


Original article

Antibiotic resistance pattern of bacteria isolated from patients with upper respiratory tract infections; a four-year study in Tripoli city

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ABSTRACT

Introduction: Respiratory tract infections have been known to be a significant health concern for mortality and morbidity since many years. This study was aimed at determining the prevalence of bacterial pathogen causing upper respiratory tract (URTIs) and the susceptibility patterns to frequently used antibiotics among patients attending Abusetta hospital in Tripoli district.

Methods: A total of 1,110 throat swabs were collected between Jan, 2011 to December, 2014 and inoculated onto Blood agar, MacConkey agar and Chocolate agar then incubated at 37 °C for 24 hours. Bacterial pathogens were determined by bacteriological culture methods and antibiotic susceptibility of the isolates was identified following Clinical Laboratory Standard Institute guidelines (CLSI).

Results: Of the 1,110 respiratory samples tested, 71.1% (n = 789) of specimens were positive cultures with the dominant bacterial pathogens being *Streptococcus pneumoniae* 43.3% (n = 342), followed by *Pseudomonas aeruginosa* 22.8% (n = 180), *Staphylococcus aureus* 13.8% (n = 109), *Escherichia coli* 6.9% (n = 55), *Enterobacter spp* 6.2% (n = 49), *Citrobacter* 4.5% (n = 36), and *Klebsiella* 2.2% (n = 18). Most isolates exhibited resistance against the commonly used antibiotics and to at least one antibiotic.

Conclusion: The level of antibiotic resistance in this study is alarming and brings to light the timely and suitable diagnosis of the common bacteria causes of URTIs and proper antibiotic administration based on susceptibility test.

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1. INTRODUCTION

Respiratory tract infections are among the most common and diverse group of infections that have continually been a major cause of morbidity and morbidity in clinical medicine [1]. Respiratory tract infections are known as any infectious disease of the upper or lower respiratory tract.

Upper respiratory tract infections (URTIs) involve the common cold, tonsillitis, laryngitis, pharyngitis, rhinitis, and otitis media. Lower respiratory tract infections (LRTIs) include acute bronchitis, and pneumonia [2]. Bacteria such as *Pseudomonas aeruginosa*, *Streptococcus pneumoniae*, *Moraxella catarrhalis*, and *Hemophilus influenzae* are among the causative agents of respiratory tract infections [3]. URTIs also cause of most antibiotic use.

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Approximately 60% of all outpatient antibiotics use is for respiratory infections, particularly for acute bacterial sinusitis for adults and acute bacterial otitis for children [4].

Antimicrobial resistance is a rising public health distress and strictly related to use of antibiotics [5]. In Egypt, 63.6 % of prescriptions for antibiotics are dispensed in primary healthcare, and the major cause were for respiratory tract infections [6]. Most URTIs are viral, self-limiting infections that can be self-treated without the use for antibiotics [7]. Excessive usage of antibiotics has caused resistance of bacteria toward antibiotics, representing a significant public health problem, mainly in light of the declining supply of newer antibiotic drugs [8]. Reducing request for antibiotics for self-limiting settings is considered as effective approach of averting additional antibiotic resistance [9]. To address the problem of antimicrobial resistance in URTIs, the current study aimed to investigate common bacterial profile in upper respiratory tract infection and their antibiotics sensitivity pattern.

2. METHODS

2.1. EXPERIMENTAL DESIGN AND SUBJECTS

This is a retrospective investigation of upper respiratory tract infections in hospital- care settings in Tripoli city. A total of 1,110 patients who had URTIs, as manifested by clinical signs and symptoms and/or chest X-ray report suggestive of URTIs admitted to Abusetta Hospital in Tripoli city, were enrolled in this study over a period of 4 year from Jan 2011 to December 2014.

Prior to enrolment, a written informed consent was obtained from each patient. In addition, patients' demographic data such as age and gender were also documented at the time of specimen gathering. Specimens were collected directly into a sterile wide mouthed container and transported to the laboratory according to standard protocol [10]. All the subjects were assured regarding privacy of disease and data. This study was ethically approved by the ethics review committee of faculty of Medical technology, University of Tripoli, Libya.

