# APOL1 Genetic Variants in Focal Segmental Glomerulosclerosis and HIV-Associated Nephropathy

Jeffrey B. Kopp,\* George W. Nelson,† Karmini Sampath,\* Randall C. Johnson,†‡ Giulio Genovese,§ Ping An, David Friedman,§ William Briggs,¶ Richard Dart,\*\* Stephen Korbet,†† Michele H. Mokrzycki,‡‡ Paul L. Kimmel,§§ Sophie Limou, Tejinder S. Ahuja, Jeffrey S. Berns,¶¶ Justyna Fryc,\*\*\* Eric E. Simon,††† Michael C. Smith,†‡‡ Howard Trachtman,§§§ Donna M. Michel, Jeffrey R. Schelling,¶¶¶ David Vlahov,\*\*\*\* Martin Pollak,§ and Cheryl A. Winkler

\*Kidney Disease Section and \$\strict{\strict}\$Division of Kidney, Urologic and Hematologic Diseases, National Institute of Diabetes and Digestive and Kidney Disease, National Institutes of Health, Bethesda, Maryland; \$\frac{1}{7}Basic Science Program Genetics Core, and \$\frac{1}{7}Basic Research Laboratory, Center for Cancer Research, SAIC-Frederick, Inc., National Cancer Institute Frederick, Frederick, Maryland; \$\frac{1}{7}Chaire de Bioinformatique, Conservatoire National des Arts et Metiers, 75003, Paris, France; \$\frac{1}{7}Beth Israel Deaconess Medical Center, Harvard Medical School, Boston, Massachusetts; \$\frac{1}{7}Oakwood University William Beaumont School of Medicine, Royal Oak, Michigan; \*\*Marshfield Clinical Research Foundation, Marshfield, Wisconsin; \$\frac{1}{7}Rush University Medical Center, Chicago, Illinois; \$\frac{1}{7}Albert Einstein College of Medicine, Bronx, New York; \$\frac{1}{1}University of Texas Medical Branch, Galveston, Texas; \$\frac{1}{1}University of Pennsylvania, Philadelphia, Pennsylvania; \*\*\*Bialystok Medical University, Bialystok, Poland; \$\frac{1}{7}Tulane University, New Orleans, Louisiana; \$\frac{1}{7}University Hospital of Cleveland, Cleveland, Ohio; \$\frac{1}{7}S\$Albert Einstein College of Medicine, Cohen Children's Medical Center of New York; \$\frac{1}{1}University, Cleveland, Ohio; and \*\*\*\*Johns Hopkins School of Public Health, Baltimore, Maryland

# **ABSTRACT**

Trypanolytic variants in APOL1, which encodes apolipoprotein L1, associate with kidney disease in African Americans, but whether APOL1-associated glomerular disease has a distinct clinical phenotype is unknown. Here we determined APOL1 genotypes for 271 African American cases, 168 European American cases, and 939 control subjects. In a recessive model, APOL1 variants conferred seventeenfold higher odds (95% CI 11 to 26) for focal segmental glomerulosclerosis (FSGS) and twenty-ninefold higher odds (95% CI 13 to 68) for HIV-associated nephropathy (HIVAN). FSGS associated with two APOL1 risk alleles associated with earlier age of onset (P = 0.01) and faster progression to ESRD (P < 0.01) but similar sensitivity to steroids compared with other subjects. Individuals with two APOL1 risk alleles have an estimated 4% lifetime risk for developing FSGS, and untreated HIVinfected individuals have a 50% risk for developing HIVAN. The effect of carrying two APOL1 risk alleles explains 18% of FSGS and 35% of HIVAN; alternatively, eliminating this effect would reduce FSGS and HIVAN by 67%. A survey of world populations indicated that the APOL1 kidney risk alleles are present only on African chromosomes. In summary, African Americans carrying two APOL1 risk alleles have a greatly increased risk for glomerular disease, and APOL1-associated FSGS occurs earlier and progresses to ESRD more rapidly. These data add to the evidence base required to determine whether genetic testing for APOL1 has a use in clinical practice.

J Am Soc Nephrol 22: 2129-2137, 2011. doi: 10.1681/ASN.2011040388

Received April 18, 2011. Accepted August 4, 2011.

Published online ahead of print. Publication date available at www.iasn.org.

Correspondence: Dr. Jeffrey Kopp, 10 Center Drive, NIH,

Bethesda, MD 20892-1268. Phone: 301-897-4541; Fax: 301-402-0014; E-mail: jeffreyk@intra.niddk.nih.gov

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African Americans have a fourfold increased risk for end-stage kidney disease (ESKD) arising from the three leading causes of chronic kidney disease (CKD): diabetic nephropathy, hypertension-attributed CKD, and glomerulonephritis. 1-3 Two admixture mapping studies identified a locus on chromosome 22 with association with focal segmental glomerulosclerosis (FSGS), HIV-associated nephropathy (HIVAN, which, histologically, is manifested as collapsing glomerulopathy), and pooled nondiabetic ESKD cases, and this was extended to hypertension-attributed ESKD.4,5 The admixture peak was centered on MYH9, encoding nonmuscle myosin IIA heavy chain, and intronic single nucleotide polymorphisms (SNPs) in dbSNP were found to be highly associated with these kidney diseases. Fine mapping and resequencing efforts did not reveal obvious causal variation.<sup>6,7</sup> Recently, the major source of genetic risk for African American nondiabetic ESRD and FSGS was localized to APOL1, encoding apolipoprotein L1 (ApoL1), 14kb from MYH9.8 FSGS and hypertension-attributed ESKD were associated with two APOL1 risk alleles: G1, comprising two missense variants (S342G and I384M), and G2, which has a 6 base pair (bp) in-frame deletion (N388del:Y389del); G1 and G2 are mutually exclusive, never occurring on the same chromosome.8 These variants were strongly associated with sporadic FSGS in either the homozygous or compound heterozygous state.8 The effect of APOL1 variants has not been determined for HIVAN, affecting approximately 10% of HIVinfected African Americans before the initiation of combined antiretroviral therapy.9

