

RESEARCH ARTICLE

Alleles of *HLA-DRB1*04* Associated with Pulmonary Tuberculosis in Amazon Brazilian Population

Dhêmerson Souza de Lima^{1☯*}, Mauricio Morishi Ogusku^{2☯}, Maisa Porto dos Santos^{2‡}, Cláudia Maria de Melo Silva^{3‡}, Vanessa Alves de Almeida^{1‡}, Irineide Assumpção Antunes^{4‡}, Antonio Luiz Boechat^{1☯}, Rajendranath Ramasawmy^{1,5,6☯}, Aya Sadahiro^{1☯}

1 Programa de Pós-Graduação em Imunologia Básica e Aplicada, Instituto de Ciências Biológicas, Universidade Federal do Amazonas (UFAM), Manaus, Amazonas, Brasil, **2** Laboratório de Micobacteriologia, Instituto Nacional de Pesquisas da Amazônia (INPA), Manaus, Amazonas, Brasil, **3** Programa de Pós-Graduação em Ciências Farmacêuticas, Faculdade de Ciências Farmacêuticas (FCF), Manaus, Amazonas, Brasil, **4** Policlínica Cardoso Fontes, Manaus, Amazonas, Brasil, **5** Fundação de Medicina Tropical Doutor Heitor Vieira Dourado (FMT/HVD), Manaus, Amazonas, Brasil, **6** Universidade Nilton Lins, Manaus, Amazonas, Brasil

☯ These authors contributed equally to this work.

‡ These authors also contributed equally to this work.

* dhemersonsouzalima@gmail.com



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Abstract

Immunogenetic host factors are associated with susceptibility or protection to tuberculosis (TB). Strong associations of HLA class II genes with TB are reported. We analyzed the *HLA-DRB1*04* alleles to identify subtypes associated with pulmonary TB and their interaction with risk factors such as alcohol, smoking, and gender in 316 pulmonary TB patients and 306 healthy individuals from the Brazilian Amazon. The *HLA-DRB1*04* was prevalent in patients with pulmonary TB ($p < 0.0001$; OR = 2.94; 95% CI = 2.12 to 4.08). Direct nucleotide sequencing of DRB1 exon 2 identified nine subtypes of *HLA-DRB1*04*. The subtype *HLA-DRB1*04:11:01* ($p = 0.0019$; OR = 2.23; 95% CI = 1.34 to 3.70) was associated with susceptibility to pulmonary TB while *DRB1*04:07:01* ($p < 0.0001$; OR = 0.02; 95% CI = 0.001 to 0.33) to protection. Notably, the interaction between alcohol and *HLA-DRB1*04:11:01* increased the risk for developing pulmonary TB ($p = 0.0001$; OR = 51.3; 95% CI = 6.81 to 386). Multibacillary pulmonary TB, the clinical presentation of disease transmission, was strongly associated with interaction to alcohol ($p = 0.0026$; OR = 11.1; 95% CI = 3.99 to 30.9), *HLA-DRB1*04:11:01* ($p = 0.0442$; OR = 2.01; 95% CI = 1.03 to 3.93) and *DRB1*04:92* ($p = 0.0112$; OR = 8.62; 95% CI = 1.63 to 45.5). These results show that *HLA-DRB1*04* are associated with pulmonary TB. Interestingly, three subtypes, *DRB1*04:07:01*, *DRB1*04:11:01* and *DRB1*04:92* of the *HLA-DRB1*04* could be potential immunogenetic markers that may help to explain mechanisms involved in disease development.

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Introduction

Tuberculosis (TB), a serious public health problem worldwide, is caused by *Mycobacterium tuberculosis* complex (MTBC) with *M. tuberculosis* being the most common [1, 2]. According to the World Health Organization (WHO), approximately one-third of the world's population is infected with TB Bacillus and two billion people are estimated to have latent TB infection with risk for development of the disease. Of notified TB cases, more than 90% occur in low and middle-income countries [3]. Brazil reported 67.966 TB cases in 2014, an incidence rate of 33.5 cases of TB/100.000 habitants. In 2014, the Amazonas state had an incidence rate of 68.4 cases of TB/100.000 habitants that is well above the national average and ranks first in relation to other states [4].

Several risk factors such as HIV-infected individuals [5, 6], diabetes mellitus [7, 8], smoking [9], alcohol [10, 11], under-nutrition [12, 13] and host immunogenetic factors [14–18] are associated with susceptibility to TB. Host genetic factors are strongly associated with the development of TB as hardly 5% to 10% of MTB-infected individuals develop the disease [19, 20]. Several studies have associated alleles of the Human Leukocyte Antigen (HLA) class II to TB [21–29]. Particularly, the following alleles *HLA-DRB1*0803*, *DRB1*0601*, *DRB1*04*, *DRB1*09*, *DRB1*10*, *DRB1*15* and *DRB1*16* are shown to be associated with susceptibility to pulmonary TB [28, 30–33]. This diversity of alleles is certainly related to the high polymorphism of the HLA system [34–36]. A limitation of these studies is the use of techniques of low-resolution typing that not identified the allele subtypes.

In preliminary studies, the generic *HLA-DRB1*04* was frequent in pulmonary TB patients (unpublished data), but this gene has many subtypes and is very important to determine which alleles are associated with the disease.

For this reason, our study aimed to identifying subtypes of *HLA-DRB1*04* in patients with pulmonary TB to correlate with risk factors for the development of disease and to explain the HLA role on the high incidence rate of TB cases in the Amazonas state.

Materials and Methods

Population Samples

A total of 622 individuals, aged 18 to 60 years, born in the Brazilian Amazon are non-indigenous and their parents and grandparents were also born and lived in the same region. All of the patients with Pulmonary TB (n = 316) participating in this study are unrelated, treatment-naïve, positive for sputum smear or culture tests and were selected at the Reference Center for Sanitary Pneumology, Policlínica Cardoso Fontes, Amazonas, Manaus, Brazil. Patients with treatment abandonment or recurrence, autoimmune diseases, cancer, diabetes, HIV, or using immunoregulatory drugs were excluded. Pregnant women were also excluded. The control group (n = 306) consisted of direct contacts of patients recently diagnosed with pulmonary TB, and were without signs and symptoms of the disease and negative for bacteriological tests.

Mycobacteriology

Bacteriological tests were performed at the Micobacteriology Laboratory, of Instituto Nacional de Pesquisas da Amazônia (INPA). The sputum samples were processed for the realization of direct or concentrated sputum smear and culture by PKO method [37, 38]. Patients were classified as multibacillary (individuals who had positive direct sputum smear or concentrated) and paucibacillary (individuals with negative sputum smear, but with positive results for the culture method).

