

# An Open-Label, Dose-Escalation Phase I Study of Anti-TYRP1 Monoclonal Antibody IMC-20D7S for Patients with Relapsed or Refractory Melanoma

Danny N. Khalil<sup>1</sup>, Michael A. Postow<sup>1</sup>, Nageatte Ibrahim<sup>2</sup>, Dale L. Ludwig<sup>3</sup>, Jan Cosaert<sup>4</sup>, Siva Rama Prasad Kambhampati<sup>3</sup>, Shande Tang<sup>3</sup>, Dmitri Grebennik<sup>5</sup>, John Sae Wook Kauh<sup>3</sup>, Heinz-Josef Lenz<sup>6</sup>, Keith T. Flaherty<sup>7</sup>, F. Stephen Hodi<sup>8</sup>, Donald P. Lawrence<sup>7</sup>, and Jedd D. Wolchok<sup>1</sup>

## Abstract

**Purpose:** Tyrosinase-related protein-1 (TYRP1) is a transmembrane glycoprotein that is specifically expressed in melanocytes and melanoma cells. Preclinical data suggest that mAbs targeting TYRP1 confer antimelanoma activity. IMC-20D7S is a recombinant human IgG1 mAb targeting TYRP1. Here, we report the first-in-human phase I/Ib trial of IMC-20D7S.

**Experimental Design:** The primary objective of this study was to establish the safety profile and the MTD of IMC-20D7S. Patients with advanced melanoma who progressed after or during at least one line of treatment or for whom standard therapy was not indicated enrolled in this standard 3 + 3 dose-escalation, open-label study. IMC-20D7S was administered intravenously every 2 or 3 weeks.

**Results:** Twenty-seven patients were enrolled. The most common adverse events were fatigue and constipation experi-

enced by nine (33%) and eight (30%) patients, respectively. There were no serious adverse events related to treatment, no discontinuations of treatment due to adverse events, and no treatment-related deaths. Given the absence of dose-limiting toxicities, an MTD was not defined, but a provisional MTD was established at the 20 mg/kg every 2-week dose based on serum concentration and safety data. One patient experienced a complete response. A disease control rate, defined as stable disease or better, of 41% was observed.

**Conclusion:** IMC-20D7S is well tolerated among patients with advanced melanoma with evidence of antitumor activity. Further investigation of this agent as monotherapy in selected patients or as part of combination regimens is warranted. *Clin Cancer Res*; 22(21); 5204–10. ©2016 AACR.

## Introduction

The incidence of melanoma in the United States has increased over the last 3 decades, with an estimated 76,100 new cases diagnosed in 2014 (1). Historically, treatment of unresectable melanoma has been challenging, as cytotoxic chemotherapy has failed to improve overall survival in this patient population. More recently, immunotherapy (2, 3) and small-molecule inhibitors

targeting BRAF and MEK (4, 5) have been shown to improve outcomes among patients with advanced melanoma. Nevertheless, many patients will either be refractory to such treatment or ultimately develop resistance to therapy and succumb to their disease. There remains a need to develop efficacious treatment options for this group of patients.

Tyrosinase-related protein-1 (TYRP1) is a transmembrane glycoprotein involved in melanin biosynthesis that is specifically expressed in melanocytes (6). Following protein translation, TYRP1 is trafficked from the endoplasmic reticulum through the Golgi apparatus to melanosomes; it is subsequently transferred to the melanocyte cell surface upon membrane fusion (7). TYRP1 is highly expressed in melanocytes and melanoma cells (8), and its expression is generally stable throughout melanoma progression (9). Given its expression pattern, TYRP1 is a promising and potentially safe therapeutic target for patients with melanoma.

The ability of therapeutic IgG1 mAbs to induce antibody-dependent cell-mediated cytotoxicity (ADCC) and complement-dependent cytotoxicity (CDC) on target cells has led to the successful development of multiple mAbs now in clinical use (10). Of note, successful targeting of cell-surface proteins that appear to be uninvolved in growth signaling (e.g., CD20 in B-cell lymphomas) highlights the importance of ADCC and CDC, as opposed to the inhibition of signaling pathways, in the anticancer activity of some therapeutic mAbs (11).

<sup>1</sup>Memorial Sloan Kettering Cancer Center, Ludwig Center for Cancer Immunotherapy, New York, New York. <sup>2</sup>Merck Research Laboratories, North Wales, Pennsylvania. <sup>3</sup>Eli Lilly and Company, New York, New York. <sup>4</sup>Sotio, Prague, Czech Republic. <sup>5</sup>Kyowa Hakko Kirin Pharma, Inc., Princeton, New Jersey. <sup>6</sup>Division of Medical Oncology, Norris Comprehensive Cancer Center, Keck School of Medicine, University of Southern California, Los Angeles, California. <sup>7</sup>Massachusetts General Hospital, Boston, Massachusetts. <sup>8</sup>Department of Medical Oncology, Dana-Farber Cancer Institute, Boston, Massachusetts.

