Open access • Posted Content • DOI:10.1101/2020.09.22.309187

Assessing whitefly diversity to infer about begomovirus dynamics in cassava in Brazil — Source link

César A. D. Xavier, Angélica Maria Nogueira, Vinicius Henrique Bello, Luís Fernando Maranho Watanabe ...+18 more authors

Institutions: Universidade Federal de Viçosa, Sao Paulo State University, Federal University of Pará, Federal University of Piauí ...+5 more institutions

Published on: 23 Sep 2020 - bioRxiv (Cold Spring Harbor Laboratory)

Topics: Begomovirus and Whitefly

Assessing whitefly diversity to infer about begomovirus dynamics in cassava in Brazil

César A.D. Xavier[1&], Angélica M. Nogueira[1€], Vinícius H. Bello[2], Luís F. M. Watanabe[2], Miguel Alves-Júnior[3], Leonardo F. Barbosa[4], José E.A. Beserra-Junior[5], Alessandra J. Boari[6], Renata F. Calegario[7], Eduardo S. Gorayeb[8], Jaime Honorato-Júnior[9], Gabriel Koch[7], Gaus S.A. Lima[10], Cristian A. Lopes[4], Raquel N. Mello[11], Késsia F. C. Pantoja[6], Fabio N. Silva[8], Roberto Ramos-Sobrinho[10#], Enilton N. Santana[12], José W.P. Silva[13], Renate Krause-Sakate[2]*, F.M. Zerbini[1]*

[1] Dep. de Fitopatologia/BIOAGRO, Universidade Federal de Viçosa, Viçosa, MG 36570-900, Brazil

[2] Dep. de Proteção Vegetal, Universidade Estadual Paulista Júlio de Mesquita Filho, UNESP-FCA, Botucatu, SP 18610-034, Brazil

[3] Lab. de Fitopatologia Agrícola e Florestal, Faculdade de Engenharia Agronômica, Universidade Federal do Pará, Altamira, PA 68372-040, Brazil

[4] Instituto Federal do Sudeste de Minas Gerais, Campus Rio Pomba, Rio Pomba, MG 36180-000, Brazil

[5] Dep. de Fitotecnia, Universidade Federal do Piauí, Teresina, PI 64049-518, Brazil

[6] Embrapa Amazônia Oriental, Belém, PA 66095-903, Brazil

[7] Dep. de Fitotecnia e Fitossanidade, Universidade Federal do Paraná, Curitiba, PR 80035-250, Brazil

[8] Centro de Ciências Agroveterinárias, Universidade do Estado de Santa Catarina, Lages, SC 88520-000, Brazil

[9] Centro Multidisciplinar do Campus de Barra, Universidade Federal do Oeste da Bahia, Barra, BA 47100-000, Brazil

[10] Centro de Ciências Agrárias/Fitossanidade, Universidade Federal de Alagoas, Rio Largo, AL 57100-000, Brazil

[11] Embrapa Arroz e Feijão, Santo Antônio de Goiás, GO 75375-000, Brazil

[12] Instituto Capixaba de Pesquisa, Assistência Técnica e Extensão Rural, CRDR-Linhares, Linhares, ES 29900-970, Brazil

[13] Lab. de Entomologia Agrícola, Faculdade de Engenharia Florestal, Universidade Federal do Pará, Altamira, PA 68372-040, Brazil

[&] Present address: Department of Entomology and Plant Pathology, North Carolina State University, Raleigh, NC 27695, USA

[€] Present address: Dep. de Proteção Vegetal, Universidade Estadual Paulista Júlio de Mesquita Filho, UNESP-FCA, Botucatu, SP 18610-034, Brazil

[#] Present address: School of Plant Sciences, University of Arizona, Tucson, AZ 85721, USA

