



# **Are We What We Eat? Impact of Diet on the Gut–Brain Axis in Parkinson's Disease**

Margherita Alfonsetti 🔍, Vanessa Castelli \* 🗅 and Michele d'Angelo \* 🕑

Department of Life, Health and Environmental Sciences, University of L'Aquila, 67100 L'Aquila, Italy; margherita.alfonsetti@guest.univaq.it

\* Correspondence: vanessa.castelli@univaq.it (V.C.); michele.dangelo@univaq.it (M.d.)

Abstract: Parkinson's disease is characterized by motor and non-motor symptoms, such as defects in the gut function, which may occur before the motor symptoms. To date, there are therapies that can improve these symptoms, but there is no cure to avoid the development or exacerbation of this disorder. Dysbiosis of gut microbiota could have a crucial role in the gut–brain axis, which is a bidirectional communication between the central nervous system and the enteric nervous system. Diet can affect the microbiota composition, impacting gut–brain axis functionality. Gut microbiome restoration through probiotics, prebiotics, synbiotics or other dietary means could have the potential to slow PD progression. In this review, we will discuss the influence of diet on the bidirectional communication between gut and brain, thus supporting the hypothesis that this disorder could begin in the gut. We also focus on how food-based therapies might then have an influence on PD and could ameliorate non-motor as well as motor symptoms.

**Keywords:** probiotics; prebiotics; synbiotics; Parkinson's disease; neurodegeneration; α-synuclein; brain; gut; microbiota; diet



Citation: Alfonsetti, M.; Castelli, V.; d'Angelo, M. Are We What We Eat? Impact of Diet on the Gut–Brain Axis in Parkinson's Disease. *Nutrients* **2022**, *14*, 380. https://doi.org/ 10.3390/nu14020380

Academic Editor: Armin Alaedini

Received: 18 December 2021 Accepted: 14 January 2022 Published: 17 January 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/).

# 1. Introduction

Parkinson's disease (PD) is a progressive neurodegenerative disorder characterized by the loss of dopaminergic neurons in the midbrain, specifically in the substantia nigra pars compacta, associated with the formation of cytoplasmatic inclusions named Lewy bodies constituted of insoluble  $\alpha$ -synuclein aggregates [1]. Nevertheless, PD pathology involves the degeneration of non-dopaminergic neurons as well. Interestingly,  $\alpha$ -synuclein deposition is observed in several peripheral organs including the gastrointestinal (GI) system (submandibular gland, stomach, and bowels) raising the question of a possible role of the gut in PD pathogenesis [2].

PD is a complex disorder with multifactorial etiology: both environmental and genetic factors participate in a common set of pathways including mitochondrial alteration [3], ROS formation [4], protein aggregation, compromised autophagy and neuroinflammation [5,6].

PD clinical diagnosis is centered on the observation of both motor and non-motor symptoms. Rest tremor, bradykinesia, rigidity and loss of postural reflexes are the cardinal PD motor symptoms. Rather, secondary motor signs include dysarthria, glabellar reflexes, dysphagia, sialorrhea, micrographia, festination, shuffling gait, hypomimia and dystonia [7]. Although PD is considered a movement disorder, it is associated with a wide spectrum of non-motor features, such as anosmia, depression, sleep disorders, gastrointestinal dysfunction and low-grade mucosal inflammation in the enteric nervous system [8]. During later stages of the disorder, additional non motor features may appear that include autonomic dysfunction (orthostatic hypotension and urogenital dysfunction), pain and cognitive deficits [9].

To date, there are treatments that could help relief these symptoms but there is no cure to control the development and progression of this disease [10]. Pharmacological

treatments include dopamine-based preparations such as levodopa, dopamine agonists and monoamine oxidase-B (MAO-B) inhibitors are usually administered as initial treatments [11]. Levodopa is the most commonly used drug, which controls some motor symptoms and counteracts dopaminergic cell loss by improving dopamine synthesis. However, this therapy has numerous adverse effects, it does not inhibit dopaminergic neurodegeneration and has no impact on non-motor symptoms [12]. Moreover, PD-associated gastrointestinal dysfunction contributes to levodopa effect fluctuations and oral treatment with levodopa needs optimal GI function to determine an ideal drug metabolism, indeed, it has been demonstrated that levodopa leads to delayed gastric emptying in healthy patients [13] and aggravates GI symptoms in PD patients [14]. In addition, further studies found that levodopa responsivity is directly correlated with the severity of  $\alpha$ -synuclein accumulation in the enteric nervous system (ENS) [15–17].

More advanced therapies represented by deep brain stimulation, MRI-guided focused ultrasound, and therapy with levodopa-carbidopa enteral suspensions can support individuals with medication-resistant tremor and dyskinesias [18,19].

#### 2. Gut-Brain Axis

The gut–brain axis is defined as the two-way communication among the central nervous system (CNS) and the ENS that bridges emotional and cognitive brain areas with outermost intestinal functions [20]. This communication comprises the CNS (brain and spinal cord), the autonomic nervous system, the ENS and the hypothalamic pituitary adrenal (HPA) axis [20].

Sympathetic and parasympathetic limbs of the autonomic system participate in afferent (vagus nerve to CNS) and efferent pathways (CNS to ENS) [21].

The hypothalamic pituitary adrenal gland is known to have roles in the adaptive responses to several stressors and it is a component of the limbic system that is crucial in emotional and memory processes [22]. Stressors together with elevated levels of proinflammatory cytokines induce the release of corticotropin-releasing factor (CRF) from the hypothalamus that activates this system. The release of adrenocorticotropic hormone (ACTH) from the pituitary gland leads to cortisol secretion from adrenal glands [23]. Cortisol is a crucial stress hormone that interacts with several organs, brain included. Therefore, activities of intestinal functional effector cells, such as immune cells, enteric neuronal cells, smooth muscle cells, interstitial cells of Cajal and enterochromaffin cells are affected by both neuronal lines and hormones [24]. Moreover, these cells are influenced by gut microbiota [25].

As mentioned above, GI dysfunction is a crucial non-motor symptom of PD, which frequently appears at the very early stage of the disorder. Some studies established that PD patients are affected by constipation for more than 20 years before the onset of motor symptoms [26]. GI symptoms, including exaggerated salivation, dysphagia, constipation, esophageal motility disorder, and gastric abnormalities, frequently occur years before motor symptoms and their incidence in a healthy population has been associated with an increased risk of developing PD [27]. Additionally, the association with GI abnormalities validates the Braak's theory that PD might initiate in the GI tract, supported by the presence of Lewy body burden in the ENS with respect to other body areas and in the CNS [28].

Observing the distribution of Lewy bodies in PD patients, Braak and his research group assumed that  $\alpha$ -synuclein pathology begins in the ENS and then travels to the brain passing through the brainstem, midbrain, basal forebrain and lastly the cortical regions [29,30].  $\alpha$ -synuclein aggregates are usually found in structures that contribute to the parasympathetic innervation to the intestine [31]. For this reason, the vagal nerve may represent the communication channel between the gut and brain. In fact, recently it has been proved that full truncal vagotomy can be related to a decreased risk in developing PD compared to highly selective vagotomy or control conditions [32,33]. Solid data are still lacking regarding this hypothesis, and several studies show contradicting evidence.

Interestingly, a clinical study showed that there is no significant association between *T. gondii* infection and idiopathic PD [34] and another work showed a significant increase in dopamine metabolism in neural cells [35]. Moreover, a very recent paper goes against Braak's theory indicating that  $\alpha$ -synuclein aggregates are found in the vagus nerve and in the stomach of PD patients, but not in normal ageing people, thus suggesting the beginning of  $\alpha$ -synuclein pathology in the brain [36].

 $\alpha$ -synuclein accumulation in the GI system is correlated with injury in the enteric neurons and feasibly triggers GI dysfunction [37]. The damage involves both myenteric and submucosal plexuses of the gut and the distribution is through the whole GI tract, starting from the esophagus to the rectum [38]. Accordingly, it has been proved that  $\alpha$ -synuclein can be retrogradely moved from the intestine to the brain in rat models. Emerging studies in vitro and in vivo showed that  $\alpha$ -synuclein can diffuse through endocytic mechanisms to neighboring neurons where it creates inclusions [38]. Another evidence showed that a PD-like pathology can be induced by oral administration of rotenone in mice, thus proving that the local effect of pesticides at the level of the ENS may be enough to induce PD signs from the ENS to the brain [39].

Since the olfactory bulbs and the ENS are continuously exposed to environmental agents throughout inhalation or ingestion, may be possible that factors such as diet, toxins, microorganisms and different environmental pathogens may determine and propagate PD pathology progression probably against a background of genetic susceptibility [40].

## 3. Gut Microbiota

The human gut is home to numerous bacteria, archaea, fungi, microbial eukaryotes and viruses/phages. This assortment of microbes is named the "gut microbiota" and their respective genes are the "microbiome" [41,42]. The typical composition of gut microbial community includes *Bacteroidetes*, *Firmicutes*, *Actinobacteria*, *Proteobacteria* and *Verrucomicrobia phyla*. *Bacteroidetes* and *Firmicutes*, in a healthy individual, represent more than the 90% of the total bacteria and their ratio depends on the host genomes and environmental features (hygiene, use of antibiotics or drugs, lifestyle and diet) [43]. Several pathologies affect the gut inducing dysbiosis. These pathologies include obesity, diabetes, diarrhea and irritable bowel syndrome [44].

The gut microbiota in adults is quite stable but the bacterial community can be easily altered by environmental factors. Dietary habits represent the main factor that influence the microbiome variety and the effects of nutrients and diet on gut microbiota were widely studied [45]. Emerging evidence shows that the interaction between diet and microbiota is highly dependent on the individual microbiota composition.

Gut microbes contribute to the digestion, absorption, metabolism and transformation of undigested macronutrients into advantageous and active molecules for human health [46]. Every macronutrient impacts the gut microbiota in a distinctive manner, and changes in macronutrient proportions, quantities and categories are associated with microbiota composition [47]. Carbohydrates represent the most effective macronutrients for modifying the gut microbiota, in particular dietary fiber [48].

Nowadays the identification and quantification of gut bacterial genera has enabled investigation of the impact of diet on gut microbiota. In a dietary intervention period ranging from 24 to 48 h it has been shown that quick variations occur in microbial composition at species and family levels (but not *phyla*) [49]. Similarly, rodent models indicate that changes in macronutrient intake can modify gut microbiota composition in a day [50–52]. These reported changes were caused not only by the diet composition, but also by intrinsic and extrinsic factors that may play a role, such as circadian rhythm and feeding behaviors [53,54]. Emerging evidence shows how specific bacteria respond to certain dietary components. Protein, fats, digestible and non-digestible carbohydrates, probiotics and polyphenols all produce changes in the microbiome with secondary consequences for host immunologic and metabolic markers.

The impact of dietary proteins on the gut microbiota was first reported in 1977. A culture-based study showed low levels of *Bifidobacterium adolescentis* and an improved number of *Bacteroides* and *Clostridia* in humans with a high beef diet compared to subjects with a meatless diet [55]. In vegetarians, it has been reported that the intake of whey and pea protein extracts increases gut-commensal *Bifidobacterium* and *Lactobacillus*, moreover, whey protein reduces the pathogenic *Bacteroides fragilis* and *Clostridium perfringens* [56]. Pea protein intake was also associated with increased intestinal short-chain fatty acid (SCFA) levels, beneficial for the maintenance of the intestinal barrier [57]. Conversely, the number of bile-tolerant anaerobes including *Bacteroides*, *Alistipes* and *Bilophila* is associated with animal-based protein intake together with a reduction in the number of the *Roseburia E. rectale* group [52,58,59].

Dietary polyphenols, such as flavonoids, anthocyanins and phenolic acids are widely investigated because of their antioxidant properties. These compounds are contained in many foods like fruits, seeds, vegetables, tea, cocoa products and wine [60] and they usually determine the enrichment of *Bifidobacterium* and *Lactobacillus genera* in the gut [61–65]. *Bifidobacterium genera* are known to have several health benefits included immune-modulation, cancer prevention and inflammatory bowel disease management [66]. Interestingly, an investigation of the antibacterial activity of fruit polyphenols demonstrated that enteropathogens such as *Staphylococcus aureus* and *Salmonella typhimurium* are particularly susceptible to these polyphenols [67]. Moreover, low levels of infective *Clostridium* species (*C. perfringens* and *C. histolyticum*) after fruit, seed, wine and tea intake were observed [62,65,68,69].

The standard Western diet is rich in saturated and trans fats but low in mono and polyunsaturated fats. A high saturated fat intake has been shown to increase the number of total anaerobic microflora and the relative quantity of *Bacteroides* and *Bilophila* [51,70]. There are no clinical studies demonstrating an alteration of gut microbiota upon high-unsaturated fat diets; however, in vivo studies (on mice) reported an increase in in Actinobacteria (*Bifidobacterium* and *Adlercreutzia*), lactic acid bacteria (*Lactobacillus* and *Streptococcus*), and Verrucomicrobia (*Akkermansia muciniphila*) [71].

Alcohol consumption could also modulate gut microbiota. Alcohol absorption occurs predominantly in the small intestine [72] and it is mostly metabolized in the liver by alcohol dehydrogenase (ADH), which converts alcohol into acetaldehyde which is lethal for tissues and gut microbes. Moreover, alcohol consumption could lead to a disruption in the microbiota homeostasis, increasing the number of gram-negative bacteria [73,74], decreasing the number of SCFA-producing bacteria [75], damaging intestinal barrier integrity through toxins produced by gram-negative bacteria [76] and increasing the permeability of the intestinal mucosa [77].

