

# Assessing transmissibility of SARS-CoV-2 lineage B.1.1.7 in England

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The SARS-CoV-2 lineage B.1.1.7, designated variant of concern (VOC) 202012/01 by Public Health England<sup>1</sup>, was first identified in the UK in late summer to early autumn 2020<sup>2</sup>. Whole-genome SARS-CoV-2 sequence data collected from community-based diagnostic testing for COVID-19 show an extremely rapid expansion of the B.1.1.7 lineage during autumn 2020, suggesting that it has a selective advantage. Here we show that changes in VOC frequency inferred from genetic data correspond closely to changes inferred by *S* gene target failures (SGTF) in community-based diagnostic PCR testing. Analysis of trends in SGTF and non-SGTF case numbers in local areas across England shows that B.1.1.7 has higher transmissibility than non-VOC lineages, even if it has a different latent period or generation time. The SGTF data indicate a transient shift in the age composition of reported cases, with cases of B.1.1.7 including a larger share of under 20-year-olds than non-VOC cases. We estimated time-varying reproduction numbers for B.1.1.7 and co-circulating lineages using SGTF and genomic data. The best-supported models did not indicate a substantial difference in VOC transmissibility among different age groups, but all analyses agreed that B.1.1.7 has a substantial transmission advantage over other lineages, with a 50% to 100% higher reproduction number.

The SARS-CoV-2 lineage B.1.1.7 spread rapidly across England between November 2020 and January 2021. This variant possesses a large number of non-synonymous substitutions of immunological importance<sup>2</sup>. The N501Y replacement on the spike protein has been shown to increase ACE2 binding<sup>3,4</sup> and cell infectivity in animal models<sup>5</sup>, and the P618H replacement on the spike protein adjoins the furin-cleavage site<sup>6</sup>. B.1.1.7 also possesses a deletion at positions 69 and 70 of the spike protein ( $\Delta 69-70$ ) that has been associated with failure of diagnostic tests using the ThermoFisher TaqPath probe, which targets the spike protein<sup>7</sup>. Although other variants with  $\Delta 69-70$  are also circulating in the UK, the absence of detection of the *S* gene target in an otherwise positive PCR test appears to be a highly specific biomarker for the B.1.1.7 lineage. Data from national community testing in November 2020 showed a rapid increase in SGTF during PCR testing for SARS-CoV-2, coinciding with a rapid increase in the frequency of B.1.1.7 observed in genomic surveillance. The B.1.1.7 lineage was designated VOC 202012/01 by Public Health England (PHE) in December 2020.

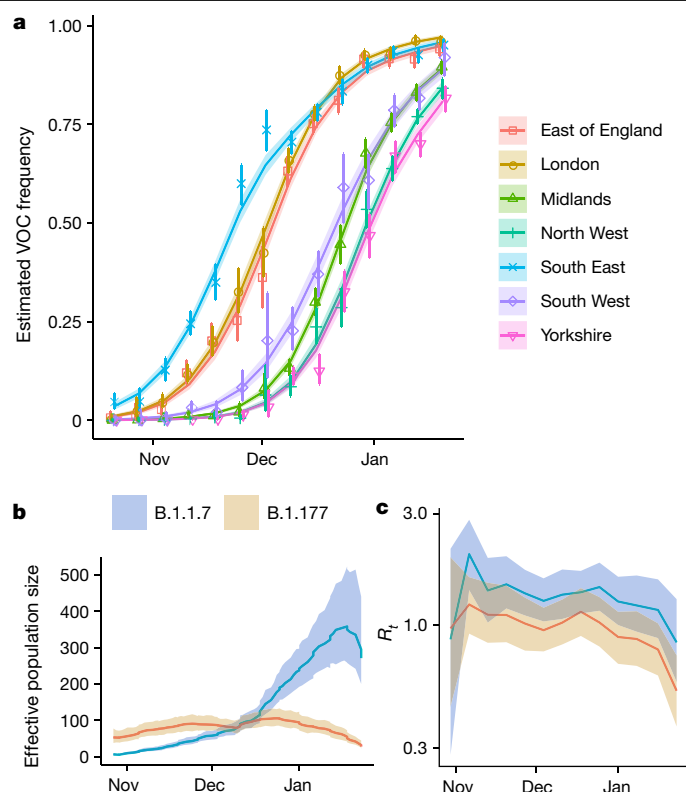
Phylogenetic studies carried out by the UK COVID-19 Genomics Consortium (COG-UK) (<https://www.cogconsortium.uk>)<sup>8</sup> provided the first indication that B.1.1.7 has an unusual accumulation of substitutions and was growing at a higher rate than other circulating lineages. We investigated time trends in the frequency of sampling VOC genomes and the proportion of PCR tests exhibiting SGTF across the UK, which we calibrated as a biomarker of VOC infection. Using multiple approaches and both genetic and SGTF data, we conclude that B.1.1.7 is associated with a higher reproduction number (*R*) than previous non-VOC lineages.

We examined whole-genome SARS-CoV-2 sequences from randomly sampled residual materials obtained from community-based COVID-19 testing in England, collected between 1 October 2020 and 16 January 2021. These data included 31,390 B.1.1.7 sequences for which the time and location of sample collection were known. Over the same period, 52,795 non-VOC genomes were collected. VOC sequences were initially concentrated in London (*n* = 9,134), the South East (*n* = 5,609), and the East of England (*n* = 4,413), but is now widely distributed across England.

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**Fig. 1 | Expansion of lineage B.1.1.7 relative to co-circulating lineages in England.** **a**, Estimated frequency of sampling the VOC (lines) over time in NHS regions ( $n = 84,185$ ). Shaded regions, 95% credible region based on Bayesian regression; points, empirical proportions of the VOC in each week; error bars, 95% CI based on binomial sampling error. **b**, Effective population size over time for lineage B.1.1.7 and estimates based on a matched sample of the most abundant co-circulating lineage, B.1.177 ( $n = 3,000$ ). Shaded regions, 95% bootstrap CI. **c**, The effective reproduction number inferred from growth of effective population size for both lineages in **b**.

Overall, we estimate the median posterior additive difference in growth rates between B.1.1.7 and co-circulating variants to be 0.69 per week (95% credible interval (CrI) 0.61–0.76) (Fig. 1a, Extended Data Fig. 1, Supplementary Methods section 2), and this difference was largest in November. However, in tandem with geographic expansion of the VOC and imposition of lockdown measures in 2021, this difference declined gradually to 0.43 per week (95% CrI 0.33–0.52) for the week ending 16 January.

The rate of genetic diversification of the VOC lineage over time allows epidemic growth rates to be estimated using phylodynamic modelling<sup>9,10</sup>. To contrast VOC and non-VOC growth patterns, we randomly sampled 3,000 VOC sequences paired with up to 3,000 non-VOC sequences and matched by week of sample collection and location (Supplementary Methods section 1). Phylodynamic modelling (Supplementary Methods section 3) of the effective population sizes of B.1.1.7 and the previously dominant non-VOC B.1.177 lineage<sup>11</sup> gave an estimated growth rate difference of 0.33 per week (95% confidence interval (CI) 0.09–0.62), and further indicated that the VOC overtook the B.1.177 lineage on 10 December (Fig. 1b), close to the date at which VOC sampling frequency exceeded 50% in England (3 December). Thus, we estimate that B.1.1.7 reached 50% frequency within 2.5 to 3 months after its emergence in England.

