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ACCELERATED PUBLICATION

An unexpected twist to the activation of IKK β : TAK1 primes IKK β for activation by autophosphorylation

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IKK β {I κ B [inhibitor of NF- κ B (nuclear factor κ B)] kinase β } is required to activate the transcription factor NF- κ B, but how IKK β itself is activated *in vivo* is still unclear. It was found to require phosphorylation by one or more ‘upstream’ protein kinases in some reports, but by autophosphorylation in others. In the present study, we resolve this controversy by demonstrating that the activation of IKK β induced by IL-1 (interleukin-1) or TNF (tumour necrosis factor) in embryonic fibroblasts, or by ligands that activate Toll-like receptors in macrophages, requires two distinct phosphorylation events: first, the TAK1 [TGF β (transforming growth factor β)-activated kinase-1]-catalysed phosphorylation of Ser¹⁷⁷ and, secondly, the IKK β -catalysed autophosphorylation of Ser¹⁸¹. The phosphorylation of Ser¹⁷⁷ by TAK1 is a priming event required for the subsequent autophosphorylation of Ser¹⁸¹, which

enables IKK β to phosphorylate exogenous substrates. We also provide genetic evidence which indicates that the IL-1-stimulated, LUBAC (linear ubiquitin chain assembly complex)-catalysed formation of linear ubiquitin chains and their interaction with the NEMO (NF- κ B essential modulator) component of the canonical IKK complex permits the TAK1-catalysed priming phosphorylation of IKK β at Ser¹⁷⁷ and IKK α at Ser¹⁷⁶. These findings may be of general significance for the activation of other protein kinases.

Key words: inhibitor of nuclear factor κ B kinase (IKK), interleukin-1 (IL-1), linear ubiquitin chain assembly complex (LUBAC), nuclear factor κ B (NF- κ B), transforming growth factor β -activated kinase-1 (TAK1).

INTRODUCTION

The canonical IKK {I κ B [inhibitor of NF- κ B (nuclear factor κ B)] kinase β } complex, consisting of the protein kinases IKK α and IKK β (also called IKK1 and IKK2) and a regulatory component called NEMO (NF- κ B essential modifier) [1,2], is one of the most studied of all protein kinases. It has featured in over 10000 papers since its discovery in 1998 due to its essential role in activating NF- κ B, a ‘master’ transcription factor that regulates many physiological processes, including innate immunity and the cellular response to DNA damage [3–5]. Nevertheless, despite the vast number of publications that have focused on this protein kinase, its mechanism of activation is still controversial.

The activation of IKK α and IKK β requires phosphorylation of the ‘activation loops’ of these protein kinases at Ser¹⁷⁶ and Ser¹⁸⁰ (IKK α) or Ser¹⁷⁷ and Ser¹⁸¹ (IKK β) [4]. The IKKs respond to many physiological stimuli, but are activated most powerfully by inflammatory stimuli, such as TLR (Toll-like receptor) agonists and the pro-inflammatory cytokines IL-1 (interleukin-1) and TNF (tumour necrosis factor). Genetic evidence indicates that the expression and activity of the TAK1 [TGF β (transforming growth factor β)-activated kinase-1; also called MAP3K7 (mitogen-activated protein kinase kinase kinase 7)] is needed for the activation of the canonical IKK complex by IL-1 and TNF in MEFs (mouse embryonic fibroblasts). These agonists fail to activate the IKKs in MEFs that do not express the TAK1 catalytic subunit [6] or that express a truncated inactive form of TAK1 [7].

IL-1 and TNF trigger TAK1 activation within minutes, a speed compatible with a role in initiating the activation of the IKKs [8]. TAK1 is also reported to phosphorylate and activate the canonical IKKs *in vitro* [9], activation being prevented by pharmacological inhibitors of TAK1 [8,10,11]. Similar lines of evidence indicate an essential role for TAK1 in activating the MKKs [MAPK (mitogen-activated protein kinase) kinases] that switch on the MAPK family members JNK1 (c-Jun N-terminal kinase 1) and JNK2 and p38 MAPKs in MEFs [8–11].

On the other hand, the canonical IKKs have been shown to be capable of phosphorylating and activating themselves *in vitro* (reviewed in [4]). For example, Met¹-linked (also called linear) ubiquitin oligomers [12] and other types of ubiquitin oligomers [13] have been reported to induce the activation of the canonical IKK complex *in vitro*, apparently in the absence of any ‘upstream’ activating protein kinase. These observations raise the alternative possibility that the role of TAK1 *in vivo* might be to stimulate the formation of these polyubiquitin chains, rather than to phosphorylate the canonical IKK complex directly. In addition, X-ray crystallographic analysis has revealed that human IKK β can adopt an open conformation that enables it to form oligomers, whereas mutagenesis studies have established that two of the surfaces that mediate oligomer formation are critical for the activation of IKK β in cells [14]. It has therefore been proposed that IKK β dimers transiently associate with one another through these interaction surfaces to promote *trans* autophosphorylation as part of their activation mechanism. Consistent with an essential

Abbreviations: BMDM, bone-marrow-derived macrophage; E, embryonic day; HA, haemagglutinin; HEK, human embryonic kidney; HOIP, HOIL1 [haem-oxidized IRP2 (iron regulatory protein 2) ubiquitin ligase 1]-interacting protein; I κ B, inhibitor of NF- κ B; IKK, I κ B kinase; IL-1, interleukin-1; JNK, c-Jun N-terminal kinase; LPS, lipopolysaccharide; LUBAC, linear ubiquitin chain assembly complex; MAPK, mitogen-activated protein kinase; M-CSF, macrophage colony-stimulating factor; MEF, mouse embryonic fibroblast; MKK, MAPK kinase; NEMO, NF- κ B essential modulator; NF- κ B, nuclear factor κ B; PP1 γ , protein phosphatase 1 γ ; TAB, TAK1-binding protein; TAK1, transforming growth factor β -activated kinase-1; TLR, Toll-like receptor; TNF, tumour necrosis factor; TRAF, TNF receptor-associated factor.

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role for autophosphorylation, we found that in IKK α -deficient MEFs the specific IKK β inhibitor BI605906 prevented the IL-1- or TNF-stimulated conversion of IKK β into the active diphosphorylated species, i.e. phosphorylated at both Ser¹⁷⁷ and Ser¹⁸¹ [8].

In the present study we report the unexpected finding that TAK1 and IKK β phosphorylate different serine residues in the activation loop of IKK β and demonstrate that the TAK1-catalysed phosphorylation of IKK β at Ser¹⁷⁷ is a priming event that enables IKK β to activate itself by phosphorylating Ser¹⁸¹. We also provide genetic evidence showing that the formation of Met¹-linked ubiquitin chains and their interaction with NEMO is needed for the TAK1-catalysed phosphorylation of Ser¹⁷⁶ (IKK α) and Ser¹⁷⁷ (IKK β), and that TAK1 activity is not required for the formation of either Lys⁶³-linked or Met¹-linked ubiquitin chains.

