

Arrival and proliferation of the invasive seaweed *Rugulopteryx okamurae* in NE Atlantic islands

João Faria¹, Afonso CL Prestes¹, Ignacio Moreu¹, Gustavo M Martins², Ana I Neto^{1†},
Eva Cacabelos³

¹*cE3c - Centre for Ecology, Evolution and Environmental Changes/ Azorean Biodiversity Group, and University of Azores, Faculty of Sciences and Technology, Department of Biology, 9501-801 Ponta Delgada, São Miguel, Azores, Portugal*

²*CIBIO - Research Centre in Biodiversity and Genetic Resources, InBIO Associate Laboratory, Pólo dos Açores – Departamento de Biologia da Universidade dos Açores, 9501-801 Ponta Delgada, Portugal*

³*MARE - Marine and Environmental Sciences Centre, Agência Regional para o Desenvolvimento da Investigação Tecnologia e Inovação (ARDITI), Edifício Madeira Tecnopolo, Piso 0, Caminho da Penteada, 9020-105 Funchal, Madeira, Portugal*

Abstract

The present study reports the recent occurrence and expansion of *Rugulopteryx okamurae* in the Azores archipelago (NE Atlantic). Morphological and molecular characters confirmed the species identification. Quick surveys around the island of São Miguel showed that it has successfully colonized the island and is quickly expanding. In some locations, *R. okamurae* is currently the dominant organism smothering all other benthic biota and posing a serious threat to the benthic ecosystems across the region. The species first record dates from 2019 near the main harbour of the island, suggesting that its introduction was driven by human-assisted transport, via boat ballast waters or adhered to ship hulls and likely originating from the Mediterranean populations that have been proliferating in recent years across the Strait of Gibraltar.

Keywords: macroalgae, oceanic islands, marine bioinvasions, non-indigenous species, Azores

The brown seaweed *Rugulopteryx okamurae* (E.Y. Dawson) I.K. Hwang, W.J. Lee & H.S. Kim 2009, which has been recently introduced in the Mediterranean (Verlaque et al. 2009) is a native and widely distributed species in the subtropical to temperate western Pacific Ocean (Lee and Kang 1986; Silva et al. 1987; Yoshida 1998). It grows in the shallow subtidal and can be present year-round as dormant rhizoidal bases. At macroscopic level, individuals of *R. okamurae* are characterized by a membranous and erect dichotomously branched thallus with smooth margins of 10-20 cm high, attached by rhizoids restricted to the basal parts of the thallus. The fronds grow into a compressed flabellum and in dense groups, and intraspecific variation is known to occur (Hwang et al. 2009). Microscopically, the thallus consists of a medulla, one cell thick centrally and two to three cells thick near the margins (Womersley 1987; De Clerck et al. 2006; Hwang et al. 2009). The reproductive structures are evenly distributed over the entire thallus surface, rather than being confined to the concavities as in the other *Rugulopteryx* species. Just like *Rugulopteryx* species, sporangia of *R. okamurae* are borne on two stalk cells. It can readily form propagules, proliferous branchlets arising on the thallus surface which grow into new plants of the same ploidy level. Although it may be confused with species of the genus *Dictyota*, such as *Dictyota pinnatifida*, *D. dichotoma*, *D. spiralis*, *D. cyanoloma* or *D. fasciola*, *R. okamurae* is a single taxonomic entity and has been described in Hwang et al. (2009).

In European waters, the non-indigenous *R. okamurae* was first detected in spring 2002 close to a small harbour in Thau Lagoon (France), a known hot spot of marine species introductions in Europe (Verlaque et al. 2009). It was more recently detected in Ceuta and Andalusian waters, southern Iberian Peninsula, in 2015 and 2016, respectively (Altamirano et al. 2016, 2017; Ocaña et al. 2016). In the Strait of Gibraltar, *R. okamurae* has expanded massively in recent years across subtidal marine hard-bottoms, causing evident ecological impacts on coastal communities (García-Gómez et al. 2018, 2020). For instance, after its first record in the area, *R. okamurae* became the most abundant species in only one year (>90% coverage at 10-20 m depth), followed by a significant change in the structure of benthic communities (García-Gómez et al. 2020; Sempere-Valverde et al. 2021). Additionally, hundreds of tons of *R. okamurae* biomass have been reported to accumulate at coastal bedrocks and/or beaches across the region with serious implications for tourism and public health (Ocaña et al. 2016, García-Gómez et al. 2018, 2020).

