

# NIH Public Access

**Author Manuscript** 

Curr Opin Allergy Clin Immunol. Author manuscript; available in PMC 2013 May 13

## Published in final edited form as:

*Curr Opin Allergy Clin Immunol.* 2010 December ; 10(6): 587–593. doi:10.1097/ACI. 0b013e32833fd5eb.

# An update on immunotherapy for food allergy

## Amy M. Scurlock, M.D. and Stacie M. Jones, M.D.

University of Arkansas for Medical Sciences, Department of Pediatrics, Division of Allergy and Immunology, Arkansas Children's Hospital, Little Rock, Arkansas

# Abstract

**Purpose of the review**—Recent investigation has resulted in significant advances toward definitive therapeutic options for food allergy. In this review, we will explore novel immunotherapeutic interventions for the active treatment of food allergy.

**Recent findings**—Because the injection route for allergen immunotherapy to foods has been associated with an unacceptable risk of severe anaphylactic reactions, use of mucosally targeted therapeutic strategies is of significant interest for food allergy. Allergen-specific immunotherapeutic approaches such as oral, sublingual, epicutaneous, and peptide immunotherapy have demonstrated efficacy in increasing threshold dose and inducing immunologic changes associated with both desensitization and oral tolerance in animal and human trials. More global immunomodulatory strategies, such as Traditional Chinese Medicine and anti-IgE therapy have been shown to effectively target the allergic response, and clinical trials are ongoing to determine the efficacy and safety in human food allergy.

**Summary**—The advent of therapies that target the mucosal immune response to promote oral tolerance have shown great promise in the treatment of food hypersensitivity. However, there is still significant risk of adverse reactions associated with these therapeutic strategies and further study is needed to carefully advance these therapeutic modalities toward general clinical implementation.

## Keywords

Food allergy; oral tolerance; immunotherapy; T-regulatory cells

# INTRODUCTION

The development of novel therapeutic modalities targeting the mucosal immune response has shown great promise in providing a definitive therapy for food allergy. Because food allergy is likely a multi-factorial disorder with both genetic and environmental influences, development of primary prevention strategies has been frustrating and at times counterproductive, making development of a definitive therapeutic option a high priority. In this review, we will examine the relationship between food hypersensitivity and oral tolerance and explore novel therapeutic approaches to modulate the food allergic response.

Corresponding Author: Stacie M. Jones, M.D., 13 Children's Way, Slot 512-13, Little Rock, Arkansas 72202, (501) 364-1060, jonesstaciem@uams.edu.

**Publisher's Disclaimer:** This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

# FOOD HYPERSENSITIVITY AND ORAL TOLERANCE

A diagnosis of food allergy is challenging for affected patients and families not only due to the medical implications, but also due to psychosocial and economic stressors. The standard of care for immediate food hypersensitivity currently includes dietary allergen restriction and ready access to emergency medications in case of accidental exposure; however, there are presently no widely available, active therapeutic options for food allergic patients. Because of the need for stringent dietary restrictions, difficulty comprehending food labels, [1;2] the continual threat of accidental ingestions,[3] and the risk of severe or fatal reactions, [4;5] a diagnosis of food allergy results in significant anxiety, psychosocial stress, economic burden, and reduced health-related quality of life.[6–11; 11]

Investigators are continually working to delineate the precise immunologic, genetic and environmental factors that promote food allergy. The current evidence indicates that food allergy is the consequence of either a failure to establish oral tolerance or an interruption of existing tolerance, resulting in dysregulated T-helper type 2 (Th2) responses and immediate hypersensitivity reactions upon antigen re-exposure. As such, aberrant regulatory T-cell (Treg) induction appears to be a key element in the development of food allergy. [12–15]

The prevalence of food allergy continues to escalate in developed countries; current estimates suggest an overall prevalence of 4% in the United States. [16\*\*] Both peanut and tree nut allergies have increased by 2-3 fold over the past decade, [17;18\*] while the prevalence of peanut allergy has tripled in the United Kingdom. [19;20] Attempts to avert the development of food allergy through primary prevention strategies such as early dietary allergen restriction and modified timing of complementary "solid" food introduction to infants have proven frustrating and possibly counter-productive. For example, approximately a decade ago, both the United Kingdom Committee on Toxicity (COT) and the American Academy of Pediatrics (AAP) recommended early dietary restriction of peanuts to avoid sensitization.[21;22] However, subsequent data suggested that early consumption of food proteins and subsequent oral tolerance induction in infants and toddlers may be a key element of preventing the development of food allergies. [23;24] Children in countries that have peanut snacks that are safe for infants have relatively low rates of peanut allergies.[23] Additionally, despite earlier introduction of peanut protein into the diet, Jewish children in Israel had a 10-fold lower prevalence of peanut allergy compared with children of similar genetic background in the United Kingdom. [24] A randomized controlled trial (Learning Early About Peanut Allergy (LEAP) Study) is underway in the United Kingdom to compare the efficacy of early peanut protein consumption versus peanut avoidance in preventing peanut allergy. Studies have shown that neither the diversity, nor the timing of introduction of complementary foods had any association with development of eczema, [25] and delayed introduction of complementary foods did not protect from asthma or atopic disease.[26] The most recent AAP recommendations for high risk infants do not endorse restriction of maternal diet during pregnancy and lactation or restriction of allergenic foods in infants after 4-6 months of age.[27] European guidelines suggest similar dietary recommendations.[28]