2.2. ISOLATION OF BACTERIAL STRAINS

1,110 throat swab specimens from patients of different age groups were taken in specialized containers for the study. The specimen was inoculated under a septic technique on Petri plates enclosing blood agar, MacConkey agar and chocolate agar media, and incubated at 37°C for 24 h. All the bacteria were isolated and identified using morphological, microscopy and biochemical tests. Only positive culture sensitivity reports were selected.

2.3. ANTIBIOTIC SUSCEPTIBILITY TEST

Antibiotic sensitivity was performed by Kirby- Bauer disc diffusion method as previously described [11]. Paper disks were impregnated with antibiotics including Amikacin (AK) (30µg), Amoxicillin (AX) (10µg), Ampicillin (AM) (10µg), Cefotaxime (CF) (30µg), Ceftriaxone (CT) (30µg), Cefuroxime (CE) (30µg), Ciprofloxacin (CP) (5µg), Erythromycin (ER) (15µg), Gentamycin (GM) (10µg), etc., and incubated at 37°C for 24 hours. The diameter of the zones of inhibition was measured and analysis of result based on CLSI guideline was performed [12].

3. RESULTS

After assessing the data collected from the hospital and the tests done in the laboratory, a scenario of antibiotic resistance pattern of bacteria was evaluated. Out of the 1,110 clinical samples studied, 632 (57.45%) were from male and 468 (42.55%) were from female. Of them, 71.1% (n = 789) of the samples examined were positive for infectious bacteria (54.62% male and 45.37% female) (Table 1).

Table 1. Gender wise distribution of patients with URTIs

Gender	No. of samples (%)	No. of positive cases (%)
Male	632 (57.45%)	431 (54.62%)
Female	468 (42.55%)	358 (45.37%)
TOTAL	1,110 (100%)	789 (100%)

URTIs: Upper respiratory tract infections.

3.1. BACTERIAL ISOLATES

The dominated species of pathogenic bacteria presented with URTIs patients attended Abusetta hospital in Tripoli has been presented in (Figure 1).

From the various inoculated samples, 71.1% (n = 789) of the samples examined were positive for infectious bacteria. The dominated species of pathogenic bacteria were identified as streptococcus pneumonia, followed by *Pseudomonas aeruginos*, *Staphylococcus aureus*, *Escherichia Coli*, *Enterobacter spp*, and *Citrobacter*. Among the identified bacterial species, the highest number 43.3% (n = 342) of isolates of *streptococcus pneumonia* were obtained, whereas lowest number were 2.2 % (n = 18) obtained of *Klebsiella*, while *Pseudomona aeruginosa*, *staphylococcus aurous*, *Escherichia Coli*, *Enterobacter spp*, *Citrobacter*, contributed 13.8% (n = 109), 6.9% (n = 55), 6.2% (n = 49), 4.5% (n = 36), of total isolates, respectively.

3.2. ANTIBIOTIC SUSCEPTIBILITY TEST

Kirby- Bauer disc diffusion method was used to test the susceptibility of isolated pathogenic bacteria against different antibiotics such as amoxicillin, amoxycylav, amikacin, ciprofloxacin, clarithromycin, cefuroxime,

doxycycline etc, ranging from (05 – 30 µg), and principles for suggesting whether the isolate is susceptible or resistant toward specific antibiotic have been given in Table 2. It was observed that the *streptococcus pneumoniae* was highly resistant to gentamycin (51 % resistant strains) and ciprofloxacin (41 % resistant strains), whereas it was least resistant to cefotixin (0.3 % resistant strains) and doxycycline (0.5 % resistant strains).

Pseudomona aeruginosa was resistant to β-lactam antibiotics such as amoxyclav, amoxicillin, ceftriaxone as well as macrolide antibiotics such as clarithromycin with percent resistance of 71, 44, 29 and 37 %, respectively (Table 2). *P. aeruginosa* was also moderately resistant to fluoroquinolone antibiotics such as ciprofloxacin, and aminoglycosides antibiotics such as gentamicin and amikacin with percent resistance of 24, 24 and 23 %, respectively.