Increasing evidence proposed that alcohol intake may directly affect the gut microbiota composition. Alcohol consumption in rats for 13 weeks led to reduced  $\alpha$ -diversity and  $\beta$ -diversity, reduced abundance of *Lactobacilli*, and enhanced *Bacteroidetes* compared to the control group [72].

#### 4. Gut Microbiota and PD

Evidence from Yang et al. [78] revealed that microbiota dysbiosis could represent a cause of PD. Specifically, in a mouse model treated with rotenone, modifications of fecal bacterial compositions displayed by a decrease in bacterial diversity and high levels of *Firmicutes* and *Bacteroidetes* preceded the instauration of  $\alpha$ -synuclein pathology [78]. In humans, gut microbiota composition varies as the PD progresses and these alterations are correlated with PD clinical symptoms [79]. Even though gut microbiota dysbiosis in PD are well known, it is still not clear whether alterations in the intestinal microbiota are either a cause or an effect of the disease.

Nowadays, it is well-established that the microbiota composition is altered in PD patients [80–84]. The first evidence is from 2015 [83], in particular, fecal samples of PD patients presented an important reduction in the number of *Prevotellaceae* (77.6%) together with an increase in the number of *Enterobacteriaceae*, compared with healthy population,

that result correlated with the severity of postural instability and gait trouble [85]. The reduction in the number of *Prevotellaceae* diminishes the levels of beneficial neuroactive SCFAs and decrease the biosynthesis of thiamine and folate, thus suggesting a cause of the lack of these vitamins in PD patients [86,87].

A decrease in *Prevotellaceae* in PD patients may be related to the reduction in mucin production defining an amplified intestinal permeability, known also as "leaky gut", that can be associated with  $\alpha$ -synuclein aggregates through the bacteria translocation and the production of bacterial products that induce inflammation [88] (i.e., gut-derived lipopolysaccharide (LPS) that can support the impairment of the blood-brain barrier [89]) and the reactive oxygen species production in the GI system thus starting the  $\alpha$ -synuclein accumulation at the level of ENS [90]. In an in vivo study, using rats injected at the level of substantia nigra with LPS, an induction of inflammation was observed, leading to damage of the nigrostriatal dopaminergic neurons, thus suggesting that this event could be implicated in neurodegeneration processes [91]. Moreover, Gorecki et al. [92] further investigated the effects of LPS in the communication between gut microbiota and  $\alpha$ -synuclein accumulation, overexpressing the gene of human  $\alpha$ -synuclein in mice. Firstly, authors showed that the levels of mucin-degrading Verrucomicrobiae and LPS-producing Gammaproteobacteria were higher in fecal samples of severe PD patients, while in mice overexpressing the human  $\alpha$ -synuclein, the number of *Verrucomicrobiae* was lower. Secondly, the researchers demonstrated that LPS exposure can alter the intestinal barrier function targeting cell membrane tight junctions. Moreover, LPS intake in in vivo studies using an  $\alpha$ -synucleinoverexpressing mouse model showed the manifestation of early motor impairment, thus supporting the hypothesis of proinflammatory gut microbiome environment as a leading cause for PD pathogenesis [92].

Recently, it has been demonstrated that gut microbiota transplants from parkinsonian mice into normal mice C57BL/6 was found to be associated with motor impairment and striatal neurotransmitter reduction. Specifically, sequencing of 16S rRNA showed that phylum *Firmicutes* and order *Clostridiales* diminished, while phylum *Proteobacteria, Turicibacterales* and *Enterobacteriales* were higher in fecal samples of parkinsonian mice, in parallel with enhanced fecal SCFAs. Notably, the fecal microbiota transplant (FMT) in MPTP-induced PD mice exerted neuroprotective effects, indeed, it was able to inhibit the activation of microglia and astrocytes in the *substantia nigra*, counteracting gut microbiome alterations, decreasing fecal SCFAs, alleviating physical impairment and increasing striatal dopamine and serotonin release [93].

Phage-related dysbiosis in PD is known, but recent findings suggest that phages can represent a leading cause of  $\alpha$ -synuclein misfolding and that lytic bacteriophages could have a pivotal role in PD onset [94]. For example, the lytic *Lactococcus* phages are more numerous in PD patients than in healthy people, which is related to a sharp decrease in *Lactococcus* bacteria [94] since these bacteria are able to release the neurotransmitter dopamine [95] and regulate gut permeability [96]. In fact, low levels of *Lactococcus* bacteria, triggered by the high amounts of strictly lytic phages in PD patients, might be involved in the generation of  $\alpha$ -synuclein misfolding [29,97,98].

A recent study demonstrated that oral administration of microbial metabolites in germ-free mice may lead to neuroinflammation causing the development of motor function alterations [99]. In particular, it is well-known that the gram-negative bacterium *Helicobacter pylori* is a leading cause of various GI problems, mainly peptic ulcers [100]. Moreover, many studies demonstrated a causal link between *Helicobacter pylori* and PD [101,102]. Numerous mechanisms were proposed to link *Helicobacter pylori* with PD pathogenesis: it could release toxins that affect the CNS or through glycosylation, generating cholesteryl glucosides with the same molecular structure of toxin cycads. These cholesteryl glucosides are neurotoxic and can go across the blood–brain barrier (BBB), leading to dopaminergic neurodegeneration [103,104]. Furthermore, *Helicobacter pylori* can activate the immune system through the activation of immune mediators, such as monocytes and determining the release of both interleukins and cytokines that may cause an important neuroinflammatory

response [105]. In line with this hypothesis, biopsies of colonic tissue form PD patients were analyzed, and it was revealed that in PD conditions there is an increased expression of pro-inflammatory cytokines, such as TNF- $\alpha$ , IFN- $\gamma$ , IL-6 and IL1- $\beta$  as well as the activation of enteric glial cells [106,107]. Neuroinflammation represents the leading cause of the disruption of the BBB, microglia activation and neurodegeneration and the presence of *Helicobacter pylori* may induce the production of autoantibodies against dopaminergic neurons that extend neuro-inflammatory processes [108]. Lastly, *Helicobacter pylori* is able to trigger apoptosis through the nitric oxide and mitochondrial Fas–FasL pathways that could lead to neurodegeneration [109].

An healthy microbiota composition is beneficial for competitive exclusion activities, immunological regulation and the production of substances such as vitamins, secondary bile acids and SCFAs [110]. Dietary fiber is used as a food source by a large number of colonic bacteria for the generation of metabolic bioproducts: in particular SCFAs [111].

Clinical studies reported a causal link between the decreased number of SCFAproducing bacteria (from the genera *Blautia*, *Coprococcus* and *Roseburia*) which contributes to the "leaky gut", and the increased number of opportunistic pathogens and carbohydratemetabolizing probiotics [112,113].

Moreover, a substantial decrease in acetate, propionate and butyrate in PD fecal samples compared with healthy controls has been observed [114]. This reduction in SCFA might have a crucial role in ENS alterations and gut dysmotility in PD. Additionally, the decrease in the number of *Prevotellaceae* and an increase in the number of *Lactobacilliceae* have been associated with lower concentrations of ghrelin in PD patients [115]. Ghrelin is a hormone produced by the gut involved in the homeostasis of the nigrostriatal dopamine function and PD patients show an impairment in ghrelin secretion [116,117].

Notably, it has been shown that carbohydrates themselves induce dopamine production from the brain by promoting the passage of tyrosine (dopamine precursor) across the BBB into cerebrospinal fluid [118]. Overall, a balanced diet of carbohydrate and protein mixture could ameliorate motor signs in PD patients [119].

Furthermore, many studies associated celiac disease, a gluten-induced GI disorder, with PD pathogenesis [120]. However, additional investigation must be conducted to clarify this association and the relevance of diet in PD.

Overall, the evidence reported suggests that gut microbiota is deeply altered in PD, as reported in clinical studies and in vivo models, and the normalization of this dysbiosis would open new therapeutic opportunities for this disorder, such as the use of nutraceutical approaches, including probiotics, prebiotics or synbiotics and microbiota transplantation approaches [115].

# 5. Diet and Gut Microbiota-Brain Axis in PD

Numerous epidemiological studies reported that diet affects (positively or negatively) the onset of neurodegenerative disorders, including PD. The PD microbiome is characterized by reduced production of SCFA and improved LPS and these alterations may promote the onset or exacerbation of PD [121]. As discussed above, diet strongly influences gut microbial composition, and the Western diet is correlated with enhanced risk for PD, while the Mediterranean diet (with high intake of dietary fiber [122])might be able to diminish PD risk [123].

In particular, studies on PD patients correlate total caloric intake of macronutrients and micronutrients with symptom severity, with greater caloric consumption related to worse symptoms [124]. Diets rich in animal saturated fat have been related to a higher risk of developing PD [125]. Other foods correlated with PD exacerbation include canned fruits and vegetables, soda, fried foods, processed food, ice cream and cheese (all typical of the Western diet). Mechanistically, this may be due to the high amount of LPS-containing bacteria in the intestinal microbiome which affects gut barrier integrity, leading to endotoxemia (i.e., systemic LPS), NLRP3 inflammasome activation, insulin resistance and mitochondrial impairment and gluconeogenesis [123]. Conversely, a "healthy" diet increased the number

of SCFA-producing bacteria and induced the release of components including glucagon-like peptide 1 (GLP-1) and brain-derived neurotrophic factor (BDNF), positively influencing PD risk [126].

The Mediterranean diet is related to a lower risk of PD onset [127]. Specific components of the Mediterranean diet are the reason of this positive effect, including fresh fruits and vegetables, nuts and other dried fruits, olive oil, wine and spices. Specifically, consumption of flavonoid-rich foods (berries, fruits, tea and wine) positively affects the risk of developing neurodegenerative disorders, including PD [128]. Polyunsaturated fatty acids (PUFA) are also inversely related to PD risk (elevated intake of  $\omega$ 3 fatty acids is linked to decreased PD risk) supporting the effects of fat consumption on the brain [129,130].

A daily diet enriched with plant carbohydrates and fiber is able to increase some particular macronutrients that PD patients lack. In contrast, a Western diet rich in refined carbohydrates and saturated fats, high fat goods and whole dairy products could lead to gut dysbiosis and may be implicated in PD pathogenesis [131,132]. Furthermore, antibiotics and microbial toxins produced by gut bacteria, comprising LPS and epoxomicin, may determine substantial variations in the gut microbiota and inflammation [133,134]. Several studies support the idea that differences in lifestyle are implicated in PD pathology. It has been proved that coffee and smoking may contribute to reducing the development of PD and this effect could be mediated by the gut microbiota. Beneficial effects of coffee and smoking could be due to the role of gut microbiota in mitigating intestinal inflammatory mechanisms [135]. Further studies showed that also red wine and tea may counteract PD predisposition [136].

While there are no therapeutic approaches that can avoid or delay PD by directly targeting the gut microbiota–brain axis, diet may influence both the gut microbiota–brain axis by modifying the microbiota composition and the neuronal functions of the ENS and CNS to ameliorate the progression of PD pathogenesis [137]. Recent investigations have shown that specific nutritional membrane precursors and cofactors are able to improve synaptic loss and membrane-related ENS and CNS impairments in PD and reduce motor and non-motor signs in preclinical studies [138,139]. The combination with the intake of prebiotic fiber may determine an amelioration in the treatment effects [126]. Moreover, oral administration of two circulating phosphatide precursors (uridine, and docosahexaenoic acid) was linked with an amelioration in dopaminergic neurotransmission, synaptic membrane formation and the density of dendritic spines [140–142].

## 6. Probiotics Interventions in PD

Numerous studies have shown that specific probiotics mixtures are able to restore gut microbiota and improve immune response [143]. Probiotics are live microorganisms that when administered in sufficient amounts can promote a restoration of gut microbiota and ameliorate immune homeostasis in the host [144].

The most commonly used probiotic bacteria are *Lactobacilli, Enterococci, Bifidobacteria,* yeasts and combinations of different beneficial bacteria [145]. Therapeutic and prophylactic effects exerted by probiotics intake are thought to be mediated throughout a broad range of mechanisms. Gut microbiota can be affected by probiotic supplement through competition with nutrients, adhesion to the intestinal epithelium, antagonism and cross-feeding [146]. Three-dimensional bacterial communities surrounded by self-produced extracellular matrices by probiotic bacteria (Biofilms) stimulate the colonization and extended duration in the GI system of the host and prevent the mucosal enrichment of pathogenic bacteria [147].

This process is caused by the release of organic acids (i.e., lactic acid by *Lactobacillus* and *Bifidobacterium* species) that lower GI pH, and bacteriocins that together can counteract pathogens' proliferation in the human GI system and urinary system [148]. Notably, treatment with *Lactobacillus* probiotics (in particular *Lactobacillus casei*) during *Helicobacter pylori* eradication therapy, ameliorated eradication efficacy, apparently through their antagonistic mechanisms against *H. pylori* [149]. Cross-feeding between probiotic bacteria and host microbiota can support the production of SCFAs such as butyrate in the gut [150].

Probiotics can also modulate a wide range of host immune functions that include both innate and adaptive (both cell-mediated and humoral) immunity. Particularly, probiotics are able to improve phagocytosis and enhance secretion of antibodies, generating increased immunological defenses against pathogens [151]. Additionally, probiotics can release a wide range of anti-inflammatory factors, downregulating pro-inflammatory cytokines [152], potentially counteracting intestinal inflammation. Moreover, probiotics can ameliorate GI barrier function [153]. For example, *Lactobacillus* and *Bifidobacterium* species can overexpress tight junction proteins and stimulate mucus secretion that can avoid the adhesion of detrimental microorganisms [154]. A growing body of evidence has demonstrated the protective effects exerted by probiotics in ameliorating intestinal epithelial integrity, counteracting barrier disruption, promoting healthy homeostasis of the mucosal immune system and blocking pathogenic bacterial proliferation [155,156].

