An integrative, multi-scale, genome -wide model reveals the phenotypic landscape of Escherichia coli

Javier Carrera, Raissa Estrela

演讲:孙隽 翻译:熊丹

Background

Materials and Methods

• Results

Discussion

Background

• The development of an integrative genome-scale model : Holy Grail

discovery of novel properties and emerging behaviors

generating and testing predictable hypotheses

• Potential

guiding experimentation

accelerating the in-depth understanding of cellular physiology

Early work

• E-cell

a modular software environment for whole-cell simulation that included organelle sub-models

More recently work

genome-scale simulations were performed to study complex phenomena

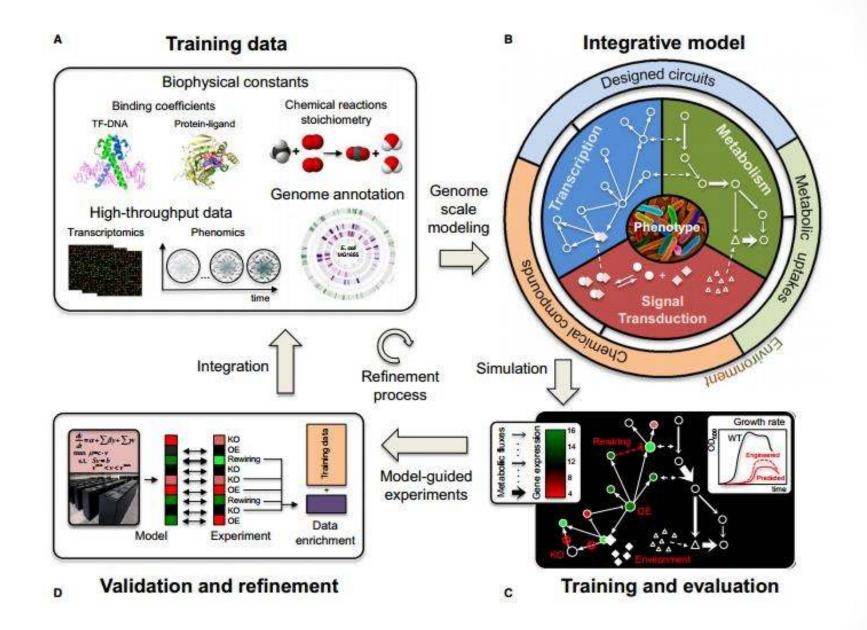
- the emergence of anticipatory behavior during evolution in varying environments
- the noise contributions of an inducible switch
- the effect of stochastic expression to metabolic variability

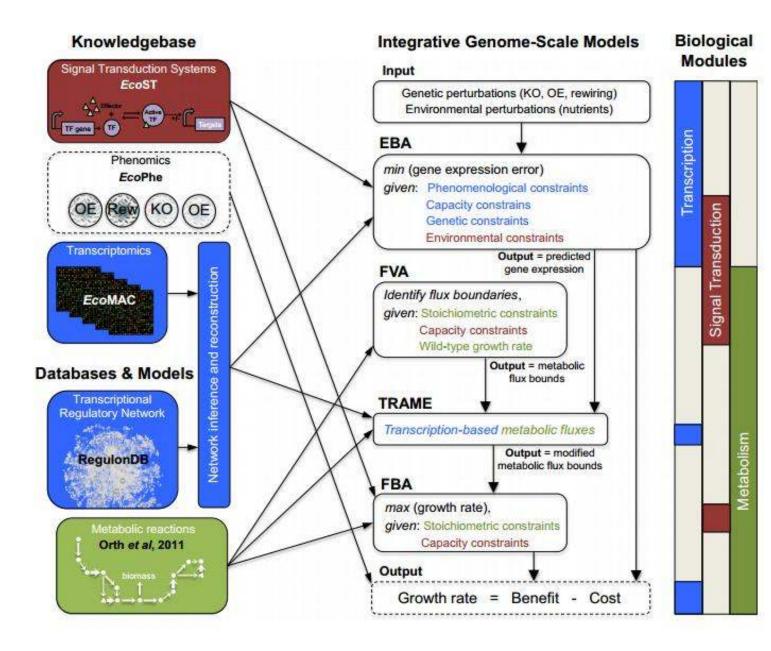
Our aim

To construct a phenomenological model for bacterial organisms that integrates multiple layers of biological organization.