**Note:** Supplementary data for this article are available at Clinical Cancer Research Online (<http://clincancerres.aacrjournals.org/>).

D.N. Khalil and M.A. Postow contributed equally to this article.

**Corresponding Author:** Jedd D. Wolchok, Memorial Sloan-Kettering Cancer Center, 1275 York Avenue, New York, NY 10065. Phone: 646-888-2315; Fax: 646-422-0453; E-mail: wolchokj@mskcc.org

doi: 10.1158/1078-0432.CCR-16-1241

©2016 American Association for Cancer Research.

### Translational Relevance

IMC-20D7S is an mAb targeting TYRP1 on melanoma cells. We find that IMC-20D7S is well tolerated among patients with advanced melanoma. Furthermore, there is evidence of antitumor activity, including a patient who achieved a complete response. In light of the possibility that it triggers an antitumor T-cell response via antibody-dependent cell-mediated phagocytosis, its efficacy may be augmented with immune checkpoint blockade. Given IMC-20D7S's safety profile and its mechanism of action, there is strong rationale for testing it in combination with checkpoint blockade therapies, such as anti-PD-1 and anti-CTLA-4 mAbs. Further investigation of IMC-20D7S in patients with melanoma is thus warranted.

IMC-20D7S is a recombinant human IgG1 mAb against TYRP1. Development of this clinical antibody is based on preclinical data showing that TA99, a murine IgG2a anti-TYRP1 mAb, localizes to subcutaneous melanoma xenografts (12) and inhibits syngeneic tumor growth in preclinical models (13). The antitumor effect was dependent on the intact antibody (7), the presence of Fc receptor (14), and natural killer (NK) cells (13), highlighting the importance of NK-mediated ADCC for this mAb.

Given the preclinical activity of TYRP1-directed mAb therapy, we conducted a phase I/Ib study of IMC-20D7S in patients with advanced melanoma. The primary objective of this study was to assess the safety of IMC-20D7S and establish an MTD. Secondary objectives were to describe the pharmacokinetic profile of IMC-20D7S, to recommend doses for subsequent clinical trials, to evaluate the immunogenicity of IMC-20D7S, and to assess progression-free survival (PFS).

## Materials and Methods

### Patient population

All enrolled patients were at least 18 years of age and had confirmed, unresectable stage III or IV melanoma with measurable disease as per RECIST 1.1. Patients who progressed after or during at least one line of treatment or for whom standard therapy was not indicated were enrolled. Other inclusion criteria included a life expectancy of at least 3 months, Eastern Cooperative Oncology Group performance status of 2 or better and adequate hematologic, renal, and hepatic function. Key exclusion criteria included ongoing grade 2 or worse side effects from prior radiation or chemotherapy, symptomatic brain or leptomeningeal disease, and ongoing immunosuppressive therapy, including steroid use. Patients were enrolled at three academic centers, and the protocol was approved by the Institutional Review Boards of the respective participating institutions. All patients provided written informed consent.

### Study design and treatment

This was an open-label, dose-escalation phase I/Ib study. IMC-20D7S injection for intravenous infusion was provided by Eli Lilly and Company. An initial dose of 5 mg/kg, administered over 60 minutes, was selected based on preclinical toxicology studies. This clinical study consisted of eval-

uating escalating doses of IMC-20D7S in two different schedules: an every 2-week schedule (Arm A) with a cycle composed of 4 weeks and an every 3-week schedule (Arm B) with a cycle composed of 6 weeks. After starting treatment for the first patient in the initial cohort (1A), a minimum of 7-day observation period elapsed until the next patient started treatment within this cohort. No waiting period was mandated in other cohorts, and no inpatient dose escalation was permitted.

This study was performed with a 3 + 3 dose-escalation study design. Within Arm A, planned dosing levels in the absence of dose-limiting toxicities (DLTs) were 5, 10, 20, and 30 mg/kg. Arm B (every 3-week dosing) was opened after the cohort receiving 10 mg/kg every 2 weeks was completed without any safety concerns. Planned dosing levels in Arm B were 10, 20, and 30 mg/kg. Patients were enrolled into both Arm A and Arm B in parallel. Cumulative DLTs across all dose levels in both arms were assessed on an ongoing basis, but dose escalation within each arm proceeded independently. Following completion of Arms A and B dose-escalating cohorts, a provisional MTD was to be defined. An expanded cohort was to be formed at the dose level defined as the MTD. At least six patients in total were to be treated at this dose level.

Patients in the dose-escalating cohorts were able to continue to receive IMC-20D7S in the absence of treatment failure, treatment intolerance, or consent withdrawal. Radiographic assessment of tumor response in both study arms was scheduled for every 6 weeks and evaluated as per RECIST v1.1. Additional imaging was performed if clinically indicated.

