

Published in final edited form as:

Nat Rev Neurol. 2013 February ; 9(2): 106–118. doi:10.1038/nrneurol.2012.263.

Apolipoprotein E and Alzheimer disease: risk, mechanisms, and therapy

Chia-Chen Liu¹, Takahisa Kanekiyo², Huaxi Xu¹, and Guojun Bu¹

¹Fujian Provincial Key Laboratory of Neurodegenerative Disease and Aging Research, Institute of Neuroscience, College of Medicine, Xiamen University, Xiamen, Fujian 361005, China

²Department of Neuroscience, Mayo Clinic, 4500 San Pablo Road, Jacksonville, Florida 32224, USA

Abstract

Apolipoprotein E (ApoE) is a major cholesterol carrier that supports lipid transport and injury repair in the brain. *APOE* polymorphic alleles are the main genetic determinants of Alzheimer disease (AD) risk: individuals carrying the $\epsilon 4$ allele are at increased risk of AD compared with those carrying the more common $\epsilon 3$ allele, whereas the $\epsilon 2$ allele decreases risk. Presence of the *APOE* $\epsilon 4$ allele is also associated with increased risk for cerebral amyloid angiopathy and age-related cognitive decline during normal ageing. ApoE–lipoproteins bind to several cell-surface receptors to deliver lipids and also to hydrophobic amyloid- β (A β) peptide, which is thought to initiate toxic events that lead to synaptic dysfunction and neurodegeneration in AD. ApoE isoforms differentially regulate A β aggregation and clearance in the brain, and have distinct functions in regulating brain lipid transport, glucose metabolism, neuronal signalling, neuroinflammation, and mitochondrial function. In this Review, we describe current knowledge on ApoE in the CNS, with a particular emphasis on the clinical and pathological features associated with carriers of different ApoE isoforms. We also discuss A β -dependent and A β -independent mechanisms that link ApoE4 status with AD risk, and consider how to design effective strategies for AD therapy by targeting ApoE.

Introduction

Alzheimer disease (AD) is a progressive neurodegenerative disease associated with cognitive decline and is the most common form of dementia in the elderly. Approximately 13% of people over the age of 65 and 45% over the age of 85 are estimated to have AD.¹ Mounting evidence from genetic, pathological, and functional studies has shown that an imbalance between the production and clearance of amyloid- β (A β) peptides in the brain results in accumulation and aggregation of A β . The toxic A β aggregates in the form of soluble A β oligomers, intraneuronal A β , and amyloid plaques injure synapses and ultimately cause neurodegeneration and dementia.^{2,3} The toxicity of A β seems to depend on the presence of microtubule-associated protein tau,⁴ the hyperphosphorylated forms of which aggregate and deposit in AD brains as neurofibrillary tangles. A β is composed of 40 or 42 amino acids and is generated through proteolytic cleavage of amyloid precursor protein (APP).⁵

Correspondence to: Guojun Bu (bu.guojun@mayo.edu).

Author contributions

All authors contributed to researching data for the article, discussion of the content, writing the article, and to review and/or editing of the manuscript before submission.

Early-onset familial AD, which typically develops before the age of 65 years and accounts for only a small portion (<1%) of AD cases,^{2, 3} is primarily caused by overproduction of A β owing to mutations in either the *APP* gene or genes encoding presenilin 1 (*PSEN1*) or presenilin 2 (*PSEN2*), essential components of the γ -secretase complexes responsible for cleavage and release of A β . The majority of AD cases occur late in life (>65 years) and are commonly referred to as late-onset AD (LOAD). Although multiple genetic and environmental risk factors are involved in LOAD pathogenesis, overall impairment in A β clearance is probably a major contributor to disease development.⁶ Genetically, the ϵ 4 allele of the apolipoprotein E (*APOE*) gene is the strongest risk factor for LOAD.^{7–9} The human *APOE* gene exists as three polymorphic alleles— ϵ 2, ϵ 3 and ϵ 4—which have a worldwide frequency of 8.4%, 77.9% and 13.7%, respectively.¹⁰ However, the frequency of the ϵ 4 allele is dramatically increased to ~40% in patients with AD.¹⁰

ApoE regulates lipid homeostasis by mediating lipid transport from one tissue or cell type to another.¹¹ In peripheral tissues, ApoE is primarily produced by the liver and macrophages, and mediates cholesterol metabolism in an isoform-dependent manner. ApoE4 is associated with hyperlipidaemia and hypercholesterolemia, which lead to atherosclerosis, coronary heart disease and stroke.^{11, 12} In the CNS, ApoE is mainly produced by astrocytes, and transports cholesterol to neurons via ApoE receptors, which are members of the low-density lipoprotein receptor (LDLR) family.⁸

ApoE is composed of 299 amino acids and has a molecular mass of ~34 kDa.¹¹ Differences between the three ApoE isoforms are limited to amino acids 112 and 158, where either cysteine or arginine is present (Figure 1a): ApoE2 (Cys112, Cys158), ApoE3 (Cys112, Arg158), and ApoE4 (Arg112, Arg158).¹¹ The single amino acid differences at these two positions affect the structure of ApoE isoforms and influence their ability to bind lipids, receptors and A β .^{13–15} Human and animal studies clearly indicate that ApoE isoforms differentially affect A β aggregation and clearance. Several A β -independent functions are also associated with ApoE isoforms. In this Review, we provide an overview of clinical evidence for the association between *APOE* genotypes and the risk of cognitive decline in AD, mild cognitive impairment (MCI) and other CNS diseases with a cognitive component, and discuss our current understanding of the mechanisms underlying ApoE actions and ApoE-targeted therapies.

***APOE* genotypes, AD and cognition**

***APOE* ϵ 4 as a strong risk factor for AD**

Genome-wide association studies have confirmed that the ϵ 4 allele of *APOE* is the strongest genetic risk factor for AD.^{16, 17} The presence of this allele is associated with increased risk for both early-onset AD and LOAD.^{18, 19} A meta-analysis of clinical and autopsy-based studies demonstrated that, compared with individuals with an ϵ 3/ ϵ 3 genotype, risk of AD was increased in individuals with one copy of the ϵ 4 allele (ϵ 2/ ϵ 4, OR 2.6; ϵ 3/ ϵ 4, OR 3.2) or two copies (ϵ 4/ ϵ 4, OR 14.9) among Caucasian subjects.¹⁰ The ϵ 2 allele of *APOE* has protective effects against AD: the risk of AD in individuals carrying *APOE* ϵ 2/ ϵ 2 (OR 0.6) or ϵ 2/ ϵ 3 (OR 0.6) are lower than those of ϵ 3/ ϵ 3.¹⁰ In population-based studies, the *APOE* ϵ 4-AD association was weaker among African Americans (ϵ 4/ ϵ 4, OR 5.7) and Hispanics (ϵ 4/ ϵ 4, OR 2.2) and was stronger in Japanese people (ϵ 4/ ϵ 4, OR 33.1) compared with Caucasian cases (ϵ 4/ ϵ 4, OR 12.5).¹⁰ *APOE* ϵ 4 is associated with increased prevalence of AD and lower age of onset.^{7, 10, 20} The frequency of AD and mean age at clinical onset are 91% and 68 years of age in ϵ 4 homozygotes, 47% and 76 years of age in ϵ 4 heterozygotes, and 20% and 84 years in ϵ 4 noncarriers,^{7, 20} indicating that *APOE* ϵ 4 confers dramatically increased risk of development of AD with an earlier age of onset in a gene dose-dependent manner (Figure 1b).

Genetic variants in the *TOMM40* (translocase of outer mitochondrial membrane 40 homologue) gene, which lies adjacent to the *APOE* gene on chromosome 19, have been implicated as a modulator of AD age-of-onset in *APOE* $\epsilon 3$ carriers.²¹ A more recent study, however, has cast doubt on the strength of this association.²² Whether the effects of *APOE* and *TOMM40* on AD risk, both genetically and functionally, are synergistic requires further investigation.

APOE and A β deposition

ApoE has an important role in A β metabolism (Figure 2). Studies show that *APOE* genotypes strongly affect deposition of A β to form senile plaques and cause cerebral amyloid angiopathy (CAA), two major hallmarks of amyloid pathology in AD brains.²³ Immunohistological evidence demonstrates that ApoE is co-deposited in senile plaques in the brains of AD patients.²⁴ The A β deposition in the form of senile plaques is more abundant in *APOE* $\epsilon 4$ carriers compared with noncarriers.^{25–27} The difference was most evident among individuals aged 50–59 years: 40.7% of *APOE* $\epsilon 4$ carriers had senile plaques compared with 8.2% of noncarriers.²⁵ In individuals with positive Pittsburgh compound B (PiB)-PET images, which indicate fibrillar aggregates of A β ,²⁸ *APOE* $\epsilon 4$ was more common than in those with negative scans (65% versus 22%) in patients with AD.²⁹

Fibrillar A β deposition is often detected in the brains of elderly, cognitively normal individuals in a manner that depends on the presence of *APOE* $\epsilon 4$, although such an association is weaker than that in patients with AD.³⁰ In addition, *APOE* $\epsilon 4$ carriers have lower cerebrospinal fluid (CSF) A β_{42} levels and higher PiB-positive imaging, which reflect the presence of cerebral amyloid deposition and serve as potential biomarkers for AD.^{31, 32} Cognitively normal *APOE* $\epsilon 4$ carriers exhibit PiB-positive imaging about 56 years of age, compared with about 76 years of age in noncarriers.³³ This difference suggests that *APOE* $\epsilon 4$ probably increases the risk of AD by initiating and accelerating A β accumulation, aggregation and deposition in the brain. Although *APOE* $\epsilon 2$ reduces the risk of dementia,³⁴ in individuals older than 90 years, both the $\epsilon 2$ and $\epsilon 4$ alleles of *APOE* increase amyloid burden compared with *APOE* $\epsilon 3$, suggesting that the protective effects of *APOE* $\epsilon 2$ against AD might not be associated with A β deposition.

APOE $\epsilon 4$ also shows an association with CAA and CAA-related haemorrhages.^{35, 36} CAA refers to the pathological condition in which amyloid spreads and deposits throughout the cerebral blood vessel walls³⁷ and is frequently detected in AD.²³ Interestingly, although *APOE* $\epsilon 2$ is protective against AD, it is a risk factor for CAA-related haemorrhage, independently of AD, possibly by predisposing vessels to vasculopathic complications of CAA.³⁶

Prediction of AD in MCI

MCI is a transitional stage between normal ageing and dementia, and is associated with increased risk of AD.³⁸ The rate at which patients with amnesic MCI (aMCI) progress to clinically diagnosable AD is 10–15% per year, in contrast to a rate of 1–2% per year among healthy elderly individuals.³⁹ The prevalence of *APOE* $\epsilon 4$ is substantially higher in both aMCI and dys-executive MCI than in control individuals.⁴⁰ Patients with MCI who harbour *APOE* $\epsilon 4$ exhibit distinct cognitive profiles, which seem to resemble those of patients in the early stages of AD.⁴¹ A case-control study reported poorer memory performance among patients with MCI who were carriers of *APOE* $\epsilon 4$ compared with noncarriers.⁴² *APOE* $\epsilon 4$ is associated with impaired memory performance and increased risk of memory decline in middle-aged (40–59 years) and elderly (60–85 years) people with MCI.^{43, 44} Furthermore, patients with MCI who are carriers of *APOE* $\epsilon 4$ experience more-rapid decline in several cognitive and functional assessments, and severity of the deficits is strongly associated with

the *APOE* $\epsilon 4$ gene dose.^{41, 45, 46} Importantly, the presence of *APOE* $\epsilon 4$ is associated with increased risk of progression from MCI to AD-type dementia.^{47–49} Among individuals with aMCI, *APOE* $\epsilon 4$ carriers tend to be younger than noncarriers, consistent with younger age of AD onset in individuals with *APOE* $\epsilon 4$.⁵⁰ These findings indicate that the *APOE* $\epsilon 4$ genotype in patients with MCI can serve as a predictive factor for determination of clinical outcome and the risk of conversion to AD.

