

RESEARCH ARTICLE

Open Access



Alpinia oxyphylla Miq. fruit extract activates IGFR-PI3K/Akt signaling to induce Schwann cell proliferation and sciatic nerve regeneration

Yung-Ming Chang^{1,2,3}, Hen-Hong Chang^{4,5,6}, Chin-Chuan Tsai^{1,2}, Hung-Jen Lin^{7,8}, Tsung-Jung Ho^{9,10}, Chi-Xin Ye¹¹, Ping-Ling Chiu³, Yueh-Sheng Chen⁹, Ray-Jade Chen¹², Chih-Yang Huang^{9,11,13*†} and Chien-Chung Lin^{14†}

Abstract

Background: It is known that the medicinal herb *Alpinia oxyphylla* Miq. is widely used as a remedy for diarrhea as well as the symptoms accompanying hypertension and cerebrovascular disorders. Moreover, it has also been reported that *Alpinia oxyphylla* Miq. has beneficial effects on anti-senescence and neuro-protection. This study focuses on the molecular mechanisms by which the *Alpinia oxyphylla* Miq. fruits promote neuron regeneration.

Methods: A piece of silicone rubber was guided across a 15 mm gap in the sciatic nerve of a rat. This nerve gap was then filled with various doses of *Alpinia oxyphylla* Miq. fruits to assess their regenerative effect on damaged nerves. Further, we investigated the role of *Alpinia oxyphylla* Miq. fruits in RSC96 Schwann cell proliferation.

Results: Our current results showed that treatment with the extract of *Alpinia oxyphylla* Miq. fruits triggers the phosphorylated insulin-like growth factor-1 receptor- phosphatidylinositol 3-kinase/serine-threonine kinase pathway, and up-regulated the proliferating cell nuclear antigen in a dose-dependent manner. Cell cycle analysis on RSC96 Schwann cells showed that, after exposure to *Alpinia oxyphylla* Miq. fruit extract, the transition from the first gap phase to the synthesis phase occurs in 12–18 h. The expression of the cell cycle regulatory proteins cyclin D1, cyclin E and cyclin A increased in a dose-dependent manner. Transfection with a small interfering RNA blocked the expression of phosphatidylinositol 3-kinase and induced down-regulation both on the mRNA and protein levels, which resulted in a reduction of the expression of the survival factor B-cell lymphoma 2.

Conclusion: We provide positive results that demonstrate that *Alpinia oxyphylla* Miq. fruits facilitate the survival and proliferation of RSC96 cells via insulin-like growth factor-1 signaling.

Keywords: *Alpinia oxyphylla* Miq. fruit extract, IGF-1, nerve regeneration, proliferation, RSC96 Schwann cell

Background

It is known that the different anatomical structures of neurons and their regenerative abilities contribute to the central and peripheral nervous systems. Neuron injury stimulates various physiological responses that facilitate nerve cell regeneration. The neurons of the central nervous system in mammals lack the myelin sheath and

therefore are incapable of regeneration. However, the neurons of the peripheral nervous system, which are surrounded by a myelin sheath, have the potential of regeneration and repair [1]. The ability of neuron regeneration is a result of intrinsic neuronal activities and other associated components, such as the Schwann cells. Schwann cells are capable of dedifferentiation, migration, proliferation, the expression of growth-promoting factors and the myelination of regenerating axons. After injury, the Schwann cells from the periphery migrate to the injured site to facilitate the repair processes [2, 3]. The growth factors that are produced by Schwann cells play an important

* Correspondence: cyhuang@mail.cmu.edu.tw

†Equal contributors

⁹School of Chinese Medicine, China Medical University, Taichung, Taiwan

¹¹Graduate Institute of Basic Medical Science, China Medical University, No 91, Hsueh-Shih Road, Taichung 404, Taiwan

Full list of author information is available at the end of the article



role in peripheral nerve repair. Therefore, enhancing Schwann cell proliferation might be a potential approach for neuron regeneration in neuron injury. On the other hand, insulin-like growth factor-1 (IGF-1) has been characterized as a biochemical marker that is secreted in response to growth hormone to stimulate tissue growth [4]. IGF-1 modulates muscle satellite cells to undergo activation, proliferation and differentiation, leading to muscle regeneration and hypertrophy [5].

Biomaterials in combination with Chinese herbal medicine have been effectively used in nerve regeneration-related research. A silicon rubber chamber filled with Schwann cells has been shown to repair and bridge a 15 mm length of abrasion in rat sciatic nerves [6]. Therefore, treating Schwann cells with Chinese herbal medicines to enhance their therapeutic potential in guiding neuron regrowth is considered a possible approach to treat nerve injury.

Alpinate Oxypiphyllae Fructus (*Alpinia oxyphylla* Miq., AOF) is a medicinal plant that is often used for treating ulcerations, gastralgia, diarrhea, dementia and tumors. Moreover, it has also been reported that AOF extracts exhibit potential neuro-protective effects against oxidative damage or neurotoxicity [7–12]. In our previous study, we evaluated whether AOF promotes RSC96 Schwann cell migration by ERK1/2, JNK and p38 signaling [13]. However, the beneficial effect of AOF on the nerve growth and regeneration facilitated by Schwann cells remains unclear.

In the present study, we compared the effects of AOF on Schwann cell proliferation and neuron regeneration in both in vivo and in vitro experiments. We examined the neuro-regenerative effect of AOF in animal models by injecting 0, 30, 60, 100, 150 or 200 mg/mL/kg concentrations of AOF into rat sciatic nerves, and in the in vitro model, we treated Schwann cells with 0, 20, 40, 60, 80, 100, 150 or 200 µg/mL of AOF.

Methods

AOF preparation

Fragments of AOF for research were purchased from the Shin-Long Pharmaceutical Company (Taichung, Taiwan, ROC). The AOF extract was prepared by boiling 150 g of AOF fragments in 600 mL of distilled water. The extract was filtered and the filtrate was concentrated under reduced pressure, and then stored at 4 °C until required. Finally, the AOF powdered extract was produced by spray drying.

Animal model and treatments

The surgery was performed as previously described [14]. Thirty-six healthy adult Sprague-Dawley rats (220 ± 20 g), were allocated randomly to six experimental groups ($n = 6$ each) and housed in silicone chambers. Each chamber

housed no more than 3 rats. The first group was the control group, and the chambers had a lumen volume of 25.5 µL, which administered saline only. The AOF groups 2–6 were administered AOF at 30, 60, 100, 150, and 200 mg/mL/kg, respectively.

All animals were housed in a standard housing room with chow and water available ad libitum. The housing room was maintained at 22 °C and 45% humidity, and kept under a regular 12 h light/dark cycle with lights on from 08:00 am to 20:00 pm.

Four weeks later, the animals were safely anesthetized using isoflurane, and the chamber connecting the nerve ends was re-exposed to examine the 15 mm gap within the chamber for evidence of successful nerve regeneration. The Institutional Animal Care and Use Committee (IACUC) of the China Medical University approved the protocols of animal use for experiment, and all animals maintained in the facilities were treated according to the principles of laboratory animal care (NIH publication).