### Tolerability and safety

The incidence and severity of adverse events were graded according to the NCI Common Terminology Criteria for Adverse Events v4.02. Treatment-emergent adverse events (TEAEs) were defined as events that occurred or worsened after the first dose of study drug. Serious adverse events (SAEs) were defined as any untoward medical occurrence, at any dose, that were life threatening, resulted in death, significant incapacity, or congenital anomaly, or that required (or extended) hospitalization, intervention to prevent permanent impairment, or intervention to prevent one of the other listed serious outcomes. DLTs were defined as any grade 3 or above toxicity that emerged during study treatment and was clearly not attributable to melanoma or comedication and was possibly, probably, or definitely related to IMC-20D7S in the judgment of the investigator. If a patient experienced a DLT, the patient would not receive further IMC-20D7S.

### Pharmacokinetics and biomarker studies

In Arm A, serial blood samples were collected prior to infusion and up to 2 weeks (336 hours) following the first (cycle 1 day 1) and fifth infusions (cycle 3 day 1). In Arm B, blood samples were collected prior to and up to 3 weeks (504 hours) following the first infusion (cycle 1 day 1) and up to 2 weeks following the fifth infusion (cycle 3 day 1). Two blood samples (pre- and 1 hour post the end of infusion) were collected for the first infusion of cycles 2, 4, and subsequent cycles. Serum concentrations of IMC-20D7S were quantified using a nonvalidated ELISA using human gp75 protein as the capture antigen and peroxidase-conjugated anti-human IgG Fcγ

**Table 1.** Patient characteristics

Cohort	1A	2A	3A	4A	1B	2B	3B	All treatment groups
Dose (mg/kg)	5	10	20	30	10	20	30	
Schedule	q2w	q2w	q2w	q2w	q3w	q3w	q3w	
N	3	3	8	3	3	4	3	27
Gender, n (%)								
Male	1 (33.3)	2 (66.7)	5 (62.5)	2 (66.7)	2 (66.7)	3 (75.0)	1 (33.3)	16 (59.3)
Female	2 (66.7)	1 (33.3)	3 (37.5)	1 (33.3)	1 (33.3)	1 (25.0)	2 (66.7)	11 (40.7)
Race, n (%)								
White	2 (66.7)	2 (66.7)	8 (100.0)	3 (100.0)	3 (100.0)	4 (100.0)	3 (100.0)	25 (92.6)
Black/AA	0	1 (33.3)	0	0	0	0	0	1 (3.7)
Asian	1 (33.3)	0	0	0	0	0	0	1 (3.7)
Ethnicity, n (%)								
H/L	0	0	1 (12.5)	0	0	0	0	1 (3.7)
Not H/L	3 (100.0)	3 (100.0)	7 (87.5)	3 (100.0)	3 (100.0)	4 (100.0)	3 (100.0)	26 (96.3)
Age (yrs)								
N	3	3	8	3	3	4	3	27
Mean	77.7	64.9	69.8	61.1	66.6	69.1	65.4	68.2
SD	5.5	4.92	8.37	11.7	4.41	5.53	18.9	9.26
Median	75	67.2	71	65.7	64.6	67.9	72.1	68.6
Min	74.1	59.2	58.3	47.8	63.6	63.8	44.1	44.1
Max	84	68.2	82.7	69.7	71.6	76.8	80.1	84
Age group, n (%)								
18 to <65	0	1 (33.3)	2 (25.0)	1 (33.3)	2 (66.7)	1 (25.0)	1 (33.3)	8 (29.6)
≥65	3 (100.0)	2 (66.7)	6 (75.0)	2 (66.7)	1 (33.3)	3 (75.0)	2 (66.7)	19 (70.4)
ECOG, n (%)								
0	3 (100.0)	1 (33.3)	5 (62.5)	0	2 (66.7)	2 (50.0)	2 (66.7)	15 (55.6)
1	0	2 (66.7)	3 (37.5)	3 (100.0)	0	2 (50.0)	1 (33.3)	11 (40.7)
≥2	0	0	0	0	1 (33.3)	0	0	1 (3.7)
Prior therapy								
Systemic	3	3	8	3	3	4	3	27
Radiotherapy	2	2	3	2	1	2	1	13
Surgery	3	3	8	3	3	4	3	27

Abbreviations: AA, African American; ECOG, Eastern Cooperative Oncology Group; H, Hispanic; L, Latino; Max, maximum; Min, minimum; q2w, every 2 weeks; q3w every 3 weeks.

as the detector antibody. IMC-20D7S concentrations were derived using SoftMax Pro software from a 4-parameter logistic regression line taken off the standard curve. Serum concentration data were analyzed by standard noncompartmental analysis using Phoenix WinNonlin 6.3.

