



# Amelioration of arthritis through mobilization of peptide-specific CD8<sup>+</sup> regulatory T cells

Jianmei W. Leavenworth,<sup>1,2</sup> Xiaolei Tang,<sup>1,2</sup> Hye-Jung Kim,<sup>1,2</sup>  
Xiaoyang Wang,<sup>1,2,3,4</sup> and Harvey Cantor<sup>1,2</sup>

<sup>1</sup>Department of Cancer Immunology and AIDS, Dana-Farber Cancer Institute, and <sup>2</sup>Department of Microbiology and Immunobiology, Division of Immunology, Harvard Medical School, Boston, Massachusetts, USA. <sup>3</sup>Perinatal Center, Department of Neuroscience and Physiology, Sahlgrenska Academy at University of Gothenburg, Gothenburg, Sweden. <sup>4</sup>Department of Pediatrics, The Third Affiliated Hospital of Zhengzhou University, Zhengzhou, People's Republic of China.

**Current therapies to treat autoimmune disease focus mainly on downstream targets of autoimmune responses, including effector cells and cytokines. A potentially more effective approach would entail targeting autoreactive T cells that initiate the disease cascade and break self tolerance. The murine MHC class Ib molecule Qa-1b (HLA-E in humans) exhibits limited polymorphisms and binds to 2 dominant self peptides: Hsp60<sub>p216</sub> and Qdm. We found that peptide-induced expansion of tetramer-binding CD8<sup>+</sup> Tregs that recognize Qa-1-Hsp60<sub>p216</sub> but not Qa-1-Qdm strongly inhibited collagen-induced arthritis, an animal model of human rheumatoid arthritis. Perforin-dependent elimination of autoreactive follicular Th (T<sub>FH</sub>) and Th17 cells by CD8<sup>+</sup> Tregs inhibited disease development. Infusion of in vitro-expanded CD8<sup>+</sup> Tregs increased the efficacy of methotrexate treatment and halted disease progression after clinical onset, suggesting an alternative approach to this first-line treatment. Moreover, infusion of small numbers of Qa-1-Hsp60<sub>p216</sub>-specific CD8<sup>+</sup> Tregs resulted in robust inhibition of autoimmune arthritis, confirming the inhibitory effects of Hsp60<sub>p216</sub> peptide immunization. These results suggest that strategies designed to expand Qa-1-restricted (HLA-E-restricted), peptide-specific CD8<sup>+</sup> Tregs represent a promising therapeutic approach to autoimmune disorders.**

## Introduction

RA is an autoimmune disorder characterized primarily by joint inflammation and erosion. The systemic nature of this disease is reflected in the involvement of multiple organ systems, including skin, lungs, and vasculature as well as diffuse tissue inflammation and increased risk of atherosclerosis (1). Although the identification of effector cytokines, including TNF, IL-1, and IL-6, that contribute to this disease has led to improved therapies, RA has remained relatively refractory to decisive intervention. More effective approaches to RA may require elimination of autoreactive T cells, which, in turn depends on a precise definition of the Tregs that may target and eliminate the pathogenic T cell subsets that induce disease.

Here we analyze the contribution of Treg and effector T cell subsets to autoimmune arthritis in the collagen-induced arthritis (CIA) animal model. This murine disease model shares several similarities with human RA, including breach of self tolerance, generation of autoantibodies, inflammatory changes in multiple joints, and erosion of bone and cartilage accompanied by pannus formation (2). Murine CIA has been used to establish the potential efficacy of several FDA-approved RA therapies, including anti-TNF Ab, IL-1 antagonists, and methotrexate (MTX) (1, 3).

The development of enlarged ectopic germinal centers (GCs) and the contribution of pathogenic autoantibodies in RA and in CIA has suggested that dysregulated follicular Th (T<sub>FH</sub>) cell responses may contribute to ectopic GC formation and production of autoantibodies (4–6). A subset of CD8<sup>+</sup> T cells, CD8<sup>+</sup> Tregs, has been shown to efficiently inhibit T<sub>FH</sub> cells through recognition of MHC class Ib molecule Qa-1 expressed by this Th subset (5). However, the potential contribution of CD8<sup>+</sup> Treg to the pathogenesis and treatment of this autoimmune disease is not well understood.

CD8<sup>+</sup> Tregs recognize the MHC class Ib molecule Qa-1b (HLA-E in humans) associated with peptide complexes that are upregulated by activated CD4<sup>+</sup> T cells (5, 7). The 2 major peptides presented by Qa-1 (HLA-E) are Qdm (B7sp in humans) and Hsp60<sub>p216</sub>. The Qdm (Qa-1 determinant modifier) peptide is derived from the leader sequence of MHC class Ia, while the Hsp60<sub>p216</sub> peptide is derived from the 60 kD heat shock protein (Hsp60, p216–224). Differential expression of these Qa-1-peptide complexes on activated cells may allow detection of abnormally stressed cells in the context of infection, autoimmunity, or cellular transformation (8–11).

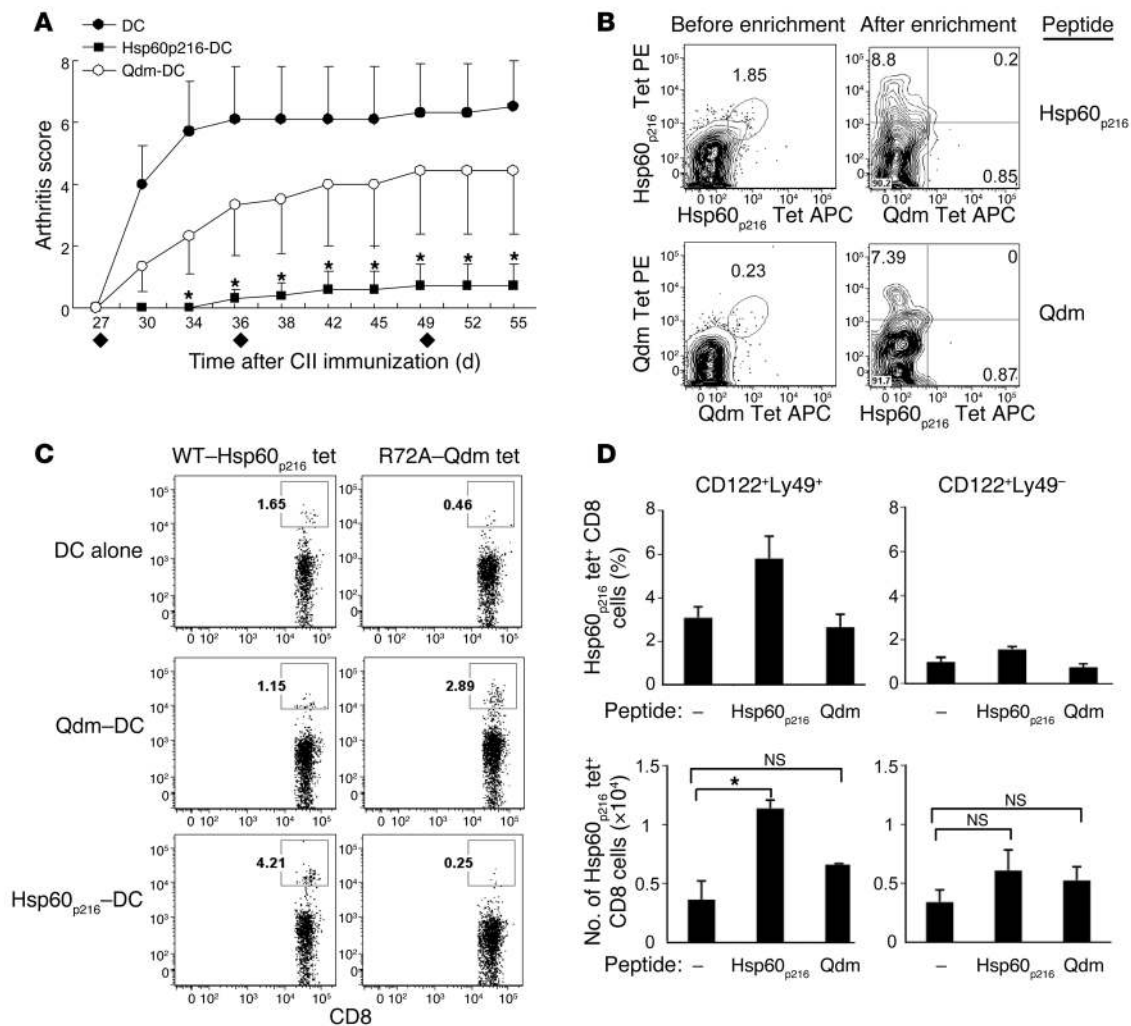
Here we investigate the impact of CD8<sup>+</sup> Tregs on the development of CIA. We find that Qa-1-restricted CD8<sup>+</sup> Tregs efficiently eliminate pathogenic T<sub>FH</sub> and Th17 cells and exert strong inhibitory effects on disease progression. Moreover, CD8<sup>+</sup> Tregs that recognize complexes of Qa-1 and Hsp60<sub>p216</sub> peptide, but not Qdm peptide, can be mobilized through peptide-based immunization to inhibit development of autoimmune arthritis. The ability to expand Qa-1-restricted CD8<sup>+</sup> Tregs based on their TCR specificity may represent a new and effective approach to treatment of autoimmune disease.