Additionally, several strains of probiotic bacteria can stimulate intestinal motility and reduce GI alterations as demonstrated by a study in aging patients, where probiotics were able to modify bowel movements, reducing symptoms such as diarrhea and constipation [157]. Moreover, probiotics have a role in alleviating symptoms connected with lactose maldigestion due to the presence of enzymes such as  $\beta$ -galactosidase and bile salt hydrolase, which ameliorate lactose digestion in the host system [158].

Notably, many studies demonstrated that it is possible to regulate brain functions by ameliorating anxiety and depression with probiotic supplement. In an in vivo model of autism spectrum disorder (ASD), Hsiao et al., [149] indicated that *Bacteroides fragilis* administration counteracted the alterations in gut permeability and ameliorated ASD symptoms [159]. Further studies on animals showed that probiotic administration (i.e., *Lactobacillus plantarum, L. rhamnosus, B. longum*) can have anti-anxiolytic and antidepressant effects and modify cognitive activity [160].

In fact, it has been shown that probiotics can release a wide variety of bioactive compounds that can impact the host and its microbiota. Particularly they can release neuroactive compounds such as oxytocin, gamma-aminobutyric acid (GABA), serotonin, tryptophan, tryptamine, noradrenaline, dopamine, and acetylcholine [161].

Intake of specific probiotics also showed positive consequences for brain performances in clinical studies. The ingestion of *Lactobacillus casei* strain *Shirota* in chronic fatigue syndrome patients could drastically counteract anxiety [162].

Studies on the administration of probiotics for PD treatment are very limited. One study reported that PD patients with chronic constipation taking fermented milk containing *Lactobacillus casei Shirota* for five weeks ameliorated fecal consistency and diminished bloating and abdominal pain [163]. Although probiotics can represent a valuable tool to counteract alterations in PD microbiota composition and ameliorate GI function by reducing gut leakiness, bacterial translocation and related inflammation in the ENS, ameliorating GI functions with probiotics might not only improve GI function and/or protection of the GI system but also increase levodopa absorption and counteract motor and cognitive impairment including anxiety, depression and memory difficulties [164,165], which are common symptoms in PD patients.

The most commonly used probiotics such as *Lactobacilli, Enterococci, Bifidobacteria*, yeasts and specific mixtures [144,145] may modulate brain function by ameliorating anxiety and depression [162]. In fact, in clinical studies and in vivo models of PD, probiotics were able to alter the composition of gut microbiota and consequently may improve gastrointestinal function, neuroinflammation and even levodopa absorption [164].

Several in vivo and in vitro models were considered to study the neuroprotective effects of probiotics and their use as a potential treatment for PD [166]. In particular, mouse models are the most commonly used. Hsieh and his research group [167] compared the motor functions upon probiotic administration vs. vehicle in a MitoPark PD mouse model. The probiotic mixture was composed of six common probiotic strains (*Bifidobacterium bifidum*, *Bifidobacterium longum*, *Lactobacillus rhamnosus*, *L. rhamnosus GG*, *Lactobacillus plantarum LP28* and *Lactococcus lactis* subsp. *Lactis*) and they observed an amelioration in motor

performances of mice treated with the probiotic. In particular, mice showed better gait, balance and coordination from the 16th week after supplementation. Additionally, upon the treatment, they displayed decreased loss of dopaminergic neurons, thus suggesting a neuroprotective effect of the probiotics [167].

Similarly, neuroprotective effects of another probiotic combination containing *L. rhamnosus GG, Bifidobacterium animalis lactis,* and *Lactobacillus acidophilus* were observed in in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)- and rotenone toxin-induced PD mouse models [168]. In these models, the probiotic supplementation promoted the butyrate production, which plays a role in rescuing nigral dopaminergic neurons from MPTP- and rotenone-induced neurotoxicity. Moreover, high levels of BDNF and glial cell line-derived neurotrophic factor (GDNF) together with the inhibition of MAO-B were detected, which can lead to increased dopamine synthesis and the promotion of dopaminergic neurons survival thus helping cell survival and cell proliferation [168].

In another study, in 6-hydroxydopamine (6-OHDA) mice, a novel probiotic mix SLAB51 (sold as Sivomixx, composed of nine bacterial strains: *Streptococcus thermophilus*, *B. longum*, *Bifidobacterium breve*, *Bifidobacterium infantis*, *L. acidophilus*, *L. plantarum*, *Lactobacillus paracasei*, *Lactobacillus delbrueckii* subsp. *Bulgaricus* and *Lactobacillus*. *brevis*) was administered [169]. Notably, this formulation was associated with neuroprotection with a reduction in dopaminergic neuronal loss in the substantia nigra and striatum, assumed to be mediated through the activation of the peroxisome proliferator-activated receptor gamma (PPAR- $\gamma$ ) by microbial metabolites, thus leading to anti-inflammatory and antioxidant effects, as well as an increase in BDNF and consequently the activation of its pro-survival pathway [169].

Moreover, probiotics can be genetically manipulated to increase their beneficial effects. In a recent study, Fang et al. used *Lactococcus lactis cremori* carrying a GLP-1 expression vector as a treatment for an MPTP mice model [170]. Treated mice showed increased expression of tyrosine hydroxylase into the nigrostriatal pathway, reduced locomotor impairment and lower inflammation, compared with the control group. Moreover, the probiotic was also able to counteract the proliferation of intestinal pathogen Enterobacteriaceae, increasing the number of probiotic *Lactobacillus* and *Akkermansia* species. Interestingly, GLP-1 can cross the BBB and binds GLP-1 receptors in the brain. This insulin-signaling pathway is crucial in neurogenesis, neuronal metabolism and synaptic plasticity [170], and GLP-1 agonists are currently treatments of clinical trials on PD patients [171].

In a *Caenorhabditis elegans*  $\alpha$ -synuclein model of PD, treatment with *Bacillus subtilis* PXN21 was associated with a reduction in  $\alpha$ -synuclein accumulation in the host [172]. *Bacillus subtilis* PXN21 could exert neuroprotective effects through modifications of host sphingolipid metabolism. This results were in line with the hypothesis that an alteration in lipid metabolism, in particular ceramides and sphingolipids, contributes to PD pathogenesis [173]. Additionally, the beneficial effect of *B. subtilis* is partially due to a biofilm formation in the gut of the model [172]. Furthermore, an in vitro study demonstrated that, by co-culturing peripheral blood mononuclear cells isolated from PD patients with probiotic species (*Lactobacillus salivarius*, *L. plantarum*, *L. acidophilus*, *L. rhamnosus*, *Bifidobacterium animalis* subsp. *lactis* and *B. breve*), the release of pro-inflammatory cytokines was inhibited, in parallel with stimulation of the release of anti-inflammatory cytokines [174]. Among the tested probiotics, *L. salivarius* and *L. acidophilus* showed leading activities. Moreover, in this study, the probiotics were able to inhibit the proliferation of potentially pathogenic bacteria such as *Escherichia coli* and *Klebsiella pneumoniae* [174].

## 7. Prebiotics Intervention in PD

Also, prebiotics represent non-digestible compounds that may be beneficial for the host by modulating the gut microbiota [175]. A prebiotic is defined as "a substrate that is selectively used by host microorganisms exerting a health benefit". Most of the prebiotic are fermentable dietary fibers but not all the dietary fibers are prebiotics. Commonly, the consumption of a high percentage of fiber in the diet promotes an in increase in bacterial

diversity and leads to an expansion and/or an increase in the activity of beneficial bacteria (i.e., *Bifidobacterium* sp., *Lactobacillus* sp., *Akkermansia* sp., *Faecalibacterium* sp., *Roseburia* sp., *Bacteroides* sp. and *Prevotella*) together with a decrease in the number of detrimental bacteria (e.g., *Enterobacteriaceae*) [176].

Examples of prebiotics include pectins, inulin, fructo-oligosaccharides (FOS) and galacto-oligosaccharides (GOS). Particularly, the chemical characteristics of fibers, such as polymerization, solubility and viscosity determine the metabolism inside the GI tract, resulting in definite microbiota transformations after the ingestion [177].

Preclinical and clinical studies showed that the intake of wholegrain food that contains  $\beta$ -glucans (soluble non-starch polysaccharides) helps the growth of *Lactobacilli* and *Bifidobacteria* in rats and humans. It has been reported that supplementation with intact cereal fibers (i.e., wholegrain cereals, barley fibers, wheat bran and rye fibers) supported the growth of *Actinobacteria*, *Bifidobacterium*, *Clostridium*, *Lachnospira*, *Akkermansia*, and *Roseburia* in humans. Finally, the consumption of resistant starch determined the proliferation of *Bifidobacterium*, *Faecalibacterium* and *Eubacterium*, while reducing the amount of *Ruminococcus* strains [178,179].

Moreover, fiber solubility also has an impact on the gut microbiome. Soluble fiber has a stronger effect on microbial composition and diversity in piglets in comparison with insoluble fiber. However, cellulose—an insoluble and non-fermentable fiber and a source of fiber in fruit and vegetables—is transformed by *Ruminococcus* and *Fibrobacter*, which for this reason are called "cellulose-degrading microbes" [180]. In vivo studies have shown that cellulose intake leads to an increase in microbial species such as *Eptostreptococcaceae*, *Clostridiaceae*, *Akkermansia*, *Parabacteroides*, *Lactobacillus*, *Clostridium*, *Eisenbergiella*, *Marvinbryantia*, *Romboutsia*, *Helicobacter*, *Enterococcus* and *Desulfovibrio* together with a lower proliferation of *Sutterellaceae*, *Lactobacillaceae* and *Coriobacteriaceae* [43].

In addition to the effects on microbiota composition, dietary fibers have a role in microbial enzymatic function and in metabolite absorption. Chemical properties, including fiber solubility and fermentability influence the degree and location of microbial fermentation and which type of metabolite is produced. Two important fibers are GOS, based on lactose and FOS, synthesized from fructose [181]. GOS and FOS arrive to the colon mainly unaltered and are mostly transformed by *Bifidobacteria*. Metabolic products such as SCFA, lactose, hydrogen, methane and carbon dioxide induce an acidic milieu in the colon, which leads to death, or reduced multiplication, of deleterious bacteria [182].

A large number of clinical studies demonstrated that lowering carbohydrate intake or wholegrain cereals reduced the amount of butyrate-producing bacteria, such as *Bifidobacteria*, as well as SCFAs themselves [183]. Soluble and fermentable fiber can intensify the microbiota enzymatic activity to transform complex carbohydrates in health-promoting SCFAs such as acetate, propionate and butyrate. These SCFAs, in particular butyrate, have been involved in colonocyte metabolism, thus helping intestinal barrier functionality, in glucose homeostasis, lipid oxidation and they have anti-inflammatory and mucosal immunomodulatory effects [177].

While insoluble fibers (i.e., cellulose) are not implicated in SCFA production, it has been observed that they probably have a role in the linoleic acid, nicotinate and nicotinamide, glycerophospholipid, glutathione and sphingolipid pathways as well as the valine, leucine and isoleucine metabolic pathways [184].

In fact, through computational-experimental framework a relationship between PD and branched-chain amino acid transferase 1 (BCAT-1) was found. This enzyme is involved in the first step of branched chain amino acid (BCAA) catabolism [185] and it has been discovered that BCAT-1 levels are usually high in PD-susceptible regions of the healthy human brain, and that its expression is lower in the substantia nigra of sporadic parkinsonian patients [186]. While this correlation has been observed, further studies on in vivo models are needed to clarify the underlying mechanisms. Moreover, glutathione exerts anti-oxidant effects reducing reactive oxygen species (ROS) [187] and glutathione S-Transferases (GSTs) enzymes catalyze the conjugation of glutathione to various electrophiles and the role of

GSTs in the protection of dopaminergic neurons has been examined using several models of Parkinson's disease [188].

Prebiotic fibers may be beneficial in the activity of the immune system, bowel mobility and constipation. For these reasons, enriching the diet with prebiotics might be beneficial for inflammation and GI alterations occurring in PD patients. Furthermore, it has been observed that GOS and FOS determine an increase in BDNF levels at the level of the dentate gyrus of the hippocampus in rats [189]. Since BDNF is a neurotrophin implicated in neuronal protection, survival and plasticity, GOS and FOS supplementation in the diet might affect brain health. Despite all the evidence reported, the use of prebiotics in patients with PD has not yet been investigated but since PD patients present a lower abundance of SCFA butyrate-producing bacteria, prebiotic fibers may be used as a supplement to correct this dysbiosis [190,191]. Notably, SCFAs are able to activate microglia, inducing T-regulatory cells to increase cytokine release to regulate neuroinflammatory mechanisms [99].

#### 8. Synbiotic Intervention in PD

The concept of synbiotic indicates food components or dietary supplements fusing probiotics and prebiotics [192]. In particular, synbiotics arise from the necessity to overcome possible probiotics survival difficulties, for this reason in these formulations the prebiotic compound must selectively promote the activity and the survival during the passage through the upper GI tract of the probiotic fraction [193]. Synbiotics are beneficial by favoring the survival and implantation of microbial supplement in the GI system. In fact, they selectively induce the proliferation and activate the metabolism of a small group of healthy bacteria. A wide range of factors such as pH, hydrogen peroxide, organic acids, oxygen and moisture stress affect probiotic viability [194]. Most commonly used probiotic strains in synbiotic formulations include *Lactobacilli*, *Bifidobacteria* spp., *S. boulardii* and *B. coagulans*, while the prebiotics used include oligosaccharides such as FOS, GOS and xyloseoligosaccharide (XOS), inulin and prebiotics from food like chicory and yacon roots. [66].

