

University of Nebraska - Lincoln

DigitalCommons@University of Nebraska - Lincoln

Agronomy & Horticulture -- Faculty Publications

Agronomy and Horticulture Department

5-11-2010

Artificial selection for determinate growth habit in soybean

Zhixi Tian

Purdue University, tian@purdue.edu

Xiaobo Wang

Chinese Academy of Agricultural Sciences, Beijing, wxbphd@163.com

Rian Lee

North Dakota State University, Rian.Lee@ndsu.edu

Yinghui Li

Chinese Academy of Agricultural Sciences, Beijing

James E. Specht

University of Nebraska-Lincoln, jspecht1@unl.edu

See next page for additional authors

Follow this and additional works at: <https://digitalcommons.unl.edu/agronomyfacpub>



Part of the [Plant Sciences Commons](#)

Tian, Zhixi; Wang, Xiaobo; Lee, Rian; Li, Yinghui; Specht, James E.; Nelson, Randall L.; McClean, Phillip E.; Qiu, Lijuan; and Ma, Jianxin, "Artificial selection for determinate growth habit in soybean" (2010). *Agronomy & Horticulture -- Faculty Publications*. 547.
<https://digitalcommons.unl.edu/agronomyfacpub/547>

This Article is brought to you for free and open access by the Agronomy and Horticulture Department at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in Agronomy & Horticulture -- Faculty Publications by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.

Authors

Zhixi Tian, Xiaobo Wang, Rian Lee, Yinghui Li, James E. Specht, Randall L. Nelson, Phillip E. McClean, Lijuan Qiu, and Jianxin Ma

Artificial selection for determinate growth habit in soybean

Zhixi Tian^{a,1}, Xiaobo Wang^{b,1}, Rian Lee^c, Yinghui Li^b, James E. Specht^d, Randall L. Nelson^e, Phillip E. McClean^{c,2}, Lijuan Qiu^{b,2}, and Jianxin Ma^{a,2}

^aDepartment of Agronomy, Purdue University, West Lafayette, IN 47907; ^bNational Key Facility for Crop Gene Resources and Genetic Improvement, Institute of Crop Sciences, Chinese Academy of Agricultural Sciences, Beijing 100081, China; ^cDepartment of Plant Sciences, and Genomics and Bioinformatics Program, North Dakota State University, Fargo, ND 58108; ^dDepartment of Agronomy and Horticulture, University of Nebraska, Lincoln, NE 68583; and ^eSoybean/Maize Germplasm, Pathology, and Genetics Research Unit, US Department of Agriculture–Agricultural Research Service, and Department of Crop Sciences, University of Illinois, Urbana, IL 61801

Edited* by Jeffrey L. Bennetzen, University of Georgia, Athens, GA, and approved April 2, 2010 (received for review January 6, 2010)

Determinacy is an agronomically important trait associated with the domestication in soybean (*Glycine max*). Most soybean cultivars are classifiable into indeterminate and determinate growth habit, whereas *Glycine soja*, the wild progenitor of soybean, is indeterminate. Indeterminate (*Dt1/Dt1*) and determinate (*dt1/dt1*) genotypes, when mated, produce progeny that segregate in a monogenic pattern. Here, we show evidence that *Dt1* is a homolog (designated as *GmTff1*) of *Arabidopsis* terminal flower 1 (*TFL1*), a regulatory gene encoding a signaling protein of shoot meristems. The transition from indeterminate to determinate phenotypes in soybean is associated with independent human selections of four distinct single-nucleotide substitutions in the *GmTff1* gene, each of which led to a single amino acid change. Genetic diversity of a minicore collection of Chinese soybean landraces assessed by simple sequence repeat (SSR) markers and allelic variation at the *GmTff1* locus suggest that human selection for determinacy took place at early stages of landrace radiation. The *GmTff1* allele introduced into a determinate-type (*tfl1/tfl1*) *Arabidopsis* mutants fully restored the wild-type (*TFL1/TFL1*) phenotype, but the *GmTff1* allele in *tfl1/tfl1* mutants did not result in apparent phenotypic change. These observations indicate that *GmTff1* complements the functions of *TFL1* in *Arabidopsis*. However, the *GmTff1* homeolog, despite its more recent divergence from *GmTff1* than from *Arabidopsis* *TFL1*, appears to be sub- or non-functionalized, as revealed by the differential expression of the two genes at multiple plant developmental stages and by allelic analysis at both loci.

comparative genomics | domestication | diversification | point mutation

Soybean (*Glycine max* L. Merr.) is one of the most economically important leguminous seed crops that provide the majority of plant proteins, and more than a quarter of the world's food and animal feed (1). It is suggested that soybean was domesticated from its annual wild relative, *G. soja* Sieb & Zucc, in China approximately 5,000 years ago (2), resulting in a multitude of soybean landraces that were adapted to various climate environments. Currently, 23,587 soybean landraces collected from 29 provinces of China are deposited in the Chinese GenBank, representing the world largest reservoir of soybean genetic diversity (3). Some of the landraces are still planted for production in several southern provinces, and some are used worldwide to develop modern soybean cultivars (2, 3).

Based on the timing of the termination of apical stem growth, most soybean cultivars can be classified into two categories of stem growth habit, commonly known as indeterminate and determinate types (4, 5). The apical meristems at the stem and branch apices in indeterminate cultivars maintain vegetative activity (i.e., produces new nodes with trifoliolate leaves) until photosynthate demand by developing seeds causes a cessation in the production of vegetative dry matter. In contrast, the apical meristems in determinate cultivars cease vegetative activity at or soon after photoperiod-induced floral induction, and then the meristems become reproductive inflorescences (6). Because determinacy is nonexistent (or rare) in

G. soja (4, 5), determinacy in the cultivated soybean is thought to be a trait associated with soybean domestication (7).

Previous studies demonstrated that the stem growth habit in soybean was primarily controlled by *Dt1* locus and that the indeterminate phenotype controlled by *Dt1/Dt1* was dominant or incompletely dominant over the determinate phenotype controlled by *dt1/dt1* (8, 9). This gene is a member of classical linkage group (LG) #5 (10), and was mapped to molecular marker linkage group (LG) L (11). Despite the monogenic inheritance pattern for the *Dt1* locus (6), a wide range in the abruptness of stem termination among soybean cultivars has also been observed, and a second gene locus, designated as *Dt2*, was reported (6). The *Dt2* allele is nearly dominant to the *dt2*, and in *Dt1/Dt1* genetic backgrounds, *Dt2/Dt2* genotypes produce semideterminate phenotypes and *dt2/dt2* genotypes produce indeterminate phenotypes. However, in *dt1/dt1* genetic backgrounds, the phenotype is determinate, because *dt1* is epistatic to *Dt2* and *dt2* (6). The *Dt2* locus was mapped to classical LG #6 (12) and from there to LG G (13). A third allele at the *Dt1* locus (*dt1-t*) has been identified that produces a phenotype that shares some characteristics of both *dt1* and *Dt2* (14).