We estimated the ratio of VOC to non-VOC reproduction numbers using a renewal equation based approach (Fig. 1c, Extended Data Fig. 2, Supplementary Methods section 4). This estimator depends on the absolute growth rate of the non-VOC, estimated using the phylodynamic model. We estimate the ratio of reproduction numbers between

25 October 2020 and 16 January 2021 to be 1.89 (95% CrI 1.43–2.65), assuming a gamma-distributed generation time with mean 6.4 days and coefficient of variation of 0.66<sup>12</sup>. This ratio is sensitive to the assumption that the generation time distribution is identical between variants. However, even if the VOC generation time is half that of previous variants, the estimated ratio of reproduction numbers was still 1.53 (95% CrI 1.27–1.79). The ratio trended downwards over time, coinciding with the increasing frequency of the VOC. By mid-January, the ratio had fallen from 1.89 to 1.54 (95% CrI 1.34–1.82) (Extended Data Fig. 2).

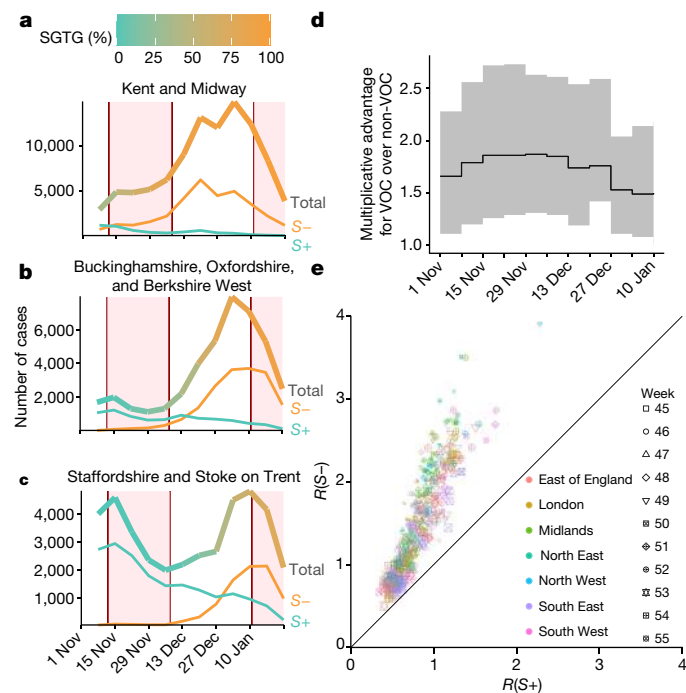
## Trends in SGTF attributed to the VOC

Infection with the VOC lineage results in a diagnostic failure on the S gene target in an otherwise positive PCR test using the ThermoFisher TaqPath assay, which is widely used for SARS-CoV-2 community PCR testing in the UK. Consequently, we gained a more detailed picture of the spatial and demographic spread of B.1.1.7 by using the much more abundant diagnostic data with SGTF than by using whole-genome sequencing only. Several SARS-CoV-2 variants can result in SGTF, but since mid-November 2020, more than 97% of PCR tests with SGTF were due to the B.1.1.7 lineage<sup>1</sup>. Approximately 35% of positive test results in UK community PCR testing use the TaqPath assay, and so provide S gene target results. Before mid-November 2020, SGTF frequency among PCR positives was a poor proxy for VOC frequency. We therefore developed a spatiotemporal model to predict the proportion of SGTF cases attributable to the VOC by area and week (Supplementary Methods section 5), here termed the true positive proportion (TPP). False positives were attributed to the S-gene-positive case (S+) category. We found that the effective population size for B.1.1.7 effective population size was highly correlated with TPP-adjusted S counts (Extended Data Fig. 3).

Figure 2a–c (and Supplementary Data 1, Extended Data Fig. 4) shows the spatiotemporal trends of SGTF cases (S–), S+ and total PCR-positive cases by National Health Service (NHS) England Sustainability and Transformation Plan (STP) areas (a geographical subdivision of NHS regions). Visually, it is clear that during the second England lockdown, when schools were open, S+ case numbers decreased but S– case numbers increased. However, during the third lockdown, when schools were closed, the incidence of both S– and S+ cases declined.

Using TPP-corrected SGTF frequencies applied to overall PHE case numbers, we jointly estimated weekly effective reproduction numbers ( $R_t$ ) values for the VOC and non-VOC in each of the 42 STP areas using a semi-mechanistic epidemiological model<sup>13</sup> (Supplementary Methods section 6). The model parametrizes VOC  $R_t$  as a multiple of non-VOC  $R_t$ . The model was fitted to case numbers obtained by multiplying overall PHE case numbers by TPP-corrected SGTF frequencies. We estimated  $R_t$  for epidemiological weeks 45–55 (1 November 2020 to 16 January 2021) (Fig. 2d), as before November there were insufficient VOC cases to reliably estimate VOC reproduction numbers across England. VOC  $R_t$  was greater than non-VOC  $R_t$  for all STP-week pairs (points above the diagonal in Fig. 2e). The estimated mean ratio of  $R_t$  for the VOC and non-VOC strains was 1.79 (95% CI 1.22–2.49) over weeks 45–55. As in the phylodynamic analysis, the multiplicative advantage in  $R_t$  for the VOC declined over the time window examined, to approximately 1.5 in week 55 (Fig. 2d).

The greater  $R_t$  estimates of the VOC, even where  $R_t$  of non-VOC variants was below 1, indicates that B.1.1.7 has a transmission advantage, and that the observed frequency trends cannot be explained solely by a reduction in the mean generation time. We repeated the joint estimation of VOC and non-VOC  $R_t$  with the assumption of a 25% reduction in the mean generation time of the VOC (Extended Data Fig. 5), and this estimated the mean ratio of  $R_t$  to be 1.60 (95% CI 1.09–2.23) over weeks 45–55. Incorporating a shorter generation time for the VOC into the model reduced, but did not eliminate, the decreasing trend in transmission advantage over time.

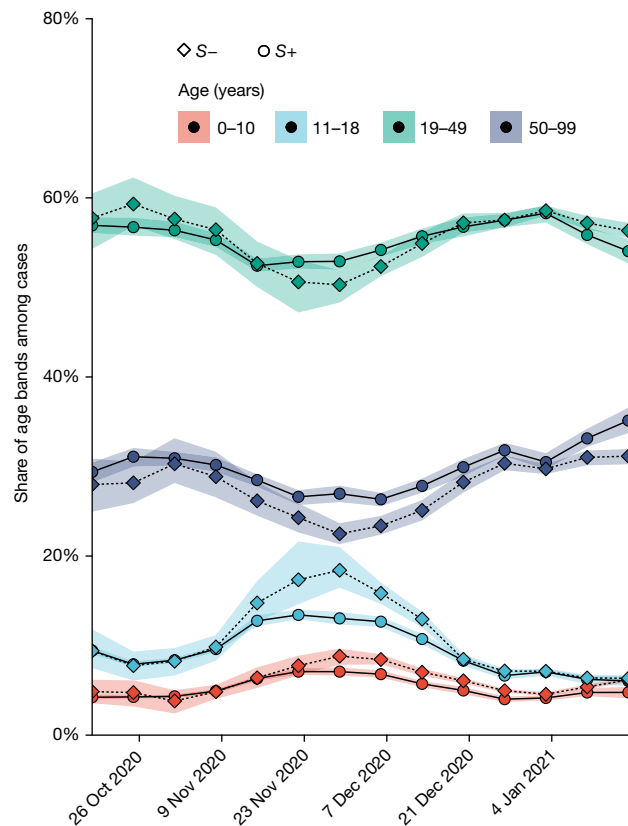


**Fig. 2 | Trends of diagnosed cases and SGTF over time and between regions, and reproduction numbers of the VOC inferred from SGTF.** **a–c**, The number of diagnosed cases over time for three English STP regions that represent a wide spectrum of outcomes in terms of time of VOC introduction into the region. Each line segment is shaded with the frequency of SGTF in each week (scale at top). Vertical shaded regions represent the times of the second and third UK lockdowns. **d**, The estimated (Bayesian posterior) multiplicative transmission advantage of the VOC over time inferred from STP-level SGTF count data. Shaded regions, 95% CrI. **e**, The reproduction number of S-gene-negative cases versus the reproduction number of S-gene-positive cases over time and among STP regions for epidemiological weeks 45–55 (1 November 2020 to 16 January 2021).

