B. P. 1993. Morphological variation in *Lolium* (Poaceae) as a measure of species relationships. Plant Syst. Evol. 188: 87-99.

Terrell, E. E. 1966. Taxonomic implications of genetics in rye grasses (*Lolium*). Bot. Rev. 32: 138-164.

Terrell, E. E. 1968. A taxonomic revision of the genus *Lolium*. U. S. D. A. Tech. Bull. No. 1392.

Vasek, F. et al. 1963. A note on taxonomic characters in *Lolium*. Madroño 17: 79-83.

PHIPPSIA

Weber, W. A. 1952. *Phippsia algida* in the United States. Rhodora 54: 141-145.

POA

Anton, A. M. & H. E. Connor. 1995. Floral biology and reproduction in *Poa* (Poeae: Gramineae). Australian J. Bot. 43(6): 577-599.

Arnow, L. A. 1981. *Poa secunda* Presl versus *P. sandbergii* Vasey (Poaceae). Syst. Bot. 6(4): 412-421.

Clausen, J. 1961. Introgression facilitated by apomixis in polyploid Poas. Euphytica 10: 87-94.

Cusick, A. W. & M. A. Vincent. 2002. *Poa bulbosa* L. ssp. *bulbosa* (Poaceae) in North America. Michigan Bot. 41(1): 19-23.

Edmondson, J. R. 1978. Infraspecific taxa in European *Poa* L. J. Linn. Soc. Bot. 76: 329-334.

Hartley, W. 1961. Studies in the origin, evolution, and distribution of the Gramineae. 4. The genus *Poa* L. Australian J. Bot. 9: 152-161.

Kellogg, E. A. 1985. Variation and names in the *Poa secunda* complex. J. Range Manag. 38(6): 516-521.

Kellogg, E. A. 1985. A biosystematic study of the *Poa secunda* complex. J. Arnold Arbor. 66: 201-242.

Kellogg, E. A. 1987. Apomixis in the *Poa secunda* complex. Amer. J. Bot. 74(9): 1431-1437.

Law, R. et al. 1977. Life-history variation in *Poa annua*. Evolution 31: 233-246.

Marsh, V. L. 1952. A taxonomic revision of the genus *Poa* of United States and southern Canada. American Midl. Nat. 47: 202-250.

Soreng, R. J. 1985. *Poa* L. in New Mexico, with a key to middle and southern Rocky Mountain species (Poaceae). Great Basin Nat. 45(3): 395-422.

- Soreng, R. J. 1991. Notes on new infraspecific taxa and hybrids in North American *Poa* (Poaceae). Phytologia 71(5): 390-413.
- Soreng, R. J. 1998. An infrageneric classification for *Poa* in North America, and other notes on sections, species, and sub-species of *Poa*, *Puccinellia*, and *Dissantherlium* (Poaceae). Novon 8(2): 187-202.
- Soreng, R. J. & S. L. Hatch. 1983. A comparison of *Poatracyi* and *P. occidentalis* (Poaceae: Poeae). Sida 10: 123-141.
- St. John, H. 1941. The status of *Poa secunda* and *Poa sand-bergii* (Gramineae) in North America. American J. Bot. 28: 78-81.
- Timm, G. 1965. Beiträge zue Biologie und Systematik von *Poa annua* L. Z. Acker- und Pflanzenbau 122: 267-294.
- Tinney, F. W. 1940. Cytology of parthenogenesis in Poa pratensis. J. Agric. Res. 60: 351-360.

PUCCINELLIA

- Choo, M. K. et al. 1994. Phylogenetic relationships among *Puccinellia* and allied genera of Poaceae as inferred from chloroplast DNA restriction site variation. American J. Bot. 81(1): 119-126.
- Church, G. L. 1949. A cytotaxonomic study of *Glyceria* and *Puccinellia*. American J. Bot. 36: 155-165.
- Clausen, R. T. 1952. Suggestion for the assignment of *Torreyochloa* to *Puccinellia*. Rhodora 54: 42-45.
- Davis, J. I. 1983. Phenotypic plasticity and the selection of taxonomic characters in *Puccinellia* (Poaceae). Syst. Bot. 8(4): 341-353.
- Davis, J. I. 1988. Genetic and environmental contributions to multivariate morphological patterns in *Puccinellia* (Poaceae). Canadian J. Bot. 66: 2436-2444.
- Davis, J. I. 1990. *Puccinellia howellii* (Poaceae), a new species from California. Madroño 37(1): 55-58.
- Davis, J. I. 1993. *Puccinellia howellii* (Poaceae) is genetically distinct. Madroño 40(4): 202-208.
- Davis, J. I. & D. H. Goldman. 1993. Isozyme variation and species delimitation among diploid populations of the *Puccinellia nuttalliana* complex (Poaceae): character fixation and the discovery of phylogenetic species. Taxon 42(2): 585-599.
- Fernald, M. L. & C. A. Weatherby. 1916. The genus *Puccinellia* in eastern North America. Rhodora 18: 1-23.
- Swallen, J. R. 1944. The Alaskan species of *Puccinellia*. J. Washington Acad. Sci. 34: 16-23.

SCLEROCHLOA

Brandenburg, D. M., J. R. Estes, & J. W. Thieret. 1991. Hard grass (*Sclerochloa dura*, Poaceae) in the United States. Sida 14(3): 369-376.

SCOLOCHLOA

Smith, A. L. 1973. Life cycle of the marsh grass, *Scolochloa festucacea*. Canadian J. Bot. 51(9): 1661-1668.

TORREYOCHLOA

- Church, R. T. 1952. The genus *Torreyochloa*. Rhodora 54: 197-200.
- Davis, J. I. 1991. A note on North American *Torreyochloa* (Poaceae), including a new combination. Phytologia 70(5): 361-365.

VULPIA

- Allard, R. W. & L. W. Kannenberg. 1968. Population studies in predominantly self-pollinated species. XI. Genetic divergence among members of the *Festuca microstachys* complex. Evolution 22: 517-528.
- Auquier, P. & C. A. Stace. 1980. Variation in flowering behaviour in *Vulpia* (Poaceae). Plant Syst. Evol. 136: 47-52.
- Barker, C. M. & C. A. Stace. 1982. Hybridisation in the genera *Vulpia* and *Festuca*: the production of artificial F1 plants. Nordic J. Bot. 2: 435-444.
- Barker, C. M. & C. A. Stace. 1986. Hybridization in the genera *Vulpia* and *Festuca* (Poaceae). Nordic J. Bot. 6(1): 1-10.
- Bulinski-Radomska, Z. & R. N. Lester. 1986. Relationships between seven species of *Vulpia* (Poaceae). Plant Syst. Evol. 153(1-2): 7-12.
- Cotton, R. & C. A. Stace. 1976. Taxonomy of the genus *Vulpia* (Gramineae): 1. Chromosome numbers and geographical distribution of the Old World species. Genetica 46(2): 235-255.
- Cotton, R. & C. A. Stace. 1977. Morphological and anatomical variation of *Vulpia* (Gramineae). Bot. Not. 130(2): 173-187.
- Henrard, J. T. 1937. A study in the genus *Vulpia*. Blumea 2: 299-326.
- Kannenberg, L. W. & R. W. Allard. 1967. Population studies in predominantly self-pollinated species. VIII. Genetic variability in the *Festuca microstachys* complex. Evolution 21: 227-240.
- Lonard, R. I. 1970. A biosystematic study of the genus *Vulpia* (Gramineae). Ph. D. dissertation. Texam A & M Univ. College Station. 167 pp.
- Lonard, R. L. & F. W. Gould. 1974. The North American species of *Vulpia* (Gramineae). Madroño 22: 217-230.
- Stace, C. A. 1976. Taxonomy of the genus *Vulpia*, Gramineae. I. Chromosome numbers and geographical distribution of the Old World species. Genetica 42: 235-255.
- Stace, C. A. 1981. Generic and infrageneric nomenclature of annual Poaceae: Poeae related to *Vulpia* and *Desmazeria*. Nordic J. Bot. 1(1): 17-26.

TRIBE BROMEAE

BROMUS

- Allred, K. W. 1993. *Bromus*, section *Pnigma*, in New Mexico, with a key to the bromegrass of the state. Phytologia 74(4): 319-345.
- Armstrong, K. C. 1981. The evolution of *Bromus inermis* and related species of *Bromus* sect. *Pnigma*. Bot. Jahr. 102(1-4): 327-443.
- Armstrong, K. C. 1991. Chromosome evolution in *Bromus*. <u>In</u>, Tsuchiya, T. & P. K. Gupta (editors). Chromosome engineering in plants. Pt. B. Elsevier. Amsterdam, The Netherlands. Pp. 363-377.
- Bartlett, E., S. J. Novak, & R. N. Mack. 2002. Genetic variation in *Bromus tectorum* (Poaceae): differentiation in the eastern United States. American J. Bot. 89(4): 602-612.
- Esnault, M. -A. & A. Huon. 1985. Application des méthodes numériques á lá systematique du genre *Bromus* L. Sect. *Genea* Dumort. Bull. Soc. Linn. Procence 37: 69-78.
- Grossmann, A. 1973. Was ist *Bromus unioloides*? Gött. Fl. Rundbr. 7: 13-19.
- Harlan, J. R. 1945. Cleistogamy and chasmogamy in Bromus carinatus Hook. & Arn. AJB 32: 66-72.
- Hill, H. D. 1965. Karyology of species of *Bromus, Festuca,* and *Arrhenatherum*. Bull. Torrey Bot. Club 92: 192-197.
- Klemmedson, J. D. & J. G. Smith. 1964. Cheatgrass (*Bromus tectorum* L.) Bot. Rev. 30: 226-262.
- Knowles, P. F. 1944. Interspecific hybridizations of *Bromus*. Genetics 29: 128-140.
- Lamson-Scribner, F. & E. D. Merrill. 1900. Studies on American grasses. The North American species of *Ceratochloa*. Bull. No. 21. U. S. D. A. Div. of Agrostoogy. Washington, D. C. 44 pp.
- Maw, C. C. 1974. A note on *Bromus unioloides* and *B. willdenowii* (Gramineae). Kew Bull. 29(2): 431-434.
- McNeill, J. 1976. Nomenclature of four perennial species of *Bromus* in eastern North America, with a proposal for the listing of *B. purgans* as a rejected name under Article 69. Taxon 24: 611-616.
- Mitchell, W. W. 1967. Taxonomic synopsis of *Bromus* section *Bromopsis* (Gramineae) in Alaska. Canadian J. Bot. 45: 1309-1313.
- Novak, S. J. & R. N. Mack. 1991. Genetic variation in *Bromus tectorum* (Poaceae): comparison between native and introduced populations. Heredity 71: 167-178.
- Novak, S. J., R. N. Mack, & D. E. Soltis. 1991. Genetic variation in *Bromus tectorum* (Poaceae): population differentiation in its North American range. Amer. J. Bot. 78(8): 1150-1161.
- Novak, S. J., R. N. Mack, & P. S. Soltis. 1993. Genetic variation in *Bromus tectorum* (Poaceae): introduction

- dynamics in North America. Canadian J. Bot. 71(11): 1441-1448.
- Pavlick, L. E. 1995. *Bromus* L. of North America. Royal British Columbia Mus. Victoria. 160 pp.
- Peterson, P. M. & A. M. Planchuelo. 1998. *Bromus catharticus* in South America (Poaceae: Bromeae). Novon 8(1): 53-60.
- Peterson, P. M. et al. 2001. Recognition of *Bromus richardsonii* and *B. ciliatus*: evidence from morphology, cytology, and DNA fingerprinting (Poaceae: Bromeae). Aliso 20(1): 21-36.
- Pillay, M. & K. W. Hilu. 1995. Chloroplast-DNA restriction site analysis in the genus *Bromus* (Poaceae). American J. Bot. 82(2): 239-249.
- Pinto-Escobar, P. 1976. Nota sobre el ejemplar tipo de "Bromus catharticus" Vahl. Caldasia 11: 9-16.
- Pinto-Escobar, P. 1986. Nota sobre *Bromus willdenovii* Kunth. Caldesia 14: 185-191.
- Raven, P. H. 1957. The introduced species of *Bromus*, sect. *Ceratochloa*, in California. Leaflts. West. Bot. 8(6): 151-154.
- Raven, P. H. 1960. The correct name for rescue grass. Brittonia 12: 219-221.
- Sales, F. 1991. A reassessment of *Bromus tectorum*: a computer analysis, synaptospermy and chorispermy. Flora et Vetetation Mundi 9: 29-41.
- Sales, F. 1991. Evolution and adaptive radiation of *Bromus* L. Sect. *Genea* Dum. (Poaceae). Ph. D. dissertation. Univ. Edinburgh.
- Sales, F. 1993. Taxonomy and nomenclature of *Bromus* sect. *Genea*. Edinburgh J. Bot. 50(1): 1-31.
- Sales, F. 1994. Evolutionary tendencies in some annual species of bromes (*Bromus* L. sec. *Genea* Dum. (Poaceae)). Bot. J. Linnean Soc. 115(3): 197-210.
- Sales, F. 1994. A reassessment of the *Bromus madritensis* complex (Poaceae): a multivariate approach. Israel J. Plant Sci. 42(3): 245-255.
- Scholz, H. 1970. Zur Systematik der Gattung *Bromus* L. subgenus *Bromus* (Gramineae). Willdenowia 6: 139-159.
- Scholz, H. 1981. Bemerkungen über *Bromus madritensis* und *B. rubens* (Gramineae). Willdenowia 11: 249-258.
- Seymour, F. C. 1966. Bromus mollis and allies in New England. Rhodora 68: 168-174.
- Shear, C. L. 1900. A revision of North American species of *Bromus* occurring north of Mexico. U. S. D. A. Agrost. Bull. 23: 5-66.
- Smith, P. M. 1970. Taxonomy and nomenclature of the brome grasses (*Bromus* s. l.). Notes from the Royal Bot. Gard., Edinburgh 30: 361-375.
- Smith, P. M. 1972. Serology and species relationships in annual bromes (*Bromus* L. sect. *Bromus*). Ann. Bot. 36: 1-30.

- Smith, P. M. 1973. Observations on some critical brome grasses. Watsonia 9: 319-332.
- Smith, P. M. 1981. Ecotypes and subspecies in annual brome-grasses (*Bromus*, Gramineae). Bot. Jahrb. Syst. 102: 497-509.
- Smith, P. M. 1986. Native or introduced: problems in the taxonomy and geography of some widely introduced annual brome-grasses. Proc. Royal Soc. Edinburgh 89B: 2723-2781.
- Smith, P. M. & F. Sales. 1993. *Bromus* L. sec. *Bromus*. Taxonomy and relationship of some species with small spikelets. Edinburgh J. Bot. 50(2): 149-171.
- Soderstrom, T. R. & J. H. Beaman. 1968. The genus *Bromus* (Gramineae) in Mexico and Central America. Publ. Michigan State Univ. Mus. Biol. Ser. 3(5): 469-519.
- Stebbins, G. L. 1947. The origin of the complex *Bromus carinatus* and its phytogeographic implications. Contr. Gray Herb. 165: 42-55.
- Stebbins, G. L. 1981. Chromosomes and evolution in the genus *Bromus* (Gramineae). Bot. Jahr. 102(1-4): 359-379.
- Stebbins, G. L., Jr. & H. A. Tobgy. 1944. The cytogenetics of hybrids in *Bromus* I. Hybrids within the section *Ceratochloa*. American J. Bot. 31: 1-11.
- Stebbins, G. L., H. A. Tobgy, & J. R. Harlan. 1944. The cytogenetics of hybrids in Bromus. II. B. carinatus & B. arizonicus. Proc. Calif. Acad. Sci. 25: 307-322.
- Thill, D. C., K. G. Beck, & R. H. Callihan. 1984. The biology of downy brome (*Bromus tectorum*). Weed Sci. 32(Suppl.) 1: 7-12.
- Tournay, R. 1961. La nomenclature des sections du genre *Bromus* L. Bull. Jard. Etat. Bruxelles 31: 289-299.
- Wagnon, H. K. 1952. A revision of the genus *Bromus*, section *Bromopsis*, of North America. Brittonia 7: 415-480.
- Wu, K. K. 1974. Ecogenetic studies on population structure of *Bromus rubens* L. and *Bromus mollis* L. Ph. D. dissertation. Univ. California, Davis.

TRIBE BRACHYPODIEAE

BRACHYPODIUM

- Catalàn, P. et al. 1995. Molecular phylogeny of the grass genus *Brachypodium* P. Beauv. based upon RFLP and RAPD analysis. Bot. J. Linnean Soc. 117(4): 263-280.
- Shi, Y., J. D. Raper, & C. Stace. 1993. Ribosomal DNA variation and its phylogenetic implication in the genus *Brachypodium* (Poaceae). Pl. Syst. Evol. 188(3-4): 125-138.
- Tateoka, T. 1968. Phytogeographic notes on the genus *Brachy-podium* P. Beauv. (Gramineae). Bol. Soc. Argentine Bot. 12(1): 44-56.

TRIBE AVENEAE

Dass, H. C. 1972. Analysis of species relationships in Aveneae by thin-layer chromatography and disc electrophoresis. Canadian J. Genet. Cytol. 14: 305-316.

SUBTRIBE AVENINAE

AMPELODESMOS

Decker, H. F. 1964. Affinities of the grass genus *Ampelodesmos*. Brittonia 16: 76-79.

APERA

McNeill, J. 1981. *Apera*, silky-bent or windgrass, an important weed genus recently discovered in Ontario, Canada. Canadian J. Plant Sci. 61: 479-485.

AVENA

- Baum, B. R. 1968. Delimitation of the genus *Avena* (Gramineae). Canadian J. Bot. 46: 121-132.
- Baum, B. R. 1968. On some relationships between *Avena sativa* and *A. fatua* (Gramineae) as studied from Canadian material. Canadian J. Bot. 46: 1013-1024.
- Baum, B. R. 1969. The use of lodicule type in assessing the origin of *Avena* fatuoids. Canadian J. Bot. 47: 931-944.
- Baum, B. R. 1974. Typification of Linnaean species of oats, *Avena*. Taxon 23: 579-583.
- Baum, B. R. 1974. Classification of the oat species (*Avena*, Poaceae) using various taxonomic methods and an information-theoretic model. Canadian J. Bot. 52: 2241-2262.
- Baum, B. R. 1975. Cladistic analysis of diploid and hexaploid oats (*Avena*, Poaceae) using numerical techniques. Canadian J. Bot. 53: 2115-2127.
- Baum, B. R. 1977. Oats, wild and cultivated -- a monograph of the genus *Avena* L. (Poaceae). Agriculture Canada. Ottawa. 463 pp.
- Coffman, F. A. 1977. Oat history, identification and classification. U. S. D. A. Tech. Bull. No. 1516. 356 pp.
- Cooper, S. R. 1979. Differentiation of *Avena* species. Seed Sci. Tech. 7(4): 517-521.
- Jain, S. K. & D. R. Marshall. 1967. Population studies in predominantly self-pollinating species. X. Variation in natural populations of *Avena fatua* and *A. barbata*. Amer. Nat. 101: 19-32.
- Jain, S. K. 1969. Comparative ecogenetics of *Avena sativa* and *A. barbata* occurring in central California. Evol. Biol. 3: 73-118.
- Ladizinsky, G. 1974. Genome relationships in the diploid oats. Chromosoma 47(1): 109-117.
- Katsiotis, A. et al. 2000. Repetitive DNA, genome and species relationships in *Avena* and *Arrhenatherum* (Poaceae). Ann. Bot. 86(6): 1135-1142.

Ladizinsky, G. 1969. New evidence on the origin of the hexaploid oats. Evolution 23: 676-684.

Ladizinsky, G. & D. Zohary. 1971. Notes on species delimitation, species relationships and polyploidy in *Avena* L. Euphytica 20: 380-395.

Marshall, D. R. et al. 1968. Phenotypic plasticity of *Avena fatua* and *A. barbata*. American Nat. 102: 457-467.

Stanton, T. R. 1955. Oat identification and classification. Tech. Bull. No. 1100. U. S. Dept. Agric. Washington, D. C. 206 pp.

ARRHENATHERUM

Hill, H. D. 1965. Karyology of species of *Bromus, Festuca,* and *Arrhenatherum*. Bull. Torrey Bot. Club 92: 192-197.

Mahmoud, A. et al. 1975. Polymorphism in *Arrhenatherum elatius* (L.) Beauv. ex J. & C. Presl. New Phytol. 75(2): 269-276.

Potztal, E. 1954. Anatomische-systematische Untersuchungen aus den Gattungen *Arrhenatherum* und *Helictotrichon*. Bot. Jahrb. 76: 281-383.

BECKMANNIA

Reeder, J. R. 1953. Affinities of the grass genus *Beckmannia* Host. Bull. Torrey Bot. Club 80(2): 187-196.

CORYNEPHORUS

Jirasek, V. & J. Chrtek. 1962. Systematische Studie über Arten der Gattung *Corynephorus* Pal. Beauv. (Poaceae). Preslia 34: 374-386.

DESCHAMPSIA

Kawano, S. 1963. Cytogeography and evolution of the *Deschampsia caespitosa* complex. Canadian J. Bot. 41: 719-742.

Koch, S. D. 1979. The relationship of three Mexican Aveneae and some new characters for distinguishing *Deschampsia* and *Trisetum* (Gramineae). Taxon 28: 225-235.

Putman, D. L. 1971. Biosystematic studies of *Deschampsia* Beauv. Ph. D. dissertation. Colorado State Univ. Ft. Collins. 97 pp.

DISSANTHELIUM

Swallen, J. R. & O. Tovar. 1965. The grass genus *Dissanthelium*. Phytologia 11: 361-376.

HELICTOTRICHON

Holub, J. 1958. Bemerkungen zur Taxonomie der Gattung *Helictotrichon* Bess. Praha 1958: 101-133.

Holub, J. 1962. Ein Beitrag zur Abgrenzung der Gattung in der Tribus Aveneae: die Gattung Avenochloa Holub. Acta Horti Bot. Prag. 1962: 75-86.

Holub, J. 1977. Notes on some species of *Avenula* and *Helicto-trichon*. Preslia 49: 203-221.

Potztal, E. 1954. Anatomische-systematische

Untersuchungen aus den Gattungen Arrhenatherum und Helictotrichon. Bot. Jahrb. 76: 281-383.

Röser, M. 1996. Ecogeography of *Helictotrichon* (Poaceae). Plant Syst. Evol. 203(3-4): 181-281.

HOLCUS

Beddows, A. R. 1971. The inter- and intraspecific relationships of *Holcus lanatus* L. and *H. mollis* L. sensu lato (Gramineae). Bot. J. Linnean Soc. 64(2): 183-198.

Zandee, M. & P. C. G. Glas. 1982. Studies in the *Holcus lanatus-Holcus mollis* complex (Poaceae, Gramineae). Proc. Koninklijke Nederlandse Akad. Van Wetenschappen 85: 413-437.

KOELERIA

Arnow, L. A. 1994. *Koeleria macrantha* and *K. pyramidata* (Poaceae): nomenclatural problems and biological distinctions. Syst. Bot. 19: 6-20.

Domin, K. 1907. Monographie der Gattung *Koeleria*. Bibl. Bot. 14: 1-354.

Looman, J. 1978. Biological flora of the Canadian prairie provinces. V. *Koeleria gracilis* Pers. Canadian J. Plant Sci. 58: 459-466.

Robertson, P. A. 1974. Morpholigical variation and chromosome numbers of North American populations of *Koeleria cristata*. Bull. Torrey Bot. Club 101: 124-129.

Shinners, L. H. 1956. Illegitimacy of Persoon's species of *Koeleria* (Gramineae). Rhodora 58: 93-96.

SPHENOPHOLIS

Erdman, K. S. 1965. Taxonomy of the genus *Sphenopholis*. Iowa State J. Sci. 39: 259-336.

Lamson-Scribner, F. 1906. The genus *Sphenopholis*. Rhodora 8: 137-146.

Terrell, E. E. et al. 1965. Natural hybrids between *Sphenopholis obtusata* and *Trisetum pennsylvanicum*. Bull. Torrey Bot. Club 92: 169-182.

TRISETUM

Chrtek, J. 1965. Bemerkungen zur Gleiderung der Gattung *Trisetum* Pers. Bot. Not. 118: 210-224.

Chrtek, J. & V. Jirasek. 1963. On the taxonomy of the genus *Trisetum* Pers. Webbia 17: 569-580.

Hultén, E. 1959. The *Trisetum spicatum* complex. Svensk Bot. Tidskr. 53: 203-228.

Father Louis-Marie O. C. 1928. The genus *Trisetum* in America. Rhodora 30: 209-228; 231-245.

Jonsell, B. 1978. Nomenclatural and taxonomic notes on *Trisetum* Pers. and *Lophochloa* Reichenb. (Gramineae). Bot. J. Linnean Soc. 76: 320-322.

Randall, J. L. & K. W. Hilu. 1986. Biosystematic studies of North American *Trisetum spicatum* (Poaceae). Syst. Bot. 11(4): 567-578.

VENTENATA

Chambers, K. L. 1985. Pitfalls in identifying *Ventenata dubia* (Poaceae). Madroño 32(2): 120, 121.

SUBTRIBE PHALARIDINAE

Riecken, W. E. 1929. A morphological study of some Phalarideae, with special reference to classification. Bull. Torrey Bot. Club 56: 409-419.

ANTHOXANTHUM

Borrill, M. 1963. Experimental studies in evolution in *Anthoxanthum* (Gramineae). Genetica 34: 183-208.

Hedberg, I. 1961. Cytotaxonomic studies in *Anthoxanthum odoratum* L. sensu lat. Svensk Bot. Tidskr. 55: 118-128.

Hedberg, I. 1990. Morphological, cytotaxonomic and evolutionary studies in *Anthoxanthum odoratum* L. s. lat. -- a critical review. Sommerfeltia 11: 97-107.

Schouten, Y. & J. F. Veldkamp. 1985. A revision of *Anthoxanthum* including *Hierochloë* (Gramineae) in Malesia and Thailand. Blumea 30: 319-351.

Valdés, B. 1973. Revisión de las especies anuales del génera *Anthoxanthum* (Gramineae). Lagascalia 3(1): 99-141.

HIEROCHLOË

Norstog, K. J. 1960. Some observations on the spikelet of *Hierochloë odorata*. Bull. Torrey Bot. Club 87: 95-98.

Reeder, J. R. & K. J. Norstog. 1961. The status of *Hierochloe nashii* and its relationship to *H. odorata*. Bull. Torrey Bot. Club 88: 77-84.

Schouten, Y. & J. F. Veldkamp. 1985. [See citation under Anthoxanthum].

Weimarck, G. 1971. Variation and taxonomy of *Hierochloë* in the northern hemisphere. Bot. Not. 124(1): 129-175.

PHALARIS

Anderson, D. E. 1961. Taxonomy and distribution of the genus *Phalaris*. Iowa State J. Sci. 36: 1-96.

Baldini, R. M. 1995. Revision of the genus *Phalaris* L. (Gramineae). Webbia 49: 265-329.

Merigliano, M. F. & P. Lesica. 1998. The native status of reed canary grass in the inland northwest, U. S. A. Nat. Areas J. 18: 223-230.

Riecken, W. E. 1929. A morphological study of some Phalarideae, with special references to classification. Bull. Torrey Bot. Club 56: 409-419.

SUBTRIBE: ALOPECURINAE

ALOPECURUS

Sieber, V. K. & B. G. Murray. 1979. The cytology of the genus *Alopecurus*. Bot. J. Linnean Soc. 79: 343-355.

AMMOPHILA

Seymour, F. C. 1966. *Ammophila champlainensis* (Gramineae), a new species in New York and Vermont. Sida 2: 349-351.

CALAMAGROSTIS

Greene, C. W. 1984. Sexual and apomictic reproduction in *Calamagrostis* (Gramineae) from eastern North America. American J. Bot. 71(3): 285-293.

Kearney, T. H. 1898. Studies on American grasses. I. A revision of the North American species of *Calamagrostis*. U. S. D. A. Div. of Agrost. Bull. 11: 7-42.

Nygren, A. 1954. Investigations on North American *Calamagrostis*. Hereditas 40: 375-397.

Nygren, A. 1958. Investigations on North American *Calamagrostis*. II. Lantbruk. Ann. 24: 363-368.

Stebbins, G. L., Jr. 1930. A revision of some North American species of *Calamagrostis*. Rhodora 32: 35-57

Wasiljew, W. N. 1960. Das System der Gattung *Calamagrostis* Roth. Fed. Repert. 63: 229-251.

AGROSTIS

Beetle, A. A. 1950. A sectional treatment for the North American species of *Agrostis*. Univ. Wyoming Publ. 15: 29-35.

Bjorkman, S. O. 1960. Studies in *Agrostis* and related genera. Symb. Bot. Uppsal. 17(1): 1-112.

Carlbom, C. G. 1967. A biosystematic study of some North American species of *Agrostis* L. and *Podagrostis* (Griseb.) Scribn. & Merr. Ph. D. dissertation. Oregon State Univ. Corvallis. 232 pp.

Hitchcock, A. S. 1905. The North American species of *Agrostis*. U. S. D. A. Bur. Plant Ind. Bull. 68: 1-68.

Jones, K. 1952. Autotetraploidy in *Agrostis canina*. Nature. 169: 159, 160.

Widen, K.-F. 1971. The genus *Agrostis* in eastern Fennoscandia. Taxonomy and distribution. Flora Fenn. 5: 1-209.

CINNA

Brandenburg, D. M. et al. 1991. Revision of the genus *Cinna* (Poaceae). Sida 14(4): 581-596.

PHLEUM

Evans, M. W. 1927. The life history of timothy. U. S. D. A. Bull. No. 1450. 55 pp.

Humphries, C. J. 1978. Notes on the genus *Phleum*. Bot. J. Linn. Soc. 76: 337-340.

Joachimiak, A. & A. Kula. 1996. Karyosystematics of the *Phleum alpinum* polyploid complex (Poaceae). Plant Syst. Evol. 203(1-2): 11-25.

Lowenstein, H. 1978. Isolation and partial characterization of three allergens of timothy pollen.

Allergy 33: 30-41.

Malley, A. & R. L. Harris. 1967. Biological properties of a non-precipitating antigen from timothy pollen extracts. J. Immunol. 99: 825-830.

Nath, J. 1967. Cytogenetical and related studies in the genus *Phleum* L. Euphytica 16: 267-282.

Weeke, B. et al. 1974. Allergens in timothy pollen identified by crossed radioimmunologelectrophoresis. Acta Allerg. 29: 409-417.

POLYPOGON

Conley, S. A. 1987. Systematic investigations in the genus *Polypogon* in northern California. M. S. thesis. California State Univ., Chico. 80 pp.

TRIBE TRITICEAE

Aniol, A. 1976. Serological studies within the tribe Triticeae. VIII. Serological affinities between genera. Genet. Pol. 17(4): 523-529.

Barkworth, M. E. 1992. Taxonomy of the Triticeae: a historic perspective. Hereditas 116(1/2): 1-14.

Barkworth, M. E. 1997. Taxonomic and nomenclatural com-ments on the Triticeae in North America. Phytologia 83(4): 302-311.

Barkworth, M. R. 2000. Changing perceptions of the Triticeae. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Colling-wood, Australia. Pp. 110-120.

Barkworth, M. E. & D. R. Dewey. 1985. Genomically based genera in the perennial Triticeae of North America: identification and membership. American J. Bot. 72: 767-776.

Barkworth, M. E. et al. 1983. New generic concepts in the Triticeae of the Intermountain Region: key and comments. Great Basin Nat. 43(4): 561-572.

Baum, B. R. 1977. Taxonomy of the tribe Triticeae (Poaceae) using various numerical techniques. I. Historical perspectives, data accumulation, and character analysis. Canadian J. Bot. 55(13): 1712-1740.

Baum, B. R. 1978. Taxonomy of the tribe Triticeae (Poaceae) using various numerical techniques. II. Classification. Canadian J. Bot. 56: 27-56.

Baum, B. R. 1978. Taxonomy of the tribe Triticeae (Poaceae) using various numerical techniques. III. Synoptic key to genera and synopses. Canadian J. Bot. 56: 374-385.

Baum, B. R. 1983. A phylogenetic analysis of the tribe Triticeae (Poaceae) based on morphological characters of the genus. Canadian J. Bot. 61: 518-535.

Baum, B. R. et al. 1987. Assessment of the genomic system of classification in the Triticeae. American J. Bot. 74: 1388-1395.

Baum, B. R. & D. B. O. Savile. 1985. Rusts (Uredinales) of Triticeae: evolution and extent of

coevolution, a cladistic analysis. Bot. J. Linn. Soc. 91: 367-394.

Bendich, A. J. & B. J. McCarthy. 1970. DNA comparisons among barley, oats, rye and wheat. Genetics 65: 545-565.

Bowden, W. M. 1958. Natural and artificial X *Elymordeum* hybrids. Canadian J. Bot. 36: 101-123.

Bowden, W. M. 1959. The taxonomy and nomenclature of the wheats, barleys, and ryes and their wild relatives. Canadian J. Bot. 37: 657-684.

Bowden, W. M. 1962. Cytotaxonomy of the native and adventive species of *Hordeum, Eremopyrum, Secale, Sitanion*, and *Triticum* in Canada. Canadian J. Bot. 40: 1675-1711.

Bowden, W. M. 1967. Taxonomy of intergeneric hybrids of the tribe Triticeae from North America. Canadian J. Bot. 45: 711-724.

Dewey, D. R. 1969. Trispecies hybrids of *Agropyron*, *Elymus*, and *Sitanion*. Bot. Gaz. 130: 203-213.

Dewey, D. R. 1983. Historical and current taxonomic perspectives of *Agropyron, Elymus*, and related genera. Crop Sci. 23(4): 637-642.

Dewey, D. R. 1984. The genomic system of classification as a guide to intergeneric hybridization with the perennial Triticeae. <u>In</u>, Gustafson, J. P. (Editor). Gene manipulation in plant improvement. Plenum Press. New York, NY. Pp. 209-279.

Dewey, D. R. 1984. Wide-hybridization and induced-polyploid breeding strategies for perennial grasses of the Triticeae tribe. Iowa State J. Res. 58(4): 383-399.

Dvorak, J. & H. -B. Zhang. 1992. Application of molecular tools for study of the phylogeny of diploid and polyploid taxa in Triticeae. Hereditas 116: 37-42.

Estes, J. R. & R. J. Tyrl. 1982. The generic concept and generic circumscription in the Triticeae: an end paper. <u>In</u>, Estes, J. R. et al (editors). Grasses and grasslands: systematics and ecology. Univ. Oklahoma Press. Norman. Pp. 145-164.

Frederiksen, S. 1993. Taxonomic studies in some annual genera of the Triticeae (Poaceae). Nordic J. Bot. 13(5): 481-493.

Frederiksen, S. & O. Seberg. 1992. Phylogenetic analysis of the Triticeae (Poaceae). Hereditas 116: 15-19.

Gupta, P. K. & B. R. Baum. 1989. Stable classification and nomenclature in the Triticeae: desirability, limitations and prospects. Euphytica 41: 191-197.

Jaradat, A. A. (editor). 1998. Triticeae III. Science Publ. Enfield, NH. 478 pp.

Jarvie, J. K. & M. E. Barkworth. 1990. Isozyme similarity in *Thinopyrum* and its relatives (Triticeae: Gramineae). Genome 33: 885-891.

Jarvie, J. K. & M. E. Barkworth. 1992. Morphological variation and genome constitution in some perennial Triticeae. Bot. J. Linn. Soc. 108: 167-180.

Jauhar, P. P. & C. F. Crane. 1989. An evaluation of

- Baum et al.'s assessment of the genomic system of classification in the Triticeae. American J. Bot. 76: 571-576.
- Kellogg, E. A. 1989. Comments on genomic genera in the Triticeae (Poaceae). American J. Bot. 76: 796-805.
- Kellogg, E. A. 1992. Tools for studying the chloroplast genome in the Triticeae (Gramineae): an EcoRI map, a diagnostic deletion, and support for *Bromus* as an outgroup. American J. Bot. 79: 186-197.
- Kellogg, E. A., R. Appels, & R. J. Mason-Gamer. 1996. When genes tell different stories: the diploid genera of Triticeae (Gramineae). Syst. Bot. 21(3): 321-347.
- Kimber, G. & Y. H. Zhao. 1983. The D genome of the Triticeae. Canadian J. Genetics and Cytol. 25: 581-589.
- Löve, A. 1984. Conspectus of the Triticeae. Feddes Repert. 95(7-8): 425-521. Nevski, S. A. 1933. Ueber das System der Tribe Hordeae Benth. Tr. Bot. Inst. Akad. Nauk SSSR, Ser. 1, 1: 9-32.
- Mason-Gamer, R. J. & E. A. Kellogg. 1996. Testing for phylogenetic conflict among molecular data sets in the tribe Triticeae (Gramineae). Syst. Biol. 45(4): 524-545.
- McIntyre, C. L. 1988. Variation at isozyme loci in Triticeae. Plant Syst. Evol. 160: 123-142.
- Melderis, A. 1978. Taxonomic notes on the tribe Triticeae (Gramineae) with special reference to the genera *Elymus* L. s. l. and *Agropyron* Gaertn. s. l. Bot. J. Linn. Soc. 76: 369-384.
- Monte, J. et al. 1993. Analysis of phylogenetic relationships in the Triticeae tribe using RFLPs. Theor. Appl. Genetics 86: 649-655.
- Pelger, S. 1993. Prolamin variation and evolution in Triticeae as recognized by monoclonal antibodies. Genome 36(6): 1042-1048.
- Peterson, G. & O. Seberg. 199x. [In press]. Phylogenetic analysis of the Triticeae (Poaceae) based on rpoA sequence data. Mol. Phylog. Evol.
- Sears, P. B. 1948. The cytology and genetics of the wheats and their relatives. Adv. in Genetics 2: 239-270.
- Stebbins, G. L., Jr. et al. 1946. Artificial and natural hybrids in the Gramineae, tribe Hordeae. I. *Elymus, Sitanion*, and *Agropyron*. American J. Bot. 33: 338-351.
- Stebbins, G. L., Jr. et al. 1946. Artificial and natural hybrids in the Gramineae, tribe Hordeae. II. *Agropyron, Elymus*, and *Hordeum*. American J. Bot. 33: 579-586.
- Stebbins, G. L. & L. A. Snyder. 1956. Artificial and natural hybrids in the Gramineae, tribe Hordeae. IX. Hybrids between western and eastern North American species. American J. Bot. 43(4): 305-312.
- Sun, G. -L., B. -H. Wu, & F. Liu. 1995. Cytogenetic and genomic relationships of *Thinopyrum elongatum* with two *Psathyrostachys* species and with *Leymus secalinus* (Poaceae). Pl. Syst. Evol. 197(1-4): 225-231.

- Wang, R. R. -C. 1990. Intergeneric hybrids between *Thinopyrum* and *Psathyrostachys* (Triticeae). Genome 33: 845-849.
- Wang, R. R.-C. 1992. Genome relationships in the perennial Triticeae based on diploid hybrids and beyond. Hereditas 116: 133-136.
- Wang, R. R.-C. et al. 1996. Genome symbols in the Triticeae (Poaceae). <u>In</u>, Proceedings second international Triticeae Symp. Utah State Univ. Design and Publ. Logan. Pp 29-34.
- West, J. G. et al. 1988. Evolution and systematic relationships in the Triticeae (Poaceae). Plant Syst. Evol. 160(1-2): 1-28.

AGROPYRON

- Beetle, A. A. 1961. The nomenclature of the crested wheatgrass complex. J. Range Manag. 14: 162.
- Bowden, W. M. 1965. Cytotaxonomy of the species and interspecific hybrids of the genus *Agropyron* in Canada and neighboring areas. Canadian J. Bot. 43: 1421-1448.
- Dewey, D. R. 1975. The origin of *Agropyron smithii*. American J. Bot. 62(5): 524-530.
- Dewey, D. R. 1986. Taxonomy of the crested wheatgrasses (*Agropyron*). <u>In</u>, Johnson, K. L. (Editor). Crested wheatgras: its values, problems, and myths. Utah State Univ. Logan. Pp. 31-41.
- Hanson, A. A. 1961. Comments on "The nomenclature of the crested wheatgrass complex." J. Range Management 14: 338, 339.
- Johnson, K. L. (editor). 1986. Crested wheatgrass: its values, problems, and myths: symposium proceedings. Utah State Univ. Logan. 348 pp.
- Jones, K. 1960. The typification of the genus *Agropyron* Gaertn. Taxon 9: 55, 56.
- Löve, A. 1982. Genetic evolution of the wheatgrasses. Biol. Zentralbl. 101: 199-212.
- Rogler, G. A. & R. J. Lorenz. 1983. Crested wheatgrass: early history in the United States. Range Manag. 36: 91-93.
- Runemark, H. & W. K. Heneen. 1968. *Elymus* and *Agropyron*, a problem of generic delimitation. Bot. Not. 121: 51-79.
- Sarkar, P. 1956. Crested wheatgrass complex. Canadian J. Bot. 34: 328-345.
- Swallen, J. R. & G. A. Rogler. 1950. The status of crested wheatgrass. Agron. J. 42: 571.
- Taylor, D. R. & L. W. Aarssen. 1988. An interpretation of pheno-typic plasticity in *Agropyron repens* (Gramineae). American J. Bot. 75(3): 401-413.
- Werner, P. A. & R. Rioux. 1977. The biology of Canadian weeds. 24. *Agropyron repens* (L.) Beauv. Canadian J. Plant Sci. 57: 905-919.

ELYMUS

Anamthawat-Jonsson, K. & S. K. Bodvarsdottir. 2001.

- Genomic and genetic relationships among species of *Leymus* (Poaceae: Triticeae) inferred from 18S-26S ribosomal genes. American J. Bot. 88(4): 553-559.
- Aniol, A. 1976. Serological studies within the tribe Triticeae. VII. Serological affinities within the genus *Elymus*. Genet. Pol. 17(3): 343-351.
- Assadi, M. & H. Runemark. 1995. Hybridisation, genomic constitution and generic delimitation in *Elymus* s. l. (Poaceae: Triticeae). Plant Syst. Evol. 194(3-4): 189-205.
- Baden, C. 1981. A taxonomic revision of *Psathyrostachys* (Poaceae). Nordic J. Bot. 11(1): 3-26.
- Barkworth, M. E. & R. J. Atkins. 1984. *Leymus* Hochst. Gramineae: Triticeae in North America: taxonomy and distribution. American J. Bot. 71(5): 609-625.
- Baum, B. R. 1979. The genus *Elymus* in Canada Bowden's generic concepts and key reappraised and relectotypification of *E. canadensis*. Canadian J. Bot. 57: 946-951.
- Baum, B. R., C. Yen. and J. -L. Yang. 1991. Roegneria: its generic limits and justification for its recognition. Canad. J. Bot. 69(2): 282-294.
- Bowden W. M. 1964. Cytotaxonomy of the species and interspecific hybrids of the genus *Elymus* in Canada and neighbouring areas. Canadian J. Bot. 42: 547-601.
- Brown, W.V. & G. A. Pratt. 1960. Hybridization and introgression in the grass genus *Elymus*. American J. Bot. 47: 669-676.
- Campbell, J. J. N. 2000. Notes on North American *Elymus* species (Poaceae) with paired spikelets. I. *E. macgregorii* sp. nov. and *E. glaucus* ssp. *mackenzii* comb. nov. J. Kentucky Acad. Sci. 61(2): 88-98.
- Church, G. L. 1954. Interspecific hybridization in eastern *Elymus*. Rhodora 56: 185-197.
- Church, G. L. 1967. Taxonomic and genetic relationships of eastern North American species of *Elymus* with setaceous glumes. Rhodora 69: 121-162.
- Gabel, M. L. 1984. A biosystematic study of the genus *Elymus* (Gramineae: Triticeae) in Iowa. Proc. Iowa Acad. Sci. 91: 140-146. Gould, F. W. 1945. Notes on the genus *Elymus*. Madroño 8: 42-47.
- Gould, F. W. 1945. Notes on the genus *Elymus*. Madroño 8: 42-47.
- Gould, F. W. 1947. Nomenclatorial changes in *Elymus* with a key to the Californian species. Madroño 9: 120-128.
- Jensen, E. R. 1972. A taxonomic study of single spikelet *Elymus* species of western United States. Ph. D. dissertation. Utah State Univ. Logan. 131 pp.
- Jensen, K. B. & S. L. Chen. 1992. An overview: systematic relationships of Elymus and Roegneria (Poaceae). Hereditas 116(1-2): 127-132.
- Jensen, K. B. et al. 1997. Cytological and molecular evidence for transferring *Elymus coreanus* from the genus *Elymus* to *Leymus* and molecular evidence for *Elymus californicus* (Poaceae: Triticeae). Int. J. Plant

- Sci. 158(6): 872-877.
- Mason-Gamer, R. J. 2001. Origin of North American *Elymus* (Poaceae: Triticeae) allotetraploids based on granule-bound starch synthase gene sequences. Syst. Bot. 26(4): 757-768.
- Melderis, A. 1978. Taxonomic notes on the tribe Triticeae (Gramineae), with special refreence to the genera *Elymus* L., sensu lato, and *Agropyron* Gaertner sensu lato. Bot. J. Linnean Soc. 76: 369-384.
- Melderis, A. & D. McClintock. 1983. The genera *Elymus* L. and *Leymus* Hochst. in Britain. Watsonia 14: 391-395.
- Orgaard, M. 1994. Intergeneric hybridization between species of *Leymus, Psathyrostachys* and *Hordeum* (Poaceae, Triticeae). Ann. Bot. 73: 471-479.
- Runemark, H. & W. K. Heneen. 1968. *Elymus* and *Agropyron*, a problem of generic delimitations. Bot. No. 121: 51-79.
- Salomon, B. & B. -R. Lu. 1992. Genomic groups, morphology, and sectional delimitation in Eurasian *Elymus* (Poaceae, Triticeae). Plant Syst. Evol. 180(1-2): 1-13.
- Stebbins, G. L. & A. Vaarama. 1954. Artificial and natural hybrids in the Gramineae, tribe Hordeae. VII. Hybrids and allopolyploids between Elymus glaucus and Sitanion spp. Genetics 39: 378-395.
- Svitashev, S. et al. 1996. A study of 28 *Elymus* species using repetitive DNA sequences. Genome 39: 1093-1101.
- Wilson, F. D. 1963. Revision of *Sitanion* (Triticeae, Gramineae). Brittonia 15: 303-323.
- Zhang, H. B. & J. Dvorak. 1991. The genome origin of tetraploid species of *Leymus* (Poaceae: Triticeae) inferred from variation in repeated nucleotide sequences. American J. Bot. 78: 871-884.

TAENIATHERUM

- Dahl, B. E. & E. W. Tisdale. 1975. Environmental factors related to medusa head distribution. J. Range Manag. 28: 463-468.
- Frederiksen, D. 1986. Revision of Taeniatherum (Poaceae). Nordic J. Bot. 6: 389-397.
- Frederiksen, S. 1994. Hybridization between *Taeniatherum caputmedusae* and *Triticum aestivum* (Poaceae). Nordic J. Bot. 14(1): 3-6.
- Frederiksen, S. and R. von Bothmer. 1986. Relationships in *Taeniatherum* (Poaceae). Canadian J. Bot. 64: 2343-2347.
- Frederiksen, S. & R. von Bothmer. 1989. Intergeneric hybridization between *Taeniatherum* and different genera of Triticeae, Poaceae. Nordic J. Bot. 9: 229-240.
- Hilken, T. O. & R. E. Miller. 1980. Medusahead (*Taeniatherum asperum* Nevski): a review and bibliography. Agric. Exp. Sta. Bull. No. 644. Oregon State Univ. Corvallis.
- Humphries, C. J. 1978. Variation in Taeniatherum

- caput-medusae (L.) Nevski. Bot. J. Linnean Soc. 76: 340-344.
- McKell, C. M., J. P. Robison, & J. Major. 1962. Ecotypic variation in medusahead, an introduced annual grass. Ecology 43: 686-698.
- Young, J. A. 1992. Ecology and management of medusa head (*Taeniatherum caput-medusae* ssp. *asperum* [Simk.] Melderis). Great Basin Nat. 52(3): 245-252.

SITANION

- Dewey, D. R. & A. H. Holmgren. 1962. Natural hybrids of *Elymus cinereus* X *Sitanion hystrix*. Bull. Torrey Bot. Club 89: 217-228.
- Smith, J. G. 1899. Studies on American grasses. A synopsis of the genus *Sitanion.* U. S. Dept. Agric. Div. of Agrostoogy. Bull. 18: 1-21.
- Wilson, F. D. 1963. Revision of *Sitanion* (Triticeae, Gramineae). Brittonia 15: 303-323.

HORDEUM

- Baden, C. & R. von Bothmer. 1994. A taxonomic revision of *Hordeum* sect. *Critesion*. Nordic J. Bot. 14(2): 117-136.
- Baum, B. R. 1987. Classification of cultivated barley (*Hordeum vulgare*). II. Elaboration of cultivar groups. Canadian J. Bot. 65: 2152-2160.
- Baum, B. R. & D. A. Johnson. 2000. The 5S rRNA gene units in the native New World annual *Hordeum* species (Triticeae: Poaceae). Canadian J. Bot. 78(12): 1590-1602.
- Baum, B. R. & L. G. Bailey. 1986. Taxonomy of the North and South American species of *Hordeum* section *Hordeastrum*. Canadian J. Bot. 64(8): 1745-1759.
- Baum, B. R. & L. G. Bailey. 1988. A taxonomic study of the annual *Hordeum depressum* and related species. Canadian J. Bot. 66(3): 401-408.
- Baum, B. R. & L. G. Bailey. 1990. Key and synopsis of North American *Hordeum* species. Canadian J. Bot. 68: 2433-2442.
- Bengtsson, B. O. 1992. Barley genetics -- not only for the beer. Trends in Genet. 8(1): 3-5.
- Bergal, P. 1948. Les lodicules et leur utilisation dans la sys-tématique du genre Hordeum. Annales des Sciences Naturelles Botanique. Series 11, 9: 187-266.
- Best, K. F., J. D. Banting, & G. G. Bowes. 1978. The biology of Canadian weeds. XXXI. *Hordeum jubatum* L. Canadian J. Plant Sci. 58: 699-708.
- Booth, T. A. & A. J. Richards. 1976. Studies in the *Hordeum murinum* aggregate. I. Morphology. J. Linnean Soc. Bot. 72: 149-159.
- Booth, T. A. & A. J. Richards. 1978. Studies in the *Hordeum murinum* aggregate. Studies in the *Hordeum murinum* L. aggregate: disc electrophoresis of seed proteins. Bot. J. Linnean Soc. 76(2): 115-126.
- Bothmer, R. von. 1986. Interspecific crosses in *Hordeum* (Poaceae). Plant Syst. Evol. 153: 49-64.

- Bothmer, R. von. 1990. Evolutionary patterns in wild species of *Hordeum*. Sommerfeltia 11: 65-72.
- Bothmer, R. von & N. Jacobsen. 1985. Origin, taxonomy, and related species. <u>In</u>, Rasmussen, D. G. (editor). Barley. Agronomy Monograph No. 26. American Soc. Agron. Madison, WI. Pp. 19-56.
- Bothmer, R. von, B. Jorgensen, & I. Linde-Laursen. 1987. Natural variation, phylogeny and genetic resources in Hordeum. Barley Genetics 5: 22-33.
- Bothmer, R. von et al. 1982. Revision of the *Hordeum pusillum* group. Nordic J. Bot. 2: 307-321.
- Bothmer, R. von et al. 1989. Variation and differentiation in *Hordeum marinum* (Poaceae). Nordic J. Bot. 9(1): 1-10.
- Bothmer, R. von et al. 1991. An ecogeographical study of the genus *Hordeum*. Systematic and ecogeographical studies on crop gene pools 7: 1-127.
- Bothmer, R. von et al. 1993. Variation and taxonomy in *Hordeum depressum* and in the *H. brachyantherum* complex (Poaceae). Nordic J. Bot. 13(1): 3-18.
- Briggs, D. E. 1978. Barley. Chapman & Hall. London, England. 612 pp.
- Covas, G. 1949. Taxonomic observations on the North American species of *Hordeum*. Madroño 10: 1-121.
- Dickson, A. D. et al. 1979. Barley: origin, botany, culture, winter hardiness, genetics, utilization, pests. Agric. Handbook No. 338. U. S. Dept. Agric. Washington, D. C. 154 pp.
- Doebley, J., R. von Bothmer, & S. Larson. 1992. Chloroplast DNA variation and the phylogeny of Hordeum (Poaceae). Amer. J. Bot. 79(5): 576-584.
- Giles, B. E. & L. P. Lefkovitch. 1986. A taxonomic investigation of the *Hordeum murinum* complex (Poaceae). Plant Syst. Evol. 153(3-4): 181-197.
- Jaaska, V. 1994. Isoenzyme evidence on the systematics of *Hordeum* sect. *Marina* (Poaceae). Plant Syst. Evol. 191(3-4): 213-226.
- Jacobsen, N. & R. von Bothmer. 1995. Taxonomy in the *Hordeum murinum* complex. Nordic J. Bot. 15(5): 449-458.
- Jorgensen, R. B. 1986. Relationships in the barley genus (*Hordeum*): an electrophoretic examination of proteins. Hereditas 104: 273-291.
- Komatsuda, T. et al. 2001. Phylogenetic analysis of *Hordeum marinum* Huds. based on nucleotide sequences linked to the vrs1 locus. Plant Syst. Evol. 227(3-4): 137-144.
- Mitchell, W. W. 1967. On the *Hordeum jubatum-H. brachyanth-erum* question. Madroño 19: 108-110.
- Morrison, J. W. 1959. Cytogenetic studies in the genus Hordeum. I. Chromosome morphology. Canadian J. Bot. 37: 527-538.
- Morrison, J. W. et al. 1959. Cytogenetic studies in the genus Hordeum. II. Interspecific and intergeneric crosses. Canadian J. Plant Sci. 39: 375-383.

Shewry, P. R. (Editor). 1992. Barley: genetics, biochemistry, molecular biology and biotechnology. Biotechnology in Agric. No. 5. CAB International. Wallingford, U. K.

AEGILOPS

Hammer, K. 1980. Zur Taxonomie und Nomenklatur der Gattung Aegilops L. Feddes Repert. 91: 225-258.

Kihara, H. 1954. Considerations on the evolution and distribution of *Aegilops* species based upon the analyser-method. Cytologia 19: 336-357.

Slageren, M. W. van. 1994. Wild wheats: a monograph of *Aegilops* L. and *Ambylopyrum* (Jaub. & Spach) Eig (Poaceae). Wangeningen Agric. Univ. Pap. 94(7): 1-512.

Waines, J. G. & D. Barnhart. 1992. Biosystematic research in *Aegilops* and *Triticum*. Hereditas 116(3): 207-212.

Witcombe, J. R. 1983. A guide to the species of *Aegilops* L. Their taxonomy, morphology and distribution. Int. Board Plant Genet. Res., Wheat Prog. Rome. 74 pp.

TRITICUM

Appels, R. 1982. The molecular cytology of wheat-rye hybrids. Intern. Rev. Cytol. 80: 93-132.

Briggle, L. W. & L. P. Reitz. 1963. Classification of *Triticum* species and of wheat varieties grown in the United States. Agric. Tech. Bull. 1278. 135 pp.

Feldman, M. & E. R. Sears. 1981. The wild gene resources of wheat. Sci. American. 244: 98-102.

Franke, R. et al. 1992. Intergeneric hybrids between *Triticum aestivum* and wild Triticeae (Poaceae). Hereditas 116: 225-231.

Harlan, J. R. & D. Zohary. 1966. Distribution of wild wheat and barley. Science 153. 1074-1080.

Helbaek, H. 1966. Commentary on the phylogenesis of *Triticum* and *Hordeum*. Econ. Bot. 20: 350-360.

Johnson, B. L. 1975. Identification of the apparent B-genome donor of wheat. Canadian J. Genetics and Cytol. 17: 21-39.

Kimber, G. 1974. A reassessment of the origin of the polyploid wheats. Genetics 78(1): 487-492.

MacKey, J. 1966. Species relationships in *Triticum*. Hereditas Suppl. 2:237-276.

Nainawatee, H. S. & N. B. Das. 1972. Species relationships in *Triticum* by protein electrophoresis. Curr. Sci. 41: 778, 779.

Rayburn, A. L. & B. S. Gill. 1988. Repeated DNA sequences in *Triticum* (Poaceae): chromosomal mapping and its bearing on the evolution of B and G genomes. Plant Syst. Evol. 159: 229-235.

Sarkar, P. & G. L. Stebbins. 1956. Morphological evidence concerning the origin of the B genome in wheat. American J. Bot. 43: 297-304.

Waines, J. G. & D. Barnhart. 1992. Biosystematic

research in Aegilops and Triticum. Hereditas 116: 207-212.

Wright, G. M. 1958. Grain in the glume of wheat. Nature 181: 1812, 1813.

Zohary, D. 1965. Colonizer species in the wheat group. $\underline{\text{In}}$, Baker, H. G. & G. L. Stebbins (Editors). The genetics of colonizing species. Pp. 403-423.

SECALE

Aniol, A. 1976. Serological studies within the tribe Triticeae: V. Serological affinity of species in the genus *Secale*. Genet. Pol. 17(2): 123-131.

Bowden, W. M. 1959. The taxonomy and nomenclature of wheats, barleys and ryes. Canadian J. Bot. 37: 651-684.

Dedio, W. et al. 1969. Numerical chemotaxonomy in the genus *Secale*. Canadian J. Bot. 47: 1175-1180.

Howlett, B. J. & A. E. Clarke. 1981. Isolation and partial characterization of two antigenic glycoproteins from rye grass pollen. Biochm. J. 197: 695-706.

Khush, G. 1962. Cytogenetic and evolutionary studies in *Secale*. II. Evolution 16: 484-496.

Khush, G. & G. L. Stebbins. 1961. Cytogenetic and evolutionary studies in *Secale*. I. Some new data on the ancestry of *S. Cereale*. American J. Bot. 48: 723-730.

Sencer, H. A. & J. G. Hawkes. 1980. On the origin of cultivated rye. Biol. J. Linn. Soc. 13: 299-313.

Stutz, H. C. 1972. On the origin of cultivated rye. American J. Bot. 59(1): 59-70.

Vences, F. J., F. Vaquero, & M. Perez de la Vega. 1987. Phylogenetic relationships in *Secale* (Poaceae): an isozymatic study. Plant Syst. Evol. 157(1-2): 33-47.

Vosa, C. G. 1974. The basic karyotype of rye (*Secale cereale*) analysed with giemsa and fluorescence methods. Heredity 33: 403-408.

X TRITICOSECALE

Baum, B. R. & P. K. Gupta. 1990. Taxonomic examination of triticale (X *Triticosecale*). Canadian J. Bot. 68: 1889-1893.

Briggle, L. W. 1969. Triticale -- a review. Crop Sci. 9: 197-202.

Hawthorn, J. 1976. Triticale -- an intergeneric hybrid. Trans. Bot. Soc. Edinburgh 42(4): 505-512.

Müntzing, A. 1979. Triticale: results and problems. Adv. Plant Brreding 10: 1-103.

Stace, C. A. 1987. Triticale: a case of nomenclatural mistreatment. Taxon 36: 445-452.

EREMOPYRUM

Frederiksen, S. 1991. Taxonomic studies in *Eremopyrum* (Poaceae). Nordic J. Bot. 11(3): 271-285.

TRIBE MELICEAE

Mejia-Saulés, T. & F. A. Bisby. 2000. Preliminary views on the tribe Meliceae (Gramineae: Pooideae). <u>In,</u> Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 83-88.

MELICA

Boyle, W. S. 1945. A cytotaxonomic study of the North American species of *Melica*. Madroño 8: 1-26.

GLYCERIA

Church, G. L. 1949. A cytotaxonomic study of *Glyceria* and *Puccinellia*. American J. Bot. 36: 155-165.

Esparza, R. R. & J. Maze. 1997. A taxonomic study of the grass genus *Glyceria* (mannagrass) in British Columbia. Canadian Field-Naturalist 111(2): 194-199.

PLEUROPOGON

But, P. 1977. Systematics of *Pleuropogon* R. Br. (Poaceae). Ph. D. dissertation. Univ. California, Berkeley. 220 pp.

But, P. P. H. 1986. Taxonomy of Oregon semaphore grass, *Lophochlaena oregona* (Poaceae). Madroño 33(2): 146, 147.

But, P. 1994. New combinations in *Pleuropogon* (Poaceae). Novon 4: 16, 17.

Löve, A. & D. Löve. 1980. Resurrection of the grass genus *Lophochlaena*. Bol. Soc. Broteriana, Ser. 2, 53(1): 563-585.

SCHIZACHNE

Boyle, W. S. 1944. Cytological evidence for the taxonomic position of *Schizachne purpurascens*. Madroño 7: 129, 130.

Swallen, J. R. 1928. The grass genus *Schizachne*. J. Washington Acad. Sci. 18: 203-206.

TRIBE STIPEAE

Barkworth, M. E. 1981. Foliar epidermes and taxonomy of North American Stipeae (Gramineae). Syst. Bot. 6(2): 136-152.

Barkworth, M. E. 1983. *Ptilagrostis* in North America and its relationship to other Stipeae (Gramineae). Syst. Bot. 8(4): 395-419.

Barkworth, M. E. 1990. *Nassella* (Gramineae, Stipeae): revised interpretation and nomenclatural changes. Taxon 39: 597-614.

Barkworth, M. E. 1993. North American Stipeae (Gramineae): taxonomic changes and other comments. Phytologia 74: 1-25.

Barkworth, M. E. & J. Everett. 1987. Evolution in the Stipeae: identification and relationships of its monophyletic taxa. <u>In</u>, Soderstrom, T. R. et al. (editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 251-

264.

Jacobs, S. W. L. et al. 2000. Relationships within the stipoid grasses (Gramineae). <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 75-82.

Soderstrom, T. R. et al. (Editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 251-264.

Renvoize, S. A. 1985. A survey of leaf-blade anatomy in grasses. VI. Stipeae. Kew Bull. 40: 731-736.

Scholz, H. 1982. Uber Mikro- und Makrohaare einiger *Piptatherum* und *Stipa*-Arten (Stipeae, Gramineae). Willdenowia 12: 235-240.

STIPA

Amme, D. & S. W. Edwards. 1988. Needlegrass notes. The Four Seasons 8(2): 28-34.

Barkworth, M. E. 1978. A taxonomic study of the large-glumed species of *Stipa* (Gramineae). occurring in Canada. Canadian J. Bot. 56: 606-625.

Barkworth, M. E. & J. Maze. 1979. Proposal to reject *Stipa columbiana* (Poaceae) and nomenclatural changes affecting three western North American species of *Stipa* (Poaceae). Taxon 28: 621-625.

Barkworth, M. E., J. McNeill, & J. Maze. 1979. A taxonomic study of *Stipa nelsonii* (Poaceae) with a key distinguishing it from related taxa in western North America. Canadian J. Bot. 57: 2539-2553.

Barkworth, M. E., J. Valdes-Reyna, & R. Q. Landers, Jr. 1989. *Stipa clandestina*: new weed threat on southwestern rangelands. Weed Tech. 3(4): 699-702.

Bartolome, J. W. & B. Gemmill. 1981. The ecological status of *Stipa pulchra* (Poaceae) in California. Madroño 28: 172-184.

Brown, W. V. 1952. The relation of soil moisture to cleistogamy in *Stipa leucotricha*. Bot. Gaz. 113: 438-444.

Dedecca, D. M. 1954. Studies on the California species of *Stipa* (Gramineae). Madroño 12: 129-139.

Hall, O. And B. L. Johnson. Electrophoretic analysis of the amphiploid of *Stipa viridula* x *Oryzopsis hymenoides* and its parent species. Hereditas 48: 530-535.

Hitchcock, A. S. 1925. The North American species of *Stipa*. Contr. U. S. Natl. Herb. 24(7): 215-262.

Jacobs, S. W. L. & J. Everett. 1996. *Austrostipa*, a new genus, and new names for Australian species formerly included in *Stipa* (Gramineae). Telopea 6(4): 579-595.

Johnson, B. L. 1962. Amphiploidy and introgression in *Stipa*. American J. Bot. 49(2): 253-262.

Love, R. M. 1946. Interspecific hybridization in *Stipa* L. I. Natural hybrids. American Natl. 80: 189-192.

Love, R. M. 1954. Interspecific hybridization in *Stipa*. II. Hybrids of *S. cernua*, *S. lepida*, and *S. pulchra*. American J. Bot. 41: 107-110.

Maze, J. 1965. Notes and key to some California species of *Stipa*. Leaflts. West. Bot. 10: 157-161.

Maze, J. 1972. Notes on the awn anatomy of *Stipa* and *Oryzopsis* (Gramineae). Syesis 5: 169-171.

ORYZOPSIS

Johnson, B. L. 1945. Cytotaxonomic studies in *Oryzopsis*. Bot. Gaz. 107: 1-32.

Johnson, B. L. 1945. Natural hybrids between *Oryzopsis hymen-oides* and several species of *Stipa*. American J. Bot. 32: 599-608.

Johnson, B. L. 1960. Natural hybrids between *Oryzopsis* and *Stipa*. I. *Oryzopsis hymenoides* x *Stipa speciosa*. American J. Bot. 47: 736-742.

Maze, J. & K. A. Robson. 1996. A new species of *Achnatherum (Oryzopsis*) from Oregon. Mardoño 43(3): 393-403.

Mehlenbacher, L. E. 1970. Floret development, embryology and systematic position of *Oryzopsis hendersonii* (Gramineae). Canadian J. Bot. 48: 1741-1758.

Voss, E. G. 1961. Which side is up? A look at the leaves of *Oryzopsis*. Rhodora 63: 285-287.

Weber, W. A. 1957. A new intergeneric natural hybrid involving *Oryzopsis* and *Stipa*. Rhodora 59: 273-277.

PTILAGROSTIS

Barkworth, M. E. 1983. *Ptilagrostis* in North America and its relationship to other Stipeae (Gramineae). Syst. Bot. 8(4): 395-419.

PIPTOCHAETIUM

Barkworth, M. E. 1987. *Piptochaetium* Presl (Gramineae, Stipeae) in North- and Mesoamerica. Taxonomic and distributional observations. Brenesia 25/26: 169-178.

Cialdella, A. M. & L. M. Giussani. 2002. Phylogenetic relationships of the genus *Piptochaetium* (Poaceae, Pooideae, Stipeae): evidence from morphological data. Ann. Missouri Bot. Gard. 89(3): 305-336.

Parodi, L. R. 1944. Revision de las graminas Australes Americanas del genero *Piptochaetium*. Revista Mus. La Plata, Secc. Bot. 6: 214-310.

Valencia, J. I. & M. Costas. 1968. Estudios citotaxonomicos sobre *Piptochaetium* (Gramineae). Bol. Soc. Argent. Bot. 12: 167-179.

TRIBE HAINARDIEAE

HAINARDIA

Gandhi, K. N. 1966. Nomenclatural novelties for the western hemisphere plants – I. [Retention of *Hainardia* over *Monerma*] Harvard Pap. Bot. 8: 63-66.]

Scholz, H. 1995. *Monerma* P. Beauv. (Poaceae) -- not an illegitimate name. Feddes Rep. 106: 169-171.

Tateoka, T. 1959. Notes on grasses. VII. Cytological evidence for the phylogenetic difference between *Lepturus* and *Monerma*. Cytologia 23: 447-451.

Tateoka, T. 1959. *Lepturus* and *Monerma*: a remarkable example of parallel development of gross morphology in grasses. Evolution 13: 418-420.

PARAPHOLIS

Runemark, H. 1962. A revision of *Parapholis* and *Monerma* in the Mediterranean. Bot. Not. 115: 1-17.

Worley, T. 1993. *Parapholis*. <u>In</u>, Hickman, J. C. (editor). The Jepson manual: higher plants of California. Univ. California Press. Berkeley. P. 1278.

3.06 - SUBFAMILY ARUNDINOIDEAE

TECHNICAL DESCRIPTION

Habit: Cane-like, reed-like, or bamboo-like; sometimes perennial herbs

Root Hairs: Equal

Leaf Epidermis: A combination of festucoid and

panicoid features

Leaf Anatomy: A combination of festucoid and bambusoid (arm cells present), with a few panicoid features; fusoid cells lacking

Spikelet: 1- to several-flowered; upper florets often reduced; in a few lower ones imperfect

Flower: Lodicules 2; stamens 3 [rarely 6]; stigmas 2, often densely and minutely plumose; flowers perfect or unisexual

Embryo: P-PF

Cytology: x = 6, 10, 11, or 12; chromosomes

small

Photosynthetic Pathway: C₃

Distribution: Cosmopolitan, especially southern

hemisphere.

SYSTEMATICS

This subfamily is a mess! It was not recognized by Hitchcock and Chase. Instead, they placed these grasses in their tribes Festuceae and Aveneae. It appears to me that the more we learn from the newer molecular studies, the less tenable our various treatments of Arundinoideae become.

The placement of *Gynerium* remains unsettled. It has been placed in its own tribe; the Grass Phylogeny Working Group left it "incertae sedis," which translates roughly from the Latin as "we don't know where the hell it goes."

TRIBE: ARUNDINEAE

Tall, coarse, rhizomatous or densely clumped perennials, often of wet sites. Inflorescence a conspicuous terminal panicle. Spikelets few- to several-flowered, often long-hairy, bisexual or unisexual, laterally compressed, disarticulating above the glumes and between the florets. See Section 4 for a new to the genera found in North America.

ARUNDO. Giant reed. Rhizomatous perennials. Culms woody, 2-6 m tall. Leaf blades broad, not pseudopetiolate. Inflorescence a terminal, plumose panicle. Spikelets several-flowered, bisexual, laterally compressed, disarticulating above the glumes and

between the florets. Glumes 2, \pm equal, 3-nerved, tapering to a point, awnless. Lemmas 3-nerved, villous on its lower half, tapering to a point or awn. Palea 2-keeled and -nerved. Rachilla glabrous. Lodicules 2; stamens 3; stigmas 2. X = 12; 2n = 60, 72, 110, and 112.

A genus of 3-6 species. *A. donax* is weedy along waterways; it is also grown as an ornamental and its is used in erosion control. It is also one of the sources of material used to make the reeds for certain wind instruments.

PHRAGMITES. Reed, common reed. Robust rhizomatous or stoloniferous perennials. Culms herbaceous to woody, to 4 [10] m tall. Leaf blades broad, not pseudopetiolate. Inflorescence a terminal, plumose panicle. Spikelets several-flowered, bisexual, disarticulating above the glumes and between the florets. Glumes 2, pointed, awnless, the first 1-nerved and the second 3-nerved; lemmas 3-nerved, awned or awnless. Rachilla with long, silky hairs. Palea 2-nerved. Lodicules 2; stamens 3; stigmas 2. X = 12; 2n = 36, 44, 46, 48, 49, 50, 51, 52, 54, and 96.

A genus of 3 cosmopolitan species. Our only North American representative is *Ph. australis* (= *Ph. communis* in H & C), often considered the most widely occurring vascular plant. It is found around lakes and along waterways over much of the continent.

GYNERIUM. Uva grass, caña brava. Robust, rhizomatous perennials to 10 m tall. Leaf blades to 2 m long; sharply serrulate. Plants dioecious. Inflorescence a conspicuous panicle. Female spikelets with a hairy callus.

A genus of one species, *G. sagittatum*, which is native to streamsides and wet places from Mexico into South America. It has become established in Florida. Stems used for construction, arrow-shafts; leaves for thatch, weaving, and basketry; inflorescences often painted some really tacky color and sold for some outrageous price in "import" shops.

SELECTED REFERENCES

Barker, N. P. et al. 1995. Polyphyly of Arundinoideae (Poaceae): evidence from *rbc*l sequence data. Syst. Bot. 20(4): 423-435.

Barker, N. P., H. P. Linder, & E. H. Harley. 1998. Sequences of the grass-specific insert in the chloroplast *rpoC2* gene elucidate generic relationships of the Arun-dinoideae (Poaceae). Syst. Bot. 23: 327-350.

Conert, H. J. 1987. Current concepts in the systematics of the Arundinoideae. \underline{In} , Soderstrom, T. R. et al. (editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 239-250.

Hilu, K. W. & A. Esen. 1990. Promalins in systematics of Poaceae subfam. Arundinoideae. Plant Syst. Evol. 173(1-2): 57-70.

Hsaio, C. et al. 1998. A molecular phylogeny of the sub-family Arundinoideae (Poaceae) based on sequences of rDNA (ITS). Australian Syst. Bot. 11: 41-52.

Renvoize, S. A. 1981. The subfamily Arundinoideae and its postion in relation to a general classification of the Gramineae. Kew Bull. 36: 85-102.

Renvoize, S. A. 1986. A survey of leaf-blade anatomy in grasses. VIII. Arundinoideae. Kew Bull. 41: 323-342.

Tucker, G. C. 1990. The genera of Arundinoideae (Gramineae) in the southeastern United States. J. Arnold Arbor. 71(2): 145-177.

TRIBE ARUNDINEAE

ARUNDO

Perdue, R. E., Jr. 1958. *Arundo donax* -- source of musical reeds and industrial cellulose. Econ. Bot. 12(4): 368-404.

MOLINIA

Jirasek, V. 1966. Uber die systematische Einordnung der Gattung *Molinia* (Gramineae). Preslia 38: 23-35.

PHRAGMITES

Clayton, W. D. 1968. The correct name of the common reed. Taxon 17: 168, 169.

Clevering, O. A. & J. Lissner. 1999. Taxonomy, chromo-some numbers, clonal diversity and population dynamics of *Phragmites australis*. Aquatic Bot. 64(3-4): 185-208.

Gorenflot, R. 1976. Le complexe polyploide du *Phragmites australis* (Cav.) Trin. ex Steud. (= *P. communis* Trin.). Bull. Soc. Bot. France 123: 261-271.

Pohl, R. W. 1985. Aerial rhizomes in *Phragmites australis*. Proc. Iowa Acad. Sci. 92: 103.

Ricciuti, E. R. & L. Line. 1983. The all too common, common reed. Audubon 85(5): 64-67.

THYSANOLAENA

Tateoka, T. 1956. Notes on grasses. II. Part 2. Peculiar features of leaf structure in the genus *Thysanolaena*. Bot. Mag. Tokyo 69: 337-339.

TRIBE GYNERIEAE

Sánchez-Ken, J. G. & L. G. Clark. 2001. Gynerieae, a new neotropical tribe of grasses (Poaceae). Novon 11(3): 350-352.

GYNERIUM

Kalliola, R. & S. A. Renvoize. 1994. One or more species of *Gynerium*? (Poaceae). Kew Bull. 49(2): 305-320.

Pohl, R. W. 1983. *Gynerium sagittatum* (cana brava, cane). <u>In</u>, Janzen, D. H. (editor). Costa Rican natural history. Univ. Chicago Press. Chicago, IL. Pp. 248, 249.

3.07 - SUBFAMILY DANTHONIOIDEAE

TECHNICAL DESCRIPTION

Habit: Caespitose perennials; reed-like in *Cortaderia*

Root Hairs:

Leaf Epidermis: Panicoid microhairs present; stomates absent or rare; silica bodies various

Leaf in Cross-section: Mesophyll non-radiate

Inflorescence: Panicle, sometimes reduced to a raceme or solitary spikelet

Spikelets: 2- to many-flowered; laterally compressed; disarticulating above the glumes

Flowers: Lodicules 2; stamens 3 (0 in unisexual

florets); stigmas 2

Embryo Formula: P - P F

Cytology: x = 9; diploids and tetraploids

Photosynthetic Pathway: C₃

Distribution: Temperate, especially southern hemi-sphere; only *Danthonia* is native to North

America

SYSTEMATICS

Hitchcock & Chase placed these grasses in their Festucoideae. More recent workers segregated them out as arundinoids. Recent research has resulted in recognizing seven clades, including the *Cortaderia* clade and the *Danthonia* clade.

TRIBE: DANTHONIEAE

Annuals or perennials herbs. Inflorescence a panicle, raceme, or occasionally reduced to a single spikelet. Spikelets few- to several-flowered, bisexual, disarticulating above the glumes and between the florets. Glumes 2, longer than the florets; lemmas bifid or toothed, awned or mucronate.

This tribe of predominantly southern hemisphere grasses is not well represented in North America. Its circum-scription remains unsettled. When C. E. Hubbard originally described the group in 1948, he separated out genera of the traditional Aveneae. Agnes Chase retained our few representatives in the Aveneae. The microcharacters of this group combine poold and panicoid features. See Section 4 for a key to the genera of North American Danthonieae.

CORTADERIA. Pampas grass. Robust, caespitose perennials. Culms to 4 m tall. Leaves mostly basal, the blades often with harsh, toothed margins. Inflorescence a terminal panicle (conspicuously

plumose in female plants). Spikelets 2-3 [5-] flowered, unisexual (the species gynodioecious), laterally compressed, disarticulating above the glumes and between the florets, the rachilla extended beyond the uppermost floret. Glumes 2, \pm equal, glabrous, 1-to 3-nerved, awnless; lemmas 3-nerved, awned or awnless, conspicuously hairy on back and base in pistillate spikelets; palea 2-nerved and keeled. Lodicules 2; stamens 0 or 3; stigmas 0 or 2. X= 9; 2n = 36, 72, 90, and 108.

A genus of 24 species, native to New Zealand and to South America. *C. selloana* is an attractive ornamental that sometimes turns weedy. *Cortaderia jubata* is a pernicious weed of coastal California. You should kill pampas grass where ever it occurs. This will not be easy. You may need a flame thrower or a small nuclear device.

DANTHONIA. Poverty-oats. Caespitose perennials. Culms to 1 m tall. Inflorescence a few-flowered panicle or raceme (sometimes reduced to a single spikelet); our North American plants also have cleistogamous spikelets hidden within the leaf sheaths. Spikelets several-flowered, bisexual, disarticulating above the glumes and between the florets. Glumes 2, 3- to 7-nerved, much longer than the florets; lemmas hairy, 7- to many-nerved (often indistinctly so), the apex 2-toothed or -cleft, with a flat, twisted, geniculate awn from its midnerve; palea 2-nerved and -keeled. Lodicules 2; stamens 3; stigmas 2.

A genus of about 20 species of mesophytic to xerophytic habitats. Several are important pasture species. *D. spicata* occurs in much of the U. S., except for the far Southwest. *D. californica* and *D. unispicata* are found over much of the West.

RYTIDOSPERMA. Hairy oat grass, hairy-danthonia, poverty grass. Plants perennial, caespitose, spreading, sometimes rhizomatous. Inflorescence a raceme or panicle. Spikelets 3- to several-flowered, bisexual, dis-articulating above the glumes and between the florets. Glumes 2, more or less equal. Lemmas 5- to 9-veined, with 2 more or less complete rows of tufts of hairs (the feature that distinguishes the genus from *Danthonia*). Lodicules 2; stamens 3; stigmas 2.

A genus of about 45 species native to Asia, Oceania, and South America. Three species have been introduced into the United States, all of them along the Pacific coast. Only *R. penicillatum* (= *Danthonia pilosa* in older literature) is well established, especially in northern California and southern Oregon.

SCHISMUS. Mediterranean grass. Annual or weak perennials, caespitose or decumbent. Inflorescence a contracted or loosely spicate panicle. Spikelets several-flowered, slightly laterally compressed, disarticulating above or below the glumes, rachilla prolonged beyond uppermost floret. Glumes 2, 3- to 7-nerved, ± equal, shorter than or equaling enclosed florets. Lemmas similar to glumes, apex bifid or emarginate, awnless, mucronate, or awned. Palea 2-nerved, rounded or acute. Lodicules 2; stamens 3; stigmas 2. X = 6.

A genus of 5 species native to Africa, and from the Mediterranean to India. *S. arabicus* (Arabian grass) and *S. barbatus* (Mediterranean grass) have become major weeds in parts of the Southwest and in the deserts of California.

SELECTED REFERENCES

Barker, N. P., C. M. Morton, & H. P. Linder. 2000. The Danthonieae: generic composition and relationships. In, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 221-229.

de Wet, J. M. 1956. Leaf anatomy and phylogeny in the tribe Danthonieae. Amer. J. Bot. 43: 175-182.

Linder, H. P. & N. P. Barker. 2000. Biogeography of the Danthonieae. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 231-238.

Verboom, G. A., H. P. Linder, & N. P. Barker. 1994. Haustorial synergids: an important character in the systematics of danthonioid grasses (Arundinoideae, Poaceae). American J. Bot. 81: 1601-1610.

CORTADERIA

Acevedo de Vargas, R. 1959. Las especies de Gramineas del genero *Cortaderia* en Chile. Bol. Mus. Nac. Hist. Nat. Chile 27: 205-246.

Astegiano, M. E., A. M. Anton, & H. E. Connor. 1995. Sinopsis del genero *Cortaderia* (Poaceae) in Argentina. Darwiniana 33(1-4): 43-51.

Connor, H. E. 1971. A naturalized *Cortaderia* (Gramineae) in California. Madroño 21: 39, 40.

Connor, H. E. 1974. Breeding systems in *Cortaderia* (Gramineae). Evol. 27(4): 663-678.

Connor, H. E. 1983. Names and types in *Cortaderia* Stapf (Gramineae). II. Taxon 32: 633, 634.

Connor, H. E. and E. Edgar. 1974. Names and types in *Cortaderia* Stapf (Gramineae). Taxon 23(4): 595-605.

Cooper, D. W. 1967. Pampas grass in Humboldt County [California]. Agron. Notes June: 19, 20.

Costas Lippmann, M. A. 1976. Ecology and reproductive biology of the genus *Cortaderia* in California. Ph. D. dissertation. Univ. of California, Berkeley. 387 pp.

Costas Lippmann, M. A. 1977. More on the weedy "pampas grass" in California. Fremontia 4(4): 25-27.

Costas Lippmann, M. A. & I. Baker. 1980. Isozyme variability in *Cortaderia selloana* and isozyme constancy in *C. jubata* (Poaceae). Madroño 27(4): 186, 187.

Cowan, B. 1976. The menace of pampas grass. Fremontia 4(2): 14-16.

Fuller, T. C. 1976. The menace of pampas grass: its history as a weed. Fremontia 4(2): 16.

Gorenflot, R. 1976. Le complexe polyploide du

Phragmites australis (Cav.) Trin. Ex Steud. (=P. communis Trin.). Bull. Soc. Bot. Du France 123: 261-271.

Hornback, B. 1994. In praise of pampas grass. Pacific Hort. 55(2): 47-52.

Madison, J. 1992. Pampas grasses: one a weed and one a garden queen. Pacific Hort. 51: 48-52.

Philipson, M. N. 1978. Apomixis in *Cortaderia jubata* (Gramineae). New Zealand J. Bot. 16: 45-59.

DANTHONIA

Baeza, P. C. M. 1996. Los géneros *Danthonia* D. C. y *Rytidosperma* Steud. (Poaceae) en América – una revisión. Sendtnera 3: 11-93.

Baum, B. R. and J. N. Findlay. 1973. Preliminary studies in the taxonomy of *Danthonia* in Canada. Canad. J. Bot. 51: 437-450.

Brock, R. D. & J. A. M. Brown. 1961. Cytotaxonomy of Australian *Danthonia*. Australian J. Bot. 8: 62-83.

Clay, K. 1983. Variation in the degree of cleistogamy within and among species of the grass *Danthonia*. Amer. J. Bot. 70(6): 835-843.

de Wet, J. M. J. 1954. The genus *Danthonia* in grass phylogeny. Amer. J. Bot. 41: 204-211.

Dobrenz, A. K. & A. A. Beetle. 1966. Cleistogenes in *Danthonia*. J. Range Manage. 19: 292-296.

Findlay, J. N. & B. R. Baum. 1974. The nomenclatural implications of the taxonomy of *Danthonia* in Canada. Canad. J. Bot. 52: 1573-1582.

RYTIDOSPERMA

Baeza, P. C. M. 1996. Los géneros *Danthonia* D. C. y *Rytidosperma* Steud. (Poaceae) en América – una revisión. Sendtnera 3: 11-93.

Connor, H.E. & E. Edgar. 1979. *Rytidosperma* Steudel (*Nothodanthonia* Zotov) in New Zealand. New Zealand J. Bot. 17: 311-337.

Linder, H. P. 1997. Nomenclatural corrections in the *Rytidosperma* complex (Danthonieae, Poaceae). Telopea 7: 269-274.

Linder, H. P. & G.A. Verboom. 1996. Generic limits in the *Rytidosperma* (Danthonieae, Poaceae) complex. Telopea 6(4): 597-627.

SCHISMUS

Conert, H. J. & A. M. Turpe. 1974. Revision der Gattung *Schismus* (Poaceae: Arundinoideae: Danthonieae). Abh. Senckenberg. Naturf. Gess. 532: 1-81.

Nicora, E. G. 1968. Observaciones sobre especies de gramineas del genero *Schismus*. Bol. Soc. Argent. Bot. 12: 312-315.

3.08 - SUBFAMILY ARISTIDOIDEAE

TECHNICAL DESCRIPTION

Habit: Annual or perennial herbs

Root Hairs:

Leaf Epidermis: 2-celled panicoid microhairs present; stomates with dome-shaped or triangular subsidiary cells; silica bodies of the festucoid, oryzoid, or panicoid type

Leaf in Cross-section: Mesophyll with radiate parenchyma

Inflorescence: An open to contracted panicle of few to many spikelets

Spikelets: 1-flowered, cylindric to laterally compressed; typically with a triple or trifid awn (the lateral branches sometimes reduced or absent)

Flowers: Lodicules 2 or 0; stamens 1-3; stigmas 2 (red or brown)

Embryo Formula:

Cytology: x = 11 or 12; diploids, tetraploids, and hexaploids

Photosynthetic Pathway: C₄

Distribution: Temperate, subtopical/tropical; often of drier sites; widespread

SYSTEMATICS

The genus *Aristida* has always been something of a problem child. Its 1-flowered spikelets led early workers to put it in Agrostideae. A host of microcharacters show that to be untenable. The Grass Phylogeny Working Group has segregated the genus into its own subfamily.

TRIBE: ARISTIDEAE

The characters of the tribe are those of the single genus that it contains.

ARISTIDA. Three-awn grass. Caespitose annuals or perennials. Culms herbaceous, the internodes hollow or solid, to 1 m tall. Inflorescence an open or contracted panicle. Spikelets 1-flowered, terete to laterally compressed, bisexual, disarticulating above the glumes. Glumes 2, thin and narrow, 1-nerved, as long as or longer than the floret; lemmas tough, terete, 3-nerved, with a sharp-pointed callus, tapering gradually to an awn column that usually bears 3 awns (the lateral ones reduced or obsolete in section *Streptachne*); palea relatively short to reduced, 2-nerved. Lodicules 2 or 0; stamens 1-3; stigmas 2, red or brown pigmented. A genus of almost 300 species native to the temperate and warmer regions of both hemispheres. A. oligantha (old field three-awn) is

found on open ground, mostly in the eastern half of the country; *A. purpurea* (purple three-awn) and *A. longiseta* (red three-awn) occur on plains and dry hills in the Southwest, especially after they have been disturbed.

SELECTED REFERENCES

TRIBE: ARISTIDEAE

Esen, A. & K. W. Hilu. 1991. Electrophoretic and immunological studies of prolamins in the Poaceae. II. Phylogenetic affinities of the Aristideae. Taxon 40: 5-17.

ARISTIDA

Allred, K. W. 1984. Morphologic variation and classification of the North American *Aristida purpurea* complex (Gramineae). Brittonia 36(4): 382-395.

Allred, K. W. 1986. Studies in the *Aristida* (Gramineae) of the southeastern United States. IV. Key and conspectus. Rhodora 88: 367-387.

Allred, K. W. 1992. The genus *Aristida* in California. Great Basin Nat. 52(1): 41-52.

Allred, K. W. & J. Valdes-Reyna. 1995. Novelties and notes in North American *Aristida* (Gramineae). Novon 5(3):

Clewell, A. F. 1989. Natural history of wiregrass (*Aristida stricta* Michx., Gramineae). Nat. Areas J. 9(4): 223-233.

Henrard, J. T. 1927. A critical revision of the genus *Aristida*. Meded. Rijks-Herb. 54: 1-220.

Hitchcock, A. S. 1924. The North American species of *Aristida*. Contr. U. S. Natl. Herb. 22(7): 517-586.

McKenzie, P. M. et al. 1989. Status of *Aristida* (Poaceae) in Puerto Rico and the Virgin Islands. Sida 13(4): 423-447.

Peet, R. K. 1993. A taxonomic study of *Aristida stricta* and *A. beyrichiana*. Rhodora 95: 25-37.

Peet, R. K. 1993. A taxonomic study of *Aristida stricta* and *A. beyrichiana*: addendum. Rhodora 95: 189, 190.

Reeder, J. R. & R. S. Felger. 1989. The *Aristida californica-glabrata* complex (Gramineae). Madroño 36(3): 187-197.

Trent, J. S. & K. W. Allred. 1990. A taxonomic comparison of *Aristida ternipes* and *Aristida hamulosa* (Gramineae). Sida 14(2): 251-261.

3.09 - SUBFAMILY CHLORIDOIDEAE

TECHNICAL DESCRIPTION

Habit: Herbaceous; culm internodes hollow or solid

Root Hairs: Equal

Leaf Epidermis: Complex; bicellular microhairs present; silica cells cross- or saddle-shaped; stomata with triangular or dome-shaped subsidiary cells

Leaf Anatomy: Vascular bundles with double parenchyma sheath, the outer often with conspicuously radiating cells

Inflorescence: Various, often 1-sided racemes or spikes

Spikelets: 1- to several-flowered; lemmas typically 3-nerved (1-nerved in *Sporobolus* and *Calamovilfa*; several-nerved in some grasses in minor tribes).

Flower: Lodicules 2; stamens 3; stigmas 2

Embryo: P + P F

Cytology: X = 9 or 10 (8 in *Erioneuron*); nucleoli

persistent

Photosynthetic Pathway: C₄

Distribution: Tropical and subtropical regions, especially of the Old World; particularly in arid and semiarid situations where there is high light intensity; best represented on this continent in the American Southwest.

SYSTEMATICS

The subfamily has also been called Eragrostoideae in the recent literature, a name that must be rejected for technical reasons. The group was not recognized by Hitchcock and Chase. It is a portion of their Festucoideae. Many of the grasses included here resided in their Festuceae and Chlorideae. The plants are essentially festucoid in spikelet structure and panicoid in many of their microcharacters.

Often there is general agreement as to what makes up the core of a subfamily; the disagreement seems to be mainly about the smaller "fringe groups." In the chloridoids, most everyone is satisfied that several smaller tribes belong here. The arguments focus on the core. Several workers recognize two large tribes, Eragrostideae and Cynodonteae (= Chlorideae of H & C); Watson & Dallwitz merge them into a unit they call the "Main Chloridoid Assemblage."

Here are the various tribes of the subfamily, as viewed by recent authors:

Clayton & Renvoize (1986): Pappophoreae, Orcuttieae, Eragrostideae, Cynodonteae

Watson & Dallwitz (1992): Pappophoreae,

Orcuttieae, "Main Assemblage"

"The Splitters:" Pappophoreae, Orcuttieae, Aeluropodieae, Unioleae, Eragrostideae, Sporoboleae, Spartineae, Cynodonteae, Zoysieae Hilu & Esen (1993): Pappophoreae, Orcuttieae, Eragrostideae

Grass Phylogeny Working Group (2001): Pappophoreae, Orcuttieae, Eragrostideae, Cynodonteae

Flora North America (2003): Pappophoreae, Orcuttieae, Eragrosideae

We will use the four tribe model, and also recognize a series of subtribes, each of which has also been viewed as tribes by various agrostologists.