# NOVEL THERAPEUTIC INTERVENTIONS TARGETING FOOD HYPERSENSITIVITY

Both allergen specific therapies that harness mucosal tolerance to abrogate the allergic response and more generalized immunomodulatory approaches are under investigation in animal and human models. The goals of these therapies are generally to induce some combination of desensitization and/or tolerance. *Desensitization* is defined as a change in threshold dose of ingested food allergen necessary to cause allergic symptoms; this state is

dependent on ongoing antigen exposure. Mechanistic markers of desensitization include increased IgG4 and reduced IgE, as well as decreased activation and release of inflammatory mediators by mast cells and basophils. In contrast, *tolerance* is the induction of long-term immunologic changes associated with the ability to ingest a food without symptoms and without ongoing therapy. Mechanisms of tolerance induction include active modulation of the immune response to promote regulatory T-cell development and immunologic skewing away from a Th2 response. (Table 1)

#### Alternative approaches to traditional injection immunotherapy

Allergen immunotherapy via the injection route has been utilized successfully for the treatment of allergic rhinoconjunctivitis and venom hypersensitivity for decades. [29] Despite its efficacy in treating allergic rhinoconjunctivitis and venom hypersensitivity, the "traditional" approach to allergen immunotherapy via the subcutaneous route is impractical and unsafe for treatment of food allergy due to an unacceptably high rate of anaphylactic reactions. [30;31] At present, multiple therapeutic alternatives to subcutaneous injection therapy are being investigated for treatment of food allergy. (Table 2)

#### **Traditional Chinese Medicine**

Most investigators have utilized an allergen-specific approach to target food allergy. However, an innovative approach with potential for the treatment of food allergy utilizes Traditional Chinese Medicine, thus providing an approach that is not allergen specific. In mouse models, Food Allergy Herbal Formula (FAHF-2) has been shown to promote tolerance and protection from anaphylaxis.[32–34\*] Human clinical trials are just beginning and hold promise for future clinical efficacy.

#### Humanized Monoclonal Anti-IgE Therapy

Omalizumab, a recombinant, humanized, monoclonal anti-IgE antibody has been utilized effectively in concert with rush immunotherapy for allergic rhinitis[35] and as an adjunctive therapy to minimize systemic immunotherapy reactions in patients with allergic asthma.[36] Anti-IgE therapy (TNX-901) was previously demonstrated to significantly increase the threshold peanut protein dose at oral food challenge from 178 mg to 2805 mg affording treated individuals with potential protection from accidental peanut ingestions.[37] Clinical trials are in progress evaluating both anti-IgE monotherapy for food allergy, as well as use of omalizumab as an adjunct to oral immunotherapy (OIT).

#### Peptide Immunotherapy

In mouse models, rectal immunization with mutated peanut protein allergens has been shown to protect mice from anaphylaxis.[38;39] Peptide immunotherapy has also been utilized with the immunodominant epitopes of OVA.[40\*] Mice treated with subcutaneous injections of peptides were protected from anaphylaxis upon OVA challenge, in addition to exhibiting decreased serum histamine levels, decreased OVA-specific IgE, reduced Th2 cytokines and increased IFN $\gamma$ . Additionally, animals that received peptide immunotherapy showed significantly higher levels of mRNA transcripts for Foxp3 and TGF $\beta$  in the intestine, suggesting modification of the local mucosal immune response in the target tissue. Studies using a similar approach are currently in early human trials.

#### **Epicutaneous Immunotherapy**

Epicutaneous immunotherapy (EPIT) has been utilized for treatment of allergic rhinitis in humans[41\*\*] and in mouse models of inhalant and food allergy.[42\*] Animal and ex vivo skin models suggest that EPIT targets Langerhans cells and dermal dendritic cells to modulate the immune response.[43] A recent pilot study of EPIT in milk allergic children

suggested that this therapy was overall well-tolerated and did not result in sensitization. [44\*\*] Clear clinical efficacy was not demonstrated in this study, likely due to the short treatment period of only 3 months, but trends toward improvement were noted in the active treatment group. It is notable that immunotherapy using this delivery system on intact skin did not result in sensitization. Phase I human trials are in progress in Europe and the United States.

#### Sublingual Immunotherapy

Sublingual immunotherapy (SLIT) has shown broad efficacy for treatment of inhalant allergies. SLIT employs a liquid concentrate administered under the tongue that is given in small, increasing doses of antigen in a controlled setting usually during an initial dose coupled with home dosing to reach a maximum tolerated maintenance dose of allergen. In clinical trials, treatment is followed by an oral food challenge with antigen or placebo to determine efficacy.

Investigators have utilized SLIT for treatment of hazelnut allergy[45;46] and in a single case for treatment of life-threatening kiwi allergy. [47;48] Trials are currently in progress evaluating the efficacy of SLIT for other food allergens including peanut and milk.

#### Oral Immunotherapy (OIT) and Specific Oral Tolerance Induction

Investigation of OIT as a therapeutic modality has yielded promising results for a variety of food allergies. OIT appears to be effective in inducing desensitization in most subjects, as well as oral tolerance in a subset of patients with food allergy. OIT generally involves the use of a powdered food protein given orally, often in a vehicle food. The usual approach to OIT involves an initial dosage escalation phase followed by observed build-up dosing to daily maintenance therapy. Therapeutic effect is evaluated by food challenge at standard points in the treatment protocol.