EXPERIMENTAL

Materials

Murine IL-1 α and TNF were purchased from Peprotech and mouse M-CSF (macrophage colony-stimulating factor) from R&D Systems. Pam₃CSK4 was from Invivogen and LPS (lipopolysaccharide) O55:B5 was from Enzo Life Science. The monophosphorylated peptide KELDQGpSLCTSFVGTLQ and the diphosphorylated peptide KELDQGpSLCTpSFVGTLQ (where pS is phosphoserine), corresponding to amino acids 171–187 of IKK β with phosphoserine at Ser¹⁷⁷ only or at both Ser¹⁷⁷ and Ser¹⁸¹ respectively, were synthesized by Pepceuticals. The IKK β inhibitor BI605906 [8] was provided by Dr Natalia Shpiro (University of Dundee, Dundee, U.K.) and the TAK1 inhibitor NG25 by Dr Nathanael Gray (Harvard Medical School, Boston, MA, U.S.A.) [11], whereas the TAK1 inhibitor 5Z-7-oxozeaenol was purchased from BioAustralis Fine Chemicals.

Protein expression and purification

The IKK β (IKK β [D166A]) was expressed as a GST fusion protein in HEK (human embryonic kidney)-293T suspension cells and, after cell lysis, was purified from the cell extracts by chromatography on glutathione–Sepharose. The GST-fusion protein was released from the glutathione–Sepharose by cleavage of the GST tag with PreScission protease. A catalytically active TAK1–TAB1 (TAK1-binding protein 1)-fusion protein [15] was expressed in insect Sf21 cells as a His₆-tagged protein and purified by chromatography on nickel-nitrilotriacetate agarose. The catalytic subunit of human PP1 γ (protein phosphatase 1 γ) was expressed in *Escherichia coli* as a GST-fusion protein, purified on glutathione–Sepharose and stored in a solution of 50 mM Tris/HCl, 0.15 M NaCl, 0.27 M sucrose, 0.03 % Brij35, 0.1 % 2-mercaptoethanol and 2 mM MnCl₂.

Antibodies

An antibody recognizing the HOIP {HOIL1 [haem-oxidized IRP2 (iron regulatory protein 2) ubiquitin ligase 1]-interacting protein} component of LUBAC (linear ubiquitin chain assembly complex) was raised in sheep and purified as described in [16]. Antibodies recognizing IKK β phosphorylated at Ser¹⁷⁷ (catalogue number 2078S) or at both Ser¹⁷⁷ and Ser¹⁸¹ (catalogue number 2697L) were obtained from Cell Signaling Technology, whereas the antibody recognizing IKK β phosphorylated at Ser¹⁸¹ was from Abcam (catalogue number AB55341). Antibodies that recognize p105/NF- κ B1 phosphorylated at Ser⁹³³ (catalogue number 4806S), GAPDH (glyceraldehyde-3-phosphate dehydrogenase; catalogue number 2118S) and all forms of p38 α MAPK

(catalogue number 9212S) and JNK (catalogue number 9258S) were from Cell Signaling Technology. Antibodies recognizing NEMO (catalogue number SC8330; Santa Cruz Biotechnology), all forms of IKK β (catalogue number DAM1774677; Millipore) and the HA (haemagglutinin) tag (catalogue number 12-013-819-001; Roche) were from the sources indicated. An antibody raised in sheep against the full-length human IKK β catalytic subunit (S189C, bleed 1) was produced and affinity purified by the Antibody Production Team of the MRC Protein Phosphorylation and Ubiquitylation Unit at Dundee (co-ordinated by Dr James Hastie). An antibody recognizing Met¹-linked ubiquitin chains was generously provided by Vishva Dixit, Genentech, U.S.A. and the antibody recognizing Lys⁶³-linked ubiquitin chains was purchased from Merck-Millipore (catalogue number 05-1313).

DNA constructs

DNA encoding IKK β (NCBI BAI45894.1) was amplified from total thymus RNA using the One Step RT PCR kit (Life Technologies). It was then cloned into pCR2.1 (Life Technologies), sequenced and sub-cloned into the NotI site of pRetrox tight HA. Mutations were created following the QuikChange Site-Directed Mutagenesis method, but using KOD Hot Start DNA Polymerase (EMD Millipore).

Cell culture, stimulation and immunoblotting

MEFs and HEK-293 cells were maintained in DMEM (Dulbecco's modified Eagle's medium) supplemented with 2 mM glutamine, 10 % (v/v) FBS, and the antibiotics streptomycin (0.1 mg/ml) and penicillin (100 units/ml). DNA constructs were transfected into HEK-293 cells using polyethyleneimine (Polysciences). BMDMs (bone-marrow-derived macrophages) were obtained by culturing bone marrow from the tibia and femurs of mice in the presence of mouse M-CSF and replating for 24 h before stimulation. Kinase inhibitors (10 mM) dissolved in DMSO, or an equivalent volume of DMSO for the control incubations, were added to the culture medium of cells grown as monolayers. After 1 h at 37°C, MEFs were stimulated with IL-1 α or TNF and BMDM with LPS or Pam₃Csk₄ (see the Figure legends). Thereafter, cells were rinsed in ice-cold PBS and extracted in lysis buffer [50 mM Tris/HCl (pH 7.4), 1 mM EDTA, 1 mM EGTA, 50 mM NaF, 5 mM sodium pyrophosphate, 10 mM sodium 2-glycerol 1-phosphate, 1 mM DTT, 1 mM sodium orthovanadate, 0.27 M sucrose, 1 % (v/v) Triton X-100, 1 μ g/ml aprotinin, 1 μ g/ml leupeptin and 1 mM PMSF]. Cell extracts were clarified by centrifugation (21 000 *g* for 10 min at 4°C) and protein concentrations determined by the Bradford assay. Cell extract protein (20 μ g) was separated by SDS/PAGE (8 % gel), transferred on to PVDF membranes and proteins detected by immunoblotting and visualized by treating the blots with enhanced chemiluminescence (Amersham).

Generation of MEFs from knockin mice

Mice in which wild-type NEMO was replaced by the polyubiquitin-binding-defective mutant NEMO[D311N] were generated by Taconic-Artemis using conventional technology and their characterization will be reported elsewhere. Primary MEFs from NEMO[D311N] mice and wild-type littermates were generated at E11.5 (embryonic day 11.5), whereas MEFs from knockin mice expressing the inactive C879S mutant of HOIP were generated at E10.5 [16]. Immortalized IKK α -deficient MEFs and wild-type control MEFs were provided by Dr Inder Verma (Salk Institute, La Jolla, CA, U.S.A.). All animals were maintained in specific pathogen-free conditions consistent with EU and U.K.