In patients with MCI, the adverse effects of *APOE* $\epsilon 4$ on cognitive functions correlate with the severity of neuronal pathology. Those who are carriers of *APOE* $\epsilon 4$ have lower CSF A β_{42} levels, higher tau levels and greater brain atrophy than do noncarriers.⁵⁰ Furthermore, patients with MCI who are PiB-positive are more likely to be *APOE* $\epsilon 4$ carriers and exhibit worse memory performance than are PiB-negative patients.⁵¹ Other findings suggest, although not without controversy,⁵² that *APOE* $\epsilon 4$ has considerable deleterious effects on memory performance⁴² and might be used to predict disease progression in combination with AD biomarkers and neuroimaging approaches.⁵³

Predicting cognitive decline in healthy cases

Healthy *APOE* $\epsilon 4$ carriers not diagnosed with MCI or AD show an accelerated longitudinal decline in memory tests, which starts around the age of 55–60 years, revealing a possible pre-MCI state in this genetic subset of individuals.^{54, 55} This memory decline, despite ongoing normal clinical status, suggests that pathological changes in AD might manifest in the brain as early as the sixth decade of life.^{56, 57} Thus, *APOE* $\epsilon 4$ is associated with cognitive decline many years before cognitive impairment becomes clinically apparent.^{56, 58} Interestingly, *APOE* $\epsilon 4$ has differential effects on memory performance depending on age. Some studies in young adults and children have found evidence of better cognitive performance in *APOE* $\epsilon 4$ carriers than in noncarriers, which could suggest antagonistic pleiotropy,^{59–61} in which *APOE* $\epsilon 4$ might offer benefits during development and early adulthood at the expense of more-rapid decline in cognitive function with ageing.⁶²

Similar to the situation in patients with MCI, *APOE* $\epsilon 4$ is associated with enhanced amyloid pathology in cognitively normal people. The proportion of PiB-positive individuals follows a strong *APOE* allele-dependent pattern ($\epsilon 4 > \epsilon 3 > \epsilon 2$),^{25, 63, 64} and *APOE* $\epsilon 4$ increases the amount of amyloid deposition in a gene-dose-dependent manner.³⁰

APOE $\epsilon 4$ and other AD risk factors

The *APOE* $\epsilon 4$ genotype combines synergistically with atherosclerosis, peripheral vascular disease, or type 2 diabetes in contributing to an increased risk of AD.^{65, 66} *APOE* $\epsilon 4$ is a risk factor for cardiovascular disease, suggesting that this allele and cerebrovascular disease might have compounding effects on cognitive decline in AD.⁶⁷ Diabetes also increases the risk of AD, and the association is particularly strong among *APOE* $\epsilon 4$ carriers.^{66, 68, 69} Patients with diabetes who are carriers of *APOE* $\epsilon 4$ have more neuritic plaques, neurofibrillary tangles and CAA than do noncarriers.⁶⁶ The combination of a diabetes-related factor—that is, hyperglycaemia, hyperinsulinaemia, and insulin resistance—and the *APOE* $\epsilon 4$ allele promotes neuritic plaque formation.⁶⁹ *APOE* $\epsilon 4$ seems to modify the risk of AD in patients with diabetes—a disease that directly or indirectly causes vascular and neuronal damage and further exacerbates AD pathology. Furthermore, recent research demonstrated that, independently of A β , ApoE4 triggers inflammatory cascades that cause neurovascular dysfunction, including blood–brain barrier breakdown, leakage of blood-derived toxic proteins into the brain and reduction in the length of small vessels.⁷⁰ This result suggests that ApoE4-associated damage to vascular systems in brain could have a key role in AD pathogenesis.

APOE and traumatic brain injury

Increasing evidence has shown that *APOE* $\epsilon 4$ is associated with poorer outcomes following traumatic brain injury (TBI), regardless of the severity of initial injury.⁷¹ A meta-analysis demonstrated that the outcome of TBI at 6 months after injury is worse in *APOE* $\epsilon 4$ carriers.⁷² TBI is associated with increased risk of AD,⁷³ and such a risk is more evident in patients with *APOE* $\epsilon 4$.⁷⁴ Only 10% of *APOE* $\epsilon 4$ noncarriers with TBI have A β plaque pathology, whereas 35% and 100% of TBI patients with one or two *APOE* $\epsilon 4$ alleles, respectively, possess A β pathology.⁷⁵ The poorer outcomes associated with ApoE4 might relate to its reduced ability to repair and remodel synapses and protect neurons upon injury compared with ApoE3.⁸ These possibilities are currently under investigation.

APOE and vascular diseases

Vascular cognitive impairment, which comprises clinical conditions with cerebrovasculature-derived cognitive disturbances including vascular dementia, is observed in approximately 8–15% of aged individuals with cognitive dysfunction in Western clinic-based series.⁷⁶ A recent meta-analysis has shown evidence of increased risk of vascular dementia in individuals with *APOE* $\epsilon 4$ compared with *APOE* $\epsilon 3$ (OR 1.72).⁷⁷ Several studies suggest that the contribution of *APOE* $\epsilon 4$ to risk of vascular cognitive impairment is independent of other vascular risk factors including hypertension, dyslipidaemia and atherogenesis,⁷⁸ whereas another report shows that age-related cognitive decline among *APOE* $\epsilon 4$ carriers is induced by brain damage owing to increased blood pressure.⁷⁹ In addition, *APOE* $\epsilon 4$ is associated with poor outcome after subarachnoid haemorrhage,⁸⁰ and is a strong risk factor for CAA-related intracranial haemorrhage.⁸¹ These results suggest that *APOE* $\epsilon 4$ is closely associated with neurovascular dysfunctions.

APOE and other types of dementia

Lewy body disease is thought to be the second most common kind of dementia comprised of a spectrum of diseases that includes Parkinson disease (PD), PD-associated dementia and dementia with Lewy bodies (DLB). Clinical and pathological features of PD and AD frequently overlap. Most studies, however, have failed to report associations between *APOE* $\epsilon 4$ and susceptibility to PD and PD-associated dementia.^{82, 83} DLB also shares clinical and pathological characteristics with AD and PD,⁸⁴ and several reports have shown that *APOE* $\epsilon 4$ increases risk of DLB.⁸⁵ Immunohistochemical analysis showed that deposition of Lewy bodies in patients with DLB who are *APOE* $\epsilon 4$ carriers is substantially more abundant than those who are noncarriers.⁸⁶ As Lewy bodies are considerably increased in the cerebral cortex of DLB patients with A β deposition,⁸⁷ the strong association between amyloid pathology and the pathology of Lewy body disease could explain why *APOE* $\epsilon 4$ increases risk of DLB. *APOE* $\epsilon 4$ might also be a risk factor for frontotemporal dementia,⁸⁸ although the pathophysiological role of ApoE in this disease requires further investigation. The *APOE* genotypes do not seem to influence the risks of Huntington disease⁸⁹ nor amyotrophic lateral sclerosis.⁹⁰

Mechanisms of ApoE isoforms in AD

APOE $\epsilon 4$ confers a gain of toxic functions, a loss of neuroprotective functions or both in the pathogenesis of AD (Figure 3).

A β metabolism and aggregation

Studies in humans and transgenic mice showed that brain A β levels and amyloid plaque loads are ApoE isoform-dependent ($\epsilon 4 > \epsilon 3 > \epsilon 2$),^{30, 63, 91} suggesting an important role of ApoE in modulating A β metabolism, aggregation, and deposition. ApoE4 is less efficient in A β clearance than is ApoE3 in young and old amyloid mouse models that express human

ApoE isoforms.⁶³ Additionally, ApoE isoforms differentially regulate cholesterol levels, which have been shown to modulate γ -secretase activity and A β production.⁹² Several studies reported an *APOE* genotype-dependent effect on CSF and brain ApoE levels ($\epsilon 4 < \epsilon 3 < \epsilon 2$) in ApoE-targeted replacement (ApoE-TR) mice, in which the mouse *ApoE* gene is replaced with human *APOE* isoforms.^{91, 93, 94} This result suggests that lower levels of total ApoE exhibited by *APOE* $\epsilon 4$ carriers might contribute to disease progression. However, whether human ApoE isoform status affects CSF and brain ApoE protein levels in healthy individuals and patients with AD remains to be established.^{95, 96}

ApoE-knockout mice clear A β from the brain faster than do control mice.⁹⁷ Stimulation of liver X receptors (LXRs)^{98, 99} or the retinoid X receptor (RXR)¹⁰⁰ facilitates A β clearance, probably by increasing ApoE levels and lipidation. Further investigation is needed to determine whether ApoE levels are directly associated with A β clearance. In addition, a recent study showed that lack of one copy of ATP-binding cassette transporter A1 (ABCA1), which shuttles lipids to ApoE, impairs A β clearance and exacerbates amyloid deposition and memory deficits in ApoE4-TR mice, but not in ApoE3-TR mice.¹⁰¹ This result suggests that ApoE isoforms exhibit differential lipidation status, which affects A β clearance in an isoform-dependent manner. Alternatively, ApoE-lipoprotein particles may sequester A β and promote cellular uptake and degradation of ApoE-A β complexes.¹⁰²

ApoE4-lipoproteins bind A β with lower affinity than do ApoE3-lipoproteins,¹⁰³ suggesting that ApoE4 might be less efficient in mediating A β clearance. In addition, ApoE might modulate A β removal from the brain to the systemic circulation by transporting A β across the blood-brain barrier. In this respect, ApoE impedes A β clearance at the blood-brain barrier in an isoform-specific fashion (ApoE4 > ApoE3 and ApoE2).¹⁰⁴ Finally, studies in microglia have shown that ApoE3 promotes enzyme-mediated degradation of A β more efficiently than does ApoE4.¹⁰⁵ Together, these studies suggest that ApoE4 inhibits A β clearance and/or is less efficient in mediating A β clearance compared with ApoE3 and ApoE2.

ApoE also seems to regulate A β aggregation and deposition. An important study showed that deletion of the mouse *ApoE* gene essentially eliminates deposition of fibrillar A β in amyloid model mice.¹⁰⁶ Given that ApoE is co-deposited with A β in human AD brains,²⁴ it is possible that ApoE promotes A β aggregation and deposition in an isoform-dependent manner. The exact mechanisms by which ApoE isoforms differentially regulate A β aggregation and deposition require further investigation.

Brain activity and atrophy

AD is associated with both functional abnormalities of the hippocampus and cortical atrophy in the memory network.^{107, 108} Patients with AD or MCI who are *APOE* $\epsilon 4$ carriers exhibit greater medial temporal lobe atrophy, particularly in the hippocampal area.^{41, 109, 110} Structural MRI studies found that, compared with noncarriers, *APOE* $\epsilon 4$ carriers have accelerated age-related loss in cortical thickness and hippocampal volume that are tightly coupled to decline in cognitive performance.^{111–113}

Functional MRI (fMRI) studies reported that ApoE4 disrupts resting state fMRI connectivity and the balance between brain networks, in the absence of amyloid pathology.^{114, 115} Furthermore, cognitively normal *APOE* $\epsilon 4$ carriers have elevated resting-state activity in the default mode network—a network that is preferentially affected early in AD—and higher hippocampal activation during memory tasks.^{116–118} Such changes have been hypothesized to represent a compensatory response by *APOE* $\epsilon 4$ carriers in which increased cognitive effort is required to achieve an equivalent level of performance to that of noncarriers.^{116, 118}

Elevated baseline activity in brain networks of *APOE* $\epsilon 4$ carriers could potentially contribute to increased $A\beta$ production, as $A\beta$ levels are regulated by neuronal activity.^{119, 120} Interestingly, in adults who do not have dementia, increased hippocampal activity was associated with reduced cortical thickness in the medial temporal lobe and brain regions that are vulnerable to AD pathology.¹²¹ Studies suggested that hippocampal hyperactivity might represent impending synaptic dysfunction and incipient cognitive decline.¹²² Interestingly, another study showed a reduction of posterior default mode network connectivity in *APOE* $\epsilon 4$ carriers in cognitively normal elderly people, implying that *APOE* $\epsilon 4$ carriers may exhibit a more rapid decline in connectivity of this network than do noncarriers as they age.¹¹⁵

¹⁸F-fluorodeoxyglucose PET imaging, which measures cerebral metabolic rates of glucose as a proxy for neuronal activity, correlates with disease progression and predicts histopathological diagnosis in AD.¹²³ Mounting evidence suggests that *APOE* $\epsilon 4$ carriers exhibit lower cerebral glucose metabolism.^{124–126} Healthy adults with *APOE* $\epsilon 4$ show altered patterns of brain metabolism both at rest and during cognitive challenges compared with noncarriers.^{126, 127} Representative studies illustrating the association of ApoE4 isoform with altered brain metabolism and activity, memory decline, and amyloid pathology in cognitively normal people are shown in Figure 4. Improved understanding of the mechanisms of ApoE4-related brain activity changes, brain atrophy and reduced metabolism should help to explain why ApoE4 is a risk factor for cognitive decline and AD.

