# **Antibacterial Activity of Coumarins**

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The antibacterial activity of coumarin *per se* and other 45 coumarin derivatives was tested against strains of *Bacillus cereus* MIP 96016, *Escherichia coli* ATCC 25922, *Pseudomonas aeruginosa* ATCC 27853, and *Staphylococcus aureus* ATCC 25923. The inhibitory effects of coumarins were affected by their substitution patterns. Osthenol (44) showed the most effective antibacterial activity against Gram-positive bacteria with MIC values ranging between 125 and  $62.5\,\mu g/ml$ . These results suggested that the prenyl chain of 44 at position 8 and the presence of OH at position 7 of the benzenic ring are required for the antibacterial activity against these strains.

Key words: Coumarins, Osthenol, Antibacterial Activity, Structure-activity Relationships (SAR)

## Introduction

Traditional antibacterial therapy is going through a crisis due to the rapidly increasing development of resistance to existing agents (Ojala et al., 2000). However, the plant kingdom constitutes a source of new chemicals, which may be important for their potential use in medicine (Alice et al., 1991). Coumarins are plant secondary metabolites compounds whose biological activity varies according to their substitution patterns. Substituted 4-(1-piperazinyl) coumarins exhibit antiplatelet aggregation activity (Di Braccio et al., 2004), 8-substituted 7-geranyloxycoumarin derivatives (specially the 8-methoxy and 8-acetoxy derivative) have anti-inflammatory activity (Curini et al., 2004) and 8-substituted 7-methoxycoumarins show potent anti-tumor promoting effects (Ito et al., 1999).

The present study reports the evaluation of the antibacterial potency of a series of simple, prenylated, furano- and pyranocoumarins with emphasis on their structure-activity relationships (SAR).

#### **Materials and Methods**

Test compounds

During the present work forty-four coumarins provided by Professor Franco Delle Monache (Instituto di Chimica, Univesità Cattolica Del Sacro Cuore, Rome, Italy) and three of a commercial source have been assayed. In addition, two coumarins obtained by simple modification were also tested. The identity of natural and semi-synthetic compounds was proved by comparison of their spectroscopic data (<sup>1</sup>H and <sup>13</sup>C NMR) with literature references (Table I).

# Antimicrobial assay

The antibacterial activity of coumarins was investigated by employing a microdilution method. The assay was carried out with four bacterial species, including the Gram-negative bacteria Escherichia coli ATCC 25922 (American Type Culture Collection) and Pseudomonas aeruginosa ATCC 27853 and the Gram-positive bacteria Bacillus cereus MIP 96016 (Departamento de Microbiologia e Parasitologia, UFSC) and Staphylococcus aureus ATCC 25923. Mueller-Hinton agar and broth (Difco Laboratories, Detroit, USA) were used for bacterial growth. The inoculum was an overnight culture of each bacterial species in Mueller-Hinton broth diluted in the same media to a final concentration of approx. 108 CFU/ml. The coumarins were dissolved in dimethyl sulfoxide (DMSO) (10% of the final volume) and diluted with Mueller-Hinton broth (Difco Laboratories) to a concentration of 2 mg/ml. Further 1:2 serial dilutions were performed by addition of Mueller-Hinton

broth to reach a final concentration range 2 to 0.0156 mg/ml.  $100 \,\mu\text{l}$  of each dilution were distributed in 96-well plates, as well as a sterility control (growth control contained Mueller-Hinton broth plus DMSO, without antimicrobial substance). Each test and growth control well was inoculated with 5  $\mu$ l of a bacterial suspension (10<sup>8</sup> CFU/ml or 10<sup>5</sup> CFU/well). All experiments were performed in duplicate and the microdilution trays were incubated at 36 °C for 18 h. Bacterial growth was detected first by optical density determination (ELISA reader, CLX800-BioTek Instruments) and later by addition of 20 µl of an alcoholic solution (0.5 mg/ml) of 2-(4-iodophenyl)-3-(4nitrophenyl)-5-phenyltetrazolium chloride (INT) (Sigma). The trays were again incubated at 36 °C for 30 min, and in those wells, where bacterial growth occurred, INT changed from yellow to purple. Any remaining yellow color indicated absence of growth. Before the addition of INT, a subculture was made from each well without apparent growth to determine MBC. MIC and MBC values were defined as the lowest concentration of each coumarin, which completely inhibited growth or yielded no viable microorganisms, respectively. The results were expressed in micrograms per millilitre. Penicillin and tetracycline were used to assess the MIC values of the reference strains (Smânia et al., 1995).