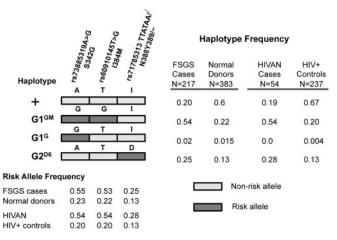
APOL1 and MYH9 are adjacent genes on Chromosome 22, in a region that shows signatures of both historical selection at the time of human and nonhuman divergence and more recent selection within certain African populations.8,10-14 APOL1 is unique to certain higher primates but is not found in nonprimate species. ApoliproteinL1 (ApolL1) lyses Trypanosoma brucei brucei, a cause of disease in a broad range of mammals; because of APOL1, humans are resistant to T. b. brucei infection. 15 Both T. b. rhodensiense and T. b. gambiense, the causes of human African sleeping sickness, have evolved independent mechanisms to avoid lysis by ApoL1. There is suggestive evidence of a recent incomplete, selective sweep of the region, which manifests as extended linkage disequilibrium in the MYH9-APOL1 region in certain African populations and high allele frequency divergence, likely due to selection acting on the APOL1 gene. 12,14 T. b. rhodesiense carries a serum resistanceassociated (SRA) protein that binds and inactivates the wild type ApoL1 protein; the G1 and G2 allelic variants, occurring near and in the SRA binding site, respectively, restore the ability of human serum to lyse T. b. rhodensiense. This appears to represent heterozygous advantage, where protection against trypanosomal infection for heterozygotes comes at the cost of increased risk of kidney disease in homozygotes or compound heterozygotes for APOL1 G1 and G2 alleles.8 These kidney risk alleles are very common (the frequency is approximately 35%) in African Americans and in Yoruba from Nigeria, but the distribution of G1 and G2

alleles has been reported only for a small number African and world populations.<sup>16</sup>

In the present report, we present a comprehensive analysis of APOL1 genetic variation in both African Americans and European Americans with FSGS and HIVAN, and correlate genotype with clinical phenotype. This is the first report to directly test the statistical association of APOL1 genetic variation with biopsy-proven, HIV-associated kidney disease (collapsing glomerulopathy) in a case control study. We also show the frequency distribution of APOL1 variants in well-defined human populations using DNA samples from the Human Genome Diversity Panel (HGDP) and HapMap. As approximately two-thirds of people with HIV disease live in sub-Saharan Africa, and the prevalence of clinically evident kidney diseases rises as patients survive longer,17 these findings have considerable relevance to understanding population differences for kidney disease and for future clinical studies to assess the efficacy of genetic screening in selecting therapy or determining prognosis.

#### **RESULTS**

To more precisely identify the effect size of the *APOL1* variants with FSGS and to determine the effect of these variants on HIVAN, *APOL1* genotypes and haplotypes were obtained for 1378 African American and European American study participants (Figure 1 and Table 1). Allele and haplotype frequencies for African American cases and controls are shown in Figure 1. Allele frequencies in the African American normal donor controls were 23% for G1 and 13% for G2. Because a subset of our



**Figure 1.** Distribution of haplotypes and risk alleles. Four *APOL1* haplotypes were observed in the African American study group: the major haplotype, denoted as (+) and comprised of three nonrisk alleles; the  $G1^{GM}$  haplotype with two missense risk alleles; the rare  $G1^{G+}$  haplotype with one missense risk allele; and the  $G2^{\Delta6}$  haplotype with the 6 bp deletion risk allele. The risk allele and haplotype frequencies are shown for FSGS and HIVAN case and control groups in African Americans, including healthy blood donors and HIV+ individuals with no kidney disease.

Table 1. Demographic characteristics of FSGS and HIVAN cases and controls

Population Group	Cases	n	Age of Onset (yr)	Controls	n	Age at Blood Donation (yr)	Total
African Americans	Primary FSGS	217	34 ± 14	Normal donors	383	44 ± 8	600
African Americans	HIVAN	54	$36 \pm 7$	HIV + hypernormals	237	$44 \pm 6$	291
European Americans	Primary FSGS	168	$33 \pm 17$	Normal donors	319	$50 \pm 9$	487
Total		439			939		1378

This table summarizes numbers and age distribution of the six groups of subjects studied, as mean ± SD. Hypernormal controls are those that have HIV disease for >8 yr and are lacking evidence of CKD (elevated creatinine or proteinuria). FSGS, focal segmental glomerulosclerosis; HIVAN, HIV-associated collapsing glomerulopathy.

FSGS cases and controls was used in the Genovese et al. study,8 we first replicated the reported associations using an independent and nonoverlapping group of primary (idiopathic) FSGS cases (n = 77) and controls (n = 310) under the recessive model for APOL1 G1 and/or G2 (OR = 42.2 (95% CI 13.1, 68.5). Next, we used the entire group of primary FSGS cases (n = 217) and controls (n = 383), as well as 54 biopsy-proven HIVAN cases and 237 hypernormal controls, to more precisely estimate the effect sizes for the alleles and their haplotypes, and to extend the associations to biopsy proven HIVAN. The G1 and G2 risk alleles are in complete negative linkage disequilibrium (LD) and never appear together on the same chromosome (Figure 1); a recombination event between the physically proximal G1 and G2 variants that would bring them together on the same chromosome has not been observed in this dataset or previously reported.8 Therefore, to determine the effect size of each of the alleles, it is necessary to stratify the analysis by the other allele (Table 2). These analyses show strong and essentially equivalent associations for individuals carrying two copies of G1, two copies of G2, or one copy of each; because of the complete negative LD, an individual can carry no more than two copies of G1 and/or G2. The odds ratios for carrying two copies of G1 and/or G2 were 17 (95% CI 11, 26) for primary FSGS and 29 (95% CI 13, 68) for HIVAN. There was a marginally significant association, with a much smaller effect size, for carriers of a single copy of the G1 allele without the G2 allele (OR 1.9, 95% CI 1.01, 3.5) for the combined FSGS and HIVAN, but no significant association for carriers of a single copy of G2. APOL1 was resequenced in all our controls and cases, but no rare or private nonsynonymous variants were observed to contribute to this modest single copy gene effect (data not shown). These associations and levels of statistical support were nearly identical in a logistic regression analysis adjusting for age of onset (data not shown).

When we did the same analyses, but without accounting for the presence or absence of a risk allele on the alternative chromosome, we noted strong artifactual associations for heterozygotes of *APOL1* G1 or G2 (Supplementary Table 1), due to the unaccounted presence of the alternative risk allele on the second haplotype. As can be seen by comparing the results in Supplementary Table 1 to those in Table 2, this apparent dominant or additive association is misleading.