Tau phosphorylation and neurotoxicity

ApoE is produced primarily by astrocytes and microglia. Neuronal ApoE expression can, however, be induced in response to stress or injury, probably for the purpose of neuronal repair and remodelling.^{128, 129} A truncated fragment of ApoE4, resulting from proteolytic cleavage of ApoE following stress or injury, increases tau hyperphosphorylation, cytoskeletal disruption and mitochondrial dysfunction.^{128, 130, 131} ApoE4 also exacerbates neurotoxicity triggered by $A\beta$ and other insults.^{128, 131}

A recent study showed that neurons in patients with temporal lobe epilepsy who harbour *APOE* $\epsilon 4$ are less resilient to the damaging hyperexcitability and more susceptible to $A\beta$ toxicity than are those in *APOE* $\epsilon 4$ carriers,¹³² suggesting that ApoE3 might confer a neuroprotective advantage over ApoE4 against neuronal stress. Interestingly, astrocyte-derived ApoE4 has neuroprotective effects against excitotoxic injuries, whereas neuronal expression of ApoE4 promotes excitotoxic cell death. This result suggests that ApoE derived from various cellular sources might exhibit different physiological and pathological activity.¹³³

Lipid metabolism

Abnormal lipid metabolism is strongly related to the pathogenesis of AD. In the CNS, ApoE mediates neuronal delivery of cholesterol, which is an essential component for axonal growth, synaptic formation and remodelling—events that are crucial for learning, memory formation and neuronal repair.^{134, 135} Brain cholesterol levels are substantially reduced in hippocampal and cortical areas in patients with AD compared with age-matched controls.¹³⁶ Preferential degradation of ApoE4 relative to ApoE3 in astrocytes has been proposed to result in low levels of ApoE in the brain and CSF and reduced capacity for neuronal delivery of cholesterol, suggesting that low levels of total ApoE exhibited by *APOE* $\epsilon 4$ carriers may directly contribute to the disease progression.⁹³ ApoE4 is also less efficient than ApoE3 in transporting brain cholesterol.¹³⁷ Moreover, ApoE4-TR mice have abnormal cholesterol levels and impaired lipid metabolism.¹³⁸ Insufficient levels of ApoE and/or impaired ApoE

function in carriers of the $\epsilon 4$ allele might, therefore, lead to aberrant CNS cholesterol homeostasis and neuronal health, which contribute to AD risk.

Synaptic plasticity and spine integrity

Synaptic failure is an early pathological feature of AD.^{139, 140} Increasing evidence demonstrates that ApoE isoforms differentially regulate synaptic plasticity and repair.^{141, 142} In AD and healthy aged controls, *APOE* $\epsilon 4$ gene dosage correlates inversely with dendritic spine density in the hippocampus.¹⁴³ ApoE4-TR mice also have lower dendritic spine density and length compared with ApoE3-TR mice.^{144, 145} ApoE3, but not ApoE4, prevents loss of synaptic networks induced by A β oligomers.¹⁴⁶ ApoE isoforms also differentially regulate dendritic spines during ageing.^{143, 147} The age-dependence of these differences implies that the effects of ApoE isoforms on neuronal integrity might relate to increased risk of dementia in aged *APOE* $\epsilon 4$ carriers.

Reduced synaptic transmission was observed in 1-month-old ApoE4-TR mice compared with ApoE3-TR mice, suggesting that ApoE4 may also contribute to functional deficits early in development, which could account for alteration of neuronal circuitry that eventually results in cognitive disorders later in life.¹⁴⁷ In addition, ApoE4 selectively impairs ApoE receptor trafficking and signalling, as well as glutamate receptor function and synaptic plasticity.¹⁴¹ Together, these findings suggest that the effect of *APOE* $\epsilon 4$ genotype on risk of AD might be mediated, at least in part, through direct effects on synaptic function.

Neuroinflammation

Neuroinflammation contributes to neuronal damage in the brain and is implicated in AD pathogenesis.¹⁴⁸ ApoE colocalizes with plaque-associated amyloid and microglia, suggesting a role for ApoE in the innate immune response in AD. Lack of ApoE in mice is associated with increased inflammation in response to A β ,^{149, 150} but ApoE isoforms might differently regulate the innate immune response.¹⁵¹ ApoE4 seems to have proinflammatory and/or reduced anti-inflammatory functions, which could further exacerbate AD pathology. For example, ApoE4-TR mice exhibit greater inflammatory responses to lipopolysaccharide compared with ApoE3-TR mice.¹⁵² In addition, young *APOE* $\epsilon 4$ carriers show an increased inflammatory response that may relate to AD risk later in life.¹⁵³ Consistent with this notion, non-steroidal anti-inflammatory drugs were shown to reduce AD risk only in *APOE* $\epsilon 4$ carriers,¹⁵⁴ suggesting that *APOE* genotype might determine the effect of anti-inflammatory medications for AD.

Neurogenesis

Hippocampal neurogenesis has an important role in structural plasticity and maintenance of brain networks. Dysfunctional neurogenesis resulting from early disease manifestations could, therefore, exacerbate neuronal vulnerability to AD and contribute to memory impairment.¹⁵⁵ ApoE is required for maintenance of the neural stem or progenitor cell pool in the adult dentate gyrus region of the hippocampus.¹⁵⁶ In ApoE-TR mice, ApoE4 inhibits hippocampal neurogenesis by impairing maturation of hilar γ -aminobutyric acid-containing interneurons, which contributes to learning and memory deficits.^{157, 158} These results demonstrate an important pathological role of ApoE4 in impairment of neurogenesis, which might contribute to AD pathogenesis.

ApoE as a therapeutic target for AD

Most therapeutic approaches for AD target the A β pathway. With the recent failure of clinical trials of drugs targeting solely A β , an urgent need exists to define new targets and develop alternative therapeutic strategies to treat AD. As *APOE* genotype determines AD

risk, and ApoE has crucial roles in cognition, ApoE might offer an attractive alternative target for AD therapy. *APOE* genotype status could be included in clinical trial enrolment criteria, as some therapies might be effective only in specific *APOE* genotypes. Here, we briefly discuss several approaches that are currently being explored (Table 1).

APOE genotype and A β immunotherapy

Recent phase III clinical trials for immunotherapy have shown that bapineuzumab, an antibody that targets the N-terminus of A β , prevents A β deposition in the brains of *APOE* ϵ 4 carriers with mild or moderate AD, but not noncarriers.^{159, 160} Bapineuzumab also lowers levels of phosphorylated tau in the CSF of both *APOE* ϵ 4 carriers and noncarriers.^{159, 160} These reports suggest that A β immunotherapy is useful to eliminate A β from the brains of patients with AD and that its effect is likely to depend on ApoE isoforms. Major adverse effects of bapineuzumab—vasogenic cerebral oedema and microhaemorrhage—occur more frequently in *APOE* ϵ 4 carriers than in noncarriers.^{159, 160} Although bapineuzumab failed to prevent cognitive and functional decline in these clinical trials, a combination of A β immunotherapy and an ApoE-targeted approach might lead to more effective therapeutic strategies.

Preventive of cognitive decline in APOE ϵ 4 carriers

A prospective study of a cognitively normal cohort showed that risk of dementia in *APOE* ϵ 4 carriers is negatively associated with high education, high level of leisure activities, and absence of vascular risk factors.¹⁶¹ A recent study demonstrated that physical exercise was strongly associated with reduced PiB-positivity in cognitively normal *APOE* ϵ 4 carriers,³¹ indicating that a sedentary lifestyle in *APOE* ϵ 4 carriers might increase the risk of amyloid deposition. Such studies indicate that high education, active leisure activities and exercise, and maintenance of vascular health could be beneficial in reducing the risk of AD and cognitive decline, particularly in *APOE* ϵ 4 carriers.

Regulation of ApoE expression

ApoE levels in CSF and plasma tend to be lower in patients with AD than in healthy individuals, although such findings remain controversial.^{162, 163} Thus, increasing the expression of ApoE in all *APOE* genotypes may prevent or slow progression of AD through acceleration of A β metabolism and promotion of ApoE functions in lipid metabolism and synaptic support. Compounds that increase brain ApoE expression can be identified through comprehensive drug screening. Given that expression of ApoE is controlled by peroxisome proliferator-activated receptor- γ and LXRs, which form complexes with RXRs,^{100, 164} agonists or antagonists of these nuclear receptors are potential candidates as ApoE modulators. Indeed, recent work has demonstrated that oral administration of an RXR agonist, bexarotene, to an amyloid mouse model decreases A β plaque deposition and improves cognitive function in an ApoE-dependent manner.¹⁰⁰ The LXR agonist TO901317 also increases ApoE levels in the brain, facilitates clearance of A β ₄₂, and reverses contextual memory deficit in amyloid mouse models.^{98, 99}

In addition to ApoE, LXRs also regulate ABCA1, which promotes cholesterol efflux.¹⁶⁵ Consequently, reduction of amyloid burden by the LXR agonist GW3965 depends on expression of ABCA1 in amyloid mouse models.¹⁶⁶ These results suggest that upregulation of lipidated ApoE might be necessary to maximize therapeutic effects in AD. These studies did not, however, assess the effect of increasing human ApoE3 or ApoE4 specifically. Because A β deposition is greater in *APP*-transgenic mice expressing mouse ApoE than in those expressing human ApoE isoforms,¹⁶⁷ further studies are needed to confirm the therapeutic effect of modulating the level of human ApoE isoforms. In addition, whether increasing ApoE4 is beneficial or harmful in AD brains remains unclear, and the effects

might depend on age and disease status. Toxic functions associated with ApoE4 suggest that lowering ApoE4 expression might be beneficial in *APOE* ϵ 4 carriers with cognitive decline during MCI and AD. Additional preclinical studies are needed to test potential beneficial or harmful effects of increasing or decreasing ApoE expression, particularly with regard to ApoE isoforms.

Blocking of ApoE-A β interaction

ApoE is required for deposition of A β fibrils in amyloid mouse models.¹⁰⁶ Recent studies have demonstrated that haploinsufficiency of human *APOE* results in significantly decreased amyloid plaque deposition in amyloid mouse models regardless of *APOE* isoform status.^{168, 169} Thus, disruption of the interaction between ApoE and A β might reduce A β aggregation and deposition, and should be considered as a therapeutic approach. A β interacts with ApoE through amino acid residues 12–28. A synthetic peptide mimicking this sequence, A β 12–28P, reduces A β deposition and ameliorates memory deficits in amyloid mouse models.¹⁷⁰ Blocking the ApoE–A β interaction using A β -mimicking peptides could, therefore, be an effective approach for treatment of AD. Screening assays can also be used to identify compounds or ApoE-specific antibodies that block ApoE–A β interaction. These approaches should be assessed carefully because they could disrupt ApoE–lipid interactions and the associated beneficial functions of ApoE.

Other ApoE-based therapeutic approaches

ApoE4 is structurally different from ApoE2 and ApoE3 owing to different domain interactions,¹³¹ and this difference probably contributes to ApoE4 isoform-specific harmful effects. Modification of the structure of ApoE4 to form an ApoE3-like molecule might, therefore, be an interesting approach to ameliorate these harmful effects. Indeed, several molecules that bind to ApoE4 and interfere with domain interactions between N- and C-termini have been found. GIND–25 (disulphonate) and GIND–105 (monosulphoalkyl) are good candidates because they decrease A β production induced by ApoE4 to a similar level induced by ApoE3.¹³¹ CB9032258 (a phthalazinone analogue) and its derivatives disrupt ApoE4 domain interaction and restore functional activities of ApoE4 in neurons.¹⁷¹

An ApoE-mimetic peptide containing the receptor-binding region suppresses neuronal cell death and calcium influx associated with N-methyl-D-aspartate exposure *in vitro*.¹⁷² COG112, a chimeric peptide containing the receptor-binding region, is also reported to improve symptoms in mouse models of multiple sclerosis¹⁷³ and sciatic nerve crush¹⁷⁴ through modulation of inflammatory responses. The effects of these peptides on AD pathogenesis are unknown, however, because they do not contain A β -interacting nor lipid-binding regions.¹³

ApoE receptors are also potential targets for AD therapy. For example, low-density lipoprotein receptor-related protein 1 and low-density lipoprotein receptor have crucial roles in brain lipid metabolism and A β clearance.^{175–177} ApoE receptor 2 and very low-density lipoprotein receptor are essential for reelin signalling, which is important for neuronal migration during development and synaptic plasticity in adult brains.¹⁷⁸ Modulation of the expression of these ApoE receptors in AD brains might, therefore, restore lipid homeostasis and synaptic plasticity, and augment A β clearance.^{8, 178} Although ApoE-based therapies are still in the early stages of development, they offer great promises in the fight against AD. Clinical trials to further evaluate therapeutic potential of ApoE-based strategies are needed, with an eventual goal to develop curative and/or protective treatments for AD.

