

Research Article

Antifatigue Activity of Liquid Cultured *Tricholoma matsutake* Mycelium Partially via Regulation of Antioxidant Pathway in Mouse

Quan Li,¹ Yanzhen Wang,^{1,2} Guangsheng Cai,¹ Fange Kong,¹ Xiaohan Wang,¹
Yang Liu,¹ Chuanbin Yang,³ Di Wang,¹ and Lirong Teng^{1,2}

¹School of Life Sciences, Jilin University, Changchun 130012, China

²Zhuhai College of Jilin University, Jilin University, Guangzhou 519041, China

³School of Chinese Medicine, Hong Kong Baptist University, Kowloon, Hong Kong

Correspondence should be addressed to Di Wang; jlwangdi@gmail.com and Lirong Teng; tenglirong@jlu.edu.cn

Received 16 October 2015; Accepted 3 November 2015

Academic Editor: Oreste Gualillo

Copyright © 2015 Quan Li et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Tricholoma matsutake has been popular as food and biopharmaceutical materials in Asian countries for its various pharmacological activities. The present study aims to analyze the antifatigue effects on enhancing exercise performance of *Tricholoma matsutake* fruit body (ABM) and liquid cultured mycelia (TM) in mouse model. Two-week *Tricholoma matsutake* treatment significantly enhances the exercise performance in weight-loaded swimming, rotating rod, and forced running test. In TM- and ABM-treated mice, some factors were observed at 60 min after swimming compared with nontreated mice, such as the increased levels of adenosine triphosphate (ATP), antioxidative enzymes, and glycogen and the reduced levels of malondialdehyde and reactive oxygen species in muscle, liver, and/or serum. Further data obtained from western blot show that CM and ABM have strongly enhanced the activation of 5'-AMP-activated protein kinase (AMPK), and the expressions of peroxisome proliferator-activated receptor γ coactivator-1 α (PGC-1 α) and phosphofructokinase-1 (PFK-1) in liver. Our data suggest that both *Tricholoma matsutake* fruit body and liquid cultured mycelia possess antifatigue effects related to AMPK-linked antioxidative pathway. The information uncovered in our study may serve as a valuable resource for further identification and provide experimental evidence for clinical trials of *Tricholoma matsutake* as an effective agent against fatigue related diseases.

1. Introduction

Regular exercise has been confirmed to protect and alleviate various diseases; however, strenuous sports are responsible for the accumulation of reactive oxygen and lipid peroxides, which damage organ tissues and further lead to fatigue [1, 2]. Fatigue, caused by intense pressure from physical and mental work [3], affects more than 20% of people worldwide. Sleep disturbance, muscle pain, headache, and impaired concentration always appear in patients suffering from chronic fatigue. Repairing the damage occurring in organs and prompting elimination of metabolites accumulation during exercise are required for its recovery. In molecular level, the activation of 5'-AMP-activated protein kinase (AMPK) inhibits the overproduction of reactive oxygen species (ROS) [4] and

suppresses anabolic ATP-consuming [5]. On the other hand, enzymatic and nonenzymatic antioxidants protect tissues from exercise-induced oxidative damage [6] and further reduce metabolite production and physical fatigue [7]. Intense exercise causes an imbalance between the body's oxidation and antioxidation systems; thus overproducing reactive oxide species (ROS) and malondialdehyde (MDA) is responsible for fatigue symptoms [8, 9]. The enhanced activities of superoxide dismutase (SOD) and glutathione peroxidase (GSH-Px) are observed combining with the antifatigue process [10].

Fatigue is becoming a serious public health problem, and what is worse current therapeutic regimen is far away from meeting the needs of patients. Since nutrient supplementation positively enhances exercise capacity, researches

attempt to seek antifatigue herbs to accelerate eliminating fatigue in human beings [11]. *Herba Rhodiolae*, commonly used for hypoxia cure by the Tibetan people [12], also enhances physical ability [13]. *Astragalus membranaceus* has been confirmed to improve exercise performance and reduce exercise-caused accumulation of the byproducts in blood [14]. *Cordyceps sinensis* improves animal exercise endurance via regulating glycogen breakdown and fatty acid oxidation which are accommodated by AMPK phosphorylation [15].

Tricholoma matsutake, which is an ectomycorrhizal symbiotic mushroom, has been popular as food and biopharmaceutical materials in Asian countries [16]. The immunomodulatory activities and anticancer effects of *Tricholoma matsutake* have been well established worldwide [17]. Various polysaccharides have been reported, which are separated from *Tricholoma matsutake* fruit body with antioxidant activity [18], antitumor effect [19], and immune regulation property [20]. However, wild fruiting body resources are scarce, and to cultivate the fruit bodies via artificial culture medium is limited. Submerged fermentation has been verified as an efficient way to produce mycelia and bioactive metabolites for fungus in amount of research groups [21, 22]. In our research group, submerged fermentation has been successfully applied to obtain mycelia of *Paecilomyces tenuipes* [23], *Marasmius androsaceus*, and *Irpex lacteus* [24]. One study shows that beta-glucan separated from liquid cultured mycelia of *Tricholoma matsutake* exerts immunostimulating activities via multiple signaling pathways linked to neural factor-kappa B activation [25].

To our knowledge, the antifatigue activities of both *Tricholoma matsutake* fruit body and liquid cultured mycelia have not been reported yet. Our present study aims to investigate the related biological activities of *Tricholoma matsutake* via mouse model. Antioxidant enzyme activities in serum and organs and AMPK activation in liver were assayed in order to further analyze its underlying mechanism.

2. Materials and Methods

2.1. *Tricholoma matsutake* Preparation. *Tricholoma matsutake* (CGMCC5.793) was cultured in a hundred-liter full-automatic fermentor (BaoXing Bioscience company, Shanghai, China) by using a defined liquid medium containing: 20 g/L sucrose, 10 g/L peptone, 10 g/L yeast extract powder, 0.5 g/L $MgSO_4 \cdot 7H_2O$, 0.5 g/L $KH_2PO_4 \cdot 3H_2O$, and 0.2 g/L Vitamin B1. The fermentation conditions were as follows: loading volume was 70 L/100 L, initial pH was 6.5, rotate speed was 150 rpm, ventilation volume was 200 L/h, culture temperature was 26°C, inoculum age was 3 days, and inoculum size was 5%. The collected *Tricholoma matsutake* mycelium (TM) was freeze-dried, and it was given directly to experimental mice. All the chemical reagents used in the submerged fermentation were obtained from Sigma-Aldrich, USA.

2.2. Antifatigue Capacity Analysis in Mouse. The experimental animal protocol used in the study was approved by Institution of Animal Ethics Committee in Jilin University.

