Apple Thinning by Photosynthetic Inhibition

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Abstract. Shading (92%) of 'Redchief Delicious' apple (*Malus domestics* Borkh.) trees for 10-day periods from 10 to 20, 15 to 25, 20 to 30, and 25 to 35 days after full bloom (DAFB) caused greater fruit abscission than shading from 5 to 15, 30 to 40, 35 to 45, or 47 to 57 DAFB. Fruit 8 to 33 mm in diameter (10 to 30 DAFB) were very sensitive to 10 days of shade, even though fruit sizes of 6 to 12 mm are considered the most sensitive to chemical thinners. In a second test, shading for 3 days caused fruit thinning; 5 days of shade in the periods 18 to 23, 23 to 28, and 28 to 33 DAFB caused greater thinning than 11 to 16 or 33 to 38 DAFB. Shading reduced photosynthesis (Pn) to about one-third that of noncovered trees. Terbacil (50 mg·liter⁻¹) + X-77 surfactant (1250 mg·liter⁻¹) applied with a hand-pump sprayer 5, 10, or 15 DAFB greatly reduced fruit set and caused some leaf yellowing, particularly in the earliest treatments. Terbacil reduced Pn by more than 90% at 72 hours after application. Shoot growth of trees defruited by shade or terbacil was equivalent to defruited or deblossomed trees; ethephon (1500 mg·liter⁻¹) inhibited tree growth and defruited trees. No terbacil residues were dectected in fruit at harvest from applications made 5, 15, 20, 25, or 30 DAFB. Eleven of 12 photosynthesis-inhibiting herbicides were also found to thin 'Redchief Delicious' apple trees. Shading caused more thinning than terbacil at the later applications, which may reflect poorer absorption and/or lesser photosynthetic inhibition than when terbacil was applied to older leaves.

Spur 'Delicious' strains of apple normally set heavy crops and are difficult to thin adequately with currently available thinning agents (Byers, 1978; Byers et al., 1982; Herrera-Aguirre and Unrath,, 1980; Unrath, 1978, 1981). High rates of naphthaleneacetic acid (NAA) or naphthaleneacetamide (NAD) may cause many dwarfed (pygmy) fruit (Byers, 1978; Byers et al., 1982; Rogers and Thompson, 1969; Rogers and Williams, 1977; Unrath 1978, 1981). Carbaryl plus lower rates of NAA (5 mg-liter⁻¹) have given excellent results in some years, but have caused serious overthinning and/or pygmy fruit development in others (Byers, 1978; Byers et al., 1982; Rogers and Williams, 1977). Combinations of ethephon plus carbaryl have over- and underthinned in some tests (Byers et al., 1982; unpublished data), but thinned adequately in others (Herrera-Aguirre and Unrath, 1980; Unrath, 1978).

Shading of apple or peach limbs or spraying trees with chemical photosynthetic inhibitors can induce fruit abscission (Byers et al., 1984, 1985; DelValle et al., 1985) without pygmy fruit development in 'Delicious' (Byers et al., 1985). Terbacil, but not shading, has caused leaf injury in some experiments, par-

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titularly when applied dilute with a hand-pump sprayer (Byers et al., 1984, 1985; DelValle et al., 1985).

The objectives of these experiments were: 1) to explore several chemical classes of photosynthetic inhibitors for apple thinning activity and degree of leaf injury, 2) to determine the most sensitive period when photosynthetic inhibition would cause apple thinning, and 3) to determine the effect of terbacil or shading on photosynthetic activity of apple leaves at rates that cause thinning.

Materials and Methods

Several studies were conducted in 1985 and 1986 on 5- and 6-year-old 'Redchief Delicious'/MM.111 trees located near Winchester, Va. These trees set a heavy crop in the fourth season and required much hand-thinning. All experiments were laid out in randomized complete-block designs. Each treatment was applied to whole, single-tree plots within six replicate blocks, except where indicated. Blocks were consecutively oriented within tree rows. Spray treatments were applied with a 5-liter stainless steel hand-pump to the point of drip. Full bloom occurred 19 Apr. 1985 and 22 Apr. 1986.