Cell culture and treatments

RSC96 Schwann cells were obtained from the American Type Culture Collection (ATCC, GA, USA) and were cultured in Dulbecco's modified Eagle's medium (DMEM) modified to contain 100 U/mL penicillin, 100 µg/mL streptomycin, 10% fetal bovine serum (FBS), 25 mM glucose, 4 mM L-glutamate and 1500 mg/L sodium bicarbonate at 37 °C under a humidified atmosphere of 5% CO₂ and 95% air. After a 4-h period of serum-free culture, the RSC96 cells were treated with the indicated concentrations of AOF for 16–24 h, and then harvested for further analysis.

Flow cytometry

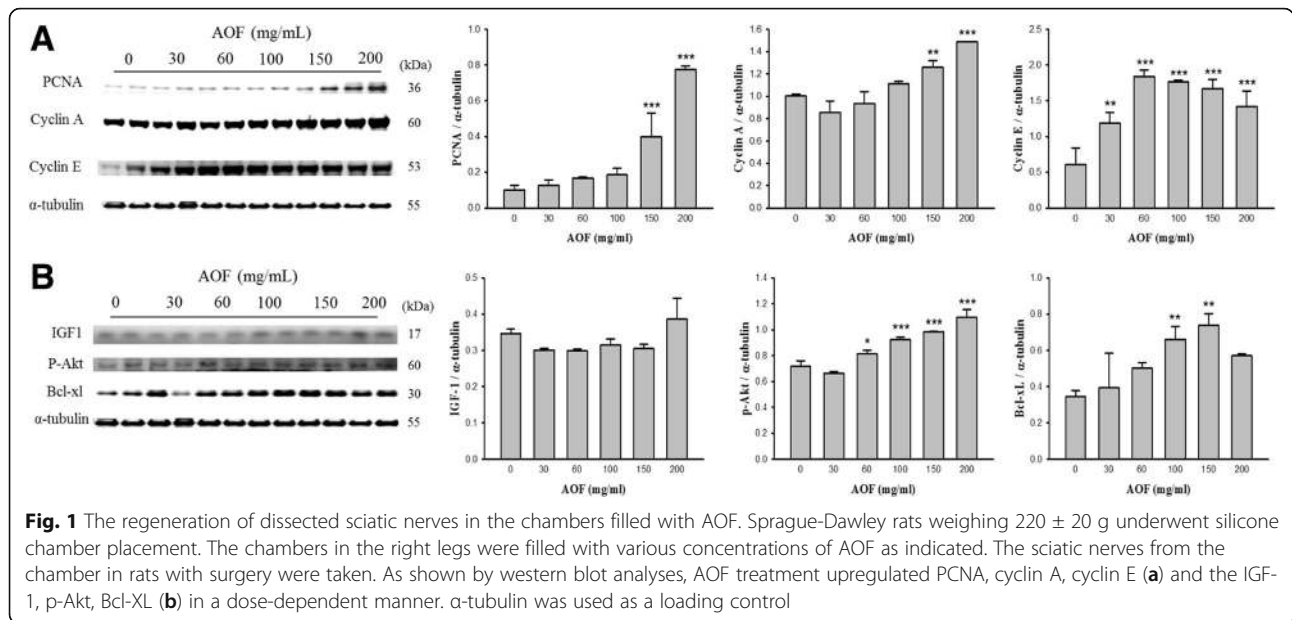
The cells were harvested and washed twice with phosphate buffered saline (PBS, pH 7.2), then fixed in 500 µL 70% (v/v) ice-cold ethanol at 4 °C overnight. The cells were then washed twice with PBS and stained with 0.005% propidium iodide (PI) at 4 °C in the dark for 30 min. To perform the cell cycle evaluation, the PI-stained cells were analyzed via a BD FACSCalibur cytometer and the data were quantified using the Modfit LT software. Each experiment was repeated three times.

Inhibitors

Two inhibitors were used in this study: AG1024 (IGF1R inhibitor; Promega, Madison, WI, USA) and LY294002 (PI3K inhibitor; Promega, Madison, WI, USA).

Migration assay

A Boyden chamber and 8-µm pore size polycarbonate membrane filters (Neuro Probes, Inc., Gaithersburg, MD, USA) were used to assess cell migration. The RSC96

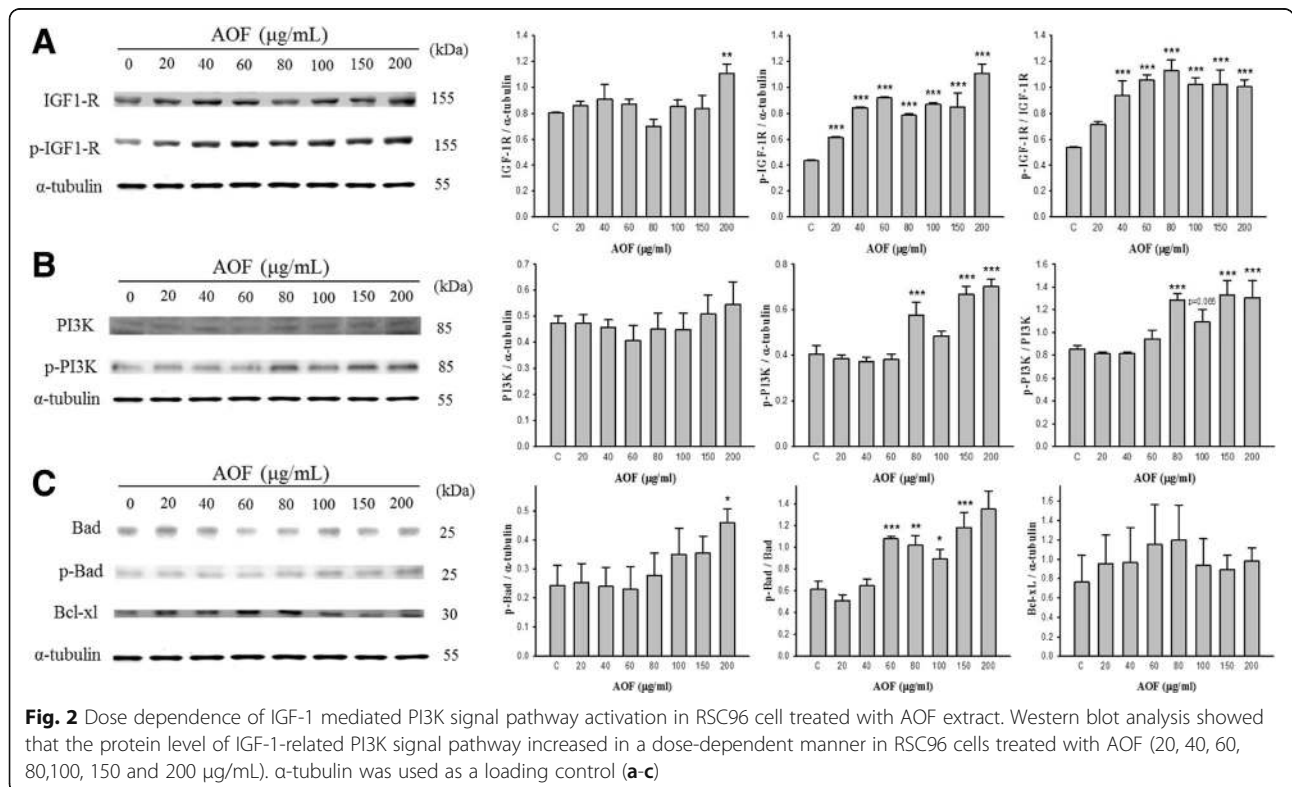


Schwann cells were added to the upper part of chamber, and the bottom chamber was filled with DMEM medium containing 10% FBS. After being incubated overnight, the cells were allowed to migrate through the filters. The cells on the membrane filter were fixed with methanol and Giemsa stain (Sigma, St. Louis, MO, USA). The number