TRIBE: ERAGROSTIDEAE

Annuals or perennials. Inflorescence typically a panicle. Spikelets typically few- to many-flowered, the lower florets fertile.

SUBTRIBE: MONANTHOCHLOINAE

Mostly grasses of seashores and saline marshy sites. Plants stoloniferous and rhizomatous, mostly dioecious. Inflorescence a condensed panicle or raceme. Leaf epidermis with papillae and sunken bicellular microhairs. These grasses have also been placed in their own tribe, Aeluropodeae.

DISTICHLIS. Salt grass. Low, rhizomatous perennials. Culms erect, rigid, the internodes solid, to 2 dm tall. Leaves often distichous. Inflorescence a reduced panicle or racemes. Spikelets several-flowered, laterally compressed, unisexual (species usually dioecious), disarticulating above the glumes and between the florets. Glumes 2, unequal, the first 1- to 5-nerved and the second 4- to 9-nerved, shorter than the florets, awnless; lemmas 9- to 11-nerved, awnless (those of the staminate spikelet thinner in texture); palea relatively long, 2-nerved, 2-keeled, these ± winged. Lodicules 2; stamens 3 or 0; stigmas 2 or 0.

A genus of 1 to a few species native to North America, with one in Australia. Beetle (1955) recognized a number of taxa. I have always found them difficult to distinguish. The treatment put forth by McVaugh (1983) seems reasonable to me. Following this more conservative view, *D. spicata* occurs along the Atlantic and Pacific coasts and in interior salt flats and marshes. *D. texana* in H & C has been transferred to the genus *Allolepis*.

MONANTHOCHLOË. Shore grass. Mat-forming rhizomatous, stoloniferous perennial. Culms herbaceous, decumbent, much-branched, to 2 dm tall. Leaves tufted, acicular, less than 1 cm long. Inflorescence reduced to a single spikelet ± concealed by upper leaf sheaths. Spikelets few-flowered, laterally compressed to rounded, unisexual (species dioecious), disarticulating below florets. Glumes 0; lemmas several-nerved, awnless; palea relatively

long, 2-keeled. Lodicules 0; stamens 0 or 3; stigmas 2 or 0.

A genus of three species, one in North America and two in South America. *M. littoralis* (shore grass) is found mostly in maritime coastal flats in southern California and in similar habitats in Texas, Louisiana, and Florida.

SWALLENIA. Eureka dune grass. Coarse, rhizomatous, perennial. Culms herbaceous to woody, to 1 m tall. Inflorescence a contracted panicle. Spikelets 3- to 7-flowered, laterally compressed, bisexual, persistent on the rachis. Glumes 2, ± equal, the lower 5- to 7-nerved and the upper 7- to 11-nerved; lemmas 5- to 7-nerved, awnless, densely hairy on lower margins; palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2. A monotypic genus, *S. alexandrae* is known only from sand dunes at two sites in Inyo Co., California. [= *Ectosperma a.* in H & C].

SUBTRIBE: ERAGROSTINAE

Annuals or perennials. Inflorescence a panicle, less often 1-sided spikes or racemes. Spikelets with several to many florets, 2 or more of them typically fertile. Glumes shorter than the florets. Lemmas 3-nerved (rarely 1-nerved), these usually conspicuous; awnless or with 1 long awn or 3 short ones. Plants of this subtribe were placed in Festuceae or Chlorideae by H & C.

ERAGROSTIS. Lovegrass, stink grass. Caespitose annuals or perennials [rarely stoloniferous]. Culms herbaceous, internodes hollow or solid, to 1 m tall. Inflorescence an open or contracted panicle. Spikelets, few- to many-flowered, the florets usually strongly overlapping, laterally compressed, bisexual [rarely unisexual and the species dioecious], disarticulating above the glumes and between the florets, the paleas persisting on the rachilla. Glumes 2, unequal, shorter than the florets, 1-nerved; lemmas 3-nerved, keeled or rounded, acute or acuminate, awnless; palea usually strongly 2-keeled, often ciliolate. Lodicules 2; stamens 1-3; stigmas 2.

A cosmopolitan genus of about 300 species, often of poorer, sandy sites. A number are weedy. *E. intermedia* (plains love grass) is a forage plant of some importance; *E. curvula* (weeping lovegrass) is an introduced forage grass in the southern states; *E. spectabilis* (purple lovegrass) occurs over much of the central and eastern sections of the United States; *E. cilianensis* (stink grass) is a widespread, malodorous weed. As treated here, the genus includes *Neeragrostis*, a small group of dioecious, mat-forming species.

TRIDENS. Purpletop, tridens. Erect, caespitose perennials. Culms herbaceous, to 1 m tall. Inflorescence an open or contracted panicle. Spikelets 3- to 12-flowered, laterally compressed, bisexual, disarticulating above the glumes and between the florets. Glumes 2, ± equal, the first 1-nerved and the second 1- to 3-nerved, shorter than the florets; lemmas broad, 3-nerved, typically hairy below, the apex bidentate, the midnerve usually extending between the teeth as a muro or short awn; palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2, often red pigmented.

A genus of 16-18 species native to the eastern and southern United States and to adjacent Mexico. In H

& C, the genus included a few species that are now assigned to *Erioneuron*. *T. flavus* (purpletop) is common in old fields and in open woods in the eastern half of the country; *T. albescens* (white tridens) of the southwest and south-central states is unusual in having glabrous lemmas; *T.* x oklahomensis is an endemic known only from a wet meadow near Stillwater, OK.

LEPTOCHLOA. Sprangletop. Caespitose annuals or perennials, often of marshy or wet sites. Culms herbaceous, to 1+ m tall. Inflorescence variously described as a panicle of racemose, unbranched branches or a compound, race-mose inflorescence whose branches bear spikes or spike-like racemes. Spikelets 2- to several-flowered, often overlapping, rounded to laterally compressed, bisexual, disarticulation above the glumes and between the florets. Glumes 2, equal of unequal, 1-nerved, shorter than the florets; lemmas 3-nerved, often minutely pubescent on the nerves, awnless, mucronate, or awned; palea shorter than lemma, 2-nerved. Lodicules 2; stamens 2 or 3; stigmas 2.

A genus of about 70 species, native to the warmer regions of both hemispheres. As treated here, the genus includes *Diplachne*. See the table below for a comparison. *L. filiformis* (red sprangletop) is a common weed in the Southwest and in the Southeast. *L. dubia* (green sprangletop) is found on dry sites in the Southwest.

DIPLACHNE VERSUS LEPTOCHLOA

Feature	Diplachne	Leptochloa
Spikelets	4-12 mm long	1-4 mm long
Florets	5-12	2-5 (7)
Caryopses	elliptical in x-s not grooved gr	triangular in x-s rooved on one side
Lemma	rounded lateral nerves extending to upper margins	strongly keeled lateral nerves r not extending to upper margins
Inflorescend	ce racemose 2.5-6 mm wide	spicate 0.5-4 mm wide

REDFIELDIA. Blowout grass. Rhizomatous perennial to 1 m. Leaf blade involute, with filiform tip. Inflorescence a large, open panicle, 1/3 to ½ length of culms, its branches flexuous. Spikelets [1-] 2- to 6-flowered, dis-articulating above glumes. Glumes 2, acuminate, 1-nerved; lemmas keeled, hairy on margins at base, 3-nerved; callus bearded. Lodicules 2; stamens 3; stigmas 2. One species, *R. flexuosa*, native to the interior sandy hills of the U. S., from SD and OK to CO, AZ.

SUBTRIBE: ELEUSININAE

ELEUSINE. Goose grass. Low, spreading, annuals. Culms herbaceous, weak, flattened, to 2 dm tall. Inflorescence a series of 2 to several \pm digitate branches clumped at the culm apex. Spikelets sessile in 2 rows, 3- to several-flowered, laterally compressed, bisexual, disarticulation above the

glumes and between the florets (except in cultivars). Glumes 2, unequal, the first 1-nerved and the second 3- to 5-nerved, shorter than the florets; lemmas 3-nerved, acute, awnless to mucronate; palea shorter than lemma, apically notched. Lodicules 2; stamens 3; stigmas 2.

A genus of 6-9 species, all but one native to the Old World. *E. indica* (goose grass) is a common weed over much of the United States; *E. coracana* (finger millet, ragi) is an important grain crop in the Old World.

DACTYLOCTENIUM. Durban grass, crowfoot grass. Annual or perennial herbs, the culms often spreading and rooting at the nodes. Inflorescence a series of paired or digitate racemes; spikelets imbricate in two rows, inserted at right angles to rachis. Spikelets 2- to several-flowered, laterally compressed, disarticulating between the first and second glume. Glumes 2, 1-nerved, the upper with an oblique awn from just below its tip; lemmas 3-nerved, strongly keeled, acute or abruptly narrowed to a short recurved awn. Lodicules 2; stamens 3; stigmas 2.

A genus of about 13 species native to the Old World, especially of dry, sandy sites. *Dactyloctenium aegyptium* is a cosmopolitan weed and is also planted for lawns and playing fields. It occurs in North America from NC to FL and the Pacific coast.

SUBTRIBE: MUHLENBERGIINAE

Annuals or perennials with well-developed panicles of small, 1-flowered spikelets. Disarticulation above the glumes. Lemmas 1- or 3-nerved. Plants of this subtribe were placed in Agrostideae by H & C. The group is often recognized as a separate tribe, Sporoboleae.

SPOROBOLUS. Dropseed. Caespitose annuals or perennials, a few rhizomatous. Culms herbaceous, internodes usually solid, to 2 m tall. Ligules ciliate (a useful feature in distinguishing the genus from *Muhlenbergia*, with which it is easily confused). Inflorescence an open or contracted panicle. Spikelets small, 1- flowered, rounded to laterally compressed, bisexual, disarticulating above the glumes. Glumes 2, unequal, 1-nerved, shorter than the floret; lemma 1-nerved, awnless; palea relatively long, sometimes splitting at maturity and thereby resembling an extra lemma; pericarp free from the seed (hence the common name). Lodicules 0 or 2; stamens 2 or 3; stigmas 2.

A genus of almost 200 species native to diverse habitats of the Old and New Worlds. *S. cryptandrus* (sand dropseed) occurs on sandy soils over much of the country; *S. heterolepis* (prairie dropseed) is found on prairies through the central portion of the United States; *S. airoides* (alkali sacaton) occurs in alkaline meadows and in valleys of the West; *S. junceus* (smut grass) [= *S. poiretii* in H & C and *S. indicus* in more recent literature] is a tropical-subtropical introduction in the Southeast; *S. virginicus* (seashore dropseed) is native to the sandy shores of the Texas Gulf coast.

MUHLENBERGIA. Muhly, nimblewill. Delicate, caespitose annuals to coarse, rhizomatous, stoloniferous perennials. Culms herbaceous, internodes solid or hollow, to 2 m tall. Ligule membranous. Inflorescence an open to contracted panicle [rarely spike-like]. Spikelets small, 1-flowered

[rarely 2-flowered and rarely keying properly!], laterally compressed, bisexual, disarticulating above the glumes. Glumes 2, equal or unequal, [0-] 1- [3-] nerved, awnless to short-awned; lemmas 3-nerved, typically with a single well-developed awn, occasionally mucronate or awn-less; palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2.

A large genus of 125-160 species native to diverse habitats of both hemispheres, especially the New World. *M. schreberi* (nimblewill) occurs in damp, shady places in the eastern half of the country; *M. emersleyi* (bull grass) is found in rocky woods and ravines of the Southwest; *M. sylvatica* (forest muhly) is a rhizomatous grass of wooded areas in the eastern and central U. S.; *M. rigens* (deer grass) of southern California was used by Native Americans in basket making.

CALAMOVILFA. Sand reedgrass. Coarse, rhizomatous perennials. Culms herbaceous, the internodes solid, to 2 m tall. Inflo-rescence an open or contracted panicle. Spikelets 1-flowered, laterally compressed, bisexual, disarticulating above the glumes. Glumes 2, unequal, 1-nerved, as long as the floret, lemmas 1-nerved, awnless, the callus bearded; palea reduced, 2-nerved. Lodicules 2; stamens 3; stigmas 2; pericarp free from the seedcoat, as in *Sporobolus*.

This small genus of 4 species is a favorite of the conservative right because all of them are native to North America. *C. longifolia* and *C. gigantea* are found in sandy habitats in the central and southwestern regions of the country.

SUBTRIBE: MUNROINAE

ERIONEURON. Fluff grass. Low, tufted, stoloniferous perennials. Culms herbaceous, to 2 dm tall. Leaf blades with cartilaginous margins. Inflorescence a compact, head-like to more open panicle. Spikelets several-flowered spikelets, laterally compressed, bisexual, disarticulating above the glumes and between the florets. Glumes 2, ± equal, 1-nerved; lemmas broad, 3-nerved, with long hairs (at least below), apex bilobed, midnerve extended as a short awn, the lateral ones as a short mucro; palea about as long as lemma, ciliate on the keels, long-hairy below. Lodicules 2; stamens 1-3; stigmas 2, white.

A genus of 5 species native to drier, often rocky sites of the American Southwest and adjacent Mexico. These taxa were included in *Tridens* by H & C. As treated here, the genus includes *Dasyochloa*. *Erioneuron pulchellum* (fluff grass) occurs on mesas, sandy washes, and rocky hills in the Southwest; *E. pilosum* (hairy tridens) is found on plains of the South Central and Southwest regions.

ERIONEURON VERSUS TRIDENS

Feature	Erioneuron	Tridens
Embryo	oval; translucent	reniform; dark brown
Stigmas	+/- white	dark purple
Lemmas	2- (3-) lobed hairy near midrib/margins	bidentate; not lobed less pubescent
Palea long-hairy below	usually glabrous; ciliate on keel	never ciliate
Habit	low, stoloniferous	tall perennials
Leaf	white margins cartilaginous	green margins not cartilaginous
x =	8	10

CRYPSIS. Prickle grass, swamp timothy. Prostrate to ascending annuals. Internodes hollow or solid. Inflores-cences terminal or axillary, ovoid to capitate, spike-like panicles, often ± enclosed by bract-like sheaths. Spikelets 1-flowered, keeled, strongly laterally compressed, disarticulating above or below the glumes. Glumes 2, acute or short-awned, 0 or 1-nerved; lemma 1-nerved; palea 1- or 2-nerved. Lodicules 0; stamens 2 or 3; stigmas 2, white. Fruit an achene, the seed free from the pericarp, as in *Sporobolus*.

A genus of 8 species, mostly native to the Middle East and Mediterranean; often found on saline soils. Three species occur in the U. S., all of them introduced.

SUBTRIBE: UNIOLINAE

UNIOLA. Sea-oats. Rhizomatous or stoloniferous perennials. Culms to 2 m tall. Inflorescence a few- to many-flowered panicle. Spikelets several-flowered (the proximal and distal florets sterile), bisexual, laterally compressed, disarticulating below the glumes. Glumes 2, shorter than the florets, 3-nerved, awnless; lemmas 3- to 10-nerved, awnless or mucronate, serrulate-keeled; palea 2-keeled, winged, serrate to ciliate. Lodicules 2; stamens 3; stigmas 2.

A small genus of 2 species native to North and South America and the Caribbean. *U. paniculata* is found on coastal sand dunes of Alabama, Florida, and the Gulf Coast. Its large, drooping panicles are popular in dried arrangements, where they are often sprayed some hideous color. The other species, *U. pittieri* is found on the beaches from Mexico to northern South America. As treated by H & C, *Uniola* included several other species now transferred to *Chasmanthium* of the Centostecoideae.

TRIBE: CYNODONTEAE

Annuals or perennials. Inflorescence typically a compound spike or raceme, the spikelets often in two rows on one side of the branch. Spikelets 1-flowered or few- to several-flowered, but only one typically fertile. Unisexual spikelets occur in some genera. Disarticulation above the glumes. Lemmas 3-nerved.

SUBTRIBE: CHLORIDINAE

CYNODON. Bermuda grass. Low, mat-forming, stolon-iferous and/or rhizomatous perennials. Culms herbaceous, to 4 dm tall. Inflorescence a series of 2 to several digitate branches, spikelets sessile, in 2 rows on a ± triangular rachis. Spikelets 1-flowered, rachilla extended beyond floret and sometimes bearing a rudiment, laterally compressed, bisexual, disarticulating above the glumes. Glumes 2, ± equal, 1-nerved, the second about as long as the floret; lemmas 3-nerved, hairy on keel and lateral nerves, awnless; palea as long as lemma, 2-nerved. Lodicules 2; stamens 3; stigmas 2, red pigmented.

A genus of 10 species native to the Old World. *C. dactylon* (Bermuda grass) is an important pasture grass, lawn grass, and aggressive weed over most of the warmer parts of North America.

CHLORIS. Windmill grass, finger grass. Caespitose annuals or perennials from rhizomes or stolons. Culms herbaceous, internodes hollow or solid. Leaf sheaths keeled. Inflorescence a series of racemose or digitate branches, each bearing 2 rows of sessile spikelets. Spikelets with 1 fertile floret and 1 or more rudimentary ones above it, laterally compressed, disarticulating above the glumes. Glumes 2, equal or unequal, the lower 1-nerved and the upper 1- to 4-nerved; lemmas [1-] 3- [5-] nerved; palea relatively long, strongly 2-nerved. Lodicules 2; stamens 3; stigmas 2.

A genus of about 70 species native to the warmer regions of the Old and New Worlds. As treated here, the genus includes the 2 *Trichloris* species in H & C, but does not include *Chloris* section *Eustachys* in H & C, which is now generally recognized as a distinct genus. *C. gayana* (Rhodes grass) has escaped from cultivation in the southern states; *C. virgata* (feather finger grass) is a common weed; *C. verticillata* (windmill grass) is native to the plains states; *C. cucullata* (hooded windmill grass) is native to the Southwest.

GYMNOPOGON. Skeleton grass. Perennials, often rhizomatous. Culms herbaceous, to 1 m tall. Leaf blades stiff, distichous. Inflorescence a series of racemose branches, the spikelets in two rows.

Spikelets 1- to 3-flowered, laterally compressed, bisexual, the rachilla extending beyond the uppermost fertile floret as slender stalk bearing a rudimentary floret, disarticulation above the glumes. Glumes 2, \pm equal, 1-nerved, the second longer than the florets; lemmas 3-nerved, usually awned; palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2.

A genus of about 15 species, all but one of them native to the New World. There are four species in North America; *G. ambiguus* is the most commonly encountered species, growing from the Pine Barrens of New Jersey south to Florida and across to Texas and Oklahoma; *G. brevifolius* is native to the southeastern Coastal Plain; two species are endemic to Florida.

SPARTINA. Cord grass, marsh grass. Caespitose, stoloniferous/rhizomatous perennials. Culms herbaceous, internodes solid or hollow, to 2 m tall. Inflorescence a series of few to many short, often appressed, racemose branches. Spikelets 1-flowered, conspicuously laterally compressed, imbricate on one side of rachis, bisexual, disarticulation below the glumes. Glumes 2, unequal, the first 1-nerved and the second 1- to 3-nerved, the second as long or longer than the floret, awned or awnless; lemma 1- or 3-nerved, keeled, awnless; palea relatively long, 2-nerved, with membranous margins. Lodicules 0; stamens 3; stigmas 2.

A genus of 16 species, one native to Europe and the remainder to the New World. *S. pectinata* (prairie cord grass) is common in wet areas over much of the country; *S. foliosa* occurs in salt marshes along the California coast; *S. densiflora*, native to South America, occurs around Humboldt Bay, California; *S. alterniflora* (smooth cord grass) is found in saline marshes along the Atlantic and Gulf coasts.

ORIGIN OF SPARTINA X TOWNSENDII

Huskins (1931)

S. stricta (2n = 56) x S. alterniflora (2n = 70) ∇

S. townsendii (2n = 126)*

Marchant (1966)

 ∇ ∇

S. X townsendii

AAB₁B₂BC

Sterile F1 hybrid

S. townsendii of 1881 description

2n = 62

10 long chromosomes

Rules out autotetraploidy

Back crosses to S. a.

 ∇ ∇

Doubling of Chromosomes

 ∇ ∇

S. x townsendii AAAABBB₁B₁B₂B₂CC 2n = 120, 122, 124 Pollen good Seed set good 20 long chromosomes

* "S. townsendii has evidently originated by chromosome doubling, following on interspecific hybridisation. It is an extremely successful new species, having spread widely from its point of origin, and has almost completely eliminated its parent species whereever it has come into competion with them. It seems to be an outstanding example of the significance of allopolyploidy in plant evolution." (C. L. Huskins, 1931)

SUBTRIBE: BOUTELOUINAE

BOUTELOUA. Grama grass. Annuals or caespitose, rhizomatous, or stoloniferous perennials. Culms herbaceous, internodes hollow or solid, to 1 m tall. Inflorescence of 1 to many short, spicate branches that are racemose along a common axis, each bearing sessile spikelets in 2 rows along a flattened or angular rachis. Spikelets with 1 fertile floret and 1-3 rudimentary ones above it, laterally compressed, disarticulation either above the glumes (subgenus Chondrosum) or at the base of a branch, the subunit of the inflorescence falling at maturity (subgenus Bouteloua). Glumes 2, equal or unequal, 1-nerved, awned or awnless; lemmas 3-nerved, the midnerve often extended as an awn, the lateral ones sometimes also awn-tipped; palea sometimes 2-awned. Lodicules 2; stamens 3; stigmas 2.

A genus of about 40-50 species native to the New World from Canada through South America; the American Southwest is a major center. *B. curtipendula* (sideoats grama) is a valuable forage grass; *B. hirsuta* (hairy grama) occurs on plains and in rocky places in the central United States; *B. gracilis* (blue grama) is found in the plains of the central and western states.

BUCHLOË. Buffalo grass. Low, stoloniferous, matforming perennials. Culms herbaceous, internodes solid, to 2 dm tall. Unisexual spikelets in separate inflorescences, typically on different plants, less frequently on the same plant; staminate inflorescence a series of 1-4 spicate branches, each bearing 2 rows of sessile, secund spikelets, well-elevated above the vegetative portion of the plants; pistillate inflorescence a bur-like head, partially hidden in ± inflated upper leaf sheaths, the outer covering of the bur a combination of thickened rachis and glumes. Staminate spikelets 2-flowered; pistillate spikelet 1-flowered. Glumes 2, the second indurate, with 3 rigid lobes, and enveloping the florets in the pistillate spikelet; lemmas 3-nerved and typically awnless; palea ± equal to lemma, 2-nerved. Lodicules 2 or 0; stamens 3 or 0; stigmas 2 or 0.

A monotypic genus native to open, drier plains of North America. *B. dactyloides*, is a very important range grass of the short-grass prairie, where it can be a dominant.

HILARIA. Galleta, tobosa, curly-mesquite. Rhizomatous, stoloniferous, or caespitose perennials. Culms herbaceous, stiff, the internodes solid, to 1 m tall. Inflorescence a balanced spike of sessile spikelets inserted in trios within ciliate, cup-like depressions along a wavy or zig-zag rachis; disarticulation below the glumes, the trio of spikelets falling as a group. Spikelets of the trio dissimilar, the lateral ones 2- to 4-flowered, staminate; the central one 1-flowered and perfect. Glumes 2, firm, united to form a false involucre, awned on one side from about the middle; lemmas 3-nerved, awned or awnless; palea relatively long, 2-nerved. Lodicules 2 or 0; stamens 3; stigmas 2.

A genus of about 10 species native to the New World; five of them are found in North America; *H. belangeri* (curly-mesquite) occurs in the arid and semiarid sites in the Southwest; *H. swallenii* is endemic to western Texas and adjacent Mexico. *H. jamesii* (galleta) is native to dry plains and deserts in the West and Southwest; *H. rigida* (big galleta) is a coarse, ± woody desert grass with a felt-like covering on its culms; *H. mutica* (tobosa) is found on drier sites in the Southwest.

SUBTRIBE: ZOYSINAE

This is small tribe of introduced grasses. The inflorescence is a contracted raceme of 1-flowered spikelets on short pedicels. Disarticulation is below the glumes. The palea is often reduced or absent. X=10.

ZOYSIA. Zoysia or zoisia. Rhizomatous/stoloniferous perennials. Inflorescence a spike, the spikelets solitary rachis. Spikelets 1-flowered; а zig-zag disarticulation below the glume. Glume 1 absent; second glume 1-nerved, mucronate or short-awned. Lemma 1-nerved, shorter than second glume and enclosed by it. Palea present or absent. Lodicules 0; stamens 2 or 3; stigmas 2. A genus of about 10 species, native to Southeast Asia and New Zealand. Three species have been introduced into the warmer, southern portions of the United States where they are grown as popular lawn grasses.

TRAGUS. Weak-stemmed annuals. Inflorescence a spike-like raceme, the spikelets in bur-like clusters of 2 to 5. Spikelets 1-flowered; disarticulation at the base of a spikelet cluster. Glumes 2, the second one in some spikelets bearing stout, hooked spines. Lemmas 3-nerved, awnless. Lodicules 2; stamens 3; stigmas 2. A genus of 7 species, 6 of them native to Africa. *T. racemosus* and *T. berteronianus* are weedy introductions in the eastern and southwestern portions of the country.

TRIBE: PAPPOPHOREAE

This small tribe of warm, dry climate grasses was treated as part of Festuceae by H & C. The inflorescence is a panicle of 3- to several-flowered spikelets. The lower florets are perfect, the upper ones staminate or neuter. Disarticulation above the glumes, the florets separating as a group. Lemmas with 9 or more nerves, the apex divided into 3-many sharp lobes or awns. X = 10. See Section 4 for a key to our North American taxa.

PAPPOPHORUM. Pappus grass. Caespitose perennials. Inflorescence a contracted to spike-like panicle. Spikelets 3- to 6-flowered; lower 1 to 3 fertile. Glumes thin, ± equal, 1-nerved, awnless. Lemmas leathery, many-nerved, these extending into unequal awns. Palea about as long as lemma. Lodicules 2; stamens 3; stigmas 2. A genus of 8 species, native to the American Southwest and South America. Pappophorum bicolor (pink pappus grass) and P. mucronulatum (whiplash pappus grass) are native to North America.

ENNEAPOGON. Spike pappus grass. Tufted perennials. Inflorescence a spike-like panicle. Spikelets several-flowered; disarticulation above the glumes. Glumes ± equal, 5- to many-nerved, awnless. Lemmas much shorter than glumes, firm, 9-nerved, these extending into plumose awns. Lodicules 2; stamens 3; stigmas 2. A genus of about 30 species, mostly of Old World xerophytic sites. *E. desvauxii* (spike pappus grass) is our only New World species. It is found in the Southwest and extends into Mexico.

COTTEA. Cottea grass, pelucilla. Tufted perennial. Leaves often pilose. Inflorescence a ± open panicle. Spikes 6- to 10-flowered; disarticulation above the glumes and between the florets. Glumes ± equal, 7-to many-nerved, awned or awnless. Lemmas 9- to 13-nerved, these extending into unequal awns. Palea slightly longer than lemma. Lodicules 2; stamens 3; stigmas 2. A monotypic genus. *C. pappophoroides* is native to Arizona, New Mexico, and Texas. It also occurs in Central and South America.

TRIBE: ORCUTTIEAE

This small tribe of California endemics was treated as part of Festuceae by H & C. A peculiar feature of these grasses is that the leaf blade and sheath are not clearly differentiated from one another. The inflorescence is a spike, spike-like raceme, or a panicle. Grasses of this tribe are restricted to mud flats and vernal pools in California and Baja California. All of them are classed as rare and/or endangered. See Section 4 for a key to our North American taxa.

NEOSTAPFIA. Colusa grass. Tufted, spreading, aromatic annual. Culm internodes solid. Leaves glandular-viscid, not clearly differentiated into blade and sheath. Inflorescence a dense, cylindric spike-like raceme, often partially enclosed in dilated upper leaf sheaths. Spikelets several-flowered; disarticulation above the glumes. Glumes 0. Lemmas fan-shaped, prominently 7- to 11-veined, awnless. Palea about as long as lemma. Lodicules 2; stamens 3; stigmas 2. A monotypic genus. *N. colusana* is endemic to vernal pool margins in four counties in California.

ORCUTTIA. Orcutt grass. Tufted, semiaquatic annuals, erect to prostrate. Leaf blade and sheath not clearly differentiated. Inflorescence a spike-like raceme. Spikelets few- to many-flowered; disarticulation above the glumes and between the florets. Glumes 2, irregularly toothed. Lemmas prominently 5-toothed, awnless. Palea about as long as lemma. Lodicules 0; stamens 3; stigmas 2. A genus of 5 species, endemic to vernal pools in California and Baja California. All taxa are rare and endangered.

TUCTORIA. Tufted annuals. Stems ascending to erect, fragile at maturity. Leaf blade and sheath not differentiated. Inflorescence spike-like, often partially

enveloped by upper leaves. Spikelets 5- to manyflowered, spirally inserted; disarticulation above the glumes and between the florets. Glumes 2, \pm equal, awnless. Lemmas 11- to 17-veined, the apex entire to minutely toothed. Palea about as long as the lemma. Lodicules 2; stamens 3; stigmas 2. A genus of 3 species, all of them endemic to vernal pools and grasslands in California and Baja California. *T. greenei* and *T. mucronata* are native to a few counties in California.

SELECTED REFERENCES

Brown, W. V. 1960. A cytological difference between the Eupanicoideae and the Chloridoideae. Southwest. Nat. 5: 7-11.

Clayton, W. D. 1982. Notes on subfamily Chloridoideae (Gramineae). Kew Bull. 37: 417-420.

Hartley, W. & C. Slater. 1960. Studies on the origin, evolution, and distribution of the Gramineae. III. The tribes of the subfamily Eragrostoideae. Australian J. Bot. 8: 256-276.

Hilu, K. W. & A. Esen. 1993. Promalin and immunological studies in the Poaceae. III. Subfamily Chloridoideae. American J. Bot. 80(1): 104-113.

Hilu, K. W. & L. A. Alice. 2000. Phylogenetic relationships in subfamily Chloridoideae (Poaceae) based on *mat*K sequences: a preliminary assessment. In, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 173-179.

Hilu, K. W. & L. A. Alice. 2001. A phylogeny of Chloridoideae (Poaceae) based on *mat*K sequences. Syst. Bot. 26(2): 386-405.

Peterson, P. M. et al. 2001. Catalogue of New World grasses (Poaceae): II. Subfamily Chloridoideae. Contr. U. S. Natl. Herb. 41: 1-255.

Phillips, S. M. 1982. A numerical analysis of the Eragrostoideae. Kew Bull. 37(1): 133-162.

Sutton, D. D. 1975. Leaf anatomy in the subfamily Eragrostoideae. Michigan Acad. 5: 373-383.

Van den Borre, A. & L. Watson. 1997. On the classifica-tion of the Chloridoideae (Poaceae). Australian Syst. Bot. 10: 491-531.

Van den Borre, A. & L. Watson. 2000. On the classification of the Chloridoideae: results from morphological and leaf anatomical data analyses. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 180-183.

TRIBE: ERAGROSTIDAE

Peterson, P. M. 2000. Systematics of the Muhlenbergiinae (Chloridoideae: Eragrostidae). <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 195-212.

Peterson, P. M., R. D. Webster, & J. Valdés-Reyna. 1995. Subtribal classification of the New World Eragrostideae (Poaceae: Chloridoideae). Sida 16(3): 529-544.

ALLOLEPIS

Soderstrom, T. & H. F. Decker. 1965. *Allolepis*: a new segregate of *Distichlis*. Madroño 18: 33-39.

BLEPHARIDACHNE

Hunziker, A. T. & A. M. Anton. 1979. A synoptical revision of *Blepharidachne* (Poaceae). Brittonia 31(4): 446-453.

BLEPHARONEURON

Reeder, J. R. 1970. Cytotaxonomy of *Blepharoneuron*. J. Colorado-Wyoming Acad. Sci. 7(1): 37.

CALAMOVILFA

Reeder, J. R. & M. A. Ellington. 1960. *Calamovilfa*: a misplaced genus of Gramineae. Brittonia 12: 71-77.

Thieret, J. W. 1966. Synopsis of the genus *Calamovilfa* (Gramineae). Castanea 31: 145-152.

CRYPSIS

Hammel, B. E. & J. R. Reeder. 1979. The genus *Crypsis* in the United States. Syst. Bot. 4(4): 267-280.

Lorch, J. 1962. A revision of *Crypsis* Ait. (Gramineae). Bull. Res. Council Israel, Sect. D., Bot. 11: 91-116.

DINEBRA

Phillips, S. M. 1973. The genus *Dinebra* Jacq. (Gramineae). Kew Bull. 28(3): 411-418.

DISTICHLIS

Beetle, A. A. 1943. The North American variations of *Distichlis spicata*. Bull. Torrey Club 70: 638-650.

Beetle, A. A. 1955. The grass genus *Distichlis*. Revista Arg. Agron. 22: 86-94.

Fassett, N. C. 1925. Notes on *Distichlis*. Rhodora 27: 67-72.

McVaugh, R. 1983. Gramineae. Flora Novo-Galiciana. Vol. 14. Univ. Michigan Press. Ann Arbor. Pp. 147, 148.

Stephenson, S. N. 1971. A putative *Distichlis* x *Monanthochloe* (Poaceae) hybrid from Baja California, Mexico. Madroño 21: 125-127.

ELEUSINE

de Wet, J. M. J. et al. 1984. Systematics and evolution of *Eleusine coracana* (Gramineae). American J. Bot. 71(4): 550-557.

Hilu, K. W. 1980. Noteworthy collections: *Eleusine tristachya*. Madroño 27: 177, 178.

Hilu, K. W. & J. L. Johnson. 1992. Ribosomal DNA variation in finger millet and wild species of *Eleusine* (Poaceae). Theoret. Appl. Genet. 83: 895-902.

Hilu, K. W. & J. L. Johnson. 1997. Systematics of

Eleusine Gaertn. (Poaceae: Chloridoideae): chloroplast DNA and total evidence. Ann. Missouri Bot. Gard. 84(4): 841-847.

Hilu, K. W. et al. 1978. Flavonoid patterns and systematics in *Eleusine*. Biochem. Syst. Ecol. 6: 247-249.

Mehra, K. L. 1963. Differentiation of the cultivated and wild *Eleusine* species. Phyton 20: 189-198.

Phillips, S. M. 1972. A survey of the genus *Eleusine* Gaertn. (Gramineae) in Africa. Kew Bull. 27: 251-270.

Rachie, K. O. & L. V. Peters. 1977. The eleusines. A review of the world literature. Int. Crops Res. Inst. for Semiarid Tropics. 129 pp.

ERAGROSTIS

Borre, A. van den & L. Watson. 1994. The infrageneric classification of *Eragrostis* (Poaceae). Taxon 43(3): 383-422.

Bush, B. F. 1903. A new genus of grass. Trans. Acad. Sci. St. Louis. 13: 175-183. [Neeragrostis]

Harvey, L. H. 1948. *Eragrostis* in North and Middle America. Ph. D. dissertation. Univ. Michigan. Ann Arbor. 269 pp.

Harvey, L. H. 1954. New entities in North and Middle America *Eragrostis* (Gramineae). Bull. Torrey Bot. Club 81(5): 405-410.

Koch, S. D. 1969. The *Eragrostis pectinacea-pilosa* complex in North and Central America. Ph. D. dissertation. Univ. Michigan. Ann Arbor. 203 pp.

Koch, S. D. 1974. The *Eragrostis pectinacea-pilosa* complex in North and Central America (Gramineae: Eragrostoideae). Illinois Biol. Monographs 48: 1-74.

Koch, S. D. 1978. Notes on the genus *Eragrostis* (Gramineae) in the southestern United States. Rhodora 80: 390-403.

Koch, S. D. & I. S. Vega. 1985. *Eragrostis mexicana, E. neo-mexicana, E. orcuttiana,* and *E. virescens*: the resolution of a taxonomic problem. Phytologia 58(6): 377-381.

Nicora, E. G. 1962. Revalidacion del genera de gramineas "Neeragrostis" de la flora Norteamericana. Rev. Argent. Agron. 29(1-2): 1-11.

Reeder, J. R. 1986. Another look at *Eragrostis tephrosanthos* (Gramineae). Phytologia 60(2): 153, 154.

Reeder, J. R. 1986. Mistaken identity in annual *Eragrostis* (Gramineae). Phytologia 60(2): 95-97.

Renvoize, S. A. 1983. A survey of leaf-blade anatomy in grasses. IV. Eragrostoideae. Kew Bull. 38: 469-478.

Sanchez Vega, I. & S. D. Koch. 1988. Estudio biosystematico de *Eragrostis mexicana, E. neomexicana, E. orcuttiana* y *E. virescens* (Gramineae: Chloridoideae). Bol. Soc. Bot. Mexico 48: 95-112.

Valdes-R., J. & S. L. Hatch. 1991. Lemma

microphotography in the Eragrostoideae (Poaceae). Sida 14: 531-549.

Witherspoon, J. T. 1975. A numerical taxonomic study of the *Eragrostis intermedia* complex (Poaceae). Ph. D. dissertation. Univ. Montana. 500 pp.

Witherspoon, J. T. 1977. New taxa and combinations in *Eragrostis* (Poaceae). Ann. Missouri Bot. Gard. 64: 324-329.

ERIONEURON

Caro, J. A. 1981. Rehabilitacion del genero *Dasyochloa* (Gramineae). Dominquezia 2: 1-17.

Reeder, J. R. & D. J. Crawford. 1970. Affinities of *Erioneuron* and *Munroa*. American J. Bot. 57: 752 (abstract).

Sanchez, E. 1983. *Dasyochloa* Willd. ex Rydberg (Poaceae). Genero monotipico de Norteamerica. Lilloa 36: 131-138.

Valdes-Reyna, J. 1985. A biosystematic study of the genus *Erioneuron* (Poaceae: Eragrostoideae). Ph. D. dissertation. Texas A & M Univ. College Station. 194 pp.

Valdes-Reyna, J. & S. L. Hatch. 1995. Anatomical study of *Erioneuron* and *Dasyochloa* (Poaceae: Chloridoideae: Eragrostideae) in North America. Sida 16(3): 413-426.

Valdes-Reyna, J. & S. L. Hatch. 1997. A revision of *Erioneuron* and *Dasyochloa* (Poaceae: Eragrostideae). Sida 17(4): 645-666.

LEPTOCHLOA

Hitchcock, A. S. 1903. The North American species of *Leptochloa*. U. S. D. A. Bureau Plant Indust. Bull. 33: 1-24.

McNeill, J. 1979. *Diplachne* and *Leptochloa* (Poaceae) in North America. Brittonia 31(3): 399-404.

Snow, N. 1996. The phylogenetic utility of lemmatal micro-morphology in *Leptochloa* s. I. and related genera in subtribe Eleusininae (Poaceae, Chloridoideae, Eragrostideae). Ann. Missouri Bot. Gard. 83(4): 504-529.

Snow, N. 1997. Phylogeny and systematics of *Leptochloa* P. Beauv. sensu lato (Poaceae: Chloridoideae). Ph. D. dissertation. Washington Univ. St. Louis, MO.

Snow, N. 1998. Nomenclatural changes in *Leptochloa* P. Beauvois sensu lato (Poaceae, Chloridoideae). Novon 8: 77-80.

Snow, N. 1998. Caryopsis morphology of *Leptochloa* sensu lato (Poaceae: Chloridoideae). Sida 18(1): 271-282.

Valls, J. F. M. 1978. A biosystematic study of *Leptochloa* with special emphasis on *Leptochloa dubia* (Gramineae: Chloridoideae). Ph. D. dissertation. Texas A & M Univ. College Station. 216 pp.

LYCURUS

Peterson, P. M. & O. Morrone. 1997. Allelic variation in

the amphitropical disjunct *Lycurus setosus* (Poaceae: Muhlenbergiinae). Madroño 44(4): 334-346.

Reeder, C. G. 1985. The genus *Lycurus* (Gramineae) in North America. Phytologia 57(4): 283-291.

Sanchez, E. & Z. Rugolo de Agrasar. 1986. Estudio taxonomico sobre el genero *Lycurus* (Gramineae). Parodiana 4: 267-310.

MICROCHLOA

Launert, E. 1966. A taxonomic revision of the genus *Microchloa* R. Brown (Gramineae, Chlorideae, Chloridinae. Senckenberg Biol. 47(4): 291-301.

MONANTHOCHLOË

Stephenson, S. N. 1971. A putative *Distichlis* x *Monanthochloe* (Poaceae) hybrid from Baja California, Mexico. Madroño 21: 125-127.

Villamil, C. B. 1969. The genus *Monanthochloë* (Gramineae). Taxonomical and morphological studies with special reference to the Argentine species. Kurtziana 5: 369-391.

MUHLENBERGIA

Anderson, M. Kat. 1996. The ethnobotany of deergrass, *Muhlenbergia rigens* (Poaceae): its uses and fire management by California Indian tribes. Econ. Bot. 50(4): 409-422.

Duvall, M. R. et al. 1994. Alliances of *Muhlenbergia* (Poaceae) within New World Eragostoideae are identified by phylogenetic analysis of mapped restriction sites from plastid DNAs. American J. Bot. 81(5): 622-629.

Fernald, M. L. 1943. Five common rhizomatous species of Muhlenbergia. Rhodora 45: 221-239.

Herrera Arrieta, Y. 1998. A revision of the *Muhlenbergia montana* (Nutt.) Hitchc. complex. (Poaceae: Chloridoideae). Brittonia 50(1): 23-50.

Kurtz, J. & D. M. Sutherland. 1977. A taxonomic investigation of some rhizomatous species of the genus *Muhlenbergia* (Gramineae). Trans. Nebraska Acad. 4: 69-72.

Mitchell, W. W. & R. W. Pohl. 1966. Variation and aneuploidy in *Muhlenbergia glomerata*. American Midl. Nat. 76(1): 211-221.

Morden, C. W. 1985. A biosystematic study of the *Muhlenbergia repens* complex (Poaceae). Ph.D. dissertation. Texas A & M Univ. College Station.

Morden, C. W. & S. L. Hatch. 1987. Anatomical study of the *Muhlenbergia repens* complex (Poaceae: Chloridoideae: Eragos-tideae). Sida 12(2): 347-359.

Morden, C. W. & S. L. Hatch. 1996. Morphological variation and synopsis of the *Muhlenbergia repens* complex (Poaceae). Sida 17(2): 349-365.

Peterson, P. M. & C. R. Annable. 1991. Systematics of the annual species of *Muhlenbergia* (Poaceae-Eragrostoideae). Syst. Bot. Monogr. 31. 109 pp.

Peterson, P. M. & L. H. Rieseberg. 1987. Flavonoids of the annual *Muhlenbergia*. Biochem. Syst. Ecol. 15(6):

647-652.

Peterson, P. M. & Y. Herrera-Arrieta. 2001. A leaf blade anatomical survey of *Muhlenbergia* (Poaceae: Muhlenbergiinae)/ Sida 19(3): 469-506.

Pohl, R. W. 1969. *Muhlenbergia*, subgenus *Muhlenbergia* (Gramineae) of North America. American Midl. Nat. 82: 512-542. Pohl, R. W. & W. Mitchell. 1965. Cytogeography of the rhizomatous species of *Muhlenbergia*. Brittonia 17: 107-112.

Soderstrom, T. R. 1967. Taxonomic study of subgenus *Podosemum* and sec. *Epicampes* of *Muhlenbergia* (Gramineae). Contr. U. S. Natl. Herb. 34(4): 75-189.

Swallen, J. R. 1947. The awnless annual species of *Muhlenbergia*. Contr. U. S. Natl. Herb. 29: 203-208.

MONROA

Anton, A. M. & A. T. Hunziker. 1978. El género *Munroa* (Poaceae): sinopsis morfológica y taxonómica. Bol. Acad. Nac. Cienc., Córdoba, Argentina 52(3-4): 229-252.

Parodi, L. R. 1934. Contribucion al estudio de las gramineas del "*Munroa."* Rev. Mus. La Plata, Secc. Bot. 34: 171-193.

Reeder, J. R. & D. J. Crawford. 1970. Affinities of *Erioneuron* and *Munroa*. American J. Bot. 57: 752 (abstract).

Sanchez, E. 1984. Estudios anatomicos en el genero *Munroa* (Poaceae. Chloridoideae, Eragrostideae). Darwiniana 25: 43-57.

REDFIELDIA

Reeder, J. R. 1976. The systematic position of *Redfieldia* (Gramineae). Madroño 23: 434-438.

SCLEROPOGON

Allred, K. W. 1989. Observations on seed dispersal and implan-tations in burro-grass (*Scleropogon brevifolius* - Gramineae). Sida 13(4): 493, 496.

Anton, A. M., H. E. Connor, & M. E. Astegiano. 1998. Taxonomy and floral biology of *Scleropogon* (Eragrostoideae: Gramineae). Plant Sp. Biol. 13(1): 35-50.

Reeder, J. R. & L. J. Toolin. 1987. *Scleropogon* (Gramineae), a monotypic genus with disjunct distribution. Phytologia 62(3): 267-275.

SPOROBOLUS

Clayton, W. D. 1965. The *Sporobolus indicus* complex. Kew Bull. 19: 287-296.

Colbry, V. L. 1957. Diagnostic characteristics of the fruits and florets of economic species of North American *Sporobolus*. Contr. U. S. Natl. Herb. 34(1): 1-24.

Ortiz-Diaz, J.-J. & A. Culham. 2000. Phylogenetic relationships of the genus *Sporobolus* (Poaceae: Eragrostideae) based on nuclear ribosomal DNA ITS sequences. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing.

Collingwood, Australia. Pp. 184-188.

Riggins, R. 1977. A biosystematic study of the *Sporobolus asper* complex. Iowa State J. Res. 51: 287-321.

Veldkamp, J. F. 1990. The true identity of *Sporobolus poiretii* (Gramineae). Taxon 39: 327, 328.

Weakley, A. S. & P. M. Peterson. 1998. Taxonomy of the *Sporobolus floridanus* complex (Poaceae: Sporobolinae). Sida 18(1): 247-270.

SWALLENIA

Anderson, D. E. 1964. Notes on the leaf epidermis and chromosome number of *Swallenia*. Madroño 17: 201-203.

Goméz-Sánchez, M., P. Dávila-Aranda, & J. Valdés-Reyne. 2001. Estudio anatómico de *Swallenia* (Poaceae: Eragrostideae: Monanthochloinae), un género monotípo de Norte América. Madroño 48(3): 152-161.

Henry, M. A. 1979. A rare grass on the Eureka dunes. Fremontia 7(2): 3-6.

Soderstrom, T. R. & H. F. Decker. 1963. *Swallenia*, a new name for the California genus *Ectosperma* (Gramineae). Madroño 17(3): 88.

Swallen, J. R. 1950. *Ectosperma*, a new genus of grasses from California. J. Washington Acad. Sci. 40: 19-21.

TRIDENS

Crooks, P. & C. L. Kucerea. 1973. *Tridens* x *oklahomensis* (*T. flavus* x *T. strictus*), an interspecific sterile hybrid in the Eragosteae (Gramineae). American J. Bot. 60(3): 262-267.

Schuckman, S. M. & C. L. Kucera. 1984. The hybrid status of *Tridens oklahomensis* (Feath.) Feath. Trans. Missouri Acad. Sci. 18: 11, 12.

Tateoka, T. 1961. A biosystematic study of *Tridens*. American J. Bot. 48: 565-573.

TRIPLASIS

Nash, G. V. 1898. Revision of the genus *Triplasis*. Bull. Torrey Club 25: 561-565.

Cheplick, G. P. 1996. Cleistogamy and seed heteromorphism in *Triplasis purpurea* (Poaceae). Bull. Torrey Bot. Club 123: 25-33.

UNIOLA

Brown, W. V. & B. N. Smith. 1974. The kranz syndrome in *Uniola* (Gramineae). Bull. Torrey Bot. Club 101: 117-120.

Harper, J. R. & E. D. Seneca. 1974. A preliminary study of flowering in *Uniola paniculata* along the North Carolina coast. Bull. Torrey Bot. Club 101: 7-13.

Yates, H. O. 1966. Revision of grasses traditionally referred to *Uniola*, I. *Uniola* and *Leptochloopsis*. Southw. Nat. 11(3): 372-394.

Yates, H. O. 1966. Morphology and cytology of *Uniola*

(Gramineae). Southw. Nat. 11(2): 145-189.

Yates, H. O. & R. B. Channel. 1973. Nomenclatural history and taxonomic status of the Linnaean genus *Uniola* (Gramineae). Tennessee Acad. Sci. J. 48(1): 12-15.

TRIBE: CYNODONTEAE

Clayton, W. D. & F. R. Richardson. 1973. Studies in the Gramineae. XXXII. The tribe Zoysieae Miq. Kew Bull. 28(1): 37-48.

AEGOPOGON

Beetle, A. A. 1948. The genus *Aegopogon* Humb. & Bonpl. Univ. Wyoming Publ. XIII: 17-23.

BOUTELOUA

Arrieta, Y. H. & M. de la Cerda Lemus. 1997. Morfometria del genero *Bouteloua* Lag. (Poaceae) de Mexico. Phytologia 83(2): 113-124.

Columbus, J. T. 1996. Lemma micromorphology, leaf blade anatomy, and phylogenetics of *Bouteloua*, *Hilaria*, and relatives (Gramineae: Chloridoideae: Boutelouinae). Ph. D. dissertation. Univ. California, Berkeley.

Columbus, J. T. 1999. Morphology and leaf blade anatomy suggest a close relationship between *Bouteloua aristidoides* and *B. (Chondrosium) eriopoda* (Gramineae: Chloridoideae). Syst. Bot. 23(4): 467-478.

Columbus, J. T. 1999. An expanded circumscription of *Bouteloua* (Gramineae: Chloridoideae): new combinations and names. Aliso 18(1): 61-65.

Columbus, J. T. et al. 2000. Phylogenetics of *Bouteloua* and relatives (Gramineae: Chloridoideae): cladistic parsimony analysis of internal transcribed spacer (nrDNA) and *trn*L-F (cp DNA) sequences. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 189-194.

Esparza Sandoval, A. R. López-Ferrari, & J. Valdes-Reyna. 2000. Revision de *Bouteloua barbata* Lagasca (Poaceae: Eragrostideae). Phytologia 80(2): 73-91.

Freter, L. E. & W. V. Brown. 1955. A cytotaxonomic study of *Bouteloua curtipendula* and *B. uniflora*. Bull. Torrey Bot. Club 82: 121-130.

Gould, F. W. 1963. *Bouteloua-Chondrosium*, one genus or two? American J. Bot. 50: 634 (abstract).

Gould, F. W. 1969. Taxonomy of the *Bouteloua repens* complex. Brittonia 21(3): 261-274.

Gould, F. W. 1980. The genus *Bouteloua* (Poaceae). Ann. Missouri Bot. Gard. 66: 348-416.

Gould, F. W. & Z. J. Kapadia. 1964. Biosystematic studies in the *Bouteloua curtipendula* complex. II. Taxonomy. Brittonia 16(2): 182-208.

Griffiths, D. 1912. The grama grasses: *Bouteloua* and related genera. Contr. U. S. Natl. Herb. 14(3): 343-428 + plates 67-83.

- Reeder, J. R. & C. G. Reeder. 1980. Systematics of *Bouteloua breviseta* and *B. ramosa* (Gramineae). Syst. Bot. 5(3): 312-321.
- Roy, G. P. 1968. A systematic study of the *Bouteloua hirsuta-B. pectinata* complex. Ph. D. dissertation. Texas A & M Univ. College Station. 82 pp.
- Roy, G. P. 1974. Geographical distribution and population differentiation of *Bouteloua hirsuta B. pectinata* complex. Trop. Ecol. 14(2): 160-166.
- Roy, G. P. & F. W. Gould. 1971. Biosystematic investigation of *Bouteloua hirsuta* and *B. pectinata*. I. Gross morphology. South-west. Nat. 15: 377-387.
- Sandoval, S. E. & Y. H. Arrieta. 1996. Revision de *Bouteloua barbata* Lagasca (Poaceae: Eragrostoideae). Phytologia 80(2): 73-91.

CHLORIS

- Anderson, D. E. 1974. Taxonomy of the genus *Chloris* (Gramineae). Brigham Young Univ. Biol. Ser. 19(2): 1-133.
- Aradarajan, G. S. & A. J. Gilmartin. 1983. Phenetic and cladistic analyses of North American *Chloris* (Poaceae). Taxon 32: 380-386.
- Brown, L. E. 1969. A biosystematic study of the *Chloris cucullata-Chloris verticillata* complex. Ph. D. dissertation. Texas A & M Univ. College Station. 119 pp.
- Fosberg, F. R. 1976. Status of the name *Chloris barbata* (L.) Swartz. Taxon 25: 176-178.
- Nash, G. V. 1898. Revision of the genera *Chloris* and *Eustachys* in North America. Bull. Torrey Club 25: 432-450.
- Varadarajan, G. S. & A. J. Gilmartin. 1983. Phenetic and cladistic analyses of North American *Chloris* (Poaceae). Taxon 32(3): 380-386.
- Varadarajan, G. S. & A. J. Gilmartin. 1983. Subgeneric classification of *Chloris* Sw. (Poaceae). Taxon 32(3): 471-473.
- Vignal, C. 1979. Etude histologique des chloridaea. I. *Chloris* Sw. Adansonia 19: 39-70.

CTENIUM

- Smith, J. G. 1896. A synopsis of the American species of *Ctenium*. Bot. Gaz. 21: 361-364.
- Young, P. A. 1970. A revision of the New World species of *Ctenium*. Master of Arts thesis. Humboldt State College. Arcata, CA. 116 pp.

EUSTACHYS

- Nash, G. V. 1898. Revision of the genera *Chloris* and *Eustachys* in North America. Bull. Torrey Club 25: 432-450.
- Jones, S. D. & J. K. Wipff. 1992. *Eustachys retusa* (Poaceae), the first report in Florida and a key to *Eustachys* in Florida. Phytologia 73(3): 274-276.
- McKenzie, P. M., L. E. Urbatsch, & C. Aulbach-Smith. 1987. *Eustachys caribaea* (Poaceae), a species new to

the United States and a key to *Eustachys* in the United States. Sida 12(1): 227-232.

CYNODON

- Caro, J. A. & E. Sanchez. 1972. Novedades en *Cynodon* (Gramineae) de América. Darwiniana 17: 510-526.
- Clayton, W. D. & J. R. Harlan. The genus *Cynodon* L. C. Rich. in tropical Africa. Kew Bull. 24: 185-189.
- de Wet, J. M. J. & J. R. Harlan. 1970. Biosystematics of *Cynodon* L. C. Richard. (Gramineae). Taxon 19(4): 565-569.
- Harlan, J. R. et al. 1970. Geographic distribution of the species of *Cynodon* L. C. Rich. (Gramineae). East African Agric. Forest J. 36: 220-226.
- Harlan, J. R. et al. 1970. A guide to the species of *Cynodon* (Gramineae). Oklahoma State Univ. Agric. Exp. Sta. Bull 673. 37 pp.
- Harlan, J. R. et al. 1970. Cytogenetic studies in *Cynodon*. Crop Sci. 10(3): 288-291.
- Hitchcock, A. S. 1901. Bermuda grass. U. S. Dept. Agric. Div. of Agrostology Circular 31: 1-6.
- Kneebone, W. R. 1966. Bermuda grass -- worldly, wily, wonderful weed. Econ. Bot. 20(1): 94-97.
- Oross, J. W. & W. W. Thomson. 1984. The ultrastructure of *Cynodon* salt glands: secreting and nonsecreting. European J. Cell Biol. 34: 287-291.
- Tripathi, R. C. et al. 1977. Cytogenetic studies and evolutionary patterns in some *Cynodon* species. Biol. Zentralbl. 96(4): 423-436.

GYMNOPOGON

- Parodi, L. R. 1945. Una neuva especies de graminea del genero *Chloris* y sus relaciones con los *Gymnopogon*. Rev. Argent. Agron. 12: 45-50.
- Smith, J. P., Jr. 1971. Taxonomic revision of the genus *Gymnopogon* (Gramineae). Iowa State J. Sci. 45(3): 319-385.

HILARIA

- Columbus, J. T. 1993. *Pleuraphis*. <u>In</u>, Hickman, J. C. (editor). The Jepson manual: higher plants of California. Univ. California Press. Berkeley. P. 1283.
- Neuenschwander, L. F. et al. 1975. Review of tobosa grass (*Hilaria mutica*). Southwestern Nat. 20(2): 255-263.
- Sohns, E. R. 1956. The genus *Hilaria* (Gramineae). J. Washington Acad. Sci. 46: 311-321.
- West, N. E. (editor). 1972. Galleta: taxonomy, ecology and management of *Hilaria jamesii* on western rangelands. Utah Agric. Serv. Sta. Bull. No. 487.

SPARTINA

Aberle, B. 1993. The biology and control of introduced *Spartina* (cordgrass) worldwide and recommendations for its control in Washington. Master's thesis. Evergreen State College. Olympia. WA.

- Anttila, C. K. et al. 2000. Reciprocal hybrid formation of *Spartina* in San Francisco Bay. Mol. Ecol. 9(3): 765-770.
- Ayres, D. R. & D. R. Strong. 2003. *Spartina foliosa* (Poaceae) a common species on the road to rarity? Madroño 50(3): 209-213.
- Callaway, J. C. & M. N. Josselyn. 1992. The introduction and spread of smooth cordgrass (*Spartina alterniflora*) in south San Francisco Bay. Estuaries 15: 218-226.
- Clifford, P. M. 2002. Dense-flowered cordgrass (*Spartina densi-flora*) in Humboldt Bay: summary and literature review. California State Coastal Commission. Oakland. 22 pp. + 2 maps.
- Coastal Conservancy. 2002. Invasive *Spartina* project. www.spartina.org
- Daehler, C. C. & D. R. Strong. 1996. Status, prediction, and prevention of introduced cordgrass (*Spartina* spp.) invasions in Pacific estuaries., USA. Biol. Conservation 78: 51-58.
- Daehler, C. C. & D. R. Strong. 1997. Hybridization between introduced smooth cordgrass (*Spartina alterniflora*; Poaceae) and native California cordgrass (*S. foliosa*) in San Francisco Bay, California, USA. American J. Bot. 84(5): 607-611.
- Dore, W. G. & C. J. Marchant. 1968. Observations on the hybrid cord-grass *Spartina* x *caespitosa* in the maritime provinces. Canadian Field Nat. 82: 181-184.
- Faber, P. 2000. Good intentions gone awry. Why should anyone bring an alien cordgrass to San Francisco Bay? Coast and Ocean 16: 14-17.
- Frenkel, R. E. 1987. Introduction and spread of cordgrass (*Spartina*) into the Pacific Northwest. North West Environ. J. 3: 152-154.
- Huskins, C. L. 1931. The origin of *Spartina townsendii*. Genetica 12: 531-538.
- Marchant, C. J. 1963. Corrected chromosome numbers for *Spartina* x *townsendii* and its parent species. Nature 199(4896): 929.
- Marchant, C. J. 1966. The cytology of *Spartina* and the origin of *S.* x townsendii. Chromomes Today 1: 71, 72.
- Marchant, C. J. 1967. Evolution in *Spartina* (Gramineae). I. The history and morphology of the genus in Britain. J. Linn. Soc. London (Bot.) 60: 1-24.
- Marchant, C. J. 1968. Evolution in *Spartina* (Gramineae). II. Chromosomes, basic relationships and the problem of S. X townsendii agg. J. Linn. Soc. London (Bot.) 60: 381-410.
- Marchant, C. J. 1968. Evolution in *Spartina* (Gramineae). III. Species chromosome numbers and their taxonomic significance. J. Linn. Soc. London (Bot.) 60: 411-417.
- McDonnell, M. J. & G. E. Crow. 1979. The typification and taxonomic status of *Spartina caespitosa* A. A. Eaton. Rhodora 81: 123-129.

- Merrill, E. D. 1902. The North American species of *Spartina*. U. S. D. A. Bur. Plant Ind. Bull. 9: 1-16.
- Mertens, T. R. 1971. Speciation in *Spartina:* a classical example erroneously reported in textbooks. BioScience 21: 420-425.
- Mobberly, D. G. 1956. Taxonomy and distribution of the genus *Spartina*. Iowa State Coll. J. Sci. 30: 471-574.
- Naidoo, G., K. L. McKee, & I. A. Mendelssohn. 1992. Anatomical and metabolic responses to waterlogging and salinity in *Spartina alterniflora* and *S. patens* (Poaceae). Amer. J. Bot. 79(7): 765-770.
- O'Brien, D. L. & D. W. Freshwater. 1999. Genetic diversity with-in tall form of *Spartina alterniflora* Loisel. along the Atlantic and Gulf coasts of the United States. Wetlands 19(2): 352-358.
- Oliver, F. W. 1925. *Spartina townsendii*: its mode of establish-ment, economic uses and taxonomic status. J. Ecol. 13: 75-91.
- Ranwell, D. S. 1967. World resources of *Spartina townsendii* (sensu lato) and economic use of *Spartina* marshland. J. Appl. Ecol. 4: 239-256.
- Silander, J. A. 1979. Microevolution and clone structure in *Spartina patens*. Science 203: 658-660.
- Spicher, D. & M. Josselyn. 1985. *Spartina* (Gramineae) in northern California: distribution and taxonomic notes. Madroño 32(3): 158-167.
- Walsh, G. E. 1990. Anatomy of the seed and seedling of *Spartina alterniflora* Lois. (Poaceae). Aquatic Bot. 38(2-3): 177-193.

TRAGUS

- Anton, A. M. 1981. The genus *Tragus* (Gramineae). Kew Bull. 36: 55-61.
- Reeder, J. R. & C. G. Reeder. 1978. *Tragus racemosus* in Arizona. Madroño 25(2): 107, 108.

WILLKOMMIA

Hitchcock, A. S. 1903. Notes on North American grasses. III. New species of *Willkommia*. Bot. Gaz. 35: 283-285.

ZOYSIA

- Clayton, W. D. & F. R. Richardson. 1973. The tribe Zoysieae Miq. Studies in Gramineae. XXXII. Kew Bull. 28: 37-48.
- Forbes, I., Jr. 1952. Chromosome numbers and hybrids in *Zoysia*. Agron. J. 44: 194-199.
- Goudswaard, P. C. 1980. The genus *Zoysia* (Gramineae) in Malesia. Blumea 26(1): 169-175.

TRIBE PAPPOPHOREAE

PAPPOPHORUM

Chase, A. 1946. Enneapogon desvauxii and Pappophorum wrightii, an agrostological detective

story. Madroño 8: 187-189.

Renvoize, S. A. 1985. A survey of leaf-blade anatomy in grasses. VII. Pommereulleae, Orcuttieae, and Pappophoreae. Kew Bull. 40: 737-744.

ENNEAPOGON

Renvoize, S. A. 1968. The Afro-Asian species of *Enneapogon* P. Beauv. (Gramineae). Kew Bull. 22: 393-401.

TRIBE ORCUTTIEAE

Reeder, J. R. 1965. The tribe Orcuttieae and the subtribes of the Pappophoreae. Madroño 18: 18-27.

Reeder, J. R. 1982. Systematics of the tribe Orcuttieae (Gramineae) and the description of a new segregate genus, *Tuctoria*. Amer. J. Bot. 69(7): 1082-1095.

Roalson, E. H. & J. T. Columbus. 1999. Glume absence in the Orcuttieae (Gramineae: Chloridoideae) and a hypothesis of intratribal relationships. Aliso 18(1): 67-70

NEOSTAPFIA

Crampton, B. 1959. The grass genera *Orcuttia* and *Neostapfia:* a study in habitat and morphological specialization. Madroño 15: 97-110.

Crampton, B. 1976. Rare grasses in a vanishing habitat. Fremontia 4(3): 22, 23.

Davy, J. B. 1898. *Stapfia*, a new genus of Meliciae, and other noteworthy grasses. Erythea 6: 109-113.

Hoover, R. F. 1940. Observations on California plants. I. Leaflts. West. Bot. 2: 273, 274. [Tabular distinction of *Neostapfia* and *Anthochloa*]

ORCUTTIA

Crampton, B. 1959. The grass genera *Orcuttia* and *Neostapfia:* a study in habitat and morphological specialization. Madroño 15: 97-110.

Crampton, B. 1976. Rare grasses in a vanishing habitat. Fremontia 4(3): 22, 23.

Griggs, F. T. 1974. Systematics and ecology of the genus *Orcuttia.* M. S. thesis. California State Univ., Chico. 69 pp.

Griggs, F. T. 1976. Life history strategies of the genus *Orcuttia* (Gramineae). <u>In</u>, Verenal pools – their ecology and conservation. Publ. No. 9. Inst. of Ecology. Univ. California, Davis. Pp. 57-63.

Griggs, F. T. 1980. Population studies in the genus *Orcuttia* (Poaceae). Ph. D. dissertation. Univ. California, Davis. 98 pp.

Hoover, R. F. 1941. The genus *Orcuttia.* Bull. Torrey Bot. Club 68: 149-156.

Hoover, R. F. 1943. Observations on California plants. I. Leaflts. West. Bot. 2: 273, 274.

Reeder, J. R. 1965. The tribe Orcuttieae and the subtribes of the Pappophoreae. Madroño 18: 18-27.

Reeder, J. R. 1980. Nomenclatural changes in *Orcuttia* (Gramineae). Phytologia 47: 221.

Stagg, C. M. 1977. The distribution of *Orcuttia californica* (Poaceae) in the vernal pools of the Santa Rosa Plateau, River-side County, California. Crossosoma Fall: 10-.

TUCTORIA

Reeder, J. R. 1982. Systematics of the tribe Orcuttieae (Gramineae) and the description of a new segregate genus, *Tuctoria*. Amer. J. Bot. 69(7): 1082-1095.

3.10 - SUBFAMILY PANICOIDEAE

TECHNICAL DESCRIPTION

Habit: Mostly herbaceous grasses; internodes often

Root Hairs: Equal

Leaf Epidermis: Complex, needle-shaped bicellular micro-hairs present; silica cells usually dumbbell-shaped, x-shaped, or saddle-shaped; stomata rhombic

Leaf Anatomy: Single sheath of parenchyma around vascular bundles (except in plants with kranz syndrome); chlorenchyma may be more or less radiating

Inflorescence: Panicles, compound racemes, rames, and spikes

Spikelet: Dorsally compressed, disarticulating below the glumes; 1 terminal perfect floret and a staminate or neuter one below it; glumes 2, 1, or absent

Flower: Lodicules short, truncate, and heavily vasculated; stamens 3; stigmas 2

Embryo: P - P P

Cytology: X = 9, 10 (rarely 5 or 8); some with

persistent nucleoli

Photosynthetic Pathways: C₃ and C₄

Distribution: Diverse habitats, abundant in the tropics and subtropics; absent from the Arctic.

SYSTEMATICS

This circumscription of this subfamily has changed little in recent years. The principal modifications at the tribal level consists of merging Melinidae with Paniceae. Tripsaceae (Maydeae), here recognized as a distinct tribe, is often merged with Andropogoneae. Many adjustments have occurred at the generic level.

TRIBE PANICEAE

Inflorescence a panicle or a series of racemose or spicate branches that bear racemes or spikes of spikelets. First glume short, sometimes missing; second glume and sterile lemma both membranous and soft; fertile floret indurate or leathery. Spikelets or clusters of them sometimes subtended by bristly or spiny involucres. See Section 4 for a key to our

SUBTRIBE SETARIINAE

The principal distinguishing feature of the subtribe is spikelets with a hard upper lemma. The surface may

be granular, wrinkled or highly polished.

PANICUM. Panic grass, panicum. Caespitose, rhizomatous or stoloniferous annuals or perennials. Culms herbaceous or woody, the internodes hollow or solid, to 4 m tall. Inflorescence an open to contracted panicle, either terminal or axillary, rarely a raceme. Spikelets 2-flowered, dorsally compressed, bisexual, disarticulating below the glumes. Lower floret sterile or sometimes staminate, its lemma similar in size and texture to the second glume. Upper floret fertile, its lemma firm to indurate, awnless, and clasping the palea with its enrolled margins. Fertile lemma and palea of similar texture. Glumes 2, the first much shorter than the second, the first 1- to 7-nerved and the second 3- to 9-nerved; sterile lemma 5- to 9-nerved, awnless, similar to second glume in size and texture; fertile lemma 3- to 11-nerved, awnless, glabrous, firm to indurate at maturity; palea of fertile floret similar to lemma in texture, and tightly clasped by it. Lodicules 2; stamens 3; stigmas 2, red pigmented.

The largest genus of Gramineae with about 500 species, native just about everywhere. Of considerable economic importance as a source of grains, pasture grasses, and weeds. As treated by H & C, the genus consisted of three subgenera: *Eupanicum, Dichanthelium*, and *Paurochaetium*. Grasses of subgenus *Paurochaetium* have a point or bristle that subtends the uppermost spikelet and they are now often placed in the genus *Setaria*. Gould elevated the subgenus *Dichanthelium* to the generic level. That opinion, while followed early on, has been increasingly rejected.

The following key may be helpful in understanding the redefinition of *Panicum*. It is modified from H & C and Gould (1979).

- Axis of branchlets extending beyond the base of the uppermost spikelet as a point or bristle -> Setaria subgenus Paurochaetium
- Axis of branchlets rarely flattened, but never pointed or bristle-like ->
- Plants annual or perennial, without a basal rosette of short, broad blades or a basal tuft of soft, linear blades; panicles open or contracted at maturity, the spikelets long- or short-pediceled (in some plants the spikelets subsessile and the primary branches spicate); lemma and palea of upper floret smooth or rugose; culms not becoming much-branched in age and with reduced branchlets and panicles; plants flowering from July to November ->

Panicum subgenus Panicum

2. Plants perennial, most species developing a basal rosette of short, broad basal blades in spring or with a tuft of soft, linear blades; panicles of main culms open, the spikelets loosely-spaced, at least some with pedicels much longer than spikelets; lemma and palea of upper floret smooth; culms of several species becoming much-branched in age to produce fascicles of reduced leafy branches and panicles; most species flowering first March to May and then throughout the growing season at irregular int Panisum subgenus Dichanthelium

Important grasses of the subgenus *Panicum* include *P. virgatum* (switch grass), a rhizomatous perennial over much of the United States; *P. capillare* (witch grass), a common plant of open ground and waste places; *P. bulbosum* (bulb panicum), an important forage grass of the Southwest; *P. miliaceum* (proso millet, broomcorn millet), an escape from cultivation; and *P. maximum* (elephant grass, Guinea grass), one of the tropical forage grasses introduced into the southern states.

Hitchcock & Chase recognized over one hundred

species in the subgenus *Dichanthelium*, many of them very difficult to distinguish from one another and known only from very localized occurrences. These grasses are especially common in the southeastern states. In their monograph, Gould and Clark performed the heroic and welcomed task of reducing the number of species to 26! *P. oligosanthes*, in the broad sense, is perhaps the most widespread species in the subgenus.

DICHANTHELIUM VERSUS PANICUM

SUBGENUS DICHANTHELIUM	SUBGENUS PANICUM	
Perennial	Annual or perennial	
Leaf blades rarely ribbed/furrowed	Pronounced ribs/furrows on upper and lower surfaces	
Autumn/winter rosettes present Cleistogamous lateral inflorescences Chasmogamous terminal inflorescences	Rosettes absent	
Diploids (mostly)	Polyploids (mostly)	
x = 9 2n = 2x = 18	x = 9 or 10 2n = 4x = 18, 20, 30, 36, 40, 72 + aneuploids	
Embryo relatively small	Embryo relatively large	
Non-kranz anatomy C ₃ [phosphoglyceric acid] Chlorenchyma irregular Numerous air spaces Outer sheath without chloroplasts	Kranz anatomy C ₄ [oxaloacetic acid, malic acid] Chlorenchyma radial Few air spaces Large, specialized chloroplasts	
Palea tip with simple papillae Papillae in regular rows	Papillae compound or clustered Papillae irregular	
Double tunica layer	Single tunica layer	

Sources:

Clark, C. & F. Gould. 1978. American J. Bot. 62(7): 743-748. Gould, F. & C. Clark. 1978. Annals Missouri Bot. Gard. 65(4): 1088-1132.

GENERA SEGREGATED FROM PANICUM

If we look in the "Species Plantarum" of Linnaeus, we will see that He recognized a genus *Panicum*. It included most everything that we now think of as a panicoid grass. Since then, botanists have carved out an impressive list of genera, most of which have stood the test of time.

Segregate Genus	Basionym in <i>Panicum</i>	
Echinochloa (1812)	Panicum crusgalli	
Setaria (1812)	Panicum viride	
Stenotaphrum (1822)	Panicum dimidiatum	
Steinchisma (1830)	Panicum hians	
Brachiaria (1853)	Panicum eruciforme	
Sacciolepis (1901)	Panicum glabrum	
Phanopyrum (1903)	Panicum gymnocarpum	
Leptoloma (1906)	Panicum cognatum	
Lasiacis (1910)	Panicum divaricatum	
Homolepis (1911)	Panicum aturense	
Paspalidium (1920)	Panicum geminatum	
Urochloa (1920)	Panicum reptans	
Dichanthelium (1974)	Panicum dichotomum	

ERIOCHLOA. Cup grass. Annuals or perennials, often of moist sites. Culms herbaceous, to 1 m tall. Inflorescence a sparingly branched, contracted panicle. Spikelets 2-flowered (the lower sterile and the upper bisexual), dorsally compressed, disarticulation below the glumes. Glumes 2, the first reduced to minute sheath or strip that is fused to the thickened ring- or cup-like callus; sterile lemma 5-nerved, similar to second glume, longer than fertile floret; fertile lemma 5-nerved, glabrous, indurate, mucronate to awned; palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2, red pigmented.

A genus of 25-30 species native to warmer parts of both hemispheres. *E. contracta* (prairie cup grass) occurs in the central and southwestern portions of the U. S.; *E. sericea* (Texas cup grass) is native to the Great Plains.

PASPALUM. Dallis grass, knot grass, bahia grass. Caespitose, rhizomatous, or stoloniferous perennials; rarely annuals. Culms herbaceous, the internodes hollow or solid, to 3 m tall. Inflorescence a series of racemose [rarely paired] branches, bearing subsessile spikelets. Spikelets 2-flowered, dorsally compressed, solitary or paired along a narrow or broadly-winged rachis, bisexual, disarticulating below the glumes. Lower floret sterile or staminate; upper floret fertile. Glumes 1 (first usually absent) or rarely 2, 3- to 6nerved, second glume and sterile lemma of lower floret similar in size and texture; sterile lemma 3- to 5-nerved, as long as the fertile lemma; fertile lemma 3- to 5-nerved, rounded on the back, firm to indurate, awnless, its back facing toward the rachis; palea broad, flat or slightly convex, its margins covered covered by the enrolled edges of the lemma. Lodicules 2; stamens 3; stigmas 2, red pigmented.

A genus of about 400 species native to warmer regions of both hemispheres. Of economic significance as a source of grains, pasture grasses, and weeds. *P. dilatatum* (Dallis grass), is an important forage grass that becomes infected by the ergot fungus; *P. distichum* (knot grass) is a plant of wet areas along both coasts and the southern half of the country.

STENOTAPHRUM. St. Augustine grass. Mat-forming, rhizomatous, stoloniferous perennials. Culms herbaceous, to 1 dm tall. Leaf sheaths compressed; blades flat, succulent. Inflorescence spike-like, the spikelets embedded in a thickened rachis. Spikelets 2-flowered (the lower staminate or sterile and the upper fertile), dorsally compressed, bisexual, disarticulation below the glumes and falling with a rachis joint. Glumes 2, unequal, the lower nerveless and the upper 5- to 9-nerved; sterile lemma 7- to 9-nerved, as long as the second glume; fertile lemma 3- to 5-nerved, chartaceous, firmer than the glumes; palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2, red or white pigmented.

A genus of 6 or 7 species native to the subtropical and tropical regions of the Old and New Worlds. *S. secundatum* (St. Augustine grass) is native to the Southeastern United States. It is a popular lawn grass that also escapes and becomes weedy.

ECHINOCHLOA. Barnyard grass, jungle-rice. Coarse, caespitose annuals or perennials. Culms herbaceous, the internodes hollow or solid, often succulent. Leaf sheaths compressed. Inflorescence a contracted to more or less open panicle, the branches simple to rebranched. Spikelets subsessile, solitary, or in irregular clusters on one side of the branch, 2-flowered (the lower sterile and the upper fertile), plano-convex, bisexual, disarticulation below the glumes. Glumes 2, unequal, the lower 0- to 3-nerved and the upper 5- to 7-nerved; sterile lemma similar to second glume, 5-nerved, awned; fertile lemma plano-convex, 5-nerved, smooth and shining, pointed, its margins enrolled below (enclosing the palea at that point), the upper portion flat; palea similar to fertile lemma in texture, narrowing to a point that is free from the lemma margins, 2-nerved. Lodicules 2; stamens 3; stigmas 2, red pigmented.

A genus of 20-40 species native to the warmer regions of both hemispheres. Of economic significance as a source of minor cereals, grains, pasture grasses, and weeds. *E. crus-galli* (barnyard grass) and *E. colona* (jungle-rice) are weedy in the United States.

AXONOPUS. Carpet grass, mat grass. Ours stoloniferous or caespitose perennials. Inflorescence a series of 2 to many subdigitate racemes, sometimes inserted on a central axis. Spikelets oblong-elliptical, disarticulating below the glume. First glume absent; rounded back of fertile lemma indurate, turned away from rachis; palea indurate. Lodicules 2; stamens 3; stigmas 2, white.

A genus of about 100 species native to the warmer regions of the New World, especially South America. One species is native to Africa. They occur in savannas, in forest clearings, and can become weedy. Some species are used for pasture and for lawns. Three species are native to the United States; *A. affinis*, common carpet grass, is the most frequently encountered.

SACCIOLEPIS. Cup scale. Annual or perennial herbs. Inflorescence a contracted [open] panicle. Spikelets laterally compressed, disarticulating below the glumes. Glumes 2, prominently ribbed, the upper one gibbous and inflated; sterile lemma 3- or 5-nerved; upper lemma dorsally compressed; fertile floret smooth, indurate, rounded. Lodicules 2; stamens 3; stigmas 2.

A genus of about 30 species native to the tropics and

subtropics, especially of Africa. They are typically found in wet sites and shallow waters. Only *S. striata*, American cupscale, is native to the U. S., from TX and OK the SE portion of the country.

SETARIA. Foxtail, bristle grass, millet. Annuals or caespitose perennials; rhizomes and stolons common. Culms herbaceous, the internodes hollow or solid, to 3 m tall. Inflorescence a dense, spike-like panicle. Spikelets awnless, but subtended by 1 to several bristles (sterile branches), 2-flowered (the lower sterile and the upper fertile), dorsally compressed, bisexual, disarticulation below the glumes, but above the bristle(s). Glumes 2, the first broad, typically about half the length of the second, second glume and sterile lemma similar in size and texture; sterile lemma 5-nerved, as long as or longer than the fertile one; fertile lemma 1- to 5-nerved, indurate, rounded at its apex, with fine to coarse transverse wrinkles; palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2, white or red pigmented.

A genus of 110-125 species native to the warmer regions of both hemispheres. Of economic importance as a source of grains, a minor cereal, pasture grasses, and weeds. Rominger recognizes three sub-genera in North America: Setaria, with non-plicate leaf blades spikelets subtended by Ptychophyllum, with plicate leave blades and all spikelets subtended by bristles; and Paurochaetium (formerly treated as a subgenus of Panicum), with only the uppermost spikelet of a branchlet bearing a bristle. Noteworthy species include *S. lutescens* (yellow foxtail), *S. viridis* (green foxtail), *S. faberi* (giant foxtail), and *S. verticillata* (bristly foxtail), often encountered in disturbed areas and as agricultural weeds. S. italica (foxtail millet, Hungarian millet, Italian millet) has been in cultivation as a food plant since prehistoric times.

SUBTRIBE: MELINIDINAE

Grasses of this subtribe have panicles of spikelets that have a reduced first glume.

MELINIS. Molasses grass, ruby grass, Natal grass. Annuals or perennials. Inflorescence an open to contracted panicle. Spikelets 2-flowered (the lower staminate or sterile and the upper fertile), rounded to dorsally compressed, disarticulation below the glumes. Glumes 2, the first a tiny scale or reduced to a rim; second glume 5- to 7-nerved, similar to sterile lemma in size and texture (silky-villous in *M. repens*), sterile lemma 3- to 5-nerved; fertile lemma 1- to 5-nerved, awned or awnless; palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2.

A genus of about 26 species, all but one of them native to the Old World. As treated here, the genus includes *Rhynchelytrum*. *M. minutiflora* (molasses grass), an important tropical pasture grass, escapes in Florida. *M. repens* [= *Tricholaena rosea* in H & C and *Rhynchelytrum r.* in most recent grass floras] (ruby grass, Natal grass) is common in the tropics and is now weedy in California, Arizona, Texas, and Florida.

SUBTRIBE: DIGITARIINAE

Grasses of this subtribe usually have racemes of spikelets, with their first glumes reduced or even absent.

DIGITARIA. Crab grass, cotton top, fall witchgrass. Caespitose annuals or rhizomatous, stoloniferous perennials. Culms herbaceous, the internodes solid or hollow, to 2 m tall. Inflorescence a series of racemose or digitate branches bearing subsessile or shortpedicellate spikelets. Spiklets in pairs or trios (rarely solitary or in 5's), alternating in 2 rows on one side of a 3-angled winged or wingless rachis. Spikelets 2-flowered (the lower sterile and the upper fertile), plano-convex, bisexual, disarticulation below the glumes. Glumes 1 [rarely 2], the first minute or absent, the second resembling the sterile lemma in size and texture, 3- to 7-nerved, glabrous to long-ciliate; sterile lemma as long as fertile floret, 3- to 7nerved, often hairy; fertile lemma more or less narrow, acute or acuminate, cartilaginous or leathery, its margins thin and flat, not clasping the palea as in other genera of the tribe; palea relatively long, 2nerved. Lodicules 2; stamens 3; stigmas 2, red pigmented.

A genus of about 300 species native to warmer regions of both hemispheres. Of economic importance as a source of grains, pasture and lawn grasses, and weeds. As treated here, the genus includes taxa assigned to *Leptoloma* and *Trichachne* in H & C. *D. sanguinalis* (hairy crab grass) and *D. ischaemum* (smooth crab grass) are major weeds around the country; *D. californica* (Arizona cottontop) [= *Trichachne c.* in H & C] is an important forage grass in the Southwest; *D. cognatum* (fall witchgrass) [= *Leptoloma c.* in H & C] is common in the East.

SUBTRIBE: CENCHRINAE

This subtribe is probably the most easily recognized because of the bristles or scales that subtend the spikelets.

PENNISETUM. Fountain grass, feathertop. Caespitose, rhizomatous, stoloniferous perennials; rarely annual. Culms herbaceous, the internodes hollow or solid, to 4 m tall. Inflorescence a dense, spike-like panicle. Spikelets 2-flowered (the lower sterile and the upper fertile), dorsally compressed, bisexual, solitary or in clusters of 2 or 3, subtended by an involucre of bristles (often plumose), these united at their bass and falling with the spikelet at disarticulation. Glumes 2, the first small or vestigial, second glume and sterile lemma similar in size and texture; sterile lemma 3- to 9-nerved, less firm than fertile lemma; fertile lemma 5- to 7-nerved, similar to or firmer than glumes in texture; palea relatively long, 2-nerved. Lodicules 2 or 0; stamens 3; stigma 2.

A genus of about 80 species native to the warmer regions of both hemispheres. Of economic importance as a source of grains, pasture and lawn grasses, ornamentals, and weeds. *P. glaucum* (pearl millet) is an important food plant in the tropics. *P. purpureum* (Napier grass, elephant grass) is an important forage plant; *P. clandestinum* (kikuyu grass) is an important pasture grass; *P. villosum* (feathertop) is grown as an ornamental, where it often escapes to become a major pest.

CENCHRUS. Sandbur. Annuals or perennials, caespitose, rhizomatous, or stoloniferous. Culms herbaceous, often weak, decumbent, internodes hollow or solid, to 1 m tall. Inflorescence a series of spike-like or racemose burs, these readily disarticulating. Spikelets 2-flowered (lower sterile and the upper fertile), dorsally com-pressed, bisexual,

hidden within burs (involucres of bristles or spines), the entire structure falling from the plant at maturity. Glumes 2, unequal, the first 1- to 5-nerved and the second 1- to 7-nerved, thin, membranous; sterile lemma 1- to 7-nerved, ± equal to fertile lemma, awnless; fertile lemma 3- to 7-nerved, thin, membranous, its apex acuminate; palea relatively long, 2-nerved. Lodicules 0; stamens 3; stigmas 2.

A genus of about 20 species native to the warmer regions of both hemispheres, but mostly American. *C. incertus* (including *C. pauciflorus* of H & C) is widespread in sandy places over much of the country; *C. myosuroides* (big sandbur) can reach 1.5 m in moist, sandy sites in the Southeast. Transitional species make the distinction between this genus and *Pennisetum* difficult.

TRIBE: ANDROPOGONEAE

Inflorescence a series of paired spikelets, evenly or unevenly pedicellate; pedicellate spikelet often reduced (sometimes missing, which can lead to misinter-pretation of the inflorescence). Spikelets 2-flowered, the lower sterile or staminate and the upper bisexual, awnless or awned from fertile lemma, the awn often easily disarticulating. Glumes thick, firm, equal in length. Fertile lemma and palea thin.

SUBTRIBE: SACCHARINAE

Inflorescence terminal, of solitary, digitate, or paniculate rames. Spikelets paired, similar; one sessile and the other pedicellate or both with pedicels. Clayton & Renvoize view this group as the most primitive because the grasses have unspecialized rachis internodes and both members of the spikelet pair are fertile.

SACCHARUM. Sugar cane, plume grass. Robust perennials. Inflorescence a conspicuous, plumose panicle to almost 1 m in length. Spikelets of the pair alike, fertile, awnless, and obscured by a tuft of long, silky hairs attached at the bases. Glumes large and firm. Sterile lemma, fertile lemma (sometimes absent), and palea membranous. *S. officinarum* (sugar cane), one of the earliest plants to be domesticated, is the source of about two-thirds of the sugar used in commerce.

As treated here, the genus includes *Erianthus*, which H & C recognized as a separate genus. These plants, which are found in the Southeast, differ from sugar cane in having a long awn arising from the fertile lemma.

MISCANTHUS. Miscanthus, eulalia. Caespitose or rhizomatous perennials, often cane- or reed-like. Culms herbaceous, the internodes solid, to 2 m tall. Inflorescence a fan-shaped panicle. Spikelets unequally pedicellate, silky hairy, 2-flowered, dorsally compressed, bisexual, disarticulation below the glumes. Glumes 2, ± equal, papery to membranous, the first 3- or 4-nerved and the second 1- to 5-nerved; sterile lemma membranous, longer than the fertile one, awnless; fertile lemma membranous, 0- to 3-nerved, with a geniculate and twisted awn or awnless; palea relatively short, nerveless. Lodicules 2; stamens 2 or 3; stigmas 2, red pigmented.

A genus of about 20 species native to the Old World, especially to Asia. *M. sinensis* is grown as an

ornamental in this country.

SUBTRIBE: ANDROPOGONINAE

Inflorescence of single, paired [digitate] rames, these aggregated into terminal or axillary compound panicles. Spikelets of the pair dissimilar. Grasses of this subtribe have 2-keeled glumes and callus inserted into hollowed tip of the internode.

ANDROPOGON. Bluestem, beard grass. Coarse, caespitose or rhizomatous perennials; less frequently annuals. Culms herbaceous, the internodes solid, to 2 m tall. Inflorescence a series of 2-several rames, the flowering culms much-branched, subtended by a spathe-like sheath in some species. Spikelets 2-flowered (the lower floret sterile and the upper bisexual), dorsally compressed. Sessile spikelet well-developed and fertile, disarticulating with a section of rachis and pedicel; pedicellate spikelet well-developed, reduced, or absent. Glumes 2, ± equal, firm, the first keeled and 1- to several-nerved and the second 1- to 3-nerved, awnless; sterile lemma hyaline, 2-nerved, ± equalling the fertile lemma; fertile lemma hyaline, narrow, 1- to 3-nerved entire or bifid, usually bearing a bent and twisted awn; palea hyaline, reduced or absent, nerveless. Lodicules 2; stamens 1-3; stigmas 2.

A genus of about 100 species native to warmer regions of the Old and New Worlds. We have a number of them in North America, mostly in the southeastern United States. As treated by H & C, Andropogon consisted of three subgenera: Arthrolophis, Amphilophis, and Schizachyrium. Each is now recognized as a distinct genus. Andropogon [= subgenus Arthrolophis] includes such common species as A. virginicus (broomsedge), a plant of sandy, sterile soils in the Southeast; A. gerardii (big bluestem), a very important forage plant of the tall grass prairie; A. hallii (sand bluestem), a close relative of big bluestem that grows in sandy places; and A. glomeratus (bushy beard grass), which grows in moist places in the Southeast and Southwest.

SCHIZACHYRIUM. Little bluestem. Caespitose or rhizomatous perennials. Culms herbaceous, the internodes hollow or solid. Flowering culms muchbranched, each terminating in a single narrow rame. Spikelets as in *Andropogon*, often closely-appressed to the rachis. Sessile spikelet fertile and awned; pedicellate spikelet rudimentary.

A genus of about 60 species native to the warmer regions of north hemispheres. *S. scoparium* (little bluestem) is an important forage plant of the tall grass prairie, where it is one of the dominants.

SUBTRIBE: ANTHISTIRIINAE

According to Clayton & Renvoize (1986: 354), this sub-tribe "... is distinguished from Andropogoniinae by a pointed callus applied obliquely to the internode tip, rather than blunt and sunk into it."

HYPARRHENIA. Thatching grass, jaraguá grass. Caespitose perennials, to 2.5 m tall. Inflorescence an elongate compound panicle, consisting of paired racemes subtended by ± conspicuous bracts. Spikelets paired, the lower pairs sterile and awnless; fertile spikelets 1 to a few on each rame, the lemma bearing a strong geniculate awn.

A genus of about 55 species native to the Old World, mainly to Africa, where they are common in savannas. Introduced in the American tropics, where it has been weedy. Two species, *H. hirta* and *H. rufa*, occur as roadside weeds in the Southwest and Southeast.

SUBTRIBE: SORGHINAE

Inflorescence terminal [axillary], of single, digitate, or paniculate rames, sometimes reduced to a trio of spike-lets or a single spikelet. Spikelets paired, dissimilar, the sessile bisexual; pedicellate spikelet male or barren, sometimes much reduced.

DICHANTHIUM. Bluestem. Caespitose, rhizomatous or stoloniferous perennials [rarely annual]. Culms herbaceous, internodes solid, to 2 m tall. Inflorescence similar to *Bothriochloa*, but lacking central groove or membranous area on its pedicels and internodes. The lower pairs of spikelets on each rame are typically sterile and awnless.

A genus of about 16 species native to the Old World tropics. The genus is not included in H & C. *D. annulatum* (Kleberg bluestem), *D. aristatum* (Angleton bluestem), and *D. sericeum* (silky bluestem) have been introduced in this country as pasture grasses and have become naturalized in Texas and Louisiana.