To test whether VOC transmissibility differed by age, we first examined the age distributions of S+ and S– cases. Case numbers were age-standardized at STP area level, and then case age distributions were calculated for each STP-week (Supplementary Methods section 7). Figure 3 shows that individuals aged 19–49 years were the only age group that was consistently over-represented among observed cases relative to their share in the population (40%), with little difference between VOC and non-VOC cases. Secondary school-aged children (11–18 years) were also over-represented among observed cases relative to their share in the population (9%), and the difference between VOC and non-VOC cases was statistically significant for three weeks in November (Fig. 3, Extended Data Fig. 6). This period coincides with the second England lockdown (5 November to 2 December 2020) when schools remained open, and the differing age distributions between variants could arise from altered contact patterns when children were at greater risk of infection from all variants compared to adults.

Next, we formulated models that incorporated a difference in VOC transmission between age groups (Supplementary Methods section 7). The models were fitted variously to genome-derived and/or SGTF-derived VOC frequencies, as well as total age-specific cases in each week and region, and compared using Bayesian leave-one-out cross-validation.

Model comparison consistently favoured models that allowed the transmission advantage to vary over time and between regions, using either genomic or SGTF data. However, models that incorporated an age effect were not significantly favoured (Extended Data Table 1). Indeed, the observed fluctuations in the age distribution are equally well captured by models that do not incorporate age-specific transmission advantages (Extended Data Fig. 6). We also used these model



**Fig. 3 | Age distribution of S-gene-positive and -negative cases over time in England.** Observed cases were age-standardized at the level of the STP area, and age distributions were calculated for each week in STP areas and then aggregated. Shaded regions, CIs computed by bootstrapping over STP areas within NHS regions for each week.

comparisons to test the hypothesis that differences in the VOC growth rates are a consequence of a reduced generation time in B.1.1.7. In principle it is possible to statistically identify such a difference, because the data cover a period during which the overall  $R_t$  has been above and below one. Models that incorporate a change in the mean generation time were sometimes favoured (Extended Data Table 1), but the estimated ratio of mean generation times was not well identified—it varied between 0.75 and 0.96, depending on the model and data being fitted to. The mean ratio of  $R_t$  between the VOC and non-VOC ranged between 1.6 and 2.01, depending on model variant. The best fit model to both SGTF and genomic data gave an estimate of 1.74 (95% CrI 1.03–2.75), which is highly consistent with the estimates obtained from the phylodynamic analysis and the direct estimation of  $R_t$  for VOC and non-VOC described above. This model also reproduces the decline in transmission advantage over time seen in our other analyses (Extended Data Fig. 7).

## Discussion

While substitutions in the B.1.1.7 lineage are associated with substantial changes in viral phenotype<sup>3–5,14</sup>, the extent to which these substitutions lead to meaningful differences in transmission between humans is unclear, and cannot be evaluated experimentally. When randomized experimental studies are not possible, observational studies provide strong evidence if consistent patterns are seen in multiple locations and at multiple times. Increasing frequency of a new lineage is consistent with a selective advantage, but changes in frequency result from founder effects, especially for genetic variants that are repeatedly introduced from overseas<sup>11,15</sup>. However, in contrast to previous variants that have achieved high prevalence, we see expansion of the VOC from within the UK.

We find some evidence that the multiplicative transmission advantage of B.1.1.7 (that is, ratio of reproduction numbers) declined in late December 2020 to January 2021, coincident with stricter social distancing, school closures, and the subsequent third England lockdown (Fig. 2d, Extended Data Figs. 2, 6). A number of mechanisms could generate this effect. First, a shorter generation time of the VOC would reduce the ratio of VOC to non-VOC growth rates for small values of the non-VOC growth rate. Thus as interventions reduce both reproduction numbers, their ratio would decline, even in the absence of any underlying change in transmission advantage. Some weak support for this hypothesis is provided by our age-specific model fits to SGTF data, where model comparison generally favours models that include a change in mean generation time (Extended Data Table 1). Second, social distancing changes human contact networks, reducing the number of people contacted per day, but increasing the duration and proximity of remaining (mostly household) contacts. In such circumstances, saturation of transmission probabilities can lead to a reduction in the transmission advantage of the VOC (Extended Data Fig. 8, Supplementary Methods section 7). The observation that secondary attack rates in contacts identified through routine national contact tracing were 30–40% higher for the VOC than for non-VOC cases<sup>16</sup> provides some support for this hypothesis, given that the large majority of contacts identified through the UK Test and Trace system are household contacts.

The data included in this study were collected as part of routine surveillance of community testing and are not representative of SARS-CoV-2 infections in England. However, previous comparisons of community case data to random household prevalence surveys have shown very strong agreement in epidemic trends<sup>17,18</sup>. Furthermore, estimates of the growth advantage of B.1.1.7 obtained during earlier iterations of this study<sup>1</sup> have largely been predictive of its subsequent spread in January, both in the UK and internationally. Independent observations of secondary attack rates inferred from UK contact tracing data have confirmed these findings<sup>19</sup>.

The substantial transmission advantage that we and others<sup>20,21</sup> have estimated has increased the challenges in controlling COVID-19. The B.1.1.7 lineage was identified quickly owing to extensive genomic surveillance in the UK, but other lineages with similar concerning features<sup>22,23</sup> have emerged almost concurrently, and lineages with similar features may be circulating undetected. Improving global genomic surveillance will be important for the control of COVID-19 in the presence of multiple emerging lineages with enhanced transmission or potential for immune escape.

## Online content

Any methods, additional references, Nature Research reporting summaries, source data, extended data, supplementary information, acknowledgements, peer review information; details of author contributions and competing interests; and statements of data and code availability are available at <https://doi.org/10.1038/s41586-021-03470-x>.

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# Article

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# Article

## Reporting summary

Further information on research design is available in the Nature Research Reporting Summary linked to this paper.

## Data availability

All data used in this study, including SGTF and genome counts aggregated by region and week and multiple sequence alignments, have been deposited in Zenodo at <https://doi.org/10.5281/zenodo.4593885>.

## Code availability

The Zenodo repository <https://doi.org/10.5281/zenodo.4593885> includes code to reproduce all figures and results presented here.

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**Competing interests** The authors declare no competing interests.