of migrated cells was counted with a counting grid in five random fields.

Western blotting

Protein samples were separated in various percentages of SDS-polyacrylamide gels (8%, 10% or 12% separating



gels with 5% stacking gels). After electrophoresis was performed, the proteins were transferred to nitrocellulose membranes. The membranes were blocked with blocking buffer (5% skim milk, 150 mM NaCl, 20 mM Tris-HCl and 0.1% Tween 20) for 1 h at room temperature, washed three times in PBS, and subsequently incubated with the appropriate primary antibodies overnight at 4 °C. Then, the membranes were washed with TBST buffer 3 times and incubated with the secondary antibody for 1 h at room temperature. The membrane data were recorded using a LAS-4000 mini (GE Healthcare Life Sciences).

siRNA transfection

Double-stranded siRNA sequences targeting PI3K mRNAs were purchased from Dharmacon (Lafayette, CO, USA). The non-specific duplex (scramble) was non-targeting and used as a control. The transfection of siRNA was carried out with a transfection reagent (PureFection™, System Biosciences, Mountain View, CA) according to the manufacturer's instructions. To assess the gene silencing efficiency, the PI3K protein level was determined using western blot.

Statistical analysis

Each experiment was performed in triplicate. The results are presented as the mean \pm SEM, and the statistical comparisons were performed by Student's *t* test. Significance was defined at the $p < 0.05$, 0.001 or the 0.0001 level.

Results

AOF promotes regenerative signaling of damaged peripheral nerves

To identify the positive effects of AOF promoting damaged nerve regeneration, the rat sciatic nerves from the chamber treated with AOF were taken by surgery, and the modulation in the expression of proliferation-related proteins was examined. AOF activated the cell cycle as evidenced by increased levels of cyclin A, cyclin E, and proliferating cell nuclear antigen (PCNA) (Fig. 1a). To investigate the role of IGF-1 signaling in AOF-induced nerve proliferation, the activities of IGF-1 signaling in the regenerated nerves were examined, and the observations showed that the protein levels of IGF-1, p-Akt and BCL-xL were increased (Fig. 1b). These findings suggest that AOF promoted nerve cell regeneration.

Dose-dependent proliferation and survival of RSC96 cells treated with AOF

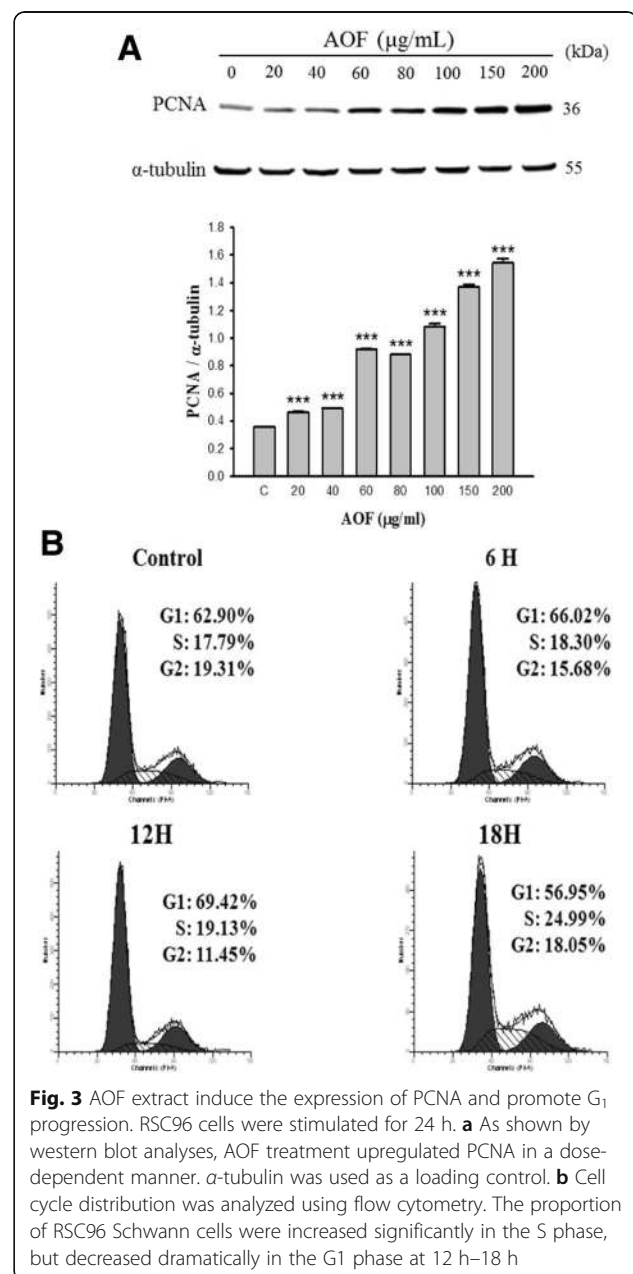
To evaluate the effect on proliferation of RSC96 Schwann cells, cell proliferation was examined after 24 h of treating with 0, 20, 40, 60, 80, 100, 150 and 200 μ g/mL of AOF extract. Western blot analysis revealed that both the survival- and proliferation-related proteins mediated by IGF-1 were markedly increased with AOF treatment (Fig. 2).

Our data demonstrate that IGF-1 expression was markedly increased with AOF treatment. Furthermore, the IGF-1 signaling-related protein expression of the PI3K and Akt were both rapidly induced in a dose-dependent manner.

These findings suggest that AOF potentially promoted Schwann cell proliferation and survival by activating the IGF-1-mediated survival signal mechanism.