BOTHRIOCHLOA. Bluestem. Caespitose, rhizomatous, or stoloniferous perennials. Culms herbaceous, the internodes solid, to 2 m tall. Inflorescence a series of a few to several rames, as in Andropogon. Pedicels and upper rachis branches with a central groove or membranous area. Sessile spikelet fertile and awned, some with a pit (depressed glandular area) on the middle or upper portion of the first glume of the sessile spikelet. Pedicellate spikelet usually well-developed, but staminate or neuter. Spikelets disarticulating with a section of rachis.

A genus of about 30-35 species native to the warmer parts of both hemispheres. This genus was treated as *Andropogon* subgenus *Amphilophis* in H & C. *B. saccharoides* (silver beard grass) and *B. barbinodis* (cane bluestem) are important forage grasses.

SORGHUM. Sorghum, milo, broomcorn, kaffir. Stout annuals or caespitose, stoloniferous, or rhizomatous perennials. Culms herbaceous, the internodes solid, to 3 m tall. Inflorescence a large, open to contracted panicle. Spikelets in trios at branchlet tips (the lateral ones pedicellate and sterile) or in pairs below (one sessile and the other pedicellate). Sessile spikelets disarticulating with a rachis segment. Glumes 2, \pm equal, 3- to several-nerved, leathery, awnless; sterile lemma membranous, 0- 2-nerved, awnless; fertile lemma membranous, 1- to 3-nerved, with a geniculate and twisted awn (deciduous in *S. halepense*); palea relatively long, 2-nerved. Lodicules 2; stamens 3; stigmas 2, red pigmented.

A genus of 30-35 species, only two of them native to the New World (none to North America). The genus is of considerable economic importance as a source of grains, fodder, and weeds. *S. bicolor* (sorghum, milo) [= *S. vulgare* in H & C] is the source of sugary juices for syrups, grain for cattle feed, millets for humans and domesticated animals, and broomcorn, from which traditional brooms are made; *S. halepense* (Johnson grass) is a tetraploid, pernicious weed.

SORGHASTRUM. Indian grass. Caespitose perennials.

Culms herbaceous, to 2 m tall. Inflorescence a terminal panicle of rames. Spikelets paired, the pedicellate greatly reduced (often represented by nothing more than a hairy pedicel). Sessile spikelet disarticulating with rachis segment remaining attached. Glumes 2, ± equal, the lower 9-nerved and the upper 5-nerved, leathery; sterile lemma membranous, 2-nerved, awnless; fertile lemma membranous, 1-nerved (?), with a stout, twisted, geniculate awn; palea often reduced or absent. Lodicules 2; stamens 3; stigmas 2.

A genus of 15-20 species native to the warmer regions of both hemispheres. *S. nutans* (Indian grass) is a native of the tall grass prairie; two other species have more restricted distribution in the Southeast.

SUBTRIBE: ROTTBOELLINEAE

Inflorescence cylindrical, spike-like. Spikelets paired, awnless, embedded in cavities or hollow rachis joints, the pedicel often fused to the rachis. Disarticulation occurs as the rachis joints separate from one another at maturity. See Section 4 for a key to our North American taxa.

ROTTBOELLIA. Itch grass, Kelly grass. Robust annual. Blades to 3 cm wide. Sheaths papillose-hispid, the hairs irritating to some individuals. Inflorescence a subcylindric raceme, its apex with abortive spikelets only. Spikelets awnless, paired (one sessile and perfect, the other pedicellate and sterile), borne at the nodes of a thickened rachis. Upper rachis joints hollow, the thickened pedicel adnate to it, thereby making the pedicellate spikelet appear sessile. A genus of 3 species native to the Old World tropics. *R. cochinchinensis* (= *R. exaltata* in H & C) is introduced in Florida. It provides fodder.

ELIONURUS. Balsamscale. Erect perennials. Inflorescence a single spike-like raceme. Spikelets awnless, paired, one sessile and the other pedicellate, along a discontinuous rachis. The sessile spikelet perfect, appressed to the concave side of the rachis; the pedicellate spikelet staminate. Spikelet pair disarticulating with a segment of the rachis. A genus of about 14 species native to America, Africa, and Australia. Our species are found in the drier areas of the Southwest and in the prairies and pine woods of the Southeast. The spelling *Elyonurus* is used in older literature.

EREMOCHLOA. Centipede grass. *Eremochloa ophuroides*, native to Southeast Asia, has been introduced into Florida and other areas in the Southeast as a popular lawn grass and for erosion control. It is a low, rhizomatous perennial that forms dense turf. It produces spike-like racemes of paired sessile/pedicellate spikelets on terminal and axillary peduncles. The rachis is not thickened. The first glume of the sessile spikelet is winged at the summit.

HACKELOCHLOA. Pitscale grass. Annuals to 1 meter. Sheaths and blades papillose-hirsute. Inflorescence a series of many solitary, spike-like racemes enclosed in spathes. Spikelets awnless, paired, one sessile and the other pedicellate. Rachis joint and pedicel fused, clasped between the edges of the first glume of the sessile spikelet. The rounded, pitted appearance of the first glume is highly diagnostic. A genus of 2 species native to the Old World and New World tropics. Hackelochloa granularis has been introduced in the Southwest and in the Southeast. It is a limited source

of forage.

HEMARTHRIA. Limpo grass. Mostly perennials. Inflorescence a single flattened, axillary raceme. Spikelets paired, the sessile one sometimes awned, its lower glume slightly winged; pedicel of stalked spikelet fused to internode. A genus of 12 species native to the warmer regions of the Old World, especially of wet sites. *H. altissima* (= *Manisuris a.* in H & C) is adventive in the United States.

COELORACHIS. Joint-tail grass. Perennials, often with broad leaf blades. Inflorescence a single cylindrical or flattened raceme. Spikelets paired, less often in trios. Pedicellate spikelet well-developed or vestigial, its stalk free from the rachis. A genus of 20 or so species native to the tropics, especially on dampsoils in savannas and grasslands. The four species found in North America were treated as species of *Manisuris* by H & C.

SUBTRIBE: ZEINAE

The grasses of this subtribe are characterized by highly modified, unisexual, dimorphic spikelets, borne within the same or in different inflorescences. This group is often treated as a distinct tribe, as in H & C, who called it Tripsaceae [= Maydeae of more recent authors].

If G. L. Stebbins was correct in concluding that grasses are most advanced of the flowering plants, then maize and its relatives may well be the most advanced of the most advanced! That's why I put them at the end of our survey.

COIX. Job's tears. Annuals or perennials. Culms herbaceous, the internodes solid, to 2 m tall. Leaf blades broad (to 4 cm). Inflorescence a raceme, but not immediately apparent as such. Staminate spikelets 2-flowered, in pairs or trios along a common rachis, protruding from an opening at apex of a very hard, white or drab, beadlike involucre. Pistillate spikelets enclosed within an involucre, typically in groups of 3 (1 fertile and 2 sterile).

A small genus of 4 or 5 species native to tropical Asia. *C. lacryma-jobi* (Job's tears) is a popular ornamental, its fruit-like involucres used in jewelry and rosaries. It has escaped from cultivation in the South.

TRIPSACUM. Gama grass. Robust, rhizomatous, caespitose perennials. Culms herbaceous, to 3+ m tall. Inflorescence a series of [1-] 2-several spike-like branches bearing pistillate spikelets on their lower portions and staminate ones above. Staminate spikelets 2-flowered, paired (1 sessile and 1 pedicellate) on one side of the rachis, disarticulating as a major segment of the inflorescence. Pistillate spikelets solitary and alternately inserted in hollow cavities of a thickened rachis, falling as separate bead-like units. Lodicules 2; stamens 0 or 3; stigmas 2.

A genus of 7-12 species native to warmer parts of the New World. *T. dactyloides* (eastern gama grass) is native to the eastern and central regions; two other species occur in Florida and Arizona.

ZEA. Maize, corn, Indian corn, teosinte. Robust annuals or perennials. Culms herbaceous, the internodes solid, to 5 m tall. Staminate spikelets

paired, unequally pedicellate, in terminal inflorescences consisting of spike-like branches. Glumes broad, thin; lemma and palea hyaline. Pistillate spikelets paired, in axillary inflorescences, either sunken in cavities of a hardened rachis or sessile on a thickened, almost woody axis, the "cob." Glumes broad, thin, rounded, and much shorter than the mature caryopsis; lemma and palea membranous and hyaline. Lodicules 0; stamens 0 or 3; stigmas 2.

A genus of 5 species native to the New World. As treated here, the genus includes *Euchlaena*. *Z. mays* ssp. *mays* (maize or corn), the only important cereal native to the New World, is unknown in the wild; *Z. mays* ssp. *mexicana* is teosinte [= *Euchlaena m.* in H & C], whose evolution is intertwined with that of maize; *Z. diploperennis* is a recently discovered perennial teosinte endemic to Mexico with great agronomic potential.

COMPARISON OF NEW WORLD "MAYDEAE"

Character	Zea	Euchlaena	Tripsacum
Growth form Number of tillers Leaf width Number leaf veins # tassel branches \$\foats\ \text{spklts. in tassel} \text{Pollen size} \text{Meiosis} \text{Lateral inflorescence} \text{Style length} \text{Styles} \text{Poitillate spikelets} \text{Rows } \polen \text{ spikelets} \text{Caryopses} \text{Rachis} # chromosome knobs	Annual Few Wide Few Intermediate No Large Early Enclosed Long Fused Paired Many Naked Solid Few	Annual Intermediate Intermediate Intermediate Many No Intermediate Intermediate Intermediate Enclosed Intermediate Fused Paired Two Enclosed Brittle Intermediate	Perennial Many Narrow Many Few* Yes Small Late Naked Short Separate Single Two Enclosed Brittle Many
Knob position X =	Internal 10	Internal & terminal 10	Terminal 9
Drought resistance	Susceptible	Intermediate	Resistant

* True of Tripsacum dactyloides

[Source: After Mangelsdorf & Reeves, 1939: 205]

SELECTED REFERENCES

Brown, W. V. 1958. Apomixis as related to geographical distribution in the panicoid grass tribes. J. S. African Bot. 24: 191-200.

Brown, W. V. & W. H. P. Emery. 1958. Apomixis in the Gramineae: Panicoideae. American J. Bot. 45: 253-263.

Esen, A. & K. W. Hilu. 1993. Promalin and immunological studies in the Poaceae. IV. Subfamily Panicoideae. Canadian J. Bot. 71: 315-322.

Giussani, L. M. et al. 2001. A molecular phylogeny of the grass subfamily Panicoideae (Poaceae) shows multiple origins of C_4 photosynthesis. American J. Bot. 88(11): 1993-2012.

Gómez-Martinez, R. & A. Culham. 2000. Phylogeny of the sub-family Panicoideae with emphasis on the tribe Paniceae: evidence from the *trn*L-F cpDNA region. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 136-140.

Quarin, C. 1992. The nature of apomixis and its origin in panicoid grasses. Apomixis Newsl. 5: 8-15.

TRIBE PANICEAE

Blake, S. T. 1959. New criteria for distinguishing genera allied to *Panicum* (Gramineae). Proc. Royal Soc. Queensland 70: 15-19.

Butzin, F. 1969. Die Begrannten Paniceae und ihre Stellung in System. Willdenowia 5: 245-270.

Butzin, F. 1970. Die systematische Gliederung der Paniceae. Willdenowia 6: 179-192.

Butzin, F. 1970. Die Blatternervatur der Paniceae in ihrer Bedeutung für die systematische Gliederung dieser Tribus. Willdenowia 6(1): 167-178.

Chandra, N. 1962. Morphological studies in the Gramineae. II. Vascular anatomy of the spikelets in the Paniceae. Proc. Indian Acad. Sci. Sect. B, 56: 217-231.

Chase, A. 1906. Notes on genera of Paniceae. I. Proc. Biol. Soc. Washington 29: 183-192.

Chase, A. 1908. Notes on genera of Paniceae. II. Proc. Biol. Soc. Washington 21: 1-10.

Chase, A. 1908. Notes on genera of Paniceae. III. Proc. Biol. Soc. Washington 21: 175-188.

Chase, A. 1911. Notes on genera of Paniceae. IV. Proc. Biol. Soc. Washington 24: 103-160.

Crins, W. J. 1992. The genera of Paniceae (Gramineae: Panicoideae) in the southeastern United States. J. Arnold Arbor. Suppl. Series 1: 171-312.

Rost, T. L. & A. D. Simper. 1975. The germination lid: a characteristic of the lemma in the Paniceae. Madroño 23: 68-72.

Webster, R. D., J. H. Kirkbride, & J. V. Reyna. 1989. New World genera of the Paniceae (Poaceae: Panicoideae). Sida 13(4): 393-416 + microcard.

Zuloaga, F. O., O. Morrone, & L. M. Giussani. 2000. A cladistic analysis of the Paniceae: a preliminary approach. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 123-135.

AMPHICARPUM

- Cheplick, G. P. & J. A. Quinn. 1982. *Amphicarpum purshii* and the "pessimistic strategy" in amphicarpic annuals with subter-ranean fruit. Oecologia 52: 327-332.
- Gray, J. R. & D. E. Fairbrothers. 1971. A clarification of some misconceptions about *Amphicarpum purshii* (Gramineae). Bull. Torrey Bot. Club 98(3): 174, 175.
- McNamara, J. & J. A. Quinn. 1977. Resource allocation and re-production in populations of *Amphicarpum purshii* (Gramineae). American J. Bot. 64: 17-23.

AXONOPUS

- Black, G. A. 1963. Grasses of the genus *Axonopus*. Adv. Front. Plant Sci. 5: 1-186.
- Hickenbick, M. C. M. et al. 1975. Cytogenetic and evolutionary relationships in the genus *Axonopus* (Gramineae). Cytologia 40: 185-204.

BRACHIARIA

- Chase, A. 1920. The North American species of *Brachiaria*. Contr. U. S. Natl. Herb. 22(1): 33-43.
- Morrone, O. & F. O. Zuloaga. 1992. Revision de las especies sudamericanas nativas e introducidas de los generos *Brachiaria* y *Urochloa* (Poaceae: Panicoideae: Paniceae). Darwiniana 31: 43-109.
- Thompson, R. A. & J. R. Estes. 1986. Anthecial and foliar micro-morphology and foliar anatomy of *Brachiaria* (Poaceae: Paniceae). American J. Bot. 73(3): 398-408.
- Veldekamp, J. F. 1996. *Brachiaria, Urochloa* (Gramineae: Paniceae) in Malesia. Blumea 41(2): 413-437.

CENCHRUS

- Caro, J. A. & E. Sanchez. 1967. Notas criticas sobre especies de *Cenchrus* (Gramineae). Kurtziana 4: 39-50.
- Chase, A. 1920. The North American species of *Cenchrus*. Contr. U. S. Natl. Herb. 22(1): 45-77.
- DeLisle, D. G. 1963. Taxonomy and distribution of the genus *Cenchrus*. Iowa State J. Sci. 37: 259-351.
- Nash, G. V. 1895. The genus *Cenchrus* in North America. Bull. Torrey Club 22: 298-302.
- Sohns, E. R. 1955. *Cenchrus* and *Pennisetum*: fascicle morphology. J. Washington Acad. Sci. 45(5): 135-143.

DIGITARIA

- Gould, F. W. 1963. Cytotaxonomy of *Digitaria* sanguinalis and *D. adscendens*. Brittonia 15: 241-244.
- Henrard, J. T. 1950. Monograph of the genus *Digitaria*. Univ. Pers Leiden. 999 pp.
- Hitchcock, A. S. 1927. The validity of the grass genus *Digitaria*. Rhodora 29: 114-116.

- Howell, J. T. 1971. Crabgrasses (*Digitaria*) in California. Wasmann J. Biol. 29(1): 101, 102.
- Nash, G. V. 1898. The genus *Syntherisma* in North America. Bull. Torrey Bot. Club 25: 289-303.
- Veldkamp, J. F. 1973. A revision of Digitaria Haller (Gramineae) in Malesia. Blumea 21: 1-80.
- Webster, R. D. 1987. Taxonomy of *Digitaria* section *Digitaria* in North America (Poaceae: Paniceae). Sida 12: 209-222.
- Webster, R. D. & S. L. Hatch. 1990. Taxonomy of *Digitaria* section *Aequiglumae* (Poaceae: Paniceae). Sida 14(2): 145-167.
- Wipff, J. K. 1990. A systematic study of Digitaria arenicola and D. cognata (Poaceae: Paniceae). M. S. thesis. Texas A & M Univ. College Station.
- Wipff, J. K. 1996. Nomenclatural combinations in *Digitaria* (Poaceae: Paniceae). Phytologia 80(5): 348, 349.
- Wipff, J. K. 2001. Nomenclatural change in the *Digitaria* cognata complex (Poaceae: Paniceae). Sida 19(4): 923, 924.
- Wipff, J. K. & S. L. Hatch. 1994. A systematic study of *Digitaria* sect. *Pennatae* (Poaceae: Paniceae) in the New World. Syst. Bot. 19(4): 613-627.

ECHINOCHLOA

- Ali, M. A. 1968. The *Echinochloa crusgalli* complex in the United States. Ph. D. dissertation. Texas A & M Univ. College Station. 99 pp.
- Gould, F. W., M. A. Ali, & D. E. Fairbrothers. 1972. A revision of *Echinochloa* in the United States. American Midl. Nat. 87: 36-59.
- Hitchcock, A. S. 1920. The North American species of *Echinochloa*. Contr. U. S. Natl. Herb. 22: 133-153.
- Norris, R. F. 1996. Morphological and phenological variation in barnyardgrass (*Echinochloa crus-galli*) in California. Weed Sci. 44: 804-814.
- Wiegand, K. M. 1921. The genus *Echinochloa* in North America. Rhodora 23: 49-65.
- Yabuno, T. 1962. Cytotaxonomic studies on the two cultivated species and the wild relatives in the genus *Echinochloa*. Cytologia 27: 296-305.
- Yabuno, T. 1966. Biosystematic study of the genus *Echinochloa*. Japanese J. Bot. 19: 277-323.

ERIOCHLOA

- Shaw, R. B. & F. E. Smeins. 1979. Epidermal characteristics of the callus in $\it Eriochloa$ (Poaceae). American J. Bot. 66(8): 907-913.
- Shaw, R. B. & F. E. Smeins. 1981. Some anatomical and morph-ological characteristics of the North American species of *Eriochloa* (Poaceae: Paniceae). Bot. Gaz. 142: 534-544.
- Shaw, R. B. & R. D. Webster. 1987. The genus *Eriochloa* (Poaceae: Paniceae) in North and Central America. Sida 12: 165-207.

LASIACIS

- Davidse, G. 1972. A systematic study of the genus *Lasiacis* (Gramineae, Panicoideae). Ph. D. dissertation. Iowa State Univ. Ames. 231 pp.
- Davidese, G. 1979. A systematic study of the genus *Lasiacis* (Gramineae: Paniceae). Ann. Missouri Bot. Gard. 65: 1133-1254.
- Hitchcock, A. S. 1920. The North American species of *Lasiacis*. Contr. U. S. Natl. Herb. 22: 13-31.

MELINIS

Zizka, G. 1988. Revision der Melinideae Hitchcock (Poaceae, Panicoideae). Biblio. Bot. 138: 1-149.

OPLISMENUS

- Davy, J. C. & W. D. Clayton. 1978. Some multiple discrimination function studies on *Oplismenus* (Gramineae). Kew Bull. 33(1): 147-158.
- Hitchcock, A. S. 1920. The North American species of *Oplismenus*. Contr. U. S. Natl. Herb. 22: 123-132.
- Kerguelen, M. 1976. *Oplismenus* Palisot de Beauvois versus *Orthopogon* R. Br. Taxon 25: 195, 196.
- Scholz, U. 1981. Monographie der Gattung *Oplismenus* (Gramineae). Phanerom. Monogr. 13: 1-218.

PANICUM

- Allred, K. W. & F. W. Gould. 1978. Geographic variation in the *Dichanthelium aciculare* complex (Poaceae). Brittonia 30(4): 497-504.
- Bacon, J. D. 1970. Taxonomy of the *Panicum oligosanthes* com-plex. Master of Science thesis. Texas A & M Univ. College Station.
- Blake, S. T. 1959. New criteria for distinguishing genera allied to *Panicum* (Gramineae). Proc. Royal Soc. Queensland 70: 15-19.
- Brown, W. V. & B. N. Smith. 1975. The genus $\it Dichanthelium$. Bull. Torrey Bot. Club 102: 10-13.
- Clark, C. F. A. 1977. Biosystematic studies of selected Texas species of *Dichanthelium* (Poaceae). Ph. D. dissertation. Texas A & M Univ. College Station. 131 pp.
- Clark, C. A. & F. W. Gould. 1975. Some epidermal characteristics of paleas of *Dichanthelium, Panicum*, and *Echinochloa*. American J. Bot. 62: 743-748.
- Clark, C. A. & F. W. Gould. 1978. *Dichanthelium* subgenus *Turfosa* (Poaceae). Brittonia 30: 54-59.
- Fernald, M. L. 1934. Realignments in the genus *Panicum*. Rhodora 36: 61-87.
- Freckmann, R. W. 1967. Taxonomic studies in *Panicum* subgenus *Dichanthelium*. Ph. D. dissertation. Iowa State Univ. Ames. 175 pp.
- Freckmann, R. W. 1978. New combinations in *Dichanthelium* (Poaceae). Phytologia 39: 268-272.
- Freckmann, R. W. 1981. Realignments in the *Dichanthelium acuminatum* complex (Poaceae). Phytologia 48: 99-110.

- Gould, F. W. 1980. The Mexican species of *Dichanthelium* (Poaceae). Brittonia 32(3): 353-364.
- Gould, F. W. & C. A. Clark. 1978. *Dichanthelium* (Poaceae) in the United States and Canada. Ann. Missouri Bot. Gard. 65: 1088-1132.
- Hansen, B. F. and R. P. Wunderlin. 1988. Synopsis of *Dichanthelium* (Poaceae) in Florida. Ann. Missouri Bot. Gard. 75(4): 1637-1657.
- Harvill, A. M., Jr. 1977. New combinations in *Dichanthelium* (Poaceae). Castanea 42(2): 177.
- Hitchcock, A. S. & A. Chase. 1910. The North American species of *Panicum*. Contr. U. S. Natl. Herb. 15: 1-396.
- Hitchcock, A. S. & A. Chase. 1915. Tropical North American species of *Panicum*. Contr. U. S. Natl. Herb. 17(6): 459-539.
- Hsu, C.-C. 1965. The classification of *Panicum* (Gramineae) and its allies, with special reference to the characters of lodicules, style-bases and lemma. J. Fac. Sci. Univ. Tokyo, Sec. III, 9: 43-150.
- Lamson-Scribner, F. & E. D. Merrill. 1901. The New England species of *Panicum*. Rhodora 3: 93-129.
- Le Blond, R. J. 2001. Taxonomy of the Dichotoma group of *Dichanthelium* (Poaceae). Sida 19(4): 821-837.
- LeLong, M. G. 1965. Studies of reproduction and variation in some *Panicum* subgenus *Dichanthelium*. Ph. D. dissertation. Iowa State Univ. Ames. 228 pp.
- LeLong, M. G. 1984. New combinations for *Panicum* and subgenus *Dichanthelium* (Poaceae) of the southeastern United States. Brittonia 36(3): 262-273.
- LeLong, M. G. 1986. A taxonomic treatment of the genus *Panicum* (Poaceae) in Mississippi. Phytologia 61: 251-269.
- Nash, G. V. 1899. The dichotomous *Panicums*: some new species. I. Bull. Torrey Bot. Club 26: 568-581.
- Ohsugi, R., T. Murata, & N. Chonan. 1982. C_4 syndrome of the species in the *Dichotomiflora* group of the genus *Panicum* (Gramineae). Bot. Mag. (Tokyo) 95: 339-347.
- Palmer, P. G. 1972. A biosystematic study of the Virgata group of *Panicum* (Gramineae) in the United States. Ph. D. dissertation. North Carolina State Univ. 69 pp.
- Palmer, P. G. 1975. A biosystematic study of the *Panicum amarum-P. amarulum* complex (Gramineae). Brittonia 27: 142-150.
- Pilger, R. 1931. Bemerkungen zu *Panicum* und verwandten Gattungen. Not. Bot. Gart. und Mus. Berlin-Dahlem 11: 237-247.
- Pizzolato, T. D. 1983. A three-dimensional reconstruction of the vascular system to the lodicules, androecium, and gynoecium of a fertile floret of *Panicum dichotomiflorum* (Gramineae). Amer. J. Bot. 70(8): 1173-1187.
- Renvoize, S. A. 1978. Studies in the Gramineae. XLIII. The genus *Panicum*, group *Lorea* (Gramineae). Kew Bull. 32: 419-428.
- Silveus, W. A. 1942. Grasses: classification and

description of species of *Paspalum* and *Panicum* in the United States. Publ. by the author. San Antonio, TX. 526 pp.

Spellenberg, R. W. 1968. Biosystematic studies in *Panicum* Group *Lanuginosum* from the Pacific Northwest. Ph. D. dissertation. Univ. Washington. Seattle. 218 pp.

Spellenberg, R. W. 1970. *Panicum shastense* (Gramineae), a sterile hybrid between *P. pacificum* and *P. scribnerianum*. Brittonia 22: 154-162.

Spellenberg, R. W. 1975. Autogamy and hybridization as evolutionary mechanisms in *Panicum* subgenus *Dichanthelium* (Gramineae). Brittonia 27: 87-95.

Spellenberg, R. W. 1975. Synthetic hybridization and taxonomy of western North American *Dichanthelium*, group *Lanuginosa* (Poaceae). Madroño 23: 134-153.

Stephenson, S. N. 1984. The genus *Dichanthelium* (Poaceae) in Michigan. Michigan Bot. 23: 107-119.

Waller, F. R. 1976. A biosystematic study of *Panicum* section *Diffusa* (Poaceae) in North America. Ph. D. dissertation. Texas A & M Univ. 132 pp.

Warmke, H. E. 1954. Apomixis in *Panicum maximum*. American J. Bot. 41: 5-11.

Zuloaga, F. O. 1987. Systematics of New World species of *Panicum* (Poaceae: Paniceae). <u>In</u>, Soderstrom, T. R. et al. (editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 287-306.

Zuloaga, F. O. 1993. Infrageneric phenetic analyses in New World *Panicum* (Poaceae: Panicoideae: Paniceae). Canadian J. Bot. 71(10): 1312-1327.

Zuloaga, F. O. & T. R. Soderstrom. 1985. Classification of the outlying species of New World *Panicum* (Poaceae: Paniceae). Smithsonian Contr. Bot. 59: 1-63.

Zuloaga, F. O., R. P. Ellis, & O. Morrone. 1993. A revision of Panicum subg. Dichanthelium sect. Dichanthelium (Poaceae: Panicoideae: Paniceae) in Mesoamerica, the West Indies, and South America. Ann. Missouri Bot. Gard. 80(1): 119-190.

Zuloaga, F. O. et al. 1998. Revisión y análisis cladistico de *Steinchisma*. Ann. Missouri Bot. Gard. 85(4): 631-656.

PASPALUM

Banks, D. J. 1963. Taxonomy of *Paspalum*, group *Setacea*. Ph. D. dissertation. Univ. Georgia. Athens.

Banks, D. J. 1966. Taxonomy of *Paspalum setaceum* (Gramineae). Sida 2: 269-284.

Chase, A. 1929. The North American species of *Paspalum*. Contr. U. S. Natl. Herb. 28: 1-310.

Fosberg, F. R. 1977. *Paspalum distichum* again. Taxon 26: 201, 202.

Morrone, O. et al. 1995. Revision del grupo Racemosa del genera *Paspalum* (Poaceae: Panicoideae: Paniceae). Ann. Missouri Bot. Gard. 82(1): 82-116.

PENNISETUM

Brunken, J. N. 1977. A systematic study of *Pennisetum*

sect. *Pennisetum* (Gramineae). American J. Bot. 64(2): 161-176.

Chase, A. 1921. The North American species of *Pennisetum*. Contr. U. S. Natl. Herb. 22(4): 209-234.

Husain, S. N. et al. 1990. Chemotaxonomic studies in the genus *Pennisetum* L. Rich. by isozyme studies. Beitr. Biol. Pflanzen 65(2): 211-222.

Kaushal, P. & J. S. Sidhu. 1993. Chemotaxonomic studies in *Pennisetum* species. Crop Improv. 20(1): 78-80

Rachie, K. O. & J. V. Majmudar. 1980. Pearl millet. The Pennsylvania State Univ. Press. University Park. 307 pp.

Schmelzer, G. H. 1997. Review of *Pennisetum* section *Brevival-vula* (Poaceae). Euphytica 97(1): 1-20.

Sohns, E. R. 1955. *Cenchrus* and *Pennisetum*: fascicle morphology. J. Washington Acad. Sci. 45(5): 135-143.

Wilvert, C. 1980. Kikiyu grass, an African invader. Pacific Hort. 41(3): 45-47.

Wipff, J. K. 2001. Nomenclatural changes in *Pennisetum* (Poaceae: Paniceae). Sida 19(3): 523-530.

SACCIOLEPIS

Judziewicz, E. J. 1990. A new South American species of *Sacciolepis* (Poaceae: Paniceae), with a summary of the genus in the New World. Syst. Bot. 15(3): 415-420.

Pohl, R. W. & N. R. Lersten. 1975. Stem aerenchyma as a character separating *Hymenachne* and *Sacciolepis* (Gramineae, Panicoideae). Brittonia 27(3): 223-227.

SETARIA

Chikara, Z. J. & P. K. Gupta. 1980. Numerical taxonomy in the genus *Setaria* (L.) Beauv. Proc. Indian Acad. Sciences: Plant Sci. 89: 401-406.

Clayton, W. D. 1979. Notes on *Setaria* (Gramineae). Kew Bull. 33(3): 501-509.

Emery, W. H. P. 1957. A cyto-taxonomic study of *Setaria macrostachya* (Gramineae) and its relatives in the southwest United States and Mexico. Bull. Torrey Bot. Club 84: 94-105.

Hitchcock, A. S. 1920. The North American species of *Chaetochloa*. Contr. U. S. Natl. Herb. 22: 155-208.

Pohl, R. W. 1962. Notes on *Setaria viridis* and *S. faberi* (Gramineae). Brittonia 14: 210-213.

Lamson-Scribner, F. & E. D. Merrill. 1900. The North American species of *Chasetochloa*. U. S. Dept. Agric. Div. of Agrostology. Bull. No. 21. 44 pp.

Reeder, J. R. 1951. *Setaria lutescens*, an untenable name. Rhodora 53: 27-30.

Rominger, J. M. 1962. Taxonomy of *Setaria* (Gramineae) in North America. Illinois Biol. Monogr. 29: 1-132.

Terrell, E. E. 1976. The correct names for pearl millet and yellow foxtail. Taxon 25(2): 297-304.

Veldekamp, J. F. 1994. Miscellaneous notes on southeast Asia Gramineae. IX. Setaria and Paspalidium. Blumea 39(1-2): 373-384.

Webster, R. D. 1993. Nomenclature of *Setaria* (Poaceae: Paniceae). Sida 15(3): 447-489.

Webster, R. D. 1995. Nomenclatural changes in *Setaria* and *Paspalidium* (Poaceae: Paniceae). Sida 16(3): 439-446.

Williams, R. D. & M. M. Schreiber. 1976. Numerical and chemotaxonomy of the green foxtail complex. Weed Sci. 24: 331-335.

STENOTAPHRUM

Busey, P. et al. 1982. Classification of St. Augustine grass. Crop Sci. 22(3): 469-473.

Sauer, J. D. 1972. Revision of *Stenotaphrum* (Gramineae: Paniceae) with attention to its historical geography. Brittonia 24: 202-222.

UROCHLOA

Morrone, O. & F. O. Zuloaga. 1992. Revision de las especies sudamericanas nativas e introducidas de los generos *Brachiaria y Urochloa* (Poaceae: Panicoideae: Paniceae). Darwinia 31: 43-109.

TRIBE ANDROPOGONEAE

Blake, S. T. 1969. Taxonomic and nomenclatural studies in the Gramineae. I. Proc. Royal Soc. Queensland 80: 55-84.

Clayton, W. D. 1972. Studies in the Gramineae. 31. The awned genera of Andropogoneae. Kew Bull. 27(3): 457-474.

Clayton, W. D. 1973. Studies in the Gramineae. 33. The awnless genera of Andropogoneae. Kew Bull. 28(1): 49-57.

Clayton, W. D. 1987. Andropogoneae. <u>In</u>, Soderstrom, T. R. et al. (Editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 307-309.

Grant, C. A. 1972. A scanning electron microscope survey of some Maydeae pollen. Grana 12: 177-184.

Harlan, J. R. et al. 1958. Studies on Old World bluestems. II. Oklahoma Agric. Exp. Sta. Bull. No. T-72.

Hartley, W. 1958. Studies on the origin, evolution, and distribution of the Gramineae. I. The tribe Andropogoneae. Australian J. Bot. 6: 116-128.

Kellogg, E. A. 2000. Molecular and morphological evolution in Andropogoneae. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 149-158.

Reeves, R. G. 1953. Comparative morphology of the American Maydeae. Texas Agric. Exp. Sta. Bull. 761. 26 pp.

Reeves, R. G. & P. C. Mangelsdorf. 1942. A proposed taxonomic change in the tribe Maydeae (family Gramineae). American J. Bot. 29(10): 815-817.

Renvoize, S. A. 1982. A survey of leaf-blade anatomy in grasses. I. Andropogoneae. Kew Bull. 37(2): 315-321.

Roberty, G. Monographie systématique des andropogonées du globe. Boissiera 9: 1-455.

Spangler, R. E. 2000. Andropogoneae systematics and generic limits in Sorghum. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 167-170.

Spangler, R. et al. 1999. Andropogoneae evolution and generic limits in *Sorghum* (Poaceae) using *ndh*F sequences. Syst. Bot. 24(2): 267-281.

ANDROPOGON

Campbell, C. S. 1980. Biosystematic studies in the *Andropogon virginicus* complex (Gramineae). Ph. D. dissertation. Harvard Univ. Cambridge, MA.

Campbell, C. S. 1982. Cleistogamy in *Andropogon* L. (Gramineae). American J. Bot. 69(10): 1625-1635.

Campbell, C. S. 1983. Systematics of the *Andropogon virginicus* complex (Gramineae). J. Arnold Arbor. 64: 171-254.

Campbell, C. S. 1983. Wind dispersal of some North American species of *Andropogon* (Gramineae). Rhodora 85(841): 65-72.

Gould, F. W. 1953. A cytotaxonomic study in the genus *Andropogon*. American J. Bot. 40: 297-306.

Gould, F. W. 1957. New *Andropogons* with a key to the native and naturalized species of section *Amphilophis* in the U. S. Madroño 14: 18-29.

Gould, F. W. 1959. The glume pit of *Andropogon barbinodis*. Brittonia 11: 182-187.

Gould, F. W. 1967. The grass genus *Andropogon* in the United States. Brittonia 19: 70-76.

Hilu, K. W. 1984. Leaf epidermis of *Andropogon* sect. *Lepto-pogon* (Poaceae) in North America. Syst. Bot. 9(2): 247-257.

Keeler, K. H. & G. A. Davis. 1999. Comparison of common ctytypes of *Andropogon gerardii* (Andropogoneae, Poaceae). American J. Bot. 86(7): 974-979.

Sorenson, J. C. 1977. *Andropogon virginicus* (broomsedge). Newsl. Hawaiian Bot. Soc. 16: 7-22.

ANTHEPHORA

Reeder, J. R. 1960. The systematic position of the grass genus *Anthephora*. Trans. American Microscop. Soc. 79: 211-218.

ARTHRAXON

Kiger, R. W. 1971. *Arthraxon hispidus* (Gramineae) in the United States: taxonomic and floristic status. Rhodora 73: 39-46.

Welzen, P. C. van. 1981. A taxonomic revision of the genus *Arthraxon* Beauv. (Gramineae). Blumea 27(1): 255-300.

Welzen, P. C. van. 1993. The phylogeny of *Arthraxon* P. Beauv. (Gramineae). Rheedea 3(2): 101-106.

BOTHRIOCHLOA

- Allred, K. W. 1979. Systematics of the *Bothriochloa* sacchar-oides complex (Poaceae). Ph. D. dissertation. Texas A & M Univ. College Station. 200 pp.
- Allred, K. W. & F. W. Gould. 1983. Systematics of the *Bothriochloa saccharoides* complex (Poaceae: Andropogoneae). Syst. Bot. 8(2): 168-184.
- Blake, S. T. 1969. Taxonomic and nomenclatural studies in the Gramineae. No. 1. [Bothriochloa]. Proc. Royal Soc. Queensland 80: 55-84.
- Celarier, R. P. & J. R. Harlan. 1958. The cytogeography of the *Bothriochloa ischaemum* complex. Gramineae. 1. Taxonomy and geographic distribution. J. Linnean Soc. Bot. 55: 755-759.
- de Wet, J. M. J. 1968. Biosystematics of the *Bothriochloa barbinodis* complex (Gramineae). American J. Bot. 55: 1246-1250.
- Gould, F. W. 1957. New North American Andropogons of subgenus *Amphilophis* and a key to those species occurring in the United States. Madroño 14: 18-29.
- Srivastava, A. K. 1980. Cytological investigations in the genus *Bothriochloa* (Gramineae). Acta Bot. Indica 8(2): 164-170.

COELORACHIS

Clayton, W. D. 1970. Studies in the Gramineae. Part 21. *Coelorachis* and *Rhytachne*. A study in numerical taxonomy. Kew Bull. 24(2): 309-314.

CYMBOPOGON

Soenarko, S. 1977. The genus *Cymbopogon* Sprengel (Gramineae). Reinwardtia 9: 225-375.

DICHANTHIUM

- Celarier, R. P. et al. 1958. Cytogeography of the *Dichanthium annulatum* complex. Brittonia 10: 59-72.
- de Wet, J. M. J. 1965. Diploid races of tetraploid *Dichanthium* species. American Nat. 99: 167-171.
- de Wet, J. M. J. & J. R. Harlan. 1968. Taxonomy of *Dichanthium*, sect. *Dichanthium* (Gramineae). Bol. Soc. Arg. Bot. 12(1): 206-227.
- de Wet, J. M. J. & J. R. Harlan. 1970. Apomixis, polyploidy, and speciation in *Dichanthium*. Evol. 24: 270-277
- Mehr, K. L., R. P. Celarier, & J. R. Harlan. 1959. Effects of environment on selected morphological characters in the *Dichanthium annulatum* complex. Proc. Oklahoma Acad. Sci. 40: 10-14.
- Singh, A. P. & K. L. Hehra. 1965. Cytological analysis of hybrids within the *Dichanthium annulatum* complex. Cytologia 30: 307-316.
- Singh, A. P. 1965. Intergeneric cross of *Dichanthium annulatum* with *Bothriochloa ischaemum*. Cytologia 30: 54-57.

EREMOCHLOA

Buitenhuis, A. G. & J. F. Veldkamp. 2001. Revision of

Eremochloa (Gramineae-Andropogoneae-Rottboelliinae). Blumea 46(2): 399-420.

HEMARTHRIA

Heuvel, E. van den & J. F. Veldkamp. 2000. Revision of *Hemarthria* (Gramineae-Andropogoneae-Rottboelliinae). Blumea 45(2): 443-475.

HETEROPOGON

- Emery, W. H. P. & W. V. Brown. 1958. Apomixis in the Gramineae: tribe Andropogoneae: *Heteropogon contortus*. Madroño 14: 238-246.
- Tothill, J. C. 1968. Variation and apomixis in *Heteropogon contortus*, Gramineae. Bol. Soc. Argentine Bot. 12: 188-201.

IMPERATA

Gabel, M. L. 1982. A biosystematic study of the genus *Imperata* (Gramineae: Andropogoneae). Ph. D. dissertation. Iowa State Univ. Ames.

MICROSTEGIUM

Fairbrothers, D. E. & J. R. Gray. 1972. *Microstegium vimineum* (Trin.) A. Camus (Gramineae) in the United States. Bull. Torrey Bot. Club 99: 97-100.

MISCANTHUS

- Freckmann, R. W. 1973. *Miscanthus* in Wisconsin. Newsletter Bot. Club Wisconsin 5: 11-16.
- Lee, Y. 1964. Taxonomic studies on the genus *Miscanthus*. 2. Anatomical patterns of leaves. Bot. Mag. Tokyo 77: 122-130.
- Lee, Y. 1964. Taxonomic studies on the genus *Miscanthus*. 4. Relationships among the section, subsection, and species. J. Japanese Bot. 39: 115-123. See also 196-204; 257-265; 289-298.

ROTTBOELLIA

Veldekamp, J. F. et al. 1986. Generic delimitation of *Rottboellia* and related genera. Blumea 31: 281-307.

SACCHARUM

- Artschwager, E. & E. W. Brandes. 1958. Sugarcane (Saccharum officinarum L.): origin, characteristics, and descriptions of representative clones. U. S. Dept. Agric. Handbook No. 122. 307 pp.
- Burner, D. M. & R. D. Webster. 1994. Cytological studies on North American species of *Saccharum* (Poaceae: Andropogoneae). Sida 16(2): 233-244.
- Daniels, J. et al. 1975. The origin of the genus *Saccharum*. Sugarcane Breed. Newsl. 36: 24-39.
- Grassl, C. O. 1968. Saccharum names and their interpreta-tion. I. S. S. C. T. Proc. $13^{\rm th}$ Congr. Taiwan. Pp. 868-875.
- Mukherjee, S. K. 1954. Revision of the genus *Saccharum*. Bull. Bot. Soc. Bengal 8: 143-148. Mukherjee, S. K. 1957. Origin and distribution of *Saccharum*. Bot. Gaz. 119: 55-61.

- Mukherjee, S. K. 1957. Origin and distribution of *Saccharum*. Bot. Gaz. 119: 55-61.
- Mukherjee, S. K. 1958. Revision of the genus *Erianthus* Michx. (Gramineae). Lloydia 21: 157-188.
- Nair, N. V. et al. 1999. Analysis of genetic diversity and phylo-geny in *Saccharum* and related genera using RAPD markers. Genet. Resources & Crop Evol. 46(1): 73-79.
- Renvoize, S. A. 1978. Studies in *Elionurus* (Gramineae). Kew Bull. 32: 665-672.
- Webster, R. D. & R. B. Shaw. 1995. Taxonomy of the native North American species of *Saccharum* (Poaceae: Andropogoneae). Sida 16(3): 551-580.
- Whalen, M. D. 1991. Taxonomy of *Saccharum* (Poaceae). Baileya 23(3): 109-125.

SCHIZACHYRIUM

- Carman, J. G. & S. L. Hatch. 1982. Aposporous apomixis in *Schizachyrium* (Poaceae: Andropogoneae). Crop Science 22: 1252-1255.
- Gandhi, K. N. 1989. A biosystematic study of the *Schizachyrium scoparium* complex. Ph. D. dissertation. Texas A & M Univ. College Station. 188 pp.
- Gandhi, K. N. & F. E. Smeins. 1996. Four new combinations and one neotypification in *Schizachyrium* (Poaceae). Harvard Pap. Bot. 8: 67, 68.
- Hatch, S. L. 1978. Nomenclatural changes in *Schizachyrium* (Poaceae). Brittonia 30(4): 496.
- Hatch, S. L. 1975. A biosystematic study of the *Schizachyrium cirratum-Schizachyrium sanguineum* complex. Ph. D. dissertation. Texas A & M Univ. College Station. 123 pp.

SORGHASTRUM

- Davila, P. D. 1988. Systematic revision of the genus *Sorghastrum* (Poaceae: Andropogoneae). Ph. D. dissertation. Iowa State Univ. Ames. 333 pp.
- Hall, D. W. 1982. *Sorghastrum* (Poaceae) in Florida. Sida 9: 302-308.

SORGHUM

- Celarier, R. P. 1958. Cytotaxonomic notes on the subsection *Halepensia* of the genus *Sorghum*. Bull. Torrey Bot. Club 85(1): 49-62.
- Celarier, R. P. 1959. Cytotaxonomy of the Andropogoneae. III. Subtribe Sorgheae, genus *Sorghum*. Cytologia 23(4): 395-418.
- DeWet, J. M. J. 1978. Systematics and evolution of *Sorghum*, sect. *Sorghum* (Gramineae). American J. Bot. 85: 477-484.
- DeWet, J. M. J. & J. P. Huckabay. 1967. The origin of *Sorghum bicolor*. II. Distribution and domestication. Evolution 21: 787-802.
- DeWet, J. M. J., J. R. Harlan, & E. G. Price. Origin of variability in the spontanea complex of *Sorghum bicolor*. American J. Bot. 57: 704-707.
- Garber, E. D. 1950. Cytotaxonomic studies in the genus

- Sorghum. Univ. California Publ. Bot. 23: 283-362.
- Harlan, J. & J. M. J. de Wet. 1972. Simplified classification of cultivated sorghum. Crop Sci. 12(2): 172-176.
- Long, B. 1930. Spikelets of Johnson grass and Sudan grass. Bot. Gaz. 89: 154-168.
- Martin, J. H. 1970. History and classification of sorghum (*Sorghum bicolor*). <u>In</u>, Wall, J. S. & W. M. Ross (editors). Sorghum production and utilization. AVI Publ. Westport, CT. Pp. 1-27.
- Mouftah, S. P. & J. D. Smith. 1968. Cytological studies on the origin of Johnson grass. Canadian J. Genet. Cytol. 11: 25-29.
- Shechter, V. 1975. Biochemical-systematics studies in *Sorghum bicolor*. Bull. Torrey Bot. Club 102: 334-339.
- Snowden, J. D. 1936. The cultivated races of sorghum. Adlard & Son, Ltd. London, England. 274 pp.
- Snowden, J. D. 1955. The wild fodder sorghums of the section Eusorghum. J. Linn. Soc. Bot. 55: 191-260.
- Spangler, R. E. 2000. Andropogoneae systematics and generic limits in Sorghum. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 167-170.
- Spangler, R. et al. 1999. Andropogoneae evolution and generic limits in *Sorghum* (Poaceae) using *ndh*F sequences. Syst. Bot. 24(2): 267-281.

THEMEDA

- Reese, W. D. & G. P. Landry. 1985. *Themeda quadrivalvis* (L.) Kuntze (Poaceae) in Louisiana. Sida 11(1): 99-102.
- Woodland, P. S. 1964. The floral morphology and embryology of *Themeda australis*. Australian J. Bot. 12: 157-172.

TRIPSACUM

- Brink, D. & J. M. J. de Wet. 1983. Supraspecific groups in *Tripsacum* (Gramineae). Syst. Bot. 8(3): 243-249.
- Burson, B. L. et al. 1990. Apomixis and sexuality in eastern gramagrass. Crop. Sci. 30: 86-89.
- Cutler, H. & E. Anderson. 1941. A preliminary survey of the genus *Tripsacum*. Ann. Missouri Bot. Gard. 28: 249-269
- de Wet, J. M. J., J. R. Gray, & J. R. Harlan. 1976. Systematics of *Tripsacum* (Gramineae). Phytologia 33(3): 203-227.
- de Wet, J. M. J. et al. 1981. Systematics of South American *Tripsacum* (Gramineae). American J. Bot. 68(2): 269-276.
- de Wet, J. M. J., J. R. Harlan, & D. E. Brink. 1982. Systematics of *Tripsacum dactyloides* (Gramineae). American J. Bot. 69(8): 1251-1257.
- de Wet, J. M. J., D. E. Brink, & C. E. Cohen. 1983. Systematics of *Tripsacum* section *Fasciculata* (Gramineae). American J. Bot. 70(8): 1139-1146.

- de Wet, J. M. J. et al. 1983. Origin of *Tripsacum andersonii* (Gramineae). American J. Bot. 70(5): 706-711.
- Farquharson, L. J. 1957. Hybridization in *Tripsacum* and *Zea.* J. Heredity 48: 295-299.
- Galinat, W. C., R. S. K. Chaganti, & F. D. Hager.1964. *Tripsacum* as a possible amphiploid of wild maize and *Manisurus*. Bot. Mus. Leaflts. Harvard Univ. 20: 289-316.
- Galinat, W. C. & F. Craighead. 1964. Some observations on the dissemination of *Tripsacum*. Rhodora 66: 371-374.
- Gray, J. R. 1974. The genus *Tripsacum* L. (Gramineae): taxonomy and chemosystematics. Ph. D. dissertation. Univ. Illinois at Urbana-Champagne. 195 pp.
- Hitchcock, A. S. 1906. Notes on North American grasses. VI. Synopsis of *Tripsacum*. Bot. Gaz. 41: 294-298.
- Maguire, M. P. 1961. Divergence in *Tripsacum* and *Zea* chromosomes. Evol. 15: 394-400.
- Newell, C. A. & J. M. J. de Wet. 1974. Morphological and cytological variability in *Tripsacum dactyloides* (Gramineae). American J. Bot. 61(6): 652-664.
- Newell, C. A. & J. M. J. de Wet. 1974. Morphology and some maize-*Tripsacum* hybrids. American J. Bot. 61: 45-53.
- Smith, J. et al. 1980. Biochemical systematics and evolution of *Zea, Tripsacum*, and related genera. Econ. Bot. 34(3): 201-218.
- Stalker, H. T., J. R. Harlan, & J. M. J. de Wet. 1977. Observations on introgression of *Tripsacum* into maize. American J. Bot. 64: 1162-1169.
- Talbert, L. E. et al. 1990. *Tripsacum andersonii* is a natural hybrid involving *Zea* and *Tripsacum*: molecular evidence. American J. Bot. 77(6): 722-726.

VETIVERIA

- Maffei, M. 2002. Vetiveria: the genus *Vetiveria*. Taylor & Francis Books. New York, NY. 191 pp.
- Molla, H. A. & D. C. Pal. 1992. Observations on *Vetiveria zizanioides* (L.) Nash (khas-khas). J. Econ. Taxon. Bot. Add. Ser. 10: 101-103.

ZEA

- Alava, R. O. 1952. Spikelet variation in *Zea mays* L. Ann. Missouri Bot. Gard. 39: 65-96.
- Anderson, E. 1944. Homologies of the ear and tassel in *Zea mays*. Ann. Missouri Bot. Gard. 31: 325-342.
- Beadle, G. W. 1939. Teosinte and the origin of maize. J. Heredity 30: 245-247.
- Benz, B. 1986. Taxonomy and evolution of Mexican maize. Ph. D. dissertation. Univ. Wisconsin. Madison. 466 pp.
- Bird, R. M. 1978. A name change for Central American teosinte. Taxon 27(4): 361-366.
- Buckler, E. S. & T. P. Holtsford. 1996. Zea systematics: ribosomal ITS evidence. Mol. Biol. Evol. 13(4): 612-622.

- Collins, G. N. 1921. Teosinte in Mexico. J. Heredity 12: 339-350.
- Collins, G. N. 1930. The phylogeny of maize. Bull. Torrey Bot. Club 57(4): 199-210.
- Culotta, E. 1991. How many genes had to change to produce corn? Science 252: 1792, 1793.
- Cutler, H. C. & M. C. Cutler. 1948. Studies on the structure of maize plants. Ann. Missouri Bot. Gard. 35: 301-316.
- de Wet, J. M. J., C. A. Newell, & D. E. Brink. 1984. Counterfeit hybrids between *Tripsacum* and *Zea*. American J. Bot. 71: 245-251.
- de Wet, J. M. J., J. R. Harlan, & C. A. Grant. 1971. Origin and evolution of teosinte (*Zea mexicana* (Schrad.) Kuntze). Euphytica 20(2): 255-265.
- de Wet, J. M. J., J. R. Harlan, & A. V. Randrianasolo. 1978. Morphology of teosintoid and trip-sacoid maize (*Zea mays* L.). American J. Bot. 65(7): 741-747.
- Doebley, J. F. 1984. Maize introgression into teosinte a reappraisal. Ann. Missouri Bot. Gard. 71: 1100-1113.
- Doebley, J. F. 1983. The maize and teosinte male inflorescence: a numerical taxonomic study. Ann. Missouri Bot. Gard. 70: 32-70.
- Doebley, J. F. 1984. The maize and teosinte male inflorescence. A numerical taxonomic study. Ann. Missouri Bot. Gard. 70: 32-70.
- Doebley, J. F. 1985. Maize introgression into teosinte. Ann. Missouri Bot. Gard. 71: 1100-1113.
- Doebley, J. 1990. Molecular systematics of *Zea* (Gramineae). Maydica 35(2): 143-150.
- Doebley, J. F. & H. H. Iltis. 1980. Taxonomy of *Zea* (Gramineae). I. A subgeneric classification with key to taxa. American J. Bot. 67(6): 982-993.
- Dorweiler, J. et al 1993. Teosinte glume architecture 1: a genetic locus controlling a key step in maize evolution. Science 262: 233-235.
- Eubanks, M. W. 2001. The mysterious origin of maize. Econ. Bot. 55: 492-514.
- Fussell, B. 1994. The story of corn. A. A. Knopf. New York, NY. 356 pp.
- Galinat, W. C. 1973. Intergenomic mapping of maize, teosinte, and *Tripsacum*. Evolution 27: 644-655.
- Galinat, W. C. 1992. Evolution of corn. Adv. Agron. 47: 203-231.
- Gaut, B. S. 1996. Evolution in the genus Zea: lessons from studies of nucleotide polymorphism. Pl. Sp. Biol. 11(1): 1-11.
- Gould, S. J. 1984. A short way to corn. Nat. Hist. 93(3): 12-20.
- Guzman Mejia, R. 1978. Redescubrimiento de *Zea perennis* (Gramineae). Phytologia 38(3): 177.
- Hitchcock, A. S. 1922. A perennial species of teosinte. J. Washington Acad. Sci. 12: 205-208.

- Iltis, H. H. 1972. The taxonomy of *Zea mays* (Gramineae). Phytologia 23(2): 248, 249.
- Iltis, H. H. 1983. From teosinte to maize: the catastrophic sexual transmutation. Science 222: 886-894.
- Iltis, H. H. & J. F. Doebley. 1980. Taxonomy of *Zea* (Gramineae). II. Subspecific categories in the *Zea mays* complex and a generic synopsis. American J. Bot. 67(6): 994-1004.
- Iltis, H. H. & J. F. Doebley. 1984. Zea -- a biosystematic odyssey. <u>In</u>, Grant, W. F. (editor). Plant biosystematics. Academic Press. New York, NY. Pp. 587-616.
- Iltis, H. H. et al. 1979. Zea diploperennis (Gramineae): a new teosinte from Mexico. Science 203: 186-188.
- Irwin, H. & E. S. Barghoorn. 1965. Identification of the pollen of maize, teosinte, and *Tripsacum* by phase contrast microscopy. Bot. Mus. Leaflts. Harvard Univ. 21: 37-58.
- Kellogg, E. A. & J. A. Birchler. 1993. Linking phylogeny and genetics: *Zea mays* as a tool for phylogenetic studies. Syst. Biol. 42: 415-439.
- Mangelsdorf, P. C. 1945. The origin and nature of the ear of maize. Bot. Mus. Leaflts. Harvard Univ. 12: 33-88.
- Mangelsdorf, P. C. 1958. The mutgenic effect of hybridizing maize and teosinte. Cold Spring Harbor Symposium on Quantitative Biology 23: 409-421.
- Mangelsdorf, P. C. 1986. The origin of corn. Sci. American 255(2): 80-86.
- Newell, C. A. & J. M. J. de Wet. 1974. Morphology of some maize-*Tripsacum* hybrids. American J. Bot. 61: 45-53.
- Nickerson, N. H. 1954. Morphological analysis of the maize ear. American J. Bot. 41: 87-92.
- Peterson, P. A. 1976. Gene expression and the evolution of unisexuality in the maize spikelet. Maydica 21: 157-164.
- Provan, J. et al. 1999. Analysis of the genus *Zea* (Poaceae) using polymorphic chloroplast simple sequence repeats. Plant Syst. Evol. 218(3-4): 245-256.
- Reeves, R. G. 1944. Chromosome knobs in relation to the origin of maize. Genetics 29: 141-147.
- Reeves, R. G. 1950. Morphology of the ear and tassel of maize. American J. Bot. 37: 697-704.
- Rhoades, M. M. 1984. The early years of maize genetics. Ann. Rev. Genetics 18: 1-29.
- Rhoades, M. M. 1986. Barbara McClintock: an appreciation. Maydica 31(1): 1-4.
- Smith, J., C. Stephen, & R. N. Lester. 1980. Biochemical systematics and evolution of *Zea, Tripsacum* and related genera. Econ. Bot. 34(3): 201-218.
- Weatherwax, P. 1916. Morphology of the flowers of *Zea mays*. Bull. Torrey Bot. Club 43: 127-144.
- Weatherwax, P. 1917. The development of the spikelets of *Zea mays*. Bull. Torrey Bot. Club 44: 483-496.

- Weatherwax, P. 1929. The morphological nature of teopod corn. J. Heredity 20: 325-330.
- Weatherwax, P. 1935. The phylogeny of *Zea mays*. American Midl. Nat. 16: 1-71.
- Westerberg, A. & J. F. Doebley. 2002. Morphological traits defining species differences in wild relatives of maize are controlled by multiple quantitative trait loci. Evolution 56: 273-283.
- Wilkes, H. G. 1985. Teosinte, the closest relative of maize. The Bussey Institution. Harvard Univ. 159 pp.

SECTION 4 - AN OVERVIEW OF U. S. GRASSES

4.01 - STATISTICAL SUMMARY

Source	Subfamilies	Tribes	Genera	Species
Vasey (1883)	2	13	114	687
Hitchcock (1935)	2	14	159	1100
H & C (1951)	2	14	169	1398
Gould (1983)	6	22	124	1053
Smith (2002)	10	27	204	1405

4.02 - SUBFAMILIES, TRIBES, & GENERA OF CONTINENTAL NORTH AMERICA

SUBFAMILY: ARISTIDOIDEAE

Tribe: Aristideae Aristida

SUBFAMILY: ARUNDINOIDEAE

Tribe: Arundineae
Arundo
Gynerium
Hakonechloa
Molinia
Phragmites
Thysanolaena

SUBFAMILY: BAMBUSOIDEAE

Tribe: Bambuseae Arundinaria Bambusa Phyllostachys Pleioblastus Pseudosasa

Tribe: Olyreae Olyra

SUBFAMILY: CENTOTHECOIDEAE

Tribe: Centotheceae Chasmanthium

SUBFAMILY: CHLORIDOIDEAE

Tribe: Cynodonteae Aegopogon Bouteloua Buchloë Chloris Ctenium Cynodon Enteropogon Eustachys Gymnopogon Hilaria Microchloa Opizia Schedonnardus Spartina Tragus WilĬkommia Zoysia

Tribe: Eragrostideae
Acrachne
Allolepis
Blepharidachne
Blepharoneuron
Calamovilfa
Cladoraphis

Dinebra Distichlis Eleusine Eragrostis Erioneuron Fingerhuthia Leptochloa Lycurus Monanthochloë Monroa Muhlenbergia Neyraudia Pogonarthria Redfieldia Scleropogon Sporobolus Śwallenia Trichoneura Tridens Triplasis Tripogon Triraphis Uniola Vaseyochloa

Dactyloctenium

Tribe: Orcuttieae Neostapfia Orcuttia Tuctoria

Tribe: Pappophoreae
Cottea
Enneapogon

Crypsis

Pappophorum

SUBFAMILY: DANTHONIOIDEAE

Tribe: Danthonieae Cortaderia

Danthonia Rytidosperma

Schismus

SUBFAMILY: EHRHARTOIDEAE

Tribe: Ehrharteae Ehrharta

Tribe: Oryzeae

Leersia Luziola Oryza Zizania Zizaniopsis

SUBFAMILY: PANICOIDEAE

Tribe: Andropogoneae

Andropogon Apluda Arthraxon Bothriochloa Chrysopogon

Coix Coelorachis Cymbopogon Dichanthium Elionurus Eremochloa Euclasta Hackelochloa Hemarthria

Heteropogon Hyparrhenia *Imperata* Ischaemum Microstegium Miscanthus **Polytrias** Rottboellia Saccharum Schizachyrium

Sorghastrum Sorghum

Themeda Trachypogon

Tripsacum Zea

Tribe: Paniceae

Alloteropsis *Amphicarpum*

Anthaenantia Anthephora Axonopus

Brachiaria Cenchrus Digitaria Echinochloa Eriochloa

Hymenachne Lasiacis

Melinis Oplismenus Panicum Paspalum Pennisetum Reimarochloa Sacciolepis Setaria Setariopsis Stenotaphrum *Urochloa*

SUBFAMILY: PHAROIDEAE

Tribe: Phareae Pharus

SUBFAMILY: POÖIDEAE

Tribe: Ampelodesmeae

Ampelodesmos

Tribe: Aveneae Agropogon Agrostis

Aĭra Alopecurus Ammophila Anthoxanthum

Apera

Arrhenatherum

Avena

Beckmannia Calamagrostis Calammophila

Cinna

Corynephorus Deschampsia Dissanthelium Gastridium Gaudinia Helictotrichon Holcus

Koeleria Lagurus Limnodea Mibora Milium Phalaris Phleum Polypogon Sphenopholis Trisetum Ventenata

Tribe: Brachypodieae

Brachypodium

Tribe: Brachyelytreae Brachyelytrum

Tribe: Bromeae **Bromus**

Tribe: Diarrheneae

Diarrhena

Tribe: Hainardieae Hainardia Parapholis Scribneria

Tribe: Meliceae

Catabrosa Glyceria Mélica Pleuropogon Schizachne

Tribe: Nardeae Nardus

Tribe: Poeae

Arctagrostis Arctophila Briza Coleanthus Cutandia Cynosurus Ďactylis Desmazeria Dupontia Elyhordeum Eremopoa Festuca Festulolium Lamarckia Lolium Phippsia Poa Puccinellia Sclerochloa Scolochloa Torreyochloa

Tribe: Stipeae Oryzopsis Piptochaetium Ptilagrostis Stipa

Tribe: Triticeae Aegilops Agropyron Dasypyrum Elymus Eremopyrum Hordeum Secale **Triticum**

Revised: 10 January 2005

4.03 - GRASS GENERA: A CHRONOLOGY

	THE :	18 [™] CENTURY	1794 1794	Jarava H. Ruiz Lopez & José Antonio Pavón Lamarckia Conrad Moench
1753 1753	Aegilops Agrostis	Carolus Linnaeus Carolus Linnaeus	1794 1796	Sorghum Conrad Moench Dichanthium Pierre Remi Willemet
1753 1753	Aira Alopecurus	Carolus Linnaeus Carolus Linnaeus	1798	Polypogon Réné Louiche Desfontaine
1753 1753 1753	Andropogon Anthoxanthun	Carolus Linnaeus		THE 19 [™] CENTURY
1753	Aristida	Carolus Linnaeus	1801	Zoysia Carl Ludwig von Willdenow
1753 1753	Arundo	Carolus Linnaeus Carolus Linnaeus	1802 1803	Ventenata Georg Ludwig Koeler Arundinaria Andre Michaux
1753	Avena Briza	Carolus Linnaeus	1803	Erianthus Andre Michaux
1753	Bromus	Carolus Linnaeus	1803	Oryzopsis Andre Michaux
1753 1753	Cenchrus	Carolus Linnaeus Carolus Linnaeus	1805 1805	Beckmannia Nicolaus Thomas Host Bouteloua Mariano Lagasca y Segura
1753	Cinna Coix	Carolus Linnaeus	1805	Bouteloua Mariano Lagasca y Segura Cynodon Louis Claude Marie Richard
1753	Cynosurus	Carolus Linnaeus	1805	<i>Danthonia</i> Augustin Pyramus De Candolle
1753	Dactylis	Carolus Linnaeus	1805	Koeleria Christian Hendrick Persoon
1753 1753	Elymus Festuca	Carolus Linnaeus Carolus Linnaeus	1805 1805	Pennisetum Louis Claude Richard Trisetum Christian Hendrick Persoon
1753	Holcus	Carolus Linnaeus	1805	Vulpia Johann Georg Gmelin
1753	Hordeum	Carolus Linnaeus	1806	Aegopogon Carl Ludwig von Willdenow
1753 1753	Lagurus	Carolus Linnaeus Carolus Linnaeus	1806 1807	Elionurus Karl Sigismund Kunth Heleochloa Nicolaus Thomas Host
1753	Lolium Melica	Carolus Linnaeus	1807	Heteropogon Christian Hendrick Persoon
1753	Milium	Carolus Linnaeus	1809	Ammophila Nicolaus Thomas Host
1753	Nardus	Carolus Linnaeus	1809	Dactyloctenium Carl Ludwig von Willdenow
1753 1753	Oryza Panicum	Carolus Linnaeus Carolus Linnaeus	1810 1810	Chondrosum Auguste Nicdise Desvaux Elytrigia Auguste Nicdise Desvaux
1753	Phalaris	Carolus Linnaeus	1810	Eustachys Auguste Nicdise Desvaux
1753	Phleum	Carolus Linnaeus	1810	Glyceria Robert Brown
1753 1753	Poa Saccharum	Carolus Linnaeus Carolus Linnaeus	1810 1810	Hemarthria Robert Brown Hierochloe Robert Brown
1753	Secale	Carolus Linnaeus	1810	Microchloa Robert Brown
1753	Stipa	Carolus Linnaeus	1810	Oplismenus A. M. F. J. Palisot de Beauvois
1753	Triticum	Carolus Linnaeus	1810	Sporobolus Robert Brown
1753 1753	Uniola Zea	Carolus Linnaeus Carolus Linnaeus	1812 1812	Achnatherum A. M. F. J. Palisot de Beauvois Anthaenantia A. M. F. J. Palisot de Beauvois
1753	Zizania	Carolus Linnaeus	1812	ArrhenatherumA. M. F. J. Palisot de Beauvois
1756	Pharus	Patrick Browne	1812	Arthraxon A. M. F. J. Palisot de Beauvois
1759 1759	Olyra Paspalum	Carolus Linnaeus Carolus Linnaeus	1812 1812	Axonopus A. M. F. J. Palisot de Beauvois Brachyelytrum A. M. F. J. Palisot de Beauvois
1759	Tripsacum	Carolus Linnaeus	1812	Brachypodium A. M. F. J. Palisot de Beauvois
1763	Apera	Michel Adanson	1812	Catabrosa A. M. F. J. Palisot de Beauvois
1763 1763	Calamagrostis Mibora	Michel Adanson Michel Adanson	1812 1812	Ceratochloa A. M. F. J. Palisot de Beauvois Corynephorus A. M. F. J. Palisot de Beauvois
1763	Phragmites	Michel Adanson	1812	Deschampsia A. M. F. J. Palisot de Beauvois
1768	Digitaria	Victor Albrecht von Haller	1812	Diarrhena A. M. F. J. Palisot de Beauvois
1768 1770	Tragus	Victor Albrecht von Haller Joseph Gaertner	1812 1812	Diplachne A. M. F. J. Palisot de Beauvois Echinochloa A. M. F. J. Palisot de Beauvois
1771	Agropyron Manisuris	Carolus Linnaeus	1812	Enneapogon A. M. F. J. Palisot de Beauvois
1775	Themeda	Pehr Forsskal	1812	Gastridium A. M. F. J. Palisot de Beauvois
1776 1776	Eragrostis	Nathanael Mattaeus von Wolf Nathanael Matthaeus von Wolf	1812	Gaudinia A. M. F. J. Palisot de Beauvois
1779	Phararoides Anthephora	J. C. D. von Schreber	1812 1812	Gymnopogon A. M. F. J. Palisot de Beauvois Gynerium A. M. F. J. Palisot de Beauvois
1779	Ehrharta	Carl Peter Thunberg	1812	Hymenachne A. M. F. J. Palisot de Beauvois
1779	Rottboellia	Carolus Linnaeus f.	1812	Leptochloa A. M. F. J. Palisot de Beauvois
1788 1788	Chloris Eleusine	Olof Swartz Joseph Gaertner	1812 1812	Melinis A. M. F. J. Palisot de Beauvois Monerma A. M. F. J. Palisot de Beauvois
1788	Leersia	Olof Peter Swartz	1812	Piptatherum A. M. F. J. Palisot de Beauvois
1789	Bambusa	Johann C. D. von Schreber	1812	Schedonorus A. M. F. J. Palisot de Beauvois
1789 1789	Crypsis Luziola	William Aiton Antoine Laurent de Jussieu	1812 1812	Schismus A. M. F. J. Palisot de Beauvois Sclerochloa A. M. F. J. Palisot de Beauvois
1789	Molinia –	Franz von Paula von Schrank	1812	Setaria A. M. F. J. Palisot de Beauvois
1789	Muhlenbergia	Johann C. D. von Schreber	1812	Triplasis A. M. F. J. Palisot de Beauvois
1789 1791	Spartina Pannonhorum	Johann C. D. von Schreber	1812 1813	Urochloa A. M. F. J. Palisot de Beauvois Ctenium Georg Wolfgang Franz Panzer
1791	Pappophorum Imperata	Johann C. D. von Schreber Domenico Maria Leone Cirillo	1815	Ctenium Georg Wolfgang Franz Panzer Cymbopogon Curt Polycarp Joachin Sprengel
1794	Hystrix	Conrad Moench	1816	Eriochloa Karl Sigismund Kunth

1816			
	Hilaria Karl Sigismund Kunth	1886	Scribneria Eduard Hackel
1816	Lycurus Karl Sigismund Kunth	1887	Blepharidachne Eduard Hackel
1817	Coleanthus Johann Heinrich Seidel	1887	Cladoraphis Adrien René Franchet
1817	Tridens J. J. Roemer & J. A. Schultes	1887	Polytrias Eduard Hackel
1817	Tripogon J. J. Roemer & J. A. Schultes	1887	Redfieldia George Vasey
1819	Critesion C. S. Rafinesque-Schmaltz	1888	Willkommia Eduard Hackel
1819	Distichlis C. S. Rafinesque-Schmaltz	1890	Calamovilfa Frank Lamson Scribner
1819		1891	
	Sitanion C. S. Rafinesque-Schmaltz		Bothriochloa Carl Ernst Otto Kuntze
1822	Chrysopogon Carl Bernard von Trinius	1891	Hackelochloa Carl Ernst Otto Kuntze
1822	Desmazeria B. C. J. Dumortier	1894	Limnodea Lyster Hoxie Dewey
1822	Rostraria Carl Bernard von Trinius	1895	Euclasta Adrien René Franchet
1022	Character de Carl Demand von Trinius		
1822	Stenotaphrum Carl Bernard von Trinius	1896	Neyraudia Joseph Dalton Hooker
1822	Vetiveria J. B. G. M. Bory de Saint-Vincents	1896	Setariopsis F. L. Scribner & C. F. Millspaugh
1823	Dupontia Robert Brown	1897	Cortaderia Otto Stapf
1823		1898	
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1823	<i>Pleuropogon</i> Robert Brown	1899	Neostapfia Joseph Burtt Davy
1824	Pleuraphis John Torrey		
1827	Ampelodesmos Johann Heinrich Friedrich Link		THE 20 [™] CENTURY
			THE 20 CENTOR
1827	Catapodium Johann Heinrich Friedrich Link		
1827	Chasmanthium Johann Heinrich Friedrich Link	1900	Sieglingia Luciano Bernardi
1827	Helictotrichon Josef August Schultes	1901	Sacciolepis George Valentine Nash
1829	Amphicarpum Karl Sigismund Kunth	1901	Sorghastrum George Valentine Nash
1029			
1829	Cottea Karl Sigismund Kunth	1903	Erioneuron George Valentine Nash
1829	Schizachyrium C. G. D. Nees von Esenbeck	1903	Neeragrostis Benjamin Franklin Bush
1829	Trachypogon C. G. D. Nees von Esenbeck	1903	Phanopyrum George Valentine Nash
1829	Trichachne C. G. D. Nees von Esenbeck	1906	Dasyochloa Per Axel Rydberg
1830			
	Alloteropsis Jan Swatopluk Presl	1906	Leptoloma Mary Agnes Chase
1830	Cathestecum Jan Swatopluk Presl	1906	Sphenopholis Frank Lamson Scribner
1830	Lophochloa Carl Ludwig Reichenbach	1907	Acrachne Emilio Chiovenda
1830	Opizia C. G. D. Nees von Esenbeck	1909	Reimarochloa Albert Spear Hitchcock
1830	Piptochaetium Jan Swatopluk Presl	1909	Schizachne Eduard Hackel
1830		1910	Lasiacis Albert Spear Hitchcock
1831	Coelorachis Adolphe Théodore Brongniart	1910	Podagrostis F. L. Scribner & E. D. Merrill
1832	Euchlaena Heinrich Adolph Schrader	1912	Hesperochloa Per Axel Rydberg
1833	Coridochloa C. G. D. Nees von Esenbeck	1919	Bromelica Oliver Atkins Farwell
1834	Fingerhuthia C. G. D. Nees von Esenbeck	1920	Acroceras Otto Stapf
1835	Thysanolaena C. G. D. Nees von Esenbeck	1920	Paspalidium Otto Stapf
1836	*	1925	
1030			
1836	Enteropogon C. G. D. Nees von Esenbeck		Pseudosasa Takenoshin Nakai
		1925	
1836	Microstegium C. G. D. Nees von Esenbeck	1933	Vaseyochloa Albert Spear Hitchcock
1836 1836	Microstegium C. G. D. Nees von Esenbeck		
1836	Microstegium C. G. D. Nees von Esenbeck Rhynchelytrum C. G. D. Nees von Esenbeck	1933 1934	Vaseyochloa Albert Spear Hitchcock Eremopoa Roman Julievich Roshevitz
1836 1837	Microstegium C. G. D. Nees von Esenbeck Rhynchelytrum C. G. D. Nees von Esenbeck Scolochloa Johann Heinrich Friedrich Link	1933 1934 1934	Vaseyochloa Albert Spear Hitchcock Eremopoa Roman Julievich Roshevitz Psathyrostachys Sergei Arsenjevic Nevski
1836 1837 1838	Microstegium C. G. D. Nees von Esenbeck Rhynchelytrum C. G. D. Nees von Esenbeck Scolochloa Johann Heinrich Friedrich Link Lophochlaena C. G. D. Nees von Esenbeck	1933 1934 1934 1934	Vaseyochloa Eremopoa Psathyrostachys Taeniatherum Albert Spear Hitchcock Roman Julievich Roshevitz Sergei Arsenjevic Nevski Sergei Arsenjevic Nevski
1836 1837 1838 1841	Microstegium C. G. D. Nees von Esenbeck Rhynchelytrum C. G. D. Nees von Esenbeck Scolochloa Johann Heinrich Friedrich Link Lophochlaena Tetrachne C. G. D. Nees von Esenbeck C. G. D. Nees von Esenbeck	1933 1934 1934 1934 1946	Vaseyochloa Eremopoa Psathyrostachys Taeniatherum Parapholis Albert Spear Hitchcock Roman Julievich Roshevitz Sergei Arsenjevic Nevski Sergei Arsenjevic Nevski Charles Edward Hubbard
1836 1837 1838 1841 1842	Microstegium Rhynchelytrum Scolochloa Lophochlaena Tetrachne Vahlodea C. G. D. Nees von Esenbeck Elias Magnus Fries	1933 1934 1934 1934 1946 1949	Vaseyochloa Eremopoa Psathyrostachys Taeniatherum Parapholis Torreyochloa Albert Spear Hitchcock Roman Julievich Roshevitz Sergei Arsenjevic Nevski Sergei Arsenjevic Nevski Charles Edward Hubbard George Lyle Church
1836 1837 1838 1841	Microstegium C. G. D. Nees von Esenbeck Rhynchelytrum C. G. D. Nees von Esenbeck Scolochloa Johann Heinrich Friedrich Link Lophochlaena Tetrachne C. G. D. Nees von Esenbeck C. G. D. Nees von Esenbeck	1933 1934 1934 1934 1946	Vaseyochloa Eremopoa Psathyrostachys Taeniatherum Parapholis Albert Spear Hitchcock Roman Julievich Roshevitz Sergei Arsenjevic Nevski Sergei Arsenjevic Nevski Charles Edward Hubbard
1836 1837 1838 1841 1842 1843	Microstegium Rhynchelytrum Scolochloa Lophochlaena Tetrachne Vahlodea Amphibromus C. G. D. Nees von Esenbeck	1933 1934 1934 1934 1946 1949	Vaseyochloa Eremopoa Psathyrostachys Taeniatherum Parapholis Torreyochloa Ectosperma Albert Spear Hitchcock Roman Julievich Roshevitz Sergei Arsenjevic Nevski Sergei Arsenjevic Nevski Charles Edward Hubbard George Lyle Church Jason Swallen
1836 1837 1838 1841 1842 1843 1843	Microstegium Rhynchelytrum Scolochloa Lophochlaena Tetrachne Vahlodea Amphibromus Phyllostachys C. G. D. Nees von Esenbeck C. G. D. Nees von Esenbeck C. G. D. Nees von Esenbeck Elias Magnus Fries C. G. D. Nees von Esenbeck C. G. D. Nees von Esenbeck P. von Siebold & J. Zuccarini	1933 1934 1934 1934 1946 1949 1950 1962	Vaseyochloa Eremopoa Psathyrostachys Taeniatherum Parapholis Torreyochloa Ectosperma Avenochloa Paseyochloa Roman Julievich Roshevitz Sergei Arsenjevic Nevski Sergei Arsenjevic Nevski Charles Edward Hubbard George Lyle Church Jason Swallen Josef Holub
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1836 1837 1838 1841 1842 1843 1843 1846 1848 1848	Microstegium Rhynchelytrum Scolochloa Lophochlaena Tetrachne Vahlodea Amphibromus Scleropoa Anisantha Leymus C. G. D. Nees von Esenbeck C. G. D. Nees von Esenbeck Elias Magnus Fries P. von Siebold & J. Zuccarini August Heinrich Grisebach Wilhelm Daniel Koch Leymus Christian Ferdinand Hochstetter Puccinellia	1933 1934 1934 1934 1946 1949 1950 1962 1963 1965 1967 1973	Vaseyochloa Eremopoa Psathyrostachys Taeniatherum Parapholis Torreyochloa Ectosperma Avenochloa SwalleniaThomas R. Soderstrom & H. Decker Hainardia Dinebra Albert Spear Hitchcock Roman Julievich Roshevitz Sergei Arsenjevic Nevski Sergei Arsenjevic Nevski Charles Edward Hubbard George Lyle Church Jason Swallen Josef Holub Werner R. Greuter Nicolaus von Jacquin
1836 1837 1838 1841 1842 1843 1843 1846 1848	Microstegium Rhynchelytrum Scolochloa Lophochlaena Tetrachne Vahlodea Amphibromus Phyllostachys Scleropoa Anisantha Leymus C. G. D. Nees von Esenbeck C. G. D. Nees von Esenbeck Elias Magnus Fries C. G. D. Nees von Esenbeck Elias Magnus Fries August Heinrich Grisebach Wilhelm Daniel Koch Christian Ferdinand Hochstetter	1933 1934 1934 1934 1946 1949 1950 1962 1963 1965 1967 1973 1974	Vaseyochloa Eremopoa Psathyrostachys Taeniatherum Parapholis Torreyochloa Ectosperma Avenochloa SwalleniaThomas R. Soderstrom & H. Decker Hainardia Albert Spear Hitchcock Roman Julievich Roshevitz Sergei Arsenjevic Nevski Sergei Arsenjevic Nevski Charles Edward Hubbard George Lyle Church Jason Swallen Josef Holub Werner R. Greuter
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Eugène Pierre Fournier

George Vasey

1886

1886

Hyparrhenia

Orcuttia

4.04 - GRASS GENERA: ALPHABETICAL

The purpose of the	nis section is to	Calammophila	Poo: Avn	Festuca	Poo: Poe
provide an alphal		Calamovilfa	Chl: Era	Festulolium	Poo: Poe
the genera of N		Catabrosa	Poo: Poe	Fingerhuthia	Chl: Era
grasses. The ge		Cataprosa Catapodium →	Desmazeria	ringernaena	CIII. LIU
the left column is	the ope that I	Catapoulum → Cathestecum →	Bouteloua	Castridium	Door Ave
				Gastridium	Poo: Avn
accept, unless yo	ou are referred	Cenchrus	Pan: Pan	Gaudinia	Poo: Avn
to another name	, as in the first	Ceratochloa →	Bromus	Glyceria	_ Poo: Mel
entry. The abbre		Chasmanthium	Cnt: Cnt	<i>Glyceria</i> (p. p.) →	
right column ind	icate the sub-	Chloris	Chl: Cyn	Gymnopogon	Chl: Cyn
family and tribe	of that genus.	Chondrosum →	Bouteloua	Gynerium	Arn: Arn
•	_	Chrysopogon	Pan: And	•	
****	****	Cinna . J	Poo: Avn	Hackelochloa	Pan: And
		Cladoraphis	Chl: Era	Hakonechloa	Arn: Arn
Achnatherum →	Stipa	Coelorachis	Pan: And	Hainardia	Poo: Hhr
Achnella →	Stipa	Coix	Pan: And	Heleochloa →	Crypsis
Acrachne	Chl: Era	Coleanthus	Poo: Poe		Poo: Avn
				Helictotrichon	
Acroceras	Pan: Pan	Coridochloa →	Alloteropsis	Hemarthria	Pan: And
Aegilopodes →	Aegilops	Cortaderia	Dan: Dnt	Hesperochloa →	Festuca
Aegilops	Poo: Trt	Corynephorus	Poo: Avn	Heteropogon	Pan: And
Aegopogon	Chl: Cyn	Cottea	Chl: Pap	Hesperostipa →	Stipa
Agrohordeum →	Elymus	Critesion →	Hordeum	Hierochloë →	Anthoxanthum
Agropogon	Pooʻ: Avn	Crypsis	Chl: Era	Hilaria	Chl: Cyn
Agropyron	Poo: Trt	Ctenium	Chl: Cyn	Hilaria (p. p.) →	Pleuraphis
Agrositanion →	Elymus	Cutandia	Poo: Poe	Holcus	Poo: Avn
	Poo: Avn		Aegilops	Hordeum	Poo: Trt
Agrostis		Cylindropyrum →			
Allalania	Poo: Avn	Cymbopogon	Pan: And	Hymenachne	Pan: Pan
Allolepis	Chl: Era	Cynodon	Chl: Cyn	Hyparrhenia	Pan: And
Alloteropsis	Pan: Pan	Cynosurus	Poo: Poe	Hystrix →	Elymus
Alopecurus	Poo: Avn				
Ammophila	Poo: Avn	Dactylis	Poo: Poe	Imperata	Pan: And
Ampelodesmos	Poo: Stp	Dactyloctenium	Chl: Era	Ischaemum	Pan: And
Amphibromus →		Danťhonia	Dan: Dnt		
Amphicarpum	Pan: Pan	Dasyochloa →	Erioneuron	Jarava →	Stipa
Andropogon	Pan: And	Dasypyrum	Poo: Trt	20.010	0 p u
Anisantha →	Bromus	Dendrocalamopsis		Koeleria	Poo: Avn
	Pan: Pan			Rueleila	POU. AVII
Anthenantia		Deschampsia	Poo: Avn	1.000.000	Door Arm
Anthephora	Pan: Pan	Desmazeria	Poo: Poe	Lagurus	Poo: Avn
Anthoxanthum	Poo: Avn	Diarrhena	Poo: Dhr	Lamarckia	Poo: Poe
Apera	Poo: Avn	Dichanthelium →	Panicum	Lasiacis	Pan: Pan
Apluda	Pan: And	Dichanthium	Pan: And	Leersia	Ehr: Ory
Arctagrostis*	Poo: Poe	Digitaria	Pan: Pan	Leptochloa	Chl: Era
Arctophila*	Poo: Poe	Dinebra →	Chl: Era	Leptoloma →	Digitaria
Aristida	Ars: Ars	Diplachne →	Leptochloa	Leucopoa →	Festuca
Arrhenatherum	Poo: Avn	Dissanthelium †	Poo: Avn	Leymus →	Elymus
Arthraxon	Pan: And	Distichlis	Chl: Era	Limnodea	Poo: Avn
Arundinaria	Bam: Bam	Dupontia*	Poo: Poe	Lolium	Poo: Poe
		Биропсіа	F00. F0C	Lophochlaena →	
Arundo	Arn: Arn	Fabina ablas	Dani Dan	Lophochica -	Pleuropogon
Austrostipa →	Stipa	Echinochloa	Pan: Pan	Lophochloa →	Koeleria
Avena	Poo: Avn	Ectosperma →	Swallenia	Ļopḥopyrum →	Elymus
Avenochloa →	Helictotrichon	Ehrharta	Poo: Ehr	Luziola	Ehr: Ory
Avenula →	Helictotrichon	Eleusine	Chl: Era	Lycurus	Chl: Era
Axonopus	Pan: Pan	Elionurus	Pan: And		
		Elyhordeum	Poo: Trt	<i>Manisuris</i> (p. p.)	→ Coelorachis
Bambusa	Bam: Bam	Elymus	Poo: Trt	<i>Manisuris</i> (p. p.)	→ Hemarthria
Beckmannia	Poo: Avn	Elysitanion →	Elymus	Melica	Poo: Mel
Blepharidachne	Chl: Era	Elytesion →	Elymus	Melinis	Pan: Pan
Blepharoneuron	Chl: Era	Elytrigia →	Elymus	Mibora	Poo: Avn
Bothriochloa	Pan: And		Chl: Pap	Microchloa	
		Enneapogon			Chl: Cyn
Bouteloua	Chl: Cyn	Enteropogon	Chl. Ero	Microstegium	Pan: And
Brachiaria	Pan: Pan	Eragrostis	Chl: Era	Milium	Poo: Avn
Brachyelytrum	Poo: Brl	Eremochloa	Pan: And	Miscanthus	Pan: And
Brachypodium	Poo: Brp	Eremopoa	Poo: Poe	Molinia	Arn: Dnt
Briza	Poo: Poe	Eremopyrum	Poo: Trt	Monanthochloë	Chl: Era
Bromelica →	Melica	Erianthús →	Saccharum	Monerma →	Hainardia
Bromopsis →	Bromus	Eriochloa	Pan: Pan	Muhlenbergia	Chl: Era
Bromus	Poo: Brm	Erioneuron	Chl: Era	Munroa	Chl: Era
Buchloë	Chl: Cyn	Euchlaena →	Zea		
_ 30,,,00	S Sy.	Euclasta	Pan: And	Nardus	Poo: Nrd
Calamagrostis	Poo: Avn	Eustachys	Chl: Cyn	Nassella →	Stipa
Calamagiostis	1 OOI AVII	Lastacitys	Cini Cyii	Hassella -	Stipa

Neeragrostis →	Eragrostis	Sporobolus	Chl: Era
Neostapfia	Chl: Orc	Steinchisma	Pan: Pan
Neyraudia	Chl: Era	Stenotaphrum	Pan: Pan
rveyradala	Cili. Liu	Stipa	Poo: Stp
Olyra †	Bam: Oly	Stiporyzopsis →	Stipa
Opizia		Swallenia	Chl: Era
	Chl: Cyn	Swallerlla	CIII. El a
Oplismenus	Pan: Pan	T'	- 1
Orcuttia	Chl: Orc	Taeniatherum →	Elymus
Oryza	Ehr: Ory	Themeda	Pan: And
Oryzopsis	Poo: Stp	Thinopyrum →	Elymus
		Thysanolaena	Arn: Arn
Panicum	Pan: Pan	Torreyochloa	Poo: Poe
Pappophorum	Chl: Pap	Trachypogon	Pan: And
Parapholis	Poo: Hhr	Tragus	Chl: Cyn
Pascopyrum →	Elymus	Trichachne →	Digitaria
Paspalidium →	Setaria	Trichloris →	Chloris
	Pan: Pan	Trichoneura	Chl: Era
Paspalum			
Pennisetum	Pan: Pan	Tridens	Chl: Era
Phalaris	Poo: Avn	Triplasis	Chl: Era
Phalaroides →	Phalaris	Tripogon	Chl: Era
Phanopyrum →	Panicum	Tripsacum	Pan: May
Pharus	Phr: Phr	Triraphis	Chl: Era
Phippsia	Poo: Poe	Trisėtum	Poo: Avn
Phle'um	Poo: Avn	Triticum	Poo: Trt
Phragmites	Arn: Arn	Tuctoria	Chl: Orc
Phyllostachys	Bam: Bam	ractoria	Cili. Oic
Pintathorum ->		Uniola	Chl. Era
Piptatherum →	Oryzopsis	Uniola	Chl: Era
Piptochaetium	Poo: Stp	Uniola (p. p.) \rightarrow C	nasmantnium
Pleioblastus	Bam: Bam	Urochloa	Pan: Pan
Pleuraphis →	Hilaria		
Pleuropogon	Poo: Mel	Vahlodea →	Deschampsia
Poa	Poo: Poe	Vaseyochloa	Chl: Era
Podagrostis →	Agrostis	Ventenata	Poo: Avn
Pogonarthria	Chl: Era	Vetiveria →	Chrysopogon
Polypogon	Poo: Avn	Vulpia →	Festuca
Polytrias	Pan: And	vaipia	7 031404
		Willkommia	Chl. Cyn
Psathyrostachys → Pseudelymus →	Elymus Elymus	vviiikoitiitiia	Chl: Cyn
PSeudeivinus →	rivitilis		
		7	D M
Pseudoroegneria →	Elymus	Zea .	Pan: May
Pseudoroegneria → Pseudosasa	<i>Elymus</i> Bam: Bam	Zizania	Ehr: Orý
Pseudoroegneria →	Elymus		
Pseudoroegneria → Pseudosasa	<i>Elymus</i> Bam: Bam	Zizania Zizaniopsis	Ehr: Orý
Pseudoroegneria → Pseudosasa Ptilagrostis Puccinellia	Elymus Bam: Bam Poo: Stp Poo: Poe	Zizania	Ehr: Ory Ehr: Ory
Pseudoroegneria → Pseudosasa Ptilagrostis	Elymus Bam: Bam Poo: Stp Poo: Poe	Zizania Zizaniopsis	Ehr: Ory Ehr: Ory
Pseudoroegneria → Pseudosasa Ptilagrostis Puccinellia Puccinellia (p. p.) →	Elymus Bam: Bam Poo: Stp Poo: Poe Torreyochloa	Zizania Zizaniopsis Zoysia	Ehr: Ory Ehr: Ory
Pseudoroegneria → Pseudosasa Ptilagrostis Puccinellia Puccinellia (p. p.) → Redfieldia	Elymus Bam: Bam Poo: Stp Poo: Poe Torreyochloa Chl: Era	Zizania Zizaniopsis	Ehr: Ory Ehr: Ory
Pseudoroegneria → Pseudosasa Ptilagrostis Puccinellia Puccinellia (p. p.) → Redfieldia Reimarochloa	Elymus Bam: Bam Poo: Stp Poo: Poe Torreyochloa Chl: Era Pan: Pan	Zizania Zizaniopsis Zoysia NOTES:	Ehr: Ory Ehr: Ory Chl: Cyn
Pseudoroegneria → Pseudosasa Ptilagrostis Puccinellia Puccinellia (p. p.) → Redfieldia Reimarochloa Rhynchelytrum →	Elymus Bam: Bam Poo: Stp Poo: Poe Torreyochloa Chl: Era Pan: Pan Melinis	Zizania Zizaniopsis Zoysia NOTES: † = presumed to	Ehr: Ory Ehr: Ory
Pseudoroegneria → Pseudosasa Ptilagrostis Puccinellia Puccinellia (p. p.) → Redfieldia Reimarochloa Rhynchelytrum → Rhytidosperma →	Elymus Bam: Bam Poo: Stp Poo: Poe Torreyochloa Chl: Era Pan: Pan Melinis Danthonia	Zizania Zizaniopsis Zoysia NOTES:	Ehr: Ory Ehr: Ory Chl: Cyn
Pseudoroegneria → Pseudosasa Ptilagrostis Puccinellia Puccinellia (p. p.) → Redfieldia Reimarochloa Rhynchelytrum → Rhytidosperma → Rostraria →	Elymus Bam: Bam Poo: Stp Poo: Poe Torreyochloa Chl: Era Pan: Pan Melinis Danthonia Koeleria	Zizania Zizaniopsis Zoysia NOTES: † = presumed to extirpated	Ehr: Ory Ehr: Ory Chl: Cyn
Pseudoroegneria → Pseudosasa Ptilagrostis Puccinellia Puccinellia (p. p.) → Redfieldia Reimarochloa Rhynchelytrum → Rhytidosperma →	Elymus Bam: Bam Poo: Stp Poo: Poe Torreyochloa Chl: Era Pan: Pan Melinis Danthonia	Zizania Zizaniopsis Zoysia NOTES: † = presumed to	Ehr: Ory Ehr: Ory Chl: Cyn
Pseudoroegneria → Pseudosasa Ptilagrostis Puccinellia Puccinellia (p. p.) → Redfieldia Reimarochloa Rhynchelytrum → Rhytidosperma → Rostraria → Rottboellia	Elymus Bam: Bam Poo: Stp Poo: Poe Torreyochloa Chl: Era Pan: Pan Melinis Danthonia Koeleria	Zizania Zizaniopsis Zoysia NOTES: † = presumed to extirpated p. p. = in part	Ehr: Ory Ehr: Ory Chl: Cyn
Pseudoroegneria → Pseudosasa Ptilagrostis Puccinellia Puccinellia (p. p.) → Redfieldia Reimarochloa Rhynchelytrum → Rhytidosperma → Rostraria →	Elymus Bam: Bam Poo: Stp Poo: Poe Torreyochloa Chl: Era Pan: Pan Melinis Danthonia Koeleria Pan: And Pan: And	Zizania Zizaniopsis Zoysia NOTES: † = presumed to extirpated p. p. = in part	Ehr: Ory Ehr: Ory Chl: Cyn
Pseudoroegneria → Pseudosasa Ptilagrostis Puccinellia Puccinellia (p. p.) → Redfieldia Reimarochloa Rhynchelytrum → Rhytidosperma → Rostraria → Rottboellia	Elymus Bam: Bam Poo: Stp Poo: Poe Torreyochloa Chl: Era Pan: Pan Melinis Danthonia Koeleria Pan: And Pan: And Pan: Pan	Zizania Zizaniopsis Zoysia NOTES: † = presumed to extirpated p. p. = in part	Ehr: Ory Ehr: Ory Chl: Cyn
Pseudoroegneria → Pseudosasa Ptilagrostis Puccinellia Puccinellia (p. p.) → Redfieldia Reimarochloa Rhynchelytrum → Rhytidosperma → Rostraria → Rottboellia Saccharum Sacciolepis	Elymus Bam: Bam Poo: Stp Poo: Poe Torreyochloa Chl: Era Pan: Pan Melinis Danthonia Koeleria Pan: And Pan: And Pan: Pan	Zizania Zizaniopsis Zoysia NOTES: † = presumed to extirpated p. p. = in part * = found in	Ehr: Ory Ehr: Ory Chl: Cyn
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Pseudoroegneria → Pseudosasa Ptilagrostis Puccinellia Puccinellia (p. p.) → Redfieldia Reimarochloa Rhynchelytrum → Rhytidosperma → Rostraria → Rottboellia Saccharum Sacciolepis Schedonorus (p. p. Schedonorus (p. p.	Elymus Bam: Bam Poo: Stp Poo: Poe Torreyochloa Chl: Era Pan: Pan Melinis Danthonia Koeleria Pan: And Pan: And Pan: Pan Chl: Cyn) → Festuca) → Lolium	Zizania Zizaniopsis Zoysia NOTES: † = presumed to extirpated p. p. = in part * = found in Canada only	Ehr: Ory Ehr: Ory Chl: Cyn
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4.05 - GRASS GENERA: SYNOPSIS

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Acrachne. One species: A. racemosa, adventive in southern California.