Our result also revealed that, *Staphylococcus aureus* was observed to be 35 % resistant against amoxicillin, 29 %

Enterobacter spp, and *Citrobacter* represent clinically important pathogens. These pathogens have also been associated with URTIs in other studies [13- 15]. Isolation of *Streptococcus spp* was within the range reported in most studies. A similar study reported a prevalence of 12.3% of this pathogen in Libya, although the pathogen was found to be in higher percentage in our study [16]. It was also reported with a prevalence of 22 % in Iceland, 14% in Brazil, Cameroon and 8% in Netherlands [17-20].

Earlier studies have highlighted the significance of establishing the suitable treatment for this infection to decrease probabilities of complications such as rheumatic fever, scarlet fever and endocarditis [21- 23]. The high prevalence of *Streptococcus spp* has also been reported of 49% in Australia, 10.8% in China, and 3.5% in Italy [24-26]. Most of the bacteria isolated during the study period including *S. pneumonia*, *P. aeruginosa*, *S. aureus*, and *Enterobacter spp* fall in either nosocomial or community-acquired classes of infection, telling that proper preventive

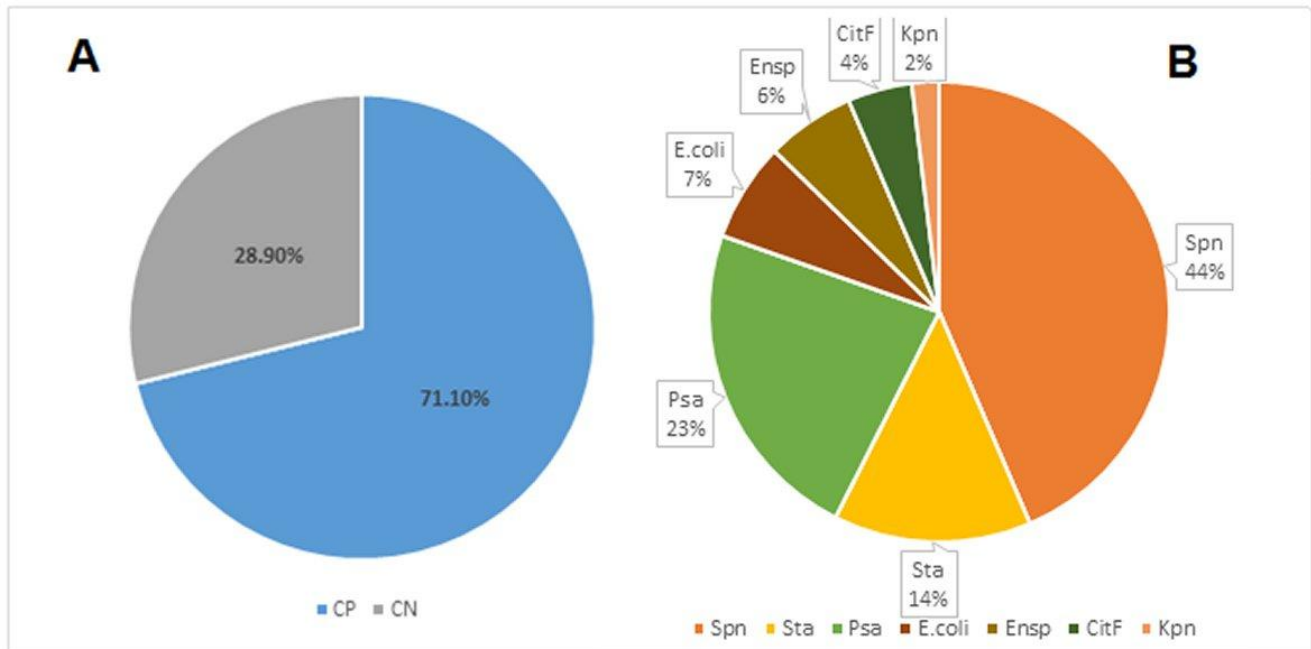


Figure 1: Detection and identification of bacterial pathogens isolated from 1,110 patients with URTIs. Fig 1A shows the rates of culture positive and culture negative specimens. Fig 1B shows the proportion of different bacterial strains among culture positive specimens.