Experiments 1, 2, and 3. Twelve photosynthesis-inhibiting chemicals used in these experiments are listed in Table 1. In 1985, 11 of these inhibitors were applied to four trees each in a randomized block design (Expt. 1), but, because of rain, a second experiment (Expt. 2) was conducted to retest those materials that may not have had sufficient time to dry before the rain (propazine, dipropetryn, metribuzin, bentazon). In 1986, 12 inhibitors were applied as described above (Expt. 3), but rates were adjusted based on leaf injury and thinning responses

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observed in 1985. The surfactant X-77 at 0.0125% (v/v) was added to all treatments in both years. When significant injury occurred in an experiment, injury was rated from 0 to 10 (0 = no injury, 4 = heavy interveinal yellowing, 5 = interveinal necrotic leaves, 10 = complete defoliation and twig injury or death).

Experiment 4. Black polypropylene shade material (92% shade) (E.C. Geiger, Harleysville, Pa.) was used to enclose whole trees for 10-day periods in the intervals 5 to 15, 10 to 20, 15 to 25, 20 to 30, 25 to 35, 30 to 40, 35 to 45, and 47 to 57 DAFB. The shade material measured 90% shade on an overcast day, but, when placed at an angle to the sun draped over the tree, 95% shade was measured using a LI-COR Model LI-85 light meter with a quantum sensor (Lincoln, Neb.). Terbacil (50 $mg \cdot liter^{-1}$) + \hat{X} -77 (2500 mg \cdot liter⁻¹), applied at 5, 10, 15, 20, 25, 30, 35, or 40 DAFB, was also compared to shade, carbaryl (900 mg·liter⁻¹) + NAA (10 mg·liter⁻¹) + a nonphytotoxic, highly refined paraffinic, 70-sec superior oil (2500 mg·liter⁻¹), ethephon (1500 mg·liter⁻¹), hand-defruited, handdeblossomed, and hand-thinned trees. At harvest, 2.5-kg samples of fruit were collected from terbacil-sprayed trees 5, 15, 20, 25, or 30 DAFB and were analyzed for residues using standard liquid chromatographic methods described by Pease et al. (1978). Fruit per tree was counted between 30 and 61 DAFB and was expressed as fruit per square centimeter of trunk crosssectional area (TCSA). A 10-fruit sample was collected from each tree near harvest and sized with a band-type caliper. Fruit color was estimated as a percentage of fruit surface showing red and fruit firmness was determined with a Magness-Taylor penetrometer (D. Ballauf, Washington, D. C.) with an ll-mmdiameter tip. Soluble solids concentration (SSC) was determined by use of a Labeco refractometer (Laboratory Equipment Co., San Francisco) from a composite juice sample from the 10 fruits noted.

The average shoot length of the five longest terminal shoots (four scaffolds plus central leader) on each tree was measured the following dormant season and expressed as a percentage of the control. Trunk circumference of each tree was measured 30

cm above the soil on 1 May 1985 and in Dec. 1985. The increment in diameter for the 1985 season was calculated and expressed as a percentage of that of the control. Return bloom was rated from 0 to 10 (0 = no bloom, 4 = enough for full crop, 10 = all spurs flowering).

A 5-kg sample of fruit was collected from trees treated with 50 mg·liter $^{-1}$ terbacil + X-77 15 DAFB. Terbacil residue analysis of fruit collected at harvest was conducted according to Pease et al. (1978). Detection levels in fruit tissues were 0.01 mg·liter $^{-1}$, with a 67% recovery rate in controlled samples.

Experiment 5. Shade material (92%) was used to enclose six whole trees for either 1, 3, 5, or 7 days 13 to 14, 12 to 15, 11 to 16, and 10 to 17 DAFB, respectively. In addition, shade material was used to enclose six trees for 5 days 18 to 23, 23 to 28, 28 to 33, or 33 to 38 DAFB. Nonthinned and hand-thinned controls were also included for comparison. Fruit diameter, average shoot length, and trunk diameters were taken as in Expt. 4.

