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# Ashwagandha (*Withania somnifera*): Role in Safeguarding Health, Immunomodulatory Effects, Combating Infections and Therapeutic Applications: A Review

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Abstract: Ashwagandha (Withania somnifera) is a well known herb possessing several health benefits. The steroidal lactones (withanolides) obtained from its roots have been implicated in a wide range of therapeutic activities and maintaining general health: Immunomodulation, combating infectious agents, anti-cancer and anti-epileptic, memory enhancer, to promote good physical and mental health, mood elevator, diuretic, general tonic and rejuvenator, stress reliever, cardiorespiratory endurance enhancer, anti-ageing, anti-oxidant, hypoglycemic, hypocholesterolemic and in common an effective adaptogen. Steroidal alkaloids and lactones are the active constituents of the plant. Withanolides as per theory occupies the receptor sites in the cell membrane thereby preventing the attachment and subsequent exertion of the effect of actual hormone. Withanolides have got analgesic and anti-inflammatory activity due to cyclooxygenase-2 inhibition property. Ashwagandha enhances nitric oxide synthatase activity of the macrophages, which in turn increases the microbial killing power of these immune cells thereby enhancing the Cell Mediated Immune (CMI) response. A glycoprotein Glycowithanolides (WSG) commonly known as W. somnifera glycoprotein is responsible for antimicrobial activity. Milk supplemented with Ashwagandha has been reported to increase total proteins and body weight and the plant alone helps in inducing tolerance and dependence. Its anti-stress and radiosensitization action; beneficial effects on cardiovascular system and sexual behavior; curative properties against neurodegenerative diseases and poisoning due to toxins and chemicals (including snake venom) has made this plant a treasure of nature. Thus the plant is an important component of many polyherbal preparations. Important for researchers and scientists is that biotechnologically advanced techniques; novel disciplines of bioinformatics and genomics can help in identifying and generating bioactive principles of the plant. All these salient health applications of this herb in biomedicine and veterinary sciences are discussed in this review focusing its potent role in maintaining sound health, immunomodulatory effects, combating infections, therapeutic usages and other beneficial applications.

**Key words:** Ashwagandha, indian ginseng, ayurveda, health, beneficial application, immunomodulation, treatment, therapy, memory enhancer, anticancer

## INTRODUCTION

Ashwagandha (Withania somnifera) is a well known herb possessing several health benefits and is an important 'Rasayana' as "Sattvic Kapha Rasayana" in Indian Ayurveda, used since centuries for its miraculous advantages (Mahima et al., 2012). Ashwagandha (Withania somnifera) is a traditional medicine with growing needs due to its remedial potentials. Chinese,

Unani, Ayurveda and Siddha literatures admire the therapeutic merits of plant-derived medicines against almost all ailments. Herbal medicines strongly involve mass appeal being safer and inexpensive. An esteemed Rishi (sage) Punarvasu Atriya was the first person who gave the teaching regarding the use of Ashwagandha that extends back over 3000 to 4000 years ago wherein its use is widely extolled as a tonic particularly for emaciation in all age group of people.

This causes enhancement of the reproductive function of both men as well as women (Mathur and Velpandian, 2009; Verma and Kumar, 2011; Mahima *et al.*, 2012; Dhama *et al.*, 2013a).

Ashwagandha is generally available in form of fine powder that can be used with water, ghee or honey (Gupta et al., 2006). The Nagori variety is the best among all Ashwagandha varieties. The health products made up of Ashwagandha (W. somnifera, Apocynaceae) becoming popular as commonly used medicinal plants. The steroidal lactones (withanolides) obtained from its roots have been implicated in a wide range of therapeutic activities and maintaining general health immunomodulation, combating infectious agents, anti-cancer, anti-epileptic, memory enhancer, to promote good physical and mental health, mood elevator, diuretic, rejuvenator, stress reliever, cardio-respiratory endurance enhancer, anti-ageing, anti-oxidant, hypoglycemic, hypocholesterolemic and in common an effective adaptogen (Scarfiotti et al., 1997; Adallu and Radhika, 2000; Hemalatha et al., 2006; Naidu et al., 2006; Mahima et al., 2012; Dhama et al., 2013a). Such alkaloids (withanolides) also work as "marker compounds/agents" for chemical standardization of Ashwagandha-based products (Dhuley, 2000; Shenoy et al., 2012).

Ashwagandha improves the memory by enhancing the brain and nervous function; promotes vigour and vitality along with cheerful sexual life and reproductive equilibrium; augments the body's pliability to stress being a powerful adaptogen; shows anxiolytic effect, has hepato-protective property, raises hemoglobin level and red blood cell count, improves energy levels, maintains mitochondrial health; has potent antioxidant properties so as to protect cellular damage caused by free radicals and improves the body's resistance against various ailments by improving the cell-mediated immunity (Scarfiotti et al., 1997; Bhattacharya and Muruganandam, Arora et al., 2004; Kuboyama et al., Harikrishnan et al., 2008; Sandhu et al., 2010). Ashwagandha is a potent adaptogen or vitalizer and has powerful antioxidant and detoxifying properties. Multiple actions of this miracle herb include antiinflammatory, analgesic, anti-stress, immunomodulatory, ant-microbial, cytoprotective, bettering anabolic activities, active against air-pollution and anti-cancer effects (Mishra et al., 2000; Kushwaha et al., 2012; Mahima et al., 2012, 2013a, b; Dhama et al., 2013a).

The present review describes Ashwagandha (W. somnifera) and its active compounds, mechanism of action and biological chemistry and classical beneficial applications of Ashwagandha in biomedicine and veterinary sciences viz., immunomodulatory effects,

activity against microbes and infection and usefulness as an alternative, chemotherapeutic agent, general health benefits, promoting vigour and vitality, stress reliever antidepressant, anti-inflammatory and adaptogenic property, effects on cardiovascular system, role in treating sexual disability, diseases and disorders, potent anti-cancer effects, reducing poisoning due to toxins/chemicals/drugs, anti-aging activities, memory enhancer, treating neurodegenerative disorders, role in development of drug tolerance and dependence.

#### CLASSIFICATION

# Ashwagandha (Withania somnifera)

#### Family-solanaceae/apocynaceae

#### Popular/common name: Indian ginseng/winter cherry:

Ashwagandha is an exceedingly valuable medicinal plant with valuable and wide therapeutic benefits in the conventional system of medicine. The plant grows in form of shrub with branching, height reaches to around 150 cm, leaves are up to 10 cm long; flowers present greenish or lurid yellow color, fruits/berries when mature are orange colored and its seeds are sown mostly during month of June or July (Khanna et al., 2006a; Dasgupta et al., 2008). Yield as well as quality of plant and its metabolites are affected by seasonal temperature, method of sowing, duration of light and dark period, depth of tillage, time of harvesting, concentration of fertilizers i.e., nitrogen, phosphorus, potassium application, effect of manure and field space present in between crops or density of plant population etc. (Kothari et al., 2003; Agarwal et al., 2004; Patel et al., 2004; Sreerekha et al., 2004; Ajay et al., 2005; Panchbhai et al., 2006).

Active constituents/compound/principle: The root of Withania somnifera has more than 35 chemical constituents (Rastogi and Mehrotra, 1998). Steroidal alkaloids and lactones (Withanolides, Withaferins): Anaferine alkaloid, anahygrine, isopelletierine, Ashwagandhanolide cuseohygrine, (dimeric thiowithanolide), chlorogenic acid, beta-Sisterol, fruit cysteine, iron, scopoletin, somniferinine, somniferiene, tropanol, withananine, withanoside IV, withanolides A-Y (Steroidal lactones) and saponins sitoindosides and acylsterylglucosides. The sitoindosides VII-X and withaferin-A are anti-stress agents which support immunomodulatory actions and have antifungal properties also (Abraham et al., 1975; Choudhary et al., 1995; Singh et al., 2006). Most of the pharmacological activities of Ashwaganda have been attributed to two main withanolides, withaferin A and withanolide D (Singh et al., 2010). Five-dehydroxy withanolide-R and with a somniferin-A are obtained from the aerial parts of

W. somnifera and effect of withaferin-A has been also seen on human blood lymphocytes. Withania somnifera is a rich source of iron (Davis and Kuttan, 2000a; Kuboyama et al., 2006; Subbaraju et al., 2006; Mirjalili et al., 2009).

Ethnopharmacological aspects: The pharmacological as well as metabolic effects of ashwagandha reveals that it has both herbal tonic as well as health food. In rats the swimming time is increased by Ashwagandha as determined by physical working capacity test (swimming endurance test). By employing such test it has been found that the weight of the heart increases relatively and the content of glycogen in myocardium increased significantly (Dhuley, 2000).

Two major classes of compounds viz., steroidal alkaloids and steroidal lactones are responsible for the wide range of beneficial effects of Ashwagandha. Withanolides are a class of compound included in the group of steroidal lactones and are responsible for antioxidant properties as well as free radical scavenging activities. Till date at least 12 alkaloids and 35 withanolides have been studied. Several studies have also revealed the antimicrobial properties of ashwagandha along with antibacterial activity against potentially dangerous like Salmonella (food poisoning causing organism). The ability of macrophage and immune cells to eat pathogens is enhanced by the root extract of Ashwagandha in comparison to macrophages (in control have not received ashwagandha group) that (Davis and Kuttan, 2000b; Anonymous, Govindarajan et al., 2005; Owais et al., 2005).

Treatment with Ashwagandha affords resistance of heart muscle of frog towards the toxic action of strophanthin-K and the duration of contractility is increased. The coagulation time is significantly increased by ashwagandha treatment resulting in attainment of normalcy after 7 days of cessation of treatment. In the blood serum of rats there is no any significant change in biochemical parameters. On the basis of such observations adaptogenic, cardioprotective as well as anticoagulant properties of ashwagandha are well understood (Dhuley, 2000).

Mechanism of action: Due to the property of helping in regulation of important physiologic processes Ashwagandha is assumed to be amphoteric. As per requirement withanolides act as important hormone precursors that has got the capability to convert into human physiologic hormones. The plant-based hormone precursor as per theory occupies the receptor sites in the cell membrane thereby preventing the attachment and

subsequent exertion of the effect of actual hormone. Small effect is exerted by the plant-based hormone if the level of original hormone is low (Misra, 2004). The anti-stress effect of ashwagandha was due to stimulation of respiratory function causing relaxation of smooth muscle along with stimulation of thyroid synthesis and secretion. Increase in dopamine receptors in the corpus callosum of brain induced by stress is suppressed by ashwagandha. Stress-induced increase in corticosterone in plasma along with blood urea nitrogen as well as blood nitric acid is also reduced. Anxiolytic effect of ashwagandha is exerted by acting as a gamma-aminobutyric acid (GABA) mimetic agent. The anticonvulsant activity by virtue of attachment to the GABA receptor is also a special feature of Ashwagandha (www.amazondiscovery.com).

Toxicological properties: Acute toxicity studies of ashwagandholine (total alkaloids from the roots of Withania somnifera) in 10% propylene glycol on the central nervous system. The acute LD<sub>50</sub> has been found to be a bit higher in rats (465 mg kg<sup>-1</sup>) than in mice (432 mg kg<sup>-1</sup>) (Mishra *et al.*, 2000). Sharada *et al.* (1993) tested acute (24 h) toxicity of alcohol extract from the roots of ashwaganda in swiss albino mice and subacute toxicity (30 days) in wistar rats. Single intraperitoneal injection of 1100 mg kg<sup>-1</sup> of the extract in mice did not produce any deaths within 24 h, but small increases in dose led to mortality. LD50 value was calculated as 1260 mg kg<sup>-1</sup> b.wt. Repeated injections of ashwagandha extract at a dose of 100 mg kg<sup>-1</sup> b.wt. for 30 days in either sex of wistar rats for subacute toxicity studies did not produce any mortality and no change in peripheral blood constituents. But, significant reductions in the weights of spleen, thymus and adrenals were observed in male rats at the end of the experiment. The acid phosphatase content of peripheral blood in both sexes showed a increase from control whereas other significant biochemical parameters were in the normal range.