Conclusions

Work summarized in this Review highlights clinical evidence for the association between *APOE* ϵ 4, AD and cognitive decline. Although the presence of *APOE* ϵ 4 does not necessarily entail disease development, this genetic isoform probably accelerates the rate of disease conversion and progression. In particular, the effects of *APOE* ϵ 4 on brain network connectivity, memory performance, and rate of cognitive decline are age-dependent in patients with AD and cognitively normal individuals. Thus, understanding the potential pathogenic link between *APOE* ϵ 4 and cognitive function might allow for earlier identification of people at risk of developing AD. In combination with other putative AD biomarkers—such as MRI scans, PiB scans, and measurements of CSF A β and tau—*APOE* allele status could add predictive value to clinical diagnosis and evaluation of treatment efficacy.

Mechanistically, ApoE4 seems to increase risk of AD and cognitive decline through both A β -dependent and A β -independent pathways. ApoE isoforms differentially regulate A β production, aggregation and clearance. Independent of A β , ApoE4 might be less efficient than ApoE3 and ApoE2 in delivering cholesterol and essential lipids for the maintenance of synaptic integrity and plasticity. In addition, ApoE is a crucial regulator of the innate immune system, and in which ApoE4 promotes proinflammatory responses that could exacerbate AD pathogenesis. Finally, ApoE isoforms have differential roles in maintaining vascular health—roles that are crucial given that defects in vascular health are strongly associated with AD. Elucidating the contribution of ApoE4 to AD pathogenesis is a considerable challenge, but one that affords the potential to assist in combating AD.

Acknowledgments

Works in authors' laboratories are supported by the NIH, the Alzheimer's Association, the American Health Assistance Foundation, and Xiamen University Research Funds. We thank Caroline Stetler and Owen Ross for critical reading of the manuscript before submission.

References

1. Alzheimer's Association. 2012 Alzheimer's disease facts and figures. *Alzheimer's dement.* 2012; 8:131–168. [PubMed: 22404854]
2. Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science.* 2002; 297:353–6. [PubMed: 12130773]
3. Blennow K, de Leon MJ, Zetterberg H. Alzheimer's disease. *Lancet.* 2006; 368:387–403. [PubMed: 16876668]
4. Roberson ED, et al. Reducing endogenous tau ameliorates amyloid β -induced deficits in an Alzheimer's disease mouse model. *Science.* 2007; 316:750–754. [PubMed: 17478722]
5. Zheng H, Koo E. Biology and pathophysiology of the amyloid precursor protein. *Mol Neurodegener.* 2011; 6:27. [PubMed: 21527012]
6. Mawuenyega KG, et al. Decreased clearance of CNS beta-amyloid in Alzheimer's disease. *Science.* 2010; 330:1774. [PubMed: 21148344]
7. Corder EH, et al. Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science.* 1993; 261:921–3. [PubMed: 8346443]
8. Bu G. Apolipoprotein E and its receptors in Alzheimer's disease: pathways, pathogenesis and therapy. *Nat Rev Neurosci.* 2009; 10:333–44. [PubMed: 19339974]
9. Huang Y, Mucke L. Alzheimer mechanisms and therapeutic strategies. *Cell.* 2012; 148:1204–22. [PubMed: 22424230]
10. Farrer, LaCLHJL., et al. Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease: A meta-analysis. *JAMA.* 1997; 278:1349–1356. [PubMed: 9343467]

11. Mahley RW, Rall SC Jr. Apolipoprotein E: far more than a lipid transport protein. *Annu Rev Genomics Hum Genet.* 2000; 1:507–37. [PubMed: 11701639]
12. Lahoz C, et al. Apolipoprotein E genotype and cardiovascular disease in the Framingham Heart Study. *Atherosclerosis.* 2001; 154:529–537. [PubMed: 11257253]
13. Frieden C, Garai K. Structural differences between apoE3 and apoE4 may be useful in developing therapeutic agents for Alzheimer's disease. *Proc Natl Acad Sci USA.* 2012; 109:8913–8. [PubMed: 22615372]
14. Chen J, Li Q, Wang J. Topology of human apolipoprotein E3 uniquely regulates its diverse biological functions. *Proc Natl Acad Sci USA.* 2011; 108:14813–8. [PubMed: 21873229]
15. Zhong N, Weisgraber KH. Understanding the association of apolipoprotein E4 with Alzheimer disease: clues from its structure. *J Biol Chem.* 2009; 284:6027–31. [PubMed: 18948255]
16. Harold D, et al. Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. *Nat Genet.* 2009; 41:1088–93. [PubMed: 19734902]
17. Lambert JC, et al. Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. *Nat Genet.* 2009; 41:1094–9. [PubMed: 19734903]
18. Chartier-Harlin MC, et al. Apolipoprotein E, epsilon 4 allele as a major risk factor for sporadic early and late-onset forms of Alzheimer's disease: analysis of the 19q13.2 chromosomal region. *Hum Mol Genet.* 1994; 3:569–74. [PubMed: 8069300]
19. Houlden H, et al. ApoE genotype is a risk factor in nonpresenilin early-onset Alzheimer's disease families. *Am J Med Genet.* 1998; 81:117–21. [PubMed: 9514597]
20. William Rebeck G, Reiter JS, Strickland DK, Hyman BT. Apolipoprotein E in sporadic Alzheimer's disease: Allelic variation and receptor interactions. *Neuron.* 1993; 11:575–580. [PubMed: 8398148]
21. Roses AD, et al. A TOMM40 variable-length polymorphism predicts the age of late-onset Alzheimer's disease. *Pharmacogenomics J.* 2010; 10:375–84. [PubMed: 20029386]
22. Cruchaga C, et al. Association and expression analyses with single-nucleotide polymorphisms in TOMM40 in Alzheimer disease. *Arch Neurol.* 2011; 68:1013–9. [PubMed: 21825236]
23. Ellis RJ, et al. Cerebral amyloid angiopathy in the brains of patients with Alzheimer's disease: the CERAD experience, Part XV. *Neurology.* 1996; 46:1592–6. [PubMed: 8649554]
24. Namba Y, Tomonaga M, Kawasaki H, Otomo E, Ikeda K. Apolipoprotein E immunoreactivity in cerebral amyloid deposits and neurofibrillary tangles in Alzheimer's disease and kuru plaque amyloid in Creutzfeldt-Jakob disease. *Brain Res.* 1991; 541:163–6. [PubMed: 2029618]
25. Kok E, et al. Apolipoprotein E-dependent accumulation of Alzheimer disease-related lesions begins in middle age. *Ann Neurol.* 2009; 65:650–657. [PubMed: 19557866]
26. Polvikoski T, et al. Apolipoprotein E, dementia, and cortical deposition of beta-amyloid protein. *N Engl J Med.* 1995; 333:1242–7. [PubMed: 7566000]
27. Schmechel DE, et al. Increased amyloid beta-peptide deposition in cerebral cortex as a consequence of apolipoprotein E genotype in late-onset Alzheimer disease. *Proc Natl Acad Sci USA.* 1993; 90:9649–53. [PubMed: 8415756]
28. Klunk WE, et al. Imaging brain amyloid in Alzheimer's disease with Pittsburgh Compound-B. *Ann Neurol.* 2004; 55:306–19. [PubMed: 14991808]
29. Barthel H, et al. Cerebral amyloid-beta PET with florbetaben (18F) in patients with Alzheimer's disease and healthy controls: a multicentre phase 2 diagnostic study. *Lancet Neurol.* 2011; 10:424–35. [PubMed: 21481640]
30. Reiman EM, et al. Fibrillar amyloid- β burden in cognitively normal people at 3 levels of genetic risk for Alzheimer's disease. *Proc Natl Acad Sci USA.* 2009; 106:6820–6825. [PubMed: 19346482]
31. Head D, et al. Exercise engagement as a moderator of the effects of APOE genotype on amyloid deposition. *Arch Neurol.* 2012
32. Prince JA, Zetterberg H, Andreasen N, Marcusson J, Blennow K. APOE ϵ 4 allele is associated with reduced cerebrospinal fluid levels of A β 42. *Neurology.* 2004; 62:2116–2118. [PubMed: 15184629]

33. Fleisher AS, et al. Apolipoprotein E ϵ 4 and age effects on florbetapir positron emission tomography in healthy aging and Alzheimer disease. *Neurobiol Aging*. 2012
34. Berlau DJ, Corrada MM, Head E, Kawas CH. APOE epsilon2 is associated with intact cognition but increased Alzheimer pathology in the oldest old. *Neurology*. 2009; 72:829–34. [PubMed: 19255410]
35. Greenberg SM, et al. Apolipoprotein E epsilon 4 is associated with the presence and earlier onset of hemorrhage in cerebral amyloid angiopathy. *Stroke*. 1996; 27:1333–7. [PubMed: 8711797]
36. Biffi A, et al. Variants at APOE influence risk of deep and lobar intracerebral hemorrhage. *Ann Neurol*. 2010; 68:934–43. [PubMed: 21061402]
37. Vinters HV. Cerebral amyloid angiopathy. A critical review. *Stroke*. 1987; 18:311–24. [PubMed: 3551211]
38. Morris JC, et al. Mild cognitive impairment represents early-stage Alzheimer disease. *Arch Neurol*. 2001; 58:397–405. [PubMed: 11255443]
39. Petersen RC, et al. Mild cognitive impairment clinical characterization and outcome. *Arch Neurol*. 1999; 56:303–308. [PubMed: 10190820]
40. Pa J, et al. Clinical-neuroimaging characteristics of dysexecutive mild cognitive impairment. *Ann Neurol*. 2009; 65:414–23. [PubMed: 19399879]
41. Farlow MR, et al. Impact of APOE in mild cognitive impairment. *Neurology*. 2004; 63:1898–1901. [PubMed: 15557508]
42. Smith GE, et al. Apolipoprotein E genotype influences cognitive ‘phenotype’ in patients with Alzheimer’s disease but not in healthy control subjects. *Neurology*. 1998; 50:355–362. [PubMed: 9484353]
43. Ramakers IHGB, et al. The association between APOE genotype and memory dysfunction in subjects with mild cognitive impairment is related to age and Alzheimer pathology. *Dement Geriatr Cogn Disord*. 2008; 26:101–108. [PubMed: 18617739]
44. Dik MG, et al. APOE- ϵ 4 is associated with memory decline in cognitively impaired elderly. *Neurology*. 2000; 54:1492–1497. [PubMed: 10751265]
45. Whitehair DC, et al. Influence of apolipoprotein E ϵ 4 on rates of cognitive and functional decline in mild cognitive impairment. *Alzheimer’s Dement*. 2010; 6:412–419. [PubMed: 20813342]
46. Cosentino S, et al. APOE ϵ 4 allele predicts faster cognitive decline in mild Alzheimer disease. *Neurology*. 2008; 70:1842–1849. [PubMed: 18401023]
47. Fleisher AS, et al. Clinical predictors of progression to Alzheimer disease in amnesic mild cognitive impairment. *Neurology*. 2007; 68:1588–1595. [PubMed: 17287448]
48. Elias-Sonnenschein LS, Vechtbauser W, Ramakers IH, Verhey FR, Visser PJ. Predictive value of APOE-epsilon4 allele for progression from MCI to AD-type dementia: a meta-analysis. *J Neurol Neurosurg Psychiatry*. 2011; 82:1149–56. [PubMed: 21493755]
49. Petersen RC, et al. Apolipoprotein E status as a predictor of the development of Alzheimer’s disease in memory-impaired individuals. *JAMA*. 1995; 273:1274–1278. [PubMed: 7646655]
50. Vemuri P, et al. Effect of apolipoprotein E on biomarkers of amyloid load and neuronal pathology in Alzheimer disease. *Ann Neurol*. 2010; 67:308–316. [PubMed: 20373342]
51. Pike KE, et al. β -amyloid imaging and memory in non-demented individuals: evidence for preclinical Alzheimer’s disease. *Brain*. 2007; 130:2837–2844. [PubMed: 17928318]
52. Bunce D, Fratiglioni L, Small BJ, Winblad B, Bäckman L. APOE and cognitive decline in preclinical Alzheimer disease and non-demented aging. *Neurology*. 2004; 63:816–821. [PubMed: 15365129]
53. Lin AL, Laird AR, Fox PT, Gao JH. Multimodal MRI neuroimaging biomarkers for cognitive normal adults, amnesic mild cognitive impairment, and Alzheimer’s disease. *Neurology research international*. 2012; 2012:907409. [PubMed: 21949904]
54. Caselli RJ, et al. Cognitive domain decline in healthy apolipoprotein E epsilon4 homozygotes before the diagnosis of mild cognitive impairment. *Arch Neurol*. 2007; 64:1306–11. [PubMed: 17846270]
55. Caselli RJ, et al. Longitudinal changes in cognition and behavior in asymptomatic carriers of the APOE ϵ 4 allele. *Neurology*. 2004; 62:1990–1995. [PubMed: 15184602]