AOF treatment stimulates G₁ phase cell cycle progression

We further investigated whether AOF stimulated the cell proliferation and related mechanisms. The levels of PCNA expression and the cell cycle distribution were



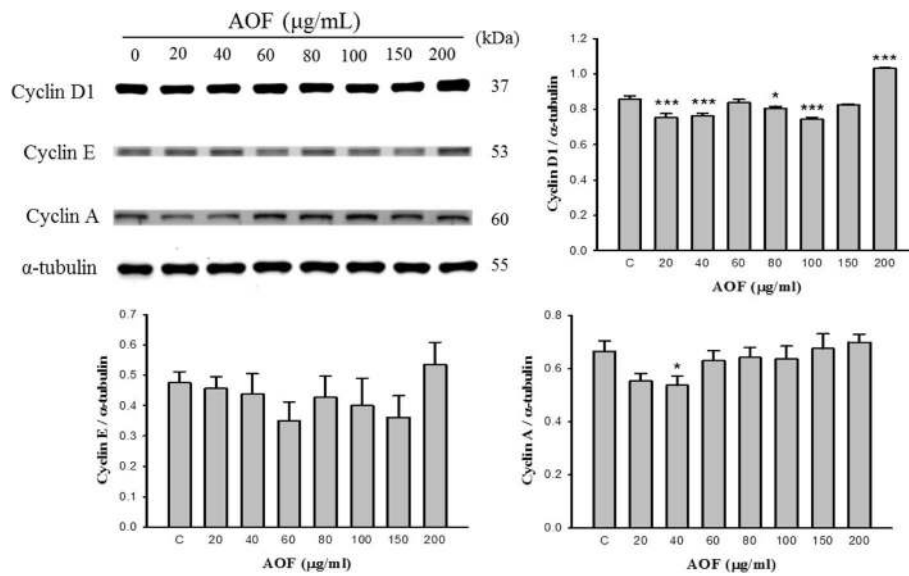


Fig. 4 AOF extract induces the expression of proteins involved in the cell cycle in a dose-dependent manner. RSC96 cells were stimulated with AOF for 24 h. The protein expressions of cell cycle regulatory proteins were determined by western blot analysis. Treatment with AOF upregulated cyclinD1, cyclinE and cyclinA in a dose-dependent manner. α-tubulin was used as a loading control

determined using western blot and flow cytometry, respectively. The results showed that PCNA levels increased in a dose-dependent manner (Fig. 3a).

Representative examples of the flow cytometry histograms showed that the cell cycle distribution of RSC96 Schwann cells were altered apparently in the time-course AOF treatment experiments. Furthermore, the proportion of RSC96 Schwann cells in the S phase increased significantly from 17.79% to 24.99%, and the proportion of cells in the G1 phase decreased dramatically from 69.42% to

56.95% at 12 h–18 h (Fig. 3b). These results suggest that AOF likely facilitated cell proliferation by accelerating the cell cycle progression.

AOF increases the level of cell cycle proliferative proteins

It is known that cell cycle progression is controlled by a regulatory network; cyclins are one of the important regulatory molecules involved in cell cycle control. To evaluate how AOF regulates the cell cycle to facilitate RSC96 Schwann cell proliferation, a western blot was

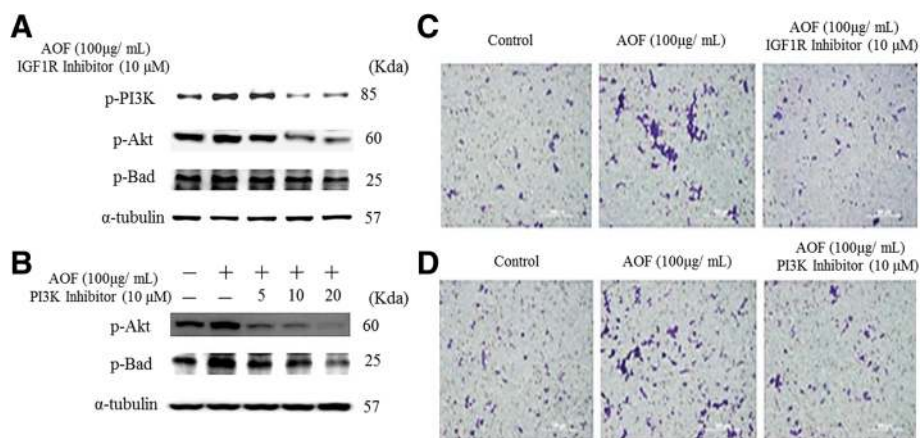


Fig. 5 AOF extract effects on Schwann cell proliferation was IGF-1signaling dependent. RSC96 cells were pretreated with IGF1R inhibitor (a) or PI3K inhibitor (b) for 1 h, and then treated with 100 µg/ml AOF extract for 24 h. Western blot analysis showed that AOF-induced the up-regulation of p-PI3K, p-Akt and p-Bad which was markedly suppressed when treated with either AG1024 (IGF1R inhibitor; Promega, Madison, WI, USA) or LY294002 (PI3K inhibitor; Promega, Madison, WI, USA) inhibitor.α-tubulin was used as a load control. After incubation with AOF extract, migration assay was performed using Boyden chambers treated with IGF1R inhibitor and PI3K inhibitor, respectively (c and d). The number of migrated cells was much more in the cells treated with AOF extract, but the migration effect was blocked when treated with IGF1R inhibitor and PI3K inhibitor

performed to characterize the key proteins of cell cycle regulation (Fig. 4). In addition, the AOF treatment induced the up-regulation of cyclin A expression in a dose-dependent manner. These results implied that AOF increases the level of cyclins, further stimulating the proliferation of RSC96 Schwann cells.

RSC96 Schwann Cell proliferation enhanced by AOF was associated with IGF-1 and PI3K/Akt signaling

Because a relationship between AOF-triggered IGF-1 signaling with cell proliferation was found in RSC96 Schwann cells, we explored whether the activation of the IGF-1R and/or the PI3K signaling pathway is necessary for the AOF-induced cell proliferation. RSC96 Schwann cells were pretreated with AG1024 and LY294002 as specific inhibitors of the IGF-1R and PI3K/Akt, respectively, followed by incubation with 100 µg/mL AOF extract for 24 h. This study demonstrated that the AOF-induced up-regulation of p-PI3K, p-Akt and p-Bad was markedly suppressed by the inhibitor treatment (Fig. 5a and b). To further confirm these results, we used the siRNA-induced knockdown of p-IGF-1R and p-PI3K proteins (Fig. 6). The results of western blot analyses demonstrated decreased expression of p-PI3K, p-Akt and p-Bad.

These results implied that AOF extract-induced Schwann cell proliferation occurs by the activation of the IGF-1R-mediated survival pathways.

Discussion

Previous reports have proposed the well model for assessing the capacity of regenerating neurons that are filled with a mixture of AOF extract and Schwann cells in a silicone rubber chamber for bridging a 15-mm rat sciatic nerve gap [14–18]. It has also been reported that Schwann cells are vital requirements in the injured nerve area, which could trigger nerve regeneration by trophic factors and the generation and release of adhesion molecules [19, 20]. AOF is a widely used traditional Chinese medicine, which has been reported to have a potential neuroprotective effect [8–12]. However, for considering the clinical application of AOF, it is necessary to determine the mechanism by which AOF-extract promotes the survival and proliferation of Schwann cells. The mechanism involved in the coordinated events is presented in Fig. 7.