Fishermen have also reported continuous hooks on fishing nets and a significant reduction in fish captures (Sempere-Valverde et al. 2019).

The present study reports the recent occurrence of *Rugulopteryx okamurae* in the Azores archipelago (NE Atlantic). The species was first observed in early 2019 on the south coast of São Miguel Island and identified as a hitherto unknown species of Dictyotales. A search for the species was conducted throughout the island in 2020 (June-July) and again in 2021 (May). Sampling was performed at several locations by snorkelling and/or scuba diving at about 5m depth. At each location, the DAFOR scale [D-dominant (>75%); A-abundant (50-75%); F-frequent (25-50%); O - occasional (10-25%); R-rare (<10%)] was applied to measure the relative abundance of the algae species along 10 min transect surveys (Fig. 1). In both field campaigns, live plants were collected, taken to the laboratory for examination, and housed as pressed and liquid specimens at the AZB Herbarium Ruy Telles Palhinha of the University of the Azores.

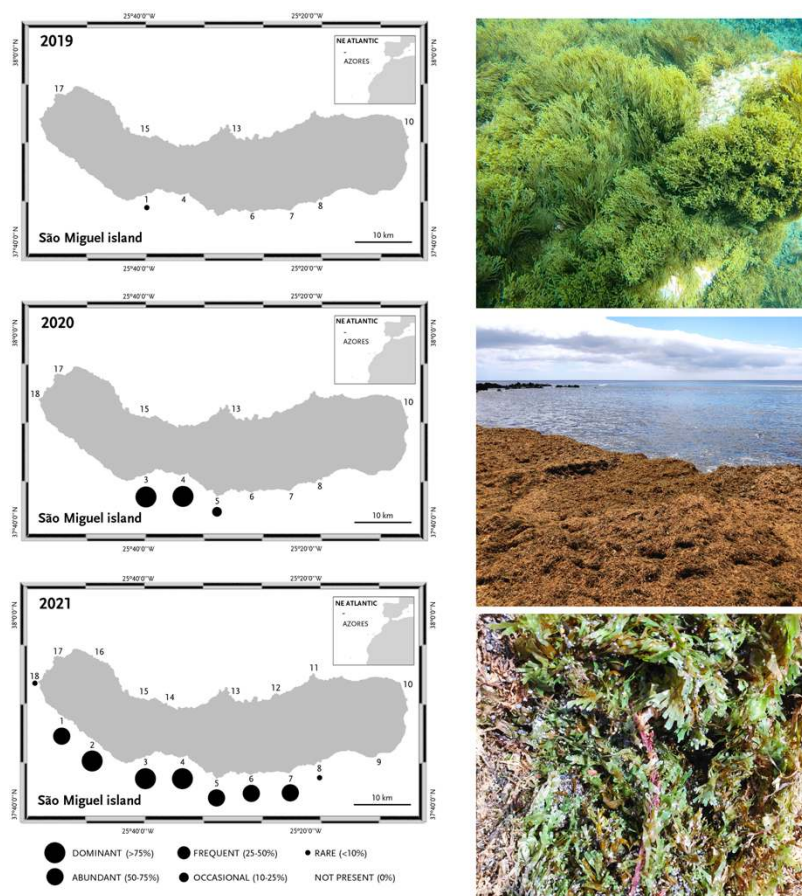


Figure 1. Map of São Miguel Island and locations (numbers) sampled for the presence of *Rugulopteryx okamurae* from 2019 to 2021. Black circles indicate the occurrence and relative abundance of the species (see Supplementary material Table S3). Images to the right show *R. okamurae* in its natural environment and as wrack cast away onto the beach.