## Results

**Expansion of Qa-1-restricted CD8<sup>+</sup> Tregs inhibits arthritis.** We reasoned that expansion of Qa-1-restricted CD8<sup>+</sup> T cells in vivo might be achieved after immunization with the 2 major peptides that are bound by Qa-1 — Qdm and Hsp60<sub>p216</sub>. The Qdm peptide, derived from the leader sequences of class Ia MHC proteins, is the dominant self peptide associated with Qa-1 at the cell surface. In addition, a peptide derived from the 60-kD heat shock protein (Hsp60, p216–224) replaces the Qdm peptide upon cellular activation and/or stress (12, 13). To selectively expand Qdm- or Hsp60<sub>p216</sub>-specific CD8<sup>+</sup> cells, we immunized mice subcutaneously with peptide-pulsed *Kb<sup>-/-</sup>Db<sup>-/-</sup>* DCs that do not

**Conflict of interest:** The authors have declared that no conflict of interest exists.

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**Figure 1**

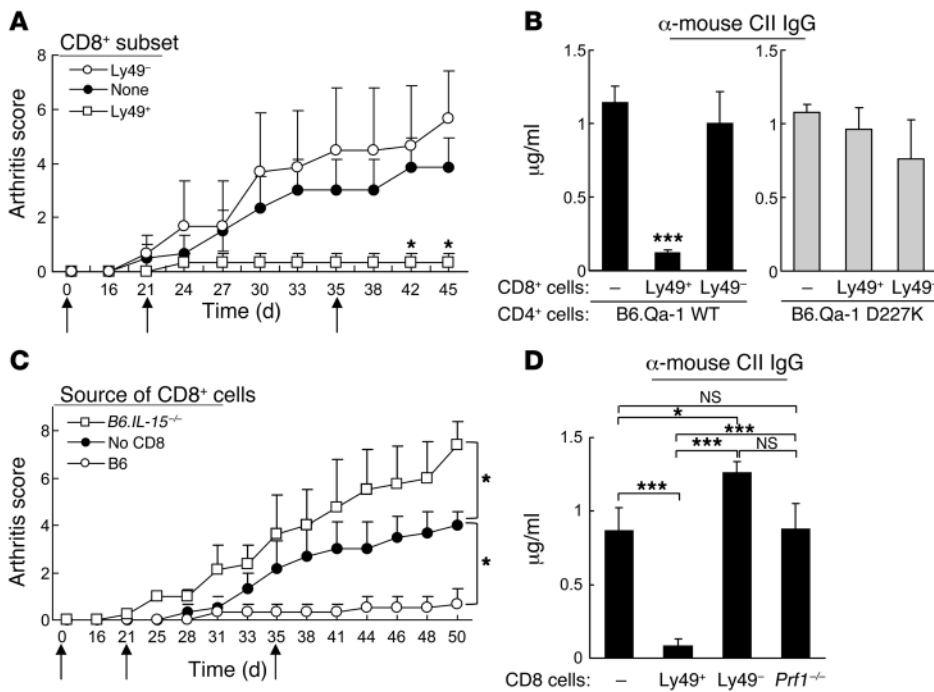
Selective expansion of Hsp60<sub>p216</sub>-specific Qa-1-restricted CD8<sup>+</sup> Tregs inhibits arthritis. (A) Arthritis was induced in B6 mice as described in Methods. Irradiated LPS-activated *Kb<sup>-/-</sup>Db<sup>-/-</sup>* BM-derived DCs were loaded with or without Hsp60<sub>p216</sub> or Qdm peptide and injected subcutaneously ( $6 \times 10^5$ /mouse) at the tail base at day 27, day 36, and day 49 (diamonds). Arthritis scores are shown for 5–6 mice per group. The group injected with DCs alone versus the group given Hsp60<sub>p216</sub>-loaded DCs differed significantly. \**P* < 0.05. (B) Flow cytometry of dLN cells from CII-immune B6 mice injected with DC-pulsed peptides as in A, after incubation with Qa-1 tetramers labeled with phycoerythrin (Tet-PE) or allophycocyanin (Tet-APC); analysis is shown before and after enrichment by magnetic bead columns for tet<sup>+</sup> CD8<sup>+</sup> cells. Numbers in plots before enrichment and after enrichment (upper left quadrant) indicate percentage of tet<sup>+</sup> CD8<sup>+</sup> cells. (C) B6 mice were immunized with either unpulsed *Kb<sup>-/-</sup>Db<sup>-/-</sup>* DCs or DCs that had been loaded with the indicated peptide as described in A. CD8<sup>+</sup> cells from dLNs from each group were analyzed at day 42 for binding to Qa-1.R72A-Qdm-tetramer or Qa-1-Hsp60<sub>p216</sub>-tetramer. Representative FACS plots are shown. Gates represent percentages of tet<sup>+</sup> CD8<sup>+</sup> cells. (D) The percentages and numbers of Hsp60<sub>p216</sub>-tet<sup>+</sup> CD8<sup>+</sup> cells that expressed the CD122<sup>+</sup>Ly49<sup>+</sup> surface profile and CD122<sup>+</sup>Ly49<sup>-</sup> surface profile are shown. \**P* < 0.05.

express class Ia MHC, but express high levels of class Ib MHC, including Qa-1. Immunization of mice with Hsp60<sub>p216</sub>- but not Qdm-pulsed *Kb<sup>-/-</sup>Db<sup>-/-</sup>* DCs during the course of CIA inhibited the development of arthritis (Figure 1A).

We then generated Qa-1-peptide tetramers (tet) to determine the frequency of peptide-specific CD8<sup>+</sup> T cells. In the case of Qa-1-Qdm tetramers, a Qa-1 protein containing a point mutation (R72A) was used to prevent binding of Qa-1-Qdm to CD94/NKG2A receptors (14) and allow specific detection of CD8<sup>+</sup> T cells that express Qa-1-restricted TCR (Supplemental Figure 1A; supplemental material available online with this article; doi:10.1172/JCI66938DS1). We labeled cells with tetramers con-

jugated to 2 different fluorophores to increase specificity, followed by magnetic enrichment of cells positive for relative Qa-1-peptide tetramers as described (refs. 10, 15, and Figure 1B). Enrichment for CD8<sup>+</sup> T cells with the Qa-1-peptide tetramers was specific, since we detected only background frequency of Qa-1-peptide tet<sup>+</sup>CD8<sup>+</sup> T cells from transgenic mice expressing a TCR specific for the unrelated OT-1 peptide (Supplemental Figure 1B). Moreover, CD8<sup>+</sup> T cells that bound to the Qa-1-Hsp60<sub>p216</sub> tetramer do not bind to the Qa-1-Qdm tetramer and vice versa (Figure 1B).