DNA extraction and PCR of the *HLA-DRB1*04* allele

Genomic DNA was extracted from peripheral blood leucocytes by using the rapid technique tetramethylammonium bromide salts (DTAB/CTAB) [39] and stored at -20°C for use in PCR. The following pair of primers: Forward 5' GT TTC TTG GAG CAG GTT AAA C 3' and Reverse 5' CCT AAA CCT TCA CCC CAA CCA C 3' was used for the amplification of the specific allele for *HLA-DRB1*04*. The PCR optimization was performed. Each reaction contains a mix of 1X Buffer (20 mM Tris-HCl pH 8.4, 50 mM KCl), 0.2 mM of each dNTPs, 2.0 mM MgCl₂, 0.2 uM of forward and reverse primers, 1.25 U of *Taq* DNA polymerase (Invitrogen) and 4 uL of genomic DNA (50 ng/uL), in a total volume of 25 uL. The PCR program was an initial denaturation at 96°C for 5 minutes, followed by 35 cycles at 96°C for 1 minute, 64.5°C for 1 minute, 72°C for 1 minute, and a final extension at 72°C for 10 minutes in a Veriti Thermal Cycler. PCR products were detected in a 1.5% agarose gel electrophoresis stained with SYBR Green and visualized in blue-light transilluminator. The electrophoresis was run in TBE buffer for 60 minutes at 130 V and 110 mA.

Purification of the PCR product and nucleotide sequencing of the *HLA-DRB1*04*

PCR products were purified using polyethylene glycol (PEG) 20%. The sequencing reaction was performed by the dideoxynucleotides method [40] using the BigDye Terminator Kit version 3.1 to sequence the fragments in both directions (forward and reverse) in the automated sequencer ABI 3130xl Genetic Analyzer (Applied Biosystems). The consensus sequences were built with Geneious software version R8 and compared with the database sequences IMGT/HLA (BLASTn) (<http://www.ebi.ac.uk/ipd/imgt/hla/>).

In silico prediction of HLA class II molecules binding ESAT-6

The predictions binding between early secreted antigenic target of 6 kDa (ESAT-6) of *Mycobacterium tuberculosis* H37Rv (accession number Rv3875) with HLA class II molecules were carried out by bioinformatics tools using analysis of algorithms ("*in silico* mapping") by the method of Artificial Neural Network (NetMHCIIpan)[41] in the Immune Epitope Database and Analysis Resource (IEDB) (http://www.iedb.org/home_v3.php). Lower percentile rank indicates higher affinity (Cut-off percentile rank ≤ 10.0).

Ethics

This study was approved by the Human Research Ethics Committee of the Universidade Federal do Amazonas (reference number: 0017.0.115.000-08) according to the Brazilian Federal laws. All study participants provided written informed consent.

Statistical analysis

Descriptive statistic was used to characterize the profile of pulmonary TB patients and controls involved in this study. For the comparison of HLA frequencies in patients and controls, 2x2 contingency tables analysis was performed using Chi-square or Fisher's Exact test. The level of significance was $p < 0.05$ and 95% confidence interval. Stepwise logistic regression was performed with $p < 0.05$ and confidence interval of 95% to assess the general characteristics of patient and control groups. Interaction analysis of *HLA-DRB1*04* gene with the general characteristics and bacterial load (multibacillary and paucibacillary) were carried out by stepwise logistic regression. The precision and accuracy (goodness of fit) of each stepwise logistic regression model were analyzed with the area under the ROC curve ($AUC \geq 0.7$) and the Hosmer-

Lemeshow test ($p > 0.05$). A formal Bonferroni correction for the number of *HLA-DRB1*04* analysed would require significance threshold of $p < 0.006$ (p_0/n , $p_0 = 0.05$, $n = 9$ *HLA-DRB1*04* subtypes). Statistics analysis were performed in GraphPad Prism version 6 and MedCalc version 15 softwares.

Results

Baseline characteristics of the study population

Of the 316 patients with pulmonary TB, 75% were classified as multibacillary and 25% as paucibacillary. Baseline characteristics of patients and controls are shown in Table 1. There were no significant differences in mean age and BCG-vaccination. There were, however, significant differences between genders. Pulmonary TB was more common in males. Alcohol and smoking were associated with the disease.

PCR and nucleotide sequencing of the *HLA-DRB1*04* allele

Of the 622 individuals participating in the study, 288 were positive for *HLA-DRB1*04*, 187 pulmonary TB patients (59.2%; 187/316) and 101 controls (33.0%; 101/306). *HLA-DRB1*04* showed strong association with susceptibility to pulmonary TB ($p < 0.0001$; OR = 2.94; 95% CI = 2.12 to 4.08). Nucleotide sequencing of the 288 *HLA-DRB1*04* allele showed nine subtypes of *HLA-DRB1*04* as listed in Table 2 when aligned with the sequences deposited in the database IMGT/HLA (BLASTn) (<http://www.ebi.ac.uk/ipd/imgt/hla/>), that also provides common alleles from different ethnicities.

Table 1. General characteristics of patients with pulmonary TB and controls.

Characteristics	Patients n = 316	Controls n = 306
Gender^a		
Male, n (%)	194 (61.4%)	142 (46.4%)
Female, n (%)	122 (38.6%)	164 (53.6%)
Mean age ± SD	36±12.6	35±10.7
BCG-vaccinated		
Yes, n (%)	212 (67.0%)	180 (58.8%)
No, n (%)	25 (8.00%)	36 (11.8%)
Data not available, n (%)	79 (25.0%)	90 (29.4%)
Alcoholic drink^b		
Yes, n (%)	87 (27.5%)	20 (6.50%)
No, n (%)	135 (42.7%)	226 (73.9%)
Data not available, n (%)	94 (29.8%)	60 (19.6%)
Smoking^c		
Yes, n (%)	69 (21.8%)	9 (3.00%)
No, n (%)	153 (48.4%)	233 (76.0%)
Data not available, n (%)	94 (29.8)	64 (21.0%)

^a Gender: ($p = 0.0002$; OR = 1.84; 95% IC = 1.33 to 2.53), calculated with Fisher's exact test

^b Alcoholic drink: Individuals who consumed ≥ 4 doses for women and ≥ 5 doses for men at the same time within the last 30 days [42]. 1 dose = 30 ml distilled (or 12 g of pure alcohol). $p < 0.0001$; OR = 7.28; 95% CI = 4.28 to 12.4, calculated with Fisher's exact test.

^c Smoking: were defined as smokers, who made use of cigarettes daily for one year or more [43]. $p < 0.0001$; OR = 11.7; 95% CI = 5.66 to 24.1, calculated with Fisher's exact test.

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Table 2. Results of the obtained nucleotide sequences and aligned with the sequences of the IMGT/HLA.

Alleles <i>HLA-DRB1*04</i>	Score	Length Obtained (IMGT)	Identities %	Positives %	Ethnic Origin IMGT/HLA
<i>DRB1*04:01:01</i>	1085	561 (801 bp)	100	100	Caucasoid, Oriental
<i>DRB1*04:02:01</i>	1085	561 (801 bp)	100	100	Caucasoid
<i>DRB1*04:03:01</i>	1085	561 (801 bp)	100	100	Caucasoid
<i>DRB1*04:04:01</i>	1085	561 (801 bp)	100	100	Caucasoid
<i>DRB1*04:05:01</i>	1085	561 (801 bp)	100	100	Caucasoid, Oriental
<i>DRB1*04:06:01</i>	1085	561 (801 bp)	100	100	Caucasoid, Oriental
<i>DRB1*04:07:01</i>	1076	561 (801 bp)	99	99	Caucasoid, Hispanic ^a
<i>DRB1*04:11:01</i>	1085	561 (801 bp)	100	100	Australian Aboriginal, H ^a , Mixed ^b
<i>DRB1*04:92</i>	1076	561 (663 bp)	99	99	Unknown ^c

^a Hispanic: People historically of mixed Mediterranean Caucasoid, American Indian race

^b Mixed: The mixture of various races

^c Unknown: Ethnic origin unknown.