A blood sample was also collected prior to study treatment to assess Fc receptor polymorphism status. We used a linear regression model, adjusted for dosage, gender, and age, to ask whether polymorphisms correlated with treatment response or duration of treatment.

### Statistical analysis

Efficacy and safety analyses were planned to be descriptive. The safety and efficacy population consisted of all patients exposed to any amount of study drug. Median PFS was determined by the Kaplan–Meier method. Data from patients in the expanded cohort were included with those of patients initially treated at the same dose in the dose-escalating cohort. With regard to Fc receptor polymorphism studies, we used linear regression models to associate the duration of treatment with candidate SNPs with adjustment for covariates (age, gender, and treatment arms). Such analyses were done using R 3.0.2 software. Significance was defined as  $P < 0.05$  for specific polymorphisms.

## Results

### Patient population and treatment

This study enrolled 27 patients between June 29, 2010, and August 20, 2012, of which 16 were men and 11 were women.

Age ranged from 44 to 84 years with a median of 67 years. Each of the seven dose-escalating cohorts included three patients with the exception of Cohort 2B, which included four patients, one of whom had withdrawn prior to completing the first treatment cycle and had been replaced per protocol. An expanded cohort of five patients was treated at 20 mg/kg every 2 weeks. Detailed patient characteristics are shown in Table 1.

Treatment duration ranged from 3 to 27 weeks with the highest number of treatment cycles (7) completed by one patient in Cohort 2A. Across all treatment groups, mean duration of treatment was 10.5 weeks. The cohort with the shortest mean treatment duration (5.3 weeks) was Cohort 1A; the longest mean treatment duration (18.7 weeks) was in Cohort 2A.

### Safety and tolerability

Fourteen patients (51.9%) experienced treatment-related adverse events. Most of these treatment-related adverse events were low grade, and no patients had grade 3 or greater treatment-related adverse events. Adverse events occurring in more than one patient in the study are shown in Table 2. The most frequent adverse events were fatigue and constipation, occurring in nine and eight patients, respectively.

A total of 12 patients experienced 21 treatment-emergent SAEs collectively (Table 2). Each treatment-emergent SAE occurred in one patient, and none were characterized as treatment related. Furthermore, none of the SAEs led to death or discontinuation of study treatment. No deaths occurred during the study or during

**Table 2.** Safety

Cohort	SAEs								All treatment groups
	1A	2A	3A	4A	1B	2B	3B		
N	3	3	8	3	3	4	3		27
No (%) of patients with SAE	1 (33.3)	1 (33.3)	4 (50.0)	2 (66.7)	0	4 (100.0)	0		12 (44.4)
No of treatment-emergent SAEs	1	3	6	4	0	7	0		
SAEs	Syncope	Abdominal pain, pyrexia, metastatic pain	Disease progression, UTI, failure to thrive, hypoglycemia, cerebral hemorrhage, mental status change	Melena, mouth hemorrhage, hematuria, urinary bladder hemorrhage			Subdural hematoma, hypophosphatemia, metastases to CNS, dyspnea, hypoxia, pleural effusion, DVT		
TEAE occurring in more than one patient per cohort									
Cohort	1A	2A	3A	4A	1B	2B	3B		
N	3	3	8b	3	3	4	3		
Event/n (%)	NA	Night sweats/2 (66.7)	Constipation/4 (50.0), fatigue/4 (50.0), arthralgia/2 (25.0), musculoskeletal pain/2 (25.0), cough/2 (25.0)	Diarrhea/2 (66.7), dry mouth/2 (66.7), hyponatremia/2 (66.7)	NA	Hypophosphatemia/2 (50.0)	Fatigue/2 (66.7), arthralgia/2 (66.7)		
TEAE occurring in more than one patient per study									
No. of patients (%)	TEAE								
9 (33.3)	Fatigue								
8 (29.6)	Constipation								
5 (18.5)	Hypophosphatemia, arthralgia								
4 (14.8)	Headache, cough, diarrhea, decreased appetite								
3 (11.1)	Abdominal pain, hyponatremia, night sweats								
2 (7.4)	Dry mouth, chills, upper respiratory tract infection, dehydration, back pain, muscle spasms, musculoskeletal pain, nodule on extremity, peripheral neuropathy, insomnia, dyspnea, pleural effusion, postnasal drip, hyperhidrosis, pruritus, deep vein thrombosis								

Abbreviations: CNS, central nervous system; DVT, deep vein thrombosis; NA, not applicable; UTI, urinary tract infection.

the protocol-required 30-day follow-up period. There were no DLTs in this study (see Supplementary Table S1 for a list of grade 3 or greater TEAE by system organ class).