Kunming (KM) mice (6 weeks, 18–22 g; $n = 20$ /group; equal numbers of males and females; purchased from Norman Bethune University of Medical Science, Jilin University, Jilin, China) (SCXK(JI)-2011-0003) were put on a 12 h light/dark cycle (lights on 07:00–19:00) at $23 \pm 1^\circ C$ with water and food available ad libitum. After one-week adaptation, the experimental mice were divided into five groups randomly and orally treated with physiological saline (control group), 0.4 g/kg of *Tricholoma matsutake* body fruit micropowder (purchased from Japan) (ABM), and TM powder at doses of 0.3 g/kg, 1.0 g/kg, and 2.0 g/kg once a day for two weeks continuously. After two hours since the last administration, following experiments were performed.

2.2.1. Weight-Loaded Swimming Test. Experimental mice were loaded with lead wire of 15% bodyweight and forced to swim in water of $22 \pm 1^\circ C$ temperature and 30 cm depth. The time from the beginning to the point at which mice failed to return to the water surface within 15 s was recorded as exhaustion time.

2.2.2. Rotating Rod Test. Three independent trainings on turning device at a speed of 15 rpm for 1 min were applied before format detection. During the rotating rod test, the total duration of experimental mice on turning device at 20 rpm was recorded.

2.2.3. Forced Running Test. Three independent trainings on a treadmill at a set speed of 15 rpm for 1 min were performed to all the mice. After practices, animals were put on the treadmill with a set speed of 20 rpm, and the total running time was recorded to evaluate their performance before exhaustion, which was identified as five-time consecutive shocks by the electrode.

2.3. Measurement of Physiological Indices Related to Fatigue. Kunming mice (6 weeks, 18–22 g; $n = 20$ /group; equal numbers of males and females) were orally treated with physiological saline, 0.4 g/kg of *Tricholoma matsutake* body fruit powder, and TM powder at doses of 0.3 g/kg, 1.0 g/kg, and 2.0 g/kg once a day for two weeks continuously. After four hours since the last administration, blood samples were collected from caudal vein in all mice. The serum levels of ATP, MDA, SOD, and GSH-Px were determined according to the procedures provided by assay kits.

After 24-hour rest, the experimental mice were forced to swim in water with $22 \pm 1^\circ C$ temperature and 30 cm depth for 60 min. Blood samples were collected from caudal vein after 10 min recess. Further, liver and muscle of sacrificed mice were dissected, and they were homogenized in double distilled water after wash three times in ice-cold physiological saline solution. The levels of ATP, SOD, GSH-Px, ROS, and MDA in liver, muscle, and/or serum were detected. The concentration of glycogen in both muscle and liver was measured. All the assay kits were purchased from Nanjing Biotechnology Co. Ltd., Nanjing, China.

TABLE 1: The effects of *Tricholoma matsutake* on fatigue and oxidant related factors in serum of mice before swimming.

	Female					Male				
	CTRL	0.3	TM (g/kg) 1	2	ABM (g/kg) 0.4	CTRL	0.3	TM (g/kg) 1	2	ABM (g/kg) 0.4
ATP (nmol/mL)	2.5 ± 0.1	2.6 ± 0.1	3.1 ± 0.1	2.9 ± 0.1	2.7 ± 0.1	2.6 ± 0.1	2.7 ± 0.1	2.9 ± 0.1	2.8 ± 0.1	2.7 ± 0.1
MDA (nmol/mL)	11.4 ± 0.4	11.2 ± 0.4	10.8 ± 0.3	11.3 ± 0.3	11.3 ± 0.3	12.7 ± 0.4	12.1 ± 0.8	11.7 ± 0.6	11.9 ± 0.3	12.3 ± 0.3
SOD (U/mL)	90.4 ± 5.3	93.0 ± 3.1	101.1 ± 3.3	99.6 ± 3.1	99.7 ± 4.8	93.2 ± 4.4	96.2 ± 3.3	93.4 ± 3.1	100.4 ± 2.5	97.8 ± 5.1
GSH-Px (μmol/mL)	156.7 ± 10.6	172.1 ± 8.3	171.0 ± 10.1	176.2 ± 10.0	174.1 ± 9.4	159.0 ± 16.7	167.6 ± 10.2	168.9 ± 12.3	169.7 ± 7.6	171.5 ± 14.4

Values were expressed as mean ± SEM ($n = 10$).

2.4. Western Blot Analysis. A part of liver tissue of each experimental mouse was homogenized in eight volumes of radioimmunoprecipitation assay (RIPA) buffer (Sigma-Aldrich, USA) which contains 1 mM Phenylmethanesulfonyl fluoride (PMSF) and 1x protease inhibitor cocktail (Sigma-Aldrich, USA). The protein concentration was detected by Bradford method, and 30 μg of sample was separated by 10% SDS-PAGE and then transferred onto a nitrocellulose membrane (0.45 μm, Bio Basic, Inc.) with the Mini-Protean two-gel electrophoresis system (Bio-Rad, USA). The blocked membranes with target strips were incubated with primary antibodies at 4°C overnight at dilution of 1:1000 as follows: phosphor-AMPK (P-AMPK), total-AMPK (T-AMPK), peroxisome proliferator activated receptor γ coactivator-1α (PGC-1α), phosphofructokinase-1 (PFK-1), and glyceraldehyde-3-phosphate dehydrogenase (GAPDH) (Cell Signaling Technology, Beverly, MA), followed by blotting with horseradish peroxidase-conjugated secondary antibodies diluted 1:2000 (Santa Cruz, USA). After visualizing with ECL detection system (GE Healthcare, UK), the intensity of target bands were quantified by using ImageJ.

2.5. Statistical Analysis. All data were expressed as mean ± SEM. A one-way analysis of variance (ANOVA) was used to detect statistical significance followed by post hoc multiple comparisons (Dunn's test) by using SPSS 16.0 software (IBM corporation, Armonk, USA). A P value < 0.05 was considered to be statistically significant.

3. Results

3.1. The Enhancing Exercise Endurance Ability of *Tricholoma matsutake*. Behavior tests are commonly applied to evaluate the antifatigue performance of drugs [26, 27]; therefore, weight-loaded swimming, rotating rod, and forced running test were taken in the present experiment. Compared to nontreated mice, both TM and ABM have significantly enhanced the exercise endurance indicated by the longer movement duration in all three behavior tests ($P < 0.05$, Figure 1). 2.0 g/kg of TM administration improved nearly twofold of residence time of both female and male mice in

all tests ($P < 0.01$, Figure 1). It seems that TM possesses more sensitive effects on athletic ability of female mice.