Investigators have utilized a standardized OIT protocol for treatment of food allergies including, most commonly, milk, egg, and fish, and described successful desensitization in 77% of treated subjects.[49] Other studies utilized specific oral tolerance induction therapy to desensitize children with IgE-mediated milk hypersensitivity.[50–53]

Buchanan and colleagues utilized a 24-month egg OIT protocol to desensitize egg allergic children; in the 7 subjects who completed the 24-month protocol, 4 of 7 passed a double blind, placebo controlled food challenge to 10 g of egg at the conclusion of the therapy, and all subjects tolerated significantly higher doses of egg protein than noted at entry into the study.[54] A subsequent report indicated that 2/21 subjects enrolled in this ongoing protocol were unable to achieve the maintenance egg protein dose due to frequent and unacceptable therapy-associated adverse reactions, highlighting that this therapy is not ready for broad implementation into routine clinical practice settings.[55] Ongoing clinical trials continue to examine the safety, efficacy, and mechanism of egg OIT.

Another study suggested that consumption of heated egg in egg-allergic individuals may have an immunomodulatory therapeutic effect. [56\*] Subjects with IgE-mediated egg allergy underwent physician-supervised oral food challenges to extensively heated egg (e.g. muffin or waffle). Subjects tolerating heated egg protein challenge integrated heated egg into their diets. Continued consumption of heated egg protein in the diet was associated with decreased skin test size, reduced egg-specific IgE levels and increased IgG4 levels.

In a recent randomized, double-blind placebo controlled cow's milk OIT trial, investigators treated nineteen children with cow's milk allergy. [57\*\*] Treatment with OIT was associated with increased median milk threshold dose inducing allergic symptoms during

oral food challenge (40 mg $\rightarrow$ 5140 mg OIT vs. 40 mg $\rightarrow$ 40 mg placebo). No significant difference in milk-specific IgE levels was detected in the treated vs. untreated groups; however, milk-specific IgG4 was significantly increased. This cohort was subsequently monitored during an open label portion of the study to evaluate the continued safety of milk OIT and the subjects' ability to tolerate gradual home dose escalation.[58\*] Six of thirteen patients undergoing follow-up food challenge tolerated the maximum cumulative dose of 16,000 mg (16 oz) of cow's milk protein without any adverse reaction. The other 7 participants tolerated doses ranging from 3000 mg to 16,000 mg, with associated clinical symptoms including oral pruritus, abdominal pain, sneezing, cough and urticaria. Significant decreases in end-point titration skin prick testing and milk specific IgE and significant increases in milk-specific IgG4 were detected following OIT.

## **Oral Immunotherapy in Peanut Allergic Subjects**

Preliminary trials of OIT in pediatric patients with peanut allergy have yielded encouraging results regarding the safety and efficacy of this therapy. Our group utilized an OIT protocol to treat children with peanut allergy and evaluate both clinical efficacy and immunologic changes in treated subjects.[59\*] Twenty-nine children completed the protocol and 27/29 (93%) ingested the maximal amount (5 g) of peanut protein during oral challenge while receiving OIT, supporting the role of OIT in effective clinical desensitization. Another group reported similar clinical effectiveness in four patients who received OIT using a similar protocol. [60\*] In our cohort, clinical effectiveness was correlated with reduced titrated skin prick test reactivity and decreased basophil activation.[59\*] Treated patients demonstrated initial increases in peanut-specific IgE which subsequently decreased by 12 and 18 months on therapy, whereas peanut-specific IgG4 increased significantly throughout the study. Additionally, a 1.5 fold increase in FOXP3+ Tregs was noted in peanut stimulated cells at 6 and 12 months on therapy. T-cell microarray data revealed down-regulation of genes in apoptotic pathways in subjects while on OIT. A related study examined the safety of OIT throughout all phases of treatment.[61\*] Allergic reactions occurred most frequently on the initial escalation day with the majority of patients requiring some form of treatment. The likelihood of allergic reactions decreased significantly during the build-up and home dosing phases; however, two subjects received epinephrine on one occasion each during home dosing. Further examination of adverse reactions during home dosing associated increased risk of reaction with concurrent illness, physical exertion following dose administration, dosing during menses, poorly controlled asthma and timing of dosing following food ingestion.[62\*]

A newly published study employed a peanut OIT protocol that included a 7-day rush OIT treatment phase, followed by a long-term build-up protocol with bi-weekly dose increases up to 0.5 grams of peanut protein, and a subsequent 8 week maintenance phase.[63\*\*] Twenty-three subjects underwent the rush OIT protocol, with 22/23 continuing the long-term treatment protocol. The median threshold dose eliciting symptoms after rush OIT was 0.15 grams, whereas, after long-term treatment the median tolerated dose was 1 gram. From a safety standpoint, the authors reported that 2.6% of the total 6137 doses elicited mild to moderate symptoms, while 1.3% of doses resulted in pulmonary obstruction; OIT was discontinued in 4 subjects. Immunologic changes while on OIT included significant increases in peanut specific IgG4, and significant decreases in Th2 cytokine production by PBMCs.

