NDT Perspectives



Are there better alternatives than haemoglobin A1c to estimate glycaemic control in the chronic kidney disease population?

Marijn Speeckaert¹, Wim Van Biesen^{1,2}, Joris Delanghe³, Robbert Slingerland^{4,5}, Andrej Wiecek⁶, James Heaf⁷, Christiane Drechsler⁸, Raluca Lacatus⁹, Raymond Vanholder¹ and Ionut Nistor^{2,9} for the European Renal Best Practice Guideline Development Group on Diabetes in Advanced CKD

¹Renal Division, Department of Internal Medicine, Ghent University Hospital, Ghent, Belgium, ²Methods Support Team ERBP, Renal Division, Ghent University Hospital, Ghent, Belgium, ³Department of Clinical Chemistry, Ghent University Hospital, Ghent, Belgium,
⁴Department of Clinical Chemistry, Isala Klinieken, Zwolle, The Netherlands, ⁵European Reference Laboratory for Glycohemoglobin,
Zwolle, The Netherlands, ⁶Department of Nephrology, Endocrinology and Metabolic Diseases, Medical University of Silesia, Katowice, Poland,
⁷Department of Nephrology B, Herlev Hospital, University of Copenhagen, Denmark, ⁸Division of Nephrology, Department of Medicine,
University of Würzburg, Würzburg, Germany and ⁹Department of Nephrology, Gr. T. Popa University of Medicine and Pharmacy, Iasi,
Romania

Correspondence and offprint requests to: Wim Van Biesen. E-mail: guidelines@era-edta.org

ABSTRACT

Background. Although measurement of haemoglobin A1c has become the cornerstone for diagnosing diabetes mellitus in routine clinical practice, the role of this biomarker in reflecting long-term glycaemic control in patients with chronic kidney disease has been questioned.

Methods. Consensus review paper based on narrative literature review.

Results. As a different association between glycaemic control and morbidity/mortality might be observed in patients with and without renal insufficiency, the European Renal Best Practice, the official guideline body of the European Renal Association-European Dialysis and Transplant Association, presents the current knowledge and evidence of the use of alternative glycaemic markers (glycated albumin, fructosamine, 1,5-anhydroglucitol and continuous glucose monitoring).

Conclusion. Although reference values of HbA1C might be different in patients with chronic kidney disease, it still remains the cornerstone as follow-up of longer term glycaemic control, as most clinical trials have used it as reference.

Keywords: diabetes, chronic kidney disease, glycaemic control, guideline

INTRODUCTION

Diabetes mellitus is the leading cause of chronic kidney disease (CKD) and is associated with an excessive (cardiovascular) morbidity and mortality [1]. Diabetic nephropathy is diagnosed in 20-40% of patients with type 1 or type 2 diabetes [2] and accounts for 30-50% of end-stage renal disease (ESRD) cases [3]. Although hyperglycaemia is the biochemical hallmark of diabetes, haemoglobin A1c (HbA1c) measurement has slowly become the cornerstone for diagnosing diabetes mellitus since its introduction in routine clinical practice in 1976 [4, 5]. For the diagnosis of diabetes, the normal range cut-off point is 48 mmol/mol (6.5%) [6]. However, some authors suggest that population-specific optimum cut-off points may be necessary in the future as an HbA1c-based diagnosis has substantially different consequences for diabetes prevalence across ethnic groups and populations [7, 8]. Differences in intracellularextracellular glucose balance, differences in red cell survival (e.g. haemolytic anaemia) and non-glycaemic genetic determinants of haemoglobin glycation are possible contributing factors of the racial and ethnic differences. For that reason, reliance on HbA1c as the sole criterion for the diagnosis of diabetes in non-Caucasians could lead to misclassification [8]. In addition to its recent role as a diagnostic marker, HbA1c is used in the assessment of the degree of metabolic control in diabetic patients and in risk prediction of vascular complications. Although one measurement gives information for the diagnosis of diabetes, HbA1c is mainly considered as a longitudinal parameter, allowing guidance of treatment in the longer term [9].

There are many advantages of using HbA1c rather than blood glucose for screening and diagnosing diabetes: less sensitivity to pre-analytical variables, lower within-subject biological variability, little/no diurnal variations and little/no influence from acute stress [10]. In contrast, the underlying challenge of HbA1c remains 2-fold: (i) accurately reflect the mean plasma glucose levels within a longer time span, taking into account different parameters such as age, ethnicity, geography, pregnancy and underlying disease [11] and (ii) accurately relate the degree of glycaemic control to important outcomes, such as death and diabetes associated morbidity.

There is conflicting evidence regarding the role of HbA1c in reflecting long-term glycaemic control in CKD patients [10, 12, 13]. In addition, the association between glycaemic control and outcome might be different in CKD versus no CKD patients. Given the unique conditions associated with the uraemic environment, there is thus a need to evaluate markers to monitor glycaemic control specifically in the CKD population.

HBA1C: BIOCHEMISTRY, METHODOLOGY AND VARIABILITY

After discovery of the heterogeneity of Hb by the deviating migration speed of sickle cell Hb in an electrical field, five sub-fractions were identified in 1958 [14]. Hb consists of ~97% adult Hb (HbA), 2.5% HbA2 and 0.5% fetal Hb (HbF). In healthy individuals, ~6% of HbA is glycated. Glycated Hb consists of HbA1a, HbA1b and HbA1c [15, 16]. In 1969, Rahbar *et al.* demonstrated elevated fast Hbs in erythrocytes of diabetes patients [17]. Trivelli *et al.* suggested a relationship between fast Hbs, mean blood glucose concentrations and long-term complications in diabetes patients [18].

Being the major form of all glycohaemoglobin species in human blood, HbA1c has been defined as the result of a nonenzymatic reaction (classical Maillard reaction) of condensation between the aldehydic group of glucose and the N-terminal amino group of the β -chain of HbA0 [*N*-(1-deoxyfructosyl) Hb] [19]. To compensate for intra- and interindividual variation in the total Hb concentration, HbA1c has been expressed as a ratio (HbA1c/total Hb) [20]. Approximately 50% of a given HbA1c value is the result of glucose exposure during the previous 30 days and 40 and 10% is the result of glucose exposure during the previous 31–90 days and 91–120 days respectively. HbA1c is neither considered dysfunctional nor harmful [21].

Different analytic systems have been developed for measurement of HbA1c, either based on difference in charge (ion-exchange chromatography, electrophoresis, capillary electrophoresis and isoelectric focusing) or structural difference (affinity chromatography, immunochemical assays and enzymatic assays). Before the 1980s, methods based on a subtle difference in iso-electric point suffered interferences from other members of the Hb family (e.g. Schiff base, carbamylated Hb and variants) and had to deal with the dominating (20-fold concentration) parent HbA0. New automated highperformance liquid chromatography (HPLC) systems (used in $\pm 60\%$ of the laboratories in Europe) provide reliable results without interference by Schiff base or carbamylated Hb. Immunochemical assays ($\pm 35\%$ of users) use antibodies against the β N-terminal glycated tetrapeptide or hexapeptide group or variation of this, but have difficulties in achieving a coefficient of variation of <2% [20]. Finally, new enzymatic tests were developed in the 2000s [22].

Due to the wide range of methods used, each with their own definition of the analyte (e.g. HbA1c, fast Hbs or total Hb) and specificity (e.g. HbF, carbamylated Hb or incomplete separation), standardization became an important topic in the 1990s. Based on the described definition of HbA1c, the International Federation of Clinical Chemistry (IFCC) Working Group on Standardization of HbA1c developed a reference measurement system for HbA1c [19, 23] by implementation of two equivalent reference methods (HPLC/mass spectrometry and HPLC/capillary electrophoresis), characterization of primary and secondary calibrators and organization of an international network of laboratories performing one or both reference procedures. Pure HbA1c and pure HbA0 were isolated from human blood and mixed in well-defined proportions to a certified primary reference material set used to calibrate the primary reference measurement system (PRMS) [24]. The results obtained on routine samples by clinical laboratories using the aligned analytical systems were traceable to the reference measurement system, obtaining standardization of HbA1c measurement. At this moment, the IFCC reference system is the only valid analytic anchor from which all other units in which HbA1c might be expressed are derived. HbA1c results could be reported in both IFCC (mmol/mol) and derived U.S. National Glycohaemoglobin Standardization Program (NGSP) (%) numbers [25, 26]. The NGSP was created to harmonize HbA1c results through the implementation of assay traceability to the ion-exchange HPLC method [27, 28] without providing a stable scientificbased anchor, originally used in the Diabetes Control and Complications Trial [29].

Unfortunately, all the main data of the trials supporting the clinical use of HbA1c have used assays aligned to the U.S. NGSP, resulting in a less specific method as approximately one-third of the chromatographic component denoted as HbA1c is not HbA1c. The conversion of analytical and clinical data from the NGSP system to the IFCC system has become possible by the so-called IFCC-NGSP 'master equation' [NGSP (%) = $0.09148 \times IFCC$ (mmol/mol) + 2.152] [28, 30, 31].