3.2. The Regulation Effects of *Tricholoma matsutake* on ATP and Glycogen. Compared to nontreated mice, no significant changes of serum levels of ATP were observed after 14-day TM and ABM administration (Table 1).

ATP, as a direct energy source for normal physiological activity, shortage leads to a variety of physiological dysfunction [28]. Compared with nontreated mice, and similar to ABM, TM dose-dependently enhanced ATP concentration in serum, muscle, and liver after 60 min of swimming, and up to 30% enhancement was noted in both male and female mice ($P < 0.01$, Figure 2).

Glycogen is the primary intracellular storable form of glucose, and its level in liver and skeletal muscles directly reflects fatigue symptoms [29]. After 60 min exercise, compared with nontreated mice, two-week TM treatment at a dose of 2.0 mg/kg has enhanced 35.6% and 28.5% muscle glycogen levels in male and female mice, respectively ($P < 0.05$, Figure 3(a)). In liver tissue, TM has increased to 48.1% and 52.5% glycogen levels in female and male mice, respectively ($P < 0.05$, Figure 3(b)). The similar improvement activities on glycogen concentration in both muscle and liver were also observed in ABM-treated mice ($P < 0.05$, Figure 3).

3.3. The Antioxidant Effects of *Tricholoma matsutake*. Antioxidant enzymes, such as SOD and GSH-Px, are responsible for cell damage and muscle fatigue. MDA is one of the end products resulting from degradation of cell membrane by radicals [30]. Before swimming, no significant changes on levels of SOD, GSH-Px, and MDA in serum, muscle, and liver were observed in both TM- and ABM-treated mice (Table 1).

Compared to nontreated group, in both female and male mice, TM significantly enhanced the activities of SOD and GSH-Px and reduced the levels of MDA in serum, muscle, and liver after 60 min swimming ($P < 0.05$, Table 2). Similarly, ABM displayed positive regulatory effects on levels of SOD, GSH-Px, and MDA in serum muscle and liver of exercised male or female mice ($P < 0.05$, Table 2). The reduced ROS levels of muscle and liver were observed in

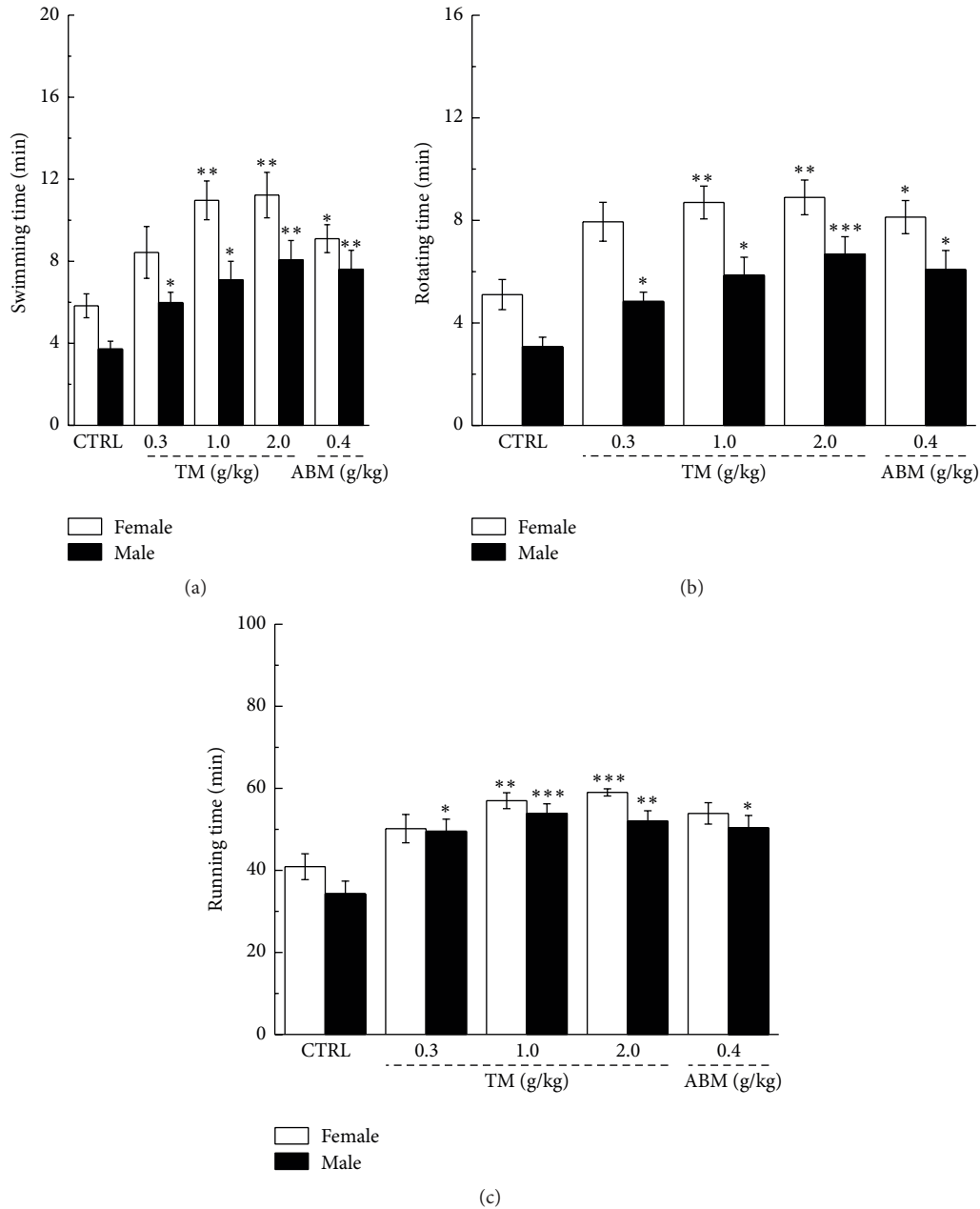


FIGURE 1: Weight-loaded swimming test (a), rotating rod test (b), and forced running test (c) were performed to verify the antifatigue effects of *Tricholoma matsutake* in mouse model. Data were expressed as mean \pm SEM ($n = 10$) and analyzed by using a one-way ANOVA followed by Dunn's test. * $P < 0.05$, ** $P < 0.01$, and *** $P < 0.001$ versus control group.

TM- and ABM-treated mice after 60 min exercise ($P < 0.05$, Table 2).

3.4. The Regulation Effects of *Tricholoma matsutake* on Protein Expression in Liver. The expressions of P-AMPK, T-AMPK, PFK-1, and PGC-1 α were detected to further analyze the preliminary molecular mechanisms under TM-mediated antifatigue effects. After 60 min swimming, over twofold enhancement of P-AMPK expression was observed in liver tissue of TM- or ABM-treated mice ($P < 0.001$, Figure 4).