Although clinical safety and mechanistic evaluation are of utmost priority when designing clinical food allergy trials, recent data suggests psychological factors that influence enrollment should be considered. Dunngalvin, et al. evaluated the factors that influence parental decision to enroll a child in a potentially risky therapeutic trial, reporting that parents of children with food allergy who elected to enroll in immunotherapy trials

perceived a significantly higher likelihood of their child having a severe reaction or dying if a food was ingested. (OR 6.753) [64\*] Participation in immunotherapy trials could be predicted with 90% accuracy using this model. These data suggest that the design of future immunotherapy trials for food allergy should focus not only on stringent clinical safety regulations, efficacy, and mechanistic evaluation, but should also consider psychological factors that influence enrollment and employ strategies to eliminate unintentional coercion, and possibly selection bias, when enrolling potentially high risk families.

# CONCLUSION

Advances in our understanding of the immunologic mechanisms underlying food allergy and of the elegant complexities of the mucosal immune response have resulted in substantial progress toward definitive therapeutic options for food allergic individuals. Current therapeutic strategies are focused on harnessing oral tolerance to modulate the allergic response using antigen specific modalities, while others, such as Traditional Chinese medicine and monoclonal anti-IgE therapy utilize a more global immunomodulatory approach. Clinical trials are ongoing to address these issues through the NIH Consortium of Food Allergy Research (CoFAR) and others with trial details available at www.clinicaltrials.gov. The current advances have brought us into an exciting era with regard to food allergy therapy. We are on the cusp of definitive therapeutic options, providing hope and optimism for food allergic patients and families. However, it should be noted that these approaches have significant associated risk and at present should only be conducted by experienced investigators in clinical trials centers. Ongoing studies will carefully move toward broader clinical application in the future.

## REFERENCES

- Altschul AS, Scherrer DL, Munoz-Furlong A, Sicherer SH. Manufacturing and labeling issues for commercial products: relevance to food allergy. J.Allergy Clin.Immunol. 2001; 108:468. [PubMed: 11544472]
- Joshi P, Mofidi S, Sicherer SH. Interpretation of commercial food ingredient labels by parents of food-allergic children. Journal of Allergy and Clinical Immunology. 2002; 109:1019–1021. [PubMed: 12063534]
- Sicherer SH, Burks AW, Sampson HA. Clinical Features of Acute Allergic Reactions to Peanut and Tree Nuts in Children. Pediatrics. 1998; 102:e6. [PubMed: 9651458]
- Bock SA, Munoz-Furlong A, Sampson HA. Further fatalities caused by anaphylactic reactions to food: 2001–2006. Journal of Allergy and Clinical Immunology. 2007; 119:1016–1018. [PubMed: 17306354]
- Clark S, Bock SA, Gaeta TJ, Brenner BE, Cydulka RK, Camargo CA. Multicenter study of emergency department visits for food allergies. Journal of Allergy and Clinical Immunology. 2004; 113:347–352. [PubMed: 14767453]
- Sicherer SH, Noone SA, Munoz-Furlong A. The impact of childhood food allergy on quality of life. Ann.Allergy Asthma Immunol. 2001; 87:461–464. [PubMed: 11770692]
- Cohen BL, Noone S, Munoz-Furlong A, Sicherer SH. Development of a questionnaire to measure quality of life in families with a child with food allergy. Journal of Allergy and Clinical Immunology. 2004; 114:1159–1163. [PubMed: 15536425]
- Bollinger ME, Dahlquist LM, Mudd K, Sonntag C, Dillinger L, McKenna K. The impact of food allergy on the daily activities of children and their families. Ann.Allergy Asthma Immunol. 2006; 96:415–421. [PubMed: 16597075]
- DunnGalvin A, de BlokFlokstra BMJ, Burks AW, Dubois EJ, Hourihane JO'B. Food allergy QoL questionnaire for children aged 0–12 years: content, construct, and cross-cultural validity. Clinical & Experimental Allergy. 2008; 38:977–988. [PubMed: 18435800]
- King RM, Knibb RC, Hourihane JO'B. Impact of Peanut Allergy on Quality of Life, Stress and Anxiety in the Family. Allergy. 2009; 64:461–468. [PubMed: 19076542]