56. Caselli RJ, et al. Longitudinal modeling of age-related memory decline and the APOE ϵ 4 effect. *N Engl J Med*. 2009; 361:255–263. [PubMed: 19605830]
57. Caselli RJ, et al. Longitudinal modeling of frontal cognition in APOE epsilon4 homozygotes, heterozygotes, and noncarriers. *Neurology*. 2011; 76:1383–8. [PubMed: 21502596]
58. Izaks GJ, et al. The association of APOE genotype with cognitive function in persons aged 35 years or older. *PLoS ONE*. 2011; 6:e27415. [PubMed: 22110642]
59. Mondadori CRA, et al. Better memory and neural efficiency in young apolipoprotein E ϵ 4 carriers. *Cerebral Cortex*. 2007; 17:1934–1947. [PubMed: 17077159]
60. Jochemsen HM, Muller M, van der Graaf Y, Geerlings MI. APOE epsilon4 differentially influences change in memory performance depending on age. The SMART-MR study. *Neurobiol Aging*. 2012; 33:832 e15–22. [PubMed: 21908077]
61. Bloss CS, Delis DC, Salmon DP, Bondi MW. Decreased cognition in children with risk factors for Alzheimer's disease. *Biol Psychiatry*. 2008; 64:904–906. [PubMed: 18722591]
62. Tuminello ER, Han SD. The Apolipoprotein E antagonistic pleiotropy hypothesis: review and recommendations. *Int J Alzheimers Dis*. 2011
63. Castellano JM, et al. Human apoE isoforms differentially regulate brain amyloid- β peptide clearance. *Sci Transl Med*. 2011; 3:89ra57.
64. Morris JC, et al. APOE predicts amyloid-beta but not tau Alzheimer pathology in cognitively normal aging. *Ann Neurol*. 2010; 67:122–131. [PubMed: 20186853]
65. Haan MN, Shemanski L, Jagust WJ, Manolio TA, Kuller L. The role of APOE epsilon4 in modulating effects of other risk factors for cognitive decline in elderly persons. *JAMA*. 1999; 282:40–6. [PubMed: 10404910]
66. Peila R, Rodriguez BL, Launer LJ. Type 2 diabetes, APOE gene, and the risk for dementia and related pathologies: The Honolulu-Asia Aging Study. *Diabetes*. 2002; 51:1256–62. [PubMed: 11916953]
67. Kalmijn S, Feskens EJ, Launer LJ, Kromhout D. Cerebrovascular disease, the apolipoprotein ϵ 4 allele, and cognitive decline in a community-based study of elderly men. *Stroke*. 1996; 27:2230–5. [PubMed: 8969786]
68. Irie F, et al. Enhanced risk for Alzheimer disease in persons with type 2 diabetes and APOE epsilon4: the Cardiovascular Health Study Cognition Study. *Arch Neurol*. 2008; 65:89–93. [PubMed: 18195144]
69. Matsuzaki T, et al. Insulin resistance is associated with the pathology of Alzheimer disease: the Hisayama study. *Neurology*. 2010; 75:764–70. [PubMed: 20739649]
70. Bell RD, et al. Apolipoprotein E controls cerebrovascular integrity via cyclophilin A. *Nature*. 2012; 485:512–516. [PubMed: 22622580]
71. Verghese PB, Castellano JM, Holtzman DM. Apolipoprotein E in Alzheimer's disease and other neurological disorders. *Lancet Neurol*. 2011; 10:241–52. [PubMed: 21349439]
72. Zhou W, et al. Meta-analysis of APOE4 allele and outcome after traumatic brain injury. *J Neurotrauma*. 2008; 25:279–90. [PubMed: 18373478]
73. Fleminger S, Oliver DL, Lovestone S, Rabe-Hesketh S, Giora A. Head injury as a risk factor for Alzheimer's disease: the evidence 10 years on; a partial replication. *J Neurol Neurosurg Psychiatry*. 2003; 74:857–62. [PubMed: 12810767]
74. Magnoni S, Brody DL. New perspectives on amyloid-beta dynamics after acute brain injury: moving between experimental approaches and studies in the human brain. *Arch Neurol*. 2010; 67:1068–73. [PubMed: 20837849]
75. Nicoll JA, Roberts GW, Graham DI. Apolipoprotein E epsilon 4 allele is associated with deposition of amyloid beta-protein following head injury. *Nat Med*. 1995; 1:135–7. [PubMed: 7585009]
76. Jellinger KA. Morphologic diagnosis of "vascular dementia" - a critical update. *J Neurol Sci*. 2008; 270:1–12. [PubMed: 18455191]
77. Yin YW, et al. Association between apolipoprotein E gene polymorphism and the risk of vascular dementia: a meta-analysis. *Neurosci Lett*. 2012; 514:6–11. [PubMed: 22381401]

78. Prince M, et al. The association between APOE and dementia does not seem to be mediated by vascular factors. *Neurology*. 2000; 54:397–402. [PubMed: 10668701]
79. Bender AR, Raz N. Age-related differences in memory and executive functions in healthy APOE varepsilon4 carriers: the contribution of individual differences in prefrontal volumes and systolic blood pressure. *Neuropsychologia*. 2012; 50:704–14. [PubMed: 22245009]
80. Lanterna LA, et al. Meta-analysis of APOE genotype and subarachnoid hemorrhage. *Neurology*. 2007; 69:766–775. [PubMed: 17709709]
81. Biffi A, et al. APOE genotype and extent of bleeding and outcome in lobar intracerebral haemorrhage: a genetic association study. *Lancet Neurol*. 2011; 10:702–9. [PubMed: 21741316]
82. Federoff M, Jimenez-Rolando B, Nalls MA, Singleton AB. A large study reveals no association between APOE and Parkinson's disease. *Neurobiol Dis*. 2012; 46:389–392. [PubMed: 22349451]
83. Ezquerro M, et al. Lack of association of APOE and tau polymorphisms with dementia in Parkinson's disease. *Neurosci Lett*. 2008; 448:20–3. [PubMed: 18930114]
84. McKeith IG, et al. Dementia with Lewy bodies. *Semin Clin Neuropsychiatry*. 2003; 8:46–57. [PubMed: 12567332]
85. Harrington CR, et al. Influence of apolipoprotein E genotype on senile dementia of the Alzheimer and Lewy body types. Significance for etiological theories of Alzheimer's disease. *Am J Pathol*. 1994; 145:1472–84. [PubMed: 7992850]
86. Mann DM, Brown SM, Owen F, Baba M, Iwatsubo T. Amyloid beta protein (abeta) deposition in dementia with Lewy bodies: predominance of A beta 42(43) and paucity of A beta 40 compared with sporadic Alzheimer's disease. *Neuropathol Appl Neurobiol*. 1998; 24:187–94. [PubMed: 9717183]
87. Pletnikova O, et al. Abeta deposition is associated with enhanced cortical alpha-synuclein lesions in Lewy body diseases. *Neurobiol Aging*. 2005; 26:1183–92. [PubMed: 15917102]
88. Seripa D, et al. The APOE gene locus in frontotemporal dementia and primary progressive aphasia. *Arch Neurol*. 2011; 68:622–8. [PubMed: 21555637]
89. Saft C, et al. Apolipoprotein E genotypes do not influence the age of onset in Huntington's disease. *J Neurol Neurosurg Psychiatry*. 2004; 75:1692–6. [PubMed: 15548484]
90. Jawaid A, et al. Does apolipoprotein E genotype modify the clinical expression of ALS? *Eur J Neurol*. 2011; 18:618–24. [PubMed: 20880000]
91. Bales KR, et al. Human APOE isoform-dependent effects on brain β -amyloid levels in PDAPP transgenic mice. *J Neurosci*. 2009; 29:6771–6779. [PubMed: 19474305]
92. Osenkowski P, Ye W, Wang R, Wolfe MS, Selkoe DJ. Direct and potent regulation of gamma-secretase by its lipid microenvironment. *J Biol Chem*. 2008; 283:22529–40. [PubMed: 18539594]
93. Riddell DR, et al. Impact of apolipoprotein E (apoE) polymorphism on brain apoE levels. *J Neurosci*. 2008; 28:11445–11453. [PubMed: 18987181]
94. Sullivan PM, et al. Reduced levels of human apoE4 protein in an animal model of cognitive impairment. *Neurobiol Aging*. 2011; 32:791–801. [PubMed: 19577821]
95. Wahrle SE, et al. Apolipoprotein E levels in cerebrospinal fluid and the effects of ABCA1 polymorphisms. *Mol Neurodegener*. 2007; 2:7. [PubMed: 17430597]
96. Beffert U, et al. Apolipoprotein E and beta-amyloid levels in the hippocampus and frontal cortex of Alzheimer's disease subjects are disease-related and apolipoprotein E genotype dependent. *Brain Res*. 1999; 843:87–94. [PubMed: 10528114]
97. DeMattos RB, et al. ApoE and clusterin cooperatively suppress Abeta levels and deposition: evidence that ApoE regulates extracellular Abeta metabolism in vivo. *Neuron*. 2004; 41:193–202. [PubMed: 14741101]
98. Riddell DR, et al. The LXR agonist TO901317 selectively lowers hippocampal Abeta42 and improves memory in the Tg2576 mouse model of Alzheimer's disease. *Mol Cell Neurosci*. 2007; 34:621–8. [PubMed: 17336088]
99. Vanmierlo T, et al. Liver X receptor activation restores memory in aged AD mice without reducing amyloid. *Neurobiol Aging*. 2011; 32:1262–72. [PubMed: 19674815]
100. Cramer PE, et al. ApoE-directed therapeutics rapidly clear beta-amyloid and reverse deficits in AD mouse models. *Science*. 2012; 335:1503–6. [PubMed: 22323736]