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The distribution of the *HLA-DRB1*04* allele subtypes in both pulmonary TB patients and controls is shown in Table 3. The most common subtypes in the study population were *HLA-DRB1*04:04:01* and *HLA-DRB1*04:11:01*. *HLA-DRB1*04:11:01* was strongly associated with susceptibility to pulmonary TB ($p = 0.0019$; OR = 2.23, 95% CI = 1.34 to 3.70). *HLA-DRB1*04:07:01* was associated with protection against the disease and was not found among the TB patients.

The stepwise logistic regression models for *HLA-DRB1*04* and pulmonary TB

Stepwise logistic regression analysis identified three risk factors. Alcohol ($p = 0.0016$; OR = 4.32; 95% CI = 1.74 to 10.7), male ($p = 0.0001$; OR = 2.69; 95% CI = 1.64 to 4.40) and smoking ($p = 0.0442$; OR = 3.16; 95% CI = 1.03 to 9.69) were associated with pulmonary TB.

To assess the influence of *HLA-DRB1*04* gene on the risk factors, another stepwise logistic regression model was performed. *HLA-DRB1*04* ($p = 0.0143$; OR = 1.92; 95% CI = 1.14 to 3.23) showed strong interaction with alcohol ($p < 0.0001$; OR = 8.43; 95% CI = 3.85 to 18.5) and increased the significant risk for development of pulmonary TB when compared to non-

Table 3. Distribution and association analysis of *HLA-DRB1*04* subtypes in pulmonary TB patients and the control group.

Alleles <i>HLA-DRB1*04</i>	Patients n = 187 (%)	Controls n = 101 (%)	<i>p</i> value ^a	OR	95% CI
<i>DRB1*04:01:01</i>	6 (3.2)	6 (5.9)	0.3544	0.525	0.165 to 1.67
<i>DRB1*04:02:01</i>	9 (4.8)	9 (8.9)	0.2039	0.517	0.198 to 1.35
<i>DRB1*04:03:01</i>	9 (4.8)	9 (8.9)	0.2039	0.517	0.198 to 1.35
<i>DRB1*04:04:01</i>	42 (22.5)	28 (27.7)	0.3185	0.755	0.434 to 1.32
<i>DRB1*04:05:01</i>	2 (1.1)	3 (3.0)	0.3479	0.353	0.058 to 2.15
<i>DRB1*04:06:01</i>	3 (1.6)	2 (2.0)	1.0000	0.807	0.133 to 4.91
<i>DRB1*04:07:01</i>	0 (0.0)	12 (11.9)	0.0001 ^b	0.019	0.001 to 0.33
<i>DRB1*04:11:01</i>	95 (50.8)	32 (31.7)	0.0019 ^b	2.23	1.34 to 3.70
<i>DRB1*04:92</i>	21 (11.2)	0 (0.0)	0.0002 ^b	26.21	1.57 to 438

^a Fisher's exact test, $p < 0.05$; OR = Odds ratio; CI = Confidence interval

^b Significant *p* values according to the Bonferroni correction ($p < 0.006$)

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Table 4. Prediction of the binding for epitopes of ESAT-6 with HLA molecules encoded by *HLA-DRB1*04:11:01* and *DRB1*04:07:01*.

Alleles <i>HLA-DRB1*04</i>	Antigens ^{Start-End}	Sequences	Core regions	Percentile rank
<i>DRB1*04:07:01</i>	ESAT-6 ₅₋₁₉	QWNFAGIEAAASAIQ	FAGIEAAAS	1.58
<i>DRB1*04:07:01</i>	ESAT-6 ₄₋₁₈	QQWNFAGIEAAASAI	FAGIEAAAS	1.96
<i>DRB1*04:07:01</i>	ESAT-6 ₆₋₂₀	WNFAGIEAAASAIQG	FAGIEAAAS	3.78
<i>DRB1*04:07:01</i>	ESAT-6 ₃₋₁₇	EQQWNFAGIEAAASA	FAGIEAAAS	4.55
<i>DRB1*04:07:01</i>	ESAT-6 ₇₋₂₁	NFAGIEAAASAIQGN	FAGIEAAAS	8.31
<i>DRB1*04:07:01</i>	ESAT-6 ₂₋₁₆	TEQQWNFAGIEAAAS	FAGIEAAAS	9.92
<i>DRB1*04:11:01</i>	ESAT-6 ₁₉₋₃₃	QGNVTSIHSLLEDEGK	VTSIHSLLD	13.28
<i>DRB1*04:11:01</i>	ESAT-6 ₁₈₋₃₂	IQGNVTSIHSLLEDEG	VTSIHSLLD	13.54
<i>DRB1*04:11:01</i>	ESAT-6 ₁₇₋₃₁	AIQGNVTSIHSLLEDE	VTSIHSLLD	16.08
<i>DRB1*04:11:01</i>	ESAT-6 ₆₂₋₇₆	ATELNALQNLARTI	LNNALQNLA	16.22
<i>DRB1*04:11:01</i>	ESAT-6 ₅₋₁₉	QWNFAGIEAAASAIQ	FAGIEAAAS	16.68
<i>DRB1*04:11:01</i>	ESAT-6 ₁₆₋₃₀	SAIQGNVTSIHSLLD	VTSIHSLLD	16.71

Percentile rank ≤ 10.0—Lower percentile rank indicates the 15-mer peptides with higher affinity to HLA

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carriers of the allele. Nevertheless, *HLA-DRB1*04* interaction to male ($p = 0.0001$; OR = 2.71; 95% CI = 1.66 to 4.43) or smoking ($p = 0.0564$; OR = 2.99; 95% CI = 0.97 to 9.26) did not affect significantly the risk of susceptibility to pulmonary TB.

Stepwise logistic regression model including the subtypes, *HLA-DRB1*04:11:01* and *HLA-DRB1*04:92* was carried out. Only the interaction of alcohol ($p = 0.0001$; OR = 51.3; 95% CI = 6.81 to 386) with *HLA-DRB1*04:11:01* ($p = 0.0265$; OR = 2.13; 95% CI = 1.09 to 4.14) showed a very high risk of susceptibility to pulmonary TB.

Stepwise logistic regression analysis on the effect of interaction of alcohol and HLA alleles on bacterial load (multibacillary and paucibacillary) showed that carriers of *HLA-DRB1*04:11:01* ($p = 0.0442$; OR = 2.01; 95% CI = 1.03 to 3.93) or *HLA-DRB1*04:92* ($p = 0.0112$; OR = 8.62; 95% CI = 1.63 to 45.5) who consume alcohol ($p = 0.0026$; OR = 11.1; 95% CI = 3.99 to 30.9) have higher risk of developing multibacillary pulmonary TB.

In silico prediction of HLA class II molecules binding ESAT-6

NetMHCIIpan was used to estimate the binding affinity of ESAT-6 epitopes to *HLA-DRB1*04:11:01* and *DRB1*04:07:01*. Low binding affinity resulted for *HLA-DRB1*04:11:01* while strong binding for *DRB1*04:07:01*. The results are shown in percentile rank values (Table 4). Importantly, the core regions are fragments of antigens that interact with HLA molecules, determining binding affinity and specificity. Prediction analysis for *HLA-DRB1*04:92* was not performed because this HLA is unavailable in IEDB database.