CP: Culture Positive; CN: Culture Negative; Spn: *Streptococcus pneumoniae*; Sta: *Staphylococcus aureus*; Psa: *Pseudomona aeruginosa*; E. coli: *Escherichia coli*; Ensp: *Enterobacter sp*; CitF: *Citrobacter freundil*; Kpn: *Klebsiella pneumoniae*.

against clarithromycin and 25 % against ciprofloxacin (Table 2). *E. coli* was observed to be highly resistant against ceftriaxone and ciprofloxacin compared to other antibiotics; it was having 40 and 38 % resistance against ceftriaxone and ciprofloxacin, respectively (Table 2). All other isolates were resistant to at least one antibiotic.

4. DISCUSSION

Isolates of *S. pneumoniae*, *P. aeruginosa*, *S. aureus*,

actions could limit bacterial involvement in URTIs. Since increasing antibiotic resistance to frequently prescribed antibiotics makes bacterial infections an interdisciplinary universal threat to public health worldwide, the study also aimed to investigate antibiotic resistance patterns for the isolated bacteria. As we have mentioned in result section, our *in vitro* antibiotic resistance study was concerning that most of the isolates except *S. pneumoniae*, *S. aureus*, and *Citrobacter freundil* showed alarming levels of resistance against the commonly used first line antibiotics. Even two of 7 multidrug resistant

isolates (*P. aeruginosa* and *E. coli*) exhibited resistance not only against the commonly used antibiotics but also against other antibiotics.

measures, the emerging resistance toward antibiotics is a growing concern among clinicians and other health professionals worldwide. The uncritical and irrational use

Table 2. Resistance pattern of the isolated bacteria to tested antibiotics

Antibiotics	Bacterial isolates						
	<i>S. pneumoniae</i> (n=342)	<i>S. aureus</i> (n=109)	<i>P. aeruginosa</i> (n=180)	<i>E. coli</i> (n=55)	<i>Enterobacter</i> <i>sp.</i> N=49)	<i>Citrobacter</i> <i>freundlii</i> (n=36)	<i>Klebsiella sp.</i> (n=18)
Amikacin	113 (33%)	11 (10.1%)	41 (23%)	9 (16%)	17 (35%)	4 (11.1%)	2 (11%)
Amoxiclav	78 (22.8%)	20 (18.3%)	128 (71.1%)	10 (18.2%)	16 (32.7%)	6 (16.7%)	8 (44.4%)
Amoxicillin	57 (16.6%)	38 (34.9%)	80 (44.4%)	10 (18.2%)	25 (51.0%)	5 (13.9%)	8 (44.4%)
Cefotaxim	1 (0.3%)	2 (1.8%)	9 (5.0%)	0	1 (2%)	0	0
Ceftriaxone	30 (8.7%)	12 (11%)	53 (29%)	22 (40%)	4 (8%)	3 (8.3%)	0
Sulfamethoxazole	3 (0.8%)	5 (4.6%)	5 (2.8%)	0	1 (2%)	0	0
Ciprofloxacin	140 (41%)	27 (24.8%)	43 (23.9%)	21 (38.2%)	9 (18.4%)	3 (8.3%)	3 (16.6%)
Clarithromycin	66 (19.2%)	32 (29.4%)	67 (37.2%)	14 (25.5%)	21 (42.9%)	5 (13.9%)	4 (22%)
Doxycycline	2 (0.5%)	0	2 (1.1%)	2 (3.6%)	0	0	0
Gentamycin	175 (51.1%)	15 (13.8%)	44 (24.4%)	8 (14.5%)	7 (14.3%)	5 (13.9%)	0
Nalidixic acid	0	1 (0.9%)	1 (0.6%)	0	0	0	0
Nitrofurantoin	0	1 (0.9%)	0	0	0	0	2 (11%)
Ceftazidim	38 (11.1%)	19	37 (20.6%)	18 (32.7%)	6 (12.2%)	7 (19.4%)	0
Moxifloxacin	5 (1.4%)	0	19 (10.6%)	0	1 (2%)	0	2 (11%)
Tobramycin	36 (10.5%)	5 (4.6%)	8 (4.4%)	3 (5.5%)	1 (2%)	4 (11.1%)	0
Polymyxin B	11 (3.2%)	1 (0.9%)	6 (3.3%)	1 (1.8%)	1 (2%)	0	0
Levofloxacin	30 (8.7%)	0	14 (7.8%)	0	1 (2%)	0	0