The positive effects exerted by synbiotic intake in clinical investigations comprise: (1) balanced gut microbiota increasing the levels of *Lactobacilli* and *Bifidobacteria*, (2) amelioration of liver function in cirrhotic patients, (3) enhancement of immune system function and (4) inhibition of bacterial translocation and decreased occurrence of nosocomial infections in patients after surgery [195].

Constipation is one of the main symptoms in PD patients affecting their quality of life. Synbiotics may be useful for PD-related non-motor side effects by ameliorating immune function, dysbiosis and bowel functions. In a clinical study, *Lactobacillus salivarius* was able to decrease inflammatory markers in healthy subjects with a higher effect in combination with prebiotics [196]. In a different investigation, treatment with synbiotic yogurt containing *Bifidobacterium animalis* and prebiotics in females with constipation, produced increased gut movement and defecation, compared to controls [197]. Moreover, it has been assessed that the daily intake of a fermented milk containing multiple probiotic strains and prebiotic fiber for four weeks was able to increase in the number of complete bowel movements in patients with PD [124].

Another main symptom of PD is small intestinal bacterial overgrowth (SIBO) and patients that are SIBO positive usually present increased motor dysfunction [198]. Khalighi et al., [199] demonstrated that the association between antibiotic treatment and synbiotic supplementation containing *Bacillus coagulans* and prebiotic ameliorated the treatment response. Moreover, it decreased abdominal pain, flatulence and diarrhea [199].

Overall, the evidence reported in this review support the potential of probiotic, prebiotic and synbiotic supplementation in PD patients.

## 9. Discussion and Conclusions

PD is a one of the most common neurodegenerative disorders, characterized by motor and non-motor sign and symptoms, including gut dysfunctions, which may appear before the motor symptoms. PD underlying mechanisms involve increased oxidative stress and neuroinflammation [200]. So far, the existing therapies can alleviate PD-associated symptoms, but there is no cure to control the development and exacerbation of this disorder. Accumulating evidence suggests a crucial role of gut microbiota and an influence on the CNS, via the gut–brain axis, mediating different pathways. In particular, a healthy microbiota is correlated with lower risk of developing CNS disorders, including PD, while microbiota dysbiosis is correlated with higher incidence of PD. Diet may influence both positively and negatively the development of neurodegenerative disorders. Specifically, the Mediterranean diet (rich in fibers, flavonoids and PUFA) has positive effects on the gut microbiome and thus may reduce the development or exacerbation of PD; on the other hand, the Western diet (rich in meat, processed food and fried food) could lead to detrimental effects on the gut–brain axis (Figure 1).



**Figure 1.** Representative scheme of the gut–brain axis. The influence of different diets and the potential effects of probiotics, synbiotics and prebiotics on gut microbiota. PD: Parkinson's Disease.

Numerous preclinical and clinical studies suggested that dietary interventions with prebiotics, probiotics or synbiotics, by modifying the microbiome composition, may improve brain health and decrease the risk of developing PD (Figure 1).

Dietary interventions are of high importance in particular at the very early stages of PD. Patients with PD may experience non-motor symptoms at early stages such as constipation, dysphagia, hyposmia and depression, that may influence dietary choices and thus may be responsible for the alterations of nutritional status reported in PD [201,202]. For example, as we mentioned above, PD patients are affected by constipation for more than 20 years before the onset of motor symptoms, therefore the use of nutraceutical interventions at this stage, including prebiotics, probiotics or symbiotics may be of high relevance [127,203].

Further studies in PD should take into consideration the role of the gut-brain axis and a deeper investigation into the underlying mechanism is required. Moreover, additional studies are needed to define the potential beneficial effects of the use of prebiotics, probiotic and synbiotics in maintaining protein and oxidative homeostasis in the ENS and to better understand the biochemical influences of these interventions on people affected by neurodegenerative diseases. Another point to be considered is whether constant exposure to probiotics, prebiotics or synbiotics affects the gut microbiome composition in a long-term way or, once the intervention is ceased, the microbiome composition would revert.

Also, it is important to design studies considering the duration of intervention, the dosages and the combination of different interventions. Finally, in this scenario, it is crucial to define the best approach based on prebiotics, probiotics or synbiotics for PD and, analyzing the specific gut microbiome composition of a single patient, could help in creating a personalized therapy. Overall, we can postulate that prebiotics, probiotics and symbiotics may represent a potential therapeutic approach for PD.

**Author Contributions:** Conceptualization, V.C. and M.d.; methodology, M.d. and V.C.; software, M.d.; writing—original draft preparation, M.A.; writing—review and editing, V.C. and M.d.; funding acquisition, V.C; figure preparation, M.A. All authors have read and agreed to the published version of the manuscript.

**Funding:** AMCEN 2020-2021: "Progetti di avvio alla ricerca" funded by University of L'Aquila granted by V.C.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

#### References

- 1. Mhyre, T.R.; Boyd, J.T.; Hamill, R.W.; Maguire-Zeiss, K.A. Parkinson's Disease. Subcell. Biochem. 2012, 65, 389–455. [CrossRef]
- Kujawska, M.; Jodynis-Liebert, J. What Is the Evidence that Parkinson's Disease Is a Prion Disorder, Which Originates in the Gut? Int. J. Mol. Sci. 2018, 19, 3573. [CrossRef] [PubMed]
- 3. Büeler, H. Impaired mitochondrial dynamics and function in the pathogenesis of Parkinson's disease. *Exp. Neurol.* **2009**, *218*, 235–246. [CrossRef]
- 4. Dias, V.; Junn, E.; Mouradian, M.M. The Role of Oxidative Stress in Parkinson's Disease. J. Parkinsons Dis. 2013, 3, 461–491. [CrossRef] [PubMed]
- Monaco, A.; Fraldi, A. Protein Aggregation and Dysfunction of Autophagy-Lysosomal Pathway: A Vicious Cycle in Lysosomal Storage Diseases. Front. Mol. Neurosci. 2020, 13, 37. [CrossRef] [PubMed]
- Troncoso-Escudero, P.; Parra, A.; Nassif, M.; Vidal, R.L. Outside in: Unraveling the Role of Neuroinflammation in the Progression of Parkinson's Disease. *Front. Neurol.* 2018, 9, 860. [CrossRef]
- 7. Váradi, C. Clinical Features of Parkinson's Disease: The Evolution of Critical Symptoms. Biology 2020, 9, 103. [CrossRef]
- Reichmann, H.; Schneider, C.; Löhle, M. Non-motor features of Parkinson's disease: Depression and dementia. *Park. Relat. Disord.* 2009, 15 (Suppl. 3), S87–S92. [CrossRef]
- 9. Idiaquez, J.; Benarroch, E.E.; Rosales, H.; Milla, P.; Ríos, L. Autonomic and Cognitive dysfunction in Parkinson's disease. *Clin. Auton. Res.* 2007, 17, 93–98. [CrossRef] [PubMed]
- Armstrong, M.J.; Okun, M.S. Diagnosis and Treatment of Parkinson Disease: A Review. JAMA 2020, 323, 548–560. [CrossRef] [PubMed]
- 11. Riederer, P.; Müller, T. Monoamine oxidase-B inhibitors in the treatment of Parkinson's disease: Clinical–pharmacological aspects. J. Neural Transm. 2018, 125, 1751–1757. [CrossRef]
- 12. Charvin, D.; Medori, R.; Hauser, R.A.; Rascol, O. Therapeutic strategies for Parkinson disease: Beyond dopaminergic drugs. *Nat. Rev. Drug Discov.* **2018**, *17*, 804–822. [CrossRef]
- 13. Waller, D.G.; Roseveare, C.; Renwick, A.G.; Macklin, B.; George, C.F. Gastric emptying in healthy volunteers after multiple doses of levodopa. *Br. J. Clin. Pharmacol.* **1991**, *32*, 691–695.
- 14. Djaldetti, R.; Baron, J.; Ziv, I.; Melamed, E. Gastric emptying in Parkinson's disease: Patients with and without response fluctuations. *Neurology* **1996**, *46*, 1051–1054. [CrossRef]
- 15. LeWitt, P.A. Levodopa therapy for Parkinson's disease: Pharmacokinetics and pharmacodynamics. *Mov. Disord.* **2015**, *30*, 64–72. [CrossRef]
- 16. Pfeiffer, R.F.; Isaacson, S.H.; Pahwa, R. Clinical implications of gastric complications on levodopa treatment in Parkinson's disease. *Park. Relat. Disord.* **2020**, *76*, 63–71. [CrossRef]
- Hardoff, R.; Sula, M.; Tamir, A.; Soil, A.; Front, A.; Badarna, S.; Honigman, S.; Giladi, N. Gastric emptying time and gastric motility in patients with Parkinson's disease. *Mov. Disord.* 2001, *16*, 1041–1047. [CrossRef] [PubMed]

- 18. Marsili, L.; Bologna, M.; Miyasaki, J.M.; Colosimo, C. Parkinson's disease advanced therapies—A systematic review: More unanswered questions than guidance. *Park. Relat. Disord.* **2020**, *83*, 132–139. [CrossRef] [PubMed]
- 19. Schlesinger, I.; Sinai, A.; Zaaroor, M. MRI-Guided Focused Ultrasound in Parkinson's Disease: A Review. *Park. Dis.* 2017, 2017, 8124624. [CrossRef] [PubMed]
- Carabotti, M.; Scirocco, A.; Maselli, M.A.; Severi, C. The gut-brain axis: Interactions between enteric microbiota, central and enteric nervous systems. *Ann. Gastroenterol.* 2015, 28, 203–209.
- 21. McCorry, L.K. Physiology of the Autonomic Nervous System. Am. J. Pharm. Educ. 2007, 71, 78. [CrossRef]
- 22. Tsigos, C.; Chrousos, G.P. Hypothalamic–pituitary–adrenal axis, neuroendocrine factors and stress. *Psychosomatics* **2002**, *53*, 865–871. [CrossRef]
- 23. Herman, J.P.; McKlveen, J.M.; Ghosal, S.; Kopp, B.; Wulsin, A.; Makinson, R.; Scheimann, J.; Myers, B. Regulation of the Hypothalamic-Pituitary-Adrenocortical Stress Response. *Compr. Physiol.* **2016**, *6*, 603–621. [CrossRef] [PubMed]
- Souza-Talarico, J.N.; Marin, M.-F.; Sindi, S.; Lupien, S.J. Effects of stress hormones on the brain and cognition: Evidence from normal to pathological aging. *Dement. Neuropsychol.* 2011, 5, 8–16. [CrossRef]
- Mayer, E.A.; Savidge, T.; Shulman, R.J. Brain–Gut Microbiome Interactions and Functional Bowel Disorders. *Gastroenterology* 2014, 146, 1500–1512. [CrossRef] [PubMed]
- 26. Poirier, A.-A.; Aubé, B.; Côté, M.; Morin, N.; di Paolo, T.; Soulet, D. Gastrointestinal Dysfunctions in Parkinson's Disease: Symptoms and Treatments. *Park. Dis.* **2016**, 2016, 6762528. [CrossRef]
- 27. Kim, J.-S.; Sung, H.-Y. Gastrointestinal Autonomic Dysfunction in Patients with Parkinson's Disease. J. Mov. Disord. 2015, 8, 76–82. [CrossRef]
- Rietdijk, C.D.; Perez-Pardo, P.; Garssen, J.; van Wezel, R.J.A.; Kraneveld, A.D. Exploring Braak's Hypothesis of Parkinson's Disease. *Front. Neurol.* 2017, *8*, 37. [CrossRef] [PubMed]
- Braak, H.; Rüb, U.; Gai, W.P.; del Tredici, K. Idiopathic Parkinson's disease: Possible routes by which vulnerable neuronal types may be subject to neuroinvasion by an unknown pathogen. J. Neural. Transm. 2003, 110, 517–536. [CrossRef] [PubMed]
- del Tredici, K.; Braak, H. Review: Sporadic Parkinson's disease: Development and distribution ofα-synuclein pathology. *Neuropathol. Appl. Neurobiol.* 2016, 42, 33–50. [CrossRef]
- Abu Aboud, O.; Donohoe, D.; Bultman, S.; Fitch, M.; Riiff, T.; Hellerstein, M.; Weiss, R.H. PPARα inhibition modulates multiple reprogrammed metabolic pathways in kidney cancer and attenuates tumor growth. *Am. J. Physiol. Physiol.* 2015, 308, C890–C898. [CrossRef] [PubMed]
- Liu, B.; Fang, F.; Pedersen, N.L.; Tillander, A.; Ludvigsson, J.F.; Ekbom, A.; Svenningsson, P.; Chen, H.; Wirdefeldt, K. Vagotomy and Parkinson disease: A Swedish register-based matched-cohort study. *Neurology* 2017, *88*, 1996–2002. [CrossRef]
- 33. Liu, Y.; Forsythe, P. Vagotomy and insights into the microbiota-gut-brain axis. *Neurosci. Res.* **2021**, *168*, 20–27. [CrossRef] [PubMed]
- Çelik, T.; Kamişli, Ö.; Babür, C.; Çevik, M.; Öztuna, D.; Altinayar, S. Is there a relationship between Toxoplasma gondii infection and idiopathic Parkinson's disease? *Scand. J. Infect. Dis.* 2010, *42*, 604–608. [CrossRef] [PubMed]
- 35. Prandovszky, E.; Gaskell, E.; Martin, H.; Dubey, J.P.; Webster, J.P.; McConkey, G.A. The Neurotropic Parasite Toxoplasma Gondii Increases Dopamine Metabolism. *PLoS ONE* **2011**, *6*, e23866. [CrossRef]
- 36. Beach, T.G.; Adler, C.H.; Sue, L.I.; Shill, H.A.; Driver-Dunckley, E.; Mehta, S.H.; Intorcia, A.J.; Glass, M.J.; Walker, J.E.; Arce, R.; et al. Vagus Nerve and Stomach Synucleinopathy in Parkinson's Disease, Incidental Lewy Body Disease, and Normal Elderly Subjects: Evidence against the "Body-First" Hypothesis. J. Park. Dis. 2021, 11, 1833–1843. [CrossRef]
- Schaeffer, E.; Kluge, A.; Böttner, M.; Zunke, F.; Cossais, F.; Berg, D.; Arnold, P. Alpha Synuclein Connects the Gut-Brain Axis in Parkinson's Disease Patients—A View on Clinical Aspects, Cellular Pathology and Analytical Methodology. *Front. Cell Dev. Biol.* 2020, *8*, 573696. [CrossRef]
- Pan-Montojo, F.; Schwarz, M.; Winkler, C.; Arnhold, M.; O'Sullivan, G.; Pal, A.; Said, J.W.; Marsico, G.; Verbavatz, J.-M.; Rodrigo-Angulo, M.; et al. Environmental toxins trigger PD-like progression via increased alpha-synuclein release from enteric neurons in mice. *Sci. Rep.* 2012, *2*, 898. [CrossRef]
- Pan-Montojo, F.; Anichtchik, O.; Dening, Y.; Knels, L.; Pursche, S.; Jung, R.; Jackson, S.; Gille, G.; Spillantini, M.G.; Reichmann, H.; et al. Progression of Parkinson's Disease Pathology Is Reproduced by Intragastric Administration of Rotenone in Mice. *PLoS* ONE 2010, 5, e8762. [CrossRef]
- 40. Cannon, J.; Greenamyre, J.T. The Role of Environmental Exposures in Neurodegeneration and Neurodegenerative Diseases. *Toxicol. Sci.* 2011, 124, 225–250. [CrossRef]
- 41. Scarpellini, E.; Ianiro, G.; Attili, F.; Bassanelli, C.; de Santis, A.; Gasbarrini, A. The human gut microbiota and virome: Potential therapeutic implications. *Dig. Liver Dis.* **2015**, *47*, 1007–1012. [CrossRef]
- 42. Thursby, E.; Juge, N. Introduction to the human gut microbiota. Biochem. J. 2017, 474, 1823–1836. [CrossRef]
- Rinninella, E.; Raoul, P.; Cintoni, M.; Franceschi, F.; Miggiano, G.A.D.; Gasbarrini, A.; Mele, M.C. What Is the Healthy Gut Microbiota Composition? A Changing Ecosystem across Age, Environment, Diet, and Diseases. *Microorganisms* 2019, 7, 14. [CrossRef]
- 44. Sekirov, I.; Russell, S.L.; Antunes, L.C.M.; Finlay, B.B. Gut Microbiota in Health and Disease. *Physiol. Rev.* **2010**, *90*, 859–904. [CrossRef]