101. Fitz NF, et al. Abca1 Deficiency Affects Alzheimer's Disease-Like Phenotype in Human ApoE4 But Not in ApoE3-Targeted Replacement Mice. *J Neurosci.* 2012; 32:13125–36. [PubMed: 22993429]
102. Kim J, Basak JM, Holtzman DM. The role of apolipoprotein E in Alzheimer's disease. *Neuron.* 2009; 63:287–303. [PubMed: 19679070]
103. LaDu MJ, et al. Isoform-specific binding of apolipoprotein E to beta-amyloid. *J Biol Chem.* 1994; 269:23403–23406. [PubMed: 8089103]
104. Deane R, et al. ApoE isoform-specific disruption of amyloid β peptide clearance from mouse brain. *J Clin Invest.* 2008; 118:4002–4013. [PubMed: 19033669]
105. Jiang Q, et al. ApoE promotes the proteolytic degradation of A β . *Neuron.* 2008; 58:681–693. [PubMed: 18549781]
106. Bales KR, et al. Lack of apolipoprotein E dramatically reduces amyloid beta-peptide deposition. *Nat Genet.* 1997; 17:263–4. [PubMed: 9354781]
107. Ries ML, et al. Magnetic resonance imaging characterization of brain structure and function in mild cognitive impairment: a review. *J Am Geriatr Soc.* 2008; 56:920–934. [PubMed: 18410325]
108. Nestor PJ, Scheltens P, Hodges JR. Advances in the early detection of Alzheimer's disease. *Nat Med.* 2004; 10 (Suppl):S34–41. [PubMed: 15298007]
109. Korf ESC, Wahlund LO, Visser PJ, Scheltens P. Medial temporal lobe atrophy on MRI predicts dementia in patients with mild cognitive impairment. *Neurology.* 2004; 63:94–100. [PubMed: 15249617]
110. Hashimoto M, et al. Apolipoprotein E ϵ 4 and the pattern of regional brain atrophy in Alzheimer's disease. *Neurology.* 2001; 57:1461–1466. [PubMed: 11673590]
111. Espeseth T, et al. Accelerated age-related cortical thinning in healthy carriers of apolipoprotein E ϵ 4. *Neurobiol Aging.* 2008; 29:329–340. [PubMed: 17161508]
112. Reiman EM, et al. Hippocampal volumes in cognitively normal persons at genetic risk for Alzheimer's disease. *Ann Neurol.* 1998; 44:288–291. [PubMed: 9708558]
113. Fennema-Notestine C, et al. Presence of APOE ϵ 4 allele associated with thinner frontal cortex in middle age. *J Alzheimers Dis.* 2011; 26:49–60. [PubMed: 21971450]
114. Sheline YI, et al. APOE4 allele disrupts resting state fMRI connectivity in the absence of amyloid plaques or decreased CSF A β 42. *J Neurosci.* 2010; 30:17035–40. [PubMed: 21159973]
115. Machulda M, JDTVP, et al. Effect of apoe ϵ 4 status on intrinsic network connectivity in cognitively normal elderly subjects. *Arch Neurol.* 2011; 68:1131–1136. [PubMed: 21555604]
116. Bookheimer SY, et al. Patterns of brain activation in people at risk for Alzheimer's disease. *N Engl J Med.* 2000; 343:450–456. [PubMed: 10944562]
117. Filippini N, et al. Distinct patterns of brain activity in young carriers of the APOE-epsilon4 allele. *Proc Natl Acad Sci USA.* 2009; 106:7209–14. [PubMed: 19357304]
118. Bondi MW, Houston WS, Eyler LT, Brown GG. fMRI evidence of compensatory mechanisms in older adults at genetic risk for Alzheimer disease. *Neurology.* 2005; 64:501–508. [PubMed: 15699382]
119. Bero AW, et al. Neuronal activity regulates the regional vulnerability to amyloid-[beta] deposition. *Nat Neurosci.* 2011; 14:750–756. [PubMed: 21532579]
120. Cirrito JR, et al. Synaptic activity regulates interstitial fluid Amyloid- β levels in vivo. *Neuron.* 2005; 48:913–922. [PubMed: 16364896]
121. Putcha D, et al. Hippocampal hyperactivation associated with cortical thinning in Alzheimer's disease signature regions in non-demented elderly adults. *J Neurosci.* 2011; 31:17680–17688. [PubMed: 22131428]
122. O'Brien JL, et al. Longitudinal fMRI in elderly reveals loss of hippocampal activation with clinical decline. *Neurology.* 2010; 74:1969–1976. [PubMed: 20463288]
123. Mosconi L, et al. Pre-clinical detection of Alzheimer's disease using FDG-PET, with or without amyloid imaging. *J Alzheimers Dis.* 2010; 20:843–854. [PubMed: 20182025]
124. Small GW, et al. Apolipoprotein E type 4 allele and cerebral glucose metabolism in relatives at risk for familial Alzheimer disease. *JAMA.* 1995; 273:942–7. [PubMed: 7884953]

125. Reiman EM, et al. Preclinical evidence of Alzheimer's disease in persons homozygous for the epsilon 4 allele for apolipoprotein E. *N Engl J Med.* 1996; 334:752–8. [PubMed: 8592548]
126. Small GW, et al. Cerebral metabolic and cognitive decline in persons at genetic risk for Alzheimer's disease. *Proc Natl Acad Sci USA.* 2000; 97:6037–6042. [PubMed: 10811879]
127. Reiman EM, et al. Correlations between apolipoprotein E ϵ 4 gene dose and brain-imaging measurements of regional hypometabolism. *Proc Natl Acad Sci USA.* 2005; 102:8299–8302. [PubMed: 15932949]
128. Huang Y. $\text{A}\beta$ -independent roles of apolipoprotein E4 in the pathogenesis of Alzheimer's disease. *Trends Mol Med.* 2010; 16:287–294. [PubMed: 20537952]
129. Aoki K, et al. Increased expression of neuronal apolipoprotein E in human brain with cerebral infarction. *Stroke.* 2003; 34:875–880. [PubMed: 12649507]
130. Brecht WJ, et al. Neuron-specific apolipoprotein E4 proteolysis is associated with increased Tau phosphorylation in brains of transgenic mice. *J Neurosci.* 2004; 24:2527–2534. [PubMed: 15014128]
131. Mahley RW, Weisgraber KH, Huang Y. Apolipoprotein E4: A causative factor and therapeutic target in neuropathology, including Alzheimer's disease. *Proc Natl Acad Sci USA.* 2006; 103:5644–5651. [PubMed: 16567625]
132. Aboud O, Mrak RE, Boop F, Griffin ST. Apolipoprotein epsilon 3 alleles are associated with indicators of neuronal resilience. *BMC medicine.* 2012; 10:35. [PubMed: 22502727]
133. Buttini M, et al. Cellular source of apolipoprotein E4 determines neuronal susceptibility to excitotoxic injury in transgenic mice. *Am J Pathol.* 2010; 177:563–9. [PubMed: 20595630]
134. Mauch DH, et al. CNS synaptogenesis promoted by glia-derived cholesterol. *Science.* 2001; 294:1354–1357. [PubMed: 11701931]
135. Pfrieger FW. Cholesterol homeostasis and function in neurons of the central nervous system. *Cell Mol Life Sci.* 2003; 60:1158–1171. [PubMed: 12861382]
136. Svennerholm L, Gottfries CG. Membrane lipids, selectively diminished in Alzheimer brains, suggest synapse loss as a primary event in early-onset form (Type I) and demyelination in late-onset form (Type II). *J Neurochem.* 1994; 62:1039–1047. [PubMed: 8113790]
137. Rapp A, Gmeiner B, Hüttinger M. Implication of apoE isoforms in cholesterol metabolism by primary rat hippocampal neurons and astrocytes. *Biochimie.* 2006; 88:473–483. [PubMed: 16376010]
138. Hamanaka H, et al. Altered cholesterol metabolism in human apolipoprotein E4 knock-in mice. *Hum Mol Genet.* 2000; 9:353–361. [PubMed: 10655544]
139. Selkoe DJ. Alzheimer's disease is a synaptic failure. *Science.* 2002; 298:789–791. [PubMed: 12399581]
140. Koffie R, Hyman B, Spires-Jones T. Alzheimer's disease: synapses gone cold. *Mol Neurodegener.* 2011; 6:63. [PubMed: 21871088]
141. Chen Y, Durakoglugil MS, Xian X, Herz J. ApoE4 reduces glutamate receptor function and synaptic plasticity by selectively impairing ApoE receptor recycling. *Proc Natl Acad Sci USA.* 2010
142. Buttini M, et al. Modulation of alzheimer-like synaptic and cholinergic deficits in transgenic mice by human apolipoprotein E depends on isoform, aging, and overexpression of amyloid β peptides but not on plaque formation. *J Neurosci.* 2002; 22:10539–10548. [PubMed: 12486146]
143. Ji Y, et al. Apolipoprotein E isoform-specific regulation of dendritic spine morphology in apolipoprotein E transgenic mice and Alzheimer's disease patients. *Neuroscience.* 2003; 122:305–315. [PubMed: 14614898]
144. Dumanis SB, et al. ApoE4 decreases spine density and dendritic complexity in cortical neurons in vivo. *J Neurosci.* 2009; 29:15317–15322. [PubMed: 19955384]
145. Wang C, et al. Human apoE4-targeted replacement mice display synaptic deficits in the absence of neuropathology. *Neurobiol Dis.* 2005; 18:390–398. [PubMed: 15686968]
146. Sen A, Alkon DL, Nelson TJ. Apolipoprotein E3 (ApoE3) but not ApoE4 protects against synaptic loss through increased expression of protein kinase C ϵ . *J Biol Chem.* 2012; 287:15947–15958. [PubMed: 22427674]

147. Klein RC, Mace BE, Moore SD, Sullivan PM. Progressive loss of synaptic integrity in human apolipoprotein E4 targeted replacement mice and attenuation by apolipoprotein E2. *Neuroscience*. 2010; 171:1265–1272. [PubMed: 20951774]
148. Bales KR, Du Y, Holtzman D, Cordell B, Paul SM. Neuroinflammation and Alzheimer's disease: critical roles for cytokine/A β -induced glial activation, NF- κ B, and apolipoprotein E. *Neurobiol Aging*. 2000; 21:427–432. [PubMed: 10858588]
149. LaDu MJ, et al. Apolipoprotein E and apolipoprotein E receptors modulate A β -induced glial neuroinflammatory responses. *Neurochem Int*. 2001; 39:427–434. [PubMed: 11578778]
150. Lynch JR, Morgan D, Mance J, Matthew WD, Laskowitz DT. Apolipoprotein E modulates glial activation and the endogenous central nervous system inflammatory response. *J Neuroimmunol*. 2001; 114:107–113. [PubMed: 11240021]
151. Keene CD, Cudaback E, Li X, Montine KS, Montine TJ. Apolipoprotein E isoforms and regulation of the innate immune response in brain of patients with Alzheimer's disease. *Curr Opin Neurobiol*. 2011; 21:920–8. [PubMed: 21907569]
152. Lynch JR, et al. APOE genotype and an ApoE-mimetic peptide modify the systemic and central nervous system inflammatory response. *J Biol Chem*. 2003; 278:48529–48533. [PubMed: 14507923]
153. Ringman, Jm; EDGDH, et al. Plasma signaling proteins in persons at genetic risk for alzheimer disease: Influence of apoe genotype. *Arch Neurol*. 2012; 69:757–764. [PubMed: 22689192]
154. Szekely CA, et al. NSAID use and dementia risk in the Cardiovascular Health Study: role of APOE and NSAID type. *Neurology*. 2008; 70:17–24. [PubMed: 18003940]
155. Mu Y, Gage F. Adult hippocampal neurogenesis and its role in Alzheimer's disease. *Mol Neurodegener*. 2011; 6:85. [PubMed: 22192775]
156. Yang CP, Gilley JA, Zhang G, Kernie SG. ApoE is required for maintenance of the dentate gyrus neural progenitor pool. *Development*. 2011; 138:4351–4362. [PubMed: 21880781]
157. Andrews-Zwilling Y, et al. Apolipoprotein E4 causes age- and Tau-dependent impairment of GABAergic interneurons, leading to learning and memory deficits in mice. *J Neurosci*. 2010; 30:13707–17. [PubMed: 20943911]
158. Li G, et al. GABAergic interneuron dysfunction impairs hippocampal neurogenesis in adult apolipoprotein E4 knockin mice. *Cell Stem Cell*. 2009; 5:634–45. [PubMed: 19951691]
159. Salloway, S. European Federation of Neurological Societies. Stockholm; Sweden: 2012.
160. Sperling, R. European Federation of Neurological Societies. Stockholm; Sweden: 2012.
161. Ferrari C, et al. How can elderly apolipoprotein E epsilon4 carriers remain free from dementia? *Neurobiol Aging*. 2012
162. Hesse C, et al. Measurement of apolipoprotein E (apoE) in cerebrospinal fluid. *Neurochem Res*. 2000; 25:511–7. [PubMed: 10823584]
163. Gupta VB, et al. Plasma apolipoprotein E and Alzheimer disease risk. *Neurology*. 2011; 76:1091–1098. [PubMed: 21422459]
164. Chawla A, et al. A PPAR gamma-LXR-ABCA1 pathway in macrophages is involved in cholesterol efflux and atherogenesis. *Mol Cell*. 2001; 7:161–71. [PubMed: 11172721]
165. Vaya J, Schipper HM. Oxysterols, cholesterol homeostasis, and Alzheimer disease. *J Neurochem*. 2007; 102:1727–37. [PubMed: 17573819]
166. Donkin JJ, et al. ATP-binding cassette transporter A1 mediates the beneficial effects of the liver X receptor agonist GW3965 on object recognition memory and amyloid burden in amyloid precursor protein/presenilin 1 mice. *J Biol Chem*. 2010; 285:34144–54. [PubMed: 20739291]
167. Fryer JD, et al. Human apolipoprotein E4 alters the amyloid-beta 40:42 ratio and promotes the formation of cerebral amyloid angiopathy in an amyloid precursor protein transgenic model. *J Neurosci*. 2005; 25:2803–10. [PubMed: 15772340]
168. Kim J, et al. Haploinsufficiency of human APOE reduces amyloid deposition in a mouse model of amyloid-beta amyloidosis. *J Neurosci*. 2011; 31:18007–12. [PubMed: 22159114]
169. Bien-Ly N, Gillespie AK, Walker D, Yoon SY, Huang Y. Reducing human apolipoprotein E levels attenuates age-dependent A β accumulation in mutant human amyloid precursor protein transgenic mice. *J Neurosci*. 2012; 32:4803–11. [PubMed: 22492035]