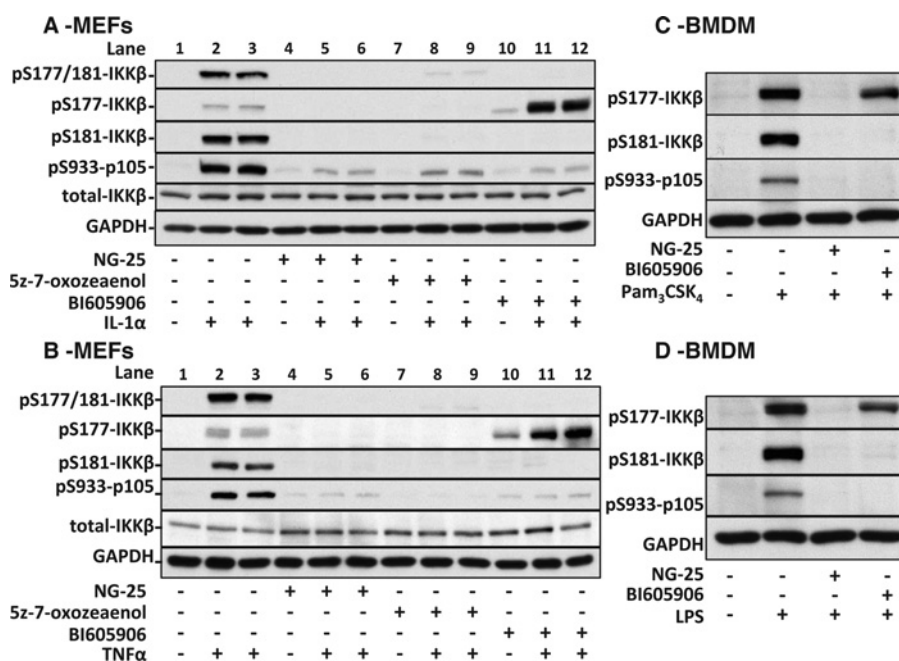


Figure 1 Effect of protein kinase inhibitors on the phosphorylation of IKK β at Ser¹⁷⁷ and/or Ser¹⁸¹ in MEFs from IKK α -deficient mice and BMDMs from knockin mice expressing catalytically inactive IKK α [S176A/S180A]

(A) MEFs from IKK α -knockout mice were incubated for 1 h without (–) or with (+) 1.0 μ M NG25, 1.0 μ M 5Z-7-oxozeaenol or 5.0 μ M BI605906, then stimulated for 10 min with 5.0 ng/ml IL-1. Cell lysates were subjected to SDS/PAGE and immunoblotting as described in the Experimental section. (B) The experiment was performed exactly as in (A) except that the cells were stimulated with 10 ng/ml TNF. (C and D) BMDMs from knockin mice expressing the catalytically inactive IKK α [S176A/S180A] mutant were incubated for 1 h without (–) or with (+) 2 μ M NG25 or 2 μ M BI605906, then stimulated for 10 min with 1 μ g/ml Pam₃CSK₄ (C) or 0.1 μ g/ml LPS (D). Cell extracts were subjected to SDS/PAGE and immunoblotted as in (A and B).

regulations. All work was performed under a U.K. Home Office project license that was awarded after recommendation by the University of Dundee Ethical Review Committee.

Retroviral transduction of IKK α -knockout MEFs

IKK α -deficient MEFs stably expressing HA-tagged empty vector (EV), wild-type HA-IKK β (WT), HA-IKK β [S177A], HA-IKK β [S177E] or HA-IKK β [D166A/S177E] were generated by retroviral transduction using an MMLV (Moloney murine leukaemia virus)-based system prepared with the VSVG (vesicular-stomatitis-virus glycoprotein) envelope protein. Retroviral particles were prepared according to the manufacturer's instructions (Clontech). Viruses encoding the gene of interest and the Tet-On protein were harvested 48 h after transfection, diluted 4-fold with fresh medium and incubated for 24 h with IKK α -deficient MEFs in the presence of 2 μ g/ml protamine sulfate (Sigma). Fresh medium containing 1 μ g/ml G418 (Tet-On) and 3 μ g/ml puromycin (gene of interest) was added to select the transduced cells. Cells were cultured for 16 h with doxycycline (0.1–1.0 μ g/ml) to induce the expression of wild-type and mutant forms of IKK β .

Immunoprecipitation and dephosphorylation of IKK β

To immunoprecipitate transfected HA-tagged IKK β , cell extract protein (40 μ g) was incubated for 60 min at 4°C with 4 μ g of anti-HA antibody, whereas for the endogenous IKK β 0.2 mg of cell extract protein was incubated with 2.5 μ g of anti-IKK β antibody. Protein G-Sepharose was added (equivalent to 10 μ l packed volume) and, after mixing for 30 min at 4°C, immune complexes were collected by brief centrifugation, washed three times in cell lysis buffer plus 0.5 M NaCl, and three times with 50 mM Tris/HCl (pH 7.5), 0.05 M NaCl and 1.0 mM DTT,

then resuspended in 0.03 ml of 50 mM Hepes, 10 mM NaCl, 2 mM DTT and 0.1 % Brij35 (pH 7.5) containing 1 mM MnCl₂. Dephosphorylation was initiated by the addition of 100 μ g of GST-PP1 γ . After 60 min at 30°C the immunoprecipitates were collected, washed three times with 1.0 ml of lysis buffer containing 0.5 M NaCl, and three times with 50 mM Tris/HCl (pH 7.5), 0.1 mM EGTA and 0.1 % 2-mercaptoethanol to remove the phosphatase.

Assay of immunoprecipitated IKK β

IKK β immunoprecipitates were assayed for IKK β activity in a 0.05 ml incubation containing 50 mM Tris/HCl (pH 7.5), 0.1 mM EGTA and 0.1 % 2-mercaptoethanol, 1.0 μ M microcystin (to inactivate any remaining traces of PP1 γ), 0.3 mM of the peptide LDDRHDSGLDSMKDEEY (corresponding to amino acid residues 26–42 of I κ B α), 10 mM magnesium acetate and 0.1 mM [γ -³²P]ATP (5 \times 10⁵ c.p.m./nmol). After incubation for 10 min at 30°C on a shaking platform, the incorporation of ³²P radioactivity into the peptide substrate was measured as described in [17].

RESULTS

IKK β is activated by TAK1 and by autophosphorylation

We initially confirmed that IL-1 or TNF stimulate the dual phosphorylation of IKK β at Ser¹⁷⁷ and Ser¹⁸¹ in IKK α -deficient MEFs, and that this was prevented by the inclusion of the IKK β inhibitor BI605906 in the culture medium (Figures 1A and 1B, top panel, compare lanes 1–3 with 10–12). In these and many earlier studies, the phospho-specific antibody used to monitor the phosphorylation of IKK β recognizes the di-phosphorylated species phosphorylated at both Ser¹⁷⁷ and Ser¹⁸¹.

It was therefore possible that BI605906 and/or pharmacological inhibitors of TAK1 had suppressed the phosphorylation of just one of the serine residues in the activation loop. To address this possibility we therefore employed antibodies that recognize IKK β phosphorylated at either Ser¹⁷⁷ or Ser¹⁸¹. These studies led to the striking and surprising observation that BI605906 suppressed the IL-1- or TNF-stimulated phosphorylation of Ser¹⁸¹, but not the phosphorylation of Ser¹⁷⁷ (Figures 1A and 1B, second and third panels from top, lanes 10–12). In contrast, two structurally unrelated inhibitors of TAK1, NG25 and 5Z-7-oxozeanin, prevented IL-1 or TNF from inducing the phosphorylation of IKK β at both Ser¹⁷⁷ and Ser¹⁸¹ in IKK α -deficient MEFs (Figures 1A and 1B, second and third lanes from top, lanes 4–9). Similar results were observed in BMDMs from knockin mice expressing the catalytically inactive IKK α [S176A/S180A] mutant (Figures 1C and 1D) [18].