It has been documented that it can be difficult to distinguish between indeterminate and determinate stem types under short photoperiod conditions or under adverse growing condition (6). As stem termination has great effects on plant height, flowering period, node production, maturity, water-use efficiency, and soybean yield (6, 15, 16), isolation and characterization of the genes associated with stem growth habit are very important for soybean germplasm assessment and breeding. In addition, characterization and analysis of these genes in soybean landraces and *G. soja* would allow us to understand the history and nature of human selection for determinacy.

The availability of the genome sequence and various “omics” tools and approaches for the model species such as *Arabidopsis* has aided the functional analyses of an increasing number of *Arabidopsis* genes and genetic pathways (17). Although the corresponding genetic pathways in other plant species are generally not known, several studies have identified the genes that are functionally conserved between model species and crops (18, 19). For

Author contributions: Z.T. and J.M. designed research; Z.T. and X.W. performed research; Z.T., R.L., Y.L., R.L.N., P.E.M., L.Q., and J.M. analyzed data; and Z.T., J.E.S., R.L.N., P.E.M., and J.M. wrote the paper.

The authors declare no conflict of interest.

*This Direct Submission article had a prearranged editor.

Freely available online through the PNAS open access option.

Data deposition: The sequences reported in this paper have been deposited in the GenBank database (accession nos. GU046912–GU047324).

¹Z.T. and X.W. contributed equally to this work.

²To whom correspondence may be addressed. E-mail: phillip.mcclean@ndsu.edu, qiu_lijuan@263.com, or maj@purdue.edu.

This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10.1073/pnas.1000088107/-DCSupplemental.

example, the *GAI* gene in *Arabidopsis* is functionally orthologous to the “Green Revolution” dwarfing gene in several cereal crops (18). It now seems clearer that the information gained from the model species can aid gene discovery and functional characterization in crops by the candidate gene approach (20), one of the applications for crop improvement that are collectively placed in the category of “plant translational genomics” (21).

Here, we used a combination of genetic linkage analysis, candidate gene association analysis, and heterologous transformation of *Arabidopsis* determinate (*tfl1/tfl*) mutants to infer the candidacy of a homolog of *Arabidopsis TFL1* in soybean for *Dt1*. In an attempt to track the history of artificial selection for determinacy, we investigated the allelic variation at the *GmTfl1* locus and its homeolog in *G. soja* accessions and in *G. max* cultivars, including a minicore collection of Chinese landraces in the context of their geographical distribution and population structure. This study illustrates how an *Arabidopsis* mutant was used as a shortcut to the characterization of natural mutations that were artificially selected in soybean.

Results

Identification of Soybean Genes Homologous to *Arabidopsis TFL1*.

The *Arabidopsis* TERMINAL FLOWER1 (*TFL1*) gene was previously identified by isolation of the recessive mutations *tfl1* in the *TFL1* gene by screening a M2 population derived from EMS-mutagenized seeds of ecotype Columbia (22). The recessive mutations resulted in the conversion of the normally indeterminate inflorescence to a determinate inflorescence condition (22–24). By BLAST searching *Arabidopsis TFL1* against the soybean (*c.v.*, Williams 82) whole genome sequence (25), we identified four soybean gene models, *Glyma03g35250.1*, *Glyma10g08340.1*, *Glyma13g22030.1*, and *Glyma19g37890.1*, that are homologous to *TFL1* (Fig. S1). Phylogenetic analysis of these genes suggest that *Glyma03g35250.1/Glyma19g37890.1* and *Glyma10g08340.1/Glyma13g22030.1* are two homeologous pairs, presumably derived from the soybean genome duplication event that occurred ~50 million years ago (MYA) (25, 26), whereas the two members of each pair likely resulted from the more recent soybean genome duplication event (i.e., allotetraploidization) (27) that took place ~13 MYA (Fig. S1) (26).

The *Dt1* locus of soybean was recently fine-mapped as a major quantitative trait locus between two simple sequence repeat (SSR) markers, Sat_099 and Satt229, on LG L (7), which is now designated as chromosome 19 (Gm19) (25). We anchored the SSR markers to the Gm19 sequence and found that *Glyma19g37890.1* (designated as *GmTfl1*) was one of the 380 annotated genes physically located between Sat_099 and Satt229 (24) (Fig. 1). This suggests that *GmTfl1* may be a candidate gene for *Dt1*. Because Williams 82 is a typical indeterminate cultivar, it is likely that *GmTfl1* is the candidate *Dt1* allele.

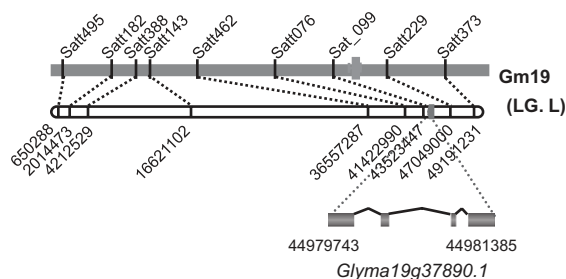


Fig. 1. Anchoring genetic markers to the genomic sequence to define the candidate *Dt1* gene. Vertical bar between Sat_099 and Satt006 on the genetic map and vertical bar on LG and chromosome sequence indicate the candidate *Dt1* gene, *Glyma19g37890.1*. Gene model was predicted and is depicted by the cartoon underneath the “chromosome.”

Allelic Variation of the Candidate Gene in the Wild and Cultivated Soybean Populations. In an attempt to address whether *GmTfl1* and the mutations, if any, that may have occurred in this gene are responsible for the conversion from an indeterminate to determinate phenotype observed in many soybean cultivars, we first sequenced the *GmTfl1* locus in a wild *G. soja* population and three soybean populations, representing genotypic groups that likely existed before and after genetic bottlenecks (e.g., domestication to produce landraces, introduction of relatively few landraces to North America, and selective breeding) (28). Fourteen unique SNPs and two insertions/deletions (indels) were detected (Table S1). Of the 14 SNPs, 10 were found in noncoding regions and four in exons. Interestingly, each of the four exonic SNPs generated a single amino acid nonsynonymous substitution (Table S1). Not a single individual genotype was found to contain more than one unique amino acid substitutions. Compared with the Williams 82 reference *GmTfl1* sequence, these four amino acid substitutions (referred as to *Gmtfl1*) were only detected in the cultivated soybeans, whereas *G. soja* genotypes were identical to Williams 82.

Association Between Determinacy and Allelic Nonsynonymous Mutations.