Furthermore, TM dose-dependently enhanced the expressions of PFK-1 and PGC-1 α in the liver of exercised mice, and the similar results were noted in ABM-treated group ($P < 0.05$, Figure 4).

4. Discussion

Herbs have become a valuable reservoir for antifatigue agents selection [31]. In our group, the polysaccharides-enriched *Cordyceps militaris* extract exhibiting exercise enhancement

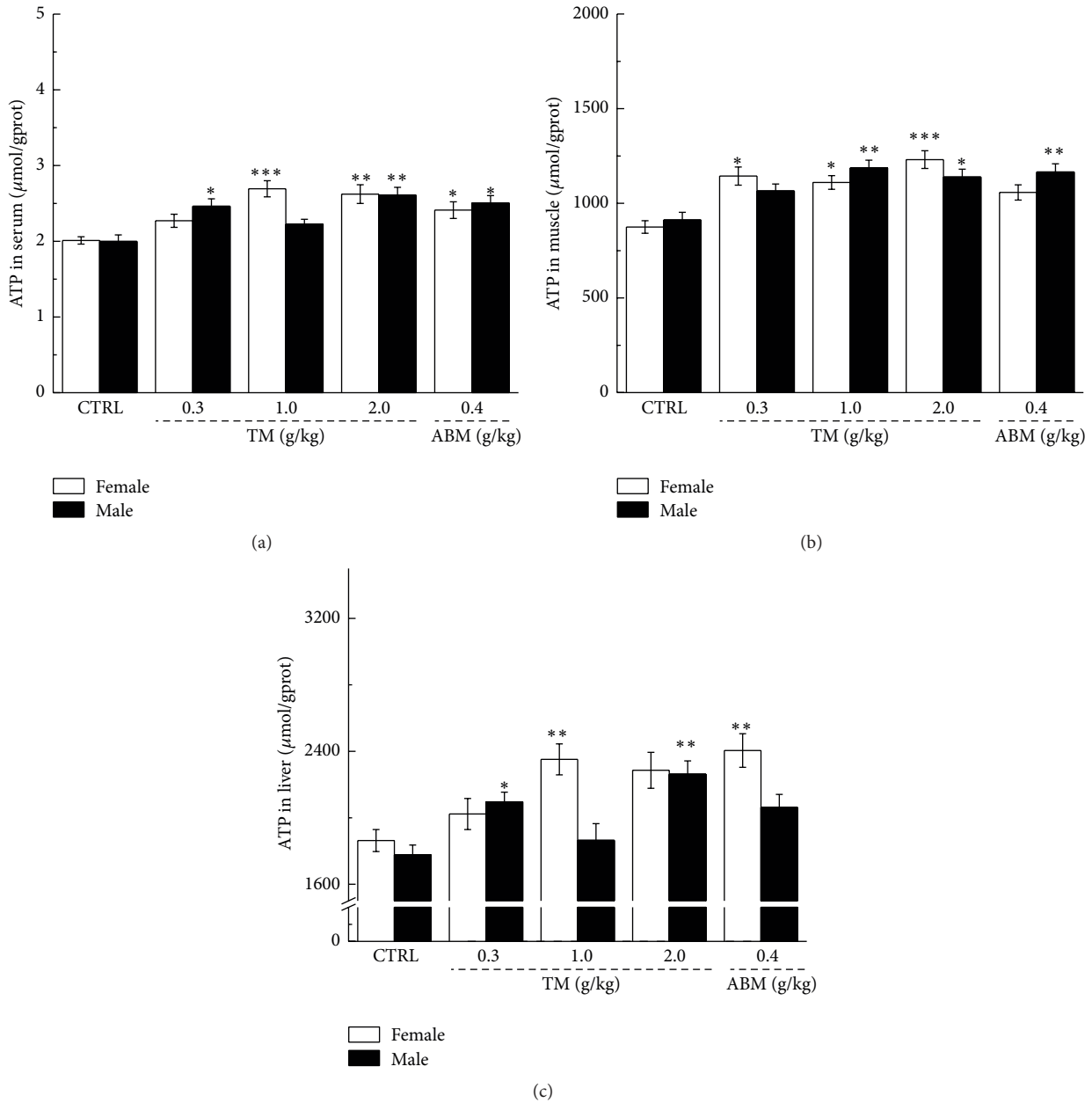


FIGURE 2: At the end of two-week administration, the levels of ATP in serum (a), muscle (b), and liver (c) in male and female mice were analyzed after 60 min of swimming. Data were expressed as mean \pm SEM ($n = 10$) and analyzed by using a one-way ANOVA. * $P < 0.05$, ** $P < 0.01$, and *** $P < 0.001$ versus control group.

activity has been successfully demonstrated via modulation AMPK-linked pathway [32]. The present study aims to investigate the antifatigue effects of *Tricholoma matsutake* liquid cultured mycelia and fruit body in mouse model. The enhanced exercise endurance of TM- and ABM-treated mice in weigh-loaded swimming, forced running, and rotating rod test revealed the antifatigue activities of *Tricholoma matsutake*. The enhanced levels of ATP and glycogen in serum, muscle, and/or liver of treated mice have contributed to TM-mediate fatigue recovery. ATP, known as a rapid energy source, can be influenced by levels of muscle $[H^+]$ and myofibrillar ATPase during exercise [33].

However, the half-life of ATP is <1 second, and it makes glycogen become an indirect energy source for ATP synthesis. Long-term endurance exercise, which is related to muscle mitochondria dysfunction, results in a reduction of muscle glycogen depletion [34]. The regulatory effect of PGC-1 α on mitochondrial biogenesis has been well reported, and exercises have influenced the expression of PGC-1 α [35]. PGC-1 α is involved in exercise-induced downregulation of the expression of glycogenolytic and glycolytic enzymes [34]. Another study reports that fatigue and exhaustion are viewed as a multicomponent biochemical process related to PFK-1 linked glycolytic pathway [36]. Although the enhanced