Why E.coli

- The wealth of data and knowledge accumulated over the years
- The easiness to culture and manipulate experimentally
- Its importance in medical and biotechnological applications





Materials and Methods

• Data

• Cellular sub-models

• Model integration

Data

• gene expression

- ➢ from GEO, ASAP database
- constructed a gene expression compendium of 4,189 genes over 2,198 arrays that were collected from 127 scientific articles

signal transduction

- A total of 328 transcription factors (TFs) and 1,357 enzymes were identified by using RegulonDB
- ➢ identify 151 instances of signal transduction systems (STSs)

• Phenomics compendium

▷ bacterial growth information for 616 of the arrays in EcoMAC by EcoPhe

Cellular sub-models

• Signal transduction model

$$y_{\rm TF} = y_{\rm TF}^{\rm wt} + \Omega (C_{\rm TF}^{\rm max} - C_{\rm TF}^{\rm min}) \chi_{\rm TF}^E \frac{\Delta n_E}{\Delta n_E^{\rm max}}$$

• Transcriptional model and EBA

$$E = \frac{1}{2} \begin{bmatrix} \bar{y}_{\text{TF}} & \bar{\varepsilon}_{\text{TF}} \end{bmatrix} \bar{\bar{H}} \begin{bmatrix} \bar{y}_{\text{TF}} \\ \bar{\varepsilon}_{\text{TF}} \end{bmatrix} + \bar{f} \begin{bmatrix} \bar{y}_{\text{TF}} \\ \bar{\varepsilon}_{\text{TF}} \end{bmatrix}$$

• Metabolic model and Transcription-based Flux Enrichment

$$PV_{\min} \le v \le PV_{\max}$$
, where $P_e = \left(\frac{y_e}{y_e^{\operatorname{wt}}}\right)^n$

Model integration

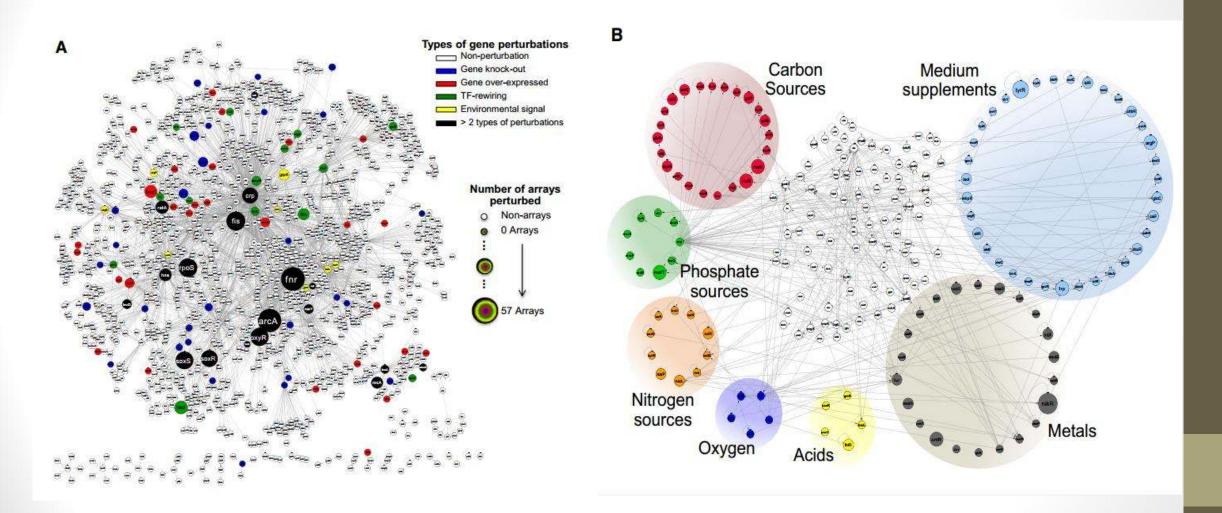
$$c = \frac{1}{N_G} \sum_{g} \left| \frac{\bar{y}_g - y_g^{\text{WT}}}{y_g^{\text{WT}}} \right|$$