Experiment 6. Net photosynthesis (Pn) was measured in the field with a portable ADC (Analytical Development Co., supplied by P.K. Morgan Instruments, Andover, Mass.) LCA-2 infrared CO₂ analyzer (LCA-2) equipped with a Parkinson leaf chamber (P.K. Morgan Instruments) that exposed 6.25 cm² of leaf to sunlight. Only bright, cloudless days were chosen for Pn measurements, which were taken between 10:00 AM and 1:00 PM and consecutively by block. Photosynthesis of three leaves on each of two trees treated with terbacil 0, 2, 24, 72, or 120 hr previously were measured at 20 DAFB.

Experiment 7. Photosynthesis of trees that were shaded with polypropylene shadecloth 5 to 15 or 10 to 20 DAFB and of nonshaded trees was measured with shade either on or off the trees at 20 DAFB (three leaves on each of two trees for each treatment).

All data were averaged for each single-tree replicate before performing LSD (0 = 0.05), Duncan's multiple range procedure (0.05), or regression analysis. General Linear Model (GLM) procedures of the Statistical Analysis System (SAS) program package (SAS Institute, 1982) were used for analysis of variance.

Chemical class	Trade Name	Common name	Chemical name	Source
Substituted ureas	Tenoran	Chloroxuron	3-[p-(p-Chlorophenoxy)phenyl]-1,1-dimethylurea	Ciba-Geigy
	Cotoran	Fluometuron	1,1-Dimethyl-3-(α, α, α -trifluoro-m-tolyl)urea	Ciba-Geigy
	Bladex	Cyanazine	2-[[4-Chloro-6-(ethylamino)-s-triazine-2-yl]amino]- 2-methyl proprionitrile	Shell Development
Uracils	Sinbar	Terbacil	3-tert-butyl-s-chloro-6-methyluracil	DuPont
Substituted Trazines Chloroazine	Princep	Simazine	2-Chloro-4,6-bis(ethylamino)-s-triazine	Ciba-Geigy
	Milogard	Propazine	2-Chloro-4,6-bis(isopropylamino)-s-triazine	Ciba-Geigy
Meth-thio	Caparol	Prometryn	2,4-bis(isopropylamino)-6-(methylthio)-s-triazine	Ciba-Geigy
	Igram	Terbutryn	2-(tert-butylamino)-4-(ethylamino)-6-(methyl- thio)-s-triazine	Ciba-Geigy
Ethylthio	Sancap	Dipropetryn	2-(Ethylthio)-4,6-bis(isopropylamino)-s-trazine	Ciba-Geigy
Asymetrical	Sencor	Metribuzin	4-Amino-6-(1,1 dimethyl ethyl)-3-(methylthio- 1,2,4-as-triazine-5-(4H)-one	DuPont
Benzothiadiazol	Basagran	Bentazon	3-isopropyl-1 <i>H</i> -2,1,3-benzothiadiazin-4(3 <i>H</i>)-one-2,2-dioxide	BASF
Pyridazinon	Pyramin	Pyrazon	5-Amino-4-chloro-2-phenyl-3(2H)-pyridazinone	BASF

Table 1. Photosynthetic inhibitors of various classes tested for thinning apple fruits.

Table 2. Effect of photosynthetic inhibitors on fruit thinning and leaf injury of 'Redchief Delicious' apples.^z

