



UPPSALA
UNIVERSITET

*Digital Comprehensive Summaries of Uppsala Dissertations
from the Faculty of Medicine 1182*

Altered proteins in the aging brain

ADILA ELOBEID



ACTA
UNIVERSITATIS
UPSALIENSIS
UPPSALA
2016

ISSN 1651-6206
ISBN 978-91-554-9482-7
urn:nbn:se:uu:diva-277214

Dissertation presented at Uppsala University to be publicly examined in Fåhraeusalen, Rudbecklaboratoriet, Dag Hammarskjölds väg 20, Uppsala, Friday, 8 April 2016 at 09:15 for the degree of Doctor of Philosophy (Faculty of Medicine). The examination will be conducted in English. Faculty examiner: Professor Thomas Brännström (Department of Medical Biosciences, Umeå University).

Abstract

Elobeid, A. 2016. Altered proteins in the aging brain. *Digital Comprehensive Summaries of Uppsala Dissertations from the Faculty of Medicine* 1182. 53 pp. Uppsala: Acta Universitatis Upsaliensis. ISBN 978-91-554-9482-7.

The classification of neurodegenerative disorders is based on the major component of the protein aggregates in the brain. The most common altered proteins associated with neurodegeneration are Hyperphosphorylated tau (HP τ), beta amyloid (A β), alpha-synuclein (α S) and transactive response DNA binding protein 43 (TDP43). In this study we assessed the incidence and the neuroanatomical distribution of proteins associated with neurodegeneration in the brain tissue of cognitively unimpaired subjects.

We demonstrated the early involvement of the Locus Coeruleus (LC) with HP τ pathology in cognitively unimpaired mid aged subjects, a finding which supports the notion that LC is an initiation site of HP τ pathology. This may suggest that development of clinical assessment techniques and radiological investigations reflecting early LC alterations may help in identifying subjects with early stages of neurodegeneration.

Furthermore, we studied a large cohort of cognitively unimpaired subjects with age at death ≥ 50 years and we applied the National Institute on Aging –Alzheimer’s disease (AD) Association (NIA-AA) guidelines for the assessment of AD related neuropathological changes. Interestingly, a considerable percentage of the subjects were classified as having an intermediate level of AD pathology. We also showed that the altered proteins; HP τ , A β , α S, and TDP43 are frequently seen in the brain of cognitively unimpaired subjects with age at death ≥ 50 years, the incidence of these proteins increased significantly with age. This finding suggests that neurodegeneration has to be extensive to cause functional disturbance and clinical symptoms. Moreover, we investigated the correlation between AD related pathology in cortical biopsies, the AD / cerebrospinal fluid (CSF) biomarkers and the Mini Mental State examination (MMSE) scores in a cohort of idiopathic Normal Pressure Hydrocephalus (iNPH) patients. We demonstrated that AD/ CSF biomarkers and MMSE scores reflect AD pathology in the cortical biopsies obtained from iNPH patients.

In conclusion, this study shows that the altered proteins associated with neurodegeneration are frequently seen in the brain tissue of cognitively unimpaired aged subjects. This fact should be considered while developing diagnostic biomarkers for identification of subjects at early stages of the disease, in order to introduce therapeutic intervention prior to the occurrence of significant cognitive impairment.

Keywords: Cognitively unimpaired subjects, Hyperphosphorylated tau, Beta amyloid, Alpha-synuclein, Transactive response DNA binding protein 43

Adila Elobeid, Department of Immunology, Genetics and Pathology, Rudbecklaboratoriet, Uppsala University, SE-751 85 Uppsala, Sweden.

© Adila Elobeid 2016

ISSN 1651-6206

ISBN 978-91-554-9482-7

urn:nbn:se:uu:diva-277214 (<http://urn.kb.se/resolve?urn=urn:nbn:se:uu:diva-277214>)

To My Dear Aunt Adila

List of Papers

This thesis is based on the following papers, which are referred to in the text by their Roman numerals.

- I Elobeid A, Soininen H, Alafuzoff I. Hyperphosphorylated tau in young and middle-aged subjects. *Acta Neuropathol.* 2012; 123(1):97-104.
- II Elobeid A, Rantakömi S, Soininen H, Alafuzoff I. Alzheimer's disease-related plaques in nondemented subjects. *Alzheimers Dement.* 2014;10(5):522-9.
- III Elobeid A, Laurell K, Cesarini KG, Alafuzoff I. Correlations between mini-mental state examination score, cerebrospinal fluid biomarkers, and pathology observed in brain biopsies of patients with normal-pressure hydrocephalus. *J Neuropathol Exp Neurol.* 2015; 74(5):470-9.
- IV Elobeid A, Libard S, Leino M, Popova S, Alafuzoff I. Altered proteins in the aging brain. Accepted for publication in the *Journal of Neuropathology and Experimental Neurology*.

Reprints were made with permission from the respective publishers.

Contents

Introduction.....	11
Neurodegenerative proteinopathies.....	11
Tauopathies.....	11
α synucleinopathies.....	16
Transactive DNA binding protein 43 related proteinopathies.....	17
Other Proteinopathies.....	18
Concomitant proteinopathies in dementia.....	19
Neurodegenerative alterations in cognitively unimpaired subjects.....	19
HP τ in cognitively unimpaired subjects.....	19
A β in cognitively unimpaired subjects.....	19
α S in cognitively unimpaired subjects.....	20
TDP43 in cognitively unimpaired subjects.....	20
Concomitant proteinopathies in cognitively unimpaired subjects.....	20
Normal Pressure Hydrocephalus.....	21
Methodological considerations.....	21
The Present Investigation.....	22
Aims of the study.....	22
Subjects and Methods.....	23
Subjects.....	23
Clinical assessment.....	23
Neuropathological assessment.....	24
Immunohistochemistry.....	24
Staging of the pathological lesions.....	24
Semi quantitative analysis of the pathological alterations.....	26
Digital analysis.....	26
CSF analysis.....	27
Statistical analysis.....	27
RESULTS.....	28
HP τ in the cognitively unimpaired subjects.....	28
A β in the cognitively unimpaired subjects.....	30
α S in the cognitively unimpaired subjects.....	31
TDP43 in the cognitively unimpaired subjects.....	31
Concomitant pathologies in the cognitively unimpaired subjects.....	31
Applicability of the neuropathological criteria.....	32

DISCUSSION	34
HP τ in the cognitively unimpaired subjects	34
A β in the cognitively unimpaired subjects	35
α S in the cognitively unimpaired subject.....	37
TDP43 in the cognitively unimpaired subjects.....	37
Concomitant pathologies in the cognitively unimpaired subjects	38
Applicability of the neuropathological criteria	39
Methodological Considerations.....	41
Clinical assessment.....	41
Case Selection.....	41
Methods	41
Conclusion.....	43
Acknowledgments	44
References	45

Abbreviations

α S	α synuclein
A β	β amyloid
AD	Alzheimer's disease
AGD	Argyrophilic Grain Disease
ALS	Amyotrophic lateral sclerosis
CAA	Cerebral Amyloid Angiopathy
CBD	Cortico Basal Degeneration
CERAD	The Consortium to Establish Registry of Alzheimer's Disease
CSF	Cerebrospinal Fluid
DLB	Dementia with Lewy Bodies
FTLD	Frontotemporal lobar degeneration
FUS	Fused in Sacroma,
HP τ	Hyperphosphorylated τ
IHC	Immunohistochemistry
iLBRP	Incidental Lewy Body related pathology
LB	Lewy Body
LC	Locus Coeruleus
MSA	Multiple System Atrophy
NFTs	Neurofibrillary Tangles
NIA-RI	The National Institute of Aging and the Regan Institute
NIA-AA	The National Institute on Aging –Alzheimer's Association
NPs	Neuritic Plaques
NTs	Neuropil Threads
PART	Primary Age Related Tauopathy
PD	Parkinson's Disease
PDD	Parkinson's Disease with Dementia
PiD	Pick's Disease
PSP	Progressive Supranuclear Palsy
SOD1	Superoxidase dismutase-1
TDP43	Transactive response DNA binding protein 43

Introduction

The incidence of neurodegenerative disorders increases with age. During life, the diagnosis of these disorders is based on certain clinical presentation; however, diagnostic biomarkers are available for some diseases. The definite diagnosis is based on the results obtained from a neuropathological assessment of the brain tissue that is carried out post mortem. The neuropathological criteria have evolved since the 1990s due to the development of assessment strategies such as immunohistochemistry (IHC), a method that visualizes altered proteins in the brain tissue. Thus, many neurodegenerative diseases are currently referred to as proteinopathies.

Neurodegenerative proteinopathies

The current classification of age-related neurodegenerative disorders is based on the altered proteins that aggregate in the brain. The most common altered proteins found in age-related neurodegenerative proteinopathies are hyperphosphorylated τ (HP τ), β amyloid ($A\beta$), α synuclein (αS), and trans active response DNA binding protein 43 (TDP43)[1,2,3,4,5,6,7,8]. The most common proteinopathies are listed in Table 1.

Tauopathies

Tauopathy is a term used to describe a group of neurodegenerative disorders, characterized by the aggregation of altered τ protein. In 1986, aggregated microtubule associated protein τ was visualized in the brain of subjects with Alzheimer's disease (AD) [9, 10]. The classification of tauopathies is based on the affected cell type (neuron or glia or both), the τ isoform (3R τ , 4R τ), and the neuroanatomical localization and distribution of the pathological deposits [11, 12,13,14].

Tauopathies with either 3R τ or 4R τ

Progressive Supranuclear Palsy (PSP)

PSP is a neurodegenerative disorder of middle and late age. The clinical characteristics of PSP include Parkinsonism, supranuclear gaze palsy, pseudobulbar palsy, frontotemporal dementia, and progressive aphasia [13, 15].

Table 1. Classification of neurodegenerative proteinopathies

Proteinopathies	Protein involved	Disorder
Tauopathies	3R τ	Pick's Disease
	4R τ	Argyrophilic Grain Disease Corticobasal Degeneration Progressive Supranuclear palsy
	3R τ +4R τ	Alzheimer's Disease Neurofibrillary tangle only dementia
α synucleinopathies	α S	Parkinson's Disease Parkinson's Disease with Dementia Dementia with Lewy bodies Multiple System Atrophy
TDP43 proteinopathies	TDP-43	Amyotrophic lateral sclerosis Frontotemporal lobar degeneration
A β proteinopathies	A β	Alzheimer's Disease
PrP proteinopathies	PrP	Creutzfeldt-Jakob disease
FUS proteinopathies	FUS	Amyotrophic lateral sclerosis Frontotemporal lobar degeneration
SOD-1 proteinopathies	SOD-1	Amyotrophic lateral sclerosis

HP τ , Hyperphosphorylated τ ; A β , β - amyloid; α S, α - synuclein; TDP43, transactive response DNA binding protein 43 kDa; FUS, fused in sarcoma; SOD-1, superoxide dismutase-1

PSP is a sporadic disease; however, familial cases with PSP like phenotype and MAPT gene mutations have been reported [16]. Neuropathologically, PSP is characterized by atrophy of the basal ganglia, subthalamic nucleus, and the brain stem. The microscopic characteristics include glial alterations such as tufted astrocytes and oligodendroglial coiled bodies and also neuronal alterations such as round/globose neurofibrillary tangles (NFTs) and neuropil threads (NTs). All these lesions are IR for HP τ isoform 4R τ . In addition, neuronal loss and gliosis are observed. These lesions are primarily seen in the central brain structures such as striatum, pallidum, subthalamic nucleus, substantia nigra, basis pontis, superior colliculi, and dentate nucleus [17, 18, 19,20].

Cortico-Basal Degeneration (CBD)

CBD is a rare neurological disorder affecting the aged. The clinical characteristics of CBD include Parkinsonism, cortical sensory loss, alien limb phenomenon, frontal lobe behavioral changes, dementia, and progressive aphasia [13, 21]. Most of the reported CBD cases are sporadic; however, cases

with MAPT gene mutations and CBD like phenotype have been reported [22, 23]. Neuropathologically, CBD is characterized by asymmetric frontoparietal atrophy that is most severe in the pre- and post-central regions, in addition to the depigmentation of the substantia nigra. The microscopic characteristics include glial alterations such as astrocytic plaques, neuropil threads, and oligodendroglial coiled bodies that are IR for HP τ , isoform 4R τ . In addition, gliosis, neuronal loss, spongiosis, and ballooned cells are seen [14, 24, 25, 26].

Argyrophilic Grain Disease (AGD)

AGD is a late onset sporadic disorder clinically characterized by progressive cognitive decline and personality changes [27]. Neuropathologically, AGD is characterized by argyrophilic grains that are IR for HP τ isoform 4R τ . These lesions are seen in both the cortical and the subcortical structures, being most frequent in the entorhinal and transentorhinal regions. These lesions are frequently seen concomitant with AD related pathology [11, 12, 27].

Pick's Disease (PiD)

PiD is a rare disorder clinically characterized by behavioral frontotemporal dementia, and progressive aphasia; the motor symptoms are less common [28, 29]. Mutations in the MAPT gene have been suggested to be associated with this disorder [30]. Neuropathologically, PiD is characterized by frontotemporal atrophy and neuronal pick bodies that are IR for HP τ , isoform 3R τ . In addition, neuronal loss, spongiosis, gliosis, and ballooned cortical neurons are seen. HP τ /IR Pick bodies are most abundant in the temporal cortex (neocortical layers II and VI) and in the hippocampus (granule cells of the dentate gyrus) [14, 31, 32, 33, 34,35].

Tauopathies with both 3R τ and 4R τ

There are three defined disorders that display both 4R τ and 3R τ : Primary Age Related Tauopathy (PART), Neurofibrillary tangle only dementia, and AD.