**Pharmacokinetics and biomarker studies**

The pharmacokinetic parameters computed for IMC-20D7S are shown in Table 3. The terminal elimination half-life ( $t_{1/2}$ )

**Table 3.** Pharmacokinetic parameters following single (cycle 1 day 1) and multiple administrations (cycle 3 day 1)

	Geometric mean (CV%) <sup>a</sup>						
	Arm A (q2w)				Arm B (q3w)		
	5 mg/kg q2w (N = 3)	10 mg/kg q2w (N = 3)	20 mg/kg q2w (N = 8)	30 mg/kg q2w (N = 3)	10 mg/kg q3w (N = 3)	20 mg/kg q3w (N = 4)	30 mg/kg q3w (N = 3)
First infusion (cycle 1 day 1)							
C <sub>max</sub> (µg/mL)	159 (30)	609 (4)	1,228.893; 1,450.289 <sup>b</sup>	1,320 (64)	580.517; 331.732 <sup>b</sup>	1,290 (15) <sup>c</sup>	1,280 (21)
t <sub>max</sub> (h) <sup>d</sup>	1.50 (1.50-1.50)	1.50 (1.00-2.00)	9.00; 1.32 <sup>b</sup>	1.52 (1.52-4.43)	2.00; 1.58 <sup>b</sup>	2.00 (1.00-2.00) <sup>c</sup>	3.65 (1.83-8.63)
AUC <sub>(0-336)</sub> (µg/h/mL)	20,900 <sup>e</sup>	34,200; 47,600 <sup>b</sup>	153,000; 135,000 <sup>b</sup>	100,000; 321,000 <sup>b</sup>	64,700 <sup>e</sup>	161,000 (43) <sup>c</sup>	152,000 (23)
Fifth infusion (cycle 3 day 1)							
C <sub>max</sub> (µg/mL)	NA	505 (22)	NA	2,501.066 <sup>e</sup>	850.041 <sup>e</sup>	1,159.235 <sup>e</sup>	NA
t <sub>max</sub> (h) <sup>d</sup>	NA	25.00 (1.00-49.00)	NA	2.63 <sup>e</sup>	2.03 <sup>e</sup>	3.67 <sup>e</sup>	NA
AUCτ (µg/h/mL)	NA	59,300; 87,400 <sup>b</sup>	NA	395,000 <sup>e</sup>	NC	NC	NA
CL <sub>ss</sub> (L/h)	NA	0.0118; 0.00744 <sup>b</sup>	NA	0.00922 <sup>e</sup>	NC	NC	NA
V <sub>ss</sub> (L)	NA	1.70 <sup>e</sup>	NA	2.51 <sup>e</sup>	NC	NC	NA
R <sub>A,Cmax</sub>	NA	0.829 (22)	NA	0.981 <sup>e</sup>	1.46 <sup>e</sup>	0.808 <sup>e</sup>	NA
R <sub>A,AUC</sub>	NA	1.24; 2.56 <sup>b</sup>	NA	1.23 <sup>e</sup>	NC	NC	NA

Abbreviations: AUC(0-336), area under the concentration-time curve from time 0 to 336 hours; AUCτ, area under the concentration-time curve during 1 dose interval (336 hours for every 2 weeks and 504 hours for every 3 weeks); CL<sub>ss</sub>, clearance at steady state (after intravenous administration); C<sub>max</sub>, maximum observed drug concentration; CV, coefficient of variation; N, number of patients dosed; n, number of observations; NA, not available; NC, not calculated; q2w, every 2 weeks; q3w every 3 weeks; R<sub>A,AUC</sub>, accumulation ratio calculated using AUC; R<sub>A,Cmax</sub>, accumulation ratio calculated using C<sub>max</sub>; t<sub>max</sub>, time of maximum observed drug concentration; V<sub>ss</sub>, volume of distribution at steady state.

<sup>a</sup>Geometric mean and geometric CV% are provided for n ≥ 3; otherwise, actual values are provided.

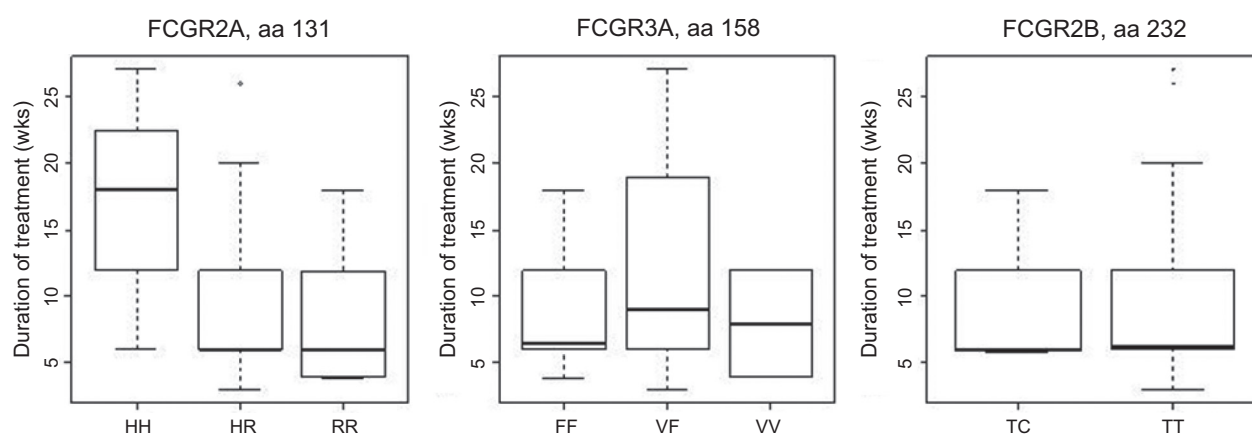
<sup>b</sup>Values separated by semicolon are provided when n = 2.