Collected specimens were identified by their morphological and anatomical characteristics using both binocular and light microscopes (see Hwang et al. 2009; Verlaque et al. 2009). Macroscopically, many individuals displayed numerous proliferous branchlets on both surfaces. Transverse sections revealed the occurrence of mature sporangia subtended by two stalk cells in most individuals, a key distinctive morphologic feature of the genus *Rugulopterix* (Fig. 2). Gametophytes were not encountered in the field.

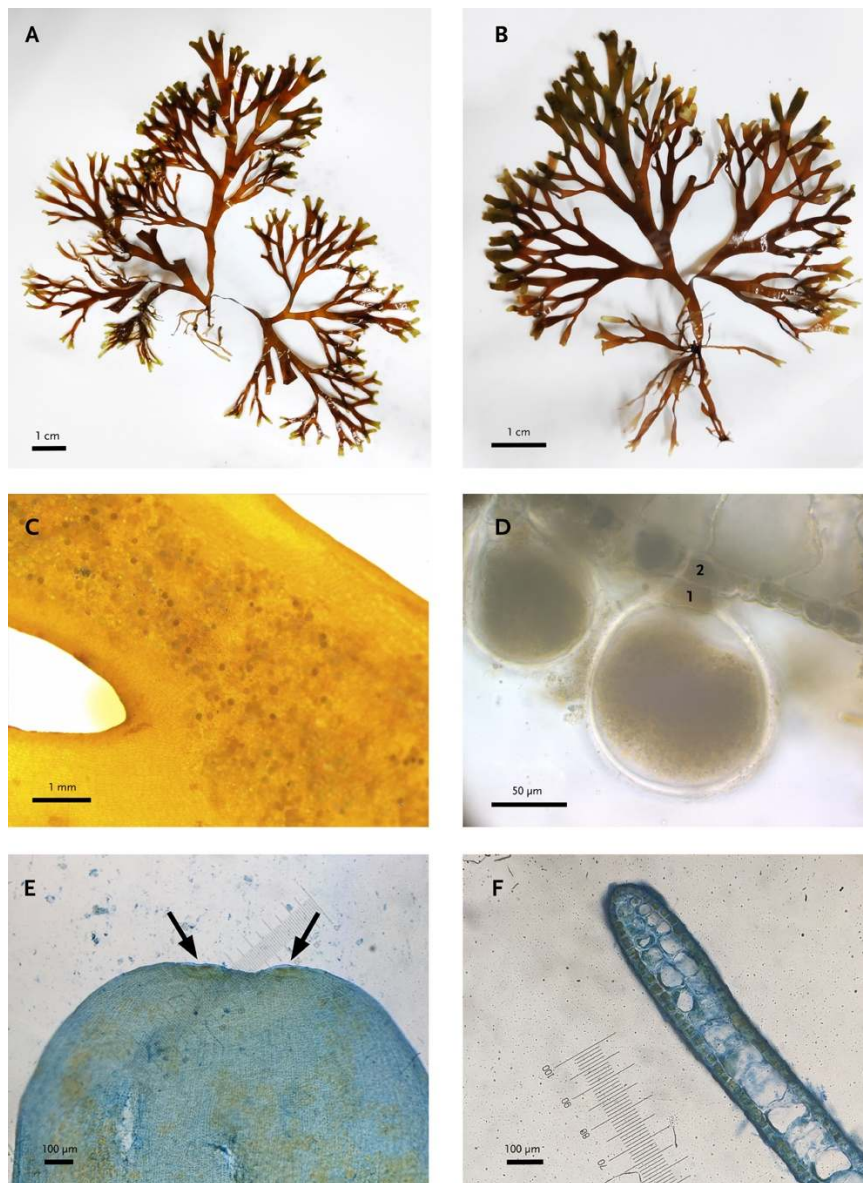


Figure 2. *Rugulopterix okamurae* from the Azores: (A and B) habit of the thallus showing the anchoring rhizoids and dichotomous branching; thallus surface (C) with sporangia (D) that are borne on 2 stalk cells (1 and 2); (E) detail of an apical portion of thallus with two apical cells; a transverse section of thallus showing (F) the margin with two layers of medullary cells. Black bars indicate scale.