Progression and distribution of NFTs and NTs

In these tauopathies (PART, NFT only dementia, and AD), the neuroanatomical distribution and progression of the NFTs and NTs follows the pattern that has been described by Braak and Braak in 1991[36]. The Braak staging is based on the topographical distribution of the silver stained NFTs and NTs. Six stages were identified in 1991, i.e., the transentorhinal and entorhinal (stages I and II), limbic (III and IV), and finally, the isocortical stages (stages V and VI) that are sequentially involved. In 2011, Braak and colleagues updated their original work, stating that the locus coeruleus (LC) is probably the initiation site for the HP τ pathology rather than the entorhinal

cortex and most importantly that this LC alteration is seen in young subjects [37, 38]. In 2008, the Brain Net Europe (BNE) consortium reported that a consensus was reached in an inter-laboratory setting, including up to 30 neuropathologists in more than 15 centers, while applying IHC and the described staging criteria for Braak stages I to VI [1].

Primary Age Related Tauopathy (PART)

In 2014, a new entity with 4 and 3 R τ was defined, i.e., PART. This entity incorporates the cognitively unimpaired aged subjects and subjects with mild cognitive impairment that display NFTs and NTs in the hippocampus and in the medial temporal lobe, displaying AD like distribution with a progression less or equal to stage IV. The neuropathological criteria also require that no or minimal A β pathology is observed [39].

Neurofibrillary tangle only dementia

This is a late onset dementia characterized neuropathologically by NFTs that follow the Braak staging [36], usually in stage IV and above, with the absence or scarcity of A β aggregates [14, 40].

Alzheimer's disease

AD is the most common cause of dementia. Based on the Diagnostic and Statistical Manual of Mental Disorders (DSM-IV), and the National Institute of Neurological Disorders and Stroke-Alzheimer's Disease and related Disorders (NINCDS-ADRDA) diagnostic criteria[41,42], the clinical diagnosis of AD requires that two or more cognitive domains being affected, including memory impairment and at least one cognitive or behavioral deficits, which include visuospatial function, abstract reasoning, executive function, mood, personality, and language abnormalities.

Neuropathologically, AD is characterized by identification of two hallmark altered proteins, i.e., HPr and A β [1, 2, 43].

Progression and distribution of the A β pathology in AD

Already in 1991, Braak and Braak defined three stages ranging from A to C [36]. Thereafter, in 2002[2], Thal and colleagues, implementing the IHC technique, proposed a five stage assessment of A β /IR: stage 1 where IR is seen in the neocortex; stage 2 in the allocortex; stage 3 in the diencephalic nuclei, the striatum, and the cholinergic nuclei of the basal forebrain; stage 4 in the subcortical nuclei, and stage 5 in the cerebellum. In 2009, the BNE reported a high agreement rate while assessing the phases of A β as proposed by Thal and colleagues [44].

Cerebral Amyloid Angiopathy (CAA)

A β can also be seen in the vessel walls of the leptomenigeal and cortical vessels; moreover, based on the type of the vessel involved, two types of

CAA are defined. In type II, the arteries, arterioles, veins, and venules are affected, whereas in Type I in addition, deposition of A β in the capillaries is observed [45]. CAA can be associated with cerebral hemorrhages [46].

Consensus regarding the neuropathological assessment of AD

The first consensus report regarding neuropathological assessment of AD was proposed by Khachaturian in 1985 as a result of a joint workshop supported by four organizations (National Institute of Aging, the American Association of Retired Persons, the National Institute of Neurology and Communicative Disorders and Stroke, and the National Institute of Mental Health). The strategy proposed was based on the counts of silver stained Neuritic Plaques (NPs) in relation to the age of the patient [47]. Later in 1991, the Consortium to Establish Registry of Alzheimer's Disease (CERAD) [48] proposed a strategy defining more in detail the brain regions to be assessed and the silver stains to be used. The criteria were based on the semi-quantitative scoring of the NPs, where the NP score is adjusted with the patient's age at death to obtain an age related NP score. This score was then related to the clinical presentation (demented/non demented) to obtain a level (probable, possible, and definite) of certainty that the clinically observed dementia was caused by the AD related pathology. The major drawback with this criterion was that the NFT pathology was not taken into account [49].

In 1991, Braak and Braak also described, by applying the silver stain, the progression of the NFT pathology from stage I to VI. In 1997, the National Institute of Aging and the Regan Institute (NIA-RI) [50] launched their recommendations, where the CERAD NP count was combined with the Braak NFT stage. Thus, by applying the silver stains, three stages were defined: stage 1 meaning a high likelihood that the dementia is caused by the AD pathology (Braak V–VI and CERAD severe), stage 2 being intermediate likelihood (Braak stages III–IV and CERAD moderate), and stage 3 low likelihood (Braak stages I–II and CERAD mild). In 2012, the NIA-RI criteria were updated to the NI-AA criteria [51, 52], implementing the modern IHC techniques. An “ABC” staging protocol was proposed, incorporating the CERAD NP score, Braak NFT (HP τ) stage, and Thal A β Phase. It was also recommended to assess comorbidities. An important point was raised, namely, that the neuropathological assessment, i.e., the staging of the observed alterations should be carried out independent of the clinical symptomatology. Thus, a stage of AD related pathology could be given both for cognitively unimpaired as well as demented subjects.

Over the last two decades, progress has been made in identifying the AD associated changes prior to the neuropathological assessment by using biomarkers such as cerebrospinal fluid (CSF) levels of A β 42, HP τ , and Total τ [53,54,55,57]. In AD patients, the CSF A β 42 concentrations decrease by about 50%; this decrease is associated with the deposition of A β in the brain. The CSF total τ increases on average by two to three fold in AD, as well as

HP τ [53, 54]. Pathological values of two or more biomarkers are considered to reliably predict mild cognitive impairment conversion to AD [55, 56, 57].

α synucleinopathies

α S is the main component of Lewy Body (LB), the hallmark lesion of Parkinson's Disease (PD), Parkinson's Disease with Dementia (PDD), Dementia with Lewy Bodies (DLB)[58,59,60,61].

PD, PDD, and DLB

PD is clinically characterized by motor signs such as bradykinesia, rigidity, resting tremors, gait instability, and non-motor signs such as hallucinations, olfactory disturbances, and sleep disturbances. If the subject develops dementia over time, the clinical diagnosis given would be PDD [62]. Contrary to PDD, DLB dementia is clinically characterized by initial signs of progressive cognitive decline. Other features of DLB include parkinsonism, and visual hallucinations [4].

The neuropathological characteristics of PD, PDD, and DLB include loss of pigmented neurons in the substantia nigra, motor nucleus of vagus, and the LC, with widespread LBs and Lewy Neurites (LN) [3, 4, 5, 63,64,65] . LBs and LNs are visualized by implementing the IHC technique and antibodies directed to the α S.

Progression and distribution of α S Pathology

Already in 1984, Kosaka and colleagues suggested that three stages of DLB pathology are identified: brain stem, limbic, or neocortical [66]. In 1996, the existence of these three stages, namely, brain stem, limbic, and neocortical was confirmed by the Consortium on DLB International Workshop [67].

In 2003, Braak and colleagues proposed a more detailed classification scheme leading to 6 stages: stage 1- involvement of the dorsal motor nucleus of the vagus, stage 2- lower raphe nucleus and LC, stage 3- Substantia nigra, stage 4- Amygdala and basal nucleus of Meynert and hippocampal CA2 sector, stage 5- Cingulate cortex and finally, stage 6- Temporal, frontal, and Parietal cortex [3]. In 2005, McKeith and colleagues launched the current, commonly used, consensus guidelines incorporating the semi quantitative score and the anatomical distribution of the LBs, while applying the IHC technique and antibodies directed to the α S [4].

McKeith's and Braak's staging systems have been applied in several studies [68, 69, 70]. In 2009, a staging protocol for the α S pathology was proposed by the BNE [5]. The protocol is a modification of the original Braak and McKeith staging systems. Instead of semi quantitative scoring of the α S pathology, a dichotomized approach was recommended. This approach was chosen due to the significant variations observed by the BNE in the staining quality and outcome of the semi quantitative counts in the inter laboratory

setting [5]. When the protocol was applied based on a dichotomized assessment, the inter observer agreement was more than 80%.

Multiple System Atrophy

MSA is a rare sporadic disease clinically characterized by autonomic failure, cerebellar ataxia, or parkinsonism not responsive to Levodopa [71]. The macroscopic features seen in MSA are various, including greyish discoloration of the putamen, atrophy of the cerebellum and pons. The microscopic hallmark lesion of MSA is the α S IR glial cytoplasmic inclusions in the oligodendrocytes [72, 73,74].

Transactive DNA binding protein 43 related proteinopathies

TDP43 is a hallmark alteration in the Amyotrophic lateral sclerosis (ALS) and Frontotemporal lobar degeneration (FTLD-TDP) [6, 75, 76]. This protein is also seen in a subset of AD patients [8, 77, 78, 79, 80,81].

Amyotrophic Lateral sclerosis

ALS is a disorder characterized by degeneration of motor neurons of the motor cortex, brain stem, and the spinal cord. The clinical symptoms of ALS include progressive muscle weakness in upper and lower limbs and progressive bulbar palsy, the involvement of respiratory muscles results in respiratory failure and death [82, 83]. In addition, ALS may present with behavioral and cognitive symptoms. Several gene mutations have been reported to contribute to the pathogenesis of ALS including SOD1, TARDBP, FUS, and C9orf72 mutations [84, 85, 86].

The currently defined neuropathological phenotypes are: ALS-TDP, ALS-FUS, and ALS-SOD1. ALS-TDP cases are mainly sporadic cases with more bulbar involvement, rapid disease progression, and earlier age of onset. TDP43 is a component of the neuronal inclusions and oligodendroglial aggregates seen in ALS-TDP [75].

FTLD-TDP

FTLD-TDP clinical characteristics include behavioral and cognitive symptoms as well as progressive aphasia. Several genes have been reported to contribute to the pathogenesis of FTLD-TDP including TARDBP, VCP, GRN, and C9orf72 mutations [87, 88, 89, 90, 91]. Macroscopically, FTLD-TDP is characterized by frontotemporal lobar atrophy. Microscopically, it is characterized by neuronal loss, gliosis, and TDP43/IR neuronal inclusions (neuronal cytoplasmic inclusions, neuronal intranuclear inclusions, and dystrophic neurites), in addition to glial cytoplasmic inclusions that are mainly oligodendroglial [92].

Progression and distribution of TDP43 pathology

Progression and distribution of TDP43 in ALS

Four stages of progression of TDP43 pathology in ALS have been described [7]. In stage I, TDP43 inclusions are detected in the motor cortex, medulla oblongata, and the spinal cord; in stage II, further spread in the prefrontal neocortex, brain stem reticular formation, and the red nucleus; in stage III, lesions are seen in the precentral and postcentral neocortex and the striatum; and in stage IV, further involvement of the anteromedial portions of the temporal lobe and hippocampus [7].

Progression and distribution of TDP43 in behavioral variant (bv) FTLTDP

Four stages of progression TDP43/IR lesions in bvFTLD were described. Stage I is characterized by the involvement of the orbital gyri, gyrus rectus, and the amygdala. In stage II, there is sequential involvement of the cingulate gyrus, temporal lobe and the striatum, red nucleus, thalamus, and the pre-cerebellar nuclei. Stage III is characterized by the sequential involvement of the motor cortex and the anterior horn of the spinal cord. In stage IV, the sequential involvement of the visual cortex is observed [93].

Progression and distribution of TDP43 in AD

Recently, Josephs and colleagues described five stages of progression of TDP43 pathology in AD. In stage I, TDP43 inclusions are detected in the amygdala; in stage II, further spread in the entorhinal cortex and subiculum; in stage III, the dentate gyrus and occipitotemporal cortex; in stage IV, lesions are also seen in the inferior temporal cortex; and in stage V, a further involvement of the frontal cortex is seen [8].

Other Proteinopathies

Other proteins known to be associated with the pathogenesis of neurodegenerative disorders include ; Fused in Sacroma (FUS) in FTLTDP and ALS, Superoxidase dismutase-1 (SOD1) in ALS, Huntingtin in Huntington's disease ,PrP in Creutzfeldt-Jakob disease[94,95,96,97,98].

Concomitant proteinopathies in dementia

The four altered proteins HP τ , A β , α S, and TDP43 are frequently seen concomitantly in demented subjects [99,100,101,102,103,104,105,106,107].

Subjects with AD related lesions display concomitant α S and TDP43IR. α S pathology was reported to be seen in 60% of the familial and sporadic AD; α S/IR is predominantly seen in the amygdala [108,109]. Moreover, TDP43 pathology is seen in up to 57% of the AD patients [8, 77, 78, 79, 80, 81].

Several previous studies have demonstrated a wide spread of A β pathology in the striatum of the PDD and DLB patients [110,111,112]. In addition, the TDP43/IR lesions were reported to be seen in 31% of the DLB cases, in 7% of the PD, and in 19% of the PDD patients [113]. Furthermore, the accumulation of the abnormal TDP43 aggregates was previously demonstrated in the hippocampal sclerosis, Huntington's disease, and PiD [114,115,116].

Neurodegenerative alterations in cognitively unimpaired subjects

HP τ in cognitively unimpaired subjects

Several previous post mortem reports have demonstrated HP τ /IR NFTs in the brain tissue of cognitively unimpaired subjects [117, 118, 119, 120, 121, 122]. Interestingly, Boyle and colleagues reported that HP τ /IR lesions were seen in 100% of a large cohort of cognitively unimpaired subjects [122]. Furthermore, post mortem reports have also demonstrated the existence of AGD and PSP related pathological alterations in clinically normal subjects [123, 124, 125]. Noteworthy, most subjects that are neuropathologically classified as PART are cognitively unimpaired or display minor cognitive changes [39].