The APOL1 G1 and G2 variants were rare in European Americans: Among 320 controls, one G2 (frequency of ap-

proximately 0.3%) and four G1 (frequency of approximately 1.3%) heterozygotes were observed. Among the 165 European American FSGS cases, two G2 heterozygotes, one G1 heterozygote, and one G1 homozygote were observed. A principal components analysis of ancestry in a previous study showed that three of these four cases had substantial African admixture (>80%).<sup>5</sup>

Onset ages for primary FSGS were highly variable. We considered that FSGS associated with a specific genetic lesion represented by homozygosity or compound heterozygosity for APOL1 risk alleles would plausibly have a different age distribution than other forms of primary FSGS, which could represent a broader spectrum of underlying causes. Among 182 African Americans for whom age of onset data were available, the mean ± SD age of onset for those with 0 or 1 APOL1 risk alleles was 37.6  $\pm$  16.0 yr (n = 64), compared with 31.7  $\pm$  11.9 yr for those with 2 APOL1 risk alleles (n = 118; P = 0.01, t test with unequal variances) (Figure 2). The variance of age on onset was 45% less in the APOL1 risk genotype (P = 0.003, F test for equality of variance). Onset ages were 39.2  $\pm$  14.4 yr (n = 22) for those with 0 APOL1 risk alleles, and  $36.7 \pm 17.0 (n = 42)$ for those with 1 *APOL1* risk allele (P > 0.2 for tests of equality of means and variances). As shown in Table 3, 70% of individuals with two APOL1 risk alleles experience onset of FSGS between the ages of 15 to 39 compared with 42% of carriers of 0 or 1 risk alleles (P = 0.0003, FET). Together these data support the idea that FSGS among APOL1 high-risk individuals is a distinct disease with earlier onset and tendency to present during the teenage years and early adulthood.

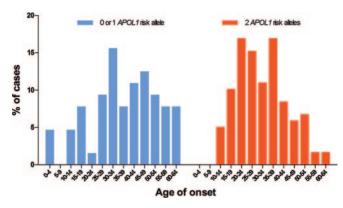
FSGS associated with nearly all Mendelian genetic mutations is resistant to glucocorticoid therapy, including those associated with the most common recessive gene, NPHS2 (podocin). Among African American FSGS cases, 56 had received at least 8 wk of glucocorticoid therapy, the minimum required to determine responsiveness. The frequency of steroid sensitivity was 29% (12/42) in African American subjects with two APOL1 risk alleles who were given an adequate trial of glucocorticoid therapy, and 33% (5/15) in subjects with zero or one APOL1 risk allele (P > 0.5). Due to limited number of informative subjects, we cannot rule out a modest influence on steroid sensitivity in carriers of two APOL1 variants.

We analyzed the duration from disease onset to ESKD, defined as estimated GFR <15 ml/min/1.73m<sup>2</sup>, chronic dialysis, or kidney transplantation among 92 African American FSGS

Table 2. Independent effects of the APOL1 G1 and G2 risk alleles

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		Nimbou	I VS. U		Z VS. U	0	Z VS. I		Recessive Model 2 vs. 1 or	81 Z VS. 1 OF
Risk Allele	Stratum	Number ()	Risk Allele	<u>е</u>	Risk Alleles	eles	Risk Alleles	eles	0 Risk Alleles	leles
		Case/Control	OR (CI)	Ь	OR (CI)	Ь	OR (CI)	Ь	OR (CI)	Ь
HIV-associate	d collapsing	glomerulopathy ( <i>n</i>	n = 54) and hype	r-normal	IIV-associated collapsing glomerulopathy ( $n = 54$ ) and hyper-normal controls ( $n = 237$ )					
G1	No G2	27/180	1.9 (0.5, 8)	0.37	47.4 (11.9, 231.5)	$4.9 \times 10^{-11}$	24.3 (6.6, 105.4)	$2.2 \times 10^{-8}$	35.2 (10.7, 132.6)	$2.3 \times 10^{-11}$
G2	No G1	11/150	1.4 (0.2, 7.5)	0.70	14.4 (1.7, 116.3)	0.007	10.1 (1.02, 105.9)	0.02	13.1 (1.7, 93.7)	0.007
G1/G2	G1/G2 <sup>b</sup>	26/113		I	44.9 (12.9, 192.1)	$2.2 \times 10^{-13}$	I	I	I	I
G1 or G2	Ħ	54/237	1.8 (0.5, 6.8)	0.42	40.4 (13.7, 148.4)	$4.10 \times 10^{-18}$	23.2 (9.5, 61.6)	$1.4 \times 10^{-16}$	29.2 (13.1, 68.5)	$6 \times 10^{-22}$
African Ameri	ican idiopathi	African American idiopathic FSGS cases ( $n=217$ ) and contro	= 217) and contro	ols $(n = 383)$	383)					
G1	No G2	125/286	1.9 (1, 3.5)	0.05	23.2 (12, 46.7)	$7.3 \times 10^{-29}$	11.6 (6.0, 23.2)	$4.7 \times 10^{-17}$	17.4 (9.8, 31.7)	$4.3 \times 10^{-30}$
G2	No G1	56/234	1.0 (0.4, 2.3)	1.0	25.1 (8.8, 83.3)	$7.8 \times 10^{-13}$	17.0 (5.1, 65.4)	$8.5 \times 10^{-8}$	25.3 (9.2, 81.7)	$1.4 \times 10^{-13}$
G1/G2	G1/G2	82/186	I	I	16.7 (8.5, 34)	$7.3 \times 10^{-21}$	I	I	I	I
G1 or G2	₹	217/383	1.4 (0.8–2.6)	0.26	20.6 (11.8–37)	$6.7 \times 10^{-38}$	14.6 (8.7, 25.1)	$6.4 \times 10^{-23}$	16.9 (11–26.5)	$1.3 \times 10^{-48}$

he APOL1 risk alleles are referred to as follows: G1, S342G mutation; G2, 6 bp deletion (N388del:Y389del). Stratum refers to analysis that included a subset of genotypes (e.g., all cases and controls that acked the G2 allele or G1 allele). The strata do not add to the total due to overlap between strata. G1/G2 compound heterozygotes were compared to subjects lacking both G1 and G2 alleles. P values were determined by Fisher exact test. As shown, the data best fit a recessive mode of inheritance, with marginal evidence for a dominant effect of the G1 allele in idiopathic FSGS.