Frequency of *HLA-DRB1*04* alleles in Worldwide Populations

Distribution of *HLA-DRB1*04* were obtained in the database "Allele Frequency Database Net" (AFND) (<http://www.allelefrequencies.net/>). The *HLA-DRB1*04* gene has the highest frequency in populations of the Americas (South = 42.0% and Central America = 33.9%). The subtype *HLA-DRB1*04:11:01* also is most common in the populations of the Americas (44.0%), although ethnic origin of this allele includes Australian aborigines, Hispanic and Mestizos (Table 2). The subtypes *HLA-DRB1*04:92* and *HLA-DRB1*04:07:01*, respectively, are more prevalent in the United States (0.0027%) and Mexico (34.5%) [44][44]. According to data provided, the subtypes associated with pulmonary TB (Table 3), are very frequent in the Americas.

Discussion

In this study, the generic *HLA-DRB1*04* allele was most common in pulmonary TB patients, showing strong association with susceptibility to disease. These data emphasize a major role of HLA system in the regulation of the immune response against pulmonary TB. A study conducted in Recife, Northeast Brazil, also identified *HLA-DRB1*04* associated with pulmonary TB [45]. A recent meta-analysis of 31 studies with 3,416 TB patients and 4,515 controls also found in the Asian population *HLA-DRB1*04* associated with susceptibility to TB [33]. Nevertheless, a study in Mexico found reduced risk for TB in carriers of *HLA-DRB1*04* (OR = 0.24; 95% CI = 0.07 to 0.84) [46].

Most of the studies used only low-resolution HLA typing to search for association with TB. We performed nucleotide sequencing to look for which subtypes of the *HLA-DRB1*04* allele are associated with susceptibility to TB. Interestingly, the subtype *HLA-DRB1*04:11:01* was strongly associated with TB in the Amazonas. Previous investigations reported other alleles subtypes within *DRB1* locus that were associated with risk to the disease among different populations such as the *DRB1*15:01* in south India [47], *DRB1*08:03* in Koreans [30], the same subtype and *DRB1*08:01* in Kazaksthan with drug resistance [48]. The HLA alleles vary among populations because of genetic background. Of note, the HLA region is the most polymorphic in human genome [49].

In contrast to other studies, we also performed interaction of the susceptibility allele *HLA-DRB1*04:11:01* with alcohol consumption, smoking and gender. Importantly, there is the consumption of alcohol worldwide, with an average of 6.2 liters of pure alcohol per year. In Brazil the average alcohol consumption is 8.7 liters. The highest consumption levels are found in Europe and the Americas. Alcohol can lead to dependence and encourage the development of diseases [50]. Alcohol consumption contributes to development of pulmonary TB, favoring increased bacterial burden in lungs, besides causing a decrease in the number of CD4+ T cells and, consequently, lower release of IFN- γ [51].

Recently, the meta-analysis of 72,684 individuals in 14 countries with high burden of tuberculosis, confirmed association with consumption of alcohol, smoking, and male gender [52]. Our study brings a further contribution to the meta-analysis study as for the first time, the interaction between risk factors of pulmonary TB disease development and *HLA-DRB1*04* gene was analyzed. The combination of *HLA-DRB1*04* gene and alcohol consumption increased twice as much the risk for pulmonary TB development. Surprisingly, the risk of TB considerably increased if there is the combination of *HLA-DRB1*04:11:01* subtype and alcohol consumption indicating the influence of genetic factor on susceptibility to TB disease. Nonetheless, the interaction of *HLA-DRB1*04* gene to smoking, and gender did not influence the risk significantly. This observation is related to the reduced number of male's smokers in the study. However, it is known that the population from the Northern region of Brazil has the lowest rate of smoking in the country, with 13.4% of smokers [42].

The strong association with *HLA-DRB1*04:11:01* subtype and susceptibility to TB, can be explained by the fact that this allele may possibly have low binding affinity as shown *in silico* prediction with epitopes of ESAT-6. ESAT-6 is an important protein released by TB bacillus. On the contrary, *HLA-DRB1*04:07:01*, associated with protection to TB disease, showed high binding affinity to six ESAT-6 epitopes as determined by the best binding [53]. Core regions of the epitopes (ESAT-6) are the fragments that interact with the molecule of HLA to determine the binding affinity and specificity [54]. The peptide residues immediately flanking the core regions contribute to HLA/peptide binding affinity [55].

These results support immunogenetic models, susceptibility and protection, and emphasize the importance the genetic background, in particular the HLA system that is essential for

response of CD4+T cell with signaling by HLA class II/peptide, co-stimulation, and cytokines. Furthermore, good interaction between HLA/peptide have a crucial role for the development of a Th1 effector response against pathogen [56]. Our data suggest that carriers of *HLA-DRB1*04:07:01* may have a good T cell response against TB bacillus while carriers of *HLA-DRB1*04:11:01* may have a poor immune response effector.

The *HLA-DRB1*04* gene are associated with Rheumatoid arthritis (RA) [57–59]. However, the subtypes reported to be associated with RA are *DRB1*0401*, *DRB1*0404*, *DRB1*0405*, *DRB1*0408*, *DRB1*0409* [60]. In our study, none of these subtypes are associated with TB. Interestingly, *HLA-DRB1*04:07:01* has been shown to increase the chance of developing RA [60] and this allele was associated with protection to TB in our study.

We also found *HLA-DRB1*04:11:01* and *DRB1*04:92*, were associated with multibacillary TB. Furthermore, there is a strong interaction of these *HLA* alleles with the alcohol consumption. These results suggest that multibacillary TB patients expressing one of these subtypes of *HLA* might be responsible for keeping the chain of disease transmission. Altogether, besides the risk factors associated with TB (socioeconomic, malnutrition, drug abuse, diabetes mellitus, HIV) [5, 61, 62], the genetic factor can also contribute to the maintenance of high rates of the disease in the Amazonas.

Different frequencies of *HLA* class II alleles are found worldwide, according to the geographic region. In South America *HLA-DRB1*04* allele (generic) is more frequent in populations of Argentina and Brazil with a mean 39.6% and 22.0%, respectively [44]. *HLA-DRB1*04:11:01* subtype is also common in populations of Brazil and Argentina, with a mean frequency of 14.0% and 20.2%, respectively [44, 63]. Both *HLA*, are mostly found in the native population [44], and may indicate a rapid evolution of *HLA-DRB1* gene in ancestral Amerindians during migration processes [64]. *HLA-DRB1*04:92* is hardly found in the world and was associated with Black population of United States with an allele frequency of 0.0027% [44, 65]. *HLA-DRB1*04:07:01* has a mean frequency of 34.5% in the Mexican population [44, 66, 67]. It is highly probable that this is the allele associated with protection in Mexico since the generic *HLA-DRB1*04* was associated with protection against pulmonary TB [46].

In conclusion, the present study showed *HLA-DRB1*04* genes and their subtypes associated with pulmonary TB. Also, we determined that interaction between alcohol consumption and *HLA* gene increased the risk for the development of TB, and was associated with multibacillary TB clinical presentation. Further studies will be needed to confirm this association so as to start unraveling the complex network of responses in TB. The relationship between *HLA* and the immune response should be taken into consideration in future studies, specifically in the management of tuberculosis treatment.