<sup>c</sup>n = 3.

<sup>d</sup>Median (range) is provided for t<sub>max</sub>.

<sup>e</sup>The value is given when n = 1.

Downloaded from http://aacrjournals.org/clincancerres/article-pdf/22/21/5204/1931581/5204.pdf by guest on 26 August 2022



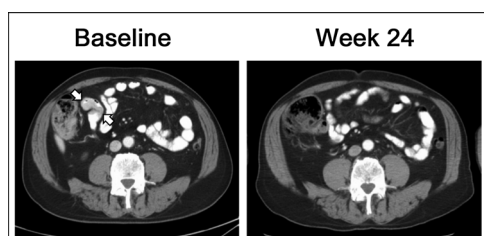
**Figure 1.** FCGR polymorphism status. FCGR2A polymorphisms correlate with duration of treatment.

could not be reliably estimated due to the limited sampling time.

Consistent with recently published data on cetuximab (15), which carries the same IgG1 backbone as IMC-20D7S, there was a significant correlation between the duration of treatment and a polymorphism in the gene encoding FcγRIIa, namely FCGR2A ( $P < 0.05$ ), while there was no discernible correlation between duration of treatment and genotype for FCGR3A or FCGR2B (Fig. 1). The dose of drug administered positively correlated with the duration of treatment ( $P < 0.05$ ). There was no significant correlation between clinical outcome and genotype.

#### Efficacy

There was one patient in Cohort 2A who had a complete response (CR) to IMC-20D7S at week 24 (Fig. 2). At baseline, this 67-year-old man had ileal metastases measuring  $3.3 \times 1.7$  cm in conglomerate dimension. His first on-treatment CT showed regression, with a CR evident by week 24. His PFS was 5.95 months. No patients had a best response of partial response. Ten patients (37%) had stable disease; their PFS values were as follows: 2.6, 3.98, 2.6, 4.4, 4.21, 2.1, 5.55, 4.3, 2.73, and 4.14 months. Twelve patients (44%) had progressive disease. Three patients (11%) were not evaluable. The disease control rate, defined as stable disease or better, was



**Figure 2.** Patient achieving CR. Resolution of ileal metastases measuring  $3.3 \times 1.7$  cm (arrows) in conglomerate dimension.

41% (Table 4). Median PFS of pooled patients from all dose levels was 2.10 months (95% confidence interval, 1.22–2.73). Six patients had a PFS beyond 3 months (4.14, 4.21, 4.3, 4.4, 5.55, and 5.95 months, respectively) with six to 13 infusions in total.

#### Discussion

Treatment with IMC-20D7S was well tolerated with doses safely escalated to 30 mg/kg every 2 weeks (Arm A) and 30 mg/kg every 3 weeks (Arm B). No MTD was determined, as there were no treatment-related SAEs, DLTs, or grade 3 toxicities. The recommended dose for further evaluation was established at 20 mg/kg given every 2 weeks based on pharmacokinetic and safety data. Although the overall objective response rate in this study was low, there was one patient with a CR. Ten patients achieved stable disease.

There are consistent data showing an association between FcR polymorphisms and function of tumor-targeting mAbs (16, 17). As FcγRIIa has been implicated in ADCC (18) and antibody-dependent cell-mediated phagocytosis (ADCP; ref. 19) and the efficacy of the 20D7S preclinical analogue, TA99, was dependent upon FcγR interactions, we hypothesized that patients' FcγRIIa polymorphisms would be relevant to clinical outcome with 20D7S. Because of the low response rate and patient numbers, however, we were unable to find an association between FcγRIIa polymorphisms and clinical response. Nevertheless, we found that FcγRIIa polymorphism status was correlated with treatment duration (Fig. 1). Further exploration of Fcγ receptor polymorphisms, in larger cohorts, is warranted.