**Figure 2.** Distribution of age of onset of primary FSGS for APOL1 risk allele groups. Age of onset of primary FSGS is presented for African Americans with 0 or 1 APOL1 risk alleles (n = 63) and with 2 APOL1 risk alleles (n = 117). Height of bars represents the percentage of subjects of each group falling in each age group. As shown, APOL1-associated FSGS tends to present at a younger age and at a narrower onset age range, with 70% of cases presenting between 15 and 39 versus 43% of carriers of 0 or 1 risk alleles (P = 0.0009) (SD of presentation age in years, 11.8 versus 16.2, P = 0.004).

**Table 3.** Onset age for primary FSGS in African Americans stratified by G1 or G2 APOL1 risk alleles

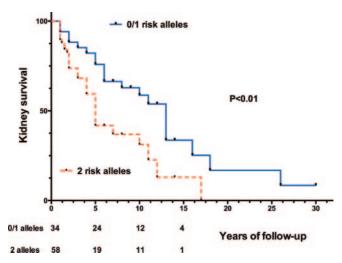
Onset Age	0 or 1 APOL1 Risk Alleles	2 APOL1 Risk Alleles
(years)	(n = 64)	(n = 118)
<15	9%	5%
15–39	42%	70%
≥ 40	48%	25%

The distribution of onset ages for primary FSGS onset is shown. P=0.0003 for the 3x2 contingency table by Fisher's exact test.

patients who were evaluated at the National Institutes of Health (NIH) Clinical Center (Figure 3). Progression to ESKD was significantly faster among subjects with two APOL1 risk alleles, with a hazard ratio = 2.3 (95% CI 1.43, 4).

Among African American subjects, there were exceptions to the reported observation that the two SNPs defining the G1 risk allele are in absolute positive linkage disequilibrium. An infrequent haplotype carrying the risk allele for G1S $^{342G}$  and the common allele at G1 $^{1384M}$  was observed. This haplotype, G1G+, carried by seven controls and seven cases, accounts for less than 1% of the *APOL1* haplotypes in the combined control groups. The common haplotype (+) containing the major frequency alleles for the two SNPs, and for the 6-bp insertion deletion, was most frequent in the controls (approximately 65%). In the HIVAN and FSGS case groups, the most frequent haplotype was G1GM (54%) and the second most frequent was G2 $^{\Delta 6}$  (25% and 28%, respectively). The G1GM and G2 $^{\Delta 6}$  haplotypes were less frequent (20 to 22%, and 13%, respectively) in both African American control groups (Figure 1).

To determine whether both G1 mutant alleles (342G and 384M) or only G1  $^{\rm S342G}$  contribute kidney disease risk, we compared the effect sizes of the four G1 haplotype combinations (+/G  $^{\rm GM}$ , +/G1  $^{\rm G+}$ , G1  $^{\rm GM}$ /G1  $^{\rm GM}$ , G1  $^{\rm GM}$ /G  $^{\rm G+}$ ) to the reference



**Figure 3.** APOL1 variants and kidney survival. The Kaplan-Meier plot depicts the kidney survival among 92 African American patients with primary FSGS, with the number of subjects at risk at each time point shown. The curves differed by the log rank test (P < 0.01). The hazard ratio for progression was 2.3 (95% CI 1.43, 4), and median survival was 5 yr for subjects with 2 APOL1 risks alleles, compared with 13 yr for subjects with 0 or 1 APOL1 risk allele. Survival curves were similar for 0 and 1 risk allele subjects (P > 0.05) and are therefore combined.

group (+/+). Due to the large effect of  $G1^{342G}$ , the relatively few individuals with the  $G1^{G+}$  haplotype were sufficient to show that this haplotype had an effect size similar to the  $G1^{GM}$  risk haplotype but different from the common (+) haplotype (P=0.02) (Figure 4). We were unable to test the risk associated with  $G1^{G+}/G2$  or the  $G1^{G+}/G1^{G+}$  genotypes (carried by one individual and by no individuals, respectively), but the  $G1^{GM}/G^{G+}$  association strongly suggests that the single  $G1^{S342G}$  allele determines the G1 pathogenic phenotype, and it is sufficient to type just  $G1^{S342G}$  and the G2 insertion/deletion to assess kidney disease risk.

Previous studies had shown that the G1 and G2 *APOL1* risk alleles were most common in western and southern Africa.<sup>8,16</sup> We determined the distribution of *APOL1* risk alleles in DNA samples from 1024 individuals from 51 discrete human populations in HDGP, and in an additional 60 Yoruba from Nigeria and 90 Luhya from Kenya in the HapMap Project. We also include 702 African and European Americans controls from the state of Maryland. These results, displayed in Figure 5 and Supplementary Table 2, indicate that the G1 and G2 variants are differentially distributed throughout Africa. The G1 allele is most frequent in West Africa, whereas the G2 allele is less frequent but more widely distributed.

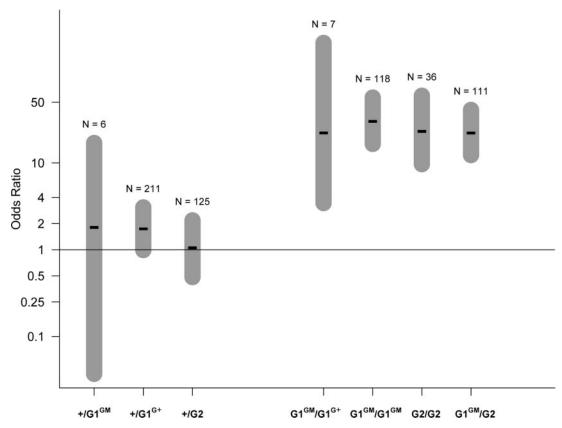
The attributable risk for homozygotes or compound heterozygotes for *APOL1* G1 and G2 risk alleles, in the recessive genetic model, is 68% for both FSGS and HIVAN. The explained fractions for G1 and G2 are 18% for FSGS and 35% for HIVAN. These are strikingly high values; for comparison, the explained fraction of smoking for lung cancer is 10 to 12%<sup>19,20</sup> Similarly, *APOL1* genotype is estimated to have a large effect

on lifetime risk of FSGS or HIVAN. For HIVAN, assuming a population lifetime risk for the development of HIVAN of 10.0% for HIV-infected African Americans who have not received antiretroviral therapy, the lifetime risks are 2.5%, 4.0%, and 50.0% in carriers of 0, 1, or 2 *APOL1* risk alleles, respectively. For FSGS, assuming an African American population lifetime risk of 0.8%,<sup>21</sup> the lifetime risks are 0.2%, 0.3%, and 4.25% for these genotypes.