## **Results and Discussion**

In the present study, a series of 45 coumarin derivatives and the parent coumarin (Table I) were tested for their antibacterial activity against 4 strains of bacteria: two Gram-positive bacteria (Staphylococcus aureus and Bacillus cereus) and two Gram-negative bacteria (Escherichia coli and Pseudomonas aeruginosa). Bacterial susceptibility to coumarins was evaluated by determining the minimal inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) and those that were active exhibited MIC values ranging from 62.5 to 2000 µg/ml (Table II). The results indicated that each compound showed more or less pronounced antibacterial potencies, affecting both Gram-positive and Gram-negative microorganisms. Among the active compounds, osthenol (44) showed the most potent activity with MIC of 62.5  $\mu$ g/ml.

A closer structure-activity relationship (SAR) was obtained from the careful examination of the

series of coumarins tested. Coumarin per se (1) exhibited broad antibacterial activity against all strains tested, but was slightly less active against B. cereus. Kayser and Kolodziej (1999) suggested that fairly high antibacterial activity of coumarin per se is due to both its lipophilic character and planar molecular structure, which contribute in penetration through bacterial cell membrane or cell walls. Within the group of mono-oxygenated coumarins, the addition of a methyl or O-methyl group at position C6 or C7 (compounds 2, 5, 13, **14**) to the aromatic nucleus of the coumarin per se core structure maintained the antibacterial activity against Gram-negative bacteria, but diminished against Gram-positive strains when compared to the parent coumarin. On the other hand, substitution of the less polar functions (OMe, Me) at C6 by an OH function (compound 3) reduced the antibacterial activity against all of the tested microorganisms (Jurd et al., 1971). Kayser and Kolodziej (1999) reported that the addition of an OH group at C7 of the coumarin per se significantly reduced the antibacterial activity against all the tested bacteria. These findings suggested that the antibacterial activity of oxygenated coumarins apparently depended on the position of polar (OH) and less polar (OMe, Me) functions at the aromatic nucleus of the coumarin structure. On the other hand, the addition of varied substitution patterns as 6- and 7-O-acetyl groups (compounds 4 and 15), halogen groups at positions C6 and C7 (compounds 6, 7 and 16), a 6-amino group (compound 8), a 6-carboxyl function (compound 9), a 6-cyano group (compound 10), as well as a 6- and 7-nitro group (compounds 12 and 17) and the addition of a 6-aldehyde group (compound 11) significantly reduced the antibacterial activity against all of the tested microorganisms when compared with the parent coumarin with MIC values ranging from 1000 up to 2000  $\mu$ g/ml.

Within the group of disubstituted coumarins, the addition of two *ortho* OH functions at C6 and C7, esculetin (compound **19**), displayed fairly high antibacterial activity, similar to coumarin *per se* in relation to Gram-positive bacteria (MIC =  $500 \mu g/ml$ ), which is probably due to the facilitated interaction with the peptidoglycan found in the cell wall. Tegos *et al.* (2002) reported that scopoletin displays a better antibacterial activity against Gram-positive bacteria rather than Gram-negative bacteria, perhaps because of Gram-negative bacteria efficient efflux pumps. Scopoletin (OMe at

Table I. Chemical structures of coumarins studied in this work.