In addition, the use of a bedside HbA1c point-of-care testing (POCT) seems attractive, but at this moment, the reliability and performance level of this approach may be questioned [32–34]. The conclusion of a systematic review and meta-analysis stated that there is insufficient evidence to date for the effectiveness of HbA1c POCT in the management of diabetes [35].

NDT PERSPECTIVES

FACTORS INFLUENCING HBA1C VALUES BESIDES GLYCAEMIA

Erythropoiesis and red cell lifespan heterogeneity

The formation of HbA1c is mainly dependent on the interaction (intensity and duration) between blood glucose concentrations and red blood cells (RBCs). On average, erythrocytes survive 117 days in men and 106 days in women. At any given time, a blood sample contains erythrocytes of different ages with a predominance of younger elements and with different degrees of exposure to hyperglycaemia [36]. An unexplained discordance between HbA1c and other measures of glycaemic control can be partly the result of differences in red cell lifespan [37]. Larger longitudinal studies should be

carried out to confirm if the observed variation in RBC survival might lead to inappropriate clinical decision-making [38] (Table 1).

A decreased erythropoiesis, due to iron or vitamin B12 deficiency or aplastic anaemia, leads to an increase in the number of circulating aged red cells and accordingly in a progressive rise in HbA1c not related to glycaemic control [39]. Iron deficiency anaemia causes an increase in HbA1c of up to 2%, which can be reversed with iron supplementation [40]. On the contrary, a decrease in HbA1c is observed after administration of erythropoietin, iron and vitamin B12 and in the case of haemolytic anaemia/reticulocytosis. Due to a reduced red cell survival, younger erythrocytes have less time exposure to ambient glycaemia and thus less glycation [41, 42].

Table 1. Comparison of the different glycaemic markers in diabetic p	patients with chronic renal failure
--	-------------------------------------

Marker	Advantages	Disadvantages
HbA1c Marker of long-term glycaemic concentra Excellent standardization of HbA1c assay Universally available PRMS. Scientific evidence on association with ou	Marker of long-term glycaemic concentrations. Excellent standardization of HbA1c assays. Universally available PRMS. Scientific evidence on association with outcomes from several	Falsely increased values with iron deficiency, vitamin B12 deficiency, decreased erythropoiesis, alcoholism, chronic renal failure, decreased erythrocyte pH, increased erythrocyte lifespan, splenectomy, hyperbilirubinaemia, carbamylated haemoglobin, alcoholism, intake of large doses of aspirin, chronic opiate use.
	trials. In comparison with blood glucose, less sensitivity to pre- analytical variables, lower within-subject biological variability, little/no diurnal variations, little/no influence from acute stress and little/no influence from common drugs which are known to influence glucose metabolism. Excellent separation of the HbA1c fraction from other	Falsely decreased values have been reported after administration of erythropoietin, iron or vitamin B12; with reticulocytosis, chronic liver disease, ingestion of aspirin, vitamin C, vitamin E, certain haemoglobinopathies, increased erythrocyte pH, a decreased erythrocyte lifespan, haemoglobinopathies, splenomegaly, rheumatoid arthritis, drugs such as antiretrovirals, ribavirin and dapsone, hypertriglyceridaemia.
	haemoglobin adducts and with no interference from carbamylated haemoglobin due to technological advances in HbA1c measurement.	Variable changes have been seen in patients with HbF, haemoglobinopathies, methaemoglobin, genetic determinants.
Glycated albumin	Measure of short-term glycaemic control (2–3 weeks). Not influenced by gender, erythrocyte lifespan, erythropoietin therapy or serum albumin concentration. Significant association with markers of vascular injury.	Values can be influenced by lipaemia, hyperbilirubinaemia, haemolysis, increased uric acid, uraemia, intake of high doses of aspirin, low serum protein concentrations/nutritional status, age, albuminuria, cirrhosis, thyroid dysfunction and smoking.
		Concentration is inversely influenced by body mass index, body fat mass and visceral adipose tissue.
		Different reference ranges depending on the applied method.
		Limited data, especially on the impact of using it as a target.
Fructosamine	Correlates with average glucose levels in the previous 10–14 days. Simple, automated analysis.	Expensive, time-consuming, not widely available. Contradictory results concerning the correlation between fructosamine and mean glucose concentrations in patients with renal failure.
		Values can be influenced by nephrotic syndrome, thyroid dysfunction, glucocorticoid administration, liver cirrhosis, icterus.
		Concentration in uraemic patients may be influenced by a number of variables other than glycaemia, including hypoalbuminaemia, hyperuricaemia.
1,5Anhydroglucitol	Reflects day-to-day changes in glucose levels.	Within-subject variation is higher than that for HbA1c. Poorer performance in identifying cases of undiagnosed diabetes in comparison to other slycasmic markers
	Retained metabolic inertness, steady-state levels in all tissues and negligible influence of sampling conditions such as collection time, body weight, age, sex and food intake of the subjects.	Influenced by traditional Chinese herbal drugs
		Limitations for use in subjects with renal tubular acidosis, advanced or ESRD.
Continuous glucose measurement		Not widely available, limited data on its clinical everyday value.
	Theoretically the most ideal marker for glycaemic control.	Exhaustion of the sensor, limited data.
	Allows examination of short-term glycaemic changes around the time of dialysis.	

Abnormally high HbA1c values have been reported in patients after a splenectomy due to an increased circulating erythrocyte lifespan. Splenomegaly, acute or chronic major blood loss, glucose-6-phosphate dehydrogenase deficiency, intensive physical activity (especially in marathon runners), rheumatoid arthritis or drugs (antiretrovirals and ribavirin) could falsely reduce the level of HbA1c even in the presence of high ambient plasma glucose [39].

Altered Hb

In the presence of haemoglobinopathies (e.g. sickle cell anaemia and thalassaemias), the correct interpretation of the measured HbA1c can be difficult. Besides the normal phenomenon (glycation of adult HbA0 to form HbA1c), other glycated products derived from HbC (African populations), HbD (Indian populations), HbE (Asian populations) or HbS (sickle cell disease) are formed in addition to or instead of HbA1c [39]. In the past, persistence of HbF leads to an overestimation of the HbA1c levels due to co-migration or co-elution with the HbA1c fraction [43-45]. However, no interference of HbF with the IFCC Reference Method has been reported, which can be explained by the absence of β chains in HbF. Only the HbA terminal hexapeptides are measured with the IFCC Reference Method [46]. Besides the genetic Hb variants, chemical alterations could also influence HbA1c results. By increasing methaemoglobin levels and decreasing erythrocyte survival, dapsone can artefactually lower HbA1c [47].

Glycation

NDT PERSPECTIVE

In the Third National Health and Nutrition Examination Survey, alcohol consumption was associated with lower HbA1c levels among 1024 adults with diabetes [48]. Those findings were confirmed in a large follow-up study of 38 564 adult patients with type 1 or 2 diabetes (Kaiser Permanente Northern California members, 1994–1997). Increasing levels of alcohol consumption predicted lower HbA1c values through a nadir at a consumption of 2–2.9 drinks/day [49]. pH levels within the erythrocyte can increase (low erythrocyte pH) or decrease (high erythrocyte pH) HbA1c. In chronic renal failure, lipid peroxidation of Hb may increase Hb glycation [50]. Chronic ingestion of aspirin and high doses of antioxidants (e.g. vitamins C and E) can lower HbA1c due to inhibition of Hb glycation [51]. It is unclear whether these phenomena could lead to a different appreciation of HbA1c in clinical practice.

Assays

Decreased HbA1c values have been reported in lipaemic blood samples. Due to a turbidity effect of triglyceride which increases the absorbance of the total Hb and unbound Hb fractions, some assays reported a decreased calculated percentage of glycated Hb [52]. Some other well-documented causes, depending on the assay used, for elevated HbA1c include hyperbilirubinaemia, carbamylated Hb and chronic opiate use [39].

HBA1C AS A MARKER OF GLYCAEMIC CONTROL IN CKD PATIENTS

In contrast to plasma glucose, HbA1c represents non-enzymatic glycosylation, which depends on the glucose concentration in the intra-erythrocyte compartment [53]. Although multiple studies found a good (positive) correlation between HbA1c and glucose concentrations in diabetic non-CKD patients [54] and CKD patients [55], the variable relationship between HbA1c and estimated average glucose (eAG) remains a potential source of concern [56], which can be partly explained by the within-subject variability in degree of Hb glycation. This 'glycation gap' is for ~70% genetically predetermined [57]. In addition, measurement of glucose has also been a major concern due to point and trend accuracy, sensitivity and specificity, device stability, calibration, lag time and traceability to the highest standard.