Moreover, DNA sequences of two chloroplast protein-encoded genes *rbcL* (ribulose biphosphate carboxylase large chain) and *psbA* (photosystem II reaction center protein D1) from six collected samples were confirmed as *R. okamurae* using the GenBank BLASTn search. Resulting sequences matched together with *R. okamurae* individuals from western Pacific Ocean (*rbcL*: GenBank accession n° MZ393479 to MZ393484; *psbA*: GenBank accession n° MZ393485 to MZ393490). DNA sequences for *rbcL* and *psbA* were amplified using primers listed in Draisna et al. (2001) and Saunders and Moore (2013), respectively (see Supplementary material for methodological details; Table S1; Table S2).

The arrival of *R. okamurae* in Azores represents its first record in the NE Atlantic islands. Although it may have been initially misidentified as a *Dictyota* spp., the phenotypic characteristics of *R. okamurae* are quite uncommon in the Azores, where most Dycytiales are smaller in size and exhibit morphological features that can be easily distinguish from *R. okamurae* (e.g. colour, branch dichotomy). A posterior analyses of video data taken in April 2019 indicates that *R. okamurae* may have been already present in the island since early that year. This finding was recorded in a location a few hundred meters from the main shipping port of São Miguel Island, pointing out for the likely vector of introduction of the species in the region. In subsequent years, the species underwent a rapid proliferation throughout the South coast of S. Miguel (Fig. 1, Table S3) dominating subtidal vegetations in both well illuminated and shaded rock surfaces. In some locations occupancy rates were close to 100%. Substantial amounts of the seaweed have also been cast away to the beach as wracks as observed during the first semester of 2021 (Fig. 2). Similar beach accumulations of this brown alga were reported at Faial Island in May 2021, but species identification requires further confirmation.

As mentioned before, and given the location where the species was first detected around the area of the main port of São Miguel, the introduction of *R. okamurae* into the Azores is likely to have been driven by human-assisted transport, via ballast waters and/or fouling (inlays in boat hulls). These are known to be two of the main introduction vectors of exotic species in the marine environment (Hewitt et al. 2009). Both routes could explain the introduction of *R. okamurae* in the region from Mediterranean populations that are proliferating across the Strait of Gibraltar, while taking advantage of the archipelago location as a known stop for transatlantic crossing. For instance, Rosas-Guerrero et al. (2018) observed that adult specimens of *R. okamurae* had survival rates between 80-

100% after being grown in dark conditions (a key condition during transport in ballast waters) for three weeks, maintaining the same survival rates when they passed to lighting conditions. Also, it has been shown that the species is capable of adhering to surfaces of very diverse nature and composition (García-Gómez et al. 2018) increasing its capacity to travel attached to boat hulls.

In São Miguel Island, *R. okamurae* is showing explosive growth and a surprising adaptation capacity. Its proliferation and high seabed occupancy rate in São Miguel may be explained by its great ability to propagate by vegetative (i.e. propagules) and asexual mechanisms (i.e. proliferous branchlets arising on the thallus surface which grow into new plants; Verlaque et al. 2009), as highlighted in the Strait of Gibraltar (Altamirano-Jeschke et al. 2017; Altamirano et al. 2019; García-Gómez et al. 2020). Also, anthropogenic disturbances (e.g. fisheries, ocean warming) and previous biological invasions (e.g. *Asparagopsis armata* Harvey 1855) (Martins et al. 2019) may have reduced niche competition in the region and favoured the arrival and spread of *R. okamurae* in such remote oceanic islands.