	$\mathbb{R}^1$	$\mathbb{R}^2$	$\mathbb{R}^3$	Source
I. Simple coumarins				
Monosubstituted				
Coumarin (1)	Н	Н	Н	Gottlieb et al., 1979
6-Methylcoumarin (2)	$CH_3$	Н	Н	Gottlieb et al., 1979
6-Hydroxycoumarin (3)	OH	Н	Н	Gottlieb et al., 1979
6-O-Acetylcoumarin (4)	$O-C_2H_3O$	Н	Н	Gottlieb et al., 1979
6-Methoxycoumarin (5)	$O-CH_3$	Н	Н	Gottlieb et al., 1979
6-Chlorocoumarin (6)	Cl	Н	Н	Gottlieb et al., 1979
6-Iodocoumarin (7)	I	Н	Н	Gottlieb et al., 1979
6-Aminocoumarin (8)	$NH_2$	Н	Н	Gottlieb et al., 1979
6-Carboxycoumarin (9)	COOH	Н	Н	Gottlieb et al., 1979
6-Cyanocoumarin (10)	CN	Н	Н	Gottlieb et al., 1979
6-Aldehydocoumarin (11)	CHO	Н	Н	Gottlieb et al., 1979
6-Nitrocoumarin (12)	$NO_2$	Н	Н	Gottlieb et al., 1979
7-Methoxycoumarin (13) (Herniarin)	Н	$O-CH_3$	Н	Sarsynthese*
7-Methylcoumarin (14)	H	CH <sub>3</sub>	Н	Gottlieb et al., 1979
7-O-Acetylcoumarin (15)	H	$O-C_2H_3O$	Н	Gottlieb et al., 1979
7-Chlorocoumarin (16)	H	Cl	Н	Gottlieb et al., 1979
7-Nitrocoumarin (17)	Н	$NO_2$	Н	Gottlieb et al., 1979
Bisubstituted				
Scopoletin (18)	O-CH <sub>3</sub>	OH	Н	Torres et al., 1979
Esculetin (19)	OH	OH	H	Fluka**
Di-O-Methyl esculetin (20)	O-CH <sub>3</sub>	O-CH <sub>3</sub>	H	Esculetin methylation
Di-O-Methyl daphnetin (21)	Н	$O-CH_3$	O-CH <sub>3</sub>	Daphnetin methylation
Trisubstituted				
Fraxetin (22)	$O-CH_3$	ОН	ОН	Sarsynthese

<sup>\*</sup> Sarsynthese Co., Genay, France.

C6 and OH at C7, 18) also showed antibacterial activity with MIC =  $1000 \mu g/ml$  for all tested bacteria. On the other hand, the addition of two OMe groups either at C6 and C7 (compound 20) or either at C7 and C8 (compound 21) decreases the antibacterial activity if compared to coumarin per se. Kayser and Kolodziej (1999) demonstrated that, in general, the introduction of an additional radical to monosubstituted coumarins does not necessarily result in a dramatic enhancement in potency. Based on these results, the addition of one or two OMe groups at the aromatic nucleus of disubstituted coumarins at positions C6, C7 or/ and C8 decreased the antibacterial activity if compared to coumarin per se. These results suggest that an OMe group at C6 reduced the antibacterial

activity of coumarin against the tested bacteria as well as two OMe groups at C7/C8.

Fraxetin (6-methoxy-7,8-dihydroxycoumarin, **22**) showed weak antibacterial activity to all tested bacteria, possibly because it lacks an additional OMe group. Kayser and Kolodziej (1999) studied trisubstituted and tetrasubstituted derivatives and the most active analogues were those with two OMe functions at positions C5/C6, C6/C7 and C5/C7 (MIC values ranging from 200 to 500  $\mu$ g/ml for standard bacteria).

Within the group of furanocoumarins, psoralene (29) presented antibacterial activity (MIC =  $1000 \,\mu\text{g/ml}$ ) against *E. coli* but was ineffective against *S. aureus*. Nonetheless, (–)-heraclenol (23) displayed similar antibacterial activity against

<sup>\*\*</sup> Fluka, Buchs, Switzerland.

Table I. (cont.)

II. Furano- and dihydrofuranocoumarins	R	$\mathbb{R}^1$	$\mathbb{R}^2$	Source
OH OH				
OH (-)-Heraclenol (23)				Trani et al., 1997
$\mathbb{R}^{1}$		O-CH <sub>3</sub>	Н	Compagnone et al., 1993
Bergaptene (24) Xanthotoxin (25) Isopimpinellin (26)		H O-CH <sub>3</sub>	O-CH <sub>3</sub> O-CH <sub>3</sub>	Compagnone et al., 1993 Trani et al., 2004
Imperatorin (27)				Trani <i>et al.</i> , 1997
Clausindine (28) Psoralene (29) Dimethyl allyl psoralene (30)	H X			Delle Monache <i>et al.</i> , 1977 Erazo <i>et al.</i> , 1990 Delle Monache <i>et al.</i> , 1976
Marmesin (31) Chalepin (32)	Н			Delle Monache <i>et al.</i> , 1989 Delle Monache <i>et al.</i> , 1977
Isoangenomalin (33)				Delle Monache et al., 1977
Oroselone (34)				Murray, 1978

Table I. (cont.)