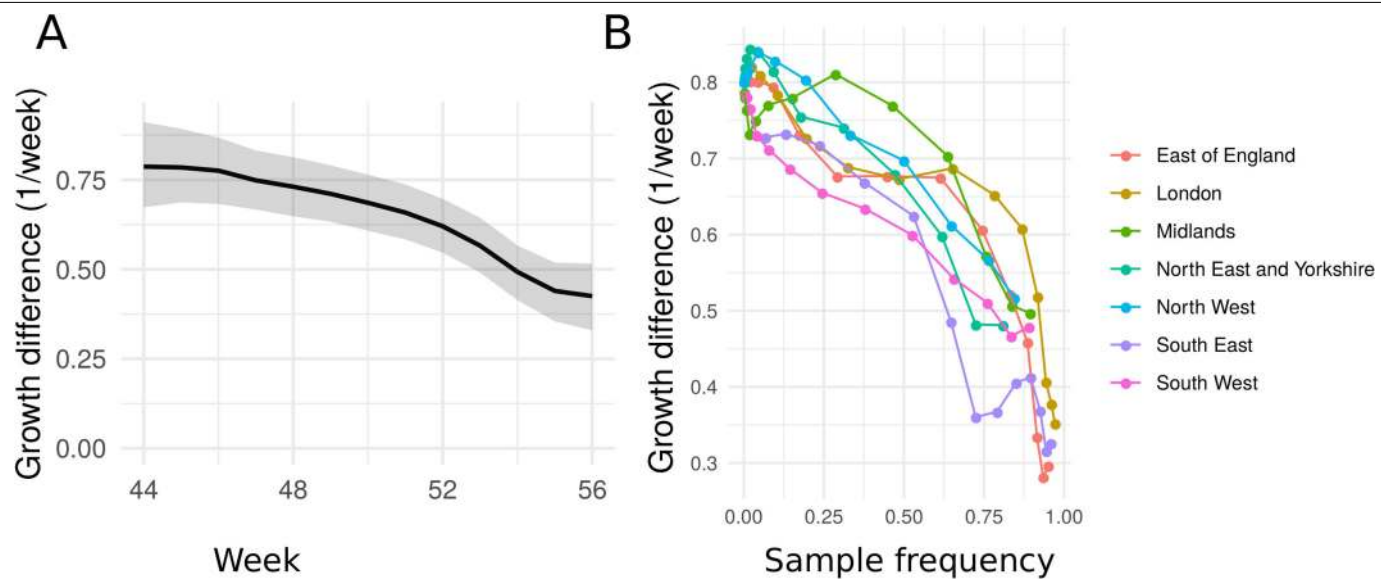
## Additional information

**Supplementary information** The online version contains supplementary material available at <https://doi.org/10.1038/s41586-021-03470-x>.

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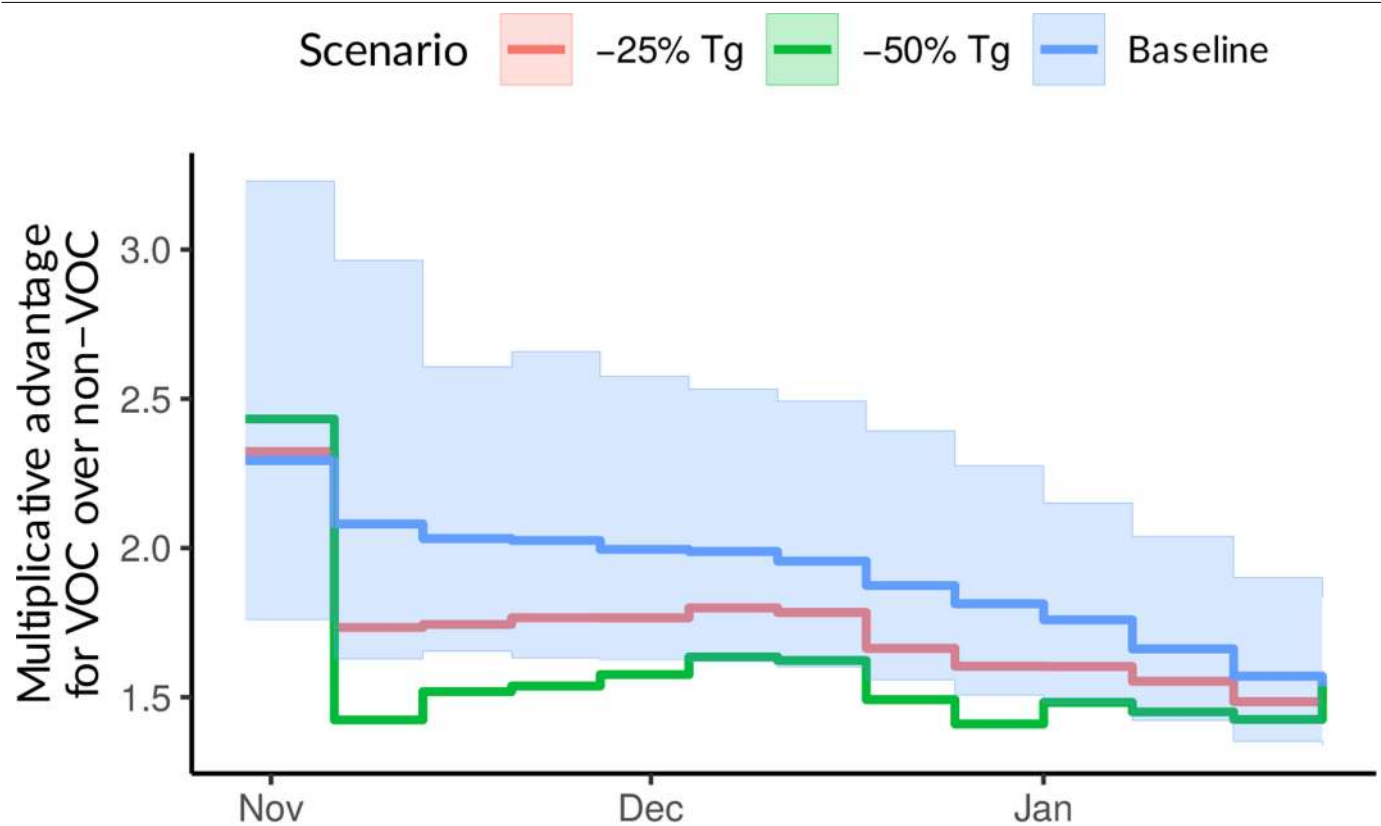
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**Extended Data Fig. 1 | Weekly growth rate of the VOC relative to other variants and relationship of VOC growth rate with VOC frequency. a,** The additive difference in growth rate between VOC and other lineages inferred from observed frequency of VOC genomes over time (Supplementary Methods