The species is known to cause serious impacts over previously established benthic communities in the Strait of Gibraltar (García-Gómez et al. 2020, 2021; Sempere-Valverde et al. 2021). *R. okamurae* not only has the potential to affect the ecological balance of the community, but also fundamental economic activities such as fishing and tourism. It can contribute to the loss of marine biodiversity and alteration of the structure of the communities, causing the physical displacement of native species due to substrate occupation and preventing the fixation of larvae or propagules of other species. This, together with the amounts of biomass observed at invaded locations, pose a significant ecological pressure to native ecosystems and local economies. This new invader should be kept monitored so that its status and potential impact on Azorean native biota can be evaluated. Given the species high level of proliferation there is a need of a rapid scientific response in providing valuable information so that appropriate preventive measures and mitigation actions can be applied by regional authorities to safeguard coastal marine habitats in Azores.

Acknowledgments: Funding was provided from National Funds through FCT—Fundação para a Ciência e Tecnologia, within the projects UID/BIA/00329/2015–2019

and UID/BIA/00329/2020–2023. ACLP was supported by PhD grant awarded by FRCT-Fundo Regional da Ciência e Tecnologia (M3.1.a/F/083/2015). In loving memory of Ana Neto.

References

Altamirano, M.J., J. De La Rosa and F.J. Martínez. 2016. Arribazones de la especie exótica *Rugulopteryx okamurae* (E.Y. Dawson) I.K. Hwang, W.J. Lee and H.S. Kim (Dictyotales, Ochrophyta) en el Estrecho de Gibraltar: primera cita para el Atlántico y España. *Algas* 52: 20.

Altamirano, M.J., J. De La Rosa, F.J.G. Martínez and A.R.G. Muñoz. 2017. Prolifera en el Estrecho un alga nunca citada en nuestro litoral de origen asiático. *Quercus* 374: 32-33.

De Clerck, O., F. Leliaert, H. Verbruggen, C.E. Lane, J.C. De Paula, D.I. Payo and E. Coppejans. 2006. A revised classification of the Dictyoteae (Dictyotales, Phaeophyceae) based on rbcL and 26S ribosomal DNA sequence data analyses. *J. Phycol.* 42: 1271–1288.

Draisna, S.G.A., W. F. Prud'homme van Reine, W.T. Stam and J.L. Olsen. 2001. A reassessment of phylogenetic relationships within the Phaeophyceae based on Rubisco large subunit and ribosomal DNA sequences. *J. Phycol.* 37: 586–603.

García-Gómez, J.C., J. Sempere-Valverde, E. Ostalé-Valriberas, M. Martínez, L. Olaya-Ponzzone, A.R. González et al. 2018. *Rugulopteryx okamurae* (E.Y. Dawson) I.K. Hwang, W.J. Lee & H.S. Kim (Dictyotales, Ochrophyta), alga exótica “explosiva” en el estrecho de Gibraltar. Observaciones preliminares de su distribución e impacto. *Almoraima* 48, 97–113.

García-Gómez, J.C., J. Sempere-Valverde, A.R. González, M. Martínez-Chacón, L. Olaya-Ponzzone, E. Sánchez-Moyano, et al. 2020. From exotic to invasive in record time: the extreme impact of *Rugulopteryx okamurae* (Dictyotales, Ochrophyta) in the strait of Gibraltar. *Sci. Total Environ.* 704: 135408.

García-Gómez J.C., M. Florido, L. Olaya-Ponzzone, J. Rey Díaz de Rada, I. Donázar-Aramendía, M. Chacón, J.J. Quintero, S. Magariño and C. Megina. 2021. Monitoring Extreme Impacts of *Rugulopteryx okamurae* (Dictyotales, Ochrophyta) in El Estrecho

Natural Park (Biosphere Reserve). Showing Radical Changes in the Underwater Seascapes. *Front. Ecol. Evol.* 9: 639161.

Hewitt, C.L., S. Gollasch and D. Minchin. 2009. The vessel as a vector – bio-fouling, ballast water and sediments. In: Rilov G. and J.A. Crooks (Ed.). *Biological invasions in marine ecosystems: Ecological, management, and geographic perspectives*. Springer-Verlag Berlin, Germany, pp. 117–131.