II. Furano- and dihydrofuranocoumarins	R	R <sup>1</sup>	R <sup>2</sup>	Source
Angelicin (35)				Erazo et al., 1990
OH Columbianetin (36)				Cuca-Suarez et al., 1998
III. Pyranocoumarins				
Hortiline (37)				Delle Monache et al., 1976
CH <sub>3</sub> O O O				Cuca-Suarez et al., 2002
Anoxanuloxyletiii (36)				
Xanthyletin (39) Dimethyl allyl xanthyletin (40)	Н			Delle Monache <i>et al.</i> , 1976 Delle Monache <i>et al.</i> , 1976
Alloxanthoxyletin (38)	Н			Delle Monache <i>et al.</i> ,

Gram-positive bacteria, but decreased activity against Gram-negative bacteria when compared to parent coumarin *per se* possibly due to oxidation of the isoprenylic chain at C8 of the furanocoumarin. *O*-Methylation at C5 (compound **24**) or C8 (xanthotoxin, **25**) also displayed the same pattern of activity for all tested strains. However, two OMe groups at C5/C8 (isopimpinellin, **26**) decreased the activity of furanocoumarins against Gram-negative and Gram-positive bacteria when

compared to the parent coumarin. A prenyloxy chain at C8 (imperatorin, 27) diminished the antibacterial potency against all tested bacteria, as well as substitutions at C3 as prenyl chains (compounds 28 and 30), substitutions at C7 as prenyl or hydroxyprenyl chain (compounds 31 and 33, respectively). It is worth noting, that chalepin (32) did not show improved antibacterial activity with the addition of an extra prenyl chain at C3 against all tested strains.

Table I. (cont.)

IV. Prenylated coumarins	R	$\mathbb{R}^1$	$\mathbb{R}^2$	Source
ROOOO				
Auraptene (41)				Delle Monache et al., 1995
ROOOO				
Phebalosin (42)	CH <sub>3</sub>			Cuca-Suarez and Delle Monache, 1991
RO O O O Balsamiferone (43)	Н	_/< _	$\prec$	Cuca-Suarez, personal communication
но				
Osthenol (44)				Cuca-Suarez et al., 1998
CH <sub>3</sub> O O O O O O O O O O O O O O O O O O O				Torres <i>et al.</i> , 1979
7- <i>O</i> -Geranyl esculetin ( <b>45</b> )				
Prenyletin (46)				Murray, 1978

Pyranocoumarins (compounds 37–40) were found to be one of the two groups of coumarin analogues that proved to be significantly less effective than all the other series of coumarins derivatives against all tested microorganisms, suggesting that the pyrano ring is not required for enhancing of antibacterial activity of the coumarin per se.

The group of prenylated coumarins also showed weak antibacterial potency compared to coumarin *per se.* Apart from compounds **41**, **42**, **43**, **45** and

**46**, which proved to be inactive against all strains, osthenol (**44**) showed the most prominent activity against Gram-positive bacteria (MIC =  $62.5 \mu g/$  ml). Osthenol is a compound with prenylation at C8 and an OH group at C7, suggesting that those groups are required for good antibacterial activity mostly against Gram-positive bacteria. The results above suggest that an OH group at C7 is an ineffective function for antimicrobial activity, while the substitution pattern at C8 is noteworthy. However, two prenyl chains at C3/C6 reduced antibac-

Table II. Antibacterial activity of coumarins against four bacterial strains.