The recognition of IKK β by the Ser¹⁷⁷ phospho-specific antibody appeared to be greatly enhanced when IKK α -deficient MEFs were incubated with BI605906 and then stimulated with IL-1 or TNF (Figures 1A and 1B, second panel from top, compare lanes 10–12 with 1–3). This observation is explained by the failure of the antibody to recognize IKK β phosphorylated at Ser¹⁷⁷ if Ser¹⁸¹ is also phosphorylated, and is not a reflection of a real increase in the phosphorylation of Ser¹⁷⁷. This was shown by immunoblotting experiments with a synthetic mono-phosphorylated peptide corresponding to amino acid residues 171–187 of IKK β containing phosphoserine at the position equivalent to Ser¹⁷⁷, and a diphosphorylated form of this peptide with phosphoserine present at both Ser¹⁷⁷ and Ser¹⁸¹ (Supplementary Figures S1A and S1B at <http://www.biochemj.org/bj/461/bj4610531add.htm>). We have encountered similar situations with other proteins in which the two sites of phosphorylation are separated by only four amino acid residues (e.g. [19]). In contrast, the antibody that recognizes the Ser¹⁸¹-phosphorylated form of IKK β detected the di-phosphorylated form of the peptide (Supplementary Figure S1C), because this antibody recognizes the epitope Cys-Thr-pSer-Phe-Val (where pSer is phospho-Ser¹⁸¹), which does not contain Ser¹⁷⁷. As expected, the antibody recognizing Ser¹⁸¹ of IKK β did not detect the mono-phosphorylated peptide containing phosphoserine only at Ser¹⁷⁷ (Supplementary Figure S1C).

The simplest interpretation of the results presented in Figure 1 was that the TAK1-catalysed phosphorylation of Ser¹⁷⁷ was a prerequisite for the subsequent IKK β -catalysed phosphorylation of Ser¹⁸¹. To investigate this hypothesis, we generated IKK α -deficient MEFs that stably expressed (under an inducible promoter) mutated forms of IKK β in which Ser¹⁷⁷ was changed to either glutamic acid (to mimic the effect of phosphorylation by introducing a negative charge) or to alanine (to prevent phosphorylation). The S177E mutant became phosphorylated at Ser¹⁸¹, even in MEFs that had not been stimulated with IL-1 or TNF, whereas the S177A mutant or wild-type IKK β did not (Figure 2A). Moreover, under these conditions, incubation with the IKK β inhibitor BI605906 induced substantial dephosphorylation of the S177E mutant at Ser¹⁸¹, whereas incubation with the TAK1 inhibitor NG25 had no effect. Furthermore, a catalytically inactive version of the S177E mutant, created by additionally mutating Asp¹⁶⁶ in the Asp-Phe-Gly motif to alanine, failed to undergo phosphorylation at Ser¹⁸¹ (Figure 2B). Taken together, these experiments demonstrated, by two independent methods, that the phospho-mimetic S177E mutation permits IKK β to autophosphorylate Ser¹⁸¹.

IKK β initiates the activation of NF- κ B *in vivo* by phosphorylating the inhibitory I κ B α component at Ser³² and Ser³⁶. The IKK β [S177E]-catalysed phosphorylation of a synthetic peptide

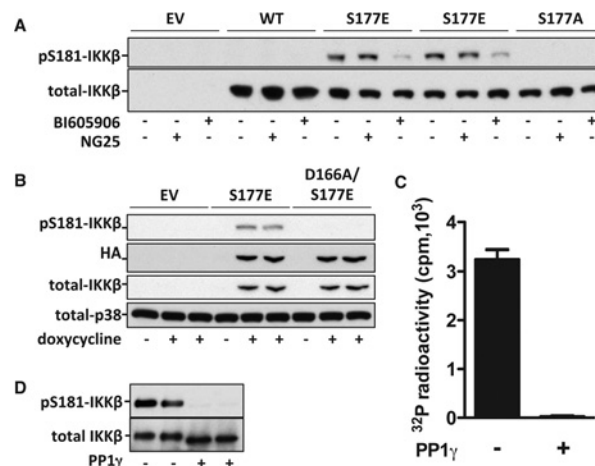


Figure 2 Expression of IKK β [S177E] induces the autophosphorylation of Ser¹⁸¹ and activation of IKK β

(A) MEFs from IKK α -deficient mice stably expressing HA-tagged wild-type IKK β (WT), IKK β [S177E] (S177E) or IKK β [S177A] (S177A) or empty vector (EV) were incubated for 16 h with 1.0 μ g/ml (WT and EV), 0.2 μ g/ml (S177A) or 0.1 μ g/ml (S177E) doxycycline to induce the expression of these proteins, and then for 1 h without (–) or with (+) 5 μ M NG25 or 5 μ M BI605906 and lysed. Extract [20 μ g (EV, S177E and S177A) or 80 μ g (WT) protein] were analysed by immunoblotting with the antibodies indicated. (B) As in (A), except that Ser¹⁸¹ phosphorylation was studied in MEFs stably expressing HA-IKK β [D166A/S177E] and HA-IKK β [S177E], and no inhibitors were present. (C and D) HA-tagged IKK β [S177E] was transfected into HEK-293 cells, immunoprecipitated from the cell extracts, incubated without (–) or with (+) PP1 γ and assayed for IKK β activity (C) or immunoblotted with antibodies that recognize IKK β phosphorylated at Ser¹⁸¹ or all forms of IKK β (D). The results in (C) are means \pm S.E.M. of duplicate determinations. Similar results were obtained in two other independent experiments.

comprising amino acid residues 26–42 of I κ B α was suppressed by BI605906 similarly to wild-type IKK β (Supplementary Figures S2 at <http://www.biochemj.org/bj/461/bj4610531add.htm>), establishing that the activity being measured was catalysed by IKK β and not by another protein kinase present in the immunoprecipitates as a contaminant. Phosphatase treatment inactivated the IKK β [S177E] mutant (Figure 2C), and this was accompanied by the dephosphorylation of Ser¹⁸¹ and a small increase in the electrophoretic mobility of IKK β (Figure 2D). These experiments established that the phospho-mimetic S177E mutation had not activated IKK β , but permitted IKK β to auto-activate by phosphorylating Ser¹⁸¹.

BI605906 is a reversible inhibitor of IKK β (Supplementary Figure S3 at <http://www.biochemj.org/bj/461/bj4610531add.htm>). To investigate whether the phosphorylation of Ser¹⁷⁷ could activate IKK β in the absence of Ser¹⁸¹ phosphorylation, we incubated IKK α -deficient MEFs with BI605906 to suppress the phosphorylation of Ser¹⁸¹ and assayed the endogenous IKK β activity after its immunoprecipitation from the extracts of IL-1-stimulated cells. These experiments showed that IKK β mainly phosphorylated at Ser¹⁷⁷ had a much lower activity than IKK β phosphorylated at both Ser¹⁷⁷ and Ser¹⁸¹ (Figure 3A). Taken together, the results presented in Figures 2 and 3 indicate that Ser¹⁷⁷ is a priming event that enables IKK β to auto-activate itself by phosphorylating Ser¹⁸¹.