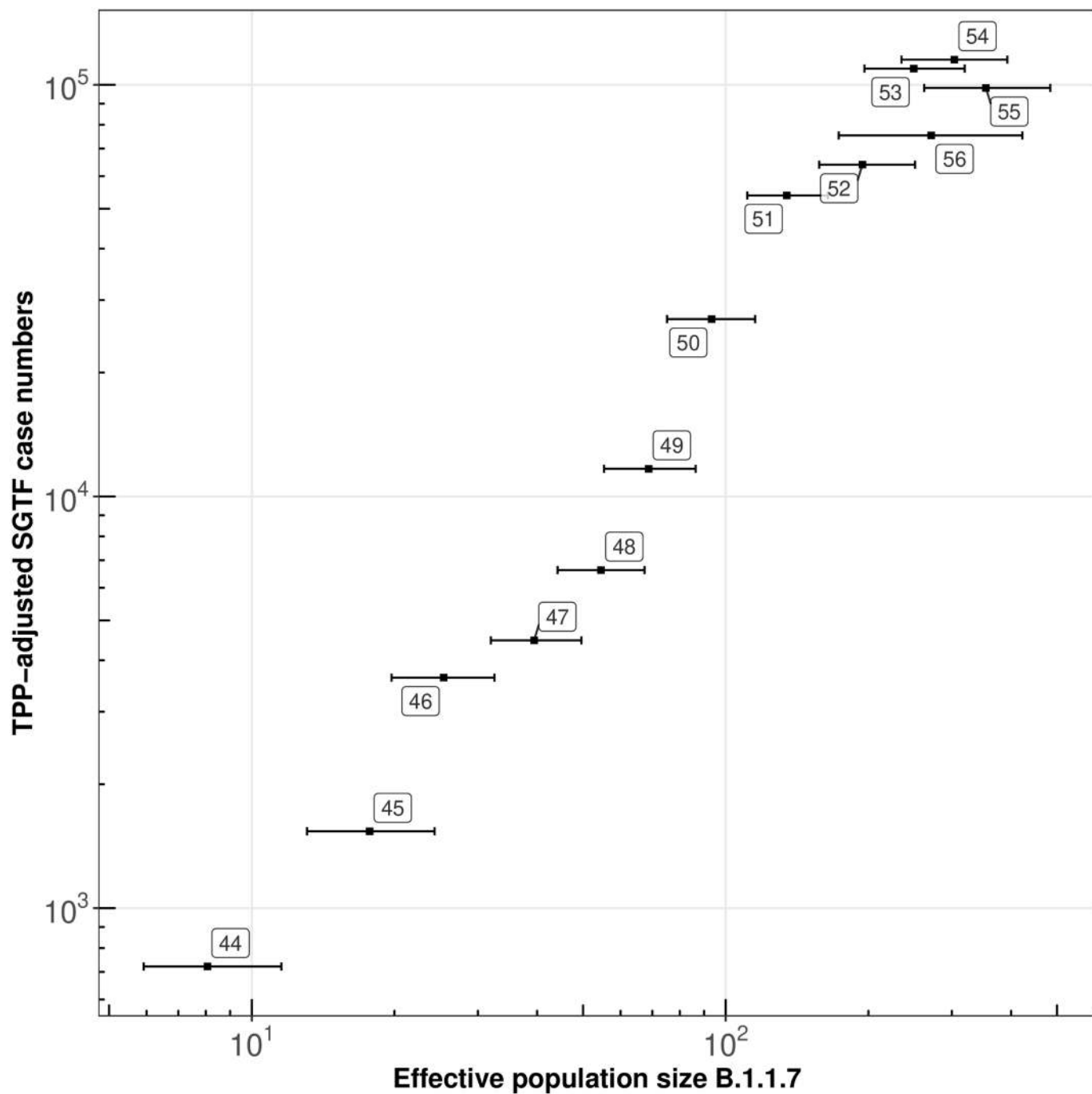
section 2). **b,** The additive difference in growth rates plotted against estimated frequency of the VOC for different NHS regions. The difference in growth rate correlates more strongly with VOC frequency than with time (data not shown). Estimates are presented for weeks 44–56 for each region.





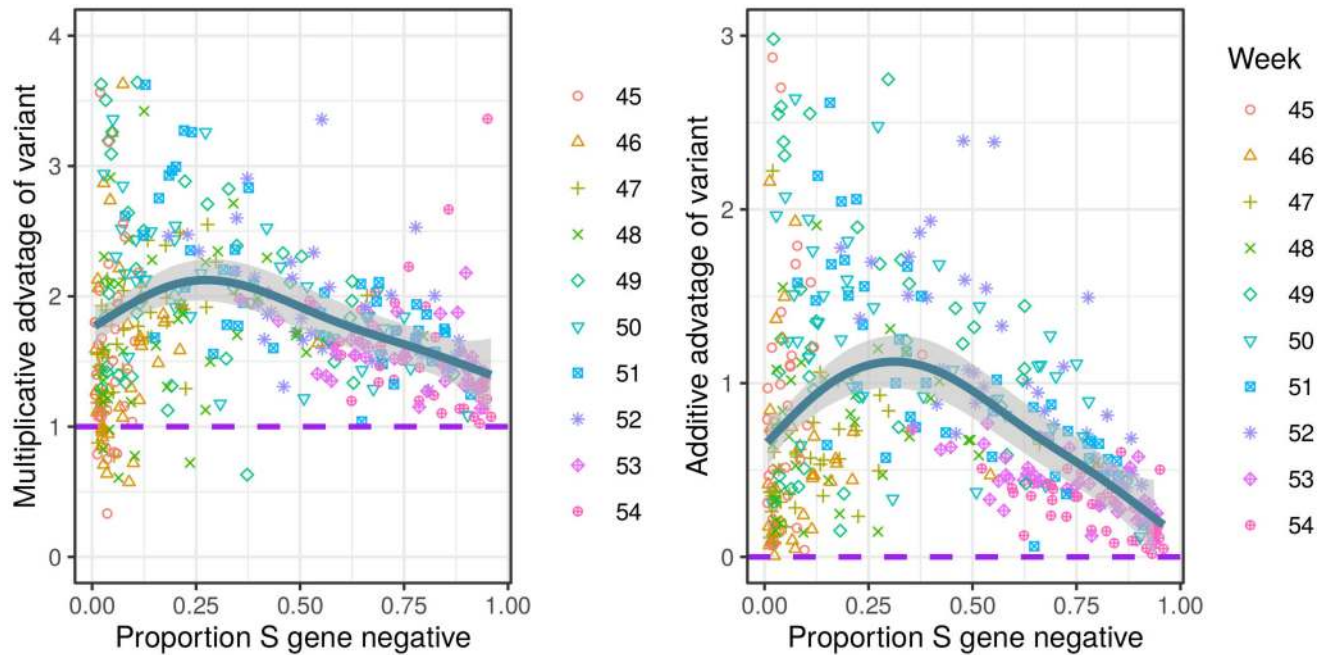
**Extended Data Fig. 2 | The ratio of the reproduction number for the VOC to that for co-circulating lineages inferred from combining estimating frequencies and phylodynamic estimates of VOC and non-VOC growth**

**rates.** Shaded region, 95% CrI. Sensitivity of estimates to differences in the mean generation time is shown in red and green (25% and 50% reduced generation time in VOC, respectively).