Coumarins	Е. с	E. coli P. aeruginosa		S. aureus		B. cereus		
	MIC <sup>a</sup>	MBC	MIC	MBC	MIC	MBC	MIC	MBC
Monosubstituted coumarin.	S							
Coumarin (1)	500 (3.42)	1000	500 (3.42)	1000	500 (3.42)	2000	1000 (6.84)	2000
6-Methylcoumarin (2)	500 (3.12)	2000	500 (3.12)	> 2000	1000 (6.24)	> 2000	1000 (6.24)	> 2000
6-Methoxycoumarin (5)	500 (2.84)	1000	500 (2.84)	> 2000	> 2000 (>11.35)	> 2000	2000 (11.35)	> 2000
6-Aminocoumarin (8)	1000 (6.21)	2000	2000 (12.41)	> 2000	> 2000 (> 12.41)	> 2000	2000 (12.41)	> 2000
7-Methoxycoumarin (13)	500 (2.84)	1000	500 (2.84)	> 2000	1000 (5.68)	> 2000	1000 (5.68)	> 2000
(Herniarin)								
7-Methylcoumarin (14)	500 (3.12)	2000	500 (3.12)	> 2000	2000 (12.49)	> 2000	1000 (6.24)	> 2000
7-O-Acetylcoumarin (15)	1000 (4.90)	1000	1000 (4.90)	> 2000	1000 (4.90)	> 2000	1000 (4.90)	> 2000
Disubstituted coumarins	4000 (# #0)	4000	1000 (5.50)	• • • • •	1000 (7.50)	• • • • •	4000 (# 40)	• • • •
Scopoletin (18)	1000 (5.20)	1000	1000 (5.20)	> 2000	1000 (5.20)	> 2000	1000 (5.20)	2000
Esculetin (19)	1000 (5.61)	1000	1000 (5.61)	2000	500 (2.81)	1000	500 (2.81)	2000
F 1 1:1 1 £	_							
Furano- and dihydrofurano coumarins	)-							
(-)-Heraclenol (23)	NT <sup>b</sup> (NT <sup>b</sup> )	NT	2000 (6.57)	> 2000	NT (NT)	NT	500 (1.64)	2000
Xanthotoxin (25)	2000 (9.25)	2000	2000 (0.37)	> 2000	1000 (4.63)	2000	500 (1.04)	1000
Chalepin (32)	2000 (9.23)	> 2000	NT (NT)	> 2000 NT	1000 (4.03)	> 2000	NT (NT)	NT
Oroselone (34)	2000 (8.84)	> 2000	2000 (8.84)	> 2000	1000 (3.18)	2000	500 (2.21)	1000
Angelicin (35)	250 (1.34)	1000	NT (NT)	NT	2000 (10.74)	> 2000	NT (NT)	NT
ringenem (35)	230 (1.54)	1000	111 (111)	111	2000 (10.74)	> 2000	111 (111)	111
Prenylated coumarins								
Osthenol (44)	2000 (8.69)	> 2000	2000 (8.69)	> 2000	62.5 (0.27)	125	62.5 (0.27)	125
Ostriciioi (44)	2000 (8.09)	> 2000	2000 (8.09)	> 2000	02.3 (0.27)	123	02.3 (0.27)	123

<sup>&</sup>lt;sup>a</sup> MIC and MBC expressed in  $\mu$ g/ml (MIC also expressed in mm).

For each observation: deviation from the mean  $d = \pm 250 \,\mu\text{g/ml}$  for compound 1 (*E. coli*), 13 (*P. aeruginosa*), 14 (*E. coli* and *S. aureus*), 32 (*E. coli*) and 35 (*E.coli*); for the remaining compounds  $d = \pm 0 \,\mu\text{g/ml}$ .

terial activity as well as the presence of OMe at C7 and an  $\alpha,\beta$ -epoxidation of C-8 prenyl group (compound **42**). These findings suggest that a prenyl group or prenyl chain or even a prenyloxy chain at C7 may contribute for the reduction of antibacterial activity of coumarin *per se* (com-

pounds **46**, **45** and **41**, respectively) as well as the addition of OH or OMe groups at C6.

Approx. 2000 MIC values have been obtained with all compounds (of Table I) except for those which are shown by the MIC values in Table II.

<sup>&</sup>lt;sup>b</sup> NT, not tested.