- 45. Singh, R.K.; Chang, H.-W.; Yan, D.; Lee, K.M.; Ucmak, D.; Wong, K.; Abrouk, M.; Farahnik, B.; Nakamura, M.; Zhu, T.H.; et al. Influence of diet on the gut microbiome and implications for human health. *J. Transl. Med.* **2017**, *15*, 73. [CrossRef] [PubMed]
- 46. Rowland, I.; Gibson, G.; Heinken, A.; Scott, K.; Swann, J.; Thiele, I.; Tuohy, K. Gut microbiota functions: Metabolism of nutrients and other food components. *Eur. J. Nutr.* 2018, 57, 1–24. [CrossRef]
- 47. Conlon, M.A.; Bird, A.R. The Impact of Diet and Lifestyle on Gut Microbiota and Human Health. *Nutrients* **2014**, *7*, 17–44. [CrossRef] [PubMed]
- 48. Yang, Q.; Liang, Q.; Balakrishnan, B.; Belobrajdic, D.P.; Feng, Q.-J.; Zhang, W. Role of Dietary Nutrients in the Modulation of Gut Microbiota: A Narrative Review. *Nutrients* 2020, 12, 381. [CrossRef] [PubMed]
- 49. Sonnenburg, J.L.; Bäckhed, F. Diet-microbiota interactions as moderators of human metabolism. *Nature* **2016**, *535*, 56–64. [CrossRef]
- 50. David, L.A.; Materna, A.C.; Friedman, J.; Campos-Baptista, M.I.; Blackburn, M.C.; Perrotta, A.; Erdman, S.E.; Alm, E.J. Host lifestyle affects human microbiota on daily timescales. *Genome Biol.* **2014**, *15*, R89. [CrossRef] [PubMed]
- 51. Wu, G.D.; Chen, J.; Hoffmann, C.; Bittinger, K.; Chen, Y.-Y.; Keilbaugh, S.A.; Bewtra, M.; Knights, D.; Walters, W.A.; Knight, R.; et al. Linking long-term dietary patterns with gut microbial enterotypes. *Science* **2011**, *334*, 105–108. [CrossRef]
- 52. David, L.A.; Maurice, C.F.; Carmody, R.N.; Gootenberg, D.B.; Button, J.E.; Wolfe, B.E.; Ling, A.V.; Devlin, A.S.; Varma, Y.; Fischbach, M.A.; et al. Diet rapidly and reproducibly alters the human gut microbiome. *Nature* **2014**, *505*, 559–563. [CrossRef]
- 53. Liang, X.; Fitzgerald, G.A. Timing the Microbes: The Circadian Rhythm of the Gut Microbiome. J. Biol. Rhythm. 2017, 32, 505–515. [CrossRef]
- Parkar, S.G.; Kalsbeek, A.; Cheeseman, J.F. Potential Role for the Gut Microbiota in Modulating Host Circadian Rhythms and Metabolic Health. *Microorganisms* 2019, 7, 41. [CrossRef] [PubMed]
- 55. Hentges, D.J.; Maier, B.R.; Burton, G.C.; A Flynn, M.; Tsutakawa, R.K. Effect of a high-beef diet on the fecal bacterial flora of humans. *Cancer Res.* **1977**, *37*, 568–571.
- 56. Wiątecka, D.; Narbad, A.; Ridgway, P.K.; Kostyra, H. The study on the impact of glycated pea proteins on human intestinal bacteria. *Int. J. Food Microbiol.* **2011**, *145*, 267–272. [CrossRef]
- Kim, C.H.; Park, J.; Kim, M. Gut Microbiota-Derived Short-Chain Fatty Acids, T Cells, and Inflammation. *Immune Netw.* 2014, 14, 277–288. [CrossRef] [PubMed]
- 58. Reddy, B.S.; Weisburger, J.H.; Wynder, E.L. Effects of High Risk and Low Risk Diets for Colon Carcinogenesis on Fecal Microflora and Steroids in Man. J. Nutr. 1975, 105, 878–884. [CrossRef]
- 59. Cotillard, A.; Kennedy, S.P.; Kong, L.C.; Prifti, E.; Pons, N.; Le Chatelier, E.; Almeida, M.; Quinquis, B.; Levenez, F.; Galleron, N.; et al. Dietary intervention impact on gut microbial gene richness. *Nature* **2013**, *500*, 585–588. [CrossRef]
- 60. Perezjimenez, J.; Neveu, V.; Vos, F.; Scalbert, A. Identification of the 100 richest dietary sources of polyphenols: An application of the Phenol-Explorer database. *Eur. J. Clin. Nutr.* **2010**, *64*, S112–S120. [CrossRef]
- 61. Eid, N.; Enani, S.; Walton, G.; Corona, G.; Costabile, A.; Gibson, G.; Rowland, I.; Spencer, J.P. The impact of date palm fruits and their component polyphenols, on gut microbial ecology, bacterial metabolites and colon cancer cell proliferation. *J. Nutr. Sci.* 2014, *3*, e46. [CrossRef] [PubMed]
- 62. Cuervo, A.; Valdés, L.; Salazar, N.; Reyes-Gavilán, C.G.D.L.; Ruas-Madiedo, P.; Gueimonde, M.; González, S. Pilot Study of Diet and Microbiota: Interactive Associations of Fibers and Polyphenols with Human Intestinal Bacteria. *J. Agric. Food Chem.* **2014**, *62*, 5330–5336. [CrossRef]
- Queipo-Ortuño, M.I.; Boto-Ordóñez, M.; Murri, M.; Gomez-Zumaquero, J.M.; Clemente-Postigo, M.; Estruch, R.; Cardona Diaz, F.; Andrés-Lacueva, C.; Tinahones, F.J. Influence of red wine polyphenols and ethanol on the gut microbiota ecology and biochemical biomarkers. Am. J. Clin. Nutr. 2012, 95, 1323–1334. [CrossRef]
- 64. Jin, J.-S.; Touyama, M.; Hisada, T.; Benno, Y. Effects of green tea consumption on human fecal microbiota with special reference to Bifidobacterium species: Effects of Green Tea on Fecal Microbiota. *Microbiol. Immunol.* **2012**, *56*, 729–739. [CrossRef]
- Tzounis, X.; Rodriguez-Mateos, A.; Vulevic, J.; Gibson, G.R.; Kwik-Uribe, C.; Spencer, J.P. Prebiotic evaluation of cocoa-derived flavanols in healthy humans by using a randomized, controlled, double-blind, crossover intervention study. *Am. J. Clin. Nutr.* 2010, 93, 62–72. [CrossRef]
- Pandey, K.R.; Naik, S.R.; Vakil, B.V. Probiotics, prebiotics and synbiotics—A review. J. Food Sci. Technol. 2015, 52, 7577–7587. [CrossRef] [PubMed]
- 67. Parkar, S.G.; Stevenson, D.E.; Skinner, M.A. The potential influence of fruit polyphenols on colonic microflora and human gut health. *Int. J. Food Microbiol.* **2008**, *124*, 295–298. [CrossRef]
- Cueva, C.; Sánchez-Patán, F.; Monagas, M.; Walton, G.E.; Gibson, G.R.; Martín-Álvarez, P.J.; Bartolomé, B.; Moreno-Arribas, M.V. In vitro fermentation of grape seed flavan-3-ol fractions by human faecal microbiota: Changes in microbial groups and phenolic metabolites. *FEMS Microbiol. Ecol.* 2013, *83*, 792–805. [CrossRef]
- 69. Lee, H.C.; Jenner, A.M.; Low, C.S.; Lee, Y.K. Effect of tea phenolics and their aromatic fecal bacterial metabolites on intestinal microbiota. *Res. Microbiol.* **2006**, *157*, 876–884. [CrossRef] [PubMed]
- Natividad, J.M.; Lamas, B.; Pham, H.P.; Michel, M.-L.; Rainteau, D.; Bridonneau, C.; da Costa, G.; van Hylckama Vlieg, J.; Sovran, B.; Chamignon, C.; et al. Bilophila wadsworthia aggravates high fat diet induced metabolic dysfunctions in mice. *Nat. Commun.* 2018, *9*, 2802. [CrossRef]