Activation of the canonical IKK complex

The experiments presented above were carried out in IKK α -deficient MEFs or in BMDMs from knockin mice expressing the catalytically inactive IKK α [S176A/S180A] mutant, because IKK α activity is unaffected by BI605906 [8]. In contrast, the

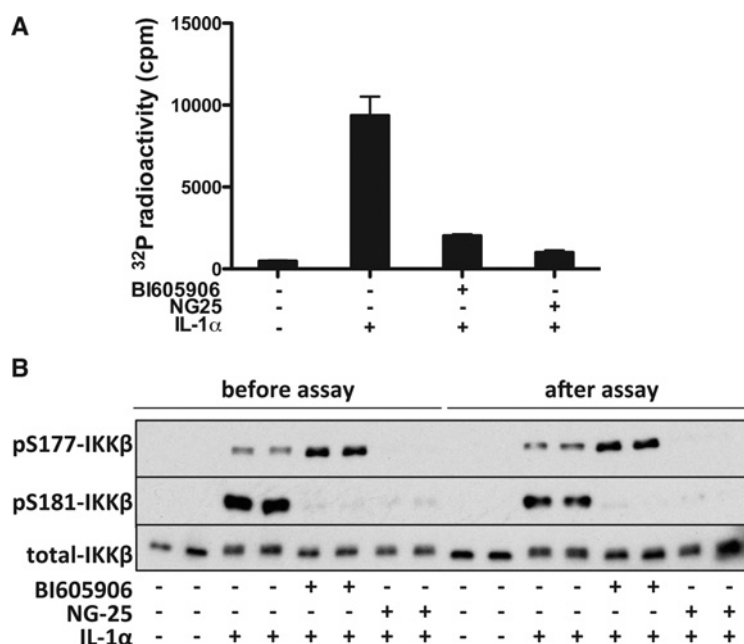


Figure 3 IKK β phosphorylated at Ser¹⁷⁷ has little activity if Ser¹⁸¹ is not phosphorylated

(A) MEFs from IKK α -deficient mice were incubated for 1 h without (–) or with (+) 5.0 μ M BI 605906 or 2 μ M NG25, then stimulated for 10 min with 5.0 ng/ml IL-1. The endogenous IKK β was immunoprecipitated from 0.2 mg of cell extract protein and assayed for activity. (B) The immunoprecipitates from (A) were denatured in SDS before and after the assay of IKK β , and aliquots of each sample were subjected to SDS/PAGE, transferred on to PVDF membranes and immunoblotted with antibodies that recognize IKK β phosphorylated at Ser¹⁷⁷ or Ser¹⁸¹ or all forms of IKK β .

phosphorylation of IKK β at Ser¹⁸¹ was only decreased slightly by BI605906 in wild-type MEFs (Supplementary Figures S4A and S4B at <http://www.biochemj.org/bj/461/bj4610531add.htm>) and was not decreased significantly in Pam₃CSK₄- or LPS-stimulated BMDMs (Supplementary Figures S4C). This suggests that in wild-type cells, in which IKK α , IKK β and NEMO form a single ternary complex, phosphorylation of IKK β at Ser¹⁸¹ can be catalysed *in trans* by IKK α if IKK β is inhibited by BI605906.

The formation of Met¹-linked ubiquitin chains and their interaction with NEMO is required for TAK1 to phosphorylate IKK α and IKK β at Ser¹⁷⁶/Ser¹⁷⁷

LUBAC is the only E3 ubiquitin ligase that catalyses the formation of Met¹-linked (linear) ubiquitin chains in IL-1-stimulated MEFs, and the formation of these ubiquitin chains is required for robust activation of the canonical IKK complex by this agonist ([16], reviewed in [3]). To investigate whether Met¹-linked ubiquitin chain formation was required for the phosphorylation of Ser¹⁷⁷, Ser¹⁸¹ or both amino acid residues, we studied the phosphorylation of each of these sites in MEFs from knockin mice in which HOIP, the catalytic subunit of LUBAC, was replaced by the inactive HOIP[C879S] mutant [16]. These experiments demonstrated that the IL-1-stimulated phosphorylation of IKK β at Ser¹⁷⁷ or Ser¹⁸¹ or IKK α at Ser¹⁷⁶ or Ser¹⁸⁰ was greatly reduced in MEFs from HOIP[C879S]-knockin mice, as was the phosphorylation of p105/NF κ B1 at Ser⁹³³, an established physiological substrate of IKK β (Figure 4A) [20].

The Met¹-linked ubiquitin chains formed by LUBAC bind to the NEMO component of the canonical IKK complex (reviewed in [3]). We therefore generated knockin mice expressing NEMO[D311N], an ubiquitin-binding-defective mutant of NEMO [21–23], and studied the phosphorylation of IKK β in MEFs from these animals. We found that the IL-1-stimulated

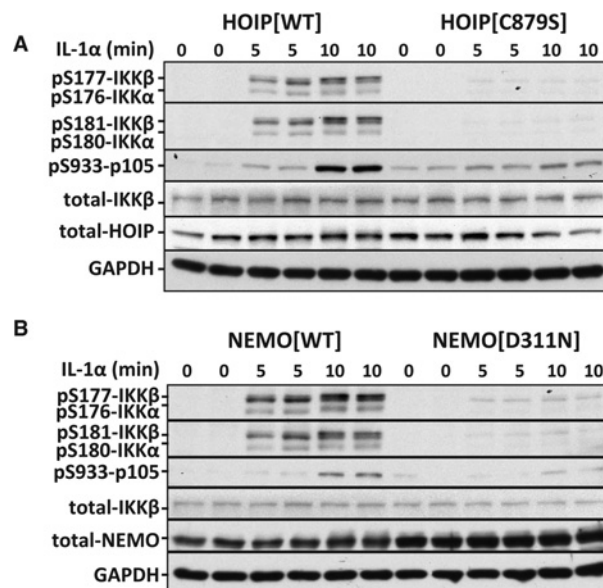


Figure 4 Met¹-linked ubiquitin chains and their interaction with NEMO are required for the IL-1-stimulated phosphorylation of IKK α and IKK β in MEFs

(A) Cells from wild-type (HOIP[WT]) or knockin mice expressing the HOIP[C879S] mutant were stimulated with 5 ng/ml IL-1 for the times indicated and lysed. The extract (20 μ g of protein) was subjected to immunoblotting and probed with the antibodies indicated. (B) As in (A) except that MEFs from NEMO[D311N]-knockin and wild-type mice were used.

phosphorylation of IKK β at Ser¹⁷⁷ or Ser¹⁸¹, or IKK α at Ser¹⁷⁶ or Ser¹⁸⁰, was impaired in MEFs expressing the NEMO[D311N] mutant (Figure 4B), similar to the results obtained in MEFs from the HOIP[C879S]-knockin mice (Figure 4A).