$$\bar{\mu} = B - c$$

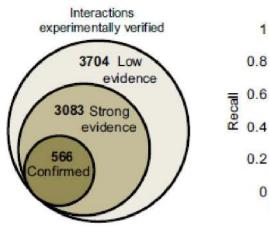
Results

- Genetic and environmental gene expression diversity
- An integrative knowledgebase as a base to regulatory network enrichment
- Expression Balance Analysis
- Phenotypic predictions in an integrated model
- Model enrichment through targeted experimentation

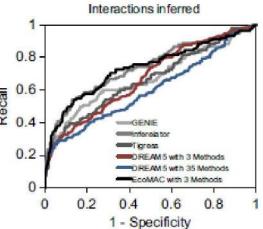
Genetic and environmental gene expression diversity

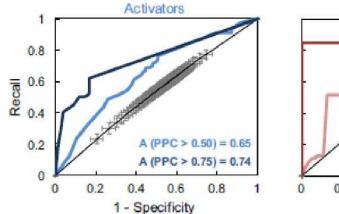


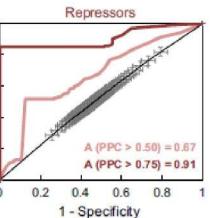
An integrative knowledgebase as a base to regulatory network enrichment

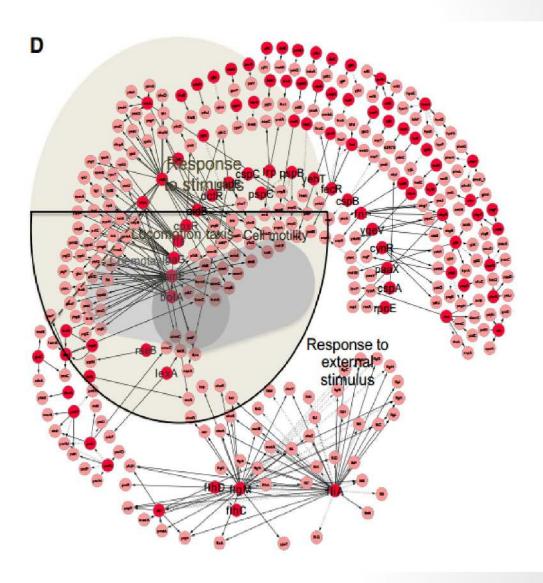


C

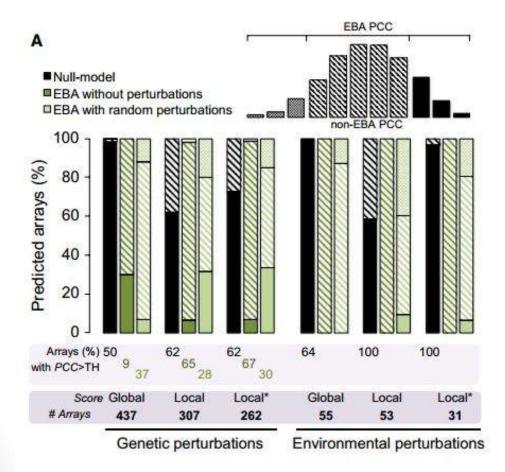