To elucidate whether *GmTfl1* is *Dt1*, and whether any or all of the mutations caused the transition from indeterminate type to determinate phenotype, we conducted an association analysis between the mutations and phenotypes using the three aforementioned soybean populations. The stem growth habit phenotypes of the soybean cultivars in these populations were obtained from the USDA Soybean Germplasm Collection database at National Plant Germplasm System (NPGS) (<http://www.ars-grin.gov/npgs/>), and some of them were directly examined in this study. Of the 89 soybean cultivars, 39 are indeterminate, 41 are determinate, and nine are semideterminate. We found that each of the 39 indeterminate cultivars exhibited the same amino acid sequence as encoded by *GmTfl1* in Williams 82, whereas none of the 41 determinate cultivars contain the Williams 82 amino acid sequence but instead possess one (or another) of the four amino acid substitutions (Fig. 2A and Table S2).

Two semideterminate cultivars in this study were found to have the *GmTfl1* allele. A previous study demonstrated that the genotype of *Dt1/Dt1* ordinarily displays an indeterminate phenotype, but in the presence of *Dt2*, a dominant allele at another locus controlling stem growth habit, the *Dt1/Dt1;Dt2/Dt2* genotype will display semideterminate phenotype (6); thus, these two semideterminate cultivars were assumed to contain both *Dt1* and *Dt2* alleles (Fig. 2A and Table S2). The other seven semideterminate cultivars were found to have *Gmtfl1* allele. Because it is generally difficult to precisely define the semideterminate phenotypes (6), these nine cultivars were not included in the association analysis below.

G. soja accessions are typically viny, and highly diverged in plant architecture and morphology from *G. max*; thus, the stem growth habit of the *G. soja* accessions included in this study was not carefully measured. All of the 20 *G. soja* accessions were found to contain the same *GmTfl1* genotype as Williams 82, which seemingly associates the indeterminacy with *G. soja* as is generally conjectured.

Thus, we observed a perfect association between the amino acid substitutions and the determinacy when *G. soja* accessions and the semideterminate cultivars were excluded. This suggests that *GmTfl1* is the *Dt1* allele, and the four single-point mutations (which could be characterized as functional SNPs) resulted in the four distinct amino acid substitutions are *dt1* alleles.

Excluding the four functional SNP variants, the *GmTfl1* alleles in the four populations were classified into two distinct types, designated as *GmTfl1-a* and *GmTfl1-b*. The four mutations were subclassifiable as *Gmtfl1-ta*, *Gmtfl1-bb*, *Gmtfl1-tb*, and *Gmtfl1-ab* (Fig. 2B). *GmTfl1-a* and *Gmtfl1-ta* share the same form, and *GmTfl1-b*, *Gmtfl1-bb*, *Gmtfl1-tb*, and *Gmtfl1-ab* share the other form, suggesting that *Gmtfl1-ta* was derived from *GmTfl1-a* whereas *Gmtfl1-bb*, *Gmtfl1-tb*, and *Gmtfl1-ab* were derived from *GmTfl1-b*. Linkage

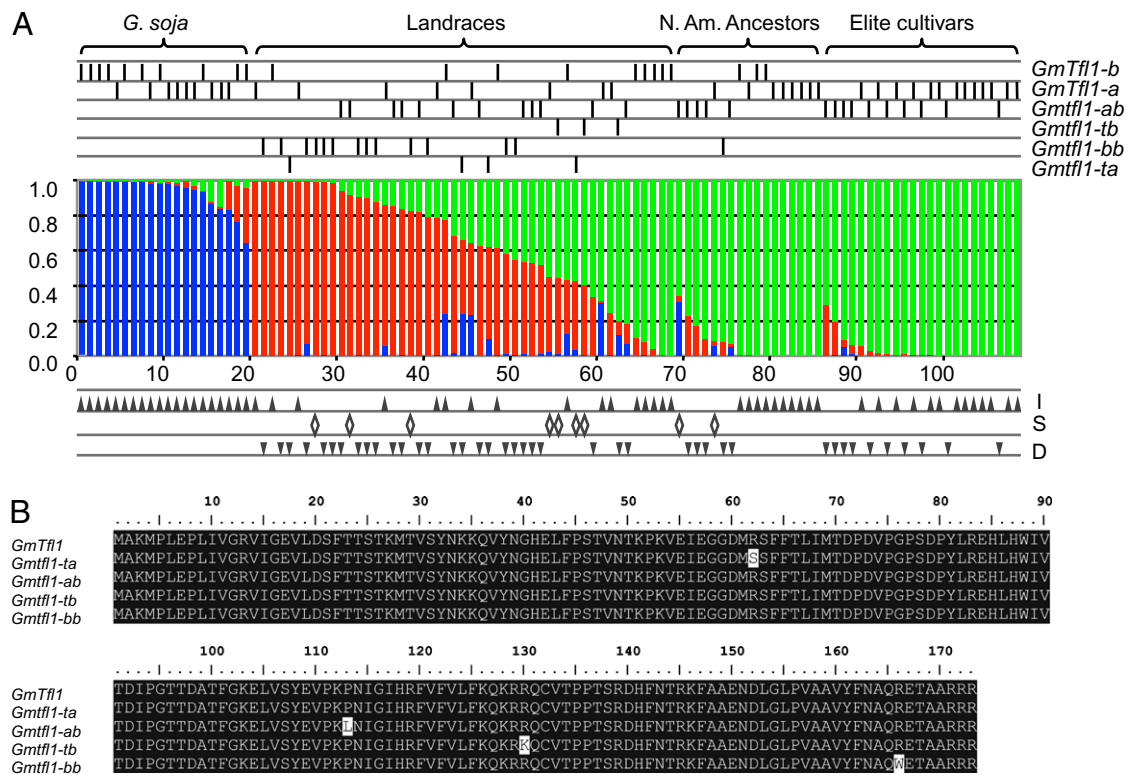


Fig. 2. Inferring the candidate gene by association analysis. (A) Distribution and association of four independent *Gmtf1* mutations with determinacy in four wild and cultivated soybean populations. The genetic structure of the populations was depicted by the vertical bars along the horizontal axis, in which the proportions of ancestry that can be attributed to each cluster were indicated by the length of each colored segment. The *GmTf1*/*GmTf1* or *Gmtf1*/*Gmtf1* genotypes of individual cultivars were marked by thin vertical bars above the plot of population structure, and their phenotypes, i.e., indeterminacy (I), semideterminacy (S), and determinacy (D), were indicated by up triangles, diamonds and down triangles, respectively. (B) Alignment of the amino acid sequences encoded by the *GmTf1* and *Gmtf1* alleles showing four single amino acid substitutions caused by four corresponding point mutations.

disequilibrium (LD) analysis showed that the SNPs and indels in the first intron (from +285 to +311) are linked with the two SNPs in the 5'UTR (−499 and −410), but the four functional SNPs did not show LD with the other sites (Fig. S24). We also sequenced *Glyma19g37900.1*, a gene flanking *GmTf1*, in six landraces that contain different alleles at the *GmTf1* locus, and found that LD exists between the SNPs detected at the *Glyma19g37900.1* locus and the nonfunctional polymorphisms (−499, −410, and +285 to +311) at the *GmTf1* locus (Fig. S2C). These observations indicate that the transition from indeterminate type to determinate type was not caused by the linked polymorphisms within the *GmTf1* locus, or between the *GmTf1* locus and its flanking gene, but by the four functional mutations. These observations further strengthen the inference that that *GmTf1* is *Dt1*.