- Sladkevicius E, Nagy E, Lack G, Guest JF. Resource implications and budget impact of managing cow milk allergy in the UK. Journal of Medical Economics. 2010; 13:119–128. [PubMed: 20092426]
- Beyer K, Castro R, Birnbaum A, Benkov K, Pittman N, Sampson HA. Human milk-specific mucosal lymphocytes of the gastrointestinal tract display a TH2 cytokine profile. Journal of Allergy and Clinical Immunology. 2002; 109:707–713. [PubMed: 11941323]
- Curotto de Lafaille MA, Kutchukhidze N, Shin S, Ding Y, Yee H, Lafaille JJ. Adaptive Foxp3+ regulatory T cell-dependent and -independent control of allergic inflammation. Immunity. 2008; 29:114–126. [PubMed: 18617425]
- Karlsson MR, Rugtveit J, Brandtzaeg P. Allergen-responsive CD4+CD25+ Regulatory T Cells in Children who Have Outgrown Cow's Milk Allergy. J.Exp.Med. 2004; 199:1679–1688. [PubMed: 15197226]
- Shreffler WG, Wanich N, Moloney M, Nowak-Wegrzyn A, Sampson HA. Association of allergenspecific regulatory T cells with the onset of clinical tolerance to milk protein. Journal of Allergy and Clinical Immunology. 2009; 123:43–52. [PubMed: 19130927]
- 16. Branum AM, Lukacs SL. Food Allergy Among Children in the United States. Pediatrics. 2009; 124:1549–1555. [PubMed: 19917585] This study evaluates food allergy prevalence and health care resource utilization using data from several national surveys and suggests increased prevalence and awareness of food allergy.
- Sicherer SH, Munoz-Furlong A, Sampson HA. Prevalence of peanut and tree nut allergy in the United States determined by means of a random digit dial telephone survey: A 5-year follow-up study. Journal of Allergy and Clinical Immunology. 2003; 112:1203–1207. [PubMed: 14657884]
- 18. Sicherer SH, Munoz-Furlong A, Godbold JH, Sampson HA. US prevalence of self-reported peanut, tree, nut sesame allergy: 11-year follow-up. Journal of Allergy and Clinical Immunology. 2010; 125:1322–1326. [PubMed: 20462634] Using a random-digit dial phone survey, these investigators estimated the prevalence of peanut, tree nut and sesame allergy and determined that peanut and tree nut allergy continue to increase.
- Grundy J, Matthews S, Bateman B, Dean T, Arshad SH. Rising prevalence of allergy to peanut in children: Data from 2 sequential cohorts. Journal of Allergy and Clinical Immunology. 2002; 110:784–789. [PubMed: 12417889]
- Hourihane JO, Aiken R, Briggs R, Gudgeon LA, Grimshaw KEC, DunnGalvin A, Roberts SR. The impact of government advice to pregnant mothers regarding peanut avoidance on the prevalence of peanut allergy in United Kingdom children at school entry. Journal of Allergy and Clinical Immunology. 2007; 119:1197–1202. [PubMed: 17353036]
- 21. Committee on Toxicity. COT report on peanut allergy. 1998. http://cot.food.gov.uk/cotreports/ cotwgreports/cotpeanutallergy
- 22. Committee on Nutrition. Hypoallergenic Infant Formulas. Pediatrics. 2000; 106:346–349. [PubMed: 10920165]
- 23. Levy Y, Broides A, Segal N, Danon YL. Peanut and tree nut allergy in children: role of peanut snacks in Israel? Allergy. 2003; 58:1206–1207. [PubMed: 14616145]
- 24. Du Toit G, Katz Y, Sasieni P, Mesher D, Maleki SJ, Fisher HR, Fox AT, Turcanu V, Amir T, Zadik-Mnuhin G, Cohen A, Livne I, Lack G. Early consumption of peanuts in infancy is associated with a low prevalence of peanut allergy. Journal of Allergy and Clinical Immunology. 2008; 122:984–991. [PubMed: 19000582]
- 25. Filipiak B, Zutavern A, Koletzko S, von Berg A, Brockow I, Grnbl A, Berdel D, Reinhardt D, Bauer CP, Wichmann HE, Heinrich J. Solid Food Introduction in Relation to Eczema: Results from a Four-Year Prospective Birth Cohort Study. The Journal of Pediatrics. 2007; 151:352–358. [PubMed: 17889067]
- 26. Zutavern A, Brockow I, Schaaf B, von Berg A, Diez U, Borte M, Kraemer U, Herbarth O, Behrendt H, Wichmann HE, Heinrich J. LISA Study Group. Timing of Solid Food Introduction in Relation to Eczema, Asthma, Allergic Rhinitis, and Food and Inhalant Sensitization at the Age of 6 Years: Results From the Prospective Birth Cohort Study LISA. Pediatrics. 2008; 121:e44–e52. [PubMed: 18166543]