Acute toxicity studies of *Withania somnifera* (L.) Dunal, WSF did not reveal any mortality and clinical signs of toxicity up to 2000 mg kg<sup>-1</sup> b.wt. Chronic administration of WSF did not cause any clinical signs of toxicity up to 1000 mg kg<sup>-1</sup> b.wt. Genotoxic study of WSF did not showed increase in percentage abnormal metaphases up to 1000 mg kg<sup>-1</sup> b.wt. Moreover, WSF was found to increase immunological response against antigenic stimuli (Sharma, 2011).

Dose-related tolerability, safety and activity of *Withania somnifera* formulation in normal individuals were evaluated in eighteen apparently healthy volunteers (12 male and 6 female) ageing about 18-30 years. The volunteers were treated with WS capsules (aqueous

extract, 8:1) daily in two divided doses with increase in daily dosage every 10 days for 30 days (750, 1000 and 1250 mg day<sup>-1</sup>×10 days). Except one volunteer, all tolerated WS without any adverse effects. One volunteer showed increased appetite, libido and hallucinogenic effects with vertigo at the lowest dose and was withdrawn from study. In six volunteers, improvement in quality of sleep was noticed. Reduction in total and LDL cholesterol, normal values in organ function tests, reduction in total body fat percentage and increase of strength of muscle activity was significant (Raut *et al.*, 2012).

Biological chemistry of Withania sominifera: Withanolide obtained from the plant possess analgesic and anti-inflammatory activity to its cyclooxygenase-2 inhibition property and Jayaprakasam, 2007). Peroxidases enzyme have been purified from this herb. Withanolides I-III and IV-V isolated from W. somnifera inhibited cholinesterase, acetylcholinesterase and butyrylcholinesterase, toxic phospholipase enzymes and therefore, are under consideration to be among the potent therapeutic candidate for treatment of Alzheimer's disease (Choudhary et al., 2004, 2005; Johni et al., 2005; Kambizi et al., 2006). Important constituent withanolides can be isolated and purified by various techniques mainly by High Performance Liquid Chromatography spectrometry (Khajuria et al., 2004; (HPLC) and Sharada et al., 2007).

Classical applications of ashwagandha: Ashwagandha attains the special name as the root smells like horse ("Ashwa") and it is believed that on guzzling it provides power of a horse. Different parts of Ashwagandha have significant therapeutic potency either as a whole plant extract or as separate constituents (Bhatt et al., 2006; Gupta and Rana, 2007). Utmost benefits comes out when fresh powder from the plant is used.

The root of Ashwagandha is very useful due to properties of being effective narcotic, diuretic, tonic, aphrodisiac, anthelmintic, astringent, antiangiogenic in case of tumor, antimutagenic, thermogenic and stimulant (Mohan *et al.*, 2004; Khanam and Devi, 2005a; Khanan *et al.*, 2006b; Mahima *et al.*, 2012).

Roots are equally effective against emaciation, constipation, debility, goiter, rheumatism, vitiated conditions of leucoderma, insomnia and nervine disorders, lead-induced DNA damage etc (Sharma *et al.*, 1985; Khanam and Devi, 2005b).

The paste of roots prepared with water effectively reduces the incidences of asthma, arthritis, rheumatoid, osteoarthritis, carbuncles, ulcers, leucorrhoea, boils, pimples, flatulent colic, piles and painful swellings as anti-arthritic agent when applied locally (Rasool and Varalakshmi, 2006b; Salve *et al.*, 2006).

The root in conjunction with other drugs is prescribed for snake venom as well as in scorpion-sting. Glycoprotein obtained from *W. somnifera* hampers activity of hyaluronidase and neurotoxic phospholipase A(2) and counteract the toxicity (Machiah and Gowda, 2006; Machiah *et al.*, 2006).

Studies demonstrated that root extract upregulates Th1-dominant polarization due to the presence of withanolide-A and hence supports the humoral immunity (HI) and Cell-mediated Immune (CMI) responses in BALB/c mice (Bani *et al.*, 2006; Malik *et al.*, 2007).

The leaves are bitter and recommended in fever and tender swellings (Kaur *et al.*, 2004).

It also shows phenomenon of cytomixis (Datta *et al.*, 2005; Kaul *et al.*, 2005).

The flowers are useful as astringent, depurative, diuretic and aphrodisiac (Singh *et al.*, 2011).

Fruits of Withania have potent inhibitory effect on peroxidation of lipid (Jayaprakasam et al., 2004).

The seeds on one side have anthelmintic property to expel the worms from the body and on another hand they are capable of increasing the sperm count and testicular growth (Abdel-Magied *et al.*, 2001).

Aqueous extract of plant can modulate the immune response of vaccine, such as DPT vaccine (Guatam *et al.*, 2004).

Ashwagandharishta prepared from seeds is used against conditions of hysteria, anxiety, memory loss etc. It also acts as a potent stimulant (Dadkar *et al.*, 1987).

**Immunomodulatory effects:** Extract of *W. somnifera* has been shown to significantly increase the Cell Mediated Immunity (CMI) in normal mice. Root extract enhances the level of interferon gamma (IFN-y), interlukin-2 (IL-2) and granulocyte macrophage colony stimulating factor (GM-CSF) in mice, suggestive of an immunopotentiating and myeloprotective effect. Ashwagandha enhances nitric oxide synthatase activity of the macrophages, which in turn increases the microbial killing power of these immune cells (Iuvone et al., 2003). It activates and mobilizes macrophages for rendering increased phagocytic activity, potentiates activity of lysosomal enzymes and acts as an anti-stress molecule and antiinflammatory agent in mice and rat (Rasool and Varalakshmi, 2006a). Immune enhancement with Ahswagandha has also been observed in mice with myelosuppression induced cyclophosphamide, by azathioprin and prednisolone. Root extract of W. somnifera has been reported to induce helper T-lymphocyte (Th1) polarised cell mediated immune response in BALB/c mice (Davis and Kuttan, 1998; Iqbal and Datta, 2006; Malik et al., 2007). Both immunostimulatory and immunosuppressive properties are present in Ashwagandha. It induces inhibition of delayed hypersensitivities (Auddy et al.Verma et al., 2012). Powdered root extract from Ashwagandha has profound effect on production of healthy white blood cells thereby it is an effective immunoregulator as well as chemoprotective agent in mice. The delayed type of hypersensitivity is also inhibited by this extract along with enhancement of phagocytic activities of macrophages while comparing with a control group. It has been found that the nitric oxide activities of the macrophages are enhanced by W. somnifera via induction of nitric oxide synthase enzyme activity. The plant is also responsible to cause senescence-specific regulation of the beta-galactosidase activity (Choudhary et al., 2004; Kiefer, 2006; Singh et al., 2010; Widodo et al., 2009).

Active against microbes and infection: Due to rapid emergence of antibiotic resistant strains of bacteria, treatment of infectious diseases is becoming challenging day by day and at the same time rapidly developing bacterial resistance is growing as a matter of global concern (Tiwari et al., 2013a). This alarming health concern particularly due to the continuous increase of immunocompromised patients demands various alternative therapeutic modalities such as bacteriophage, panchgavya, cytokine, herbal therapy and others (Tiwari and Hirpurkar, 2011; Mahima et al., 2012; Dhama et al., 2013b, c, d; Tiwari et al., 2013b, c, d).

Herbal therapy is an ancient revered therapy which is again gaining the momentum in lieu of need of alternative novel therapies and with least or no side effects this therapy is rapidly speeding the steps (Mahima et al., 2012; Dhama et al., 2013b). Though morphological, biochemical, functional and genetic variation exist but as a whole variety of herbs form a bouquet of safe, sound and easily available medicine (Dhar et al., 2006; Bandyopadhyay et al., 2007; Kuniar et al., 2007; Mahima et al., 2012). In regards to Ashwagandha besides other biological health promoting effects, the herb has also been found to illustrate antibacterial, antifungal and anti-viral effects.

A glycoprotein Glycowithanolides (WSG), commonly known as *Withania somnifera* glycoprotein, 28 kDa isolated from the *W. somnifera* root has demonstrated potent antimicrobial activity against the pathogenic fungi and bacteria. WSG protein put forth fungistatic effect in terms of inhibiting fungal spore germination and reduction of hyphal growth of *Fusarium oxysporum*, *F. verticilloides* and *Aspergillus flavus*. Antibacterial

effect has also been seen against *Clvibacter michiganensis* subsp. *Michiganensis bacteria*. *In vitro* antibacterial property of *Withania* plant in laboratory plant cell culture is also on hand. These findings persuade further studies to explore wide horizons of WSG as a budding therapeutic agent against various fungi and bacteria (Girish *et al.*, 2006; Jamil *et al.*, 2007; Kulkarni *et al.*, 2007).

W. somnifera plant has potent antibacterial property even against multidrug resistant (MDR) strains of microbes as withaferin and withanolides are the chief compounds. Withania has been found to be effective in the growth of several bacteria inhibiting Neisseria gonorrhoea, Escherichia coli, Salmonella, Pseudomonas fluorescens, P. aeruginosa, Bacillus subtilis, Listeria monocytogenes, Staphylococcus aureus and methicillin-resistant Staphylococcus aureus or Oxacillin resistant S. aureus (MRSA or ORSA) (Akinyemi et al., 2004; Owais et al., 2005; Kambizi and Afolayan, 2008; Mehrotra et al., 2011; Sundaram et al., 2011; El-Boshy et al., 2013). Ashwagandha has been shown to provide immunoprotection against Escherichia coli infection in Guinea pigs, Listeria monocytogenes infection in mice and Bordetella pertussis infection (Teixeira et al., 2006). It is a potent inducer of inhibiting Delayed Type of Hypersensitivities (DTH). Clinical health benefits of Ashwagandha have been reported in groups of Human Immunodeficiency Virus (HIV) infected patients and to treat the cases of genital herpes occurring due to herpes simplex virus type 1 and 2 (HSV1 and-2) (Kambizi et al., 2007). Ashwagandha is a potent antidepressant with the property to strengthen immunity against cold, flu and other common infections. Recently, ashwagandha has been found to ameliorate the effects of chicken infectious anaemia virus induced clinical parameters (haematological changes), pathology and pathogenesis in virus infected chicks, indicating protective potential of this herb in immunosuppressive viral disease of poultry (Latheef et al., 2013a, b).

General health benefits: Milk supplemented with Ashwagandha has been reported to increase total proteins and body weight (Venkataraghavan et al., 1980). It has a rejuvenative effect on the body mainly on the reproductive and nervous systems and is used to improve vitality and aid in recovery after chronic illness (Bhattacharya et al., 1987). Aphale et al. (1998) reported that the combination of Asgand (Withania somnifera) and Ginseng (Panax ginseng) was significantly increased the body weight, food consumption, liver weight and improved haematopoiesis when administered orally for 90 days using three doses in rats.