Limitations

The number of individuals with the *HLA-DRB1*04:07:01* or *HLA-DRB1*04:92* is small, therefore, require more research with larger numbers of patients with pulmonary TB and controls. Regarding the alcohol consumption and smoking, we did not assess the amount and frequency of consumption. Nevertheless, it is important to emphasize that the results of the statistical analysis were significant, with strong associations with pulmonary TB.

Supporting Information

S1 Fig. ROC curve for stepwise logistic regression model with alcoholic drink, smoking and gender (male) associated with pulmonary TB (AUC = 0.734).
(TIF)

S2 Fig. ROC curve of stepwise logistic regression model for variables alcoholic drink, gender (male) and their interaction to *HLA-DRB1*04* gene, associated with pulmonary TB (AUC = 0.745).

(TIF)

S3 Fig. ROC curve of stepwise logistic regression model for alcoholic drink and their interaction to *HLA-DRB1*04:11:01*, associated with pulmonary TB (AUC = 0.732).

(TIF)

S4 Fig. ROC curve of stepwise logistic regression model for alcoholic drink, *HLA-DRB1*04:11:01* and *HLA-DRB1*04:92* associated with bacterial load (AUC = 0.743).

(TIF)

S1 Table. Stepwise logistic regression analysis with general characteristics of the population.

(DOCX)

S2 Table. Stepwise logistic regression for variables alcoholic drink, gender (male) and smoking, including the *HLA-DRB1*04* gene.

(DOCX)

S3 Table. Stepwise logistic regression analysis for alcohol and *HLA-DRB1*04:11:01* allele associated with pulmonary TB.

(DOCX)

S4 Table. Stepwise logistic regression analysis for the alcoholic drink, *HLA-DRB1*04:11:01* and *DRB1*04:92* associated with the bacterial load (multibacillary).

(DOCX)

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Author Contributions

Conceived and designed the experiments: DSL AS MMO IAA. Performed the experiments: DSL MMO MPS CMMS VAA. Analyzed the data: DSL ALB AS RR. Contributed reagents/materials/analysis tools: AS MMO. Wrote the paper: DSL AS RR MMO ALB. Designed primers: DSL.

References

1. BRASIL. Manual Nacional de Vigilância Laboratorial da Tuberculose e outras Micobactérias. Brasília: Ministério da Saúde. 2008; 1:434.
2. Forrellad MA, Klepp LI, Giofré A, Sabio y García J, Morbidoni HR, Santangelo MdIP, et al. Virulence factors of the Mycobacterium tuberculosis complex. Virulence. 2013; 4(1):3–66. doi: [10.4161/viru.22329](https://doi.org/10.4161/viru.22329) PMID: [PMC3544749](https://pubmed.ncbi.nlm.nih.gov/22329/).
3. WHO. Global Tuberculosis report 2012. 2012.

4. BRASIL. Boletim Epidemiológico. Detectar, tratar e curar: desafios e estratégias brasileiras frente à tuberculose. Ministério da Saúde. 2015; 46(9).
5. Bekker L- G, Wood R. The Changing Natural History of Tuberculosis and HIV Coinfection in an Urban Area of Hyperendemicity. *Clinical Infectious Diseases*. 2010; 50(Supplement 3):S208–S14. doi: [10.1086/651493](https://doi.org/10.1086/651493)
6. Getahun H, Gunneberg C, Granich R, Nunn P. HIV Infection—Associated Tuberculosis: The Epidemiology and the Response. *Clinical Infectious Diseases*. 2010; 50(Supplement 3):S201–S7. doi: [10.1086/651492](https://doi.org/10.1086/651492)
7. Dobler CC, Flack JR, Marks GB. Risk of tuberculosis among people with diabetes mellitus: an Australian nationwide cohort study. *BMJ Open*. 2012; 2(1). doi: [10.1136/bmjopen-2011-000666](https://doi.org/10.1136/bmjopen-2011-000666)
8. Goldhaber-Fiebert JD, Jeon CY, Cohen T, Murray MB. Diabetes mellitus and tuberculosis in countries with high tuberculosis burdens: individual risks and social determinants. *International journal of epidemiology*. 2011; 40(2):417–28. doi: [10.1093/ije/dyq238](https://doi.org/10.1093/ije/dyq238) PMID: [21252210](https://pubmed.ncbi.nlm.nih.gov/21252210/)
9. Gambhir HS, Kaushik RM, Kaushik R, Sindhvani G. Tobacco smoking-associated risk for tuberculosis: a case-control study. *International Health*. 2010; 2(3):216–22. doi: [10.1016/j.inhe.2010.07.001](https://doi.org/10.1016/j.inhe.2010.07.001) PMID: [24037702](https://pubmed.ncbi.nlm.nih.gov/24037702/)
10. Cois A, Ehrlich R. Problem drinking as a risk factor for tuberculosis: a propensity score matched analysis of a national survey. *BMC Public Health*. 2013; 13(1):871. doi: [10.1186/1471-2458-13-871](https://doi.org/10.1186/1471-2458-13-871)
11. Lonroth K, Williams B, Stadlin S, Jaramillo E, Dye C. Alcohol use as a risk factor for tuberculosis—a systematic review. *BMC Public Health*. 2008; 8(1):289. doi: [10.1186/1471-2458-8-289](https://doi.org/10.1186/1471-2458-8-289)
12. Bhargava A, Chatterjee M, Jain Y, Chatterjee B, Kataria A, Bhargava M, et al. Nutritional Status of Adult Patients with Pulmonary Tuberculosis in Rural Central India and Its Association with Mortality. *PloS one*. 2013; 8(10):e77979. doi: [10.1371/journal.pone.0077979](https://doi.org/10.1371/journal.pone.0077979) PMID: [24205052](https://pubmed.ncbi.nlm.nih.gov/24205052/)
13. Cegielski JP, Arab L, Cornoni-Huntley J. Nutritional Risk Factors for Tuberculosis Among Adults in the United States, 1971–1992. *American Journal of Epidemiology*. 2012. doi: [10.1093/aje/kws007](https://doi.org/10.1093/aje/kws007)
14. Amirzargar AA, Yalda A, Hajabolbaghi M, Khosravi F, Jabbari H, Rezaei N, et al. The association of HLA-DRB, DQA1, DQB1 alleles and haplotype frequency in Iranian patients with pulmonary tuberculosis. *The international journal of tuberculosis and lung disease: the official journal of the International Union against Tuberculosis and Lung Disease*. 2004; 8(8):1017–21. Epub 2004/08/13. PMID: [15305487](https://pubmed.ncbi.nlm.nih.gov/15305487/).
15. Lü J, Pan H, Chen Y, Tang S, Feng Y, Qiu S, et al. Genetic polymorphisms of IFNG and IFNGR1 in association with the risk of pulmonary tuberculosis. *Gene*. 2014; 543(1):140–4. doi: <http://dx.doi.org/10.1016/j.gene.2014.03.042> PMID: [24680779](https://pubmed.ncbi.nlm.nih.gov/24680779/)
16. Ma M-j, Xie L-p, Wu S-c, Tang F, Li H, Zhang Z-s, et al. Toll-like receptors, tumor necrosis factor- α , and interleukin-10 gene polymorphisms in risk of pulmonary tuberculosis and disease severity. *Human immunology*. 2010; 71(10):1005–10. doi: <http://dx.doi.org/10.1016/j.humimm.2010.07.009> PMID: [20650298](https://pubmed.ncbi.nlm.nih.gov/20650298/)
17. Sahiratmadja E, Baak-Pablo R, de Visser AW, Alisjahbana B, Adnan I, van Crevel R, et al. Association of polymorphisms in IL-12/IFN- γ pathway genes with susceptibility to pulmonary tuberculosis in Indonesia. *Tuberculosis*. 2007; 87(4):303–11. <http://dx.doi.org/10.1016/j.tube.2007.02.001>. PMID: [17392024](https://pubmed.ncbi.nlm.nih.gov/17392024/)
18. Zhang Z, Zhu H, Pu X, Meng S, Zhang F, Xun L, et al. Association between tumor necrosis factor alpha-238G/a polymorphism and tuberculosis susceptibility: a meta-analysis study. *BMC infectious diseases*. 2012; 12(1):328. doi: [10.1186/1471-2334-12-328](https://doi.org/10.1186/1471-2334-12-328)
19. Bellamy R. Genetics and pulmonary medicine bullet 3: Genetic susceptibility to tuberculosis in human populations. *Thorax*. 1998; 53(7):588–93. doi: [10.1136/thx.53.7.588](https://doi.org/10.1136/thx.53.7.588) PMID: [9797760](https://pubmed.ncbi.nlm.nih.gov/9797760/)
20. Selvaraj P, Alagarasu K, Harishankar M, Vidyarani M, Nisha Rajeswari D, Narayanan PR. Cytokine gene polymorphisms and cytokine levels in pulmonary tuberculosis. *Cytokine*. 2008; 43(1):26–33. doi: [10.1016/j.cyto.2008.04.011](https://doi.org/10.1016/j.cyto.2008.04.011) PMID: [18522869](https://pubmed.ncbi.nlm.nih.gov/18522869/).
21. Dubaniewicz A, Moszkowska G, Szczerkowska Z. Frequency of DRB1-DQB1 two-locus haplotypes in tuberculosis: preliminary report. *Tuberculosis*. 2005; 85(4):259–67. doi: [10.1016/j.tube.2004.12.003](https://doi.org/10.1016/j.tube.2004.12.003) PMID: [15958261](https://pubmed.ncbi.nlm.nih.gov/15958261/).
22. El Baghdadi J, Grant AV, Sabri A, El Azbaoui S, Zaidi H, Cobat A, et al. Human genetics of tuberculosis. *Pathologie-biologie*. 2013; 61(1):11–6. doi: [10.1016/j.patbio.2013.01.004](https://doi.org/10.1016/j.patbio.2013.01.004) PMID: [23399414](https://pubmed.ncbi.nlm.nih.gov/23399414/); PubMed Central PMCID: [PMC3740217](https://pubmed.ncbi.nlm.nih.gov/PMC3740217/).
23. Goldfeld AE. Genetic susceptibility to pulmonary tuberculosis in Cambodia. *Tuberculosis*. 2004; 84(1–2):76–81. doi: [10.1016/j.tube.2003.08.007](https://doi.org/10.1016/j.tube.2003.08.007) PMID: [14670348](https://pubmed.ncbi.nlm.nih.gov/14670348/)
24. Lombard Z, Dalton DL, Venter PA, Williams RC, Bornman L. Association of HLA-DR, -DQ, and vitamin D receptor alleles and haplotypes with tuberculosis in the Venda of South Africa. *Human immunology*. 2006; 67(8):643–54. Epub 2006/08/19. doi: [10.1016/j.humimm.2006.04.008](https://doi.org/10.1016/j.humimm.2006.04.008) PMID: [16916662](https://pubmed.ncbi.nlm.nih.gov/16916662/).