This tetramer-based analysis revealed that expansion of Qa-1-Hsp60<sub>p216</sub>-specific CD8<sup>+</sup> T cells as detected by Qa-1-Hsp60<sub>p216</sub>



**Figure 2** Infusion of CD122<sup>+</sup>CD44<sup>hi</sup>Ly49<sup>+</sup> CD8<sup>+</sup> Tregs inhibits arthritis development. **(A)** Purified CD25<sup>-</sup> CD4<sup>+</sup> cells ( $1 \times 10^6$ ) and B cells ( $2 \times 10^6$ ) from arthritic mice were cotransferred into *Rag2*<sup>-/-</sup>*Prf1*<sup>-/-</sup> mice at day 0 with or without CD122<sup>+</sup>CD44<sup>hi</sup>Ly49<sup>+</sup> or Ly49<sup>-</sup> CD8<sup>+</sup> cells ( $1.5 \times 10^5$ ) FACS-sorted from cCII-immunized B6 mice. Mice were immunized at day 0 and boosted at days 21 and 35 (black arrows) with cCII. Arthritis scores are shown for 5 mice/group. \**P* < 0.05 = group (none) versus group (Ly49<sup>+</sup> CD8). Representative data from 5 independent experiments are shown. **(B)** Adoptive transfer of B6.WT or B6.D227K CD4<sup>+</sup> cells and WT B cells with or without Ly49<sup>+</sup> CD8<sup>+</sup> cells or Ly49<sup>-</sup> CD8<sup>+</sup> cells. Arthritis was induced as in **A**. Serum titers of anti-mouse CII IgG were measured at day 45. **(C)** Adoptive transfer of B6.WT CD4<sup>+</sup> cells and B cells with or without CD8<sup>+</sup> Treg or CD44<sup>hi</sup> CD8<sup>+</sup> cells ( $1.5 \times 10^5$ ) from *B6.IL15*<sup>+/-</sup> or *B6.IL15*<sup>-/-</sup> mice. Arthritis was induced as in **A** and scores are shown. **(D)** Adoptive transfer of CD4<sup>+</sup> cells and B cells with  $1.5 \times 10^5$  CD122<sup>+</sup>CD44<sup>hi</sup>Ly49<sup>+</sup> or CD122<sup>+</sup>CD44<sup>hi</sup>Ly49<sup>-</sup> CD8<sup>+</sup> cells or CD8<sup>+</sup> cells from cCII-immunized WT or *Prf1*<sup>-/-</sup> mice as described in **A**. Serum titers of anti-mouse CII IgG were measured at day 45. \**P* < 0.05; \*\*\**P* < 0.001.

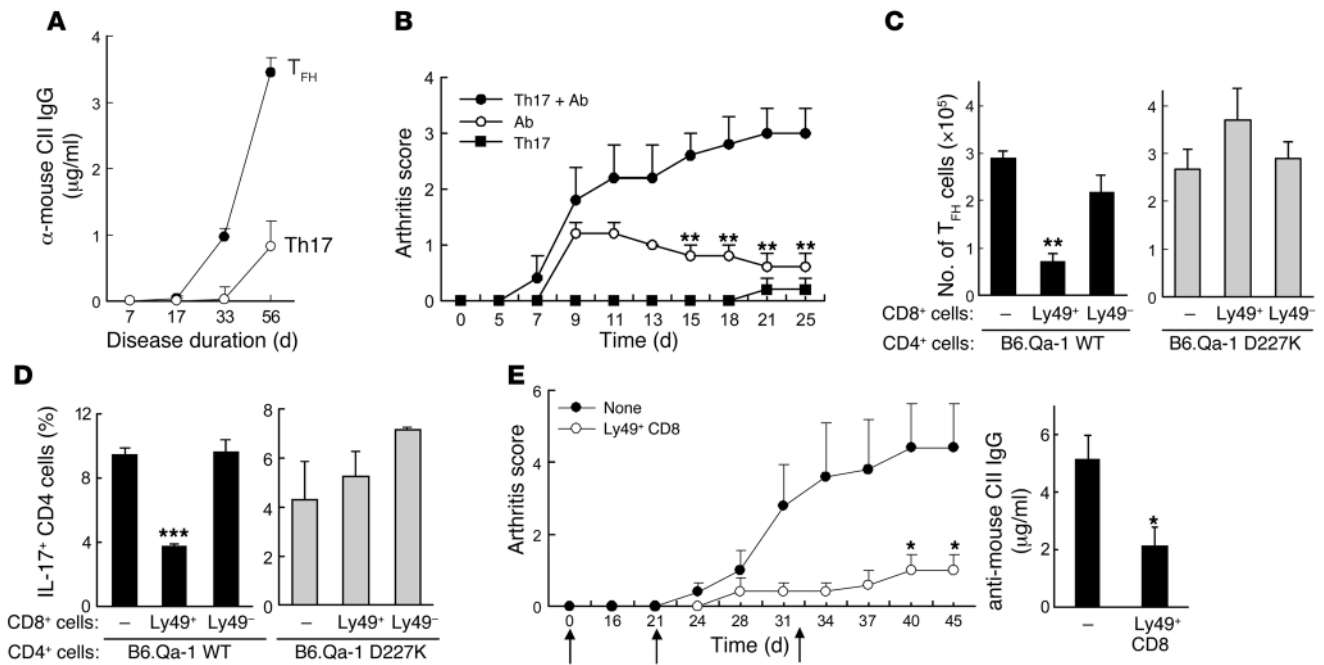
tetramers after immunization with Hsp60<sub>p216</sub>-pulsed DCs inhibited the development of CIA (Figure 1, C and D). Although immunization with Qdm-pulsed *Kb*<sup>-/-</sup>*Db*<sup>-/-</sup> DCs resulted in increased numbers of Qa-1 R72A-Qdm tet<sup>+</sup> cells that include both CD122<sup>+</sup>Ly49<sup>+</sup> and CD122<sup>+</sup>Ly49<sup>-</sup> populations (ref. 16, Figure 1C, and Supplemental Figure 1, C and D), this immunization did not inhibit disease development (Figure 1A). Although T cells specific for several other Hsp60 epitopes have been detected in juvenile idiopathic arthritis patients along with expansion of CD4<sup>+</sup> Tregs (17), Hsp60<sub>p216</sub> immunization did not change the frequency of Foxp3<sup>+</sup>CD4<sup>+</sup> cells (Supplemental Figure 1E). We also noted that Hsp60<sub>p216</sub> immunization resulted in expansion of Hsp60<sub>p216</sub>-tet<sup>+</sup> CD8<sup>+</sup> cells within the CD122<sup>+</sup>Ly49<sup>+</sup> (16) but not the CD122<sup>+</sup>Ly49<sup>-</sup> CD8 subset (Figure 1D). These results suggest that Hsp60<sub>p216</sub>-Qa-1-restricted CD8<sup>+</sup> cells belonging to the CD122<sup>+</sup>Ly49<sup>+</sup>CD8<sup>+</sup> subset may inhibit autoimmune arthritis.