**Anti-stress agent:** Aswagandha is a potent anti-stress agent. It checks stress induced changes in adrenal function and augments protein synthesis. Ashawagandha with anti-stress activity is effective in increasing the physical endurance, plasma corticosterone level, sexual vigour, more sperm count, phagocytic index, cardiac activity, augmenting level of Th-1 cytokines, rising T lymphocytes proliferation and preventing stress induced ulcer, carbon tetrachloride (CCl<sub>4</sub>) induced hepatotoxicity and mortality (Ilayperuma et al., 2002; Tomi et al., 2005; Khan et al., 2006; Al-Qirim et al., 2007). Experimental studies in rats and mice showed same aforesaid effects when pre-treated with the crude form of Ashawagandha. Several studies have indicate the potent clinical and beneficial use of Ashwagandha (W. somnifera) in various health related issues viz., in the prevention and treatment of cyclophosphamide induced urotoxicity, protection of gonads in case of carbendazim toxicity, many stress induced diseases like arteriosclerosis, early ageing, arthritis, hypertension and malignancy due to its potent antistress, vitalizing and rejuvenating properties (Scarfiotti et al., 1997; Davis and Kuttan, 2000b; Bhattacharya and Muruganandam, 2003; Singh et al., 2011).

Rodents received a mild electric shock to their feet for a period of 21 days, resulting in stress induced effects like hyperglycemia, increase in plasma corticosterone levels, glucose intolerance, gastric ulcerations, male sexual dysfunction, immunosuppression and mental depression. Ashwagandha was given to the animals one hour before the electric shock and it is effectively reduces chronic stress in rodents (Bhattacharya and Muruganandam, 2003). The anti-anxiety effect of Ashwagandha was due to GABA-like activity, inhibitory neurotransmitter in the brain. It decreases the neuron activity and inhibits nerve cells from over firing, results in calming effect (Mehta *et al.*, 1991).

Ashwagandha has been used to stabilize mood in patients with behavioural disturbances. It has an anti-depressant and anti-anxiety effect in rodents when compared to the anti-depressant drug imipramine and the anti-anxiety drug lorazepam (Ativan) (Archana and Namasivayam, 1999). Ashwagandha is one of the most widespread tranquillizers used in India (Singh et al., 2010). Bhattacharya and Muruganandam (2003) compared the ability of Withania somnifera and Panax ginseng to relieve chronic stress syndrome in a rat model. They showed that both Ashwagandha and Panax ginseng decreased the frequency and severity of stress-induced ulcers, reversed stress-induced immunosuppression. reversed stress-induced inhibition of male sexual behaviour and inhibited the effects of chronic stress on retention of learned tasks, but only the Withania extract increased peritoneal macrophage activity.

Anti-oxidant activity: Root powder of *W. somnifera* has been reported to prevent Cadmium-induced oxidative stress in chicken and lead-induced oxidative damage in mouse (Chaurasia *et al.*, 2000; Mahadik *et al.*, 2008; Bharavi *et al.*, 2010). Plant extract protects myocardial cells from adverse effect of infarction or cardiac necrosis and stroke in rats which has molecular basis and high regeneration power as well (Arya *et al.*, 2004; Gupta *et al.*, 2004; Mohanty *et al.*, 2004; Sivanesan, 2007). *Withania* has showed antiulcer property and antioxidant activity also in rats along with improved calcification of bone in calcium-deficient ovariectomized rats (Sisodia and Bhatnagar, 2004; Bhatnagar *et al.*, 2005; Nagareddy and Lakshmana, 2006; Sumantran *et al.*, 2007a).

Withania somnifera has powerful antioxidants. It increases the levels of three natural antioxidant enzymes like superoxide dismutase, catalase and glutathione peroxidise in the brain of rats (Dhuley, 2000). Active principles of Withania somnifera root have antioxidant effects like anti-stress, cognition-facilitating, anti-inflammatory and anti-aging effects (Bone, 1996).

Anti-inflammatory effects: Ashwagandha acts as an effective anti-inflammatory agent and relives the symptoms of arthritis and variety of rheumatologic conditions. Naturally, it has much higher steroidal content than that of hydrocortisone (Anbalagan and Sadique, 1981). Begum and Sadique (1988) demonstrated that rats treated with powder of *Withania somnifera* root orally for 3 days, 1 h before injection of inflammatory agent produced anti-inflammatory responses comparable to that of hydrocortisone sodium succinate.

Effect on cardiovascular system: In humans, assessment of the hypoglycemic as well as diuretic and hypocholesterolemic effects of Ashwagandha root revealed that the treatment of subjects suffering from type 2 diabetes and mildly hypercholesterolemic can be initiated for a period of 30 days with a powder extract which results in a decrease in the glucose level in blood, comparable to that of an oral hypoglycemic drug. Significant increase in volume and sodium content in urine and decrease in cholesterol as well as triglycerides and low density lipoproteins in serum have also been observed (Bhattacharya and Muruganandam, 2003).

**Hepatoprotective activity:** Withaferin A has significant hepatoprotective effect in Ccl₄-induced hepatotoxicity in rats at a dose of 10 mg kg<sup>-1</sup> b.wt. (Rastogi and Mehrotra, 1998; Khare, 2007).

**Hypothyroid activity:** An aqueous extract of dried Ashwaganda root was administered to mice daily for 20 days to test thyroid activity. Significant increase in serum T4 levels indicates the stimulating effect at the glandular level via its effect on cellular antioxidant systems. These results indicate ashwaganda is a useful treating agent for hypothyroidism (Panda and Kar, 1998).

Anti-hyperglycaemic effect: Transina is a commercial preparation, which contains active ingredient of Ashwaganda and other components. It decreases streptozocin (STZ) induced hyperglycaemia in rats due to its pancreatic islet free radical scavenging activity (Bhattacharya et al., 2001).

Musculotropic activity: Alkaloids of Ashwaganda have muscle relaxant and antispasmodic effects against several spasmogens on bronchial, blood vascular, intestinal, uterine and tracheal muscles. This smooth muscle relaxant activity of the alkaloids was similar to that of papaverine which is a direct musculotropic agent (Anonymous, 1982).

Effect on sexual behaviour: Impairment in libido and sexual performance, sexual vigour as well as dysfunction in penile erection can be corrected by root extract of W. somnifera. The roots contain Fe, K, Mg and Ni which plays significant role in the diuretic, aphrodisiac activity and in the treatment of spermatopathia and seminal depletion. On cessation of treatment, these effects are partially reversible attributed and are hyperprolactinemic, Gamma Amino Butyric Acid (GABA), serotonergic or sedative activities of the extract instead of changes in levels of testosterone. Male sexual competence is detrimentally affected by roots of W. somnifera and thereby is contradictory. GABA mimetic activities of W. somnifera roots as well as serotonergic systems strongly depresses the libido (Ilayperuma et al., 2002).

Anti-cancer effects: The anti-cancer importance of the Ashwagandha plant has been well documented in a number of experimental studies and its extracts have potential use in cancer chemotherapy (Nath et al., 2005; Winters, 2006; Yang et al., 2007; Mathew et al., 2010; Dhama et al., 2013a). Literature reveals that Ashwagandha can be used as synergizer to support conventional chemo or radiation therapy due to its long term tumor growth inhibition property. Roots have been found to hamper the cellular growth and attachment of Chinese Hamster Ovary (CHO) carcinoma cells and thereby exerting the anti-tumor effect. This plant has also been found effective against uterine fibroids, dermatosarcoma, prostate cancer in

humans, urethane induced lung-adenoma in mice, neuroblastomas in humans, ascitic lymphoma, benzopyrene induced lung cancer in male Swiss Albino rat and leukaemia in humans (Singh et al., 1986; Devi, 1996; Christina et al., 2004; Senthilnathan et al., 2006a, b; Winters, 2006; Senthil et al., 2007; Sumantran et al., 2007b; Srinivasan et al., 2007; 2010; Kataria *et al.*, Widodo et al., 2013) (www.fibroids-and-endometriosis-help.com). Anticarcinogenic effects are mainly on account of decreased expression of nuclear factor-kappa-B, suppression of intercellular Tumor Necrosis Factor (TNF) and potentiation of apoptotic signaling in cancerous cells of animals or cell lines (Singh et al., 2010; Dhama et al., 2013e). Withaferin A is shown to inhibit umbilical vein endothelial cell (HUVEC) that sprouts three-dimensional collagen-I matrix at doses relevant to the inhibitory activity of NF-kappa B. In HUVECS, Withaferin A inhibits proliferation of cell at significant doses that are lower than those that are required for cell line of tumor origin via cyclin D1 expression inhibition. On the basis of these findings, it is proposed that in HUVECS NF-kappa B inhibition by Withaferin A occurs by interference of proteosome pathway mediated by ubiquitin. This is evident from the increase in the level of poly-ubiquinated proteins. Moreover, the finding that a potent anti-angiogenic activity is exerted by Withaferin A in vivo at lower doses than that required to induce anti-tumor activity in vivo highlights the use of this natural product obtained from W. somnifera to treat or prevent cancer (Mohan et al., 2004; Bargagna-Mohan et al., 2005; Ichikawa et al., 2006; Rao and Naresh, 2010; Dhama et al., 2013a).

A study regarding revealing the effect of W. somnifera root extracts on cell cycle and angiogenesis, as an anti-angiogenic compound showed Withaferin A and Withanolide D to inhibit growth of cancer (Maitra et al., 2003; Mathur et al., 2006). Ashwagandha plant extract inhibited benzo (a) pyrene-induced forestomach papillomagenesis, carrageenin induced air pouch granuloma and DMBA-induced skin papillomagenesis with up to 60 and 92% and 45 and 71% inhibition in tumor incidence and multiplicity, respectively in mice (Padmavathi et al., 2005). Genesis of papilloma of skin induced by 7, 12-dimethylbennzanthracene is inhibited by W. somnifera. In mice, however during the study this plant does not show any toxic effect apparently (Padmavathi et al., 2005). Sometimes, Ashwagandha (Indian ginseng) may produce interference in the immunoassay of serum digoxin level measurement (Dasgupta et al., 2007).

W. somnifera reduces tumor cell proliferation and mitigate undesirable side effects, hence increases overall animal survival time. It potentially supports radiation therapy and reduces the side effects produced by chemotherapeutic agents such as cyclophosphamide and paclitaxel without interfering with the tumor-reducing actions of the drugs. W. somnifera has been suggested to act as a novel complementary therapy in the field of oncology (Visavadiya and Narasimhacharya, 2007).

Recent in vitro studies in India has shown that the extract of the plant disrupt the ability of cancer cells to reproduce and is a significant step in fighting cancer. In addition it has been indicated by laboratory analysis that anti-angiogenic activity of ashwagandha extract against supporting unbridled growth. new blood vessels Oral administration ashwagandha extract effectively Inhibits the experimentally induced stomach cancers in laboratory animals. It reduces the incidence of tumor by 60% and multiplicity of tumor by 92%. Incidence and multiplicity of tumor are inhibited by 45 and 71%, respectively when study has been carried out in rodent model of skin cancer (Christina 2004; Mathur et al., 2004; Padmavathi et al., 2005).

Anti-cancer activity of Ashwagandha has been attributed to Hypothalamic Pituitary Adrenal (HPA) axis and the neuroendocrine system. It modulates the activity of cytotoxic lymphocytes (CTL) for reducing the tumour and cancerous growth. Augmentation of the Natural Killer (NK) cell activity reduces the tumor growth and incidences and increases serum T3 and T4 in mice (Panda and Kar, 1998) Compared to doxorubicin, Ashwagandha showed better efficacy in arresting growth of breast and colon cancer cell line (Jayaprakasam et al., 2003). Extracts of Ashwagandha also possess potent antioxidant and detoxifying properties (Rasool and Varalakshmi, 2007). Withaferin A stoutly exerts I kappa B kinase beta hyperphosphorylation for inhibition of its kinase action thereby causing death of cancerous cells (Kaileh et al., 2007; Wang et al., 2012).