Neurotrophic factors, including IGFs, are a set of growth factors that influence the growth and regeneration of neurons [21–25]. According to recent reports, IGF-1 plays an important role in cell survival and Schwann cell proliferation [4, 26]. Regenerating peripheral nerves show high IGF-1 expression in their advancing growth cone. The IGF-1 expression activates the PI3K/Akt signaling pathway and is associated with the proliferation of Schwann cells and regeneration of peripheral nerves [27–32].

AOF extract also induced cell proliferation via activating the PI3K/Akt signaling pathway in RSC96 cells. The

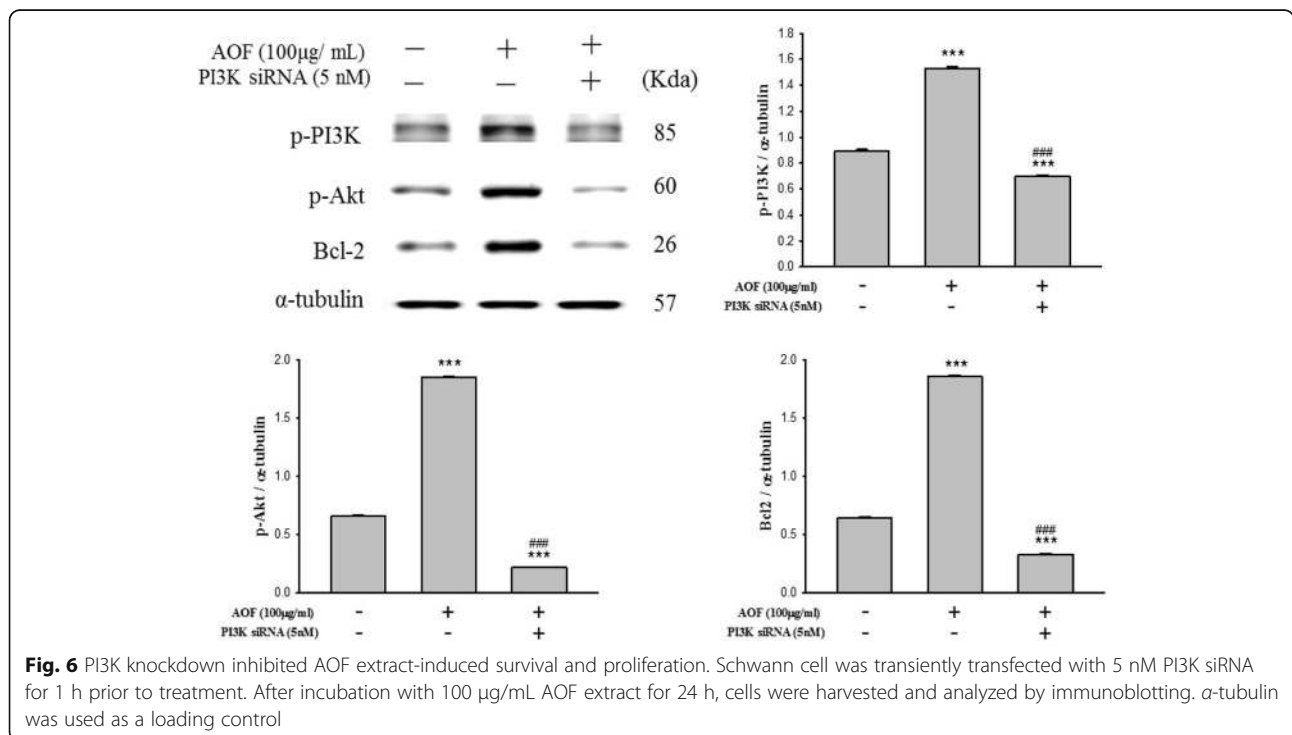
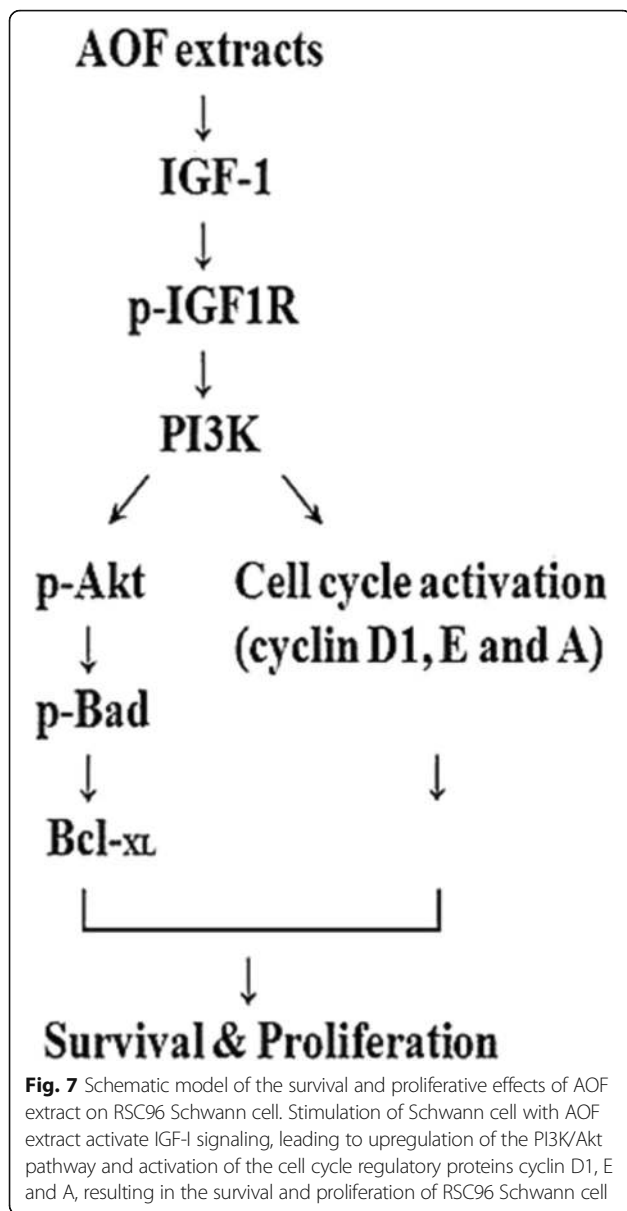


Fig. 6 PI3K knockdown inhibited AOF extract-induced survival and proliferation. Schwann cell was transiently transfected with 5 nM PI3K siRNA for 1 h prior to treatment. After incubation with 100 µg/mL AOF extract for 24 h, cells were harvested and analyzed by immunoblotting. α-tubulin was used as a loading control



PI3K signaling induced by IGF-1 attenuates Schwann cell apoptosis by caspase activation and is also known to facilitate the transition of cell cycle from the G_1 to S phase [31, 33]. Inhibition of PI3K effectively inhibits the IGF-1-induced anti-apoptotic and neuro-protective effects, and exhibits the significant role of PI3K in the induced survival of Schwann cells [29, 30]. Bioactive compounds that induce the up-regulation of IGF-1 can potentially activate the PI3K/Akt signaling and lead to modulating the expression of regulatory proteins (Bcl-2, p-Bad, PCNA and cyclins) to favor Schwann cell survival and proliferation during nerve regeneration. We show a beneficial effect in the up-regulation of the IGF-1-related survival pathway and the stimulation of cell proliferation in cultured RSC96 cells following AOF treatment.