- 71. Caesar, R.; Tremaroli, V.; Kovatcheva-Datchary, P.; Cani, P.D.; Bäckhed, F. Crosstalk between Gut Microbiota and Dietary Lipids Aggravates WAT Inflammation through TLR Signaling. *Cell Metab.* **2015**, *22*, 658–668. [CrossRef] [PubMed]
- 72. Lee, E.; Lee, J.-E. Impact of drinking alcohol on gut microbiota: Recent perspectives on ethanol and alcoholic beverage. *Curr. Opin. Food Sci.* **2021**, *37*, 91–97. [CrossRef]
- Mutlu, E.A.; Gillevet, P.M.; Rangwala, H.; Sikaroodi, M.; Naqvi, A.; Engen, P.A.; Kwasny, M.; Lau, C.K.; Keshavarzian, A. Colonic microbiome is altered in alcoholism. *Am. J. Physiol. Liver Physiol.* 2012, 302, G966–G978. [CrossRef]
- 74. Schnabl, B.; Brenner, D. Interactions between the Intestinal Microbiome and Liver Diseases. *Gastroenterology* **2014**, *146*, 1513–1524. [CrossRef] [PubMed]
- 75. Wang, H.; Yan, Y.; Yi, X.; Duan, Y.; Wang, J.; Li, S.; Luo, L.; Huang, T.; Inglis, B.; Li, X.; et al. Histopathological Features and Composition of Gut Microbiota in Rhesus Monkey of Alcoholic Liver Disease. *Front. Microbiol.* **2019**, *10*, 165. [CrossRef]
- 76. Ferrere, G.; Wrzosek, L.; Cailleux, F.; Turpin, W.; Puchois, V.; Spatz, M.; Ciocan, D.; Rainteau, D.; Humbert, L.; Hugot, C.; et al. Fecal microbiota manipulation prevents dysbiosis and alcohol-induced liver injury in mice. *J. Hepatol.* 2017, 66, 806–815. [CrossRef] [PubMed]
- 77. Lee, J.-E.; Ha, J.S.; Park, H.-Y.; Lee, E. Alteration of gut microbiota composition by short-term low-dose alcohol intake is restored by fermented rice liquor in mice. *Food Res. Int.* **2020**, *128*, 108800. [CrossRef]
- Yang, X.; Qian, Y.; Xu, S.; Song, Y.; Xiao, Q. Longitudinal Analysis of Fecal Microbiome and Pathologic Processes in a Rotenone Induced Mice Model of Parkinson's Disease. *Front. Aging Neurosci.* 2018, 9, 441. [CrossRef]
- 79. Li, W.; Wu, X.; Hu, X.; Wang, T.; Liang, S.; Duan, Y.; Jin, F.; Qin, B. Structural changes of gut microbiota in Parkinson's disease and its correlation with clinical features. *Sci. China Life Sci.* **2017**, *60*, 1223–1233. [CrossRef]
- 80. Santos, S.F.; de Oliveira, H.L.; Yamada, E.S.; Neves, B.; Pereira, A., Jr. The Gut and Parkinson's Disease—A Bidirectional Pathway. *Front. Neurol.* **2019**, *10*, 574. [CrossRef]
- Breen, D.P.; Halliday, G.M.; Lang, A.E. Gut–brain axis and the spread of α-synuclein pathology: Vagal highway or dead end? Mov. Disord. 2019, 34, 307–316. [CrossRef]
- 82. Sun, M.-F.; Shen, Y.-Q. Dysbiosis of gut microbiota and microbial metabolites in Parkinson's Disease. *Ageing Res. Rev.* 2018, 45, 53–61. [CrossRef]
- 83. Scheperjans, F.; Derkinderen, P.; Borghammer, P. The Gut and Parkinson's Disease: Hype or Hope? *J. Park. Dis.* **2018**, *8*, S31–S39. [CrossRef]
- Clairembault, T.; Leclair-Visonneau, L.; Neunlist, M.; Derkinderen, P. Enteric glial cells: New players in Parkinson's disease? Enteric Glia in PD. *Mov. Disord.* 2015, 30, 494–498. [CrossRef]
- 85. Scheperjans, F.; Aho, V.; Pereira, P.A.; Koskinen, K.; Paulin, L.; Pekkonen, E.; Haapaniemi, E.; Kaakkola, S.; Eerola-Rautio, J.; Pohja, M.; et al. Gut microbiota are related to Parkinson's disease and clinical phenotype. *Mov. Disord.* **2015**, *30*, 350–358. [CrossRef]
- Brown, C.T.; Davis-Richardson, A.G.; Giongo, A.; Gano, K.A.; Crabb, D.B.; Mukherjee, N.; Casella, G.; Drew, J.C.; Ilonen, J.; Knip, M.; et al. Gut Microbiome Metagenomics Analysis Suggests a Functional Model for the Development of Autoimmunity for Type 1 Diabetes. *PLoS ONE* 2011, 6, e25792. [CrossRef]
- 87. Forsyth, C.B.; Shannon, K.M.; Kordower, J.H.; Voigt, R.M.; Shaikh, M.; Jaglin, J.A.; Estes, J.D.; Dodiya, H.B.; Keshavarzian, A. Increased intestinal permeability correlates with sigmoid mucosa alpha-synuclein staining and endotoxin exposure markers in early Parkinson's disease. *PLoS ONE* **2011**, *6*, e28032. [CrossRef]
- Caputi, V.; Giron, M.C. Microbiome-Gut-Brain Axis and Toll-Like Receptors in Parkinson's Disease. *Int. J. Mol. Sci.* 2018, 19, 1689. [CrossRef] [PubMed]
- Banks, W.A.; Gray, A.M.; Erickson, M.A.; Salameh, T.S.; Damodarasamy, M.; Sheibani, N.; Meabon, J.S.; Wing, E.E.; Morofuji, Y.; Cook, D.G.; et al. Lipopolysaccharide-induced blood-brain barrier disruption: Roles of cyclooxygenase, oxidative stress, neuroinflammation, and elements of the neurovascular unit. *J. Neuroinflammation* 2015, *12*, 223. [CrossRef] [PubMed]
- 90. Lei, Q.; Wu, T.; Wu, J.; Hu, X.; Guan, Y.; Wang, Y.; Yan, J.; Shi, G. Roles of α-synuclein in gastrointestinal microbiome dysbiosisrelated Parkinson's disease progression (Review). *Mol. Med. Rep.* **2021**, *24*, 734. [CrossRef] [PubMed]
- 91. Castaño, A.; Herrera, A.J.; Cano, J.; Machado, A. Lipopolysaccharide intranigral injection induces inflammatory reaction and damage in nigrostriatal dopaminergic system. *J. Neurochem.* **2002**, *70*, 1584–1592. [CrossRef] [PubMed]
- 92. Gorecki, A.M.; Preskey, L.; Bakeberg, M.C.; Kenna, J.E.; Gildenhuys, C.; MacDougall, G.; Dunlop, S.A.; Mastaglia, F.L.; Akkari, P.A.; Koengten, F.; et al. Altered gut microbiome in Parkinson's disease and the influence of lipopolysaccharide in a human α-synuclein over-expressing mouse model. *Front. Neurosci.* **2019**, *13*, 839. [CrossRef]
- Sun, M.-F.; Zhu, Y.-L.; Zhou, Z.-L.; Jia, X.-B.; Xu, Y.-D.; Yang, Q.; Cui, C.; Shen, Y.-Q. Neuroprotective effects of fecal microbiota transplantation on MPTP-induced Parkinson's disease mice: Gut microbiota, glial reaction and TLR4/TNF-α signaling pathway. *Brain Behav. Immun.* 2018, 70, 48–60. [CrossRef] [PubMed]
- 94. Tetz, G.; Brown, S.M.; Hao, Y.; Tetz, V. Parkinson's disease and bacteriophages as its overlooked contributors. *Sci. Rep.* **2018**, *8*, 10812. [CrossRef]
- 95. Strandwitz, P.; Kim, K.H.; Terekhova, D.; Liu, J.K.; Sharma, A.; Levering, J.; McDonald, D.; Dietrich, D.; Ramadhar, T.R.; Lekbua, A.; et al. GABA-modulating bacteria of the human gut microbiota. *Nat. Microbiol.* **2019**, *4*, 396–403. [CrossRef] [PubMed]
- 96. Darby, T.M.; Owens, J.; Saeedi, B.; Luo, L.; Matthews, J.D.; Robinson, B.S.; Naudin, C.; Jones, R.M. Lactococcus Lactis Subsp. cremoris Is an Efficacious Beneficial Bacterium that Limits Tissue Injury in the Intestine. *iScience* 2019, 12, 356–367. [CrossRef] [PubMed]

- Asano, Y.; Hiramoto, T.; Nishino, R.; Aiba, Y.; Kimura, T.; Yoshihara, K.; Koga, Y.; Sudo, N. Critical role of gut microbiota in the production of biologically active, free catecholamines in the gut lumen of mice. *Am. J. Physiol. Liver Physiol.* 2012, 303, G1288–G1295. [CrossRef]
- 98. Scheperjans, F. Gut microbiota, 1013 new pieces in the Parkinson's disease puzzle. *Curr. Opin. Neurol.* **2016**, *29*, 773–780. [CrossRef] [PubMed]
- Sampson, T.R.; Debelius, J.W.; Thron, T.; Janssen, S.; Shastri, G.G.; Ilhan, Z.E.; Challis, C.; Schretter, C.E.; Rocha, S.; Gradinaru, V.; et al. Gut Microbiota Regulate Motor Deficits and Neuroinflammation in a Model of Parkinson's Disease. *Cell* 2016, 167, 1469–1480. [CrossRef] [PubMed]
- Kusters, J.G.; van Vliet, A.H.M.; Kuipers, E.J. Pathogenesis of Helicobacter pylori Infection. *Clin. Microbiol. Rev.* 2006, 19, 449–490.
   [CrossRef]
- Dardiotis, E.; Tsouris, Z.; Mentis, A.-F.A.; Siokas, V.; Michalopoulou, A.; Sokratous, M.; Dastamani, M.; Bogdanos, D.; Deretzi, G.; Kountouras, J.H. pylori and Parkinson's disease: Meta-analyses including clinical severity. *Clin. Neurol. Neurosurg.* 2018, 175, 16–24. [CrossRef]
- Çamcı, G.; Oğuz, S. Association between Parkinson's Disease and Helicobacter Pylori. J. Clin. Neurol. 2016, 12, 147–150. [CrossRef]
   [PubMed]
- McGee, D.J.; Lu, X.-H.; Disbrow, E.A. Stomaching the Possibility of a Pathogenic Role for Helicobacter pylori in Parkinson's Disease. J. Park. Dis. 2018, 8, 367–374. [CrossRef] [PubMed]
- 104. Testerman, T.L. Beyond the stomach: An updated view of Helicobacter pylori pathogenesis, diagnosis, and treatment. *World J. Gastroenterol.* **2014**, *20*, 12781–12808. [CrossRef] [PubMed]
- Alvarez-Arellano, L.; Maldonado-Bernal, C. Helicobacter pylori and neurological diseases: Married by the laws of inflammation. World J. Gastrointest. Pathophysiol. 2014, 5, 400–404. [CrossRef]
- 106. Gundersen, V. Parkinson's Disease: Can Targeting Inflammation Be an Effective Neuroprotective Strategy? Front. Neurosci. 2020, 14, 580311. [CrossRef]
- 107. Marogianni, C.; Sokratous, M.; Dardiotis, E.; Hadjigeorgiou, G.M.; Bogdanos, D.; Xiromerisiou, G. Neurodegeneration and Inflammation—An Interesting Interplay in Parkinson's Disease. *Int. J. Mol. Sci.* **2020**, *21*, 8421. [CrossRef]
- 108. Suwarnalata, G.; Tan, A.H.; Isa, H.; Gudimella, R.; Anwar, A.; Loke, M.F.; Mahadeva, S.; Lim, S.-Y.; Vadivelu, J. Augmentation of Autoantibodies by Helicobacter pylori in Parkinson's Disease Patients May Be Linked to Greater Severity. *PLoS ONE* 2016, 11, e0153725. [CrossRef]
- Shibayama, K.; Doi, Y.; Shibata, N.; Yagi, T.; Nada, T.; Iinuma, Y.; Arakawa, Y. Apoptotic Signaling Pathway Activated by Helicobacter pylori Infection and Increase of Apoptosis-Inducing Activity under Serum-Starved Conditions. *Infect. Immun.* 2001, 69, 3181–3189. [CrossRef]
- Koh, A.; de Vadder, F.; Kovatcheva-Datchary, P.; Bäckhed, F. From Dietary Fiber to Host Physiology: Short-Chain Fatty Acids as Key Bacterial Metabolites. Cell 2016, 165, 1332–1345. [CrossRef]
- 111. Deehan, E.C.; Duar, R.M.; Armet, A.M.; Perez-Muñoz, M.E.; Jin, M.; Walter, J. Modulation of the Gastrointestinal Microbiome with Nondigestible Fermentable Carbohydrates to Improve Human Health. *Microbiol. Spectr.* **2017**, *5*. [CrossRef]
- 112. Kho, Z.Y.; Lal, S.K. The Human Gut Microbiome–A Potential Controller of Wellness and Disease. *Front. Microbiol.* **2018**, *9*, 1835. [CrossRef]
- 113. Parada Venegas, D.; de la Fuente, M.K.; Landskron, G.; González, M.J.; Quera, R.; Dijkstra, G.; Harmsen, H.J.M.; Faber, K.N.; Hermoso, M.A. Short Chain Fatty Acids (SCFAs)-Mediated Gut Epithelial and Immune Regulation and Its Relevance for Inflammatory Bowel Diseases. *Front. Immunol.* 2019, 10, 277. [CrossRef]
- 114. Baert, F.; Matthys, C.; Maselyne, J.; van Poucke, C.; van Coillie, E.; Bergmans, B.; Vlaemynck, G. Parkinson's disease patients' short chain fatty acids production capacity after in vitro fecal fiber fermentation. *NPJ Park. Dis.* **2021**, *7*, 72. [CrossRef] [PubMed]
- 115. Queipo-Ortuño, M.I.; Seoane, L.M.; Murri, M.; Pardo, M.; Gomez-Zumaquero, J.M.; Cardona, F.; Casanueva, F.; Tinahones, F.J. Gut Microbiota Composition in Male Rat Models under Different Nutritional Status and Physical Activity and Its Association with Serum Leptin and Ghrelin Levels. *PLoS ONE* 2013, *8*, e65465. [CrossRef]
- 116. Bayliss, J.A.; Andrews, Z.B. Ghrelin is neuroprotective in Parkinson's disease: Molecular mechanisms of metabolic neuroprotection. *Ther. Adv. Endocrinol. Metab.* 2013, 4, 25–36. [CrossRef] [PubMed]
- 117. Unger, M.M.; Möller, J.C.; Mankel, K.; Eggert, K.M.; Bohne, K.; Bodden, M.; Stiasny-Kolster, K.; Kann, P.H.; Mayer, G.; Tebbe, J.J.; et al. Postprandial ghrelin response is reduced in patients with Parkinson's disease and idiopathic REM sleep behaviour disorder: A peripheral biomarker for early Parkinson's disease? *J. Neurol.* 2011, 258, 982–990. [CrossRef] [PubMed]
- 118. Wurtman, R.J.; Wurtman, J.J.; Regan, M.M.; McDermott, J.M.; Tsay, R.H.; Breu, J.J. Effects of normal meals rich in carbohydrates or proteins on plasma tryptophan and tyrosine ratios. *Am. J. Clin. Nutr.* **2003**, *77*, 128–132. [CrossRef] [PubMed]
- Berry, E.M.; Growdon, J.H.; Wurtman, J.J.; Caballero, B. A balanced carbohydrate: Protein diet in the management of Parkinson's disease. *Neurology* 1991, 41, 1295. [CrossRef] [PubMed]
- Vinagre-Aragón, A.; Zis, P.; Grunewald, R.A.; Hadjivassiliou, M. Movement Disorders Related to Gluten Sensitivity: A Systematic Review. Nutrients 2018, 10, 1034. [CrossRef]
- Mischley, L.K.; Lau, R.C.; Bennett, R.D. Role of Diet and Nutritional Supplements in Parkinson's Disease Progression. Oxidative Med. Cell. Longev. 2017, 2017, 6405278. [CrossRef]