Falsely decreased HbA1c values

HbA1c readings can be falsely low in patients on either form of dialysis (haemodialysis or peritoneal dialysis), questioning the accuracy of the HbA1c assay in diabetic patients with severely reduced renal function [58]. Besides the average glucose concentrations, HbA1c is also determined by lifespan of RBCs [59, 60], the use of recombinant human erythropoietin [61], intravenous iron replacement treatment [62], the uraemic environment itself, blood pH and blood transfusions. Iron replacement therapy and erythropoietin-stimulating agents result in a fall in HbA1c, independent of glycaemic changes [58, 62]. Some caution in the interpretation of HbA1c alone with regard to glycaemia management is thus warranted.

Falsely increased HbA1c values

The production of carbamylated Hb depends on the duration and severity of renal failure. Carbamylated Hb is formed by non-enzymatic condensation of cyanate with the N-terminal valine of Hb [63]. Carbamylation is a physiologic process that can alter protein structure and function, inducing significant pathophysiological perturbations [64]. Previous studies described a clinically relevant overestimation of glycated Hb by chromatography, but not by immunochemical measurement, which can be attributed predominantly to incomplete separation of the carbamylated Hb fraction and the HbA1c fraction [65, 66]. Technological advances in HbA1c measurement (e.g. newer ion-exchange HPLC assay methods, specific immunoassays or affinity chromatography) showed however an improved separation of the HbA1c fraction from other Hb adducts with no interference from carbamylated Hb [67, 68], thus allowing a correction for carbamylated Hb interference by simply subtracting carbamylated Hb from the measured HbA1c value.

Debate on the pros and cons of using HbA1c in CKD patients

The relationship between HbA1c and glucose is in advanced CKD more complex because of a wide variability in Hb, poor nutritional status and inflammation [69]. In addition, these underlying comorbidities might also hamper the prognostic value of HbA1c. Current guidelines recommend HbA1c as the preferred biomarker of glycaemic control in CKD patients with a target of <53 mmol/mol (7.0%) to prevent or delay progression of the microvascular complications of diabetes, including diabetic nephropathy [70]. However, these guidelines mostly focus on early stages of CKD. In diabetic patients with advanced CKD, it is suggested that aiming at a too intensive glycaemic control [HbA1c level <48 mmol/mol (6.5%)] may be associated with increased mortality [71]. Similar to diabetes patients without CKD [72], observational cohort studies of diabetic patients with advanced CKD, peritoneal dialysis or haemodialysis demonstrate a U- or J-shaped curve of HbA1c versus mortality [71, 73, 74]. However, after adjustment for potential confounding factors (demographics, dialysis vintage, dose, comorbidity, anaemia and surrogates of malnutrition and inflammation), patients with an HbA1c > 86 mmol/mol (10%) had all-cause and cardiovascular death a hazard ratio of 1.41 (95% CI: 1.25-1.60) in comparison to an HbA1c in the 31-42 mmol/mol (5-6%) range (P < 0.001) [75]. Similarly, the hazard ratio for cardiovascular death was significantly increased to 1.73 (95% CI: 1.44-2.08). In addition, an inferior survival was reported in diabetic haemodialysis and peritoneal dialysis patients with extremes of glycaemia [76]. The association between high HbA1c values [≥64 mmol/mol (8%)] and all-cause mortality was particularly robust in individuals with higher Hb levels (>11 g/dL) [74]. Subgroup analyses showed that the HbA1c threshold for higher all-cause mortality was lower [HbA1c \geq 53 mmol/mol (7%)] in Caucasians, men and patients with albumin level of >3.8 g/dL. These findings may illustrate the possible interaction of factors related to protein-energy wasting, inflammation and anaemia with indices of glycaemic control [77]. In a large-scale and contemporary cohort of 54757 diabetic maintenance haemodialysis patients, a time-averaged HbA1c of >64 mmol/mol (8%) or time-averaged serum glucose of >200 mg/dL appeared to be associated with a higher all-cause and cardiovascular mortality [74]. Other small studies have reported that a poor glycaemic control is a predictor of cardiovascular morbidity and mortality for type 2 diabetics with advanced CKD [78, 79]. In 2872 kidney transplant recipients, poor glycaemic control [HbA1c > 64 mmol/mol (8%)] during the preceding haemodialysis period appeared to be associated with higher all-cause and cardiovascular mortality [80]. A recent meta-analysis, investigating the relationship between HbA1c and risk of death in diabetic haemodialysis patients, showed that the HbA1c level remains a useful clinical tool in predicting mortality risk. In this study consisting of nine observational studies [12, 74, 76, 81-86] and one secondary analysis [87] of a randomized trial (n = 83684 participants), baseline HbA1c levels of >69 mmol/mol (8.5%) were associated with a 29% increase in the adjusted risk of death compared with the reference group with HbA1c levels of 48-57 mmol/mol (6.5-7.4%). Mean HbA1c levels <36 mmol/mol (5.4%) were associated with a small, but non-significant increase in mortality, which could be explained by the heterogeneity of this subgroup. There was a similar association between mean HbA1c level and mortality risk in both incident and prevalent patients. Based on their findings, the authors proposed an HbA1c

target of <69 mmol/mol (8.5%) in diabetic haemodialysis patients [88].

Other observational studies do not confirm the link between HbA1c values and survival in ESRD patients [75, 84, 89–91]. In a cohort study of 24 875 haemodialysis patients with type 1 or 2 diabetes mellitus, only a weak correlation with mean random glucose values and HbA1c and no correlation between HbA1c and subsequent 12-month mortality risk was observed [89]. This study was criticized however for a short follow-up period, non-time-dependent survival models and lack of stratified analyses [92]. Also in peritoneal dialysis patients, baseline and time-averaged follow-up HbA1c did not correlate with patient and peritoneal dialysis technique survival [90]. Of note, the study only included 91 patients, which may have been far too few to reach a statistical significance.

Despite the evidence showing an association between HbA1c and outcomes, several trials did not show a benefit of targeting lower HbA1c values [93], probably because the potential advantages (prevention from diabetes associated comorbidity) might not outweigh the disadvantages (risk of hypoglycaemia) in advanced CKD patients. Physicians are encouraged to individualize glycaemic targets based on potential risks and benefits in diabetic ESRD patients [76, 77, 94].

ARE THERE BETTER ALTERNATIVE MARKERS OF GLYCAEMIC CONTROL?

Glycated albumin

Glycated albumin is gaining interest as a potential marker of glycaemic control [95]. Glycated albumin is a ketoamine formed from a non-enzymatic oxidation of albumin by glucose [96]. As the half-life of albumin is \sim 15 days, glycated albumin is a measure of short-term glycaemic control (2–3 weeks) [97] and as such, it could act as an intermediate-term index of glycaemic control (Table 1).

Analytical methods. Boronate-affinity chromatography (followed by tandem mass spectrometric detection), ionexchange chromatography, HPLC, immunoassay-related techniques (e.g. enzyme-linked immunosorbent assays or radio-immunoassays), Raman spectroscopy, refractive index measurements, capillary electrophoresis and other electrophoretic and enzymatic assays (e.g. ketoamine oxidase) can be used for measuring the glycated albumin concentration. This involves calculation of the glycated albumin peak area to the total albumin peak area [95, 98, 99]. A method for conversion between HbA1c and glycated albumin using a measurement error model has been published [100].

Measurement of glycated albumin is not influenced by gender, erythrocyte lifespan and erythropoietin therapy; for serum albumin concentration, conflicting results are reported [58, 101–103]. However, results can be impacted by age, nutritional status [104], albuminuria, cirrhosis, thyroid dysfunction and smoking. Glycated albumin is inversely influenced by body mass index, body fat mass and visceral adipose tissue [105, 106]. Accelerated albumin catabolism accompanied by chronic micro-inflammation, which occurs for example in the

Glycated albumin versus HbA1c. It has been suggested that the relationship between HbA1c and serum glucose concentration is altered as the GFR declines in diabetic subjects with advanced CKD, whereas the glycated albumin assay is not impacted by Stage 3 (after transplantation) or Stage 4 CKD [109]. However, this study had several important limitations. In patients with diabetic nephropathy (CKD Stage 3 or 4) and overt proteinuria, glycated albumin values may be lower relative to plasma glucose levels as a result of an increased turnover in albumin metabolism [96]. As albuminuria typically falls with decreasing glomerular filtration rates, this effect might be mitigated in dialysis patients [110].

A better correlation between glycated albumin and glycaemic status (measured by casual plasma glucose or average blood glucose level) has been reported in patients on haemodialysis or peritoneal dialysis in comparison to HbA1c [58, 102, 109, 111, 112]. Opponents argue that ESRD is characterized by an abnormal albumin homeostasis and that the serum albumin threshold at which risk of death increases varies by dialysis modality [113–115]. In hypoalbuminaemia, plasma protein glycation is increased [116]. However, glycated albumin seems to reflect the percentage of albumin that is glycated regardless of the total serum albumin concentration [110], although further large-scale studies with dialysis patients are needed to substantiate this observation.

