

Anticancer activities of constituents of kava (*Piper methysticum*)

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ABSTRACT

Crude extracts of kava (*Piper methysticum* G. Forster, Piperaceae) showed good activity against ovarian tumour and leukaemia cancer cell lines. Bioassay-guided isolation resulted in the isolation of six known kava lactones and two flavokavains. The structure of the compounds were elucidated by spectroscopic techniques and by comparison with data in the literature.

Keywords: Kava, *Piper methysticum*, Piperaceae, Lactones, Toxicity, Bioassay, Ovarian tumour, Leukaemia, NMR.

1 INTRODUCTION

Kava (*Piper methysticum* G. Forster, Piperaceae) has been cultivated in the Oceania islands of the South Pacific for over 3,000 years (Hocart *et al.* 1993), and has a long history of being used as a remedy for the treatment of gonorrhoea, rheumatism, bronchitis, asthma as well as stomach aches and headaches (Weiner and White 1976). The kava lactones are believed to be responsible for biological activity which include local anaesthetic properties (Meyer and May 1964), sedative (Klohs *et al.* 1959; Meyer 1962), analgesic (Bruggemann and Meyer 1963), anticonvulsive (Meyer 1964; Meyer and Meyerburg 1964; Meyer 1965), antispasmodic (Meyer 1965), antimycotic (Hansel *et al.* 1966), anxiolytic (Pittler and Ernst 2000; Scherer 1998; Volz and Kiesser 1997), and central muscular relaxing effects (Baum *et al.* 1998). This has led to its popular use in Europe and North America for the treatment of anxiety disorders (Singh and Blumenthal 1997). Some nineteen lactones have been isolated from kava with 6 major and 13 minor constituents (Shao *et al.* 1998) Also known to be present in trace amounts are the alkaloids: pipermethysticine (Smith 1979), *N*-cinnamoylpyrrolidine and *N*-methoxycinnamoylpyrrolidine (Achenbach and Wittman 1970), 3 α ,4 α -epoxy-5 β -pipermethystin and awaine (Dragull *et al.* 2003). Also known to be present are three flavokavains, flavokavain A-C (Achenbach and Wittman 1970; Dutta *et al.* 1973). The study (Steiner 2000) linking the low occurrences of some forms of cancer with kava drinking prompted us into this investigation. A screen of a crude extract of kava had shown good activity against ovarian and leukaemia cell lines. A bioassay-guided isolation on this extract led to the isolation of six known kavalactones and two flavokavains. The paper describes the isolation, identification and the anticancer activities of the compounds.

2 MATERIAL AND METHODS

2.1 PLANT MATERIAL

A kava plant was collected from the village of *Navakasali*, on the island of *Vanua Levu*, Fiji Islands. The plant was identified at the South Pacific Regional Herbarium at the University of the South Pacific. The samples were cleaned thoroughly in fresh water, cut into smaller pieces and sun dried. The dried samples were pounded to powdered form and stored in a plastic bag, sealed and sent by mail to the University of Aberdeen,

Scotland where it was stored in a freezer at -20 °C until extracted.

2.2 GENERAL EXPERIMENTAL PROCEDURES

Low resolution electrospray ionization mass spectra (LREIMS) were obtained on a Finnigan Masslab Navigator and high resolution mass data (HREIMS) were obtained on a Finnigan MAT-95. ¹H, ¹³C and all NMR 2D experiments were recorded on a Varian Unity INOVA 400 MHz spectrometer, in CDCl₃ solution. Chemical shifts are reported in parts per million (δ) downfield relative to residual CHCl₃ at 7.27 ppm. HPLC separations were carried out using a Spectra Physics P100 isocratic pump and a Waters reversed phase (ODS, 250 x 10 mm) column and monitored using a Hewlett Packard HP 1050 Series Variable Wavelength UV Detector. UV and IR were taken on a Perkin Elmer Lambda 15 UV/VIS spectrophotometer and Ati Mattson Genesis Series FTIR machine respectively.