A β in cognitively unimpaired subjects

Many previous studies have demonstrated the existence of the A β aggregates in the brain tissue of cognitively unimpaired subjects [118, 120, 121, 122, 126, 127]. Interestingly, A β pathology was reported to be seen in up to 82% of a large cohort of cognitively unimpaired subjects [122].

α S in cognitively unimpaired subjects

Several post mortem studies have documented the incidence of α S pathology in the cortical and subcortical regions in cognitively unimpaired subjects. Incidental LB related pathology (iLBRP) is seen in 8.3 to 31% of cognitively unimpaired subjects [128,129,130,131].

The clinical importance of iLBRP in cognitively unimpaired subjects was discussed in several previous studies [132,133,134,135]. It was suggested that cognitively unimpaired subjects with iLBRP might represent preclinical PD cases [132,133]. Interestingly, Delle Donne and colleagues [136] demonstrated nigral cell loss in subjects displaying iLBRP, a finding which supports that iLBRP might represent preclinical PD. In line with this, previous post mortem reports demonstrated an incidence of iLBRP in subjects dis-

playing non motor clinical features of PD such as olfactory dysfunction [137] and rapid eye movement sleep disorder [138,139].

TDP43 in cognitively unimpaired subjects

The incidence of TDP43 pathology in the brain of cognitively unimpaired subjects was previously reported [113,124,140]. TDP43/IR lesions were reported to be observed in the amygdala and/or the hippocampus in 36% of 110 cognitively unimpaired subjects [140]. Interestingly, Kovacs and colleagues studied 51 cognitively unimpaired subjects, and TDP43 pathology was reported to be seen in 7% of the subjects [124]. A lower incidence of TDP43 pathology in cognitively unimpaired subjects (3%) has also been reported [113].

Concomitant proteinopathies in cognitively unimpaired subjects

The existence of mixed pathologies in the brain tissue of aged non demented subjects have been demonstrated in a number of post mortem reports [121,122,123,124,140,141]; however, these reports are fewer when compared with the number of studies investigating mixed pathologies in demented subjects [99,100,101,102,103,104,105,106,107].

Several reports have shown concomitant AD related hallmark lesions (HP τ and A β) in cognitively unimpaired subjects [117, 118, 119, 120, 121, 122]. Furthermore, studies have demonstrated concomitant HP τ , A β , and α S pathology in non-demented subjects [121,122,124]. Some studies have reported concomitant HP τ , A β , and TDP43 pathology [124] and HP τ and TDP43 or α S and TDP43 [140] in the brain tissue of cognitively unimpaired subjects.

Normal Pressure Hydrocephalus

Idiopathic normal pressure hydrocephalus (iNPH) is a disorder characterized by cognitive impairment, gait disturbance, and urinary incontinence [142]. The cognitive impairment in iNPH is potentially reversible by ventriculo-peritoneal shunt operation; however, shunt irresponsiveness can be attributed to the existence of comorbid neurodegenerative pathology. AD related pathology was reported to be seen in the cortical biopsies obtained from patients with iNPH [143,144,145,146].

Methodological considerations

The reported incidences of neurodegenerative proteinopathies in cognitively unimpaired subjects are quite various. This variation is probably due to issues such as various number of study subjects included, selection bias, age range of the study subjects, and the level of clinical assessment carried out. Furthermore, the methods used while assessing the pathology are of major significance; The post mortem delay, fixation time, selected antibody, pre-treatment used, and detection system implemented have all been reported to significantly influence the outcome [147,148,149,150,151,152].

The Present Investigation

Aims of the study

General Aim

To assess the incidence and distribution of the altered proteins common in age related neurodegenerative disorders in a well characterized post mortem cohort of cognitively unimpaired subjects.

The specific aims of the study are to investigate the following:

- The initiation site of HP τ pathology in cognitively unimpaired middle aged subjects (Study I).
- The extent of AD related pathology seen in aged cognitively unimpaired subjects and the applicability of the recent National Institute on Aging – Alzheimer’s Association (NIA-AA) criteria (Study II).
- The incidence of A β and HP τ pathology in cortical biopsies obtained from iNPH patients and to study whether the clinical parameters and available biomarkers reflect the brain pathology (Study III).
- The incidence and the extent of the most common altered proteins seen in the aging brain (HP τ , A β , α S, and TDP43) in a well characterized, large, cohort of cognitively unimpaired aged subjects (Study IV).

Subjects and Methods

Subjects

The brain tissue assessed in this study was either obtained post mortem (study I, II, and IV) or during a surgical procedure (study III). General demographics and selection criteria of the study subjects are given in Table 2 and 3, respectively. Ethical permission was obtained from the ethical committee at Kuopio University Hospital and from the regional ethical committee in Uppsala.

Table 2. General description of the study subjects

Study	Center	Year	Samples	N	Age range	Gender (M/F)
Study I	1	1992-2008	3	95	22-50	55/40
Study II	1	1992-2000	3	192	55-98	87/105
Study III	2	2010-2013	4	111	54-89	67/44
Study IV	2	2009-2014	3	296	50-102	185/111

Pathology department at 1, Kuopio University Hospital, 2, Uppsala University Hospital; F, Female; M, Male; 3, postmortem brain material; 4, surgical samples; N, number of subjects

Clinical assessment

The clinical data were obtained retrospectively from the hospital records blinded to the neuropathological findings (study I, II, and IV). Information regarding the cognitive status was also obtained retrospectively, but there was no neuropsychological tests carried out. Some of the subjects might thus have displayed mild cognitive impairment, but the presence of dementia was not registered in the medical records (study I, II, and IV).

iNPH cases were pre and post operatively assessed by a multidisciplinary iNPH team including a neurologist, a neurosurgeon, a trained physiotherapist, and an occupational therapist. The cognitive status was assessed by applying Mini Mental State Examination (MMSE). Subjects with a MMSE score ≥ 24 were considered as being cognitively unimpaired, and subjects with a MMSE score ≤ 23 were considered as being demented (study III).

Table 3. Selection criteria of included subjects

Study	Inclusion criteria
Study I	Age at death \leq 50 years, cognitively unimpaired subjects, no concomitant brain disease (infections, primary or secondary brain tumor).
Study II	Age at death $>$ 50 years, cognitively unimpaired subjects, presence of neuritic plaques in modified Bielschowsky staining in the neocortex.
Study III	Clinical diagnosis of idiopathic normal pressure hydrocephalus, availability of cortical brain biopsy and lab and clinical data.
Study IV	Age at death \geq 50 years, cognitively unimpaired subjects.

Neuropathological assessment

The brains were weighed, evaluated for grossly detectable lesions and vessel abnormalities, and immersed in 10% buffered formalin for at least one week. Thereafter, the brains were sampled in a standardized manner (Table 4) (Study I, II, IV).

In study III, a frontal cortical biopsy was obtained during the surgical operation and placed in 10% buffered formalin and then embedded in paraffin.

Immunohistochemistry

For the IHC, seven μ m thick sections were used. The details regarding the antibodies used are given in Table 5. For detection, a poly HRP-IHC detection kit with Romulin-3-amino-9-ethylcarbazol (AEC) chromogen was used (study I, IV). The streptavidin-biotin complex was visualized using vector red (vector red alkaline phosphatase substrate kit I, Zymed; Cat. No.SK-5100) for A β and DAB (Liquid DAB substrate Kit, Zymed; Cat. No.00-2014) for ubiquitin (Ubq), and HP τ (study II). For detection, the Dako EnVision FLEX detection system was used (study III and IV). The details regarding the neuroanatomical regions assessed applying IHC are given in Table 6.

Staging of the pathological lesions

Modified Bielschowsky (mBky) silver impregnation method was used for the assessment the score of NPs (study II,IV). A CERAD NP score and the level of AD neuropathological changes were given as recommended by the NIA-RI [50] (study II) and NIA-AA criteria [51, 52] (study II and IV). HP τ Braak stage [1] (study I, II, and IV), A β Thal phase [2] (study II and IV), α S stage [3,4,5] (study IV), and TDP43 stage [8] (study IV) were assessed as recommended.

Table 4. Brain regions sampled and assessed

Brain region	HE	Immunohistochemistry			
		Study I	Study II	Study III	Study IV
Frontal Cortex, gyrus medius	X*			X	X
Temporal Cortex, gyrus medius	X*	X	X		X
Gyrus cinguli, anterior	X				X
Parietal Cortex, inferior	X*	X	X		X
Pre-post central cortex	X				
Occipital Cortex	X*	X	X		X
Hippocampus anterior	X	X	X		X
Hippocampus posterior	X	X	X		X
Basal forebrain, incl. amygdala	X	X	X		X
Striatum	X				
Thalamus	X				
Mesencephalon incl. Substantia Nigra	X		X		X
Pons, incl locus coeruleus	X	X			X
Medulla, incl nucleus vagus	X				X
Vermis	X				
Cerebellum	X		X		X

* modified Bielshowsky silver stain

Table 5. Immunohistochemical stains

Antibody	Clone	Source	Dilution	Pre-treatment	Study
HP τ	AT8	Thermo Scientific	1:500	-	I,II,III,IV
A β	6F/3D	Dako	1:100	80 % Formic Acid, 6 hours	I,II,III,IV
Ubq	-	Dako	1:200	Citrate Buffer* Citrate Buffer* + 80 % Formic Acid,	II
α S	KM51	NovoCastra	1:100	5min	IV
TDP43	11-9	Cosmo Bio	1:5000	Citrate Buffer*	IV

HP τ , hyperphosphorylated τ ; A β , β -amyloid; Ubq, Ubiquitin; α S, α -synuclein; TDP43, trans- active response DNA binding protein 43 kDa; *Autoclave

Table 6. Neuroanatomical regions assessed applying immunohistochemistry

Brain region	Immunohistochemical staining			
	HP τ	A β	α S	TDP43
Frontal Cortex, gyrus medius				X
Temporal Cortex, gyrus medius	X		X	X
Gyrus cinguli, anterior			X	
Parietal Cortex, inferior		X	X	
Pre-post central cortex				
Occipital Cortex	X			
Hippocampus anterior	X			X
Hippocampus posterior	X	X	X	X
Basal forebrain, incl. amygdala		X	X	X
Striatum				
Thalamus				
Mesencephalon incl. Substantia Nigra		X	X	
Pons, incl locus coeruleus	X			
Medulla, incl nucleus vagus			X	X
Vermis				
Cerebellum		X		

HP τ , hyperphosphorylated τ ; A β , β -amyloid; α S, α -synuclein; TDP43, transactive response DNA binding protein 43 kDa

Semi quantitative analysis of the pathological alterations

Semi quantitative assessments of NPs in the HP τ , Ubq, and the mBky stain were carried out for study II. For each stain, the section was first scanned at magnification x40 to select the grey matter area with the most severe involvement. Then, in each sample within the selected area, three microscopic fields were randomly chosen at 100 magnifications. The number of lesions was counted within each field and scored as 0- none, 1- 1 to 5 lesions, 2- 6 to 20 lesions, and 3- \geq 21 lesions (Study II).

In study III, semi quantitative assessment of A β and HP τ /IR lesions was performed. Results were reported semi quantitatively in three levels: absent, sparse or extensive/IR.

Digital analysis

The digital quantification of the A β /IR and HP τ /IR load was carried out as follows; all slides with IR were scanned using the Aperio slide scanner. The Aperio image analysis positive pixel count (PCC) version 9.1 was used. The PCC was applied in the grey matter region. The percentage of grey matter

area covered with the A β /IR and HP τ /IR was calculated as the stained area fraction (study III).

CSF analysis

According to the protocol from the manufacturer and using a commercial ELIZA kit, the levels of A β 42 and HP τ and the total τ in the CSF were measured (Study III).

Statistical analysis

IBM SPSS statistics was used. The correlation between the studied variables was assessed using Spearman correlation (study I, II, III, and IV). For assessment of the statistical difference between the studied groups, Mann-Whitney-U test and Kruskal-Wallis H test were applied (study III, IV). Logistic regression analysis was used to assess the relation between the variables (study III). Receiver operating characteristic analysis was used to determine the HP τ /A β 42 cut off for identifying subjects with the IR lesions (study III).

RESULTS

HP τ in the cognitively unimpaired subjects

Thirty-three percent of the 95 cognitively unimpaired subjects with age at death ranging from 22 to 50 years displayed HP τ /IR lesions in the cortical and the subcortical structures (study I). HP τ pathology was seen in the LC in 28 out of the 95 cognitively unimpaired subjects. Three out of these 28 subjects displayed concomitant HP τ pathology in the hippocampus (study I).

HP τ /IR lesions were visualized in the cortical and the subcortical structures in 98% of the 296 subjects with age at death ranging from 50 to 102 years (study IV). HP τ pathology was seen in the LC in 95% of the total cohort (study IV). The incidence of HP τ /IR lesions increased with age (study I, II, and IV).

PART

Fifty two percent of the subjects fulfilled the criteria for definite PART (study IV). These subjects lacked A β aggregates and displayed HP τ pathology with the Braak stages ranging from (a-IV).

HP τ in the cortical biopsies

HP τ pathology was observed in 25% of the 111 cortical biopsies obtained from the iNPH patients (study III) (Figure 1, 2). Lower preoperative MMSE scores corresponded with higher stained area fraction of HP τ in the cortical biopsies. HP τ in the CSF correlated significantly with the HP τ stained area fraction in the biopsies.

HP τ /IR NPs in cognitively unimpaired subjects

One hundred and ninety-two cognitively unimpaired subjects were included in study II. Sixty two percent of these subjects displayed HP τ /IR NPs in the temporal cortex.