**Infusion of CD8<sup>+</sup> Tregs inhibits arthritis development.** The above findings indicated that expansion of Hsp60<sub>p216</sub>-specific Qa-1-restricted CD8<sup>+</sup> cells (but not control Qa-1-restricted peptide) was associated with disease inhibition. Infusion of the fewer than 5% of CD8<sup>+</sup> T cells expressing the triad of surface receptors that characterize

CD8<sup>+</sup> Tregs (CD122<sup>+</sup>CD44<sup>+</sup>Ly49<sup>+</sup>; triad<sup>+</sup>) into adoptive hosts that had been given CD25<sup>-</sup>CD4<sup>+</sup> T cells and B cells from chicken type II collagen-immune (cCII-immune) donors abolished disease progression (Figure 2A) and reduced titers of anti-CII Ab (Figure 2B). In contrast, infusion of CD122<sup>+</sup>CD44<sup>+</sup>Ly49<sup>-</sup> CD8<sup>+</sup> cells failed to inhibit disease progression. The inhibitory activity of Ly49<sup>+</sup>CD8<sup>+</sup> T cells depended on recognition of Qa-1 expressed by CD4<sup>+</sup> Th cells, since CD4<sup>+</sup> Th cells expressing a Qa-1 point mutation (Qa-1 D227K) that impaired recognition by CD8<sup>+</sup> cells (5, 18) were resistant to CD8<sup>+</sup> Treg-dependent inhibition (Figure 2B).

**CD8<sup>+</sup> Treg activity depends on perforin and IL-15 expression.** To further investigate the mechanism of suppression by Ly49<sup>+</sup>CD8<sup>+</sup> Tregs, we asked whether suppression by the Ly49<sup>+</sup>CD8<sup>+</sup> cells depended on IL-15. Transfer of CD8<sup>+</sup> cells from *IL-15*<sup>-/-</sup> donors failed to inhibit disease severity; indeed, an increase in disease intensity was noted (Figure 2C). Suppression also reflected perforin-dependent elimination, since purified *Prf1*-deficient CD8<sup>+</sup> T cells did not reduce anti-mouse collagen Ab responses (Figure 2D). In sum, IL-15 was essential to the development of CD122<sup>+</sup>CD44<sup>+</sup>Ly49<sup>+</sup> CD8<sup>+</sup> Tregs, and intracellular perforin was required for expression of suppressive activity.

**CD8<sup>+</sup> Tregs inhibit arthritis through Qa-1-dependent elimination of pathogenic T<sub>FH</sub> and Th17 cells.** We then investigated the cellular targets of CD8<sup>+</sup> Tregs. Expansion of collagen-reactive CD4<sup>+</sup> T cells is associated with the development of CIA. Examination of effector CD4<sup>+</sup> T cells during the course of CIA revealed that both CD4<sup>+</sup> T<sub>FH</sub> (ICOS<sup>+</sup>CXCR5<sup>+</sup> or ICOS<sup>+</sup>BTLA<sup>+</sup>) and CD4<sup>+</sup>IL-17<sup>+</sup> cells in all LNs (not only LNs draining the site of cCII immunization) increased 10- to 20-fold during disease progression (Supplemental Figure 2, A and B), while IFN-γ<sup>+</sup>CD4<sup>+</sup> cells were increased to a lesser extent (~5-fold) (Supplemental Figure 2). Analysis of cellular infiltrates in arthritic joints revealed that approximately 50% of CD4<sup>+</sup> cells expressed the ICOS and CXCR5 pair, characteristic of T<sub>FH</sub> cells (Supplemental Figure 2). Th17 cells were also detected in inflamed joints, albeit at lower frequencies (Supplemental Figure 2C). Increased numbers of T<sub>FH</sub> cells were associated with expansion of GC B cells and increased titers of IgG anti-CII autoantibody, and adoptive transfer of T<sub>FH</sub> (but not Th17) cells along with B cells induced robust anti-CII autoantibody production (Figure 3A and Supplemental Figure 2). Although transfer of large numbers of T<sub>FH</sub> or Th17 cells did not induce disease in adoptive *Rag2*<sup>-/-</sup>*Prf1*<sup>-/-</sup> hosts, transfer of both Th subsets induced strong disease (19).



**Figure 3**

Inhibition of arthritis by CD8<sup>+</sup> Tregs is associated with targeting of pathogenic T<sub>FH</sub> and Th17 cells. (A) T<sub>FH</sub> cells (ICOS<sup>+</sup>CXCR5<sup>+</sup>BTLA<sup>+</sup> CD4<sup>+</sup> cells) directly from cCII-immune donors and cCII-specific Th17 cells differentiated in vitro were transferred along with B cells into *Rag2*<sup>-/-</sup>*Prf1*<sup>-/-</sup> hosts at day 0. Arthritis was induced as described in Methods. Serum titers of anti-mouse CII IgG at the indicated time points are shown. (B) Anti-collagen II Ab (1 mg) was injected intraperitoneally into *Rag2*<sup>-/-</sup>*Prf1*<sup>-/-</sup> hosts with or without cotransfer of in vitro-generated cCII-specific Th17 cells (2 × 10<sup>6</sup>) before all recipients were immunized with cCII. Arthritis scores are shown for 5 mice/group. \*\**P* < 0.01, group (Ab alone) versus group (Th17 + Ab). Data are representative of 5 independent experiments. (C and D) Adoptive transfer of B6.WT or B6.D227K CD4<sup>+</sup> cells and WT B cells with or without Ly49<sup>+</sup>CD8<sup>+</sup> cells or Ly49<sup>-</sup>CD8<sup>+</sup> cells as in Figure 2A. The effect on splenic T<sub>FH</sub> cells (C) or percentage of IL-17<sup>+</sup> CD4<sup>+</sup> cells (D) at day 45 is shown. (E) ICOS<sup>+</sup>CXCR5<sup>+</sup>BTLA<sup>+</sup>CD25<sup>-</sup> (T<sub>FH</sub>) CD4<sup>+</sup> cells (2 × 10<sup>6</sup>) sorted from arthritic mice (score = 12) and cCII-specific Th17 cells generated as in A (2 × 10<sup>6</sup>) were cotransferred with B cells (4 × 10<sup>6</sup>) into *Rag2*<sup>-/-</sup>*Prf1*<sup>-/-</sup> hosts with or without CD122<sup>+</sup>CD44<sup>+</sup>Ly49<sup>+</sup> CD8<sup>+</sup> cells (1.5 × 10<sup>6</sup>) followed by arthritis induction by cCII injections on days 0, 21, and 32 (black arrows). Arthritis scores are shown on left. On right, serum anti-mouse CII IgG titers at day 45 are shown. \**P* < 0.05; \*\**P* < 0.01; \*\*\**P* < 0.001.