Cell proliferation is regulated at various levels, and the G_1/S cell cycle transition is one of the most important requirements for DNA replication and mitosis.

Progression through the G_1 phase into the S phase requires cyclin D and cyclin E activity; whereas, Cyclin A activity is required for the DNA replication of S phase and mitosis initiation [34–36]. Our current results showed that the G_1/S transition of cell cycle progression was induced by AOF extract treatment (Fig. 3b). The time course study revealed that the AOF extract treatment is effective in cyclin A up-regulation, subsequent promotion of DNA replication and the increasing cell number in the S phase, which eventually results in cell proliferation. However, the expressions of cyclin D1 and cyclin E did not show any significant change. In certain cells, such as hematopoietic cells, the IGF expression acts as an inhibitor of cell death [37, 38]. However, IGF-1 has also been shown to stimulate cell cycle progression via G_1 or the G_0/G_1 transition in cultured fibroblasts and mammary epithelial cells [39–41]. We also provide data demonstrating that the cell cycle is driven by cyclins with corresponding changes in IGF-1 levels. AOF-induced alterations of the cell cycle in RSC96 cells may serve as critical determinants for nerve cell proliferation and IGF-1-related cell survival. Despite the fact that we cannot exclude other minor candidates involved in the related pathways, we clearly showed that certain proteins of the IGF-1-related signal pathway were up-regulated following AOF extract treatment.

Systematic survey on literatures shows that over 80 chemical constituents which were identified from *A. oxyphylla* including diarylheptanoids, flavonoids, polyphenols, sesquiterpenes, steroids, volatile oil and their glycosides, etc. [42–48]. Emerging studies evaluated that the abundance of nine secondary metabolites differentially concentrated in seeds and fruit capsules of *A. oxyphylla*, including sesquiterpenes (e.g., nootkatone), diarylheptanoids (e.g., oxyphyllacinol and yakuchinone A and B) and flavonoids (e.g., chrysin, izalpinin, tectochrysin, kaempferide and apigenin-4',7-dimethylether) [49, 50]. Protocatechuic acid (PCA), one of the major metabolite of complex polyphenols, which has been identified in the kernels of AOF [9, 51, 52], it also has been reported that PCA process positive effect on antioxidant, anti-inflammatory, and antiapoptosis [8–11, 53]. Our recent studies revealed that PCA promoted RSC96 Schwann cells regeneration and migration by activating ERK1/2, JNK1/2 and p38 MAPK pathways [54]. It also been reported that PCA effectively promoted RSC96 cell survival and proliferation by activating IGF-IR-PI3K-Akt signaling [55]. The protective effects associated with the chemical constituents of PCA correlates with the pharmacological activities of AOF as defined in the present study.

Conclusion

In summary, it was found that AOF extract significantly increases the IGF-1-related PI3K/Akt pathway in RSC96 Schwann cells, and upregulates the cell cycle regulatory proteins cyclin A, cyclin D1 and cyclin E. Therefore, AOF extract might be effective for RSC96 Schwann cell proliferation and survival.

Abbreviations

AKT: Protein kinase B; AOF: *Alpinia oxyphylla* Miq; Bad: Bcl-2-associated death promoter; Bcl-2: B-cell lymphoma-2; Bcl-xL: B-cell lymphoma-extra large; DMEM: Dulbecco's modified eagle's medium; FBS: Fetal bovine serum; IGF-1: Insulin-like growth factor-1; IGF-1R: Insulin-like growth factor-1 receptor; PBS: Phosphate buffered saline; PCNA: Proliferating cell nuclear antigen; PI: Propidium iodide; PI3K: Phosphoinositide 3 kinase; SDS-PAGE: Sodium dodecyl sulfate polyacrylamide gel electrophoresis

Acknowledgments

The authors would like to thank all of the colleagues and students who contributed to this study.

Funding

This study is supported in part by the Taiwan Ministry of Health and Welfare Clinical Trial and Research Center of Excellence (MOHW105-TDU-B-212-133,019), and also supported by the China Medical University under the Aim for Top University Plan of the Ministry of Education, Taiwan.

Availability of data and materials

The datasets supporting the conclusions of this article are included within this article.

Authors' contributions

The authors' contributions were as follows: Y-MC and C-YH designed the experiments; H-HC, C-CT, H-JL and C-XY acquired and analyzed the results; C-CT contributed material support; T-JH, P-LC and Y-SC interpreted the results; R-JC and C-CL performed statistical analysis; and Y-MC and C-YH prepared and edited the manuscript. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Consent for publication

Not applicable.

Ethics approval

All procedures in the animal studies complied with the standards for the care and use of experimental animals and were approved by the China Medical University for Accredited Laboratory Animal Care, according to the regulations and standards of the National Science Council of Health of the Republic of China.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

¹The School of Chinese Medicine for Post-Baccalaureate, I-Shou University, Kaohsiung 840, Taiwan. ²Chinese Medicine Department E-DA Hospital, Kaohsiung 824, Taiwan. ³PT Biotechnology Co., Ltd., Taichung 433, Taiwan. ⁴Research Center for Chinese Medicine & Acupuncture, China Medical University, Taichung 404, Taiwan. ⁵Departments of Chinese Medicine, China Medical University Hospital, Taichung 404, Taiwan. ⁶School of Post-Baccalaureate Chinese Medicine, College of Chinese Medicine, China Medical University, Taichung 404, Taiwan. ⁷Division of Chinese Internal Medicine, China Medical University Hospital, Taichung 404, Taiwan. ⁸School of Post-Baccalaureate Chinese Medicine, China Medical University, Taichung 404, Taiwan. ⁹School of Chinese Medicine, China Medical University, Taichung, Taiwan. ¹⁰Chinese Medicine Department, China Medical University Beigang Hospital, Beigang, Yunlin 651, Taiwan. ¹¹Graduate Institute of Basic

Medical Science, China Medical University, No 91, Hsueh-Shih Road, Taichung 404, Taiwan. ¹²Department of Surgery, School of Medicine, College of Medicine, Taipei Medical University, Taipei, Taiwan. ¹³Department of Health and Nutrition Biotechnology, Asia University, Taichung, Taiwan. ¹⁴Orthopaedic Department Armed Forces General Hospital, Taichung, Taiwan.