#### **DISCUSSION**

We investigated the association of APOL1 genetic variation in FSGS and HIVAN and have several findings. First, we show, for the first time, that APOL1 G1 and G2 alleles are strongly associated with HIVAN and confirm their association with primary FSGS; the associations are best described by a recessive model. We have more precisely determined the true effect sizes for these alleles for primary FSGS and HIVAN, with OR of 17 and 29, respectively, in a recessive model. These are the strongest effect sizes discovered, to date, for common variants with a complex disease.<sup>22</sup> Second, we show that the G1<sup>384M</sup> allele is probably not required for pathogenesis, as the  $\mathrm{G1}^{\mathrm{G+}}$  haplotype containing just the  $\mathrm{G1}^{\mathrm{342G}}$  allele can complement the  $\mathrm{G1}^{\mathrm{GM}}$ haplotype containing both variant alleles; we lack data for the other possible combinations (G1<sup>G+</sup>/ G1<sup>G+</sup> or G1<sup>G+</sup>/ G2). Third, we found no significant differences in glucocorticoid sensitivity in subjects who had received an adequate therapeutic trial. While the numbers of subjects are relatively small, they provide no evidence to support withholding glucocorticoid therapy in individuals carrying two APOL1 risk alleles. Fourth, we show that the age of onset is significantly earlier for G1 and/or G2 homozygotes or compound heterozygotes. Fifth, we have shown that kidney survival is significantly shorter for individuals with primary FSGS who are carrying two APOL1 risk alleles. Sixth, explained fractions for APOL1 variants are notably high, at 18% for FSGS and 35% for HIVAN; the variants increase the risk of developing primary FSGS and HIVAN by approximately twentyfold compared with individuals lacking two risk alleles. We note that estimates for HIVAN were based on data from the era before the widespread use of effective antiviral therapy, and that the epidemiology of HIV-associated kidney disease has and continues to change. Finally, we show that both the G1 and G2 risk alleles are widely distributed across sub-Saharan Africa, but both kidney disease risk alleles are absent from populations outside of sub-Saharan Africa and in populations without African ancestry.

The mechanism by which *APOL1* variants cause kidney disease remains unknown. Apolipoprotein L1 is a circulating protein, a component of high-density lipoprotein (HDL) class 3, particularly the dense subset (class 3a), and is largely absent from other classes. HDL3 has the particular function of protecting low-density lipoprotein (LDL) particles from oxidation.<sup>23</sup> Whether apolipoproteinL1 has a role in this antioxidant function is unknown. Likewise, it remains to be determined

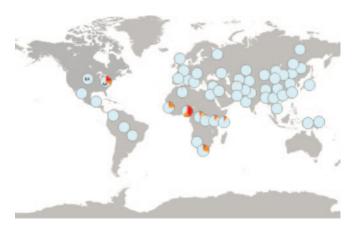


**Figure 4.** Odds ratios for the effects of the APOL1 G1 and G2 alleles. Odds ratios with confidence intervals for association of different G1–G2 genotypes with combined FSGS/HIVAN cases compared with subjects carrying no G1 or G2 risk alleles (+/+). Heterozygous carriers of the risk alleles do not show significant association with FSGS/HIVAN cases, with the exception that combined +/  $G1^{GM}$  and +/  $G1^{G+}$  was marginally significant (OR 1.8, P=0.02). By contrast, dual heterozygous and homozygous individuals showed a consistent association. In this analysis, the rare  $G1^{G+}$  haplotype is distinguished from the common  $G1^{GM}$  haplotype and  $G1^{G+}$  is able to complement  $G1^{GM}$ , resulting in disease association for this genotype. A meaningful odds ratio for  $G1^{G+}/G2$ , carried by one subject, could not be determined.

whether circulating apoL1, kidney-expressed apoL1, or both contribute to kidney injury. As noted above, the G1 and G2 variants are located in the C-terminal domain (residues 342 to 398) of apolipoproteinL1, the binding region for the trypanosomal SRA.15 The function of this domain in human physiology is unknown, but the fact that both pathogenic variants occur in this domain may be a clue to kidney disease pathogenesis. Structural studies will be required to determine how these variants may alter domain structure. The data presented here indicate that I384M mutation does not contribute to pathogenesis. Until APOL1 variants are directly implicated in kidney disease by experimental techniques such as expression in transgenic animals or injection of recombinant protein into animals, it remains possible that kidney injury is due not to APOL1 variants but to another gene in linkage disequilibrium. However, only a few SNPs in the region are in moderate to strong linkage disequilibrium ( $r^2 > 0.6$ ) with G1 and G2, and these are six or more orders of magnitude less statistically significant than the APOL1 variants. The 1000 Genomes Project has not yet revealed plausible alternative candidates to APOL1 G1 and G2 in the region.24

APOL1 genetic variation is an attractive candidate for personalized medicine. Our data suggest a striking increase in disease prevalence among APOL1 risk variant homozygotes. We estimate the lifetime incidence for HIVAN of 50% among HIV-infected African Americans with two APOL1 risk alleles who are not receiving antiviral therapy. Future studies will be required to demonstrate whether genetic testing could play a role in the initiation of antiretroviral therapy in an individual who does not otherwise meet the criteria. Before such a recommendation can be made, an appropriate controlled trial is needed. We estimate the lifetime incidence for primary FSGS among African Americans with two APOL1 risk alleles to be 4%. Some important questions remain, including whether the initial phase of APOL1-associated FSGS may involve a sustained period of microalbuminuria or how best to treat these individuals when they are glucocorticoid-resistant.