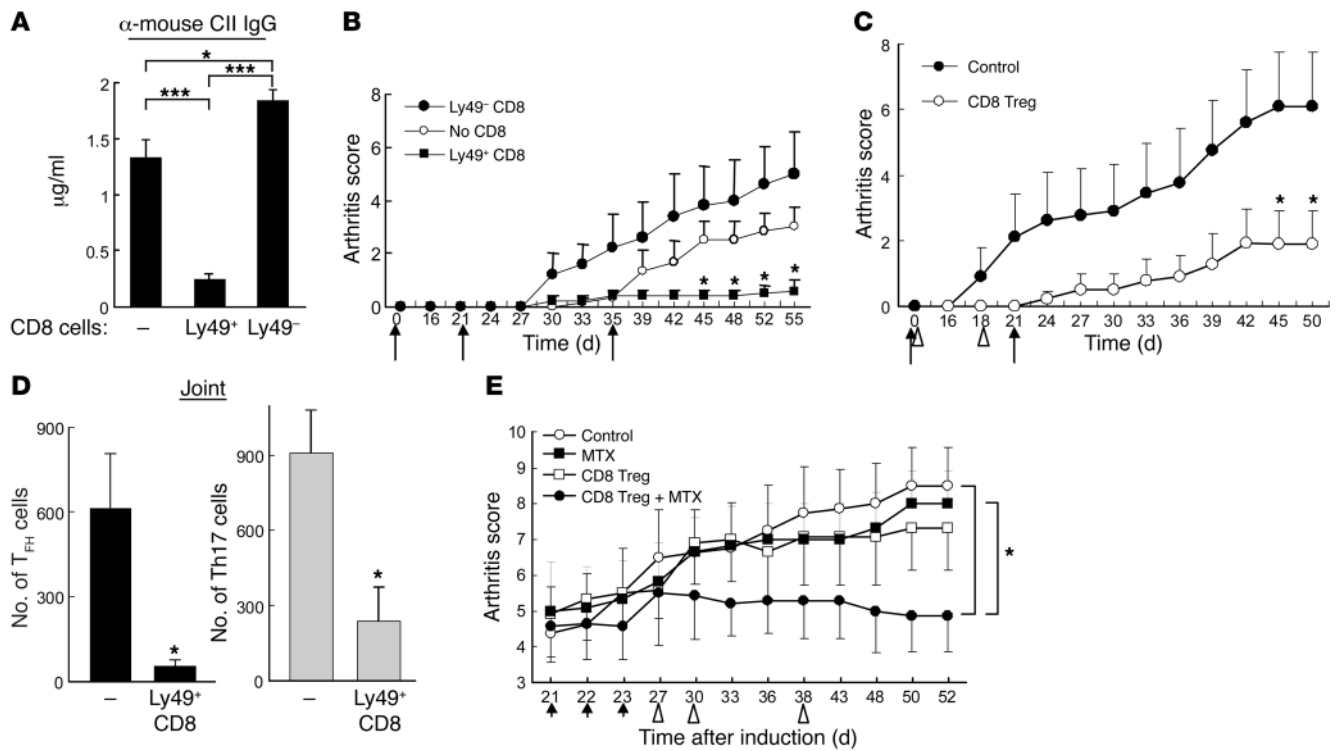
We tested the hypothesis that disease might reflect a T<sub>FH</sub>-induced autoantibody response combined with a Th17-dependent inflammatory reaction. Indeed, mice given Th17 cells and autoantibody (mainly anti-CII Ab) in place of T<sub>FH</sub> cells developed severe and sustained arthritis (Figure 3B). These data suggest that CIA initiation in intact animals may reflect a synergistic interaction in which a Th17-mediated inflammatory response enhances a T<sub>FH</sub>-dependent autoantibody production.

These results also suggested that inhibition of the T<sub>FH</sub> and Th17 cell response might interrupt early events in the autoimmune disease process. Indeed, reduced arthritis and autoantibody titers in recipients of CD8<sup>+</sup> Tregs were associated with diminished numbers of T<sub>FH</sub> and Th17 cells (Figure 3, C and D), but not IFN-γ<sup>+</sup> (Th1) cells in adoptive hosts (Supplemental Figure 3). Elimination of T<sub>FH</sub> and Th17 cells required perforin expression by Ly49<sup>+</sup>CD8<sup>+</sup> Tregs (Supplemental Figure 4) and depended on Qa-1 expression by CD4<sup>+</sup> Th target cells, since CD4<sup>+</sup> Th cells expressing a Qa-1 point mutation (Qa-1 D227K) that impaired recognition by CD8<sup>+</sup> cells (5, 18) were resistant to elimination (Figure 3, C and D).

To further define the mechanism of Treg-dependent inhibition of arthritis, we transferred Ly49<sup>+</sup>CD8<sup>+</sup> Tregs into *Rag2*<sup>-/-</sup>*Prf1*<sup>-/-</sup> hosts reconstituted with isolated T<sub>FH</sub> cells (ICOS<sup>+</sup>CXCR5<sup>+</sup>BTLA<sup>+</sup>) and Th17 cells. Cotransfer of T<sub>FH</sub> and Th17 cells induced clinical arthritis; infusion of Ly49<sup>+</sup>CD8<sup>+</sup> Tregs diminished disease progres-

sion and significantly reduced anti-CII Ab titers (Figure 3E), supporting the conclusions drawn from experiments using unseparated CD4<sup>+</sup> cells (Figure 3, C and D).

*Transfer of in vitro IL-15-expanded CD8<sup>+</sup> Tregs inhibits CIA.* We further defined the contribution of IL-15 to the suppressive activity of CD8<sup>+</sup> Tregs in experiments that analyzed the effects of incubation of CD8<sup>+</sup> Tregs with low concentrations of IL-15/IL-15Rα complexes (IL-15C) (20, 21). Ly49<sup>+</sup> CD8<sup>+</sup> T cells incubated in vitro for 1 week in the presence of 10 ng ml<sup>-1</sup> IL-15C maintained their surface phenotype (Supplemental Figure 5) after vigorous expansion: approximately 3 × 10<sup>6</sup> CD122<sup>+</sup>CD44<sup>+</sup>Ly49<sup>+</sup> CD8<sup>+</sup> cells were generated within 7 days from an initial seeding of 10<sup>5</sup> CD8<sup>+</sup> cells that expressed the same triad of surface receptors. Moreover, these in vitro-expanded CD8<sup>+</sup> Tregs abolished anti-mouse collagen Ab responses (Figure 4A) and efficiently inhibited disease development in *Rag2*<sup>-/-</sup>*Prf1*<sup>-/-</sup> hosts given cCII-immune CD4<sup>+</sup> T cells (Figure 4B). The inhibitory activity of IL-15C-incubated CD8<sup>+</sup> Tregs was at least as potent as that of nonincubated CD8<sup>+</sup> Tregs (Figure 4B compared with Figure 2A), suggesting that infusion of in vitro IL-15-expanded CD8<sup>+</sup> Tregs may represent an effective therapeutic approach to CIA. Moreover, transfer of in vitro-expanded CD8<sup>+</sup> Tregs into unirradiated C57BL/6 (B6) mice followed by immunization with cCII suppressed arthritis progression (Figure 4C and Supplemental Figure 6A) and reduced T<sub>FH</sub> and Th17 cell numbers (Figure 4D and Supplemental Figure 6B) in



**Figure 4**

Transfer of in vitro IL-15C–expanded CD8<sup>+</sup> Tregs inhibits CIA. (A) CD122<sup>+</sup>CD44<sup>hi</sup>Ly49<sup>+</sup> or CD122<sup>+</sup>CD44<sup>hi</sup>Ly49<sup>-</sup> CD8<sup>+</sup> cells sorted from cCII-immunized B6 mice were incubated in 10 ng ml<sup>-1</sup> IL-15C × 1 week. Purified CD25<sup>-</sup> CD4<sup>+</sup> and B cells from arthritic donors were transferred with or without the indicated CD8<sup>+</sup> cells (0.6 × 10<sup>5</sup>) into Rag2<sup>-/-</sup>Prf1<sup>-/-</sup> mice followed by arthritis induction at day 0 and boost at days 21 and 35 (black arrows). Serum titers of anti-mouse CII IgG are shown. \*\*\*P < 0.001. (B) Arthritis scores are shown for 5–6 mice/group. Group (no CD8) vs. group (Ly49<sup>+</sup> CD8), \*P < 0.05. Data represent 3 independent experiments. (C) In vitro–expanded CD122<sup>+</sup>CD44<sup>hi</sup>Ly49<sup>+</sup> CD8<sup>+</sup> cells (2 × 10<sup>5</sup>) were transferred into B6 mice at days 0 and 18 (triangles). cCII immunization and boosting at days 0 and 21 (black arrows) and arthritis scores over 50 days are shown. (D) Numbers of T<sub>FH</sub> and IL-17–expressing CD4<sup>+</sup> cells from inflamed joints in C are shown for 5–6 mice/group. (E) 0.75 mg/kg MTX was injected into B6 mice from day 21 to day 23 (arrows) after development of arthritis (average score of 4.5). In vitro IL-15C–expanded CD122<sup>+</sup>CD44<sup>hi</sup>Ly49<sup>+</sup> CD8<sup>+</sup> cells (2.5 × 10<sup>5</sup>) were transferred into mice at days 27, 30, and 38 (triangles). Arthritis scores are shown for 5–8 mice/group. Group (control) or group (MTX) versus group (CD8<sup>+</sup> Treg + MTX) difference: \*P < 0.05.

both spleen (data not shown) and inflamed joints, reflecting a broad elimination of potential effector Th activity.