<u></u>	1985 ^y				1986 ^y					
Inhibitor	Concn ^{y,x} (mg·liter ⁻¹)	cross-s		Injury rating (0–10)	Concn ^{x,w} (mg·liter)	Fruit/cm ² cross-sectional area of trunk ^v (FB + 51), Expt. 3	Injury rating (0–10) (FB + 23)	Injury rating (0–10) (FB +34)	Average shoot length ^u (% of control)	Trunk diameter increment ^u (% of control)
Control		4.7 a	2.9 a			6.0 a	0.0 a	0.0 a	100 abc	100 abc
Terbutryn	200	2.4 bc		2	100	1.0 ef	3.5 e	2.3 e	127 c	156 de
Terbacil	50	2.0 c		2	50	3.9 b	1.8 bc	0.0 a	113 bc	130 bcde
Simazine	50	5.1 a		0	2000	3.6 bc	0.2 a	0.0 a	111 bc	134 bcde
Prometryn	50	3.0 bc		2	100	1.6 def	2.3 cd	2.3 e	127 c	181 e
Chloroxuron	200	4.0 a		0	2000	2.1 de	1.2 b	1.2 bc	115 bc	120 bcd
Fluometuron	50	2.5 bc		0	100	2.1 de	2.8 de	0.0 a	122 c	157 de
Propazine	200	3.9 ab	2.5 ab	0	2000	2.2 de	0.3 a	1.0 b	100 abc	151 cde
Dipropetryn	100	4.0 ab	1.4 bc	0	4000	0.4 f	2.7 d	4.3 f	112 bc	132 bcde
Metribuzin	100	2.3 bc	0.7 c	0	200	2.3 de	2.8 de	0.0 a	128 c	172 de
Bantazon	200	5.3 a	1.6 c	0	400	2.5 cd	2.8 de	1.3 cd	121 c	163 de
Cyanazine	50	2.6 bc		0	100	2.6 cd	3.0 de	1.6 d	115 bc	133 bcde
Pyrazon					200	5.9 a	0.3 a	0.0 a	79 a	63 a
Hand-thinned						2.8 bcd	0.0 a	0.0 a	121 c	141 bcde

^zMean separation within columns by Duncan's multiple range test (P = 0.05).

^yFull bloom occurred 19 Apr. 1985 and 22 Apr. 1986.

*X-77 at 0.0125% (v/v) was added as a surfactant.

"Whole apple trees were sprayed with a hand-pump sprayer: Expt. 1, 2 May 1985 (FB + 13 days); Expt. 2, 6 May 1985 (FB + 17); and Expt. 3, 7 May 1986 (FB + 15). Fruit size at treatment was 6.5 ± 0.23 , 12.5 ± 0.47 , 8.8 ± 0.41 mm for each experiment, respectively.

*Fruit counts were made 17 June 1985 (FB + 59 days) on apples and 12 June 1986 (FB + 51 days).

"Average shoot length for five terminals (four scaffolds plus central leader) was 38.9 cm; mean trunk diameter increment for 1986 was 0.40 cm.

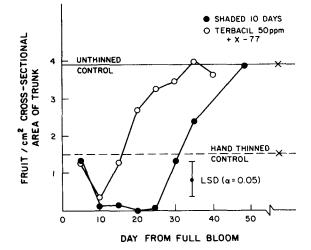


Fig. 1. Effect of terbacil sprays or shading of whole trees of 'Redchief Delicious' on fruit set. Each point represents six treated trees. Shaded trees were enclosed with 92% shade material for 10 days [the point was placed at the beginning date after full bloom (DAFB), i.e., at 5 days = shade (5 to 15 days)], sprayed with terbacil at 5 DAFB, hand-thinned, or unthinned controls. (Expt. 4, 1985). Vertical bar represents LSD, P = 0.05.

Results

In 1985, terbutryn, terbacil, prometryn, fluometuron, metribuzin, bentazon, and cyanazine caused significant fruit thinning (Table 2, Expts. 1 and 2). In 1986, all of the inhibitors except pyrazon caused thinning (Table 2, Expt. 3). Propazine, simazine, terbacil, fluometuron, and metribuzin seemed to cause less injury than other materials for the degree of thinning achieved. Where fruit thinning occurred, both shoot length and trunk diameter increased in spite of injury.

Experiment 4. In 1985, shading 5 to 15 DAFB was not as effective in reducing fruit set as shading 10 to 20, 15 to 25, 20 to 30, or 25 to 35 DAFB (Fig. 1). Shading trees 30 to 40 DAFB and later became progressively less effective in reducing fruit set and, by 47 to 57 DAFB, shading was completely ineffective.