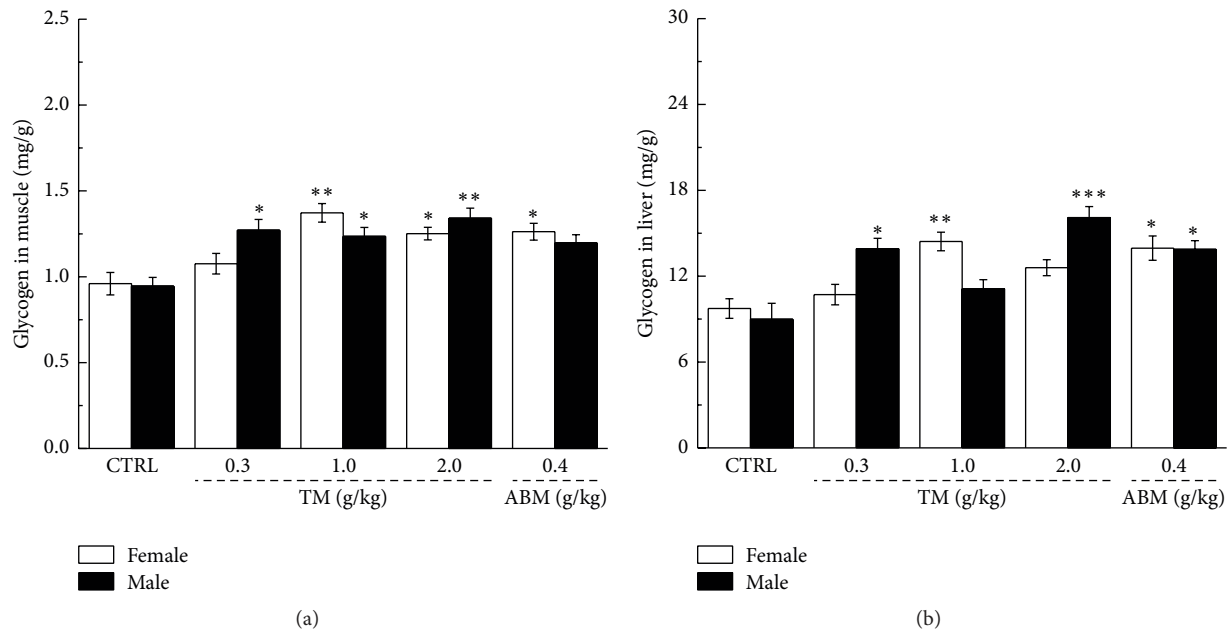


FIGURE 3: Two-week *Tricholoma matsutake* and ABM administration strikingly enhanced the levels of glycogen in both muscle (a) and liver (b) of exercise fatigued mouse. Data were expressed as mean \pm SEM ($n = 10$) and analyzed by using a one-way ANOVA followed by Dunn's test. * $P < 0.05$, ** $P < 0.01$, and *** $P < 0.001$ versus control group.

expression of both PGC-1 α and PFK-1 were observed in the liver of TM- and ABM-treated mice after 60 min exercise, it is hard to conclude that they are involved in the antifatigue effects. Based on our data, the enhanced ATP and glycogen levels in serum and organs by TM and ABM contribute to improve exercise endurance.

The regulatory effects on ROS, MDA, SOD, and GSH-Px in organs and/or serum of TM and ABM in mice after 60 min exercise indicated the important role of oxidative system in preventing exercise-induced fatigue. The accumulation of oxygen free radicals is one of the risk factors which are responsible for oxidative stress and muscle fatigue [37], and they can be cleared by antioxidant enzymes. Antioxidants can successfully prevent or reduce oxidative stress and further improve exercise performance [38]. Proteins, separated from *Panax quinquefolium*, improve behavioural alterations linked to chronic fatigue via inhibition oxidative damage [10]. Meanwhile, *Cordyceps militaris* displays antifatigue property via scavenging ROS and enhancing activities of SOD and GSH-Px [32]. MDA is recognized as the end product of lipid peroxidation, which can be used to assess the degree of the lipid peroxidation for free radical [39]. SOD prevents lipid peroxidation via catalyzing the conversion of superoxide into hydrogen peroxide and oxygen [37]. GSH forms a formidable defense with glutathione peroxidase to prevent lipid peroxides [40]. All these work together to prevent lipid peroxidation and further protect cells from oxidative injury via suppressing hyperlevel of MDA and ROS and enhancing SOD and GSH-Px activities, which are involved in TM- and ABM-mediated antifatigue effect on enhancing exercise endurance.

AMPK serves as the key factor on regulation of glucose and lipid metabolism [41] and helps to maintain the levels of ATP in various conditions [41]. The activated AMPK via falling cellular energy status enhanced ATP generation, whilst inhibiting ATP consumption [42]. AICAR, an AMPK agonist, significantly enhanced running endurance in animals [43]. On the other hand, except for the fatigue situation, during oxidative stress, the activated AMPK successfully promoted cell survival by inhibiting free radical accumulation [44]. Liraglutide reduced oxidative stress by the alteration of AMPK/SREBP1 pathway in Raw264.7 cells [45]. AMPK contributed to the antioxidant activity via regulating the levels of SOD and GSH [46, 47]. Both TM and ABM improved the activities of AMPK by enhancing the expression of its phosphorylated form in liver of 60 min exercised mice. Collectively, the antifatigue effects of *Tricholoma matsutake* may be related to the alteration of AMPK-linked antioxidant pathway.

Although the regulatory effects of TM and ABM on male and female mice were monitored during the experiments, we failed to conclude the different performances on animals of different genders. In the separated experiment, we found that the modulation of sex hormones contribute to the sex differences in response to *Cordyceps militaris*-mediated antifatigue property [32]. Our further study will focus on the regulatory effects of *Tricholoma matsutake* on sex hormones and the relationship between them and antifatigue activities.

In conclusion, our data indicate that TM and ABM elevate the endurance capacity at least in part via activating AMPK-linked antioxidant pathway, which provides experimental

TABLE 2: The antioxidant effects of *Tricholoma matsutake* in exercise fatigue mice.