In addition to the four populations analyzed above, we sequenced the *GmTf1* locus in 17 previously described determinate soybean cultivars (Table S3). All of the 17 cultivars were found to be *Gmtf1* mutations (two *Gmtf1-ta*, two *Gmtf1-tb*, and 13 *Gmtf1-ab*), a result consistent with the association analysis above. We also sequenced the *GmTf1* locus in three semideterminate isogenic lines that share the Clark (an indeterminate cultivar) genetic background but differ from Clark at the *Dt2* locus (6), and did detect the *GmTf1* allele in all these isogenic lines (Table S3).

***GmTf1* Complements the Functions of *TFL1* in *Arabidopsis*.** To validate the function of *GmTf1* for indeterminacy (vs. *Gmtf1* for determinacy) we introduced the Williams 82 *GmTf1* allele into the *Arabidopsis* determinate mutant (*tfl1-1*) (24) (Materials and Methods), and obtained two transgenic lines, one of which is shown in Fig. 3C. The absence of *Arabidopsis TFL1* allele and the presence of the

soybean *GmTf1* allele in the transgenic lines were confirmed by PCR analysis and sequencing of PCR fragments (Fig. 3 G and I). The transgenic (*GmTf1*) lines (Fig. 3 C and G) showed the same phenotypes as the wild-type *Arabidopsis* (i.e., indeterminate and late flowering). Because the transgene (*GmTf1*) is a combination of the *Arabidopsis TFL1* promoter and the protein coding sequence (CDS) of the *GmTf1* allele, the conversion of the transgenic lines from the mutant type (determinate and early flowering) to the wild type would be interpreted that the transgene in the *Arabidopsis* (*tfl1/tfl1*) mutant fully complements the functions of *TFL1* observed in the wild-type *Arabidopsis*.

The question remained whether the nonsynonymous substitutions (*Gmtf1* alleles) detected at the *GmTf1* locus in the cultivated soybean have no or diminished functions relative to the *GmTf1* allele for indeterminacy. To address this question, we introduced the *Gmtf1-ab*, the predominant allele detected in the cultivated soybean populations (Fig. 2A), into the *Arabidopsis tfl1-1* mutants (Materials and Methods), and obtained eight transgenic (*Gmtf1*) lines. The absence of the *Arabidopsis TFL1* allele and the presence of the soybean *Gmtf1-ab* allele in the eight transgenic lines were confirmed by PCR analysis and sequencing of PCR fragments (Fig. 3 H and J). We found that each of the eight lines showed phenotypes nearly identical to that of the *Arabidopsis tfl1-1* mutant. The phenotypes of one of the eight lines are illustrated in Fig. 3 D and H.

Evolutionary Diversification between *GmTf1* and its Homeolog. Since *GmTf1* and *Glyma03g35250.1* are thought to be a homeologous pair (Fig. 1B), it would be interesting to track the evolutionary divergence between *GmTf1* and *Glyma03g35250.1*. We thus sequenced the *Glyma03g35250.1* locus in the same populations used

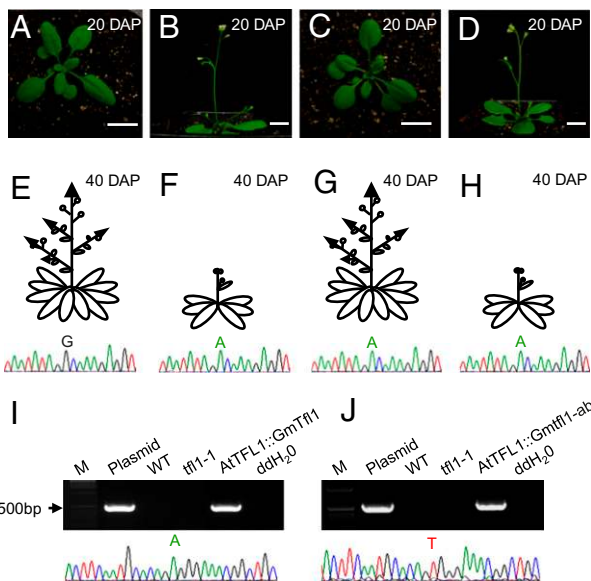


Fig. 3. Functional analysis of *GmTff1* and *Gmtff1* alleles in the *Arabidopsis tff1* mutants. (A) Wild-type *Arabidopsis* (*TFL1/TFL1*), (B) *tff1-1* mutant, (C) *tff1-1* mutant with transgene *GmTff1*. (D) *tff1-1* mutant with transgene *Gmtff1-ab*. (E–H) Cartoons of growths of the wild-type (*TFL1*), *tff1* mutant, the *GmTff1* transgenic line, and *Gmtff1-ab* transgenic line, as shown in A, B, C, and D, respectively. Curves and letters beneath the cartoons illustrate a single nucleotide difference (G and A) between *Arabidopsis TFL1* and *tff1-1* alleles detected in the three lines by sequencing. (I and J) Confirmation of presence of soybean *GmTff1* and *Gmtff1-ab* alleles, marked by a single nucleotide (A and T), respectively, in the transgenic *Arabidopsis tff1* lines by PCR and sequencing of PCR fragments.

for the analysis of the *GmTff1* locus. Five SNPs were detected at the *Glyma03g35250.1* locus in the *G. soja* population, but none were found in the cultivated populations (Table S1 and Fig. S2B). The level of nucleotide diversity at both *GmTff1* and *Glyma03g35250.1* loci (Table 1) is lower than the average in the *G. soja* population estimated based on a set of gene fragments (28). In addition, nonsynonymous substitutions were not found at either locus in the *G. soja* population (Table 1), suggesting that both genes have undergone purifying selection. However, *GmTff1* and *Glyma03g35250.1* exhibited a substantial difference in diversity in the cultivated soybean populations. For example, the *Glyma03g35250.1* allele was invariant among all of the members of the soybean landrace population, whereas the nonsynonymous substitutions at the *GmTff1* locus that resulted in the four *Gmtff1* alleles were observed in the same population (Table 1). Together, these observations suggest that the fixation of the four *Gmtff1* alleles in cultivated

determinate soybean would be the outcome of deliberate human selection during the development of soybean landraces.