- 27. Greer FR, Sicherer SH, Burks AW. the Committee on Nutrition and Section on Allergy and Immunology. Effects of Early Nutritional Interventions on the Development of Atopic Disease in Infants and Children: The Role of Maternal Dietary Restriction, Breastfeeding, Timing of Introduction of Complementary Foods, and Hydrolyzed Formulas. Pediatrics. 2008; 121:183–191. [PubMed: 18166574]
- 28. Agostoni C, Decsi T, Fewtrell M, Goulet O, Kolacek S, Koletzko B, Michaelsen KF, Moreno L, Puntis J, Rigo J, Shamir R, Szajewska H, Turck D, van Goudoever J. ESPGHAN Committee on Nutrition. Complementary Feeding: A Commentary by the ESPGHAN Committee on Nutrition. Journal of Pediatric Gastroenterology & Nutrition. 2008; 46:99–110. [PubMed: 18162844]
- Francis JN, James LK, Paraskevopoulos G, Wong C, Calderon MA, Durham SR, Till SJ. Grass pollen immunotherapy: IL-10 induction and suppression of late responses precedes IgG4 inhibitory antibody activity. Journal of Allergy and Clinical Immunology. 2008; 121:1120–1125. [PubMed: 18374405]
- 30. Oppenheimer JJ, Nelson HS, Bock SA, Christensen F, Leung DY. Treatment of peanut allergy with rush immunotherapy. 1992; 90:256–262.
- Nelson HS, Lahr J, Rule R, Bock SA, Leung DY. Treatment of anaphylactic sensitivity to peanuts by immunotherapy with injections of aqueous peanut extract. J.Allergy Clin.Immun. 1997; 99:744–751. [PubMed: 9215240]
- 32. Srivastava KD, Kattan JD, Zou ZM, Li JH, Zhang L, Wallenstein S, Goldfarb J, Sampson HA, Li XM. The Chinese herbal medicine formula FAHF-2 completely blocks anaphylactic reactions in a murine model of peanut allergy. Journal of Allergy and Clinical Immunology. 2005; 115:171–178. [PubMed: 15637565]
- 33. Qu C, Srivastava K, Ko J, Zhang TF, sampson HA, Li XM. Induction of tolerance after establishment of peanut allergy by the food allergy herbal formula-2 is associated with upregulation of interferon-gamma. Clin.Exp.Allergy. 2007; 37:846–855. [PubMed: 17517098]
- 34. Srivastava KD, Qu C, Zhang T, Goldfarb J, Sampson HA, Li XM. Food Allergy Herbal Formula-2 silences peanut-induced anaphylaxis for a prolonged posttreatment period via IFN-gamma-producing CD8+ T cells. Journal of Allergy and Clinical Immunology. 2009; 123:443–451. [PubMed: 19203662] Using a mouse model of peanut allergy, FAHF-2 demonstrated prolonged clinical efficacy in protection from anaphylaxis associated with mechanistic changes including increased IFN-gamma production from CD8+ T-cells.
- 35. Casale TB, Busse WW, Kline JN, Ballas ZK, Moss MH, Townley RG, Mokhtarani M, Seyfert-Margolis V, Asare A, Bateman K, Deniz Y. the Immune Tolerance Network Group. Omalizumab pretreatment decreases acute reactions after rush immunotherapy for ragweed-induced seasonal allergic rhinitis. Journal of Allergy and Clinical Immunology. 2006; 117:134–140. [PubMed: 16387596]
- Massanari M, Nelson H, Casale T, Busse W, Kianifard F, Geba GP, Zeldin RK. Effect of pretreatment with omalizumab on the tolerability of specific immunotherapy in allergic asthma. Journal of Allergy and Clinical Immunology. 2010; 125:383–389. [PubMed: 20159249]
- Leung DYM, Sampson HA, Yunginger JW, Burks AW Jr, Schneider LC, Wortel CH, Davis FM, Hyun JD, Shanahan WR Jr. the TNX. Effect of Anti-IgE Therapy in Patients with Peanut Allergy. N Engl J Med. 2003; 348:986–993. [PubMed: 12637608]
- 38. Li XM, Srivastava K, Huleatt JW, Bottomly K, Burks AW, Sampson HA. Engineered Recombinant Peanut Protein and Heat-Killed Listeria monocytogenes Coadministration Protects Against Peanut-Induced Anaphylaxis in a Murine Model. J.Immunol. 2003; 170:3289–3295. [PubMed: 12626588]
- Li XM, Srivastava K, Grishin A, Huang CK, Schofield B, Burks W, Sampson HA. Persistent protective effect of heat-killed Escherichia coli producing "engineered," recombinant peanut proteins in a murine model of peanut allergy. Journal of Allergy and Clinical Immunology. 2003; 112:159–167. [PubMed: 12847493]
- 40. Yang M, Yang C, Mine Y. Multiple T cell epitope peptides suppress allergic responses in an egg allergy mouse model by the elicitation of forkhead box transcription factor 3- and transforming growth factor-ß-associated mechanisms. Clin.Exp.Allergy. 2010; 40:668–678. [PubMed: 20082619] These investigators demonstrated that peptide immunotherapy was efficacious in promoting both systemic and local mucosal tolerance in egg-allergic mice.