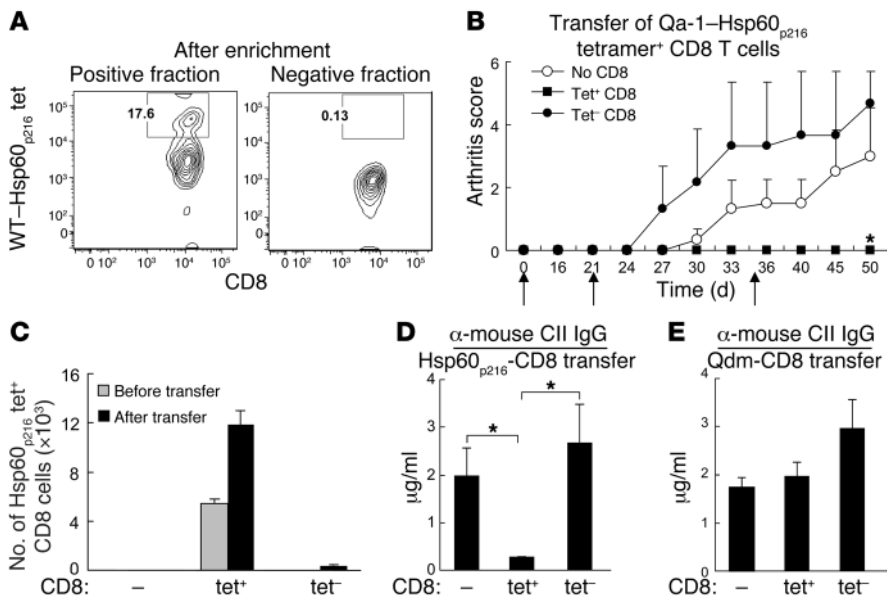
*Combination of CD8<sup>+</sup> Tregs and MTX exerts therapeutic effects on ongoing disease.* MTX, the disease-modifying anti-rheumatic drug (22, 23), is thought to prevent arthritis through inhibition of inflammation and joint destruction at the level of synoviocytes (24). In contrast, CD8<sup>+</sup> Tregs interrupt the autoimmune pathway much earlier (e.g., see Figure 2). We therefore studied the combined effects of an MTX pulse to block joint inflammation followed by infusion of CD8<sup>+</sup> Tregs at later time points to prevent new damage inflicted by a continuing systemic autoimmune response. Attenuation of disease activity was noted using an MTX pulse of 1.5 mg/kg over a short time period (from day 21 to day 23) (Supplemental Figure 7, A and B), while doses lower than 0.75 mg/kg had minimal effects on disease progression (Supplemental Figure 7, A–C). Coadministration of MTX at 0.75 mg/kg with CD8<sup>+</sup> Tregs resulted in a marked reduction in disease severity (Figure 4E). The synergistic therapeutic effect of this combination therapy may depend, in part, on the failure of low doses of MTX to diminish the numbers of CD122<sup>+</sup>Ly49<sup>+</sup> CD8<sup>+</sup> cells (Supplemental Figure 7 and ref. 16). Together, these results suggest the potential contribution of therapy using CD8<sup>+</sup> Tregs and MTX to ameliorate ongoing disease.

*Transfer of CD8<sup>+</sup> Tregs specific for Qa-1–Hsp60<sub>p216</sub> inhibits arthritis.*

The IL-15 dependence of CD8<sup>+</sup> Tregs allowed us to directly test the hypothesis raised by the results of Figure 1: Hsp60<sub>p216</sub>–Qa-1–restricted CD8<sup>+</sup> cells mediate CD8<sup>+</sup> Treg activity. We separated Qa-1–Hsp60<sub>p216</sub>–tet<sup>+</sup> CD8<sup>+</sup> cells from the tet<sup>-</sup> fraction by consecutive FACS sorting and microbead selection (Figure 5A). After in vitro expansion of enriched tet<sup>-</sup> CD8<sup>+</sup> cells by IL-15C, adoptive transfer of small numbers (5 × 10<sup>4</sup>/mouse) of Qa-1–Hsp60<sub>p216</sub>–tet<sup>+</sup> CD8<sup>+</sup> cells, but not Qa-1 R72A–Qdm–tet<sup>+</sup> CD8<sup>+</sup> cells, into Rag2<sup>-/-</sup>Prf1<sup>-/-</sup> hosts along with cCII-immune CD4 and B cells inhibited autoantibody production and halted arthritis progression (Figure 5, B–E). Analysis of CD8<sup>+</sup> cells in these adoptive hosts revealed expansion of Qa-1–Hsp60<sub>p216</sub>–tet<sup>+</sup> CD8<sup>+</sup> cells after transfer of tet<sup>+</sup> but not tet<sup>-</sup> CD8<sup>+</sup> cells (Figure 5C). Moreover, transfer of Qa-1 R72A–Qdm tet<sup>+</sup> CD8<sup>+</sup> cells did not inhibit autoantibody production (Figure 5E), consistent with the failure of Qdm immunization to suppress CIA (Figure 1).

**Discussion**

Clinical approaches to the treatment of arthritis have focused mainly on downstream elements of this disease process, while sparing the initiating autoimmune events including induction



**Figure 5** Transfer of Qa-1-Hsp60<sub>p216</sub> restricted CD8<sup>+</sup> Treg inhibits arthritis. (A) Hsp60<sub>p216</sub>-tet<sup>+</sup> and Hsp60<sub>p216</sub>-tet<sup>-</sup> CD8<sup>+</sup> cells were sorted from cCII-immune B6 mice that had been immunized 2 weeks earlier with *Kb<sup>-/-</sup>Db<sup>-/-</sup>* DCs loaded with Hsp60<sub>p216</sub> before incubation in IL-15C (10 ng ml<sup>-1</sup>) × 10 days. After incubation with Hsp60<sub>p216</sub> tetramer and enrichment by anti-PE microbeads, they were subjected to FACS analysis. Incubation of CD8<sup>+</sup> cells that were initially tet<sup>+</sup> resulted in a substantial increase and enrichment of tet<sup>+</sup> CD8<sup>+</sup> cells; identical incubation of CD8<sup>+</sup> cells that were tet<sup>-</sup> did not result in significant levels of tet<sup>+</sup> cells after incubation in IL-15C. Hsp60<sub>p216</sub>-tet<sup>+</sup> or Hsp60<sub>p216</sub>-tet<sup>-</sup> fraction of CD8<sup>+</sup> cells was transferred into *Rag2<sup>+/-</sup>Prf1<sup>+/-</sup>* mice at day 0 along with CD4 and B cells from arthritic mice. Mice were immunized at day 0 and boosted at days 21 and 35 (black arrows) with cCII. Arthritis scores (B), numbers of splenic Hsp60<sub>p216</sub>-tet<sup>+</sup> CD8<sup>+</sup> cells at day 34 (C), and anti-mouse CII IgG titers at day 30 (D) after adoptive transfer are shown for 3–5 mice per group. Group (tet<sup>-</sup> CD8) versus group (tet<sup>+</sup> CD8), \**P* < 0.05. (E) By comparison, transfer of Qdm-tet<sup>+</sup> (or tet<sup>-</sup>) CD8<sup>+</sup> cells had no significant effect on the response of CD4<sup>+</sup> and B cells from arthritic mice using the same conditions (D).

of autoantibodies and inflammatory responses by autoimmune Th cells. Here, we describe a strategy that depends on targeting of pathogenic T<sub>FH</sub> and Th17 cells by CD8<sup>+</sup> Tregs.