170. Sadowski MJ, et al. Blocking the apolipoprotein E/amyloid-beta interaction as a potential therapeutic approach for Alzheimer's disease. *Proc Natl Acad Sci USA*. 2006; 103:18787–92. [PubMed: 17116874]
171. Chen HK, et al. Small molecule structure correctors abolish detrimental effects of apolipoprotein E4 in cultured neurons. *J Biol Chem*. 2012; 287:5253–66. [PubMed: 22158868]
172. Aono M, et al. Protective effect of apolipoprotein E-mimetic peptides on N-methyl-D-aspartate excitotoxicity in primary rat neuronal-glial cell cultures. *Neuroscience*. 2003; 116:437–45. [PubMed: 12559098]
173. Li FQ, et al. Apolipoprotein E-derived peptides ameliorate clinical disability and inflammatory infiltrates into the spinal cord in a murine model of multiple sclerosis. *J Pharmacol Exp Ther*. 2006; 318:956–65. [PubMed: 16740622]
174. Li FQ, Fowler KA, Neil JE, Colton CA, Vitek MP. An apolipoprotein E-mimetic stimulates axonal regeneration and remyelination after peripheral nerve injury. *J Pharmacol Exp Ther*. 2010; 334:106–15. [PubMed: 20406857]
175. Kanekiyo T, et al. Heparan sulphate proteoglycan and the low-density lipoprotein receptor-related protein 1 constitute major pathways for neuronal amyloid- β uptake. *J Neurosci*. 2011; 31:1644–1651. [PubMed: 21289173]
176. Liu Q, et al. Neuronal LRP1 knockout in adult mice leads to impaired brain lipid metabolism and progressive, age-dependent synapse loss and neurodegeneration. *J Neurosci*. 2010; 30:17068–17078. [PubMed: 21159977]
177. Kim J, et al. Overexpression of low-density lipoprotein receptor in the brain markedly inhibits amyloid deposition and increases extracellular abeta clearance. *Neuron*. 2009; 64:632–644. [PubMed: 20005821]
178. Herz J, Chen Y. Reelin, lipoprotein receptors and synaptic plasticity. *Nat Rev Neurosci*. 2006; 7:850–859. [PubMed: 17053810]

Biographies

Chia-Chen Liu is a Postdoctoral Research fellow in the Department of Neuroscience at Mayo Clinic College of Medicine, Jacksonville, FL, USA. She is also a visiting scientist in the Institute of Neuroscience at Xiamen University in Xiamen, China. She gained her Ph.D. from Washington University School of Medicine, St. Louis, MO, USA. Her current research interests focus on apolipoprotein E and its receptors in Alzheimer diseases and vascular dementia.

Takahisa Kanekiyo is an Instructor in the Department of Neuroscience at Mayo Clinic College of Medicine, Jacksonville, FL, USA. He gained his M.D. and Ph.D. from Osaka University Graduate School of Medicine, Japan, and engaged in postdoctoral training at Washington University School of Medicine, St. Louis, MO, USA. His research interests focus on apolipoprotein E and its receptors in neurovascular diseases including Alzheimer diseases and cerebral amyloid angiopathy.

Huaxi Xu is an adjunct Professor in the Institute of Neuroscience at Xiamen University in Xiamen, China. His research focuses on the molecular and cellular mechanisms underlying the pathogenesis of Alzheimer disease. His major contributions include dissecting the cellular trafficking pathways for amyloid precursor protein (APP) and presenilins, and identifying cellular factors that regulate APP processing and neuronal apoptosis. He serves as Co-Editor-in-Chief of *Molecular Neurodegeneration*.

Guojun Bu is a Professor of Neuroscience at Mayo Clinic College of Medicine, Jacksonville, FL, USA. He is also an adjunct Professor in the Institute of Neuroscience at Xiamen University in Xiamen, China. Since 1994, he has led a research programme studying the biological and pathological functions of Apolipoprotein E (ApoE) and ApoE

receptors using cellular and animal models, with a specific focus on the pathogenesis of Alzheimer disease. His group has defined a critical role for the ApoE receptor lipoprotein receptor-related protein 1 in brain ApoE metabolism and in the clearance of amyloid peptides. He founded and serves as the Editor-in-Chief of *Molecular Neurodegeneration*.

Key Points

The $\epsilon 4$ allele of the apolipoprotein E (*APOE*) gene is the main genetic risk factor for Alzheimer disease (AD)

APOE $\epsilon 4$ carriers have enhanced AD pathology, accelerated age-dependent cognitive decline and worse memory performance than do noncarriers.

Numerous structural and functional brain changes associated with AD pathogenesis are detected in *APOE* $\epsilon 4$ carriers before clinical symptoms become evident.

ApoE affects amyloid- β ($A\beta$) clearance, aggregation and deposition in an isoform-dependent manner.

ApoE4 also contributes to AD pathogenesis by $A\beta$ -independent mechanisms that involve synaptic plasticity, cholesterol homeostasis, neurovascular functions, and neuroinflammation.

ApoE-targeted AD therapy should focus on restoration of the physiological function of ApoE through increased expression and lipidation, and inhibition of the detrimental effects of ApoE4.

Review criteria

This Review was based on searches of the PubMed database using the following terms: “apolipoprotein E”, “cognitive decline”, “Alzheimer disease”, “amyloid beta”, “synaptic plasticity”, “cerebral amyloid angiopathy”, “mild cognitive impairment”, “cholesterol”, “brain activity”, “cerebrovascular diseases”, “brain metabolism”, “neurogenesis”, “brain atrophy”, “neuroinflammation”, “tau” and “traumatic brain injury”. Only articles published in English were retrieved. Full-text papers were available for most of the articles that were chosen for review, and the references of these articles were searched for further relevant material.

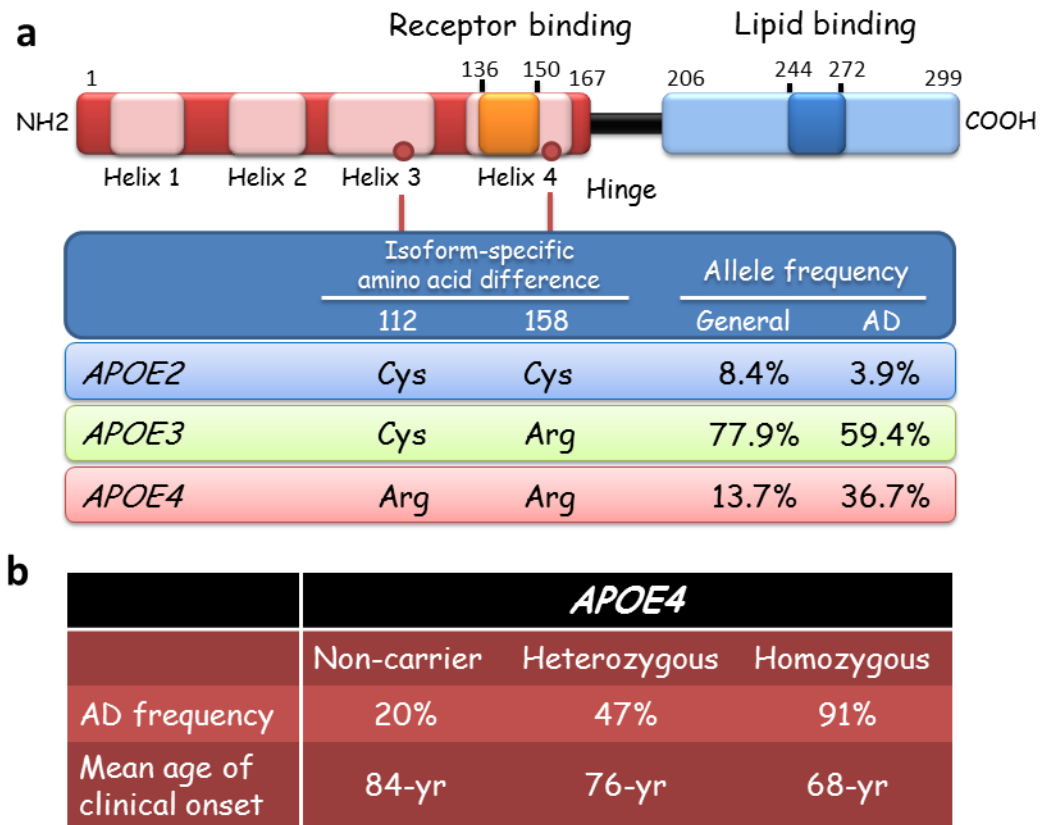


Figure 1. *APOE* $\epsilon 4$ is a major genetic risk factor for Alzheimer disease

(a) The ApoE2, E3, and E4 isoforms, which are encoded by the $\epsilon 2$, $\epsilon 3$ and $\epsilon 4$ alleles of the *APOE* gene, respectively, differ from one another at amino acid residues 112 and/or 158 (red circles). ApoE has two structural domains: the N-terminal domain, which contains the receptor-binding region (residues 136–150), and the C-terminal domain, which contains the lipid-binding region (residues 244–272); the two domains are joined by a hinge region. A meta-analysis demonstrated a significant association between the $\epsilon 4$ allele of *APOE* and AD.¹⁰ (b) *APOE* $\epsilon 4$ increases the risk of AD and lowers the age of disease onset in a gene-dose-dependent manner.^{7, 20} Abbreviations: AD, Alzheimer disease; ApoE, Apolipoprotein E.