Differential Expression of *GmTff1* and its Homeolog. To shed lights on the functional diversity of *GmTff1* and *Glyma03g35250.1*, we compared their expression pattern. Quantitative RT-PCR was used to profile the expressions of *GmTff1* and *Glyma03g35250.1*, in the indeterminate cultivar Williams 82 in different tissues and at different developmental stages, *GmTff1* was mainly expressed in young roots, young leaves and flowers seven day after flowering (7DAF), whereas *Glyma03g35250.1* was mainly expressed in young roots, young stems and buds (Fig. S3). Given that *Arabidopsis TFL1* is involved in inflorescence meristem development pathway (22, 24), high-level of expression of *GmTff1* in flowers 7DAF is expected. Thus, the lack of expression of *Glyma03g35250.1* at this stage may be considered as evidence that the *Glyma03g35250.1* was subfunctionalized or neofunctionalized. This inference is echoed by the analysis of allelic variation at both *GmTff1* and *Glyma03g35250.1* loci in the soybean populations. It can be deduced that neither *Glyma03g35250.1* nor the other pair of homeologous genes (*Glyma13g22030.1*, *Glyma10g08340.1*), homologous to *TFL1*, are potential candidates for the *Di2* locus, as these three loci are not located on LG G (chromosome 18), where the *Di2* was mapped.

Timing and Nature of Artificial Selection for the *Gmtff1* Alleles. None of the four *Gmtff1* alleles identified in *G. max* were detected in the *G. soja* population analyzed in this study. To search for evidence of the history of the human selection with respect to the *Gmtff1* alleles, we sequenced the *GmTff1* locus in a minicore collection of 195 soybean landraces, which were selected based on the genetic structure of a core collection of 1,863 landraces that maximally represent the 23,587 Chinese soybean landraces deposited in the Crop GenBank at the Chinese Academy of Agricultural Sciences. We subsequently analyzed the distribution of the four *Gmtff1* alleles in the core collection of landraces with respect to their genetic diversity and geographic distribution. The *Gmtff1* alleles were seen in all of the major branches of the Neighbor-Joining tree of the 195 landraces constructed based on 59 SSR markers (3) (Fig. 4A). It is noticeable that *Gmtff1-ta* and *Gmtff1-tb* were found in a highly diverged group of (seven) landraces that are the most closely related to *G. soja*, a wild accession used as an outgroup, and six of these seven landraces show “semi-wild” phenotypes, such as viny stems and dark brown seed coat (3). These data indicate that the human selection for determinacy must have occurred before the radiation of all of the lineages of these Chinese landraces, either just after or during the major domestication transition. Although the *Gmtff1* landrace alleles were found in all of the three large soybean-growing ecological regions, referred to as Northern eco-region (NR), Huang-Huai eco-region (HR), and Southern eco-region (SR), which were subclassified into NESp and NSp,

Table 1. Nucleotide diversity per base pair $\times 10^3$ in *GmTff1* and *Glyma03g35250.1*

	gDNA		CDS		Synonymous		Nonsynonymous	
	π	θ	π	θ	π	θ	π	θ
<i>GmTff1</i>								
All	1.86	1.21	1.39	1.46	0.00	0.00	1.83	1.91
Elite cultivars	1.86	0.98	0.98	0.52	0.00	0.00	1.29	0.68
Landraces	1.78	1.05	1.78	1.61	0.00	0.00	2.34	2.11
<i>G. soja</i>	1.65	1.28	0.00	0.00	0.00	0.00	0.00	0.00
<i>Glyma03g35250.1</i>								
All	0.15	0.49	0.20	0.36	0.87	1.57	0.00	0.00
Elite cultivars	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
Landraces	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
<i>G. soja</i>	0.66	0.73	0.85	0.54	3.65	2.32	0.00	0.00

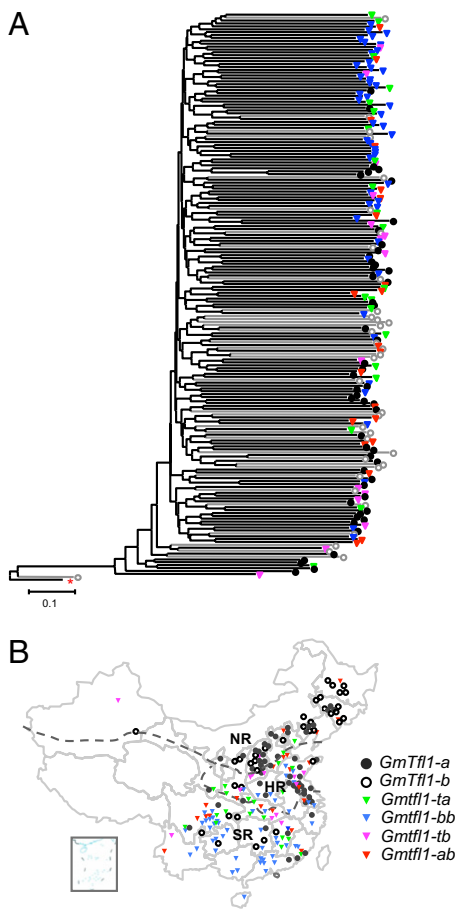


Fig. 4. Allelic mutations at the *GmTfl1* locus in the context of genetic diversity and eco-geographic distribution of a core collection of soybean landraces. (A) Phylogenetic relationship of the landraces assessed by 59 SSR markers and the types of alleles (*GmTfl1* or *GmTfl1*) detected in individual landraces. (B) Geographical distribution of the landraces in the soybean growing eco-regions or subregions in China.

HSp and HSu, and SR, CSp, SSp, SAu, and SSu subregions, respectively (3), the four *GmTfl1* alleles are mainly present in the landraces distributed in SR, *GmTfl1* is mainly found in the NR, and *GmTfl1* and *GmTfl1* are nearly equally distributed in HR (Fig. 4B, Table S4, and Fig. S4).

Discussion

Functional Conservation and Divergence of *TFL1* Homologs Within and Among Species. We demonstrated that the soybean *GmTfl1* gene is the functional homolog of the *Arabidopsis TFL1* gene by a comparative genomics approach. When *GmTfl1* was introduced into the *Arabidopsis tfl1* mutants, it fully restored the wild-type phenotypes, which are controlled by *TFL1* in the wild-type *Arabidopsis*. The functional homeolog of *TFL1* has been found in *Antirrhinum* (29), *Solanum lycopersicum* (30), and *Pisum sativum* (31), suggesting that the common mechanism underlies indeterminacy in these species.