2.3 CHEMICALS

All extraction and partitioning solvents such as methanol, dichloromethane, hexane, ethyl acetate and *s*-butanol were laboratory grade (Sigma Aldrich). All HPLC solvents were HPLC grade (Rathburn Chemicals Ltd). Size exclusion column chromatography was performed on a Sephadex LH-20 (Sigma Aldrich) column. Flash column chromatography was performed on a Flash 40i system (Flash 40S, 32-63 μ m, 60Å cartridge). Normal and reversed phase TLC were carried out on silica gel and C-18 pre-coated plates (Merck) respectively.

2.4 EXTRACTION AND ISOLATION

Traditional preparation of kava involves extraction with water. The closest solvent to this is MeOH, which should extract most of the polar compounds. The use of dichloromethane was to improve the extraction efficiency of all polar as well as semi-polar compounds. The extraction was performed as follows: The powdered kava (500g dry weight) was extracted with MeOH (3x) and CH₂Cl₂ (3x), the solvent was removed under reduced pressure, and the extracts were combined. The crude oil was partitioned between water and CH₂Cl₂. The aqueous layer was then extracted with *s*-BuOH to give a yellow coloured oil (WB). The solvent was removed from the CH₂Cl₂ layer and the resulting oil was partitioned between

n-hexane and 10% aqueous MeOH. The *n*-hexane fraction was dried to give a brownish-yellow coloured oil (FH).

The MeOH layer was then phase adjusted to 50% aqueous MeOH and extracted with CH₂Cl₂ to give a dark brownish oil (FD). The 50% aqueous MeOH fraction was dried to give a light brownish coloured oil (FM). Interest was focused on the dichloromethane (FD) and hexane (FH) fractions, which were found to be highly active

against ovarian tumour and leukaemia cancer cells. In addition, these fractions possessed interesting high and low field ¹H and ¹³C NMR resonances. The FH fraction was subjected to normal phase flash chromatography using a mixture of hexane and ethyl acetate (80/20) as solvent, yielding 27 fractions. The column was eluted further by ethyl acetate (F-EtOAc) and finally methanol (F-MeOH).

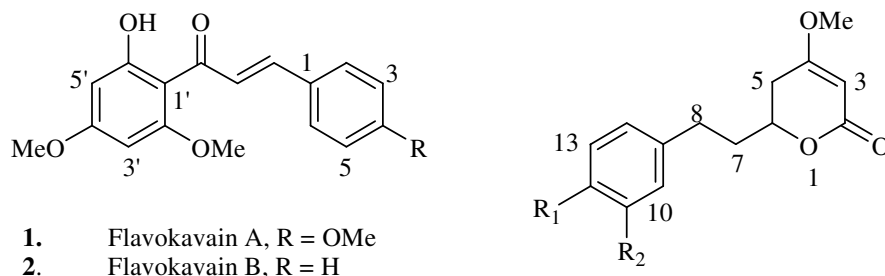


Figure 1. Structure of kava flavokavains and lactones.

No.	Compound	R ₁	R ₂	C5-C6	C7-C8
3.	dihydromethysticin	O-CH ₂ -O	—	—	—
4.	7,8 dihydrokavain	H	H	—	—
5.	kavain	H	H	—	==
6.	demethoxyyangonin	H	H	==	==
7.	<i>cis</i> -yangonin	OMe	H	==	==
8.	<i>trans</i> -yangonin	OMe	H	==	==

After monitoring by TLC, similar fractions were pooled together and then purified by HPLC. The fraction (F12-18) was purified by normal phase HPLC using a mixture of hexane and ethyl acetate (80/20) to afford 14.2 mg of flavokavain A (**1**) while the fraction F6-11 yielded 8.2 mg of flavokavain B (**2**). The fraction (F-EtOAc) was purified by reversed phase C-18 HPLC using a mixture of

water, methanol and acetic acid (70/30/0.1) as solvent to yield 6.3 mg of dihydromethysticin (**3**) and 8.4 mg of 7,8-dihydrokavain (**4**). The FD fraction was purified as above to afford 21.6 mg of kavain (**5**), 11.6 mg of demethoxyyangonin (**6**), 9.3 mg of compound of *cis*-yangonin (**7**) and 8.4 mg of *trans*-yangonin (**8**).