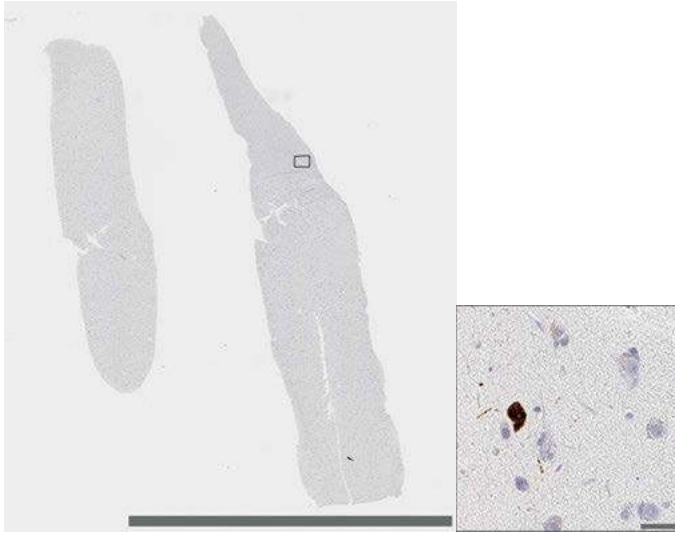


Figure 1. Sparse hyperphosphorylated τ immunoreactive lesions seen in a frontal cortical biopsy obtained from a subject with normal pressure hydrocephalus diagnosis (scale bar =6mm); inset magnification x20 (scale bar=100 μ m), note the solitary tangle with few surrounding neurites.

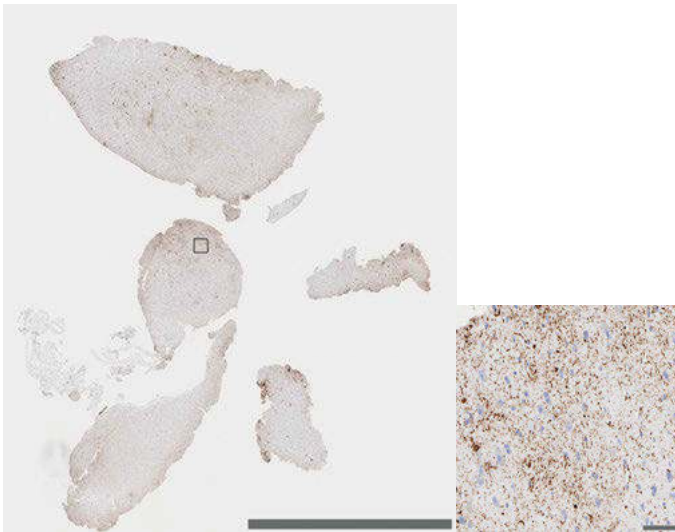


Figure 2. Extensive hyperphosphorylated τ immunoreactive lesions seen in a frontal cortical biopsy obtained from a subject with normal pressure hydrocephalus diagnosis (scale bar =4mm); inset magnification x20 (scale bar=100 μ m), note the numerous tangles and neurites

A β in the cognitively unimpaired subjects

In study I, 7 out of the 95 cognitively unimpaired subjects (7%) displayed A β aggregates in the cortex. In study IV, the cortical A β aggregates were seen in 47% of the total cohort. CAA was observed in the parietal cortex in 15% of the subjects. Six percent displayed type I, and 9% displayed type II CAA (study IV).

A β in the cortical biopsies

A β aggregates were observed in 44% of the assessed 111 biopsies obtained from the iNPH patients (study III) (Figure 3, 4). Lower preoperative MMSE scores corresponded with higher stained area fraction of A β in the cortical biopsies. Moreover, A β 42 in the CSF correlated significantly with the A β stained area fraction in the cortical biopsies.

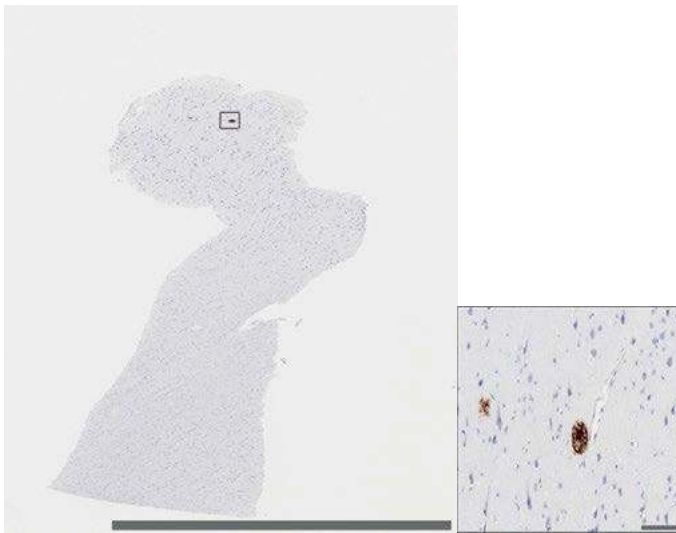


Figure 3. Sparse β -amyloid Immunoreactive lesions seen in a frontal cortical biopsy obtained from a subject with normal pressure hydrocephalus diagnosis (scale bar =3mm); inset magnification x20 (scale bar=100 μ m), note the solitary dense plaque

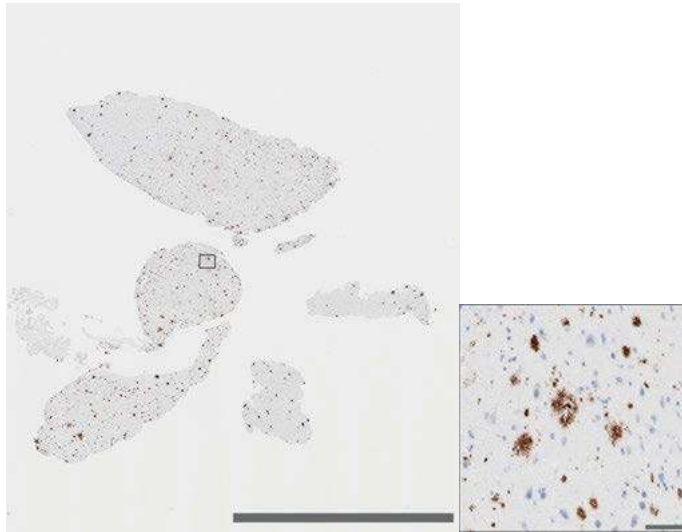


Figure 4. Extensive β -amyloid Immunoreactive lesions seen in a frontal cortical biopsy obtained from a subject with normal pressure hydrocephalus diagnosis (scale bar =4mm); inset magnification x20 (scale bar=100 μ m), note the numerous β -amyloid immunoreactive lesions

α S in the cognitively unimpaired subjects

α S/IR lesions were visualized in 19% of the 296 cognitively unimpaired subjects in study IV.

TDP43 in the cognitively unimpaired subjects

TDP43/IR was observed in 36% of the total cohort in study IV. The most frequently affected region was the medulla.

Concomitant pathologies in the cognitively unimpaired subjects

Thirty-one out of the 95 cognitively unimpaired subjects, with age ranging from 22 to 50 years, displayed HP τ /IR, but concomitant A β /IR was observed only in one subject that was in Braak stage II (study I). Concomitant HP τ and A β /IR were seen in 46% of the 296 cognitively unimpaired subjects, with age ranging from 50–102 (study IV). Concomitant HP τ and TDP43/IR was observed in 35% the cognitively unimpaired subjects and 19% of displayed concomitant HP τ and α S/IR. Out of the 296 subjects, 11% displayed concomitant A β /HP τ and α S/IR, and 15% displayed concomitant A β /HP τ and TDP43/IR. All four altered proteins (HP τ /A β / α S /TDP43) were detected simultaneously in 5% of the total cohort.

Concomitant pathologies in the cortical biopsies

Concomitant A β and HP τ /IR were seen in 22% of the 111 biopsies obtained from the iNPH patients (study III).

Applicability of the neuropathological criteria

Braak stages of HP τ pathology

Thirty one out of the 95 cognitively unimpaired subjects, with age at death ranging from 22–50 years, displayed HP τ /IR lesions in cortical and subcortical structures. Eighty percent of these subjects were in Braak stages a – b; twenty percent of the subjects were in Braak stage I–II (study I).

Seventy three percent of the 192 cognitively unimpaired subjects included in study II, with age at death ranging from 55–98, were in Braak stage I – II, 13% were in Braak stage III- IV, and 1% in Braak stage V.

In study IV, Ninety-five percent of the 296 cognitively unimpaired subjects, with age at death ranging from 50 to 102 years were given a Braak stage. Forty six percent were in Braak stages a – b, 34% were in Braak stage I – II, 14% were in Braak stage III- IV, and 1% in Braak stage V. A significant correlation was noted between the Braak stage of HP τ pathology and the age at death ($r = 0.34$, $p = 0.001$) (study IV).

Thal Phases of A β pathology

Thirty Four percent of the cognitively unimpaired subjects with A β /IR lesions were in Thal phase 1, 6% were in Thal phase 2-3 and 7% were in Thal phase 4-5 .A significant correlation was noted between the Thal phase and the age at death ($r = 0.32$, $p = 0.001$) (study IV).

BNE stages of α S pathology

In study IV, Most of the subjects with α S/IR lesions were in McKeith mid-brain/limbic stage of α S pathology [4] i.e. (BNE stage 1 to 5) [5]. A significant correlation was noted between the age at death and the α S/BNE stage ($r = 0.2$, $p = 0.001$).

Joseph stages of TDP43 Pathology

Twenty three percent of the subjects with AD related pathology fulfilled the criteria for Joseph stages of TDP43 pathology from stage 1 to 5. A significant correlation was noted between the age at death and the TDP43 Joseph stage ($r = 0.3$, $p = 0.001$) (study IV).

NIAA criteria

Fifteen percent of the cognitively unimpaired subjects were classified as having an intermediate level of AD pathology based on NIAA criteria, and 85% were classified as having a low level of AD pathology. Ninety four

subjects displayed HP τ /IR NPs in the temporal cortex but were classified as having a low level of AD related pathology based on NIAA criteria (study II). In study IV, 12% of the subjects were classified as having an intermediate level of AD related pathology based on the NIA-AA criteria and only one case was classified as having a high level of AD pathology.

DISCUSSION

Our results indicate that the altered proteins (HP τ , A β , α S, and TDP43) are frequently seen in the brain tissue of cognitively unimpaired aged subjects, and the incidence increases with age. This suggests that even if neurodegeneration is common, it has to be extensive enough to cause a functional disturbance. Assessment of only one type of pathology is certainly not enough, as a substantial number of subjects seem to display mixed pathologies.

HP τ in the cognitively unimpaired subjects

τ Protein is a microtubule associated protein which has a fundamental role in axoplasmic transport. HP τ is the hallmark lesion of tauopathies [11, 12, 13,14]; however, HP τ /IR lesions can also be seen in aged non-demented subjects[117,118,119,120,121,122]. The transentorhinal cortex was presumed to be the initiation site for the HP τ pathology. The HP τ alteration is presumed to progress in an orderly manner as was described by Braak and Braak in 1991[36], reaching the occipital cortex in the end stage. In stages V and VI, the occipital cortex is involved and this stage is usually seen in subjects with symptoms of dementia, whereas stages I to III tend to be associated with the cognitively unimpaired subjects. Braak stage IV has been reported to be associated both with demented and with cognitively unimpaired subjects.

We observed HP τ /IR solely in LC in 28% of the cognitively unimpaired subjects lacking HP τ /IR in transentorhinal/entorhinal cortex (study I). This observation supports the notion that the LC might be the initiation site for the HP τ pathology, a hypothesis proposed by Braak and colleagues in 2011[37, 38]. There are a few studies that have shed light on this issue relating to the initiation site in the LC, as most neuropathologists assess the hippocampal formation for the HP τ pathology. One study opposing the LC as being the initiation site looked at the extent of the pathology in the LC in relation to HP τ Braak stages [153,154]. The observation of the LC being involved early on suggests that the clinical assessment strategies and the biomarkers detecting the LC malfunction should be investigated. This is particularly of importance, as any therapeutic intervention should be initiated before the development of significant cognitive impairment.

HP τ /IR lesions were seen in the cortical and the subcortical structures in 33% of the cognitively unimpaired subjects with age at death ranging from 22–50 years (study I) and in 98% of the subjects with age at death ranging from 50 to 102 years (study IV). The methods used in both studies are similar as well as the brain regions assessed. The difference in the obtained results is explained by the difference in the age at death of the included subjects in studies I and IV and emphasizes the significant influence of age on the incidence of HP τ /IR. The results are in line with previous reports show-

ing that the HP τ /IR was seen in 30% of cognitively unimpaired subjects with mean age at death of 53 ± 2 years [121] and in 99% of the aged subjects [37]. This observation highlights the significance of age on the incidence of the HP τ pathology in the aging brain. In line with this, in 2013, Boyle and colleagues reported an incidence of 100% HP τ pathology while studying cognitively unimpaired subjects with mean age at death of 87 ± 7 [122].

A major factor influencing the incidence of HP τ pathology is the selection bias. In study I, all subjects with trauma or brain tumors or pharmaceutical treatments that are reported to be associated with tau hyperphosphorylation were excluded [155,156,157,158]. This probably explains the significant difference in the incidence of HP τ pathology in the LC, reported by us (29%) when compared with (90%) reported by Braak and colleagues [38]. Another aspect that should always be taken into consideration is the section thickness. Braak and colleagues used 100 μ m thick sections; Boyle and colleagues used 20 μ m thick sections when compared to our routine section thickness of 7 μ m (study I, IV).