In conclusion, *APOL1* genetic variation is a powerful contributor to risk for FSGS and HIVAN. The attributable risk and explained fraction for *APOL1* variants are quantitatively comparable to the role of smoking in non-small-cell lung cancer risk. While only 12 to 13% of African Americans carry two



**Figure 5.** Worldwide frequency distribution of *APOL1* variants G1 and G2. Genotypes of G1 and G2 are determined for 51 populations in the Centre d'Etude du Polymorphisme Humain (CEPH) HGDP, for the HapMap Luhya population from Kenya (International HapMap Project [Phase II]), and for African Americans (AA) and European American (EA) controls in the NIH FSGS study cohort. The allele frequencies of G1 (red), G2 (orange), and wild-type alleles (light blue) in each population are depicted in pie charts overlaid upon a world map. Allele frequencies for G1 and G2 for each population are shown in Supplementary Table 2.

APOL1 risk alleles, these individuals have an estimated 4% lifetime risk for FSGS, and when they develop FSGS, they tend to progress to ESKD rapidly. Further studies will be required to determine whether early detection could alter the clinical course, and thus provide a rationale for genetic screening in the context of personalized medicine.

# **CONCISE METHODS**

## **Study Participants**

The study includes three case-control groups, two of which were selfdescribed as African American and one of which was self-described as European American, as shown in Table 1. Cases included African Americans with biopsy-proven primary FSGS (n = 217) or biopsyproven HIVAN, defined histologically as HIV-associated collapsing glomerulopathy (n = 54), and European Americans with biopsyproven primary FSGS (n = 168). The cases and controls have been previously described.<sup>5,25</sup> Primary FSGS and HIVAN cases were enrolled in the NIH FSGS Genetic Study from 22 academic medical centers in the United States. Institutional review boards at each collaborating medical center approved study protocols and each subject provided written informed consent. Renal diagnoses at each institution were made according to standard pathologic criteria. For simplicity, primary collapsing glomerulopathy was classified under primary FSGS. We excluded subjects judged to have postadaptive FSGS as a result of reflux nephropathy, reduced renal mass, sickle cell nephropathy, or morbid obesity and FSGS due to medications.

Onset age was defined as the age of proteinuria onset. Glucocorticoid sensitivity was defined as complete or partial remission in subjects who had received at least 8 wk of daily or alternate-day glucocorticoid therapy.<sup>26</sup> Glucocorticoid sensitivity was evaluable for 56 African Americans with FSGS (26% of the total number of cases); the other subjects did not receive the required course of glucocorticoids.

# **DNA Samples for Global Diversity**

DNA samples from the HGDP, the Foundation Jean Dausset-CEPH, Paris, France,<sup>27</sup> the Yoruba trios from Nigeria, and the Luhya samples from Kenya, both sets taken from the International HapMap Project, were used to determine G1 and G2 allele frequencies; the DNA and cell lines were obtained from the Coriell Institute (Camden, New Jersey). Genotypes were determined for 1024 individuals from 51 distinct ethnic groups from HGDP, and 60 Yoruba parents and 90 Luhya for G1 (rs73885319 (S342G) and rs60910145 (I384M), and the G2 6 bp insertion/deletion (indel) (rs71785313).

### **Terminology**

For APOL1 haplotypes, we use the terms  $G1^{G+}$  (harboring the minor frequency risk allele at S342G only) and  $G1^{GM}$  (harboring the minor frequency risk alleles S342G plus I384M). The G2 haplotype harboring the 6 bp in-frame deletion is termed  $G2^{\Delta6}$ . For APOL1 alleles, we use the terms G1, the risk allele for S342G that occurs on two G1 haplotypes ( $G1^{G+}$  and  $G1^{GM}$ ), and G2 for the deletion allele (Figure 1). We use "+" to designate the most frequent (nonrisk) state for haplotypes and alleles.

### Genotyping

APOLI SNPs (rs73885319 and rs60910145) and the G2 indel (rs717185313) were genotyped by TaqMan assays (ABI, Foster City, California). Each SNP was tested for deviations from Hardy-Weinberg expectations (HWE) among control subjects using a  $\chi^2$  goodness-of-fit test. All of the tested SNPs confirmed to HWE in the normal donor control groups.

#### Statistical Methods

Fisher's exact test (FET) was used to compare individuals with one or two risk alleles (G1 or G2) *versus* zero or one risk alleles (+/+), and also for the recessive model comparing individuals with G1/G1, G2/G2, or G1/G2 genotypes to individuals with +/G1, for primary FSGS and HIVAN. A similar analysis was performed comparing the  $G1^{GM}$  and  $G1^{G+}$  haplotypes and their diplotypes.

For the analysis of age of onset of FSGS, an *F* test (*R* function var.test) was used to test for inequality of the variances of the two groups with and without *APOL1* risk genotypes; a Welch two-sample *t* test was used to compare the means (*R* function t.test). Difference in kidney survival was analyzed with the log-rank test (Mantel-Cox), using Prism (Graph Pad, San Diego, California). For attributable risk, explained fractions, and lifetime estimate risks, population contingency tables were extrapolated from case-control data by scaling the number of controls to yield risks of 0.8% for FSGS and 10% (among untreated HIV infected subjects) for HIVAN, based on estimates of African American population lifetime risk.<sup>20,28,29</sup>

# **ACKNOWLEDGMENTS**

The authors acknowledge the assistance of Dr. Susan Leitman, National Institutes of Health (NIH) Clinical Center, for assistance with

recruitment of blood donors, and Dr. Greg Germino and Dr. Rebekah Rasooly, National Institute of Diabetes and Digestive and Kidney Disease (NIDDK), for critical reviews of the manuscript.

We thank E. Binns-Roemer and Y. Zhou for excellent technical support and Bailey Kessing for database support. This project has been funded, in whole or in part, with federal funds from the National Cancer Institute, National Institutes of Health, under contract HHSN261200800001E and the intramural Program of NIDDK, NIH. The content of this publication does not necessarily reflect the views or policies of the Department of Health and Human Services, nor does mention of trade names, commercial products, or organizations imply endorsement by the U.S. government.

Author contributions: J.B.K. and C.A.W. conceived the study and drafted the paper, with assistance from D.F., M.P., and G.W.N; G.W.N. performed statistical analysis, with help from R.C.J., G.G., P.A., and S.L; J.F. provided clinical samples; J.B.K., K.S., W.B., R.D., S.K., M.M., P.L.K., T.S.A., J.S.B., E.S., M.C.S., H.T., D.M.M., J.R.S., and D.V. provided patient materials, under the auspices of a multicenter study.