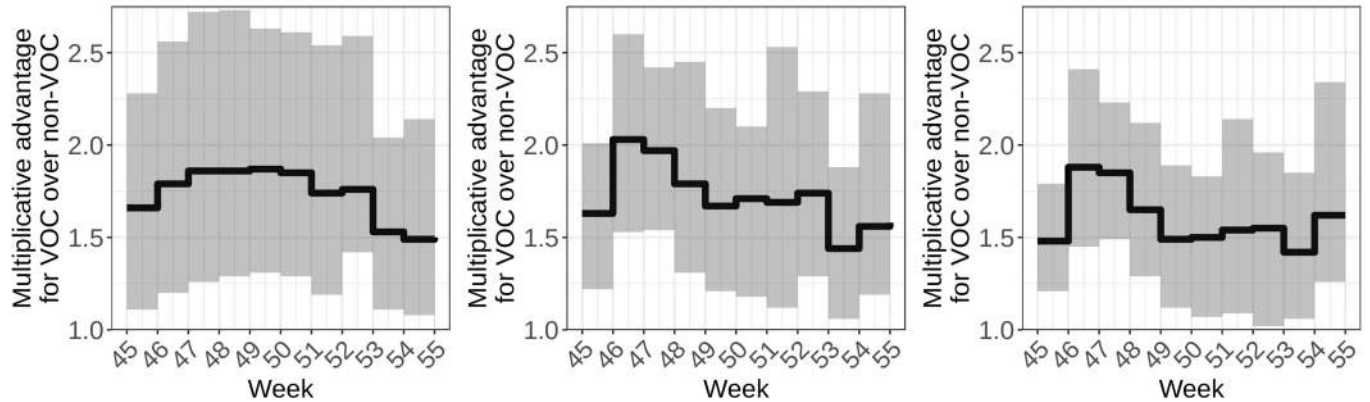
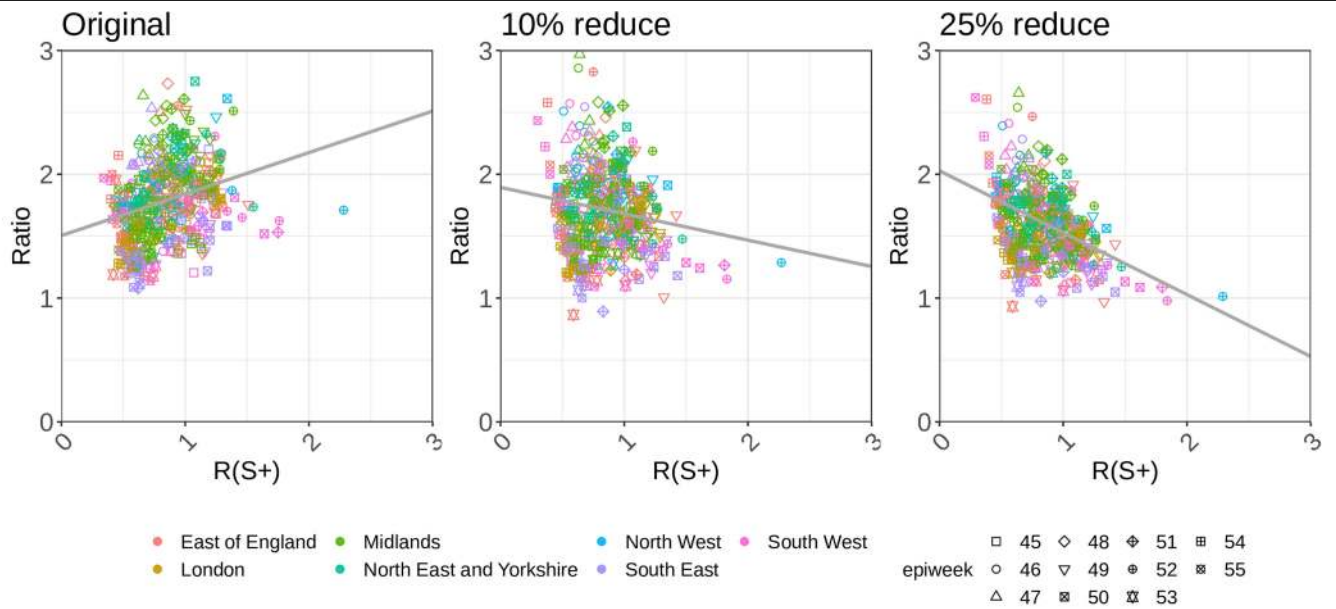
**Extended Data Fig. 3 | Weekly numbers of S-gene-positive samples with TPP correction plotted against the effective population size of the VOC.** Point labels indicate week of data collection. Effective sample size is taken on

the final day of the corresponding epidemiological week. Error bars, 95% bootstrap CIs.



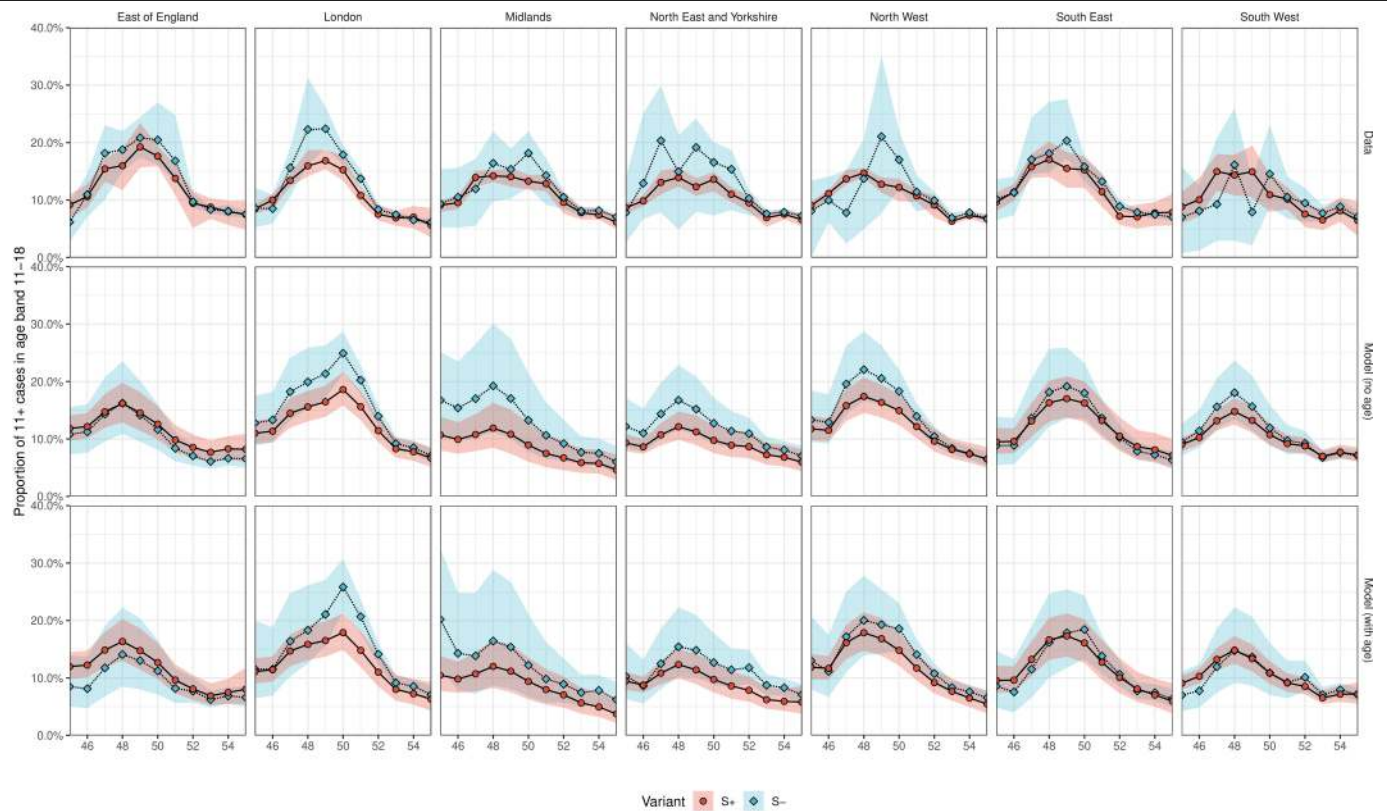
**Extended Data Fig. 4 | Empirical data analysis of the advantage in weekly growth factors (cases in week  $t+1$  divided by cases in week  $t$ ) for the VOC and non-VOC lineages (Supplementary Methods section 5).** Each point represents either the ratio (left) or difference (right) of weekly growth factors for the VOC versus the non-VOC for an NHS England STP area and week, using the raw SGTf data shown in Fig. 2 (not adjusting for TPP). Colours and shapes

differentiate epidemiological weeks. Numbers above 1 on the left plot and above 0 on the right plot show a transmission advantage. The blue line represents the mean advantage for a particular proportion of VOC among all cases, and the grey shading the 95% asymptotic CrI ( $\pm 2\sigma$ ). Scatter at low frequencies largely reflects statistical noise resulting from low counts.