Fifty two percent of the subjects fulfilled the criteria for definite PART, i.e., displayed HP τ pathology (Braak stage a–IV) but lacked the A β aggregates (study IV). PART was defined in 2014 when the question was raised if all subjects with AD related pathology indeed develop AD, or maybe there are those who do not progress to full blown dementia [39]. Noteworthy, definite PART subjects were significantly younger when compared to subjects with AD related pathology (study IV). Thus, PART subjects might have developed AD related pathology if they had lived longer.

HP τ /IR lesions were seen in 25% of the 111 cortical biopsies obtained from the iNPH patients (study III). This percentage is in line with the percentage of 20% reported by Pyykko and colleagues in their cohort of iNPH patients [159]. Lower preoperative MMSE scores corresponded with higher HP τ stained area fraction in the cortical biopsies (study III). These findings are in line with a previous report [160]. In iNPH patients, HP τ in the CSF correlated significantly with the HP τ stained area fraction in the cortical biopsies, indicating that the biomarkers reflect the pathology seen in the brain (study III).

A β in the cognitively unimpaired subjects

A β protein is a product of the proteolytic cleavage of the Amyloid Precursor Protein (APP) by β & γ –secretase [161]. Based on the post mortem studies [2] A β aggregates are initially observed in the cortex in aged subjects. Sequentially, other brain regions are involved, and five stages of the progression of the A β pathology are defined, i.e., involvement of the neocortex, allocortex, the diencephalic nucleus, basal forebrain, subcortical nuclei, and finally, the cerebellum. Furthermore, the A β deposits in the walls of the vessels in the brain, i.e., CAA. Two types have been described, Type: I in-

involvement of all the vessel types including capillaries and type II: involvement of all the vessel types except the capillaries [45].

A β aggregates were seen in 7% of the 95 cognitively unimpaired subjects with age at death ranging from 22–50 years (study I). In study IV, the incidence of the A β aggregates in the cognitively unimpaired subjects was 47%. The difference in the obtained results in study I vs. study IV is attributed to the difference in the age range of the included subjects, i.e., the age at death of the subject in study IV was ≥ 50 years. Interestingly, Braak and colleagues [37] studied a large clinically unselected cohort with age at death ranging from 0–100 years. They reported that the A β aggregates were seen in 44% of their study subjects; a finding in line with our results (study IV). Noteworthy, the majority of subjects in the study by Braak and colleagues had age at death > 50 years, and quite many of them displayed advanced HP τ Braak stages. However, our study in principle is not fully comparable with the study carried out by Braak and colleagues.

The significance of the methods used in the assessment of the A β pathology is an important factor that should be taken into account. Issues that are of interest are the brain regions assessed, tissue processing, section thickness (range from 3 μ m to 100 μ m), pretreatment -both compound and time, and certainly the antibody used [150,151]. In line with this, while applying 20 μ m thick sections, Boyle and colleagues [122] reported that A β aggregates were seen in 82% of their study subjects, with mean age at death 87 ± 7 . This percentage is certainly higher than 47% observed by us when assessing a cohort with mean age at death of 76 ± 1 year (study IV). Interestingly, however, the incidence of A β was 62% in subjects within the age range of 80–89 and 80% in subjects older than 90 years at death (study IV). Thus, the age of the cohort and not only the thickness of the sections (7 μ m vs. 20 μ m) are of significance and explain the different percentages obtained in these two studies. Furthermore, the pretreatments used for the assessment of the A β /IR are of importance. Already in 1998, Kraszpulski and colleagues reported that a 6 hour formic acid pretreatment significantly increases the extent of the detected A β /IR aggregates in the post mortem brain tissue [151]. The various pretreatments implemented explain the difference in the results obtained by Kovacs and colleagues [124]. Specifically, A β /IR was seen in 39% of the subjects with age at death ranging from 77 to 87 years [124], when compared to our 59% incidence of A β /IR in subjects with the same age range (77 to 87 years) in study IV.

A β /IR was observed in 44% of the cortical biopsies obtained from the 111 iNPH patients (study III), a finding in line with previous reports [146,159]. Interestingly, this observation is close to the 47% A β /IR observed in our post mortem study of subjects with mean age at death of 75 ± 1 year (study IV). Similar percentages have also been previously reported [37]. Noteworthy, the relatively high incidence of A β aggregates in the cortical biopsies obtained from the iNPH patients emphasizes the importance of follow up of the

iNPH patients to identify cases that might develop AD with time. Lower preoperative MMSE scores corresponded with a high A β stained area fraction in the biopsies (study III). These results are in line with a previous report [160]. Thus, assessment of the brain tissue obtained from an iNPH patient has a diagnostic value. These findings also emphasize the importance of long-term follow-up studies of the iNPH patients. Moreover, A β in the CSF correlated significantly with the A β stained area fraction in the cortical biopsies (study III). This finding shows that the CSF biomarkers reflect the brain pathology.

Fifteen percent of the 296 cognitively unimpaired subjects with age at death ranging from 50–102 years displayed CAA. Our results are in line with previous reports, as Kovacs and colleagues [124] reported 25% incidence of CAA within the age range of 77–87 years compared with 28% in our study within the same age group.

α S in the cognitively unimpaired subject

α S is a 140 amino acid protein that is present predominantly in the presynaptic nerve terminals. The protein plays a significant role in dopamine metabolism, proteasome function and has chaperon activity; however, it may form pathological insoluble fibrils [162,163]. α S is the main component in LBs, the hallmark lesion of α -synucleinopathies: PD, PDD, and DLB [58,59,60,61]. α S/IR lesions have been described to be first seen in the brain stem structures, from where it progresses to the limbic structures and finally, the neocortex [3,4]. Interestingly, recent observations indicate that α S is also seen in the peripheral organs, suggesting that this neurodegeneration is a systemic disorder [164].

α S/IR was seen in the brain in 19% of the 296 cognitively unimpaired subjects in study IV. It has been suggested that incidental α S pathology represents preclinical PD or DLB [132,133]. Thus, this group of subjects (19%) might represent preclinical stages of α synucleinopathy. α S/IR has previously been reported to be seen in 8%–31% of the cognitively unimpaired subjects [128,129,130,131]. This variability in the incidence of α S pathology can be attributed to the case selection, brain regions assessed, and most importantly, the methods used for the detection of α S/IR.

TDP43 in the cognitively unimpaired subjects

TDP43 is a protein of 414 amino acids length and is predominantly a nuclear protein. This protein plays a role in the transcriptional repression, RNA metabolism, and gene splicing [165].

TDP43/IR lesions are seen in ALS, FTLN[6,75,76], and in AD patients [8,77,78,79,80,81]. Currently, four stages of progression of TDP43 pathology have been described in ALS, involving initially the medulla and sequen-

tially involving the red nucleus, prefrontal neocortex, pre and post central neocortex and striatum, and finally involving the temporal cortex and hippocampus [7]. Furthermore, four stages of progression TDP43/IR lesions in bvFTLD were described. Stage I is characterized by the involvement of the orbital gyri, gyrus rectus, and the amygdala. In stage II, the involvement of the cingulate gyrus, temporal lobe, and the striatum, red nucleus, thalamus, and the pre-cerebellar nuclei is seen. Stage III is characterized by the sequential involvement of the motor cortex and the anterior horn of the spinal cord. In stage IV, the involvement of the visual cortex is observed [93]. Moreover, five stages of progression of TDP43 pathology were described in AD patients, initially starting in the amygdala and sequentially involving the entorhinal cortex, subiculum, occipitotemporal cortex, inferior temporal cortex, and finally reaching the frontal cortex [8].

There are few studies regarding the incidence of TDP43 pathology in cognitively unimpaired subjects. TDP43/IR was seen in 36% of the 296 cognitively unimpaired subjects with age at death ranging from 50 to 102 when the medulla, amygdala and hippocampal formation were screened. The most frequently affected region was the medulla (study IV). In line with our results, Arnold and colleagues reported an incidence of 34% for TDP43/IR in cognitively unimpaired subjects [140]. Noteworthy, they applied 40 μ m thick sections when compared with our 7 μ m sections, and they assessed the amygdala and the hippocampus, whereas we assessed the amygdala, hippocampus, and the medulla (study IV). Thin neurites that could have been seen in the 40 μ m thick sections can easily be missed in a section of 7 μ m thickness. Thus, this might indicate that the TDP43 pathology is even more common than reported by us and by Arnold and colleagues. Thus, the incidence of TDP43 is influenced by both the brain regions assessed and the methods applied.

Our findings regarding the incidence of TDP43 pathology in cognitively unimpaired subjects are also in line with the findings reported in a recent study conducted in aging research center in Japan by Uchino and colleagues [166], in this study the medulla, amygdala, and the hippocampus were assessed and the most frequently affected region was the hippocampus. Interestingly, in study IV the most frequently affected region was the medulla. This finding may suggest that the neuroanatomical distribution of TDP43/IR lesions might be genetically i.e., racially determined.

Concomitant pathologies in the cognitively unimpaired subjects

Mixed pathologies in the brain tissue in aged cognitively unimpaired subjects have been previously reported [121,122,123,124,140]. The most frequent combination observed is A β + HP τ . This combination is also frequently referred to as the AD related pathology. In the large cognitively unimpaired cohort, A β + HP τ were seen in 46% of the subjects, with age at death

ranging from 50 to 102 years (study IV). Noteworthy, only 1% of the younger subjects with age at death ranging from 22 to 50 years displayed concomitant A β and HP τ pathology (study I). A β + HP τ pathology was seen in 22% of the 111 cortical biopsy specimens (study III). This is a finding in line with what was previously reported by Pyykko and colleagues [159]. Remarkably, a clinical follow-up study of 63 iNPH patients revealed that as many as 16% had displayed HP τ + A β pathology in cortical biopsies, and half of them developed AD during the follow-up time. Thus, the findings warrant a clinical follow-up of subjects with concomitant A β and HP τ pathology.

Nineteen percent of the cognitively unimpaired subjects displayed concomitant HP τ and α S/IR (study IV), in line to what was reported by Kovacs and colleagues [124]. Concomitant HP τ and TDP43/IR was observed in 35% of the cognitively unimpaired subjects (study IV). Arnold and colleagues [140] reported HP τ + TDP43 to be seen in only 15% of their study material. This difference is probably primarily due to the difference in the brain regions assessed by Arnold and colleagues.

Applicability of the neuropathological criteria

Braak stages of HP τ pathology

In study I, 33% of cognitively unimpaired subjects with age at death ranging from 22–50 years displayed HP τ /IR lesions, 80% of these subjects were in Braak stages a – b and 20% were in Braak stage I–II. In study IV, Forty six percent of the cognitively unimpaired subjects with age at death ranging from 50-102 years were in Braak stages a – b, and 34% were in Braak stage I – II. The difference in the obtained results might be explained by the difference in the age at death of the included subjects in study I and IV.

Thal Phases of A β pathology

Most of the cognitively unimpaired subjects displaying A β /IR were in Thal phase 1. As expected, only 7% of the total cohorts were in advanced Thal phases 4 – 5 (study IV).

BNE stages of α S pathology

Most of the subjects with α S/IR lesions were in McKeith midbrain/limbic stage of α S pathology [4] i.e. (BNE stage 1 to 5) [5]. This observation is in line with a previous study on cognitively unimpaired subjects [131].

Joseph stages of TDP43 pathology

Recently, Joseph and colleagues identified five stages of progression of TDP43 pathology in AD cases [8]. However, Joseph staging of TDP43/IR has not been reported for cognitively unimpaired subjects previously. Interestingly, twenty three percent of the cognitively unimpaired subjects with

AD related pathology in study IV were given a Joseph stage of TDP43 pathology from stage 1 to 5.

NIAA Criteria

In study II, fifteen percent of the cognitively unimpaired subjects were classified as having an intermediate level of AD related pathology based on NIAA criteria [51, 52]. In line with this, 12% of the subjects were classified as having an intermediate level of AD related pathology based on the NIA-AA criteria and only one case was classified as having a high level of AD related pathology (study IV).

Methodological Considerations

Clinical assessment

Normal cognitive function is one of the main selection criteria in our studies (I, II, and IV). Noteworthy, detailed neuropsychological tests were not carried out, and the information regarding the cognitive status of the included subjects was obtained retrospectively from the medical records. Thus, some of the included subjects might have displayed a mild degree of cognitive impairment during life; however, none of the study subjects displayed clinical symptoms of dementia based on the medical records. Furthermore, some of the cases with more severe HP τ pathology, i.e., Braak stages ≥ 4 , were hospitalized for some time prior to death; thus, clinical symptoms of dementia should certainly have been reported.

Case Selection

Our selection of cases was as unbiased as possible. In all subjects that were referred for post mortem examination due to various diseases, a neuropathological assessment was also carried out. The cohort includes subjects with risk factors known to cause protein alterations in the brain i.e., confounding variables other than aging. Subjects with disorders such as cerebrovascular diseases, diabetes, certain pharmaceutical treatments, and brain trauma reported to cause deposition of altered proteins associated with neurodegeneration were not excluded [155,156,157,158,167,168]. In order to reliably assess the influence of the above-mentioned factors on the neurodegenerative protein alteration, multi-center studies on large cohorts are required.

Methods

Anatomical Regions assessed

Regarding the incidence of HP τ pathology, we assessed the LC (study I, IV). The assessment of this region was recommended in 2011. Thus, this region has not been frequently assessed. Furthermore, we assessed TDP43/IR in the medulla, also a region that has not been frequently assessed in cognitively unimpaired subjects. A significant influence of the region assessed regarding incidence was noted.

Thickness of the sections

In the current study, we used 7 μ m thick sections, whereas the assessment of protein alteration has also been carried out on sections with a thickness of 20, 40, and even 100 μ m. The section thickness certainly influences the obtained results.