Glycated albumin apparently has a better association with different parameters of microvascular (kidney disease, retinopathy) and macrovascular disease (pulse wave velocity) as compared with HbA1c [103, 117–119] and also with mortality [82, 120, 121]. New prospective studies are necessary to provide evidence that improving glycaemic control ameliorates glycated albumin and decreases mortality, micro- and macro-angiopathy.

Glycated albumin appears to be superior in accuracy as a marker of glycaemic control compared with HbA1c in patients with diabetic nephropathy [122]. However, given the limited data, the absence of interventional outcome studies based on glycated albumin and the expensive and laborious methodology, it seems premature to abandon HbA1c in favour of glycated albumin [91, 123].

Fructosamine

NDT PERSPECTIVE

Fructosamine (1-amino-1-deoxy-D-fructose) represents a clinically accessible measure of non-enzymatic glycation of proteins in the same compartment as plasma glucose and should integrate plasma glucose fluctuations [53]. It is formed when the carbonyl group of glucose reacts with an amino group of circulating serum proteins and is a measure of serum ketoamines.

Analytical methods. Several assays have been designed to quantify fructosamine: boronate-affinity chromatography, phenylhydrazine procedure, furosine procedure, colorimetric

methods and enzymatic methods [124-126]. The fructosamine level correlates best with average glucose levels in the previous 10-14 days [127]. Being a measure of total glycated serum proteins with glycated albumin accounting for ~90% of these proteins, fructosamine concentrations may be influenced by serum protein concentrations and profile of different proteins [128]. Simply correcting for total protein may not accurately compensate for variations in protein half-life and reaction to serum glucose concentrations [129]. In addition, fructosamine is influenced by the concentration of bilirubin and low-molecular-weight substances (e.g. urea and uric acid) coexisting in the plasma [130]. Fructosamine is not altered by disorders of Hb metabolism, but is affected by disorders in protein turnover, such as dysproteinaemias [131]. Serum fructosamine concentration may also be determined by some reducing activities caused by unknown factors other than glycated proteins [132]. Reference values depend upon age, gender, sample population and applied test method.

Fructosamine versus HbA1c. Contradictory results have been reported with respect to the correlation between fructosamine and mean glucose concentrations in patients with renal failure [55, 56, 133–136]. The relationship between the fructosamine level and glycaemic control was good in type 2 diabetic patients with CKD Stages 3–4. However, calculation of eAG from fructosamine level may underestimate mean blood glucose levels in those patients [56].

Consistent discordances between HbA1c and fructosamine have been reported, which has been called the previous mentioned 'glycation' gap, defined as actual HbA1c minus HbA1c predicted from fructosamine [53, 137]. In a study of 23 diabetic haemodialysis patients, HbA1c appeared to correlate most accurately with measured blood glucose, whereas fructosamine and glycated plasma proteins correlated poorly with glycaemic control [55]. Fructosamine is considered a reliable marker of medium-term integrated blood glucose in diabetics on maintenance haemodialysis by some [133], but not by others [135, 136]. In a study of 100 diabetic haemodialysis patients with a follow-up of 3 years, albumin corrected fructosamine was as reliable as HbA1c for glycaemic control in diabetic patients on haemodialysis and might be advantageous for patients with serum glucose in a desirable therapeutic range (<8.3 mmol/L). The value of fructosamine as a glycaemic index in CAPD diabetic patients has also been demonstrated [138, 139].

In a cohort study of 9704 white women (>65 years of age), elevated serum fructosamine levels were found to be associated with cardiovascular mortality [140]. In contrast to HbA1c, albumin corrected fructosamine correlated with morbidity (hospitalizations and infections) in diabetic haemodialysis patients [141]. Again, the relatively small sample size needs to be taken into account and the prognostic role of fructosamine in dialysis patients has to be further investigated [96].

1,5-Anhydroglucitol

1,5-Anhydroglucitol (1,5-AG) is a non-metabolizable glucose analogue found in plasma after ingestion. It is characterized by urinary excretion, filtration via the glomeruli at a

rate of 5–10 g/L each day and very high tubular reabsorption (>99%), which is inhibited by glucose during periods of hyperglycaemia [142, 143]. As a consequence, 1,5-AG levels in blood respond within 24 h [144] and repetition of hyperglycaemic episodes decreases dramatically the normal steadystate concentration. The 1,5-AG values reflect hyperglycaemic exposure over approximately a 1-week period [145]. Measurement of 1,5-AG could play a role in diabetes monitoring as an adjunct to continuous monitoring of plasma glucose and HbA1c measurement, especially as a unique short-term marker for excursions of hyperglycaemia beyond the glucosuric threshold [146].

Analytical methods. Several methods for 1,5-AG measurement have been evaluated: gas chromatography, gas chromatography/mass spectrometry [147], liquid chromatography/mass spectrometry [148] and enzymatic methods [149]. The 1,5-AG measurement has several advantages: retained metabolic inertness, steady-state levels in all tissues, no influence of anaemia, haemoglobinopathy or liver disease and negligible influence of sampling conditions such as collection time, body weight, age and gender [150, 151]. Serum 1,5-AG levels are influenced by the intake of traditional Chinese herbal drugs [152] and dairy products [153].

1,5-Anhydroglucitol versus HbA1c. As a subanalysis of the Atherosclerosis Risk in Communities study, a cross-sectional comparison of 1,5-AG, fructosamine and glycated albumin with HbA1c and fasting glucose measurements in 1719 participants was conducted. Although a strong correlation was found between 1,5-AG and HbA1c, 1,5-AG performed worse (AUC: 0.74; 95% CI: 0.69-0.78) for identifying cases of undiagnosed diabetes in comparison to the other glycaemic markers [154]. In the same population, fructosamine, glycated albumin and 1,5-AG were strongly associated with future diabetes risk, even after adjustment for HbA1c or fasting glucose [155]. The poorer performance of 1,5-AG in identifying diabetes cases is consistent with the fact that 1,5-AG concentrations are substantially lowered only when circulating glucose concentrations are very high [156]. Of note, 1,5-AG may be useful in the setting of overt hyperglycaemia [154, 155]. It may function as an alternative index for some subtypes of diabetes and as a warning sign of diabetes complications [151].

In patients with CKD, serum 1,5-AG decreases due to a decrease in reabsorption, independently of glucose excretion. Serum and/or urinary 1,5-AG can be a useful marker for renal tubular dysfunction because its reabsorption system is more vulnerable than the glucose reabsorption system [157]. In a recent cross-sectional study of 269 subjects with type 2 diabetes (57 in control, 111 in CKD Stages 1–2, 78 in Stage 3 and 23 in Stages 4–5), 1,5-AG levels did not appear to be influenced by mild or moderate renal dysfunction. This suggests that 1,5-AG could be a reliable glycaemic marker in type 2 diabetes with CKD Stages 1–3. Associations between logarithmic transformed 1,5-AG and HbA1c or fasting plasma glucose were insignificant for CKD Stages 4–5 [158]. Impaired renal function and removal of 1,5-AG by dialysis may contribute to its decreased concentration in patients with ESRD [159]. So

1,5-AG has severe limitations for use in subjects with renal tubular acidosis, uraemia or ESRD [158].

Continuous glucose monitoring system

In patients undergoing dialysis, the use of continuous subcutaneous glucose monitors (CGM) is probably the only method to correctly evaluate glycaemic control. The evaluation of short-term glycaemic undulations around the time of dialysis is possible and results are unaffected by urea, RBC lifespan and RBC production [160]. Using CGM over a 2-day period, significantly higher glucose profiles were reported on the day off dialysis than the day on dialysis [161]. This increased glycaemic variability may represent an adjunctive risk factor for cardiovascular complications [162]. Using the CGM system as reference, mean glucose concentration correlated weakly with HbA1c, but did not correlate at all with fructosamine in haemodialysis patients, in contrast with non-dialysis patients. The CGM system could represent a major advance in assessing glycaemic control in dialysis patients, although future studies should evaluate if this method can be used to indicate necessary adjustments in diabetes treatment or will result in lower mortality rates [163]. In this context, the detection of hypoglycaemic episodes by CGM may be particularly relevant for the prevention of morbidity and mortality in diabetic patients with kidney disease.

CONCLUSION

Due to the availability of relatively inexpensive and routinely measured HbA1c assays and the inconsistent or limited data to prove superiority of other glycaemic markers (glycated albumin, fructosamine, 1,5-AG and continuous glucose monitoring) at this moment, HbA1c is the reference standard for glycaemic monitoring in diabetic patients with CKD (see data extraction table in supplementary data table online). It remains an open question whether and how the HbA1c target should be individualized in patients with advanced CKD, based on age, comorbidity, life expectancy and the presence of risk factors for the occurrence of hypoglycaemia. Continuous subcutaneous glucose monitoring seems to be well suited to correctly evaluate glycaemic control in diabetic patients undergoing dialysis as a 20-40 min time delay can be important. Prospective studies testing pre-specified diabetes control targets based on glycated albumin and continuous glucose measurement remain to be performed in order to determine whether morbidity and mortality would be reduced with intensive glycaemic control using these measurements as reference target.