Radiosensitization action and activities: Studies have revealed that a good natural source of a potent and relatively safe radio sensitizer/chemotherapeutic agent is Ashwagandha. The radiosensitizing effect of *W. somnifera* has been studied *in vivo* on the B16F1 mouse melanoma. Volume doubling time (the time required for a quantity to double in size) and growth delay in dose dependent manner has been observed by treating 100 mm<sup>3</sup> tumors with intraperitonial injection of 10-60 mg kg<sup>-1</sup> of Withaferin A. On the other hand, there has been a significant enhancement in tumor response due to gamma

irradiation locally followed by injection of Withaferin A at the rate of 30-50 mg kg<sup>-1</sup>. Such study has shown that the plant extract (Withaferin A) is effective mostly when injected intraperitonially 1 h prior to irradiation and thereby Withaferin A significantly enhance the radiation response of melanoma (Uma *et al.*, 2000; Diwanay *et al.*, 2004; Rao and Naresh, 2010).

## Reducer of poisoning due to toxins and chemicals/drugs:

Cancer is likely induced by chemotherapeutic agents like cyclophosphamide and cadmium and the side effects produced by chemotherapy as well as radiotherapy are sometimes more hazardous than the disease proper. Significant reduction in cyclophosphamide induced leucopenia has been observed due to administration of W. somnifera. When W. somnifera and cyclophosphamide together are used for treatment purposes, cellulairty of bone marrow increases significantly compared to treatment alone. Alpha-esterase cyclophophamide positive cells increase in number in the bone marrow of animals treated with cyclophosphamide due to administration of Ashwagandha because of stem cell proliferation (Kumar et al., 2011; Rahal et al., 2013). As per studies so far, toxicity induced by cadmium has not been reported to be regulated by any plant. But lipid peroxidation based results indicate that cadmium induced toxicity can be reduced by Ashwagandha, thereby indicating the potential of this plant to regulate metal induced toxicity (Panda et al., 1997). Ashwagandha is also known to significantly reduce the ochratoxin A induced suppression of chemotactic activity as well as interleukin IL-1 and TNF-alpha along with Asparagus racemosus (Satavari), Tinospora cordofolia and Picrorhiza kurroa (Katuki) (Mahima et al., 2012; Chakraborty and Pal, 2012; Dhama et al., 2013e). Tissue venom like hyaluronidase destroys the integrity of extracellular matrix thereby helping to spread toxin. It is an interesting finding that W. somnifera is a source of a hyaluronidase inhibitor, glycowithanolide (WSG) glycoprotein inhibits the hyaluronidase activity completely at a concentration of 1:1 w/w of snake venom to WSG, which could be help provide protection in case of Cobra (Naja naja) and Viper (Daboia russelii) bites. It is a scientific approach to use the Ashwagandha plant extract externally as an antidote to victims of snake bite in rural India. The Naja naja venom has got phospholipase A2 (PL-A2) activity which can be neutralized by glycoprotein isolated from W. somnifera known as antitoxin-PLA2. This has got implications in novel therapeutic reagent development as well as for treating snake evenomations along with implication in snake biology (Machiah et al., 2006).

Anti-aging activities: Double-blind clinical trial was conducted to test the anti-aging properties of Ashwagandha in a group of 101 healthy males, ageing about 50-59 years were given at a dosage of 3 g daily for one year. The volunteers showed significant improvement in hemoglobin, red blood cell count, hair melamin, seated stature, improvement in sexual performance, decrease in serum cholesterol and nail calcium was preserved (Bone, 1996; Ilayperuma et al., 2002).

Role against neurodegenerative disorders: Ashwagandha can be used as neuro-regenerative agent to treat Alzheimer's, Parkinson's, Huntington's and other neurodegenerative diseases at any stage of the disease as it can significantly reverse the neuritic atrophy and synaptic loss, along with GABA mediated anticonvulsant effect, GABA mimetic effect and promoting formation of dendrites due to therapeutic activity of glycowithanolides withaferin-A and sitoindosides VII-X present in the roots of Ashwagandha (Schliebs et al., 1997; Abbas et al., 2004, 2005; Ahmad et al., 2005). Ashwagandha enhances regeneration of the neurons along with reconstruction of synapse thereby acting as memory enhancer. The expression of Brain Derived Neurotropic Factor (BDNF) as well as Glial Fibrillary Acidic Protein (GFAP) is reversed by treatment with extract of Ashwagandha (Konar et al., 2011).

Alkaloids from the roots of Ashwagandha showed prolonged hypotensive, bradycardiac and respiratory stimulant activities in dogs. Hypotensive effect was mainly due to autonomic ganglion blocking action and was augmented by the depressant action on higher cerebral centres. In experimental ammals, total alkaloids produced a taming and a mild depressant effect like tranquillizer-sedative type on the CNS (Rastogi and Mehrotra. Systemic administration 1998). Ashwagandha root extract led to slightly enhanced acetylcholinesterase (ACHE) activity in the lateral septum and globus pallidus. Ashwagandha root extract affects mainly in the cortical and basal forebrain cholinergic signal transduction cascade. It increases cortical muscarinic acetylcholine receptor capacity leads to cognition-enhancing and memory-improving effects in animals and humans (Schliebs et al., 1997).

**Anticonvulsant activity:** Ashwagandha root extract significantly reduces the jerks in 70% animals and clonus in 10% animals caused by pentylene tetrazole (PTZ) induced convulsions when administered with dose of 100 mg kg<sup>-1</sup> and it was evident from EEG wave pattern (Kulkarni and George, 1996). It also showed reduction in severity of motor seizures induced by electrical

stimulation in right basilateral amygdaloid nuclear complex through bipolar electrodes. The protective effect was due to GABAergic mediation of Ashwagandha (Kulkarni *et al.*, 1993).

Role of Ashwagandha in development of drug tolerance and dependence: Drug addiction has become a major health problem worldwide and is a very costly affair to be managed. In drug addict individuals, tolerance as well as withdrawal signs are observed due to chronic treatment with benzodiazepine as well as ethanol or opioids that can be blocked by BR-16A (Mentat) which contains W. somnifera as one of its important ingredient. The morphine induced analgesic effect is significantly attenuated when the Ashwagandha plant extract is administered repeatedly for a period of 9 days. As per assessment by naloxone precipitation, withdrawal signs of development of dependence to opiate viz., morphine withdrawal jumps can be suppressed by W. somnifera. Studies have revealed that there is no dependence liability of the plant even upon cessation abruptly which have got clinical implications without production of long-term tolerance and withdrawal effect (Kulkarni and Sharma, 1994; Kulkarni and Ninan, 1997; Kest et al., 2002).

Biotechnological techniques as a stand-pillar in **upliftment of herbal therapy:** Medicinal plants are broadly used to deal with an array of health problems. The major impediments in the study of therapeutic herbal plants consist of erroneous identification and less yield of active principles such as medicinally important withanolides in case of Ashwagandha. Biotechnological advance techniques such as deoxyribonucleotide (DNA) based techniques like Polymerase Chain Reaction (PCR), stochastic algebraic modelling language (SAMPL), Restriction Fragment Length Polymorphism (RFLP), Amplified Fragment Length Polymorphism (AFLP), Random Amplified Polymorphic DNA (Rapd), High Performance Liquid Chromatography (HPLC), molecular cloning and sequencing of any target gene can help to resolve uncertainties in identification of appropriate plants and their constituents (Sharma et al., 2007). The novel disciplines of bioinformatics and genomics involving recombinant DNA techniques, in vitro organ and tissue/cell culture methods can be employed to produce bioactive alkaloids such as withanolides under definite conditions (Patra et al., 2004; Jha et al., 2005; Negi et al., 2006; Supe et al., 2006; Wadegaonkar et al., 2006; Titanji et al., 2007).

Commercial preparations: Ashwagandha is the principal component of many polyherbal preparations viz., Immu-21, Amrit<sup>®</sup>, Su-Ruksh<sup>®</sup>, Ashwagandha<sup>®</sup> and

ImmuPlus® etc (Sangwan et al., 2004). Immu-PlusR has been reported to stimulate blastogenic capacity of T and B cells and increases antibody titer in dogs. Polyherbal preparations like Ashwagandharist act as a nervine tonic and Himalaya ashwagandha is a monoherbal extract used for management of stress. Stresswin is used for reduction of anxiety, strain as well as stress. Stresscom is a monoherbal extract that relieves anxiety. Himalaya massage oil is a polyherbal preparation used for relaxation of body along with relief from stress. It is a good immunopotentiating agent when used along with vaccines, like in pups it improves immune stimulation when used in conjunction with parvovirus and rabies virus vaccines (Chauhan, 1999). Similar beneficial immunomodulatory effects have been observed in poultry birds while using with infectious bursal disease and Newcastle disease vaccines (Dhote et al., 2005). Not only this but in few fishes also such as Labeo rohita, famous as Indian major carp, 'ImmuPlus' has helped in increasing immunity and providing enhanced resistance against the diseases at different growth stages of fish life (Kumari et al., 2007). Immunomodulatory effects of Immu-21<sup>R</sup> have been reflected in modest improvements in conditions of HIV patients (Singh et al., 2001).

#### CONCLUSION

The uses of botanical medicines continue to grow with the expansion of modern medicine. The revered herb (Ashwagandha) potentiates the immune functions, enhances the longevity and facilitates the restoration of homeostasis by reducing the stress. Along with these, the role of Withania to exert beneficial effect against anxiety as well as cognitive and neurological disorders, inflammation and Parkinson's disease are quiet noteworthy. Ashwagandha is a potent stress reliever and antidepressant with the property to strengthen immunity against cold, flu and other common infections. Root powder is useful in treatment of acute rheumatoid arthritis. The roots are used as potent diuretic and aphrodisiac, increases sexual performance and help to maintain vigour and vitality. The plant is also a good anti-inflammatory agent and is useful in graft-vs-host reaction. Ashwagandha extract supports antioxidant immunomodulant activities. Withaferin A has powerful effects. It possesses metastatic antitumor angiogenetic properties of decresing order. Withania increases the body's ability to withstand stress of all types signifying anti-stressor adaptogenic property. Roots and leaves of Withania exhibit marked antibacterial activity against S. aureus, Neisseria gonorrhoea and anti-fungal activity against Candida albicans. These

facts indicate that W. somnifera can be regarded as a fine natural source of a potent and relatively safe radiosensitizer/chemotherapeutic agent. However the aforesaid benefits are documented in the literature but still multi-disciplinary evaluation is required with systematic approach before large scale commercialization of this miraculous herb. Proper caution should be taken while using this plant along with drugs that have anectodal effect (importantly barbiturates). The dose regimen should be given equal importance as in large dosage the plant extract causes gastrointestinal upset as well as diarrhea and vomition and may also have abortifacient effect (so better to avoid during pregnancy). Working hand-in-hand with oncologists as experienced natural medicine practitioners can effectively increase the therapeutic efficiency of Ashwagandha as well as decrease the side effects of W. somnifera when used for conventional treatments. In order to determine whether W. somnifera can duplicate the immunomodulatory and haematopietic activities in humans, optimal dosage for achieving these effects must be determined for which more research works are mandatory. Because of its wide pharmacological activities, Ashwagandha is considered as an important component of various polyherbal preparations. Thus the plant has got immense practical applicability in biomedicine as well as veterinary medicine focusing its potent role in the maintenance of sound health.

#### REFERENCES

Abbas, S.S., V. Singh, M. Bhalla and N. Singh, 2004. Clinical study of organic Ashwagandha in cases of parkinsonism, neuropathy, paralysis and uterine tumours (fibroids and other tumours) including Cutaneous Endodermal carcinoma. Proceedings of the National Seminar on Eco-Friendly Herbs of Ayurveda in Healthcare of Mankind: A Strategy for Scientific Evaluation an Uniform Standardization, January 9-11, 2004, Lucknow, pp: 81.