Although the efficacy of IMC-20D7S as a single agent was limited, IMC-20D7S may have greater clinical efficacy in combination with other immunotherapeutic approaches, such as checkpoint (e.g., CTLA-4, PD-1, PD-L1) blockade. In principle, as tumor-targeted mAb therapeutics like IMC-20D7S can induce a tumor-specific T-cell response via ADCP (20–22), the induced T-cell response may theoretically be augmented with checkpoint blockade. The favorable toxicity profile of 20D7S makes it an attractive candidate for use in such combinations in subsequent clinical trials.

**Table 4.** Response

Cohort	1A	2A	3A	4A	1B	2B	3B	All treatment groups
N	3	3	8	3	3	4	3	27
Best overall response, n (%)								
CR	0	1 (33.3)	0	0	0	0	0	1 (3.7)
PR	0	0	0	0	0	0	0	0
SD	0	2 (66.7)	2 (25.0)	0	1 (33.3)	2 (50.0)	3 (100.0)	10 (37.0)
Non-CR/non-PD <sup>a</sup>	1 (33.3)	0	0	0	0	0	0	1 (3.7)
PD	2 (66.7)	0	5 (62.5)	2 (66.7)	2 (66.7)	1 (25.0)	0	12 (44.4)
Not evaluable	0	0	1 (12.5)	1 (33.3)	0	1 (25.0)	0	3 (11.1)
Disease control rate (CR + PR + SD)	0.00%	100.00%	25.00%	0	33.30%	50	100.00%	40.70%
95% CI (%)	0.0, 70.8	29.2, 100.0	3.2, 65.1	0.0, 70.8	0.8, 90.6	6.8, 93.2	29.2, 100.0	22.4, 61.2
P	0.1000							

Abbreviations: CI, confidence interval; PR, partial response; SD, stable disease.

<sup>a</sup>One patient in this cohort had nontarget disease only, and as per protocol was characterized as non-CR/non-PD.

### Disclosure of Potential Conflicts of Interest

M.A. Postow reports receiving commercial research grants from Bristol-Myers Squibb. J.S.W. Kauh holds ownership interest (including patents) in Eli Lilly & Co. F.S. Hodi reports receiving commercial research grants from Bristol-Myers Squibb and is a consultant/advisory board member for Genentech, Merck, and Novartis. J.D. Wolchok is a consultant/advisory board member for Lilly. No potential conflicts of interest were disclosed by the other authors.

### Authors' Contributions

**Conception and design:** D.L. Ludwig, D. Grebennik, H.-J. Lenz, J.D. Wolchok  
**Development of methodology:** J. Cosaert, D. Grebennik, H.-J. Lenz, D.P. Lawrence, J.D. Wolchok

**Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.):** M.A. Postow, N. Ibrahim, S. Tang, J.S.W. Kauh, H.-J. Lenz, K.T. Flaherty, F.S. Hodi, D.P. Lawrence, J.D. Wolchok

**Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis):** D.N. Khalil, M.A. Postow, N. Ibrahim, S.R.P. Kambhampati, S. Tang, D. Grebennik, J.S.W. Kauh, H.-J. Lenz, K.T. Flaherty, F.S. Hodi, D.P. Lawrence, J.D. Wolchok

**Writing, review, and/or revision of the manuscript:** D.N. Khalil, M.A. Postow, N. Ibrahim, D.L. Ludwig, S.R.P. Kambhampati, S. Tang, D. Grebennik, J.S.W. Kauh, H.-J. Lenz, K.T. Flaherty, F.S. Hodi, D.P. Lawrence, J.D. Wolchok

**Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases):** D.N. Khalil, D.L. Ludwig, S. Tang, J.S.W. Kauh, H.-J. Lenz, D.P. Lawrence

**Study supervision:** N. Ibrahim, J. Cosaert, D. Grebennik, J.S.W. Kauh, D.P. Lawrence, J.D. Wolchok

**Other (design and medical monitoring of the study):** D. Grebennik

### Acknowledgments

We are grateful for the support of patients and their families. Nicholas Cimaglia provided thorough research assistance.

### Grant Support

This work was supported by an NIH grant P30CA008748. D. Khalil is currently supported by a fellowship in Clinical/Translational Cancer Research from the American Association for Cancer Research and Amgen, in Clinical Investigation from the American Philosophical Society, and through a Young Investigator Award from the Conquer Cancer Foundation and the American Society of Clinical Oncology.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received May 15, 2016; revised July 17, 2016; accepted August 4, 2016; published OnlineFirst October 19, 2016.