25. Magira EE, Papasteriades C, Kanterakis S, Toubis M, Roussos C, Monos DS. HLA-A and HLA-DRB1 amino acid polymorphisms are associated with susceptibility and protection to pulmonary tuberculosis in a Greek population. *Human immunology*. 2012; 73(6):641–6. doi: <http://dx.doi.org/10.1016/j.humimm.2012.03.008> PMID: [22504415](https://pubmed.ncbi.nlm.nih.gov/22504415/)
26. Mishra G, Kumar N, Kaur G, Jain S, Tiwari PK, Mehra NK. Distribution of HLA-A, B and DRB1 alleles in Sahariya tribe of North Central India: An association with pulmonary tuberculosis. *Infection, genetics and evolution: journal of molecular epidemiology and evolutionary genetics in infectious diseases*. 2013. doi: [10.1016/j.meegid.2013.08.019](https://doi.org/10.1016/j.meegid.2013.08.019) PMID: [23994122](https://pubmed.ncbi.nlm.nih.gov/23994122/).
27. Qu HQ, Fisher-Hoch SP, McCormick JB. Knowledge gaining by human genetic studies on tuberculosis susceptibility. *Journal of human genetics*. 2011; 56(3):177–82. doi: [10.1038/jhg.2010.164](https://doi.org/10.1038/jhg.2010.164) PMID: [21179108](https://pubmed.ncbi.nlm.nih.gov/21179108/); PubMed Central PMCID: PMC3169186.
28. Shi GL, Hu XL, Yang L, Rong CL, Guo YL, Song CX. Association of HLA-DRB alleles and pulmonary tuberculosis in North Chinese patients. *Genetics and molecular research: GMR*. 2011; 10(3):1331–6. doi: [10.4238/vol10-3gmr1132](https://doi.org/10.4238/vol10-3gmr1132) PMID: [21751159](https://pubmed.ncbi.nlm.nih.gov/21751159/).
29. Wang LX, Nagata T, Tsujimura K, Uchijima M, Seto S, Koide Y. Identification of HLA-DR4-restricted T-cell epitope on MPT51 protein, a major secreted protein derived from *Mycobacterium tuberculosis* using MPT51 overlapping peptides screening and DNA vaccination. *Vaccine*. 2010; 28(8):2026–31. doi: [10.1016/j.vaccine.2009.10.063](https://doi.org/10.1016/j.vaccine.2009.10.063) PMID: [20188259](https://pubmed.ncbi.nlm.nih.gov/20188259/).
30. Kim HS, Park MH, Song EY, Park H, Kwon SY, Han SK, et al. Association of HLA-DR and HLA-DQ genes with susceptibility to pulmonary tuberculosis in Koreans: preliminary evidence of associations with drug resistance, disease severity, and disease recurrence. *Human immunology*. 2005; 66(10):1074–81. Epub 2006/01/03. doi: [10.1016/j.humimm.2005.08.242](https://doi.org/10.1016/j.humimm.2005.08.242) PMID: [16386650](https://pubmed.ncbi.nlm.nih.gov/16386650/).
31. Ruggiero G, Cosentini E, Zanzi D, Sanna V, Terrazzano G, Matarese G, et al. Allelic distribution of human leucocyte antigen in historical and recently diagnosed tuberculosis patients in Southern Italy. *Immunology*. 2004; 111(3):318–22. doi: [10.1111/j.1365-2567.2004.01811.x](https://doi.org/10.1111/j.1365-2567.2004.01811.x) PMID: [PMC1782420](https://pubmed.ncbi.nlm.nih.gov/PMC1782420/).
32. Duarte R, Carvalho C, Pereira C, Bettencourt A, Carvalho A, Villar M, et al. HLA class II alleles as markers of tuberculosis susceptibility and resistance. *Revista portuguesa de pneumologia*. 2011; 17(1):15–9. Epub 2011/01/22. PMID: [21251479](https://pubmed.ncbi.nlm.nih.gov/21251479/).
33. Tong X, Chen L, Liu S, Yan Z, Peng S, Zhang Y, et al. Polymorphisms in HLA-DRB1 Gene and the Risk of Tuberculosis: A Meta-analysis of 31 Studies. *Lung*. 2015; 193(2):309–18. Epub 2015/03/20. doi: [10.1007/s00408-015-9692-z](https://doi.org/10.1007/s00408-015-9692-z) PMID: [25787085](https://pubmed.ncbi.nlm.nih.gov/25787085/).
34. Guha P, Srivastava SK, Bhattacharjee S, Chaudhuri TK. Human migration, diversity and disease association: a convergent role of established and emerging DNA markers. *Frontiers in genetics*. 2013; 4:155. doi: [10.3389/fgene.2013.00155](https://doi.org/10.3389/fgene.2013.00155) PMID: [23950760](https://pubmed.ncbi.nlm.nih.gov/23950760/); PubMed Central PMCID: PMC3738866.
35. Rojas-Villarraga A, Botello-Corzo D, Anaya JM. HLA-Class II in Latin American patients with type 1 diabetes. *Autoimmunity reviews*. 2010; 9(10):666–73. Epub 2010/06/22. doi: [10.1016/j.autrev.2010.05.016](https://doi.org/10.1016/j.autrev.2010.05.016) PMID: [20561992](https://pubmed.ncbi.nlm.nih.gov/20561992/).
36. Robinson J, Halliwell JA, Hayhurst JD, Flicek P, Parham P, Marsh Steven GE. The IPD and IMGT/HLA database: allele variant databases. *Nucleic Acids Research*. 2015; 43(D1):D423–D31. doi: [10.1093/nar/gku1161](https://doi.org/10.1093/nar/gku1161)
37. Salem JI, Marója MF, Carvalho FF, Lima MO, Litaiff LRL, Cardoso MSL, et al. Valor relativo do exame direto, após concentração e por cultivo de escarro no diagnóstico bacteriológico da tuberculose pulmonar no Amazonas. *J Pneumol*. 1990; 16:133–6.
38. Salem JI, Carvalho CM, Ogusku MM, Maia R, Ruffino-Neto A. PKO—Alternative method for isolating mycobacteria from sputum. *Acta Amazonica* 2007;37 419–24.
39. Gustincich S, Manfioletti G, Del Sal G, Schneider C, Carninci P. A fast method for high-quality genomic DNA extraction from whole human blood. *BioTechniques*. 1991; 11(3):298–300, 2. Epub 1991/09/01. PMID: [1931026](https://pubmed.ncbi.nlm.nih.gov/1931026/).
40. Sanger F, Nicklen S, Coulson AR. DNA sequencing with chain-terminating inhibitors. *Proceedings of the National Academy of Sciences of the United States of America*. 1977; 74(12):5463–7. Epub 1977/12/01. PMID: [271968](https://pubmed.ncbi.nlm.nih.gov/271968/); PubMed Central PMCID: PMC431765.
41. Nielsen M, Lundegaard C, Blicher T, Peters B, Sette A, Justesen S, et al. Quantitative predictions of peptide binding to any HLA-DR molecule of known sequence: NetMHCIIpan. *PLoS computational biology*. 2008; 4(7):e1000107. Epub 2008/07/08. doi: [10.1371/journal.pcbi.1000107](https://doi.org/10.1371/journal.pcbi.1000107) PMID: [18604266](https://pubmed.ncbi.nlm.nih.gov/18604266/); PubMed Central PMCID: PMC431765.
42. BRASIL. Pesquisa Nacional de Saúde 2013. Percepção do estado de saúde, estilos de vida e doenças crônicas. Brasil, Grandes Regiões e Unidades da Federação. IBGE. 2014.
43. den Boon S, van Lill SWP, Borgdorff MW, Verver S, Bateman ED, Lombard CJ, et al. Association between smoking and tuberculosis infection: a population survey in a high tuberculosis incidence area. *Thorax*. 2005; 60(7):555–7. doi: [10.1136/thx.2004.030924](https://doi.org/10.1136/thx.2004.030924) PMID: [15994262](https://pubmed.ncbi.nlm.nih.gov/15994262/)