- Makki, K.; Deehan, E.C.; Walter, J.; Bäckhed, F. The Impact of Dietary Fiber on Gut Microbiota in Host Health and Disease. *Cell Host Microbe* 2018, 23, 705–715. [CrossRef] [PubMed]
- 123. Jackson, A.; Forsyth, C.B.; Shaikh, M.; Voigt, R.M.; Engen, P.A.; Ramirez, V.; Keshavarzian, A. Diet in Parkinson's Disease: Critical Role for the Microbiome. *Front. Neurol.* **2019**, *10*, 1245. [CrossRef]
- 124. Barichella, M.; Cereda, E.; Cassani, E.; Pinelli, G.; Iorio, L.; Ferri, V.; Privitera, G.; Pasqua, M.; Valentino, A.; Monajemi, F.; et al. Dietary habits and neurological features of Parkinson's disease patients: Implications for practice. *Clin. Nutr.* 2017, 36, 1054–1061. [CrossRef]
- 125. Liddle, R.A. Parkinson's disease from the gut. Brain Res. 2018, 1693, 201–206. [CrossRef]
- 126. Perez-Pardo, P.; Kliest, T.; Dodiya, H.B.; Broersen, L.M.; Garssen, J.; Keshavarzian, A.; Kraneveld, A.D. The gut-brain axis in Parkinson's disease: Possibilities for food-based therapies. *Eur. J. Pharmacol.* **2017**, *817*, 86–95. [CrossRef]
- 127. Maraki, M.I.; Yannakoulia, M.; Stamelou, M.; Stefanis, L.; Xiromerisiou, G.; Kosmidis, M.H.; Dardiotis, E.; Hadjigeorgiou, G.M.; Sakka, P.; Anastasiou, C.A.; et al. Mediterranean diet adherence is related to reduced probability of prodromal Parkinson's disease. *Mov. Disord.* **2019**, *34*, 48–57. [CrossRef]
- 128. Castelli, V.; Grassi, D.; Bocale, R.; D'Angelo, M.; Antonosante, A.; Cimini, A.; Ferri, C.; Desideri, G. Diet and Brain Health: Which Role for Polyphenols? *Curr. Pharm. Des.* **2018**, *24*, 227–238. [CrossRef] [PubMed]
- 129. Bousquet, M.; Calon, F.; Cicchetti, F. Impact of omega-3 fatty acids in Parkinson's disease. *Ageing Res. Rev.* **2011**, *10*, 453–463. [CrossRef]
- Kamel, F.; Goldman, S.; Umbach, D.M.; Chen, H.; Richardson, G.; Barber, M.R.; Meng, C.; Marras, C.; Korell, M.; Kasten, M.; et al. Dietary fat intake, pesticide use, and Parkinson's disease. *Park. Relat. Disord.* 2014, 20, 82–87. [CrossRef]
- 131. Fava, F.; Gitau, R.; Griffin, B.A.; Gibson, G.R.; Tuohy, K.; Lovegrove, J.A. The type and quantity of dietary fat and carbohydrate alter faecal microbiome and short-chain fatty acid excretion in a metabolic syndrome 'at-risk' population. *Int. J. Obes.* **2013**, *37*, 216–223. [CrossRef]
- 132. Ascherio, A.; Schwarzschild, M.A. The epidemiology of Parkinson's disease: Risk factors and prevention. *Lancet Neurol.* **2016**, *15*, 1257–1272. [CrossRef]
- Davey, K.J.; Cotter, P.; O'Sullivan, O.; Crispie, F.; Dinan, T.; Cryan, J.F.; Mahony, S.O. Antipsychotics and the gut microbiome: Olanzapine-induced metabolic dysfunction is attenuated by antibiotic administration in the rat. *Transl. Psychiatry* 2013, *3*, e309. [CrossRef]
- 134. Parashar, A.; Udayabanu, M. Gut microbiota: Implications in Parkinson's disease. Park. Relat. Disord. 2017, 38, 1–7. [CrossRef]
- 135. Derkinderen, P.; Shannon, K.M.; Brundin, P. Gut feelings about smoking and coffee in Parkinson's disease. *Mov. Disord.* 2014, 29, 976–979. [CrossRef] [PubMed]
- 136. Mills, C.E.; Tzounis, X.; Oruna-Concha, M.-J.; Mottram, D.S.; Gibson, G.R.; Spencer, J.P.E. In vitro colonic metabolism of coffee and chlorogenic acid results in selective changes in human faecal microbiota growth. *Br. J. Nutr.* 2015, 113, 1220–1227. [CrossRef] [PubMed]
- 137. Yang, D.; Zhao, D.; Shah, S.Z.A.; Wu, W.; Lai, M.; Zhang, X.; Li, J.; Guan, Z.; Zhao, H.; Li, W.; et al. The Role of the Gut Microbiota in the Pathogenesis of Parkinson's Disease. *Front. Neurol.* **2019**, *10*, 1155. [CrossRef]
- Perez-Pardo, P.; de Jong, E.M.; Broersen, L.M.; van Wijk, N.; Attali, A.; Garssen, J.; Kraneveld, A.D. Promising Effects of Neurorestorative Diets on Motor, Cognitive, and Gastrointestinal Dysfunction after Symptom Development in a Mouse Model of Parkinson's Disease. *Front. Aging Neurosci.* 2017, *9*, 57. [CrossRef] [PubMed]
- 139. Wurtman, R.J. Enhancing Synaptogenesis in Diseases Characterized by Deficiencies in Brain Synapses. *Front. Psychiatry* **2010**, *1*, 147. [CrossRef]
- 140. Wang, L.; Pooler, A.M.; Albrecht, M.A.; Wurtman, R.J. Dietary Uridine-5'-Monophosphate Supplementation Increases Potassium-Evoked Dopamine Release and Promotes Neurite Outgrowth in Aged Rats. J. Mol. Neurosci. 2005, 27, 137–146. [CrossRef]
- 141. Cansev, M.; Ulus, I.H.; Wang, L.; Maher, T.J.; Wurtman, R.J. Restorative effects of uridine plus docosahexaenoic acid in a rat model of Parkinson's disease. *Neurosci. Res.* **2008**, *62*, 206–209. [CrossRef]
- 142. Tanaka, K.; Farooqui, A.A.; Siddiqi, N.J.; Alhomida, A.S.; Ong, W.-Y. Effects of Docosahexaenoic Acid on Neurotransmission. *Biomol. Ther.* 2012, 20, 152–157. [CrossRef]
- 143. Yan, F.; Polk, D. Probiotics and immune health. Curr. Opin. Gastroenterol. 2011, 27, 496–501. [CrossRef] [PubMed]
- 144. Reid, G.; Younes, J.A.; van der Mei, H.C.; Gloor, G.B.; Knight, R.; Busscher, H.J. Microbiota restoration: Natural and supplemented recovery of human microbial communities. *Nat. Rev. Genet.* **2011**, *9*, 27–38. [CrossRef] [PubMed]
- 145. Varankovich, N.; Nickerson, M.T.; Korber, D.R. Probiotic-based strategies for therapeutic and prophylactic use against multiple gastrointestinal diseases. *Front. Microbiol.* **2015**, *6*, 685. [CrossRef]
- 146. Sanders, M.E.; Merenstein, D.J.; Reid, G.; Gibson, G.R.; Rastall, R.A. Probiotics and prebiotics in intestinal health and disease: From biology to the clinic. *Nat. Rev. Gastroenterol. Hepatol.* **2019**, *16*, 605–616. [CrossRef]
- 147. Salas-Jara, M.J.; Ilabaca, A.; Vega, M.; García, A. Biofilm Forming Lactobacillus: New Challenges for the Development of Probiotics. *Microorganisms* 2016, 4, 35. [CrossRef]
- 148. Bron, P.A.; van Baarlen, P.; Kleerebezem, M. Emerging molecular insights into the interaction between probiotics and the host intestinal mucosa. *Nat. Rev. Genet.* **2011**, *10*, 66–78. [CrossRef] [PubMed]
- Yu, M.; Zhang, R.; Ni, P.; Chen, S.; Duan, G. Efficacy of Lactobacillus-supplemented triple therapy for H. pylori eradication: A meta-analysis of randomized controlled trials. *PLoS ONE* 2019, 14, e0223309. [CrossRef]

- Onrust, L.; Ducatelle, R.; van Driessche, K.; De Maesschalck, C.; Vermeulen, K.; Haesebrouck, F.; Eeckhaut, V.; van Immerseel, F. Steering Endogenous Butyrate Production in the Intestinal Tract of Broilers as a Tool to Improve Gut Health. *Front. Vet. Sci.* 2015, 2, 75. [CrossRef]
- 151. Maldonado Galdeano, C.; Cazorla, S.I.; Lemme Dumit, J.M.; Vélez, E.; Perdigón, G. Beneficial Effects of Probiotic Consumption on the Immune System. *Ann. Nutr. Metab.* **2019**, *74*, 115–124. [CrossRef] [PubMed]
- 152. Cristofori, F.; Dargenio, V.N.; Dargenio, C.; Miniello, V.L.; Barone, M.; Francavilla, R. Anti-Inflammatory and Immunomodulatory Effects of Probiotics in Gut Inflammation: A Door to the Body. *Front. Immunol.* **2021**, *12*, 578386. [CrossRef]
- Zhao, X.; Zhou, J.; Liang, W.; Sheng, Q.; Lu, L.; Chen, T.; Chen, J.; Tan, K.; Lv, Z. Probiotics mixture reinforces barrier function to ameliorate necrotizing enterocolitis by regulating PXR-JNK pathway. *Cell Biosci.* 2021, 11, 20. [CrossRef]
- 154. Liu, Q.; Yu, Z.; Tian, F.; Zhao, J.; Zhang, H.; Zhai, Q.; Chen, W. Surface components and metabolites of probiotics for regulation of intestinal epithelial barrier. *Microb. Cell Factories* **2020**, *19*, 23. [CrossRef] [PubMed]
- 155. Ait-Belgnaoui, A.; Durand, H.; Cartier, C.; Chaumaz, G.; Eutamene, H.; Ferrier, L.; Houdeau, E.; Fioramonti, J.; Bueno, L.; Theodorou, V. Prevention of gut leakiness by a probiotic treatment leads to attenuated HPA response to an acute psychological stress in rats. *Psychoneuroendocrinology* **2012**, *37*, 1885–1895. [CrossRef] [PubMed]
- 156. Corridoni, D.; Pastorelli, L.; Mattioli, B.; Locovei, S.; Ishikawa, D.; Arseneau, K.O.; Chieppa, M.; Cominelli, F.; Pizarro, T.T. Probiotic Bacteria Regulate Intestinal Epithelial Permeability in Experimental Ileitis by a TNF-Dependent Mechanism. *PLoS ONE* 2012, 7, e42067. [CrossRef]
- Zaharoni, H.; Rimon, E.; Vardi, H.; Friger, M.; Bolotin, A.; Shahar, D. Probiotics improve bowel movements in hospitalized elderly patients—The proage study. J. Nutr. Health Aging 2011, 15, 215–220. [CrossRef] [PubMed]
- 158. Plaza-Diaz, J.; Ruiz-Ojeda, F.J.; Gil-Campos, M.; Gil, A. Mechanisms of Action of Probiotics. *Adv. Nutr. Int. Rev. J.* 2019, 10, S49–S66. [CrossRef] [PubMed]
- 159. Hsiao, E.Y.; McBride, S.W.; Hsien, S.; Sharon, G.; Hyde, E.R.; McCue, T.; Codelli, J.A.; Chow, J.; Reisman, S.E.; Petrosino, J.F.; et al. Microbiota Modulate Behavioral and Physiological Abnormalities Associated with Neurodevelopmental Disorders. *Cell* 2013, 155, 1451–1463. [CrossRef]
- 160. Salami, M. Interplay of Good Bacteria and Central Nervous System: Cognitive Aspects and Mechanistic Considerations. *Front. Neurosci.* **2021**, *15*, 613120. [CrossRef]
- Kim, N.; Yun, M.; Oh, Y.J.; Choi, H.-J. Mind-altering with the gut: Modulation of the gut-brain axis with probiotics. J. Microbiol. 2018, 56, 172–182. [CrossRef] [PubMed]
- 162. Rao, A.V.; Bested, A.C.; Beaulne, T.M.; A Katzman, M.; Iorio, C.; Berardi, J.M.; Logan, A.C. A randomized, double-blind, placebocontrolled pilot study of a probiotic in emotional symptoms of chronic fatigue syndrome. *Gut Pathog.* 2009, 1, 6. [CrossRef] [PubMed]
- 163. Cassani, E.; Privitera, G.; Pezzoli, G.; Pusani, C.; Madio, C.; Iorio, L.; Barichella, M. Use of probiotics for the treatment of constipation in Parkinson's disease patients. *Minerva Gastroenterol. Dietol.* **2011**, *57*, 117–121. [PubMed]
- 164. Liang, S.; Wang, T.; Hu, X.; Luo, J.; Li, W.; Wu, X.; Duan, Y.; Jin, F. Administration of Lactobacillus helveticus NS8 improves
- behavioral, cognitive, and biochemical aberrations caused by chronic restraint stress. *Neuroscience* 2015, 310, 561–577. [CrossRef]
  165. van Kessel, S.P.; El Aidy, S. Contributions of Gut Bacteria and Diet to Drug Pharmacokinetics in the Treatment of Parkinson's Disease. *Front. Neurol.* 2019, 10, 1087. [CrossRef]
- 166. D'Angelo, M.; Cimini, A.; Castelli, V.; Quintiliani, M.; Benedetti, E.; Cifone, M.G. The emerging role of probiotics in neurodegenerative diseases: New hope for Parkinson's disease? *Neural Regen. Res.* **2021**, *16*, 628–634. [CrossRef]
- 167. Hsieh, T.-H.; Kuo, C.-W.; Hsieh, K.-H.; Shieh, M.-J.; Peng, C.-W.; Chen, Y.-C.; Chang, Y.-L.; Huang, Y.-Z.; Chen, C.-C.; Chang, P.-K.; et al. Probiotics Alleviate the Progressive Deterioration of Motor Functions in a Mouse Model of Parkinson's Disease. *Brain Sci.* 2020, 10, 206. [CrossRef]
- 168. Srivastav, S.; Neupane, S.; Bhurtel, S.; Katila, N.; Maharjan, S.; Choi, H.; Hong, J.T.; Choi, D.-Y. Probiotics mixture increases butyrate, and subsequently rescues the nigral dopaminergic neurons from MPTP and rotenone-induced neurotoxicity. *J. Nutr. Biochem.* **2019**, *69*, 73–86. [CrossRef]
- 169. Castelli, V.; d'Angelo, M.; Lombardi, F.; Alfonsetti, M.; Antonosante, A.; Catanesi, M.; Benedetti, E.; Palumbo, P.; Cifone, M.G.; Giordano, A.; et al. Effects of the probiotic formulation SLAB51 in in vitro and in vivo Parkinson's disease models. *Aging* 2020, 12, 4641–4659. [CrossRef]
- Fang, X.; Tian, P.; Zhao, X.; Jiang, C.; Chen, T. Neuroprotective effects of an engineered commensal bacterium in the 1-methyl-4phenyl-1, 2, 3, 6-tetrahydropyridine Parkinson disease mouse model via producing glucagon-like peptide-1. *J. Neurochem.* 2019, 150, 441–452. [CrossRef]
- 171. Mulvaney, C.A.; Duarte, G.S.; Handley, J.; Evans, D.J.; Menon, S.; Wyse, R.; Emsley, H.C. GLP-1 receptor agonists for Parkinson's disease. *Cochrane Database Syst. Rev.* 2020, 7, CD012990. [CrossRef]
- 172. Goya, M.E.; Xue, F.; Quevedo, C.S.T.; Arnaouteli, S.; Riquelme-Dominguez, L.; Romanowski, A.; Brydon, J.; Ball, K.L.; Stanley-Wall, N.R.; Doitsidou, M. Probiotic Bacillus subtilis Protects against α-Synuclein Aggregation in C. elegans. *Cell Rep.* 2020, 30, 367–380. [CrossRef] [PubMed]
- 173. Huebecker, M.; Moloney, E.; van der Spoel, A.C.; Priestman, D.A.; Isacson, O.; Hallett, P.J.; Platt, F.M. Reduced sphingolipid hydrolase activities, substrate accumulation and ganglioside decline in Parkinson's disease. *Mol. Neurodegener.* 2019, *14*, 40. [CrossRef] [PubMed]