Abbas, S.S., M. Bhalla and N. Singh, 2005. A clinical study of organic Ashwagandha in some cases of uterine tumors (fibroids) and dermatofibrosarcoma. Proceedings of the Workshop on Essential Medicines, Adverse Drug Reactions and Therapeutic Drug Monitoring, August 2005, Lucknow, pp: 143-144.

Abdel-Magied, E.M., H.A. Abdel-Rahman and F.M. Harraz, 2001. The effect of aqueous extracts of *Cynomorium coccineum* and *Withania somnifera* on testicular development in immature Wistar rats. J. Ethnopharmacol., 75: 1-4.

- Abraham, A., I. Kirson, D. Lavie and E. Glotter, 1975. The withanolides of *Withania somnifera* chemotypes I and II. Phytochemistry, 14: 189-194.
- Adallu, B. and B. Radhika, 2000. Hypoglycemic, diuretic and hypocholesterolemic effect of winter cherry (*Withania somnifera*, Dunal) root. Indian J. Exp. Biol., 38: 607-609.
- Agarwal, M., P. Singh and M.K. Agarwal, 2004. Effect of sowing dates and spacing on yield attributes and root yield of ashwagandha, *Withania somnifera*. J. Med. Aromatic Plant Sci., 26: 473-474.
- Ahmad, M., S. Saleem, A.S. Ahmad, M.A. Ansari, S. Yousuf, M.N. Hoda and F. Islam, 2005. Neuroprotective effects of *Withania sominifera* on 6-hydroxydopamine induced Parkinsonism in rats. Human Exp. Toxicol., 24: 137-147.
- Ajay, R.P., K.S. Reddy, S. Ramana and B. Maji, 2005. Effect of nitrogen and farm yard manure on physiological parameters in Ashwagandra (*Withania somnifera*) under vertisol soil type. Indian J. Plant Physiol., 10: 389-393.
- Akinyemi, K.O., U.E. Mendie, S.T. Smith, A.O. Oyefolu and A.O. Coker, 2004. Screening of some medicinal plants used in south-west Nigerian traditional medicine for anti-Salmonella typhi activity. J. Herb. Pharmocother., 5: 45-60.
- Al-Qirim, T.M., A. Zafir and N. Banu, 2007. Comparative anti-oxidant potential of *Rauwolfia serpentina* and *Withania somnifera* on cardiac tissues. FASEB J., 21: 271-271.
- Anbalagan, K. and J. Sadique, 1981. Influence of an Indian medicine (Ashwagandha) on acute-phase reactants in inflammation. Indian J. Exp. Biol., 19: 245-249.
- Anonymous, 1982. The Wealth of India. Vol. 10, Publications and Information Directorate, CSIR, New Delhi, pp. 580-585.
- Anonymous, 2004. Monograph. Withania somnifera. Alternative Med. Rev., 9: 211-214.
- Aphale, A.A., A.D. Chibba, N.R. Kumbhakarna, M. Mateenuddin and S.H. Dahat, 1998. Subacute toxicity study of the combination of ginseng ginseng) (Panax and ashwagandha (Withania somnifera) in ras: Α safety assessment. Indian J. Physiol. Pharmacol., 11: 299-302.
- Archana, R. and A. Namasivayam, 1999. Antistressor effect of *Withania somnifera*. J. Ethnopharmacol., 64: 91-93.
- Arora, S., S. Dhillon, G. Ram and A. Nagpal, 2004. The in vitro antibacterial synergistic activities of Withania somnifera extracts. Fitoterapia, 75: 385-388.

- Arya, D.S., I. Mohanty, S.K. Ojha, M. Nandave and S.K. Gupta, 2004. Cardioprotective effects of hydro-alcoholic extract of *Withania somnifera* on isoproterenol-induced myocardial infarction in rats. J. Mol. Cellular Cardiol., 37: 279-279.
- Auddy, B., J. Hazra, A. Mitra, B. Abedon and S. Ghosal, 2008. A standardized *Withania sominifera* extract significantly reduces stress related parameters in chronically stressed humans: A double blind study, randomized, placebo-controlled study. JANA, 11: 50-56.
- Bandyopadhyay, M., S. Jha and D. Tepfer, 2007. Changes in morphological phenotypes and withanolide composition of Ri-transformed roots of *Withania somnifera*. Plant Cell Rep., 26: 599-609.
- Bani, S., M. Gautam, F.A. Sheikh, B. Khan and N.K. Satti et al., 2006. Selective Th1 up-regulating activity of Withania somnifera aqueous extract in an experimental system using flow cytometry. J. Ethnopharmacol., 107: 107-115.
- Bargagna-Mohan, P., R. Gambaro and R. Mohan, 2005. Withanolides: A new class of angiogenesis inhibitors. Abstracts Papers Am. Chem. Soc., 229: 161-161.
- Begum, V.H. and J. Sadique, 1988. Long term effect of herbal drug *Withania somnifera* on adjuvant induced arthritis in rats. Indian J. Exp. Biol., 26: 877-882.
- Bharavi, K., A.G. Reddy, G.S. Rao, A.R. Reddy and S.V. Rao, 2010. Reversal of cadmium-induced oxidative stress in chicken by herbal adaptogens *Withania somnifera* and *Ocimum sanctum*. Toxicol. Int., 17: 59-63.
- Bhatnagar, M., S.S. Sisodia and R. Bhatnagar, 2005.

  Antiulcer and antioxidant activity of Asparagus racemosus WILLD and Withania somnifera DUNAL in rats. Ann. N. Y. Acad. Sci., 1056: 261-278.
- Bhatt, P., D. Swarup, R. Ranjan and R.C. Patra, 2006. Evaluation of extracts of *Tinospora cordifolia* stem, *Terminalia arjuna* bark, *Withania somnifera* root and juice of *Allium sativum* cloves for amelioration of iron overload. Indian J. Anim. Sci., 76: 366-369.
- Bhattacharya, S.K., R.K. Goel, R. Kaur and S. Ghosal, 1987. Anti-stress activity of sitoindosides VII and VIII, new acylsterylglucosides from *Withania somnifera*. Phytotherapy Res., 1: 32-37.
- Bhattacharya, A., S. Ghosal and S.K. Bhattacharya, 2001. Anti-oxidant effect of *Withania somnifera* glycowithanolides in chronic footshock stress-induced perturbations of oxidative free radical scavenging enzymes and lipid peroxidation in rat frontal cortex and striatum. J. Ethnopharmacol., 74: 1-6.

- Bhattacharya, S.K. and A.V. Muruganandam, 2003. Adaptogenic activity of *Withania somnifera*: An experimental study using a rat model of chronic stress. Pharmacol. Biochem. Behav., 75: 547-555.
- Bone, K., 1996. Clinical Applications of Ayuvedic and Chinese Herbs. Phytotherapy Press, Queensland, Australia, pp. 137-141.
- Chakraborty, S. and S.K. Pal, 2012. Plants for cattle health: A review of ethno-veterinary herbs in veterinary health care. Ann. Ayurvedic Med., 1: 144-152.
- Chauhan, R.S., 1999. Effect of immuplus on humoral and cell mediated immunity in dogs. J. Immunol. Immunopathol., 1: 54-57.
- Chaurasia, S.S., S. Panda and A. Kar, 2000. Withania somnifera root extract in the regulation of lead-induced oxidative damage in the male mouse. Pharmacol. Res., 41: 663-666.
- Choudhary, M.I., Dur-E-Shahwar, Z. Parveen, A.Jabbar, I. Ali and Atta-ur-Rahman, 1995. Antifungal steroidal lactones from withania coagulance. Phytochemistry, 40: 1243-1246.
- Choudhary, M.I., S. Yousuf, S.A. Nawaz, S. Ahmed and Atta-ur-Rahman, 2004. Cholinesterase inhibiting withanolides from *Withania somnifera*. Chem. Pharmaceutical Bull. (Tokyo), 52: 1358-1361.
- Choudhary, M.I., S.A. Nawaz, Z. Ul-Haq, M.A. Lodhi and M.N. Ghayur et al., 2005. Withanolides, A new class of natural cholinesterase inhibitors with calcium antagonistic properties. Biochem. Biophys. Res. Commun., 334: 276-287.
- Christina, A.J.M., D.G. Joseph, M. Packialakshmi, R. Kothai, S.J.H. Robert, N. Chidambaranathan and M. Ramasamy, 2004. Anticarcinogenic activity of *Withania somnifera* dunal against Dalton's Ascitic lymphoma. J. Ethnopharmacol., 93: 359-361.
- Dadkar, V.N., N.U. Ranadive and H.L. Dhar, 1987. Evaluation of antistress (adaptogen) activity of *Withania somnifera* (Ashwagandha). Indian J. Clin. Biochem., 2: 101-108.
- Dasgupta, A., E. Kang, M. Olsen, J.K. Actor and P. Datta, 2007. Interference of Asian, American and Indian (Ashwagandha) ginsengs in serum digoxin measurements by a fluorescence polarization immunoassay can be minimized by using a new enzyme-linked chemiluminescent immunosorbent or turbidimetric assay. Arch. Pathol. Lab. Med., 131: 619-621.
- Dasgupta, A., G. Tso and A. Wells, 2008. Effect of Asian ginseng, Siberian ginseng and Indian Ayurvedic medicine Ashwagandha on serum digoxin measurement by Digoxin III, a new digoxin immunoassay. J. Clin. Lab. Anal., 22: 295-301.

- Datta, A.K., M. Mukherjee and M. Iqbal, 2005. Persistent cytomixis in *Ocimum basilicum* L. (Lamiaceae) and *Withania somnifera* (L.) Dun. (Solanaceae). Cytologia, 70: 309-313.
- Davis, L. and G. Kuttan, 1998. Suppressive effect of cyclophosphamide-induced toxicity by *Withania somnifera* extract in mice. J. Ethnopharmacol., 62: 209-214.
- Davis, L. and G. Kuttan, 2000a. Effect of Withania somnifera on cyclophosphamide-induced urotoxicity. Cancer Lett., 148: 9-17.
- Davis, L. and G. Kuttan, 2000b. Immunomodulatory activity of *Withania somnifera*. J. Ethnopharmacol., 71: 193-200.
- Devi, P.U., 1996. *Withania somnifera* Dunal (Ashwagandha): potential plant source of a promising drug for cancer chemotherapy and radiosensitization. Indian J. Exp. Biol., 34: 927-932.
- Dhama, K., S. Chakraborty and R. Tiwari, 2013a. Panchgavya therapy (Cowpathy) in safeguarding health of animals and humans: A review. Res. Opin. Anim. Vet. Sci., 3: 170-178.
- Dhama, K., S. Chakraborty, M.Y. Wami, R. Tiwari and R. Barathidasan, 2013b. Cytokine therapy for combating animal and human diseases: A review. Res. Opin. Anim. Vet. Sci., 3: 195-208.
- Dhama, K., S. Chakraborty, Mahima, M.Y. Wani and A.K. Verma *et al.*, 2013c. Novel and emerging therapies safeguarding health of humans and their companion animals: A review. Pak. J. Biol. Sci., 16: 101-111.
- Dhama, K., S. Mani, S. Chakraborty, R. Tiwari, A. Kumar, P. Selvaraj and R.B. Rai, 2013d. Herbal remedies to combat cancers in humans and animals-a review. Int. J. Curr. Res., 5: 1908-1919.
- Dhama, K., S.K. Latheef, H.A. Samad, S. Chakraborty, R. Tiwari, A. Kumar and A. Rahal, 2013e. Tumor necrosis factor as mediator of inflammatory diseases and its therapeutic targeting: A review. J. Med. Sci., 13: 226-235.
- Dhar, R.S., V. Verma, K.A. Suri, R.S. Sangwan and N.K. Satti et al., 2006. Phytochemical and genetic analysis in selected chemotypes of Withania somnifera. Phytochemistry, 67: 2269-2276.
- Dhote, B.S., G.K. Singh and R.S. Chauhan, 2005. Effect of immuplus (a herbal immunomodulator) on paraspecific immune responses in chicks. ISAH Warsaw Poland, 2: 60-65.
- Dhuley, J.N., 2000. Adaptogenic and cardioprotective action of Ashwagandha in rats and frogs. J. Ethnopharmacol., 70: 57-63.