SUPPLEMENTARY DATA

Supplementary data are available online at http://ndt.oxford journals.org.

ACKNOWLEDGEMENTS

This paper was drafted as a preparation for the upcoming ERBP guideline on management of patients with diabetes and advanced kidney disease. This guideline development group consists of: Henk Bilo, Davide Bolignano, Cecile Couchoud, Adrian Covic, Louis Coentrao, Johan De Sutter, Christiane Drechsler, Luigi Gnudi, David Goldsmith, James Heaf, Olle Heimburger, Kitty Jager, Hakan Nacak, Ionut Nistor, Maria Soler, Charlie Tomson, Liesbeth Vanhuffel, Wim Van Biesen, Steven Van Laecke, Laurent Weekers, Andrzej Wiecek.

CONFLICT OF INTEREST STATEMENT

The declaration of interest forms of C.D., I.N., W.V.B., R.V. and A.W. can be found on the webpage of ERBP: www. european-renal-best-practice.org; M.S., R.L., J.D. and R.S. declared no interests with regard to this paper. The present text is based upon the information available to the guideline development group at the moment of the preparation of this publication. It has been designed to provide information and assist decision-making, but is not intended to define a standard of care or to improve an exclusive course of treatment. Individual decision-making is essential in the approach to any disease and thus also diabetes and advanced chronic kidney disease. Variations in practice are inevitable when physicians take into account individual patient needs, available resources and limitations specific for a geographic area, country, institution or type of practice. In addition, evidence may change over time as new information becomes available, so that practice may be modified subsequently. Every practitioner using this text is responsible for its application to any particular clinical situation.

REFERENCES

NDT PERSPECTI

- Mehdi U, Toto RD. Anemia, diabetes, and chronic kidney disease. Diabetes Care 2009; 32: 1320–1326
- 2. American Diabetes Association. Standards of medical care in diabetes 2012. Diabetes Care 2012; 35: S11–S63
- 3. Collins AJ, Foley RN, Herzog C *et al.* US Renal Data System 2010 Annual Data Report. Am J Kidney Dis 2011; 57: e1–e526
- Koenig RJ, Peterson CM, Jones RL et al. Correlation of glucose regulation and hemoglobin AIc in diabetes mellitus. N Engl J Med 1976; 295: 417–420
- American Diabetes Association, European Association for the Study of Diabetes, International Federation of Clinical Chemistry and Laboratory Medicine *et al.* Consensus statement on the worldwide standardisation of the HbA1c measurement. Diabetologia 2007; 50: 2042–2043
- World Health Organization. Use of Glycated Haemoglobin (HbA1c) in the Diagnosis of Diabetes Mellitus, 2011. http://www.who.int/diabetes/ publications/report-hba1c_2011.pdf (16 January 2014, date last accessed)
- Hutchinson MS, Joakimsen RM, Njølstad I *et al*. Glycated hemoglobin in diagnosis of diabetes mellitus and pre-diabetes; validation by oral glucose tolerance test. The Tromsø OGTT Study. J Endocrinol Invest 2012; 35: 835–840
- Herman WH, Cohen RM. Racial and ethnic differences in the relationship between HbA1c and blood glucose: implications for the diagnosis of diabetes. J Clin Endocrinol Metab. 2012; 97: 1067–1072

- 9. Adler A, Casula A, Steenkamp R *et al.* Association between glycemia and mortality in diabetic individuals on renal replacement therapy in the United Kingdom. Diabetes Care 2013; (in press)
- Lippi G, Targher G. Glycated hemoglobin (HbA1c): old dogmas, a new perspective? Clin Chem Lab Med 2010; 48: 609–614
- Kilpatrick ES, Rigby AS, Atkin SL. Variability in the relationship between mean plasma glucose and HbA1c: implications for the assessment of glycemic control. Clin Chem 2007; 53: 897–901
- Ansari A, Thomas S, Goldsmith D. Assessing glycemic control in patients with diabetes and end-stage renal failure. Am J Kidney Dis 2003; 41: 523–531
- Okada T, Nakao T, Matsumoto H et al. Association between markers of glycemic control, cardiovascular complications and survival in type 2 diabetic patients with end-stage renal disease. Intern Med 2007; 46: 807–814
- 14. Allen DW, Schroeder WA, Balog J. Observations on the chromatographic heterogeneity of normal adult and fetal hemoglobin: a study of the effects of crystallization and chromatography in the heterogeneity and isoleucine content. J Am Chem Soc 1958; 80: 1628–1634
- Shaklai N, Garlick RL, Bunn HF. Nonenzymatic glycosylation of human serum albumin alters its conformation and function. J Biol Chem 1984; 259: 3812–3817
- Lenters-Westra E, Schindhelm RK, Bilo HJ et al. Haemoglobin A1c: Historical overview and current concepts. Diabetes Res Clin Pract 2013; 99: 75–84
- Rahbar S, Blumenfeld O, Ranney HM. Studies of an unusual hemoglobin in patients with diabetes mellitus. Biochem Biophys Res Commun 1969; 36: 838–843
- Trivelli LA, Ranney HM, Lai HT. Hemoglobin components in hemoglobin components in patients with diabetes mellitus. N Eng J Med 1971; 284: 353–357
- Jeppsson JO, Kobold U, Barr J *et al.* Approved IFCC reference method for the measurement of HbA1c in human blood. Clin Chem Lab Med 2002; 40: 78–89
- Weykamp C, John WG, Mosca A. A review of the challenge in measuring hemoglobin A1c. J Diabetes Sci Technol 2009; 3: 439–445
- 21. Henrichs HR. HbA1c-glycated Hemoglobin and Diabetes Mellitus, 1st ed. Bremen: UNI-MED, 2009
- 22. Liu L, Hood S, Wang Y *et al.* Direct enzymatic assay for % HbA1c in human whole blood samples. Clin Biochem 2008; 41: 576–583
- Hoelzel W, Miedema K. Development of a reference system for the international standardization of HbA1c/glycohemoglobin determinations. J Int Fed Clin Chem 1996; 9: 62–67
- Weykamp C, John WG, Mosca A *et al.* The IFCC Reference Measurement System for HbA1c: a 6-year progress report. Clin Chem 2008; 54: 240–248
- 25. Consensus Committee. Consensus statement on the worldwide standardization of the hemoglobin A1c measurement: the American Diabetes Association, European Association for the Study of Diabetes, International Federation of Clinical Chemistry and Laboratory Medicine, and the International Diabetes Federation. Diabetes Care 2007; 30: 2399–2400
- Hanas R, John G.; International HbA(1c) Consensus Committee. 2010 Consensus statement on the worldwide standardization of the hemoglobin A(1c) measurement. Clin Chem Lab Med 2010; 48: 775–776
- Little RR, Rohlfing CL, Sacks DB. Status of hemoglobin A1c measurement and goals for improvement: from chaos to order for improving diabetes care. Clin Chem 2011; 57: 205–214
- Braga F, Panteghini M. Standardization and analytical goals for glycated hemoglobin measurement. Clin Chem Lab Med 2013; 51: 2064
- 29. [No authors listed] The Diabetes Control and Complications (DCCT) Research Group. Effect of intensive therapy on the development and progression of diabetic nephropathy in the Diabetes Control and Complications Trial. Kidney Int 1995; 47: 1703–1720
- 30. Hoelzel W, Weykamp C, Jeppsson JO *et al.* IFCC reference system for measurement of hemoglobin A1c in human blood and the national standardization schemes in the United States, Japan, and Sweden: a method-comparison study. Clin Chem. 2004; 50: 166–174