#### **DISCLOSURES**

None.

### **REFERENCES**

- Coresh J, Byrd-Holt D, Astor BC, Briggs JP, Eggers PW, Lacher DA, Hostetter TH: Chronic kidney disease awareness, prevalence, and trends among U.S. adults, 1999 to 2000. J Am Soc Nephrol 16: 180–188, 2005
- Satko SG, Sedor JR, Iyengar SK, Freedman BI: Familial clustering of chronic kidney disease. Semin Dial 20: 229–236, 2007
- United States Renal Data Systems 2010 Annual Data Report: Volume

   Atlas of Chronic Renal Disease in the United States, National Institutes of Health, National Institute of Diabetes and Digestive and Kidney Diseases, Bethesda, MD, 2010
- 4. Kao WH, Klag MJ, Meoni LA, Reich D, Berthier-Schaad Y, Li M, Coresh J, Patterson N, Tandon A, Powe NR, Fink NE, Sadler JH, Weir MR, Abboud HE, Adler SG, Divers J, Iyengar SK, Freedman BI, Kimmel PL, Knowler WC, Kohn OF, Kramp K, Leehey DJ, Nicholas SB, Pahl MV, Schelling JR, Sedor JR, Thornley-Brown D, Winkler CA, Smith MW, Parekh RS: MYH9 is associated with nondiabetic end-stage renal disease in African Americans. *Nat Genet* 40: 1185–1192, 2008
- Kopp JB, Smith MW, Nelson GW, Johnson RC, Freedman BI, Bowden DW, Oleksyk T, McKenzie LM, Kajiyama H, Ahuja TS, Berns JS, Briggs W, Cho ME, Dart RA, Kimmel PL, Korbet SM, Michel DM, Mokrzycki MH, Schelling JR, Simon E, Trachtman H, Vlahov D, Winkler CA: MYH9 is a major-effect risk gene for focal segmental glomerulosclerosis. Nat Genet 40: 1175–1184, 2008
- Behar DM, Rosset S, Tzur S, Selig S, Yudkovsky G, Bercovici S, Kopp JB, Winkler CA, Nelson GW, Wasser WG, Skorecki K: African ancestry allelic variation at the MYH9 gene contributes to increased susceptibility to non-diabetic end-stage kidney disease in Hispanic Americans. Hum Mol Genet 19: 1816–1827, 2010
- Nelson GW, Freedman BI, Bowden DW, Langefeld CD, An P, Hicks PJ, Bostrom MA, Johnson RC, Kopp JB, Winkler CA: Dense mapping of MYH9 localizes the strongest kidney disease associations to the region of introns 13 to 15. Hum Mol Genet 19: 1805–1815, 2010
- 8. Genovese G, Friedman DJ, Ross MD, Lecordier L, Uzureau P, Freed-

- man BI, Bowden DW, Langefeld CD, Oleksyk TK, Uscinski Knob AL, Bernhardy AJ, Hicks PJ, Nelson GW, Vanhollebeke B, Winkler CA, Kopp JB, Pays E, Pollak MR: Association of trypanolytic ApoL1 variants with kidney disease in African Americans. *Science* 329: 841–845, 2010
- Shahinian V, Rajaraman S, Borucki M, Grady J, Hollander WM, Ahuja TS: Prevalence of HIV-associated nephropathy in autopsies of HIVinfected patients. Am J Kidney Dis 35: 884–888, 2000
- 10. Frazer KA, Ballinger DG, Cox DR, Hinds DA, Stuve LL, Gibbs RA, Belmont JW, Boudreau A, Hardenbol P, Leal SM, Pasternak S, Wheeler DA, Willis TD, Yu F, Yang H, Zeng C, Gao Y, Hu H, Hu W, Li C, Lin W, Liu S, Pan H, Tang X, Wang J, Wang W, Yu J, Zhang B, Zhang Q, Zhao H, Zhou J, Gabriel SB, Barry R, Blumenstiel B, Camargo A, Defelice M, Faggart M, Goyette M, Gupta S, Moore J, Nguyen H, Onofrio RC, Parkin M, Roy J, Stahl E, Winchester E, Ziaugra L, Altshuler D, Shen Y, Yao Z, Huang W, Chu X, He Y, Jin L, Liu Y, Sun W, Wang H, Wang Y, Xiong X, Xu L, Waye MM, Tsui SK, Xue H, Wong JT, Galver LM, Fan JB, Gunderson K, Murray SS, Oliphant AR, Chee MS, Montpetit A, Chagnon F, Ferretti V, Leboeuf M, Olivier JF, Phillips MS, Roumy S, Sallee C, Verner A, Hudson TJ, Kwok PY, Cai D, Koboldt DC, Miller RD, Pawlikowska L, Taillon-Miller P, Xiao M, Tsui LC, Mak W, Song YQ, Tam PK, Nakamura Y, Kawaguchi T, Kitamoto T, Morizono T, Nagashima A, Ohnishi Y, Sekine A, Tanaka T, Tsunoda T, Deloukas P, Bird CP, Delgado M, Dermitzakis ET, Gwilliam R, Hunt S, Morrison J, Powell D, Stranger BE, Whittaker P, Bentley DR, Daly MJ, de Bakker PI, Barrett J, Chretien YR, Maller J, McCarroll S, Patterson N, Pe'er I, Price A, Purcell S, Richter DJ, Sabeti P, Saxena R, Schaffner SF, Sham PC, Varilly P, Stein LD, Krishnan L, Smith AV, Tello-Ruiz MK, Thorisson GA, Chakravarti A, Chen PE, Cutler DJ, Kashuk CS, Lin S, Abecasis GR, Guan W, Li Y, Munro HM, Qin ZS, Thomas DJ, McVean G, Auton A, Bottolo L, Cardin N, Eyheramendy S, Freeman C, Marchini J, Myers S, Spencer C, Stephens M, Donnelly P, Cardon LR, Clarke G, Evans DM, Morris AP, Weir BS, Mullikin JC, Sherry ST, Feolo M, Skol A, Zhang H, Matsuda I, Fukushima Y, Macer DR, Suda E, Rotimi CN, Adebamowo CA, Ajayi I, Aniagwu T, Marshall PA, Nkwodimmah C, Royal CD, Leppert MF, Dixon M, Peiffer A, Qiu R, Kent A, Kato K, Niikawa N, Adewole IF, Knoppers BM, Foster MW, Clayton EW, Watkin J, Muzny D, Nazareth L, Sodergren E, Weinstock GM, Yakub I, Birren BW, Wilson RK, Fulton LL, Rogers J, Burton J, Carter NP, Clee CM, Griffiths M, Jones MC, McLay K, Plumb RW, Ross MT, Sims SK, Willey DL, Chen Z, Han H, Kang L, Godbout M, Wallenburg JC, L'Archeveque P, Bellemare G, Saeki K, An D, Fu H, Li Q, Wang Z, Wang R, Holden AL, Brooks LD, McEwen JE, Guyer MS, Wang VO, Peterson JL, Shi M, Spiegel J, Sung LM, Zacharia LF, Collins FS, Kennedy K, Jamieson R, Stewart J: A second generation human haplotype map of over 3.1 million SNPs. Nature 449: 851-861, 2007
- Voight BF, Kudaravalli S, Wen X, Pritchard JK: A map of recent positive selection in the human genome. PLoS Biol 4: e72, 2006
- Oleksyk TK, Nelson GW, An P, Kopp JB, Winkler CA: Worldwide distribution of the MYH9 kidney disease susceptibility alleles and haplotypes: evidence of historical selection in Africa. PLoS One 5: e11474, 2010
- Grossman SR, Shylakhter I, Karlsson EK, Byrne EH, Morales S, Frieden G, Hostetter E, Angelino E, Garber M, Zuk O, Lander ES, Schaffner SF, Sabeti PC: A composite of multiple signals distinguishes causal variants in regions of positive selection. Science 327: 883–886, 2010
- Genovese G, Tonna SJ, Knob AU, Appel GB, Katz A, Bernhardy AJ, Needham AW, Lazarus R, Pollak MR: A risk allele for focal segmental glomerulosclerosis in African Americans is located within a region containing APOL1 and MYH9. Kidney Int 78: 698–704, 2010
- Vanhamme L, Paturiaux-Hanocq F, Poelvoorde P, Nolan DP, Lins L, Van Den Abbeele J, Pays A, Tebabi P, Van Xong H, Jacquet A, Moguilevsky N, Dieu M, Kane JP, De Baetselier P, Brasseur R, Pays E: Apolipoprotein L-I is the trypanosome lytic factor of human serum. Nature 422: 83–87, 2003
- Tzur S, Rosset S, Shemer R, Yudkovsky G, Selig S, Tarekegn A, Bekele E, Bradman N, Wasser WG, Behar DM, Skorecki K: Missense muta-