Acroceras. One species: A. oryzoides.

Aegilops. Goat grass. Three species: Ae. cylindrica, Ae. ovata, and Ae. triuncialis.

Aegopogon. One species: Ae. tenellus.

X Agropogon. One species: X A. littoralis, an intergeneric hybrid involving Agrostis stolonifera and Polypogon monspeliensis.

Agropyron. Crested wheatgrass. One to three species: (a much smaller genus than in H & C; most species have been transfered to *Elymus*). *A. desertorum*, *A. cristatum*, and *A. pectiniforme*.

Agrostis. Bent, bent grass, hair grass, redtop, tickle grass. About 40 species.

Aira. Hair grass, tickle grass. Three species: *A. caryophyllea, A. elegans*, and *A. praecox*.

Allolepis. One species: A. texana, native to TX.

Alloteropsis. One species: A. cimicina. FL.

Alopecurus. Foxtail, meadow foxtail. Eleven species.

Ammophila. Beach grass, European beach grass. Three species: *A. arenaria* (a European species introduced to control erosion on coastal sand dunes), *A. breviligulata*, and *A. champlainensis* (restricted to the Great Lakes region).

Ampelodesmos. Dis grass. One species, *A. mauritanica*, a Mediterranean species sometimes grown as an ornamental has escaped in Napa Co., CA.

Amphicarpum. Goober grass. Two species: *A. muhlenbergii* and *A. purshii*, are unusual in having underground cleistogamous spikelets.

Andropogon. Bluestem, beard grass, broom-sedge. About 14 species.

Anthaenantia. Silky scale. Two species: A. rufa and A. villosa.

Anthephora. One species: A. hermaphrodita.

Anthoxanthum. Vernal grass, sweet vernal grass. Two species: *A. aristatum* and *A. odoratum*.

Apera. Silky bent grass, wind grass. Two species: *A. spica-venti* and *A. interrupta*.

Apluda. Mauritian grass. One species: *A. mutica*, intro-duced in MD where it may not be persisting.

Arctagrostis. Polar grass. One species: A. latifolia, which is native to AK and Canada, but not found in the conterminous U. S.

Arctophila. Pendant grass. One species, A. fulva, which is native to AK and elsewhere, but not found in the conterminous U. S.

Aristida. Three-awn, arrow feather. Thirty-six species.

Arrhenatherum. Oat-grass. One species: *A. elatius*. Eurasia. Some plants have a series of small, bulb-like corms.

Arthraxon. One species: A. hispidus.

Arundinaria. Cane, switch cane. Three species: *A. gigantea*, with two subspecies and a hybrid between them (*A. g.* ssp. x *macrosperma*) of the eastern and southeastern U. S. are our only native woody bamboos. *A. pumila*, an ornamental bamboo from Japan, is naturalized in CA. *A. simonii* is naturalized in NV.

Arundo. Reed, giant reed. One species: *A. donax*. Robust perennial typically found along rivers, streams, and waterways.

 $\boldsymbol{Avena}.$ Oat, wild oat. Ten species, all native to the Old World.

Axonopus. Carpet grass. Three species: A. affinis, A. compressus, and A. furcatus.

Bambusa. Hedge bamboo, common bamboo. Two species: *B. glauscens* and *B. vulgaris* have escaped from cultivation and are now established in FL.

Beckmannia. Slough grass. One species: *B. syzigachne*.

Blepharidachne. Eyelash grass. Two species: *B. bigelovii* and *B. kingii*.

Blepharoneuron. Pine dropseed. One species: *B. tricholepis*.

Bothriochloa. Silver bluestem. Fifteen species.

Bouteloua. Grama grass, sideoats grama. Twentytwo species. Includes *Cathestecum*.

Brachyelytrum. Short-husk. Two species: *B. erectum* and *B. septentrionale*.

Brachypodium. False brome. Three species (all European): *B. distachyon* occurs widely. *B. sylvaticum* is naturalized in Benton Co., OR. *B. pinnatum* occurs along the California coast.

Briza. Quaking grass, rattlesnake grass. Three species: *B. maxima* (often used in dried arrangements), *B. media*, and *B. minor*. European introductions.

Bromus. Brome, chess, cheat grass, rescue grass, ripgut, chess. Forty-five species.

Buchloë. Buffalo grass. One species: *B. dactyloides*, a native of the short grass and mixed grass prairies.

Calamagrostis. Reed grass, reed bent grass, bluejoint. Thirty-seven species.

X Calammophila. Reed grass. Two species, hybrids between *Calamagrostis* and *Ammophila*.

Calamovilfa. Sand reed. Five species.

Catabrosa. Brook grass. One species: C. aquatica.

Cenchrus. Sandbur. Twelve species.

Chasmanthium. Wild-oats, spike grass. Five species. The conspicuously flattened spikelets of *C. latifolium* are sometimes used in dried arrangements.

Chloris. Finger grass, windmill grass, feather grass, Rhodes grass. Twenty-four species.

Chrysopogon. Beard grass, khus-khus, vetiver grass. Three species: introduced grasses, mostly in the south-eastern U. S.

Cinna. Wood reed. Three species: *C. arundinacea*, *C. bolanderi*, and *C. latifolia*.

Cladoraphis. Bristly love grass. One species: *C. cyperoides*, adventive in OR. Southern Africa.

Coelorachis. Joint grass, joint-tail, thintail. Four species. Treated as *Manisuris* in H & C.

Coix. Job's tears. One species: *C. lacryma-jobi*, established in FL and LA. Its grey-white, seed-like pistillate involucres are found in necklaces, rosaries, and junk jewelry.

Coleanthus. One species: *C. subtilis*, adventive in OR and WA.

Cortaderia. Pampas grass. Two species: *C. jubata* has become a major pest in northern CA. *C. dioica* is a popular ornamental.

Corynephorus. Gray hair grass. One species: *C. canescens*. Europe.

Cottea. One species: *C. pappophoroides*, native from AZ to TX.

 $\it Crypsis$. Swamp timothy, prickle grass. Three species: $\it C.~alopecuroides,~C.~schoenoides,~and~C.~vaginiflora.$ Eurasia & Africa. [= $\it Heleochloa$ in H & C].

Ctenium. Toothache grass. Two species: *Ct. aromaticum* and *Ct. floridanum*. Grasses of the southeastern U. S., the latter endemic to FL.

Cutandia. One species: *C. memphitica*, aMediterranean introduction naturalized in CA. Doubtfully persisting.

Cymbopogon. Citronella grass. Three species: *C. citratus*, *C. nardus*, and *C. refractus*. Source of essential oil used in cooking and perfumes, etc.

Cynodon. Bermuda grass, star grass. Five species: *C. dactylon* is a popular lawn grass.

Cynosurus. Dogtail. Two species: *C. cristatus* and *C. echinatus*. European weeds.

Dactylis. Orchard grass, cocksfoot. One species: *D. glomerata*. Europe.

Dactyloctenium. Crowfoot grass. Two species: *D. aegyptium* naturalized along the Gulf and Atlantic coastal plains. *D. radulans*, from Australia, escaped in Tucson, AZ.

Danthonia. Poverty-oats, oat-grass. Thirteen species.

Dasypyrum. Mosquito grass. One species: *D. villosum*, a Mediterranean introduction.

Deschampsia. Hair grass. Seven species: all of them native.

Desmazeria. Fern grass. One species: *D. rigida*, a European introduction.

Diarrhena. Beak grain. One species: *D. americana*, native to eastern and central states.

Dichanthium. Bluestem. Three species: *D. annulatum*, *D. aristatum*, and *D. sericeum*, Old World pasture grasses found in TX and LA.

Digitaria. Crab grass, witch grass, Arizona cottontop. About thirty species.

Dinebra. Viper grass. One species: *D. retroflexa*, adventive in CA & NC. Tropical Africa and Asia.

Dissanthelium. One species: *D. californicum,* native to San Clement, Santa Catalina Islands, and Baja California, presumed extinct in the United States (last collected in 1912).

Distichlis. Salt grass. One species: *D. spicata*, found typically in saline soils and coastal salt marshes.

Dupontia. Tundra grass. One speices: *D. fischeri*, native to AK and elsewhere, but not found in the conterminous U. S.

Echinochloa. Barnyard grass, jungle-rice, water grass, cockspur. Eight species.

Ehrharta. Veldt grass. Three species: *E. calycina*, *E. erecta*, and *E. longiflora* (all from S. Africa) naturalized in CA.

Eleusine. Goose grass, African millet, three-spike grass, yard grass. Three species: *E. coracana, E. indica* and *E. tristachya* are naturalized in scattered locations.

Elionurus. Balsam scale. Two species: *E. barbiculmis* and *E. tripsacoides*.

X *Elyhordeum*. Five species. An intergeneric hybrid between *Elymus* and *Hordeum*.

Elymus. Rye grass, wild-rye, quack grass, squirreltail, bottlebrush grass. Genus much larger or much smaller than in H & C, depending upon taxonomic treatment followed. About fifty-five species.

Enneapogon. Spike-pappus grass, 9-awned pappus grass. One species: *E. desvauxii*.

Enteropogon. Umbrella grass. Three species, two of them introduced from the Old World.

Eragrostis. Love grass, stink grass, pony grass. About 56 species.

Eremochloa. Centipede grass. One species: *E. ophuroides*, naturalized in TX and LA. Historic collection of *E. ciliaris* from San Francisco bay area.

Eremopyrum. Annual wheat grass. Three species: *E. bonepartis* and *E. orientale* are naturalized in NY. *E.*

triticeum is naturalized in the western U. S. Old World introductions.

Eriochloa. Cup grass. Eleven species.

Erioneuron. Fluff grass. Three species: *E. avenaceum*. *E. pilosum* and *E. pulchellum* are grasses of drier sites in the southwestern U. S.

Eustachys. Chickenfoot grass, finger grass, windmill grass. Seven species.

Festuca. Fescue, annual fescue, rye grass, darnel. As treated here, genus includes annual species often segregated into *Vulpia*, *Lolium* and X *Festulolium*. About 47 species.

Festulolium. Three species, hybrids between *Festuca* and *Lolium*.

Fingerhuthia. Zulu-fescue. One species: *F. africana,* adventive in AZ. Southern Africa, Asia.

Gastridium. Nit grass. One species: *G. ventricosum*, a European introduction, widely naturalized.

Gaudinia. One species: *G. fragilis*, a European introduction, found only in Sonoma Co., CA.

Glyceria. Manna grass, fowl meadow grass. Nineteen species.

Gymnopogon. Skeleton grass, beard grass. Three species: mostly eastern half of country; *G. floridana* endemic to FL and adjacent GA.

Gynerium. Uva grass. One species: *G. sagittatum*, a popular ornamental in dried arrangements. Escaping in FL.

Hackelochloa. One species: *H. granularis*, is naturalized in AZ, NM, and LA.

Hainardia. Thintail. One species: *H. cylindrica*, introduced from Europe.

Hakonechloa. Hakone grass, Japanese forest grass. One species: *H. macra*, escaped from cultivation in UT

Helictotrichon. Spike-oat, alpine-oat. Three species: *H. hookeri, H. mortonianum*, and *H. pubescens*.

Hemarthria. Limpo grass. One species: *H. altissima*, endemic to TX.

Heteropogon. Tanglehead. Two species: *H. contortus* and *H. melanocarpus*, occur in AZ to TX.

Hilaria. Tabosa, curly-mesquite, galleta. XXX Two species: *H. jamesii* and *H. rigida* native, mostly from CA to UT, WY, and TX.

Holcus. Velvet grass. Two species: *H. mollis* (annual) and *H. lanatus* (perennial). Introduced from Europe and Africa.

Hordeum. Barley, squirreltail. Nine species.

Hymenachne. One species: *H. amplexicaulis*, found in FL.

Hyparrhenia. Thatching grass, jaragua grass. Two species: *H. rufa* and *H. hirta*. Neither may persist in

North America.

Imperata. Cogon grass, satin tail.Three species: *I. cylindrica*, *I. brasiliensis*, and *I. brevifolia*.

Ischaemum. Muraina grass. Two species, introduced from Asia.

Koeleria. June grass. Two species: *K. macrantha* (a common native perennial) and *K. gerardii* (an introduced annual).

Lagurus. Hare's tail. One species, *L. ovatus*, a Mediterranean introduction. Sometimes used in dried arrange-ments.

Lamarckia. Golden top. One species: *L. aurea*, introduced from the Mediterranean.

Lasiacis. Small cane, tibisee. Two species: *L. divaricata* and *L. ruscifolia* (both restricted to FL).

Leersia. Cutgrass, white grass, catchfly grass. Five species. Grasses of moist woods and stream banks.

Leptochloa. Sprangletop. Twelve species.

Limnodea. Ozark grass. One species, *L. arkansana*. This genus endemic to North America.

Lolium. Rye grass, darnel. Six species, all introduced.

Luziola. Southern water grass. Three species: *L. bahiensis*, *L. carolinensis*, and *L. peruviana*. Aquatic grasses found mostly in Southeast.

Lycurus. Wolftail, Texas timothy. One species: *L. phleoides*, of drier sites in Southwest.

Melica. Onion grass, melic. Eighteen species. Some have small, onion-like bulbs.

Melinis. Two species: *M. minutiflora* (molasses grass) a Brazilian introduction; *M. repens* (ruby grass, Natal grass), an African introduction, from CA to TX. Includes taxa traditionally assigned to *Rhynchelytrum*.

Mibora. Early sand grass. One species: *M. minima*, a European introduction naturalized in MA.

Microchloa. Small grass. One species: *M. kunthii*, endemic to Cochise Co., AZ.

Microstegium. One species: *M. vimineum*, an Asian introduction.

Milium. Wood-millet. One species: *M. effusum*, a Eurasia introduction.

Miscanthus. Eulalia, plume grass, zebra grass. Four species: all Old World introductions, have become naturalized. Popular ornamentals.

Molinia. Moor grass. One species: *M. caerulea*, a Eurasian introduction.

Monanthochloë. Shore grass, key grass. One species: *M. littoralis*, Atlantic, Gulf, and California coasts. Its needle-like leaves are unique among N. American grasses.

Monroa. False buffalo grass. One species: *M. squarrosa*, native from California across Great Plains.

Muhlenbergia. Muhly, scratch grass, hair grass, nimble will, bull grass, satin grass, deer grass, ararejo grass. Seventy-three species.

Nardus. Moor mat grass. One species: *N. stricta*, a European introduction.

Neostapfia. Colusa grass. One species: *N. colusana*, endemic to vernal pools in California.

Neyraudia. Burma-reed. Two species: *N. arundinacea* and XXX introduced in CA and FL.

Olyra. One species: *O. latifolia*, known only in N. America from questionable historic collection from Tampa Bay area, FL.

Opizia. One species: *O. stolonifera*, occurs in Florida.

Oplismenus. Basket grass, wood grass. One species: O. hirtellus, naturalized in TX and LA.

Orcuttia. Orcutt grass. Five species: all endemic to vernal pools in California; one variety in Baja California. Other species cited in H & C are now in *Tuctoria*.

Oryza. Rice. Two species: O. sativa (cultivated rice), persists around paddies in CA, AR, MS, LA and FL. O. rufipogon naturalized in CA and FL.

Oryzopsis. Rice-grass, mountain-rice. Eleven species. Some species cited in H & C are now placed in *Stipa*.

Panicum. Panic grass, witch grass, millet, vine-mesquite, maiden cane. The largest genus of grasses in North America, which alone makes for a grand challenge.

Pappophorum. Pappus grass. Two species: *P. bicolor* and *P. mucronulatum* found in AZ and TX.

Parapholis. Sickle grass, hard grass. Two species: *P. incurva* and *P. strigosa* (known in N. America from recent collections around Humboldt Bay, in Humboldt Co., CA).

Paspalum. Knot grass, dallis grass, Vasey grass, paspalum, bahia grass. Forty-four species.

Pennisetum. Fountain grass, feather grass, buffel grass, Napier grass, elephant grass, kikuyu grass, pearl millet. Thirteen species. Pernicious weeds and graceful ornamentals.

Phalaris. Canary grass, reed canary grass, May grass. Eleven species.

Pharus. Creeping leafstalk grass. One species: *Ph. lap-pulaceus*; FL.

Phippsia. Ice grass, snow grass. One species: *Ph. algida*; native to CO, WY, and MT.

Phleum. Timothy. Six species: *Ph. pratense*, a Eurasian pasture grass now occurs widely in N. America. *Ph. alpinum* native to mid- and high altitude grasslands. Three other species naturalized locally, especially in OR.

Phragmites. Common reed, carrizo. One species: *Ph. australis*, common along water ways. Some authors suggest this species the most widely occurring vascular plant.

Phyllostachys. Bamboo. Two species: *Ph. aurea* (golden bamboo, fishpole bamboo) and *Ph. bambusoides* (timber bamboo, madake) escaped from cultivation in CA, NM, LA, and FL.

Piptochaetium. Piñon rice grass, needle grass. Six species: native species occur mostly in FL and Southwest.

Pleioblastus. Dwarf bamboo. Two species: *P. humilis* and *P. simonii* are escaped Japanese bamboos.

Pleuropogon. Semaphore grass. Five species: mostly grasses of Pacific coast; two endemic to CA.

Poa. Blue grass, winter grass, spear grass, mutton grass. About 80 species.

Pogonarthria. Herringbone grass. One species: *P. squarrosa*, introduced in AZ.

Polypogon. Beard grass, rabbitfoot grass, water bent grass. Five species: two native.

Polytrias. One species: *P. praemorsa*, introduced from Java.

Pseudosasa. Arrow bamboo, metake. One species: *P. japonica*, naturalized in FL.

Ptilagrostis. Porter's needle grass. One subspecies, *P. mongholica* ssp. *porteri*, endemic to CO.

Puccinellia. Alkali grass. Thirty-two species.

Redfieldia. Blowout grass. One species: *R. flexuosa*, native to sandy sites.

Reimarochloa. Florida reimar grass. One species: *R. oligostachya*, native to FL.

Rottboellia. Itch grass. One species: *R. cochinchinensis*, naturalized in TX, LA and FL.

Rytidosperma. Hairy oat grass, hairy danthonia. Three introduced species.

Saccharum. Sugar cane, noble cane. Ten species: *S. saccharum* persists around sugar cane fields in FL. *S. spontaneum* (wild Asian sugar cane) also naturalized in FL. Includes *Erianthus*.

Sacciolepis. Cupscale. Two species: *S. striata* (native) and *S. indica*, naturalized in TX and perhaps in GA.

Schedonnardus. Wire grass, tumble grass, Texas crab grass. One species: *S. paniculatus*.

Schismus. Mediterranean grass. Two species: *S. arabicus* and *S. barbatus*, Old World introductions in CA and AZ.

Schizachne. False melic. One species, *S. purpurascens*, native.

Schizachyrium. Bluestem. Eleven species.

Sclerochloa. Hard grass. One species: *S. dura*, a European introduction.

Scleropogon. Burro grass. One species: *S. brevifolius*, native to CA to CO and TX.

Scolochloa. Marsh grass, prickle grass. One species: *S. festucacea*.

Scribneria. Scribner's grass. One species: *S. bolanderi*, native along Pacific coast from BC to CA.

Secale. Rye. Two species: *S. cereale*, cultivated rye, escapes widely. *S. montanum*, an Asian introduction, established in WA and CA.

Setaria. Foxtail, millet, bristle grass. Twenty-seven species.

Setariopsis. One species: *S. auriculata*, naturalized in AZ.

Sorghastrum. Indian grass, wood grass. Four species: *S. apalachicolense*, endemic to FL; *S. nutans*, dominant of tall grass prairie.

Sorghum. Sorghum, sorgo, sorgho, durra, Johnson grass, Kafir-corn, Sudan grass, chicken-corn, broomcorn. Two species: *S. bicolor*, source of important forage and seed crops; *S. halepense* (Johnson grass), a pernicious weed, especially in agricultural areas.

Spartina. Cord grass, slough grass, marsh hay. Thirteen species: common grasses of prairies, marshes, and coastal sites.

Sphenopholis. Wedge grass, bunch grass. Five species: all native.

Sporobolus. Dropseed, smut grass, sacaton, poverty grass. Twenty-eight species.

Stenotaphrum. St. Augustine grass. One species: *S. secundatum*, popular lawn grass of Atlantic and Gulf coastal states, where it escapes.

Stipa. Needle grass, porcupine grass, winter grass, sleepy grass. Thirty-eight species.

Swallenia. Eureka Valley Dune grass. One species: *S. alexanderae*, endemic to sand dunes in Inyo Co., CA.

Themeda. Kangaroo grass. One species: *T. quadrivalvis* naturalized in LA.

Thysanolaena. Tiger grass. One species: *T. latifolia*, escaped ornamental from Southeast Asia.

Torreyochloa.

Trachypogon. Crinkle awn. Two species: *T. montufari* and *T. secundus*, the latter indigenous from AZ to TX.

Tragus. Bur grass. Two species: *T. berteronianus* and *T. racemosus* (Old World introductions), naturalized in AZ, NM, and TX.

Trichoneura. Silveus grass. One species: *T. elegans*, native to TX.

Tridens. Purpletop, redtop, tridens. Twelve species. Some species in H & C now in *Erioneuron*.

Triplasis. Sand grass. Two species: *T. americana* and *T. purpurea*.

Tripogon. One species: T. spicatus, native to TX.

Tripsacum. Gama grass. Four species: *T. dacyloides* grows in prairies; *T. floridanum* endemic to FL.

Triraphis. Purpleheads. One species: *T. mollis*, naturalized in TX.

Trisetum. Yellow-oats. Thirteen species: all but one native.

Triticum. Wheat. One species: *T. aestivum* established around agricultural fields.

Tuctoria. Tuctoria. Two species: *T. greenei* and *T. mucronata*, endemic to a few counties in CA.

Uniola. Sea-oats. One species: *U. paniculata*, native to coastal dunes of Atlantic and Gulf states. Often used in dried arrangements because of large, conspicuously flattened spikelets.

Urochloa. Signal grass, Guinea grass, brown-top millet, panicum, Pará grass. Fifteen species.

Vaseyochloa. Texas grass. One species: *V. multi-nervosa*, endemic to TX.

Ventenata. One species: *V. dubia*, Eurasian introduction naturalized in OR and CA.

Willkommia. One species: *W. texana*, endemic to San Patricio and Kleberg counties in TX.

Zea. Maize, corn, Indian corn, teosinte. One species: Zea mays; two subspecies of teosinte naturalized in SC and FL. Maize does not persist.

Zizania. Wild-rice, water-rice. Three species: *Z. texana* endemic to Hayes Co., TX.

Zizaniopsis. Water-millet, Texas wild-rice, marsh millet, southern wild-rice. One species: *Z. miliacea*, aquatic grass of southern U. S.

Zoysia. Zoysia, zoisia, matronella grass. Two species: *Z. materella* and *Z. tenuifolia*, popular lawn grasses.

Revised: 10 January 2005

XXX

4.06 - GRASSES: COMMON NAMES

Acapulco grass adlay African foxtail African joint-tail African millet African-timothy agarista agropiro alkali grass alkali grass alpine-oat (grass)* annual wheat grass aparejo (grass) Arizona cottontop arrocillo arrowfeather Asian broom grass avena loco avenilla

Bahia grass balsam scale bamboo bamboo bamboo bamboo bamboo bamboo grass banderilla banderita barb grass barba negra barbas de indio barbwire grass barley barley barnyard grass basket grass beach grass beach grass beach wire grass beak grain beard grass beggartick grass bent (grass) bent grass Bermuda grass billion dollar grass birdseed grass birdwood grass birdwood grass biscuit grass blackoat grass blady grass blowout grass blue bunch grass blue grass

bluejoint

bluejoint

bluestem

bluestem

bluestem

bluestem

Bouteloua dimorpha Coix lacryma-jobi Cenchrus ciliaris Hemarthria altissima Eleusine corocana Setaria sphacelata Cynodon dactylon Elymus spp. Puccinellia spp. Torreyochloa spp. Helictotrichon spp. Eremopyrum spp. Muhlenbergia spp. Digitaria californica Paspalum paniculatum Aristida purpurascens Thysanolaena latifoliai Avena fatua Avena spp.

Paspalum notatum Elionurus tripsacoides Bambusa spp. Phyllostachys spp. Pseudosasa spp. Pleioblastus spp. Dendrocalamus latiflorus Thysanolaena maxima Bouteloua spp. Bouteloua curtipendula Hainardia cylindrica Heteropogon contortus Andropogon bicornis Cymbopogon refractus Hordeum spp. Elyhordeum spp. Echinochloa spp. Oplismenus hirtellus Ammophila spp. Panicum amarum Dactyloctenium aegyptium Diarrhena americana Andropogon spp. Bothriochloa laguroides Schizachyrium spp. Agropogon spp. Polypogon spp. Gymnopogon spp. Chrysopogon spp. Aristida orcuttiana Agrostis spp. Apera spp. Cynodon spp. Echinochloa frumentacea Phalaris canariensis Cenchrus biflorus Pennisetum setigerum Paspalum vaginatum Piptochaetium avenaceum Imperata cylindrica Redfieldia flexuosa Festuca idahoensis Poa spp. Elymus smithii Calamagrostis spp. Schizachyrium spp. Andropogon spp. Bothriochloa spp.

bluestem (grass) bottlebrush (grass) branching foxtails bristle grass bristly love grass bristly spear grass brome [-o] bronco grass brook grass broom-corn broom-sedge broomcorn millet browntop browntop millet Brunswick grass buckskin grass buffalo bunch grass buffalo grass buffel grass bugseed grass bull grass bull grass bunch grass bunch grass bur grass Burma reed burrero burro grass buryseed chloris bush grass button grass

California grass canary grass candy grass cane cane cane canoe grass canyon grass caña brava cape grass Carib grass Carib grass carpet grass carrizo carrizo carrizo chico carrycillo Catalina grass catchfly grass centeno silvestre centipede grass cheat (grass) chess chicken foot grass chicken-corn chloris Christmas grass citronella grass club-awn grass cockfoot cockspur grass cogon grass cola de zorra cola de zorra Colorado grass Columbia grass Columbus grass

Dichanthium spp. Elymus spp. Eustachys floridana Setaria spp. Cladoraphis cyperoides Piptochaetium setosum Bromus spp. Bromus tectorum Catabrosa aquatica Sorghum bicolor Andropogon glomeratus Panicum miliaceum Microstegium vimineum Urochloa ramosa Paspalum nicorae Urochloa arizonica Festuca campestris Buchloë dactyloides Pennisetum' ciliare Alloteropsis cimicina Paspalum boscianum Muhlenbergia emersleyi Sphenopholis spp. Festuca spp. Tragus spp. Neyraudia reynaudiana Scleropogon brevifolius Scleropogon brevifolius Enteropogon chlorideus Calamagrostis epigeois Dactyloctenium radulans

Urochloa mutica Phalaris spp. Eragrostis cilianensis Gynerium sagittatum Lasiacis spp. Arundinaria spp. Paspalum acuminatum Eriochloa lemmonii Gynerium sagittatum Tribolium obliterum Eriochloa spp. Echinochloa polystachya Axonopus spp. Arundo donax Phragmites australis Hymenachne amplexicaulis Olyra latifolia Dissanthelium californicum Leersia spp. Elymus canadensis Eremochloa spp. Bromus spp. Bromus spp. Eustachys caribaea Sorghum bicolor Chloris spp. Themeda arquens Cymbopogon citratus Corynephorus canescens Dactylis glomerata Echinochloa spp. Imperata cylindrica Hordeum jubatum Elymus elymoides Urochloa texana Paspalum fimbriatum Sorghum almum

Elymus smithii

Colusa grass common reed coral-panicum cord grass corn corn grass corn grass cotta grass cotton grass cottontop couch grass couch grass crab grass creeping leafstalk grass crinkle-awn (grass) crowfoot (grass) crown grass crypsis cup grass cup scale curly mesquite cut grass Dallis grass darnel deer grass desert grass dichanthium

dis grass
doddering dillies
dogtail grass
dogtooth grass
dogtown grass
door mat grass
double comb grass
dropseed
dropseed
dune grass
durra

elephant grass English blue grass espiga negra eulalia Eureka Valley dune grass eyelash grass

fairy grass fakahatchee grass false bluestem false brome false buffalo grass false citronella grass false manna grass false melic false needle grass false oat false Rhodes grass false rice grass feather grass feather grass feathertop feathertop feathertop fern grass fescue finger grass finger grass finger grass fish-on-a-pole grass flag grass flat crab grass flat-crab grass flechilla

Neostapfia colusana Phragmites australis Setaria chapmanii Spartina spp. Zea mays ssp. mays Setaria barbata Setaria barbata Cottea pappophorioides Digitaria californica Digitaria spp. Elymus repens Cynodon dactylon Digitaria spp. Pharus lappulaceus Trachypogon spp. Dactyloctenium aegyptium Paspalum spp. Crypsis spp. Eriochloa spp. Sacciolepis ssp. Hilaria spp. Leersia spp. Paspalum dilatatum Lolium spp. Muhlenbergia rigens Blepharidachne bigelovii Dichanthium spp.

Ampelodesmos mauritanica
Briza media
Cynosurus spp.
Cynodon aethiopicus
Aristida purpurea
Nardus stricta
Dactyloctenium geminatum
Sporobolus spp.
Blepharoneuron tricholepis
Elymus mollis
Sorghum bicolor

Pennisetum purpureum Festuca pratensis Hilaria belangeri Miscanthus spp. Swallenia alexandrae Blepharidachne kingii

Miscanthus nepalensis Tripsacum dactyloides Schizachyrium spp. Brachypodium spp. Monroa squarrosa Cymbopogon nardus Torreyochloa californica Schizachne purpurascens Oryzopsis kingii Trisetum spp. Chloris pluriflora Stipa papposa Pennisetum spp. Stipa neomexicana Calamagrostis epigeios Pennisetum villosum Calamagrostis epigeios Catapodium rigida Festuca spp. Digitaria spp. Eustachys spp. Chloris spp. Chasmanthium latifolium Sporobolus compositus Axonopus furcatus Axonopus furcatus Stipa spp.

Florida grass
Florida rhaphis
fluff grass
fluff grass
fly-away grass
fountain grass
fowl meadow grass
foxtail
foxtail
foxtail millet
fragile grass
fragile-oat
French oat grass
funeral grass

galleta gama grass German millet giant cut grass giant reed giant rye grass Glenwood grass goat grass golden-timothy goldentop goober grass goose grass goose grass gordura grader grass grama (grass) gray hair grass Guatemala grass guinea grass gyp grass

Habana oat grass hair grass hair grass hair grass hair grass hair grass hairy oat grass hairy-nerve grass Hakone grass hard grass hard grass Harding grass hare's tail heath grass hedgehog grass herd grass herringbone grass Hilo grass hoe grass hog millet holy grass hurrah grass hurricane sour grass

ice grass
Indian corn
Indian grass
Indian millet
Indian rice grass
Indian woodoats
Italian millet
itch grass

Japanese forest grass Japanese lawn grass jaraguá grass Java (-nese) grass Job's tears Cynodon transvaalensis
Chrysopogon pauciflorus
Erioneuron pulchellum
Tridens spp.
Agrostis hiemalis
Pennisetum spp.
Glyceria spp.
Alopecurus spp.
Setaria italica
Aegopogon spp.
Gaudinia fragilis
Gaudinia fragilis
Stipa arida

Hilaria spp. Tripsacum spp. Setaria italica Zizaniopsis miliacea Arundo donax Festuca gigantea Sacciolepis indica Aegilops spp. Setaria sphacelata Lamarckia aurea Amphicarpum spp. Eleusine indica Acrachne racemosa Melinis minutiflora Themeda quadrivalvis Bouteloua spp. Corynephorus canescens Tripsacum laxum Urochloa spp. Sporobolus nealleyi

Themeda quadrivalvis Muhlenbergia spp. Deschampsia spp. Aira spp. Corynephorus canescens Agrostis spp. Rytidosperma spp. Trichoneura elegans Hakonechloa macra Sclerochloa dura Parapholis spp. Phalaris aquatica Lagurus ovatus Danthonia decumbens Cenchrus echinatus Agrostis alba Pogonarthria squarrosa Paspalum conjugatum Muhlenbergia porteri Panicum miliaceum Anthoxanthum spp. Paspalum setaceum Bothriochloa pertusa

Phippsia algida
Zea mays ssp. mays
Sorghastrum spp.
Stipa hymenoides
Stipa hymenoides
Chasmanthium latifolium
Setaria italica
Rottboellia cochinchinensis

Hakonechloa macra Zoysia japonica Hyparrenhia rufa Polytrias amaura Coix lacryma-jobi Johnson grass joint grass joint head joint-tail June grass June grass jungle-rice

Kafir-corn Sorghum bicolor kangaroo grass Themeda quadrivalvis Bouteloua kayii Kay's grass Kelly grass Rottboellia cochinchinensis Monanthochloë littoralis key grass khas-khas Chrysopogon zizanioides khus-khus Chrysopogon zizanioides kikuyu grass Pennisetum clandestinum Dichanthium annulatum Kleberg grass Klein grass Panicum coloratum Paspalum notatum knot grass kodo millet Paspalum scrobiculatum Koeler's grass Koeleria asiatica koeleria Koeleria spp. Korean temple grass Zoysia japonica kyasuma grass Pennisetum pedicellatum

Sorghum halepense

Arthraxon hispidus

Echinochloa colona

Eragrostis capillaris

Pharus lappulaceus

Pennisetum orientale

Ctenium aromaticum

Cymbopogon citratus

Hemarthria altissima

Brachyelytrum aristosum

Urochloa spp.

Paspalum lividum

Eragrostis spp.

Bromus hordeaceus

Bouteloua juncea

Coelorachis spp.

Coelorachis spp.

Poa pratensis

Koeleria spp.

lace grass
lamilla
Laurisa grass
leafstalk grass
lemon grass
lemon grass
limpo grass
liverseed grass
long-awned wood grass
longtom
lop grass
love grass

millet

millet

millet

millet grass

lyme grass Elymus spp. Madagascar grass Neyraudia arundinacea Phyllostachys bambusoides madake maicillo Tripogon spicatus maiden cane Amphicarpum spp. maiden cane Panicum hemitomon maize Zea mays ssp. mays Manila grass Zoysia spp. Glyceria spp. manna grass Ammophila arenaria marram grass marram grass Calammophila baltica marsh grass Scolochloa festucacea Spartina patens marsh hay marsh millet Zizaniopsis miliacea mary grass Setaria barbata Zoysia tenuifolia Mascarene grass Nardus stricta mat grass mat grass Axonopus spp. matronella grass Zoysia spp. Mauritian grass Apluda mutica Phalaris caroliniana may grass meadow grass Poa spp. Mediterranean grass Schismus spp. medusa head (grass) Elymus caput-medusae Melica spp. melic Memphis grass Cutandia memphitica metake Pseudosasa japonica Mexican teosinte Zea perennis millet Paspalum spp. millet Setaria spp. millet Urochloa spp. millet Pennisetum spp.

milo
mission grass
molasses grass
moor grass
moor mat grass
mosquito grass
moss grass
mountain bunch grass
mountain-rice
mountain-rice
muhly
Munro grass
muraina grass
mutton grass

Napier grass nard grass Natal grass navajita needle grama needle grass needle grass needle grass needle-and-thread (grass) nimblewill nit grass noble cane North Africa grass

oat
oat grass
oat grass
oat grass
oat grass
oat grass
oat meal grass
oil grass
one-glumed hard grass
onion grass
orange grass
orchard grass
Orcutt grass
Ozark grass

paja mansé palisade grass palm grass pampas grass pangola grass panic grass panic grass panicum panicum pappus grass pappus grass Pará grass paspalidium paspalum pasto chino pata de gallo pato de gallo peanut grass pearl millet pendant grass pigeon grass pili grass pine grass pine grass pit grass pitscale grass plume grass plume grass plush grass poison darnel

Sorghum bicolor
Pennisetum polystachyon
Melinis minutiflora
Molinia caerulea
Nardus strictus
Dasypyrum villosum
Coleanthus subtilis
Festuca arizonica
Stipa webberi
Oryzopsis spp.
Muhlenbergia spp.
Panicum rigidulum
Ischaemum spp.
Poa fendleriana

Pennisetum purpureum
Cymbopogon nardus
Melinis repens
Bouteloua spp.
Bouteloua aristidoides
Stipa spp.
Piptochaetium spp.
Aristida spp.
Stipa comata
Muhlenbergia schreberi
Gastridium ventricosum
Saccharum officinarum
Ventenata dubia

Avena spp.
Trisetum spp.
Piptochaetium spp.
Danthonia spp.
Arrhenatherum spp.
Leersia lenticularis
Cymbopogon iwarancusa
Hainardia cylindrica
Melica spp.
Ctenium aromaticum
Dactylis glomerata
Orcuttia spp.
Limnodea arkansana

Paspalum quadrifarium Urochloa brizantha Setaria palmifolia Cortaderia spp. Digitaria decumbens Panicum spp. Steinchisma hians Urochloa spp. Panicum spp. Enneapogon desvauxii Pappophorum spp. Urochloa mutica Setaria spp. Paspalum spp. Buchloë dactyloides Cynodon dáctylon Chloris cucullata Amphicarpum purshii Pennisetum americanum Arctophila fulva Setaria pumila Heteropogon contortus Festuca arizonica Calamagrostis spp. Hackelochloa granularis Hackelochloa granularis Miscanthus spp. Saccharum spp. Chloris radiata Lolium temulentum Arctagrostis latifolia

polar grass

Panicum spp.

Eleusine spp.

Milium effusum

Echinochloa spp.

Eragrostis hypnoides Muhlenbergia asperifolia pony grass scratch grass popotillo Andropogon spp. Scribner's grass Scribneria bolanderi Elymus mollis popotillo colorado Schizachyrium scoparium sea lyme grass popotillo del pinar Blepharoneuron tricholepis sea-oat Uniola paniculata Pleuropogon spp. porcupine grass Stipa spp. semaphore grass Porter's needle grass Ptilagrostis porteri shore grass Monanthochloë littoralis Sporobolus spp. short husk (grass) Brachyelytrum spp. poverty grass poverty-oats Danthonia spp. Siberian grass Eremopoa persica Parapholis spp. prickle grass Crypsis spp. sickle grass prickle grass Scolochloa festucacea signal grass Brachiaria eruciformis signal grass silk grass Urochloa spp. prickle-fescue Scolochloa festucacea proso millet Panicum miliaceum Stipa hymenoides silk reed pull-and-be-damned Paspalum lividum Neyraudia reynaudiana puna grass Stipa brachychaeta silky scale Anthaenantia spp. purple needle grass Triraphis mollis silky-bent Apera spp. purple heads Triraphis mollis silver beard grass Bothriochloa spp. Tridens flavus Miscanthus spp. purpletop silver grass Silveus grass Trichoneura elegans quack grass Elymus repens skeleton grass Gymnopogon spp. quack grass Elyhordeum spp. skunk grass Eragrostis cilianensis quaking grass Briza spp. sleepy grass Stipa robusta slender grass quick grass Elymus repens Leptochloa panicea ssp. mucronata quitch grass Elýmus repens slough grass Spartina spp. slough grass Beckmannia syzigachne small cane Lasiacis spp. rabbit foot grass Polypogon monspeliensis ragi Eleusine corocana small carpet grass Arthraxon hispidus Microchloa kunthii rat's tail Sporobolus jacquemontii small grass rattlesnake chess Bromus briziformis smut grass Sporobolus indicus Briza maxima snow grass Phippsia spp. rattlesnake grass Ravenna grass Saccharum ravennae soap grass Cymbopogon refractus Tridens flavus sorghum Sorghum bicolor redtop redtop Agrostis spp. sorgo [-gho] Sorghum bicolor reed (grass) reed (grass) sour grass Bothriochloa pertusa Cinna spp. Phragmites australis sour grass Digitaria insularis reed (grass) Calamagrostis spp. South African bluestem Hyparrhenia hirta reed bent grass Calamagrostis spp. southern wild-rice Zizaniopsis miliacea Poa spp. reed canary grass Phalaris arundinacea spear grass reed grass Calammophila spp. spider grass Aristida ternipes Reimarochloa oligostachya spike bur grass Tragus spp. reimar grass Elionurus barbiculmis rescue grass Bromus catharticus spike grass Chloris spp. spike grass Chasmanthium spp. Rhodes grass rice Oryza sativa spike-oat Helictotrichon hookeri Piptochaetium spp. Tuctoria spp. spiral grass rice grass Oryzopsis spp. Stipa spp. rice grass sprangletop Leptochloa spp. rice grass squirreltail Hordeum spp. ring grass Muhlenbergia spp. squirreltail Elymus spp. Bromus rigidus St. Augustine grass Stenotaphrum secundatum ripgut grass river grass Echinochloa polystachya star grass Cynodon plectostachyus Chasmanthium latifolium Stebbins' grass Ehrharta calycina river-oats rooi grass Themeda triandra stiff brome Brachypodium spp. rosette grass stiff grass Panicum spp. Catapodium rigidum ruby grass Melinis repens stink grass Eragrostis cilianensis rush grass Sporobolus spp. stipa Stipa spp. rye Secale spp. strand-wheat Elymus arenarius Lolium spp. Sudan grass Sorghum bicolor rye grass rye grass Elymus spp. Sudan negro Sorghum almum Saccharum spp. sugar cane sabi grass Urochloa mosambicensis summer grass Alloteropsis cimicina sacaton Sporobolus spp. swamp oat grass Sphenopholis pensylvanica salt and pepper grass Deschampsia caespitosa swamp-timothy Crypsis schoenoides salt grass Allolepis texana Glyceria fluitans sweet grass salt grass Distichlis spp. sweet grass Glyceria septentrionalis sweet grass Anthoxanthum ssp. salt grass Sporobolus airoides salt meadow grass Leptochloa panicoides switch cane Arundinaria gigantea sand bunch grass Panicum virgatum Stipa hymenoides switch grass sand bur(r) Cenchrus spp. Triplasis spp. sand grass tabosa Hilaria mutica sand grass Mibora minima talquezal Paspalum virgatum sand reed Calamovilfa spp. tanglehead Heteropogon spp. sandbur grass Cenchrus spp. tanner grass Urochloa arrecta Cenchrus gracillimus Eragrostis tef sandspur grass teff satin grass Muhlenbergia racemosa teosinte Zea mays satintail Imperata spp. Texas crab grass Schedonnardus paniculatús

Texas grass

Panicum bulbosum

Schismus spp.

schismus

Texas grass Vaseyochloa multinervosa Texas millet Urochloa texana Texas wild-rice Zizaniopsis miliacea Texas winter grass Stipa leucotricha Texas-panicum Urochloa texana Texas-timothy Lycurus phleoides thatch grass Hyparrhenia hirta thatching grass Hyparrhenia spp. thimble grass Fingerhuthia africana thintail grass Coelorachis spp. Hainardia cylindrica Paspalum distichum thintail grass Thompson grass three-awn grass Aristida spp. three-spike grass Eleusine tristachya tibisee Lasiacis spp. tick grass Eragrostis echinochloidea tickle grass Aira spp. tickle grass Agrostis spp. tiger grass Thysanolaena latifolia timothy Phleum spp. tobosa Hilaria mutica tobosa menudo Hilaria belangeri toothache grass Ctenium spp. torpedo grass Panicum repens tres barbas Aristida oligantha tridens Tridens spp. tridens Erioneuron spp. trisetum Trisetum spp. triticale Triticosecale rimpaui trompetilla Hymenachne amplexicaulis tuctoria Tuctoria spp. Schedonnardus paniculatus tumble grass tumble grass Eragrostis spectabilis tundra grass Dupontia fischeri tussock grass Stipa neesiana umbrella grass Enteropogon spp.

umbrella grass Enteropogon spp. uniola Chasmanthium spp. uva (grass) Gynerium sagittatum

Anthoxanthum spp. vanilla grass Vasey grass Paspalum urvillei Ehrharta spp. veldt grass velvet grass Holcus spp. vernal grass Anthoxanthum spp. vetiver grass Chrysopogon zizanioides vine-mesquite Panicum obtusum viper grass Dinebra retroflexa

wallaby grass Rytidosperema spp. wallaby grass Danthonia spp. water grass Echinochloa spp. water grass Luziola spp. water millet Zizaniopsis miliacea water-rice Zizania spp. waterside-reed Pennisetum macrourum wedge grass Sphenopholis spp. weeping grass Ehrharta calycina West Indian marsh grass Hymenachne amplexicaulis wheat Triticum spp. wheat grass Agropyron spp. wheat grass Elymus spp. white grass Leersia virginica whorl grass Catabrosa aquatica wild cane Gynerium sagittatum wild rye X Elyhordeum ssp. wild rye Elymus spp. wild-oats Chasmanthium spp. wild-rice Zizania ssp. Muhlenbergia glomerata wild-timothy wind grass Apera interrupta windmill grass Eustachys spp. windmill grass Chloris spp.

winter grass

Stipa spp. winter grass wire grass Eleusine indica Schedonnardus paniculatus wire grass wire stem Muhlenbergia mexicana witch grass Ďigitaria spp. witch grass Panicum spp. wolftail Lycurus phleoides wood grass Oplismenus hirtellus wood grass Sorghastrum spp. wood reed Cinna spp. wood-millet Milium effusum wood-oats Chasmanthium spp. woolly grass Erioneuron spp.

yard grass yellow-oat Yorkshire fog

zacata ovillo Dactylis glomerata zacate araña Muhlenbergia porteri zacate burro Scleropogon brevifolius zacate colorado Heteropogon contortus zacate del amor Eragrostis curvula zacate gigante Leptochloa dubia zacate guta Panicum obtusum zacate Ilorón Eragrostis curvula Tripogon spicatus zacate maicero zacate pelillo Chloris pluriflora zacate punta blanca Digitaria californica zacate triguillo Elymus elymoides zacatón alcalino Sporobolus airoides zacatón arenoso Sporobolus cryptanthus Miscanthus sinensis zebra grass zig-zag grass Panicum dichotomiflorum Zoysia spp. zoisia Zoysia spp. zoysia Zulu-fescue Fingerhuthia africana

* (grass) indicates that the word grass is sometimes used as part of the common name.

Revised: 12 January 2005

Eleusine indica

Trisetum spp.

Holcus lanatus

Poa annua

4.07 - THE H & C MANUAL: AN UPDATE

Why are we using a book that first appeared in 1935 and then revised in 1951? Half a century is a long time, even in systematics! The answer is amazingly simple -- nothing has come along to take its place. Hitchcock & Chase remains the only comprehensive treatment of the grasses of the United States. The late Frank Gould, John Kartesz, and I have compiled checklists for the United States and for North America, but they are nothing more than that.

But, help is on the way! In 2003, Oxford University Press published the first of a two-volume treatment of the grasses of North America. It will be part of the larger "Flora of North America" project. The FNA is a first-class operation. The other volume is scheduled to appear in 2006. Now the bad news. The volumes will cost about \$100 each! There is a chance that a one volume, scaled-down version may become available.

Just how out-of-date are the names of subfamilies, tribes, genera, and individual grasses? In this section of the syllabus you will see a comparison of the subfamilies, tribes, and genera used in The Manual (left entry) versus today's version of the truth (right entry). You will also have access to a notebook in the lab that will permit you to look up any species name used in The Manual to see if it is still current. Of course, I am the person who compiled this list and other experts would not always agree with me.

TWO VERY DIFFERENT VIEWS

HITCHCOCK & CHASE (The Old Testament)

Subfamily: Festucoideae

Tribe: Bambuseae Tribe: Festuceae Tribe: Hordeae Tribe: Aveneae Tribe: Agrostideae Tribe: Zoysieae Tribe: Chĺorideae Tribe: Phalarideae Tribe: Oryzeae Tribe: Zizanieae

Subfamily: Panicoideae Tribe: Melinideae Tribe: Paniceae Tribe: Andropogoneae Tribe: Tripsaceae

> **GRASS PHYLOGENY GROUP** (The New Testament)

Subfamily: Bambusoideae

Tribe: Bambuseae Tribe: Olyreae Subfamily: Pharioideae Tribe: Phareae

Subfamily: Ehrhartoideae

Tribe: Oryzeae Tribe: Ehrharteae

Subfamily: Centothecoideae

Tribe: Centotheceae

Subfamily: PooideaeTribe: Triticeae
Tribe: Brachypodieae Tribe: Bromeae Tribe: Poeae

Tribe: Ampelodesmeae

Tribe: Meliceae Tribe: Stipeae Tribe: Nardeae Tribe: Brachyelytreae Tribe: Diarrheneae Subfamily: Arundinoideae

Tribe: Arundineae Subfamily: Danthonioideae

Tribe: Danthonieae Subfamily: Aristidoideae Tribe: Aristideae

Subfamily: Chloridoideae

Tribe: Pappophoreae Tribe: Orcuttieae Tribe: Cynodonteae Tribe: Eragrostideae Tribe: Leptureae Subfamily: Panicoideae

Tribe: Paniceae Tribe: Andropogoneae

SUBFAMILIES

As you will see from the table below, the subfamily Panicoideae of H & C came through unscathed, but look at what happened to their Festucoideae. One large subfamily became nine.

Festucoideae

Pooideae Pharoideae Bambusoideae Erhartoideae Centothecoideae Chloridoideae Aristidoideae Arundinoideae Danthonioideae

Panicoideae Panicoideae

TRIBES

At the tribe level, three things have happened. Some have been merged (Melinidae with Paniceae), some have been been split into smaller tribes (look at Festuceae!), and some are again unchanged.

Agrostideae Aveneae

Stipeae Eragrostideae Brachyelytreae Áristideae Andropogoneae

Andropogoneae Aveneae Bambuseae Chlorideae

Festuceae

Aveneae Bambuseae Cynodonteae Eragrostideae Poeae Eragrostideae

Orcuttieae Pappophoreae Arundineae Danthonieae Aeluropodieae Diarrheneae Bromeae Aveneae Meliceae Ampelodesmeae Triticeae Triticeae Brachypodieae Hainardieae Poeae Paniceae Oryzeae Paniceae Aveneae

Maydeae

Cynodonteae

Oryzeae

Hordeae

Melinidae Oryzeae Paniceae Phalarideae Tripsaceae Zizanieae Zoysieae

GENERA

The genera below are arranged as they are in H & C. The entry in the right-hand column tells you whether the genus is still recognized and its taxonomic position.

AVENEAE

Unchanged Aira Unchanged Arrhenatherum Unchanged Avena Corynephorus Unchanged Danthonia Danthonioideae: Danthonieae Deschampsia Unchanged Helictotrichon Unchanged Unchanged Holcus Unchanged Danthonioideae: Danthonieae Koeleria Schismus Danthonia Sieglingia Sphenopholis Unchanged Unchanged Trisetum

BAMBUSEAE

Arundinaria Bambusoideae: Bambuseae

FESTUCEAE

Ampelodesmos Unchanged Chloridoideae: Eragrosteae Blepharidachne Brachypodium Pooideae: Triticeae Briza Unchanged Pooideae: Bromeae Unchanged **Bromus** Catabrosa Cortaderia Danthonioideae: Danthonieae Chloridoideae: Pappophoreae Cottea Cutandia Unchanged Cynosurus Unchanged Unchanged Dactvlis Unchanged Desmazeria Pooideae: Diarrheneae Diarrhena Pooideae: Aveneae Dissanthelium Chloridoideae: Aeluropodieae Distichlis Enneapogon Chloridoideae: Pappophoreae Eragrostis Chloridoideae: Eragrosteae Unchanged Festuca Glyceria Pooideae: Meliceae

Hesperochloa Unchanged Lamarckia Unchanged Pooideae: Meliceae Melica Arundinoideae: Arundineae Molinia Chloridoideae: Aeluropodieae Monanthochloe Monroa Chloridoideae: Eragrosteae Chloridoideae: Eragrosteae Chloridoideae: Eragrosteae Neostapfia Neyruadia Orćuttia Chloridoideae: Orcuttieae Pappophorum Chloridoideae: Pappophoreae Arundinoideae: Arundineae Pooideae: Meliceae Phragmites Pleuropogon Unchanged Poa Unchanged Puccinellia Chloridoideae: Eragrosteae Pooideae: Meliceae Redfieldia Schizachne Sclerochloa Unchanged Unchanged [Desmazeria] Scleropoa Scleropogon Chloridoideae: Eragrosteae Scolochloa Unchanged Swallenia Chloridoideae: Aeluropodieae Chloridoideae: Eragrosteae Tridens Chloridoideae: Eragrosteae Chloridoideae: Eragrosteae **Triplasis** Uniola Chloridoideae: Eragrosteae Vaseyochloa

AGROSTIDEAE

Agrostis Unchanged Alopecurus Unchanged Ammophila Unchanged Unchanged Apera Aristida Aristidoideae: Aristideae Blepharoneuron Chloridoideae: Eragrosteae Pooideae: Brachyelytreae Brachyelytrum Calamagrostis Uńchanged Calamovilfa Chloridoideae: Eragrosteae Cinna Unchanged Pooideae: Poeae Chloridoideae: Eragrosteae Coleanthus Crypsis Gastridium Unchanged Chloridoideae: Eragrosteae [Crypsis] Heleochloa Lagurus Unchánged Unchanged Limnodea Lycurus Chloridoideae: Eragrosteae Pooideae: Stipeae Milium Chloridoideae: Eragrosteae Pooideae: Stipeae Pooideae: Poeae Muhlenbergia Oryzopsis Phíppsia Unchanged Phleum Pooideae: Stipeae Piptochaetium Unchanged Polypogon Sporobolus Chloridoideae: Eragrostideae Stipa Pooideae: Stipeae

ANDROPOGONEAE

CHLORIDEAE Phalaris Aveneae

Beckmannia Pooideae: Aveneae Chloridoideae: Cynodonteae Bouteloua Chloridoideae: Cynodonteae Buchloe Chloridoideae: Cynodonteae Chloridoideae: Cynodonteae Chloridoideae: Cynodonteae Chloridoideae: Cynodonteae Chloridoideae: Cynodonteae Cathestecum Chloris Ctenium Cynodon Dactyloctenium Chloridoideae: Eragrostideae Chloridoideae: Eragrostideae Chloridoideae: Cynodonteae Chloridoideae: Eragrostideae Eleusine Gymnopogon Léptochloa Chloridoideae: Cynodonteae Microchloa Chloridoideae: Cynodonteae
us Chloridoideae: Cynodonteae
Chloridoideae: Cynodonteae
Chloridoideae: Cynodonteae
Chloridoideae: Cynodonteae Munroa Schedonnardus Spartina Trichloris Chloridoideae: Eragrostideae Chloridoideae: Eragrostideae Trichoneuron Tripogon Chloridoideae: Cynodonteae Wilkommia

TRIPSACEAE

CoixAndropogoneaeEuchlaenaZeaTripsacumAndropogoneaeZeaAndropogoneae

ZIZANIEAE

Hydrochloa Luziola
Luziola Erhartoideae: Oryzeae
Pharus Pharoideae: Phareae
Zizania Erhartoideae: Oryzeae
Zizaniopsis Erhartoideae: Oryzeae

ZOYSIEAE

Aegopogon Chloridoideae: Cynodonteae Hilaria Chloridoideae: Cynodonteae Tragus Chloridoideae: Cynodonteae Coysia Chloridoideae: Cynodonteae

HORDEAE

Aegilops Unchanged Agropyron Unchanged Unchanged Elymus Unchanged Hordeum Unchanged Hvstrix Pooideae: Poeae Lolium Pooideae: Hainardieae Monerma Pooideae: Hainardieae Parapholis Scribneria Pooideae: Hainardieae Secale Unchanged Sitanion Unchanged Triticum Unchanged

MELINIDAE

Melinis Paniceae

ORYZEAE

Leersia Erhartoideae: Oryzeae Oryza Erhartoideae: Oryzeae

PANICEAE

Amphicarpum

Unchanged Anthaenantia Unchanged **Axonopus** Brachiaria Unchanged Unchanged Cenchrus Unchanged Digitaria Echinochloa Unchanged Eriochloa Unchanged Unchanged Lasiacis Digitaria Leptoloma Bambusoideae: Olyreae Olyra Unchanged Oplismenus Unchanged Panicum Paspalum Unchanged Unchanged Pennisetum Reimarochloa Unchanged Rhynchelytrum Melinis Sacciolepis Unchanged Unchanged Setaria Stenotaphrum Unchanged Digitaria Trichachne

PHALARIDEAE

Anthoxanthum Aveneae Hierochloe Anthoxanthum

Unchanged

4.08 - FORAGE GRASSES

There is another vastly important group of grasses -those that we feed to our domesticated animals,
especi-ally horses, dairy cattle and beef cattle.

Forage is the general term for plants consumed by
livestock. In the broad sense, the term includes
pasture and browse plants, straw, hay, and silage.
Another way of looking at it is that we use dairy and
beef cattle to transform plants into meat, milk, and
other dairy products. In the United States alone,
forage crops constitute a multibillion dollar industry.
It is estimated that more than half of the earth's land
surface is devoted to pastures and meadows used for
grazing by farm animals.

The high cellulose levels of grass stems and leaves make these tissues relatively difficult for most animals to digest. However, the bacteria that inhabit the intestinal tracts of both ruminants and nonruminants carry out a fermentation process that reduces the cellulose to simpler compounds. We also create an environment in which anaerobic fermentation can occur when we put silage into a silo. For all practical purposes, chopped up plant material is pickled by being bathed in organic acids that are produced by the bacteria. If done properly, silage can be stored for years. It certainly doesn't sound very appetizing, does it? Do you like sauerkraut? How is it prepared?

Although literally thousands of plant species can provide palatable food for our domesticated animals, all of the important forage plants are either grasses or legumes. Most of them are Old World introductions, especially from Europe and Africa.

SELECTED REFERENCES

Barnes, R. F. et al. 1995. Forages. Vol. 1. An introduction to grassland agriculture. Fifth edition. Iowa State Univ. Press. Ames. 516 pp.

Heath, M., D. Metcalfe, & R. Barnes (editors). 1973. Forages: the science of grassland agriculture. Third edition. Iowa State Univ. Press. Ames.

Hodgson, H. J. 1976. Forage crops. Sci. American 234(2): 61-68.

Janick, J. et al. 1974. Plant science: an introduction to world crops. Second edition. W. H. Freeman. San Francisco, CA. Pp. 443-454.

Judd, B. I. 1979. 1979. Handbook of tropical forage grasses. Garland STPM Press. New York, NY. 116 pp.

Looman, J. 1983. 111 range and forage plants of the Canadian prairies. Publ. No. 1751. Agriculture Canada. Ottawa. 255 pp.

Phillips Petroleum Co. 1963. Pasture and range plants. Bartlesville, OK. 176 pp.

Simpson, B. B. & M. C. Ogorzaly. 1995. Economic botany: plants in our world. Second edition. McGraw-Hill. New York, NY. Pp. 198-200.

Sprague, H. B. (editor). 1974. Grasslands of the United States: their economic and ecologic importance. Iowa State Univ. Press. Ames.

Stechman, J. V. 1977. Common western range plants. Second edition. Vocational Education Prod. California Polytechnic State Univ. San Luis Obispo. 164 pp.

Stubbendieck, J., S. L. Hatch, & C. H. Butterfield. 1992. North American range plants. Fourth edition. Univ. Nebraska Press. Lincoln. 493 pp.

FORAGE GRASSES

Bahia grass [Paspalum notatum] Bent grass [Agrostis spp.] Bermuda grass [Cynodon dactylon] Big bluestem [Andropgon gerardii] Black grama [Bouteloua eriopoda]

Blue grama grass [Bouteloua gracilis]
Brome grasses [Bromus spp.]
Buffalo grass [Buchloë dactyloides]
Bush muhly [Muhlenbergia porteri]
Crested wheat grass [Agropyron cristatum]

Dallis grass [Paspalum dilatatum] Fescues [Festuca spp.] Gama grass [Tripsacum dactyloides] Hair grass [Deschampsia cespitosa] Harding grass [Phalaris aquatica]

Indian grass [Sorghastrum nutans]
June grass [Koeleria macrantha]
Kentucky bluegrass [Poa pratensis]
Little bluestem [Schizachyrium scoparium]
Love grasses [Eragrostis spp.]

Maiden cane [Panicum hemitomon]
Onion grass [Melica bulbosa]
Orchard grass [Dactylis glomerata]
Pangola grass [Digitaria decumbens]
Pearl millet [Pennisetum americanum]

Redtop [Agrostis stolonifera] Reed canary grass [Phalaris arundinacea] Rye grasses [Lolium spp.] Sidoats grama [Bouteloua curtipendula] Sorghum [Sorghum bicolor]

Spike trisetum [Trisetum spicatum]
Sudan grass [Sorghum sudanense]
Timothy [Phleum pratense]
Wheat grasses [Agropyron spp.]
Wild oats [Avena fatua]
Wild ryes [Elymus spp.]

Used especially in the southeastern U. S. Excellent forage; common in western states Also an aggressive weed and lawn grass Provides excellent forage Important southwestern species

Best while immature Very important in dry, cool regions Native grass of cool, dry prairies Native southwestern species Good in cool, dry areas

Well adapted to Cotton Belt states and California's C. Valley
Well adapted to warm summers
Excellent forage; southeastern states
Used in northern and western states
Across the southern states

Excellent forage; eastern and central states Commonly used, except in the Southeast One of the best and most palatable Best when immature Widely used in southern Great Plains

Native grass of the Southeast coastal plain Used especially in Northwestern states; native Does well in cool, humid regions Popular in Florida; native to Africa Also a major human food in Africa and India

One of the best wetland forage grasses
Well adapted to wet areas
Winter/irrigated pasture in southern states and California
Provides forage over much of the country
Also used for grain, silage, and syrup

Used in the West and in the Northeast
Hybrids with sorghum widely used
Eurasian; widely planted
Good in cool, dry regions
Commonly used, except in southeastern states
Natives of the Pacific Northwest

[With thanks to Prof. K. O. Fulgham for his assistance.]

4.09 - WEEDY GRASSES

"What is a weed? A plant whose virtues have not yet been discovered." (Ralph Waldo Emerson)

Weeds are of great economic importance, mostly in the negative sense. It is estimated that weeds cost the American farmer several billion dollars each year by reducing both the quantity and the quality of crops produced. Their damage causes a loss as large as insect injury and disease combined. Another reason for studying weeds is their intimate association with our own species. Many of them are essentially our wards and they would perish without our encouragement. As Edgar Anderson said, "... the history of weeds is the history of man."

A DEFINITION

There are many definitions of a weed. Inherent in most of them is the idea that a plant is a weed if it is growing where we do not wish it to be. The picture of a well-manicured lawn dotted with dandelions comes easily to mind. There are problems with this approach. If I am growing irises, then a rose that appears in my garden is a weed. Bermuda grass is a highly prized lawn grass in much of the southern United States. Elsewhere it tends to live in disturbed areas. Is Bermuda grass a weed?

A good botanical definition of a weed is that of Herbert Baker, a botanist at the University of California at Berkeley. A plant is a weed, "... if, in any specified geographical area, its populations grow entirely or predominately in situations markedly disturbed by man (without, of course, being deliberately cultivated plants)." Remember that disturbed sites include not only relatively undesirable vacant lots and roadsides, but also our prime agricultural lands. Some weeds invade one or the other; some live in both.

Weeds are such a problem in the agricultural states that there is legislation against them. Many states have weed laws that require the farmer to use varying degrees of control against weedy plants. The "primary noxious weeds" are considered so bad that the land owner is required to destroy them if he discovers them on his property.

CHARACTERISTICS

There are certain biological features that many weedy plants have. Many grasses are excellent weeds because they:

- can persist from year to year in an area;
- reproduce vegetatively, by such means as rhizomes or stolons that allow the plants to spread quickly and efficiently;
- have seeds that can germinate in many different environments;
- have high seed production;
- set seed in a wide variety of conditions;

- have rapid seedling growth;
- have a "general purpose" set of genes that will enable the plants to compete very effectively against native plants when they are competing on disturbed sites;
- often contain multiple genomes derived through hybridization;
- are self-pollinated or substitute some asexual means of reproduction for a sexual one; and
- may be unpalatable or even toxic to livestock or herbivores.

If we were to construct the "perfect weedy grass," it would:

- be physically attractive
- mimic a crop
- be a perennial
- reproduce both sexually and asexually
- have long-lived seeds
- produce numerous seeds over a long period
- germinate its seeds early in the growing season
- germinate its seeds in many environments
- have rapid seedling growth
- be unpalatable to livestock
- have chemical/physical defense mechanisms
- thrive in disturbed habitats.

Can we say anything positive about weeds? Certainly! In ruined and abandoned areas, weeds make up much of the flora. Many of the more attractive plants that city folks see these days are weeds. They also retard or prevent erosion along many of our roadsides.

NOXIOUS GRASSES & SEDGES OF CALIFORNIA*

Aegilops cylindricaJointed goat grass [B]Aegilops ovataOvate goat grass [B]Aegilops triuncialisBarb goat grass [B]Elymus caput-medusaeMedusa head grass [C]Elymus repensQuack grass [B]

Cenchrus echinatus
Cenchrus incertus
Cenchrus longispinus
Cypodon
Cyperus esculentus

Southern sand bur [C]
Coast sand bur [C]
Mat sand bur [C]
Bermuda grass [C]
Yellow nut grass [B]

Cyperus rotundus Purple nut grass [B]
Heteropogon contortus Tanglehead [A]
Imperata cylindrica Satintail [B]
Muhlenbergia schreberi Nimblewill [B]
Oryza rufipogon Perennial wild red rice [B]

Panicum antidotale Blue panic grass [B]
Pennisetum clandestinum Kikuyu grass [C]
Setaria faberi Giant foxtail [B]
Sorghum halepense Johnson grass [C]
Stipa brachychaeta Puna grass [A]

Ratings:

- A = Subject to state enforced action involving eradication, quarantine, containment, rejection, or other holding action at state or county level
- B = Subject to eradication, containment, control, or other holding action at the discretion of the county commissioner
- C = Not subject to state enforced action, except to retard spread

[Source: California Dept. of Food & Agriculture]

GRASSES: EXOTIC PEST PLANT LIST

A. Pest Plants of Greatest Ecological Concern:

Aegilops triuncialis (barbed goat grass) (slender wild oat) Avena barbata (wild oat) Avena fatua var. fatua Brachypodium distachyon (false brome) Bromus diandrus (ripgut brome) Lolium multiflorum (Italian ryegrass) (Mediterranean grass) Schismus arabicus Schismus barbatus (Mediterranean grass)

B. Potential to Spread Explosively:

Spartina anglica (cord grass) Spartina densiflora (dense-flowered cord grass) Spartina patens (salt-meadow cord grass)

C. Most Invasive Pest Plants (Widespread):

Ammophila arenaria
Arundo donax
Bromus tectorum
Cortaderia jubata
Cortaderia selloana
Elymus caput-medusae
Pennisetum setaceum
(European beach grass)
(giant reed)
(cheat grass)
(Andean pampas grass)
(pampas grass)
(medusa head)
(fountain grass)

D. Most Invasive Pest Plants (Regional):

Bromus rubens (red brome)
Ehrharta calycina (veldt grass)
Spartina alterniflora (smooth cord grass)

E. Wildland Plants of Lesser Invasiveness:

Ehrharta erecta (veldt grass)
Festuca arundinacea (tall fescue)
Holcus lanatus (velvet grass)
Phalaris aquatica (Harding grass)

[Source: California Exotic Pest Council]

FEDERALLY LISTED GRASSES [Listed as of 08 September 2000]

Avena sterilis (animated oat)
Chrysopogon aciculatus (pilipiliula)
Digitaria abyssinica (African couch grass)
Digitaria velutina (velvet finger grass)
Imperata brasiliensis (Brazilian satintail)

Imperata cylindrica (cogon grass)

Ischaemum rugosum Nassella trichotoma Leptochloa chinensis Oryza longistaminata

Oryza punctata Oryza rufipogon Paspalum scrobiculatum Pennisetum clandestinum Pennisetum macrourum

Pennisetum pedicellatum Pennisetum polystachion Rottboellia cochinchinensis Saccharum spontaneum Setaria pallide-fusca Urochloa panicoides (murain grass) (serrated tussock) (Asian sprangletop) (red rice)

(red rice) (red rice) (Kodo millet) (kikuyu grass) (African feather grass)

> (kyasuma grass) (mission grass) (itch grass) (wild sugarcane) (cattail grass)

A CHECKLIST OF WEEDY GRASSES OF THE UNITED STATES

Aegilops cylindrica Aegilops geniculata Aegilops triuncialis Agropyron cristatus Agrostis canina Agrostis gigantea Agrostis stolonifera Agrostis tenuis Alopecurus aequalis Alopecurus geniculatus Alopecurus myosuroides Alopecurus rendlei Andropogon gerardii Andropogon glomeratus Andropogon ternarius Andropogon virginicus Anthoxanthum aristatum Anthoxanthum odoratum Apera spica-venti Aristida adscensionis Aristida longiseta Aristida oligantha Arrhenatherum elatius Arundo donax Avena barbata Avena fatua Avena sterilis Axonopus affinis Axonopus compressus

Bothriochloa saccharoides Bouteloua aristidoides Bouteloua gracilis Brachiaria fasciculata Brachiaria mutica Brachiaria plantaginea Brachiaria platyphylla Brachiaria reptans Brachiaria texana Brachypodium distachyon Briza maxima Briza media Briza minor Bromus arvensis Bromus catharticus Bromus commutatus Bromus diandrus Bromus erectus Bromus hordeaceus Bromus inermis Bromus japonicus Bromus lanceolatus Bromus rigidus Bromus rubens Bromus secalinus Bromus sterilis Bromus tectorum

Catapodium rigidum
Cenchrus biflorus
Cenchrus brownii
Cenchrus ciliaris
Cenchrus echinatus
Cenchrus incertus
Cenchrus longispinus
Cenchrus myosuroides
Cenchrus pauciflorus
Cenchrus tribuloides
Chloris gayana
Chloris polydactylon

Chloris virgata Coix lacryma-jobi Cynodon dactylon Cynosurus cristatus Cynosurus echinatus

Dactyloctenium aegyptium Deschampsia cespitosa Deschampsia flexuosa Dichanthium annulatum Dichanthium aristatum Digitaria ciliaris Digitaria filiformis Digitaria horizontalis Digitaria ischacne Digitaria longiflora Digitaria sanguinalis Digitaria violaceus

Echinochloa colona Echinochloa crus-galli Echinochloa crus-pavonis Eleusine indica Elymus caninus Elymus caput-medusae Elymus repens Eragrostis barrelieri Eragrostis cilianensis Eragrostis curvula Eragrostis mexicana Eragrostis minor Eragrostis pectinacea Eragrostis pilosa Eragrostis tenella Eragrostis unioloides Eragrostis virescens Eriochloa gracilis Eriochloa punctata

Festuca arundinacea Festuca ovina Festuca rubra

Heteropogon contortus Holcus lanatus Holcus mollis Hordeum jubatum Hordeum leporinum Hordeum murinum Hordeum pusillum

Imperata brasiliensis Imperata cylindrica

Koeleria phleoides Koeleria pyramidata

Lagurus ovatus
Lamarckia aurea
Leersia hexandra
Leersia oryzoides
Leptochloa fascicularis
Leptochloa filiformis
Leptochloa scabra
Leptochloa unnervia
Leptochloa virgata
Lolium multiflorum
Lolium perenne
Lolium persicum
Lolium remotum

Lolium temulentum

Melinis repens Microstegium vimineum Miscanthus floridus Muhlenbergia schreberi

Oplismenus hirtellus Oryza rufipogon Oryzopsis miliacea

Panicum antidotale Panicum capillare Panicum clandestinum Panicum dichotomiflorum Panicum dichotomiflorum Panicum gattingeri Panicum maximum Panicum miliaceum Panicum obtusum Panicum repens Panicum trichoides Panicum virgatum Paspalidium geminatum Paspalum ciliatifolium Paspalum conjugatum Paspalum dilatatum Paspalum distichum Paspalum fluitans Paspalum laeve Paspalum lividum Paspalum notatum Paspalum paspaloides Paspalum plicatulum Paspalum scrobiculatum Paspalum urvillei Paspalum vaginatum Paspalum virgatum Pennisetum americanum Pennisetum clandestinum Pennisetum polystachyon Pennisetum purpureum Pennisetum villosum Phalaris aquatica

Phalaris arundinacea Phalaris brachystachya Phalaris canariensis Phalaris minor Phalaris phalaroides Phragmites australis Poa annua Poa bulbosa Poa compressa Poa pratensis Poa trivialis Polypogon monspeliensis Polypogon viridis

Rottboellia cochinchinensis

Schizachyrium scoparium Setaria barbata Setaria faberi Setaria glauca Setaria gracilis Setaria italica Setaria lutescens Setaria palmifolia Setaria sphacelata Setaria verticillata Setaria viridis Sorghum bicolor Sorghum halepense Sporobolus airoides Sporobolus juncea Sporobolus neglectus Sporobolus pyramidalis Sporobolus vaginiflorus Sporobolus virginicus Stenotaphrum secundatum

Tragus berteronianus Tragus racemosus

Vulpia bromoides Vulpia myuros

Zoysia matrella

MAJOR WEEDY GRASSES OF THE WORLD

Arundinoideae

Arundineae

Phragmites australis

Phragmites karka

Chloridoideae

Cynodon dactylon

Eragrostideae
Dactyloctenium aegyptium
Eleusine indica
Leptochloa chinensis
Leptochloa panicea

Ehrhartoideae

Oryzeae Leersia hexandra

Panicoideae

Paniceae

Axonopus compressus Brachiaria mutica Cenchrus echinatus Digitaria abysinnica Digitaria ciliaris Digitaria sanguinalis Echinochloa colona Echinochloa crus-galli Imperata cylindrica Ischaemum rugosum Panicum maximum Panicum repens Paspalum conjugatum Paspalum dilatatum Pennisetum clandestinum Pennisetum pedicellatum Pennisetum polystachion Pennisetum purpureum Rottboellia cochinchinensis Setaria verticillata Setaria viridis Sorghum halepense

Pooideae

Aveneae
Avena fatua

Poeae

Lolium temulentum

[Source: Chapman & Peat, 1992: 86]

SELECTED REFERENCES

GENERAL REFERENCES

Baker, H. G. 1962. Weeds -- native and introduced. J. California Hort. Soc. 23: 97-104.

Baker, H. G. 1965. Characteristics and modes of origin of weeds. <u>In</u>, Baker, H. G. & G. L. Stebbins (editors). Genetics of colonizing species. Pp. 147-168.

Baker, H. G. 1974. The evolution of weeds. Ann. Rev. Ecol. Syst. 5: 1-24.

Behrendt, S. & M. Hanf. 1979. Grass weeds in world agriculture: identification in the flowerless state. Ludwigshafen, Basf Aktiengesellschaft. 159 pp.

Crockett, L. J. 1977. Wildly successful plants: a handbook of North American weeds. Collier Books. New York, NY. 268 pp.

deWet, J. M. J. & J. R. Harlan. 1975. Weeds and domesticates: evolution in the man-made habitat. Econ. Bot. 29: 99-107.

Frenkel, R. E. 1970. Ruderal vegetation along certain California roadsides. Univ. California Publ. Geogr. 20: 1-163.

Häfliger, E. & H. Scholz. 1980. Grass weeds 1: weeds of the sub-family Panicoideae. Ciba-Geigy. Basle, Switzerland. 142 pp. + plates.

Häfliger, E. & H. Scholz. 1981. Grass weeds 2: weeds of the sub-families Chloridoideae, Pooideae,

- Oryzoideae. Ciba-Geigy. Basle, Switzerland. 137 pp. + plates.
- Harlan, J. R. 1965. The possible role of weed races in the evolution of cultivated plants. Euphytica 14: 173-176.
- Harlan, J. R. & J. M. J. deWet. 1965. Some thoughts about weeds. Econ. Bot. 19: 16-24.
- Holm, L. G. 1971. The role of weeds in human affairs. Weed Sci. 19: 485-490.
- Holm, L. G. et al. 1977. The world's worst weeds, distribution and biology. Univ. Press Hawaii. Honolulu, HI. 609 pp.
- Mack, R. N. 1990. Catalog of woes. Natural History 3/90: 44-53.
- Muenscher, W. C. 1960. Weeds. Second edition. Macmillan. New York, NY. 560 pp.
- Reed, C. F. 1970. Selected weeds of the United States. Agric. Handbook No. 366. U. S. Dept. Agric. Washington, D. C. Pp. 32-90.
- Reed, C. F. 1977. Economically important weeds. Agriculture Handbook No. 498. U. S. Dept. Agric. Washington, D. C. 746 pp.
- Robbins, W. W., M. K. Bellue, & W. S. Ball. 1951. Weeds of California. State of California. Sacramento. 547 pp.
- Whitson, T. D. 1996. Weeds of the West. Fifth edition. Western Soc. Weed Sci. Newark, CA. 630 pp.
- Wilkinson, R. E. & H. E. Jacques. 1979. How to know the weeds. Third edition. W. C. Brown. Dubuque, IA. 235 pp.

PARTICULAR GRASSES

- Aptekar, R. 1999. The ecology and control of European beach-grass (*Ammophila arenaria*). Ph. D. dissertation. Univ. California, Davis.
- Barkworth, M. R. et al. 1989. *Stipa clandestina*: new weed threat on southwestern rangelands. Weed Technol. 3: 699-702.
- Billings, W. D. 1990. *Bromus tectorum*, a biotic cause of eco-system impoverishment in the Great Basin. <u>In</u>, Woodwell, G. M. (editor). The earth in transition. Cambridge Univ. Press. Cambridge, England. Pp. 301-322.
- Buell, A. C., A. Pickart, & J. D. Stuart. 1995. Introduction, history and invasion patterns of *Ammophila arenaria* on the north coast of California. Cons. Biol. 9: 1587-1593.
- Callaway, J. C. & M. N. Josselyn. 1992. The introduction and spread of smooth cordgrass (*Spartina alterniflora*) in south San Francisco Bay. Estuaries 15: 218-226.
- Daehler, C. C. & D. R. Strong. 1996. Status, prediction, and pre-vention of introduced cordgrass (*Spartina* spp.) invasions in Pacific estuaries, U. S. A. Biol. Conserv. 78: 51-58.
- Fuller, T. C. 1976. Pampas grass: its history as a

- weed. Fremontia 4: 16.
- Mack, R. N. 1981. Invasion of *Bromus tectorum* into western North America: an ecological chronicle. Agroecosystems 7: 145-165.
- Marks, M., B. Lapin, & J. Randall. 1994. *Phragmites australis (P. communis)*: threats, management, and monitoring. Nat. Areas J. 14(4): 285-292.
- Morrow, L. A. & P. W. Stahlman. 1984. The history and distribu-tion of downy brome (*Bromus tectorum*) in North America. Weed Sci. 32(Supplement) 1: 2-6.
- Schery, R. W. 1965. The migration of a plant: Kentucky bluegrass followed settlers to the New World. Nat. Hist. 74(Dec.): 43, 44.
- Sigg, J. 1996. *Ehrharta erecta*: sneak attack in the making? CalEPPC Newsletter 4(3): 8, 9.
- Young, J. A. 1992. Ecology and management of medusa head (*Taeniatherum caput-medusae* ssp. *asperum* [Simk.] Melderis). Great Basin Nat. 52(3): 245-252.
- Wiedemann, A. M. & A. Pickart5. 1996. The *Ammophila* problem on the northwest coast of North America. Landscape Urban Planning 34: 287-299.
- Wilvert, C. 1980. Kikiyu grass, an African invader. Pacific Hort. 41(3): 45-47.

4.10 - ORNAMENTAL & LAWN GRASSES

Grasses have, of course, long been used for lawns. Less appreciated has been their use as more showy ornamentals. You may have noticed how many more grasses are now being offered for landscaping purposes and that there are now a number of books on ornamental grasses available.

HERBACEOUS ORNAMENTALS

Agrostis nebulosa (cloud grass)
Alopecurus lanatus (woolly foxtail grass)
Alopecurus pratensis (foxtail grass)
Ampelodesmos mauritanicus
Andropogon gerardii (big bluestem)

Andropogon glomeratus
Andropogon ternarius
Anthoxanthum odoratum
Apera spica-venti
Aristida purpurea

(bushy bluestem)
(split beard bluestem)
(velvet grass)
(silky bent)
(purple three-awn grass)

Arrhenatherum elatius
Arundo donax
Arundo pliniana
Avena sterilis
Beckmannia syzigachne

(tall oat grass)
(giant reed, carrizo)
(arrow-reed)
(animated oat)
(slough grass)

Bothriochloa barbinodis
Bouteloua curtipendula
Bouteloua gracilis
Briza maxima
Briza media

(cane bluestem)
(side-oats grama)
(blue grama)
(giant quaking grass)
(common quaking grass)

Bromus brizaeformis (rattlesnake chess)
Bromus ramosus (wood brome)
Calamagrostis acutiflora (feather reed grass)
Calamagrostis arundinacea(fall-blooming reed grass)
Calamagrostis canescens (purple small reed)

Calamagrostis epigejos (bush grass)
Chasmanthium latifolium (wild-oats)
Chloris virgata (finger grass)
Coix lacryma-jobi (Job's tears)
Cortaderia fulvida (erect-plumed tussock grass)

Cortaderia jubata (purple pampas grass)
Cortaderia selloana (pampas grass)
Cymbopogon citratus (lemon grass)
Cynodon dactylon (Bermuda grass)
Dactylis glomerata (orchard grass)

Deschampsia caespitosa
Deschampsia flexuosa
Elymus arenarius
Elymus hystrix
Elymus magellanicus
Eragrostis curvula

(tufted hair grass)
(crinkled hair grass)
(blue lyme grass)
(bottlebrush grass)
(Magellan blue grass)
(weeping love grass)

Eragrostis spectabilis
Eragrostis suaveolens
Eragrostis tef
Eragrostis trichodes
Erianthus contortus
Erianthus giganteus
Erianthus ravennae

(purple love grass)
(teff)
(sand love grass)
(bent awn plume grass)
(sugar cane plume grass)
(ravenna grass)

Festuca geniculata

Festuca gigantea (giant fescue)
Festuca ovina (sheep fescue)
Glyceria maxima (reed manna grass)
Gynerium sagittatum (uva grass)
Helictotrichon sempervirens (blue oat grass)

Holcus mollis
Hordeum jubatum
Koeleria brevis
Koeleria glauca
Lagurus ovatus

(velvet grass)
(foxtail barley, squirreltail)
(blue hair grass)
(large blue hair grass)
(hare's tail, rabbit-tail grass)

Lamarckia aurea(golden top)Melica altissima(Siberian melic)Melica ciliata(hairy melic)Melica uniflora(wood melic)Melinis repens(ruby grass, Natal grass)

Mibora minima (sand bent)
Milium effusum (wood millet)
Miscanthus giganteus (Chinese silver grass)
Miscanthus nepalensis (Himalaya fairy grass)
Miscanthus sacchariflorus (eulalia)

Miscanthus sinensis (eulalia)
Miscanthus transmorrisonens(evergreen miscanthus)
Molinia caerulea (purple moor grass)
Muhlenbergia dumosa (bamboo muhly)
Muhlenbergia emersleyi (bull grass)

Muhlenbergia filipes(purple muhly)Muhlenbergia rigens(deer grass)Oplismenus hirtellus(basket grass)Oplismenus imbecillicus(basket grass)Oryza sativa(rice)

Oryzopsis miliacea (smilo grass, rice grass)
Panicum miliaceum (common millet, proso m.)
Panicum virgatum (switch grass)
Pennisetum alopecuroides (Chinese pennisetum)
Pennisetum caudatum(white-flowering feather grass)

Pennisetum latifolium
Pennisetum macrostachyum
Pennisetum orientale
Pennisetum setaceum
Pennisetum villosum

(Uruguay pennisetum)
(Oriental feather grass)
(fountain grass)
(feathertop)

Phalaris arundinacea(reed canary grass)Phalaris canariensis(canary grass)Phragmites australis(common reed)Phragmites macra(hakone grass)Polypogon monspeliensis(annual beard grass)

Saccharum officinarum
Schizachyrium scoparium
Sesleria caerulea
Setaria italica
Setaria lutescens
Setaria palmifolia
Setaria poiretiana
(sugar cane)
(blue moor grass)
(foxtail millet)
(foxtail millet)
(yellow foxtail)
(palm grass)

Sorghum bicolor (sorghum, African millet)
Sorghastrum nutans (Indian grass)
Spartina pectinata (prairie cord grass)
Spodiopogon sibericus (frost grass)
Stenotaphrum secundatum (St. Augustine grass)

Stipa arundinacea (pheasant's tail grass)

Stipa calamagrostis Stipa capillata Stipa gigantea Stipa hymenoides

(feather grass) (golden oats) (Indian rice grass)

Stipa ichu Stipa pennata Stipa ramosissima Stipa splendens Stipa tenacissima (Peruvian feather grass) (European feather grass) (pillar of smoke) (chee grass) (esparto)

Themeda triandra Thysanolaena maxima Tripsacum dactyloides Uniola latifolia Zea mays Zizania aquatica (Japanese themeda) (tiger grass) (eastern gama grass) (sea-oats) (maize, corn) (wild-rice)

ORNAMENTAL BAMBOOS

Arundinaria amabilis Arundinaria disticha Arundinaria humilis (Tonkin bamboo) (dwarf fernleaf bamboo)

Arundinaria simonii (Simon bamboo, medake) Arundinaria variegata (dwarf whitestripe bamboo)

Bambusa arundinacea (giant thorny bamboo)
Bambusa beecheyana (Beechey bamboo)
Bambusa glaucescens (hedge bamboo)
Bambusa oldhan(timber bamboo, Oldham's bamboo)
Bambusa pervariabilis

Bambusa polymorpha Bambusa textilis Bambusa tulda Bambusa tuldoides

Bambusa tuldoides (punting pole bamboo) Bambusa ventricosa (Buddha's belly bamboo)

Bambusa vulgaris Chimonobambusa falcata Chimonobambusa marmorea (marbled bamboo) Chimonobambusa quadrangularis (square-stem b.) Dendrocalamus asper

Dendrocalamus latiflorus (sweet bamboo)
Dendrocalamus strictus (Calcutta bamboo, male b.)
Phyllostachys aurea (fishpole bamboo)
Phyllostachys aureosulcata (yellowgroove bamboo)
Phyllostachys bambusoides (madake, timber bamboo)
Phyllostachys dulcis (sweetshoot bamboo)
Phyllostachys flexuosa
Phyllostachys eyeri (Meyer's bamboo)

Phyllostachys nigra(black bamboo, Henon's bamboo)
Phyllostachys nuda

Phyllostachys pubescens Phyllostachys viridi-glaucescens

Phyllostachys viridis

cens (moso bamboo) glaucescens

Phyllostachys vivax Pseudosasa japonica

Pseudosasa japonica (metake, arrow bamboo)
Sasa disticha (dwarf fernleaf bamboo)
Sasa palmata (palmate bamboo)
Sasa tessellata

Sasa veitchii Semiarundinaria fastuosa Semiarundinaria murieiae Semiarundinaria nitida Shibataea kumasaca Yoshania aztecorum (kumazasa) (narihira bamboo)

LAWN GRASSES

Agrostis palustris Agrostis tenuis Cynodon dactylon Eremochloa ophuroides Festuca arundinacea (creeping bent grass) (colonial gent grass) (Bermuda grass) (centipede grass) (tall fescue)

Festuca longifolia Festuca rubra Lolium multiflorum Lolium perenne Paspalum notatum (hard fescue) (red fescue) (annual rye grass) (perennial rye grass) (Bahia grass)

Paspalum vaginatum Poa pratensis Poa trivialis Stenotaphrum secundatum Zoysia matrella

(seashore paspalum) (Kentucky blue grass) (rough-stalk blue grass) (St. Augustine grass) (zoysia)

SELECTED REFERENCES

Bell, M. 2000. The gardener's guide to growing temperate bamboos. David & Charles. Devon, England. 159 pp.