### References

- American Cancer Society. Cancer Facts & Figures 2014. Atlanta, GA: American Cancer Society; 2014.
- Robert C, Schachter J, Long GV, Arance A, Grob JJ, Mortier L, et al. Pembrolizumab versus ipilimumab in advanced melanoma. *N Engl J Med* 2015;372:2521-32.
- Larkin J, Chiarion-Sileni V, Gonzalez R, Grob JJ, Cowey CL, Lao CD, et al. Combined nivolumab and ipilimumab or monotherapy in untreated melanoma. *N Engl J Med* 2015;373:23-34.
- Long GV, Stroyakovskiy D, Gogas H, Levchenko E, de Braud F, Larkin J, et al. Dabrafenib and trametinib versus dabrafenib and placebo for Val600 BRAF-mutant melanoma: a multicentre, double-blind, phase 3 randomised controlled trial. *Lancet* 2015;386:444-51.
- Larkin J, Ascierto PA, Dréno B, Atkinson V, Liszkay G, Maio M, et al. Combined vemurafenib and cobimetinib in BRAF-mutated melanoma. *N Engl J Med* 2014;371:1867-76.
- Ghanem G, Fabrice J. Tyrosinase related protein 1 (TYRP1/gp75) in human cutaneous melanoma. *Mol Oncol* 2011;5:150-5.
- Takechi Y, Hara I, Naftzger C, Xu Y, Houghton AN. A melanosomal membrane protein is a cell surface target for melanoma therapy. *Clin Cancer Res* 1996;2:1837-42.
- Tai T, Eisinger M, Ogata S, Lloyd KO. Glycoproteins as differentiation markers in human malignant melanoma and melanocytes. *Cancer Res* 1983;43:2773-9.
- Journe F, Id Boufker H, Van Kempen L, Galibert M-D, Wiedig M, Salès F, et al. TYRP1 mRNA expression in melanoma metastases correlates with clinical outcome. *Br J Cancer* 2011;105:1726-32.
- Sliwkowski MX, Mellman I. Antibody therapeutics in cancer. *Science* 2013;341:1192-8.
- Boross P, Leusen JHW. Mechanisms of action of CD20 antibodies. *Am J Cancer Res* 2012;2:676-90.
- Welt S, Mattes MJ, Grando R, Thomson TM, Leonard RW, Zanzonico PB, et al. Monoclonal antibody to an intracellular antigen images human melanoma transplants in nu/nu mice. *Proc Natl Acad Sci U S A* 1987;84:4200-4.
- Hara I, Takechi Y, Houghton AN. Implicating a role for immune recognition of self in tumor rejection: passive immunization against the brown locus protein. *J Exp Med* 1995;182:1609-14.
- Clynes R, Takechi Y, Moroi Y, Houghton A, Ravetch JV. Fc receptors are required in passive and active immunity to melanoma. *Proc Natl Acad Sci U S A* 1998;95:652-6.

15. Liu G, Tu D, Lewis M, Cheng D, Sullivan LA, Chen Z, et al. Fc- $\gamma$  receptor polymorphisms, cetuximab therapy, and survival in the NCIC CTG CO.17 trial of colorectal cancer. *Clin Cancer Res*. 2016;22:2435–44.
16. Bekaii-Saab TS, Roda JM, Guenterberg KD, Ramaswamy B, Young DC, Ferketich AK, et al. A phase I trial of paclitaxel and trastuzumab in combination with interleukin-12 in patients with HER2/neu-expressing malignancies. *Mol Cancer Ther* 2009;8:2983–91.
17. Srivastava RM, Lee SC, Andrade Filho PA, Lord CA, Jie HB, Davidson HC, et al. Cetuximab-activated natural killer and dendritic cells collaborate to trigger tumor antigen-specific T-cell immunity in head and neck cancer patients. *Clin Cancer Res* 2013;19:1858–72.
18. Derer S, Glorius P, Schlaeth M, Lohse S, Klausz K, Muchhal U, et al. Increasing Fc $\gamma$ RIIa affinity of an Fc $\gamma$ RIII-optimized anti-EGFR antibody restores neutrophil-mediated cytotoxicity. *MAbs* 2014;6:409–21.
19. Petricevic B, Laengle J, Singer J, Sachet M, Fazekas J, Steger G, et al. Trastuzumab mediates antibody-dependent cell-mediated cytotoxicity and phagocytosis to the same extent in both adjuvant and metastatic HER2/neu breast cancer patients. *J Transl Med* 2013;11:307.
20. DiLillo DJ, Ravetch JV. Differential Fc-receptor engagement drives an anti-tumor vaccinal effect. *Cell* 2015;161:1035–45.
21. Hilchey SP, Hyrien O, Mosmann TR, Livingstone AM, Friedberg JW, Young F, et al. Rituximab immunotherapy results in the induction of a lymphoma idiotype-specific T-cell response in patients with follicular lymphoma: Support for a "vaccinal effect" of rituximab. *Blood* 2009; 113:3809–12.
22. Abès R, Gélizé E, Fridman WH, Teillaud JL. Long-lasting antitumor protection by anti-CD20 antibody through cellular immune response. *Blood* 2010;116:926–34.