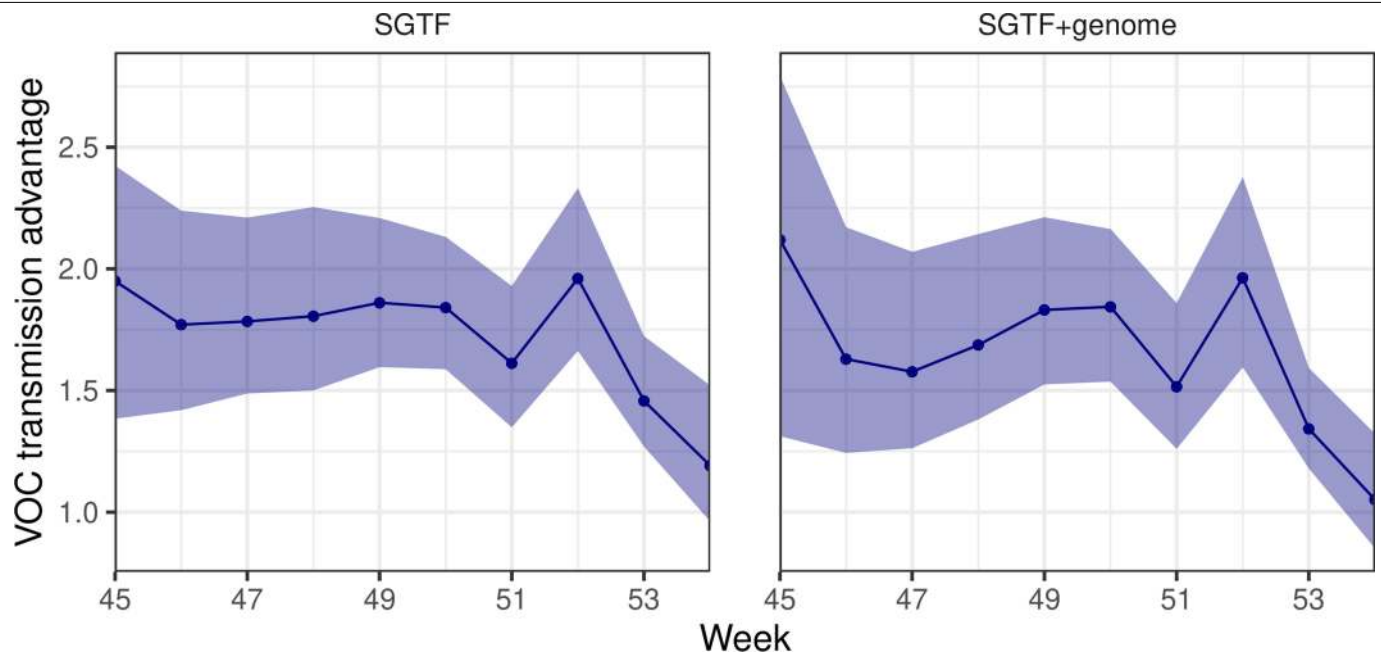
**Extended Data Fig. 5 | Sensitivity of reproduction number estimates at regional level to differences in the mean generation time in the VOC (0–25% reduction in the mean, while holding coefficient of variation constant).** Top, scatter plots of estimated ratio of S– to S+ reproduction numbers plotted against the reproduction number of S-gene-positive cases

over time and among STP regions. Grey lines, linear regression—if changes in the generation time were able to completely explain temporal variation in the ratio, we would expect the slope to be zero. Bottom, ratio of reproduction number over time for each assumption about the mean generation time of the VOC. Shaded region, 95% CrI.



**Extended Data Fig. 6 | Proportion of all cases in individuals over 10 years of age that were in 11-18-year-olds, stratified by SGTF status, NHS regions and week of testing.** Top, data; middle, predicted trends from a semi-mechanistic model (Supplementary Methods section 7) fitting a time- and region-varying

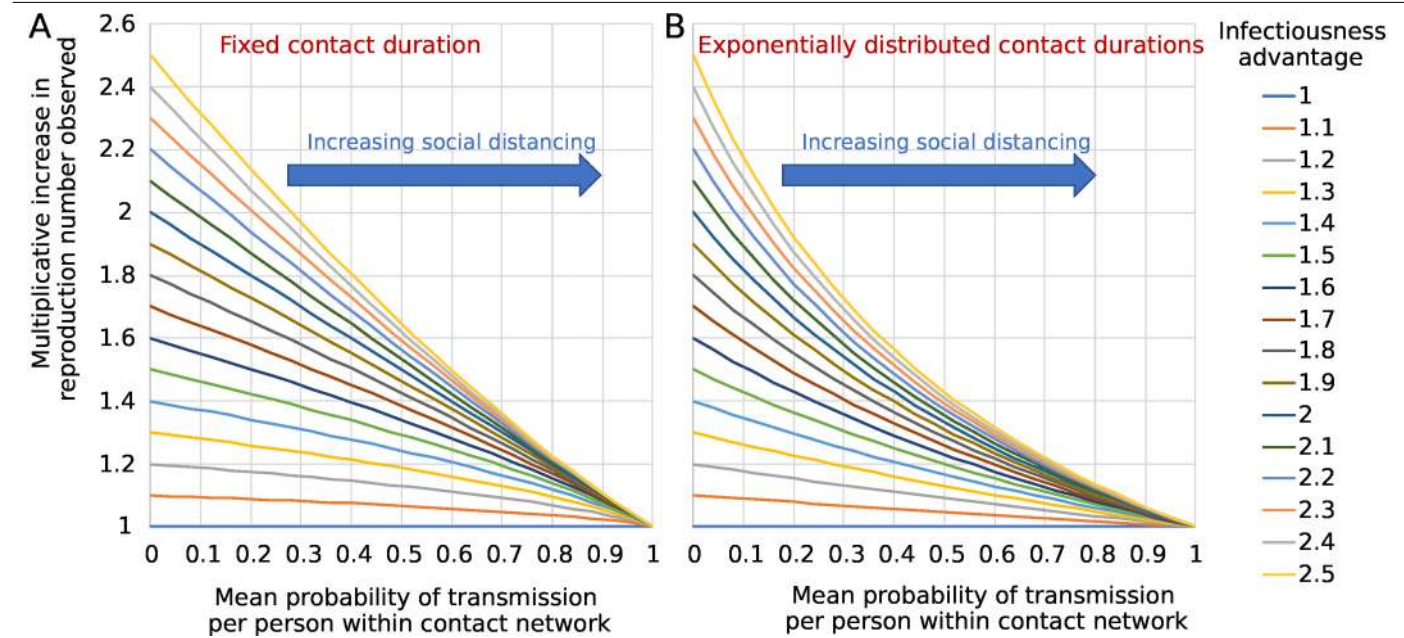
transmission advantage, but no age variation in transmission advantage; bottom, predicted trends from the model fitting a transmission that which varies by week, region and age group. Shaded region, 95% CrI.