Chatto, B. 1976. Grasses and grass-like plants. The Garden 101: 448-453.

Cusack, V. 1999. Bamboo world: the growing and use of clumping bamboos. Kangaroo Press. East Roseville, Australia. 224 pp.

Darke, R. 1994. For your garden: ornamental grasses. Little, Brown. Boston, MA. 72 pp.

Darke, R. 1994. Manual of grasses. The new Royal Horticultural Society dictionary. Timber Press. Portland, OR. 169 pp.

Darke, R. 1999. The color encyclopedia of ornamental grasses. Timber Press. Portland, OR. 325 pp.

Darke, R. 2004. Timber Press pocket guide to ornamental grasses. Timber Press. Portland, OR. 226 pp.

Dunmire, J. R. 1995. Bamboo in the garden. Pacific Hort. 56(1): 34-44.

Feldman, F. 1989. Sunset lawns & ground covers. Lane Publ. Co. Menlo Park, CA. Pp. 19-26.

Feesey, M. T. 1983. Ornamental grasses and bamboos. Royal Hort. Soc. London, England.

Greenlee, J. & D. Fell. 1992. The encyclopedia of ornamental grasses. Rodale Press. Emmaus, PA. 186 pp.

Grounds, R. 1979. Ornamental grasses. Van Nostrand Reinhold. New York, NY. 216 pp.

Grounds, R. 1998. The plantfinder's guide to ornamental grasses. Timber Press. Portland, OR. 166 pp.

Grounds, R. 2002. Grasses and bamboos. DK Publ. New York, NY. 80 pp.

Haubrich, R. 1981. Handbook of bamboos cultivated in the United States. J. American Bamboo Soc.

Haubrich, R. 1982. The sasas. J. American Bamboo Soc. 2: 24-38.

Hodge, W. H. & D. A. Bisset. 1957. Running bamboos for hedges. Nat. Hort. Mag. 36(4): 335-339.

Holmes, R. (editor). 1997. Taylor's guide to ornamental grasses. Houghton Mifflin. Boston, MA. 309 pp.

King, M. & P. Oudolf. 1998. Gardening with grasses. Timber Press. Portland, OR. 152 pp.

Kingsbury, N. 2000. Grasses and bamboos. Watson-Guptill. New York, NY. 111 pp.

Lawson, A. H. 1968. Bamboos. A gardener's guide to their cultivation in temperate climates. Taplinger. New York, NY. 192 pp.

Loewer, H. P. 1977. Growing and decorating with grasses. Walker Publ. New York, NY. 128 pp.

Loewer, H. P. 1995. Ornamental grasses. Better Homes and Gardens Books. Des Moines, IA. 132 pp.

Meredith, T. J. 2001. Bamboos for gardens. Timber Press. Portland, OR. 408 pp.

Oakes, A. J. 1993. Ornamental grasses and grass-like plants. Second edition. Krieger Publ. Co. Malabar, FL. 614 pp.

Ottesen, C. 1989. Ornamental grasses: the amber wave. McGraw-Hill. New York, NY. 230 pp.

Pesch, B. B. (editor). 1988. Ornamental grasses. Brooklyn Bot. Gard. Record 44, No. 3.

Reinhardt, T. A., M. Reinhardt, & M. Moskowitz. 1995. Ornamental grasses. Friedman/Fairfax. New York, NY. 127 pp.

Shor, G. 1999. Bamboo species source list no. 19. American Bamboo Soc. Albany, NY. 27 pp.

Taylor, N. J. 1992. Ornamental grasses, bamboos, rushes & sedges. Ward Lock. London, England. 96 pp.

Thomas, G. S. 1957. Bamboos. Royal Hort. Soc. 82(6): 247-255.

Young, R. A. 1945-1946. Bamboos for American horticulture. Nat. Hort. Mag. 24: 171-196, 274-291; 25: 40-64, 352-365.

Young, R. A. & J. R. Haun. 1961. Bamboo in the United States: description, culture, and utilization. Handbook No. 193. U. S. Dept. Agric. Washington, D. C. 74 pp.

4.11 - ENDEMIC GRASSES

Agrostis ampla (western bent grass) Agrostis aristiglumis (awned bent grass) Agrostis blasdalei var. blasdalei (Blasdale's bent grass) Agrostis blasdalei var. marinensis (Marin bent grass) Agrostis clivicola var. clivicola (coastal bluff bent grass)	OR, CA Marin Co., CA CA CA CA
Agrostis clivicola var. punta-reyesensis (Pt. Reyes bent grass)	CA
Agrostis densiflora (California bent grass)	OR, CA
Agrostis exarata var. minor (spike bent grass)	NM
Agrostis hallii (Hall's bent grass)	OR, CA
Agrostis hendersonii (Henderson's bent grass)	OR, CA
Agrostis hooveri (Hoover's bent grass)	CA
Agrostis howellii (Howell's bent grass)	OR
Agrostis lepida (Sequoia bent grass)	CA
Agrostis longiligula	???
Agrostis microphylla var. major	???
Alopecurus aequalis var. sonomensis (Sonoma foxtail)	CA
Alopecurus howellii (Howell's meadow foxtail)	WA, OR
Amphicarpum amphicarpon (annual peanut grass)	eastern U. S.
Amphicarpum muhlenbergianum (blue maiden cane)	southeastern U. S.
Andropogon arctatus (pinewoods bluestem)	AL, FL
Andropogon floridanus (Florida bluestem)	GA, AL, FL
Andropogon glomeratus var. glaucopsis (purple bluestem)	southeastern U. S.
Andropogon gyrans var. stenophyllus (Elliott's bluestem)	TX to NC
Andropogon liebmannii (Liebmann's bluestem)	southeastern U. S.
Andropogon ternarius var. cabanisii	FL
Andropogon tracyi (Tracy's bluestem) Andropogon virginicus var. decipiens (broom-sedge bluestem) Andropogon virginicus var. glaucus (chalky bluestem) Anthaenantia rufa (purple silky scale)	southeastern U. S. southeastern U. S. NJ to TX southeastern U. S.

Anthaenantia villosa (green silky scale)	southeastern U. S.
Aristida condensata (big three-awn) Aristida desmantha (sand three-awn) Aristida dichotoma var. curtissii (Curtiss's three-awn) Aristida dichotoma var. dichotoma (churchmouse three-awn) Aristida gyrans (corkscrew three-awn)	southeastern U. S. central U. S. central & southeastern U. S. central & eastern U. S. GA, FL
Aristida lanosa var. lanosa (woolly sheathed three-awn) Aristida mohrii (Mohr's three-awn) Aristida palustris (long leaf three-awn) Aristida patula (tall three-awn) Aristida purpurascens var. tenuispica (Hillsboro three-awn)	eastern U. S. southeastern U. S. southeastern U. S. FL NC to MS
Aristida purpurascens var. virgata (pinewoods three-awn) Aristida ramosissima (s-curve three-awn) Aristida rhizomorpha (Florida three-awn) Aristida schiediana var. orcuttiana (single three-awn) Aristida simpliciflora (Chapman's three-awn)	southeastern U. S. central & eastern U. S. FL southeastern U. S. southeastern U. S.
Aristida tuberculosa (seaside three-awn) Arundinaria gigantea ssp. gigantea (switch cane) Arundinaria gigantea ssp. x macrosperma (switch cane) Arundinaria gigantea ssp. tecta (switch cane) Blepharidachne kingii (King's eyelash grass)	eastern U. S. DE to TX, FL VA to AL southeastern U. S. CA, ID, UT, NV
Bothriochloa exaristata (awnless bluestem) Bothriochloa hybrida (hybrid bluestem) Bouteloua kayi (Kay's grass) Brachyelytrum erectum (bearded shorthusk) Bromus berterianus var. excelsus (Chilean chess) Bromus frondosus (weeping brome) Bromus grandis (tall brome) Bromus kalmii (Kalm's brome)	CA, TX, LA TX, LA TX, LA Brewster Co., TX eastern & central U. S. CA, AZ ??? southwestern U. S. CA +++ ??? a-central & northeastern U. S.
Bromus laevipes (woodland brome) Bromus luzonensis (hoary brome) Bromus marginatus (mountain brome) Bromus maritimus (maritime brome) Bromus orcuttianus var. hallii (Orcutt's brome)	WA, OR, NV, CA WA, OR, CA, ID, NV western & central U. S. OR, CA CA
Bromus polyanthus var. paniculatus (Great Basin brome) Bromus polyanthus var. polyanthus (Colorado brome) Bromus pseudolaevipes (woodland brome) Bromus suksdorfii (Suksdorf's brome) Calamagrostis bolanderi (Bolander's reed grass)	southwestern U. S. western U. S. CA WA, OR, ID, CA, NV CA
Calamagrostis breweri (Brewer's reed grass) Calamagrostis cainii (Cain's reed grass) Calamagrostis californica Calamagrostis coarctata (Nuttall's reed grass) Calamagrostis fernaldii (Fernald's reed grass)	OR, CA TN, NC ??? eastern & southeastern U. S. Piscataquis Co., ME
Calamagrostis foliosa (leafy reed grass) Calamagrostis howellii (Howell's reed grass) Calamagrostis koelerioides (tufted pine grass) Calamagrostis x lactea (bluejoint) Calamagrostis muriana (Muir's reed grass)	CA WA, OR western U. S. WA, OR, CA CA
Calamagrostis ophiditis (serpentine reed grass) Calamagrostis perplexa (wood reed grass) Calamagrostis porteri ssp. inseparata (Porter's reed grass) Calamagrostis porteri ssp. porteri (Porter's reed grass) Calamagrostis scopulorum (Jones' reed grass)	CA Tompkins Co., NY eastern U. S. eastern U. S. western U. S.
Calamagrostis tweedyi (Tweedy's reed grass) Calammophila don-hensonii (Henson's reed grass) Calamovilfa arcuata (Cumberland sand reed) Calamovilfa brevipilis var. brevipilis (pine barren sand reed) Calamovilfa brevipilis var. calvipes	WA, MT, ID MI OK, AR, AL, TN NJ to SC VA
Calamovilfa curtisii (Florida sand reed) Calamovilfa gigantea (big sand reed) south- Chasmanthium laxum var. laxum (spike grass)	FL -central & southwestern U. S. southeastern U. S.

Chasmanthium laxum var. sessiliflorum (long leaf uniola)	southeastern U. S.
Chasmanthium nitidum (shiny uniola)	southeastern U. S.
Chasmanthium ornithorhynchum (bird bill uniola) Chloris x subdolichostachya (short-spike windmill grass) Chloris texensis (Texas windmill grass) Cinna bolanderi (Bolander's reed grass)	southeastern U. S. AZ to KS, LA TX CA
Coelorachis cylindrica (Carolina joint-tail)	southeastern U. S.
Coelorachis rugosa (wrinkled joint-tail) Coelorachis tessellata (lattice joint-tail) Coelorachis tuberculosa (Florida joint-tail) Ctenium aromaticum (toothache grass) Ctenium floridanum (Florida toothache grass)	southeastern U. S. LA to FL southeastern U. S. VA to FL, LA GA, FL
Danthonia epilis (Carolina oat grass) Danthonia sericea (downy oat grass) Digitaria arenicola (sand witch grass) Digitaria cognata (fall witch grass)	southeastern U. S. eastern U. S. TX eastern & central U. S.
Digitaria filiformis var. filiformis (Caribbean crab grass)	southeastern U. S. ???
Digitaria floridana (Florida crab grass) Digitaria horizontalis (Jamaica crab grass) Digitaria leucocoma Digitaria pauciflora (two-spike finger grass) Digitaria pubiflora (western witch grass)	Hernando Co., FL southeastern U. S. ??? Lake Co., FL Dade Co., FL southwestern U. S.
Digitaria simpsonii (Simpson's	Southwestern of Si
Dissanthelium californicum (Catalina grass) Elyhordeum californicus	CA (CFP)
Elyhordeum iowense (Iowa barley) Elyhordeum piperi (Piper's barley)	MT, ND, NB, ÏA WA
Elyhordeum stebbinsianum (Stebbins' barley) Elymus ambiguus var. ambiguus (Rocky Mountain lyme grass) Elymus arenicola (sand lyme grass) Elymus xaristatus (purple wheat grass) Elymus arizonicus (Arizona wild rye)	WA, OR, CA UT, MT to NM WA, OR, MT, ID western U. S. CA, AZ, NM, TX
Elymus californicus (California bottlebrush) Elymus elymoides ssp. hordeoides (western squirreltail) Elymus flavescens (sand lyme grass) Elymus laevis (California wild rye) Elymus x multiflorus	CA WA, OR, ID, CA, NV WA, OR, ID, SD CA CA, CFP ?
Elymus multisetus (big wild rye)	western U. S.
Elymus pacificus (Pacific wild rye) Elymus salinus ssp. mojavensis (Salina wild rye) Elymus salinus ssp. salinus (Salina wild rye) Elymus sierrae (Sierra wild rye)	CA CA, AZ western U. S. CA, NV
Elymus simplex (smooth wild rye) Elymus stebbinsii ssp. septentrionalis (Stebbins' wild rye)	western U. S. CA
Elymus stebbinsii ssp. stebbinsii (Stebbins' wild rye) Elymus trachycaulus ssp. sierrus Elymus villosus var. arkansanus (hairy wild rye)	CA CA Source ??? central & eastern U. S.
Eragrostis bahiensis (bahia love grass) Eragrostis lutescens (six week's love grass)	LA, AL, FL WA, CA, NV, CO, AZ, NM
Eragrostis pectinacea var. miserrima (desert love grass) Eragrostis pectinacea var. tracyi (Tracy's love grass) Eragrostis refracta (coastal love grass)	TX to FL FL southeastern U. S.
Eragrostis swallenii (Swallen's love grass) Eragrostis trichodes (sand love grass) Eriochloa michauxii var. michauxii (long leaf cup grass) Eragrostis michauxii var. simpsonii (Simpson's cup grass) Eustachys floridana (Florida finger grass)	TX central & eastern U. S. southeastern U. S. FL AL, GA, FL
Eustachys glauca (salt marsh windmill grass)	southeastern U. S.
Eustachys neglecta (four-spike windmill grass) Festuca arizonica (Arizona fescue) Festuca brachyphylla ssp. coloradensis (alpine fescue)	AZ, CO, NM, TX western U. S.

Festuca californica var. californica (California fescue)	OR, CA
Festuca calligera (southwestern fescue) Festuca dasyclada (Utah fescue) Festuca elliotea (squirrel-tail six-weeks grass) Festuca elmeri (Elmer's fescue) Festuca howellii (Howell's fescue)	southwestern U. S. UT, CO central & southeastern U. S. WA, OR, CA WA, OR, CA
Festuca kingii (spiked fescue) Festuca ligulata (Guadalupe fescue) Festuca minutiflora (small-flowered fescue) Festuca occidentalis (western fescue) Festuca paradoxa (clustered fescue)	western U. S. TX western U. S. western U. S. central & eastern U. S.
Festuca parishii (California fescue) Festuca prolifera (proliferous fescue) Festuca rubra ssp. arctica (red fescue) Festuca rubra ssp. deniuscula (red fescue) Festuca sororia (ravine fescue)	CA ME, NM (Canada only ???) NH (+ ?) OR, CA AZ, CO, UT, NM
Festuca thurberi (Thurber's fescue) Festuca versuta (Texas fescue) Festuca viridula (green leaf fescue) Festuca washingtoniana (Washington fescue) Glyceria acutiflora (creeping manna grass)	AZ, NM, CO, UT, WY OK, TX western U. S. WA eastern U. S.
Glyceria nubigena (Great Smoky Mountain manna grass) Glyceria septentrionalis var. arkansana (Arkansas manna grass Gymnopogon brevifolius (slim skeleton grass) south- Gymnopogon chapmanianus (Chapman's skeleton grass) Helictotrichon mortonianum (Morton's alpine-oat)	NC, TN s)OK, TX, southeastern U. S. central & southeastern U. S. GA, FL UT, CO, NM
Hilaria jamesii (galleta) Hordeum arizonicum (Arizona barley) Hordeum brachyantherum ssp. californicum (California barley) Leersia hexandra (southern cut grass) Leersia lenticularis (catchfly grass)	western U. S. CA, AZ, NM OR, CA, NV southeastern U. S. eastern U. S.
Melica aristata (awned melic) Melica bulbosa var. inflata Melica californica var. californica (California melic) Melica californica var. nevadensis (California melic) Melica fugax (little onion grass)	WA, OR, CA, NV, KT WA, CA OR, CA CA WA, OR, CA, ID, NV
Melica geyeri var. aristulata (Geyer's onion grass) Melica geyeri var. geyeri (Geyer's onion grass) Melica imperfecta (coast range melic) Melica mutica (two-flowered melic) Melica porteri var. laxa (Porter's melic)	CA OR, CA, NV CA (CFP) eastern U. S. AZ, NM, TX
Melica porteri var. porteri (Porter's melic) Melica stricta var. albicaulis (nodding onion grass) Melica stricta var. stricta (nodding onion grass) Melica subulata var. pammelii (Pammel's onion grass) Melica torreyana (Torrey's melic)	UT, CO, AZ, NM, TX CA OR, CA, NV, UT ID, MT, WY CA
Muhlenbergia bushii (nodding muhly) Muhlenbergia californica (California muhly) Muhlenbergia expansa (cutover muhly) Muhlenbergia filiculmis (slim stem muhly) Muhlenbergia jonesii (Modoc muhly)	central & eastern U. S. CA [extinct] southeastern U. S. southwestern U. S. CA
Muhlenbergia pungens (sandhill muhly) Muhlenbergia reverchonii (seep muhly) Muhlenbergia sericea (dune hair grass) Muhlenbergia thurberi (Thurber's muhly) Muhlenbergia torreyana (New Jersey muhly)	western & central U. S. OK, TX southeastern U. S. NV, UT, CO, AZ, NM northeastern U. S.
Muhlenbergia villiflora var. villosa (hairy muhly) Muhlenbergia virescens (screw-leaf muhly) Neostapfia colusana (Colusa grass) Orcuttia californica (California orcutt grass) Orcuttia inaequalis (San Joaquin Valley orcutt grass)	NM, TX AZ, NM CA CA (CFP) CA

Orcuttia pilosa (hairy Orcutt grass) Orcuttia tenuis (slender Orcutt grass) Orcuttia viscida (Sacramento Orcutt grass) Oryzopsis contracta (contracted rice grass) Oryzopsis hendersonii (Henderson's rice grass)	CA CA Sacramento Co., CA MT, WY, CO WA, OR, ID
Oryzopsis kingii (King's mountain-rice) Oryzopsis porteri (Porter's needle grass) Oryzopsis wallowaensis (Wallowa rice grass) Oryzopsis webberi (Webber's rice grass) Panicum acuminatum var. columbianum	CA CO OR western U. S. northeastern U. S.
Panicum acuminatum var. consanguineum (blood panic grass) Panicum acuminatum var. sericeum Panicum anceps var. anceps (beaked panic grass) south-central Panicum anceps var. rhizomatum (beaked panic grass) Panicum boscii (Bosc's panic grass)	southeastern U. S. western U. S. I & southeastern U. S. southeastern U. S. eastern U. S.
Panicum brachyantherum (pimple panic grass) Panicum capillare var. hillmanii (Hillman's panic grass) CA, I Panicum chamaelonche var. breve Panicum consanguineum (Kunth's panic grass) Panicum dichotomiflorum var. puritanorum	OK, TX, AR, LA, MS A, south-central U. S. FL southeastern U. S. eastern U. S.
Panicum dichotomum var. glabrifolium (fall panic grass) Panicum dichotomum var. lucidum (fall panic grass) Panicum dichotomum var. mattamuskeetense Panicum dichotomum var. nitidum Panicum ensifolium var. curtifolium	FL & southeastern U. S. MA to FL southeastern U. S. southeastern U. S.
Panicum ensifolium var. breve Panicum gymnocarpon (savanna) Panicum malacophyllum (soft-leaved panic grass) Companicum mohavense (Mohave panic grass) Panicum philadelphicum ssp. lithophilum	FL southeastern U. S. entral & eastern U. S. AZ, NM GA, NC, SC
Panicum philadelphicum ssp. philadelphicum (Philadelphia panic gras Panicum ovale var. pseudopubescens (egg-leaf panic grass) c Panicum perlongum (long-sheath panic grass) Panicum ravenelii (Ravenell's panic grass) eastern Panicum rigidulum var. abscissum (cut throat panic grass)	
Panicum rigidulum var. combsii (Comb's panic grass) Panicum rigidulum var. elongatum Panicum rigidulum var. pubescens Panicum rigidulum var. rigidulum (redtop panic grass) Panicum scabriusculum (tall swamp panic grass)	southeastern U. S. eastern U. S. southeastern U. S. CA, OR, eastern U. S. southeastern U. S.
Paspalum bifidum (pitchfork paspalum) Paspalum distichum (knot grass) western, south-cei Paspalum floridanum var. floridanum (Florida paspalum) Paspalum laeve (field paspalum) Paspalum lividum (longtom)	southeastern U. S. ntral, eastern U. S. ?? eastern U. S. eastern U. S. southeasternU. S.
Paspalum separatum Paspalum setaceum var. longepedunculatum (bare stem paspalum) Paspalum setaceum var. muhlenbergii (hurrah grass) c Paspalum setaceum var. psammophila (sand paspalum) Paspalum setaceum var. supinum (supine paspalum)	TX ??? southeastern U. S. entral & eastern U. S. eastern U. S. southeastern U. S.
Phalaris californica (California canary grass) Phalaris lemmonii (Lemmon's canary grass) Piptochaetium avenacioides (Florida needle grass) Pleuropogon californicus var. californicus (California semaphore grass) Pleuropogon californicum var. davyi (Davy's semaphore grass)	OR, CA (CFP) CA FL CA CA CA
Pleuropogon hooverianus (Hoover's semaphore grass) Pleuropogon oreganus (Oregon semaphore grass) Poa arctica ssp. aperta (arctic blue grass) Poa arctica ssp. grayana (Gray's blue grass) Poa arnowiae (Arnow's blue grass)	CA OR CO, WY, NM MT to NM, UT ID, UT

Poa atropurpurea (San Bernardino blue grass)	CA
Poa autumnalis (autumn blue grass)	OK, TX
Poa bolanderi (Bolander's blue grass)	WA, OR, CA, ID, UT
Poa chambersii (Chambers' blue grass)	Lane Co., OR
Poa chapmaniana (Chapman's blue grass)	central & eastern U. S.
Poa curtifolia (Little Mountain blue grass) Poa cusickii ssp. cusickii (Cusick's blue grass) Poa cuspidata (early blue grass) Poa douglasii (sand dune blue grass) Poa x fibrata	WA, OR, CA, MT, ID, NV southeastern U. S. CA ???
Poa hartzii ssp. alaskana (Hartz's blue grass)	AK
Poa keckii (Keck's blue grass)	CA
Poa liebergii (Leiberg's blue grass)	WA, OR, ID
Poa macroclada	ID, MT, CO
Poa x multnomae (Multnomah Falls blue grass)	OR
Poa napensis (Napa blue grass)	Napa Co., CA
Poa occidentalis (New Mexico blue grass)	AZ, CO, NM, TX
Poa paludigena (bog blue grass)	northeastern U. S.
Poa piperi (Piper's blue grass)	OR, CA
Poa pringlei (Pringle's blue grass)	OR, CA
Poa reflexa (nodding blue grass)	western U. S.
Poa rhizomata (timber blue grass)	OR, CA
Poa sierrae (Sierra blue grass)	CA
Poa stebbinsii (Stebbins' blue grass)	CA
Poa tenerrima (delicate blue grass)	CA
Poa tracyi (Tracy's blue grass)	CO, NM
Poa unilateralis ssp. pachypholis	WA
Poa wolfii (Wolf's blue grass)	eastern U. S.
Puccinellia howellii (Howell's alkali grass)	Shasta Co., CA
Puccinellia parishii (Parish's alkali grass)	CA, AZ, NM
Puccinellia simplex (little alkali grass)	CA, UT
Puccinellia sublaevis (smooth alkali grass)	AK
Redfieldia flexuosa (blowout grass)	western & central U. S.
Saccharum alopecuroides (silver plume grass)	southeastern U. S.
Saccharum baldwinii (narrow-plume plume grass	southeastern U. S.
Saccharum brevibarbe var. brevibarbe Saccharum brevibarbe var. contortum (bent-awr Saccharum coarctum (bunched plume grass) Schizachyrium maritimum (seashore false bluest Schizachyrium niveum (pine scrub false bluester	southeastern U. S. tem) LA, AL, MS, FL
Schizachyrium rhizomatum (Florida false blueste Schizachyrium scoparium var. divergens (little b Schizachyrium scoparium var. neomexicanum (li Schizachyrium scoparium var. stoloniferum (little Schizachyrium sericatum (little bluestem)	luéstemouth-central & southeastern U. S. ttle bluéstem) AZ, NM, TX
Scribneria bolanderi (Scribner's grass) Setaria reverchonii ssp. firmula [ssp. not in K & Sorghastrum elliottii (slender Indian grass) Sorghastrum secundum (lop-sided Indian grass) Spartina bakeri (sand cord grass)	south-central & southeastern U. S.
Spartina cynosuroides (big cord grass)	MA to TX gulf coast
Sphenopholis filiformis (long-leaf wedge grass)	southeastern U. S.
Sphenopholis longiflora (Texas wedge grass)	TX, AR, LA
Sphenopholis x pallens (wedge grass)	eastern U. S.
Sphenopholis pensylvanica (swamp wedge grass	eastern & southeastern U. S.
Sporobolus clandestinus (hidden dropseed) Sporobolus compositus var. drummondii (meado Sporobolus compositus var. macer (Mississippi d Sporobolus coromandelianus (whorled dropseed) Sporobolus floridanus (Florida dropseed)	ropseed) south-central U. S.

ΑZ

Sporobolus interruptus (black dropseed)

Sporobolus pinetorum (Carolina dropseed) NC, SC, GA Sporobolus silveanus (Silveus' dropseed) OK, TX, LA NC, SC, GA, AL Sporobolus teretifolius (wire-leaf dropseed) Sporobolus texanus (Texas dropseed) south-central & southwestern U. S. Sporobolus tharpii (Tharp's dropseed) Padre Island, TX Stipa arida (funeral grass) southwestern Ú. S. Stipa californica (California needle grass) Stipa cernua (nodding needle grass) WA, OR, CA, ID, NV CA Stipa coronata (giant stipa) CA (CFP) CA (CFP) Stipa diegoensis (San Diego needle grass) Stipa x latiglumis (Yosemite needle grass) Stipa lemmonii var. pubescens (Crampton's needle grass) Stipa lepida (small-flowered neèdle grass) CA, AZ (MX ???) Stipa lettermanii (Letterman's needle grass) western U.S. Stipa lobata (little-awn needle grass) AZ, NM, TX Stipa nevadensis (Nevada rice grass) western U.S. CA (Canada ??) Stipa occidentalis var. occidentalis (western needle grass) Stipa perplexa (New Mexico rice grass) UT, AZ, NM, TX Stipa pinetorum (pine woods needle grass) western U.S. Stipa porteri (Porter's needle grass) Stipa pulchra (purple needle grass) CA (CFP) western & central U. S. Stipa scribneri (Scribner's needle grass) Stipa shoshoneana (Shoshone needle grass) ID, NV Stipa stillmanii (Stillman's needle grass) Stipa thurberiana (Thurber's needle grass) western U.S. Stipa webberi (Webber's mountain-rice) western U.S. Inyo Co., CA OR, CA, NV Swallenia alexandrae (Eureka Valley dune grass) Torreyochloa erecta (spiked alkali grass) Tridens ambiguus (pine barrens tridens) NC to TX Tridens buckleyanus (Buckley's tridens) Tridens carolinianus (Carolina fluff grass) NC to LA *Tridens congestus* (pink tridens) TX KS to TX, southeastern U. S. Tridens flavus var. chapmanii (Chapman's tridens) Tridens muticus var. elongatus (rough tridens) southwest & south-central U. S. Tridens strictus (long-spike tridens) south-central & southeastern U. S. Triplasis americana (perennial sand grass) southeastern U. S. Tripsacum floridanum (Florida gama grass) Trisetum orthocahaetium (bitter root false oat) Missoula Co., MT Trisetum projectum Tuctoria greenei (awnless spiral grass) Tuctoria mucronata (prickly spiral grass) Solano Co., CA Vaseyochloa multinervosa (Texas grass) Hayes Co., TX Zizańia texana (Texas wild-rice)

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4.12 - INTRODUCED GRASSES WITH LIMITED DISTRIBUTION*

Acrachne racemosa (goose grass)	Riverside Co., CA
Aegilops crassa (Persian goat grass)	NY
Aegilops geniculata (ovate goat grass)	CA, VA, NY
Aegopogon cenchroides (Guatemalan fragile grass)	, CA
Agropyron orientale (oriental wheat grass)	NY
Agropyron squarrosum	NY
Agrostis tandilensis (Argentine bent grass)	CA
Aira caryophyllea var. cupiana (silver hair grass)	CA
Alloteropsis cimicina (bug seed grass)	MD, FL
Alopecurus creticus (Cretan meadow foxtail)	PA (historic)

Alopecurus rendlei (Rendle's meadow foxtail) Ampelodesmos mauritanica (dis grass or it is dat grass?) Amphibromus neesii (wallaby grass) Amphibromus scabrivalvis (swamp wallaby grass) Andropogon bicornis (barbas de indio)	PA CA CA CA, LA FL
Anthephora hermaphrodita (old field grass) Apluda mutica (Mauritian grass) Avena occidentalis (western oat) Avena strigosa (black oat) Bambusa multiplex (hedge bamboo)	FL MD CA CA, MA FL
Bambusa oldhamii (Oldham's bamboo) Bambusa vulgaris (common bamboo) Brachypodium caespitosum (false brome) Brachypodium phoenicoides (Mediterranean false brome) Brachypodium sylvaticum (slender false brome)	TX SC, FL CA CA Benton Co., OR
Bromus alopecuros Bromus hordeaceus ssp. pseudothominei (soft brome) Bromus lepidus (slender soft brome) Cenchrus biflorus (Indian sandbur) Chloris barbata (swollen finger grass)	Source ??? MO, NY NY, CT, MA AL, NY TX, LA
Chloris elata (many-flowered windmill grass) Chloris pectinata (comb windmill grass) Chloris radiata (radiate windmill grass) Chloris truncata (Australian finger grass) Chloris ventricosa (plump windmill grass)	MS, FL SC (historic) OR (extirpated?), FL CA, SC SC, VA
Chrysopogon fulvus (red false beard grass) Chrysopogon pauciflorus (Florida rhaphis) Chrysopogon zizanioides (khus-khus) Cladoraphis cyperoides (bristly love grass) Coleanthus subtilis (moss grass)	FL TX, NC, FL LA OR (historic) WA, OR
Cortaderia jubata (purple pampas grass) Crypsis alopecuroides (foxtail prickle grass) Crypsis schoenoides (swamp prickle grass) Crypsis vaginiflora (modest prickle grass) Cutandia memphitica (Memphis grass)	OR, CA WA, OR, CA OR, CA, UT CA, NV, ID CA
Cymbopogon iwarancusa (oil grass) Cymbopogon nardus (citronella grass) Cynodon aethiopicus (Ethiopian dogtooth grass) Cynodon dactylon var. aridus (couch grass) Cynodon dactylon var. x magennisii (Magennis' Bermuda grass)	FL FL TX, FL AZ TX, AL
Cynodon aristiglumis Cynodon nlemfuensis var. nlemfuensis (African Bermuda grass) Cynodon nlemfuensis var. robustus (African Bermuda grass) Cynodon plectostachyus (star grass) Dactylis glomerata var. aschersoniana (orchard grass)	Source ??? TX TX CA NY
Dactyloctenium radulans (button grass) Dasypyrum villosum (mosquito grass) Dendrocalamus latiflorus (giant bamboo) Dichanthium annulatum (Kleberg bluestem) Dichanthium sericeum (Queensland bluestem)	AZ, MA, SC PA (historic) NC TX, LA, FL TX, FL
Digitaria longiflora (Indian crab grass) Digitaria milanjiana (Madagascar finger grass) Digitaria nuda (naked crab grass) Digitaria setigera (East Indian crab grass) Dinebra retroflexa (viper grass)	FL TX, FL FL CA, MD, NC
Echinochloa crusgalli ssp. spiralis (barnyard grass) Echinochloa esculenta (Japanese millet) Echinochloa oryzicola (late barnyard grass) Echinochloa oryzoides (early barnyard grass) Echinochloa utilis	CA MO CA CA, LA Source ???
Ehrharta calycina (weeping veldt grass)	CA, TX

Ehrharta erecta (panic veldt grass) Ehrharta longiflora (long-flowered veldt grass) Eleusine coracana ssp. africana (finger millet, ragi) Elyhordeum atlanticus (Russian quack grass)	CA CA SC OR, CA
Elymus pycnanthus (tick grass) Elymus racemosus (giant wild rye) Enneapogon cenchroides (soft feather pappus grass) Enneapogon mollis (soft feather pappus grass) Enteropogon dolichostachyus	OR, NB, TX WA, OR (?), WY AZ AZ SC
Enteropogon prieurii (Prieuri's umbrella grass) Eragrostis airoides (darnel love grass) Eragrostis atrovirens (thalia love grass) Eragrostis ciliaris var. laxa (gopher tail love grass) Eragrostis cylindriflora (tube-flowered love grass)	NC, AL (historic) Brazos Co., TX FL FL MD
Eragrostis echinochloidea (African love grass) Eragrostis elongata (long love grass) Eragrostis gangetica (slim-flowered love grass) Eragrostis plana (South African love grass) Eragrostis scaligera (tender love grass)	AZ, MD DC, SC, FL LA, MS, FL SC FL
Eragrostis setifolia (never-fail love grass) Eragrostis unioloides (Chinese love grass) Eremochloa ciliaris (centipede grass) Eriochloa fatmensis (tropical cup grass) Eriochloa leersioides (sharp cup grass)	SC MD, GA, FL San Francisco Co., Ca (historic) CA, AZ, MS CA (historic)
Eriochloa polystachya (Carib grass) Eriochloa pseudoacrotricha (vernal cup grass) Eustachys distichophylla (weeping love grass) Eustachys paspaloides var. caribaea Festuca arvernensis (blue fescue)	TX, MS, FL TX, MS CA, TX (?), GA, FL Source ??? CA (?), NM
Festuca ciliata (fringed six-weeks grass) Festuca gigantea (giant fescue) Festuca heteromalla (spreading fescue) Festuca rigescens Festulolium loliaceum	PA MI, CT, NY WI AZ (historic) NY, SC
Fingerhuthia africana (zulu-fescue) Gaudinia fragilis (fragile-oat) Glyceria declinata (waxy manna grass) Gynerium sagittatum (uva grass, wild cane) Hakonechloa macra (Hakone grass, Japanese forest grass)	AZ Sonoma Co., CA CA, LA, NY FL UT
Hemarthria altissima (limpo grass) Hordeum bulbosum (bulbous barley) Hyparrhenia hirta (thatch grass) Hyparrhenia rufa (jaraguá grass) Ischaemum indicum (Indian muraina grass)	TX, FL CA CA, TX, FL FL MD
Ischaemum rugosum (ribbed muraina grass) Lamarckia aurea (goldentop) Leptochloa chloridiformis (Argentine sprangletop) Leptochloa digitata (cane sprangletop) Lolium temulentum ssp. remotum (poison darnel)	TX, MD CA, AZ, TX TX (historic) SC ND (historic)
Mibora minima (early sand grass) Milium vernale (spring millet grass) Miscanthus floridulus (giant Chinese silver grass) Muhlenbergia diversiglumis Neyraudia arundinacea (Madagascar grass)	NY, MA ID MO, AR Galveston Co., TX CA (unverified)
Neyraudia reynaudiana (Burma reed) Opizia stolonifera (Acapulco grass) Oplismenus hirtellus var. hirtellus (wood grass) Oplismenus hirtellus var. undulatifolius (basket grass) Oryza rufipogon (wild red rice)	FL FL MD CA, FL
Oryza sativa var. fatua (rice) Panicum alatum var. alatum (winged panic grass)	FL CA, AZ

Panicum alatum var. longiflorum (winged panic grass) Panicum bergii (Berg's panic grass) Panicum napaliense (Napal panic grass)	CA TX, GA, AL NM
Panicum paludosum (aquatic panic grass) Pappophorum pappiferum (limestone pappus grass) Parapholis strigosa (hard grass) Paspalum almum (Comb's crown grass) Paspalum coryphaeum (emperor crown grass)	MD (Peterson et al. 2001: 179) Humboldt Co., CA TX, LA FL
Paspalum fimbriatum (fringed crown grass) Paspalum malacophyllum (ribbed crown grass) Paspalum modestum (water paspalum) Paspalum nicorae (Brunswick grass) Paspalum notatum var. latiflorum	FL TX TX, LA AL, GA, FL TX (Source ???)
Paspalum wrightii (Wright's paspalum) Pennisetum advena (purple feather grass) Pennisetum clandestinum (kikuyu grass) Pennisetum flaccidum (Himalayan fountain grass) Pennisetum latifolium (Uruguay fountain grass)	TX CA, TX CA, AZ TX CA
Pennisetum macrourum (African feather grass) Pennisetum nervosum (bent-spike fountain grass) Pennisetum orientale (laurisa grass) Pennisetum pedicellatum ssp. unispiculum (kyasuma grass) Pennisetum petiolare (petioled fountain grass)	CA CA, TX TX FL IA
Pennisetum polystachyon ssp. setosum (West Indian penniset Pennisetum purpureum (Napier grass, elephant grass) Phalaris aquatica (Harding grass) Phalaris coerulescens Phleum arenarium (sand timothy)	um) AZ, NV, FL CA, TX, FL OR, CA CA OR, MA, NY
Phleum paniculatum (British timothy) Phleum subulatum (Italian timothy) Phragmites karka (tall reed) Phyllostachys dulcis (sweet shoot bamboo) Phyllostachys flexuosa (zig-zag bamboo)	OR, NY OR, MA, PA TX MA MD
Phyllostachys meyeri (Meyer's bamboo) Phyllostachys rubromarginata (red-margined bamboo) Phyllostachys viridiglauscens (green-wax golden bamboo) Piptochaetium setosum (bristly spear grass) Piptochaetium stipoides var. purpurascens	MD SC NJ CA CA
Pleioblastus humilis (dwarf bamboo) Pleioblastus simonii (Simon's bamboo) Poa chaixii (Chaix's spear grass) Pogonarthria squarrosa (herringbone grass) Polypogon australis (Chilean beard grass)	CA NV, GA MN, NY Cochise Co., CA WA, CA, NM
Polypogon imberbis (bear grass) Polytrias amaura (Java grass) Puccinellia rupestris (British alkali grass) Rytidodosperma biannulare (wallaby grass) Rytidosperma penicillatum (hairy wallaby grass)	CA FL WA, PA, NY OR, CA OR, CA
Rytidosperma racemosum (wallaby grass) Rytidosperma semiannulare (wallaby grass) Saccharum bengalense (wild sugar cane) Saccharum spontaneum (wild Asian sugar cane) Schizachyrium sanguineum var. sanguineum (crimson false bl	CA CA Zuloaga et al. 2003: 551 FL uestem) AL, FL
Secale montanum (wild rye) Setaria barbata (Mary grass) Setaria megaphylla (big leaf bristle grass) Setaria nigrirostris (black bristle grass) Setaria pumila ssp. pallidefusca (yellow bristle grass)	WA, CA MS, FL LA, FL OR (historic) OR, LA
Setaria rariflora (Brazilian bristle grass) Setaria setosa (West Indian bristle grass) Setaria sphacelata (golden-timothy)	AL, FL NJ, AL, FL CA, AL, FL

Setariopsis auriculata Sorghum bicolor ssp. arundinaceum (wild sorghum)	Pima Co., AZ CA, FL
Sporobolus creber Stipa capensis (Mediterranean needle grass) Stipa manicata (Ecuador needle grass) Stipa neesiana (Uruguayan tussock grass) Stipa papposa (false rice grass)	Glenn Co., CA CA CA AL ????
Stipa plumosa (South American rice grass) Themeda arguens (Christmas grass) Themeda quadrivalvis var. helferi (kangaroo grass) Themeda quadrivalvis var. quadrivalvis (grader grass) Themeda triandra (rooi grass)	CA VA, MD KS CA, LA, FL TX
Thysanolaena latifolia (tiger grass) Tragus australianus (Australian bur grass) Tragus berteronianus (spike bur grass) Tragus heptaneuron (Kenyan bur grass) Tribolium obliterum (cape grass)	??? SC AZ, NM, TX SC Monterey Co., CA
Triraphis mollis (purple heads) Trisetum aureum (golden false oat) Triticum spelta (spelt wheat) Triticum turgidum (rivet wheat) Urochloa arrecta (African signal grass)	TX NJ (historic) KT, VT NY FL
Urochloa brizantha (palisade signal grass) Urochloa mosambicensis (sabi grass) Urochloa oligobrachiata (few-bracted liverseed grass) Urochloa piligera (hairy signal grass) Urochloa platytaenia	TX TX FL FL Source ???
Urochloa subquadripara (two-fingered guinea grass) Urochloa villosa (two-ranked liverseed grass) Zea mays ssp. parviglumis (teosinte) Zea perennis (Mexican teosinte) Zoysia matrella var. matrella (Manila temple grass) Zoysia tenuifolia (Mascarene grass)	FL MD, VA FL SC AL, GA, FL LA, FL

 $^{^{}st}$ I define limited distribution as those grasses found in no more than three states. This is, of course, an entirely arbitrary decision. You can easily see which ones are found in only one or two states, if you should prefer a narrower limit.

Revised: 02 January 2005

SECTION 5 - GRASS IDENTIFICATION

5.01 - SPECIMEN COLLECTION & PREPARATION

The principal reason for collecting is to provide permanent, representative specimens of plants for future study. In the case of smaller vascular plants, such as annual herbs, the specimen often consists of one to several complete individuals. In larger plants, such as trees or shrubs, a specimen usually consists of representative portions of vegetative and reproductive material.

Many specimens collected by students in university botany classes or by the serious amateur will eventually become housed in an herbarium, a permanent collection of pressed and dried plant specimens. Here the plants will be examined by botanists interested in such matters as distribution, verification of determinations, blooming and fruiting times, general morphological features, and anatomical details. Herbarium specimens are frequently loaned to experts doing monographic work and duplicates are often exchanged among herbaria.

EQUIPMENT

The following items are useful: field press, plant press, plastic bags, digger, clippers, pocket knife, compass, altimeter, coin envelopes, pickling fluids, camera, and notebook. While none of them is absolutely essential, having the proper collecting gear close at hand can result in greater efficiency and better specimens. By a "digger," I mean any of a variety of implements, such as a geologist's pick, a dandelion digger, a gardener's trowel or even a large screwdriver.

Although specimens may be stored temporarily in plastic bags or other containers, they should be pressed as soon as possible. Pressing flattens the plants so that they do not curl or wrinkle and it also brings the plant parts into direct contact with newspapers and indirectly with blotters and corrugates, thereby beginning the drying process. There are two types of plant presses. One is the temporary field press. It is usually small, light-weight, and easy to carry in a pack. You do not buy a field press; you make your own out of cardboard or pressboard end pieces, newspapers, and perhaps a few blotters, the whole thing being bound up by a strap or belt. Those of you who are backers will find that you can accommodate an amazing number of plants in a field press. Specimens will last for a few days in such a temporary press until you can transfer them to a regular press.

A standard plant press (12" X 18") is too bulky and heavy to carry about in the field. Although you can construct your own, most of them are purchased, usually at great price, from one of the biological supply houses. A regular plant press has wooden or light metal end pieces called frames. Between the two

frames is a series of blotters, and corrugates or ventilators arranged in a particular sequence. Two common arrangements are repeating units made up of corrugate-blotter-blotter-corrugate, and corrugate-blotter-blotter-blotter-corrugate. In the first plan, a specimen in a single fold of newspaper is inserted between the two blotters (corrugate-blotter-specimen-blotter-corrugate). In the second option, two specimens are inserted (corrugate-blotter-specimen-blotter-specimen-blotter-specimen-blotter-specimen-blotter-specimen-blotter-specimen about a foot or so of pressing material in it.

COLLECTING SPECIMENS

You should be guided by one overriding consideration, to collect and prepare a permanent specimen that is as much like the living plant as possible, given the constraints of the pressing and drying techniques. Flower color may fade or change and three dimensional forms are flattened, but a wealth of scientific information and even a certain aesthetic quality remain intact.

Always keep in mind that the specimen that you collect must be determined eventually. Most keys and descriptions rely heavily upon the structure of the flower and fruit. Collecting herbaceous plants in the vegetative state is probably futile. I suggest that you collect extra flowering and fruiting material for use during the identification process. In this way the specimen itself can remain intact. This additional material should be submitted as a part of the specimen. It will be placed in a fragment folder and mounted on the herbarium sheet along with the plant. As you become more familiar with the genera, you will learn what plant features are critical for accurate determination.

With herbaceous plants, it is also standard practice to gather underground parts. The nature of the root system or subterranean stems may be critical. "Top-snatching" is a dreadful habit. Roots and other underground plant parts should be cleaned carefully to remove soil or mud.

A major problem facing the inexperienced collector is what constitutes enough plant material to make an acceptable specimen. In the case of small annuals, a specimen is not a single plant, but a few to many, depending upon their size. A single larger annual or smaller perennial is usually sufficient. With experience comes the almost unconscious habit of deciding that a particular plant will make a suitable specimen because it will fit on an herbarium sheet of 12" X 18". However, many larger herbs and most woody plants are too large to accommodate within these limits. Special techniques are used here. These will be discussed later.

PRESSING SPECIMENS

Plants are first placed in a single fold of newsprint. One of the most common errors is to assume that if a single fold of newspaper is good, then an entire section will be just that much better. All you accomplish is retarding the drying process by having several layers of wet newsprint. Tabloid newspaper, such as "The National Inquirer" or "The Lumberjack" are just the right size. If you use a full-sized newspaper, then tear it down the middle to yield two single fold sections of about 11" X 15". Do not exceed this size or the plant specimens may not fit on the herbarium sheet. Do not use slick, clay-finish newsprint from magazines or catalogues. It will not absorb moisture from the specimen.

Annuals and small perennials fit nicely in the newspaper and present no particular problem. But, some herbs are too tall and/or broad to be accommodated properly. If the problem is mainly one of height, consider folding the plant. This works well if it is no more than about a half meter tall. Fold the plant in such a way that the parts do not obscure one another. Too much bulk may also impair proper drying. Make sharp bends, not gently rounded ones. These may be held in place during the drying process by using **flexostats**. You make you own by cutting and index card or computer card into segments about 4 cm x 8 cm. Cut a slit about 3 cm long in each and slip the "knee" of the plant through the opening. After the plant has dried, remove the flexostat and reuse it. Still larger plants may be subdivided into two or more sections. Such a suite of specimens is often the most practical method of collecting larger herbaceous plants.

It is important that you put only one kind of plant inside the newspaper. The collection number (see below) for that particular plant should be written prominently along one margin. This will assist you later in sorting material and in finding a particular specimen. Some arranging of plant parts and trimming can now be done. Leaves and stems should be positioned so that they do not overlap unnecessarily. Leaf blades should be turned so that some of them have the upper surface exposed, while in others the lower surface may be seen in the final specimen. Specimen quality can often be improved by some judicious pruning of excess bulk. If parts are removed, leave a short stub so that it is evident what has occurred. This is also a good time to get rid of the dirt or mud trapped in the roots. It can ruin the specimen and label if allowed to remain.

This process of putting specimens in a single fold of newsprint, trimming and arranging, and assigning collection numbers (see below) is done until the press is filled or you have run out of plants. The plant press is now closed by tightening the straps, belts or ropes. It must be cinched tight enough to flatten the specimens and bring them into firm contact with the pressing materials. Presses will loosen as the plant dry. Tighten the straps from time to time.

FIELD DATA

At the same time that it is vital to collect and prepare adequate plant specimens, it is just as critical to take down the necessary field data. Without them, the specimens are scientifically worthless. Data may be

recorded in permanent notebooks carried into the field or written in temporary pocket notebooks. Either method has its advantages and disadvantages. The important point is, however, to write down your field data, rather than relying on your memory.

The collection site is probably the single most important bit of data. This should be as precise as possible. I suggest the following sequence:

- 1) 2) 3) 4) county or parish
- quadrangle name
- tier, range, and section (or latitude and longitude)
- reference to a more or less permanent location, such as towns, highways, rivers, particularly those that can be found on ordinary highway maps

Quadrangle names, tier, range and section coordinates, and latitude/longitude are found on topographic maps available from the United States Geological Survey. Some of this information may also be found on U.S. Forest Service and Bureau of Land Management maps. The new hand-held GPS devices allow for very accurate site data.

Other data that you should enter in your records include:

- 1) habitat information (vegetation type, associated species, geology of the site, soil type, etc.)
- 2) 3) 4) remarks on the frequency of the plant at that site
- remarks on the plant itself (size, color, odor, etc.)
- collection date
- personal collection number for that plant specimen

The personal collection number is the number that you assign to this particular specimen. A different collection number is given to each different collection of a particular kind of plant that you make during your career as a field botanist. Your first collection bears the number "1". You will now use a different number anytime you collect a new plant at this site, anytime you move from one location to another, or anytime you collect on a different day. Perhaps a few illustrations will help to clarify this matter.

- If I collect ten different kinds of plants at a certain site, I will have ten collection numbers.
- If I collected each of the ten plants in duplicate or triplicate, I would still have only ten collection numbers, each in duplicate or triplicate. This is the only situation in which a collection number is used more than once.
- If I move to a second site and collect five more plants, I will have five more collection numbers. This is true whether or not any or all of the five plants duplicate species collected at the first site. New numbers are assigned because this a different collection site.
- If I should return to any of these sites at a later date, all of the plant collections made at that time would get new numbers.

It is not uncommon to be unsure whether two plants belong to the same or different species or varieties. If in doubt, assign them different numbers. Should they later turn out to be the same thing, combine them under the first collection number. If what you thought in the field to be duplicates are later determined as two or more different taxa, then call one of them 682 and the other 682A or 682A and 682B.

DRYING SPECIMENS

Once plants have been put into the plant press, they must be dried. Presses may be left out in the sunlight or they may be strapped into rooftop racks on automobiles, much to the curiosity of fellow motorists. But the usual method is to put the plant press in an electric or steam drier. These are found on most college campuses.

How long should plants remain in the drier? Until the plants are dry and no longer. The length of time will depend upon the kind of drier, the types of plants collected, the arrangement of pressing materials in the press, and how many other presses are in the drier. While 48 hours is often sufficient for most plants, it is critical to check the presses. Are the newspapers still slightly damp? Does the plant still feel and smell wet? Will your thumbnail leave an impression in a stem or petiole? If the answer is "yes" to any of these questions, then the plant needs to remain in the drier. If you take them out too soon, the plants will mold. If, however, they are dried too long, they will discolor badly and become very brittle. Remember to check the straps periodically. Presses will loosen during the drying process and curling of plant parts can occur.

LABELS

If specimens are to be deposited in an herbarium or submitted as part of a class requirement, they must be accompanied by a label that gives the pertinent collection data for that plant. Labels should be made from high quality paper, preferably 100% rag content bond paper. Most herbaria supply them to collectors. Label information should be typed. Permanent ink is an acceptable alternative. do not use ballpoint pen or soft lead pencil. Labels should provide at least the following information:

- 1) scientific name of the plant
- 2) location data
- 3) location date
- 4) your name
- 5) collection number for the specimen

The scientific name, for purposes of completeness and accuracy, must include the authority. Location data has already been discussed. Dates should be presented as 12 March 1979 or March 12, 1979, not 3-12-79. In the last example, the date may be read in at least four different ways. Use your first name or initials, not just your last name, unless you are Carolus Linnaeus, Willis Lynn Jepson, Asa Gray, or some other equally famous dead botanist. Put the collection number beside your name.

In addition to these essential elements, you may also wish to provide habitat data, along with commentary on the plant itself. A series of sample labels is appended to this handout. Once completed, the labels are slipped inside the newspaper with the plant

specimen. Do not glue, tape, or staple either the plant or the label to the newspaper. Both will eventually be removed and mounted on an herbarium sheet for permanent reference.

THE ETHICS OF PLANT COLLECTING

While there are certainly valid educational and scientific reasons for plant collecting, important questions should be considered before taking specimens. Will the collecting of this plant contribute to educational or scientific advancement? What will be the impact on the population of the removal of this plant?

In 1993, California Native Plant Society adopted a set of guidelines regarding plant collecting for educational/scientific purposes. The following points are based on that statement.

- It is illegal to collect plants along a highway right-ofway, in national parks, national monuments or national forests, state parks, or most local parks without a collecting permit, which may be obtained from the appropriate supervising agency.
- It is legal and permissible to collect plants on private lands, provided that permission of the landowner is obtained.
- It is the responsibility of an instructor to ensure that students are made aware of rare plants that are endemic to the area in which collecting is to take place, and to caution students against collecting these plants.
- *Collecting should be limited to the taking of as little plant material as necessary to allow identification.
- *Collecting should be done inconspicuously. Casual observers may not understand the reasons for such activities and may feel they can do likewise.
- The Society encourages all botany instructors to use common plants, especially weedy or garden species, for demonstrating collecting techniques, structures, and taxonomic features.
- The primary justification for collecting plants for herbaria is that they contribute to increased knowledge of the California flora. Repeated collecting in well known areas may serve no useful purpose. While it is important to document the distribution of plants, including rare species, it is critical to evaluate the impact of collecting.
- A key to ensuring preservation of California's diverse flora and fauna is to develop a public informed of their value. For this reason, CNPS encourages limited and discriminating collection of plants as part of the educational process.

SOME FAMOUS LAST WORDS

"No, let's not stop here. There will be lots more of them down the road."

"No reason to collect this. It's just a weed."

"I really shouldn't take the last one, but"

"This stuff lasts forever in a plastic bag. We'll press all these plants when we get back to campus."

"Sure, dump everything in the same plastic bag. We can sort them out later."

"Are you kidding? Why number them now? We'll never get these specimens mixed up."

"Let's not take the time. We'll be able to find all of these locations on the maps when we get back home."

"No, this plant press isn't too high."

"Why anchor this stuff down? There's no wind."

"Oh well, you probably didn't need the rhizome anyway."

"Get that one. It will fit in the press!"

SELECTED REFERENCES

Brayshaw, T. C. 1973. Plant collecting for the amateur. Museum Methods Manual 1. British Columbia Provincial Museum. Victoria. 15 pp.

DeWolf, G. P., Jr. 1968. Notes on making an herbarium. Arnoldia 28: 69-111.

Follette, W. 1977. To reduce the impact on native plants. CNPS guidelines on collecting, plant sales, and field trips. Fremontia 4(4): 20-22.

Gould, F. W. 1983. The herbarium and the preparation and handling of grass specimens. \underline{In} , Gould, F. W. & R. B. Shaw. Grass systematics. Second edition. Texas A & M Univ. Press. College Station. Pp. 359-363.

Huxley, A. 1976. The ethics of plant collecting. Fremontia 4(2): 17-21.

Radford, A. E. et al. 1974. Vascular plant systematics. Harper & Row. New York, NY. Pp. 387-398.

Smith, C. E., Jr. 1971. Preparing herbarium specimens of vascular plants. Bull. No. 348. U. S. Dept. of Agric. Washington, D. C. 29 pp.

Smith, J. P., Jr. 1977. Collection and preparation of vascular plant specimens. <u>In</u>, Vascular plant families. Mad River Press. Eureka, CA. Pp. 282-287.

5.02 - GRASS FLORAS & CHECKLISTS

GRASS NAMES (Authors, Where Published, Synonyms, etc.)

Chase, A. & C. Niles. 1962. Index to grass species. Three vols. G. K. Hall. Boston, MA.

Hitchcock, A. S. 1951. Synonymy. <u>In</u>, Hitchcock, A. S. Manual of the grasses of the United States. Revised by A. Chase. U. S. Dept. Agric. Misc. Publ. No. 200. Washington, D. C. Pp. 796-980.

Judziewicz, E. J. et al. 2000. Catalogue of New World grasses (Poaceae). I. Subfamilies Anomochlooideae, Bambusoideae, Ehrhartoideae, and Pharoideae. Contr. U. S. Natl. Herb. 39: 1-128.

Peterson, P. M. et al. 2001. Catalogue of New World grasses (Poaceae): II. Subfamily Chloridoideae. Contr. U. S. Natl. Herb. 41: 1-255.

Soreng, R. J. et al. 2000 ->. Catalogue of New World grasses (Poaceae).

http://mobot.mobot.org/W3T/Search/nwgc.html

Soreng, R. J. et al. 2003. Catalogue of New World grasses (Poaceae). IV. Subfamily Pooideae. Contr. U. S. Natl. Herb. 48: 1-730.

Zuloaga, F. O. et al. Catalogue of New World grasses (Poaceae). III. Subfamiles Panicoideae, Aristidoideae, Arundinoideae, and Donthonioideae. Contr. U. S. Natl. Herb. 46: 1-662.

GENERAL REFERENCES

Baker, F. W. G. & P. J. Terry (editors). 1991. Tropical grassy weeds. CAB International. Wallingford, England. 203 pp.

Behrendt, S. & M. Hanf. 1979. Grass weeds in world agriculture: identification in the flowerless state. Ludwigshafen, Basf Aktiengesellschaft. 159 pp.

Bews, J. W. 1929. The world's grasses, their differentiation, distribution, economics and ecology. Longman, Green & Co. London, England. 408 pp.

Clayton, W. D. & S. A. Renzoize. 1986. Genera graminum: grasses of the world. Kew Bull. Addt. Series XIII. Royal Botanic Gardens, Kew. Her Majesty's Stationery Office. London, England. 389 pp.

Gould, F. W. & R. B. Shaw. 1983. Grass systematics. Second edition. Texas A & M Univ. Press. College Station. 397 pp.

Hackel, E. 1890. The true grasses. Translated from Die natürlichen Pflanzenfamilien by F. Lamson-Scribner and E. A. Southworth. Henry Holt & Co. New York, NY. 228 pp.

Pilger, R. 1956. Gramineae II. \underline{In} , Die natürlichen Pflanzenfamilien, Melchoir H. & E. Werdermann (editors). Band 14d. Duncker & Humblot. Berlin. 225 pp.

Pilger, R. 1960. Gramineae III. <u>In</u>, Die natürlichen Pflanzenfamilien, H. Harms & J. Mattfeld (editors). Band 14e. Duncker & Humblot. Berlin. 208 pp.

Potztal, E. 1956. Gramineae III. <u>In</u>, Die natürlichen Pflanzenfamilien, Melchoir H. & E. Werdermann (editors). Band 14e. Duncker & Humblot. Berlin.

Roshevits, R. Yu. 1937. Grasses. An introduction to the study of fodder and cereal grasses. Translated from the Russian. Indian National Sci. Documentation Centre. New Delhi. Two vols. 635 pp.

Watson, L. & M. J. Dallwitz. 1992. The grass genera of the world. CAB International. Wallingford, England. 1038 pp.

NORTH AMERICA

Barkworth, M. E. & K. M. Capels. 2000. Grasses in North America: a geographic perspective. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 331-350.

Barkworth, M. E., K. M. Capels, S. Long, & M. P. Piep (editors). 2003. Flora of North America north of Mexico. Vol. 25. Magnoliophyta: Commelinidae (in part): Poaceae, part 2. Oxford Univ. Press. New York, NY. 783 pp.

Beal, W. J. 1887-1896. The grasses of North America. Two vols. Henry Holt & Co. New York, NY. 457 pp. + 706 pp.

Hitchcock, A. S. 1931. Poaceae. North American Flora 17(4): 289-354

Hitchcock, A. S. 1935. Poaceae. North American Flora 17(5): 355-418; 17(6): 419-482.

Hitchcock, A. S. 1937. Poaceae. North American Flora 17(7): 483-542.

Hitchcock, A. S., J. R. Swallen, & A. Chase. 1939. Poaceae. North American Flora 17(8): 543-638.

Nash, G. V. 1909. Poaceae. North American Flora 17(1): 77-98.

Nash, G. V. 1912. Poaceae. North American Flora 17(2): 99-196.

Nash, G. V. & A. S. Hitchcock. 1915. Poaceae. North American Flora 17(3): 197-288.

Stubbendieck, J., S. L. Hatch, & C. H. Butterfield. 1997. North American range plants. Fifth edition. Univ. Nebraska Press. Lincoln. 499 pp.

CANADA

Aiken, S. G., L. L. Consaul, & M. J. Dallwitz. 1996. Grasses of the Canadian Arctic Archipelago: A DELTA database for interactive identification and illustrated information retrieval. Canadian J. Bot. 74(11): 1812-1825.

Aiken, S. G. & S. J. Darbyshire. 1983. Grass genera of western Canadian cattle rangelands. Monograph No. 29. Research Branch. Agriculture Canada. Ottawa. 173 pp.

Best, K. F., J. Loman, & J. B. Campbell. 1971. Prairie grasses identified and described by vegetative

characters. Agric. Canada Publ. 1413. Ottawa. 239 pp.

Dore, W. G. & A. E. Roland. 1942. The grasses of Nova Scotia. Proc. Nova Scotia Inst. Sci. 20: 177-288.

Douglas, G. W. et al. 2001. Illustrated flora of British Columbia. Vol. 7. Monocotyledons (Orchidaceae through Zozteraceae). Min. Environment, Lands and Parks.

Hubbard, W. A. 1969. The grasses of British Columbia. British Columbia Prov. Mus. Handbook No. 9. Victoria, BC. 205 pp.

Looman, J. 1982. Prairie grasses identified and described by vegetative characters. Publ. No. 1413. Agriculture Canada. Canadian Gov. Publ. Centre. Ottawa. 244 pp.

McLachlan, K. I. et al. 1989. Grasses of the Queen Elizabeth Islands. Canadian J. Bot. 67(7): 2088-2105.

Nowosad, F. S. et al. 1946. The identification of certain native and naturalized hay and pasture grasses by their vegetative characteristics. Macdonald College Tech. Bull. No. 16. Macdonald College. Quebec, Canada. 78 pp.

Oldham, M. J. et al. 1995. New and noteworthy Ontario grasses (Poaceae) records. Michigan Bot. 34(3): 105-132.

Swallen, J. R. 1938. Additions to the grass flora of British Columbia. J. Washington Acad. Sci. 28(1): 6-11.

Stewart, H. & R. J. Hebda. 2000. Grasses of the Columbia Basin of British Columbia. Min. Forests Res. Program. Royal British Columbia Mus. Victoria, Canada.

Watson, L., S. G. Aiken, M. J. Dallwitz, L. P. Lefkowitch, & M. Dube. 1986. Canadian grass genera: keys and descriptions in English and French from an automated data bank. Canadian J. Bot. 64(1): 53-70.

UNITED STATES (GENERAL)

Alderson, J. & W. C. Sharp. 1995. Grass varieties in the United States. CRC Press. Boca Raton, FL. 296 pp.

Brown, L. 1979. Grasses, an identification guide. Peterson Nature Library. Houghton Mifflin Co. New York, NY. 240 pp.

Griffiths, D. 1915. Native pasture grasses of the United States. Prof. Paper No. 201. U. S. Dept. of Agric. Washington, D. C. 52 pp.

Hanson, A. A. 1965. Grass varieties in the United States. Revised edition. United States Dept. of Agric. Handbook No. 170. 102 pp.

Hitchcock, A. S. 1921. A manual of farm grasses. Publ. by the author. Washington, D. C. 175 pp.

Hitchcock, A. S. 1936. The genera of grasses of the United States. Revised. United States Dept. of Agric. Bull. 772: 1-302.

Hitchcock, A. S. 1951. Manual of the grasses of the

United States. Second edition revised by Agnes Chase. Misc. Publ. No. 200. U. S. Dept. of Agric. Washington, D.C. 1051 pp.

Knobel, E. 1980. Field guide to the grasses, sedges and rushes of the United States. Second revised edition by M.E. Faust. Dover Publ. New York, NY. 83 pp.

Lamson-Scribner, F. 1896. Useful and ornamental grasses. Bull. 3. Div. Agrostology. U. S. Dept. Agric. Washington, D. C. 119 pp.

Lamson-Scribner, F. 1898. American grasses. I. Bull. 7. Div. Agrostology. U. S. Dept. Agric. Washington, D. C. 331 pp.

Lamson-Scribner, F. 1901. American grasses. II. Div. Agrostology. U. S. Dept. Agric. Washington, D. C. 349 pp.

Lamson-Scribner, F. 1900. American grasses. III. Div. Agrostology. U. S. Dept. Agric. Washington, D. C. 197 pp.

Pohl, R. W. 1978. How to know the grasses. Third edition. W. C. Brown Co. Dubuque, IA. 200 pp.

Smith, J. P., Jr. 1981. A key to the genera of grasses of the conterminous United States. Sixth edition. Mad River Press. Eureka, CA. 81 pp.

Smith, J. P., Jr. 1996. A checklist of North American grasses. Misc. Publ. No. 4. Humboldt State Univ. Herbarium. Arcata, CA. 57 pp.

Terrell, E. E. 1970. Checklist of scientific names of introduced agricultural grasses. Agric. Res. Serv. 34-166. United States Dept. of Agric. Washington, DC. 000 pp.

Vasey, G. 1883. The grasses of the United States: being a synopsis of the tribes and genera, with descriptions of the genera, and a list of the species. U. S. Dept. Agric. Special Report No. 63. Washington, D. C. 47 pp.

Weintraub, F. C. 1953. Grasses introduced into the United States. United States Dept. of Agric. Handbook No. 58. 79 pp.

Young, R. A. & J. R. Haun. 1961. Bamboo in the United States: description, culture, and utilization. Key to genera by F. A. McClure. United States Dept. of Agric. Handbook No. 193. 74 pp.

UNITED STATES (REGIONAL)

Angelo, R. & D. E. Boufford. 1998. Atlas of the flora of New England: Poaceae. Rhodora 100: 101-233.

Cronquist, A. et al. 1977. Intermountain flora. Columbia Univ. Press. New York, NY. 6: 175-464.

Fernald, M.L. 1950. Gray's manual of botany. Eighth edition. American Book Co. New York, NY. Pp. 94-236.

Gleason, H. A. 1963. New Britton and Brown illustrated flora of the northeastern United States and adjacent Canada. Hafner Publ. Co. New York, NY. 1: 96-246.

Gleason, H. A. & A. Cronquist. 1991. Manual of vascular plants of northeastern United States and adjacent Canada. Second edition. New York Bot. Gard. Bronx, NY. Pp. 737-818.

Gould, F. W. 1951. Grasses of the southwestern United States. Univ. Arizona Biol. Sci. Bull. No. 7, 22(1): 1-343.

Hafenrichter, A. L. et al. 1979. Grasses and legumes for soil conservation in the Pacific Northwest and Great Basin states. U. S. Dept. Agric. Handbook. No. 339.

Hitchcock, A. S. 1933. Poaceae. <u>In</u>, Small, J.K. Manual of the southeastern flora. Univ. North Carolina Press. Chapel Hill. Pp. 30-139.

Hitchcock, A. S. 1955. Poaceae. \underline{In} , Abrams, L. An illustrated flora of the Pacific states. Stanford Univ. Press. Stanford, CA. 1: 103-255.

Hitchcock, C. L. & A. Cronquist. 1973. Flora of the Pacific Northwest: an illustrated manual. Univ. Washington Press. Seattle, WA. Pp. 602-674.

Hitchcock, C. L. et al. 1969. Vascular plants of the Pacific Northwest. Univ. Washington Press. Seattle. 1: 385-725.

Leithead, H. L. et al. 1971. 100 Native forage grasses in 11 southern states. Agric. Handbook No. 389. United States Dept. of Agric. Washington, D. C. 216 pp.

Phillips, C. E. 1962. Some grasses of the Northeast: a key to their identification by vegetative characters. Univ. Delaware Agric. Exp. Stat. Field Manual No. 2. 77 pp.

Seymour, F. C. 1982. Flora of New England. Second edition. C.E. Tuttle. Rutland, VT. Pp. 54-107.

Sutherland, D. 1986. Poaceae. Great Plains Flora Associ-ation. Flora of the Great Plains. Univ. Press of Kansas. Lawrence. Pp. 1113-1235.

Weber, W. A. 1976. Rocky Mountain flora. Fifth edition. Colorado Associated Univ. Press. Boulder, CO. Pp. 385-425.

Yarlett, L. L. 1996. Common grasses of Florida and the Southeast. Florida Native Plant Soc. Spring Hill. 167 pp.

UNITED STATES (BY STATE)

ALABAMA

Banks, D. J. 1965. A checklist of the grasses (Gramineae) of Alabama. Castanea 30: 84-96.

ALASKA

Scribner, F. L. & E. D. Merril. 1910. The grasses of Alaska. Contr. U. S. Natl. Herb. 13: 47-92.

Welsh, S. L. 1974. Anderson's flora of Alaska and adjacent parts of Canada. Brigham Young Univ. Press. Provo, UT. 724 pp.

ARIZONA

Arizona Native Plant Society. 1993. Desert grasses. Arizona Native Plant Society. 41 pp.

Copple, R. F. & C. P. Pase. 1978. A vegetative key to some common Arizona range grasses. Gen. Tech. Rep. RM-53. 106 pp.

Humphrey, R. R. 1997. Arizona range grasses. Revised by G. B. Ruyle & D. J. Young. Coop. Ext. Coll. Agric. Univ. Arizona. Tucson.

Lehr, J. H. 1978. A catalogue of the flora of Arizona. Desert Bot. Gard. Phoenix, AZ. Pp. 9-25.

Swallen, J. R. 1960. Gramineae. <u>In</u>, Kearney, T. H. & R. H. Peebles. Arizona flora. Univ. California Press. Berkeley. Pp. 70-145.

ARKANSAS

Moore, D. M. 1961. Revised and annotated catalogue of the grasses of Arkansas. Arkansas Acad. Sci. Proc. 15: 9-25.

Robinson, A., Jr. 1964. Report on the Gramineae of the Arkansas Salem Plateau. Trans. Kansas Acad. Sci. 67: 460-469.

Smith, E. B. 1988. An atlas and annotated list of the vascular plants of Arkansas. Second edition. Publ. by the author. Fayetteville, AR. Pp. 421-477.

CALIFORNIA

Beetle, A. A. 1947. Distribution of the native grasses of California. Hilgardia 17: 309-357.

Crampton, B. 1961. The endemic grasses of the California floral province. Leaflts. Western Bot. 9(9&10): 154-158.

Crampton, B. 1974. Grasses in California. Univ. California Press. Berkeley. 178 pp.

Hickman, J. C. (editor). 1993. The Jepson manual: higher plants of California. Univ. California Press. Berkeley. Pp. 1218-1303.

Sampson, A. W. & A. Chase. 1927. Range grasses of California. Univ. California Agric. Exp. Stat. Bull. 430: 1-94.

Smith, J. P., Jr. 2004. An annotated checklist of California grasses. Misc. Publ. No. 5. Humboldt State University Herbarium. Arcata, CA. 63 pp.

COLORADO

Harrington, H. D. 1946. Grasses of Colorado. Colorado A. & M. College. Ft. Collins. 167 pp.

Harrington, H. D. 1954. Manual of the plants of Colorado. Swallow Press. Denver, CO. Pp. 38-116.

Harrington, H. D. & L. W. Durrell. 1944. Key to some Colorado grasses in vegetative condition. Colorado Agric. Exp. Stat. Tech. Bull. 33: 1-86.

Weber, W. A. & R. C. Witman. 1996. Colorado flora: western slope. Revised edition. Colorado Assoc. Univ. Press. Boulder. Pp. 244-277.

Weber, W. A. & R. C. Witman. 1996. Colorado flora: eastern slope. Revised edition. Univ. Press of Colorado. Niwot. Pp. 263-301.

Weber, W. A. & R. C. Wittman. 1992. Catalog of the Colorado flora: a biodiversity baseline. Univ. Press of Colorado. Niwot. Pp. 108-118.

FLORIDA

Clewell, A. F. 1985. Guide to the vascular plants of the Florida panhandle. Florida State Univ. Press. Gainsville. Pp. 106-167.

Hall, D. W. 1978. The grasses of Florida. Ph.D. dissertation. Univ. Florida. University Microfilms International. Ann Arbor, MI. 498 pp.

Long, R. W. & O. Lakela. 1971. A flora of tropical Florida. Univ. Miami Press. Coral Gables, FL. Pp. 132-202.

Ward. D. B. 1968. Checklist of the vascular flora of Florida. Part 1. Tech. Bull. 726, Agric. Exp. Stations. Univ. Florida. Gainsville. 72 pp.

Wunderlin, R. P. & B. F. Hansen. 2003. Guide to the vascular plants of Florida. Second edition. Univ. Press of Florida. Gainesville. 787 pp.

Yarlett, L. L. 1996. Common grasses of Florida and the Southeast. Florida Native Plant Soc. Spring Hill. 167 pp.

GEORGIA

Duncan, W. H. & J. T. Kartesz. 1980. Vascular flora of Georgia. An annotated checklist. Univ. Georgia Press. Athens. Pp. 9-23.

Jones, S. B., Jr. & N. C. Coile. 1988. The distribution of the vascular flora of Georgia. The Herbarium. Dept. of Botany. Univ. Georgia. Athens. 230 pp.

HAWAI'I

Haselwood, E. L. & G. G. Motter (editors). 1983. Handbook of Hawaiian weeds. Second edition revised and expanded by R. T. Hirano. Univ. Hawai'i Press. Honolulu. 491 pp.

Hitchcock, A. S. 1922. Grasses of Hawaii. Mem. Bernice P. Bishop Mus. 8: 101-230.

Rotar, P. P. 1968. Grasses of Hawaii. Univ. Hawai'i Press. Honolulu. 355 pp.

Wagner, W. L., D. R. Herbst, & S. H. Sohmer. 1990. Manual of the flowering plants of Hawai'i. Bishop Mus. Spec. Publ. No. 82. Univ. Hawai'i Press and Bishop Mus. Press. Honolulu. 2: 1481-1604.

Whitney, L. D. E. Y. Hosaka, & J. C. Ripperton. 1939. Grasses of the Hawaiian ranges. Hawaii Agric. Exp. Sta. Bull. No. 82. 148 pp.

IDAHO

Davis, R. J. 1952. Flora of Idaho. W.C. Brown Co. Dubuque, IA. Pp. 80-131.

ILLINOIS

Glassman, S. F. 1964. Grass flora of the Chicago

region. Amer. Midland Naturalist 72: 1-49.

Mohlenbrock. R. H. 2002. The illustrated flora of Illinois: Grasses: *Bromus* to *Paspalum*. Second edition. Southern Illinois Univ. Press. Carbondale. 404 pp.

Mohlenbrock. R. H. 2001. The illustrated flora of Illinois: Grasses: *Panicum* to *Danthonia*. Second edition. Southern Illinois University Press. Carbondale. 455 pp.

Mohlenbrock, R. H. 2002. Vascular flora of Illinois. Southern Illinois Univ. Press. Carbondale. 736 pp.

Mosher, E. 1918. The grasses of Illinois. Univ. Illinois Agric. Exp. Stat. Bull. 205: 257-425.

INDIANA

Crovello, J. A., C. A. Keller, & J. T. Kartesz. 1983. The vascular plants of Indiana: a computer based checklist. American Midl. Nat. and Univ. Notre Dame Press. Notre Dame, IN. Pp. 50-56.

Deam, C. C. 1929. Grasses of Indiana. W. B. Burford. Indianapolis, IN. 356 pp.

Deam, C. C. 1940. Flora of Indiana. Dept. of Conservation. Indianapolis, IN. Pp. 93-181.

IOWA

Eilers, L. J. & D. M. Roosa. 1994. The vascular plants of Iowa: an annotated checklist and natural history. Univ. Iowa Press. Iowa City. Pp. 169-185.

Pohl, R. W. 1966. The grasses of Iowa. Iowa State J. Sci. 40(4): 341-566.

KANSAS

Gates, F. C. 1937. Grasses in Kansas. Report of the Kansas State Board of Agric. 55(220-A): 1-349.

Barkley, T. H. 1968. A manual of the flowering plants of Kansas. Kansas State Univ. Endowment Assoc. Manhattan. 402 pp.

McGregor, R. L., R. E. Brooks, & L. A. Hauser. 1976. Checklist of Kansas vascular plants. Tech. Publ. State Biol. Surv. Kansas 2: 1-168.

Ohlenbusch, P. D. 1976. Range grasses of Kansas. Coop. Ext. Serv. Kansas State Univ. Manhattan. 20 pp.

KENTUCKY

Browne, E. T., Jr. & R. Athey. 1992. Vascular plants of Kentucky: an annotated checklist. Univ. Press of Kentucky. Lexington. Pp. 11-24.

LOUISIANA

Allen, C. M., D. A. Newman, & H. H. Winters. 2004. Grasses of Louisiana. Third edition. Allen's Native Ventures. Pitkin, LA. 374 pp.

MAINE

Ogden, E. C., F. H. Steinmetz, & F. Hyland. 1948. Checklist of the vascular plants of Maine. Bull. Joss. Bot. Soc. 8: 1-70.

MARYLAND

Brown, M. L. & R. G. Brown. 1984. Herbaceous plants of Maryland. Port City Press. Baltimore, MD. 1127 pp.

Norton, J. B. S. 1931. Maryland grasses. Maryland Agric. Exp. Stat. Bull. 323: 251-326.

MASSACHUSETTS

Ahmadjian, V. 1979. Flowering plants of Massachusetts. Univ. Massachusetts Press. Amherst. 582 pp.

MICHIGAN

Moore, W. O. 1977. Some very common grasses. Michigan Bot. 16(4): 167-188.

Voss, E. G. 1972. Michigan flora: a guide to the identification and occurrence of the native and naturalized seed-plants of the state. Part I: Gymnosperms and monocots. Cranbrook Inst. Sci. Bull. 55. Pp. 109-244.

MINNESOTA

Cholewa, A. F. 2003. Checklist of the flora of Minnesota. www.cbs.umn.edu/herbarium/checklis2.htm

Ownbey, G. & T. Morley. 1991. Vascular plants of Minnesota: a checklist and atlas. Univ. Minnesota Press. Minneapolis. Pp. 37-46.

MISSISSIPPI

Bennett, H. W., R. O. Hammons, & W. R. Weissinger. 1950. The identification of 76 grasses by vegetative morphology. Mississippi State College Agric. Exp. Stat. Tech. Bull. 31: 1-108.

Lowe, E. N. 1921. Plants of Mississippi. Bull. Mississippi State Geol. Survey 17: 1-292.

MISSOURI

Gereau, R. E. 1987. Grasses of Missouri: an annotated checklist. Missouriensis 8: 49-70.

Kucera, C. L. 1998. The grasses of Missouri. Second edition. Univ. Missouri Press. Columbia. 305 pp.

Steyermark, J. A. 1963. Flora of Missouri. Iowa State Univ. Press. Ames. Pp. 68-254.

Yatskievych, G. & J. Turner. 1990. Catalog of the flora of Missouri. Monographs in Syst. Bot. Vol. 37. Missouri Bot. Gard. St. Louis. Pp. 249-275.

MONTANA

Dorn, R. D. 1984. Vascular plants of Montana. Mountain West Publ. Co. Cheyenne, WY. 276 pp.

Hitchcock, C. L. 1937. A key to the grasses of Montana based upon vegetative characters. J. S. Swift Co. St. Louis, MO. 30 pp.

Booth, W. E. 1950. Flora of Montana (Part I, conifers and monocots). Montana State College Research Foundation. Bozeman. 232 pp.

NEBRASKA

Bessey, C. E. 1905. The grasses of Nebraska. Annual Report State Board Agric. 1904: 175-205.

Frolik, A. L. & F. D. Keim. 1938. Common native grasses of Nebraska. Univ. Nebraska Exp. Stat. Circular 59. 52 pp.

Keim, F. D., G. W. Beadle, & A. L. Frolik. 1932. The identification of the more important prairie hay grasses of Nebraska by their vegetative characters. Univ. Nebraska Agric. Exp. Stat. Res. Bull. 65: 1-40.

Petersen, N. F. 1923. Flora of Nebraska: ferns, conifers and flowering plants of the state, with keys for their identification. Third edition. Publ. by the author. Lincoln, NB. 220 pp.

Sutherland, D. M. 1975. A vegetative key to Nebraska grasses. <u>In</u>, Prairie, a multiple view. Univ. North Dakota Press. Grand Forks. Pp. 283-316.

Sutherland, D. M. 1984. Vegetative key to the grasses of the Sand Hills of Nebraska. Trans. Nebraska Acad. 12: 23-60.

Wilcox, E. M., G. K. K. Link, & V. W. Pool. 1915. A handbook of Nebraska grasses. Nebraska Agric. Exp. Stat. Bull. 148: 1-120.

NEVADA

Kartesz, J. 1987. A flora of Nevada. Ph. D. dissertation. Univ. Nevada, Reno. 3: 1556-1681.

Swallen, J. R. 1940. Gramineae of Nevada. Contr. Toward a Flora of Nevada. No. 1. 91 pp.

NEW HAMPSHIRE

Hodgdon, A. R., G. E. Crow, & F. L. Steele. 1979. Grasses of New Hampshire. I. Tribes Poeae (Festuceae) and Triticeae (Hordeae). Station Bulletin 512. New Hampshire Agric. Exp. Sta. Durham. 53 pp.

Pease, A. S. 1964. A flora of northern New Hampshire. New England Bot. Club. Cambridge, MA. 278 pp.

NEW JERSEY

Stone, W. 1973. The plants of southern New Jersey. Quarterman. Boston, MA.

NEW MEXICO

Allred, K. W. 1997. A field guide to the grasses of New Mexico. Second edition. Agric. Exp. Stat. New Mexico State Univ. Las Cruces. 259 pp.

Allred, K. W. 1999. Eponymy of New Mexico grass names. Desert Plants 15(1): 3-10.

Allred, K. W. 2003. A working index of New Mexico vascular plant names, interim version of 13 October 2003. http://web.nmsu.edu/~kallred/herbweb/

Allred, K. W., S. L. Hatch, & R. J. Soreng. 1986. Verified checklist of the grasses of New Mexico. New Mexico Agric. Exper. Stat. Res. Rept. 579. 47 pp.

Allred, K. W., S. L. Hatch, & R. J. Soreng. 1988. New Mexico grasses: a synopsis of the classification and a key to genera. New Mexico J. Sci. 28(1): 21-43.

Allred, K. W., S. L. Hatch, & R. J. Soreng. 1990. New Mexico grass types and a selected bibliography for New Mexico grasses. Great Basin Nat. 50(1): 73-82.

Barnard, C. M. & L. D. Potter. 1984. New Mexico grasses: a vegetative key. Univ. New Mexico Press. Albuquerque. 157 pp.

Ivey, R. D. 2003. Flowering plants of New Mexico. Fourth edition. Publ. by author. Albuquerque. 573 pp.

Martin, W. C. & C. R. Hutchins. 1980. A flora of New Mexico. J. Cramer. 1: 96-317.

NEW YORK

Mitchell, R. S. & G. C. Tucker.1997. Revised checklist of New York state plants. Contributions to a flora of New York state checklist IV. Bull. No. 490. New York State Mus. Pp. 304-335.

Smith, S. J. 1965. Checklist of the grasses of New York state. New York State Museum and Science Service Bull. No. 403. 44 pp.

NORTH CAROLINA

Blomquist, H. L. 1948. The grasses of North Carolina. Duke Univ. Press. Durham, NC. 276 pp.

Radford, A. E., H. E. Ahles, & C. R. Bell. 1968. Manual of the vascular flora of the Carolinas. Univ. North Carolina Press. Chapel Hill. Pp. 55-167.

NORTH DAKOTA

Stevens, O. A. 1963. Handbook of North Dakota plants. North Dakota Inst. for Regional Studies. Fargo. 324 pp.

OHIO

Braun, E. L. 1967. The vascular flora of Ohio. Vol. 1. The Monocotyledoneae. Ohio State Univ. Press. Columbus. 464 pp.

Cooperrider, T. S., A. Cusick, & J. Kartesz. 2001. Seventh catalog of vascular plants. Ohio State Univ. Press. Columbus. 256 pp.

Schaffner, J. H. 1917. The grasses of Ohio. Ohio Biol. Surv. Bull. 2(5): 255-329.

Weishaupt, C. G. 1971. Vascular plants of Ohio: a manual for use in field and laboratory. Third edition. Kendall/Hunt Publ. Dubuque, IA. 292 pp.

Weishaupt, C. G. 1985. Descriptive key to the grasses of Ohio based on vegetative characteristics. Ohio Biol. Survey. 99 pp.

OKLAHOMA

Estes, J. R. & R. J. Tyrl. 1976. Classification of Oklahoma grasses. Ann. Oklahoma Acad. Sci. 6: 141-161

Featherly, H. I. 1946. Manual of grasses of Oklahoma. Oklahoma State Univ. Bull. 43(21): 1-137.

Taylor, R. J. & C. E. S. Taylor. 1989. An annotated list of the ferns, fern allies, gymnosperms and flowering plants of Oklahoma. Biol. Dept. Herbarium. Southeastern Oklahoma State Univ. Durant. Pp. 15-24.

Waterfall, U. T. 1979. Keys to the flora of Oklahoma. Sixth edition. Published by the author. Student Union Bookstore. Oklahoma State University. Stillwater. Pp. 26-53.

OREGON

Peck, M. E. 1961. A manual of the higher plants of Oregon. Second edition. Oregon State Univ. Press. Corvallis. Pp. 77-132.

PENNSYLVANIA

Gress, E. M. 1924. The grasses of Pennsylvania. Bull. Penn. Dept. Agric. 384(7): 1-245.

Pohl, R. W. 1947. A taxonomic study on the grasses of Pennsylvania. Amer. Midl. Nat. 38(3): 513-604.

Rhoades, A. F. & T. A. Block. 2000. The plants of Penn-sylvania. Morris Arboretum and Univ. Pennsylvania Press. Philadelphia. Pp. 891-986.

RHODE ISLAND

Palmatier, E. A. 1952. Flora of Rhode Island. Univ. Rhode Island. Kingston. 75 pp.

SOUTH CAROLINA

Redford, A. E., H. E. Ahles, & C. R. Bell. 1968. Manual of the vascular flora of the Carolinas. Univ. North Carolina Press. Chapel Hill. Pp. 55-167.

SOUTH DAKOTA

Van Bruggen, T. n. d. A key for the identification of grasses of South Dakota. Mimeographed. Botany Department. Univ. South Dakota. 27 pp.

Van Bruggen, T. 1985. Vascular plants of South Dakota. Second edition. Iowa State University Press. Ames. 476 pp.

TENNESSEE

Chester, E. W. et al. 1993. Atlas of Tennessee vascular plants. Vol. 1. Pteridophytes, gymnosperms, angiosperms: monocots. Misc. Publ. No. 9. Center for Field Biology. Austin Peay State Univ. Clarksville, TN.

De Selm, H. R., B. E. Wofford, R. Kral, & E. W. Chester. 1994. An annotated list of grasses (Poaceae, Gramineae) of Tennessee. Castanea 59(4): 338-353.

Wofford, B. E. & R. Kral. 1993. Checklist of the vascular plants of Tennessee. Sida Bot. Misc. 10. 66 pp.

TEXAS

Correll, D. S. & M. C. Johnston. 1970. Gramineae. <u>In</u>, Manual of the vascular plants of Texas. Texas Research Foundation. Renner. Pp. 103-261.

Gould, F. W. 1975. The grasses of Texas. Texas A & M Univ. Press. College Station. 653 pp.

Gould, F. W. 1978. Common Texas grasses. Texas A & M Univ. Press. College Station. 267 pp.

Gould, F. W. & T. W. Box. 1965. Grasses of the Texas coastal bend. Texas A & M Univ. Press. College Station. 186 pp.

Hatch, S. L. & J. Pluhar. 1992. Texas range plants. Texas A & M Univ. Press. College Station. 344 pp.

Hatch, S. L., J. L. Schuster, & D. L. Drawe. 1999. Grasses of the Texas Gulf prairies and marshes. Texas A & M Univ. Press. College Station. 355 pp.

Highnight, K. W., J. K. Wipff, & S. L. Hatch. 1988. Grasses (Poaceae) of the Texas Cross Timbers and prairies. MP-1657. Texas Agric. Exper. Stat. College Station. 174 pp.

Johnston, M. C. 1990. The vascular plants of Texas. A list up-dating the "Manual of the vascular plants of Texas." Second edition. Publ. by the author. Austin, TX

Jones, S. D., J. K. Wipff, & P. M. Montgomery. 1997. Vascular plants of Texas. Univ. Texas Press. Austin. Pp. 235-264.

Lonard, R. I. 1988. Phytogeography of the grasses of the lower Rio Grande Valley, Texas. <u>In</u>, Davis, A. & G. Stanford (editors). The prairie: roots of our culture, foundation of our economy. Paper No. 01-07. Proc. Tenth North American Prairie Conf. Native Prairie Assoc. Texas. Dallas. 3 pp.

Lonard, R. I. 1993. Guide to grasses of the Lower Rio Grande Valley, Texas. Univ. Texas-Pan American Press. Edinburg. 240 pp.

Powell, A. M. 1994. Grasses of the Trans-Pecos and adjacent areas. Univ. Texas Press. Austin. 377 pp.

Shinners, L. H. 1954. Notes on North Texas grasses. Rhodora 56: 25-38.

Silveus, W. A. 1933. Texas grasses. Publ. by the author. San Antonio, TX. $782\ pp.$

Tharp, B. J. 1952. Texas range grasses. Univ. Texas Press. Austin. 125 pp.

Turner, B. L., H. Nichols, G. Denny, & O. Doron. 2003. Atlas of the vascular plants of Texas. Sida, Misc. No. 24. Two vols. 888 pp.

UTAH

Flowers, S. 1959. Common grasses of Utah. Third edition. Univ. Utah Press. Salt Lake City. 122 pp.

Harrison, B. F. 1939. An annotated list of Utah grasses. Proc. Utah Acad Sci. 16: 23-35.

Parker, K. G., L. R. Mason, & J. F. Valentine. 1979. Utah grasses. Extension Circular 384. Cooperative Extension Service. Utah State Univ. Provo. 69 pp.

Welsh, S. et al. 2003. A Utah flora. Third edition. Brigham Young Univ. Provo, UT. Pp. 716-805.

VERMONT

Seymour, F. C. 1969. The flora of Vermont. Fourth edition. Agric. Exp. Sta. Bull. 660. Univ. Vermont. Burlington. 393 pp.

Zika, P. F. 1990. Range extensions of some grasses in Vermont. Rhodora 92(870): 80-89.

VIRGINIA

Massey, A. B. 1961. Virginia flora. Virginia Agric. Exp. Sta. Tech. Bull. 152. Blacksburg. 258 pp.

WASHINGTON

Creso, I. 1984. Vascular plants of western Washington. Publ. by the author. Tacoma, WA. 532 pp.

St. John, H. 1963. Flora of southeastern Washington and adjacent Idaho. Third edition. Outdoor Pictures. Escon-dido, CA. 583 pp.

WEST VIRGINIA

Core, E. L., E. E. Berkley, & H. A. Davis. 1944. West Virginia grasses. West Virginia Agric. Exp. Stat. Bull. 313. 96 pp.