Received: 23 June 2016 Accepted: 17 March 2017

Published online: 31 March 2017

Reference

- Bunge RP. Expanding roles for the Schwann cell: ensheathment, myelination, trophism and regeneration. *Curr Opin Neurobiol.* 1993;3(5):805–9.
- Oudega M, Xu XM. Schwann cell transplantation for repair of the adult spinal cord. *J Neurotrauma.* 2006;23(3–4):453–67.
- Navarro X, Vivo M, Valero-Cabre A. Neural plasticity after peripheral nerve injury and regeneration. *Prog Neurobiol.* 2007;82(4):163–201.
- Le Roith D, Butler AA. Insulin-like growth factors in pediatric health and disease. *J Clin Endocrinol Metab.* 1999;84(12):4355–61.
- Philippou A, Halapas A, Maridaki M, Koutsilieris M. Type I insulin-like growth factor receptor signaling in skeletal muscle regeneration and hypertrophy. *J Musculoskelet Neuronal Interact.* 2007;7(3):208–18.
- Shen JL, Chen YS, Lin JY, Tien YC, Peng WH, Kuo CH, Tzang BS, Wang HL, Tsai FJ, Chou MC, et al. Neuron Regeneration and Proliferation Effects of Danshen and Tanshinone IIA. *Evid. Based Complement. Alternat. Med.* 2011; 2011:378907.
- Chang YM, Tsai CT, Wang CC, Chen YS, Lin YM, Kuo CH, Tzang BS, Chen RJ, Tsai FJ, Huang CY. Alpinate oxyphyllae fructus (*Alpinia Oxyphylla* Miq) extracts inhibit angiotensin-II induced cardiac apoptosis in H9c2 cardiomyoblast cells. *Biosci Biotechnol Biochem.* 2013;77(2):229–34.
- Shi GF, An LJ, Jiang B, Guan S, Bao YM. *Alpinia* protocathechuic acid protects against oxidative damage in vitro and reduces oxidative stress in vivo. *Neurosci Lett.* 2006;403(3):206–10.
- Koo BS, Lee WC, Chang YC, Kim CH. Protective effects of alpiniae oxyphyllae fructus (*Alpinia oxyphylla* MIQ) water-extracts on neurons from ischemic damage and neuronal cell toxicity. *Phytother Res.* 2004;18(2):142–8.
- Yu X, An L, Wang Y, Zhao H, Gao C. Neuroprotective effect of *Alpinia oxyphylla* Miq. fruits against glutamate-induced apoptosis in cortical neurons. *Toxicol Lett.* 2003;144(2):205–12.
- An LJ, Guan S, Shi GF, Bao YM, Duan YL, Jiang B. Protocatechuic acid from *Alpinia oxyphylla* against MPP+–induced neurotoxicity in PC12 cells. *Food Chem Toxicol.* 2006;44(3):436–43.
- Shui G, Bao YM, Bo J, An LJ. Protective effect of protocatechuic acid from *Alpinia oxyphylla* on hydrogen peroxide-induced oxidative PC12 cell death. *Eur J Pharmacol.* 2006;538(1–3):73–9.
- Chang YM, Ye CX, Ho TJ, Tsai TN, Chiu PL, Tsai CC, Lin YM, Kuo CH, Tsai FJ, Tsai CH, et al. *Alpinia oxyphylla* Miquel fruit extract activates MAPK-mediated signaling of PAs and MMP2/9 to induce Schwann cell migration and nerve regeneration. *Int J Artif Organs.* 2014;37(5):402–13.
- Chen YS, Liu CJ, Cheng CY, Yao CH. Effect of bilobalide on peripheral nerve regeneration. *Biomaterials.* 2004;25(3):509–14.
- Shoji N, Umeyama A, Takemoto T, Ohizumi Y. Isolation of a cardiotonic principle from *Alpinia oxyphylla*. *Planta Med.* 1984;50(2):186–7.
- Cheng CY, Yao CH, Liu BS, Liu CJ, Chen GW, Chen YS. The role of astragaloside in regeneration of the peripheral nerve system. *J Biomed Mater Res A.* 2006;76(3):463–9.
- Chen HT, Tsai YL, Chen YS, Jong GP, Chen WK, Wang HL, Tsai FJ, Tsai CH, Lai TY, Tzang BS, et al. Dangshen (*Codonopsis pilosula*) activates IGF-I and FGF-2 pathways to induce proliferation and migration effects in RSC96 Schwann cells. *Am. J. Chin. Med.* 2010;38(2):359–72.
- Liu SP, Hsu CY, Fu RH, Huang YC, Chen SY, Lin SZ, Shyu WC. *Sambucus williamsii* induced embryonic stem cells differentiated into neurons. *Biomedicine.* 2015;5(1):3.
- Jessen KR, Mirsky R. Schwann cells and their precursors emerge as major regulators of nerve development. *Trends Neurosci.* 1999;22(9):402–10.
- Fawcett JW, Keynes RJ. Peripheral-Nerve Regeneration. *Annu Rev Neurosci.* 1990;13:43–60.
- Ishii DN, Glazner GW, Whalen LR. Regulation of Peripheral-Nerve Regeneration by Insulin-Like Growth-Factors. *Ann N Y Acad Sci.* 1993;692:172–82.
- Savignat M, De-Doncker L, Vodouhe C, Garza JM, Lavalle P, Libersa P. Rat nerve regeneration with the use of a polymeric membrane loaded with NGF. *J Dent Res.* 2007;86(11):1051–6.