The IL-1-stimulated phosphorylation of JNK1/JNK2 and p38 α MAPK in MEFs from HOIP[C879S] or NEMO[D311N] mice was similar to wild-type MEFs (Supplementary Figure S5 at <http://www.biochemj.org/bj/461/bj4610531add.htm>), but was suppressed by the TAK1 inhibitors NG25 or 5Z-7-oxozeanol (Supplementary Figure S6 at <http://www.biochemj.org/bj/461/bj4610531add.htm>). These control experiments indicated that activation of the TAK1 complex was unimpaired in MEFs from HOIP[C879S]- or NEMO[D311N]-knockin mice. The TAK1 inhibitor NG25 did not affect the IL-1-stimulated formation of Lys⁶³-linked ubiquitin chains significantly and actually enhanced Met¹-linked ubiquitin chain production in IKK α -deficient MEFs (Supplementary Figure S7 at <http://www.biochemj.org/bj/461/bj4610531add.htm>). Thus TAK1 activity is not required for the IL-1-stimulated formation of Lys⁶³-linked or Met¹-linked ubiquitin chains and NG25 does not suppress the phosphorylation of IKK β by preventing formation of the ubiquitin chains. The enhanced formation of Met¹-linked ubiquitin chains in the presence of NG25 implies the existence of a TAK1-dependent feedback control mechanism for restricting the formation of these ubiquitin chains.

Finally, it should be noted that although TAK1 phosphorylates the IKK β -NEMO complex at Ser¹⁷⁷ in IKK α -deficient MEFs, the active TAK1 catalytic subunit is capable of phosphorylating a catalytically inactive mutant of the IKK β catalytic subunit at Ser¹⁸¹, as well as Ser¹⁷⁷, *in vitro* (Supplementary Figure S8 at <http://www.biochemj.org/bj/461/bj4610531add.htm>). It is therefore possible that the interaction of NEMO with IKK β in the canonical IKK complex and/or the recruitment of the TAK1 complex to Lys⁶³-linked ubiquitin chains are factors that prevent TAK1 from phosphorylating Ser¹⁸¹ in cells.

DISCUSSION

In the present study, we have clarified the mechanism by which the canonical IKK complex is activated. Unexpectedly, we discovered that the activation of IKK β requires two sequential phosphorylation events. The activation process is initiated by the TAK1-catalysed phosphorylation of IKK β at Ser¹⁷⁷, which is a priming event that permits IKK β to phosphorylate itself at Ser¹⁸¹, which is needed before IKK β can phosphorylate exogenous substrates, such as I κ B α (Figure 5). We have shown that this mechanism of activation operates in IL-1- or TNF-stimulated MEFs and in TLR-stimulated BMDMs indicating that is likely to be of general significance. However, the identity of the 'priming' kinase may vary from cell to cell.

The mutation of Ser¹⁷⁷ of IKK β to glutamic acid (to mimic the effect of phosphorylation by introducing a negative charge) permitted the IKK β catalytic subunit to autophosphorylate at Ser¹⁸¹ and this induced activation even in cells that had not been stimulated with IL-1 or TNF. Interestingly, the other two members of the IKK subfamily of protein kinases, IKK ϵ and TBK1 {TANK [TRAF (TNF receptor-associated factor)-associated NF- κ B activator]-binding kinase 1}, both possess a glutamic acid at position 168 in their activation loops, which is the amino acid residue equivalent to Ser¹⁷⁶/Ser¹⁷⁷ of IKK α / β , and they are activated by the phosphorylation of Ser¹⁷², the site equivalent to Ser¹⁸⁰/Ser¹⁸¹ of IKK α / β [8]. These features explain why these IKK-related kinases are not activated directly by TAK1 *in vivo* and why they are instead activated by the canonical IKK complex and by autophosphorylation in response to IL-1 [8]. Once activated, IKK-related kinases restrict the activity of the canonical IKKs by phosphorylating inhibitory sites on the canonical IKKs,

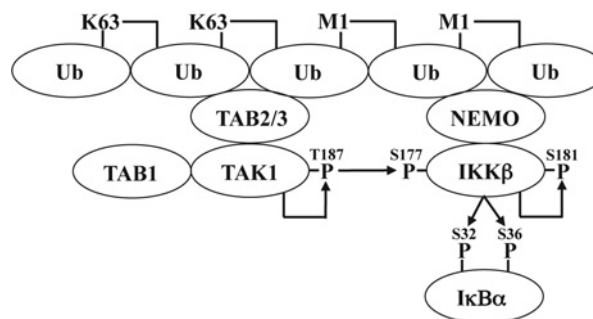


Figure 5 Proposed mechanism for the activation of IKK β by IL-1 in IKK α -deficient MEFs

IL-1 stimulates the formation of hybrid ubiquitin chains in which LUBAC-generated Met¹-linked ubiquitin oligomers are attached covalently to TRAF6-generated Lys⁶³-linked ubiquitin oligomers. The binding of Lys⁶³-linked ubiquitin to TAB2 or TAB3 activates the TAK1 complex by inducing autophosphorylation of the catalytic subunit at Thr¹⁸⁷ [31]. The Met¹-linked ubiquitin chains interact with NEMO permitting TAK1 to phosphorylate IKK β at Ser¹⁷⁷. Phosphorylation of Ser¹⁷⁷ allows autophosphorylation of Ser¹⁸¹. The activated IKK β can then phosphorylate I κ B α . Sites of phosphorylation are denoted by 'P'.

which is critical to prevent autoimmune nephritis in mice [8,24].

The activation of the canonical IKK complex by IL-1 does not just require the phosphorylation of serine residues in the activation loop, but also the formation of a hybrid ubiquitin chain containing both Lys⁶³-linked and Met¹-linked ubiquitin oligomers [16]. The Lys⁶³-linked ubiquitin chains interact with the TAB2 and TAB3 components of TAK1 complexes, inducing the auto-activation of TAK1 [9,13,25], whereas the Met¹-linked ubiquitin chains formed by the action of the E3 ubiquitin ligase LUBAC [16,26] interact with NEMO [27,28] and are critical for activation of the canonical IKK complex [12,16,29]. Nearly all of the Met¹-linked ubiquitin chains formed in response to IL-1 are attached covalently to Lys⁶³-linked ubiquitin chains, which may facilitate the TAK1-dependent activation of canonical IKK complex by recruiting both protein kinases to the same ubiquitin chains [16]. In the present study, we found that the IL-1-stimulated phosphorylation of IKK α /IKK β at Ser¹⁷⁶/Ser¹⁷⁷, and hence the phosphorylation of Ser¹⁸⁰/Ser¹⁸¹, was suppressed in MEFs that were unable to produce Met¹-linked ubiquitin chains or that expressed a ubiquitin-binding-defective mutant of NEMO (Figure 4). Thus the formation of Met¹-linked ubiquitin chains and their interaction with NEMO are both needed for TAK1 to phosphorylate IKK α /IKK β at Ser¹⁷⁶/Ser¹⁷⁷ and so enable the IKKs to complete the activation process by phosphorylating Ser¹⁸⁰/Ser¹⁸¹ (Figure 5).