Terbacil (50 mg·liter⁻¹) + X-77 (1250 mg·liter⁻¹) also caused fruit abscission during the same periods as shading, but shade caused more fruit abscission and for later periods than this particular concentration of terbacil (Fig. 1). Where crop loads were greatly reduced (terbacil at 10 DAFB), fruit size was increased, but, in general, shading or terbacil did not influence fruit size at harvest (Table 3). Since control trees were not excessively loaded with fruit, fruit size and the differences in tree growth between controls and thinned treatments were not as large as expected, although hand-thinning caused a significant increase in fruit size. Carbaryl + NAA + 70-sec oil did not defruit trees, but ethephon almost did. Ethephon-treated trees had fruit that were much smaller than expected, since their crop load was much lower than the hand-thinned treatment and fruit size of the hand-thinned trees was larger than the ethephon-treated trees. Furthermore, fruit size of the 50 mg·liter⁻¹ terbacil (10 DAFB) treatment was much larger than ethephon-treated fruit, although both treatments had similar crop loads. Average terminal shoot length was greatly reduced by ethephon (1500 mg·liter⁻¹), while return bloom was equivalent to defruited and deblossomed trees. Mean shoot length and TCSA of shaded and terbacil-treated trees were inversely related to crop load. Terminal shoot growth

Treatment ^z	Concn ^y (mg·liter ⁻¹)	Timing* (DAFB)	Fruit/cm ² cross-sectional area of trunk (FB + 30)	Fruit diameter (cm)	Average shoot length ^w (% of control)	Return bloom ^v (0–10)	Trunk diameter increment (1985) ^w (% of control)
Control			3.90	7.42	100	2.8	100
Hand-thinned		41	1.51	7.98	122	4.3	114
Shade (92%)		5-15	1.36	7.57	111	3.2	114
		10-20	0.10		122	3.2	118
		15-25	0.16		122	4.5	133
		20-30	0.00		129	4.7	137
		25-35	0.08		129	5.3	138
		30-40	1.29	7.54	116	4.0	106
		35-45	2.41	7.44	87	2.5	82
		47–57	3.86	7.57	89		79
Ferbacil	50	5	1.31	7.75	119	3.3	99
	50	10	0.36	8.13	124	3.0	108
	50	15	1.29	7.54	121	3.5	111
	25	15	3.13	7.52	101	3.2	93
	50	20	2.72	7.57	110	3.5	128
	50	25	3.26	7.65	106	3.8	93
	50	30	3.45	7.67	103	2.7	99
	50	35	4.00	7.75	103	3.0	124
	200	35	2.87	7.47	105	4.0	99
	50	40	3.61	7.42	92	3.2	104
Carbaryl +	900		1.74	7.92	129	6.8	125
NAA +	10						
70-sec oil	2500	15					
Ethephon	1500	15	0.09	7.24	33	9.7	89
Defruited		15	0.24	8.31	123	8.0	155
Deblossomed		5	0.08		129	9.0	156
LSD (P = 0.05)			0.88	0.37	18.25	2.4	45.05

Table 3. Effect of shading, terbacil, and other growth regulators on 'Redchief Delicious' apple fruit set, fruit set, fruit size, and tree growth (Expt. 4, 1985).

^zX-77 at 0.0125% (v/v) was added to terbacil treatments.

^yAll spray treatments were applied with a hand-pump sprayer. Fruit size was 8.95 ± 0.32 mm on 4 May (FB + 15 days). *Full bloom occurred 19 Apr. 1985.

"Average shoot length for five terminals (four scaffolds plus central leader) was 49.4 cm; trunk diameter increment was 0.765 cm for the controls.

*Return bloom was rated 17 Apr. 1986 at 25% bloom open (0 = no bloom; 5 = enough for full crop; 10 = all spurs flowering).

Table 4. Effect of length and time of shade on 'Redchief Delicious' fruit thinning and fruit size (Expt. 5, 1986).

Treatment	Duration of shade (days)	Timing ^z (DAFB)	Fruit/cm ² cross-sectional area of trunk (FB + 61)	Fruit diameter (FB + 122) (cm)	Average shoot length ^y (% of control)	Trunk diameter increment 1986 (% of control)
No shade			5.15	6.68	100	100
Shaded	1	13–14	4.31	6.78	106	107
	3	12-15	2.86	7.32	98	136
	5	11–16	1.92	7.52	109	163
	7	10–17	1.97	7.52	106	135
	5	18-23	0.70	7.52	124	178
	5	23–28	0.27	7.80	97	155
	5	28-33	0.60	7.37	124	182
	5	33-38	1.41	7.11	94	132
Hand-thinned		60	2.42	7.11	112	149
LSD (P = 0.05)			0.92	0.31	18	33