44. González-Galarza Faviel F, Takeshita Louise YC, Santos Eduardo JM, Kempson F, Maia Maria Helena T, Silva Andrea Luciana Soares d, et al. Allele frequency net 2015 update: new features for HLA epitopes, KIR and disease and HLA adverse drug reaction associations. *Nucleic Acids Research*. 2015; 43(D1):D784–D8. doi: [10.1093/nar/gku1166](https://doi.org/10.1093/nar/gku1166)
45. Lucena-Silva N, Baliza MD, Martins AE, Deghaide NH, Teixeira KM, Rodrigues LC, et al. Relatedness and HLA-DRB1 typing may discriminate the magnitude of the genetic susceptibility to tuberculosis using a household contact model. *Journal of epidemiology and community health*. 2010; 64(6):513–7. Epub 2009/08/21. doi: [10.1136/jech.2008.086801](https://doi.org/10.1136/jech.2008.086801) PMID: [19692729](https://pubmed.ncbi.nlm.nih.gov/19692729/).
46. Terán-Escandón D, Terán-Ortiz L, Camarena-Olvera A, González-Avila G, Vaca-Marín MA, Granados J, et al. Human Leukocyte Antigen-Associated susceptibility to pulmonary tuberculosis: Molecular Analysis of Class II Alleles by DNA Amplification and Oligonucleotide Hybridization in Mexican Patients. *CHEST*. 1999; 115:428–33. PMID: [10027443](https://pubmed.ncbi.nlm.nih.gov/10027443/)
47. Ravikumar M, Dheenadhayalan V, Rajaram K, Shanmuga Lakshmi S, Paul Kumaran P, Paramasivan CN, et al. Associations of HLA-DRB1, DQB1 and DPB1 alleles with pulmonary tuberculosis in south India. *Tubercle and Lung Disease*. 1999; 79(5):309–17. PMID: [10707259](https://pubmed.ncbi.nlm.nih.gov/10707259/)
48. Kuranov AB, Kozhamkulov UA, Vavilov MN, Belova ES, Bismilda VL, Alenova AH, et al. HLA-class II alleles in patients with drug-resistant pulmonary tuberculosis in Kazakhstan. *Tissue antigens*. 2014; 83(2):106–12. Epub 2014/01/09. doi: [10.1111/tan.12279](https://doi.org/10.1111/tan.12279) PMID: [24397488](https://pubmed.ncbi.nlm.nih.gov/24397488/).
49. Vandiedonck C, Knight JC. The human Major Histocompatibility Complex as a paradigm in genomics research. *Briefings in functional genomics & proteomics*. 2009; 8(5):379–94. doi: [10.1093/bfpg/elp010](https://doi.org/10.1093/bfpg/elp010) PMID: [19468039](https://pubmed.ncbi.nlm.nih.gov/19468039/); PubMed Central PMCID: [PMC2987720](https://pubmed.ncbi.nlm.nih.gov/PMC2987720/).
50. WHO. Global status report on alcohol and health 2014. World Health Organization. 2014.
51. Mason CM, Dobard E, Zhang P, Nelson S. Alcohol Exacerbates Murine Pulmonary Tuberculosis. *Infection and immunity*. 2004; 72(5):2556–63. doi: [10.1128/IAI.72.5.2556-2563.2004](https://doi.org/10.1128/IAI.72.5.2556-2563.2004) PMID: [PMC387844](https://pubmed.ncbi.nlm.nih.gov/PMC387844/).
52. Patra J, Jha P, Rehm J, Suraweera W. Tobacco Smoking, Alcohol Drinking, Diabetes, Low Body Mass Index and the Risk of Self-Reported Symptoms of Active Tuberculosis: Individual Participant Data (IPD) Meta-Analyses of 72,684 Individuals in 14 High Tuberculosis Burden Countries. *PloS one*. 2014; 9(5):e96433. doi: [10.1371/journal.pone.0096433](https://doi.org/10.1371/journal.pone.0096433) PMID: [PMC4008623](https://pubmed.ncbi.nlm.nih.gov/PMC4008623/).
53. IEDB. Immune Epitope Database and Analysis Resource. 2015.
54. Jones EY, Fugger L, Strominger JL, Siebold C. MHC class II proteins and disease: a structural perspective. *Nature reviews Immunology*. 2006; 6(4):271–82. doi: [10.1038/nri1805](https://doi.org/10.1038/nri1805) PMID: [16557259](https://pubmed.ncbi.nlm.nih.gov/16557259/).
55. Godkin AJ, Smith KJ, Willis A, Tejada-Simon MV, Zhang J, Elliott T, et al. Naturally Processed HLA Class II Peptides Reveal Highly Conserved Immunogenic Flanking Region Sequence Preferences That Reflect Antigen Processing Rather Than Peptide-MHC Interactions. *The Journal of Immunology*. 2001; 166(11):6720–7. doi: [10.4049/jimmunol.166.11.6720](https://doi.org/10.4049/jimmunol.166.11.6720) PMID: [11359828](https://pubmed.ncbi.nlm.nih.gov/11359828/)
56. Joffre O, Nolte MA, Sporri R, Reis e Sousa C. Inflammatory signals in dendritic cell activation and the induction of adaptive immunity. *Immunol Rev*. 2009; 227(1):234–47. Epub 2009/01/06. doi: [10.1111/j.1600-065X.2008.00718.x](https://doi.org/10.1111/j.1600-065X.2008.00718.x) PMID: [19120488](https://pubmed.ncbi.nlm.nih.gov/19120488/).
57. Boechat NdO, Ogusku MM, Boechat AL, Sadahiro A. Interaction between Smoking and HLA-DRB1*04 Gene Is Associated with a High Cardiovascular Risk in Brazilian Amazon Patients with Rheumatoid Arthritis. *PloS one*. 2012; 7(8):e41588. doi: [10.1371/journal.pone.0041588](https://doi.org/10.1371/journal.pone.0041588) PMID: [22912672](https://pubmed.ncbi.nlm.nih.gov/22912672/)
58. Mourad J, Monem F. HLA-DRB1 allele association with rheumatoid arthritis susceptibility and severity in Syria. *Revista Brasileira de Reumatologia*. 2013; 53:51–6.
59. Lin L, Chen Y, Xiao Z, Huang S, Yang Z. The association of HLA-DRB1 alleles with rheumatoid arthritis in the Chinese Shantou population: a follow-up study. *Biochemistry and cell biology = Biochimie et biologie cellulaire*. 2007; 85(2):227–38. Epub 2007/05/31. doi: [10.1139/o06-204](https://doi.org/10.1139/o06-204) PMID: [17534404](https://pubmed.ncbi.nlm.nih.gov/17534404/).
60. Zanelli E, Breedveld FC, de Vries RR. HLA class II association with rheumatoid arthritis: facts and interpretations. *Human immunology*. 2000; 61(12):1254–61. Epub 2001/02/13. PMID: [11163080](https://pubmed.ncbi.nlm.nih.gov/11163080/).
61. Reed GW, Choi H, Lee SY, Lee M, Kim Y, Park H, et al. Impact of Diabetes and Smoking on Mortality in Tuberculosis. *PloS one*. 2013; 8(2):e58044. doi: [10.1371/journal.pone.0058044](https://doi.org/10.1371/journal.pone.0058044) PMID: [23469139](https://pubmed.ncbi.nlm.nih.gov/23469139/)
62. Lawn SD, Zumla AI. Tuberculosis. *The Lancet*. 2011; 378(9785):57–72. [http://dx.doi.org/10.1016/S0140-6736\(10\)62173-3](http://dx.doi.org/10.1016/S0140-6736(10)62173-3).
63. Tsuneto LT, Probst CM, Hutz MH, Salzano FM, Rodriguez-Delfin LA, Zago MA, et al. HLA class II diversity in seven Amerindian populations. Clues about the origins of the Ache. *Tissue antigens*. 2003; 62(6):512–26. Epub 2003/11/18. PMID: [14617035](https://pubmed.ncbi.nlm.nih.gov/14617035/).
64. Petzl-Erler ML, McDevitt HO. Molecular analysis of the HLA-DRB genes in two tribes of Brazilian Indians. *Human immunology*. 1994; 41(3):180–4. Epub 1994/11/01. PMID: [7868373](https://pubmed.ncbi.nlm.nih.gov/7868373/).