CD8<sup>+</sup> Treg treatment was particularly effective at blunting the autoantibody response in CIA (e.g., Figures 2 and 3). Since CD8<sup>+</sup> Tregs target T<sub>FH</sub> cells and Th17 cells, the major Th subsets that induce the humoral and inflammatory components of CIA, we asked whether infusion of CD8<sup>+</sup> Tregs might therefore confer durable inhibition of this autoimmune disorder. Transfer of CD122<sup>+</sup>CD44<sup>+</sup>Ly49<sup>+</sup> CD8<sup>+</sup> cells, which represents less than 5% of the total CD8<sup>+</sup> cell population and carries essentially all CD8<sup>+</sup> Treg activity (16), resulted in reduction of the numbers of T<sub>FH</sub> and Th17 cells and robust inhibition of disease. The finding that CD8<sup>+</sup> Tregs inhibit T<sub>FH</sub> and Th17 CD4<sup>+</sup> cells but do not target Th1 cells suggests direct targeting of Th subsets, consistent with previous findings (5). However, the possibility that activated DCs that upregulate the Qa-1-peptide complexes may also be targeted by CD8<sup>+</sup> Tregs cannot be excluded. Although definition of the precise target cells requires further analysis, the low frequency of CD8<sup>+</sup> Tregs found in arthritic synovia (<1% of total T cells) suggests that the inhibitory activity of CD8<sup>+</sup> Tregs is exerted within the systemic lymphoid organs rather than arthritic joints.

Although defective CD4<sup>+</sup> (Foxp3<sup>+</sup>) Treg activity has been reported in CIA (25–27), regulatory CD4<sup>+</sup> cells may not result in sustained disease inhibition (28, 29), consistent with our find-

ings (Supplemental Figure 8). Possibly, impairment of CD4<sup>+</sup> Treg activity in the context of an inflammatory milieu associated with CIA may limit this approach (30). Although the recently defined CXCR5<sup>+</sup> follicular CD4<sup>+</sup> Treg (T<sub>FR</sub>) can suppress GC responses (31, 32), whether this CD4<sup>+</sup> Treg subset can target T<sub>FH</sub> cell activity and reduce CIA remains to be determined.

Incubation of CD122<sup>+</sup>Ly49<sup>+</sup> CD8<sup>+</sup> cells with IL-15/IL-15R $\alpha$  complexes induced robust expansion of these cells, which retained potent disease inhibitory activity, possibly reflecting the ability of 2 canonical CD8<sup>+</sup> Treg surface receptors – Ly49F and CD122 – to reduce the threshold for IL-15 activation (33). Although the contribution of IL-15 to the development and regulation of autoimmune disease has not been clearly defined (34–38), low doses of IL-15 may allow selective expansion of CD8<sup>+</sup> Tregs in vivo. A precedent for such an approach is suggested by recent reports that low-dose IL-2 treatment selectively enhances CD4<sup>+</sup> Tregs in vivo and can suppress chronic graft-versus-host disease (GVHD) and HCV-related vasculitis after BM transplants (39–41).

We designed Qa-1 R72A-Qdm and Qa-1-Hsp60<sub>p216</sub> tetramers to detect and enrich CD8<sup>+</sup> T cells specific for these Qa-1-associated ligands. Although immunization of mice with DCs pulsed

with the relevant peptide resulted in selective expansion of CD8<sup>+</sup> T cells specific for either Qa-1-Qdm or Qa-1-Hsp60<sub>p216</sub>, only immunization with the latter peptide efficiently inhibited the development of CIA. Moreover, transfer of a small number of Qa-1-Hsp60<sub>p216</sub> tetramer<sup>+</sup> CD8<sup>+</sup> cells, but not Qa-1(R72A)-Qdm tetramer<sup>+</sup> CD8<sup>+</sup> cells, inhibited disease progression in adoptive hosts. The CD8<sup>+</sup> Treg-mediated suppression we observed following transfer of CD8<sup>+</sup> Tregs was selectively exerted on the response to self antigens, since immune responses to foreign proteins (e.g., NP-KLH) were spared (Supplemental Figure 9). Autoreactive T<sub>FH</sub> and Th17 subsets that are chronically activated by self antigen (e.g., type II collagen) may upregulate stress-related proteins, including Hsp60, and present peptides derived from them in association with Qa-1 to Qa-1-restricted CD8<sup>+</sup> T cells. Further studies are required for a better understanding of the functional difference between Qa-1-Hsp60<sub>p216</sub> and Qa-1-Qdm-specific CD8<sup>+</sup> T cells. Possibly, differential expansion of Ly49<sup>+</sup> CD8<sup>+</sup> T cells by the 2 self peptides may be relevant to their distinct levels of suppressive activity (Figure 1). Inhibitory signaling associated with engagement of the CD94/NKG2A receptor by the Qa-1-Qdm complex expressed by CD4<sup>+</sup> target cells may dampen TCR-dependent activation of CD8<sup>+</sup> Tregs (19, 42). The Hsp60-related peptides may also include the Hsp60 signal peptide (Hsp60sp) derived from the leader sequence of Hsp60<sub>p10-18</sub>, since a subset of CD8<sup>+</sup> Tregs that recognizes the



Qa-1-Hsp60sp complex suppresses autoimmunity (43). Further definition of this subset of CD8<sup>+</sup> Tregs requires successful construction of Qa-1-Hsp60sp tetramers.

Analysis of the activity of tetramer<sup>+</sup> CD8<sup>+</sup> Tregs depended in part on immunization using peptide-pulsed DCs. Since CD8<sup>+</sup> Treg are memory cells according to function and surface phenotype (CD122<sup>+</sup>CD44<sup>+</sup>Ly49<sup>+</sup>), they may also be expanded by antigen in the absence of cell-dependent costimulatory activity. For example, peptide-Qa-1-coated (HLA-E-coated) nanoparticles (NP) may represent an effective and simplified method for CD8<sup>+</sup> Treg expansion that does not require cellular therapy. Moreover, since both Qa-1 and HLA-E MHC class Ib genes exhibit a limited polymorphism, in contrast with the extensive polymorphisms of MHC class Ia molecules (e.g., HLA-E expression is confined to 2 alleles that differ at a single amino acid), peptide-HLA-E-based immunotherapy should not require extensive individualized peptide-MHC design.

In the present studies, expansion of CD8<sup>+</sup> cells specific for Hsp60<sub>p216</sub> (but not a control self peptide) was associated with efficient inhibition of autoimmune arthritis, a reduction of the T<sub>FH</sub>/Th17 pathogenic Th pair, and diminished autoantibody to mouse collagen. Whether autoreactive Th cells that induce other forms of autoimmunity also can be targeted by Hsp60<sub>p216</sub>-specific CD8<sup>+</sup> Tregs is under investigation. The development of therapeutic strategies based on specific expansion and activation of peptide-specific CD8<sup>+</sup> Tregs represents a new and potentially effective approach to the treatment of autoimmune disease.

## Methods

**Mice.** B6, B6.*Prf1*<sup>-/-</sup>, *Rag2*<sup>-/-</sup>*Prf1*<sup>-/-</sup>, *IL-15*<sup>-/-</sup>, *Kb*<sup>-/-</sup>*Db*<sup>-/-</sup> (Taconic Farms), OT-1 TCR transgenic (Jackson Laboratory), and B6.Qa-1 D227K mice (backcrossed for 11 generations) (18) were housed in pathogen-free conditions.