²Full bloom (FB) occurred 22 Apr. 1986. Fruit size was $8.2 \pm 0.4 \text{ mm} (+14 \text{ DAFB})$; $10.0 \pm 0.4 \text{ mm} (+17 \text{ DAFB})$; $15.2 \pm 0.9 \text{ mm} (+24 \text{ DAFB})$; $24.0 \pm 0.8 \text{ mm} (+33 \text{ DAFB})$; $30.0 \pm 0.6 \text{ mm} (+38 \text{ DAFB})$. ³Average shoot length for five terminals (four scaffolds plus central leader) was 46.9 cm; trunk diameter increment was 0.48 cm.

for the defruited terbacil treatments was equivalent to the control, hand-thinned, deblossomed, and defruited trees. Fruit color, firmness, and SSC were not affected by shade or terbacil. Residue analysis of fruit from terbacil (50 mg·liter $^{-1}$) + X-77 trees showed non-detectable levels of terbacil in the fruit. The legal tolerance set by the U.S. Environmental Protection Agency is

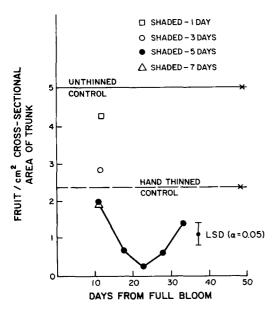


Fig.2. Effect of shading whole trees of 'Redchief Delicious' on fruit set. Each point represents fruit set on six shaded trees enclosed with 92% shade material for the period indicated, hand-thinned, and unthinned controls. [The point was placed at the beginning date of each shade period (Expt. 5, 1986).] Vertical bar represents LSD, P =0.05.

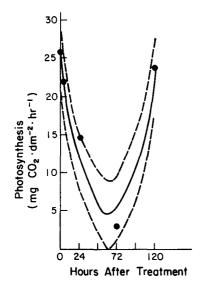


Fig. 3. Effect of terbacil on net photosynthesis of 'Redchief Delicious' trees sprayed at various times before measurement (y = 24.9 $- 0.6.57x + 0.005 x^2$) ($R^2 = 0.899$) (Expt. 6, 1986). Dotted lines represent confidence limit belts, P = 0.05.

currently $0.1 \text{ mg}\cdot\text{g}^{-1}$ fresh weight for terbacil on fruit when used as a herbicide in apple orchards.

Experiment 5. In 1986, shading trees 12 to 15 DAFB for 3 days caused fruit thinning, even though the most and about equally sensitive periods to shade were 18 to 23, 23 to 28, and 28 to 33 DAFB (Fig. 2, Table 4). Fruit sizes on nonshaded trees in the 1986 experiments were 8.2 ± 0.4 mm (14 DAFB); 10.0 \pm 0.4 mm (17 DAFB); 15.2 \pm 0.9 mm (24 DAFB); 24.0 \pm 0.8 mm (33 DAFB); 30.0 \pm 0.6 (38 DAFB); and 33.5 \pm 1.2 mm (41 DAFB). These data are interesting because fruit at the normal thinning stage (10 mm, 7 DAFB), as well as larger

Experiment 6. Twenty days after full bloom, Pn of trees treated with terbacil (50 mg·liter $^{-1}$) + X-77 (1250 mg·liter $^{-1}$) 72 hr before measurement was reduced to =10% of nontreated trees, but trees treated 120 hr before measurement were photosynthesizing at near-normal levels (Fig. 3).

Experiment 7. Leaf Pn measured 20 DAFB on trees shaded 5 to 15 DAFB was less than the control, and some leaves showed obvious red pigmentation (Table 5). Shade removal after 10 days may have caused light-induced destruction of chlorophyll for the subsequent 5 days, possibly leading to the visible expression of redness of some leaves. Before shade removal, the Pn of trees shaded 10 to 20 DAFB was equal to that of trees shaded on the 20th DAFB, about one-third of the control. After removal, the Pn of trees shaded 10 to 20 DAFB was similar to that of the control.