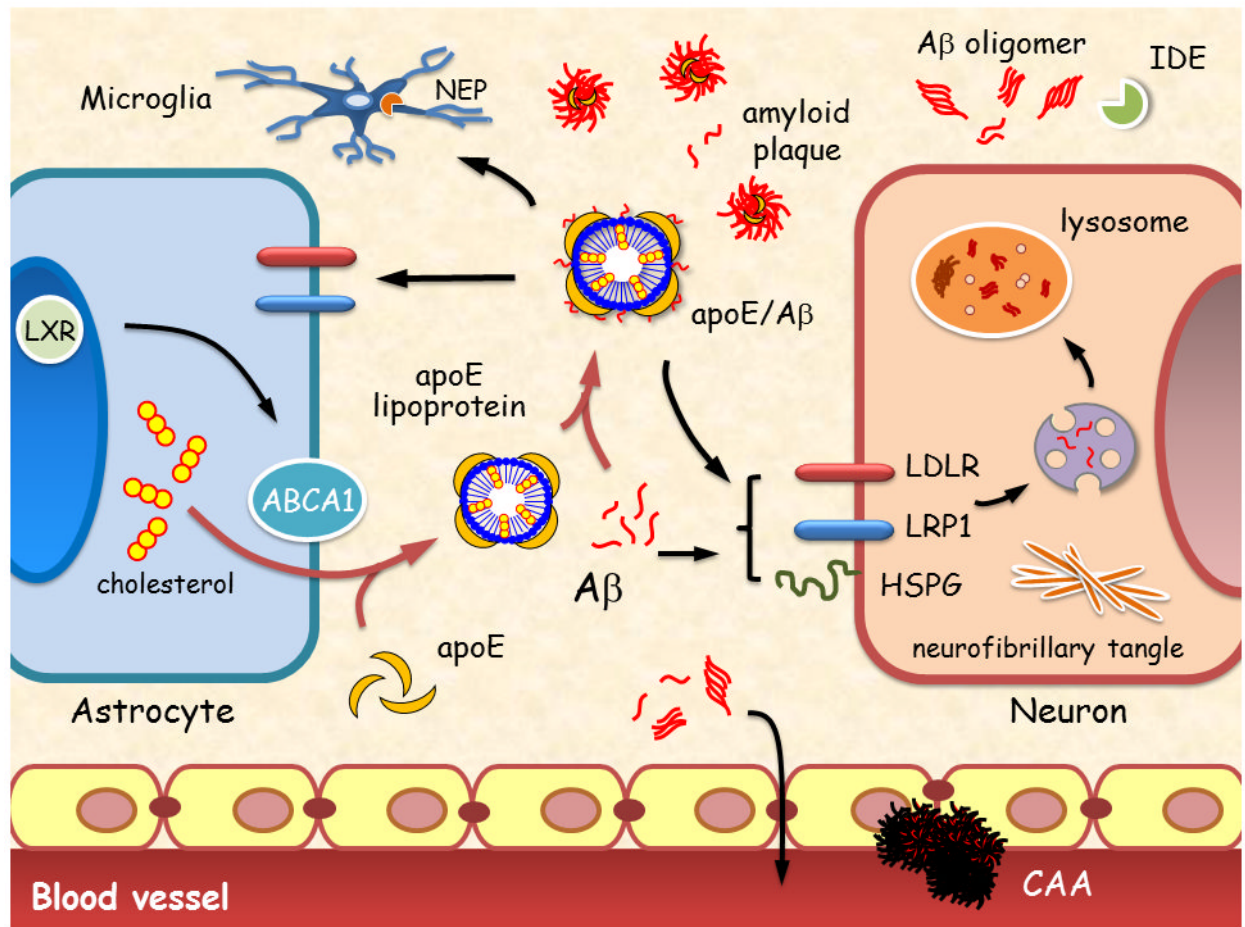


Figure 2. Apolipoprotein E and amyloid- β metabolism in the brain

The main A β clearance pathways include receptor-mediated uptake by neurons and glia, drainage into interstitial fluid or through the BBB, and proteolytic degradation by IDE and neprilysin. Impaired clearance of A β can cause A β accumulation in brain parenchyma, leading to formation of neurotoxic A β oligomers and amyloid plaques. A β accumulation in the perivascular region leads to CAA, which disrupts blood vessel function. ApoE is primarily synthesized by astrocytes and microglia, and is lipidated by the ABCA1 transporter to form lipoprotein particles. Lipidated ApoE binds to soluble A β and facilitates A β uptake through cell surface receptors, including LRP1, LDLR, and HSPG^{175, 177} in a manner that probably depends on ApoE isoform and its level of lipidation. ApoE facilitates binding and internalization of soluble A β by glial cells, disrupts A β clearance at the BBB in an isoform-dependent manner (ApoE4 > ApoE3 > ApoE2) and influences CAA pathogenesis. Abbreviations: A β , amyloid- β ; ABCA1, ATP-binding cassette A1; BBB, blood-brain barrier; CAA, cerebral amyloid angiopathy; HSPG, heparan sulphate proteoglycan; IDE, insulin-degrading enzyme; LDLR, low-density lipoprotein receptor; LRP1, low-density lipoprotein receptor-related protein 1; LXR, liver X receptor.

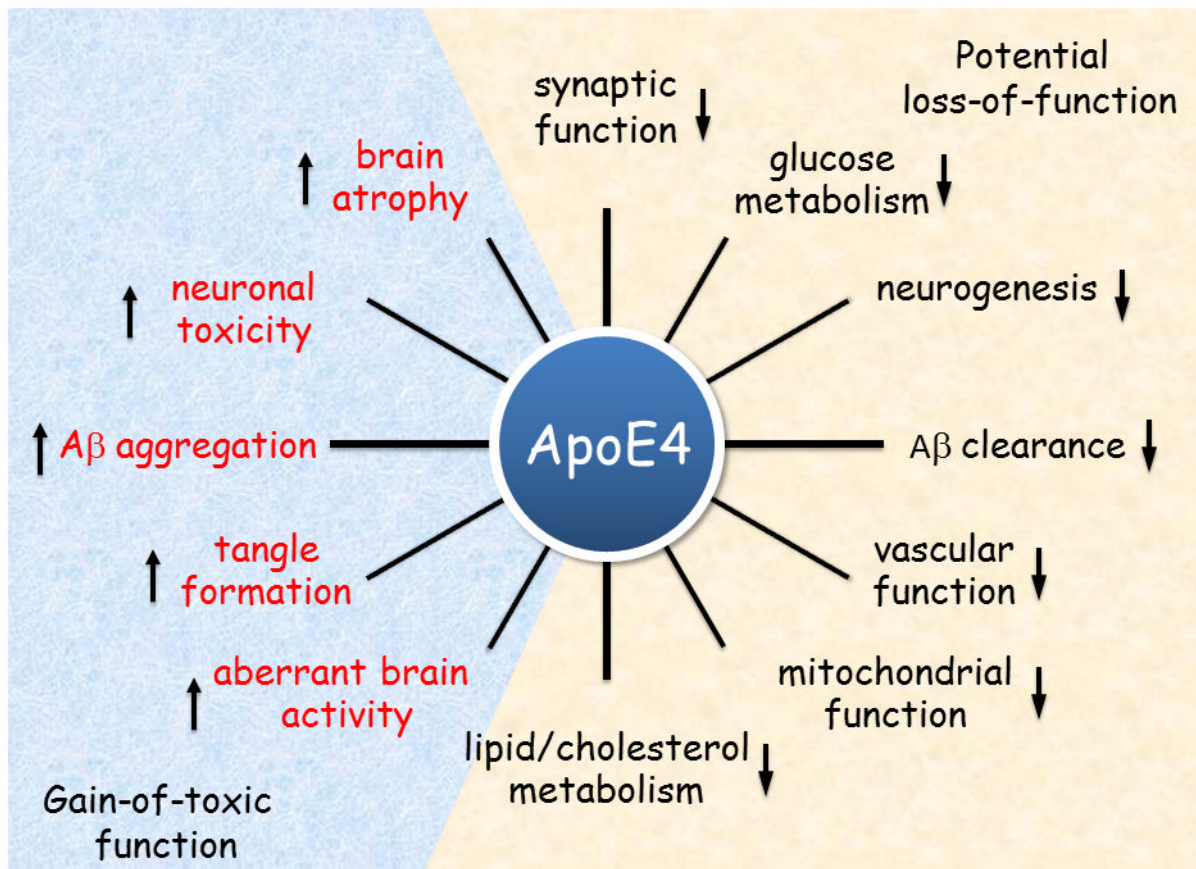


Figure 3. The role of Apolipoprotein E4 in Alzheimer disease pathogenesis

ApoE4 confers toxic gain of function, loss of neuroprotective function or both in the pathogenesis of Alzheimer disease. Key functional differences between ApoE4 and ApoE3 are illustrated. Abbreviations: A β , amyloid- β ; ApoE, Apolipoprotein E.

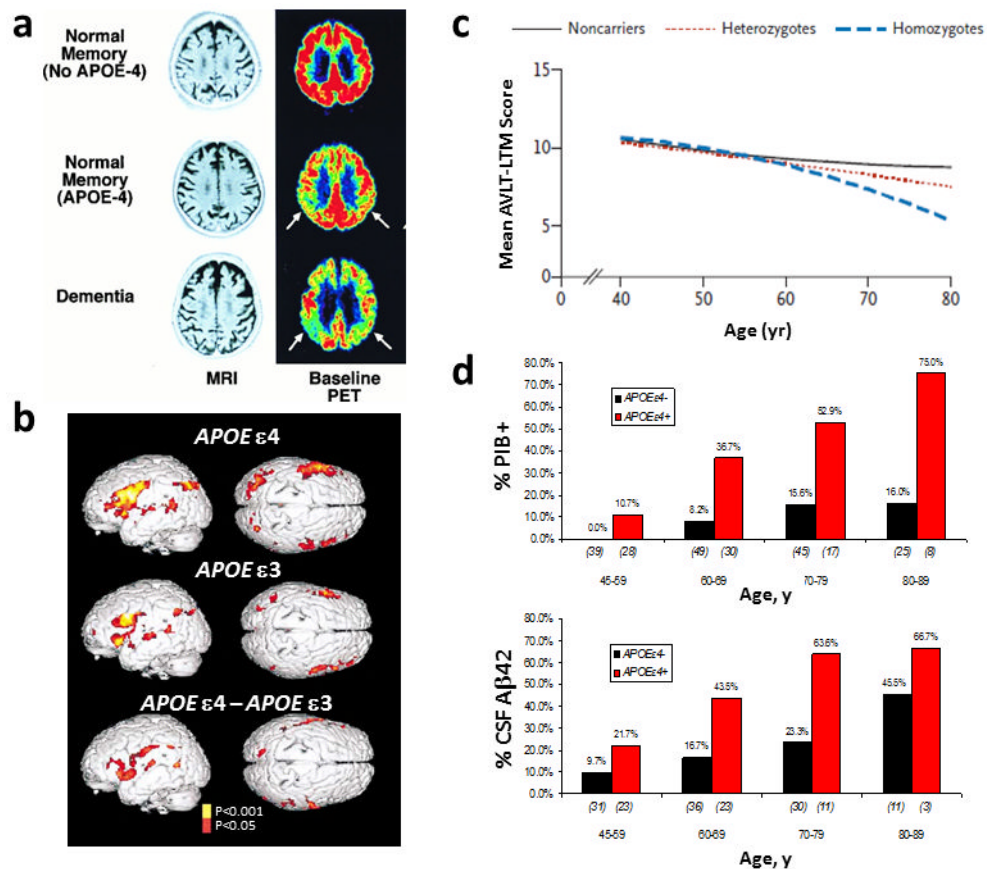


Figure 4. Abnormal brain function and enhanced neuropathology and memory decline in cognitively normal *APOE* ϵ 4 carriers

(a) ^{18}F -fluorodeoxyglucose PET images show that cognitively normal *APOE* ϵ 4 carriers have lower glucose metabolism than do noncarriers. (b) *APOE* ϵ 4 carriers exhibit a greater increase in functional MRI signal in brain regions associated with task performance, and show increases in additional regions compared with *APOE* ϵ 3 carriers. (c) Age-related memory decline occurs more rapidly in *APOE* ϵ 4 carriers than noncarriers, starting from age 55–60 years. (d) *APOE* ϵ 4 carriers show increased cerebral A β deposition which persists in greater frequencies with age compared with noncarriers. Increased PiB binding and reduced CSF A β ₄₂ levels reflect cerebral amyloid deposition. Abbreviations: A β , amyloid- β ; APOE, apolipoprotein E; CSF, cerebrospinal fluid; PiB, Pittsburgh compound B. Part a, is modified, with permission from the National Academy of Science, USA © Small, G. W. *et al. Proc. Natl Acad. Sci. USA* **97**, 6037–6042 (2000). Part b, is modified, with permission from the Massachusetts Medical Society © Bookheimer, S. Y. *et al. N. Engl. J. Med.* **343**, 450–456 (2000). Part c, is modified, with permission from the Massachusetts Medical Society © Caselli *et al. N. Engl. J. Med.* **361**, 255–263 (2009). Part d is modified, with permission, from John Wiley and Sons © Morris, J. C. *et al. Ann. Neurol.* **67**, 122–131 (2010).

Table 1

ApoE-targeted strategies for treatment of Alzheimer disease

Strategy	Rationale	Examples
Pharmacological approaches		
Modulate ApoE levels	Promotes A β clearance, lipid homeostasis and synaptic function	LXR/RXR agonists, small molecules
Increase ABCA1 expression	Promotes ApoE lipitation and stabilizes ApoE, thereby decreasing amyloid deposition	LXR/RXR agonists, small molecules
Disrupt ApoE-A β interaction	Reduces A β aggregation and deposition	A β 12-28P, small molecule inhibitors, ApoE-specific antibody
Conversion of ApoE4 to ApoE3	Increases ApoE3-associated protective functions and decreases ApoE4-related toxic effects	Small compounds (i.e., disulphonate and (monosulphoalkyl))
Restore ApoE functions	Increases ApoE protective functions and decreases neuroinflammation	ApoE-mimetic peptide
Blockade of ApoE fragmentation	Decreases tau pathology, and toxicity to mitochondria	Inhibitors for specific proteinases involved in ApoE fragmentation
Increase LRP1 and/or LDLR levels	Enhances A β clearance, cholesterol transport and synaptic plasticity	Small molecules
Increase apoE receptor 2 and/or VLDLR levels	Increases ApoE signalling and synaptic plasticity	Small molecules
Restore brain vascular integrity	Eliminates ApoE4-mediated blood-brain barrier breakdown and leakage of blood-derived toxic molecules	Cyclosporine A
Nonpharmacological approaches		
APOE genotyping prior to immunotherapy	Helps to predict clinical outcome for A β -targeted or other therapies	A β immunotherapy might be more effective in APOE ϵ 4 carriers or noncarriers
Preventive care	Maintains healthy brain vasculature function	Physical exercise, intellectual activities (e.g., puzzles), social connections (e.g., calling family and friends), healthy diet

Abbreviations: A β , amyloid- β ; ABCA1, ATP-binding cassette transporter A1; AD, Alzheimer disease; ApoE, apolipoprotein E; LDLR, low-density lipoprotein receptor; LRP1, low-density lipoprotein receptor-related protein 1; LXR, liver X receptor; RXR, retinoid X receptor; VLDLR, very-low-density lipoprotein receptor.