	CTRL	Female TM (g/kg)		CTRL	0.3	2	ABM (g/kg) 0.4	0.3	Male TM (g/kg)		2	ABM (g/kg) 0.4
		0.3	1						1	2		
Serum												
MDA (nmol/mL)	15.5 ± 0.4	14.3 ± 0.7	13.9 ± 0.3*	13.4 ± 0.4**	17.2 ± 0.5	14.2 ± 0.5*	13.7 ± 0.4*	14.2 ± 0.5*	14.8 ± 0.5*	14.7 ± 0.4**	14.7 ± 0.4**	14.7 ± 0.4**
SOD (U/mL)	105.8 ± 4.0	120.7 ± 2.1*	133.2 ± 6.5**	139.2 ± 5.5	106.7 ± 3.0	118.9 ± 4.9	141.4 ± 5.9**	118.9 ± 4.9	141.3 ± 3.5**	136.1 ± 3.2**	137.8 ± 3.3***	137.8 ± 3.3***
GSH-Px (μmol/mL)	357.9 ± 8.4	390.3 ± 4.8*	427.5 ± 10.4***	418.1 ± 15.9*	340.1 ± 11.8	417.2 ± 12.5**	404.5 ± 8.3**	417.2 ± 12.5**	397.2 ± 12.1*	415.1 ± 14.3**	401.3 ± 9.1**	401.3 ± 9.1**
Muscle												
MDA (nmol/mgprot)	63.1 ± 4.1	51.04 ± 3.1	46.7 ± 2.1*	41.3 ± 3.4**	65.3 ± 2.4	53.1 ± 2.9	48.7 ± 3.4	53.1 ± 2.9	49.5 ± 3.5*	44.6 ± 3.9**	50.8 ± 3.2*	50.8 ± 3.2*
SOD (U/mgprot)	877 ± 3.1	106.9 ± 5.4	111.5 ± 5.7*	130.9 ± 4.4***	86.6 ± 4.5	104.5 ± 6.8	114.1 ± 5.1**	104.5 ± 6.8	111.5 ± 5.3*	127.8 ± 6.0***	114.1 ± 4.6**	114.1 ± 4.6**
GSH-Px (μmol/gprot)	369.2 ± 29.2	469.6 ± 48.2	559.8 ± 476	550.4 ± 38.8**	371.2 ± 27.8	499.5 ± 42.5	514.9 ± 32.1*	499.5 ± 42.5	591.6 ± 45.9**	558.6 ± 46.7*	538.9 ± 42.5	538.9 ± 42.5
ROS (FI/gprot)	15300.7 ± 516.3	14335.0 ± 386.2	12461.6 ± 378.2**	13037.7 ± 527.4*	14294.1 ± 401.6	13532.2 ± 591.9	14277.3 ± 548.6	13532.2 ± 591.9	14396.8 ± 464.7	12162.4 ± 689.7*	11669.1 ± 690.7*	11669.1 ± 690.7*
Liver												
MDA (nmol/mgprot)	12.2 ± 0.7	9.8 ± 0.5	8.7 ± 0.4**	8.8 ± 0.5*	11.8 ± 0.4	9.6 ± 0.7	9.7 ± 0.6	9.6 ± 0.7	9.3 ± 0.6	8.8 ± 0.7*	8.5 ± 0.7*	8.5 ± 0.7*
SOD (U/mgprot)	270.5 ± 17.8	369.8 ± 18.6*	387.5 ± 22.7*	303.9 ± 18.3	240.9 ± 19.2	316.7 ± 16.5	317.9 ± 17.6	316.7 ± 16.5	365.4 ± 22.6**	296.5 ± 15.6	335.9 ± 17.4*	335.9 ± 17.4*
GSH-Px (μmol/gprot)	1013.5 ± 50.9	1244.7 ± 67.5	1264.0 ± 99.3	1361.5 ± 60.4**	929.2 ± 32.1	1169.0 ± 55.1*	1371.9 ± 74.1*	1169.0 ± 55.1*	1098.0 ± 44.4	1365.9 ± 91.8**	1100.8 ± 77.9	1100.8 ± 77.9
ROS (FI/gprot)	1516.2 ± 53.9	1494.2 ± 33.6	1209.1 ± 55.3**	1356.8 ± 46.9	1606.6 ± 75.9	1446.9 ± 46.8	1360.7 ± 30.1	1446.9 ± 46.8	1360.7 ± 62.6	1278.4 ± 51.4*	1286.3 ± 52.7*	1286.3 ± 52.7*

After 60 min swimming, the oxidative stress factors in serum, muscle, and liver of all experimental mice were detected. Data were expressed as mean ± SEM (n = 10). *P < 0.05, **P < 0.01, and ***P < 0.001 versus control group.

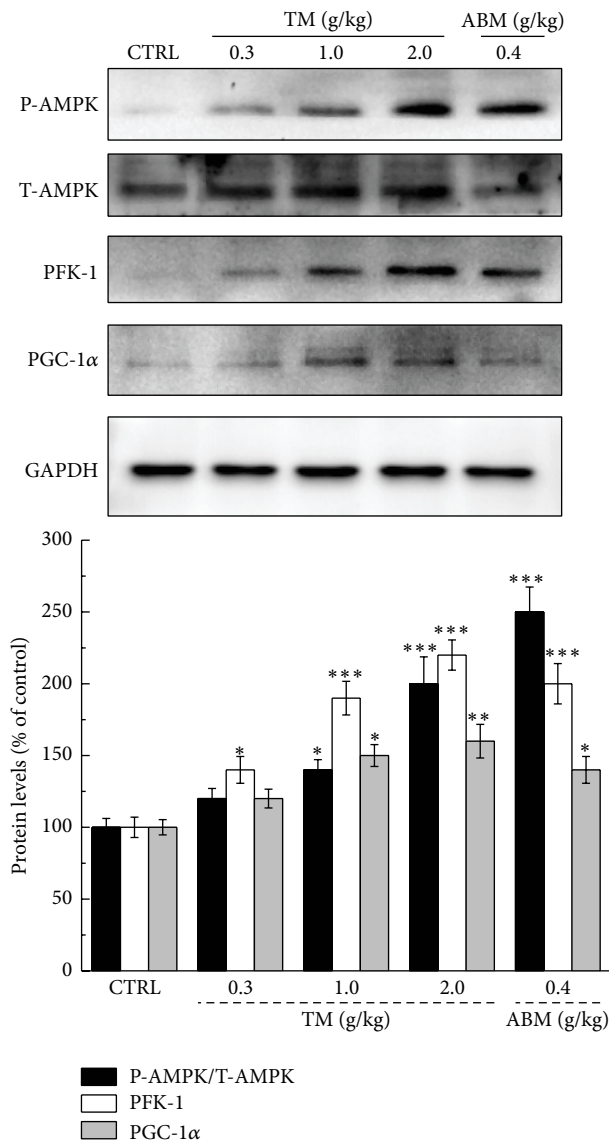


FIGURE 4: At the end of two-week *Tricholoma matsutake* and ABM administration, a 60 min swimming was performed. After exercise, the levels of P-AMPK, PFK-1, and PGC-1 α in liver tissue were detected by western blot. The intensity of target bands was quantified by using ImageJ. Quantification data of P-AMPK, PFK-1, and PGC-1 α were normalized by T-AMPK and GAPDH and expressed as a percent of that from corresponding control mice. Data are expressed as mean \pm SEM ($n = 4$). * $P < 0.05$, ** $P < 0.01$, and *** $P < 0.001$ versus control group.

evidence in supporting the clinical use of *Tricholoma matsutake* as an effective agent against fatigue related diseases.

Conflict of Interests

The authors have declared that there is no conflict of interests.

Authors' Contribution

Quan Li and Yanzhen Wang equally contributed to this project.