**Extended Data Fig. 7 | The overall multiplicative transmission advantage of the VOC over time estimated using a semi-mechanistic model with a VOC transmission advantage that varies by week and region, but not with age (Supplementary Methods section 7). The model fitted one transmission**

advantage parameter per week and per NHS region to SGTF (STP-level) counts by NHS STP area (left; finer scale than region) and SGTF counts by STP area and VOC genome counts by NHS region (right). Solid lines, medians; shading, 95% CrI pooled over all regions.





**Extended Data Fig. 8 | Multiplicative increase in reproduction number seen for different intrinsic infectiousness advantages and probability of transmission per contact for the non-VOC. a, Fixed contact duration.**

**b, Exponentially distributed contact durations.** See Supplementary Methods section 7.

**Extended Data Table 1 | Tabulation of parameter estimates and model assessment using Pareto-smoothed importance sampling leave-one-out cross validation (PSIS-LOO CV) and widely applicable information criterion (WAIC)**

| Covariates in reproduction number ratio | Data            | Mean generation time ratio | Mean reproduction number ratio | LOO          | WAIC         |
|---|-----------------|----------------------------|--------------------------------|--------------|--------------|
| None                                    | SGTF            | 1 (fixed)                  | 1.92 (1.80,2.06)               | -9021        | -8850        |
| None                                    | SGTF            | 0.78 (0.72,0.85)           | 1.61 (1.48,1.74)               | -9035        | -8867        |
| Age                                     | SGTF            | 1 (fixed)                  | 1.94 (1.80,2.24)               | -9026        | -8855        |
| Age                                     | SGTF            | 0.77 (0.72,0.84)           | 1.60 (1.47,1.74)               | -9028        | -8862        |
| Time, region                            | SGTF            | 1 (fixed)                  | 1.76 (1.10,2.24)               | <b>-9013</b> | <b>-8850</b> |
| Time, region                            | SGTF            | 0.89 (0.82,0.98)           | 1.62 (1.12,2.04)               | -9019        | <b>-8860</b> |
| Age, time, region                       | SGTF            | 1 (fixed)                  | 1.81 (1.19,2.30)               | -9024        | <b>-8860</b> |
| Age, time, region                       | SGTF            | 0.77 (0.70,0.85)           | 1.55 (1.22,1.88)               | -9018        | -8856        |
| None                                    | SGTF and genome | 1 (fixed)                  | 1.86 (1.74,2.00)               | -9701        | -9523        |
| None                                    | SGTF and genome | 0.80 (0.73,0.87)           | 1.60 (1.48,1.75)               | -9703        | -9525        |
| Age                                     | SGTF and genome | 1 (fixed)                  | 1.85 (1.73,2.00)               | -9699        | -9517        |
| Age                                     | SGTF and genome | 0.79 (0.72,0.86)           | 1.59 (1.46,1.74)               | -9693        | -9518        |
| Time, region                            | SGTF and genome | 1 (fixed)                  | 1.66 (0.98,2.38)               | -9654        | <b>-9494</b> |
| Time, region                            | SGTF and genome | 1.00 (0.90,1.11)           | 1.66 (0.98,2.39)               | <b>-9650</b> | <b>-9489</b> |
| Age, time, region                       | SGTF and genome | 1 (fixed)                  | 1.69 (1.00,2.34)               | -9676        | -9514        |
| Age, time, region                       | SGTF and genome | 0.92 (0.82,1.02)           | 1.61 (1.02,2.19)               | -9684        | -9523        |
| None                                    | Genome          | 1                          | 1.74 (1.61,1.89)               | <b>-964</b>  | <b>-940</b>  |
| None                                    | Genome          | 0.89 (0.76,1.06)           | 1.65 (1.50,1.84)               | <b>-961</b>  | <b>-940</b>  |
| Time, region                            | Genome          | 1                          | 1.71 (1.26,2.20)               | <b>-929</b>  | <b>-905</b>  |
| Time, region                            | Genome          | 0.95 (0.82,1.11)           | 1.66 (1.25,2.16)               | <b>-938</b>  | <b>-915</b>  |

Bold indicates no significant difference in the estimated predictive accuracy among the best-performing models within each subset of data (SGTF counts, SGTF and genome counts, genome counts).

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*Our web collection on [statistics for biologists](#) contains articles on many of the points above.*

### Software and code

Policy information about [availability of computer code](#)

#### Data collection

Genomic analyses We examined whole genome SARS-CoV-2 sequences from randomly sampled residual materials obtained from community-based COVID-19 testing in England. Sequence alignments and phylogenetic trees are available for download in the code repository. Sample weights were used to select sequences for phylodynamic modelling and to adjust counts when estimating the frequency of the variant of concern B.1.1.7 (VOC, hereafter). Weights were assigned to sequenced samples according to their local authority and their collection date. Each weight was proportional to the ratio of diagnosed cases to the number of sequence samples (reciprocal sequencing coverage rate) in each week and local authority. Case counts and sequence counts were summed over a fourteen day moving window in order to smooth over small case counts and sequence counts. Confirmed cases used for weighting were obtained using the UK government COVID-19 dashboard API (<https://api.coronavirus.data.gov.uk>). Code to compute sequence sample weights is available in the sequencing\_coverage package (v1.0) <https://git.io/Jqcve>. SGTF analyses Data on SGTF among national community testing was obtained from the 3 largest PCR testing laboratories and integrated into the PHE Second Generation Surveillance System (SGSS) database. We also obtained the linelist of "pillar 2" (community) PCR-positive cases from Public Health England and linked this to the SGTF data. Application of SGTF as a diagnostic for the VOC provides a large advantage over genomic sequencing in terms of cost, speed, and the sample size of available test results. We extracted 585,165 S target positive (S+) and 548,649 S target negative (S-) test results collected for weeks 43 to 56 and examined the potential to use SGTF cases (S-) as a biomarker for the VOC lineage. While the tests are not a representative sample of infections over this time period, they are a representative sample of tests within a given region and week and thus provide information about the relative abundance of the VOC versus other variants over time and between regions.

#### Data analysis

A large variety of open source R packages were used and described in the supporting information including version numbers. The exact code is provided in a Zenodo repository enabling reproducibility of all results and figures. Phylogenetic analysis made use of BEAST 1.10.4. Citations are provided for all packages.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research [guidelines for submitting code & software](#) for further information.

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All data used in this study including SGTF and genome counts aggregated by region and week and multiple sequence alignments have been deposited in Zenodo <https://doi.org/10.5281/zenodo.4593885>

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## Ecological, evolutionary & environmental sciences study design

All studies must disclose on these points even when the disclosure is negative.

|                                   |  |
|-----------------------------------|--|
| Study description                 | Genomic epidemiological analysis of the spread of a SARS-CoV-2 variant   |
| Research sample                   | SARS CoV 2 Genomic data compiled by COG-UK consortium; community case data provided by Public Health England.  |
| Sampling strategy                 | All genetic sequences and case records were utilized   |
| Data collection                   | N/A  |
| Timing and spatial scale          | All of England October 2020-January 2021   |
| Data exclusions                   | No data were excluded  |
| Reproducibility                   | The analysis is completely reproducible using code and data in the given repository  |
| Randomization                     | The study involved observational data analysis that did not involve randomization. There was no original data collection. There were no experiments. |
| Blinding                          | This study involved observational data analysis and did not require blinding.  |
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