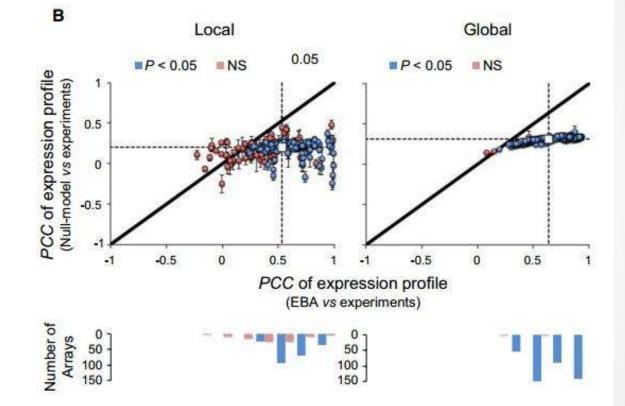




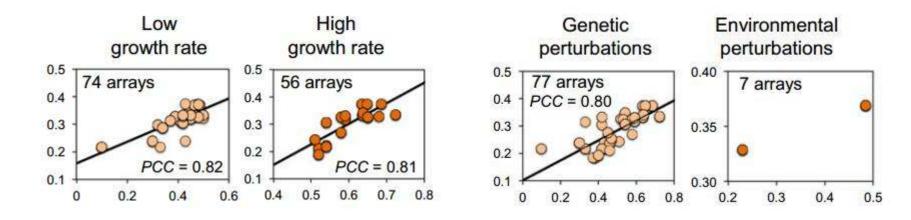


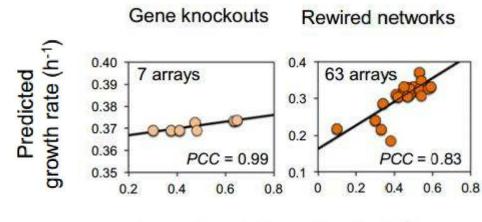
Expression Balance Analysis





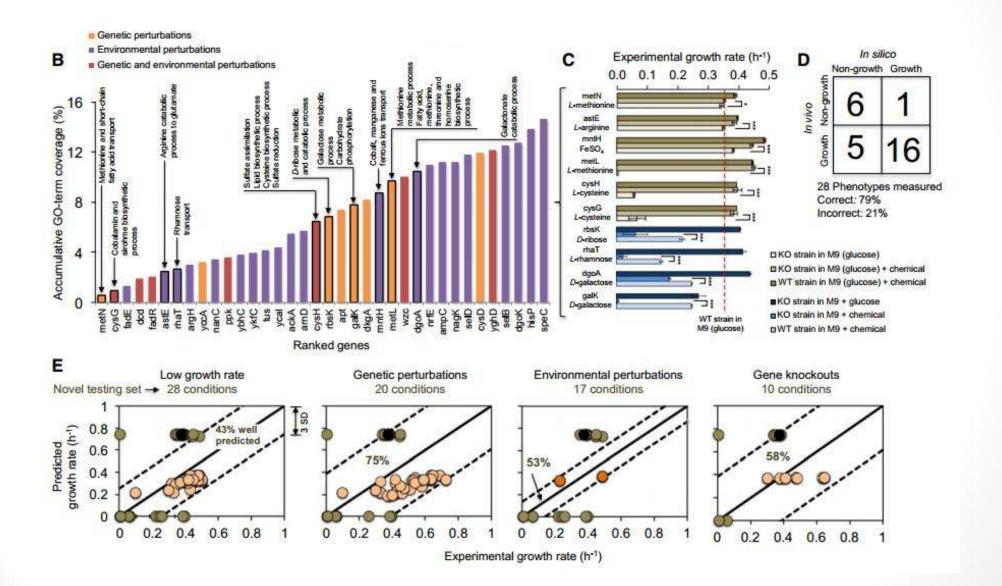
Phenotypic predictions in an integrated model





Experimental growth rate (h⁻¹)

Model enrichment through targeted experimentation



Discussion

Advantage

≻ the creation of a signal transduction network (EcoST)

> its integration to the transcriptional and metabolic network through constraint modeling

• Disadvantage

≻Coverage

> the severe bias to negative samples in the ground truth



• Binding site

• Funtion

Thanks for your attention