Soybean *GmTfl1* was found to play the same roles as *Arabidopsis TFL1* in determining the inflorescence commitment and architecture (24) in the transgenic *Arabidopsis tfl1* mutant, but it does not seem to delay the commitment to inflorescence development in soybean. This is reflected by a general lack of correlation between the flowering time (i.e., late flowering and early flowering) and stem growth habit (i.e., indeterminacy and determinacy) of soybean cultivars. In addition, our expression data and allelic analysis at the *GmTfl1* and *Glyma03g35250.1* loci indicate that this

pair of homeologs has been sub- or neo-functionalized, likely after their duplication through allotetraploidization.

Natural Selection vs. Artificial Selection. Despite their functional divergence, *GmTfl1* and *Glyma03g35250.1* both appear to have undergone purifying selection, which is partly reflected by the lack of nonsynonymous substitutions at either *GmTfl1* or *Glyma03g35250.1* loci in the natural population of *G. soja* (Table 1). Our data revealed a total of four unique nonsynonymous substitutions in the domesticated soybean landraces, each of which led to the conversion of soybean stem habit from indeterminacy to determinacy. By contrast, no mutations present in *G. soja* at the *Glyma03g35250.1* locus were detected in the cultivated soybean populations. Given that more than 80% rare alleles presented in the *G. soja* population were eliminated through the bottleneck of soybean domestication (28), the appearance and maintenance of the *GmTfl1* alleles at such a high frequency in the soybean populations, which are currently absent in the *G. soja* population, must be assumed to be the outcome of deliberate artificial selection. Because only several semideterminate cultivars were identified in the populations investigated, *Dr2* was unlikely an allele associated with soybean domestication.

Artificial Selection, Linkage Disequilibrium, and Genetic Bottleneck. Although 50% of the *G. soja* genetic diversity was reduced through the bottleneck of soybean domestication (28), it appears that selection for the *GmTfl1* mutations did not cause apparent erosion of diversity. This was inferred by the observation that both indeterminate and determinate landraces in the minicore collection exhibited the similar levels of genetic diversity (Fig. S4). Instead, four *GmTfl1* alleles were observed among cultivated soybean, whereas *G. soja* only contained the *GmTfl1* allele. Genetic bottle necks are thought to reduce genetic diversity and increase LD (28). We found that LD in the *GmTfl1* locus and the flanking regions (Fig. S2C) is extremely high, but LD was decayed at *GmTfl1* alleles. This suggests that the selection for the *GmTfl1* alleles has had little effects on the genes linked to the *GmTfl1* locus. We found that *GmTfl1-ta* and *GmTfl1-tb* were absent in North American Ancestors, and *GmTfl1-bb* and *GmTfl1-a* were further eliminated from the Elite Cultivars developed in the USA, reflecting the effects of genetic bottlenecks created by soybean germplasm introduction and modern breeding (28).

Radiation and Adaptation of *GmTfl1/GmTfl1* Alleles to Local Eco-Regions. The domestication of soybean is hypothesized to have occurred in China, but there is no consensus about where within China it might have occurred. An early study proposed that soybean was domesticated in the Northeastern (NE) subregion within NR (32). However, a recent analysis of genetic structure and diversity of a core collection of Chinese soybean landraces demonstrated that the landraces collected from the region between 32.0 and 40.5°N, and 105.4 and 122.2°E along the central and downstream parts of the Yellow River (HSu subregion within HR) display the highest genetic diversity. This molecular data were used as evidence for the hypothesis that the cultivated soybean originated in the Yellow River region (3). Our observations are generally consistent with the latter hypothesis for a few reasons. First, *GmTfl1-b* was found to be the predominant allele in the *G. soja* accessions from the NESp subregion, but not a single *GmTfl1* allele derived from *GmTfl1-b* (i.e., *GmTfl1-bb*, *GmTfl1-tb*, and *GmTfl1-ab*) were detected in the landraces from this subregion. Second, because indeterminate cultivars were highly desirable in the NESp subregion, the determinate alleles were unlikely to be deliberately selected by humans and from there widely spread to other eco-regions, at least during or after the domestication event. Next, compared with all other subregions, the HSu subregion contains landraces that display the highest level of allelic variation at the *GmTfl1* locus (Fig. S4).

Regardless of the origin of the cultivated soybean, it is clear that the *GmTfl1* and *GmTfl1* alleles spread rapidly, fixed, and adapted to local eco-regions or subregions. The *GmTfl1* allele was favored

in the NR region, whereas *Gmtfl1* alleles were favored in the SR, and thus formed a middle region (i.e., HR) with *Gmtfl1* and *Gmtfl1* alleles fairly evenly distributed (Fig. 4 and Fig S3). Under the assumption that each landrace is homozygous at the *Gmtfl1* locus, which is highly supported by the high quality of nucleotides at the mutation sites, it is estimated that the frequencies of *Gmtfl1* and *Gmtfl1* in the landraces collected from the three major eco-regions, NR, HR, and SR, are 0.18 and 0.82, 0.50 and 0.50, and 0.81 and 0.19, respectively. We still do not know whether the *Gmtfl1* mutations were selected after the domestication event or integral to the process of domestication, but it is obvious the artificial selection of the natural *Gmtfl1* mutations played a central role in shaping the radiation of initially developed landraces. Because the determinate phenotype is shorter and thus more lodging-resistant in fertile production areas, its appearance during or after domestication probably resulted in an ancient “green revolution” in soybean cultivation in the southern parts of ancient China.

Materials and Methods

Plant Materials. The *G. soja* population and the three soybean populations previously described by Hyten et al. (28), and the 17 determinate cultivars and the *Dt2* isogenic lines listed in Table S3 were obtained from United States Department of Agriculture Soybean Germplasm Collection. The collection of Chinese soybean landraces previously described by Li et al. (3) were obtained from Chinese Academy of Agricultural Sciences (CAAS). The *ttf1-1* mutant was obtained from the Arabidopsis Biological Resource Center (ABRC).

DNA Isolation, PCR, and Sequencing. Genomic DNA isolation, PCR primer design, PCR amplification, PCR fragment purification, and sequencing of PCR fragments were conducted as described (33). Primers used for PCR amplification of *GmTfl1*, and *Glyma03g35250.1* were listed in Table S5.

Sequence Alignments, Genetic Structure, Linkage Disequilibrium, and Phylogenetic Analysis. The alignments of nucleotide and amino acid sequences were performed using MUSCLE (34). The observed nucleotide diversity (π) was calculated using DnaSP (35). The SNP data (28) and SSR marker data (3) were used to analyze the genetic structures of *G. soja* and *G. max* populations using the software package STRUCTURE (36). LD was evaluated using TASSEL (37). Neighbor-joining phylogenetic relationship of the minicore collection of Chinese landraces was analyzed by PowerMarker (38), rooted using a *G. soja* accession as an outgroup, and visualized by MEGA (39).