Staining methods and tissue processing

We applied the IHC technique on formalin fixed paraffin embedded tissue. It has previously been reported that many factors should be taken into consideration, as they are known to alter the staining outcome. Issues of major importance include post mortem delay, fixation time, storage time of sections to be stained, pretreatments, and the choice of antibody used [147,148,149,150,151,152]. All methods that were applied have been shown by the Brain Net Europe to lead to reliable results, even in an inter laboratory setting.

Conclusion

- We demonstrated the early involvement of LC with the HP τ pathology in a relatively large cohort of cognitively unimpaired mid aged subjects (study I). The early involvement of the LC region with the HP τ /IR might be reflected in the clinical presentation of the affected subjects. Development of clinical assessment techniques and radiological investigations that reflect early functional alterations in the LC might assist in identifying subjects with early stages of neurodegeneration; thus, early therapeutic intervention could be applied prior to the occurrence of significant cognitive impairment.
- The recently updated guidelines by the National Institute on Aging – Alzheimer’s Association (NIA-AA) [51, 52] for the assessment of AD related neuropathological changes were indeed applicable in the cognitively unimpaired subjects (study II). Noteworthy, however a substantial number of subjects with HP τ /IR NPs in the temporal cortex were classified as subjects with a low level of AD related changes and were not identified as a risk group (study II). Our recommendation is to assess the HP τ /IR NPs in the temporal cortex in order to identify all subjects at risk.
- We showed that the AD CSF biomarkers and the MMSE scores reflect AD related pathology seen in the cortical biopsies obtained from the iNPH patients (study III). Twenty-two percent of the 111 iNPH patients included in study III displayed AD related pathology in the cortical biopsies. Thus, long-term follow-up of subjects with A β and HP τ pathology in the cortical biopsies is warranted. Furthermore, subjects displaying A β and/or HP τ /IR in the biopsies could be selected for inclusion in future pharmaceutical intervention trials toward these protein alterations.
- We demonstrated a relatively high incidence of neurodegenerative proteinopathies in the aged cognitively unimpaired subjects (IV). Interestingly, a substantial number of subjects fulfilled the criteria for definite PART. Another large group of subjects displayed AD related pathology. Whether PART subjects represent a different entity when compared to subjects with AD related pathology should be further investigated. The high incidence of TDP43 was a surprise, particularly, the frequent involvement of the medulla. The involvement of the medulla should be further studied in cognitively unimpaired subjects.

Acknowledgments

I would like to express my appreciation and gratitude to the patients who participated in this study and I would like to thank their families.

I would like to express my appreciation and gratitude to the study participants who donated their brains and I would like to thank their families.

I would like to express my gratitude to my supervisor Irina Alafuzoff for the immense knowledge and the excellent guidance.

I would like to thank my co supervisor Lena Welsh ,and I would like to express my sincere appreciation and gratitude to the neuropathology research group members; Sevetlana Popova, Sylwia Libard , Marina Leino, Maud Salomonsson, Karin Staxäng and Olivera Casar-Borota. I would like also to thank Katarina Laurell, Kristina Giuliana Cesarini, Hilkka Soininen and Sanna RantaKomi.

I gratefully acknowledge the support of L'ORÉAL-UNESCO for Women in Science program. I am honored to be a recipient of this fellowship. And I would like to express my deepest gratitude to Ahfad University for Women-Sudan and to Professor Gasim Badri and Dr. Amna Badri.

Special thanks to my dear aunt Adila Salih Elobeid to whom this dissertation is dedicated and to my dear uncle Adil Haseeb. Words cannot express how grateful I am.

I would like to express my sincere appreciation and love to my dear mother suad. Thank you for the support, love, encouragement, and endless patience. My sister Hado. Words cannot express how grateful I am for all of the sacrifices that you've made. Thank you for the encouragement, love and support.

I would like to express my sincere appreciation to my dear sisters Balsomi, Hayoota and Lulu .thank you for the love and support and for always being there for me.

References

1. Braak H, Alafuzoff I, Arzberger T, Kretschmar H, Del Tredici K. Staging of Alzheimer disease-associated neurofibrillary pathology using paraffin sections and immunocytochemistry. *Acta Neuropathol.* 2006; 112(4):389-404.
2. Thal DR, Rüb U, Orantes M, Braak H. Phases of A beta-deposition in the human brain and its relevance for the development of AD. *Neurology.* 2002; 58(12):1791-800.
3. Braak H, Del Tredici K, Rüb U, de Vos RA, Jansen Steur EN et al. staging of brain pathology related to sporadic Parkinson's disease. *Neurobiol Aging.* 2003; 24(2):197-211.
4. McKeith IG, Dickson DW, Lowe J, Emre M, O'Brien JT et al .Diagnosis and management of dementia with Lewy bodies: third report of the DLB Consortium. *Neurology.* 2005; 65(12):1863-72.
5. Alafuzoff I, Ince PG, Arzberger T, Al-Sarraj S, Bell J et al. Staging/typing of Lewy body related alpha-synuclein pathology: a study of the BrainNet Europe Consortium. *Acta Neuropathol.* 2009; 117(6):635-52.
6. Neumann M, Sampathu DM, Kwong LK, Truax AC, Micsenyi MC et al. Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Science.* 2006; 314(5796):130-3.
7. Brettschneider J, Del Tredici K, Toledo JB, Robinson JL, Irwin DJ et al .Stages of pTDP-43 pathology in amyotrophic lateral sclerosis. *Ann Neurol.* 2013; 74(1):20-38.
8. Josephs KA, Murray ME, Whitwell JL, Parisi JE, Petrucelli L et al. Staging of TDP-43 pathology in Alzheimer's disease. *Acta Neuropathol.* 2014; 127(3):441-50.
9. Kosik KS, Joachim CL, SelkoeDJ. Microtubule-associated protein tau is a major antigenic component of paired helical filaments in Alzheimer disease. *Proc Natl AcadSci U S A.* 1986; 83(11):4044-8.
10. Grundke-Iqbal I, Iqbal K, Quinlan M, Tung YC, Zaidi MS et al. Microtubule-associated protein tau. A component of Alzheimer paired helical filaments. *J Biol Chem.* 1986; 261(13):6084-9.
11. Lee VM, Goedert M, Trojanowski JQ. Neurodegenerative tauopathies. *Annu Rev Neurosci.* 2001; 24:1121-59.
12. Tolnay M, Probst A. The neuropathological spectrum of neurodegenerative tauopathies. *IUBMB Life.* 2003; 55(6):299-305.
13. Williams DR. Tauopathies: classification and clinical update on neurodegenerative diseases associated with microtubule-associated protein tau. *Intern Med J.* 2006; 36(10):652-60.
14. Kovacs GG. Invited review: Neuropathology of tauopathies: principles and practice. *NeuropatholApplNeurobiol.* 2015; 41(1):3-23.
15. Litvan I. Diagnosis and management of progressive supranuclear palsy. *Semin Neurol.* 2001; 21(1):41-8.
16. Borroni B, Agosti C, Magnani E, Di Luca M, Padovani A. Genetic bases of Progressive Supranuclear Palsy: the MAPT tau disease. *Curr Med Chem.* 2011; 18(17):2655-60.
17. Hauw JJ, Verny M, Delaère P, Cervera P, He Y, DuyckaertsC .Constant neurofibrillary changes in the neocortex in progressive supranuclear palsy. Basic differences with Alzheimer's disease and aging. *Neurosci Lett.* 1990; 119(2):182-6.
18. Komori T. Tau-positive glial inclusions in progressive supranuclear palsy, corticobasal degeneration and Pick's disease. *Brain Pathol.* 1999;9(4):663-79.

19. Yamada T, McGeer PL, McGeer EG. Appearance of paired nucleated, Tau-positive glia in patients with progressive supranuclear palsy brain tissue. *Neurosci Lett.* 1992; 135(1):99-102.
20. Sergeant N, Watzel A, Delacourte A. Neurofibrillary degeneration in progressive supranuclear palsy and corticobasal degeneration: tau pathologies with exclusively "exon 10" isoforms. *J Neurochem.* 1999; 72(3):1243-9.
21. Stover NP, Watts RL. Corticobasal degeneration. *Semin Neurol.* 2001; 21(1):49-58.
22. Di Maria E, Tabaton M, Vigo T, Abbruzzese G, Bellone E et al. Corticobasal degeneration shares a common genetic background with progressive supranuclear palsy. *Ann Neurol.* 2000; 47(3):374-7.
23. Houlden H, Baker M, Morris HR, MacDonald N, Pickering-Brown S et al. Corticobasal degeneration and progressive supranuclear palsy share a common tau haplotype. *Neurology.* 2001; 56(12):1702-6.
24. Iwatsubo T, Hasegawa M, Ihara Y. Neuronal and glial tau-positive inclusions in diverse neurologic diseases share common phosphorylation characteristics. *Acta Neuropathol.* 1994; 88(2):129-36.
25. Feany MB, Mattiace LA, Dickson DW. Neuropathologic overlap of progressive supranuclear palsy, Pick's disease and corticobasal degeneration. *J Neuropathol Exp Neurol.* 1996; 55(1):53-67.
26. Dickson DW, Bergeron C, Chin SS, Duyckaerts C, Horoupian D et al. Office of Rare Diseases of the National Institutes of Health. neuropathologic criteria for corticobasal degeneration. *J Neuropathol Exp Neurol.* 2002; 61(11):935-46.
27. Tolnay M, Clavaguera F. Argyrophilic grain disease: a late-onset dementia with distinctive features among tauopathies. *Neuropathology.* 2004; 24(4):269-83.
28. McKhann GM, Albert MS, Grossman M, Miller B, Dickson D et al. Clinical and pathological diagnosis of frontotemporal dementia: report of the Work Group on Frontotemporal Dementia and Pick's Disease. *Arch Neurol.* 2001; 58(11):1803-9.
29. Hodges JR, Davies RR, Xuereb JH, Casey B, Broe M et al. Clinicopathological correlates in frontotemporal dementia. *Ann Neurol.* 2004; 56(3):399-406.
30. Rizzini C, Goedert M, Hodges JR, Smith MJ, Jakes R et al. Tau gene mutation K257T causes a tauopathy similar to Pick's disease. *J Neuropathol Exp Neurol.* 2000; 59(11):990-1001.
31. Tolnay M, Probst A. Review: Tau protein pathology in Alzheimer's disease and related disorders. *Neuropathol Appl Neurobiol.* 1999; 25(3):171-87.
32. Probst A, Tolnay M, Langui D, Goedert M, Spillantini MG. Pick's disease: hyperphosphorylated tau protein segregates to the somatoaxonal compartment. *Acta Neuropathol.* 1996; 92(6):588-96.
33. Sergeant N, David JP, Lefranc D, Vermersch P, Watzel A et al. Different distribution of phosphorylated tau protein isoforms in Alzheimer's and Pick's diseases. *FEBS Lett.* 1997; 412(3):578-82.
34. Mailliot C, Sergeant N, Bussi re T, Caillet-Boudin ML, Delacourte A et al. Phosphorylation of specific sets of tau isoforms reflects different neurofibrillary degeneration processes. *FEBS Lett.* 1998; 433(3):201-4.
35. Dickson DW. Pick's disease: a modern approach. *Brain Pathol.* 1998; 8(2):339-54.
36. Braak H, Braak E. Neuropathological staging of Alzheimer-related changes. *Acta Neuropathol.* 1991; 82(4):239-59.
37. Braak H, Thal DR, Ghebremedhin E, Del Tredici K. Stages of the pathologic process in Alzheimer disease: age categories from 1 to 100 years. *J Neuropathol Exp Neurol.* 2011; 70(11):960-9.

38. Braak H, Del Tredici K. The pathological process underlying Alzheimer's disease in individuals under thirty. *Acta Neuropathol.* 2011; 121(2):171-81.
39. Crary JF, Trojanowski JQ, Schneider JA, Abisambra JF, Abner EL et al. Primary age-related tauopathy (PART): a common pathology associated with human aging. *Acta Neuropathol.* 2014; 128(6):755-66.
40. Jellinger KA, Bancher C. Senile dementia with tangles (tangle predominant form of senile dementia). *Brain Pathol.* 1998; 8(2):367-76.
41. American Psychiatric Association (1994) Diagnostic and statistical manual of mental disorders, American Psychiatric Association, Washington DC.
42. McKhann G, Drachman D, Folstein M, Katzman R, Price D et al. Clinical diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's disease. *Neurology.* 1984; 34(7):939-44.
43. Alafuzoff I, Pikkarainen M, Al-Sarraj S, Arzberger T, Bell J et al. Interlaboratory comparison of assessments of Alzheimer disease-related lesions: a study of the BrainNet Europe Consortium. *J Neuropathol Exp Neurol.* 2006; 65(8):740-57.
44. Alafuzoff I, Thal DR, Arzberger T, Bogdanovic N, Al-Sarraj S et al. Assessment of beta-amyloid deposits in human brain: a study of the BrainNet Europe Consortium. *Acta Neuropathol.* 2009; 117(3):309-20.
45. Thal DR, Ghebremedhin E, Rüb U, Yamaguchi H, Del Tredici K et al. Two types of sporadic cerebral amyloid angiopathy. *J Neuropathol Exp Neurol.* 2002; 61(3):282-93.
46. Mehndiratta P, Manjila S, Ostergard T, Eisele S, Cohen ML et al. Cerebral amyloid angiopathy-associated intracerebral hemorrhage: pathology and management. *Neurosurg Focus.* 2012; 32(4):E7.
47. Khachaturian ZS. Diagnosis of Alzheimer's disease. *Arch Neurol.* 1985; 42(11):1097-105.
48. Mirra SS, Heyman A, McKeel D, Sumi SM, Crain BJ et al. The Consortium to Establish a Registry for Alzheimer's Disease (CERAD). Part II. Standardization of the neuropathologic assessment of Alzheimer's disease. *Neurology.* 1991; 41(4):479-86.
49. Geddes JW, Tekirian TL, Soultanian NS, Ashford JW, Davis DG et al. Comparison of neuropathologic criteria for the diagnosis of Alzheimer's disease. *Neurobiol Aging.* 1997; 18(4 Suppl):S99-105.
50. Hyman BT, Trojanowski JQ. Consensus recommendations for the postmortem diagnosis of Alzheimer disease from the National Institute on Aging and the Reagan Institute Working Group on diagnostic criteria for the neuropathological assessment of Alzheimer disease. *J Neuropathol Exp Neurol.* 1997; 56(10):1095-7.
51. Hyman BT, Phelps CH, Beach TG, Bigio EH, Cairns NJ et al. National Institute on Aging-Alzheimer's Association guidelines for the neuropathologic assessment of Alzheimer's disease. *Alzheimers Dement.* 2012; 8(1):1-13.
52. Montine TJ, Phelps CH, Beach TG, Bigio EH, Cairns NJ et al. National Institute on Aging-Alzheimer's Association guidelines for the neuropathologic assessment of Alzheimer's disease: a practical approach. *Acta Neuropathol.* 2012; 123(1):1-11.
53. Hulstaert F, Blennow K, Ivanoiu A, Schoonderwaldt HC, Riemenschneider M et al. Improved discrimination of AD patients using beta-amyloid (1-42) and tau levels in CSF. *Neurology.* 1999; 52(8):1555-62.
54. Blennow K, Hampel H. CSF markers for incipient Alzheimer's disease. *Lancet Neurol.* 2003; 2(10):605-13.