- Diwanay, S., D. Chitre and B. Patwardhan, 2004. Immunoprotection by botanical drugs in cancer chemotherapy. J. Ethnopharmacol., 90: 49-55.
- El-Boshy, M.E.S., O.M. Abdalla, A. Risha and F. Moustafa, 2013. Effect of *Withania somnifera* extracts on some selective biochemical, hematological and immunological parameters in guinea pigs experimental infected with *E. coli.* ISRN Vet. Sci., Vol. 2013. 10.1155/2013/153427
- Girish, K.S., K.D. Machiah, S. Ushanandini, K. Harish Kumar and S. Nagaraju et al., 2006. Antimicrobial properties of a non-toxic glycoprotein (wsg) from Withania somnifera (ashwagandha). J. Basic Microbiol., 47: 365-374.
- Govindarajan, R., M. Vijayakumar and P. Pushpangadan, 2005. Antioxidant approach to disease management and the role of 'Rasayana' herbs of Ayurveda. J. Ethnopharmacol., 99: 165-178.
- Guatam, M., S. Diwanay, S. Gairola, Y.S. Shinde, S.S. Jadhay and B.K. Patwardhan, 2004. Immune response modulation to DPT vaccine by aqeous extract of *Withanis somnifers* in experimental system. Int. Immunopharmacol., 4: 841-849.
- Gupta, S.K., I. Mohanty, K.K. Talwar, A. Dinda and S. Joshi *et al.*, 2004. Cardioprotection from ischemia and reperfusion injury by Withania somnifera: A hemodynamic, biochemical and histopathological assessment. Mol. Cell. Biochem., 260: 39-47.
- Gupta, M.S., H.N. Shivaprasad, M.D. Kharya and A.C. Rana, 2006. Immunomodulatory activity of the ayurvedic formulation Ashwagandha Churna. Pharm. Biol., 44: 263-265.
- Gupta, G.L. and A.C. Rana, 2007. *Withania somnifera* (Ashwagandha): A review. Plant Rev., 1: 129-136.
- Harikrishnan, B., P. Subramamian and S. Subash, 2008. Effect of Withania somnifera root powder on the levels of circulatory lipid peroxidation and liver marker enzymes in chronic hyperammonemia. E-J. Chem., 5: 872-877.
- Hemalatha, S., A.K. Wahi, P.N. Singh and J.P. Chansouria, 2006. Hypolipidemic activity of aqueous extract of *Withania coagulans* dunal in Albino rats. Phytother. Res., 20: 614-617.
- Ichikawa, H., Y. Takada, S. Shishodia, B. Jayaprakasam, M.G. Nair and B.B. Aggarwal, 2006. Withanolides potentiate apoptosis, inhibit invasion and abolish osteoclastogenesis through suppression of nuclear factor-êB (NF-êB) activation and NF-êB-regulated gene expression. Mol. Cancer Ther., 5: 1434-1445.
- Ilayperuma, I., W.D. Ratnasooriya and T.R. Weerasooriya, 2002. Effect of *Withania somnifera* root extract on the sexual behaviour of male rats. Asian J. Androl., 4: 295-298.

- Iqbal, M. and A.K. Datta, 2006. A report on a paracentric inversion in *Withania somnifera* (L.) Dun. Indian J. Gen. Plant Breed., 66: 73-74.
- Iuvone, T., G. Esposito, F. Capasso and A.A. Izzo, 2003.
  Induction of nitric oxide synthatase expression by Withania somnifera in macrophages. Life Sci., 72: 1617-1625.
- Jamil, A., M. Shahid, M.M. Khan and M. Ashraf, 2007. Screening of some medicinal plants for isolation of antifungal proteins and peptides. Pak. J. Bot., 39: 211-221.
- Jayaprakasam, B., Y. Zhang, N.P. Seeram and M.G. Nair, 2003. Growth inhibition of human tumor cell lines by withanolides from *Withania somnifera* leaves. Life Sci., 74: 125-132.
- Jayaprakasam, B., G.A. Strasburg and M.G. Nair, 2004.
  Potent lipid peroxidation inhibitors from Withania somnifera fruits. Tetrahedron, 60: 3109-3121.
- Jha, S. M. Banyopadhyay, K.N. Chaudhuri, S. Ghosh and B. Ghosh, 2005. Biotechnological approaches for the production of forskolin, withanolides, colchicine and tylophorine. Plant Genet. Resour., 3: 101-115.
- Johri, S., U. Jamwal, S. Rasool, A. Kumar, V. Verma and G.N. Qazi, 2005. Purification and characterization of peroxidases from *Withania somnifera* (AGB 002) and their ability to oxidize IAA. Plant Sci., 169: 1014-1021.
- Kaileh, M., W.V. Berghe, A. Heyerick, J. Horion and J. Piette et al., 2007. Withaferin a strongly elicits IkappaB kinase beta hyperphosphorylation concomitant with potent inhibition of its kinase activity. J. Biol. Chem., 282: 4253-4264.
- Kambizi, L., P.O. Adebola and A.J. Afoayan, 2006. Effects of temperature, pre-chilling and light on seed germination of *Withania somnifera*: A high value medicinal plant. South African J. Bot., 72: 11-14.
- Kambizi, L., B.M. Goosen, M.B. Taylor and A.J. Afolayan, 2007. Anti-viral effects of aqueous extracts of *Aloe ferox* and *Withania somnifera* on herpes simplex virus type 1 in cell culture. S. Afr. J. Sci., 103: 359-360.
- Kambizi, L. and A.J. Afolayan, 2008. Extracts from *Aloe ferox* and *Withania somnifera* inhibit *Candida albicans* and *Neisseria gonorrhoea*. Afr. J. Biotechnol., 7: 12-15.
- Kataria, H., R. Wadhwa, S.C. Kaul and G. Kaur, 2013.
  Withania somnifera water extract as a potential candidate for differentiation based therapy of human neuroblastomas. PLoS One, Vol. 8. 10.1371/journal.pone.0055316

- Kaul, M.K., A. Kumar and A. Sharma, 2005. Reproductive Biology of *Withania somnifera* L. Dunal. Curr. Sci., 88: 1375-1377.
- Kaur, K., G. Rani, N. Widodo, A. Nagpal, K. Taira, S.C. Kaul and R. Wadhwa, 2004. Evaluation of the anti-proliferative and anti-oxidative activities of leaf extract from *in vivo* and *in vitro* raised Ashwagandha. Food Chem. Toxicol., 42: 2015-2020.
- Kest, B., C.A. Palmese, E. Hopkins, M. Adler, A. Juni and J.S. Moqil, 2002. Naloxone-precipitated withdrawal jumping in 11 inbred mouse strains: Evidence for common genetic mechanisms in acute and chronic morphine physical dependence. Neuroscience, 115: 463-469.
- Khajuria, R.K., K.A. Suri, R.K. Gupta, N.K. Satti, M. Amina, O.P. Suri and G.N. Qazi, 2004. Separation, identification and quantification of selected withanolides in plant extracts of *Withania somnifera* by HPLC-UV(DAD)-positive ion electrospray ionisation-mass spectrometry. J. Separation Sci., 27: 541-546.
- Khan, B., S.F. Ahmad, S. Bani, A. Kaul and K.A. Suri *et al.*, 2006. Augmentation and proliferation of T lymphocytes and Th-1 cytokines by *Withania somnifera* in stressed mice. Int. Immunopharmacol., 6: 1394-1403.
- Khanam, S. and K. Devi, 2005a. Antimutagenic activity of ashwagandha. J. Nat. Remedies, 5: 126-131.
- Khanam, S. and K. Devi, 2005b. Effect of *Withania somnifera* root extract on lead-induced DNA damage. J. Food Agric. Environ., 3: 31-33.
- Khanna, P.K., A. Kumar, A. Ahuja and M.K. Kaul, 2006a. Biochemical composition of roots of Withania somnifera (L.) dunal. Asian J. Plant Sci., 5: 1061-1063.
- Khanna, P.K., A. Kumar, A. Ahuja and M.K. Kaul, 2006b. Seed protein characterization for morphotype identification in *Withania somnifera* (L.) Dunal. Indian J. Plant Physiol., 11: 321-324.
- Khare, C.P., 2007. Indian Medicinal Plants: An Illustrated Dictionary. Springer, New Delhi, India, pp. 717-718.
- Kiefer, D., 2006. Ashwagandha: Stress reduction, neural protection and a lot more from an ancient herb. Life Extension Magazine. http://www.lef.org/magazine/mag2006/jun2006 report ashwa 01.htm.
- Konar, A., N. Shah, R. Singh, N. Saxena, S.C. Kaul, R. Wadhwa and M.K. Thakur, 2011. Protective role of ashwagandha leaf extract and its component withanone on scopolamine-induced changes in the brain and brain-derived cells. PloS One, Vol. 6. 10.1371/journal.pone.0027265

- Kothari, S.K., C.P. Singh, Y.V. Kumar and K. Singh, 2003. Morphology, yield and quality of ashwagandha (*Withania somnifera* L. Dunal) roots and its cultivation economics as influenced by tillage depth and plant population density. J. Horticul. Sci. Biotechnol., 78: 422-425.
- Kuboyama, T., C. Tohda and K. Komatsu, 2005a. Neuritic regeneration and synaptic reconstruction induced by withanolide A. Br. J. Pharmacol., 144: 961-971.
- Kuboyama, T., C. Tohda and K. Komatsu, 2006b. Withanoside IV and its active metabolite, sominone, attenuate A beta (25-35) induced neurodegeneration. Eur. J. Neurosci., 23: 1417-1426.
- Kulkarni, S.K., A. Sharma, A. Verma and M.K. Ticku, 1993. GABA receptor mediated anticonvulsant action of Withania somnifera root extract. Indian Drugs, 7: 305-312.
- Kulkarni, S.K. and A. Sharma, 1994. Reversal of diazepam withdrawal induced hyperactivity in mice by BR-16A (Mentat), a herbal preparation. Ind. J. Exp. Biol., 32: 886-888.
- Kulkarni, S.K. and B. George, 1996. Anticonvulsant action of Withania somnifera (Aswaganda) root extract against pentylenetetrazol-induced kindling in mice. Phytotherapy Res., 95: 447-449.
- Kulkarni, S.K. and I. Ninan, 1997. Inhibition of morphine tolerance and dependence by *Withania somnifera* in mice. J. Ethnopharmacol., 57: 213-217.
- Kulkarni, A.A., S.M. Kelkar, M.G. Watve and K.V. Krishnamurthy, 2007. Characterization and control of endophytic bacterial contaminants in vitro cultures of Piper spp., Taxus baccata subsp wallichiana and Withania somnifera. Can. J. Microbiol., 53: 63-74.
- Kumar, A., M.K. Kaul, M.K. Bhan, P.K. Khanna and K.A. Suri, 2007. Morphological and chemical variation in 25 collections of the Indian medicinal plant, *Withania somnifera* (L.) Dunal (Solanaceae). Gen. Resources Crop Evol., 54: 655-660.
- Kumar, S., P. Gupta, S. Sharma and D. Kumar, 2011. A review on immunostimulatory plants. J. Chinese Integr. Med., 9: 117-128.
- Kumari, J., P.K. Sahoo and S.S. Giri, 2007. Effects of polyherbal formulation 'ImmuPlus' on immumity and disease resistance of Indian major carp, *Labeo rohita* at different stages of growth. Indian J. Exp. Biol., 45: 291-298.
- Kushwaha, S., S. Roy, R. Maity, A. Mallick and V.K. Soni et al., 2012. Chemotypical variations in Withania somnifera lead to differentially modulated immune response in BALB/c mice. Vaccine, 30: 1083-1093.