- Panteghini M, John WG. Implementation of haemoglobin A1c results traceable to the IFCC reference system: the way forward. Clin Chem Lab Med 2007; 45: 942–944
- Lenters-Westra E, Slingerland RJ. Six of eight hemoglobin A1c point-ofcare instruments do not meet the general accepted analytical performance criteria. Clin Chem 2010; 56: 44–52
- Leca V, Ibrahim Z, Lombard-Pontou E *et al.* Point-of-care measurements of HbA(1c): simplicity does not mean laxity with controls. Diabetes Care 2012; 35: e85
- Little RR, Lenters-Westra E, Rohlfing CL *et al.* Point-of-care assays for hemoglobin A(1c): is performance adequate? Clin Chem 2011; 57: 1333–1334
- 35. Al-Ansary L, Farmer A, Hirst J *et al.* Point-of-care testing for HbA1c in the management of diabetes: a systematic review and metaanalysis. Clin Chem 2011; 57: 568–576
- Jeffcoate SL. Diabetes control and complications: the role of glycated haemoglobin, 25 years on. Diabet Med 2004; 21: 657–665
- Cohen RM, Franco RS, Khera PK *et al.* Red cell life span heterogeneity in hematologically normal people is sufficient to alter HbA1c. Blood 2008; 112: 4284–4291
- Leslie RD, Cohen RM. Biologic variability in plasma glucose, hemoglobin A1c, and advanced glycation end products associated with diabetes complications. J Diabetes Sci Technol 2009; 3: 635–643
- Gallagher EJ, Le Roith D, Bloomgarden Z. Review of hemoglobin A(1c) in the management of diabetes. J Diabetes 2009; 1: 9–17
- El-Agouza I, Abu SA, Sirdah M. The effect of iron deficiency anaemia on the levels of haemoglobin subtypes: possible consequences for clinical diagnosis. Clin Lab Haematol 2002; 24: 285–289
- Jiao Y, Okumiya T, Saibara T *et al*. Abnormally decreased HbA1c can be assessed with erythrocyte creatine in patients with a shortened erythrocyte age. Diabetes Care 1998; 21: 1732–1735
- 42. Uzu T, Hatta T, Deji N *et al.* Target for glycemic control in type 2 diabetic patients on hemodialysis: effects of anemia and erythropoietin injection on hemoglobin A(1c). Ther Apher Dial 2009; 13: 89–94
- 43. Little RR, Rohlfing CR, Hanson S et al. Effects of hemoglobin E and D traits on glycated hemoglobin (HbA1c) Measurements by twenty-three methods. Clin Chem 2008; 54: 1277–1282
- 44. Rohlfing C, Connolly S, England J *et al.* The effect of elevated fetal hemoglobin on HbA1c results: five common HbA1c methods compared to the IFCC reference method. Am J Clin Path 2008; 129: 811–814
- 45. Little RR, Rohlfing CL, Hanson SE *et al*. The effect of increased fetal hemoglobin on 7 common Hb A1c assay methods. Clin Chem 2012; 58: 945–946
- 46. Little RR, Roberts WL. A review of variant hemoglobins interfering with hemoglobin A1c measurement. J Diabetes Sci Technol. 2009; 3: 446–451
- Albright ES, Ovalle F, Bell DS. Artificially low hemoglobin A1c caused by use of dapsone. Endocr Pract 2002; 8: 370–372
- Mackenzie T, Brooks B, O'Connor G. Beverage intake, diabetes, and glucose control of adults in America. Ann Epidemiol 2006; 16: 688–691
- 49. Ahmed AT, Karter AJ, Warton EM *et al.* The relationship between alcohol consumption and glycemic control among patients with diabetes: the Kaiser Permanente Northern California Diabetes Registry. J Gen Intern Med 2008; 23: 275–282
- Selvaraj N, Bobby Z, Sridhar MG. Increased glycation of hemoglobin in chronic renal failure patients and its potential role of oxidative stress. Arch Med Res 2008; 39: 277–284
- Camargo JL, Stifft J, Gross JL. The effect of aspirin and vitamins C and E on HbA1c assays. Clin Chim Acta 2006; 372: 206–209
- Garrib A, Griffiths W, Eldridge P *et al.* Artifactually low glycated haemoglobin in a patient with severe hypertriglyceridaemia. J Clin Pathol 2003; 56: 394–395
- Cohen RM, Holmes YR, Chenier TC *et al.* Discordance between HbA1c and fructosamine: evidence for a glycosylation gap and its relation to diabetic nephropathy. Diabetes Care 2003; 26: 163–167
- Rohlfing CL, Wiedmeyer HM, Little RR *et al.* Defining the relationship between plasma glucose and HbA(1c): analysis of glucose profiles and HbA(1c) in the Diabetes Control and Complications Trial. Diabetes Care 2002; 25: 275–258

- Joy MS, Cefalu WT, Hogan SL *et al.* Long-term glycemic control measurements in diabetic patients receiving hemodialysis. Am J Kidney Dis 2002; 39: 297–307
- Chen HS, Wu TE, Lin HD *et al*. Hemoglobin A(1c) and fructosamine for assessing glycemic control in diabetic patients with CKD stages 3 and 4. Am J Kidney Dis 2010; 55: 867–887
- Bloomgarden ZT, Inzucchi SE, Karnieli E *et al.* The proposed terminology 'A(1c)-derived average glucose' is inherently imprecise and should not be adopted. Diabetologia 2008; 51: 1111–1114
- Inaba M, Okuno S, Kumeda Y *et al.* Glycated albumin is a better glycemic indicator than glycated hemoglobin values in hemodialysis patients with diabetes: effect of anemia and erythropoietin injection. J Am Soc Nephrol 2007; 18: 896–903
- 59. Ly J, Marticorena R, Donnelly S. Red blood cell survival in chronic renal failure. Am J Kidney Dis 2004; 44: 715–719
- 60. Vos FE, Schollum JB, Coulter CV *et al.* Red blood cell survival in patients on long-term dialysis. Am J Kidney Dis 2011; 58: 591–598
- Ng JM, Jennings PE, Laboi P *et al*. Erythropoetin treatment significantly alters measured glycated haemoglobin (HbA1c). Diabet Med 2008; 25: 239–240
- 62. Ng JM, Cooke M, Bhandari S *et al.* The effect of iron and erythropoietin treatment on the A1C of patients with diabetes and chronic kidney disease. Diabetes Care 2010; 33: 2310–2313
- Nigen AM, Bass BD, Manning JM. Reactivity of cyanate with valine-1 (alpha) of hemoglobin. A probe of conformational change and anion binding. J Biol Chem 1976; 251: 7638–7643
- 64. Jaisson S, Lorimier S, Ricard-Blum S *et al.* Impact of carbamylation on type I collagen conformational structure and its ability to activate human polymorphonuclear neutrophils. Chem Biol 2006; 13: 149–159
- 65. Weykamp CW, Penders TJ, Siebelder CW et al. Interference of carbamylated and acetylated haemoglobins in assays of glycohaemoglobin by HPLC, electrophoresis, affinity chromatography and enzyme immunoassay. Clin Chem 1993; 39: 138–142
- 66. Weykamp CW, Penders TJ, Muskiet FA *et al.* Influence of haemoglobin variants and derivatives on glycohaemoglobin determinations, as investigated by 102 laboratories using 16 methods. Clin Chem 1993; 39: 1717–1723
- 67. Chachou A, Randoux C, Millart H *et al.* Influence of in vivo hemoglobin carbamylation on HbA1c measurements by various methods. Clin Chem Lab Med 2000; 38: 321–326
- Meijs MF, Dijkhorst-Oei LT, van Loo R *et al.* Does carbamylated hemoglobin still affect the analysis of HbA(1c) in uremic and hyperglycemic patients?. Clin Chem Lab Med 2008; 46: 1791–1792
- Agarwal R, Light RP. Relationship between glycosylated hemoglobin and blood glucose during progression of chronic kidney disease. Am J Nephrol 2011; 34: 32–41
- National Kidney Foundation. KDOQI Clinical Practice Guideline for Diabetes and CKD: 2012 Update. Am J Kidney Dis 2012; 60: 850–886. [Erratum in: Am J Kidney Dis 2013; 61: 1049.]
- Shurraw S, Hemmelgarn B, Lin M *et al.* Association between glycemic control and adverse outcomes in people with diabetes mellitus and chronic kidney disease: a population-based cohort study. Arch Intern Med 2011; 171: 1920–1927
- Currie CJ, Peters JR, Tynan A *et al.* Survival as a function of HbA(1c) in people with type 2 diabetes: a retrospective cohort study. Lancet 2010; 375: 481–489
- Williams ME, Lacson E, Jr, Teng M *et al.* Extremes of glycemic control (HbA1C) increase hospitalization risk in diabetic hemodialysis patients in the USA. Am J Nephrol 2009; 29: 54–61
- Ricks J, Molnar MZ, Kovesdy CP *et al.* Glycemic control and cardiovascular mortality in hemodialysis patients with diabetes: a 6-year cohort study. Diabetes 2012; 61: 708–715
- 75. Kalantar-Zadeh K, Kopple JD, Regidor DL *et al.* A1C and survival in maintenance hemodialysis patients. Diabetes Care 2007; 30: 1049–1055
- 76. Williams ME, Lacson E, Jr, Wang W et al. Glycemic control and extended hemodialysis survival in patients with diabetes mellitus: comparative results of traditional and time-dependent Cox model analyses. Clin J Am Soc Nephrol 2010; 5: 1595–1601