Acknowledgments

This work was supported by the Natural Science Foundation of China (Grant no. 81402955) and Science and Technology Key Project in Jilin Province of China (Grant no. 20130201006ZY and 20150203002NY).

References

- [1] C. K. Roberts and R. J. Barnard, "Effects of exercise and diet on chronic disease," *Journal of Applied Physiology*, vol. 98, no. 1, pp. 3–30, 2005.
- [2] R. R. Jenkins, "Exercise, oxidative stress, and antioxidants: a review," *International Journal of Sport Nutrition*, vol. 3, no. 4, pp. 356–375, 1993.
- [3] M. Tanaka, Y. Baba, Y. Kataoka et al., "Effects of (–)-epigallocatechin gallate in liver of an animal model of combined (physical and mental) fatigue," *Nutrition*, vol. 24, no. 6, pp. 599–603, 2008.
- [4] S. M. Shin and S. G. Kim, "Inhibition of arachidonic acid and iron-induced mitochondrial dysfunction and apoptosis by oltipraz and novel 1,2-dithiole-3-thione congeners," *Molecular Pharmacology*, vol. 75, no. 1, pp. 242–253, 2009.
- [5] D. G. Hardie and M. L. J. Ashford, "AMPK: Regulating energy balance at the cellular and whole body levels," *Physiology*, vol. 29, no. 2, pp. 99–107, 2014.
- [6] M. Dékány, V. Nemeskéri, I. Györe, I. Harbula, J. Malomsoki, and J. Pucso, "Antioxidant status of interval-trained athletes in various sports," *International Journal of Sports Medicine*, vol. 27, no. 2, pp. 112–116, 2006.
- [7] G. C. Bogdanis, P. Stavrinou, I. G. Fatouros et al., "Short-term high-intensity interval exercise training attenuates oxidative stress responses and improves antioxidant status in healthy humans," *Food and Chemical Toxicology*, vol. 61, pp. 171–177, 2013.
- [8] K. P. Skenderi, M. Tsironi, C. Lazaropoulou et al., "Changes in free radical generation and antioxidant capacity during ultramarathon foot race," *European Journal of Clinical Investigation*, vol. 38, no. 3, pp. 159–165, 2008.
- [9] C. Y. Wu, R. Chen, X. S. Wang, B. Shen, W. Yue, and Q. Wu, "Antioxidant and anti-fatigue activities of phenolic extract from the seed coat of *Euryale ferox* Salisb. and identification of three phenolic compounds by LC-ESI-MS/MS," *Molecules*, vol. 18, no. 9, pp. 11003–11021, 2013.
- [10] B. Qi, L. Liu, H. Zhang et al., "Anti-fatigue effects of proteins isolated from *Panax quinquefolium*," *Journal of Ethnopharmacology*, vol. 153, no. 2, pp. 430–434, 2014.
- [11] C. Xu, J. Lv, Y. M. Lo, S. W. Cui, X. Hu, and M. Fan, "Effects of oat β -glucan on endurance exercise and its anti-fatigue properties in trained rats," *Carbohydrate Polymers*, vol. 92, no. 2, pp. 1159–1165, 2013.
- [12] X. Liu, W. Zhu, S. Guan et al., "Metabolomic analysis of anti-hypoxia and anti-anxiety effects of Fu Fang Jin Jing Oral Liquid," *PLoS ONE*, vol. 8, no. 10, Article ID e78281, 2013.
- [13] A. Panossian and H. Wagner, "Stimulating effect of adaptogens: an overview with particular reference to their efficacy following single dose administration," *Phytotherapy Research*, vol. 19, no. 10, pp. 819–838, 2005.
- [14] T.-S. Yeh, H.-L. Chuang, W.-C. Huang, Y.-M. Chen, C.-C. Huang, and M.-C. Hsu, "Astragalus membranaceus improves exercise performance and ameliorates exercise-induced fatigue in trained mice," *Molecules*, vol. 19, no. 3, pp. 2793–2807, 2014.