**CIA induction and assessment.** cCII (MD Bioscience) was dissolved in 0.01 M acetic acid at a concentration of 4 mg ml<sup>-1</sup> by stirring overnight at 4°C. All mice used were males between the age of 8 and 12 weeks. To induce CIA, B6 mice were injected intradermally (i.d.) at the base of the tail with 150 µg of cCII emulsified in CFA (supplemented with 4 mg ml<sup>-1</sup> *Mycobacterium tuberculosis*) and boosted at day 21 with 100 µg of cCII emulsified in IFA. A cocktail of 4 monoclonal Abs (1 mg; MD Bioscience) binding to cCII was used to induce arthritis, as shown in Figure 3B. For adoptive transfers, CD4<sup>+</sup> (CD25<sup>-</sup>CD4<sup>+</sup> Treg depleted) and B cells were purified as described previously (5) from arthritic mice and then intravenously injected into *Rag2*<sup>-/-</sup>*Prf1*<sup>-/-</sup> hosts along with FACS-sorted CD8<sup>+</sup> or CD4<sup>+</sup> Tregs with indicated surface phenotype before immunization at day 0 and boosting at days 21 and 35. Clinical assessment of CIA was performed every 2–3 days each week, and scoring was as follows: 0, normal; 1, mild swelling and/or erythema confined to the mid-foot or ankle joint; 2, moderate edematous swelling extending from the ankle to the metatarsal joints; and 3, pronounced swelling encompassing the ankle, foot, and digits. Each limb was graded, allowing a maximum score of 12 per mouse.

**Measurement of Abs against CII.** Serum levels of anti-mouse CII IgG were measured by ELISA after serum collection at the indicated days after initial cCII immunization. Briefly, 96-well ELISA microplates were coated with mouse CII (Chondrex) at 5 µg ml<sup>-1</sup> dissolved in dilution buffer (Chondrex) at 100 µl/well at 4°C overnight. 100 µl of diluted serum sample was incubated for 2 hours at room temperature. The plates were washed with PBST (0.05% Tween-20 in PBS) 5 times, followed by addition of peroxidase-conjugated goat anti-mouse IgG at 1:50,000 (Sigma-Aldrich) at 100 µl/well. After 1 hour incubation at room temperature and wash, the final color development was achieved by adding TMB

substrate (BD Bioscience) to each well at 100 µl/well, and absorbance was measured at 405 nm at the appropriate time.

**Flow cytometry.** Spleen and LNs were excised, and single-cell suspensions were prepared. Draining LNs (dLN) included popliteal and inguinal LNs; non-draining LNs (non-dLN) were axillary, cervical, and mesenteric LNs. For isolation of intraarticular cells, paw pieces were isolated and digested with collagenase/dispase (Roche) for 1 hour at 37°C followed by filtration to yield single-cell suspensions. Cells were incubated with Fc block for 15 minutes followed by staining with various Abs against surface markers or Qa-1 tetramers. CD45 marker was included to gate leukocytes from joints for further analysis. For intracellular cytokine staining, cells were restimulated with leukocyte activation cocktail (BD Bioscience) for 5 hours, stained with surface markers, fixed, and permeabilized, followed by incubation with indicated Abs. Cells were acquired on a FACSCanto II using FACSDiva software (BD Biosciences) and analyzed with FlowJo software (Tristar). Mouse-specific Abs to CD8α, TCR-β, CD44, CD45, CD62L, CD25, B220, Fas, CXCR5, IFN-γ, IL-17, ICOS, IgM, Ly49A<sup>B6</sup>, Ly49G2, Ly49F, Ly49C/I, and NK1.1 were purchased from BD Bioscience. Abs to Ly49C/I/F/H, CD122, BTLA, IL-21, NKG2A<sup>B6</sup>, and Foxp3 were purchased from eBioscience.

**In vitro differentiation of Th17 subsets and adoptive transfer.** Cells from spleen and LNs were collected from B6 mice immunized with 150 µg cCII/CFA, and CD4<sup>+</sup>CD25<sup>-</sup> cells were purified and enriched using negative selection. 2 × 10<sup>5</sup> ml<sup>-1</sup> CD4<sup>+</sup> cells were stimulated with 100 µg ml<sup>-1</sup> cCII in the presence of 2 × 10<sup>6</sup> irradiated total splenocytes. For differentiation of CD4<sup>+</sup> cells to Th17 cell phenotype, the following cytokine cocktail was added to cultures: 3 ng ml<sup>-1</sup> TGF-β, 20 ng ml<sup>-1</sup> rIL-6, 20 ng ml<sup>-1</sup> rIL-23, 10 µg ml<sup>-1</sup> anti-IL-12 Ab, 10 µg ml<sup>-1</sup> anti-IFN-γ Ab, 10 µg ml<sup>-1</sup> anti-IL-4 Ab. At day 5, live CD4<sup>+</sup> cells were harvested from cultures by Percoll gradient centrifugation and used for adoptive transfer. Confirmation of Th17 phenotype was performed using FACS analysis of IL-17 expression or using RT-PCR for detection of transcription factor *Rorc* (19).

**Generation of WT and R72A mutant Qa-1 tetramers.** A standard tetramer generation protocol was used, as described previously (44, 45). Briefly, an R72A mutant Qa-1 construct was generated using the QuikChange II Site-Directed Mutagenesis Kit (Stratagene) on a WT Qa-1 construct that contained biotin-binding sites for conjugation of monomers to form tetramers. Constructs were then used to transform Rosetta 2 (DE3) competent cells, and inclusion bodies were purified and stored at -80°C. For analysis of refolding capacity of each individual peptide, candidates were refolded with either WT or R72A mutant Qa-1 heavy chain in the presence of β<sub>2</sub>m. The refolding solution was then analyzed by size exclusion chromatography for a distinctly refolded Qa-1/β<sub>2</sub>m/peptide peak that was collected as a monomer and stored at -80°C before conjugation with streptavidin-phycoerythrin or streptavidin-allophycocyanin to form tetramers prior to each use.

**Enrichment for tetramer-positive cells.** Single-cell suspensions from spleen and LNs of cCII-immune B6 mice were incubated with anti-CD8α, anti-TCR-β and PE-labeled Qa-1 tetramers for 30 minutes at room temperature followed by sorting for tetramer-positive and tetramer-negative CD8α<sup>+</sup>TCRβ<sup>+</sup> cells on a BD FACSAria cell sorter (BD Bioscience). Sorted cells were incubated with IL-15C (10 ng ml<sup>-1</sup>) for 10 days before further enrichment of tetramer-positive cells as described (10). Briefly, cultured cells were washed and incubated with PE-labeled Qa-1 tetramers for 30 minutes at room temperature followed by washing and resuspension in sorter buffer. Anti-PE microbeads (Miltenyi Biotec) were added to each sample and incubated for 15 minutes at 4°C. Cells were washed, and PE-labeled cells were isolated by passage over an LS magnetic column (Miltenyi Biotec) followed by reanalysis of tetramer staining and adoptive transfer.

**Statistics.** Statistical analyses were performed using 2-tailed Student's *t* test or Mann-Whitney test for comparison of 2 conditions. Error bars denote mean ± SEM. A *P* value of < 0.05 was considered to be statistically significant.



**Study approval.** All experiments involving vertebrates were performed in compliance with federal laws and institutional guidelines and approved by the Dana-Farber Cancer Institute Animal Care and Use Committee.

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Address correspondence to: Harvey Cantor, Dana-Farber Cancer Institute, 450 Brookline Ave., Boston, Massachusetts 02215 USA. Phone: 617.632.3348; Fax: 617.632.4630; E-mail: Harvey\_Cantor@dfci.harvard.edu.

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