Table 1. Anticancer activities of kava crude fractions and isolated compounds.

Sample	K562 (μg/mL) leukaemia	A2780 ovarian tumour
WB	>100	48.16
FD	0.85	0.43
FH	0.70	0.66
flavokavain A (1)	2.04	1.32
flavokavain B (2)	0.95	0.56
dihydromethysticin (3)	6.43	5.15
7,8-dihydrokavain (4)	9.15	4.87
kavain (5)	5.35	2.54
demethoxy-yagonin (6)	2.88	3.79
<i>cis</i> -yagonin (7)	0.42	0.75
<i>trans</i> -yagonin (8)	1.41	2.39

2.5 MTT ASSAY

The anti-tumour assay was performed at the Paterson Institute for Cancer Research, at the Christie Hospital in Manchester, UK. Tests were performed using leukaemia and ovarian tumour cell lines. The K562 human leukaemia and the A2780 ovarian cell lines were cultured as

described in the literature (McGowan and Fox 1988). Cytotoxicity tests were carried out using the MTT assay described in the literature (Mossmann 1983). Cells were treated with the drug in 96 well plates in antibiotic free RPMI medium containing 10% foetal calf serum. Drugs were dissolved in dimethylsulphoxide (DMSO) and were

added by serial dilution. Drug treatment lasted 5 days, the duration of the assay. The IC₅₀ value was calculated by reference to a standard curve constructed for control cells.

3 RESULTS AND DISCUSSION

Structures of all the compounds were confirmed by interpretations of 1D, 2D NMR and HRESIMS data and by comparison with spectroscopic data in the literature (Dharmaratne *et al.* 2002). The anticancer activity of the eight compounds is shown in Table 1 with IC₅₀ values ranging between 0.42-9.15 µg/mL. It is reasonable to suggest that the anticancer activity observed in the FD and FH crude fractions is due to the presence of *cis*-yagonin and flavokavain B. The relatively high activity of one of the isomeric forms of yagonin is interesting from a structure-activity perspective. *Cis*-yagonin was an order of magnitude more potent than its geometric isomer, as well as the compound demethoxyyagonin which was found by a previous study (Sotheeswaran *et al.* 2002) to inhibit the release of the tumour necrosis factor α (TNF α). The mode of activity of flavokavain A has been studied and is known to induce apoptosis in bladder cancer cells by involvement of Bax protein-dependent and mitochondria-dependent apoptotic pathway (Zi and Simonaeu 2005).

4 CONCLUSION

This study has substantiated the findings of previous studies about the anticancer activities of kava. The most active compounds against leukaemia and ovarian tumour are *cis*-yagonin and flavokavain B.

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REFERENCES

Achenbach, H. and Wittman, G. 1970. Tetrahedron Letters **37**, 3259-3262.
 Baum, S. S., Hill, R. and Rommelspacher, H. 1998. Effect of kava extract and individual kavapyrones on neurotransmitter levels in the nucleus accumbens of rats. *Progress in Neuro-Psychopharmacology and Biological Psychiatry* **22**, 1105-1120.
 Bruggemann, F. and Meyer, H. J. 1963. Die analgetische Wirkung der Kava-Inhaltstoffe Dihydrokavain und Dihydromethysticin. *Arzneimittelforschung*, **13**, 407-409.
 Dharmaratne, E. R. W., Nanayakkara, N. P. D. and Khan, I. A. 2002. Kavalactones from *Piper methysticum*, and their ¹³C NMR spectroscopic analyses. *Phytochemistry* **59**, 429-433.
 Dragull, K., Yoshida, W. Y. and Tang, C.S. 2003. *Phytochemistry* **63**, 193-198.