- tions in the APOL1 gene are highly associated with end stage kidney disease risk previously attributed to the MYH9 gene. *Hum Genet* 128: 345–350, 2010
- Arendse CG, Wearne N, Okpechi IG, Swanepoel CR: The acute, the chronic and the news of HIV-related renal disease in Africa. Kidney Int 78: 239–245, 2010
- Ruf RG, Lichtenberger A, Karle SM, Haas JP, Anacleto FE, Schultheiss M, Zalewski I, Imm A, Ruf EM, Mucha B, Bagga A, Neuhaus T, Fuchshuber A, Bakkaloglu A, Hildebrandt F: Patients with mutations in NPHS2 (podocin) do not respond to standard steroid treatment of nephrotic syndrome. J Am Soc Nephrol 15: 722–732, 2004
- Nilsson S, Carstensen JM, Pershagen G: Mortality among male and female smokers in Sweden: A 33 year follow up. J Epidemiol Community Health 55: 825–830, 2001
- O'Brien SJ, Nelson GW: Human genes that limit AIDS. Nat Genet 36: 565–574, 2004
- 21. Kitiyakara C, Kopp JB, Eggers P: Trends in the epidemiology of focal segmental glomerulosclerosis. *Sem Neph* 23: 172–182, 2003
- Hindorff LA, Sethupathy P, Junkins HA, Ramos EM, Mehta JP, Collins FS, Manolio TA: Potential etiologic and functional implications of genome-wide association loci for human diseases and traits. *Proc Natl Acad Sci U S A* 106: 9362–9367, 2009
- Davidson WS, Silva RA, Chantepie S, Lagor WR, Chapman MJ, Kontush A: Proteomic analysis of defined HDL subpopulations reveals particle-specific protein clusters: relevance to antioxidative function.
   Arterioscler Thromb Vasc Biol 29: 870–876, 2009
- 24. The 1000 Genomes Project Consortium: A map of human genome

- variation from population-scale sequencing. *Nature* 467: 1061–1073, 2010
- McKenzie LM, Hendrickson SL, Briggs WA, Dart RA, Korbet SM, Mokrzycki MH, Kimmel PL, Ahuja TS, Berns JS, Simon EE, Smith MC, Trachtman H, Michel DM, Schelling JR, Cho M, Zhou YC, Binns-Roemer E, Kirk GD, Kopp JB, Winkler CA: NPHS2 variation in sporadic focal segmental glomerulosclerosis. J Am Soc Nephrol 18: 2987–2995, 2007
- Troyanov S, Wall CA, Miller JA, Scholey JW, Cattran DC: Focal and segmental glomerulosclerosis: Definition and relevance of a partial remission. J Am Soc Nephrol 16: 1061–1068, 2005
- Li JZ, Absher DM, Tang H, Southwick AM, Casto AM, Ramachandran S, Cann HM, Barsh GS, Feldman M, Cavalli-Sforza LL, Myers RM: Worldwide human relationships inferred from genome-wide patterns of variation. Science 319: 1100–1104, 2008
- Kopp JB, Winkler CA, Nelson GW: MYH9 genetic variants associated with glomerular disease: What is the role for genetic testing? Semin Nephrol 30: 409-417, 2010
- Nelson GW, O'Brien SJ: Using mutual information to measure the impact of multiple genetic factors on AIDS. J Acquir Immune Defic Syndr 42: 347–354, 2006

See related editorial, "Apolipoprotein L1 and the Genetic Basis for Racial Disparity in Chronic Kidney Disease," on pages 1955–1958.

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