Strausbaugh, P. D. & E. L. Core. 1978. Flora of West Virginia. Second Edition. Seneca Books, Grantsville, WV. Pp. 66-151.

WISCONSIN

Fassett, N. C. 1951. Grasses of Wisconsin. Univ. Wisconsin Press. Madison. 173 pp.

Freckmann, R. W. 1972. Grasses of central Wisconsin. Reports on the Fauna and Flora of Wisconsin. Report No. 6 Mus. of Nat. Hist. Univ. Wisconsin. Stevens Point. 81 pp.

WYOMING

Dorn, R. D. 1977. Manual of the vascular plants of Wyoming. Garland Publ. Co. New York. 1: 692-797.

Dorn, R. D. 2001. Poaceae. \underline{In} , Vascular plants of Wyoming. Third edition. Mountain West Pub. Cheyenne, WY. Pp. 261-290.

Hallsten, G. P., Q. D. Skinner, & A. A. Beetle. 1987. Grasses of Wyoming. Third edition. Research J. 202. Agric. Exp. Stat. Univ. Wyoming. Laramie. 432 pp.

Porter, C. L. 1974. Poaceae (Gramineae). A flora of Wyoming. Part 3. Univ. Wyoming Agric. Exp. Stat. Bull. 418. 80 pp.

MEXICO, CENTRAL AMERICA & THE CARIBBEAN

Beetle, A. A. 1974. Noteworthy grasses from Mexico. I. Phytologia 27(6): 441-444.

Beetle, A. A. 1974. Noteworthy grasses from Mexico. II. Phytologia 28(4): 313-318.

Beetle, A. A. 1975. Noteworthy grasses from Mexico. III. Phytologia 30(5): 344-349.

Beetle, A. A. 1977. Noteworthy grasses from Mexico. IV. Phytologia 35(3): 221-223.

Beetle, A. A. 1977. Noteworthy grasses from Mexico. V. Phytologia 37(4): 317-407. [Checklist of 1070 spp.]

Beetle, A. A. 1977. Noteworthy grasses from Mexico.

VI. Phytologia 38(2): 173-176.

Beetle, A. A. 1981. Noteworthy grasses from Mexico. VII. Phytologia 47(5): 379-383.

Beetle, A. A. 1981. Noteworthy grasses from Mexico. VIII. Phytologia 48(2): 189-193.

Beetle, A. A. 1981. Noteworthy grasses from Mexico. IX. Phytologia 49(1): 33-43.

Beetle, A. A. 1982. Noteworthy grasses from Mexico. X. 52(1): 11-17.

Beetle, A. A. 1983. Noteworthy grasses from Mexico. XI. Phytologia 54(1): 1-5.

Beetle, A. A. 1983. Las gramineas de Mexico. Vol. I. Secretaria de Agricultura y Recursos Hidraulicos COTECOCA. Editorial Calypso. Mexico, D. F. Mexico. 260 pp.

Beetle, A. A. 1986. Noteworthy grasses from Mexico. XII. Phytologia 59(4): 287-289.

Beetle, A. A. 1987. Noteworthy grasses from Mexico. XIII. Phytologia 63(4): 209-297.

Beetle, A. A. 1987. Las gramineas de Mexico. Vol. II. Secretaria de Agricultura y Recursos Hidraulicos COTECOCA. Editorial Calypso. Mexico, D. F. Mexico. 344 pp.

Beetle, A. A. 1991. Las gramineas de Mexico. Vol. III. Secretaria de Agricultura y Recursos Hidraulicos COTECOCA. Editorial Calypso. Mexico, D. F. Mexico. 336 pp.

Beetle, A. A. & D. J. Gordon. 1991. Gramineas de Sonora. Sec. Agric. y Recursos Hidraulicos. Gobierno del Estado de Sonora. Hermosillo, Mexico.

Catasus Guerra, L. 1997. Las gramineas (Poaceae) de Cuba. I. Fontqueria 46(1/2): 1-259.

De La Cerda-Lemus, M. & Y. Herrara-Arrieta. 1995. New Poaceae distribution records for Aguascalientes, Mexico. Madrono 42(3): 396-405.

Gould, F. W. 1979. A key to the genera of Mexican grasses. Texas Agric. Exp. Stat. Misc. Publ. No. 1422. 46 pp.

Gould, F. W. 1979. Poaceae. <u>In</u>, Flora of the Lesser Antilles, Leeward and Windward Islands. Arnold Arbor. Harvard Univ. 3: 25-220.

Gould, F. W. & R. Moran. 1981. The grasses of Baja California, Mexico. Mem. 12. San Diego Soc. Nat. Hist. San Diego, CA. 140 pp.

Hitchcock, A. S. 1909. Catalogue of the grasses of Cuba. Contr. U. S. Natl. Herbarium 12: 183-258.

Hitchcock, A. S. 1913. Mexican grasses in the U. S. National Herbarium. Contr. U. S. Natl. Herb. 17: 181-389.

Hitchcock, A. S. 1930. The grasses of Central America. Contr. U. S. Natl. Mus. 24: 557-802.

Hitchcock, A. S. 1936. Manual of the grasses of the West Indies. Misc. Publ. No. 243. U. S. Dept. Agric. Washington, D. C. 439 pp.

- Judziewicz, E. J. & R. W. Pohl. 1984. Grasses of La Selva, Costa Rica. Contr. Univ. Wisconsin Herb. 1(3): 1-86.
- Lenz, L. W. 1992. Poaceae. <u>In</u>, An annotated catalogue of the plants of the Cape Region, Baja California Sur, Mexico. Cape Press. Claremont, CA. Pp. 12-21.
- Matuda, E. 1958. Las gramineas del Estado de Mexico. Toluca. 83 pp.
- McVaugh, R. 1983. Flora Novo-Galiciana. A descriptive account of the vascular plants of western Mexico. Vol. 14. Gramineae. Univ. Michigan Press. Ann Arbor. 436 pp.
- Mejia Saules, M. T. & P. D. Davila Aranda. 1992. Gramineas utiles de Mexico. Cuadernos del Inst. Biologia. 298 pp.
- Pohl, R. W. 1972. Keys to the genera of grasses of Costa Rica. Rev. Biol. Trop. 20: 189-219.
- Pohl, R. W. 1976. The genera of native bamboos of Costa Rica. Rev. Biol. Trop. 24(2): 243-249.
- Pohl, R. W. 1980. Gramineae. <u>In</u>, Burger, W. (editor). Flora Costaricensis. Fieldiana Botany, N. S. No. 4. Publ. No. 1313. Field Mus. Nat. Hist. Chicago, IL. 608 pp.
- Pohl, R. W. 1982. Additions and notes on the grass flora of Costa Rica. Brenesia 19/20: 617, 618.
- Reeder, J. R. 1969. Las gramineas dioicas de Mexico. Bol. Soc. Bot. Mexico. 30: 121-126.
- Renvoize, S. A. 1984. The grasses of Bahia. Royal Bot. Gard. Kew, England. 301 pp.
- Spellman, D. L., J. D. Dwyer, & G. Davidse. 1975. Gramineae. <u>In</u>, A list of the Monocotyledoneae of Belize including an historical introduction to plant collecting in Belize. Rhodora 77: 124-132.
- Swallen, J. R. 1934. Grasses of the Yucutan Peninsula. Carnegie Inst. Publ. 436: 325-355.
- Swallen, J. R. 1938. Additions to the grass flora of British Honduras. J. Washington Acad. Sci. 28: 6-11.
- Swallen, J. R. 1943. Gramineae. In, Flora of Panama. Pt. II, Fascicle I. Annals of the $\overline{\text{M}}$ issouri Bot. Gard. 30(2): 104-280.
- Swallen, J. R. 1955. Flora of Guatemala. Part II. Grasses of Guatemala. Fieldiana: Botany 24(2): 1-390
- Swallen, J. R. & E. Hernandez X. 1961. Clave de los generos Mexicanos de gramineas. Bol. Soc. Bot. Mexico. 26: 52-118.
- Valdez Reyna, J. 1977. Gramineas de Coahuila. Univ. Autonomo Agraria Aontonio Narro. Monographia Tecnico Cientifica 3(11): 884-1018.
- Valdez Reyna, J. 1977. Grasses of Chihuahua, Mexico. Thesis. Range Management Section. Plant Science Division. Univ. Wyoming. Laramie. 257 pp.

SOUTH AMERICA

- Cabrera, A. L. 1970. Gramíneas. Flora de la Provincia de Buenos Aires. Inst. Nac. Tecn. Agropec. Buenos Aires. 4(2): 1-624.
- Caro, J. A. 1982. Sinopsis taxonomica de las gramineas Argentinas. Domingguezia 4: 1-51.
- Febres, Z. L. de. 1963. Las gramineas del Distrito Federal. Inst. Bot. Direccion de Recursos Naturales Renovables. Ministerio de Agric. y Cria. Caracas, Venezuela.
- Hitchcock, A. S. 1922. Grasses of British Guiana. Contr. U. S. Natl. Herb. 22: 439-515.
- Hitchcock, A. S. 1927. Grasses of Ecuador, Peru, and Bolivia. Contr. U. S. Natl. Herb. 24: 291-556.
- Judziewicz, E. 1990. Poaceae. Flora of the Guianas. Series A. Fascicle 8. 725 pp.
- Killeen, T. J. 1990. The grasses of Chiquitanaia, Santa Cruz, Bolivia. Ann. Missouri Bot. Gard. 77(1): 125-201.
- Laegaard, S. & T. Delgado. 1999. Claves para tribus y generos de gramineas ecuatorianas. Herbario LOJA 4: 1-45.
- Macbride, J. F. 1936. Gramineae. Flora of Peru. Field Mus. Nat. Hist. Bot. Series 13: 96-261.
- Nicora, E. G., Z. E. Rugolo de Agrasar. 1987. Los generos de Gramineas de America Austral: Argentina, Chile, Uruguay y areas limitrofes de Bolivia, Paraguay y Brasil. Hemisferio Sur. Buenos Aires, Argentina. 611 pp.
- Parodi, L. R. 1918. Las chlorideas de la Republica de Argentina. Rev. Fac. Agron. y Vet. 2: 233-335.
- Parodi, L. R. 1922. Alguna gramineas mal conocidas o nuevas para la flora Argentina. Physis 6: 101-104.
- Parodi, L. R. 1922. Las gramineas de la region de Concordia (Argentina). Rev. Fac. Agron. y Vet. Univ. Nac. Buenos Aires. 4: 24-101.
- Parodi, L. R. 1928. Notas sobre Gramineas de la flora Argentina. Physis 9: 12-45.
- Parodi, L. R. 1935. Notas sobre gramineas Argentinas. Physis 11: 497-500.
- Parodi, L. R. 1937. Gramineas Argentinas neuvas o criticas. Not. Mus. La Plata 2: 1-16.
- Parodi, L. R. 1938. Gramineas Austroamericanas. Not. Mus. Univ. Nac. La Plata 3: 15-33.
- Parodi, L. R. 1943. Gramineas Austroamericanas. Part II. Not. Mus. Univ. Nac. La Plata 8: 75-100.
- Parodi, L. R. 1948. Gramineas Argentinas neuvas o criticas. Part II. Rev. Argentine Agron. 15: 53-61.
- Rambo S. J., B. 1984. Gramineae Riograndenses. Pesquisas Botanica No. 36. 191 pp.
- Renvoize, S. A. 1998. Gramíneas de Bolivia. Royal

Botanic Gardens. Kew, England. 646 pp.

Rosengurtt, B. et al. 1960. Caracteres vegetivos y forrojeros de 175 gramineas del Uruguay. Rev. Fac. Agron. Montevideo 47: 1-168.

Rosengurtt, B., B. R. Arrillaga de Maffei, & P. Izaguire de Artuco. 1970. Gramíneas Uruguyas. Univ. de la Rep. Montevideo. 489 pp.

Smith, . B. & D. C. Wasshausen. 1977. Genera of grasses of Brazil. Bradea 2(32): 221-228.

Standley, P. C. 1936. Gramineae. <u>In</u>, Macbride, J. F. Flora of Peru. Pt. I. Field Mus. Nat. Hist. Botanical Series. 13: 96-261.

Tovar, O. 1993. Las Gramineas (Poaceae) del Peru. Ruizia 13: 1-480.

Turpe, A. M. 1975. Los generos de gramineas de las Provincia de Tucuman. Op. Lilloana No. 24. 203 pp.

Zuloaga, F. O. et al. 1994. Catalogo de la familia Poaceae en la Republica Argentina. Monographs in Syst. Bot. No. 47. Missouri Bot. Gard. St. Louis, MO. 178 pp.

Zuloaga, F. O. et al. 1994. Flora del Paraguay. Gramineae. V. Panicoideae, Paniceae, Acroceras Stapf - Panicum L. Conservatoire et Jardin Botaniques de la Valle de Geneve and Missouri Botanical Garden. 327 pp.

GREAT BRITAIN & EUROPE

Dumortier, B. C. J. 1824. Observations sur les graminees de la flore belgique. J. Castereman. Turnay. 153 pp. + 16 plates.

Hubbard, C. E. 1984. Grasses: a guide to their structure, identification, uses, and distribution in the British Isles. Third edition. Revised by J. C. E. Hubbard. Penguin Books. Harmondsworth, England. 476 pp.

Kerguelen, M. 1975. Les Gramineae (Poaceae) de la flora Française. Essai de mise au point taxonomique et nomenclaturale. Lejuenia N. S. 75: 1-343.

Parnell, R. MDCCCXLII. The grasses of Scotland. Blackwood & Sons. Edinburgh, Scotland. 152 pp. + LXVI plates.

Pratt, A. 1858(?). The British grasses and sedges. Soc. Promoting Christian Knowledge. London, England. 136 pp. + plates.

Rose, F. 1989. Colour identification guide to the grasses, sedges, rushes and ferns of the British Isles and northwestern Europe. Viking. London, England. 239 pp.

Ryves, T. B., E. J. Clement, & M. C. Foster. 1996. Alien grasses of the British Isles. Bot. Soc. British Isles. London, England. 181 pp.

Tutin, T. G. (Editor). 1980. Gramineae. <u>In</u>, Tutin, T. G. et al. Flora Europaea. Cambridge Univ. Press. Cambridge, England. 5: 118-267.

AFRICA

Bennett, K. ER. 1980. Keys to Zimbabwean grass species. Kirkia 11(2): 169-286.

Bogdan, A. V. 1958. A revised list of Kenyan grasses. Government Printer. Nairobi, Kenya.

Chippindall, L. K. A. 1955. A guide to the identification of grasses in South Africa. <u>In</u>, Meredith, D. (editor). The grasses and pastures of South Africa. Central News Agency.

Chippindall, L. K. A. & A. O. Crook. 1976-1978. 240 grasses of southern Africa. M. O. Collins. Salisbury, South Africa.

Cope, T. A. 1985. Key to Somali grasses. Natl. Herbarium. Mogadishu, Somalia. 77 pp.

Cope, T. A. & H. A. Hosni. 1992. A key to Egyptian grasses. Royal Botanic Gardens. Kew, England. 75 pp.

Costa, E., T. Martins, & F. Monteiro. 2004. A checklist of Angola grasses/checklist das Poaceae de Angola. SABONET Report Series No. 28. 24 pp.

Froman, B. & S. Persoon. 1974. An illustrated guide to the grasses of Ethiopia. Chilalo Agric. Dec. Unit. Asella, Ethiopia. 504 pp.

Gibbs Russell, G. E. et al. 1990. Grasses of southern Africa. Bot. Surv. S. Africa No. 58. Bot. Inst. Pretoria, South Africa. 437 pp.

Harker, K. W. & D. Napper. 1961. An illustrated guide to the grasses of Uganda. Government Printing Office. Entebbe, Uganda. 63 pp.

Ibrahim, K. M. & C. H. S. Kabuye. 1987. An illustrated manual of Kenya grasses. Food Agric. Admin. United Nations. Rome, Italy. 765 pp.

Innes, R. R. 1977. A manual of Ghana grasses. Land Resources Div. Min. Overseas Dev. Surbiton, England. 265 pp.

Kabelo, M. & D. Mafokate. 2004. A checklist of Botswana grasses. SABONET Report Series No. 24. 16 pp.

Koechlin, J. 1962. Flore du Gabon: Graminées. Paris, France. 292 pp.

Launert, E. 1971. Gramineae. Flora Zambesica 10(1): 1-152.

Lightfoot, C. 1975. Common veld grasses of Rhodesia. Second edition. Nat. Res. Board Rhodesia. Gov. Printer. Salisbury, Rhodesia. 131 pp.

Lin, W.-C. 1967. The species and distribution of bamboos in the Republic of Malagasy (Madagascar), East Africa. Taiwan For. Res. Inst. Spec. Bull. No. 4: 1-34.

Loew, J. 1989. The flora of Nigeria: grasses. Second edition. Ibadan Univ. Press. Ibadan, Nigeria. 326 pp.

Napper, D. 1965. Grasses of Tanganyika. Bull. Bo. 18. Min. Agric. Forests and Wildlife. Gov. Printer. Dares Salaam. 146 pp.

Oudtshoorn, F. van. 1999. Guide to grasses of South Africa. Briza Publ. Arcadia, S. Africa. 302 pp.

Oudshoorn, F. van. 1999. Guide to the grasses of southern Africa. Briza Publ. Arcadia, S. Africa. 288 pp.

Scholz, H. 1974. Liste der Gräser Libyens. Willdenowia 7(2): 419-458.

Sherif, A. S. & M. A. Siddiqi. 1988. Flora of Libya: 145. Poaceae. National Herbarium. Al Faateh Univ. 356 pp.

Simon, B. K. 1971. Rhodesian and Zambian grass lists. Kirkia 8(1): 3-83.

Stanfield, D. P. 1970. Grasses. In, Stanfield, D. P. & J. Lowe (Editors). The flora of Nigeria. Ibadan Univ. Press. Ibadan, Nigeria. 118 pp.

Stapf, O. 1897. Gramineae. In, Thiselton-Dyer, W. T. Flora Capensis. 7: 310-750; 760-765.

Stapf, O. & C. E. Hubbard. 1934. Gramineae. \underline{In} , Thiselton-Dyer, W. T. Flora of tropical Africa. Vol. 9. \overline{L} . Reeve. London, England.

Tadros, T. M. 1971. Atlas of the common grasses of Tanzania. Pt. 1. Publ. No. 1. Botany Dept. Herbarium. Univ. Dar es Salaam. 64 pp.

Tadros, T. M. 1973. Atlas of the common grasses of Tanzania. Pt. 2. Publ. No. 1. Botany Dept. Herbarium. Univ. Dar es Salaam. 54 pp.

van Oudtshoorn, F. 1992. Guide to grasses of South Africa. Briza Publ. Arcadia, South Africa. 301 pp.

Zoon, A. P., M. van der. 1993. Graminees du Cameroun. Wageningen Afric. Univ. Wageningen, The Netherlands. Two vols. 650 pp.

ASIA

Ahmad, S. & R. R. Stewart. 1958. Grasses of West Pakistan. Pt. 1. Subfamily Panicoideae. Biol. Soc. Pakistan Monograph No. 3. 151 pp.

Bor, N. L. 1938. A list of the grasses of Assam. Indian Forest Records, N. S. Botany 1(3): 47-102.

Bor, N. L. 1940. Gramineae. <u>In</u>, The flora of Assam. Prabasi Press. Calcutta.

Bor, N. L. 1960. The grasses of Burma, Ceylon, India, and Pakistan (excluding Bambuseae). Pergamon Press. New York, NY. 767 pp.

Bor, N. L. 1970. Gramineae. <u>In</u>, Rechinger, K. H. Flora Iranica. Akademische Druck und Verlag Sanstalt. Graz, Austria. 573 pp. + 72 plates.

Christopher, J. 1978. Studies of the cytology and phylogeny of South Indian grasses. IV. Subfamily Panicoideae, tribes Andropogoneae and Maydeae. Cytologia 43(2): 273-288.

Cope, T. A. 1985. A key to the grasses of the Arabian Peninsula. Studies in the flora of Arabia. Pt. 15. Arab Bur. Educ. Gulf States. Riyadh, Saudia Arabia. 82 pp.

Dabadghao, P. M. & K. A. Shankarnarayan. 1973. The grass cover of India. Indian Council of Agric. Res. New

Delhi, India. 713 pp.

Dransfield, S. & E. A. Widjaja (Editors). 1995. Plant resources of South-East Asia No. 7. Bamboos. Backhuys Publ. Leiden, The Netherlands. 189 pp.

Edgar, E. & E. S. Gibb. 1996. Checklist of the pooid grasses naturalized in New Zealand: 4. Tribe Poeae. New Zealand J. Bot. 34(2): 147-152.

Gamble, J. S. 1896. The Bambuseae of British India. Ann. Royal Bot. Gard. (Calcutta) 7(1): 1-133.

Gilliland, H. B. et al. 1971. A revised flora of Malaya. Vol. III. Grasses of Malaya. Government Printing Office. Singapore. 319 pp. + 36 plates.

Grubov, V. I. (editor). 2001. Plants of Central Asia. Vol. 4: Gramineae (grasses). 315 pp.

Hodd, T. & P. Hodd. 1982. Grasses of western India. Bombay Nat. Hist. Soc. 167 pp.

Hsu, C.-C. 1971. A guide to the Taiwan grasses with keys to subfamilies, tribes, genera and species. Taiwania 16(2): 199-341.

Hsu, C.-C. 1975. Taiwan grasses. Taiwan Provincial Educ. Assoc. Taipei. 884 pp.

Keng, Y. L. 1933. The grasses of China. Ph. D. dissertation. George Washington Univ. 424 pp.

Koyama, T. 1987. Grasses of Japan and its neighboring regions: an identification manual. Kodansha. Tokyo, Japan. 570 pp.

Lazarides, M. The tropical grasses of Southeast Asia. Lubrecht & Cramer. Monticello, New York. 350 pp.

Lee, Y. N. 1966. Manual of Korean grasses. Ewha Womans Univ. Press. Seoul, Korea. 300 pp.

Lin, W.-C. 1961. Study of the classification of Bambusaceae in Taiwan. Bull. Taiwan For. Res. Inst. 69: 1-144.

Lin, W.-C. 1968. The bamboos of Thailand (Siam). Taiwan For. Res. Inst. Spec. Bull. No. 6: 1-52.

Moulik, S. 1997. The grasses and bamboos of India. Two vols. Scientific Publ. Jodhpur, India. 700 pp.

Mouroi, H. 1956. The Japanese bamboos. Inoue Book Co. Tokyo. 340 pp.

Nakai, T. 1933. Bambusaceae in Japan proper (II). J. Japanese Bot. 9(2): 77-95.

Ohwi, J. 1971. Contributions to the flora of south-east Asia: 5. Gramineae and Cyperaceae of Thailand. Tonan Ajia Kenkyu 9(2): 194-219.

Reveal, J. L. 2001. Grasses of Pakistan. Spring Canyon Enterprises. Montrose, CO. 140 pp.

Senaratna, J. E. 1956. The grasses of Ceylon. Government Press. Columbo, Ceylon. 229 pp. + 50 plates.

Shukla, U. 1996. The grasses of north-eastern India. Scientific Publ. Jodphur, India. 404 pp.

Stapleton, C. M. A. 1994. The bamboos of Nepal and

Bhutan. Part I. Edinburgh J. Bot. 51(1): 1-.

Tolmachev, A. I. & J. G. Packer. 1996. Gramineae. Flora of the Russian Arctic. Vol. 1. Polypodiaceae - Gramineae. Univ. Alberta Press. Edmonton, Canada. Pp. 87-306.

Tsvelev, N. N. 1968. Sistema zlakov (Poaceae) Fory SSSR. Bot. Zhur. 53(3): 301-312.

Tsvelev, N. N. 1976. Zlaki SSSR. Akademiya Nauk SSSR Botanicheskii Institut Imeni V. L. Komarova. Leningrad. 788 pp.

Tzvelev, N. N. 2002. Plants of Central Asia (plant collections from China and Mongolia). Vol. 4. Gramineae. Science Publ. Enfield, NH. 328 pp.

Tzvelev, N. N. (editor). 2003. Vascular plants of the Russian Far East. Vol. 1. Lycopodiophyta, Juncaceae, Poaceae (Gramineae). Science Publ. Enfield, NH. 544 pp.

Wong, K. M. 1995. The bamboos of peninsular Malaysia. Malayan Forest Records No. 41. Forest Res. Inst. Malaysia. Sabah, Malaysia. 200 pp.

AUSTRALIA & OCEANIA

Burbidge, N. T. 1966. Australian grasses. Vol. 1. Australian Capital Territory and southern tablelands of New South Wales. Angus & Robertson. Sydney, Australia. 158 pp.

Burbidge, N. T. 1968. Australian grasses. Vol. 2. Northern tablelands of New South Wales. Angus & Robertson. Sydney, Australia. 167 pp.

Burbidge, N. T. 1970. Australian grasses. Vol. 3. The east coast from southeast Queensland to Victoria. Angus & Robertson. Sydney, Australia. 219 pp.

Clifford, H. T. & B. K. Simon. 1981. The biogeography of Australian grasses. <u>In</u>, Keast, A. (Editor). Ecological biogeography of Australia. Second edition. W. Junk. The Hague. Pp. 538-554.

Dobson, A. T. 1975. A guide to the New Zealand grass genera. Mauri ORA 3: 111-123.

Edgar, E., H. E. Connor, & J. E. Shand. 1991. Checklist of oryzoid, arundinoid, and chloridoid grasses naturalized in New Zealand. New Zealand J. Bot. 29: 117-129.

Edgar, E. & E. S. Gibb. 1996. Checklist of the pooid grasses naturalized in New Zealand: 4. Tribe Poeae. New Zealand J. Bot. 34(2): 147-152.

Fosberg, F. R. 1939. Notes on Polynesian grasses. Occas. Pap. Bernice P. Bishop Mus. 15(3): 37-48.

Fosberg, F. R. & M.-H. Sachet. 1982. Micronesian Poaceae: critical and distributional notes. Micronescia 18(2): 45-102.

Gardner, C. A. 1952. Flora of western Australia. Vol. 1, Pt. 1. Gramineae. Government Press. Perth, Australia.

Holttum, R. E. 1967. The bamboos of New Guinea. Kew Bull. 21(2): 263-292.

Lampe, C. A. et al. 1990. Grasses of Australia: a field guide. Inkata Press. Melbourne, Australia. 310 pp.

Lazarides, M. 1970. The grasses of central Australia. Australian Natl. Univ. Press. Canberra. 282 pp.

Mitchell, M. 2002. Native grasses. Identification handbook for temperate Australia. Third edition. Landlink Press. Collingwood. 43 pp.

Morris, D. I. 1991. The grasses of Tasmania. Tasmanian Herbarium. Hobart. 167 pp.

Parham, J. W. 1955. The grasses of Fiji. Bull. No. 30. Dept. Agric. Gov. Press. Suva, Fiji. 166 pp.

Reeder, J. R. & C. G. Reeder. 1971. Gramineae. Flora of the Galápagos Islands. Stanford Univ. Press. Stanford, CA. Pp. 823-892.

Ripperton, J. C. et al. 1933. Range grasses of Hawaii. Hawaii Agric. Exp. Stat. Bull. 65: 1-58.

Romanowski, N. 1993. Grasses, bamboos, and related plants in Australia. 166 pp.

Sharp, D. & B. K. Simon. 2002. AusGrass: grasses of Australia. CSIRO Publ. Collingwood. CD-ROM + 38 pp. booklet.

Simon, B. K. 1980. A check-list of Queensland grasses. Queensland Dept. Primary Industries. Brisbane, Australia. 89 pp.

Simon, B. K. 1989. Studies in Australian grasses: 4. Taxonomic and nomenclatural studies in Australian Andropogoneae. Austrobaileya 3(1): 79-99.

Simon, B. K. 1993. A key to Australian grasses. Second edition. Queensland Dept. Primary Industries. Brisbane, Australia. 206 pp.

Simon, B. K. & P. Latz. 1994. A key to the grasses of the Northern Territory, Australia. N. Territory Bot. Bull. No. 17. 71 pp.

Sykes, W. R. 1996. Checklist of bamboos (Poaceae) naturalized in New Zealand. New Zealand J. Bot. 34(2): 153-156.

Tothill, J. C. & J. B. Hacker. 1979. Grasses of southeast Queensland. Univ. Queensland Press. Brisbane.

Tothill, J. C. & J. B. Hacker. 1983. The grasses of southern Queensland. Trop. Grassland Soc. Australia. Univ. Queensland Press. St. Lucia, Australia. 475 pp.

Vickery, J. W. 1975. Gramineae. Flora NSW 19(Suppl. Pt. 1, Pt. 2): 125-306.

Watson, L. & H. T. Clifford. 1976. The major groups of Australasian grasses: a guide to sampling. Australian J. Bot. 24(4): 489-507.

Watson, L. & M. J. Dallwitz. 1980. Australian grass genera: anatomy, morphology, and keys. Australian Natl. Univ. Research School Biol. Sci. Canberra. 209 pp.

Wheeler, D. J. B., S. W. L. Jacobs, & B. E. Norton.1982. Grasses of New South Wales. Monograph No. 3. Univ. New England. Armidale, Australia. 295 pp.

Whitney, L. D. et al. 1939. Grasses of the Hawaiian ranges. Hawaii Agric. Exp. Stat. Bull. 82: 1-148.

Wilson, A. J. G. (editor). 1994. Poaceae. <u>In</u>, Flora of Australia. 49: 442-499.

Zotov, V. D. 1963. Synopsis of the grass subfamily Arundinoideae in New Zealand. New Zealand J. Bot. 1:

78-136.

Zotov, V. D. 1965. Grasses of the subantarctic islands of the New Zealand region. Rec. Dom. Mus. (Wellington) 5(15): 101-146.

5.03 - DICHOTOMOUS KEYS

INTRODUCTION

A key is a logical device that assists you in the identification -- not the classification -- of an unknown organism. The keys used in botany usually take the form of a series of paired statements that describe various aspects of the plants treated therein. Each statement is called a **lead**; the two leads together constitute a **couplet**. The leads of a couplet are written in such a way that they are contrasting or contradictory propositions, as in:

1. Florets 1 Agrostis 1. Florets 2 or more . . . Festuca

You must decide in each case which of the two statements best describes the unknown plant. These decisions then determine a somewhat serpentine pathway through a series of subsequent couplets. Because you are presented with a series of paired statements, the key is called a **dichotomous key** (Greek, in two + to cut).

TYPES OF KEYS

Dichotomous keys may be simple or complex, long or short, utilize a wide combination of floral and vegetative features or be restricted to statements relating to a particular aspect of the plants, such as leaf features. A key is termed **natural** if it is constructed in such a way that it shows evolutionary affinities or **artificial** if there is no concern shown for relationships. Most modern keys are artificial ones.

Two types of keys are recognized on the basis of their mechanical structure. In the **bracket key** the leads of a couplet are always together. Each statement ends in the name of a taxon or in the number or letter of the next couplet that is to be read, as in the example below. The "Key to the U. S. Species of *Spartina*" in this syllabus is an example of a bracket key.

In the **indented key** or **yoked key** a subordinate lead is indented beneath the statement preceding it. Each type of key has its advantages. The indented key is easier to follow and allows the user to more easily grasp the subgroupings and logic of the key. The bracket key is preferred by editors and publishers because it is easier to set up and requires less space. The keys in Hitchcock & Chase are examples of indented keys.

WRITING A DICHOTOMOUS KEY

I suggest the following steps be done when you are attempting to write a dichotomous key.

- Carefully define the taxa to be included. If nothing else, make a checklist of the plants that you must consider.
- ☼ Give the key a descriptive title, such as "A Key to the Grasses of Humboldt County, California". This will immediately alert the potential user of the appropriateness of the key.
- Examine the plants to be treated. Determine a series of "key characters" that will discriminate among the taxa. Remember that you are primarily interested in how the plants are different from one another. These traits should be stable, easily observed features that are usually present through much of the year. Most keys employ macroscopic features or those that are visible under low magnification. Try to minimize the use of ephemeral traits.
- Prepare a comparison chart, putting the taxa to be keyed along one axis and the characters along the other. A particular feature that looked promising may not be that useful once this chart is completed. During the writing of the key statements, you may have to return to this comparison chart and add additional columns of features before all taxa can be distinguished.
- Examine the comparison chart to discover a trait that will divide the plants into two groups of about the same size. There may be several possible characters that you can use. With experience you will learn to weigh the relative merits of characters. Once this feature is chosen, it is defined in terms to two opposing character states. This will be the first couplet of the key. This process of selecting features continues until all of the taxa have been incorporated into the key.
- It is critical that the key be strictly dichotomous. For each statement there can be only one alternative lead.
- Select the key format (indented or bracket) that you wish to use. Do not attempt to hybridize these two structures. The internal logic of the key must be maintained consistently through the entire series of statements.
- Select key characters that are in opposition to one another and that are mutually exclusive. The

- unknown plant should fit under one, and only one, of the contradictory propositions.
- Use parallel construction within a couplet. If the first lead describes leaf shape, then so must the other lead of the pair.
- The initial word of each lead should be the name of the plant part being described. This is followed by the adjective, as in "Plants annual" or "Leaves hairy," rather than "Annual plants" or "Leaves hairy."
- Omit verbs. They are not necessary. Although it will look a bit awkward at first, you will quickly get use to it.
- The statements in a key may be numbered or lettered to make it easier to find the other member of a couplet. This is seldom a problem in shorter keys, but can become a frustration in longer keys that run to several pages. Once you have used a number or letter for a particular couplet, do not reuse it.
- Whenever possible, phrase the leads in a positive form. Avoid the use of "not as above" as the contrasting statement of a couplet. It provides no information about the plants falling on that side of the dichotomy.
- Avoid ill-defined characters. What is meant by the terms "tall," broad," and "rough?" Use precise botanical terminology.
- It is often useful to include measurements. Avoid overlapping limits in variation, as in "Leaves 3-7 cm long" vs. "Leaves 5-12 cm long". Under which lead is a plant with a leaf 7 cm long?
- The simplest keys usually contrast one feature in a statement. Better keys employ two or more features, separated by semicolons, as in:
 - 1. Inflorescence a panicle; glumes awned
 - 1. Inflorescence a spike; glumes awnless
- Sometimes it may be necessary to bring a plant out in more than one place in a key if it has two or more character states for a particular trait.

SOME HINTS IN USING KEYS

- Make sure that you are using the right key. It is both a waste of time and an embarrassment to spend several hours attempting to identify a plant in the wrong key.
- Read the key very carefully. If it is a good key, the author has spent a great deal of time selecting the right words. Much frustration results from misreading. There is a world of difference in the meaning of "and" and "or." They are not interchangeable.
- Watch for weasel words, such as "mostly" and "usually." Most of us who write keys use these words as avenues of escape when we do not want to be pinned down.
- Read both leads of the couplet before making your decision. The first lead may sound pretty good, but

- the second half may be perfect.
- Do not base your decision on a single observation, particularly when you are being asked about measurements.
- If neither lead seems to make any sense at all, you have probably made an error in keying and should not be in that section. Go back a step or two and check yourself.
- Check to see if there is a glossary at the back of the key. Authors vary in their use of certain terms and you will have to get use to the eccentricities of the writer.
- Do not assume that a key says something that it does not. In the second lead below, the author has not said that the leaves are alternate.
 - 1. Plant annual; leaves opposite
 - 1. Plants perennial
- If you are not confident about which lead to take, try both of them. One pathway should get you into difficulty fairly quickly.
- If one side of a dichotomy will take you to a relatively small number of plants, check out their descriptions or look at drawings. This additional knowledge may be helpful to you. Also, as you gain more knowledge of the flora you will be able to eliminate certain leads because they will take you to plants that you know.
- Learn to weigh the relative values of characters used in keys. Features of flower and fruit tend to be more important than those of plant height, for instance.
- When you have arrived at a determination, you should check it against a technical description, illustration, or specimen.

FEATURES USED IN GRASS KEYS

Growth Form (Habit)

Annual
Herbaceous perennial
Woody perennial
Tufted
Caespitose
Rhizomatous
Stoloniferous
Bulbous

Culms

Round or flattened Solid or hollow Erect or decumbent

Leaves

Blades Flat Rolled Bow-shaped tips Nature of margins

Sheaths Rounded Flattened Auricles Present Absent

Inflorescence

Terminal or axillary Open or contracted Included or exserted Balanced or 1-sided Involucre present or absent Type Panicle Raceme Spike Digitate Solitary

Intact at maturity (continuous) Shattering at maturity (discontinuous)

Spikelets Similar or dissimilar Number per node 1, 2, 3, etc. Fascicles Position relative to rachis Inserted edgewise Inserted flatwise Sunken in corky or fleshy rachis Covered by involucre Subtended by spines/bristles Compression Lateral Dorsal Terete Disarticulation Above glumes Below glumes Falling singly Falling in groups

Falling with rachis segment

Glumes

Number (2, 1, 0) United or separate Size relative to one another Size relative to lemma(s) Glabrous or variously hairy Nerve number Nature of apex Awned or awnless

Floret

Number Sexuality Perfect Pistillate Staminate Degree of reduction All ± similar Upper reduced Lower reduced Upper and lower reduced

Lemma

Size relative to glumes Size relative to palea Texture Membranous Papery Indurate Nerve number Nerves converging or parallel Glabrous or variously hairy Nature of apex Acute Rounded Bifid Truncate Sterile or fertile Awned or awnless Awn attachment Nature of callus Glabrous Bearded Cobwebby

Palea

Present or absent Texture Size relative to lemma Nerve number Awn absent or present Wings present or absent

Rachilla

Extension Beyond uppermost floret Not extended Rudiments Ending in rudiment Not ending in rudiment Glabrous or variously hairy

SECTION 6 - SELECTED TOPICS

6.01 - SCIENTIFIC & COMMON NAMES OF GRASSES

Grasses often have two names. One is a common name used by most of us in everyday circumstances when we make reference to a weed growing in the yard, an ornamental, or to one of the widely grown cereal crops. Grasses also have scientific names (or Latin names, as they are sometimes called) used by scientists and by the "serious" amateur. In this course, you and I will be using both common names and scientific names. You will need to be able to communicate with more than one audience.

COMMON NAMES

It would be foolish for me to maintain that common names have no value. They are the only names known to most of us. These names are often simple, easy to remember, descriptive, colorful, pleasing to the ear, and easy to pronounce. Given this impressive list of advantages, why do we not simply use common names for grasses and be done with it? There are several reasons why scientists do not use them.

- A grass may have more than one common name. Stipa hymenoides is commonly called Indian ricegrass, Indian millet, silk grass, and sand bunch grass.
- The same common name may be used for more than one plant. We all know corn when we see it. You may be surprised to learn that in other English-speaking countries, their corn is what we call wheat.
- Many common names are confusing and misleading. Kentucky bluegrass is not blue, nor is it native to Kentucky. Broomcorn is not a kind of corn, but a variety of sorghum. The heavenlybamboo is not a kind of bamboo, but a member of the barberry family.
- Because there are no universally accepted rules for giving common names to grasses, we cannot say that a particular one is **the** correct common name.
- Common names do not provide an indication of close relationship among the plants that share the name. Sour-grass, arrow-grass, blue-eyed grass, grass (marijuana), and China-grass are not kinds of grasses, nor are they are related to one another.
- Probably the most serious difficulty is that most grasses do not have common names. We have used only a small portion of the half million or so kinds of plants to the extent that common names have been applied to them. This is a problem for authors of field guides, for consultants who write environmental impact statements, and for staff members in various state and federal agencies who must prepare material for general

consumption. Authors have attempted to compensate for this lack of common names by inventing them, usually by translating the scientific name into English. The advantage of Orcutt's brome over *Bromus orcuttianus* or the spicate trisetum over *Trisetum spicatum* is not immediately apparent to me.

A word or two about the spelling of the common names of grasses. You will notice inconsistencies from one text to another. For instance, *Stipa comata* is variously called needle and thread, needle and thread grass, needle-and-thread grass, and needleandthread grass. The last spelling seems terribly awkward. Some floras list needlegrass; others, needle grass. Some authors capitalize common names (Giant Needle Grass); others do not (giant needle grass). *Sorghum halepense* is Johnson grass, but *Tuctoria greenei* is Greene's tuctoria. Apostrophes come and go.

SCIENTIFIC NAMES

Although scientific names may cause you some discomfort, their advantages to the botanist are compelling.

- ☼ There is a single, universally recognized name for each plant. Because they are used by botanists all over the world, scientific names facilitate the free transfer of ideas and information. Consider the difficulties that would arise if the botanists in the United States, England, Germany, Russia, China, etc. each had their own independent set of names for the plants of their countries.
- The same scientific name may not be used for more than one kind of plant. Once it has been published, that name cannot be used again for any other plant.
- Scientific names are given according to an "International Code of Botanical Nomenclature." These regulations are reviewed every four years at International Botanical Congresses. There is, therefore, a legally correct scientific name.
- ☼ Inherent in our system of scientific names is the concept of evolutionary or genetic relationship. When we place einkorn wheat, emmer wheat, and bread wheat in the same genus (*Triticum*), we do so because we have concluded that the morphological, anatomical, genetic, and chemical traits that they share suggest that they are closely related. Because there is a set of features associated with the name, it has predictive value. The better we have circumscribed the taxa, the higher the value.

There are some difficulties with scientific names. They can be difficult to pronounce, especially if you did not

learn to divide words into syllables early on in your education. You might note, however, that such familiar and easily pronounced common names as aster, rhododendron, magnolia, chrysanthemum, petunia, and begonia are also the first part of the scientific names of these plants. My own experience in teaching undergraduates to use scientific names is that once you can get past the psychological barrier that these are terribly long words that only those who have had a strong background in Latin and Greek can pronounce, then you will become much more comfortable with them and begin using them rather easily.

WHY ALL THE NAME CHANGES?

One of the more frustrating features of scientific names, especially for someone who is just learning about them, is that they are changed from time to time. Just when you think that you have become familiar with the scientific names for a particular group of plants, someone will publish a new revision of the group and you discover that some of the names have been changed. These changes come about for several reasons. As new information about the anatomy, chemistry, and genetics of plants becomes known, it may cause botanists to rethink the evolutionary relationships among the plants being studied. These changes may require us to revise the scientific names to reflect the new level of information now available to us. Sometimes names are changed, not for biological reasons, but because someone studying a group may discover that the name given to a particular plant has to be rejected because it violated some provision of the International Code of Botanical Nomenclature. For example, the name may not have been properly published in a scientific journal. One 19th century botanist was a public school principal who handed out printed copies of his newly described plants to his students at graduation each spring. This is not exactly

Examples point out one of the important operating principles in plant classification. As new information becomes available and as errors are discovered, we make adjustments and corrections. What appears to be a fine scheme of classification today may be modified drastically or even discarded completely at some point in the future.

COMPONENTS

If we examine the botanical works of the 15th and 16th centuries, we see that the name of a plant was often a **polynomial**, a lengthy series of descriptive words, typically in Latin, as in "Convolvulus argentateus foliis ovatis divisis basi truncatis: laciniis intermediis duplo longioribus." These phrase names became increasingly awkward because the discovery of a new kind of plant required that the existing polynomial be slightly modified so that it could be distinguished from the older one.

A new way of naming plants was developed over two centuries ago to replace the polynomials. It was popularized by Carolus Linnaeus, the leading botanist of his time. This system was based upon the principle that each plant (or animal for that matter, because they are named according to the same scheme) is given a scientific name that consists of two components, both of them parts of the taxonomic

hierarchy mentioned above. The first element of the scientific name is the **genus** (or generic name), as in *Triticum*, the genus of wheat. The plural of genus is **genera**, not genuses. The second element is the **specific epithet**, as in *aestivum*, the particular kind of wheat called bread wheat. This second element of the scientific name is often incorrectly called the "species." It is the genus and specific epithet together that form the species name. *Triticum aestivum* is the species name of bread wheat. Because the name of a plant or animal is the combination of these two words, the scientific name is called a **binomial** and we call this scheme of giving technical names to organisms the **Binomial System of Nomenclature**.

The binomial, for reasons of completeness and accuracy, is followed by the name (typically abbreviated) of the person or persons who first published that name for the plant. For example, in the scientific name *Zea mays* L., the "L." stands for Linnaeus. This part of the scientific name is the **authority**.

It is sometimes necessary to move the name of a plant from one genus to another, usually because more recent research has demonstrated that the plant was incorrectly assigned to a particular genus. For instance, in 1753 Linnaeus published the name Panicum dactylon for the plant that we now call Bermuda grass. In 1805, Christian Persoon transferred the epithet (or "moved the species," as we say more informally) from Panicum to the new genus Cynodon. The scientific name of Bermuda grass then becomes Cynodon dactylon (L.) Persoon. The person whose name is in the parentheses first published the specific epithet for the plant. The name after the parentheses is that of the person who transferred it into the genus where it now resides.

It is often useful to recognize and to name variation below the species level. The two most widely used are the **subspecies** (abbreviated ssp.) and the **variety** (abbreviated var.). These names also have authorities, as in *Bromus vulgaris* (Hook.) Shear var. *robustus* Shear. If the subspecies or varietal name is a repeat of the specific epithet, then the authority is not repeated, as in *Zea mays* L. ssp. *mays*.

An additional explanation is needed for the term variety. For reasons that are obvious, we have developed many different cultivated strains of a particular crop plant or ornamental. There are literally thousands of different kinds of rice. There are probably hundreds of different kinds of tuberous begonias. In general parlance, we often call these varieties. However, for purposes of formal nomenclature, these variations are considered too minor and often too short-lived to warrant giving them a scientific name. The variety of botanical nomenclature is not used in these instances. Instead, we employ the term **cultivar** (abbreviated cv.). A kind of sorghum used to make molasses in the Southwest by American Indians is *Sorghum bicolor* cv. 'Apache Red Cane.'

Many of our economic plants are of hybrid origin, that is they result from the accidental or purposeful crossing of two closely related plants. This can be reflected in the scientific name of the plant by inserting an "X." If the X occurs before the generic name, then the plant is considered the result of a cross between two plants in different genera. X Agropogon is an intergeneric hybrid between Agrostis and Polypogon. If the X occurs between the generic

name and the specific epithet, then the plant is the product of a cross between two species in the same genus, as in *Tridens* x *oklahomensis*.

THE ORIGIN OF NAMES

Most of the words that make up scientific names are derived from Latin or Greek, although there is no requirement that they must be. Modern names and even nonsensical ones have been used. Many students, however, believe that there must be some requirement that scientific names be as long and unpronounceable as possible. This reveals a certain lack of scholarship. Even a rudimentary knowledge of etymology is very helpful in understanding the composition of scientific names. The following examples may be helpful.

Commemorative Names: Lamarckia, Scribneria, and Orcuttia are named after J. B. A. P. Monnet de Lamarck (the famous French naturalist), Frank L. Scribner (a noted American agrostologist), and Charles Russell Orcutt (a San Diego botanist). Agnesia is named after Agnes Chase.

Classical/Aboriginal Names: Agrostis, Bromus, Festuca, and Poa are all ancient Latin names for grasses.

Geographical Names: anglicus (from England), gallicus (from France), canadensis (from Canada), australis (southern)

Habitat: arenarius (growing in sand), campestris (of the fields), fluviatilis (of the rivers), riparius (of the river banks), sativus (cultivated), littoralis (of the seashore).

Growth Form: arboreus (tree), repens (creeping), scandens (climbing), pusillus (insignificant).

Structural Feature: amabilis (lovely in appearance), bulbosum (having a swollen part), gracilis (slender), mollis (soft hairy), scoparius (broom-like).

Use: esculentus (edible), officinalis (recognized as medically important), textilis (having useful fibers)

DERIVATION OF GENERIC NAMES OF GRASSES

[Gk., awned scale] Achnatherum Aegilops [L., ancient name for wheat] Aegopogon [Gk., goat + beard Agropyron [Gk., wild + wheat] [Gk., a kind of grass, pasture] [Gk., a kind of grass] Agrostis Aira [Gk., different + scale Allolepis [Gk., fox + tail] Alopecurus Ammophila [Gk., sand + loving]
Ampelodesmos [Gk., grape leaves + to tie together]
Amphicarpum [Gk., double + fruit-bearing] Andropogon [Gk., man + beard] [Gk., flower + contrary] [Gk., flower + to bear] Anthaenantia Anthephora [Gk., yellow + flower Anthoxanthum Apera [Gk., not maimed] Äpluda [L., chaff] [L., arctic + a kind of grass] Arctagrostis

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Arctophila
                                [L., arctic + G., to love]
                                  [L., a stout hair, awn]
Aristida
Arrhenatherum
                                      [Gk., male + awn]
Arthraxon
                                       [Gk., joint + axis
                                       [L., a reed grass]
Arundo
Avena
                                       [L., oats]
[Gk., axis + foot]
Axonopus
Beckmannia
                      [J. Beckmann, German botanist]
Blepharidachne
                                  [Gk., eyelash + chaff]
Blepharoneuron
                                 [Gk., eyelash + nerve
              [Gk., pit + grass]
[C. and E. Boutelou, Spanish botanists]
Bothriochloa
Bouteloua
Brachiaria
                                                [L., arm]
                                     [Gk., short + husk]
Brachyelytrum
                                       [Gk., thick + foot]
Brachypodium
                          [Gk., a kind of nodding grain]
Briza
Bromus
                    [Gk., food, ancient name for oats]
Buchloe
                                  [Gk., buffalo + grass]
Calamagrostis
                                      [Gk., a reed grass]
Calamovilfa
                     [Gk., reed + Vilfa, a grass genus]
Catabrosa
                                         Gk., devouring
Cathestecum
                                        [Gk., stationary]
                                 [Gk., an ancient name]
Cenchrus
Chloris
                              [Gk., Goddess of flowers
                                  [Gk., golden + beard]
[Gk., a kind of grass]
[Gk., a kind of palm]
Chrysopogon
Cinna
Coix
Coleanthus
                                  [Gk., sheath + flower]
                                     [Sp., cutting]
[Gk., club-bearing]
Cortaderia
Corynephorus
                                            [Gk., hidden]
Crypsis
Ctenium
                                     [Gk., a small comb]
                                     [Gk., boat + beard]
Cymbopogon
Cynodon
                                      [Gk., dog + tooth]
                                        [Gk., dog + tail]
Cynosurus
                             [Gk., finger]
[Gk., finger + small comb]
Dactylis
Dactyloctenium
Danthonia
                        [E. Danthione, French botanist]
                  [Gk., hairy + grass]
[J. Deslongchamps, French botanist]
[J. B. Desmazieres, French botanist]
Dasyochloa
Deschampsia
Desmazeria
Diarrhena
                                     [Gk., twice + male]
Dichanthelium
                                [Gk., twice + flowering]
                                       [L., finger]
[Arabic, little tail]
Digitaria
Dinebra
Dissanthelium
                              [Gk., two + small flower]
Dupontia
Distichlis
                                       [Gk., two-ranked]
Echinochloa
                               [Gk., hedgehog + grass]
                       [J. F. Ehrhart, German botanist]
Ehrharta
Eleusine
                        [Gk., Eleusis, an ancient town]
                                   [Gk., to roll + tail]
[Gk., a kind of millet]
Elionurus
Elymus
Elytrigia
                                    [Elymus + Triticum]
Enneapogon
                                     [Gk., nine + beard
                                 [G., intestine + beard
Enteropogon
                                     [Gk., love + grass]
Eragrostis
Eremochloa
                               [Gk., centipede + grass]
                                    [Gk., wool + flower]
Erianthus
                                   [L., woolly + grass]
[Gk., woolly + nerve]
Eriochloa
Erioneuron
Euchlaena
                             [Gk., true or well + cloak]
Festuca
                 [L., classical name for a weedy grass]
Fingerhuthia
                  [K. A. Fingerhuth, German botanist]
                             [Gk., a small pouch or sac]
Gastridium
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[J. F. P. G. Gaudin, French botanist]

[Gk., sweet]

[Gk., naked + beard]

Gaudinia

Glyceria

Gymnopogon

Gynerium [Gk., female + wool] Hackelochloa [E. Hackel + Gk., grass] Hainardia [P. Hainardi, Swiss phytogeographer]	Polypogon [Gk., many + beard] Pseudoroegneria [Gk., false + a name for Elymus] Ptilagrostis [Gk., feather + grass] Puccinellia [B. Puccinelli, Italian botanist]
Hakonechloa [Mt. Hakon (in Japan) + Gk., grass] Heleochloa [Gk., marsh + grass] Helictotrichon [Gk., twisted + bristle] Hemarthria [Gk., blood (?) + joint] Hesperostipa [Gk., western + tow] Heteropogon [Gk., different + beard]	Redfieldia [J. H. Redfield, Philadelphia businessman] Reimarochloa [J. A. H. Reimarus + Gk., grass] Rhynchelytrum [L., beak + scale] Rottboellia [C. F. Rottboell, Danish botanist]
Hierochloe [Gk., holy + grass] Hilaria [A. de St. Hilaire, French botanist] Hordeum [L., classical name for barley] Holcus [L., a kind of grass] Hydrochloa [Gk., water + grass] Hyparrhenia [Gk., below + male]	Saccharum [L., sugar] Sacciolepis [Gk., small bag + scale] Schedonnardus [Gk., near + Nardus, a grass genus] Schismus [Gk., split] Schizachne [Gk., split + chaff] Schizachyrium [Gk., split + chaff]
Hystrix[Gk., porcupine]Imperata[F. Imperato, Italian naturalist]Ischaemum[G., hip-socket joint]	Sclerochloa[Gk., hard + grass]Scleropogon[Gk., hard + beard]Scolochloa[Gk., prickle + grass]Scribneria[F. L. Scribner, American botanist]Secale[L., classical name for rye]
Koeleria [G. L. Koeler, German botanist]	Setaria [L., bristle] Setariopsis [L., Setaria + resembling]
Lagurus [Gk., hair + tail] Lamarckia [J. B. Lamarck, French naturalist] Lasiacis [Gk., woolly + point] Leersia [J. D. Leers, German apothecary] Leptochloa [Gk., slender + grass] Leptoloma [Gk., thin + border] Leymus [Anagram of Elymus] Limnodea [Alteration of Limnas, a grass genus] Lolium [L., classical name for darnel] Luziola [Luzula, a genus of sedges + resembling] Lycurus [Gk., wolf + tail]	Sieglingia Sitanion Sorghastrum Sorghum Spartina Sphenopholis Sporobolus Stenotaphrum Stipa Swallenia Sitanion [Gk., a kind of grain] [It., sorgho] [Gk., sedge + scale] [Gk., seed + to throw] [Gk., narrow + trench] [In tow] [I
Manisuris[Gk., necklace + tail]Melica[L., honey]Melinis[Gk., a kind of millet]Microchloa[Gk., small + grass]Microstegium[Gk., small + cover]Miscanthus[Gk., stalk + flower]Molinia[J. I. Molina, Jesuit missionary-botanist]Monanthochloe[Gk., one + flower + grass]Monerma[Gk., one + support]Monroa[W. Munro, English botanist]Muhlenbergia[H. L. E. Muhlenberg, Penn. botanist]Nassella[L., a kind of basket]Neostapfia[Gk., new + O. Stapf, British botanist]	Taeniatherum Themeda Thysanolaena Torreyochloa Trachypogon Tragus Tribolium Trichachne Trichloris Trichoneura Tripogon Tripogon Tripogon Tripogon Tripogon Tripogon Tripogon Tripogon Tripsacum Tripsacum Tripsetum [Gk., ribbon + awn] [G., fringe + cloak] [Gk., rough + beard] [Gk., rough + beard] [Gk., he-goat] Thrichery [Gk., hair + chaff] [Gk., hair + chaff] [Gk., hair + nerve] [Gk., hair + nerve] [Gk., three + tooth] [Gk., three + beard] [Gk., three + beard] Tripsacum [Uncertain: perhaps Gk., to rub] Trisetum
Neyraudia [Anagram of Reynaudia]	Tuctoria [Anagram of Orcuttia]
Olyra [Gk., an ancient name for a kind of grain] Oplismenus [Gk., armed] Orcuttia [C. Orcutt, California botanist] Oryza [Gk., classical name for rice] Oryzopsis [Gk., rice + resembling] Panicum [L., ancient name for common millet]	Uniola [L., a kind of grass] Urochloa [Gk., tail + grass] Vaseyochloa [G. Vasey, American botanist + grass] Ventenata [P. Ventenat, French botanist] Vetiveria [Tamil name for this grass] Vulpia [J. S. Vulpius, German botanist]
Pappophorum [Gk., pappus + bearing] Parapholis [Gk., beside + scale]	Willkommia [H. M. Willkomm, German botanist]
Pascopyrum[L., pasture + Gk., wheat]Paspalum[Gk., a kind of grass]Pennisetum[L., feather + bristle]Phalaris[Gk., a grass with shiny spikelets]Pharus[Gk., mantle or cloth]Phippsia[C. J. Phipps] XXX	Zea [Gk., a kind of grain] Zizania [Gk., a weed of grain fields] Zizaniopsis [Gk., Zizania + resembling] Zoysia [K. von Zois, German botanist]
Phleum [Gk., a marsh reed] Phragmites [Gk., growing in hedges along streams]	PRONUNCIATION
Piptatherum[Gk., falling + bristle]Piptochaetium[Gk., falling + hair]Pleuraphis[Gk., side + needle]Pleuropogon[Gk., side + beard]Poa[Gk., ancient name]Pogonarthria[G., beard + a joint]	The International Code of Botanical Nomenclature states that scientific names of plants are to be treated as Latin words, regardless of their origin. A few of the more scholastically inclined botanists will argue,

therefore, that we ought to pronounce scientific names according to the strict rules of the sounds of vowels and consonants in Latin and that great care should be taken in accenting the proper syllable. But, there are traditional English, reformed academic, and Church Latin versions of Latin to choose from, each with its own set of rules for pronunciation.

Most American botanists pronounce the scientific names of plants as though they were English words. Some of us follow the rules in Latin for determining which syllable is accented; most of us do not. Many of us pronounce scientific names the way we were taught as under-graduates (if any formal discussion occurred) or more commonly we imitate the way our professors said them when we took their classes. These become the familiar and "correct" way to pronounce the scientific names of plants.

The following is an attempt on my part to present a basic guide to pronouncing vowels, consonants, and diphthongs, together with some of the rules for accenting syllables.

- The letters of the Latin alphabet are basically the same as ours, except that J, U, and W did not occur in the classical version.
- Each syllable will contain a vowel or a double vowel combination (ae, au, ei, oe, or ui). The latter are called diphthongs.
- Pronounce all of the syllables. Secale is "see-kale," not "see-kale."
- Final vowels are long, with the exception of a. If a word ends in two vowels (unless they are a diphthong), they are sounded separately. The epithet angustifolia is pronounced "an-gust-i-fo-liah."
- The diphthongs "ae" and "oe" have the sound "e," as in beat; "au" has the sound of "aw," as in the word awful; "ei" usually has the sound "i," as in site; "eu" has the sound of "u," as in neuter; and "ui" has the ui-sound in the word ruin.
- The "oi" in the ending "-oides" is treated as a diphthong by most American botanists and we give it the sound that "oi" has in the word oil. This habit is considered close to barbaric by English and Europeans who are much more persnickety about such matters. Because these two vowels do not form a diphthong, they should be pronounced separately, so that the ending "-oides" has the sound "-o-e-deez."
- A single consonant is placed with the following vowel, as in "pa-ter." Double consonants are separated, as in "am-mi." If there are two or more consonants, the first one is usually put with the preceding vowel, as in "an-gli-cus."
- B, d, f, h, l, m, n, p, qu, and z are pronounced the same in Latin and English.
- The consonants c and g are soft (that is, have the sounds of "s" and "j") if they are followed by ae, e, i, oe, or y. Otherwise, the c is pronounced like a "k" and the g is also hard, as in "go." The s is always pronounced as it is in the word "so," not as a "z." An initial x is pronounced as a "z," not "ekz." Xerochloa is pronounced "zero-o-chlo-a," not "ek-zero-chlo-a."

The first letter is silent in words beginning with cn, ct, gn, mn, pn, ps, pt, and tm.

Accenting the proper syllable can be tricky. Sometimes the author of a flora or other manual may provide assistance by including an accent mark. Most do not. If included, they are for the convenience of the reader and they are not part of the scientific name itself. If you must determine which syllable to accent, the following rules may be helpful.

Words of two syllables are always accented on the first syllable. The genus of Kentucky bluegrass is "Po-a." In words of three or more syllables, the last syllable is never accented. The stress will fall either on the next to the last syllable (the penultimate syllable), as in "ar-ven-sis," or on the third from the last syllable (antepenultimate), as in "an-gli-cus."

No matter how long the word, the accent can never be to the left of the antepenultimate syllable. Deciding between these two options is a difficult choice. Accent the penultimate syllable if it ends in a consonant, diphthong, or in a long vowel.

Commemorative names (patronyms) present a special problem because giving them the proper accenting can render the person's name unrecognizable. The epithet *jamesii* should be pronounced "ja-**me**-se-i," not "j**amz**-e-i." Most American botanists ignore this rule.

There is a somewhat less scholarly approach that you might find useful.

- Pronounce all of the syllables.
- Say them as you would any English syllables.
- Put the accent where you think it sounds best.
- Try to be consistent.

A LITTLE ETYMOLOGY

PREFIXES

a- without angusti- narrow apo- separate **bi-** two **brachy-** short **brevi-** short chori- separate **cleisto-** closed, hidden con- with echino- spiny eu- true, typical, good ex- without gyno- female **halo-** salt homo- the same **hyper-** above hypo- below in- in, within
inter- between lati- broad, wide longi- long macro- large, great meso- in the middle micro- small mono- one

multi- many neo- new ob- inverse, upside down parvi- small pauci- few penta- five **peri-** around, about phyllo- leaf poly- many **pseudo-** false **sub-** below, somewhat sym- with **syn-** growing together tetra- four tri- three uni- one xero- dry

SUFFICES

-aceus likeness, resemblance
-chloa grass
-flora flowered
-formis having the form of
-ifera bearing or yielding
-oides resembling
-phylla relating to leaves
-pogon beard
-seta bristle, stiff hair
-stachys an ear of grain

EPITHETS

alba - white **arenaria** - growing in sandy places **aristata** - bearing an awn or bristle arundinacea - reed-like arvensis - of cultivated lands canadensis fasciculatus - clump or bundle filiformis - thread-shaped foliosa - leafy formosus - handsome **glauca -** bluish-green; gray **gracilis** - slender **inermis** - without spines or prickles miliacea - millet-like (a minor cereal) **mollis** - pubescent nigra - black occidentalis - west, western orientalis - east, eastern palustris - growing in swamps and marshes **pilosa** - hairy **pratensis-** growing in meadows pumila - dwarf **repens** - creeping *rubra -* red sativa - sown or planted speciosa - showy tenuis - thin, fine, slender verticillata - a whorl or circular arrangement virginica vulgaris - common, ordinary

WRITING SCIENTIFIC NAMES

There are a few simple rules that must be followed in writing scientific names. The genus is always capitalized; the specific epithet should not be. The rules of nomenclature allow them to be if they are commemorative, as in *Elymus smithii* (a relative, no doubt) or if the epithet was once itself a generic

name, as in *Arundo Donax*, the giant reed grass. Even in such instances, however, the rules discourage capitalization.

The generic name and specific epithet are underlined when they appear in handwritten or typed material. They are put in italics or bold-face in printed text. The authority is always capitalized, but it is not underlined or otherwise set off from the remainder of the text.

THE TAXONOMIC HIERARCHY

The taxonomic hierarchy is the series of categories that have been arranged in a particular sequence to show relationships with one another. The names of these categories and their sequence are set by the ICBN. It is the official list of groups into which plants are classified. Any one of these categories, at any level, is called a **taxon** (plural, **taxa**). The sequence of taxa and their standard endings is as follows:

Category	Ending	Example
Division or Phylum Class Subclass Order Family-aceae Subfamily Tribe Subtribe Genus * Subgenus Section Species	-phyta -opsida -idae -ales -oideae -eae -inae * *	Magnoliophyta Liliopsida Commelinidae Poales Poaceae Panicoideae Andropogoneae Tripsacinae Zea Zea Zea Zea mays
Subspecies Variety	*	mexicana mexicana

* No standard ending

There are no standardized endings for taxa at or below the rank of genus. There are, however, grammatical consid-erations. Their terminations must agree in gender and in number. This explains why *Tridens pulchellus* becomes *Erioneuron pulchellum* when the epithet is transferred from one genus to the other.

My reason for including this brief discussion of the taxonomic hierarchy is that the generic and species names are two levels of this hierarchy. In this course, we will use the names of subfamilies, tribes, genera, and species frequently.

GRAMINEAE VERSUS POACEAE

Before leaving the subject of the naming of grasses, an explanation of the family name of the true grasses is in order. In older references, grasses seem to belong to a family called Gramineae, while in more recent publications the family name is Poaceae. This might suggest that the "old name" Gramineae has been replaced by the "new name" Poaceae, probably because Poaceae is considered the correct name and Gramineae is now incorrect for some reason.

One of the basic principles of botanical nomenclature is that we are to use the first validly and effectively published name for a plant or group of plants (genus,

family, etc.). The name Gramineae was published by A. L. Jussieu in 1789; Poaceae by Barnhart in 1895. Therefore, following this principle of priority of publication, Gramineae is the correct name for the grass family.

The problem arises because another section of the International Code of Botanical Nomenclature (ICBN) requires that family names end in the suffix -aceae, which Gramineae obviously does not. Nor do Compositae, Cruciferae, Labiatae, Leguminosae, Umbelliferae, and Palmae. What do all of these families have in common? They are among our best known, most easily recognized, and most economically important plant families. These names were in use long before we had any international rules to govern such matters.

Which name, Gramineae or Poaceae, is correct? They are both correct and they may be used interchangeably. The ICBN makes an exception in these cases and allows two valid names for the same group. Article 18.5 of the ICBN states, "The following names, sanctioned by long usage, are treated as validly published: ... Gramineae (Poaceae; type *PoaL.*)...." Article 18.6 then goes on to say that, "The use, as alternatives, of the names in parentheses in Art. 18.5 is authorized."

The same holds for these first published family names and their equivalents with an -aceae ending:

Compositae/Asteraceae Cruciferae/ Brassicaceae Guttiferae/Clusiaceae Labiatae/ Lamiaceae Leguminosae/Fabaceae Umbelliferae/Apiaceae Palmae/Arecaceae.

My personal preference is to use Gramineae. It was the first name to be legally published for the family. It is the name used in the Code itself. Poaceae is the alternate name for Gramineae, and not the other way around.

SELECTED REFERENCES

Boorstin, D. J. 1983. The invention of species. <u>In</u>, The discoverers: a history of man's search to know his world and himself. Random House. New York, NY. Pp. 429-435.

Clifford, H. T. 1996. Etymological dictionary of grasses. CD-ROM. World Biodiversity Database. Expert Center for Taxonomic Identification. Amsterdam.

Coombes, A. J. 1985. Dictionary of plant names: the pronunciation, derivation and meaning of botanical names, and their common-name equivalents. Timber Press. Portland, OR. 207 pp.

Estes, J. R. & R. J. Tyrl. 1987. Concepts of taxa in the Poaceae. <u>In</u>, Soderstrom, T. R. et al. (editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 325-333.

Gledhill, D. 1989. The names of plants. Second edition. Cambridge University Press. Cambridge, England. 159 pp.

Greuter, W. et al. 2000. International code of

botanical nomenclature. Adopted by the Sixteenth International Botanical Congress, St. Louis, Missouri, July-August 1999. Koeltz Scientific Books. Königstein, Germany. 474 pp.

Hyam, R. & R. Pankhurst. 1995. Plants and their names: a concise dictionary. Oxford Univ. Press. Oxford, England. 545 pp.

McVaugh, R., R. Ross, and F. A. Stafleu. 1968. An annotated glossary of botanical nomenclature. Regnum Vegetabile Vol. 56. International Bureau for Plant Taxonomy and Nomenclature of the International Association for Plant Taxonomy. Utrecht, The Netherlands. 31 pp.

Neal, B. 1992. Gardener's Latin. Algonquin Books. Chapel Hill, NC. 136 pp.

Smith, J. P., Jr. 1977. The taxonomic hierarchy and scientific names. <u>In</u>, Vascular plant families. Mad River Press. Eureka, CA. Pp. 12-20.

Stearn W. T. 1992. Botanical Latin: history, grammar, syntax, terminology and vocablary. Fourth edition. David & Charles. Devon, England. 546 pp.

Trehane, P. et al. 1995. International code of nomenclature for cultivated plants. Sixth edition. Quarterjack Publ. Dorset, England. 175 pp.

Weber, W. A. 1986. Pronunciation of scientific names. Madroño 33: 234, 235.

6.02 - GREAT MOMENTS IN AGROSTOLOGY

- 1703 J. Ray publishes *Methodus Graminum,* Juncorum et Cyperorum Specialis
- 1709 J. Scheuchzer publishes *Agrostographiae Helveticae Prodromus*
- 1737 C. Linnaeus publishes Genera Plantarum
- 1753 C. Linnaeus publishes Species Plantarum
- 1811 A. J. Gaudin publishes Agrostologia Helvetica
- 1812 P. de Beauvois publishes *Essai d'une Nouvelle Agrostographie*
- 1822 C. B. Trinius publishes Clavis Agrostographiae
- 1829 C. G. Nees von Esenbeck publishes *Agrostologia Brasiliensis*
- 1835 C. S. Kunth publishes *Distribution Methodique* de la Famille des Graminees.
- 1855 Charles Darwin identifies his first grass!
- 1855 E. G. Steudel publishes *Synopsis Plantarum Glumacearum*
- 1881 G. Bentham publishes "Notes on Gramineae"
- 1881 E. Hackel publishes "Untersuchungen über die Lodiculae der Gräser"
- 1883 C. S. Kunth publishes Agrostographia...
- 1883 G. Vasey publishes Grasses of the United States
- 1883 G. Bentham publishes treatment of Gramineae in *Genera Plantarum*
- 1884 G. Vasey publishes *Agricultural Grasses of the United States*
- 1887 W. J. Beal publishes Grasses of North America
- 1887 E. Hackel publishes treatment of Gramineae in Engler & Prantl's *Die Natürlichen Pflanzenfamilien*
- 1891 G. Vasey publishes Grasses of the Southwest
- 1892 G. Vasey publishes Grasses of the Pacific Slope
- 1892 G. Vasey publishes Monograph of the Grasses of the United States and British America
- 1892 E. Bruns publishes "Der Grasembryo"
- 1896 W. J. Beal publishes *Grasses of North America*, Vol. II
- 1896 A. Grob publishes *Bieträge zur Anatomie der Epidermis der Gramineenblätter*
- 1897 F. Scribner publishes American Grasses
- 1903 G. V. Nash publishes treatment of Gramineae in J. K. Small's Flora of the Southeastern United

States

- 1910 A. S. Hitchcock & A. Chase publish "The North American Species of *Panicum*"
- 1913 E.-G. Camus publishes Les Bambusées
- 1917 E. A. Bessey publishes "Phylogeny of the Grasses"
- 1917 O. Stapf publishes first installment of Gramineae in *Flora of Tropical Africa*
- 1922 A. Chase publishes First Book of Grasses
- 1927 C. Niles publishes Beauvois' Agrostographie
- 1929 J. W. Bews publishes *The World's Grasses*
- 1931 N. P. Avdulov publishes "Kariosistematicheskoye Issledovaniye Semeystva Zlakov"
- 1932 H. Prat publishes "L' épiderme des Graminées..."
- 1932 M. K. Elias publishes "Grasses and Other Plants from the Tertiary Rocks of Kansas and Colorado"
- 1934 A. Arber publishes *The Gramineae*
- 1935 P. Weatherwax publishes "Phylogeny of Maize"
- 1935 A. S. Hitchcock publishes *Manual of the Grasses* of the *United States*
- 1936 H. Prat publishes "La Systématique des Graminées"
- 1939 P. Mangelsdorf & R. G. Reeves publish *The Origin of INdian Corn and Its Relatives*
- 1939 G. W. Beadle publishes "Teosinte and the Origin of Maize"
- 1939 R. Pilger publishes "Zur Morphologie des Ahrchens der Gramineen"
- 1951 A. Chase publishes revised edition of *Manual of the Grasses*
- 1954 R. Pilger publishes "Das System der Gramineae"
- 1954 T. Tateoka publishes "On the Systematic Significance of Starch Grains of Seeds in Poaceae"
- 1955 A. Beetle publishes "The Four Subfamilies of the Gramineae"
- 1956 G. L. Stebbins publishes "Cytogenetics and Evolution of the Grass Family"
- 1956 G. L. Stebbins publishes "Taxonomy and Evolution of Genera, with Special References to the Family Gramineae"
- 1958 W. V. Brown publishes "Leaf Anatomy in Grass

Systematics"

- 1960 C. R. Metcalf publishes *Anatomy of the Monocotyledons, Vol. I. Gramineae*
- 1960 H. Prat publishes "Vers une Classification Naturelle des Graminées"
- 1960 W. V. Brown publishes "The Morphology of the Grass Embryo"
- 1960 T. Tateoka publishes "Cytology in Grass Systematics: a Critical Review"
- 1960 N. L. Bor publishes *Grasses of Burma, Ceylon, India and Pakistan*
- 1961 G. Ledyard Stebbins & Beecher Crampton publish "A Suggested Revision of the Grass Genera of Temperate North America"
- 1961 H. T. Clifford publishes "Floral Evolution in the Family Gramineae"
- 1961 H. J. Conert publishes *Die Systematik und Anatomie der Arundineae*
- 1962 Agnes Chase & Cornelia Niles publish *Index to Grass Species*
- 1966 F. A. McClure publishes *The Bamboos: a Fresh Perspective*
- 1968 F. W. Gould publishes Grass Systematics
- 1972 G. L. Stebbins publishes "The Evolution of the Grass Family"
- 1973 F. A. McClure publishes "Genera of Bamboos Native to the New World..."
- 1975 F. W. Gould publishes The Grasses of Texas
- 1976 N. N. Tsvelev publishes Zlaki SSSR
- 1978 F. W. Gould & C. A. Clark publish "Dichanthelium (Poaceae) in the United States and Canada
- 1978 R. W. Pohl publishes How to Know the Grasses
- 1979 H. E. Conner publishes "Breeding Systems in the Grasses: a Survey"
- 1979 H. H. Iltis et al. publishes "Zea diploperennis (Gramineae): a new Teosinte from Mexico"
- 1980 C. E. Calderón & T. R. Soderstrom publish "The Genera of Bambusoideae (Poaceae) of the American Continent..."
- 1980 R. W. Pohl publishes treatment of Gramineae in Flora Costaricensis
- 1981 L. Watson & M. J. Dallwitz publish "An Automated Data Bank for Grass Genera"
- 1983 H. H. Iltis publishes "The Catastrophic Sexual Transmutation Theory..."
- 1986 Derek Clayton & S. Renvoize publish *Genera Graminum: Grasses of the World*
- 1987 Thomas Soderstrom et al. publish Grass

Systematics and Evolution

- 1989 N. N. Tzvelev publishes *The System of Grasses* ... and Their Evolution
- 1992 L. Watson & M. J. Dallwitz publish *The Grass Genera of the World*
- 1996 L. G. Clark & R. W. Pohl publish *Agnes Chase's First Book of Grasses (4th edition)*
- 1998 R. J. Soreng & J. I. Davis publish *Phylogenetics* and Character Evolution in the Grass Family
- 1999 E. J. Rudziewicz et al. publish *American Bamboos*
- 2000 Jacobs & Everett publish *Grass Systematics and Evolution*
- 2001 Grass Phylogeny Working Group publishes new system of subfamilies and tribes
- 2003 Second volume of the treatment of Gramineae published in *Flora of North America north of Mexico*

6.03 - WHO WERE HITCHCOCK & CHASE?

Albert Spear Hitchcock was born in Michigan in 1865, grew up in Kansas and Nebraska, and went to Iowa State Agricultural College (now Iowa State University), where he graduated in 1884. [1865 to 1884 = 19 years!] While there he studied under the eminent American botanist, Charles Edwin Bessey. He was then a faculty or staff member at Iowa State, the State University of Iowa, the Missouri Botanical Garden, Washington University, and Kansas State Agricultural College (now Kansas State University). In 1901 he moved to Washington, D. C. as the Assistant Chief of the Division of Agrostology in the U.S. Department of Agriculture. He would spend the remainder of his career working there, becoming the head of the grass collection of the United States National Herbarium, and one of this country's most respected systematic botanists. Willis Lynn Jepson, the distinguished University of California botanist and pre-eminent expert on the state's flora, once inscribed a book to Dr. Hitchcock, calling him an "eager explorer, farseeing botanist, and wise promoter of scientific research in America." Dr. Hitchcock died in 1935, on board a ship returning from an International Botanical Congress in Europe.

His best known work, the "Manual of the Grasses of the United States," was published only months before his death. The first printing sold out in a matter of weeks. I have been told that the two editions of The Manual are the top-selling government publications in history. The U. S. Government Printing Office finally had to give up on reprinting the second edition because the plates had worn out! The comprehensive nature of the work, its keys and illustrations, made it the "Bible" for people needing to know about grasses. Its system of subfamilies and tribes, and the names of individual grasses, would dominate regional and state floras for decades.

One year after A. S. Hitchcock moved to Washington, D. C., so did Mary Agnes Chase, as a scientific illustrator for the Department of Agriculture. In 1905, she started to work for Dr. Hitchcock. Being a person of great intelligence and sensitivity, she fell in love with grasses. She became Hitchcock's scientific collaborator and was a major force behind the publication of The Manual in 1935. She retired as Senior Botanist at the Smithsonian in 1939, having become the successor to A. S. Hitchcock. She stayed on as an unsalaried research scientist until her death in 1963. One of her greatest accomplishments was the revision of The Manual, which appeared in 1951. Many of us, wanting to give her the recognition that she richly deserved, always call it "Hitchcock and Chase."

In her later years, Mrs. Chase could have been the type specimen of the little old granny. But as a young woman she was an activist for women's causes – not

at all the shy and retiring lady botanist. On one or two occasions she was put in jail for her political beliefs.

ALBERT SPEAR HITCHCOCK

- 1891. Hitchcock, A. S. A catalogue of the Anthophyta and Pteridophyta of Ames, Iowa. St. Louis Acad. Sci. 5(3): 477-532.
- 1894. Hitchcock, A. S. A key to the genera of Manhattan plants, based upon fruit characters.... Mercury Publ. Manhattan, KS. 35 pp.
- 1894. Hitchcock, A. S. A key to the spring flora of Manhattan.... Mercury Publ. Manhattan, KS. 35 pp.
- 1896. Hitchcock, A. S. The grasses of Kansas. Trans. Kansas Acad. Sci. 14: 135-149.
- 1899. Hitchcock, A. S. & G. L. Clothier. Native agricultural grasses of Kansas. Hitchcock, A. S. Kansas Exp. Stat. Bull. 87: 1-29.
- 1903. Hitchcock, A. S. North American species of *Leptochloa*. U. S. Dept. Agric. Bull. No. 33. 24 pp. + 6 plates.
- 1905. Hitchcock, A. S. North American species of *Agrostis*. U. S. Dept. Agric. Bull. No. 68. 68 pp.
- 1905. Hitchcock, A. S. The identification of Walter's grasses. Ann. Rep. Missouri Bot. Gard. 16: 31-56.
- 1906. Hitchcock, A. S. Notes on North American grasses. VI. Synopsis of *Tripsacum*. Bot. Gaz. 41: 294-298.
- 1908. Hitchcock, A. S. Types of American grasses: a study of the American species of grasses described by Linnaeus, Gronovius, Sloane, Swartz, and Michaux. Contr. U. S. Natl. Herb. 12(3): 113-158.
- 1909. Hitchcock, A. S. Catalogue of the grasses of Cuba. Contr. U. S. Natl. Herb. 12(6): 183-258.
- 1909. Hitchcock, A. S. & A. Chase. Directions for preparing herbarium specimens of grasses. U. S. Dept. Agric. Bureau Plant Industry Circular No. 442. 4 pp.
- 1910. Hitchcock, A. S. & A. Chase. The North American species of *Panicum*. Contr. U. S.

- Natl. Herb. 15: 1-396.
- 1913. Hitchcock, A. S. Mexican grasses in the United States National Herbarium. Contr. U. S. Natl. Herb. 17(3): 181-389.
- 1914. Hitchcock, A. S. A text-book of grasses with special reference to the economic species of the United States. Macmillan Co. New York, NY. 276 pp.
- 1915. Hitchcock, A. S. & A. Chase. Tropical North American species of *Panicum*. Contr. U. S. Natl. Herb. 17(6): 459-539.
- 1915. Hitchcock, A. S. New or noteworthy grasses. American J. Bot. 2: 299-310.
- 1915. Nash, G. V. & A. S. Hitchcock. Poaceae. North American Flora 17(3): 197-288.
- 1917. Hitchcock, A. S. & A. Chase. Grasses of the West Indies. Contr. U. S. Natl. Herb. 18(7): 261-471.
- 1918. Hitchcock, A. S. Generic types with special reference to the grasses of the United States. American J. Bot. 5: 248-253.
- 1919. Hitchcock, A. S. & P. C. Standley. Flora of the District of Columbia and vicinity. Contr. U. S. Natl. Herb. 21: 1-329.
- 1919. Hitchcock, A. S. A botanical trip to Mexico. Sci. Monthly 8: 129-145; 216-238.
- 1920. Hitchcock, A. S. The genera of grasses of the United States with special reference to the economic species. U. S. Dept. Agric. Bull. No. 772. 307 pp.
- 1920. Hitchcock, A. S. North American species of *Ichnanthus*. Contr. U. S. Natl. Herb. 22(1): 1-11 + 9 plates.
- 1920. Hitchcock, A. S. North American species of *Lasiacis*. Contr. U. S. Natl. Herb. 22(1): 13-31 + 15 plates.
- 1920. Hitchcock, A. S. North American species of *Isachne*. Contr. U. S. Natl. Herb. 22(3): 115-121 + 8 plates.
- 1920. Hitchcock, A. S. North American species of *Oplismenus*. Contr. U. S. Natl. Herb. 22(3): 123-132.
- 1920. Hitchcock, A. S. North American species of *Echinochloa*. Contr. U. S. Natl. Herb. 22(3): 133-153.
- 1920. Hitchcock, A. S. North American species of

- Chaetochloa. Contr. U. S. Natl. Herb. 22(3): 155-208.
- 1921. Hitchcock, A. S. A manual of farm grasses. Publ. by the author. Washington, D. C. 175 pp.
- 1922. Hitchcock, A. S. Grasses of British Guiana. Contr. U. S. Natl. Herb. 22(6): 439-515.
- 1922. Hitchcock, A. S. The grasses of Hawaii. Mem. Bernice P. Bishop Mus. 8(3): 101-230.
- 1923. Hitchcock, A. S. Poaceae. <u>In</u>, Abrams, L. R. An illustrated flora of the Pacific states. 1: 103-255.
- 1923. Hitchcock, A. S. *Dissanthelium*, an American genus of grasses. J. Washington Acad. Sci. 13: 223-225.
- 1924. Hitchcock, A. S. The North American species of *Aristida*. Contr. U. S. Natl. Herb. 22(7): 517-586.
- 1925. Hitchcock, A. S. The North American species of *Stipa*. Contr. U. S. Natl. Herb. 24(7): 215-262.
- 1925. Hitchcock, A. S. Synopsis of the South American species of *Stipa*. Contr. U. S. Natl. Herb. 24(7): 215-289.
- 1925. Hitchcock, A. S. Gramineae. <u>In</u>, Jepson, W. L. A manual of the flowering plants of California. Revised by A. Chase. Univ. California Press. Berkeley. Pp. 72-144.
- 1925. Hitchcock, A. S. Methods of descriptive systematic botany. John Wiley & Sons. New York, NY. 216 pp.
- 1927. Hitchcock, A. S. The grasses of Ecuador, Peru and Bolivia. Contr. U. S. Natl. Herb. 24(8): 291-556.
- 1927. Hitchcock, A. S. New species of grasses from Central America. Proc. Biol. Soc. Washington 40: 82, 83.
- 1927. Hitchcock, A. S. Conservation of the names of the grass genera. American J. Bot. 14: 526-531.
- 1927. Hitchcock, A. S. The validity of the grass genus *Digitaria*. Rhodora 29: 114-116.
- 1928. Hitchcock, A. S. New species of grasses from the United States. Proc. Biol. Soc. Washington 41: 157-164.
- 1929. Hitchcock, A. S. Grasses of Canton and

- vicinity. Lingman Sci. J. 7: 177-265.
- 1929. Hitchcock, A. S. The relation of grasses to man. South Africa J. Sci. 26: 133-138.
- 1930. Hitchcock, A. S. The grasses of Central America. Contr. U. S. Natl. Herb. 24(9): 557-762.
- 1930. Hitchcock, A. S. Four new grasses. J. Washington Acad. Sci. 20: 381-384.
- 1930. Hitchcock, A. S. Fifteen new species of grasses, six from Africa, nine from China. Proc. Biol. Soc. Washington 43: 89-96.
- 1931. Hitchcock, A. S. Poaceae. North America Flora 17(4): 289-354.
- 1932. Hitchcock, A. S. The grasses of the Muhlenberg herbarium. Bartonia 14: 27-52.
- 1934. Hitchcock, A. S. Location of type specimens. Mimeographed. Smithsonian Inst. Washington, D. C. 19 pp.
- 1935. Hitchcock, A. S. Grasses, what they are and where they live. Ann. Rep. Smithsonian Inst. 1934: 297-312.
- 1935. Hitchcock, A. S. Poaceae. North American Flora 17(5): 355-418; 17(6): 419-482.
- 1935. Hitchcock, A. S. Location of type specimens. Mimeographed. Smithsonian Inst. Washington, D. C. 28 pp.
- 1935. Hitchcock, A. S. Manual of the grasses of the United States. U. S. Dept. Agric. Misc. Publ. No. 200. 1040 pp.
- 1935. Hitchcock, A. S. Gramineae. <u>In</u>, Christophersen, E. Flowering plants of Samoa. Bull. 128 Bernice P. Bishop Museum. Honolulu, HI. Pp. 6-15.
- 1936. Hitchcock, A. S. The genera of grasses of the United States with special reference to the economic species. Revised edition by A. Chase. U. S. Dept. Agric. Bull. No. 772. 302 pp.
- 1936. Hitchcock, A. S. Manual of the grasses of the West Indies. U. S. Dept. Agric. Misc. Publ. No. 243. 439 pp.
- 1937. Hitchcock, A. S. Poaceae. North American Flora 17(7): 483-542.
- 1939. Hitchcock, A. S., J. R. Swallen, & A. Chase. Poaceae. North American Flora 17(8): 543-638.

1951. Hitchcock, A. S. Manual of the grasses of the United States. Revised edition by A. Chase. U. S. Dept. Agric. Misc. Publ. No. 200. 1051 pp.

MARY AGNES CHASE

- 1903. Chase, A. & Millspaugh, C. F. Plantae Yucatanae. Field Mus. Bot. 3: 15-84.
- 1904. Chase, A. & Millspaugh, C. F. Plantae Yucatanae. Field Mus. Bot. 3: 85-151.
- 1904. Chase, A. The North American allies of *Scirpus lacustris*. Rhodora 6: 65-71.
- 1906. Chase, A. Notes on genera of Paniceae. I. Proc. Biol. Soc. Washington 19: 183-192.
- 1908. Chase, A. Notes on genera of Paniceae. II. Proc. Biol. Soc. Washington 21: 1-10.
- 1908. Chase. A. Notes on genera of Paniceae. II. Proc. Biol. Soc. Washington 21: 175-118.
- 1908. Chase, A. Notes on cleistogamy in grasses. Bot. Gaz. 45: 135, 136.
- 1908. Chase, A. Text-figures in Gray's new manual of botany. Rhodora 10: 207, 208.
- 1910. Hitchcock, A. S. & A. Chase. The North American species of *Panicum*. Contr. U. S. Natl. Herb. 17(6): 1-396.
- 1911. Chase, A. The subterranean organs of *Cinna arundinacea*. Rhodora 13: 9, 10.
- 1911. Chase, A. Notes on genera of Paniceae. IV. Proc. Biol. Soc. Washington 24: 103-160.
- 1911. Chase, A. Ornithological observations on cleisto-gamy. Rhodora 13: 76.
- 1911. Chase, A. Amateur botanical illustrating. Rhodora 13: 93-95.
- 1914. Chase. A. An unwelcome invader. *Bromus villosus* Forsk. (*Bromus maximus* Desf.) Rhodora 16: 166.
- 1914. Chase, A. Field notes on the climbing bamboos of Porto Rico. Bot. Gaz. 58: 277-279.
- 1915. Hitchcock, A. S. & A. Chase. The tropical North American species of *Panicum*. Contr. U. S. Natl. Herb. 17(6): 459-539.
- 1916. Chase, A. The structure of the spikelet of *Aphanelytrum.* Bot. Gaz. 61: 340-343.
- 1917. Hitchcock, A. S. & A. Chase. Grasses of the

- West Indies. Contr. U. S. Natl. Herb. 18(7): 261-471.
- 1917. Chase, A. Rev. E. J. Hill. Rhodora 19: 61-69.
- 1918. Chase, A. Axillary cleistogenes in some American grasses. American J. Bot. 5: 256-258.
- 1919. Chase, A. Some causes of confusion in plant names. J. Forestry 17(2): 159-162.
- 1920. Chase, A. The North American species of *Brachiaria*. Contr. U. S. Natl. Herb. 22(1): 33-44.
- 1920. Chase, A. The North American species of *Cenchrus*. Contr. U. S. Natl. Herb. 22(1): 45-77.
- 1921. Chase, A. The North American species of *Pennisetum*. Contr. U. S. Natl. Herb. 22(4): 209-234.
- 1921. Chase, A. The Linnaean concept of pearl millet.American J. Bot. 8: 41-49.
- 1921. Chase, A. Gramineae. <u>In</u>, Standley, P. C. Flora of Glacier National Park. Contr. U. S. Natl. Herb. 22(5): 280-294.
- 1922. Chase, A. First book of grasses: the structure of grasses explained for beginners. Macmillan. New York, NY. 121 pp.
- 1923. Chase, A. Visit to European herbaria. Smithsonian Misc. Coll. 74: 80-82.
- 1923. Chase, A. The identification of Raddi's grasses. J. Washington Acad. Sci. 13: 167-179.
- 1923. Chase, A. *Pennisetum vulcanicum* Chase. J. Washington Acad. Sci. 13: 363.
- 1923. Chase, A. *Paspalum botteri, Syntherisma fiebrigii* <u>In</u>, Standley, P. C. New species of plants of Salvador. J. Washington Acad. Sci. 13: 436.
- 1924. Chase, A. A new species of *Panicum* found in alfalfa seed. J. Washington Acad. Sci. 14: 343-345.
- 1924. Chase, A. *Actachne*, a cleistogamous grass of the high Andes. J. Washington Acad. Sci. 14: 364-366.
- 1925. Chase, A. Collecting grasses in Brazil. J. New York Bot. Gard. 26: 196-198.
- 1925. Chase, A. 1925. <u>In</u>, Niles, C. D. A bibliographic study of Beauvois' agrostographie. Contr. U.