Discussion

Fruit abscission caused by shading or terbacil treatment suggests a brief period after bloom when the fruits are extremely sensitive to the application of a photosynthetic inhibitor or a Imitation of light by shading. However, the sensitive period for shade-thinning apparently extends well-past the 10-mm-diameter stage of fruit development; i.e., past the period when fruit are most susceptible to "thinning by growth-regulator and carbamate chemical thinning agents.

All of the Pn inhibitors tested in these studies were originally selected for their persistence as herbicides. Other Pn inhibitors that are more short-lived may have potential as thinning agents, with less risk of over-thinning and leaf injury. Many of the persistent Pn inhibitors, such as terbacil, are normally used as root-absorbed herbicides and may not be appropriate for foliar application. Since all of the Pn inhibitors from the five classes tested were active as thinning agents, more-suitable thinning agents may be found among other Pn inhibitors.

Shading limbs of 20-year-old 'Starkrimson' trees in a previous study (Byers et al., 1985) indicated that fewer fruit abscised from shading 10 days (16 to 26 DAFB) than shading younger, whole trees of 'Redchief' in these experiments. When limbs in the previous studies (Byers et al., 1985) were shaded 26 to 36 DAFB, no thinning occurred, but whole trees were almost completely defruited when shaded 25 to 35 DAFB. These results suggest that shading whole trees was more effective than shading limbs on old trees. We believe limb experimental units were less-responsive to shading treatments than whole 'Redchief' trees and a significant compensatory effect is likely in limb experiments. However, the whole-tree shading experiment was on much younger trees than the limb-shading experiment (Byers et al., 1985). An additional experiment would be required to test the compensatory effect when single limbs are shaded.

Since photosynthetic inhibitors or short periods of shading can dramatically reduce set, we suspect that cloudy periods as short as 3 days, or even less, may greatly affect fruit set under natural conditions. The combined effect of a chemical thinner

Table 5.	Effect of	of shading a	and remov	al of shad	e material	on photosynthes	s of 'Red	1-
chief D	elicious'	apple leav	es (Expt.	7, 1985). ²				

Treatment ^y	Photosynthesis ^x (mg CO ₂ /dm ² per hr) (FB + 20)	Leaf color
No shade	26 a	Green
Shade $FB + 5$ to $+ 15$ (shade off $FB + 15$)	17 b	Red leaf pigments prominent
Shade $FB + 10$ to 20 (shade on $FB + 20$)	8 c	Green
Shade $FB + 10$ to $+ 20$ (shade off $FB + 20$)	23 a	Green
Shade $FB + 20$ (shade on $FB + 20$)	7 c	Green

²Mean separation in columns by Duncan's multiple range test (P = 0.05). ^yFull bloom on 19 Apr. 1985.

*Photosynthesis at 20 DAFB taken in the field on three leaves on each of two replicate trees per treatment.

and environmental shading should be extensively investigated. Our measurements of photosynthetic photon flux (PPF) levels show that PPF at Winchester was reduced by 85% to 90% of full sun on a typical cloudy day, and during rainy days it was even less. An understanding of the interactions between cloudy weather, fruit set, chemical thinning application methods, and stage of fruit physiological development is critical for reproducible thinning results.

Apple fruit abscission after fertilization and during June drop is considered to result from the competition for essential metabolites among individual fruitlets, and between fruitlets and vegetative shoots (Abbott, 1960; Quinlan and Preston, 1971; Wardlaw, 1968). Schneider and Lasheen (1973) and Schneider (1975, 1977) showed that NAA thinning sprays decreased the amount of reducing sugars in young apple fruitlets. Weinbaum and Simons (1974) also showed reduced starch deposition in seed tissue; this reduction was correlated with impending seed abortion in NAA-treated apples. Apparently, the first effect of hormone thinners is a reduced level of photosynthate reaching the developing fruit. Second, the most sensitive period for NAAinduced abscission in apples is 10 to 20 DAFB. Our data show that chemical photosynthetic-inhibitor activity and natural June drop are initiated at about the same critical period as hormone spray-thinning. Shade-thinning appears to be effective for a longer period than terbacil, NAA, or carbaryl applications. The mechanism leading to June drop or NAA-, carbaryl-, or ethyleneinduced abscission may be the same as that caused by photosynthetic inhibition, but absorption of chemicals as leaves age may limit their effect later in the season.

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