Hwang, I.K., W.J. Lee, H.S. Kim and O. De Clerck. 2009. Taxonomic reappraisal of *Dilophus okamurae* (Dictyotales, Phaeophyta) from the western Pacific Ocean. *Phycologia* 48:1–12.

Lee I.K. and J.A. Kang. 1986. A check list of marine algae in Korea. *Korean J. of Phycol.* 1: 311–325.

Martins G.M., E. Cacabelos, J. Faria, N.M. Álvaro, A.C.L. Prestes and A.I. Neto. 2019. Patterns of distribution of the invasive alga *Asparagopsis armata* Harvey: a multi-scaled approach. *Aquat. Invasions* 14(4): 582–593.

Ocaña, O., J. Afonso-Carrillo and E. Ballesteros. 2016. Massive proliferation of a dictyotalean species (Phaeophyceae, Ochrophyta) through the Strait of Gibraltar (Research note). *Rev. Acad. Canar. Cienc.* 28: 165–170.

Rosas-Guerrero, J., Y.E.Meco and M. Altamirano. 2018. Could *Rugulopteryx okamurae* (Dictyotales, Ochrophyta) have been introduced by ballast waters? *Algas* 54: 52.

Saunders, G.W. and T.E. Moore. 2013. Refinements for the amplification and sequencing of red algal DNA barcode and RedToL phylogenetic markers: a summary of current primers, profiles and strategies. *Algae* 28: 31–43.

Sempere-Valverde, J.J., J. Carlos García, E.O. Valribras et al. 2019. Expansion of the exotic brown algae *Rugulopteryx okamurae* (e.y. Dawson) I.K. Hwang, W.J. Lee & H.S. Kim in the strait of Gibraltar. *Proceedings of the 1st Mediterranean Symposium on the Non-Indigenous Species* (Antalya, Turkey, 18 January 2019). SPA/RAC, Tunis, 116 p.

Sempere-Valverde, J., E. Ostalé, M. Maestre, R. González, H. Bazairi and F. Espinosa. 2021. Impacts of the non-indigenous seaweed *Rugulopteryx okamurae* on a Mediterranean coralligenous community (Strait of Gibraltar): the role of long-term monitoring. *Ecol. Indic.* 121: 107135.

Silva, P.C., E.G. Menez and R.L. Moe. 1987. Catalog of the benthic marine algae of the Philippines. *Smithson. Cont. Mar. Sci.* 27: 1–179.

Verlaque, M., F. Steen and O. De Clerck. 2009. *Rugulopteryx* (Dictyotales, Phaeophyceae), a genus recently introduced to the Mediterranean. *Phycologia* 48: 536–542.

Womersley, H.B.S. 1987. *The marine benthic flora of southern Australia*. Part II. South Australian Government Printing Division.

Yoshida, T. 1998. *Marine Algae of Japan*. Uchida Rokakuho Publication, Tokyo.

Supplementary Material

Molecular protocol

Specimens used in molecular analyses are listed in Table S1. DNA was extracted using the DNeasy Plant Mini Kit (Qiagen) following the manufacturer's instructions.

Extracted DNA was used as a PCR template consisting of 2 µl DNA, 25 µl MyTaq HS Mix (Bioline), 18 µl sterile H₂O, and 0.6 µM of each primer (Table S2) for a total reaction volume of 50 µl. PCR protocols were as follows: i) *rbcL*: 96 °C for 3 min, 27 cycles of 96 °C for 1 min, 42 °C for 2 min, 72 °C for 2 min, followed by a final extension step of 72 °C for 10 min; ii) *psbA*: 94 °C for 2 min, 5 cycles of 94 °C for 30 s, 45 °C for 30 sec, 72 °C for 1 min, 35 cycles of 94 °C for 30 s, 46.2 °C for 30 sec, 72 °C for 1 min followed by a final extension step of 72 °C for 7 min. Amplification products were electrophoresed on a 1.5% agarose gel and positive PCR amplicons were sent to STABVIDA for purification and sequencing. Resulting sequences were edited to eliminate ambiguities and aligned with Bioedit v7.2 (Hall 1999).