- 77. Duong U, Mehrotra R, Molnar MZ *et al.* Glycemic control and survival in peritoneal dialysis patients with diabetes mellitus. Clin J Am Soc Nephrol 2011; 6: 1041–1048
- Wu MS, Yu CC, Yang CW *et al.* Poor pre-dialysis glycaemic control is a predictor of mortality in type II diabetic patients on maintenance haemodialysis. Nephrol Dial Transplant 1997; 12: 2105–2110
- Morioka T, Emoto M, Tabata T *et al.* Glycemic control is a predictor of survival for diabetic patients on hemodialysis. Diabetes Care 2001; 24: 909–913
- Molnar MZ, Huang E, Hoshino J et al. Association of pretransplant glycemic control with posttransplant outcomes in diabetic kidney transplant recipients. Diabetes Care 2011; 34: 2536–2541
- Adler A, Casula A, Steenkamp R *et al.* Association between Glycaemia and Mortality in Diabetic Individuals on Renal Replacement Therapy in the United Kingdom. Oral Presentation at: American Diabetes Association Conference; June 8–12, 2012; Philadelphia, PA
- Freedman BI, Andries L, Shihabi ZK et al. Glycated albumin and risk of death and hospitalizations in diabetic dialysis patients. Clin J Am Soc Nephrol 2011; 6: 1635–1643
- Oomichi T, Emoto M, Tabata T *et al.* Impact of glycemic control on survival of diabetic patients on chronic regular hemodialysis: a 7-year observational study. Diabetes Care 2006; 29: 1496–1500
- Hayashino Y, Fukuhara S, Akiba T *et al.* Diabetes, glycaemic control and mortality risk in patients on haemodialysis: the Japan Dialysis Outcomes and Practice Pattern Study. Diabetologia 2007; 50: 1170–1177
- Shurraw S, Majumdar SR, Thadhani R et al. Glycemic control and the risk of death in 1,484 patients receiving maintenance hemodialysis. Am J Kidney Dis 2010; 55: 875–884
- 86. Sturm G, Lamina C, Zitt E *et al.* Association of HbA1c values with mortality and cardiovascular events in diabetic dialysis patients. The INVOR study and review of the literature. PLoS ONE 2011; 6: e20093
- Drechsler C, Krane V, Ritz E *et al.* Glycemic control and cardiovascular events in diabetic hemodialysis patients. Circulation 2009; 120: 2421–2428
- Hill CJ, Maxwell AP, Cardwell CR *et al.* Glycated hemoglobin and risk of death in diabetic patients treated with hemodialysis: a meta-analysis. Am J Kidney Dis 2013 doi: 10.1053/j.ajkd.2013.06.020
- Williams ME, Lacson E, Jr, Teng M *et al.* Hemodialyzed type I and type II diabetic patients in the US: Characteristics, glycemic control, and survival. Kidney Int 2006; 70: 1503–1509
- Sekercioglu N, Dimitriadis C, Pipili C et al. Glycemic control and survival in peritoneal dialysis patients with diabetes mellitus. Int Urol Nephrol 2012; 44: 1861–1869
- Shima K, Chujo K, Yamada M *et al.* Lower value of glycated haemoglobin relative to glycaemic control in diabetic patients with end-stage renal disease not on haemodialysis. Ann Clin Biochem 2012; 49: 68–74
- 92. Kalantar-Zadeh K. A critical evaluation of glycated protein parameters in advanced nephropathy: a matter of life or death: A1C remains the gold standard outcome predictor in diabetic dialysis patients. Counterpoint. Diabetes Care 2012; 35: 1625–1628
- Hemmingsen B, Lund SS, Gluud C et al. Intensive glycaemic control for patients with type 2 diabetes: systematic review with meta-analysis and trial sequential analysis of randomised clinical trials. BMJ 2011; 343: d6898
- Freedman BI, Shenoy RN, Planer JA et al. Comparison of glycated albumin and hemoglobin A1c concentrations in diabetic subjects on peritoneal and hemodialysis. Perit Dial Int 2010; 30: 72–79
- Furusyo N, Hayashi J. Glycated albumin and diabetes mellitus. Biochim Biophys Acta 2013; 1830: 5509–5514
- 96. Zheng CM, Ma WY, Wu CC *et al.* Glycated albumin in diabetic patients with chronic kidney disease. Clin Chim Acta 2012; 413: 1555–1561
- Schleicher ED, Olgemöller B, Wiedenmann E et al. Specific glycation of albumin depends on its half-life. Clin Chem 1993; 39: 625–628
- Paroni R, Ceriotti F, Galanello R et al. Performance characteristics and clinical utility of an enzymatic method for the measurement of glycated albumin in plasma. Clin Biochem 2007; 40: 1398–1405
- 99. Anguizola J, Matsuda R, Barnaby OS *et al.* Review: glycation of human serum albumin. Clin Chim Acta 2013; 425C: 64–76

- 100. Tahara Y. Analysis of the method for conversion between levels of HbA1c and glycated albumin by linear regression analysis using a measurement error model. Diabetes Res Clin Pract 2009; 84: 224–229
- 101. Shima K, Ito N, Abe F *et al.* High-performance liquid chromatographic assay of serum glycated albumin. Diabetologia 1988; 31: 627–631
- 102. Peacock TP, Shihabi ZK, Bleyer AJ et al. Comparison of glycated albumin and hemoglobin A(1c) levels in diabetic subjects on hemodialysis. Kidney Int 2008; 73: 1062–1068
- 103. Selvin E, Francis LM, Ballantyne CM et al. Nontraditional markers of glycemia: associations with microvascular conditions. Diabetes Care 2011; 34: 960–967
- Okada T, Nakao T, Matsumoto H *et al.* Influence of age and nutritional status on glycated albumin values in hemodialysis patients. Intern Med 2009; 48: 1495–1499
- 105. Koga M, Matsumoto S, Saito H et al. Body mass index negatively influences glycated albumin, but not glycated hemoglobin, in diabetic patients. Endocr J 2006; 53: 387–391
- 106. Wang F, Ma X, Hao Y *et al.* Serum glycated albumin is inversely influenced by fat mass and visceral adipose tissue in Chinese with normal glucose tolerance. PLoS ONE 2012; 7: e51098
- 107. Koga M, Otsuki M, Matsumoto S *et al.* Negative association of obesity and its related chronic inflammation with serum glycated albumin but not glycated hemoglobin levels. Clin Chim Acta 2007; 378: 48–52
- 108. Kohzuma T, Yamamoto T, Uematsu Y *et al.* Basic performance of an enzymatic method for glycated albumin and reference range determination. J Diabetes Sci Technol 2011; 5: 1455–1462
- 109. Freedman BI, Shihabi ZK, Andries L et al. Relationship between assays of glycemia in diabetic subjects with advanced chronic kidney disease. Am J Nephrol 2010; 31: 375–379
- 110. Freedman BI. A critical evaluation of glycated protein parameters in advanced nephropathy: a matter of life or death: time to dispense with the hemoglobin A1C in end-stage kidney disease. Diabetes Care 2012; 35: 1621–1624
- 111. Nagayama H, Inaba M, Okabe R et al. Glycated albumin as an improved indicator of glycemic control in hemodialysis patients with type 2 diabetes based on fasting plasma glucose and oral glucose tolerance test. Biomed Pharmacother 2009; 63: 236–240
- 112. Kim JK, Park JT, Oh HJ *et al.* Estimating average glucose levels from glycated albumin in patients with end-stage renal disease. Yonsei Med J 2012; 53: 578–586
- 113. Lamb E, Venton TR, Cattell WR *et al.* Serum glycated albumin and fructosamine in renal dialysis patients. Nephron 1993; 64: 82–88
- 114. Spiegel DM, Breyer JA. Serum albumin: a predictor of long-term outcome in peritoneal dialysis patients. Am J Kidney Dis 1994; 23: 283–285
- 115. Mehrotra R, Duong U, Jiwakanon S et al. Serum albumin as a predictor of mortality in peritoneal dialysis: comparisons with hemodialysis. Am J Kidney Dis 2011; 58: 418–428
- 116. Bhonsle HS, Korwar AM, Kote SS *et al.* Low plasma albumin levels are associated with increased plasma protein glycation and HbA1c in diabetes. J Proteome Res 2012; 11: 1391–1396
- 117. Lu L, Pu LJ, Xu XW *et al.* Association of serum levels of glycated albumin, C-reactive protein and tumor necrosis factor-alpha with the severity of coronary artery disease and renal impairment in patients with type 2 diabetes mellitus. Clin Biochem 2007; 40: 810–816
- 118. Kumeda Y, Inaba M, Shoji S *et al.* Significant correlation of glycated albumin, but not glycated haemoglobin, with arterial stiffening in haemodialysis patients with type 2 diabetes. Clin Endocrinol (Oxf) 2008; 69: 556-561
- 119. Yamada S, Inaba M, Okada S *et al.* Association of glycated albumin, but not glycated hemoglobin, with calcaneus quantitative ultrasound in male hemodialysis patients with type 2 diabetes mellitus. Metabolism 2010; 59: 390–394
- Fukuoka K, Nakao K, Morimoto H *et al.* Glycated albumin levels predict long-term survival in diabetic patients undergoing haemodialysis. Nephrology (Carlton) 2008; 13: 278–283
- 121. Murea M, Moran T, Russell GB *et al.* Glycated albumin, not hemoglobin A1c, predicts cardiovascular hospitalization and length of stay in diabetic patients on dialysis. Am J Nephrol 2012; 36: 488–496