Antibiotic sensitivity studies exhibited *Streptococcus spp* to be highly resistant to gentamycin and ciprofloxacin; however, it was least resistant to cefotaxim and doxycycline. Similar studies have also reported resistance of *Streptococcus spp* to gentamycin and ciprofloxacin [27,28]. In previous studies, cefotaxim was considered as the best antibiotic against *Streptococcus pneumoniae* [29]. However, a disappointing antibiotic sensitivity pattern of *streptococcus spp* was observed for commonly prescribed antimicrobial including amoxiclav, amoxicillin, and clarithromycin with resistance rate of 22.8%, 16.6%, and 19.2%, respectively. The resistance pattern of *Streptococcus spp* has been reported in other studies [30]. Furthermore, the rate bacterial resistance to penicillin has also been reported in previous studies [31,32]. Inappropriate widespread use of these antibiotics has been suggested as an explanation to this resistance.

The main cause favoring the appearance of antibiotic resistance is their widespread use. *Pseudomonas aeruginosa* exhibited low rate of susceptibility to amoxiclav (71.4%) amoxicillin (44.4%), and ciprofloxacin (24%) which are frequently dispensed over-the-counter in pharmacies, contrasts with the noticeable levels of sensitivities of the isolates to doxycycline and cefotaxim which are less commonly used, thus suggesting an association between antibiotic use and the level of drug resistance reported in this study as previously suggested in another study [33]. Despite the improvements in healing and defensive

of antibiotics have donated to the emergence of resistance, which may turn out to be a principal cause of morbidity and mortality in the developing countries.

A precious antibiotic treatment is in demand to overcome this critical concern, but the shortage of evidence regarding the principle causes of antibiotic sensitivity patterns in countries like Libya have made it difficult. The information about antibiotic resistance profile reported here in this study is expected to promote awareness among physicians, community pharmacist, as well as among policy makers of community and private sectors in the country.

5. CONCLUSION

In summary, URTIs incidence, as well as pathogen exposure rates, were high in Abusetta hospital. *S. pneumoniae* and *P. aeruginosa* were the predominant bacterial causative agents, with amoxiclav and amoxicillin being resistance to most identified pathogenic bacteria in our population sample. There is calls for a crucial need to impose evidence-based guidelines and the delivery of educational program to promote better antibiotic dispensing practices that can be worth in improving current resistant pattern in Libya. Although the number of isolates used may be small to draw significant assumptions on the susceptibility patterns, they, however, deliver baseline data for future investigations, particularly considering the thing

that no such data have been reported in this vicinity notwithstanding the very high rate of antibiotic misuse. These findings are, therefore, of clinical and epidemiological importance.