- S. Natl. Herb. 24: 135-214.
- 1925. Chase, A. Poaceae. <u>In</u>, Tidestrom, I. Flora of Utah and Nevada. Contr. U. S. Natl. Herb. 25: 61-98.
- 1926. Chase, A. Botanical expedition to Brazil. Explorations and field-work of the Smithsonian Institution in 1925. Smithsonian Misc. Coll. 78. Pp. 48-54.
- 1927. Chase, A. New grasses from Panama. J. Washington Acad. Sci. 17: 142-147.
- 1927. Chase, A. Eastern Brazil through an agrostologist's spectacles. Smithsonian Rep. 1926: 383-403.
- 1927. Chase, A. *Calamagrostis leonardi* and *Leptochloa monticola*. <u>In</u>, Leonard, E. C. Fourteen new species of plants from Hispaniola. J. Washington Acad. Sci. 17: 72, 73.
- 1927. Chase, A. & A. W. Sampson. Range grasses of California. Univ. California Exp. Stat. Bull. No. 430. 94 pp.
- 1928. Chase, A. Native ornamental grasses. Natl. Hort. Mag. 7: 9-11.
- 1929. Chase, A. The North American species of *Paspalum*. Contr. U. S. Natl. Herb. 28: 1-310.
- 1931. Chase, A. [Description of] *Paspalum longum.*<u>In</u>, Johnston, I. M. Flora of the Revillagigedo
 Islands. Proc. California Acad. Sci. IV. 20: 52,
 53.
- 1933. Chase, A. A new grass of Texas. J. Washington Acad. Sci. 23: 137, 138.
- 1935. Chase, A. Studies in the Gramineae of Brazil. J. Washington Acad. Sci. 25: 187-190.
- 1935. Chase, A. [Description of] Paspalum redundans, P. galagageium, and P. galagageium var. minoratum. In, Hitchcock, A. S. New species of grasses from the Galapagos and the Revillagigedao Islands. Proc. California Acad. Sci. IV. 21: 297, 299, 300.
- 1937. Chase, A. Notes on types of North American grasses. American J. Bot. 24: 33-35.
- 1937. Chase, A. First book of grasses. Second edition. W. A. Silveus. San Antonio, TX. 125 pp.
- 1937. Chase, A. *Arthraxon hispidus* var. *cryptatherus* (Hack.) Honda in Pennsylvania. Rhodora 39:

100.

- 1937. Chase, A. New species of *Paspalum* from tropical America. J. Washington Acad. Sci. 27: 143-146.
- 1938. Chase, A. The carpet grasses. J. Washington Acad. Sci. 28: 178-182.
- 1938. Chase, A. New grasses from Oregon. J. Washington Acad. Sci. 28: 51-55.
- 1939. Hitchcock, A. S., J. R. Swallen, & A. Chase. Poaceae. North American Flora 17(8): 543-638.
- 1939. Chase, A. Papuan grasses collected by L. J. Brass. J. Arnold Arboretum 20: 304-316.
- 1939. Chase, A. Genera 130-136 of Poaceae. North American Flora 17: 568-579.
- 1939. Chase, A. *Paspalum setiglume*. <u>In</u>, Gleason, H. A. & E. P. Killip. Flora of Mount Auyan-Tepui. Brittonia 3: 150-152.
- 1940. Chase, A. Report to the Minister of Agriculture of Venezuela. Forage Crop Gaz. 4: 7-10.
- 1940. Chase, A. Studying the grasses of Venezuela. Explor. Field-Work Smithsonian Inst. 1940: 61-66.
- 1943. Chase, A. Papuan grasses collected by L. J. Brass. 2. J. Arnold Arboretum 24: 77-89.
- 1943. Chase, A. New grasses from South America. J. Washington Acad. Sci. 33: 316, 317.
- 1944. Chase, A. Grasses of Brazil and Venezuela. Agriculture in the Americas 4: 123-126.
- 1946. Chase, A. *Enneapogon desvauxii* and *Pappophorum wrightii*, an agrostological detective story. Madroño 8: 187-189.
- 1950. Chase, A. *Pogonatherum* Beauv. J. Arnold Arboretum 31: 130-132.
- 1951. Chase, A. Manual of the grasses of the United States. Revised edition. Misc. Publ. No. 200. U. S. Dept. of Agric. Washington, D. C.
- 1951. Sampson, A. W., D. W. Hedrick, & A. Chase. California grasslands and range forage grasses. Univ. California Exp. Stat. Report No. 724. Pp. 5-131.
- 1952. Chase, A. New species of grasses from Venezuela. J. Washington Acad. Sci. 42: 122-124.

- 1954. Chase, A. First book of grasses. Third edition. Smithsonian Inst. Washington, D. C. 127 pp.
- 1962. Chase, A. & C. Niles. Index to grass species. Three vols. G. K. Hall. Boston, MA.

SELECTED REFERENCES

Chase, A. 1936. Obituary. Albert Spear Hitchcock. Science 83: 222-224.

Fosberg, F. R. & J. R. Swallen. 1959. Agnes Chase. Taxon 8(5): 145-151.

Henson, P. M. 1996. [Agnes Chase]. \underline{In} , Clark, L. G. & R. W. Pohl. Agnes Chase's first book of grasses. Fourth edition. Smithsonian Inst. Washington, D. C. Pp. xiii-xviii.

Isely, D. 1994. Albert Spear Hitchcock (1865-1935). One hundred and one botanists. Iowa State Univ. Press. Ames. Pp. 296-298.

Isely, D. 1994. Mary Agnes Chase (1869-1963). One hundred and one botanists. Iowa State Univ. Press. Ames. Pp. 303-305.

Stieber, M. T. 1980. Chase, Mary Agnes. <u>In</u>, Notable American women: the modern period. Belknap Press. Harvard Univ. Cambridge, MA. Pp. 146-148.

6.04 - EVOLUTION OF GRASSES

"In my opinion, the climax of flowering-plant evolution is represented by the grasses..(G. Ledyard Stebbins)

EVOLUTIONARY TRENDS IN THE GRASSES*

VEGETATIVE FEATURES

Habit: perennial →	annual
Elongate rhizome: absent →	present
Seedling leaves: short →	elongate
Ligule: membranous →	hairs
Pseudopetiole: absent →	present
Bicellular hairs: present →	absent
Silica cells: quadrate/elongate →	dumbbell-shaped
Stomates: lozenge-shaped →	oblong
Kranz anatomy: absent →	present

SPIKELET AND INFLORESCENCE

Panicle branches: single → pairs, trios, clusters Peduncle: well developed → short/absent Florets per spikelet: several → one Disarticulation below glumes: no → yes Glumes: awnless → awned Glumes: shorter or equal to → longer than lemmas Sterile basal lemmas: absent → present Lemma callus: blunt → elongate, pointed Lemma texture: similar → dissimilar Lemmas: awned → awnless Awn: straight, single → bent, twisted or trifid Awn: terminal → dorsal or basal Lemma apex: tapering → notched, bilobed, toothed Lemma veins: convergent → parallel

FLORET AND CARYOPSIS

Florets: bisexual/mixed →	unisexual
Palea: 2-partite →	1-partite
Lodicule number: 3 →	2
Lodicules: vascular →	nonvascular
Lodicule apex: thin →	thick, truncate
Stamen number: six →	three
Style branches: three →	two
Stigmas: elevated →	sessile
Embryo/caryopsis ratio: <1/3 →	>1/3
Embryo internode: short →	elongate/absent
Starch grains: compound →	simple

NUMBER OF ADVANCED CHARACTER STATES IN REPRESENTATIVE GRASSES

Bambuseae	Bambusa	02
Streptochaeteae	Streptochaeta	04

Genus

Tribe

Oryzeae Ampelodesmeae	Oryza Ampelodesmos	09 11
Eragrosteae	Eragrostis	11
Ehrharteae	Ehrharta	11
Danthonieae	Danthonia	12
Poeae	Festuca	12
Arundineae	Arundo	13
Chlorideae	Chloris	14
Olyreae	Olyra	16
Stipeae	Stipa	16
Triticeae	Hordeum	17
Paniceae	Panicum	17
Aveneae	Avena	18
Agrostideae	Agrostis	18
Andropogoneae	Schizachyrium	19

*From Stebbins, G. L. Major trends of evolution in the Poaceae and their possible significance. <u>In</u>, Estes, J. R., R. J. Tyrl, & J. N. Brunken (editors). Grasses and grasslands. Univ. Oklahoma Press. Norman. Pp. 3-36.

MAJOR CLADES

Evidence is presented to support the recognition of several major clades within the family:

- Streptochaeta & Phareae (early-diverging lineage)
- PACC clade (Panicoideae, Arundinoideae, Chlorid-oideae, Centothecoideae)
- BOP clade (Bambusoideae, Oryzoideae, Pooideae)

[Source: Grass Phylogeny Working Group, 2000]

SELECTED REFERENCES

Anderson, E. & G. L. Stebbins. 1954. Hybridization as an evolutionary stimulus. Evolution 8: 378-388.

Bessey, E. A. 1917. Phylogeny of the grasses. Ann. Rep. Michigan Acad. Sci. 19: 239-245.

Catalán, P., E. A. Kellogg, & R. G. Olmstead. 1997. Phylo-geny of Poaceae subfamily Pooideae based on chloroplast *ndhF* gene sequences. Mol. Phylo. Evol. 8: 150-166.

Chapman, G. P. (editor). 1992. Grass evolution and domestication. Cambridge Univ. Press. New York, NY. 352 pp.

Advanced States

Clayton, W. D. 1981. Evolution and distribution of grasses. Ann. Missouri Bot. Gard. 68: 5-14.

Clifford, H. T. 1969. Attribute correlations in the Poaceae (grasses). Bot. J. Linnean Soc. 62: 59-67.

Coughenour, M. B. 1985. Graminoid responses to grazing by large herbivores: adaptations, exaptations, and interacting processes. Ann. Missouri Bot. Gard. 72(4): 852-863.

Davis, J. I. & R. J. Soreng. 1993. Phylogenetic structure in the grass family (Poaceae) as inferred from chloroplast DNA restriction site variation. American J. Bot. 81: 1444-1454.

Devos, K. M. & M. D. Gale. 2000. Genome relationships: the grass model in current research. Plant Cell 12: 637-646.

De Wet, J. M. J. 1987. Hybridization and polyploidy in the Poaceae. <u>In</u>, Soderstrom, T. R. et al. (editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 188-194.

Duvall, M. R. & B. R. Morton. 1996. Molecular phylogenetics of Poaceae: an expanded analysis of *rbcL* sequence data. Mol. Phylo. Evol. 5: 352-358.

Grass Phylogeny Working Group. 2000. A phylogeny of the grass family (Poaceae), as inferred from eight character sets. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publ. Collingwood, Australia. Pp. 3-7.

Hilu, K. W. 1985. Trends of variation and systematics of Poaceae. Taxon 34(1): 102-114.

Hilu, K. W. 1987. Chloroplast DNA in the systematics and evolution of the Poaceae. <u>In</u>, Soderstrom, T. R. et al. (editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 65-72.

Hsiao, C., S. W. L. Jacobs, N. J. Chatterton, & K. H. Asay. 1999. A molecular phylogeny of the grass family (Poaceae) based on the sequences of nuclear ribosomal DNA (ITS). Australian Syst. Bot. 11(5/6): 667-688

Hunziker, H. H. & G. L. Stebbins. 1987. Chromosomal evolution in the Gramineae. <u>In</u>, Soderstrom, T. R. et al. (editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 179-187.

Jacobs, S. W. L. & J. Everett (editors). 2000. Grasses: systematics and evolution. CSIRO Publ. Collingwood, Australia. 406 pp.

Kahler, A. L. & S. C. Price. 1987. Isozymes in population genetics, systematics, and evolution of grasses. <u>In</u>, Soder-strom, T. R. et al. (editors). Grass systematics and evolution. Smithsonian Inst. Press.

Washington, D. C. Pp. 97-106.

Kellogg, E. A. & H. P. Linder. 1995. Phylogeny of the Paoles. <u>In</u>, Rudall, P. J. et al. (editors). Monocotyledons: systematics and evolution. Royal Botanic Gard. Kew, England. Pp. 511-542.

Renvoize, S. A. & W. D. Clayton. 1992. Classification and evolution of the grasses. <u>In</u>, Chapman, G. P. (editor). Grass evolution and domestication. Cambridge Univ. Press. Cambridge, England. Pp. 3-37.

Sharma, M. L. 1979. Some considerations on the phylogeny and chromosomal evolution in grasses. Cytologia 44: 679-685.

Smith, D. B. & R. B. Flavell. 1974. The relatedness and evolution of repeated nucleotide sequences in the genomes of some Gramineae species. Biochem. Gen. 12: 243-256.

Soreng, R. J. & J. I. Davis. 1998. Phylogenetics and character evolution in the grass family (Poaceae): simultaneous analysis of morphological and chloroplast DNA restriction site character setes. Bot. Rev. 64(1): 1-85.

Stebbins, G. L. 1949. The evolutionary significance of natural and artificial polyploids in the family Gramineae. Proc. 8th Inter. Congr. Genet., 1948. Pp. 461-485.

Stebbins, G. L. 1956. Taxonomy and the evolution of genera, with special reference to the family Gramineae. Evol. 10: 235-245.

Stebbins, G. L. 1956. Cytogenetics and evolution of grasses. American J. Bot. 43: 890-906.

Stebbins, G. L. 1972. The evolution of the grass family. <u>In</u>, Younger & McKell. Pp. 1-17.

Stebbins, G. L. 1975. The role of polyploid complexes in the evolution of North American grasslands. Taxon 24: 67-80.

Stebbins, G. L. 1981. Coevolution of grasses and herbivores. Ann. Missouri Bot. Gard. 68: 75-86.

Stebbins, G. L. 1985. Polyploidy, hybridization and the invasion of new habitats. Ann. Missouri Bot. Gard 72: 824-382.

Stebbins, G. L. 1987. Grass systematics and evolution: past, present and future. <u>In</u>, Soderstrom, T. R. et al. (editors). Pp. 359-367.

Tsvelev, N. N. 1969. "Some questions about the evolution of grasses." Bot. Zhurn. 54: 361-373. [In Russian].

6.05 - DOMESTICATION OF GRASSES

"History celebrates the battlefields whereon we meet our death, but scorns to speak of the plowed fields whereby we thrive; it knows the names of the King's bastards, but cannot tell us the origin of wheat. That is the way of human folly." (Jean Henri Fabre)

"The value of the grasses to the human race is incalculable; they affect and support virtually every facet of human existence." (Richard W. Pohl, 1987)

The great civilizations, both past and present, have been based upon agriculture. These agricultural systems, in turn, have been founded upon a handful of cereals or grains. The great civilizations of the Near and Middle East, notably those of Greece, Rome, and Egypt, were based primarily on wheat; as were those of Europe and later North America. The welldeveloped agriculture of the Maya, Aztecs, and Incas rested on maize. The great societies of China, India, and the Far East were based upon rice. While all of the great civilizations cultivated many different kinds of plants for a variety of purposes, it is almost impossible to overestimate the importance of the cereals. We devote 70% of our farmland to growing cereals and we derive about 50% of our calories from them. As a group they are, without question, the most important source of our food and they have been throughout our entire cultural history.

All of the **true cereals** belong to Gramineae. It is common to recognize maize (corn), rice, and wheat as the major cereals. Barley, rye, and oats are the best known of the minor cereals. In addition to the true cereals is an artificial group of plants called the **false cereals**. They are characterized by small, grain-like fruits. Sunflower and buckwheat "seeds" are perhaps the best known examples.

There are several features of cereals that make them useful to us. They are annuals, which means that we can rely on getting a crop in a relatively short time. They are also adaptable and efficient producers of food. Cereals are very nutritious. Grains can be easily harvested, cleaned, and processed.

The most important part of the cereal plant is its fruit, the seed-like **caryopsis**. It is more commonly known as a **grain** or a **berry**. It contains a single seed whose outer coat is fused to the inner wall of the fruit. The outer layer of the grain (ovary wall and seed coat) are often called **bran**; the embryo within the grain is the **germ**.

THE PROCESS OF DOMESTICATION

The monumental event that is often called the single most significant occurrence in human cultural evolution happened a scant 10,000 years ago. That

event is the domestication of plants and animals. To emphasize how recent this was in the scheme of things, I will switch time scales. Assume that the entire history of the universe can be collapsed into a single year.

Jan. 01: Creation of the Universe
Sep. 25: Origin of life on earth
Dec. 20: Plants colonize the land
Dec. 28: First flowering plants appear
Dec. 31: 10:30 p.m. - First humans
Dec. 31: 11:00 p.m. - Use of tools

Dec. 31: 11:59 p.m. - Domestication

(agriculture)

What do we mean by cultivation and domestication of plants and what are the processes involved? To **cultivate** means to care for a plant; to till the soil, water, weed, and prune. To **domesticate** means to bring into the household; to alter, especially genetically from the wild state. We have domesticated a hundred or so plants and 50 or so animals, such as the dog, pig, cattle, horse, water buffalo, goat, sheep, and chicken.

Domestication is really directed evolution, which in turn is based upon two basic phenomena: **variation**, the concept that not all individuals are the same and that some are better adapted for survival than others, and **natural selection**, the view that nature selects for those individuals that are best adapted to reproduce the species. Natural selection has been largely replaced by **artificial selection** -- by people selecting those individuals that we want to preserve. This has been done consciously and unconsciously.

Domestication involves three important steps:

- moving seeds, grains, etc. from their native habitats and planting them in new areas;
- removing selective pressures and thereby allowing more variants to survive; and
- selecting for characteristics that are useful to us, but not necessarily for the plant under its natural conditions.

THE CHANGES

Some changes in plants that have occurred as a result of domestication include:

- spread into a greater diversity of environments and a wider geographic range;
- flowering and fruiting simultaneously;
- reduction or loss of dispersal mechanisms;
- conversion from perennials to annuals;
- absence of normal pollinators;
- loss of defense mechanisms (thorns, awns, etc.);

- increased palatability;
- development of seedless fruits;
- reproduction by vegetative means;
- increase or decrease in plant size;
- change in chromosome number;
- increased susceptibility to disease;
- loss of seed dormancy;
- ⋄ loss of photoperiod controls;
- change from self-incompatability to self-compatability
- conversion of flower parts from one series to another.

WHERE DID IT OCCUR?

The Near East Complex

Avena sativa oats
Hordeum vulgare barley
Secale cereale rye
Triticum aestivum bread wheat
Triticum monococcum einkorn wheat
Triticum turgidum emmer wheat

The Asian Complex

Brachiaria ramosa anda horra Coix lacryma-jobi Job's tears Digitaria cruciata raishan Digitaria sanguinalis manna Echinochloa colona shama Echinochloa frumentacea Japanese millet Orvza sativa rice Proso millet Panicum miliaceum Panicum sumatrense sawan Paspalum scrobiculatum khodo millet Setaria glauca korali Setaria italica foxtail millet

The African Complex

Brachiaria deflexa animal fonio Digitaria exilis fonio Digitaria iburua black fonio Eleusine coracana finger millet Eragrostis tef teff Oryza glaberrima African rice Pennisetum americanum pearl millet Sorghum bicolor sorghum

The New World Complex

Bromus mango
Bromus unioloides
Panicum sonorum
Setaria geniculata
Zea mays
mango
tuca
sauwi
brittle grass
maize (corn)

[After deWet, J. M. J., 1981]

WHEN WERE THEY DOMESTICATED?

9000	Barley
9000	Emmer wheat
7500	Rice
7500	Rye
7000	Einkorn wheat
7000	Sugar cane
7000	Durum wheat
6000	Bread wheat
6000	Finger millet
5500	Maize
5500	Foxtail millet
4500	Sorghum
1500	African rice
1000	Millets
1000	Oats
1000	Maize (larger ears)
1972	Wild rice

Why Did It Take So Long?

The overriding question about the domestication of plants is why did it take so long for us to make so simple a "discovery" or to take this step. A number of theories have been put forth:

- While we lived by hunting, fishing, and gathering we had too little time for such cultural luxuries.
- Domestication became a necessity after dramatic shifts in climate.
- ☼ For thousands of years, we would be satisfied just to meet our basic needs for food, shelter, and clothing. Domestication occurred as the culmination of an ever increasing differentiation and specialization of human communities.
- Some plants and animals may have been domesticated as parts of religious ceremonies.
- No particular motive or advance was required; only the revelation that seeds can be sown to produce plants when and where desired ("The Eureka! Model").
- There is no single explanation; all of them have contributed to our understanding of the problem ("The No-Model Model").

The Worst Mistake in History?

You should be aware that not everyone is convinced that the domestication of plants and animals has been such a fine thing. But certainly we are now better off than the people in the Middle Ages? The cavemen? The apes? Jared Diamond (1987) argues the following:

- We are now much more dependent upon a few high carbohydrate crops, such as rice and the potato.
- We are more susceptible to famine and crop failure.
- Studies show an increase in tooth enamel defects associated with malnutrition, an increase in irondeficiency anemia, an increase in bone lesions, and until recently a decrease in life expectancy.
- The population densities that are now possible with agriculture encourage the spread of parasites and infectious disease.
- Agriculture and led to deep class divisions and accentuated the inequality of the sexes.

SELECTED REFERENCES

Bates, L. S. & C. W. Deyoe. 1973. Wide hybridization and cereal improvement. Econ. Bot. 27: 401-412.

Bright, S. W. J. & P. R. Shewry. 1983. Improvement of protein quality in cereals. CRC Critical Rev. Plant Sci. 1(1): 49-93.

Chapman, G. P. (editor). 1992. Grass evolution and domestication. Cambridge Univ. Press. New York, NY. 352 pp.

Davies, M. S. & G. C. Hillman. 1992. Domestication of cereals. <u>In</u>, Chapman, G. P. (editor). Grass evolution and domestication. Cambridge Univ. Press. Cambridge, England. Pp. 199-224.

Harlan, J. R. 1992. Origins and processes of domestication. <u>In</u>, Chapman, G. P. (editor). Grass evolution and domestication. Cambridge Univ. Press. Cambridge, England. Pp. 159-175.

Harlan, J. R., J. M. J. deWet, & E. G. Price. 1973. Comparative evolution of cereals. Evolution 27: 311-325.

Henry, R. J. 1985. A comparison of the non-starch carbohydrates in cereal grains. J. Sci. Food Agric. 36(12): 1243-1253.

Hillman, G. C. & M. S. Davies. 1990. Domestication rates in wild-type wheats and barley under primitive cultivation. Biol. J. Linnean Soc. 39(1): 39-78.

Johnson, V. A., J. W. Schmidt, & P. J. Mattern. 1968. Cereal breeding for better protein impact. Econ. Bot. 22: 16-25.

Lazenby, A. 1975. The evolution of the temperate cereals. Austr. Field Crops 1: 1-36.

Lee, B. 1989. Cereal transformation. Plants Today

2(1): 9-11.

Leonard, W. H. & J. H. Martin. 1963. Cereal crops. Macmillan. New York, NY. 825 pp.

Paterson, A. H. et al. 1995. Convergent domestication of cereal crops by independent mutations at corresponding genetic loci. Science 269: 1714-1718.

Raloff, J. 1998. Can grain yields keep pace? Science News 152(7): 105, 106.

Sage, R. F. 1995. Was low atmospheric CO_2 during the Pleistocene a limiting factor for the origin of agriculture? Global Change Biol. 1: 93-100,

Scade, J. 1975. Cereals. Oxford Univ. Press. London, England. 70 pp.

Wet, J. M. J. de. 1975. Evolutionary dynamics of cereal domestication. Bull. Torrey Bot. Club 102: 307-312.

Wet, J. M. J. de. 1981. Species concepts and systematics of domesticated cereals. Kulturpflanze 29: 177-198.

Wet, J. M. J. de. 1992. The three phases of cereal domestication. <u>In</u>, Chapman, G. P. (editor). Grass evolution and domestication. Cambridge Univ. Press. Cambridge, England. Pp. 176-198.

Zohary, D. 1971. Origin of south-west Asiatic cereals: wheats, barley, oats and rye. <u>In</u>, Davis, P. H. et al. (editors). Plant life in southwest Asia. Pp. 235-263.

6.06 - THE MAJOR & MINOR CEREALS

First, a short detour into the field of economic botany. There is a convention of long standing that calls for recognizing two groups of cereals – true and false. **True cereals** are the grains derived from plants of Gramineae. All others, such as buckwheat, sunflower, grains-of-paradise, etc., are **false cereals** or **pseudocereals**. We will ignore the false cereals.

It is also customary to distinguish two groups of true cereals – major and minor. The **major cereals** are wheat, rice, and maize (corn). The **minor cereals** are all of the remaining true cereals such as barley, oats, and rye.

WHEAT

This is most widely cultivated crop plant and also one of our oldest. Archeological remains dating to 6700 B. P. have been found in Jarmo, Iraq. These were relatively primitive wheats. But, even the advanced bread wheat is known from 5000 B. C. from the Nile Valley. Wheat was brought to the New World by the Spanish in 1529. It has been in cultivated in the United States since about 1602.

There are about 14-16 commonly recognized species of wheat. They fall easily into three groups, differing in chromosome number and morphology. In Triticum, x = 7. Two of the wheats are diploid (2x = 14); eight are tetraploids (4x = 28); and six are hexaploids (6x = 28)= 42). A more detailed summary is presented below. The diploid and tetraploid wheats are of little economic importance; it is the hexaploids that we use. The evolution of these 6X wheats is a fascinating story of hybridization between primitive wheats and weedy relatives (goat grasses of the genus Aegilops), followed by what was described for many years as a "spontaneous doubling of chromosome number". It always seemed to me that there was something mystical in that phrase. Most researchers now believe that the change from diploid to tetraploid came about through the union of unreduced gametes. This process occurred without our assistance. We just took advantage of the results. I will go into this in more detail in lecture.

Today there are many different cultivars of wheat available. They are classified in several ways. The **winter wheats** are planted in the fall, remain dormant during the winter, and then mature in the early summer. Winter wheats are grown from Texas to South Dakota. **Spring wheat** is planted in the spring and matures that same summer. It is adapted for growing seasons as short as 90 days. Spring wheat is used in the northern regions of the U. S. and Canada.

Wheat is the most important cereal for bread making

because of the nature of the protein in its grains. Glutenin and gliadin are sticky proteins that can hold a paste or dough together in a mass. Together they form **gluten**. If a mixture of wheat flour and water is exposed to the air for any length of time, it will become infected by naturally occurring microorganisms, including yeasts. They produce gases as part of their life cycle. The proteins in wheat flour have the ability to trap these gas bubbles within the dough. The result is leavened bread.

WILD AND DOMESTICATED WHEATS

Ploidy: Scientific Name	Genome(s)
Diploids $[2n = 2x = 14]$	
Triticum boeticum (wild einkorn) Triticum monococcum (einkorn)	AA AA
Tetraploids $[2n = 4x = 28]$	
Triticum dicoccoides (wild emmer variticum dicoccon(emmer wheat) Triticum durum (durum or macaron Triticum turgidum (poulard or rivet Triticum polonicum (Polish wheat) Triticum carthlicum (Persian wheat Triticum timopheevii Triticum araraticum	AABB AABB AABB AABB
Hexaploids $[2n = 6x = 42]$	
Triticum spelta (spelt wheat) Triticum macha (macha wheat) Triticum vavilovii (Vavilov's wheat) Triticum compactum (club wheat) Triticum sphaerococcum (shot whe Triticum aestivum (bread wheat)	AABBDD

This summary is modified after Simmons, N. W. (editor). 1976. Evolution of crop plants. Longman. London, England. P. 121. The nomenclature for the various wheat species follows Terrell, et al. (1986).

EVOLUTION OF MODERN HEXAPLOID WHEATS

PHASE I: DIPLOID TO TETRAPLOID

Triticum boeoticum		Angilans spaltaides
(Wild einkorn wheat)		Aegilops speltoides (Goat grass)
(Wild ellikolli Wileat)		` ,
[2n = 2x = 14]	Χ	[2n = 2x = 14]
[Genome: AA]		[Genome: BB]
	∇	
	∇	
	∇	

Sterile F, Hybrid [2n = 2x = 14][Genomes: AB]

Unreduced Gametes ("Chromosome Doubling")

Triticum dicoccoides (Wild emmer wheat) [2n = 4x = 28]

[Genomes: AABB]

 ∇ ∇

Domestication

 ∇

Triticum dicoccum (Cultivated emmer wheat)

[2n = 4x = 28][Genomes: AABB]

PHASE II: TETRAPLOID TO HEXAPLOID

Triticum dicoccum (Cultivated emmer wheat)

[2n = 4x = 28]

[Genomes: AABB] Χ Aegilops squarrosa (Goat grass) [2n = 2x = 14][Genomes: DD]

Sterile Hybrid

[2n = 3x = 21][Genomes: ABD]

> ∇ ∇

Unreduced Gametes ("Chromosome Doubling")

> ∇ ∇

Triticum aestivum (Bread wheat)

[2n = 6x = 42]

[Genomes: AABBDD]

 ∇ ∇

Domestication

Hulled/Free-threshing cultivars

Recent domestication/genetic engineering

RICE

Rice is the principal food for about 60% of the world's population. It has been cultivated in southeast Asia for at least 5000 years. Literally thousands of cultivars have been developed, 8000 of them in India alone. Rice was introduced into America in 1647.

Unlike wheat, most kinds of rice are diploid (2n = 2x)= 24). It is usually grown in a swampy field known as a paddy. This helps to explain why so much rice is raised in the monsoon belt where heavy seasonal rainfall is used. In most instances, rice seeds are not planted directly in the paddies. Instead there are nurseries where seedlings are started and then transferred. The seedlings are planted in small bunches, each clump about 4-16" from the next one. Most cultivated strains require flooding, this being accomplished by taking advantage of the monsoons and by the skillful manipulation of dikes in the paddies.

At maturity, most rice plants are 4-6 ft. tall. In the Mekong Valley, some deep water varieties reach 20 ft. Once the plants have flowered, the water level is reduced and finally the supply is shut off entirely and the fields allowed to dry. When the plants begin to wither, it is time to harvest the crop. In the Old World, the harvesting and threshing processes are done by hand. In the U.S. and other technologically advanced countries, much of this is done by machine. In this country, Arkansas, Louisiana, and California are the main rice growing states.

THE PROCESS

Oryza rufipogon

Wild: Asia Perennial 2n = 2x = 24

Genome: AA

Oryza nivara

Wild: India, Asia, Oceania Annual

2n = 2x = 24Genome: AA

Domestication

Oryza sativa Cultivated: widespread Annual

> 2n = 2x = 24Genome: AA

We commonly recognize three types of rice based upon the length of the grain:

- long: tropical rices; not too soft nor starchy; grains 7-8 mm long, the length prized by the connoisseur
- medium: commonly grown in the U. S.; somewhat softer; grains averaging about 6.6 mm long
- short: grown in the more northern climates, often planted in Japan; even more starchy; grains averaging about 5.5 mm long

MAIZE

First, a word about the common name of *Zea mays*. In this country, we usually call this plant corn or Indian corn. Maize is a better common name (and a perfectly legitimate one) because corn is used by English-speaking peoples around the world for several other cereals.

There are three features of maize that make it different from wheat, rice, or any other cereal. It is the only important cereal that is native to the New World. Second, maize as we known it today is considerably different in appearance from its wild ancestors. The progenitors of the other cereals are basically the same in general appearance as their modern derivatives. Maize is strikingly distinct. And third, maize is unique among the major cereals in having separate male and female flowers borne on entirely different parts of the plant. The male flowers are found on the branches of the tassel, while the female flowers are clustered in the ear, the complex fruiting structure that bears an even number of rows of caryopses, or kernels as they are commonly called.

There are six main types of maize in use today:

- flint: kernel made of hard starch; in use by Native Americans at the time of Columbus; widely used in the northern corn belt;
- dent: kernel of hard starch, capped by soft starch that dries to leave a small depression in the top of the grain; economically the most important maize; much used in the corn belt;
- flour: kernel consists almost entirely of soft starch; used by the Native Americans of the Southwest and those in South America for hand grinding;
- sweet: kernels with high sugar content, consumed while immature; most widely grown for human consumption here and in Europe;
- pop: kernels lacking soft starch, cells burst upon heating because of high water content of central cells; related in flint corn;

pod: peculiar type with comparatively little economic importance; kernel enclosed by bracts; considered by some to be the ancestor of modern maize.

Maize has many uses. About 90% of the crop goes into livestock food. In this country, maize is relatively unimportant as a food for humans. This is not the case in many other areas of the world, especially Africa. It is inferior to wheat and to some other cereals in its protein content. This means that maize flour products are less tasty than those made from rye or wheat. Maize flour, however, has been the mainstay of many peoples in Central and South America. Other important products from maize include corn starch, corn oil, alcoholic beverages, and silage.

HYBRID MAIZE

One of the great developments in modern agricultural genetics is hybrid corn. The basic principle involved is that stable inbred lines can be crossed with one another to produce uniform plants with higher yields that combined the desirable features of the two parental lines. Modern hybrid corn involves a double crossing. During the first year, inbred line A is crossed with line B. Self-pollination is prevented during these crosses by removing the male flowers from one strain (detasseling), thereby rendering the plants effectively female. The manual emasculation of plants, once a common summer job for young people living in the Corn Belt, has all but disappeared because of another important advance -- genetically controlled male sterility in maize. In separate fields, lines C and D are similarly crossed. The seeds from the A x B and from the C x D crosses are planted. These mature into AB and CD individuals. These are then crossed during the second year, yielding the double cross ABCD hybrid seed. It is planted the third year to produce tremendous yields of high-quality seed. The ABCD seed is not true-breeding and must, therefore, be purchased regularly.

CYTOPLASMIC MALE STERILITY

In 1938, Paul Mangelsdorf, a Harvard botanist who devoted his life to the study of maize, discovered a sweet corn variety in Texas that was male sterile. The male flowers of its tassel had shriveled anthers that did not produce fertile pollen grains. Investigation of this plant revealed that the sterility was under genetic control, as opposed to some short-lived environmental problem, such as drought. Sterile sex cells typically result from chromosomal abnormalities, either in their number or structure. However, in this case the corn plant produced sterile pollen when a sterility factor [S] in the cytoplasm of the cell was present at the same time that it had a double recessive gene [rf] in its nucleus. This same kind of phenomenon was first found in onions, and is now known to occur in several crop plants. One possible explanation is that the cytoplasmic sterility is caused by viruses that can survive only if the rf rf condition exists. If the gene is

present in the Rf state, fertility is restored. Cytoplasm without the sterility factor is designated N, for "normal." The cytoplasmic factor passes from one generation to the next only via the egg. To summarize:

S rf rf = male sterile plants S Rf rf = male fertile plants S Rf Rf = male fertile plants N rf rf = male fertile plants N rf rf = male fertile plants

Therefore, by using an inbred line that contains the S rf rf genetic combination, male sterile plants are produced. The corn plants are rendered functionally female. The difficulty in finding enough workers and their cost made the male-sterile strains a very attractive alternative to manual detasseling. Within twenty years, practically all of the maize grown in the United States incorporated the male sterility factor first found in the Texas corn plants. Once again we had made one of our major crop plants more genetically similar to one another, with all of the advantages and disadvantages associated with that uniformity. The bill came due in the summer of 1970. Our corn fields were invaded by a fungus (Helminthosporium maydis), which causes the southern leaf blight. The disease spread rapidly, moving from Florida northward at about 150 km per day. By the end of the summer, the blight had covered much of the eastern and central United States. It devastated the Texas male-sterile hybrids, causing more than a \$1 billion loss in the corn crop. Today's maize cultivars are based upon normal cytoplasm and are detasseled by hand.

"JUMPING GENES"

James Watson, who shared the Nobel Prize with Francis Crick for their discovery of the structure of DNA, said, "There are really three main figures in the history of genetics -- the three M's: Mendel, Morgan, and McClintock." Gregor Johann Mendel (1822-1884), an Austrian monk, is often called the father of genetics. His work on the changes that he observed from one generation to the next in pea plants that he grew in the monastery garden is well known -- a standard fixture in all highschool and college texts in biology and genetics. Thomas Hunt Morgan (1866-1945), of Columbia University, along with his wife (Lillian) and his students did his research on the fruit fly (Drosophila melanogaster). His lab was the first to demonstrate that genes were located chromosomes in the cell nucleus, that genes were located at specific sites on a chromosome, and that traits were passed from parent to offspring through genes. Morgan won the Nobel Prize in 1933 for these fundamental discoveries.

The third "M" is Barbara McClintock (1902-1992). She earned her bachelor's degree in botany from Cornell University, where she was also awarded her master's and doctorate. In 1931, McClintock identified the ten

chromosomes of maize, and she co-authored with Harriet Creighton the first paper to describe the genetic phenomenon of crossing-over. In 1944, McClintock identified the seven chromosomes of the bread mold, *Neurospora*, and began her research on mobile genes and controlling elements in maize. She had observed that some plants have leaves with different patterns of pigment in them. In maize, some kernels were white, some solid purple, and some had speckles of purple on otherwise white kernels.

After years of detailed study, McClintock developed a theory to explain what she had seen. The differences in pigmentation of corn kernels was caused by some genes moving from one site on a chromosome to another location, or from one chromosome to another. Further, it appeared to her that other genes acted as switches that turn a gene on and off during plant development. McClintock presented the results of her work in 1951 at a Cold Spring Harbor Symposium. The reaction was mixed. Most of her colleagues failed to understand her work, others rejected it outright, and others thought that poor Barbara had been out in the sun too long playing her corn plants. One said, that "... he had never heard anything as ridiculous." Another, "I understand that you're doing something that's very strange. I don't want to hear a word about it." At the other end of the spectrum, the distinguished Caltech geneticist Alfred Sturtevant said, "I didn't understand one word she said, but if she says it is so, it must be so!"

What Barbara McClintock had proposed was heresy! Everyone knew that a chromosome was like a necklace and the beads were genes. This bead is always next to that bead in a necklace; this gene is always next to that gene on a chromosome. And she was saying that it ain't necessarily so. In her now classic paper, McClintock concluded that the best explanation for what she was seeing was that a gene did, in fact, actually move from one site on a chromosome to the site of the gene that controlled pigment color. She called it Ds, the dissociator gene. Ds would instruct the color gene. The Ds gene, in turn, was controlled by an activator, Ac.

McClintock called these mobile genetic units **transposons**. Time Magazine called them "jumping genes." It explained McClintock's theory in terms of three characters -- a painter, a boss, and a policeman. The painter is the structural gene that makes a kernel have a particular color. The boss (Ds or dissociator gene) can tell the painter to paint or not to paint. The boss must follow the directions of the police officer (Ac or activator gene), who can tell the boss to let the painter do his job or not. The officer can tell the boss to have the painter stop and then later resume painting. Depending on the interaction of the painter, boss, and policeman, the kernel will be pigmented, speckled, or colorless.

On 10 October 1983, McClintock learned from the radio that she had won the Nobel Prize in Physiology

or Medicine. The folks in Stockholm had tried to call her at home, but she didn't have a telephone. She, Marie Curie in 1911, and Dorothy Hodgkin in 1964 are the only three women to receive an unshared Nobel in any field.

McClintock's long-time friend and champion, Marcus Rhoades, said of her work:

"One of the remarkable things about Barbara McClintock's surpassingly beautiful investigations is that they came solely from her own labors. Without technical help of any kind she has by virtue of her boundless energy, her complete devotion to science, her originality and ingenuity, and her quick and high intelligence made a series of significant discoveries unparalleled in the history of cytogenetics. A skilled experimentalist, a master at interpreting cytological detail, a brilliant theoretician, she has had an illuminating and pervasive role in the development of cytology and genetics."

Transposable elements have since been found in many plants and animals. They are best known in maize, fruit flies, yeasts, and humans.

MAIZE RELATIVES

It is also important not to over emphasize the uniqueness and remoteness of maize. It does have close relatives, although you might not recognize them as such on casual inspection.

Gama grass (*Tripsacum* **ssp.).** There are about seven species of gama grasses found from the central United States to southern Brazil. All of them are perennials. The male and female flowers are separated from one another, but do not occur in the tassel/ear configuration in corn.

Teosinte. There are three kinds of teosinte, all occurring in Mexico and Central America. Traditionally, teosinte has been placed in its own genus (*Euchlaena*), but in more recent works the species have been put in *Zea*. The male flowers are borne in a tassel at the top of the plant; the female flowers are borne on a spike on the lower parts of the plant.

POSSIBLE ANCESTORS

A great deal of research and speculation has been focused on the ancestor or ancestors of modern maize. Here are the major players.

Corn Grass. This plant is an anomalous grass with numerous slender leaves, numerous tillers, and small ears. There is a long spathe, more characteristic of *Coix* than of maize. These differences are supposedly the result of a single dominant gene. The plant really does not look like maize, but it was once proposed as an ancestor. It has few, if any, proponents these days.

Teosinte. When first proposed by Ascherson, the

hypothesis was that maize was a domesticated form of teosinte. The theory was later modified by Harshberger and Collins to say that one parent of maize was teosinte, but that it had a second parent of unknown identity. George Beadle discovered that teosinte kernels will pop; a reason to preserve it as a food plant. The polystichous nature of the maize ear could be the result of fusion of teosinte spikes. The advocates of teosinte provided the only real challenge to the pod corn theory as the leading explanation of corn's ancestor.

Common Ancestry.According to this theory, maize originated from a perennial, wild, maize-like ancestor. This plant is now extinct. The pre-maize, in turn, had an ancestor in common with both teosinte and gama grass. That grandparent is also now extinct. This theory is largely untestable because the principal players are extinct. They cannot be tested or measured.

Pod Corn. This theory was developed about sixty years ago by Paul Mangelsdorf and R. G. Reeves, two of the great names in this area. Pod corn is still very much with us today. It is a peculiar, primitive looking maize with well-developed papery bracts surrounding the individual kernels. We grow it in this country as a curiosity; in South America it remains a food plant of limited importance. The pod corn theory has three basic premises:

- modern maize originated from a wild form of pod corn indigenous to the lowlands of Central or South America;
- *teosinte is the product of natural hybridization between maize and gama grass; and
- *many of the strains of maize that we see today are the result of past hybridization between *Zea* and *Tripsacum*.

NATURE OF EVIDENCE

Archeological/ethnobotanical remains. Although archeological remains of maize are fairly common in the New World, they have never been found in the Old World. This fact helps us to conclude that maize is indigenous to the New World. Several of the important digs are:

Bat Cave (New Mexico). The floor of this cave is littered with six feet of garbage, coprolites (fossil poop), debris, and tiny cobs of maize about 5600 years old. The cobs are 2-3 cm long, with kernels partly enclosed by bracts. Anatomically, it is a pod corn or a pop corn.

Swallow Cave (New Mexico). At the lowest levels are tiny cobs similar to those in the Bat Cave. The prehistoric context of the cobs indicates their great age.

Coxcatlán Cave (Tehuacán Valley of Mexico). The cave system consists of 28 superimposed levels. The upper 14 have well-preserved cobs. The cave was occupied from 10,000 to 2300 B. C. and then again from 900 B. C. to A. D. 1500. The cave faces a broad alluvial plain where wild and cultivated maize could have grown. The oldest remains bridge the gap between wild maize and the earliest stages of domesticated maize.

La Perra Cave (Tamaulipas, Mexico). The oldest remains are from about 2500 B. C. They show the signs of crossing with gama grass at the lower levels, but show "tripsacoid" features at the upper levels.

Purron Cave (Tehuacán Valley of Mexico). This cave was inhabited from about 200 B. C. to A. D. 1500. The surrounding soils ar fertile and there was an abundance of water.

Pollen. During the excavations required for a new office building in Mexico City, grass pollen was discovered in cores taken at a depth of about 70 meters. The pollen was about 80,000 years old. Detailed studies made by Elso Barghoorn, a paleobotanist at Harvard University, showed that the pollen was that of maize. Above the six meter level was copious pollen of modern, cultivated maize. Barghoorn's work again provides support for the New World origin of maize. The plants that produced this pollen were alive and well thousands of years before we migrated into the Valley of Mexico. Second, this discovery tended to support the pod corn theory of Mangelsdorf and Reeves. The ancestor of modern maize as a more primitive kind of maize.

Genetic. On the fourth longest chromosome of pod corn is a gene designated Tu. In pure pod corn the gene is present in the homozygous dominant state (TuTu). It is heterozygous in modern pod corn (Tutu) and it is homozygous recessive (tutu) in modern maize. The change from TuTu to tutu is associated with many changes in pod corn, including a number that have made the plant much more useful to us. These have also rendered it unable to live in the wild. The changes include:

- reduction in prominence of the tassel;
- increase in the development of the ear;
- changing the tassel from principally female to male;
- decrease in the length and weight of the bracts surround the kernels; and
- increase in the size and weight of the axis of the ear.

Walter Galinat (1983, 1985) reports that the gene Tr controls the two- versus four-ranked ears. The gene Pd controls single versus paired spikelets. Ab determines the presence or absence of abscission layers in the ear. The gene Tu codes for soft outer

glumes and a soft rachis.

Molecular. John Doebley of the Univ. of Minnesota has studied isozyme variation in maize and teosinte. He has looked at 13 enzyme systems encoded by 21 loci. If teosinte were the ancestor of maize, we would expect to find considerable similarity at the molecular level. His studies show that maize and teosinte are indistinguishable.

TEOSINTE VERSUS MAIZE

- In teosinte the female spikelets are solitary; they are paired in maize;
- In teosinte the central spike of the female inflorescence is two-ranked; it is many-ranked in maize;
- In teosinte the rachis shatters at maturity; it is non-shattering in maize;
- The caryopsis is encased in teosinte; naked in maize.

At first, the differences seem striking. As Paul Mangelsdorf noted, "If maize has originated from teosinte, it represents the widest departure of a cultivated plant from its wild ancestor which still comes within man's purview...

However, according to Doebley (1990), perhaps as few as five genes account for 50-80% of the differences between maize and teosinte. "If you took those five regions from maize and put them into teosinte, the thing you'd have in front of you would be called maize." He estimates that these dramatic changes could have occurred in less than 1000 years.

An alternative explanation, wonderfully entitled the "Cataclysmic Sexual Transmutation Theory," was advanced by Hugh Iltis of the Univ. of Wisconsin. He thinks of an ear of corn as "... that magnificently monstrous enigmatic anomaly...." His argument builds on the fact that the maize ear and the central spike of the maize tassel are homologous. Both are polystichous and both are governed by the same genes. The central spike of the tassel is often feminized into an ear by abnormal environments or by disease, such as corn smut. Iltis argues that the maize ear is a feminized tassel reduced to its terminal spike. Therefore, the teosinte tassel could be transformed into a maize ear in one giant step. This would explain the lack of connecting links in the archaeological record.

PUTTING IT ALL TOGETHER

The salient points appear to be:

Maize is native to the New World, probably Mexico or Central America.

- The direct ancestor of maize is teosinte.
- "... teosinte is not a hybrid of maize and *Tripsacum*. Perhaps I may be permitted to enjoy some degree of satisfaction in the fact that it is colleagues and not my critics who have shown that this part of our tripartite hypothesis is no longer tenable." (Paul Mangelsdorf, 1974)
- The transformation of teosinte into maize began about 8000 B. P.
- Archaeological evidence suggests that we first began to cultivate maize about 5200-3400 B. C.
- "The mystery of maize is not such a mystery after all, and the romance has been exaggerated." (Jack Harlan, 1992)
- Early Native Americans discovered that they could accelerate the forces of natural selection by physical separation of selected plant types.
- Use of irrigated garden plots or isolated plots meant that these forms would not be genetically swamped by wild populations.
- They selected the most useful variants out of large populations of teosinte.
- "... American Indians had already developed two or three hundred races of corn, essentially all we have today. Theirs was the most extraordinary achievement in plant breeding in all of man's existence, including his most recent history.... Perhaps most remarkable of all were the earliest steps when man (or, more likely, woman) first began to influence the evolution of what I believe to be the wild ancestor of corn." (George W. Beadle, 1972)

THE PROCESS

Mangelsdorf & Reeves

 $\begin{array}{c} \text{Pod corn} \\ \nabla \\ \text{Wild maize} \\ \nabla \\ \text{Domestication} \\ \nabla \\ \text{Modern maize} \end{array}$

 $\begin{array}{c} \text{Maize x grama grass} \\ \nabla \\ \text{Teosinte} \end{array}$

Beadle & Galinat

Teosinte ∇ Domestication ∇ Primitive maize ∇ Domestication ∇ Modern maize

THE MINOR CEREALS

BARLEY

Hordeum vulgare ranks number fourth in terms of world-wide annual production. Along with wheat, it was one of the first plants that we domesticated. All of the cultivated species are diploids (2n = 2x = 14). Barley differs from wheat, maize, and rice in having three spikelets per node. If all three develop, the spike has the appearance of having six rows of grains (the 6-rowed barleys); if the two lateral spikelets are rudimentary, then the spike appears to have two rows of spikelets (the 2-rowed barleys). The two bracts immediately surrounding the grain are fused to it. The grain is pearled, rubbed against abrasive disks to remove the hulls and some of the outer layers of the grain, during the processing for human consumption. The chief use of barley is as animal food. It is a relatively unimportant food for humans. About onethird of the crop is used for making malt used in brewing, flavoring, cereals, icings, coffee substitutes, infant foods, flours, medicinal syrups, candies, and industrial fermentations.

THE PROCESS

Hordeum spontaneum

Wild barley
Fertile Crescent ∇ Domestication ∇ Hordeum vulgare var. distichum

2-rowed barley ∇ Domestication ∇ Hordeum vulgare var. hexastichum

6-rowed barley

RYE

Secale cereale is a plant of cool, non-humid regions. It is grown chiefly in northern Europe. In the United States, North and South Dakota and Nebraska grow

the most rye. The species is diploid (2n = 2x = 14). It is now unknown in the wild. Most rye is fed to cattle. We use it to make flour for "rye bread" or the famous black bread (Schwartz brot) of Germany, Poland, and Russia. Most of our U. S. rye bread has a very high wheat flour content. Rye is also used to make whisky and industrial alcohol. Ergot (Claviceps purpurea) is an important fungal parasite of rye. It causes tremendous crop losses and poisoning in both cattle and humans. Consumption of contaminated grain over a period of time leads to a gangrenous loss of tissues because of constriction of blood vessels, especially in the extremities of the body. Larger doses can have pronounced effects on the central nervous system.

THE PROCESS

OATS

The origin of oats (*Avena* spp.) is obscure. There are few references to it in the ancient literature; none, for instance, in the Old Testament. It may have become domesticated in the cultivated fields of barley or of some other crop. It is now grown in temperate regions, chiefly Europe and North America.

As in wheat, there are diploid, tetraploid, and hexaploid oats. We use the hexaploids more than the others. The diploids include *A. brevis* (slender oats), *A. strigosa* (sand oats), *A. wiestii* (desert oats), and *A. nudibrevis*. The tetraploids are *A. barbata* (slender oats) and *A. abyssinica* (Abyssinian oats). The hexaploids include *A. fatua* var. *fatua* (wild oats), *A. fatua* var. *sativa* (cultivated oats), *A. sterilis* (wild red oats), *A. byzantina* (red oats), and *A. nuda* (naked oats).

Until quite recently, oats were not widely appreciated, even though they are very nutritious (protein content of 13.8%). Oats are used to make flour, rolled oats, and even as a beverage (avena). The crop is often rotated with corn. Iowa is the leading U. S. producer.

SORGHUM

The U. S. is the leading producer of sorghum (Sorghum bicolor + other spp.). Annual world-wide production now stands at 57 million metric tons of this increasingly popular grain. The species are believed to be Asian or African in origin. Sorghum was introduced into the U. S. in the mid-1800's. The grains are small and difficult to process. We use the various species mostly for forage and silage, but in the Old World the grains are often eaten like rice or made into an unleavened bread. All of the species are diploids (2n

= 2x = 40), except for Johnson grass, a very aggressive tetraploid weed.

There are four commonly recognized groups of sorghum species, based upon their use:

- syrup or sorgos, whose stem juices are abundant and sweet;
- broomcorn, used to make old-style commercial brooms;
- grain sorghums, such as kaffir, milo, and durra; and
- grass sorghums, such as Sudan grass, Tunis grass, and Johnson grass, grown for forage and silage.

WILD RICE

Zizania aquatica is native to North America. The common name is confusing, because it is not really a kind of rice (Oryza). The plants are robust aquatics. As in maize, the two sexes are found in separate spikelets on different parts of the plant. Native Americans gathered the grains by boat. Until recently, wild rice has eluded cultivation with most of the crop coming from Minnesota. It is now being cultivated, including in some of our northern California counties.

T' EF OR TEFF

This popular grain, derived from *Eragrostis abyssinica*, is native to northeastern Africa. Its major production site is Ethiopia, where it is more popular than all other cereal grains combined. The very small grains (1/32 in. in diameter) are typically fermented for a day or so and then made into pancakes. Teff has become popular among natural food enthusiasts in this country in recent years. The plants are also an excellent source of fodder.

JOB'S TEARS

Although native to southeastern Asia, Coix lacryma-jobi, named after the righteous sufferer in the Old Testament parable, is now very common through all of the tropical and subtropical regions of the world where its grains are used as food. It is not highly regarded, even though it has a very high protein content. Many of you will have seen these grains because they are also used to make rosaries and tourist trinkets.

MILLETS

This is a group name for an artificial assemblage of grasses that have very small grains. Most of the common ones belong to the genera *Pennisetum, Setaria, Panicum,* and *Eleusine*. For the most part, the various species are native to tropical and subtropical areas of the Old World. They are especially well

adapted to poorer soils. Most of us in Europe or North America have never eaten any of the millets (unless, of course, we frequent hippie cooperative food stores) and probably do not appreciate the role that they play in the diet of about one-third of the world's people. We see millet relatives as roadside weeds or as constituents in bird seed mix. They are also used for forage.

TRITICALE

Both its common name and generic name (X *Triticosecale*) suggest the origin of this cereal. It is a human-mediated hybrid between wheat (*Triticum*) and rye (*Secale*). Natural hybrids had been known for many years, but their offspring were usually sterile. Genetic work began in the late 1930's to produce a synthetic hybrid that would combine the useful features of these two important cereals. In 1965, the Centro Internacional de Mejoramiento de Maize y Trigo launched a major effort to make triticale a major food crop. It is the only artificial cereal to date that has had a major impact.

THE PROCESS

Secale cereale Rye RR	Χ	<i>Triticum durum</i> Durum wheat AABB
	<i>Triticosecale</i> ABR	
Secale cereale Rye RR	X V	Triticum aestivum Breadwheat AABBCC
	<i>Triticosecale</i> ABCR	

THE MINOR CEREALS

Common Name [Scientific Name]

Comments

acha [Digitaria exilis] Adlay [Coix lacryma-jobi] African millet [Eleusine coracana] barley [Hordeum vulgare] barnyard grass [Echinochloa crus-galli]

broom millet [Panicum miliaceum] browntop [Brachiaria ramosa] bulrush millet [Pennisetum americanum] channel millet [Echinochloa turnerianum] club wheat [Triticum compactum]

common millet [Panicum miliaceum] durum wheat [Triticum durum] einkorn wheat [Triticum monococcum] emmer wheat [Triticum dicoccon] finger millet [Eleusine coracana]

fonio [Digitaria elixis]
foxtail millet [Setaria italica]
German millet [Setaria italica]
guinea grass [Panicum maximum]
hog millet [Panicum miliaceum]

Hungarian millet [Setaria italica] Hungry-rice [Digitaria eximis] Italian millet [Setaria italica] Japanese millet [Echinochloa crusgalli] Job's tears [Coix lacryma-jobi]

kans [Saccharum spontaneum] koda millet [Paspaum commersonii] little millet [Panicum sumatrense] manna grass [Glyceria spp.] naked oats [Avena nuda]

oats [Avena spp.]
pearl millet [Pennisetum glaucum]
perennial teosinte [Zea mays ssp. diploperennis]
Polish wheat [Triticum polonicum]
proso millet [Panicum miliaceum]

ragi [Eleusine coracana]
rye [Secale cereale]
sanwa millet [Echinochloa frumentacea]
shama millet [Echinochloa colona]
sorghum [Sorghum bicolor]

tartarian oats [Avena orientalis] teff [Eragrostis abyssinica] teosinte [Zea mays ssp. mexicana] triticale [X Triticosecale spp.] wild rice [Zizania spp.] African; quite palatable and nutritious
See Job's tears
Widely used in China, India, and Africa
Old World; one of the ancient cereals
Known to us also as an agricultural weed

Cultivated especially in the Old World A relative of the *Panicum* cereals A relative of elephant and Napier grass A relative of our barnyard grass Grown mostly in Chile, USA, and India

In use since prehistoric times; Eurasia High in gluten; used to make spaghetti Primitive diploid, 1-seeded wheat Ancient Mediterranean wheat; still used Important cereal in Africa and India

> Used in tropical Africa Native to India; Near East & China See foxtail millet A perennial grass of tropical areas See common millet

> Old World; now widely cultivated West Africa; now unknown in wild See Hungarian millet See barnyard grass SE Asia; ornamental use in jewelry

Sugar cane relative grown in Africa Old World; relative of Dallis and bahia grass Grown extensively in India Used especially in North America Upland regions of China

Hexaploids most important Highly nutritious; hybrids grown in USA Recently discovered in Mexico S. Europe and n. Africa, not Poland Ancient; grown mostly in USSR and Asia

> See finger millet Probably native to southeast Asia Used primarily as cereal in Far East Old World; now also a widespread Ancient cereal of Asia and Africa

One-sided spikelet clusters Ethiopia and African highlands A close relative of maize Artificial wheat/rye hybrid Native to North America; recently domesticated

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SELECTED REFERENCES

WHEAT

Beardsley, T. 1991. A nitrogen fix for wheat. Sci. Amer. 264(3): 32.

Bell, G. D. H. 1987. The history of wheat cultivation. <u>In</u>, Lupton, F. G. H. Wheat breeding: its scientific basis. Chapman & Hall. London. Pp. 31-49.

Briggle, L. W. 1969. Triticale -- a review. Crop Science 9: 197-201.

Evans, L. T. & W. J. Peacock (editors). 1981. Wheat science: today and tomorrow. Cambridge Univ. Press. 290 pp.

Farbiani, G. & C. Lintas (editors). 1988. Durum wheat: chemistry and technology. American Assoc. Cereal Chemists. St. Paul, MN. 332 pp.

Feldman, M. & E. R. Sears. 1981. The wild gene resources of wheat. Sci. Amer. 244(1): 98-109.

Friggens, P. 1975. Triticale: world's first man-made crop. Reader's Digest 107(Dec.): 33-36.

Gregory, R. S. 1975. The commercial production of triticale. Span 18: 65, 66.

Harlan, J. R. 1967. A wild wheat harvest in Turkey. Archaeology 20: 197-201.

Harlan, J. R. & D. Zohary. 1966. Distribution of wild wheats and barley. Science 513: 1074-1080.

Helbaek, H. 1966. Commentary on the phylogensis of *Triticum* and *Hordeum*. Econ. Bot. 20: 350-360.

Hillman, G. C. & M. S. Davies. 1990. Measured domestication rates in crops of wild type wheats and barley and the archaeological implications. J. World Prehist. 4: 157-222.

Huen, M. et al. 1997. Site of einkorn wheat domestication identified by DNA fingerprinting. Science 278: 1312-1314.

Hulse, J. H. & D. Spurgeon. 1974. Triticale. Sci. Amer. 231: 72-81.

Inglett, G. E. (editor). 1973. Wheat: production and utilization. AVI Publ. Co. Westport, CT. 500 pp.

Jenkins, J. A. 1966. The origin of cultivated wheat. Canadian J. Genet. Cytol. 8: 220-232.

Kerby, K. & J. Kuspira. 1987. The phylogeny of the polyploid wheats, *Triticum aestivum* (bread wheat) and *Triticum turgidum* (macaroni wheat). Genome 29: 722-737.

Kimber, G. 1974. A reassessment of the origin of the polyploid wheats. Genetics 78(1): 487-492.

Kimber, G. E. & E. R. Sears. 1987. Evolution in the genus Triticum and the origin of cultivated wheat. \underline{In} , Heyne, E. G. (editor). Wheat and wheat improvement. Second edition. Pp. 154-164.

Kimber, G. & M. Feldman. 1987. Wild wheat: an introduction. Univ. Missouri College of Agric. Special Report No. 353. Columbia. 142 pp.

Kuckuck, H. 1970. Primitive wheats. <u>In</u>, Frankel, O. H. and E. Bennett. Genetic resources in plants. Pp. 249-266.

Mangelsdorf, P. C. 1953. Wheat. Sci. American 189(July): 50-59.

Merker, A. 1992. The Triticeae in cereal breeding. Hereditas 116: 277-280.

Peterson, R. F. 1965. Wheat: botany, cultivation, and utilization. Leonard-Hill. London. 422 pp.

Pomeranz, Y. 1973. From wheat to bread: a biochemical study. Amer. Sci. 61: 683-691.

Quisenberry, K. S. & L. P. Reitz (editors). 1967. Wheat and wheat improvement. American Soc. Agron. Madison, WI. 560 pp.

Riley, R. 1965. Cytogenetics and the evolution of wheat. \underline{In} , Hutchinson, J. (editor). Essays on crop plant evolution. Cambridge Univ. Press. London. Pp. 103-122.

Sears, E. R. 1948. The cytology and genetics of the wheats and their relatives. Adv. in Genetics 2: 239-270

Sears, E. R. 1977. The origin and future of wheat. \underline{In} , Seigler, D. S. (editor). Crop resources. Academic Press. New York, NY. Pp. 193-196.

Smith, N. 1983. Triticale: the birth of a new cereal. New Sci. 97: 98, 99.

Spicer, A. 1975. Bread. The social, nutritional and agricultural aspects of wheaten bread. Appl. Sci. Essex, England. 358 pp.

Zohary, D. 1970. Wild wheats. <u>In</u>, Frankel, O. H. & E. Bennett (editors). Genetic resources in plants. Pp. 239-248.

RICE

Anonymous. 1973. Rice in the United States: varieties and production. Agric. Handbook No. 289. U. S. Dept. Agric. Washington, D. C. 154 pp.

Association of Japanese Agricultural Scientific

Societies (editors). 1975. Rice in Asia. Univ. Tokyo Press. Tokyo. 600 pp.

Chang, T.-T. 1976. The origin, evolution, cultivation, dissemination, and diversification of Asian and African rices. Euphytica 25(2): 425-441.

Chang, T.-T. 1984. Conservation of rice genetic resources: luxury or necessity? Science 224: 251-256.

Crawford, R. 1991. Gene mapping Japan's number one crop. Science 252: 1611.

Grist, D. H. 1986. Rice. Sixth edition. Longman. London, England. 599 pp.

Hanks, L. M. 1972. Rice and man: agricultural ecology in Southeast Asia. Aldine & Atherton. Chicago, IL. 174 pp.

Hargrove, T. R. et al. 1988. Twenty years of rice breeding. BioScience 38(10): 675-681.

Harris, D. R. 1974. Rice and man in Southeast Asia. Geogr. Rev. 64: 140-142.

Nayar, N. M. 1973. Origin and cytogenetics of rice. Genetics 17: 153-292.

Normile, D. 1997. Yangtze seen as earliest rice site. Science 275: 309.

Oka, H.-I. 1984. Origin of cultivated rice. Elsevier Science Publ. Co. New York, NY. 380 pp.

Sampath, S. 1964. The species ancestral to cultivated rice. Curr. Sci. 33: 205-207.

Vaughan, D. A. & L. A. Sitch. 1991. Gene flow from the jungle to farmers. BioScience 41(1): 22-28.

MAIZE

Anderson, E. 1945. What is Zea mays? Chron. Bot. 9: 88-92.

Anderson, E. 1947. Corn before Columbus. Pioneer Hi-Bred Corn Company. Des Moines, IA. 24 pp.

Anderson, E. & H. C. Cutler. 1950. Methods of corn popping and their historical significance. Southwest. J. Anthrop. 6: 303-308.

Anderson, E. & R. H. Barlow. 1943. The maize tribute of Montezuma's empire. Annals Missouri Bot. Gard. 30: 413-418.

Anderson, E. & W. L. Brown. 1952. The history of the common maize varieties of the United States corn belt. Agric. Hist. 26: 2-8.

Barghoorn, E. S., M. K. Wolf, & K. H. Clisby. 1954. Fossil maize from the Valley of Mexico. Bot. Mus.

Leaflts. Harvard Univ. 16: 229-240.

Beadle, G. W. 1939. Teosinte and the origin of maize. J. Heredity 30: 245-247.

Beadle, G. W. 1972. The mystery of maize. Bull. Field Mus. Nat. Hist. 43(10): 2-11.

Beadle, G. W. 1977. The origins of *Zea mays*. <u>In</u>, Reed, C. A. (editor). Origins of agriculture. Mouton Publ. The Hague. Pp. 615-635.

Beadle, G. W. 1978. Teosinte and the origin of maize. <u>In</u>, Walden, D. B. (editor). Maize breeding and genetics. John Wiley & Sons. New York, NY. Pp. 113-128.

Beadle, G. W. 1980. The ancestry of corn. Sci. American 242(1): 112-119.

Bennetzen, J. et al. 2001. Genetic evidence and the origin of maize. Latin American Antiq. 12: 84-86.

Benz, B. F. 1986. Taxonomy and evolution of Mexican maize. Ph. D. dissertation. Univ. Wisconsin. Madison. 466 pp.

Benz, B. F., L. Sanchez-Velasquz, & F. J. Santana-Michel. Ecology and ethnobotany of *Zea diploperennis*. Maydica 35: 85-98.

Bird, R. M. 1980. Maize evolution from 500 B. C. to the present. Biotropica 12: 30-41.

Bird, R. M. 1984. South American maize in Central America? <u>In</u>, Stone, D. (editor). Pre-Columbian plant migration. Papers Peabody Mus. Archaeol. & Ethnol. 76: 39-65.

Brown, W. L. & E. Anderson. 1947. The northern flint corns. Annals Missouri Bot. Gard. 34: 1-28.

Brush, S. B., M. B. Corrales, & E. Schmidt. 1988. Agricultural development and maize diversity in Mexico. Human Ecol. 16(3): 307-328.

Bush, M. B., D. R. Piperno, & P. A. Colinvaux. 1989. A 6,000 year history of Amazonian maize cultivation. Nature 340: 303, 305.

Callen, E. O. 1967. The first New World cereal. American Antiq. 32: 535-538.

Carter, G. F. & E. Anderson. 1945. A preliminary survey of maize in the southwestern United States. Annals Missouri Bot. Gard. 32: 297-322.

Collins, G. N. 1912. The origin of maize. J. Washington Acad. Sci. 2: 520-530.

Collins, G. N. 1918. Maize, its origin and relationships. J. Washington Acad. Sci. 8: 42, 43.

- Collins, G. N. 1921. Teosinte in Mexico. J. Heredity 12: 339-350.
- Collins, G. N. 1925. The "metamorphosis" of *Euchlaena* into maize. J. Heredity 16: 378-380.
- Collins, G. N. 1931. The phylogeny of maize. Bull. Torrey Bot. Club 57: 199-210.
- Collins, G. N. & J. H. Kempton. 1920. A teosinte-maize hybrid. J. Agric. Res. 19: 1-37.
- Contreras, G., L. G. Elias, & R. Bressani. 1980. Limitations of corn (*Zea mays*) and common beans (*Phaseolus vulgaris*) diets as protein and calorie sources. Qualitas Plantarum 30(2): 145-153.
- Culotta, E. 1991. How many genes had to change to produce corn? Science 252: 1792, 1793.
- Curtin, L. S. M. 1968. Preparation of sacred corn meal in the Rio Grande pueblos. Southwest Mus. Leaflts. No. 32. 15 pp.
- Cutler, H. C. & L. W. Blake. 1971. Travels of corn and squash. <u>In</u>, Riley, C. L. et al. (editors). Man across the sea. Univ. Texas Press. Austin. Pp. 366-375.
- Dellaporta, S. L. & A. Calderon-Urrea. 1994. The sex determination process in maize. Science 266: 1501-1504.
- de Wet, J. M. J. & J. R. Harlan. 1971. Origin of maize: the tripartite hypothesis. Euphytica 21(2): 271-279.
- de Wet, J. M. J. & J. R. Harlan. 1974. *Tripsacum*-maize inter-action: a novel cytogenetic system. Genetics 78: 493-502.
- de Wet, J. M. J. & J. R. Harlan. 1976. Cytogenetic evidence for the origin of teosinte (*Zea mays* ssp. *mexicana*). Euphytica 25(2): 447-455.
- de Wet, J. M. J., J. R. Harlan, & C. A. Grant. 1971. Origin and evolution of teosinte (*Zea mexicana*). Euphytica 20: 255-265.
- de Wet, J. M. J. et al. 1972. Cytology of maize-tripsacum intro-gression. American J. Bot. 59: 1026-1029.
- de Wet, J. M. J. et al. 1972. Introgression from *Tripsacum* into *Zea* and the origin of maize. Caryologia 25: 25-31.
- de Wet, J. M. J. et al. 1978. The origin of tripsacoid maize (*Zea mays* L.) Evolution 32(2): 233-244.
- Doebley, J. F. 1985. Maize introgression into teosinte -- a reappraisal. Annals Missouri Bot. Gard. 71: 1100-1113.
- Doebley, J. F. 1990. Molecular evidence and the

- evolution of maize. Econ. Bot. 44(3: Suppl.): 6-27.
- Doebley, J. 1992. Mapping the genes that made maize. Trends in Genetics 8(9): 302-307.
- Doebley, J. F. 1996. Genetic dissection of the morpholoigcal evolution of maize. Aliso 14(4): 297-304.
- Doebley, J. F. & A. Stec. 1991. Genetic analysis of the morphological difference between maize and teosinte. Genetics 129: 285-295.
- Doebley, J. F. & G. Nabhan. 1989. Further evidence regarding gene flow between maize and teosinte. Maize Genetics Coop. Newsletter 63: 107, 108.
- Doebley, J. F. et al. 1988. The origin of cornbelt maize: the isozyme evidence. Econ. Bot. 42(1): 120-131.
- Doolittle, W. E. & C. D. Frederick. 1991. Phytoliths as indicators of prehistoric maize (*Zea mays* ssp. *mays*, Poaceae) cultivation. Plant Syst. Evol. 177(3-4): 175-184.
- Eddy, F. W. 1964. Metates & manos: the basic corn grinding tools of the Southwest. Popular Series Pamphlet No. 1. Museum of New Mexico Press. Santa Fe. s. p.
- Erwin, A. T. 1950. The origin and history of pop corn. Econ. Bot. 4(3): 294.
- Eubanks, M. W. 2001. The mysterious origin of maize. Econ. Bot. 55(4): 492-514.
- Eubanks, M. W. 2001. The origin of maize: evidence for *Tripsacum* ancestry. Plant Breeding Reviews 20: 15-61.
- Eubanks, M. W. 2001. An interdisciplinary perspective on the origin of maize. Latin American Antiq. 12: 91-98
- Eyre-Walker, A. et al. 1998. Investigation of the bottleneck lead-ing to the domestication of maize. Proc. Natl. Acad. Sci. 95: 4441-4446.
- Finan, J. J. 1948. Maize in the great herbals. Annals Missouri Bot. Gard. 35: 149-191.
- Fussell, B. 1994. The story of corn. A. A. Knopf. New York, NY. 356 pp.
- Galinat, W. C. 1961. Corn's evolution and its significance for breeding. Econ. Bot. 15: 320-325.
- Galinat, W. C. 1963. Form and function of plant structures in the American Maydeae and their significance for breeding. Econ. Bot. 17: 51-59.
- Galinat, W. C. 1965. The evolution of corn and culture

in North America, Econ. Bot. 19: 350-357.

Galinat, W. C. 1966. The evolution of glumeless sweet corn. Econ. Bot. 20: 441-445.

Galinat, W. C. 1971. The origin of maize. Annual Rev. Genet. 5: 447-478.

Galinat, W. C. 1971. The evolution of sweet corn. Res. Bull. No. 591. Univ. Massachusetts Agric. Exp. Sta. Amherst. 20 pp.

Galinat, W. C. 1974. The domestication and genetic erosion of maize. Econ. Bot. 28: 31-37.

Galinat, W. C. 1975. The evolutionary emergence of maize. Bull. Torrey Bot. Club 102: 313-324.

Galinat, W. C. 1977. The origin of corn. <u>In</u>, Sprague, Pp. 1-47.

Galinat, W. C. 1983. The origin of maize as shown by key morphological traits of its ancestor, teosinte. Maydica 28: 121-138.

Galinat, W. C. 1985. Domestication and diffusion in maize. <u>In</u>, Ford, R. I. (editor). Prehistoric food production in North America. Univ. Michigan. Ann Arbor. Pp. 245-282.

Galinat, W. C. 1985. The missing link between teosinte and maize. A review. Maydica 30: 137-160.

Galinat, W. C. 1988. The origin of corn. Agronomy 18: 1-31.

Galinat, W. C. 1992. Evolution of corn. Adv. Agron. 47: 203-231.

Galinat, W. C. 1995. The origin of maize: grain of humanity. Econ. Bot. 49(1): 3-12.

Galinat, W. C. 2001. A reconstruction of a possible role of crucial observations leading to a rapid domestic transformation of wild teosinte into the first maize. Econ. Bot. 55(4): 570-574.

Goette, S. et al. 1994. Toward reconstructing ancient maize: experiments in processing and charring. J. Ethnobiol. 14(1): 1-21.

Goodman, M. M. 1988. The history and evolution of maize. CRC Critical Rev. Plant Sci. 7(3): 197-220.

Goodman, M. M. & W. L. Brown. 1988. Races of corn. Agronomy 18: 33-79.

Gould, S. J. 1984. A short way to corn. Nat. Hist. 93(3): 12, 14, 16, 18, 20.

Guzman Mejia, R. 1978. Redescubrimiento de *Zea perennis* (Gramineae). Phytologia 38(3): 177.

Hallauer, A. (editor). 1994. Specialty corns. CRC Press. Boca Raton, FL. 410 pp.

Hardemann, N. P. 1981. Shucks, shocks, and hominy blocks: corn as a way of life in pioneer America. Louisiana State University Press. Baton Rouge. 271 pp.

Harpstead, D. D. 1971. High-lysine corn. Sci. Amer. 225(2): 34-42.

Harshberger, J. W. 1893. Maize, a botanical and economic study. Contr. Bot. Lab. Univ. Pennsylvania 1: 75-202.

Harshberger, J. W. 1904. A study of the fertile hybrids produced by crossing teosinte and maize. Contr. Bot. Lab. Univ. Pennsylvania 2: 231-235.

Hatt, G. 1951. The Corn Mother in America and Indonesia. Anthropos 46: 853-914.

Hilton, H. & B. Gaut. 1998. Speciation and domestication in maize and its wild relatives. Evidence from the globulin-1 gene. Genetics 150: 863-872.

Iltis, H. H. 1972. The taxonomy of *Zea mays* (Gramineae). Phytologia 23(2): 248, 249.

Iltis, H. H. 1983. From teosinte to maize: the catastrophic sexual transmutation. Science 222: 886-894.

Iltis, H. H. 1987. Maize evolution and agricultural origins. <u>In</u>, Soderstrom, T. R. et al. (editors). Grass systematics and evolution. Smithsonian Inst. Press. Washington, D. C. Pp. 195-213.

Iltis, H. H. et al. 1979. Zea diploperennis (Gramineae): a new teosinte from Mexico. Science 203: 186-188.

Iltis, H. H. & J. F. Doebley. 1984. Zea -- a biosystematic odyssey. <u>In</u>, Grant, W. F. (editor). Plant biosystematics. Academic Press. Montreal, Canada. Pp. 587-616.

Inglett, G. E. (editor). 1982. Maize. Recent progress in chemistry and technology. Papers from a symposium, Prague, June 1982. Academic Press. New York, NY. 252 pp.

Jeffreys, M. D. W. 1967. Pre-Columbian maize in southern Africa. Nature 215: 695-697.

Jeffreys, M. D. W. 1971. Pre-Columbian maize in Asia. E. Anthrop. 9: 21-28.

Jeffreys, M. D. W. 1971. Pre-Columbian maize in Asia. In, Riley, C. L. et al. (editors). Man across the sea. Univ. Texas Press. Austin. Pp. 376-400.

Johannessen, C. L. & A. Z. Parker. 1989. Maize ears

sculptured in 12th and 13th century A. D. India as indicators of Pre-Columbian diffusion. Econ. Bot. 43(2): 164-180.

Johannessen, S. & C. A. Hastorf. 1989. Corn and culture in central Andean prehistory. Science 244: 690-692.

Johannessen, S. & C. A. Hastorf (editors). 1994. Corn and culture in the prehistoric New World. Westview Press. Boulder, CO. 623 pp.

Johanessen, C. L., M. R. Wilson, & W. A. Davenport. 1970. The domestication of maize: process or event? Geogr. Review 60: 393-413.

Kahn, E. J. 1984. Profiles: the staffs of life. I. The golden thread. The New Yorker X:46-88.

Kato, T. A. 1984. Chromosome morphology and the origin of maize and its races. Evol. Biol. 17: 219-253.

Katz, S. H., M. L. Hediger, & L. A. Valleroy. 1974. Traditional maize processing techniques in the New World. Science 184: 765-773.

Kempton,, J. H. 1919. The ancestry of maize. J. Washington Acad. Sci. 9: 3-11.

Kempton, J. H. 1938. Maize -- our heritage from the Indian. Smithsonian Report for 1937. Smithsonian Institution. Washington, D. C. Pp. 385-408.

Levings III, C. S. 1996. Infertility treatment: a nuclear restorer gene in maize. Science 272: 1279, 1280.

Linn, A. 1973. Corn: the New World's secret weapon and the builder of its civilization. Smithsonian 4: 58-65.

Long, A. et al. 1989. First direct AMS dates on early maize from Tehuacan, Mexico. Radiocarbon 31: 1035-1040.

MacNeish, R. S. 1955. Ancient maize and Mexico. Archaeology 8(2): 108-115.

MacNeish, R. S. 1985. The archaeological record in the problem of the domestication of corn. Maydica 30: 171-178.

Mangelsdorf, P. C. 1945. The origin and nature of the ear of maize. Bot. Mus. Leaflts. Harvard Univ. 12: 33-88.

Mangelsdorf, P. C. 1947. The origin and evolution of maize. Adv. in Genetics 1: 161-207.

Mangelsdorf, P. C. 1948. The role of pod corn in the origin and evolution of maize. Annals Missouri Bot. Gard. 35: 377-406.

Mangelsdorf, P. C. 1950. The mystery of corn. Sci.

Amer. (July): 1-6.

Mangelsdorf, P. C. 1958. Ancestor of corn. Science 128: 1313-1320.

Mangelsdorf, P. C. 1958. Reconstructing the ancestor of corn. Proc. American Phil. Soc. 102: 454-463.

Mangelsdorf, P. C. 1960. Reconstructing the ancestor of corn. Smithsonian Report for 1959. Smithsonian Institution. Washington, D. C. Pp. 495-507.

Mangelsdorf, P. C. 1961. Introgression in maize. Euphytica 10: 157-168.

Mangelsdorf, P. C. 1964. Corn in the Old World. Science 145:659.

Mangelsdorf, P. C. 1965. The evolution of maize. \underline{In} , Hutchinson, J. Essays on crop plant evolution. Cambridge University Press. Cambridge, England. Pp. 23-49.

Mangelsdorf, P. C. 1974. Corn: its origin, evolution, and improvement. Harvard University Press. Cambridge, MA. 262 pp.

Mangelsdorf, P. C. 1983. The mystery of corn: new perspectives. Proc. American Phil. Soc. 127: 215-247.

Mangelsdorf, P. C. 1983. The search for wild corn. Maydica 28: 89-96.

Mangelsdorf, P. C. 1986. The origin of corn. Sci. American 255(2): 80-86.

Mangelsdorf, P. C. & D. L. Oliver. 1951. Whence came maize to Asia? Bot. Mus. Leaflts. Harvard Univ. 14: 263-291.

Mangelsdorf, P. C. & R. G. Reeves. 1935. A trigeneric hybrid of *Zea, Tripsacum*, and *Euchlaena*. J. Heredit. 26: 129-140.

Mangelsdorf, P. C. & R. G. Reeves. 1938. The origin of maize. Proc. Natl. Acad. Sci. 24: 303-312.

Mangelsdorf, P. C. & R. G. Reeves. 1939. The origin of Indian corn and its relatives. Texas Agric. Exper. Station Bull. No. 574. 315 pp.

Mangelsdorf, P. C. & R. G. Reeves. 1959. The origin of corn. I. Pod corn, the ancestral form. Bot. Mus. Leaflts. Harvard Univ. 18: 329-356.

Mangelsdorf, P. C. & R. G. Reeves. 1959. The origin of corn. II. Teosinte, a hybrid of corn and tripsacum. Bot. Mus. Leaflts. Harvard Univ. 18: 357-387.

Mangelsdorf, P. C. & R. G. Reeves. 1959. The origin of corn. III. Modern races, the product of teosinte introgression. Bot. Mus. Leaflts. Harvard Univ. 18: 389-411.

Mangelsdorf, P. C. & R. G. Reeves. 1959. The origin of corn. IV. Place and time of origin. Bot. Mus. Leaflts. Harvard Univ. 18: 413-427.

Mangelsdorf, P. C., R. S. MacNeish, & W. C. Galinat. 1964. Domestication of corn. Science 143: 538-545.

Mangelsdorf, P. C., R. S. MacNeish, & W. C. Galinat. 1967. Prehistoric wild and cultivated maize. <u>In</u>, Byers, D. S. (editor). Prehistory of the Tehuacan Valley. Univ. Texas Press. Austin. 1: 178-200.

Mangelsdorf, P. C. et al. 1978. Fossil pollen and the origin of corn. Bot. Mus. Leaflts. Harvard Univ. 26(7): 237-255.

Martienssen, R. 1997. The origin of maize branches out. Nature 386: 443, 445.

Montgomery, E. G. 1906. What is an ear of corn? Pop. Sci. Monthly 68: 55-62.

Morrison, G. 1947. Hybrid corn -- science in practice. Econ. Bot. 1(1): 5-19.

Nault, L. R. & W. R. Findley. 1983. *Zea diploperennis*, a primitive relative, offers new traits to improve corn. Desert Plants 3: 203-205.

Onion, D. K. 1964. Corn in the culture of the Mohawk Indians. Econ. Bot. 18: 60-66.

Ortiz, A. 1989. Some cultural meanings of corn in aboriginal North America. Northeast Indian Quart. 6: 64-73.

Pearsall, D. M. 1978. Early movement of maize between Mesoamerica and South America. J. Steward Anthrop. Soc. 9: 41-75.

Raloff, J. 1993. Corn's slow path to stardom. Sci. News 143(16): 248-250.

Randolf, L. F. 1952. New evidence on the origin of maize. American Natl. 86: 193-202.

Randolf, L. F. 1959. The origin of maize. Ind. J. Genet. Plant Breed. 19: 1-12.

Randolf, L. F. 1976. Contributions of wild relatives of maize to the evolutionary history of domesticated maize: a synthesis of divergent hypotheses. I. Econ. Bot. 30: 321-345.

Reeves, R. G. & P. C. Mangelsdorf. 1959. The origin of corn. V. A critique of current theories. Bot. Mus. Leaflts. Harvard Univ. 18: 428-440.

Rhoades, M. M. 1984. The early years of maize genetics. Ann. Rev. Genetics 18: 1-29.

Rhoades, R. E. 1993. The golden grain: corn. Natl. Geogr. 183(6): 92-117.

Roush, W. 1996. Corn: a lot of change from a little DNA. Science 272: 1873.

Sehgal, S. M. 1985. Paul C. Mangelsdorf. Maydica 30(2): 125-136.

Shaver, D. L. 1967. Perennial maize. J. Heredit. 58: 271-273.

Smith, B. D. 1995. Maize. <u>In</u>, Smith, B. D. The emergence of agriculture. Sci. American Library. New York, NY. Pp. 150-160.

Sprague, G. F. & J. W. Dudley (editors). 1988. Corn and corn improvement. Third edition. American Soc. Agron. Madison, WI. 986 pp.

Swartz, H. M. & A. B. Flood. The corntinental theory of flat and depressed areas: on the relationship between corn and topography. J. Irreproducible Results 35(5): 16, 17; 19. [A very funny spoof that demonstrates that corn flattens the land on which it grows!]

Tatum, L. A. 1971. The southern corn leaf blight epidemic. Science 171: 1113-1116.

Tsukada, M. & J. R. Rowley. 1964. Identification of modern and fossil maize pollen. Grana Palynol. 5(3): 406-412.

Walden, H. T. 1966. Native inheritance. The story of corn in America. Harper & Row. New York, NY. 199 pp.

Wang, R.-L. et al. 1999. The limits of selection during maize domestication. Nature 398: 236-239.

Watson, S. A. & P. E. Ranstad (editors). 1987. Corn: chemistry and technology. American Assoc. Cereal Chemists. St. Paul, MN. 605 pp.

Weatherwax, P. 1918. The origin of maize. Bull. Torrey Club 45: 309-342.

Weatherwax, P. 1919. The ancestry of maize -- a reply to criticism. Bull. Torrey Bot. Club 46: 275-278.

Weatherwax, P. 1923. The story of the maize plant. Univ. Chicago Press. Chicago, IL. 247 pp.

Weatherwax, P. 1935. Phylogeny of maize. American Midl. Natl. 16: 1-71.

Weatherwax, P. 1945. Early contacts of European science with the Indian corn plant. Proc. Indiana Acad. Sci. 54: 169-178.

Weatherwax, P. 1950. The history of corn. Sci. Monthly 71: 50-60.

Weatherwax, P. 1954. Indian corn in Old America. Macmillan & Co. New York, NY. 253 pp.

Weatherwax, P. 1955. History and origin of corn. I.

Early history of corn and theories as to origin. <u>In</u>, Sprague, G. F. Corn and corn improvement. Academic Press. Pp. 1-16.

Whitehead, D. R. & E. J. Langham. 1965. Measurement as a means of identifying fossil maize pollen. Bull. Torrey Bot. Club 92(11): 7-20.

Wilkes, H. G. 1972. Maize and its wild relatives. Science 177: 1071-1077.

Wilkes, H. G. 1977. Hybridization of maize and teosinte, in Mexico and Guatemala and the improvement of maize. Econ. Bot. 31(3): 254-293.

Wilkes, H. G. 1977. The origin of corn -- studies of the last hundred years. <u>In</u>, Seigler, D. S. (editor). Crop resources. Academic Press. New York, NY. Pp. 211-223.

Wilkes, H. G. 1979. Mexico and Central America as a centre for the origin of agriculture and the evolution of maize. Crop Improvement [India] 6: 1-18.

Wilkes, H. G. 1985. Teosinte, the closest relative of maize revisited. Maydica 30: 209-224.

Will, G. F. & G. H. Hyde. 1917. Corn among the Indians of the Upper Missouri. William Harvey Miner. Co. St. Louis, MO.

Xolocotzi, E. H. 1985. Maize and man in the greater Southwest. Econ. Bot. 39(4): 416-431.

Yen, D. E. 1959. The use of maize by the New Zealand Maoris. Econ. Bot. 13(4): 319-327.

MINOR CEREALS

Andrews, D. J. & K. A. Kumar. 1992. Pearl millet for food, feed, and forage. Adv. Agron. 48: 89-139.

Baum, B. R. 1987. Classification of cultivated barley (*Hordeum vulgare*). II. Elaboration of cultivar groups. Canadian J. Bot. 65: 2152-2160.

Bothmer, R. von et al. 1991. An ecogeographical study of the genus *Hordeum*. Systematic and ecogeographical studies on crop genepools 7: 1-127.

Briggs, D. E. 1978. Barley. Chapman & Hall. London, England. 612 pp.

Brunken, J. et al. 1977. The morphology and domestication of pearl millet. Econ. Bot. 31(2): 163-174.

de Wet, J. M. J. & E. A. Oelke. 1978. Domestication of American wild rice (*Zizania aquatica* L., Gramineae). J. Agric. Tradit. Bot. Appl. 25: 67-84.

de Wet, J. M. J. & J. R. Harlan. 1971. The origin and domestication of *Sorghum bicolor*. Econ. Bot. 25: 128-

135.

de Wet, J. M. J. et al. 1979. Origins and evolution of foxtail millets (*Setaria italica*). J. Agric. Tradit. Bot. Appl. 26: 53-64.

de Wet, J. M. J. et al. 1983. Domestication of sawa millet (*Echinochloa colona*). Econ. Bot. 37(3): 283-291.

Dickson, A. D. et al. 1979. Barley: origin, botany, culture, winter hardiness, genetics, utilization, pests. Agric. Handbook No. 338. U. S. Dept. Agric. 154 pp.

Doggett, H. 1989. Sorghum. Second edition. John Wiley & Sons. New York, NY. 512 pp.

Grassl, C. O. 1968. *Saccharum* names and their interpretation. I. S. S. C. T. Proc. 13th Congress. Taiwan. Pp. 868-875.

Hayes, P. M., R. E. Stucker, & G. G. Wandrey. 1989. The domestication of American wildrice (*Zizania palustris*, Poaceae). Econ. Bot. 43(2): 203-214.

Hilu, K. W. & J. M. J. de Wet. 1976. Domestication of *Eleusine coracana*. Econ. Bot. 30: 199-208.

Jain, S. K. & D. K. Banerjee. 1974. Preliminary observations on the ethnobotany of the genus *Coix*. Econ. Bot. 28: 38-42.

Ladizinsky, G. 1969. New evidence on the origin of the hexaploid oats. Evolution 23: 676-684.

Mehra, K. L. 1963. Differentiation of the cultivated and wild *Eleusine* species. Phyton 20: 189-198.

Mengesha, M. H. 1966. Chemical composition of teff (*Eragrostis tef*) compared with that of wheat, barley and grain sorghum. Econ. Bot. 20: 268-273.

Nabhan, G. & J. M. J. de Wet. 1984. *Panicum sonorum* in Sonoran desert agriculture. Econ. Bot. 38(1): 65-82.

Rachie, K. O. & J. V. Majmudar. 1980. Pearl millet. Pennsylvania State Univ. Press. University Park. 307 pp.

Rachie, K. O. & L. V. Peters. 1977. The eleusines: a review of the world literature. Crops Res. Inst. for the Semiarid Tropics. 129 pp.

Sampson, D. R. 1954. On the origin of oats. Bot. Mus. Leaflts. Harvard University 16: 265-303.

Sencer, H. A. & J. G. Hawkes. 1980. On the origin of cultivated rye. Biol. J. Linnean Soc. 13: 299-313.

Shapley, D. 1973. Sorghum: "miracle" grain for the world protein shortage? Science 182: 147, 148.

Shewry, P. R. (editor). 1992. Barley: genetics, biochemistry, molecular biology and biotechnology. Biotechnology in Agricul-ture No. 5. CAB International. Wallingford, U. K.

Sokolov, R. 1993. The teff also rises. Nat. Hist. 102(3): 96, 98, 99.

Spencer, H. A. & J. G. Hawkes. 1980. On the origins of cultivated rye. Biol. J. Linnean Soc. 13: 299-313.

Standt, G. 1961. The origin of cultivated barleys: a discussion. Econ. Bot. 15(3): 205–212.

Stutz, H. C. 1972. On the origin of cultivated rye. American J. Bot. 59(1): 59-70.

Vinall, H. M. 1917. Foxtail millet: its culture and utilization in the United States. Farmers Bull. No. 793. U. S. Dept. of Agric. 28 pp.

Welch, R. W. (editor). 1995. The oat crop: production and utili-zation. Chapman & Hall. New York, NY. 584 pp.

6.07 - GRASSLANDS OF THE WORLD

Water

Grasslands are plant communities in which grasses are dominant. Major grassland areas occur in North America, South America, Africa, Asia, and Australia. Most of them are associations with more than one dominant species. Grasslands are often found in the interior of continents, where rain falls mainly in the summer months. The distribution of grasslands is dependent upon certain climatic, edaphic, and humangenerated factors.