- 41. Senti G, Graf N, Haug S, Rnedi N, von Moos S, Sonderegger T, Johansen P, Knndig TM. Epicutaneous allergen administration as a novel method of allergen-specific immunotherapy. Journal of Allergy and Clinical Immunology. 2009; 124:997–1002. [PubMed: 19733905] This study demonstrates the safety and clinical efficacy of a novel epidermal delivery system in treating grass pollen allergy in adult patients.
- 42. Mondoulet L, Dioszeghy V, Ligouis M, Dhelft V, DuPont C, Benhamou PH. Epicutaneous immunotherapy on intact skin using a new delivery system in a murine model of allergy. Clin.Exp.Allergy. 2010; 40(4):659–667. [PubMed: 20002446] This proof of concept study demonstrated that EPIT on intact skin was efficacious in mouse models of inhalant and food allergy.
- 43. Sparber F, Tripp CH, Hermann M, Romani N, Stoitzner P. Langerhans cells and dermal dendritic cells capture protein antigens in the skin: Possible targets for vaccination through the skin. Immunobiology. 2010 In press.
- 44. Dupont C, Kalach N, Soulaines P, Legoue-Morillon S, Piloquet H, Benhamou PH. Cow's milk epicutaneous immunotherapy in children: A pilot trial of safety, acceptability, and impact on allergic reactivity. Journal of Allergy and Clinical Immunology. 2010; 125:1165–1167. [PubMed: 20451043] This study demonstrates that EPIT is safe, well-tolerated, and does not induce sensitization in milk allergic children. A trend toward clinical efficacy was noted, but did not reach statistical significance most likely due to the short observation period.
- 45. Enrique E, Pineda F, Malek T, Bartra J, Basagana M, Tella R, Castello JV, Alonso R, de Mateo JA, Cerda-Trias T, San Miguel-Moncin Mdel M, Monzon S, Garcia M, Palacios R, Cistero-Bahima A. Sublingual immunotherapy for hazelnut food allergy: A randomized, double-blind, placebo-controlled study with a standardized hazelnut extract. Journal of Allergy and Clinical Immunology. 2005; 116:1073–1079. [PubMed: 16275379]
- Enrique E, Malek T, Pineda F, Palacios R, Bartra J, Tella R, Basagana M, Alonso R, Cistero-Bahima A. Sublingual immunotherapy for hazelnut food allergy: a follow-up study. Ann.Allergy Asthma Immunol. 2008; 100:283–284. [PubMed: 18429351]
- Mempel M, Rakoski J, Ring J, Ollert M. Severe anaphylaxis to kiwi fruit: Immunologic changes related to successful sublingual allergen immunotherapy. Journal of Allergy and Clinical Immunology. 2003; 111:1406–1409. [PubMed: 12789247]
- Kerzl R, Simonowa A, Ring J, Ollert M, Mempel M. Life-threatening anaphylaxis to kiwi fruit: Protective sublingual allergen immunotherapy effect persists even after discontinuation. Journal of Allergy and Clinical Immunology. 2007; 119:507–508. [PubMed: 17125821]
- 49. Patriarca G, Nucera E, Roncallo C, Pollastrini E, Bartolozzi F, DePasquale T, Buononomo A, Gasbarrini G, DiCampali C, Schiavino D. Oral desensitizing treatment in food allergy: clinical and immunological results. Aliment.Pharmacol.Ther. 2003; 17:459–465. [PubMed: 12562461]
- Staden U, Rolinck-Werninghaus C, Brewe F, Wahn U, Niggemann B, Beyer K. Specific oral tolerance induction in food allergy in children: efficacy and clinical patterns of reaction. Allergy. 2007; 62:1261–1269. [PubMed: 17919140]
- Meglio P, Bartone E, Plantamura M, Arabito E, Giampietro PG. A protocol for oral desensitization in children with IgE-mediated cow's milk allergy. Allergy. 2004; 59:980–987. [PubMed: 15291907]
- 52. Staden U, Blumchen K, Blankenstein N, Dannenberg N, Ulbricht H, Dobberstein K, Ziegert M, Niggemann B, Wahn U, Beyer K. Rush oral immunotherapy in children with persistent cow's milk allergy. Journal of Allergy and Clinical Immunology. 2008; 122:418–419. [PubMed: 18602681]
- 53. Longo G, Barbi E, Berti I, Meneghetti R, Pittalis A, Ronfani L, Ventura A. Specific oral tolerance induction in children with very severe cow's milk-induced reactions. Journal of Allergy and Clinical Immunology. 2008; 121:343–347. [PubMed: 18158176]
- Buchanan AD, Green TD, Jones SM, Scurlock AM, Christie L, Althage KA, Steele PH, Pons L, Helm RM, Lee LA, Burks AW. Egg oral immunotherapy in nonanaphylactic children with egg allergy. Journal of Allergy and Clinical Immunology. 2007; 119:199–205. [PubMed: 17208602]
- Burks AW, Jones SM. Egg oral immunotherapy in nonanaphylactic children with egg allergy: Follow-up. Journal of Allergy and Clinical Immunology. 2008; 121:270–271. [PubMed: 17889930]