- Alice C. B., Vargas V. M. F., Silva G. A. A.B., Siqueira N. C. S., Shapoval E. E. S., Gleye J., Henriques J. A. P., and Henriques A. T. (1991), Screening of plants used in south Brazilian folk medicine. J. Ethnopharm. 35, 165–171.
- Compagnone R., Rodrigues M. C., and Delle Monache F. (1993), Coumarins from *Pilocarpus racemosus*. Fitoterapia **64**, 557.
- Cuca-Suarez L. E. and Delle Monache F. (1991), Constituents of *Murraya exótica* adapted in Colombia. Rev. Latinoam. Quim. **22**, 38–40.
- Cuca-Suarez L. E., Martinez J. C., and Delle Monache F. (1998), Constituentes químicos de *Zanthoxylum monophyllum*. Ve. Col. Quim. **27**, 17–27.
- Cuca-Suarez L. E., Menichini F., and Delle Monache F. (2002), Tetranortriterpenoids and dihydrocinnamic acid derivatives from *Hortia colombiana*. J. Braz. Chem. Soc. **13**, 339–344.
- Curini M., Epifano F., Maltese F., Marcotullio M. C., Tubaro A., Altinier G., Gonzales S. P., and Rodriguez J. (2004), Synthesis and anti-inflammatory activity of natural and semisynthetic geranyloxycoumarins. Bioorg. Med. Chem. Lett. 14, 2241–2243.
- Delle Monache F., Marletti F., Marin Bertolo G. B., De Mello J. F., and De Lima O. G. (1976), Coumarins of *Hortia Arbórea*: hortiline and hortiolone. Gazz. Chim. Ital. **106**, 681–689.
- Delle Monache F., Valera G. C., Marini Bertolo G. B., De Mello J. F., and De Lima O. G. (1977), Coumarins of *Hortia arbórea* II hortiolone and hortinone. Gazz. Chim. Ital. **107**, 399–402.
- Delle Monache F., Delle Monache G., De Moraes Souza M. A., Da Salete Cavalcanti M., and Chiappeta A. (1989), Isopentenylindole derivatives and other components of *Esembeckia leiocarpa*. Gazz. Chim. Ital. **119**, 435–439.
- Delle Monache F., Trani M., Yunes R. A., and Falkenberg D. (1995), (-)-Lunacrinol from *Esembeckia hieronim*. Fitoterapia **66**, 474.
- Di Braccio M., Grossi G., Roma G., Signorello M. G., and Leoncini G. (2004), Synthesis and *in vitro* inhibitory activity on human platelet aggregation of novel properly substituted 4-(1-piperazinyl) coumarins. Eur. J. Med. Chem. **39**, 397–409.
- Erazo S., Garcia R., and Delle Monache F. (1990), Bakuchiol and other compounds from *Psoralea glandulosa*. Rev. Latinoam. Quim. **21**, 62.

- Gottlieb H. E., Alves De Lima R., and Delle Monache F. (1979), <sup>13</sup>C NMR of 6- and 7- substituted coumarins. Correlations with Hammett constants. J. Chem. Soc. Perkin Trans. II, 435–437.
- Ito C., Itoigawa M., Furukuda H., Tokuda H., Okuda Y., Mukainaka T., Okuda M., and Nishino H. (1999), Anti-tumor-promoting on Epstein-Barr virus activation assay. Cancer Lett. **138**, 87–92.
- Jurd L., Corse J., King A. D., Bayne Jr. H., and Mihara K. (1971), Antimicrobial properties of 6,7-dihydroxy-, 7,8-dihydroxy-, 6-hydroxy- and 8-hydroxycoumarins. Phytochemistry **10**, 2971–2974.
- Kayser O. and Kolodziej H. (1999), Antibacterial activity of simple coumarins: structural requirements for biological activity. Z. Naturforsch. **54c**, 169–174.
- Murray R. D. H. (1978), Naturally occurring coumarins. Fortschr. Chem. Org. Naturst. **35**, 200–209.
- Ojala T., Remmes S., Haansuu P., Vuorela H., Hiltunen R., Haahtela K., and Vuorla P. (2000), Antimicrobial activity of some coumarin containing herbal plants growing in Finland. J. Ethnopharm. **73**, 299–305.
- Smânia A. Jr., Delle Monache F., Smânia E. F., Gil M. L., Benchetrit L. C., and Cruz F. S. (1995), Antibacterial activity of a substance produced by the fungus *Pycnoporus sanguineus*. J. Ethnopharm. 45, 177– 181.
- Tegos G., Stermitz F. R., Lomovskayd O., and Lewis K. (2002), Multidrug pump inhibitors uncover remarkable activity of plant antimicrobials. Antimicrob. Agents Chemother. **46**, 3133–3141.
- Torres R., Delle Monache F., Marini Bertolo G. B., and Cassels B. K. (1979), Coumarins and cinnamic acid from *Gymnophyton isatidicardum*. J. Nat. Prod. **42**, 532–533.
- Trani M., Delle Monache F., Delle Monache G., Yunes R. A., and Falkenberg D. (1997), Dihydrochalcones and coumarins of *Esembeckia grandiflora* subsp. *grandiflora*. Gazz. Chim. Ital. **127**, 415–418.
- Trani M., Carbonetti A., Delle Monache G., and Delle Monache F. (2004), Dihydrochalcones and coumarins of *Esembeckia grandiflora* subsp. *brevipetiolata*. Fitoterapia **75**, 99–102.