- 174. Magistrelli, L.; Amoruso, A.; Mogna, L.; Graziano, T.; Cantello, R.; Pane, M.; Comi, C. Probiotics May Have Beneficial Effects in Parkinson's Disease: In vitro Evidence. *Front. Immunol.* **2019**, *10*, 969. [CrossRef]
- 175. Gibson, G.R.; Roberfroid, M.B. Dietary Modulation of the Human Colonic Microbiota: Introducing the Concept of Prebiotics. *J. Nutr.* **1995**, *125*, 1401–1412. [CrossRef]
- 176. Davani-Davari, D.; Negahdaripour, M.; Karimzadeh, I.; Seifan, M.; Mohkam, M.; Masoumi, S.J.; Berenjian, A.; Ghasemi, Y. Prebiotics: Definition, Types, Sources, Mechanisms, and Clinical Applications. *Foods* **2019**, *8*, 92. [CrossRef]
- 177. Holscher, H.D. Dietary fiber and prebiotics and the gastrointestinal microbiota. *Gut Microbes* **2017**, *8*, 172–184. [CrossRef] [PubMed]
- 178. Belobrajdic, D.P.; Hino, S.; Kondo, T.; Jobling, S.; Morell, M.K.; Topping, D.L.; Morita, T.; Bird, A. High wholegrain barley β-glucan lowers food intake but does not alter small intestinal macronutrient digestibility in ileorectostomised rats. *Int. J. Food Sci. Nutr.* 2016, 67, 678–685. [CrossRef]
- Gill, S.K.; Rossi, M.; Bajka, B.; Whelan, K. Dietary fibre in gastrointestinal health and disease. *Nat. Rev. Gastroenterol. Hepatol.* 2021, 18, 101–116. [CrossRef]
- Flint, H.J.; Scott, K.P.; Duncan, S.; Louis, P.; Forano, E. Microbial degradation of complex carbohydrates in the gut. *Gut Microbes* 2012, *3*, 289–306. [CrossRef]
- 181. Liu, F.; Li, P.; Chen, M.; Luo, Y.; Prabhakar, M.; Zheng, H.; He, Y.; Qi, Q.; Long, H.; Zhang, Y.; et al. Fructooligosaccharide (FOS) and Galactooligosaccharide (GOS) Increase Bifidobacterium but Reduce Butyrate Producing Bacteria with Adverse Glycemic Metabolism in healthy young population. *Sci. Rep.* 2017, *7*, 11789. [CrossRef]
- 182. Kovács, Z.; Benjamins, E.; Grau, K.; Ur Rehman, A.; Ebrahimi, M.; Czermak, P. Recent developments in manufacturing oligosaccharides with prebiotic functions. In *Biotechnology of Food and Feed Additives*; Zorn, H., Czermak, P., Eds.; Springer: Berlin/Heidelberg, Germany, 2013; Volume 143, pp. 257–295. ISBN 978-3-662-43760-5.
- Jefferson, A.; Adolphus, K. The Effects of Intact Cereal Grain Fibers, Including Wheat Bran on the Gut Microbiota Composition of Healthy Adults: A Systematic Review. Front. Nutr. 2019, 6, 33. [CrossRef]
- 184. Nie, C.; He, T.; Zhang, W.; Zhang, G.; Ma, X. Branched Chain Amino Acids: Beyond Nutrition Metabolism. *Int. J. Mol. Sci.* 2018, 19, 954. [CrossRef]
- Valerio, A.; D'Antona, G.; Nisoli, E. Branched-chain amino acids, mitochondrial biogenesis, and healthspan: An evolutionary perspective. *Aging* 2011, *3*, 464–478. [CrossRef] [PubMed]
- 186. Yao, V.; Kaletsky, R.; Keyes, W.; Mor, D.E.; Wong, A.K.; Sohrabi, S.; Murphy, C.T.; Troyanskaya, O.G. An integrative tissue-network approach to identify and test human disease genes. *Nat. Biotechnol.* **2018**, *36*, 1091–1099. [CrossRef]
- 187. Mailloux, R.; McBride, S.L.; Harper, M.-E. Unearthing the secrets of mitochondrial ROS and glutathione in bioenergetics. *Trends Biochem. Sci.* **2013**, *38*, 592–602. [CrossRef]
- 188. Smeyne, M.; Smeyne, R.J. Glutathione metabolism and Parkinson's disease. Free Radic. Biol. Med. 2013, 62, 13–25. [CrossRef]
- Savignac, H.M.; Corona, G.; Mills, H.; Chen, L.; Spencer, J.P.; Tzortzis, G.; Burnet, P.W. Prebiotic feeding elevates central brain derived neurotrophic factor, N-methyl-d-aspartate receptor subunits and d-serine. *Neurochem. Int.* 2013, 63, 756–764. [CrossRef] [PubMed]
- Keshavarzian, A.; Green, S.J.; Engen, P.A.; Voigt, R.M.; Naqib, A.; Forsyth, C.B.; Mutlu, E.; Shannon, K.M. Colonic bacterial composition in Parkinson's disease. *Mov. Disord.* 2015, 30, 1351–1360. [CrossRef]
- 191. Unger, M.M.; Spiegel, J.; Dillmann, K.-U.; Grundmann, D.; Philippeit, H.; Bürmann, J.; Faßbender, K.; Schwiertz, A.; Schäfer, K.H. Short chain fatty acids and gut microbiota differ between patients with Parkinson's disease and age-matched controls. *Park. Relat. Disord.* 2016, 32, 66–72. [CrossRef] [PubMed]
- de Vrese, M.; Schrezenmeir, J. Probiotics, Prebiotics, and Synbiotics. *Adv. Biochem. Eng. Biotechnol.* 2008, 111, 1–66. [CrossRef]
   [PubMed]
- Cencic, A.; Chingwaru, W. The Role of Functional Foods, Nutraceuticals, and Food Supplements in Intestinal Health. *Nutrients* 2010, 2, 611–625. [CrossRef] [PubMed]
- Romeo, J.; Nova, E.; Warnberg, J.; Gómez-Martínez, S.; Ligia, L.E.D.; Marcos, A. Immunomodulatory effect of fibres, probiotics and synbiotics in different life-stages. *Nutr. Hosp.* 2010, 25, 341–349.
- 195. Zhang, M.-M.; Cheng, J.-Q.; Lu, Y.-R.; Yi, Z.-H.; Yang, P.; Wu, X.-T. Use of pre-, pro- and synbiotics in patients with acute pancreatitis: A meta-analysis. *World J. Gastroenterol.* **2010**, *16*, 3970–3978. [CrossRef]
- 196. Rajkumar, H.; Kumar, M.; Das, N.; Kumar, S.N.; Challa, H.R.; Nagpal, R. Effect of Probiotic Lactobacillus salivarius UBL S22 and Prebiotic Fructo-oligosaccharide on Serum Lipids, Inflammatory Markers, Insulin Sensitivity, and Gut Bacteria in Healthy Young Volunteers: A Randomized Controlled Single-Blind Pilot Study. J. Cardiovasc. Pharmacol. Ther. 2014, 20, 289–298. [CrossRef]
- 197. de Paula, J.A.; Carmuega, E.; Weill, R. Effect of the ingestion of a symbiotic yogurt on the bowel habits of women with functional constipation. *Acta Gastroenterol. Latinoam.* **2008**, *38*, 16–25.
- 198. Tan, A.H.; Mahadeva, S.; Thalha, A.M.; Gibson, P.R.; Kiew, C.K.; Yeat, C.M.; Ng, S.W.; Ang, S.P.; Chow, S.K.; Tan, C.T.; et al. Small intestinal bacterial overgrowth in Parkinson's disease. *Park. Relat. Disord.* **2014**, *20*, 535–540. [CrossRef]
- 199. Khalighi, A.; Khalighi, M.; Behdani, R.; Jamali, J.; Khosravi, A.; Kouhestani, S.; Radmanesh, H.; Esmaeelzadeh, S.; Khalighi, N. Evaluating the efficacy of probiotic on treatment in patients with small intestinal bacterial overgrowth (SIBO)—A pilot study. *Indian J. Med Res.* 2014, 140, 604–608. [PubMed]

- 200. Castelli, V.; Benedetti, E.; Antonosante, A.; Catanesi, M.; Pitari, G.; Ippoliti, R.; Cimini, A.; D'Angelo, M. Neuronal Cells Rearrangement During Aging and Neurodegenerative Disease: Metabolism, Oxidative Stress and Organelles Dynamic. *Front. Mol. Neurosci.* 2019, 12, 132. [CrossRef]
- Lin, T.-K.; Chang, Y.-Y.; Chen, N.-C.; Liou, C.-W.; Lan, M.-Y.; Chen, Y.-F.; Tsai, C.-L. Nutritional Status Associated with Molecular Biomarkers, Physiological Indices, and Clinical Severity in Parkinson's Disease Patients. *Int. J. Environ. Res. Public Health* 2020, 17, 5727. [CrossRef] [PubMed]
- 202. Ma, K.; Xiong, N.; Shen, Y.; Han, C.; Liu, L.; Zhang, G.; Wang, L.; Guo, S.; Guo, X.; Xia, Y.; et al. Weight Loss and Malnutrition in Patients with Parkinson's Disease: Current Knowledge and Future Prospects. *Front. Aging Neurosci.* 2018, 10, 1. [CrossRef] [PubMed]
- Molsberry, S.; Bjornevik, K.; Hughes, K.C.; Healy, B.; Schwarzschild, M.; Ascherio, A. Diet pattern and prodromal features of Parkinson disease. *Neurology* 2020, 95, e2095–e2108. [CrossRef] [PubMed]