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7. REFERENCES

1. Malosh RE, Martin ET, Ortiz JR, Monto AS. The risk of lower respiratory tract infection following influenza virus infection: A systematic and narrative review. *Vaccine*. 2018;36(1):141-7. doi: 10.1016/j.vaccine.2017.11.018.
2. Wang L, Qiao X, Ai L, Zhai J, Wang X. Isolation of antimicrobial resistant bacteria in upper respiratory tract infections of patients. *3 Biotech*. 2016;6(2):166. doi: 10.1007/s13205-016-0473-z.
3. Assane D, Makhtar C, Abdoulaye D, Amary F, Djibril B, Amadou D, et al. Viral and Bacterial Etiologies of Acute Respiratory Infections among Children Under 5 Years in Senegal. *Microbiol Insights*. 2018;11:1-5. doi: 10.1177/1178636118758651.
4. Jong EC, Stevens DL. *Netter's Infectious Diseases*. Saunders: 2011.
5. Raft CF, Bjerrum L, Arpi M, Jarløv JO, Jensen JN. Delayed antibiotic prescription for upper respiratory tract infections in children under primary care: Physicians' views. *Eur J Gen Pract*. 2017;23(1):190-5. doi: 10.1080/13814788.2017.1347628.
6. Sabry NA, Farid SF, Dawoud DM. Antibiotic dispensing in Egyptian community pharmacies: an observational study. *Res Social Adm Pharm*. 2014;10(1):168-84. doi: 10.1016/j.sapharm.2013.03.004.
7. Peters S, Rowbotham S, Chisholm A. Managing self-limiting respiratory tract infections: a qualitative study of the usefulness of the delayed prescribing strategy. *Br J Gen Pract*. 2011;61(590):e579-e89. doi: 10.3399/bjgp11X593866.
8. Arason VA, Kristinsson KG, Sigurdsson JA. Do antimicrobials increase the carriage rate of penicillin resistant pneumococci in children? Cross sectional prevalence study. *BMJ*. 1996;313(7054):387-91. doi: 10.1136/bmj.313.7054.387.
9. Atia AE, Abired AN. Antibiotic prescribing for upper respiratory tract infections by Libyan community pharmacists and medical practitioners: An observational study. *Libyan J Med Sci*. 2017;1:31-5. doi: 10.4103/LJMS.LJMS_14_17.
10. Collee JG, Fraser AG, Marmion BP, Simmons A. Mackie & McCartney *Practical Medical Microbiology*, 14th ed. Churchill Livingstone: 2007.
11. Bauer AW, Kirby WN, Sheris JC, Tuck M. Antibiotic susceptibility testing by standardised single disc method. *Am J Clin Pathol*. 1966;36:493-6.
12. National Committee for Clinical Laboratory Standards, *Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically*, 3rd Edition, approved standard (NCCLS, Pennsylvania, Document M7-A3, 1997).
13. Khan S, Priti S, Ankit S. Bacteria Etiological Agents Causing Lower Respiratory Tract Infections and Their Resistance Patterns. *Iran Biomed J*. 2015;19(4):240-6. doi: 10.7508/ibj.2015.04.008.
14. Josphat M, John M, Anthony K. Antimicrobial susceptibility patterns of bacteria associated with upper respiratory tract infections in Kitui, Kenya. *Ethiop Med J*. 2017;55(2):121-7.
15. Dutta A, Dutta SE, Mazumdar S, Chatterjee M, Sarkar A. Bacteriological Profile of Recurrent Upper Respiratory Tract Infection in Children Attending a Tertiary Care Hospital. *Int J Curr Microbiol App Sci*. 2017;6(8):2561-7.
16. Eldeeb A, Khashan E. Microbiological Study on Respiratory Tract Infections in Libya. *Egypt J Hosp Med*. 2006;24:442-59.
17. Magnúsdóttir BT, Jónsson JS, Kristinsson KG. Prevalence of *Streptococcus pyogenes* and methicillin-resistant *Staphylococcus aureus* in the pharynx of healthy children in the town of Gardabaer. *Laeknabladid*. 2008;94:447-51.
18. Mouro A, Luci B, Tabacow H, Marines D, Valle M, Jacyr P. Prevalence of upper respiratory tract infections at a tertiary care hospital in the city of São Paulo. *Einstein (Sao Paulo)*. 2010;8:197-9. doi: 10.1590/S1679-45082010AO1348.
19. Akoachere JF, Ndip RN, Chenwi EB, Ndip LM, Njock TE, Anong DN. Antibacterial effect of *Zinziberofficinale* and *Garcinia kola* on respiratory tract patho-gens. *East Afr Med J*. 2002;79(11):588-91. doi: 10.4314/eamj.v79i11.8804.
20. Van Gageldonk-Lafeber AB, Marianne AB, Marie-Louise AH, Marcel FP, Aad IM, Berry W. Risk factors for acute respiratory tract infections in general practitioner patients in The Netherlands: a case-control study. *BMC Infect Dis*. 2007;7:35. doi: 10.1186/1471-2334-7-35.