There are three major kinds of grasslands, with many subdivisions and transitional types:

- Savannas are dominated by high, coarse grasses and more or less widely scattered, low trees. Turf is seldom formed. They are often found in the tropical and subtropical regions, often in areas characterized by a short dry season. Typical examples include the llano of Venezuela, the Campo of Brazil, and the vast grasslands of much of Africa.
- Steppes are treeless grasslands of low relief. They may be arid or semiarid and occur in warm or cool climates. Steppes are dominated by sodforming grasses, tufted tall grasses, or short grass vegetation. They are characterized by a protracted cold season. Typical examples include the grasslands of the southern republics of the former Soviet Union and the Great Plains east of the Rocky Mtns. in the United States.
- Prairies are treeless grasslands of low relief, climatically and vegetationally intermediate between savannas and short-grass steppes. Prairies are less humid that savannas and are less arid than steppes. They are usually absent from tropical/subtropical regions. Their deep, dark soils are covered with sod-forming tall-grasses. Typical examples include the Trans-Mississippi Valley in the United States, the Black Earth Belt of Russia, and the Pampa of Argentina.

THE EARTH'S COVER (mi²)

Ice caps	5,830,000
Forests: Tropical Rain Forest Temperate Rain Forest Deciduous Forest Coniferous Forest Dry or Monsoon Forest Thorn Forest Sclerophyll Brushland Total	3,800,000 550,000 6,500,000 7,600,000 2,000,000 340,000 1,180,000 21,970,000
Deserts: Desert Shrub/Grass Salt Desert Desert (Hot & Dry) Tundra (Cold) Total	10,600,000 30,000 2,400,000 4,400,000 17,430,000
Grasslands: High Grass Savanna Tall Grass Savanna Tall Grass Short Grass Desert Grass Savanna Total	2,800,000 3,900,000 1,580,000 1,200,000 2,300,000 12,570,000

[After Shantz, 1954]

139,150,000

Forests are 42% of the earth's vegetation cover, deserts are 34%, and grasslands are 24%. With adjustments to recognize the extent of grasslands in forests and deserts: forests = 40%; grasslands = 27%; and deserts = 33%.

GRASSLANDS OF NORTH AMERICA

Grassland Type	%	Hectares
Tallgrass Prairie	19	57,351,100
Mixed-grass Prairie	19	56,617,400
Shortgrass Prairie	21	61,522,300
Coastal Prairie	01	3,800,000
California Grassland	03	9,200,000
Palouse Prairie	22	64,471,600
Fescue Prairie	08	25,500,000
Desert Grassland	07	20,756,500
Total		299,222,900

[After Sims & Risser in Barbour & Billings, 2000]

TALL GRASS PRAIRIE

The Tall Grass Prairie, True Prairie, or Bluestem Prairie occurs at the eastern edge of the grassland formation, from southern Manitoba to south-central and eastern Texas, and eastward into Ohio. Characteristic grasses include:

Andropogon gerardii	Big bluestem
Sorghastrum nutans	Indian grass
Panicum virgatum	Switch grass
Schizachyrium scoparium	Little bluestem
Spartina pectinata	Prairie cord grass
Sporobolus heterolepis	Prairie dropseed
Koeleria macrantha	June grass
Bouteloua curtipendula	Sideoats grama
Bouteloua gracilis	Blue grama
Bouteloua hirsuta	Hairy grama
Stipa spartea	Porcupine needle grass

MIXED GRASS PRAIRIE

The Mixed Grass Prairie or Mixed Prairie is the largest grassland association. It occurs from Canada to south-central Texas; from western Nebraska to the Rocky Mountains. Rainfall (10-27") comes mainly in the spring and in the early summer. A summer drought is expected periodically. The characteristic grasses include:

Elymus smithii	
Elymus dasystachys	
Schizachyrium scoparium	
Stipa spartea	
Stipa comata	ſ

Western wheat grass Thickspike wheat grass Little bluestem Porcupine needle grass Needle-and-thread grass

Koeleria macrantha	June grass
Calamovilfa longifolia	Prairie sandreed
Sporobolus cryptandrus	Sand dropseed
Bouteloua gracilis	Blue grama
Carex spp.	Sedges

The Sandhills of Nebraska is a recognizably distinct component, characterized by:

Redfieldia flexuosa	Blowout grass
Stipa hymenoides	Indian rice grass
Eragrostis trichoides	Sand love grass
Stipa comata	Porcupine needle grass
Calamovilfa longifolia	Prairie sandreed
Schizachyrium scoparium	Little bluestem
Andropogon hallii	Sand bluestem
Muhlenbergia pungens	Sandhill muhly

SHORT GRASS PRAIRIE

The Short Grass Prairie or Short Grass Steppe occupies about 280,000 sq. km in the Central Plains of the United States. Its northern limit is close to the Colorado-Wyoming boundary, extending south into Texas; from the foothills of the Rocky Mtns. on the west to the Oklahoma Panhandle on the east. Rainfall is from about 300-500 mm annually. The characteristic grasses include:

Bouteloua gracilis	Blue grama
Stipa comata	Needle-and-thread grass
Elymus smithii	Western wheat grass
Buchloë dactyloides	Buffalo grass
Aristida purpurea	Purple three-awn grass
Hilaria mutica	Tobosa
Bouteloua hirsuta	Hairy grama

DESERT GRASSLAND

The Desert Grassland occurs in the American Southwest and in adjacent north-central Mexico. It is the hottest, driest of our grasslands, receiving 11-17" of precipitation each year. Snow is a factor at higher elevations. Characteristic low-elevation grasses include:

Black grama
Tobosa
Poverty three-awn
Purple three-awn
Bush muhly
Sand dropseed

At higher elevations:

Hilaria belangeri	Curly mesquite
Bouteloua gracilis	Blue grama
Bouteloua hirsuta	Hairy grama
Bouteloua eriopoda	Black grama
Hilaria mutica	Tobosa
Stipa neomexicana	New Mexico feather grass
Bouteloua curtipendula	Sideoats grama

CALIFORNIA PRAIRIE

The California Prairie or Pacific Prairie covers about 10 million hectares in California and Baja California. It occurs as open grassland and as an understory. It consists of two elements – one occurring along the North Coast at medium and higher elevations and the other in the Great Central Valley, southern Coast Range, and extending into Mexico.

Northern Component: pre-European settlement

Danthonia californicaCalifornia oat grassDeschampsia cespitosaTufted hair grassFestuca occidentalisWestern fescueFestuca idahoensisIdaho fescueFestuca rubraRed fescueCalamagrostis nutkaensisPacific reed grass

Northern Component: post-European settlement

Festuca arundinacea
Holcus lanatus
Velvet grass
Anthoxanthum occidentale
Lolium multiflorum
Arrhenatherum elatius
Dactylis glomerata

Tall fescue
Velvet grass
Sweet vernal grass
Italian rye grass
Tall oat grass
Orchard grass

Central Valley: pre-European settlement

Stipa pulchra Purple needle grass
Stipa cernua Nodding needle grass
Elymus glaucus Blue wild rye
Poa scabrella Pine bluegrass
Muhlenbergia rigens Deer grass

Central Valley: post-European settlement

Avena fatua var. fatua Wild oat Avena barbata Slender oat Bromus mollis Soft cheat grass Bromus diandrus Ripgut grass Bromus rubens Foxtail brome Hordeum murinum Mouse barley Little barley Hordeum pusillum Festuca myuros Foxtail fescue Festuca megalura Foxtail fescue

Erodium cicutariumFilareeErodium botrysFilaree

PALOUSE PRAIRIE

The Palouse Prairie, named after a Native American people, occurs from southern British Columbia southward to central Oregon and eastward into Idaho and Montana. Minor examples are found in Utah and in Nevada. The Palouse Prairie is a relatively arid grassland with about 8-25" of precipitation each year, much of it as snow. It occupies sites between sagebrush steppe on the drier side and ponderosa pine or Douglas-fir on the wetter side. Characteristic grasses include:

Elymus spicatus Festuca idahoensis Elymus cinereus Poa secunda Stipa comata Stipa occidentalis Koeleria macrantha Elymus pauciflorus Elymus smithii Bluebunch wheat grass
Idaho fescue
Ashy wild rye
Sandberg's bluegrass
Needle-and-thread grass
Western needle grass
June grass
Pacific wild rye
Western wild rye

FESCUE PRAIRIE

The Fescue Prairie is found on the northern and northwestern sides of the Mixed Grass Prairie, from central Saskatchewan into Alberta and southward into northern Montana. Its sole dominant is *Festuca scabrella*, which can account for 50% or more of the vegetative cover. It is richer in forbs than is the adjacent Mixed Grass Prairie. Characteristic grasses include:

Festuca scabrellaRough fescueElymus subsecundusBearded wheat grassDanthonia intermediaTimber oat grassHelictotrichon hookeriSpike-oatStipaPorcupine needle grassFestuca idahoensisIdaho fescue

COASTAL PRAIRIE

The Coastal Prairie occurs along the Gulf of Mexico, from southwestern Louisiana through Texas, into northeastern Mexico. It is a region of high humidity and rainfall (26-34"). The mild climate and long growing season favor the growth of subtropical grasses. There has also been an influx of taxa from the Short Grass Prairie. Characteristic grasses include:

Bothriochloa saccharoides Silver blustem Stipa leucotricha Texas needle grass Schizachyrium scoparium var. littorale

Seacoast bluestem
Andropogon gerardii Big bluestem
Heteropogon contortus Tanglehead
Sorghastrum nutans Indian grass
Paspalum plicatulum Brownseed paspalum
Spartina spartinae Gulf cord grass

THE KÜCHLER CLASSIFICATION

A. W. Küchler (1964) drew a distinction between **real vegetation**, which comprised all of the types of vegetation present at the time observations were made and **potential natural vegetation**, which he defined as "the vegetation that would exist today if man were removed from the scene and if the resulting plant succession were telescoped into a single moment." He recognized 116 vegetation types in the conterminous United States (i. e., the lower 48 states). The following contain one or more grasses as dominants.

Juniper Steppe Woodland. Dominants: *Elymus spicatus, Artemisia tridentata, Juniperus occidentalis.* Other components: *Artemisia, Balsamorhiza, Elymus, Festuca idahoensis, Lithospermum, Lupinus, Poa secunda, Purshia.* Occurrence: East of Cascade Range.

Fescue-Oatgrass. Dominants: Carex tumulicoa, Danthonia californica, Deschampsia holciformis, Festuca idahoensis. Other components: Agrostis hallii, Brodiaea, Calamagrostis nutkaensis, Calochortus, Chrysopsis, Grindelia, Iris, Lupinus, Pteridium, Ranunculus, Sanicula, Sisyrinchium, Stipa lepida. Occurrence: Western slopes of northern Coast Ranges, California.

California Steppe. Dominants: Stipa cernua, Stipa pulchra. Other components: Aristida divaricata, Elymus glaucus, Elymus triticoides, Eschscholzia, Gilia, Lupinus, Orthocarpus, Plagiobothrys, Poa scabrella, Sisyrinchium, Stipa coronata, Stipa lepida, and introduced annual species of Avena, Bromus, Festuca. Occurrence: Central Valley of California and some coastal regions south of San Francisco.

Fescue-Wheatgrass. Dominants: *Elymus spicatus, Festuca idahoensis.* Other components: *Achillea, Artemisia, Collinsia, Hieracium, Lupinus, Potentilla, Rosa, Symphoricarpos.* Occurrence: Eastern Washington and northwestern Idaho.

Wheatgrass-Bluegrass. Dominants: *Elymus spicatus, Festuca idahoensis, Poa secunda.* Other components: *Achillea, Astragalus, Chrysothamnus, Draba, Festuca microstachys, Lithophragma, Lupinus, Plantago, Stellaria.* Occurrence: Washington, Oregon, and northwestern Idaho.

Alpine Meadows and Barren. Dominants: Agrostis, Carex, Deschampsia cespitosa, Festuca viridula, Luzula spicata, Phleum alpinum, Poa, Trisetum spicatum. Other components: Achillea, Antennaria, Aquilegia, Arenaria, Castilleja, Draba, Erigeron, Oxyria, Penstemon, Phacelia, Phlox, Polemonium Polygonum, Potentilla, Salix, Saxifraga, Selaginella, Sibbaldia, Sieversia, Solidago. Occurrence: Rocky Mtns., Cascade Range, Sierra Nevada.

Grama-Galleta Steppe. Dominants: Bouteloua gracilis, Hilaria jamesii. Other components: Andropogon hallii, Schizachyrium scoparium, Artemisia, Astragalus, Atriplex, Bouteloua curtipendula, Bouteloua hirsuta, Ephedra, Opuntia, Stipa hymenoides, Tetradymia, Yucca. Occurrence: Northern Arizona and New Mexico.

Grama-Tobosa Prairie. Dominants: *Bouteloua gracilis, Hilaria mutica.* Other components: *Bouteloua curtipendula, Bouteloua eriopoda, Bouteloua hirsuta, Erioneuron pilosum, Gutierrezia, Muhlenbergia, Opuntia, Yucca.* Occurrence: Western Texas.

Sagebrush Steppe. Dominants: *Elymus spicatus, Artemisia tridentata*. Other components: *Artemisia,*

Balsamorhiza, Elymus, Festuca idahoensis, Lithospermum, Lupinus, Phlox, Poa nevadensis, Poa secunda, Purshia, Stipa hymenoides. Occurrence: Pacific Northwest, eastward to Rocky Mtns.

Wheatgrass-Needlegrass Shrubsteppe. Dominants: Elymus smithii, Artemisia tridentata, Poa arida, Stipa comata. Other components: Artemisia, Atriplex, Carex, Elymus spicatus, Eurotia, Koeleria macrantha, Sarcobatus. Occurrence: Montana and Wyoming.

Galleta-Three Awn Shrubsteppe. Dominants: Aristida longiseta, Artemisia filifolia, Ephedra viridis, Hilaria jamesii. Other components: Aster, Berberis, Bouteloua gracilis, Chrysopsis, Chrysothamnus, Ephedra, Euploca, Franseria, Helianthus, Mentzelia, Muhlenbergia pungens, Munroa squarrosa, Oenothera, Poliomintha, Quercus, Sphaeralcea, Sporobolus cryptandrus, Stephanomeria, Stipa hymenoides. Occurrence: Southeastern Utah.

Grama-Tobosa Shrubsteppe. Dominants: Bouteloua eriopoda, Hilaria mutica, Larrea divaricata. Other components: Acacia, Andropogon barbinodis, Aristida divaricata, Aristida glabrata, Aristida hamulosa, Aristida longiseta, Astragalus, Baileya, Bouteloua, Gutierrezia, Hilaria belangeri, Hilaria jamesii, Mentzelia, Muhlenbergia porteri, Opuntia, Prosopis, Sphaeralcea, Sporobolus airoides, Sporobolus cryptandrus, Sporobolus flexuosus, Yucca, Zinnia. Occurrence: Southeastern Arizona and southern New Mexico.

Mesquite Savanna. Dominants: Hilaria belangeri, Hilaria mutica, Prosopis juliflora. Other components: Acacia, Andropogon barbinodis, Aristida glauca, Aristida purpurea, Aristida wrightii, Bouteloua curtipendula, Bouteloua rigidiseta, Buchloë dactyloides, Juniperus, Quercus. Occurrence: Western Texas.

Mesquite-Acacia Savanna. Dominants: Acacia rigidula, Andropogon littoralis, Prosopis juliflora, Setaria macrostachya. Other components: Acacia, Bothriochloa saccharoides, Aristida, Bouteloua filiformis, Buchloë dactyloides, Cenchrus myosuroides, Chloris, Condalia, Heteropogon contortus, Hilaria belangeri, Opuntia, Pappophorum bicolor, Paspalum, Quercus, Schizachyrium scoparium (eastern part). Occurrence: Southern Texas.

Mesquite-Live Oak Savanna. Dominants: Andropogon littoralis, Prosopis juliflora, Quercus virginiana var. maritima. Other components: Aristida roemeriana, Brachiaria ciliatissima, Heteropogon contortus, Panicum virgatum, Paspalum, Sorghastrum nutans, Trachypogon secundus. Occurrence: Southern Texas.

Foothills Prairie. Dominants: *Elymus spicatus, Festuca idahoensis, Festuca scabrella, Stipa comata.* Other components: *Achillea, Agropyron, Artemisia,*

Bouteloua gracilis, Carex, Elymus smithii, Eriogonum, Koeleria macrantha, Penstemon, Poa secunda. Occurrence: Western Montana.

Grama-Needlegrass-Wheatgrass. Dominants: Elymus smithii, Bouteloua gracilis, Stipa comata. Other components: Artemisia, Carex, Chrysopsis, Elymus spicatus, Gutierrezia, Koeleria macrantha, Liatris, Poa secunda, Muhlenbergia cuspidata, Schizachyrium scoparium, Sporobolus cryptandrus, Stipa viridula. Occurrence: Montana and Wyoming.

Grama-Buffalo Grass. Dominants: Bouteloua gracilis, Buchloë dactyloides. Other components: Aristida purpurea, Bouteloua curtipendula, Bouteloua hirsuta, Elymus elymoides, Elymus smithii, Gaura, Grindelia, Haplopappus, Lycurus phleoides, Muhlenbergia torreyi, Opuntia, Plantago, Psoralea, Ratibida, Senecio, Sphaeralcea, Sporobolus cryptandrus, Yucca, Zinnia. Occurrence: Eastern parts of New Mexico and Colorado, southeastern Wyoming, western parts of Nebraska, Kansas, Oklahoma, and Texas.

Wheatgrass-Needlegrass. Dominants: Bouteloua gracilis, Elymus smithii, Stipa comata, Stipa viridula (not in Colorado). Other components: Antennaria, Artemisia, Aster, Echinacea, Elymus trachycaulus, Koeleria macrantha, Liatris, Psoralea, Solidago, Stipa spartea. Colorado only: Chrysothamnus, Festuca, Muhlenbergia, Tetradymia. Occurrence: Western North Dakota and South Dakota, eastern Montana and Wyoming, and Colorado.

Wheatgrass-Bluestem-Needlegrass. Dominants: Andropogon gerardii, Elymus smithii, Stipa spartea. Other components: Elymus trachycaulus, Schizachyrium scoparium, Artemisia, Aster, Bouteloua curtipendula, Bouteloua gracilis, Echinacea, Koeleria cristata, Liatris, Psoralea, Rosa, Solidago, Stipa comata, Stipa viridula. Occurrence: North Dakota, eastern South Dakota, and Nebraska.

Wheatgrass-Grama-Buffalo Grass. Dominants: Bouteloua gracilis, Buchloë dactyloides, Elymus smithii. Other components: Schizachyrium scoparium, Artemisia, Aster, Bouteloua curtipendula, Carex, Echinacea, Juniperus, Liatris, Solidago, Stipa comata. Occurrence: South Dakota.

Bluestem-Grama Prairie. Dominants: Schizachyrium scoparium, Bouteloua curtipendula, Bouteloua gracilis. Other components: Ambrosia, Amorpha, Andropogon gerardii, Buchloë dactyloides, Clematis, Dalea, Echinacea, Elymus smithii, Erysimum, Heodeoma, Liatris, Oenothera, Panicum virgatum, Paronychia, Psoralea, Scutellaria, Sorghastrum nutans, Sporobolus asper, Stenosiphon. Occurrence: Central and western Kansas, extending into adjacent areas of Nebraska, Colorado, and Oklahoma.

Sandsage-Bluestem. Dominants: *Schizachyrium scoparium, Andropogon hallii, Artemisia filifolia,*

Bouteloua hirsuta. Other components: Bouteloua gracilis, Buchloë dactyloides, Calamovilfa longifolia, Eragrostis trichodes, Helianthus, Hordeum jubatum, Panicum virgatum, Redfieldia flexuosum, Sporobolus cryptandrus, Stipa comata, Yucca. Occurrence: Southwestern Nebraska, western Kansas, eastern Colorado, and northwestern Oklahoma.

Shinnery. Dominants: *Schizachyrium scoparium, Quercus mohriana.* Other components: *Acacia, Andropogon hallii, Aristida, Artemisia, Bouteloua gracilis, Bouteloua hirsuta, Buchloë dactyloides, Celtis, Cenchrus, Eriogonum, Juniperus, Prosopis, Prunus, Quercus, Rhus, Sorghastrum nutans, Sporobolus cryptandrus, Yucca.* Occurrence: Panhandle of Texas and adjacent parts of New Mexico and Oklahoma.

Sea Oats Prairie. Dominants: Andropogon littoralis, Uniola paniculata. Other components: Batis, Carex, Croton, Elionurus tripsacoides, Ipomoea, Juncus, Lycium, Paspalum monostachyum, Sesuvium, Xanthoxylum. Occurrence: Islands off southern Texas coast.

Northern Cordgrass Prairie. Dominants: *Distichlis spicata, Spartina alterniflora, Spartina patens.* Other components: *Gerardia, Juncus, Limonium, Plantago, Salicornia, Triglochin.* Other components: Along Atlantic coast from New England southwards to North Carolina.

Bluestem Prairie. Dominants: Andropogon gerardii, Schizachyrium scoparium, Panicum virgatum, Sorghastrum nutans. Other components: Amorpha, Antennaria, Aster, Baptisia, Bouteloua curtipendula, Erigeron, Galium, Helianthus, Koeleria macrantha, Liatris, Panicum leibergii, Panicum scribnerianum, Phlox, Psoralea, Ratibida, Rosa, Silphium, Solidago, Sporobolus heterolepis, Stipa spartea. Occurrence: North Dakota and Minnesota southward to Oklahoma.

Nebraska Sandhills Prairie. Dominants: Andropogon gerardii, Andropogon hallii, Schizachyrium scoparium, Calamovilfa longifolia, Stipa comata. Other components: Artemisia, Asclepias, Carex, Eragrostis trichodes, Erigeron, Gilia, Panicum virgatum, Dalea, Sporobolus cryptandrus, Stipa hymenoides. Occurrence: Nebraska and southern South Dakota.

Blackland Prairie. Dominants: *Schizachyrium scoparium, Stipa leucotricha.* Other components: *Andropogon gerardii, Bothriochloa saccharoides, Aristida purpurea, Bouteloua curtipendula, Bouteloua hirsuta, Bouteloua rigidiseta, Buchloë dactyloides, Panicum virgatum, Sorghastrum nutans, Sporobolus asper.* Occurrence: Texas.

Bluestem-Sacahuista Prairie. Dominants: Andropogon littoralis, Spartina spartinae. Other components: Andropogon glomeratus, Andropogon virginicus, Aristida purpurea, Aristida roemeriana, Buchloë dactyloides, Carex, Paspalum monostachyum, Paspalum plicatulum, Schizachyrium tenerum.

Occurrence: Coastal plains of Texas and Louisiana.

Southern Cordgrass Prairie. Dominants: *Spartina alterniflora*. Other components: *Carex, Distichlis spicata, Juncus, Mariscus, Panicum hemitomon, Panicum repens, Phragmites australis, Sagittaria, Scirpus, Spartina cynosuroides, Spartina patens, Spartina spartinae, Typha, Zizaniopsis miliacea.* Occurrence: Southeastern Texas and southern Louisiana.

Palmetto Prairie. Dominants: Aristida stricta, Serenoa repens. Other components: Andropogon, Aristida spiciformis, Axonopus compressus, Axonopus furcatus, Lyonia, Paspalum distichum, Sabal, Vaccinium. Occurrence: Central Florida.

Oak Savanna. Dominants: Andropogon gerardii, Schiza-chyrium scoparium, Quercus macrocarpa. Other components: Amphicarpa bracteata, Calamovilfa longifolia, Carya, Comandra, Euphorbia, Fraxinus, Monarda, Panicum leibergii, Quercus, Rosa, Sorghastrum nutans, Sporobolus heterolepis, Stipa spartea. Occurrence: Wisconsin, Minnesota, North Dakota.

Cedar Glades. Dominants: Celtis laevigata, Juniperus virginiana, Quercus stellata, Sporobolus neglectus, Sporobolus vaginiflorus, Ulmus alata. Other components: Andropogon gerardii, Arenaria, Bouteloua curtipendula, Bumelia, Carya, Celtis, Cercis, Cheilanthes, Croton, Forestiera, Leavenworthia, Palafoxia, Dalea, Pleurochaete, Psoralea, Quercus, Rhus, Schizachyrium scoparium, Sedum, Symphoricarpos. Occurrence: Tennessee, Alabama, Missouri, and Arkansas.

Cross Timbers. Dominants: Schizachyrium scoparium, Quercus marilandica, Quercus stellata. Other components: Andropogon gerardii, Bouteloua curtipendula, Bouteloua hirsuta, Carya, Celtis, Elymus canadensis, Eragrostis spectabilis, Eragrostis trichodes, Panicum scribnerianum, Panicum virgatum, Sorghastrum nutans, Sporobolus asper, Stipa leucotricha, Ulmus. Occurrence: Texas to Kansas.

Mesquite-Buffalo Grass. Dominants: Buchloë dactyloides, Prosopis juliflora. Other components: Acacia, Aristida purpurea, Aristida roemeriana, Bouteloua gracilis, Bouteloua hirsuta, Bouteloua trifida, Condalia, Juniperus, Quercus, Schedonnardus paniculatus, Yucca. Occurrence: Northwestern Texas and sw. Oklahoma.

Juniper-Oak Savanna. Dominants: Schizachyrium scoparium, Juniperus ashei, Quercus virginiana. Other components: Andropogon gerardii, Aristida glauca, Aristida intermedia, Aristida purpurea, Bouteloua curtipendula, Bouteloua hirsuta, Buchloë dactyloides, Cercis, Erioneuron pilosum, Fraxinus, Hilaria belangeri, Leptochloa dubia, Panicum obtusum, Quercus, Sorghastrum nutans, Sporobolus. Occurrence: Central Texas.

Mesquite-Oak Savanna. Dominants: *Schizachyrium scoparium, Prosopis juliflora, Quercus.* Other components: *Aloysia, Andropogon barbinodis, Aristida intermedia, Aristida purpurea, Bouteloua curtipendula, Bouteloua hirsuta, Bouteloua rigidiseta, Brayodendron, Buchloë dactyloides, Juniperus, Quercus, Ulmus.* Occurrence: Central Texas.

Fayette Prairie. Dominants: *Schizachyrium scoparium, Buchloë dactyloides.* Other components: *Aristida purpurea, Aristida roemeriana, Bothriochloa saccharoides, Bothriochloa tenerium, Paspalum dilatatum, Paspalum plicatulum, Stipa leucotricha.* Occurrence: Southern Texas.

Live Oak-Sea Oats. Dominants: Quercus virginiana var. maritima, Uniola paniculata. Other components: Baccharis, Cenchrus tribuloides, Croton, Ilex, Iva, Juncus, Myrica, Opuntia, Panicum amarum, Sabal, Salsola, Serenoa, Spartina alterniflora, Spartina patens, Yucca. Occurrence: Eastern and Gulf coasts from North Carolina to Alabama.

Cypress Savanna. Dominants: Aristida affinis, Aristida patula, Taxodium distichum. Other components: Acer, Annona, Blechnum, Cyperus, Hypercium, Ilex, Leersia hexandra, Magnolia, Mariscus, Myrica, Persea, Rhynchospora, Salix, Saururus, Spartina bakeri, Stillingia aquatica, Taxodium, Tillandsia, Utricularia. Occurrence: Southwestern Florida.

NATIONAL GRASSLANDS

California:

Butte Valley National Grassland Macdoel, CA	18,000 acres
Colorado:	
Comanche National Grassland Springfield, CO	435,028 acres
Pawnee National Grassland Greeley, CO	193,060 acres
Idaho:	
Curlew National Grassland Malad, ID	47,749 acres

Kansas:

Cimarron National Grassland 108,175 acres Elkhart, KS

Nebraska:

Oglala National Grassland 94,316 acres Chadron, NE

New Mexico:

Kiowa/Rita Blanca National Grasslands136,417 acres Clayton, NM

North Dakota:

Little Missouri National Grassland 525,000 acres

Medora Ranger District Dickinson, ND

Little Missouri National Grassland 503,000 acres McKenzie Ranger District Watford City, ND

Cheyenne National Grassland 70,000 acres Lisbon, ND

Oklahoma:

McClellan Creek/Black Kettle Natl. Grasslands Cheyenne, OK 32,749 acres

Oregon:

Crooked River National Grassland 111,379 acres Madras, OR

South Dakota:

Buffalo Gap National Grassland c. 600,000 acres Fall River Ranger District Hot Springs, SD

Buffalo Gap National Grassland 591,727 acres Wall Ranger District Wall, SD

Cedar River/Grand River National Grasslands Lemmon, SD 161,809 acres

Fort Pierre National Grassland 115,997 acres Pierre, SD

Texas:

Lyndon B. Johnson/Caddo Natl. Grasslands Decatur, TX 38,095 acres

Wyoming:

Thunder Basin National Grassland 572,211 acres Douglas, WY

GRASSLANDS OF SOUTH AMERICA

Grasslands cover about 2 million sq. mi. (or about one-third) of the continent. There are three well-defined and separate regions. The **llano** and savanna of the Orinoco Basin occur mostly in Venezuela and in the Guianas. The common grasses include Andropogon condensatus, Cymbopogon rufus, Sporobolus junceus, Trachypogon plumosus, Panicum maximum, and Aristida spp. The **campo** and savanna of upland Brazil is a region of extreme wet and dry seasons. The grasses are typically tall and coarse. Common species include Melinis minutiflora, Panicum maximum, and Hyparrhenia spp. The **pampa** and prairies of Uruguay and Argentina are broad, level plains and plateaus. The regions may be subhumid, semiarid, or arid. The grass flora is rich. Various species of Stipa, Poa, Sporobolus, Bromus, Paspalum, Bothriochloa, Panicum, Aristida, Hordeum, Melica, Eragrostis, Briza, Axonopus, and Cortaderia are typical.

GRASSLANDS OF AFRICA

Grasses cover about 20% of the continent. Two very different kinds of grasslands are present. The **high-grass savanna** is characterized by tall grasses and low trees. It occurs next to the tropical rain forests

centered in the Congo Basin. Common grasses include Andropogon schimperi, Hyparrhenia rufa, Pennisetum benthamii, Imperata cylindrica, and Pennisetum purpurascens. Sorghum, millets, and rice are also widely cultivated in the region. The **tall-grass savanna** is characterized by tall grasses and various species of Acacia, a shrubby legume. The grasses are about 1-2 m tall; the trees are scattered. It occurs in the eastern and southern parts of the continent. Common grasses include various species of Andropogon, Hyparrhenia, Themeda, and on the drier sites Aristida, Chloris, Melinis, and Digitaria are found.

GRASSLANDS OF ASIA

The Asian grasslands are mostly steppes. They are similar to those found on this continent in their general climatic features, and soils. The steppe region lies on either side of ca. 50° N latitude and extends in an east-west direction. The soils vary from deep, dark fertile soils to sandy, hard clays and to sterile alkaline soils in the South and in the East. The region west of the Volga River, including the Republic of Ukraine, is a rich agricultural area. Common native grasses include various species of *Stipa, Koeleria, Festuca, Poa, Hierochloë, Agropyron, Elymus, Calamagrostis, Aegilops, Haynaldia, Hordeum,* and *Setaria*.

GRASSLANDS OF AUSTRALIA

A huge interior desert with thinly populated grasslands making up a large portion of its rim occurs here. There are also important agricultural areas in New South Wales, Victoria, and in Queensland. The common grasses there (and probably the least well known to us at the generic level) include Astrebla, Dichanthium, Danthonia, Themeda, Iseilema, Poa, Eulalia, Spinifex, Triodia, Imperata, Melinis, Bromus, Festuca, Avena, and Hordeum.

SELECTED REFERENCES

Allred, K. W. 1982. Cousins to the south: amphitropical disjunctions in southwestern grasses. Des. Plants 3: 98-107.

Amme, D. 2004. Stewardship of a changed landscape: grassland heritage. BayNature 4(2): 20-25.

Anderson, R. C. 1990. The historic role of fire in the North American grasslands. <u>In</u>, Collins, S. L. & L. Wallace (editors). Fire in North American tallgrass prairies. Univ. Oklahoma Press. Norman. Pp. 8-18.

Axelrod, D. I. 1985. Rise of the grassland biome, central North America. Bot. Rev. 51(2): 163-201.

Bailey, A. W. 2000. Future of temperate natural grasslands in the northern hemisphere. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO Publishing. Collingwood, Australia. Pp. 361-368.

Baker, H. G. 1976. Invasion and replacement in California and neotropical grasslands. In, Wilson, J. R. (editor). Plant relations in pastures. CSIRO. Canberra, Australia. Pp. 368-384.

Baker, H. G. 1989. Sources of the naturalized grasses

- and herbs in California grasslands. <u>In</u>, Huenneke, L. F. & H. Mooney (editors). Grassland structure and function: California annual grassland. Kluwer Acad. Publ. Dordrechth. Pp. 29-45.
- Baker, M. J. (editor). 1993. Grasslands for our world. SIR Publ. Wellington, New Zealand. 865 pp.
- Barbour, M. & V. Whitworth. 1992. California's grassroots: native or European? Pacific Discovery 45: 8-15.
- Barnard, C. (editor). 1964. Grasses and grasslands. Macmillan & Co. New York, NY. 269 pp.
- Barry, W. J. 1972. Central Valley prairie: Vol. 1. California prairie ecosystem. California State Dept. Parks and Recreation. Sacramento. 82 pp.
- Billings, W. D. 1990. Grasslands. <u>In</u>, Woodwell, G. M. (editor). The earth in transition. Cambridge Univ. Press. Cambridge, England.
- Biswell, H. H. 1956. Ecology of California grasslands. J. Range Manag. 9: 19-24.
- Blumer, M. A. 1992. Some myths about California grasslands and grazers. Fremontia 20(3): 22-27.
- Borchert, J. R. 1950. The climate of the central North American grassland. Ann. Assoc. American Geogr. 40: 1-39.
- Box, T. W. & F. W. Gould. 1958. An analysis of the grass vegetation of Texas. Southw. Nat. 3: 124-129.
- Brown, D. E. 1982. Plains and Great Basin grasslands. Desert Plants 4: 115-121.
- Burcham, L. T. 1957. California rangeland: an historico-ecological study of the range resource of California. Dept. Nat. Res. State of California. Sacramento.
- Burcham, L. T. 1981. California rangelands in historical perspective. Rangelands 3: 95-104.
- Burkhart, A. 1975. Evolution of grasses and grasslands in South America. Taxon 14(1): 53-66.
- Carpenter, J. R. 1940. The grassland biome. Ecol. Monogr. 10: 617-684.
- Clayton, W. D. 1975. Chorology of the genera of Gramineae. Kew Bull. 30: 111-132.
- Clayton, W. D. & T. A. Cope. 1980. The chorology of Old World species of Gramineae. Kew Bull. 35: 135-171.
- Clayton, W. D. & T. A. Cope. 1980. The chorology of North American species of Gramineae. Kew Bull. 35: 567-576.
- Cole, M. M. 1986. The savannas: biogeography and geobotany. Academic Press. London, England. 438 pp.
- Cope, T. A. 2000. Chorology of grasses a review. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO. Collingwood, Australia. Pp. 325-330.
- Cope, T. A. & B. K. Simon. 1995. The chorology of Australasian grasses. Kew Bull. 50: 367-378.

- Coughenour, M. B. 1985. Graminoid responses to grazing by large herbivores: adaptations, exaptations, and interacting processes. Ann. Missouri Bot. Gard. 72: 852-863.
- Coupland, R. T. 1961. A reconsideration of grassland classification in the northern Great Plains of North America. J. Ecol. 49: 135-167.
- Coupland, R. T. (editor). 1979. Grassland ecosystems of the world: analysis of grasslands and their uses. Cambridge Univ. Press. Cambridge, England. 401 pp.
- Coupland, R. T. (editor). 1992. Ecosystems of the world 8A. Natural grasslands: introduction and western hemisphere. Elsevier. Amsterdam, The Netherlands. 469 pp.
- Coupland, R. T. (editor). 1993. Ecosystems of the world 8B. Natural grasslands: eastern hemisphere and resume. Elsevier. Amsterdam, The Netherlands.
- Coupland, R. T. & T. C. Brayshaw. 1953. The fescue grassland in Saskatchewan. Ecology 34: 386-405.
- Cross, R. A. 1980. Distribution of sub-families of Gramineae in the Old World. Kew Bull. 35: 279-289.
- D'Antonio, C. M. & P. M. Vitousek. 1992. Biological invasions by exotic grasses: the grass/fire cycle and global change. Ann. Rev. Ecol. Syst. 23: 63-87.
- Dix, R. L. 1964. A history of biotic and climatic changes within the North American grassland. <u>In</u>, Crisp, D. J. (editor). Grazing in terrestrial and marine environments: a symposium of the British Ecological Soc. Blackwell Sci. Publ. Oxford, U. K. Pp. 71-89.
- Dodd, J. D. 1983. Grassland associations in North America. <u>In</u>, Gould, F. W. & R. B. Shaw. Grass systematics. Second edition. Texas A & M Univ. Press. College Station. Pp. 343-357.
- Dyer, M. I. et al. 1982. The role of herbivores in grasslands. <u>In</u>, Estes, J. R., R. J. Tyrl, & J. N. Brunken (editors). Grasses and grasslands: systematics and ecology. Univ. Oklahoma Press. Norman. Pp. 255-292.
- Edwards, S. W. 1992. Observations on the prehistory and ecology of grazing in California. Fremontia 20(1): 3-11.
- England, R. C. & A. DeVos. 1969. Influence of animals on the pristine conditions on the Canadian grasslands. J. Range Mgt. 22: 87-94.
- Estes, J. R., R. J. Tyrl, & J. N. Brunken (editors). 1982. Grasses and grasslands: systematics and ecology. Univ. Oklahoma Press. Norman. 312 pp.
- Freudenberger, D. O., B. E. Fish, & J. E. Keeley. 1987. Dis-tribution and stability of grasslands in the Los Angeles basin. Bull. S. California Acad. Sci. 86: 13-26.
- Gerlach, J., A. Dyer, & K. Rice. 1998. Grassland and foothill wood-land ecosystems of the Central Valley. Fremontia 26(4): 19-43.
- Groves, R. H. 2000. Temperate grasslands of the southern hemisphere. <u>In</u>, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO. Collingwood, Australia. Pp. 356-360.
- Hamilton, J. G. 1997. Changing perceptions of pre-

- European grasslands in California. Madroño 44(4): 311-333.
- Hartley, W. 1950. The global distribution of tribes of Gramineae in relation to historical and environmental factors. Australian J. Agric. Res. 1: 355-373.
- Heady, H. F. 1977. Valley grassland. <u>In</u>, Barbour, M. G. & J. Major (editors). Terrestrial vegetation of California. John Wiley & Sons. New York, NY. Pp. 491-514.
- Herrara, C. M. 1982. Grasses, grazers, mutualism, and coevolution: a comment. Oikos 38: 254-258.
- Hornby, R. J. 1992. Grasslands. <u>In</u>, Groombridge, B. Global biodiversity: status of the earth's living resources. Chapman & Hall. London, England. Pp. 280-292.
- Huenneke, L. F. & H. A. Mooney (editors). 1989. Grassland structure and function: California annual grassland. Kluwer Acad. Publ. Dordrecht. The Netherlands.
- Humphrey, R. R. 1953. The desert grassland, past and present. J. Range Mgt. 6: 159-164.
- Humphrey, R. R. 1958. The desert grassland: a history of vegetational change and an analysis of causes. Bot. Rev. 24: 193-252.
- Jackson, L. E. 1985. Ecological origins of California's Mediterranean grasses. J. Biogeogr. 12: 349-361.
- Joern, A. & K. Keeler. 1994. The changing prairie: North American grasslands. Oxford Univ. Press. New York, NY. 304 pp.
- Johnston, M. C. 1963. Past and present grasslands of southern Texas and northeastern Mexico. Ecology 44: 456-466.
- Klapp, E. 1964. Features of grassland theory. J. Range Manag. 17: 309-322.
- Küchler, A. W. 1964. Manual to accompany the map "potential natural vegetation of the conterminous United States. Special Publ. No. 36. American Geogr. Soc. New York, NY. 39 + 116 pp.
- Larson, F. 1940. The role of bison in maintaining the short grass plains. Ecology 21: 113-121.
- Lewis, J. K. 1971. The grassland biome: a synthesis of structure and function. <u>In</u>, French, N. R. (editor). Preliminary analysis of structure and function in grasslands. Science Series No. 10. Range Science Dept. Colorado State Univ. Fort Collins. Pp. 317-387.
- Looman, J. 1969. The fescue grasslands of western Canada. Vegetatio 19: 128-145.
- Looman, J. 1980. The vegetation of the Canadian Prairie Provinces. II. The grasslands. Pt. 1. Phytocoenologia 8: 153-190.
- Looman, J. 1981. The vegetation of the Canadian Prairie Provinces. II. The grasslands. Pt. 2. Phytocoenologia 9: 1-26.
- Malin, J. C. 1947. The grassland of North America: prolegomena to its history. Edward Bros. Ann Arbor, MI. 398 pp.

- Love, R. M. 1970. The rangelands of the western U. S. Sci. American 222(2): 88-96.
- MacFadden, B. J. 1997. Origin and evolution of the grazing guild in New World terrestrial mammals. TREE 12(5): 182-187.
- McClaran, M. P. & T. R. Van Devender (editors). 1995. The desert grassland. Univ. Arizona Press. Tucson, 346 pp.
- McMillan, C. 1959. The role of ecotypic variation in the distribution of the central grassland of North America. Ecol. Monog. 29: 285-308.
- McNaughton, S. J. 1968. Structure and function in California grasslands. Ecology 49: 962-972.
- Moss, E. H. & J. A. Campbell. 1947. The fescue grassland of Alberta. Canad. J. Res. C. 25: 209-227.
- Numata, M. (editor). 1979. Ecology of grasslands and bamboo-lands in the world. Gustav Fischer. Jena.
- Orth, M. L. P. 1980. Distribution of the grasses (Gramineae) in relation to plate tectonics. Masters thesis. Oklahoma State Univ. Stillwater.
- Owen, D. F. & R. G. Wiegert. 1982. Grasses and grazers: is there a mutualism? Oikos 27: 258, 259.
- Pagani, M., K. H. Freeman, & M. A. Arthur. 1999. Late Miocene atmospheric CO_2 concentrations and the expansion of C^4 grasses. Science 285: 876-879.
- Risser, P. G. 1988. Diversity in and among grasslands. \underline{In} , Wilson, E. O. (editor). Biodiversity. National Acad. \underline{Press} . Washington, D. C. Pp. 176-180.
- Risser, P. G. et al. 1981. The true prairie ecosystem. Hutchinson Ross. Stroudsburg, PA. 557 pp.
- Root, R. A. & J. R. Habeck. 1974. A study of high elevation grassland communities in western Montana. American Midl. Nat. 87: 109-121.
- Roseveare, G. M. 1948. The grasslands of Latin America. Bull. No. 36. Imperial Bur. Pastures and Field Crops. Aberystwyth. 291 pp.
- Rzedowski, J. 1975. An ecological and phytogeographical analysis of the grasslands of Mexico. Taxon 24(1): 67-80.
- Sauer, C. O. 1950. Grassland climax, fire, and man. J. Range Management 3: 16-21.
- Scholz, H. 1975. Grassland evolution in Europe. Taxon 24(1): 81-90.
- Scholz, H. 1975. Grassland evolution in Europe. Taxon 14(1): 81-90.
- Shantz, H. L. 1954. The place of grassland in the earth's cover of vegetation. Ecology 35: 143-145.
- Shaw, R. B. 2000. Tropical grasslands and savannas. In, Jacobs, S. W. L. & J. Everett (editors). Grasses: systematics and evolution. CSIRO. Collingwood, Australia. Pp. 351-355.
- Shreve, F. 1942. Grassland and related vegetation in northern Mexico. Madrono 6: 190-198.

Silvertown, J. W. 1982. No evolved mutualism between grasses and grazers. Oikos 38: 253, 254.

Simons, G. (editor). 1985. Plant Earth: grasslands and tundra. Time-Life Books. New York, NY. 176 pp.

Sims, P. L. & P. G. Risser. 2000. Grasslands. In, Barbour, M. G. & W. D. Billings (editors). North American terrestrial vegetation. Second edition. Cambridge Univ. Press. Pp. 323-356.

Singh, J. S., W. K. Laurenroth, & D. G. Milchunas. 1983. Geography of grassland ecosystems. Progr. Phys. Geogr. 7: 46-80.

Smith, J. M. B. 1975. Mountain grasslands of New Guinea. J. Biogeogr. 2: 27-44.

Soriano, A. 1979. Distribution of the grasses and grasslands of South America. <u>In</u>, Numata, M. (editor). Ecology of grasslands and bamboolands in the world. Gustav Fischer. Jena. Pp. 84-91.

Stebbins, G. L. 1975. The role of polyploid complexes in the evolution of North American grasslands. Taxon 24(1): 91-106.

Stebbins, G. L. 1981. Coevolution of grasses and herbivores. Ann. Missouri Bot. Gard. 68: 75-86.

Stoddard, L. A. 1941. The Palouse grassland association in northern Utah. Ecology 22: 158-163.

Teeri, J. A. & L. G. Stowe. 1976. Climatic patterns and the distribution of C4 grasses in North America. Oecologia 23: 1-12.

Tilman, D. & J. A. Downing. 1994. Biodiversity and stability in grasslands. Nature 367: 363-365.

Tisdale, E. W. 1947. The grasslands of southern British Columbia. Ecology 28: 346-382.

Transeau, E. N. 1935. The prairie peninsula. Ecology 16: 423-437.

Van Dyne, G. M. 1971. The U. S. IBP grassland biome study – an overeview. <u>In</u>, French, N. R. (editor). Preliminary analysis of structure and function in grasslands. Science Series No. 10. Range Sci. Dept. Colorado State Univ. Fort Collins.

Wagner, F. H. 1989. Grazers, past and present. <u>In</u>, Huenneke, L. F. & H. Mooney (editors). Grassland structure and function in California annual grassland. Kluwer. Dordrecht. Pp. 151-162.

Weaver, J. E. 1954. North American prairie. Johnson Publ. Co. Lincoln, NB. 348 pp.

Weaver, J. E. 1958. Summary and interpretation of underground development in natural grassland communities. Ecol. Monogr. 28: 55-78.

Weaver, J. E. & F. W. Albertson. 1956. Grasslands of the Great Plains. Johnson Publ. Co. Lincoln, NB. 395 pp.

Weaver, J. E. & T. J. Fitzpatrick. 1934. The prairie. Ecol. Monogr. 4: 109-295.

Weaver, T. 1979. Climates of fescue grasslands of mountains in the western United States. Great Basin Nat. 39(3): 284-288.

Wedin, D. A. 1992. Grasslands: a common challenge. Restoration and Management Notes 10(2): 137-143.

West, N. E. 1993. Biodiversity of rangelands. J. Range Mgt. 46: 2-13.

Wester, L. 1981. Composition of native grasslands in the San Joaquin Valley, California. Madroño 28(4): 231-241.

Whyte, R. O. 1968. Grasslands of the monsoon. Faber & Faber. London, U. K. 325 pp.

6.08 - TOXIC GRASSES

Because the grass family is the source of so many important food plants, it may come as a surprise to learn that it is also the source of a number of poisonous plants. The victims of grass toxicity are wild and domesticated animals, humans, and even other plants. Symptoms range from those that are mildly irritating to death. Mechanisms of poisoning include:

- plant parts that cause mechanical injury;
- absorption of toxins from the soils where grasses are growing;
- manufacture of one or more toxins by the grass itself;
- acting as a host to a fungus that makes the toxins.

MECHANICALLY INJURIOUS*

Some grasses are toxic only in the broadest sense because they are armed with stout awns that can cause mechanical injury. The sites of penetration, often around in eyes, snout, or the soft parts of the mouth cavity, can become infected and then further complications may occur. Such grasses do not actually produce toxic substances.

Scientific (Common) Name	Cause
Aristida spp. (three-awns) Avena fatua (wild oat) Bromus spp. (bromes) Cenchrus spp. (sand burs) Hordeum spp. (barleys) Leersia spp. (cut grasses) Setaria spp. (foxtails, bristle grasses) Spartina spp. (cord grasses) Stipa spp. (needle grasses)	awns awns spines awns leaf margins awns leaf margins awns

NITRATE/NITRITE INTOXICATION*

Others species, especially cereal crops and agricultural weeds are toxic not because they contain a poison, but because they absorb it from fertilizer-rich soils and then sequester in the plant body.

Scientific Name	Common Name
Avena sativa Cynodon spp. Echinochloa frumentacea Hordeum jubatum Hordeum vulgare Lolium spp. Pennisetum glaucum Sorghum spp. Triticum aestivum Zea mays	Oat Bermuda grasses Sanwa millet Foxtail barley Barley Rye grasses Pearl millet Sorghums Bread wheat Maize, corn

CYANOGENIC GLYCOSIDES*

√ General Features:

- Found widely in plant kingdom
- Common in roses, grasses, legumes, spurges
- HCN glycoside (sugar + toxic component)
- Intact glycoside harmless
- Activated by chewing, crushing, freezing, etc.
- Pure HCN very toxic
- Readily absorbed on skin; dangerous when inhaled

√ Symptoms:

- Acts at cellular level
- Blocks release of oxygen from red blood cells
- Instantaneous collapse (large doses)
- Weakness, giddiness, headache
- Nausea & vomiting
- Coma
- Death from cellular asphyxiation

Scientific Name Common Name

Agrostis stolonifera
Andropogon spp.
Avena sativa
Bambusa spp.
Bothriochloa spp.
Bouteloua gracilis

Creeping bent grass
Bluestems
Coat
Bamboos
Bamboos
Both World bluestems
Blue grama

Bouteloua hirsuta Hairy grama
Briza spp. Quaking grasses
Catabrosa aquatica Brook grass
Chloris truncata Australian finger grass
Cortaderia spp. Pampas grasses

Cymbopogon spp. Lemon grasses
Cynodon spp. Bermuda grass, star grasses
Dactyloctenium aegyptium Egyptian crowfoot
Danthonia semiannularis Oat grass
Eleusine coracana African millet

Eleusine indica Goose grass
Elymus spp. Wild ryes
Festuca spp. Fescues
Glyceria canadensis Canadian manna grass
Glyceria grandis American manna grass

Glyceria septentrionalis Eastern manna grass Holcus lanatus Velvet grass Hordeum vulgare Barley Lagurus ovatus Hare's-tail grass

Lamarckia aurea Goldentop

Leersia hexandraClubhead cutgrassLeptochloa dubiaGreen sprangletopLolium perennePerennial rye grass

Melica altissimaOnion grassMolinia caeruleaPurple moor grassOryza sativaRicePanicum maximumGuinea grassPanicum muticumPara grassPoa pratensisKentucky bluegrass

Secale cerealeRyeSorghastrum nutansIndian grassSorghum x almumColumbus grassSorghum bicolorSorghum, Sudan grassSorghum halepenseJohnson grassStipa robustaSleepy grass

Tridens flavusPurpletopTriticum aestivumBread wheatZea maysMaize, corn

PHOTOSENSITIZATION*

Several grasses are implicated in this syndrome. Sensitive animals are poisoned when they eat grasses that contain certain pigments that react with sunlight to form toxins that can cause damage to their skin and underlying tissues. Probably the best known example of this phenomenon is found in our local weedy Klamath weed or St. John's wort (*Hypericum perforatum*), a member of Guttiferae.

Scientific Name Common Name

Avena sativa Oat
Cenchrus incertus Southern sandbur
Echinochloa crusgalli Barnyard grass
Eriochloa contracta Prairie cup grass
Hordeum murinum Wall barley

Hordeum vulgareBarleySecale cerealeAnnual ryeSetaria italicaFoxtail milletSorghum spp.SorghumTriticum aestivumBread wheat

GRASS TETANY*

Also known as grass staggers, this syndrome appears to be associated with ionic imbalances in the blood serum after eating large amounts of lush growth. Low magnesium levels are typical. Animals suffering from grass tetany first show signs of excitement, poor coordination, and anorexia, followed by cardiovascular involvement, convulsions, coma, and death.

Scientific Name Common Name

Agropyron spp.Wheat grassesAvena sativaOatBromus spp.BromesDactylis glomerataOrchard grassElymus spp.Wild ryes

Festuca spp. Fescues
Hordeum spp. Barleys
Lolium perenne Perennial rye grass
Phalaris spp. Canary grasses
Phleum pratense Timothy
Secale cereale Annual rye

ERGOTISM*

√ Chronic or Gangrenous Ergotism

- Small amounts over long period of time
- Constriction of blood vessels
- Death of tissues
- Loss of extremities

√ Acute or Convulsive Ergotism

- Larger amounts quickly consumed
- Central nervous system
- Crawling sensation on skin
- ☼ Tingling of skin; fingers
- Tinnitus aurium
- Headache
- Vertigo
- Vomiting & diarrhea
- Hallucinations
- Painful muscular contractions
- Epileptiform seizures

Scientific Name

Common Name

Agrostis spp. Bent grasses Bromus spp. Brome grasses Dactylis glomerata Orchard grass Elymus spp. Wild ryes Koeleria spp. June grasses Phalaris spp. Canary reed grasses Bluegrasses Poa spp. Secale cereale Rye

GRASSES + ENDOPHYTIC FUNGI

Interestingly, the best known cases of poisoning from ingesting grasses are caused by parasitic fungi that live on the plants. It is the fungus that makes the toxin, not the grass. The so-called endophytic fungi are the subject of much recent research, particularly those that infect fescue grasses. The following grasses have been found to serve as hosts to such fungi.

Scientific Name Common Name

Agropyron spp. (s. l.)

Agrostis spp.

Andropogon spp.

Cynodon spp.

Elymus spp.

Wheat grasses
Bent grasses
Bluestems
Bermuda grass, star grasses
Wild ryes

Festuca spp.FescuesGlyceria spp.Manna grassesHolcus spp.Velvet grassesHilaria spp.Tobosas, galletasHystrix spp.Bottlebrush grasses

Lolium spp.Rye grassesPaspalum spp.Paspalum, water grassesPoa spp.BluegrassesSphenopholis spp.Wedgescales

*After Burrows, G. E. & R. J. Tyrl. 2001. Toxic plants of North America. Iowa State Univ. Press. Ames. Pp. 872-888.

SELECTED REFERENCES

Anas, K. et al. 1998. A survey concerning the equine fescue toxicosis malady. J. Equine Vet. Sci. 18: 631-637.

Bacon, C. W. 1986. Ergot toxicity from endophyte-infected grasses: a review. Agron. J. 78: 106-116.

Bacon, C. W. 1995. Toxic endophyte-infected tall fescue and range grasses: historic perspectives. J. Animal Sci. 73: 861-870.

Ball, D. M., J. F. Pedersen, & G. D. Lacefield. 1993. The tall-fescue endophyte. American Sci. 81(4): 370-379.

Bourke, C. A. 1994. The evidence against the existence of so-called convulsive ergotism in ruminants. <u>In</u>, Colegate, S. M. & P. R. Dorling (editors). Plant-associated toxins. CAB International. Wallingford, U. K. Pp. 387-392.

Burrows, G. E. & R. J. Tyrl. 2001. Poaceae. <u>In</u>, Toxic plants of North America. Iowa State Univ. Press. Ames. Pp. 870-977.

Casabuono, A. C. & A. B. Pomilio. 1997. Alkaloids from endophye-infected *Festuca argentina*. J. Ethnopharm. 57(1): 1-10.

Cheplick, G. P. 1997. Efects of endophytic fungi on the phenotypic plasticity of *Lolium perenne* (Poaceae). American J. Bot. 84(1): 34-40.

Clay, K. 1988. Fungal endophytes of grasses: a defensive mutualism between plants and fungi. Ecology 69: 10-16.

Clay, K. 1990. Fungal endophytes of grasses. Ann. Rev. Ecol. Syst. 21: 275-297.

Clay, K. 1993. The ecology and evolution of endophytes. Agric., Ecosystems & Environ. 44: 39-64.

Clay, K. & J. Holah. 1999. Fungal endophyte symbiosis and plant diversity in successional fields. Science 285: 1742-1744.

Cross, D. L., L. M. Redmond, & J. R. Strickland. 1995. Equine fescue toxicosis: signs and solutions. J. Animal Sci. 73: 899-908.

Epstein, W., K. Gerber, & R. Karler. 1964. The hypnotic constituent of *Stipa vaseyi* sleepy grass. Experientia 20: 390.

Fichte, B. E. 1972. Bermuda grass tremors. Farm Qtr. March: 62, 63.

Frey, M. et al. 1997. Analysis of a chemical plant defense mechanism in grasses. Science 277: 696-699.

Fuller, T. C. & E. McClintock. 1986. Poaceae. <u>In</u>, Poisonous plants of California. Univ. California Press. Berkeley. Pp. 292-304.

Gallagher, C. H., J. H. Koch, & H. Hoffman. 1966. Diseases of sheep due to ingestion of *Phalaris tuberosa*. Australian Vet. J. 42: 279-286.

Goodwin, D. E. 1967. Ergot poisoning of cattle grazing Dallisgrass. J. American Vet. Med. Assoc. 151: 204, 205.

Grunes, D. L., P. R. Stout, & J. R. Brownell. 1970.

- Grass tetany of ruminants. Adv. Agron. 22: 331-374.
- Hemken, R. W., J. A. Jackson, Jr., & J. A. Boling. 1984. Toxic factors in tall fescue. J. Animal Sci. 58:
- Kingsbury, J. M. 1964. Gramineae. \underline{In} , Poisonous plants of the United States and Canada. Prentice-Hall. Englewood Cliffs, NJ. Pp. 475-500.
- Leuchtmann, A. 1992. Systematics, distribution, and host specificity of grass endophytes. Nat. Toxins 1: 150-162.
- Olofsdotter, M. 2001. Rice a step toward use of allelo-pathy. Agron. J. 93(1): 3-8.
- Mantle, P. G. & R. H. C. Penny. 1981. Tremorgenic mycotoxins and neurological disorders a review. Vet. Ann. 21: 51-62.
- Petroski, R. J., R. G. Powll, & K. Clay. 1992. Alkaloids of *Stipa robusta* (sleepygrass) infected with an *Acremonium* endophyte. Na.t Toxins 1: 84-88.
- Porter, J. K. 1995. Analysis of endophyte toxins: fescue and other grasses toxic to livestock. J. Animal Sci. 73: 871-880.
- Powell, R. G. & R. J. Petroski. 1992. Alkaloid toxins in endophyte-infected grass. Nat. Toxins 1: 163-170.
- Pratley, J. E. 1996. Allelopathy in annual grasses. Plant Protection Qtr. 11: 213, 214.
- Rice, E. L. 1972. Allelopathic effects of *Andropogon virginicus* and its persistence in old fields. American J. Bot. 59: 752-755.
- Rowe, L. D. 1989. Photosensitization problems in livestock. Vet. Clin. North Amer. Food Animal Pract. 5(2): 301-323.
- Saha, D. C., M. A. Jackson, & J. M. Johnson-Cicalese. 1988. A rapid staining method for detection of endophytic fungi in turf and forage grasses. Phytopathol. 78: 237-239.
- Schardel, L. 1996. *Epichloe* species: fungal symbionts of grasses. Ann. Rev. Phytopathol. 34: 109-130.
- Selmar, D., Z. Irandoost, & V. Wray. 1996. Dhurrin-6'-glucoside, a cyanogenic diglucoside from *Sorghum bicolor*. Phytochem. 43: 569-572.
- Siegel, M. R., G. C. M. Latch, & M. C. Johnson. 1985. *Acremonium* fungal endophytes of tall fescue and perennial ryegrass: significance and control. Plant Dis. 69: 179-183.
- Siegel, M. R. et al. 1987. Fungal endophytes of grasses. Ann. Rev. Phytopathol. 25: 293-315.
- Smalley, H. E. & H. R. Crookshank. 1976. Toxicity studies on sleepy grass, *Stipa robusta*, (Vasey) Scribn. Southwest Vet. 29: 35-39.
- Strain, G. M., C. L. Seger, & W. Flory. 1982. Toxic Bermuda grass tremor in a goat: an electroencepahlographic study. American J. Vet. Res. 43: 158-162.
- Strickland, J. R., J. W. Oliver, & D. L. Cross. 1993. Fescue toxicosis and its impact on animal agriculture. Vet. Human Toxicol. 35: 454-464.

- Stuedemann, J. A. & J. S. Hoveland. 1988. Fescue endophyte: history and impact on animal agriculture. J. Prod. Agric. 1: 39-44.
- Taylor, M. C., W. F. Loch, & M. Ellersieck. 1985. Toxicity in pregnant pony mares grazing Kentucky-31 fescue pastures. Nutr. Rep. Int. 31: 787-795.
- Thompson, F. N. & J. K. Porter. 1990. Tall fescue toxicosis in cattle: could there be a public health problem here? Vet. Human Toxicol. 32(Suppl.): 51-57.
- Tinnin, R. O. & C. H. Muller. 1971. The allelopathic potential of *Avena fatua*: influence on herb distribution. Bull. Torrey Bot. Club 98: 243-250.
- Tinnin, R. O. & C. H. Muller. 1972. The allelopathic influence of *Avena fatua*: the allelopathic mechanism. Bull. Torrey Bot. Club 99: 287-292.
- Van Rensburg, S. J. & B. Altenkirk. 1974. *Claviceps purpurea* ergotism. <u>In</u>, Mycotoxins. Elsevier. Amsterdam. Pp. 69-96.
- Vogel, K. P., F. A. Haskins, & H. J. Gorz. 1987. Potential for hydrocyanic acid poisoning of livestock by Indiangrass. J. Range Manage. 40: 506-509.
- White, J. F., Jr. 1987. Widespread distribution of endophytes in the Poaceae. Plant Dis. 71: 340-342.
- White, J. F., Jr. & P. M. Halisky. 1992. Association of systemic fungal endophytes with stock-poisoning grasses. <u>In</u>, James, L. F. et al. (editors). Poisonous plants: Proc. Third Intern. Symp. Iowa State Univ. Press. Ames. Pp. 574-578.
- Woods, L. W., J. Bradley Jones, & P. G. Mantle. 1966. An outbreak of gangrenous ergotism in cattle. Vet. Rec. 78: 742-749.

ALLERGIC REACTIONS

- Howlett, B. J. et al. 1981. Immunofluorescent localization of two water-soluble glycoproteins including the major allergen from pollen of rye grass (*Lolium perenne*). Histochem. J. 13: 461-480.
- Howlett, B. J. & A. E. Clarke. 1981. Isolation and partial character-ization of two antigenic glycoproteins from rye grass pollen. Biochem. J. 197: 695-706.
- Johnson, P. & D. G. Marsh. 1965. The isolation and characterization of allergens from the pollen of rye grass (*Lolium perenne*). J. Europ. Polymers 1: 63-77.
- Lowenstein, H. 1978. Isolation and partial characterization of three allergens of timothy pollen. Allergy 33: 30-41.
- Malley, A. & R. L. Harris. 1967. Biological properties of a non-precipitating antigen from timothy pollen extracts. J. Immunol. 99: 825-830.
- Weeke, B. et al. 1974. Allergens in timothy pollen identified by crossed radioimmunologelectrophoresis. Acta Allerg. 29: 409-417.

SECTION 7.0 - G L O S S A R Y

- A -

abaxial: the side of a structure facing away from an axis

achene: a dry, single-seeded indehiscent fruit whose seed coat and fruit wall separate from one another, as in the sedges

acicular: needle-shaped, as in the leaves of *Monanthochloë*

acuminate: gradually tapering to an extended point

acute: sharp-pointed

adaxial: the side of a structure facing toward an axis

adnate: the fusion of unlike parts

adventitious: originating from mature tissues rather than meristematic ones, as in aerial roots that arise from a location other than the primary root system or aerial bulbs

adventitious embryony: a type of apomixis (q. v.) in which the embryo arises in the nucellus or in the integument, rather than within the embryo sac

adventive: a plant that is introduced accidentally

aerial: growing above ground, rather than in the soil

agamospermy: the production of seed without the prior fusion of gametes

aleurone: the outermost protein-rich layer(s) of endosperm

allopolyploid: a type of polyploid that contains genomes that are different from one another, often from two or more species

amphiploid: an allopolyploid that behaves as though it were a diploid

androecium: the male portion of a flower, consisting of one or more stamens

andromonoecious: the condition of having both bisexual and male flowers in the same inflorescence, as in most panicoid spikelets

anemophily: wind-mediated pollination

aneuploid: the condition of having a chromosome number that is not an exact multiple of the base number for that organism

annual: living for a single growing season

anther: the sac-like, pollen-producing part of a stamen

anthesis: the phase during which a flower is fully opened and pollination occurs

anthoecium: the collective term for the lemma and palea

antrorse: directed upwards, as in barbs on an awn

apex: the upper or distal end of a structure; plural,
apices

aphyllopodic: the condition of having bladeless lower leaves, as in some sedges

apiculate: an apex that bears a short, typically
flexible point

apomixis: a type of reproduction that involves the organs and processes typically associated with sexual reproduction, but which does not involve the actual fusion of egg and sperm nuclei; used more loosely as a synonym for asexual reproduction

apospory: a type of apomixis in which the embryo sac is derived from a cell of the inner integument

apposed: when similar parts occur close to one another or side by side

appressed: lying against a surface or, in the case of inflorescence branches, against a central or principal axis

arista: an awn or beard

aristate: an apex that tapers to a very narrow, elongate, bristle-like point; awned

arm cell: a leaf mesophyll cell type, characteristic of bambusoid grasses, in which the internal partitions or septae are incomplete

ascending: growing upward, obliquely at first and then erect, as in certain grass stems

asexual: any form of reproduction that does not involve the union of egg and sperm

attenuate: gradually narrowed to a slender point

auricles: the paired, ear-shaped appendages at the apex of the sheath in some grasses

autopolyploid: an organism with three or more chromosome sets that are \pm identical to one another; often the result of doubling of chromosomes or through unreduced gametes

awl-shaped: the leaf or bract shape characterized by a gradual taper from the base to a sharp point

awn: a substantial hair or bristle that arises from the apex or back of glumes or lemmas [very rarely paleas]; awned, having awns

axile: the interior angle formed by a stem and the petiole or pedicel that it bears

axillary inflorescence: an inflorescence that arises from a lateral position on a culm, as opposed to one that is terminal

axis: the central stem of an inflorescence

- B -

balanced: having spikelets ± equally inserted on both sides of a central axis

basifixed: said of an anther that is attached to a filament by its base, as opposed to being attached at its midpoint

beak: a prominent sterile elongation of a caryopsis

beard: a line or tuft of hairs

beautiful: of or pertaining to grasses, especially native ones

berry: a multi-seeded, indehiscent fruit in which the fruit wall is fleshy throughout; common examples include the tomato and grape; in the grass family, berries are found in certain bamboos

bifid: two-cleft or two-lobed, as in the apex of a lemma or glume

bisexual: a flower, floret, or spikelet that bears both male and female reproductive structures; the term perfect is also used for this condition

blade: the flattened, expanded portion of a leaf

body: the portion of a glume, lemma, or palea, exclusive of awns or teeth

B. P.: Before Present

bract: a reduced leaf; glumes, lemmas, and paleas are all considered bracts

bractlet: a small bract

bran: the outer layers of a cereal grain that are removed during the grinding process

bristle: a short, stiff hair; a sterile branch

bulb: an underground plant structure consisting of a series of overlapping leaf bases attached to a much-reduced stem axis; many bulbs are actually corms (q. v.)

bulbils: small axillary bulbs that replace more typical florets or spikelets, as in *Poa bulbosa*

bulblet: a small bulb

bulliform cells: the comparatively large, thin-walled, colorless epidermal cells of the intercostal zone of the grass leaf blade

bundle sheath: the layer of tissue that surrounds the vascular bundle; not all grasses have bundle sheaths

- C -

C₃ **grasses**: those grasses, often found in the cooler, temperate regions, whose photosynthetic pathway has a 3-carbon compound (3-phosphoglycerate) as its first detectable sugar precursor

 $\mathbf{C_4}$ grasses: those grasses, often subtropical and tropical, whose photosynthetic pathway has a 4-

carbon compound (malate or aspartate) as its first sugar precursor

caducous: falling off early

caespitose: occurring in clumps or tufts; also spelled cespitose

callus: a hardened, often pointed, base of a lemma or floret

canescent: a vestiture type characterized by gray pubescence

capillary: hair-like, as in delicate panicle branches or awns

capitate: aggregated into a dense, head-like cluster, as in an arrangement of spikelets

capsule: a dry, dehiscent fruit that opens by means of slits, lids, pores, or teeth as in the rush family

carinate: having a longitudinal ridge on the dorsal surface, as in the keel of a ship

cartilaginous: having a hard, tough cartilage-like texture

caryopsis: a dry, single-seeded, indehiscent fruit whose seed coat and fruit wall are inseparable; the fruit type of the grasses

cauline: pertaining to a stem, as in cauline leaves that are inserted along a stem, as opposed to its base

chartaceous: having a papery or tissue-like appearance, as in certain leaf and bract margins

chasmogamous: characterized by pollination and fertilization of open flowers or florets

chorology: the study of the distribution and composition of the components of a flora

ciliate: fringed with marginal hairs

ciliolate: as in ciliate, but the hairs minute

cleistogamous: the condition in which flowers or florets remain closed and are typically self-pollinated

cleistogene: a spikelet with self-compatible flowers that remains hidden within basal leaf sheaths

closed sheath: a sheath in which the two edges are fused with one another to form a continuous cylinder around the culm, as in brome grasses and orchard grass

coleoptile: the sheath that covers the shoot apex in the monocot embryo

coleorhiza: the sheath that covers the primary root in the monocot embryo

collar: the band of tissue located at the junction of the blade and sheath of the grass leaf; sometimes differentiated by its lighter color

column: the lower, sometimes twisted portion of an awn; the fused bases of awns, as in *Aristida*

coma: a tuft of hairs; comose, having a coma

complex: a group of closely related, difficult to differentiate, taxa, as in the *Festuca microstachys* complex

compound raceme: an inflorescence in which the peduncle bears two or more branches, each bearing a raceme of spikelets

compound rame: an inflorescence type in which the peduncle bears two or more branches, each bearing a rame of spikelets

compound spike: an inflorescence type in which the peduncle bears two or more branches, each bearing a spike of spikelets

compressed: flattened, as if pressure had been applied to a structure from the back or sides

continuous: not breaking apart; remaining intact, as in the central axis of an inflorescence at maturity

contracted: narrowed, as opposed to open or spreading

convolute: rolled longitudinally, with one edge completely within the other, as in a rolled up leaf blade

cordate: heart-shaped, as in the shape of certain leaf

coriaceous: leathery, as in the texture of certain lemmas or glumes

corm: a dense, vertical, underground stem surrounded by dry, papery leaf bases; often loosely called a bulb

corrugated: wrinkled

cosmopolitan: common to all or to most of the world

costal: the region on a grass leaf that is above the strands of vascular tissue (nerves)

culm: the stem of a grass plant

cultivar: a cultivated variety; a cultivated strain of a

crop plant or of an ornamental

cuneate: wedge-shaped

cv.: cultivar

- D -

deciduous: falling from a plant at the end of a season

decumbent: said of stems that lie on the ground, but whose ends are upturned

denticulate: minutely toothed

depauperate: not fully developed, stunted; often the result of growing on an impoverished site

diaspore: from the Greek word for dispersion, a unit of plant dispersal; examples include caryopses, grains with husks remaining attached, groups of spikelets, etc.

diffuse: widely or loosely spreading, as in inflorescence branches

digitate: having parts that radiate from a central point, as do the fingers of a hand

dimorphic: having two different shapes, as in the glumes of *Koeleria*

dioecious: a species in which staminate and pistillate flowers or spikelets occur on separate plants, as in buffalo grass

diploid: the chromosome complement found in vegetative cells of the plant body; typically expressed in terms of "2n," as in 2n = 14

disarticulation: the separating or disjoining of spikelet parts from one another or of portions of an inflorescence axis from one another

distal: at the end opposite the point of attachment, as opposed to proximal (q. v.)

distichous: attached in two vertical rows, as in leaves on a stem or spikelet bracts on a rachilla

divaricate: spread very far apart, as in inflorescence branches

divergent: spread apart from one another, as in divaricate, but less so

dorsal: relating to or attached to the back of an organ, the side that is turned away from the axis

dorsally compressed: flattened, as if pressure had been brought to bear on the back of bract

- E -

elliptical: in the form of a flattened circle

emarginate: a leaf or bract with a shallow notch at its apex

endemic: confined to a particular region, applied especially when the area is relatively small

endosperm: the nutritive tissue within the seed that originates from the fusion of polar nuclei and sperm nucleus

entire: said of a margin of a leaf or bract that lacks lobes or teeth

entomophily: insect-mediated pollination

epiblast: the small, nonvasculated flap of tissue that occurs on the side opposite the scutellum in some, but not all, grass embryos

erose: said of a margin that appears to have been gnawed or worn away

euploid: having a chromosome number that is an exact multiple of the base chromosome number

exserted: protruding beyond or out of another structure, as in an inflorescence from a sheath

extirpate: to eliminate or destroy; literally, to pull up by the roots

extravaginal branching: a type of branching in which the shoot breaks through the leaf sheath

- F -

fascicle: a tight cluster or clump, as in leaves, axillary stems, or spikelets

fertile lemma: a lemma that encloses a flower

filament: the delicate stalk that supports an anther

filiform: thread-like

fimbriate: fringed, as in a bract margin

flabellate: fan-shaped

floret: a subunit of a spikelet, consisting of a lemma, palea, and flower; sometimes incorrectly defined as only the flower itself

forb: any herbaceous plant that is not a grass or does not appear grass-like

fusoid cells: the large, colorless mesophyll cells found in the leaves of most bambusoid grasses

- G -

gamete: a sex cell; the egg or sperm

gametophytic apomixis: a kind of reproduction in which a diploid embryo sac is produced because reduction division did not occur during the meiotic cycle

geniculate: said of structure that is bent sharply, as in certain grass stems and awns

genome: all of the genetic information found in a single complete set of chromosomes in an organism

genus: a group of related species; the first component of a scientific name

gibbous: having a pouch-like enlargement on one side of a structure, as at the base of a glume

glabrous: without hairs

gland: a secretory structure; used more broadly for any warty protuberance; glandular, having or bearing glands

glaucous: having a blue-gray or sea-green color; also used for a whitish waxy covering that can be easily rubbed off

globose: almost spherical

glume: a sterile bract at the base of a spikelet; most grasses have two such structures, some have one, a few have none

glutinous: covered with a sticky exudate

grain: the fruit of the grass family; see caryopsis

graminoid: grass-like

gynoecium: the female portion of a flower, consisting of the seed-producing components (carpels)

- H -

H & C: an abbreviation for Hitchcock & Chase, the authors of the Manual of the Grasses of the United

States

habit: the general appearance of a plant

haploid: the chromosome complement found in the nuclei of gametes; often expressed by the letter n, as in n=7

herb: an annual, biennial, or perennial plant whose stems die back to the ground at the end of the growing season because they lack the firmness of sufficient secondary growth

herbaceous: having the features of an herb

hirsute: having coarse, ± erect hairs

hispid: having long, rigid, bristly hairs; hispidulous, minutely hispid

hulls: the bracts of the grass spikelet, especially the lemma and palea

hyaline: having a colorless, thin, translucent or transparent texture

hybrid: a plant or animal that is the offspring of a cross between two or more strains, breeds, varieties, species, or genera; hybrids occur spontaneously in nature and they are created in the garden and laboratory

hybridization: the natural or artificial methods by which hybrids are created

hygroscopic: absorbing or attracting water; water sensitive, as in the grass lodicules

- I -

imbricate: overlapping one another, as in bracts of the spikelet

included: occurring within, as opposed to protruding from

indigenous: native to a region

indurate: hard, as in texture

inflorescence: the flowering portion of a grass plant; the arrangement of spikelets on a culm

innovation: a basal, typically vigorous offshoot

inserted: joined to or placed on, as in leaves on a stem or bracts on the spikelet axis

intercostal: the region between the nerves or veins of vascular tissue on a leaf or bract

internode: the region between two consecutive nodes on a stem

intravaginal branching: a type of branching in which the side shoot emerges from the top of the leaf sheath, as opposed to breaking through the side of the sheath

introduced: purposefully brought into a region, as in the case of a crop plant or an ornamental

involucre: an organized set of bracts or of branchlets that surrounds or forms a series or set beneath a spikelet, group of spikelets, or floret

involute: with both edges rolled longitudinally inward toward the midpoint of a leaf or bract

- J -

joint: the node of a grass stem; a point where articulation or disarticulation occurs

- K -

keel: a prominent ridge or rib, as seen in some glumes, lemmas, or paleas

kranz syndrome: the set of anatomical and physiological traits that are found in those plants that have the C_4 photosynthetic pathway

- L -

lanceolate: a leaf blade or bract that is narrow and tapers on both ends and that is widest above the middle; not to be confused with Lancelot, the most famous of King Arthur's knights

laterally compressed: flattened, as if pressure had been brought to bear on the sides of a bract, as opposed to the back (dorsally compressed)

lemma: one of the two bracts enclosing the grass flower, the other being the palea; the lemma is typically the larger bract

lenticular: lens-shaped, as in the appearance of certain seeds or fruits

leptomorph rhizome: the type of slender rhizome found in certain bamboos, in which each node as a bud and a whorl of roots

lignified: woody

ligule: the membranous flap or series of hairs at the junction of the sheath and blade of the grass leaf

linear: several to many times longer than wide, as in the typical blade of a grass leaf

 $\begin{tabular}{ll} \textbf{lodicule}: the reduced perianth of the grass flower; \\ these tiny, mitten-shaped structures are all that remains of the calyx \\ \end{tabular}$

- M -

macrohair: any of the larger, easily seen surface hairs on a plant structure

malt: germinated cereal grain, often barley, used as an enzyme source in brewing and distilling

membranous: soft, thin, and pliable, as in the texture of a glume or lemma

meristem: the region of actively dividing cells of the stem or root apex; the meristematic region of the grass leaf occurs at its base, thereby permitting the plants to survive grazing, fires, and lawn mowers

mesocotyl: that portion of the grass embryo axis that occurs between the node where the scutellum and the coleoptile are attached

metabolically challenged: dead

microhair: any of the more or less microscopic hairs

that occur on the surface of plant parts; in the grasses, they are of diagnostic significance

midrib: the central rib of a leaf or bract

minute: small, as in the size of your vocabulary if you had to use the glossary for this term

monoecious: said of a species in which the staminate and pistillate flowers or spikelets occur on the same plant, as in maize or Job's tears

monotypic genus: a genus of only one species

mucro: a short, sharp point or extension, as seen at the tip of lemma or glume; mucronate: bearing a mucro

muricate: a surface characterized by short, hard, tubercular outgrowths

- N -

 ${f n}:$ the chromosome number found in the nuclei of sex cells; in diploid organisms ${f n}$ equals the haploid chromosome number

native: originating naturally in a particular region; occurring in an area before the arrival of humans, especially European explorers, traders, etc.

naturalized: not native to a particular area, but now well established and maintaining itself without our assistance

nerve: a vein or strand of vascular tissue, appearing as ridges on the surface of glumes, lemmas, or paleas

neuter: lacking reproductive structures; sterile
node: the point or region on a stem where a leaf or
bract is attached

nut: a dry, hard, indehiscent, 1-seeded fruit

- 0 -

oblique: slanting or unequal-sided

oblong: much longer than broad, with the sides \pm parallel, as in the shape of certain leaf blades

obovate: of the shape of an inverted egg

obtuse: blunt in form; also dull in perception or intellect, as in the people who find grasses ugly and boring

oides: a suffix meaning resembling

open sheath: a sheath in which the two edges touch one another or overlap, but are not fused to form a collar or cylinder

ovary: the seed-bearing portion of a flower

- P -

pachymorph rhizome: the short, thick type of rhizome found in some bamboos, in which lateral buds typically produce only additional rhizomes

palea: one of two bracts enclosing the grass flower, the other being the lemma; typically the smaller and more delicate of the two

pampas: the vast open grasslands of South America, especially those in Argentina and Uruguay, dominated by taller bunch grasses in the east and by shorter grasses and shrubs in the drier southern and western portions

panicle: an elongate or rounded, much-branched inflorescence in which the spikelets are attached on the outermost branchlets

papillate: bearing small pimple- or nipple-like protuber-ances; also papillose

parallel: extending in the same direction and equidistant, as in the veins of most grass blades

parthenogenesis: meaning "virgin birth," a kind of apomixis in which the embryo develops from an egg cell that was not fertilized

pearl: to remove the outer layer(s) of a grain by exposing them to abrasive surfaces that grind away the tissues

pectinate: having units, such as spikelets, closely inserted next to one another, as in the teeth of a comb

pedicel: the stalk that supports a spikelet; see also peduncle

pedicellate: borne on a stalk (pedicel)

peduncle: the stalk that supports an inflorescence of spikelets; see also pedicel

pendent: hanging down

perennial: living for several to many years, often blooming and dying back at the end of each growing season; see also annual

perfect: a flower, floret, or spikelet that bears both male and female reproductive structures; the term bisexual is also used for this condition

pericarp: the fruit wall

perigynium: the membranous sac or sheath that surrounds the gynoecium or achene in some sedges

persistent: not breaking apart, as in an inflorescence axis or rachilla that remains intact at maturity

petiole: the stalk that supports a leaf blade

plano-convex: a structure that is flat on one side and rounded on the other, as in the fertile floret of certain panicoid grasses

Pohlstoffe: a distilled water, methanol, detergent (Aerosol OT) mixture used to soften dried plant specimens to facilitate their being examined; named after the late R. W. Pohl, the eminent and eccentric American agrostologist

pilose: covered with soft distinct hairs

pistillate: a flower, floret, spikelet, or plant that bears only female reproductive structures

pitted: having small cavities or depressions; also referred to as punctate plicate: folded into pleats, typically lengthwise

plumose: feather-like, as in the awn of certain needle-grasses with prominent hairs

polygamous: a plant that bears both bisexual and unisexual flowers or spikelets

polyploid: an organism whose nuclei contain three or more sets of chromosomes

p. p.: the abbreviation of the Latin phrase *pro parte*, meaning "in part;" often used to mean some, but not all species in a genus, as in *Panicum* p. p.

prairie: the extensive level or somewhat undulating grasslands of central North America, characterized by rich soils and tall, sod-forming grasses

prophyllum: the first leaf of a lateral branch; also spelled prophyll

proliferated: the term applied to a spikelet or an inflorescence when some portion has been modified into bulblets or other vegetative structures

prop roots: the aerial roots at the base of a maize plant that provide mechanical support for the stem

prostrate: lying flat on the ground

pseudogamy: a kind of apomixis in which the embryo develops without the egg cell being fertilized, but which requires sperm nuclei to fertilize the polar nuclei for the embryo and the endosperm to develop

pseudospikelet: literally a "false spikelet," it is the structure found in some bamboos in which a single true spikelet is subtended by several bracts

puberulent: minutely pubescent; downy, the hairs
soft, straight, and erect

pubescent: said of any plant structure that is hairy, especially if the hairs are short and soft

pulvinus: the swollen or enlarged base of a leaf sheath or of an inflorescence branch in some grasses; pulvini are associated with the movement of these structures

- Q -

q. v.: the abbreviation of a Latin phrase meaning "which see," which is the author's way of telling you that the word or topic is explained elsewhere in the text

- R -

raceme: an elongate arrangement of stalked spikelets attached along an unbranched central axis

rachilla: the unbranched central axis of a spikelet; not the central axis of an inflorescence of spikelets; rhachilla is the English spelling

rachilla extension: the portion of a rachilla that extends beyond the insertion of the uppermost floret; often appearing as a bristle

rachis: the unbranched central axis of a spike, raceme, or rame; the primary axis of a panicle

rame: an elongate arrangement of stalked and unstalked spikelets borne in repeating pairs or trios along an unbranched axis; the inflorescence type characteristic of the bluestem grasses and their relatives

rank: a vertical row, as seen when looking down on a plant; often expressed in terms of 2-ranked, 3-ranked, etc., which would indicate the number of rows

reflexed: turned or bent abruptly downward or backward

repent: creeping or sprawling plants or stems, often rooting at the nodes; also referred to as trailing; when the accent is placed on the other syllable, you feel sorry or contrite about a past action

retrorse: directed backward or downward, as in barbs on a bristle

rhizomatous: rhizome-bearing

rhizome: an underground, horizontal stem that bears reduced, scaly leaves

rosette: a dense, circular cluster of basal leaves

rudiment: a small, very poorly developed floret

rugose: wrinkled, as in the surface of a bract

runner: a stolon (q. v.), especially a slender one

- S -

savanna(h): a type of subtropical or tropical grassland characterized by coarse grasses and scattered trees

scabrous: covered with short, stiff hairs, so as to be rough to the touch

scarious: thin, dry, membranous, and non-green, as in margins of certain leaves and bracts

scutellum: the organ of the grass embryo that is located between the endosperm and the embryo axis; it is often interpreted as the grass cotyledon

secund: with florets or spikelets turned toward one side only, usually as a result of torsion along an axis

segmental allopolyploid: a polyploid of hybrid origin in which the chromosome sets of the parents are not identical, nor are they that different from one another, such that the plants sometimes behave as though they are autopolyploids and sometimes as allopolyploids

sessile: not stalked; seated on or attached directly to another plant part

seta: a bristle; setaceous means bristle-like

sheath: the lower portion of a grass leaf that surrounds the stem

silica bodies: crystals of silicon dioxide that occur in specialized epidermal cells of the grass leaf; their shape is of diagnostic significance

silica cells: the shorter epidermal cells of the grass leaf and stem that contain silica deposits

s. l.: the abbreviation of the Latin phrase *sensu lato*, meaning "in the broad sense"

somatic cells: the vegetative cells of the plant body, as opposed to the gametes or sex cells

sp.: species, in the singular

spathulate: having a large bract that is attached beneath and that often ± surrounds an inflorescence, as in the bracts of certain bluestems

species: a kind of plant or animal whose distinctiveness is seen in morphological, anatomical, cytological, chemical, and genetic discontinuities presumably brought about by reproductive isolation; thought by many zoologists to be real biological entities and by many botanists to be convenient constructs of the human mind

species name: a binomial consisting of the genus and the specific epithet, as in *Zea mays*

specific epithet: the second element of a binomial (the *mays* of *Zea mays*)

spike: an elongate arrangement of sessile spikelets borne on an unbranched central axis

spikelet: the basic unit of the grass inflorescence, typically consisting of two glumes and one or more florets

spp.: species, in the plural

spreading: oriented outward and more or less diverging from the point of origin

s. s.: the abbreviation of the Latin phrase *sensu stricto*, meaning "in the narrow or restricted sense"

ssp.: abbreviation for subspecies; sspp., plural

stamen: the pollen-producing organ of a flower, consisting of an anther and a filament

staminate: said of a flower, floret, spikelet, or plant that bears only stamens

stem: the plant axis that bears leaves, flowers, and fruits; principally aerial, but sometimes subterranean in the form of rhizomes, bulbs, etc.

steppe: any of the extensive, often tree-less, semiarid grasslands of Eurasia, Africa, and the Americas dominated by short bunch grasses

sterile: lacking reproductive parts

sterile lemma: a lemma that does not enclose a flower; often all that remains of a reduced floret

stigma: the region of the female reproductive structure (carpel) that is receptive to pollen; in grasses, the feathery portion that sits atop the ovary

stipe: a stalk

stipitate: stalked, as in the florets of certain grasses

stolon: an aerial, horizontal stem, often rooting at the nodes, that bears ordinary foliage leaves, as in Bermuda grass; often called runners

stoloniferous: stolon-bearing

striate: marked with fine, longitudinal, parallel lines,

grooves, or ridges

strict: close together, straight, and upright, as in

certain stems

strigose: characterized by short, stiff, appressed

hairs

sub: Latin prefix, meaning below, inferior to, almost,

or somewhat

subglobose: almost spherical, as in the shape of a

floret or a grain

suborbicular: almost circular in outline

subtend: to be below another plant part in point of attachment, as in a set of bracts attached beneath a

spikelet

sucker: a vegetative shoot that originates from below

ground

- T -

tabashir: silica deposits within the culms of certain bamboos; the term is little known outside the world of

crossword puzzles

tawny: dull brownish-yellow

taxon: a taxonomic group of any rank; plural, taxa

terete: round, as seen in cross-section; spherical is

not a synonym

terminal: uppermost, as in a floret in a spikelet or an

inflorescence on a stem

tetraploid: a cell or an organism in which the nuclei

contain four sets of chromosomes

throat: the adaxial surface at the junction of the

sheath and blade of the grass leaf

tiller: a ± erect basal branch or sucker shoot

tomentose: covered by dense, soft, woolly hairs

transverse: in a cross-wise direction, as across the

face or surface of a plant part

trichome: any hair-like outgrowth of the epidermis

truncate: with a squared-off or chopped-off apex or base, as in the appearance of certain leaves or bracts

tubercule: a warty protuberance

tufted: in bunches or clumps, as in the stems of a

grass plant

turgid: swollen

- U -

uncinate: hooked, as in certain hairs

unilateral: one-sided; situated on one side only

unisexual: a flower, spikelet, or plant that bears

either stamens or carpels, but not both

utricle: an indehiscent, 1-seeded, bladdery fruit;

found in certain bamboos

- V -

var.: variety

vein: a strand of vascular tissue in a leaf or bract, seen as a ridge on the surface; in the grasses, veins

are also called nerves

veld: a type of tree-less grassland best developed in

southern Africa

venation: the arrangement of veins on a leaf or bract

ventral: pertaining to or attached to the inner side of an organ; the side that faces toward a central axis

verrucose: a surface, as on a lemma, covered by

warty protuberances

versatile: said of an anther that is attached at its midpoint, such that it moves \pm freely on the apex of

the filament

verticil: a whorl or circular arrangement of parts

around a central axis; verticillate, whorled

vestigial: rudimentary, poorly developed, much-

reduced in size

vestiture: any surface feature on a plant, such as

hairs, spines, scales, wax, etc.

villous: covered with shaggy, soft, but unmatted

hairs

vivipary: in grasses, the term is restricted to a form of asexual reproduction in which bracts of the spikelet

are modified into leafy or tailed bulb-like structures, as in certain blue grasses, fescues, and wheat grasses

viscid: sticky, gummy

- W -

wanting: lacking, missing

weed: an undesirable, worthless, aggressive kind of plant that has a set of biological features that often allows it to out-compete native species and crop

plants in a particular area

winged: having a wing, a membranous lateral

extension of an organ, as in a winged glume or inflorescence branch

- X -

x: the designation of the number of chromosomes that constitutes the basic set for a particular

organism, as in x = 5

- Z -

zygote: the fertilized egg

SOURCES

Chapman, G. P. 1996. The biology of grasses. CAB International. Wallingford, U. K. Pp. 237-249.

Jackson, B. D. 1928. A glossary of botanic terms with their derivation and accent. Fourth edition. Duckworth & Co., Ltd. London, England. 481 pp. [A wonderful reference!]

Harrington, H. D. 1977. How to identify grasses and grasslike plants. The Swallow Press. Chicago, IL. Pp.85-142.

Smith, J. P., Jr. 1977. Vascular plant families. Mad River Press. Eureka, CA. Pp. 288-312.

Stearn, W. T. 1992. Botanical Latin. Fourth edition. David & Charles. Devon, England. 546 pp. [The standard reference]

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