65. Gragert L, Madbouly A, Freeman J, Maiers M. Six-locus high resolution HLA haplotype frequencies derived from mixed-resolution DNA typing for the entire US donor registry. *Human immunology*. 2013; 74(10):1313–20. doi: [10.1016/j.humimm.2013.06.025](https://doi.org/10.1016/j.humimm.2013.06.025) PMID: [23806270](https://pubmed.ncbi.nlm.nih.gov/23806270/).
66. Vargas-Alarcon G, Gamboa R, Zuniga J, Hernandez-Pacheco G, Ramos-Kuri M, Castillo E, et al. HLA-DR4 allele frequencies on Indian and Mestizo population from Mexico. *Human immunology*. 2000; 61(3):341–4. Epub 2000/02/26. PMID: [10689126](https://pubmed.ncbi.nlm.nih.gov/10689126/).
67. Garcia-Ortiz JE, Sandoval-Ramirez L, Rangel-Villalobos H, Maldonado-Torres H, Cox S, Garcia-Sepulveda CA, et al. High-resolution molecular characterization of the HLA class I and class II in the Tarahumara Amerindian population. *Tissue antigens*. 2006; 68(2):135–46. Epub 2006/07/27. doi: [10.1111/j.1399-0039.2006.00636.x](https://doi.org/10.1111/j.1399-0039.2006.00636.x) PMID: [16866883](https://pubmed.ncbi.nlm.nih.gov/16866883/).