23. Grumbles RM, Sesodia S, Wood PM, Thomas CK. Neurotrophic Factors Improve Motoneuron Survival and Function of Muscle Reinnervated by Embryonic Neurons. *J Neuropath Exp Neurol*. 2009;68(7):736–46.
24. Geremia NM, Pettersson LME, Hasmatali JC, Hryciw T, Danielsen N, Schreyer DJ, Verge VMK. Endogenous BDNF regulates induction of intrinsic neuronal growth programs in injured sensory neurons. *Exp Neurol*. 2010;223(1):128–42.
25. Ishii DN, Glazner GW, Pu SF. Role of Insulin-Like Growth-Factors in Peripheral-Nerve Regeneration. *Pharmacol Therapeut*. 1994;62(1–2):125–44.
26. Koriyama Y, Homma K, Sugitani K, Higuchi Y, Matsukawa T, Murayama D, Kato S. Upregulation of IGF-I in the goldfish retinal ganglion cells during the early stage of optic nerve regeneration. *Neurochem Int*. 2007;50(5):749–56.
27. Hansson HA. Insulin-like growth factors and nerve regeneration. *Ann N Y Acad Sci*. 1993;692:161–71.
28. Pu SF, Zhuang HX, Ishii DN. Differential spatio-temporal expression of the insulin-like growth factor genes in regenerating sciatic nerve. *Brain Res Mol Brain Res*. 1995;34(1):18–28.
29. Mairet-Coello G, Tury A, DiCicco-Bloom E. Insulin-Like Growth Factor-1 Promotes G(1)/S Cell Cycle Progression through Bidirectional Regulation of Cyclins and Cyclin-Dependent Kinase Inhibitors via the Phosphatidylinositol 3-Kinase/Akt Pathway in Developing Rat Cerebral Cortex. *J Neurosci*. 2009; 29(3):775–88.
30. Leininger GM, Backus C, Uhler MD, Lentz SJ, Feldman EL. Phosphatidylinositol 3-kinase and Akt effectors mediate insulin-like growth factor-I neuroprotection in dorsal root ganglia neurons. *FASEB J*. 2004;18(11):1544–.
31. Hu CL, Cowan RG, Harman RM, Quirk SM. Cell cycle progression and activation of Akt kinase are required for insulin-like growth factor I-mediated suppression of apoptosis in granulosa cells. *Mol Endocrinol*. 2004;18(2):326–38.
32. Campana WM, Darin SJ, O'Brien JS. Phosphatidylinositol 3-kinase and Akt protein kinase mediate IGF-I- and prosaptide-induced survival in Schwann cells. *J Neurosci Res*. 1999;57(3):332–41.
33. Delaney CL, Cheng HL, Feldman EL. Insulin-like growth factor-I prevents caspase-mediated apoptosis in Schwann cells. *J Neurobiol*. 1999;41(4):540–8.
34. Sherr CJ. G1 Phase Progression - Cycling on Cue. *Cell*. 1994;79(4):551–5.
35. Florenes VA, Maelandsmo GM, Faye R, Nesland JM, Holm R. Cyclin A expression in superficial spreading malignant melanomas correlates with clinical outcome. *J Pathol*. 2001;195(5):530–6.
36. Walker DH, Maller JL. Role for Cyclin-a in the Dependence of Mitosis on Completion of DNA-Replication. *Nature*. 1991;354(6351):314–7.
37. Williams GT, Smith CA, Spooncer E, Dexter TM, Taylor DR. Haemopoietic colony stimulating factors promote cell survival by suppressing apoptosis. *Nature*. 1990;343(6253):76–9.
38. Liu LC, Su CH, Wang HC, Chang WS, Tsai CW, Maa MC, Tsai CH, Tsai FJ, Bau DT. Contribution of personalized Cyclin D1 genotype to triple negative breast cancer risk. *Biomedicine*. 2014;4:3.
39. Clemmons DR, Vanwyk JJ, Pledger WJ. Sequential Addition of Platelet Factor and Plasma to Balb-C3t3 Fibroblast-Cultures Stimulates Somatomedin-C Binding Early in Cell-Cycle. *P Natl Acad Sci-Biol*. 1980;77(11):6644–8.
40. Olashaw NE, Vanwyk JJ, Pledger WJ. Control of Late G0/G1 Progression and Protein Modification by Smc/Igf-I. *Am J Phys*. 1987;253(4):C575–9.
41. Stull MA, Richert MM, Loladze AV, Wood TL. Requirement for IGF-I in epidermal growth factor-mediated cell cycle progression of mammary epithelial cells. *Endocrinology*. 2002;143(5):1872–9.
42. Miyazawa M, Nakamura Y, Ishikawa Y. Insecticidal sesquiterpene from *Alpinia oxyphylla* against *Drosophila melanogaster*. *J Agric Food Chem*. 2000; 48(8):3639–41.
43. Morikawa T, Matsuda H, Toguchida I, Ueda K, Yoshikawa M. Absolute stereostructures of three new sesquiterpenes from the fruit of *Alpinia oxyphylla* with inhibitory effects on nitric oxide production and degranulation in RBL-2H3 cells. *J Nat Prod*. 2002;65(10):1468–74.
44. Shoji N, Umeyama A, Asakawa Y, Takemoto T, Nomoto K, Ohizumi Y. Structural determination of nootkatol, a new sesquiterpene isolated from *Alpinia oxyphylla* Miquel possessing calcium-antagonistic activity. *J Pharm Sci*. 1984;73(6):843–4.
45. Lv XQ, Luo JG, Wang XB, Wang JS, Luo J, Kong LY. Four new sesquiterpenoids from the fruits of *Alpinia oxyphylla*. *Chem Pharm Bull (Tokyo)*. 2011;59(3):402–6.
46. Qing ZJ, Yong W, Hui LY, Yong LW, Long LH, Ao DJ, Xia PL. Two new natural products from the fruits of *Alpinia oxyphylla* with inhibitory effects on nitric oxide production in lipopolysaccharide-activated RAW264.7 macrophage cells. *Arch Pharm Res*. 2012;35(12):2143–6.
47. Hou L, Ding G, Guo B, Huang W, Zhang X, Sun Z, Shi X. New sesquiterpenoids and a diterpenoid from *Alpinia oxyphylla*. *Molecules*. 2015;20(1):1551–9.
48. Song W, Li Y, Wang J, Li Z, Zhang J. Characterization of nucleobases and nucleosides in the fruit of *Alpinia oxyphylla* collected from different cultivation regions. *Drug Test Anal*. 2014;6(3):239–45.
49. Chen F, Li HL, Tan YF, Guan WW, Zhang JQ, Li YH, Zhao YS, Qin ZM. Different accumulation profiles of multiple components between pericarp and seed of *Alpinia oxyphylla* capsular fruit as determined by UFLC-MS/MS. *Molecules*. 2014;19(4):4510–23.
50. Chen F, Li HL, Tan YF, Li YH, Lai WY, Guan WW, Zhang JQ, Zhao YS, Qin ZM. Identification of known chemicals and their metabolites from *Alpinia oxyphylla* fruit extract in rat plasma using liquid chromatography/tandem mass spectrometry (LC-MS/MS) with selected reaction monitoring. *J Pharm Biomed Anal*. 2014;97:166–77.
51. Vitaglione P, Donnarumma G, Napolitano A, Galvano F, Gallo A, Scalfi L, Fogliano V. Protocatechuic acid is the major human metabolite of cyanidin-glucosides. *J Nutr*. 2007;137(9):2043–8.
52. Pimpao RC, Dew T, Figueira ME, McDougall GJ, Stewart D, Ferreira RB, Santos CN, Williamson G. Urinary metabolite profiling identifies novel colonic metabolites and conjugates of phenolics in healthy volunteers. *Mol Nutr Food Res*. 2014;58(7):1414–25.
53. Semaming Y, Pannengpetch P, Chattipakorn SC, Chattipakorn N. Pharmacological properties of protocatechuic Acid and its potential roles as complementary medicine. *Evid Based Complement Alternat Med*. 2015; 2015:593902.
54. Ju DT, Kuo WW, Ho TJ, Paul CR, Kuo CH, Viswanadha VP, Lin CC, Chen YS, Chang YM, Huang CY. Protocatechuic Acid from *Alpinia oxyphylla* Induces Schwann Cell Migration via ERK1/2, JNK and p38 Activation. *Am J Chin Med*. 2015;43(4):653–65.
55. Ju DT, Liao HE, Shibu MA, Ho TJ, Padma W, Tsai FJ, Chung LC, Day CH, Lin CC, Huang CY. Nerve Regeneration Potential of Protocatechuic Acid in RSC96 Schwann Cells by Induction of Cellular Proliferation and Migration through IGF-IR-PI3K-Akt Signaling. *Chin J Physiol*. 2015;58(6):412–9.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at
www.biomedcentral.com/submit