The activation of many protein kinases requires the phosphorylation of two amino acid residues within their activation loops. For example, similar to the canonical IKK complex, the seven members of the MKK family undergo dual phosphorylation at Ser/Thr-Xaa-Xaa-Xaa-Ser/Thr (where Xaa is any amino acid residue) sequences, enabling them to activate their cognate MAPKs. Similarly, most MAPKs are activated by the dual phosphorylation of a threonine and a tyrosine residue that are located in Thr-Xaa-Tyr within their activation loops. Although the activation of many MKKs and MAPKs is thought to be catalysed by a single protein kinase, the present study has shown that the requirement for one 'upstream' protein kinase does not exclude the possibility that a second protein kinase is also required. Indeed, we have shown that the activation of JNK requires the MKK7-catalysed phosphorylation of the threonine and the MKK4-catalysed phosphorylation of the tyrosine residue within

the Thr-Xaa-Tyr motif [30]. The activation of a kinase by two different 'upstream' kinases provides additional opportunities for signal integration if each activating kinase responds to distinct physiological cues. We suggest that this situation may be a more frequent occurrence than has hitherto been realized, and that this is a neglected area that merits further attention.

AUTHOR CONTRIBUTION

Philip Cohen, Kristopher Clark and Jiazhen Zhang designed the experiments. Jiazhen Zhang performed the experiments. Philip Cohen and Jiazhen Zhang wrote the paper. Toby Lawrence provided the bone marrow from the knockin mice expressing catalytically inactive IKK α .

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SUPPLEMENTARY ONLINE DATA

An unexpected twist to the activation of IKK β : TAK1 primes IKK β for activation by autophosphorylation

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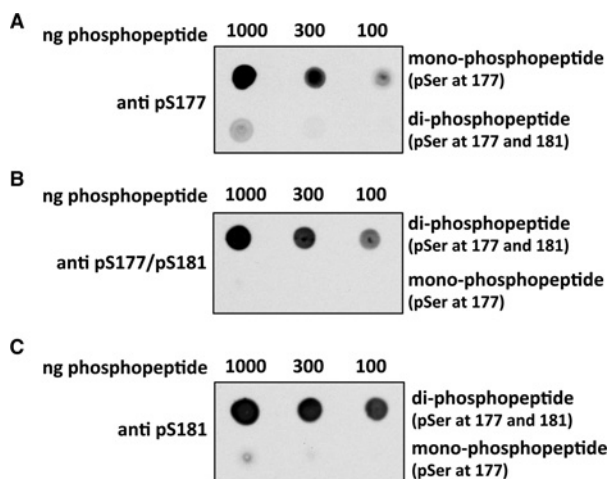


Figure S1 Phosphorylation of Ser¹⁸¹ interferes with the recognition of phospho-Ser¹⁷⁷ by the phospho-specific antibody that recognizes this site

(A) The indicated amounts of phosphopeptides corresponding to amino acid residues 171–187 of IKK β phosphorylated at Ser¹⁷⁷ only or at both Ser¹⁷⁷ and Ser¹⁸¹ were spotted on to nitrocellulose membranes and probed with the phospho-specific antibody recognizing phospho(p)-Ser¹⁷⁷. (B) Same as (A), except that the peptides were immunoblotted with the antibody that only recognizes the di-phosphorylated form of IKK β phosphorylated at both Ser¹⁷⁷ and Ser¹⁸¹. (C) Same as (A), except that the phosphopeptides were immunoblotted with the antibody that recognizes IKK β phosphorylated at Ser¹⁸¹.

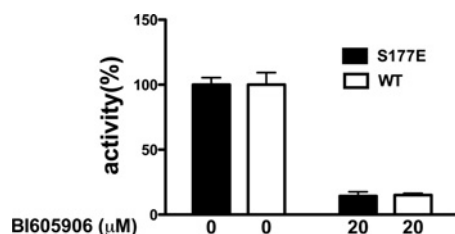


Figure S2 Effect of BI605906 on the activity of wild-type and mutant IKK β

HA-tagged wild-type IKK β (WT) or the IKK β [S177E] mutant (S177E) were expressed in HEK-293 cells, immunoprecipitated from the cell extracts using an anti-HA antibody and assayed for activity in the absence or presence of BI605906. The activities are plotted as a percentage of that obtained in the absence of inhibitor. Results are means \pm S.E.M. of duplicate determinations. Similar results were obtained in another independent experiment.

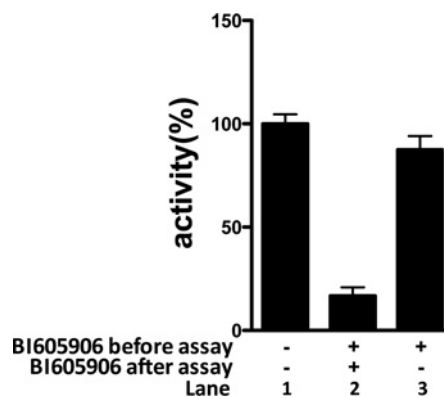


Figure S3 BI605906 is a reversible inhibitor of IKK β

MEFs from IKK α -deficient mice were stimulated for 10 min with 5.0 ng/ml IL-1 and the cells were lysed. The endogenous IKK β was immunoprecipitated from 0.2 mg of cell extract protein and incubated for 1 h at 30 °C without (–, lane 1) or with (+, lanes 2 and 3) 20 μ M BI605906. In lane 3 only, the immunoprecipitates were washed extensively to remove BI605906. All the immunoprecipitates were then assayed for IKK β activity. The Figure shows that IKK β activity was full restored after washing away the inhibitor. Results are means \pm S.E.M. of duplicate determinations. Similar results were obtained in another independent experiment.

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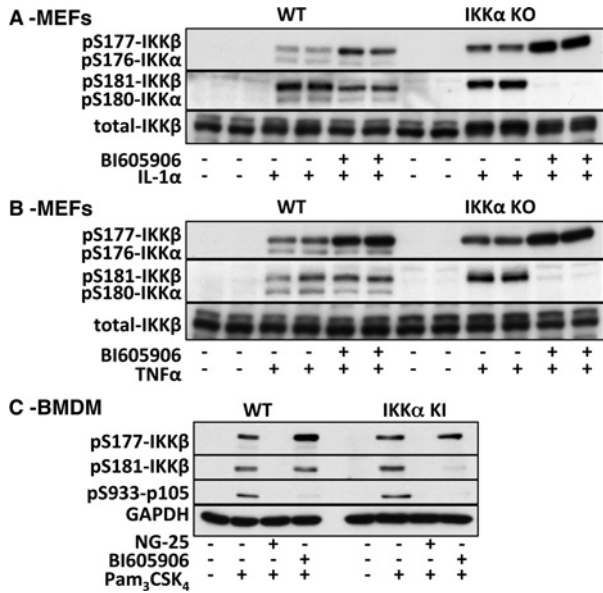


Figure S4 Effect of inhibitors on agonist-stimulated phosphorylation of IKK β at Ser¹⁷⁷ and Ser¹⁸¹ in MEFs and BMDM

(**A** and **B**) MEFs from wild-type (WT) or IKK α -knockout (KO) mice were incubated for 1 h without (–) or with (+) 5.0 μ M BI 605906 and then stimulated for 10 min with 5.0 ng/ml IL-1 (**A**) or 10 ng/ml TNF (**B**). Following cell lysis, cell extract (20 μ g of protein) was denatured in SDS, subjected to SDS/PAGE, and immunoblotted with antibodies that recognize IKK α and IKK β phosphorylated at Ser¹⁷⁶ and Ser¹⁷⁷ respectively, or with antibodies that recognize IKK α and IKK β phosphorylated at Ser¹⁸⁰ or Ser¹⁸¹ respectively. The membranes were also immunoblotted with antibodies that recognize all forms of IKK β . (**C**) Same as (**A** and **B**), except that BMDMs from wild-type (WT) mice and knockin (KI) mice expressing the catalytically inactive mutant of IKK α were used and the cells were stimulated for 10 min with 1.0 μ g/ml Pam₃CSK₄. An antibody recognizing GAPDH (glyceraldehyde-3-phosphate dehydrogenase) was used as a loading control.