Plasmid Construction and Transformation. The promoter region of *Arabidopsis TFL1* (the same as that of *ttf1*) was fused with the CDS of *GmTfl1* (amplified from an indeterminate soybean cultivar Williams 82) or *Gmtfl1-ab* (amplified from a determinate soybean cultivar Young), and inserted to pCAMBIA1391 vector (CAMBIA). Then the constructs were introduced into the *Arabidopsis ttf1-1* mutants by the floral dip procedure (40). The absence of the *Arabidopsis TFL1* allele and the presence of the *GmTfl1* or *Gmtfl1-ab* constructs were confirmed by PCR and sequencing of PCR fragments. Primers used are listed in Table S5. All *Arabidopsis* plants were grown at 24 °C under the condition of 16 h of 120 $\mu\text{E}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ light and 8 h of dark.

RNA Extraction and Expression Analysis. Total RNA isolation, cDNA synthesis, and RT and quantitative PCR were conducted as previously described (41). The soybean *Actin11* gene was used as control. Primers used are listed in Table S5.

ACKNOWLEDGMENTS. We thank Dr. Enrico Coen for providing the wide-type *Arabidopsis* and the *ttf1-1* mutant seeds, Dr. Michael Zanis for insightful comments, and the anonymous reviewers for their constructive suggestions. This work was supported by the Purdue University faculty startup funds, the Indiana Soybean Alliance, the US National Science Foundation Plant Genome Research Program (DBI-0822258), the National Natural Science Foundation of China (30490251), the State Key Basic Research and Development Plan of China (973) (2004CB117203 and 2010CB125900), the State High-Tech (863) Plan (2006AA10A110, 2006AA102164, 2006AA10A111), and the National Key Technologies R&D Program in the 11th Five-Year Plan (2006BAD13B05).

- Graham PH, Vance CP (2003) Legumes: Importance and constraints to greater use. *Plant Physiol* 131:872–877.
- Carter T, Nelson R, Sneller C, Cui Z (2004) *Soybeans: Improvement Production and Uses* (Am Soc of America, Crop Sci Soc of America, Soil Sci Soc of America, Madison, WI), pp 303–416.
- Li Y, et al. (2008) Genetic structure and diversity of cultivated soybean (*Glycine max* (L.) Merr.) landraces in China. *Theor Appl Genet* 117:857–871.
- Ting CL (1946) Genetic studies on the wild and cultivated soybeans. *J Am Soc Agron* 38:381–398.
- Nagata T (1950) *Studies on the Characteristics of Soybean Varieties* (Jap Soybean Assoc, Tokyo), p 115.
- Bernard RL (1972) Two genes affecting stem termination in soybeans. *Crop Sci* 12: 235–239.
- Liu B, et al. (2007) QTL mapping of domestication-related traits in soybean (*Glycine max*). *Ann Bot (Lond)* 100:1027–1038.
- Woodworth CM (1932) Genetics and breeding in the improvement of the soybean. *Illinois Agr Exp Sta Bull* 384:297–404.
- Williams LF (1950) *Structure and Genetic Characteristics of the Soybean* (Interscience, New York).
- Weiss MG (1970) Genetic linkage in soybeans: Linkage groups V and VI. *Crop Sci* 10: 469–470.
- Shoemaker RG, Specht JE (1995) Integration of the soybean molecular and classical linkage groups. *Crop Sci* 35:436–446.
- Muehlbauer GJ, Specht JE, Staswick PE, Graef GL, Thomas-Compton MA (1989) Application of the near-isogenic line gene mapping technique to isozyme markers. *Crop Sci* 29:1548–1553.
- Cregan PC, et al. (1999) An integrated genetic linkage map of the soybean genome. *Crop Sci* 39:1464–1490.
- Thompson JA, Bernard RL, Nelson RL (1997) A third allele at the soybean *dt1* locus. *Crop Sci* 37:757–762.
- Heatherly LG, Smith JR (2004) Effect of soybean stem growth habit on height and node number after beginning bloom in the midsouthern USA. *Crop Sci* 44:1855–1858.
- Specht J, et al. (2001) Soybean response to water: A QTL analysis of drought tolerance. *Crop Sci* 41:493–509.
- Swarbreck D, et al. (2008) The *Arabidopsis* Information Resource (TAIR): Gene structure and function annotation. *Nucleic Acids Res* 36(Database issue):D1009–D1014.
- Peng J, et al. (1999) ‘Green revolution’ genes encode mutant gibberellin response modulators. *Nature* 400:256–261.
- Hayama R, Yokoi S, Tamaki S, Yano M, Shimamoto K (2003) Adaptation of photoperiodic control pathways produces short-day flowering in rice. *Nature* 422: 719–722.
- Pflieger S, Lefebvre V, Causse M (2001) The candidate gene approach in plant genetics: A review. *Mol Breed* 7:275–291.
- Salentijn EMJ, et al. (2007) Plant translational genomics: From model species to crops. *Mol Breed* 20:1–13.
- Shannon S, Meeks-Wagner DR (1991) A mutation in the *Arabidopsis TFL1* gene affects inflorescence meristem development. *Plant Cell* 3:877–892.
- Alvarez J, Guli CL, Yu X-H, Smyth DR (1992) Terminal flower: A gene affecting inflorescence development in *Arabidopsis thaliana*. *Plant J* 2:103–116.
- Bradley D, Ratcliffe O, Vincent C, Carpenter R, Coen E (1997) Inflorescence commitment and architecture in *Arabidopsis*. *Science* 275:80–83.
- Schmutz J, et al. (2010) Genome sequence of the paleopolyploid soybean (*Glycine max* (L.) Merr.). *Nature* 463:178–183.
- Shoemaker RC, Schlueter J, Doyle JJ (2006) Paleopolyploidy and gene duplication in soybean and other legumes. *Curr Opin Plant Biol* 9:104–109.
- Gill N, et al. (2009) *Molecular and chromosomal evidence for allopolyploidy in soybean, Glycine max (L.) Merr* (Plant Physiol).
- Hyten DL, et al. (2006) Impacts of genetic bottlenecks on soybean genome diversity. *Proc Natl Acad Sci USA* 103:16666–16671.
- Carpenter R, et al. (1995) Control of flower development and phyllotaxy by meristem identity genes in antirrhinum. *Plant Cell* 7:2001–2011.
- Pnueli L, et al. (1998) The SELF-PRUNING gene of tomato regulates vegetative to reproductive switching of sympodial meristems and is the ortholog of *CEN* and *TFL1*. *Development* 125:1979–1989.
- Foucher F, et al. (2003) DETERMINATE and LATE FLOWERING are two TERMINAL FLOWER1/CENTRORADIALIS homologs that control two distinct phases of flowering initiation and development in pea. *Plant Cell* 15:2742–2754.
- Hymowitz T (1970) On the domestication of the soybean. *Econ Bot* 24:408–421.
- Ma J, Bennetzen JL (2004) Rapid recent growth and divergence of rice nuclear genomes. *Proc Natl Acad Sci USA* 101:12404–12410.
- Edgar RC (2004) MUSCLE: Multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res* 32:1792–1797.
- Rozas J, Sánchez-DelBarrio JC, Messeguer X, Rozas R (2003) DnaSP, DNA polymorphism analyses by the coalescent and other methods. *Bioinformatics* 19:2496–2497.
- Pritchard JK, Stephens M, Donnelly P (2000) Inference of population structure using multilocus genotype data. *Genetics* 155:945–959.
- Bradbury PJ, et al. (2007) TASSEL: Software for association mapping of complex traits in diverse samples. *Bioinformatics* 23:2633–2635.
- Liu K, Muse SV (2005) PowerMarker: An integrated analysis environment for genetic marker analysis. *Bioinformatics* 21:2128–2129.
- Tamura K, Dudley J, Nei M, Kumar S (2007) MEGA4: Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0. *Mol Biol Evol* 24:1596–1599.
- Clough SJ, Bent AF (1998) Floral dip: A simplified method for Agrobacterium-mediated transformation of *Arabidopsis thaliana*. *Plant J* 16:735–743.
- Lin H, et al. (2009) DWARF27, an iron-containing protein required for the biosynthesis of strigolactones, regulates rice tiller bud outgrowth. *Plant Cell* 21:1512–1525.