- [15] R. Kumar, P. S. Negi, B. Singh, G. Ilavazhagan, K. Bhargava, and N. K. Sethy, "Cordyceps sinensis promotes exercise endurance capacity of rats by activating skeletal muscle metabolic regulators," *Journal of Ethnopharmacology*, vol. 136, no. 1, pp. 260–266, 2011.
- [16] H. Tong, X. Liu, D. Tian, and X. Sun, "Purification, chemical characterization and radical scavenging activities of alkali-extracted polysaccharide fractions isolated from the fruit bodies of *Tricholoma matsutake*," *World Journal of Microbiology & Biotechnology*, vol. 29, no. 5, pp. 775–780, 2013.
- [17] A. T. Borchers, A. Krishnamurthy, C. L. Keen, F. J. Meyers, and M. E. Gershwin, "The immunobiology of mushrooms," *Experimental Biology and Medicine*, vol. 233, no. 3, pp. 259–276, 2008.
- [18] X. Ding, J. Tang, M. Cao et al., "Structure elucidation and antioxidant activity of a novel polysaccharide isolated from *Tricholoma matsutake*," *International Journal of Biological Macromolecules*, vol. 47, no. 2, pp. 271–275, 2010.
- [19] L. You, Q. Gao, M. Feng et al., "Structural characterisation of polysaccharides from *Tricholoma matsutake* and their antioxidant and antitumour activities," *Food Chemistry*, vol. 138, no. 4, pp. 2242–2249, 2013.
- [20] H. Hoshi, H. Iijima, Y. Ishihara, T. Yasuhara, and K. Matsunaga, "Absorption and tissue distribution of an immunomodulatory α -D-glucan after oral administration of *Tricholoma matsutake*," *Journal of Agricultural and Food Chemistry*, vol. 56, no. 17, pp. 7715–7720, 2008.
- [21] T. Saha, S. Sasmal, S. Alam, and N. Das, "Tamarind kernel powder: a novel agro-residue for the production of cellobiose dehydrogenase under submerged fermentation by *Termitomyces clypeatus*," *Journal of Agricultural and Food Chemistry*, vol. 62, no. 15, pp. 3438–3445, 2014.
- [22] Z. Zhou, Z. Yin, and X. Hu, "Corn cob hydrolysate, an efficient substrate for *Monascus* pigment production through submerged fermentation," *Biotechnology and Applied Biochemistry*, vol. 61, no. 6, pp. 716–723, 2014.
- [23] L. Du, J. Song, H. Wang et al., "Optimization of the fermentation medium for *Paecilomyces tenuipes* N45 using statistical approach," *African Journal of Microbiology Research*, vol. 6, no. 32, pp. 6130–6141, 2012.
- [24] Z. Na, L. Quan, W. Juan et al., "Application of near infrared spectroscopy in quality control of *Irpex lacteus*," *Journal of Food, Agriculture & Environment*, vol. 12, no. 2, pp. 1213–1219, 2014.
- [25] J. Y. Kim, S. E. Byeon, Y. G. Lee et al., "Immunostimulatory activities of polysaccharides from liquid culture of pine-mushroom *Tricholoma matsutake*," *Journal of Microbiology and Biotechnology*, vol. 18, no. 1, pp. 95–103, 2008.
- [26] D. Z. Kang, H. D. Hong, K. I. Kim, and S. Y. Choi, "Anti-fatigue effects of fermented *Rhodiola rosea* extract in mice," *Preventive Nutrition and Food Science*, vol. 20, no. 1, pp. 38–42, 2015.
- [27] J. A. Mercer, B. T. Bates, J. S. Dufek, and A. Hreljac, "Characteristics of shock attenuation during fatigued running," *Journal of Sports Sciences*, vol. 21, no. 11, pp. 911–919, 2003.
- [28] S.-J. Tan, N. Li, F. Zhou et al., "Ginsenoside Rb1 improves energy metabolism in the skeletal muscle of an animal model of postoperative fatigue syndrome," *Journal of Surgical Research*, vol. 191, no. 2, pp. 344–349, 2014.
- [29] L. D. Gershen, B. E. Prayson, and R. A. Prayson, "Pathological characteristics of glycogen storage disease III in skeletal muscle," *Journal of Clinical Neuroscience*, vol. 22, no. 10, pp. 1674–1675, 2015.
- [30] I. Nallamuthu, A. Tamatam, and F. Khanum, "Effect of hydroalcoholic extract of *Aegle marmelos* fruit on radical scavenging activity and exercise-endurance capacity in mice," *Pharmaceutical Biology*, vol. 52, no. 5, pp. 551–559, 2014.
- [31] C. T. Horng, J. K. Huang, H. Y. Wang, C. C. Huang, and F. A. Chen, "Antioxidant and antifatigue activities of polygonatum alte-lobatum hayata rhizomes in rats," *Nutrients*, vol. 6, no. 11, pp. 5327–5337, 2014.
- [32] Y. Dong, S. Hu, C. Liu et al., "Purification of polysaccharides from *Cordyceps militaris* and their antihypoxic effect," *Molecular Medicine Reports*, vol. 11, no. 2, pp. 1312–1317, 2015.
- [33] L. Zhang and N. Wan, "Advances in the research of sport fatigue caused by the action of free radical lipid oxidation," *Chinese Journal of Laboratory Diagnosis*, vol. 10, pp. 1104–1108, 2006.
- [34] S. H. Kim, J. H. Koh, K. Higashida, S. R. Jung, J. O. Holloszy, and D.-H. Han, "PGC-1 α mediates a rapid, exercise-induced downregulation of glycogenolysis in rat skeletal muscle," *Journal of Physiology*, vol. 593, no. 3, pp. 635–643, 2015.
- [35] K. Baar, A. R. Wende, T. E. Jones et al., "Adaptations of skeletal muscle to exercise: rapid increase in the transcriptional coactivator PGC-1," *The FASEB Journal*, vol. 16, no. 14, pp. 1879–1886, 2002.
- [36] G. P. Dobson, W. S. Parkhouse, and P. W. Hochachka, "Regulation of anaerobic ATP-generating pathways in trout fast-twitch skeletal muscle," *American Journal of Physiology—Regulatory Integrative and Comparative Physiology*, vol. 253, no. 1, part 2, pp. R186–R194, 1987.
- [37] A. Whaley-Connell, P. A. McCullough, and J. R. Sowers, "The role of oxidative stress in the metabolic syndrome," *Reviews in Cardiovascular Medicine*, vol. 12, no. 1, pp. 21–29, 2011.
- [38] T.-T. Peternej and J. S. Coombes, "Antioxidant supplementation during exercise training: beneficial or detrimental?" *Sports Medicine*, vol. 41, no. 12, pp. 1043–1069, 2011.
- [39] M. H. El-Maghrabey, N. Kishikawa, K. Ohyama, and N. Kuroda, "Analytical method for lipoperoxidation relevant reactive aldehydes in human sera by high-performance liquid chromatography-fluorescence detection," *Analytical Biochemistry*, vol. 464, pp. 36–42, 2014.
- [40] L. Sun, W. Shen, Z. Liu, S. Guan, J. Liu, and S. Ding, "Endurance exercise causes mitochondrial and oxidative stress in rat liver: effects of a combination of mitochondrial targeting nutrients," *Life Sciences*, vol. 86, no. 1–2, pp. 39–44, 2010.
- [41] R. B. Ceddia, "The role of AMP-activated protein kinase in regulating white adipose tissue metabolism," *Molecular and Cellular Endocrinology*, vol. 366, no. 2, pp. 194–203, 2013.
- [42] M. Ríos, M. Foretz, B. Viollet et al., "Lipoprotein internalisation induced by oncogenic AMPK activation is essential to maintain glioblastoma cell growth," *European Journal of Cancer*, vol. 50, no. 18, pp. 3187–3197, 2014.
- [43] V. A. Narkar, M. Downes, R. T. Yu et al., "AMPK and PPAR δ agonists are exercise mimetics," *Cell*, vol. 134, no. 3, pp. 405–415, 2008.
- [44] M. G. Bonini and B. N. Gantner, "The multifaceted activities of AMPK in tumor progression—why the 'one size fits all' definition does not fit at all?" *IUBMB Life*, vol. 65, no. 11, pp. 889–896, 2013.
- [45] Y.-G. Wang and T.-L. Yang, "Liraglutide reduces oxidized LDL-induced oxidative stress and fatty degeneration in Raw 264.7 cells involving the AMPK/SREBP1 pathway," *Journal of Geriatric Cardiology*, vol. 12, no. 4, pp. 410–416, 2015.

- [46] H.-I. Lee, K. W. Yun, K.-I. Seo, M.-J. Kim, and M.-K. Lee, "Scopoletin prevents alcohol-induced hepatic lipid accumulation by modulating the AMPK-SREBP pathway in diet-induced obese mice," *Metabolism: Clinical and Experimental*, vol. 63, no. 4, pp. 593–601, 2014.
- [47] Y. Zhao, Y. Sun, Y. Ding et al., "GL-V9, a new synthetic flavonoid derivative, ameliorates DSS-induced colitis against oxidative stress by up-regulating Trx-1 expression via activation of AMPK/FOXO3a pathway," *Oncotarget*, vol. 6, no. 28, pp. 26291–26307, 2015.



Hindawi
Submit your manuscripts at
<http://www.hindawi.com>