- Latheef, S.K., K. Dhama, M.Y. Wami, H.A. Samad and R. Barathidasan *et al.*, 2013a. Ameliorative effects of four herbs (*Withania somnifera*, *Tinospora cordifolia*, *Azadirachta indica* and E care Se herbal) on the pathogenesis of chicken infectious anaemia virus. Int. J. Curr. Res., 5: 2327-2331.
- Latheef, S.K., K. Dhama, M.Y. Wani, H.A. Samad, R. Tiwari and S.D. Singh, 2013b. Ameliorative effects of Withania somnifera, Azadirachta indica, Tinospora cordifolia and E care Se herbal preparations on chicken infectious anaemia virus induced haematological changes in chicks and their live body weights. South Asian J. Exp. Biol., 3: 172-182.
- Machiah, D.K. and T.V. Gowda, 2006a. Purification of a post-synaptic neurotoxic phospholipase A2 from *Naja naja* venom and its inhibition by a glycoprotein from *Withania somnifera*. Biochimie., 88: 701-710.
- Machiah, D.K., K.S. Girish and T.V. Gowda, 2006b. A glycoprotein from a folk medicinal plant, Withania somnifera, inhibits hyaluronidase activity of snake venoms. Comp. Biochem. Physiol. C. Toxicol. Pharmacol., 143: 158-161.
- Mahadik, K.R., C.L. Gopu, S.S. Gilda, A.R. Paradkar and K.R. Mahadik, 2008. Comparative evaluation of antioxidant potential of Ashwagandha arishta and self generated alcoholic preparation of Withania somnifera dunal. Planta Med., 74: 288-288.
- Mahima, A. Rahal, R. Deb, S.K. Latheef and H.A. Samad *et al.*, 2012. Immunomodulatory and therapeutic potentials of herbal, traditional/indigenous and ethnoveterinary medicines. Pak. J. Biol. Sci., 15: 754-774.
- Mahima, A.M. Ingle, A.K. Verma, R. Tiwari and K. Karthik *et al.*, 2013a. Immunomodulators in day to day life: A review. Pak. J. Biol. Sci., 16: 826-843.
- Mahima, V.A.K., R. Tiwari, K. Karthik, S. Chakraborty, R. Deb and K. Dhama, 2013b. Nutraceuticals from fruit and vegetables at a glance: A review. J. Biol. Sci., 13: 38-47.
- Maitra, R., M.S. Kang and S. Sharma, 2003. Identification of a natural product, *Withania somnifera* L. dunal extract, as an antiangiogenic agent, a potential candidate for chemoprevention therapy. Proceedings of the American Association for Cancer Research Annual Meeting, Volume 44, July 11-14, 2003, Pittsburgh, pp. 604.
- Malik, F., J. Singh, A. Khajuria, K.A. Suri and N.K. Satti, 2007. A standardized root extract of Withania somnifera and its major constituent withanolide-A elicit humoral and cell-mediated immune responses by up regulation of Th1-dominant polarization in BALB/c mice. Life Sci., 80: 1525-1538.

- Mathew, T., Z. Mathew and K. Dhama, 2010. Plants and herbs for the treatment of cancer in human and animals. Proceedings of the National Seminar on Advances in Animal Cancer Research in India: Diagnosis, Treatment and Clinical Management, June 15-16, 2010, Indian Veterinary Research Institute, Izatnagar (U.P.)., India Souvenir, pp. 85-86.
- Mathur, S., P. Kaur, M. Sharma, A. Katyal, B. Singh, M. Tiwari and R. Chandra, 2004. The treatment of skin carcinoma, induced by UV B radiation, using 1-oxo-5beta, 6beta-epoxy-witha-2-enolide, isolated from the roots of *Withania somnifera*, in a rat model. Phytomedicine, 11: 452-460.
- Mathur, R., S.K. Gupta, N. Singh, S. Mathur, V. Kochupillai and T. Velpandian, 2006. Evaluation of the effect of *Withania somnifera* root extracts on cell cycle and angiogenesis. J. Ethnopharmacol., 105: 336-341.
- Mathur, R. and T. Velpandian, 2009. Medicinal plant-based health products: Where is the medicinal constituent? Ind. J. Pharmacol., 41: 205-206.
- Mehrotra, V., S. Mehrotra, V. Kirar, R. Shyam, K. Misra, A.K. Srivastava and S.P. Nandi, 2011. Antioxidant and antimicrobial activities of aqueous extract of *Withania somnifera* against methicillin-resistant *Staphylococcus aureus*. J. Microbiol. Biotech. Res., 1: 40-45.
- Mehta, A.K., P. Binkley, S.S. Gandhi and M.K. Ticku, 1991. Pharmacological effects of Withania somnifera root extract on GABAA receptor complex. Indian J. Med. Res., 94: 312-315.
- Mirjalili, M.H., E. Moyano, M. Bonfill, R.M. Cusido and J. Palazon, 2009. Steroidal Lactones from Withania somnifera, an ancient plant for novel Medicine. Molecules, 14: 2373-2393.
- Mishra, L.C., B.B. Singh and S. Dagenais, 2000. Scientific basis for the therapeutic use of *Withania somnifera* (Aswagaandha): A review. Altern. Med. Rev., 5: 334-346.
- Misra, B., 2004. Ashwagandha-bhavprakash nigantu (Indian Materia Medica) varanasi. Chaukhambha Bharti Academy, pp: 393-394.
- Mohan, R., H.J. Hmmers, P. Bargagna-Mohan, X.H. Zhan and C.J. Herbstritt *et al.*, 2004. Withaferin A is a potent inhibitor of angiogenesis. Angiogenesis, 7:115-122.
- Mohanty, I., D.S. Arya, A. Dinda, K.K. Talwar, S. Joshi and S.K. Gupta, 2004. Mechanisms of cardioprotective effect of *Withania somnifera* in experimentally induced myocardial infarction. Basic Clin. Pharmacol. Toxicol., 94: 184-190.

- Nagareddy, P.R. and M. Lakshmana, 2006. *Withania somnifera* improves bone calcification in calcium-deficient ovariectomized rats. J. Pharm. Pharmacol., 58: 513-519.
- Naidu, P.S., A. Singh and S.K. Kulkarni, 2006. Effect of Withania somnifera root extract on reserpine induced orofacial dyskinesia and cognitive dysfunction. Phytotherapy Res., 20: 140-146.
- Nair, M.G. and B. Jayaprakasam, 2007. Cyclooxygenase-2 inhibitory withanolide compositions and method. Board of Trustees of Michigan State University, USA.
- Nath, A., P. Sinha and P. Sinha, 2005. Avenues for potential therapeutic use of anticancer properties of Vinca rosea, Withania somnifera and Ocimum sanctum. Indian J. Med. Res., 121: 43-44.
- Negi, M.S., V. Sabharwal, N. Wilson and M.S. Lakshmikumaranl, 2006. Comparative analysis of the efficiency of SAMPL and AFLP in assessing genetic relationships among *Withania somnifera* genotypes. Curr. Sci., 91: 464-471.
- Owais, M., K.S. Sharad, A. Shehbaz and M. Saleemuddin, 2005. Antibacterial efficacy of *Withania somnifera* (Ashwagandha) an indigenous medicinal plant against experimental murine salmonellosis. Phytomedicine, 12: 229-235.
- Padmavathi, B., P.C. Rath, A.R. Rao and R.P. Singh, 2005. Roots of Withania somnifera inhibit forestomach and skin carcinogenesis in mice. Evidence-Based Complement Altern. Med., 2: 99-105.
- Panchbhai, D.M., B.R. Bachkar, P.L. Deokar and S.G. Wankhade, 2006. Effect of nitrogen and phosphorus on root yield and quality of ashwagandha (*Withania somnifera* Dunal.). Adv. Plant Sci., 19: 89-92.
- Panda, S., P. Gupta and A. Kar, 1997. Protective role of Ashwagandha in cadmium induced hepatotoxicity and nephrotoxicity in male mouse. Curr. Sci., 72: 546-547.
- Panda, S. and A. Kar, 1998. Changes in thyroid hormone concentrations after administration of ashwagandha root extract to adult male mice. J. Pharm. Pharmacol., 50: 1065-1068.
- Patel, D.H., P.N. Upadhyay, K.V. Patel, J.B. Patel and B.K. Patel, 2004. Effect of method of sowing, time of harvesting and nitrogen application on dry root yield of Aswagandha (*Withania somnifera*). J. Med. Aromatic Plant Sci., 26: 288-292.
- Patra, D.D., K. Singh, H.O. Misra, A.K. Gupta, J. Singh, S.C. Singh and S.P.S. Khanuja, 2004. Agrotechnologies of Ashwagandha (Withania somnifera). J. Med. Aromatic Plant Sci., 26: 332-335.

- Rahal, A., A.H. Ahmad, A. Kumar, Mahima and A.K. Verma *et al.*, 2013. Clinical drug interactions: A holistic view. Pak. J. Biol. Sci., 16: 751-758.
- Rao, P.M. and K. Naresh, 2010. Aswagandha (*Withania somnifera*)-Ayurvedic bequest for the patients of cancer: An update on current research. Int. J. Res. Ayurveda Pharm., 1: 234-238.
- Rasool, M. and P. Varalakshmi, 2006a. Immunomodulatory role of *Withania somnifera* root powder on experimental induced Inflammation: An *in vivo* and *In vitro* study. Vascular Pharmacol., 44: 406-410.
- Rasool, M. and P. Varalakshmi, 2006b. Suppressive effect of Withania somnifera on experimental gouty arthritis: An in vivo and in vitro study. Chem. Biol. Interact., 164: 174-180.
- Rasool, M. and P. Varalakshmi, 2007. Protective effect of Withania somnifera root powder in relation to lipid peroxidation, antioxidant status, glycoproteins and bone collagen on adjuvantinduced arthritis. Fundamental Clin. Pharmacol., 21: 157-164.
- Rastogi, R.P. and B.N. Mehrotra, 1998. Compendium of Indian Medicinal Plants. Vol. 6, Central Drug Research Institute, New Delhi, India.
- Raut, A.A., N.N. Rege, F.M. Tadvi, P.V. Solanki and K.R. Kene *et al.*, 2012. Exploratory study to evaluate tolerability, safety and activity of Ashwagandha (*Withania somnifera*) in healthy volunteers. J. Ayurveda Integrative Med., 3: 111-114.
- Salve, B.U., V.B. Chauhan, V.S. Kasture and S.B. Kasture, 2006. Effect of methanolic extract of Azadirachta indica leaves and Withania somnifera roots on some animal models of asthma. Indian J. Pharmacol., 38: 85-85.
- Sandhu, J.S., B. Shah, S. Shenoy, S. Chauhan, G.S. Lavekar and M.M. Padhi, 2010. Effects of Withania somnifera (Ashwagandha) and Terminalia arjuna (Arjuna) on physical performance and cardiorespiratory endurance in healthy young adults. Int. J. Ayurveda Res., 1: 144-149.
- Sangwan, R.S., N.D. Chaurasiya, L.N. Misra, P. Lal and G.C. Uniyal *et al.*, 2004. Phytochemical variability in commercial herbal products and preparations of *Withania somnifera* (Ashwagandha). Curr. Sci., 86: 461-465.
- Scarfiotti, C., F. Fabris, B. Cestaro and A. Giuliani, 1997. Free radicals, atherosclerosis, ageing and related dysmetabolic pathologies: Pathological and clinical aspects. Eur. J. Cancer Prevention, 6: S31-S36.