Supporting Information

Tian et al. 10.1073/pnas.1000088107

SI Materials and Methods

Genetic Structure Analysis. The SNP data previously reported by Hyten et al. (1) and the SSR marker data described by Li et al. (2) were used to analyze the genetic structures of *G. soja* and *G. max* populations using the software package STRUCTURE (3). Admixture model and independent allele frequency model were used to analyze the dataset without prior population information. Five runs of STRUCTURE were done for each number of populations (K) (set from 1 to 10). Burn-in time and replication number were both set up at 100,000 per run.

Plasmid Construction and Transformation. A 2,500-bp upstream sequence upstream of start codon (ATG) of *AtTFL1* from *Arabidopsis* ecotype Colombia and the coding sequences of *GmTff1* or *Gmtff1-ab* from soybean indeterminate cultivar Williams 82 or determinate cultivar Young cDNA were amplified using KOD hot start DNA polymerase (Novagen, catalog no. 71086–3) with primers shown in Table S5. The PCR fragments were ligated to T-easy vector (Promega, catalog no. A1360) and sequenced. The clone containing 2,500-bp upstream of *TFL1* was digested using *Bgl*II and *Spe*I, and then ligated to *Bgl*II and *Spe*I digested pCAMBIA1391 vector (CAMBIA), forming pCAMBIA1391-TFL1-pro. The clone with *GmTff1* or *Gmtff1-ab* coding sequences were digested using *Spe*I and *Pml*II, and then ligated to *Spe*I and *Pml*II digested pCAMBIA1391-TFL1-pro, forming pCAMBIA1391-*AtTFL1*-*GmTff1* or pCAMBIA1391-*AtTFL1*-*Gmtff1-ab*, respectively, which contain a 2,455-bp upstream region of *AtTFL1* from *Arabidopsis* and the coding sequences of the *GmTff1* or *Gmtff1-ab* alleles from soybean. *Arabidopsis* transformation was performed by the floral dip procedure (4). The seeds were col-

lected from the infiltrated plants and selected in Murashige and Skoog medium containing 50 µg/mL kanamycin. Kanamycin-resistant plants were transferred to soil 9 day later and grown in a growth chamber. To confirm the transgenic lines, primer sets of Williams 82-*GmTff1*-CDS or Young-*Gmtff1-ab*-CDS were used (Table S5), and the *AtTFL1* or *tff1-1* alleles in wild type Colombia, *tff1-1* mutant, and the transgenic lines were also sequenced. All of the *Arabidopsis* plants were grown at 24 °C in growth chambers under LD condition (16 h of 120 µE·m⁻²·s⁻¹ light and 8 h of dark).

RNA Extraction and Expression Analysis. Total RNA was isolated using RNeasy Plant Mini (Qiagen, catalog no. 74903), treated with DNase I (Invitrogen, catalog no. 18068–015) and then used for cDNA synthesis with an RT kit (Invitrogen, catalog no. 18064–014) according to the manufacturer's instructions. cDNAs were resuspended in 50 µl water, and 1 µl was used per PCR. The PCR products were loaded on 2% (wt/vol) agarose gels. Real-time PCR experiments were performed in a total volume of 20 µl with 1 µl of the RT reactions, 2 mM gene-specific primers, and 10 µl SYBR Green Master mix (Applied Biosystems, catalog no. 4309155) with ABI Step One Plus real-time PCR machine (Applied Biosystems). The PCR program consisted of a first step of denaturation and Taq activation (95 °C for 10 min) followed by 45 cycles of denaturation (94 °C for 10 s), annealing (60 °C for 15 s), and extension (72 °C for 15 s). At the end, amplified products were denatured (95 °C), re-natured (65 °C), and progressively denatured (step from 65 to 95 °C over 30 min or 0.1 °C/s for the fusion curve analysis). The soybean *Actin11* gene was used as the internal control. The primer sequences used for PCR and sequencing are listed in Table S5.

- Hyten DL, et al. (2006) Impacts of genetic bottlenecks on soybean genome diversity. *Proc Natl Acad Sci USA* 103:16666–16671.
- Li Y, et al. (2008) Genetic structure and diversity of cultivated soybean (*Glycine max* (L.) Merr.) landraces in China. *Theor Appl Genet* 117:857–871.
- Pritchard JK, Stephens M, Donnelly P (2000) Inference of population structure using multilocus genotype data. *Genetics* 155:945–959.
- Clough SJ, Bent AF (1998) Floral dip: A simplified method for Agrobacterium-mediated transformation of *Arabidopsis thaliana*. *Plant J* 16:735–743.
- Schmutz J, et al. (2010) Genome sequence of the paleopolyploid soybean (*Glycine max* (L.) Merr.). *Nature* 463:178–183.

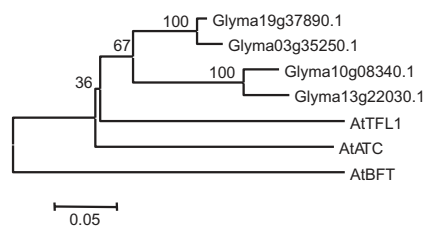


Fig. S1. Neighbor-Joining phylogenetic tree of the *AtTFL1*-*GmTff1* homologous genes constructed based on predicted amino acid sequences. *AtATC* and *AtBFT* are the other two homologous genes most closely related to *TFL1* in *Arabidopsis*.