NDT PERSPECTIVE

- 122. Little RR, Rohlfing CL, Tennill AL *et al.* Measurement of Hba(1C) in patients with chronic renal failure. Clin Chim Acta 2013; 418: 73–76
- 123. Chujo K, Shima K, Tada H *et al.* Indicators for blood glucose control in diabetics with end-stage chronic renal disease: GHb vs. glycated albumin (GA). J Med Invest 2006; 53: 223–228
- 124. Armbruster DA. Fructosamine: structure, analysis, and clinical usefulness. Clin Chem 1987; 33: 2153–2163
- Schleicher ED, Vogt BW. Standardization of serum fructosamine assays. Clin Chem 1990; 36: 136–139
- 126. Montagnana M, Paleari R, Danese E et al. Evaluation of biological variation of glycated albumin (GA) and fructosamine in healthy subjects. Clin Chim Acta 2013; 423: 1–4
- True MW. Circulating biomarkers of glycemia in diabetes management and implications for personalized medicine. J Diabetes Sci Technol 2009; 3: 743–747
- Van Dieijen-Visser MP, Seynaeve C, Brombacher PJ. Influence of variations in albumin or total-protein concentration on serum fructosamine concentration. Clin Chem 1986; 32: 1610
- 129. Vos FE, Schollum JB, Coulter CV et al. Assessment of markers of glycaemic control in diabetic patients with chronic kidney disease using continuous glucose monitoring. Nephrology (Carlton) 2012; 17: 182–188
- Goldstein DE, Little RR, Lorenz RA *et al.* Tests of glycemia in diabetes. Diabetes Care 1995; 18: 896–909
- 131. Mosca A, Carenini A, Zoppi F *et al.* Plasma protein glycation as measured by fructosamine assay. Clin Chem 1987; 33: 1141–1146
- Schleicher ED, Mayer R, Wagner EM *et al.* Is serum fructosamine assay specific for determinations of glycated serum proteins? Clin Chem 1988; 34: 320–323
- 133. Baker JR, O'Connor JP, Metcalf PA *et al*. Clinical usefulness of estimation of serum fructosamine concentration as a screening test for diabetes mellitus. Br Med J (Clin Res Ed) 1983; 287: 863–867
- 134. Morgan LJ, Marenah CB, Morgan AG et al. Glycated haemoglobin and fructosamine in non-diabetic subjects with chronic renal failure. Nephrol Dial Transplant 1990; 5: 868–873
- 135. Nunoi K, Kodama T, Sato Y et al. Comparison of reliability of plasma fructosamine and glycosylated hemoglobin assays for assessing glycemic control in diabetic patients on hemodialysis. Metabolism 1991; 40: 986–989
- 136. Morgan L, Marenah CB, Jeffcoate WJ et al. Glycated proteins as indices of glycaemic control in diabetic patients with chronic renal failure. Diabet Med 1996; 13: 514–519
- Cohen RM, Smith EP. Frequency of HbA1c discordance in estimating blood glucose control. Curr Opin Clin Nutr Metab Care 2008; 11: 512–517
- 138. Coronel F, Macia M, Cidoncha A *et al*. Fructosamine levels in CAPD: its value as glycemic index. Adv Perit Dial 1991; 7: 253–256
- 139. Lee SY, Chen YC, Tsai IC *et al.* Glycosylated hemoglobin and albumincorrected fructosamine are good indicators for glycemic control in peritoneal dialysis patients. PLoS ONE 2013; 8: e57762
- 140. Browner WS, Pressman AR, Lui LY *et al.* Association between serum fructosamine and mortality in elderly women: the study of osteoporotic fractures. Am J Epidemiol 1999; 149: 471–475
- 141. Mittman N, Desiraju B, Fazil I *et al.* Serum fructosamine versus glycosylated hemoglobin as an index of glycemic control, hospitalization, and infection in diabetic hemodialysis patients. Kidney Int Suppl 2010; 117: S41–S45
- 142. Yamanouchi T, Tachibana Y, Akanuma H et al. Origin and disposal of 1,5-anhydroglucitol, a major polyol in the human body. Am J Physiol 1992; 263: E268–E273
- 143. Yamanouchi T, Akanuma Y. Serum 1,5-anhydroglucitol (1,5 AG): new clinical marker for glycemic control. Diabetes Res Clin Pract 1994; 24: S261–S268

- 144. Buse JB, Freeman JL, Edelman SV *et al.* Serum 1,5-anhydroglucitol (GlycoMark): a short-term glycemic marker. Diabetes Technol Ther 2003; 5: 355–363
- 145. Stettler C, Stahl M, Allemann S *et al.* Association of 1,5-anhydroglucitol and 2-h postprandial blood glucose in type 2 diabetic patients. Diabetes Care 2008; 31: 1534–1535
- 146. Nerby CL, Stickle DF. 1,5-anhydroglucitol monitoring in diabetes: a mass balance perspective. Clin Biochem 2009; 42: 158–167
- 147. Niwa T, Yamamoto N, Maeda K et al. Gas chromatographic-mass spectrometric analysis of polyols in urine and serum of uremic patients. Identification of new deoxyalditols and inositol isomers. J Chromatogr 1983; 277: 25-39
- Niwa T, Dewald L, Sone J *et al.* Quantification of serum 1,5-anhydroglucitol in uremic and diabetic patients by liquid chromatography/mass spectrometry. Clin Chem 1994; 40: 260–264
- Yahuuchi M, Masuda M, Katoh K *et al.* Simple enzymatic method for determining 1,5-anhydro-D-glucitol for diagnosis of diabetes mellitus. Clin Chem 1989; 35: 2039–2043
- Dungan KM. 1,5-anhydroglucitol (GlycoMark) as a marker of shortterm glycemic control and glycemic excursions. Expert Rev Mol Diagn 2008; 8: 9–19
- 151. Kim WJ, Park CY. 1,5-Anhydroglucitol in diabetes mellitus. Endocrine 2013; 43: 33-40
- 152. Kawasaki T, Yamanouchi T, Kashiwabara A *et al*. The influence of traditional Chinese herbal drugs on serum 1,5-anhydroglucitol levels. Diabetes Res Clin Pract 2000; 50: 97–101
- 153. Koga M, Murai J, Saito H *et al.* Habitual intake of dairy products influences serum 1,5-anhydroglucitol levels independently of plasma glucose. Diabetes Res Clin Pract 2010; 90: 122–125
- 154. Juraschek SP, Steffes MW, Selvin E. Associations of alternative markers of glycemia with hemoglobin A(1c) and fasting glucose. Clin Chem 2012; 58: 1648–1655
- 155. Juraschek SP, Steffes MW, Miller ER, 3rd *et al.* Alternative markers of hyperglycemia and risk of diabetes. Diabetes Care 2012; 35: 2265–2270
- 156. Saudek CD, Derr RL, Kalyani RR. Assessing glycemia in diabetes using self-monitoring blood glucose and hemoglobin A1c. JAMA 2006; 295: 1688–1697
- 157. Shimizu H, Shouzu A, Nishikawa M *et al.* Serum concentration and renal handling of 1,5-anhydro-D-glucitol in patients with chronic renal failure. Ann Clin Biochem 1999; 36: 749–774
- 158. Kim WJ, Park CY, Lee KB *et al.* Serum 1,5-anhydroglucitol concentrations are a reliable index of glycemic control in type 2 diabetes with mild or moderate renal dysfunction. Diabetes Care 2012; 35: 281–286
- Emoto M, Tabata T, Inoue T *et al.* Plasma 1,5-anhydroglucitol concentration in patients with end-stage renal disease with and without diabetes mellitus. Nephron 1992; 61: 181–186
- 160. Jung HS, Kim HI, Kim MJ et al. Analysis of hemodialysis-associated hypoglycemia in patients with type 2 diabetes using a continuous glucose monitoring system. Diabetes Technol Ther 2010; 12: 801–807
- 161. Kazempour-Ardebili S, Lecamwasam VL, Dassanyake T et al. Assessing glycemic control in maintenance hemodialysis patients with type 2 diabetes. Diabetes Care 2009; 32: 1137–1142
- 162. Mirani M, Berra C, Finazzi S *et al*. Inter-day glycemic variability assessed by continuous glucose monitoring in insulin-treated type 2 diabetes patients on hemodialysis. Diabetes Technol Ther 2010; 12: 749–753
- 163. Riveline JP, Teynie J, Belmouaz S et al. Glycaemic control in type 2 diabetic patients on chronic haemodialysis: use of a continuous glucose monitoring system. Nephrol Dial Transplant 2009; 24: 2866–2871

Received for publication: 3.12.2013; Accepted in revised form: 2.1.2013