- Schliebs, R., A. Liebmann, S.K. Bhattacharya, A. Kumar, S. Ghosal and V. Bigl, 1997. Systemic administration of defined extracts from *Withania somnifera* (Indian Ginseng) and Shilajit differentially affects cholinergic but not glutamatergic and GABAergic markers in rat brain. Systemic administration of defined extracts from *Withania somnifera* (Indian Ginseng) and Shilajit differentially affects cholinergic but not glutamatergic and GABAergic markers in rat brain. Neurochem. Int., 30: 181-190.
- Senthil, V., S. Ramadevi, V. Venkatakrishnan, P. Giridharan, B.S. Lakshmi, R.A. Vishwakarma and A. Balakrishnan, 2007. Withanolide induces apoptosis in HL-60 leukemia cells via mitochondria mediated cytochrome c release and caspase activation. Chem. Biol. Interactions, 167: 19-30.
- Senthilnathan, P., R. Padmavathi, S.M. Banu and D. Sakthisekaran, 2006a. Enhancement of antitumor effect of paclitaxel in combination with immunomodulatory *Withania somnifera* on benzo(a)pyrene induced experimental lung cancer. Chem. Biol. Interactions, 159: 180-185.
- Senthilnathan, P., R. Padmavathi, V. Magesh and D. Sakthisekaran, 2006b. Stabilization of membrane bound enzyme profiles and lipid peroxidation by *Withania somnifera* along with paclitaxel on benzo(a)pyrene induced experimental lung cancer. Mol. Cell. Biochem., 292: 13-17.
- Sharada, A.C., E.F. Solomon and U.P. Devi, 1993. Toxicity of *Withania somnifera* root extract in rats and mice. Pharm. Biol., 31: 205-212.
- Sharada, M., A. Ahuja, K.A Suri, S.P. Vij, R.K. Khajuria, V. Verma and A. Kumar, 2007. Withanolide production by in vitro cultures of Withania somnifera and its association with differentiation. Bio. Planta, 51: 161-164.
- Sharma, S., S. Dahanukar and S.M. Karandikar, 1985. Effects of long-term administration of the roots of Ashwagandha and shatavari in rats. Indian Drugs, 29: 1339-1339.
- Sharma, L.K., B.R. Madina, P. Chaturvedi, R.S. Sanawan and R. Tuli, 2007. Molecular cloning and characterization of one member of 3 beta-hydroxy sterol glucosyltransferase gene family in *Withania somnifera*. Arch. Biochem. Biophys., 460: 48-55.
- Sharma, R., 2011. Toxicity evaluation of a novel anticancer formulation of *Withania somnifera* (L.) Dunal. Ph.D. Thesis, Department of Biotechnology, Guru Nanak Dev University, Amritsar, India.

- Shenoy, S., U. Chaskar, J.S. Sandhu and M.M. Paadhi, 2012. Effects of eight-week supplementation of Ashwagandha on cardiorespiratory endurance in elite Indian cyclists. J. Ayurveda Integr Med., 3: 209-214.
- Singh, N., S.P. Singh, R. Nath, D.R. Singh, M.L. Gupta, R.P. Kohli and K.P. Bhargava, 1986. Prevention or Urethane-induced lung adenomas by Withania somnifera (L.) Dunal in albino mice. Int. J. Crude Drug Res., 24: 99-100.
- Singh, S., K. Veena, S. Singhal and N. Singh, 2001. An Indian herbal immunomodulator: Highly effective in the treatment of HIV/AIDS. Proceedings of the 6th International Conference on AIDS Asia Pacific, October 5-10, 2001, Melbourne, Australia, pp. 5-10.
- Singh, A.K., R. Varshney, M. Sharma, S.S. Agarwal and K.C. Bansal, 2006. Regeneration of plants from alginate-encapsulated shoot tips of *Withania somnifera* (L.) Dunal, a medicinally important plant species. J. Plant Physiol., 163: 220-223.
- Singh, G., P.K. Sharma, R. Dudhe and S. Singh, 2010. Biological activities of *Withania somnifera*. Ann. Biol. Res., 1: 56-63.
- Singh, N., M. Bhalla, P. de Jager and M. Gilca, 2011. An overview on Ashwagandha: A rasayana (Rejuvenator) of ayurveda. Afr. J. Tradit Complement Altern Med., 8: 208-213.
- Sisodia, S.S. and M. Bhatnagar, 2004. Antiulcer activity of *Withania somnifera* in stress induced gastric ulcer model. Ind. J. Pharmacol., 36: 271-272.
- Sivanesan, I., 2007. Direct regeneration from apical bud explants of *Withania somnifera* Dunal. Indian J. Biotechnol., 16: 125-127.
- Sreerekha, M.V., K.V. Patel, R. Bhatnagar and S. Sriram, 2004. Distribution of total withanolides in various plant parts of Ashwagandha (*Withania somnifera*) accessions as influenced by light and dark reaction cycle. J. Med. Aromatic Plant Sci., 26: 681-683.
- Srinivasan, S., R.S. Ranga, R. Burikhanov, S.S. Han and D. Chendil, 2007. Par-4-dependent apoptosis by the dietary compound Withaferin A in prostate cancer cells. Cancer Res., 67: 246-253.
- Subbaraju, G.V., M. Vanisree, C.V. Rao, C. Sivaramakrishna, P. Sridhar, B. Jayaprakasam and M.G. Nair, 2006. Ashwagandhanolide, a bioactive dimeric thiowithanolide isolated from the roots of *Withania somnifera*. J. Nat. Prod., 69: 1790-1792.
- Sumantran, V.N., A. Kulkarni, S. Boddul, T. Chinchwade and S.J. Koppikar *et al.*, 2007a. Chondroprotective potential of root extracts of *Withania somnifera* in osteoarthritis. J. Biosci., 32: 299-307.

- Sumantran, V.N., S. Boddul, S.J. Koppikar, M. Dalvi, A. Wele, V. Gaire and U.V. Wagh, 2007b. Differential growth inhibitory effects of W. somnifera root and E. officinalis fruits on CHO cells. Phytother Res., 21: 496-499.
- Sundaram, S., P. Dwivedi and S. Purwar, 2011. In vitro evaluation of antibacterial activities of crude extracts of Withania somnifera (Ashwagandha) to bacterial pathogens. Asian J. Biotechnol., 3: 194-199.
- Supe, U., F. Dhote and M.G. Roymon, 2006. *In vitro* plant regeneration of *Withania somnifera*. Plant Tissue Culture Biotechnol., 16: 111-115.
- Teixeira, S.T., M.C. Valadares, S.A. Goncalves, A. de Melo and M.L.S. Queiroz, 2006. Prophylactic administration of Withania somnifera extract increases host resistance in Listeria monocytogenes infected mice. Int. Immunopharmacol., 6: 1535-1542.
- Titanji, V.P., A.A. Ngwa and M. Ngemenya, 2007. Applications of biotechnology techniques to the study of medicinal plants. Afr. J. Med. Sci., 36: 23-29.
- Tiwari, R. and S.D. Hirpurkar, 2011. Therapeutic potential of lytic phages against chronic wound infections. Indian Vet. J., 88: 1375-1377.
- Tiwari, R., K. Dhama, S. Chakraborty, A. Kumar, A. Rahal and S. Kapoor, 2013a. Bacteriophage therapy for safeguarding animal and human health: A review. Pak. J. Biol. Sci., (In Press).
- Tiwari, R., S. Chakraborty and K. Dhama, 2013b. Miracle of herbs in antibiotic resistant wounds and skin infections: Treasure of nature-a review/perspective. Pharma Sci. Monitor, 4: 214-248.
- Tiwari, R., S. Chakraborty, K. Dhama, M.Y. Wani, A. Kumar and S. Kapoor, 2013c. Wonder world of phages: Potential biocontrol agents safeguarding biosphere and health of animals and humans-current scenario and perspectives. Pak. J. Biol. Sci., (In Press)
- Tiwari, R., S. Chakraborty, K. Dhama, S. Rajagunalan and S.V. Singh, 2013d. Antibiotic resistance-an emerging health problem: Causes worries challenges and solutions-a review. Int. J. Curr. Res., 5: 1880-1892.
- Tomi, H., M. Yoshida and K. Kishida, 2005. Withania somnifera Dunal extracts for increasing male sperm count. Nippon Shinyaku Co., Ltd., USA.
- Uma, D.P., R. Kamath and B.S. Rao, 2000. Radiosensitization of a mouse melanoma by Withaferin A: *In vivo* studies. Ind. J. Exp. Biol., 38: 432-437.

- Venkataraghavan, S., C. Seshadri, T.P. Sundaresan, R. Revathi, V. Rajagopalan and K. Janaki, 1980. The comparative effect of milk fortified with Aswagandha, Aswagandha and Punarnava in children-a double-blind study. J. Res. Ayur Sid., 1: 370-385.
- Verma, S.K. and A. Kumar, 2011. Therapeutic uses of *Withania somnifera* (ashwagandha) with a note on withanolides and its pharmacological actions. Asian J. Pharm. Clin. Res., 4: 1-4.
- Verma, S.K., A. Shaban, R. Purohit, M.L. Chimata, G. Rai and O.P. Verma, 2012. Immunomodulatory activity of *Withania somnifera* (L.). J. Chem. Pharm. Res., 4: 559-561.
- Visavadiya, N.P. and A.V. Narasimhacharya, 2007. Hypocholesteremic and antioxidant effects of *Withania somnifera* (Dunal) in hypercholesteremic rats. Phytomedicine, 14: 136-142.
- Wadegaonkar, P.A., K.A. Bhagwat and M.K. Rai, 2006. Direct rhizogenesis and establishment of fast growing normal root organ culture of *Withania somnifera* dunal. Plant Cell. Tissue Organ. Culture, 84: 223-225.
- Wang, H.C., Y.L. Tsai, Y.C. Wu, F.R. Chang, M.H. Liu, W.Y. Chen and C.C. Wu, 2012. Withanolides-induced breast cancer cell death is correlated with their ability to inhibit heat protein 90. PLoS One, Vol. 7. 10.1371/journal.pone.0037764
- Widodo, N., N. Shah, D. Priyandoko, T. Ishii, S.C. Kaul and R. Wadhwa, 2009. Declaration of senescence in normal human fibroblasts by withanone extracted from ashwagandha leaves. J. Gerontol. A Biol. Sci. Med. Sci., 64: 1031-1038.
- Widodo, N., D. Priyandoko, N. Shah, R. Wadhwa and S.C. Kaul, 2010. Selective killing of cancer cells by Ashwagandha leaf extract and its component Withanone involves ROS signaling. PLoS One, Vol. 5. 10.1371/journal.pone.0013536
- Winters, M., 2006. Ancient medicine, modern use: *Withania somnifera* and its potential role in integrative oncology. Altern. Med. Rev., 11: 269-277.
- Yang, H., G. Shi and Q.P. Dou, 2007. The tumor proteasome is a primary target for the natural anticancer compound withaferin a isolated from Indian Winter Cherry. Mol. Pharmacol., 71: 426-437.