Table S1. List of specimens collected in São Miguel island used for molecular analyses. Vouchers are deposited in the AZB Herbarium Ruy Telles Palhinha at the University of the Azores.

Sample code	Location	Date
SMG-20-07	Caloura	27-07-2020
SMG-20-11	Lagoa	28-07-2020
SMG-20-13	Baixa d'Ouro	29-07-2020
SMG-20-14	Caloura	27-07-2020
SMG-21-04	S. Roque	17-03-2021
SMG-21-24	Pópulo	17-05-2021

Table S2. List of oligonucleotides used in this study.

Primer name	Sequence (5' to 3')	Reference
<i>rbcL</i> 68F	TGCCWAAATGGGRWAYTGGGATGC	Draisna <i>et al.</i> 2001
S3R	AAACATCCTTGTGTA ACTCTC	Draisna <i>et al.</i> 2001
<i>psbA</i> F1	ATGACTGCTACTTTAGAAAGAC	Saunders and Moore 2013
<i>psbA</i> R2	TCATGCATWACTTCCATACCTA	Saunders and Moore 2013

Table S3. Field locations surveyed at São Miguel Island in 2019-2021. At each location, the DAFOR scale [D-dominant (>75%); A-abundant (50-75%); F-frequent (25-50%); 0 - occasional (10-25%); R-rare (<10%)] was applied to measure the relative abundance of the algae species along 10 min transect surveys.

Code	Location	Coordinates (decimal)	2019	2020	2021
1	Feteiras	37.803488, -25.802394	Not present	Not present	Abundant
2	Relva	37.768793, -25.747248	Not present	Not present	Dominant
3	São Roque	37.743929, -25.639247	Rare	Dominant	Dominant
4	Lagoa	37.741149, -25.573575	Not present	Dominant	Dominant
5	Caloura	37.707311, -25.506511	Not present	Occasional	Abundant
6	Vila Franca do Campo	37.713330, -25.437622	Not present	Not present	Abundant
7	Ponta Garça	37.713824, -25.373018	Not present	Not present	Abundant
8	Ribeira Quente	37.727905, -25.314917	Not present	Not present	Rare
9	Faial da Terra	37.739026, -25.196121	Not present	Not present	Not present
10	Nordeste	37.845703, -25.146526	Not present	Not present	Not present
11	Fenais da Ajuda	37.861622, -25.314817	Not present	Not present	Not present
12	Maia	37.835565, -25.393074	Not present	Not present	Not present
13	Ribeirinha	37.835735, -25.483861	Not present	Not present	Not present
14	Calhetas	37.824471, -25.606583	Not present	Not present	Not present
15	São Vicente	37.834168, -25.668119	Not present	Not present	Not present
16	Bretanha	37.899115, -25.750651	Not present	Not present	Not present
17	Mosteiros	37.899121, -25.814042	Not present	Not present	Not present
18	Ferraria	37.858455, -25.852884	Not present	Not present	Rare

References:

Draisna, S.G.A., W. F. Prud'homme van Reine, W.T. Stam and J.L. Olsen. 2001. A reassessment of phylogenetic relationships within the Phaeophyceae based on Rubisco large subunit and ribosomal DNA sequences. *J. Phycol.* 37: 586–603.

Hall, T.A. 1999. BioEdit: A User-Friendly Biological Sequence Alignment Editor and Analysis Program for Windows 95/98/NT. *Nucleic Acids Symp. Ser.* 41: 95-98.

Saunders, G.W. and T.E. Moore. 2013. Refinements for the amplification and sequencing of red algal DNA barcode and RedToL phylogenetic markers: a summary of current primers, profiles and strategies. *Algae* 28: 31–43.