55. Zetterberg H, Wahlund LO, Blennow K. Cerebrospinal fluid markers for prediction of Alzheimer's disease. *Neurosci Lett*. 2003; 352(1):67-9.
56. Hampel H, Mitchell A, Blennow K, Frank RA, Brettschneider S et al. Core biological marker candidates of Alzheimer's disease - perspectives for diagnosis, prediction of outcome and reflection of biological activity. *J Neural Transm*. 2004;111(3):247-72.
57. Parnetti L, Lanari A, Silvestrelli G, Saggese E, Reboldi P. Diagnosing prodromal Alzheimer's disease: role of CSF biochemical markers. *Mech Ageing Dev*. 2006; 127(2):129-32.
58. Spillantini MG, Schmidt ML, Lee VM, Trojanowski JQ, Jakes R et al. Alpha-synuclein in Lewy bodies. *Nature*. 1997; 388(6645):839-40.
59. Spillantini MG, Goedert M. The alpha-synucleinopathies: Parkinson's disease, dementia with Lewy bodies, and multiple system atrophy. *Ann N Y Acad Sci*. 2000; 920:16-27.
60. Jellinger KA. Neuropathological spectrum of synucleinopathies. *Mov Disord*. 2003; 18 Suppl 6:S2-12.
61. McCann H, Stevens CH, Cartwright H, Halliday GM. α -Synucleinopathy phenotypes. *Parkinsonism Relat Disord*. 2014; 20 Suppl 1:S62-7.
62. Gelb DJ, Oliver E, Gilman S. Diagnostic criteria for Parkinson disease. *Arch Neurol*. 1999; 56(1):33-9.
63. Fearnley JM, Lees AJ. Ageing and Parkinson's disease: substantia nigra regional selectivity. *Brain*. 1991;114 (Pt 5):2283-301.
64. Ma SY, Rinne JO, Collan Y, R ytt  M, Rinne UK. A quantitative morphometrical study of neuron degeneration in the substantia nigra in Parkinson's disease. *J Neurol Sci*. 1996; 140(1-2):40-5.
65. Forno LS. Neuropathology of Parkinson's disease. *J Neuropathol Exp Neurol*. 1996; 55(3):259-72.
66. Kosaka K, Yoshimura M, Ikeda K, Budka H. Diffuse type of Lewy body disease: progressive dementia with abundant cortical Lewy bodies and senile changes of varying degree--a new disease? *Clin Neuropathol*. 1984; 3(5):185-92.
67. McKeith IG, Galasko D, Kosaka K, Perry EK, Dickson DW et al. Consensus guidelines for the clinical and pathologic diagnosis of dementia with Lewy bodies (DLB): report of the consortium on DLB international workshop. *Neurology*. 1996 ;47(5):1113-24.
68. Leverenz JB, Hamilton R, Tsuang DW, Schantz A, Vavrek D et al. Empiric refinement of the pathologic assessment of Lewy-related pathology in the dementia patient. *Brain Pathol*. 2008; 18(2):220-4.
69. Parkkinen L, Pirttil  T, Alafuzoff I. Applicability of current staging/categorization of alpha-synuclein pathology and their clinical relevance. *Acta Neuropathol*. 2008; 115(4):399-407.
70. Zaccai J, Brayne C, McKeith I, Matthews F, Ince PG et al. Patterns and stages of alpha-synucleinopathy: Relevance in a population-based cohort. *Neurology*. 2008 25; 70(13):1042-8.
71. Gilman S, Wenning GK, Low PA, Brooks DJ, Mathias CJ et al. Second consensus statement on the diagnosis of multiple system atrophy. *Neurology*. 2008;71(9):670-6.
72. Lantos PL. The definition of multiple system atrophy: a review of recent developments. *J Neuropathol Exp Neurol*. 1998; 57(12):1099-111.
73. Ahmed Z, Asi YT, Sailer A, Lees AJ, Houlden H et al. The neuropathology, pathophysiology and genetics of multiple system atrophy. *Neuropathol Appl Neurobiol*. 2012; 38(1):4-24.

74. Trojanowski JQ, Revesz T. Proposed neuropathological criteria for the post mortem diagnosis of multiple system atrophy. *Neuropathol Appl Neurobiol.* 2007; 33(6):615-20.
75. Arai T, Hasegawa M, Akiyama H, Ikeda K, Nonaka T et al. TDP-43 is a component of ubiquitin-positive tau-negative inclusions in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *BiochemBiophys Res Commun.* 2006; 351(3):602-11.
76. Neumann M. Molecular neuropathology of TDP-43 proteinopathies. *Int J Mol Sci.* 2009; 10(1):232-46.
77. Higashi S, Iseki E, Yamamoto R, Minegishi M, Hino H et al. Concurrence of TDP-43, tau and alpha-synuclein pathology in brains of Alzheimer's disease and dementia with Lewy bodies. *Brain Res.* 2007; 12(1184):284-94.
78. Arai T, Mackenzie IR, Hasegawa M, Nonaka T, Niizato K et al. Phosphorylated TDP-43 in Alzheimer's disease and dementia with Lewy bodies. *Acta Neuropathol.* 2009; 117(2):125-36.
79. Kadokura A, Yamazaki T, Lemere CA, Takatama M, Okamoto K. Regional distribution of TDP-43 inclusions in Alzheimer disease (AD) brains: their relation to AD common pathology. *Neuropathology.* 2009; 29(5):566-73.
80. King A, Sweeney F, Bodi I, Troakes C, Maekawa S et al. Abnormal TDP-43 expression is identified in the neocortex in cases of dementia pugilistica, but is mainly confined to the limbic system when identified in high and moderate stages of Alzheimer's disease. *Neuropathology.* 2010; 30(4):408-19.
81. Tremblay C, St-Amour I, Schneider J, Bennett DA, Calon F. Accumulation of transactive response DNA binding protein 43 in mild cognitive impairment and Alzheimer disease. *J Neuropathol Exp Neurol.* 2011; 70(9):788-98.
82. Gordon PH. Amyotrophic Lateral Sclerosis: An update for 2013 Clinical Features, Pathophysiology, Management and Therapeutic Trials. *Aging Dis.* 2013; 4(5):295-310.
83. Wijesekera LC, Leigh PN. Amyotrophic lateral sclerosis. *Orphanet J Rare Dis.* 2009 .3;4:3.
84. Ince PG, Highley JR, Kirby J, Wharton SB, Takahashi H et al. Molecular pathology and genetic advances in amyotrophic lateral sclerosis: an emerging molecular pathway and the significance of glial pathology. *Acta Neuropathol.* 2011; 122(6):657-71.
85. Lill CM, Abel O, Bertram L, Al-Chalabi A. Keeping up with genetic discoveries in amyotrophic lateral sclerosis: the ALSod and ALSGene databases. *Amyotroph Lateral Scler.* 2011; 12(4):238-49.
86. Al-Chalabi A, Jones A, Troakes C, King A, Al-Sarraj S et al. The genetics and neuropathology of amyotrophic lateral sclerosis. *Acta Neuropathol.* 2012; 124(3):339-52.
87. Benajiba L, Le Ber I, Camuzat A, Lacoste M, Thomas-Anterion C et al. TARDBP mutations in motor neuron disease with frontotemporal lobar degeneration. *Ann Neurol.* 2009; 65(4):470-3.
88. Neumann M, Mackenzie IR, Cairns NJ, Boyer PJ, Markesbery WR et al. TDP-43 in the ubiquitin pathology of frontotemporal dementia with VCP gene mutations. *J Neuropathol Exp Neurol.* 2007; 66(2):152-7.
89. Baker M, Mackenzie IR, Pickering-Brown SM, Gass J et al. Mutations in progranulin cause tau-negative frontotemporal dementia linked to chromosome 17. *Nature.* 2006; 442(7105):916-9.
90. Chen-Plotkin AS, Martinez-Lage M, Sleiman PM, Hu W, Greene R et al. Genetic and clinical features of progranulin-associated frontotemporal lobar degeneration. *Arch Neurol.* 2011; 68(4):488-97.

91. Annakaisa Haapasalo A, Remes A. Genetic and Molecular Aspects of Frontotemporal Lobar Degeneration. *Curr Genet Med Rep*; 2015;(3)8–18.
92. Mackenzie IR, Neumann M, Baborie A, Sampathu DM, Du Plessis D et al. A harmonized classification system for FTL-D-TDP pathology. *Acta Neuropathol*. 2011; 122(1):111-3.
93. Brettschneider J, Del Tredici K, Irwin DJ, Grossman M, Robinson JL et al. Sequential distribution of pTDP-43 pathology in behavioral variant frontotemporal dementia (bvFTD). *Acta Neuropathol*. 2014; 127(3):423-39.
94. Vance C, Rogelj B, Hortobágyi T, De Vos KJ, Nishimura AL et al. Mutations in FUS, an RNA processing protein, cause familial amyotrophic lateral sclerosis type 6. *Science*. 2009; 323(5918):1208-11.
95. Neumann M, Rademakers R, Roeber S, Baker M, Kretzschmar HA et al. A new subtype of frontotemporal lobar degeneration with FUS pathology. *Brain*. 2009; 132(Pt 11):2922-31.
96. Kabashi E, Valdmanis PN, Dion P, Rouleau GA. Oxidized/misfolded superoxide dismutase-1: the cause of all amyotrophic lateral sclerosis? *Ann Neurol*. 2007; 62(6):553-9.
97. Walker FO. Huntington's disease. *Lancet*. 2007; 369(9557):218-28.
98. Head MW, Ironside JW. Review: Creutzfeldt-Jakob disease: prion protein type, disease phenotype and agent strain. *Neuropathol Appl Neurobiol*. 2012; 38(4):296-310.
99. Armstrong RA, Lantos PL, Cairns NJ. Overlap between neurodegenerative disorders. *Neuropathology*. 2005; 25(2):111-24.
100. Schneider JA, Arvanitakis Z, Bang W, Bennett DA. Mixed brain pathologies account for most dementia cases in community-dwelling older persons. *Neurology*. 2007; 11; 69(24):2197-204.
101. Jellinger KA, Attems J. Prevalence and impact of vascular and Alzheimer pathologies in Lewy body disease. *Acta Neuropathol*. 2008; 115(4):427-36.
102. Kovacs GG, Alafuzoff I, Al-Sarraj S, Arzberger T, Bogdanovic N et al. Mixed brain pathologies in dementia: the BrainNet Europe consortium experience. *Dement Geriatr Cogn Disord*. 2008; 26(4):343-50.
103. Jellinger KA, Attems J. Prevalence of dementia disorders in the oldest-old: an autopsy study. *Acta Neuropathol*. 2010; 119(4):421-33.
104. Attems J, Jellinger K. Neuropathological correlates of cerebral multimorbidity. *Curr Alzheimer Res*. 2013; 10(6):569-77.
105. Dugger BN, Adler CH, Shill HA, Caviness J, Jacobson S et al. Concomitant pathologies among a spectrum of parkinsonian disorders. *Parkinsonism Relat Disord*. 2014; 20(5):525-9.
106. Rahimi J, Kovacs GG. Prevalence of mixed pathologies in the aging brain. *Alzheimers Res Ther*. 2014;6(9):82.
107. Jellinger KA, Attems J. Challenges of multimorbidity of the aging brain: a critical update. *J Neural Transm*. 2015; 122(4):505-21.
108. Lippa CF, Fujiwara H, Mann DM, Giasson B, Baba M et al. Lewy bodies contain altered alpha-synuclein in brains of many familial Alzheimer's disease patients with mutations in presenilin and amyloid precursor protein genes. 1998. *Am J Pathol* .153: 1365-1370.
109. Hamilton RL. Lewy bodies in Alzheimer's disease: a neuropathological review of 145 cases using alpha-synuclein immunohistochemistry. *Brain Pathol*. 2000; 10(3):378-84.
110. Jellinger KA, Attems J. Does striatal pathology distinguish Parkinson disease with dementia and dementia with Lewy bodies? *Acta Neuropathol*. 2006;112(3):253-60.