21. Bisno AL. Acute pharyngitis. *N Engl J Med.* 2003;344(3):205-11. doi: 10.1056/NEJM200101183440308.
22. Choby BA. Diagnosis and treatment of streptococcal pharyngitis. *Am Fam Physician.* 2009;79(5):383-90.
23. McGregor KF, Spratt BG, Kalia A, Bennett A, Bilek N, Beall B. Multilocus sequence typing of *Streptococcus pyogenes* representing most known emm types and distinctions among subpopulation genetic structures. *J Bacteriol.* 2004;186(13):4285-94. doi: 10.1128/JB.186.13.4285-4294.2004.
24. Watson K, Carville K, Bowman J, Jacoby P, Riley TV, Leach AJ, et al. Upper respiratory tract bacterial carriage in Aboriginal and non-Aboriginal children in a semi-arid area of Western Australia. *Pediatr Infect Dis J.* 2006;25(9):782-90. doi: 10.1097/01.inf.0000232705.49634.68.
25. Sung RY, Ling JM, Fung SM, Oppenheimer SJ, Crook DW, Lau JT, et al. Carriage of *Haemophilus influenzae* and *Streptococcus pneumoniae* in healthy Chinese and Vietnamese children in Hong Kong. *Acta Paediatr.* 1995;84(11):1262-7. doi: 10.1111/j.1651-2227.1995.tb13545.x.
26. Principi N, Marchisio P, Schito GC, Mannelli S. Risk factors for carriage of respiratory pathogens in the nasopharynx of healthy children. *Pediatr Infect Dis J.* 1999;18(6):517-23. doi: 10.1097/00006454-199906000-00008.
27. Eliopoulos GM, Wennersten C, Zigelboim-Daum S, Reiszner E, Goldmann D, Moellering RC. High-level resistance to gentamicin in clinical isolates of *Streptococcus (Enterococcus) faecium*. *Antimicrob Agents Chemother.* 1988;32(10):1528-32. doi: 10.1128/aac.32.10.1528.
28. Canton R, Morosini M, Enright MC., Morrissey I. Worldwide incidence, molecular epidemiology and mutations implicated in fluoroquinolone-resistant *Streptococcus pneumoniae*: data from the global PROTEKT surveillance programme. *J. Antimicrob. Chemother.* 2003;52(6):944-52. doi: 10.1093/jac/dkg465.
29. ZhiHua F. Efficacy observation of cefotaxime/sulbactam in treatment of severe respiratory infection. *Evaluation and Analysis of Drug-Use in Hospitals of China.* 2015;15(12):1575-7.
30. El-Mahmood AM, Isa H, Mohammed A, Tirmidhi AB. Antimicrobial susceptibility of some respiratory tract pathogens to commonly used antibiotics at the Specialist Hospital, Yola, Adamawa State, Nigeria. *J Clin Med Res.* 2010; 2:135-42.
31. Passali D, Lauriello M, Passali GC, Passali FM, Bellusi L. Group A *Streptococcus* and its antibiotic resistance. *Acta Otorhinolaryngol Ital.* 2007;27:27-32.
32. Nevio CS. Pyogenesis well and alive. *BC Medical Journal.* 2009;51:122-7.
33. Anab F, Syed B, Sheikh A, Shaheen P, Sabahat J. Antimicrobial susceptibility pattern of clinical isolates of *Pseudomonas aeruginosa* isolated from patients of lower respiratory tract infections. *SpringerPlus.* 2012;1(1):70. doi: 10.1186/2193-1801-1-70.