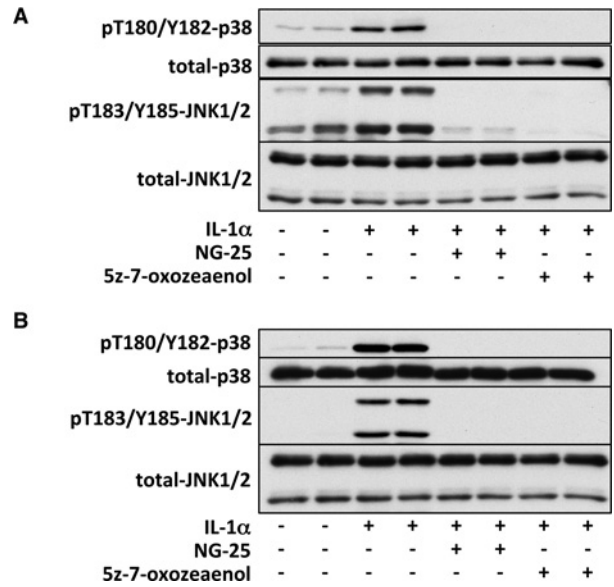


Figure S6 TAK1 inhibitors suppress the IL-1-stimulated phosphorylation of MAPKs in MEFs from HOIP[C879S]- and NEMO[D311N]-knockin mice

(**A**) As in Figure S5, except that before stimulation with IL-1, MEFs from mice expressing the HOIP[C879S] mutant were incubated for 1 h without (–) or with (+) the TAK1 inhibitors NG25 (1.0 μ M) or 5Z-7-oxozeaenol (1.0 μ M) before stimulation with 5.0 ng/ml IL-1 for the times indicated. (**B**) Same as (**A**), except that the experiment was performed with MEFs from mice expressing the polyubiquitin-binding-deficient mutant NEMO[D311N].

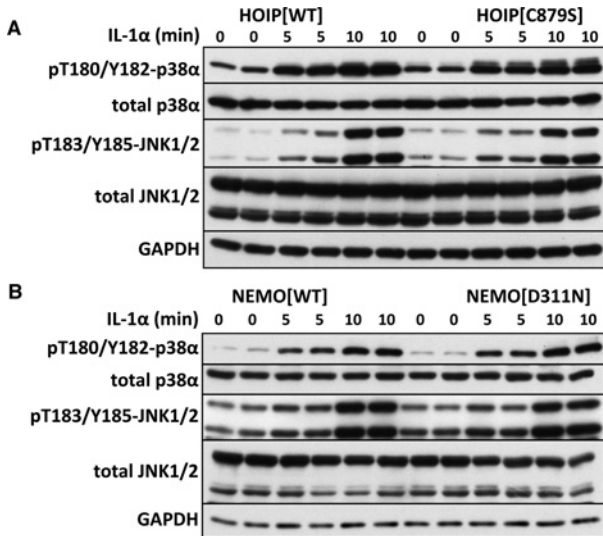


Figure S5 Phosphorylation of MAPKs is unimpaired in MEFs from HOIP[C879S] and NEMO[D311N] mice

(**A**) MEFs from wild-type mice (HOIP[WT]) or knockin mice expressing the inactive HOIP[C879S] mutant were stimulated with 5.0 ng/ml IL-1 for the times indicated. After cell lysis, 20 μ g of cell extract protein was denatured in SDS, subjected to SDS/PAGE and immunoblotted with the antibodies indicated. (**B**) Same as (**A**), but using MEFs from mice expressing the polyubiquitin-binding-deficient mutant of NEMO (NEMO[D311N]). The antibody recognizing pT183/Y185 of JNK1/2 was from Invitrogen and the antibody recognizing pT180/Y182 of p38 α MAPK was from Cell Signaling Technology.

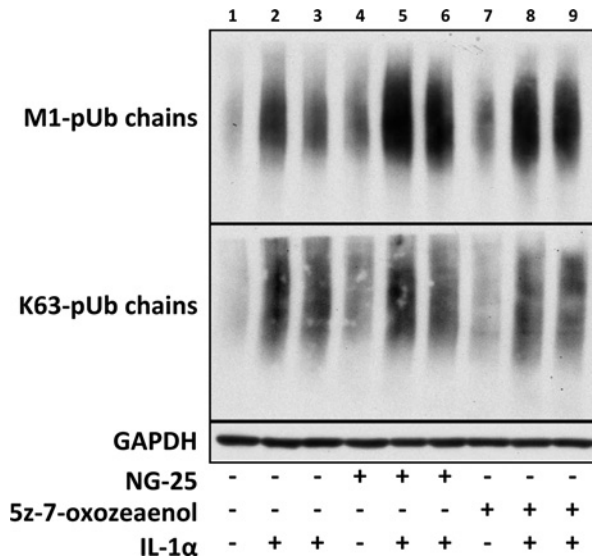


Figure S7 Effect of TAK1 inhibition on the IL-1-stimulated formation of Lys⁶³-linked and Met¹-linked ubiquitin chains

MEFs were incubated for 1 h with (+) or without (-) 2 μ M NG25 or 1 μ M 5Z-7-oxozeaenol, then stimulated for 10 min with 5 ng/ml IL-1 α and lysed. The Met¹-linked and Lys⁶³-linked ubiquitin chains present in 2 mg of cell extract protein were captured on Halo-NEMO [1], released by denaturation in SDS and immunoblotted with antibodies that recognize Met¹-linked or Lys⁶³-linked ubiquitin chains specifically. The same cell extracts (20 μ g of protein) were immunoblotted with an anti-GAPDH antibody as a loading control.

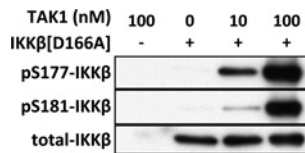


Figure S8 TAK1 phosphorylates IKK β at Ser¹⁷⁷ and Ser¹⁸¹ *in vitro*

Catalytically inactive IKK β [D166A] (0.8 μ M) was incubated for 3 min at 30 °C with the indicated concentrations of the active TAK1-TAB1 fusion protein in 50 mM Tris/HCl (pH 7.5), 0.1 mM EGTA, 2 mM DTT, 10 mM magnesium acetate and 0.1 mM ATP. Reactions were terminated by denaturation in SDS and, after SDS/PAGE and transfer on to PVDF membranes, proteins were immunoblotted with antibodies that recognize IKK β phosphorylated at Ser¹⁷⁷ or Ser¹⁸¹ or antibodies recognizing all forms of IKK β .

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