111. Kalaitzakis ME, Graeber MB, Gentleman SM, Pearce RK. Striatal beta-amyloid deposition in Parkinson disease with dementia. *J Neuropathol Exp Neurol.* 2008;67(2):155-6.
112. Kalaitzakis ME, Pearce RK. The morbid anatomy of dementia in Parkinson's disease. *Acta Neuropathol.* 2009;118(5):587-98.
113. Nakashima-Yasuda H, Uryu K, Robinson J, Xie SX, Hurtig H et al. Comorbidity of TDP-43 proteinopathy in Lewy body related diseases. *Acta Neuropathol.* 2007;114(3):221-9.
114. Amador-Ortiz C, Lin WL, Ahmed Z, Personett D, Davies P et al. TDP-43 immunoreactivity in hippocampal sclerosis and Alzheimer's disease. *Ann Neurol.* 2007; 61(5):435-45.
115. Freeman SH, Spires-Jones T, Hyman BT, Growdon JH, Frosch MP. TAR-DNA binding protein 43 in Pick disease. *J Neuropathol Exp Neurol.* 2008; 67(1):62-7.
116. Schwab C, Arai T, Hasegawa M, Yu S, McGeer P L. Colocalization of transactivation-responsive DNA-binding protein 43 and huntingtin in inclusions of Huntington disease. *J Neuropathol Exp Neurol.* 2008;67(12):1159-65
117. Price JL, Davis PB, Morris JC, White DL. The distribution of tangles, plaques and related immunohistochemical markers in healthy aging and Alzheimer's disease. *Neurobiol Aging.* 1991; 12(4):295-312.
118. Price JL, Morris JC. Tangles and plaques in nondemented aging and "preclinical" Alzheimer's disease. *Ann Neurol.* 1999;45(3):358-68.
119. Price JL, McKeel DW Jr, Buckles VD, Roe CM, Xiong C et al. Neuropathology of nondemented aging: presumptive evidence for preclinical Alzheimer disease. *Neurobiol Aging.* 2009; 30(7):1026-36.
120. Bennett DA, Wilson RS, Boyle PA, Buchman AS, Schneider JA. Relation of neuropathology to cognition in persons without cognitive impairment. *Ann Neurol.* 2012; 72(4):599-609.
121. Aho L, Karkola K, Juusela J, Alafuzoff I. Heavy alcohol consumption and neuropathological lesions: a post-mortem human study. *J Neurosci Res.* 2009;87(12):2786-92.
122. Boyle PA, Yu L, Wilson RS, Schneider JA, Bennett DA. Relation of neuropathology with cognitive decline among older persons without dementia. *Front Aging Neurosci.* 2013;9: 5-50.
123. Knopman DS, Parisi JE, Salviati A, Floriach-Robert M, Boeve BF et al. Neuropathology of cognitively normal elderly. *J Neuropathol Exp Neurol.* 2003; 62(11):1087-95.
124. Kovacs GG, Milenkovic I, Wöhrer A, Höftberger R, Gelpi E et al. Non-Alzheimer neurodegenerative pathologies and their combinations are more frequent than commonly believed in the elderly brain: a community-based autopsy series. *Acta Neuropathol.* 2013;126(3):365-84.
125. Dugger BN, Hentz JG, Adler CH, Sabbagh MN, Shill HA et al. Clinicopathological outcomes of prospectively followed normal elderly brain bank volunteers. *J Neuropathol Exp Neurol.* 2014; 73(3):244-52.
126. Delaère P, Duyckaerts C, Masters C, Beyreuther K, Piette F et al. Large amounts of neocortical beta A4 deposits without neuritic plaques nor tangles in a psychometrically assessed, non-demented person. *Neurosci Lett.* 1990;116(1-2):87-93.
127. Delaère P, He Y, Fayet G, Duyckaerts C, Hauw JJ. Beta A4 deposits are constant in the brain of the oldest old: an immunocytochemical study of 20 French centenarians. *Neurobiol Aging.* 1993;14(2):191-4.

128. Tsuboi Y, Ahlskog JE, Apaydin H, Parisi JE, Dickson DW .Lewy bodies are not increased in progressive supranuclear palsy compared with normal controls. *Neurology*. 2001;57(9):1675-8.
129. Parkkinen L, Soininen H, Laakso M, Alafuzoff I. Alpha-synuclein pathology is highly dependent on the case selection. *Neuropathol Appl Neurobiol*. 2001;27(4):314-25.
130. Jellinger KA. Lewy body-related alpha-synucleinopathy in the aged human brain. *J Neural Transm*. 2004;111(10-11):1219-35.
131. Markesbery WR, Jicha GA, Liu H, Schmitt FA. Lewy body pathology in normal elderly subjects. *J Neuropathol Exp Neurol*. 2009;68(7):816-22.
132. Gibb WR, Lees AJ. The relevance of the Lewy body to the pathogenesis of idiopathic Parkinson's disease. *J Neurol Neurosurg Psychiatry*. 1988;51(6):745-52.
133. Del Tredici K, Rüb U, De Vos RA, Bohl JR, Braak H .Where does parkinson disease pathology begin in the brain?. *J Neuropathol Exp Neurol*. 2002 ;61(5):413-26.
134. Frigerio R, Fujishiro H, Ahn TB, Josephs KA, Maraganore DM et al. Incidental Lewy body disease: do some cases represent a preclinical stage of dementia with Lewy bodies? *Neurobiol Aging*. 2011;32(5):857-63.
135. Donaghy PC, McKeith IG. The clinical characteristics of dementia with Lewy bodies and a consideration of prodromal diagnosis. *Alzheimers Res Ther*. 2014;6(4):46.
136. DelleDonne A, Klos KJ, Fujishiro H, Ahmed Z, Parisi JE et al. Incidental Lewy body disease and preclinical Parkinson disease. *Arch Neurol*. 2008;65(8):1074-80.
137. Ross GW, Abbott RD, Petrovitch H, Tanner CM, Davis DG et al .Association of olfactory dysfunction with incidental Lewy bodies. *Mov Disord*. 2006;21(12):2062-7.
138. Boeve BF, Dickson DW, Olson EJ, Shepard JW, Silber MH et al .Insights into REM sleep behaviour disorder pathophysiology in brainstem-predominant Lewy body disease. *Sleep Med*. 2007;8(1):60-4.
139. Uchiyama M, Isse K, Tanaka K, Yokota N, Hamamoto M et al. Incidental Lewy body disease in a patient with REM sleep behaviour disorder. *Neurology*. 1995;45(4):709-12.
140. Arnold SJ, Dugger BN, Beach TG. TDP-43 deposition in prospectively followed, cognitively normal elderly individuals: correlation with argyrophilic grains but not other concomitant pathologies. *Acta Neuropathol*. 2013; 126(1):51-7.
141. White L. Brain lesions at autopsy in older Japanese-American men as related to cognitive impairment and dementia in the final years of life: a summary report from the Honolulu-Asia aging study. *J Alzheimers Dis*. 2009; 18(3):713-25.
142. Relkin N, Marmarou A, Klinge P, Bergsneider M, Black PM. Diagnosing idiopathic normal-pressure hydrocephalus. *Neurosurgery*. 2005; 57(3 Suppl):S4-16; discussion ii-v. Review.
143. Savolainen S, Paljärvi L, Vapalahti M. Prevalence of Alzheimer's disease in patients investigated for presumed normal pressure hydrocephalus: a clinical and neuropathological study. *Acta Neurochir* . 1999; 141(8):849-53.
144. Golomb J, Wisoff J, Miller DC, Boksay I, Kluger A et al. Alzheimer's disease comorbidity in normal pressure hydrocephalus: prevalence and shunt response. *J Neurol Neurosurg Psychiatry*. 2000; 68(6):778-81.

145. Leinonen V, Koivisto AM, Savolainen S, Rummukainen J, Tamminen JN et al. Amyloid and tau proteins in cortical brain biopsy and Alzheimer's disease. *Ann Neurol.* 2010;68(4):446-53.
146. Leinonen V, Koivisto AM, Savolainen S, Rummukainen J, Sutela A et al. Post-mortem findings in 10 patients with presumed normal-pressure hydrocephalus and review of the literature. *Neuropathol Appl Neurobiol.* 2012;38(1):72-86.
147. Chandana R, Mythri RB, Mahadevan A, Shankar SK, Srinivas Bharath MM. Biochemical analysis of protein stability in human brain collected at different post-mortem intervals. *Indian J Med Res.* 2009; 129(2):189-99.
148. Pikkarainen M, Martikainen P, Alafuzoff I. The effect of prolonged fixation time on immunohistochemical staining of common neurodegenerative disease markers. *J Neuropathol Exp Neurol.* 2010; 69(1):40-52.
149. Ferrer I, Santpere G, Arzberger T, Bell J, Blanco R et al. Brain protein preservation largely depends on the postmortem storage temperature: implications for study of proteins in human neurologic diseases and management of brain banks: a BrainNet Europe Study. *J Neuropathol Exp Neurol.* 2007;66(1):35-46.
150. Ramos-Vara JA, Miller MA. When tissue antigens and antibodies get along: revisiting the technical aspects of immunohistochemistry--the red, brown, and blue technique. *Vet Pathol.* 2014;51(1):42-87.
151. Kraszpulski M, Soininen H, Riekkinen P Sr, Alafuzoff I. Pitfalls in the quantitative estimation of beta-amyloid immunoreactivity in human brain tissue. *Histochem Cell Biol.* 1998;110(4):439-45.
152. Ramos-Vara JA, Webster JD, DuSold D, Miller MA. Immunohistochemical evaluation of the effects of paraffin section storage on biomarker stability. *Vet Pathol.* 2014;51(1):102-9.
153. Attems J, Thal DR, Jellinger KA. The relationship between subcortical tau pathology and Alzheimer's disease. *Biochem Soc Trans.* 2012;40(4):711-5.
154. Attems J, Thomas A, Jellinger K. Correlations between cortical and subcortical tau pathology. *Neuropathol Appl Neurobiol.* 2012;38(6):582-90.
155. Ikonovic MD, Uryu K, Abrahamson EE, Ciallella JR, Trojanowski JQ et al. Alzheimer's pathology in human temporal cortex surgically excised after severe brain injury. *Exp Neurol.* 2004;190(1):192-203.
156. Guise S, Braguer D, Remacle-Bonnet M, Pommier G, Briand C. Tau protein is involved in the apoptotic process induced by anti-microtubule agents on neuroblastoma cells. *Apoptosis.* 1999;4(1):47-58.
157. Guise S, Braguer D, Carles G, Delacourte A, Briand C. Hyperphosphorylation of tau is mediated by ERK activation during anticancer drug-induced apoptosis in neuroblastoma cells. *J Neurosci Res.* 2001;63(3):257-67.
158. Höglinger GU, Lannuzel A, Khondiker ME, Michel PP, Duyckaerts C et al. The mitochondrial complex I inhibitor rotenone triggers a cerebral tauopathy. *J Neurochem.* 2005;95(4):930-9.
159. Pyykkö OT, Lumela M, Rummukainen J, Nerg O, Seppälä TT et al. Cerebrospinal fluid biomarker and brain biopsy findings in idiopathic normal pressure hydrocephalus. *PLoS One.* 2014;9(3):e91974.
160. Seppälä TT, Nerg O, Koivisto AM, Rummukainen J, Puli L et al. CSF biomarkers for Alzheimer disease correlate with cortical brain biopsy findings. *Neurology.* 2012;78(20):1568-75.
161. Wilson CA, Doms RW, Lee VM. Intracellular APP processing and A beta production in Alzheimer disease. *J Neuropathol Exp Neurol.* 1999;58(8):787-94.
162. Jakes R, Spillantini MG, Goedert M. Identification of two distinct synucleins from human brain. *FEBS Lett.* 1994;345(1):27-32.

163. Bendor JT, Logan TP, Edwards RH. The function of α -synuclein. *Neuron*. 2013;79(6):1044-66.
164. Beach TG, Adler CH, Sue LI, Vedders L, Lue L et al. Multi-organ distribution of phosphorylated alpha-synuclein histopathology in subjects with Lewy body disorders. *Acta Neuropathol*. 2010; 119(6):689-702.
165. Buratti E, Baralle FE. Multiple roles of TDP-43 in gene expression, splicing regulation, and human disease. *Front Biosci*. 2008; 13:867-78.
166. Uchino A, Takao M, Hatsuta H, Sumikura H, Nakano Y et al. Incidence and extent of TDP-43 accumulation in aging human brain. *Acta Neuropathol Commun*. 2015; 20; 3:35.
167. Skoog I, Lernfelt B, Landahl S, Palmertz B, Andreasson LA et al. 15-year longitudinal study of blood pressure and dementia. *Lancet*. 1996; 347(9009):1141-5.
168. de Toledo Ferraz Alves TC, Ferreira LK, Wajngarten M, Busatto GF. Cardiac disorders as risk factors for Alzheimer's disease. *J Alzheimers Dis*. 2010; 20(3):749-63.

Acta Universitatis Upsaliensis

*Digital Comprehensive Summaries of Uppsala Dissertations
from the Faculty of Medicine 1182*

Editor: The Dean of the Faculty of Medicine

A doctoral dissertation from the Faculty of Medicine, Uppsala University, is usually a summary of a number of papers. A few copies of the complete dissertation are kept at major Swedish research libraries, while the summary alone is distributed internationally through the series Digital Comprehensive Summaries of Uppsala Dissertations from the Faculty of Medicine. (Prior to January, 2005, the series was published under the title "Comprehensive Summaries of Uppsala Dissertations from the Faculty of Medicine".)

Distribution: publications.uu.se
urn:nbn:se:uu:diva-277214



ACTA
UNIVERSITATIS
UPSALIENSIS
UPPSALA
2016