Dutta, C. P., Lala, P., Ray, K. and Chatterjee, A. 1973. Constitution of flavokavain C. *Indian Journal of Chemistry* **11**, 509-510.
 Hansel, R., Weiss, D. and Schmidt, B. 1966. Zwei Chalkonepigmente aus *Piper methysticum* Forst. Zu Fraze der biosynthesis der Kawalaktone. *Planta Medica* **14**, 1-9.
 Hocart, C. H., Fankhauser, B. and Buckle, D. W. 1993. Chemical archeology of kava, a potent brew. *Rapid Communication in Mass Spectrometry* **7**, 219-224.
 Klohs, M. W., Keller, F., Williams, R. E., Toekes, M. I. and Cronheim, G. E. 1959. A chemical and pharmacological investigation of *Piper methysticum*, Forster. *Journal of Medicinal and Pharmacological Chemistry* **1**, 95-103.
 McGowan, A. T. and Fox, B. W. 1988. Structure and biochemical comparison of the anti-mitotic agents colchicine, combrestatin A-4 and amphethinile. *Anti-cancer Drug Design* **3**, 249.
 Meyer, H. J. 1962. Pharmakologie der wirksamen Prinzipien des Kawa-Rhizoms (*PiperMethysticum* Forster). *Archives Internationales de Pharmacodynamie et de Therapie* **138**, 505-536.
 Meyer, H. J. 1964. Untersuchungen uber den antikonvulsiven Wirkungstyp der Kawa Pyrone Dihydromethysticin und Dihydrokawain mit Hihe chemische induzierter Krampfe. *Archives Internationales de Pharmacodynamie et de Therapie* **150**, 118-131.
 Meyer, H. J. 1965. Spasmolytische Effeckte von Dihydromethysticin, einem Wirkstoff aus *Piper methysticum* Forst. *Archives Internationales de Pharmacodynamie et de Therapie* **154**, 448-467.
 Meyer, H. J. and May, H. U. 1964. Lokalanaesthetische Eigenschaften natuerlicher Kawapyrone. *Klinische Wochenschrift* **42**, 407.
 Meyer, H. J. and Meyer-Burg, J. 1964. Hemmung des Elektrokramfes durch die Kawa-Pyrene Dihydromethysticin and Dihydrokawain. *Archives Internationales de Pharmacodynamie et de Therapie* **148**, 97-110.
 Mosmann, T. 1983. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *Journal of Immunological Method* **65**, 55-63.
 Pittler, M. H. and Ernst, E. 2000. Efficacy of kava extract for treating anxiety: sytematic review and metal analysis. *Journal of Clinical Psychopharmacology* **20**, 84-89.
 Scherer, I. 1998. Kava-kava extract in anxiety disorders: An outpatient observational studies. *Advances in Therapy* **15**, 261-269.
 Shao, Y., he, K., Zheng, B. and Zheng, Q. 1998. Reversed-phase high-performance liquid chromatographic method for quantitave analysis of the six major kavalactones in *Piper methysticum*. *Journal of Chromatography A* **825**, 1-8.
 Singh, Y. N. and Blumenthal, M. 1997. Kava: an overview. Distribution mythology, culture, chemistry and pharmacology of the South Pacific's most favourite herb. *Herbalgram* **39**, 33-56.
 Smith, R. M. 1979. Pipermethystine, a novel pyridine alkaloid from *Piper methysticum*. *Tetrahedron* **35**, 437-438.

- Sotheeswaran, S., Fujiki, H. and Gunatilaka, A. A. L. 2002. Anticancer activity studies on kava (*Piper methysticum*). In: Proceedings of the Pacific kava research symposium. Forum Secretariat, Suva, Fiji.
- Steiner, G. G. 2000. The correlation between cancer incidence and kava consumption. *Hawaii Medical Journal* **59**, 420-422.
- Volz, H. P. and Kiesser, M. 1997. Kava extract WS 1490 versus placebo in anxiety disorders- a randomized placebo-controlled 25 week outpatient trial. *Pharmacopoeia* **30**, 1-5.
- Weiner, M. A. and White, A. 1976. *Secrets of Fijian Medicine*, University of California, Berkley, California.
- Zi, X. and Simoneau, A. R. 2005. Flavokawain A, a novel chalcone from kava extract, induces apoptosis in bladder cancer cells by involvement of Bax protein-dependent and mitochondria-dependent apoptotic pathway and suppresses tumour growth in mice. *Cancer Research* **65**, 3479-3486.