- 56. Lemon-Mule H, Sampson HA, Sicherer SH, Shreffler WG, Noone S, Nowak-Wegrzyn A. Immunologic changes in children with egg allergy ingesting extensively heated egg. Journal of Allergy and Clinical Immunology. 2008; 122:977–983. [PubMed: 18851876] This study demonstrates that extensively heated egg protein ingestion by a subset of egg allergic children has immunomodulatory effects that are comparable to results seen with acquisition of tolerance to egg protein.
- 57. Skripak JM, Nash SD, Rowley H, Brereton NH, Oh S, Hamilton RG, Matsui EC, Burks AW, Wood RA. A randomized, double-blind, placebo-controlled study of milk oral immunotherapy for cow's milk allergy. Journal of Allergy and Clinical Immunology. 2008; 122:1154–1160. [PubMed: 18951617] These investigators utilized a standardized OIT protocol to treat milk-allergic children and demonstrated increased median threshold dose to elicit clinical symptoms as well as immunologic changes consistent with effective specific immunotherapy.
- 58. Narisety SD, Skripak JM, Steele P, Hamilton RG, Matsui EC, Burks AW, Wood RA. Open-label maintenance after milk oral immunotherapy for IgE-mediated cow's milk allergy. Journal of Allergy and Clinical Immunology. 2009; 124:610–612. [PubMed: 19665770] This long-term follow-up study monitored a cohort of milk-allergic individuals who had received OIT and demonstrated that these subjects could tolerate gradually increasing home doses and that long-term therapy resulted in decreased IgE and increased IgG4.
- 59. Jones SM, Pons L, Roberts JL, Scurlock AM, Perry TT, Kulis M, Shreffler WG, Steele P, Henry KA, Adair M, Francis JM, Durham S, Vickery BP, Zhong X, Burks AW. Clinical efficacy and immune regulation with peanut oral immunotherapy. Journal of Allergy and Clinical Immunology. 2009; 124:292–300. [PubMed: 19577283] This study was the first to couple evaluation of clinical efficacy and immunologic changes in subjects undergoing peanut OIT. Effective OIT was associated with decreases in IgE, increased IgG4, Treg induction, as well as down-regulation genes in apoptosis pathways.
- 60. Clark AT, Islam S, King Y, Deighton J, Anagnostou K, Ewan PW. Successful oral tolerance induction in severe peanut allergy. Allergy. 2009; 64:1218–1220. [PubMed: 19226304] These investigators demonstrated clinical efficacy of peanut OIT in a small cohort of peanut allergic patients.
- 61. Hofmann AM, Scurlock AM, Jones SM, Palmer KP, Lokhnygina Y, Steele PH, Kamilaris J, Burks AW. Safety of a peanut oral immunotherapy protocol in children with peanut allergy. Journal of Allergy and Clinical Immunology. 2009; 124:286–291. [PubMed: 19477496] This study establishes that peanut OIT is a safe and overall well-tolerated therapy when performed by experienced investigators in controlled research settings with appropriate safety precautions in place.
- 62. Varshney P, Steele PH, Vickery BP, Bird JA, Thyagarajan A, Scurlock AM, Perry TT, Jones SM, Burks AW. Adverse reactions during peanut oral immunotherapy home dosing. Journal of Allergy and Clinical Immunology. 2009; 124:1351–1352. [PubMed: 19913285] w Data from this study defines five scenarios associated with increased risk of adverse reaction during home immunotherapy dosing, providing important data to inform the design and safety of future immunotherapy trials.
- 63. Blumchen K, Ulbricht H, Staden U, Dobberstein K, Beschorner J, de Oliveira LCL, Shreffler WG, Sampson HA, Niggemann B, Wahn U, Beyer K. Oral peanut immunotherapy in children with peanut anaphylaxis. Journal of Allergy and Clinical Immunology. 2010; 126(1):83.e1–91.e1. [PubMed: 20542324] These investigators utilized a 7-day rush OIT protocol, followed by gradual dose escalation, resulting in clinical efficacy and immunologic changes consistent with desensitization and tolerance.
- 64. DunnGalvin A, Chang WC, Laubach S, Steele PH, Dubois AEJ, Burks AW, Hourihane JO. Profiling Families Enrolled in Food Allergy Immunotherapy Studies. Pediatrics. 2009; 124:e503– e509. [PubMed: 19706573] This study suggests that in addition to evaluation of safety and efficacy, the design of future immunotherapy trials should also take into consideration the psychological factors that influence parents' decisions to enroll their children in potentially risky trials to avoid inadvertent coercion or selection bias.

### Table 1

Comparison of immunologic changes in food allergy vs. allergen-specific immunotherapy

	Food allergy	Effective immunotherapy
Serum IgE	↑	$\downarrow$
Serum lgG4	_/↓	↑
Th2 cytokine production	↑	$\downarrow$
Mast cell/basophil reactivity	↑	$\downarrow$
Regulatory T-cell activation	$\downarrow$	<b>↑</b>

			Route of		
Therapy	Model	Model Allergen	administration	Clinical response	Immunologic effects
Allergen-specific therapies					
Oral immunotherapy	Human	Milk [32–36,37••,38•], egg [39], peanut [40•, 41•], fish [32], other [32]	Oral	Clinical desensitization, <sup>↑</sup> threshold dose	$U$ = specific IgE, $\downarrow$ PST reactivity, $\downarrow$ basophil activation, $\uparrow$ lgG4, $\uparrow$ FOXP3 <sup>+</sup> T cells, down regulation of apoptosis genes, <i>trials ongoing</i> <sup>a</sup>
Sublingual immunotherapy	Human	Human Hazelnut [42,43], kiwi [44,45], milk, peanut	Sublingual	Clinical desensitization, <sup>↑</sup> threshold dose	↑ lgG4,↑ IL-10,↓ PST reactivity, <i>Trials</i> ongoing <sup>a</sup>
Epicutaneous immunotherapy	Mouse	Peanut [46•], egg [46•], aeroallergens [46•]	Epicutaneous	↓ Airway hyper responsiveness	↑ lgG2a, ↓ lgE/lgG2a ratio
	Human	Milk [47••]	Epicutaneous	Trend toward $\hat{\uparrow}$ cumulative tolerated dose	No increase in IgE noted during 3 months of treatment, <i>Trials ongoing</i> <sup>4</sup>
Peptide immunotherapy	Mouse	Peanut [48,49], egg [50•]	Rectal, subcutaneous	Protection from anaphylaxis	↑ IFN $\gamma$ , ↑ IGFβ, ↓ Th2 cytokines
	Human	Peanut	Rectal	Trials ongoing <sup>a</sup>	Trials ongoing <sup>a</sup>
Allergen nonspecific therapies					
Traditional Chinese Medicine (FAHF-2)	Mouse	Peanut [51,52•]	Oral	Protection from anaphylaxis	$\ell$ Peanut-specific IgE, $\uparrow$ IgG2a levels, $\downarrow$ Th2 cytokines, $\uparrow$ IFNy by CD8 <sup>+</sup> T cells
	Human		Oral	Phase I trials	Trials ongoing <sup>a</sup>
Monoclonal anti-lgE	Human	Peanut [53], milk	Subcutaneous	1 Threshold dose	Trials ongoing <sup>a</sup>

**NIH-PA Author Manuscript** 

**NIH-PA** Author Manuscript

**NIH-PA** Author Manuscript

Table 2