* Corresponding author: Renate Krause-Sakate

Phone: (+55-14) 3880-7487

E-mail: renate.krause@unesp.br

* Corresponding author: F. Murilo Zerbini

Phone: (+55-31) 3612-2423

E-mail: zerbini@ufv.br

Abstract

Plant virus ecology is strongly dependent on that of its vector. The necessity of a competent vector for transmission is a primary ecological factor driving the host range expansion of plant arthropod-borne viruses, with vectors playing an essential role in promoting disease emergence. Cassava begomoviruses severely constrain cassava production in Africa. Curiously, begomoviruses have never been reported in cassava in South America, the center of origin for this crop. It has been hypothesized that the absence of a competent begomoviruses vector that efficiently colonizes cassava is the reason why begomoviruses have not emerged in South America. To test this hypothesis, we performed a country-wide whitefly diversity study in cassava in Brazil. Adults and/or nymphs of whiteflies were collected from sixty-six cassava fields across twelve states representing the main agroecological zones of the country. A total of 1,385 individuals were genotyped based on partial mitochondrial cytochrome oxidase I (mtCOI) sequences. A high species richness was observed, with five previously described species and two putative new ones. The most prevalent species were *Tetraleurodes acaciae* and *Bemisia tuberculata*, representing over 75% of the analyzed individuals. Although we detected, for the first time, the presence of *Bemisia tabaci* Middle East-Asia Minor 1 (*Bt*MEAM1) colonizing cassava in Brazil, it was not prevalent. The species composition varied across regions, with fields in the Northeast region showing a higher diversity. These results expand our knowledge of whitefly diversity in cassava and support the hypothesis that begomovirus epidemics have not occurred in cassava in Brazil due to the absence of competent vector populations. However, they indicate an ongoing adaptation process of *Bt*MEAM1 to cassava, increasing the likelihood of begomovirus emergence in this crop in the near future.

Keywords: whitefly diversity, cassava, *Bemisia tabaci* complex, *Tetraleurodes acaciae*, *Bemisia tuberculata***,** begomovirus

Introduction

Cassava (*Manihot esculenta* Crantz) is a perennial shrub of the Euphorbiaceae family with great economic and social importance, especially in Africa, Asia, and Latin America. Currently, cassava is the third most important source of calories after rice and corn and is a staple food for more than one billion of people living mainly in developing countries (1). Although the botanical and geographical origin of *M. esculenta* is still debated, studies based on genetic markers and archaeological evidence suggest that domesticated cassava originated from the wild relative progenitor *M. esculenta* ssp. *flabellifolia* in the Amazon basin, with the domestication center located at the southern border of the Amazon in Brazil (2-5). After its introduction in west Africa by Portuguese traders during the 16th century, cassava quickly disseminated throughout tropical Africa and Asia (6). Currently, the African continent is the world's biggest cassava producer, followed by Asia and South America (FAO, 2017). Due to its high resilience to adverse environmental conditions, especially drought, high yield per unit of land and low level of management and inputs required during its life cycle, cassava is a suitable crop for poor and small farmers, partially ensuring food security in many African countries (7-9).

Nevertheless, cassava may be affected by several pathogens and pests. Whiteflies (Hemiptera: Aleyrodidae) are one of the major constraints to its production in developing countries (10). Whiteflies comprise a diverse group of phloem-feeding insects, with more than 1,500 species assigned to 126 genera of which over 20 species have been reported to colonize cassava worldwide (11). In addition to the direct damage due feeding in the plant phloem, whiteflies cause indirect damage by deposition of honeydew, favoring the growth of sooty mold fungi on the leaf surface, and mainly by transmission of a broad range of viruses (12). Currently, species included in the genera *Aleurodicus*, *Aleurothrixus*, *Bemisia* and *Trialeurodes* have been shown to constitute

effective vectors of plant viruses classified in five families (12-15). *Aleurodicus dispersus* and *Aleurothrixus trachoides* each transmit only one virus from the genera *Ipomovirus* and *Begomovirus*, respectively, while *Trialeurodes vaporariorum* and *T. abutilonea* transmit a few viruses included in the genera *Crinivirus* and *Torradovirus*. On the other hand, the *Bemisia tabaci* complex comprises one of the most important group of plant virus vectors, transmitting over 430 viruses, the majority included in the genus *Begomovirus* (12, 16) but including also viruses classified in the genera *Carlavirus*, *Crinivirus*, *Ipomovirus*, *Polerovrius* and *Torradovirus* (12, 17-20).

Over the last decade, advances in the use of molecular markers has led to a deep reappraisal of the taxonomic status of *B. tabaci* (21, 22). Based on molecular phylogeny of the mitochondrial cytochrome oxidase I (mtCOI) gene, it has been proposed that *B. tabaci* consists of a complex of more than 40 cryptic (morphologically indistinguishable) species (21-25). Partial or complete reproductive isolation and biological and ecological differences among distinct species within the complex support the proposed classification (18, 26-28). The global dissemination of polyphagous and invasive species, such as *B. tabaci* Middle East-Asia Minor 1 (*Bt*MEAM1) and *B. tabaci* Mediterranean (*Bt*MED), have caused major changes in the epidemiology of crop-infecting begomoviruses such as *Tomato yellow leaf curl virus* (TYLCV), currently present in all the main tomato producing areas of the world (29-31). In addition, the dissemination of polyphagous whiteflies has favored the transfer of indigenous begomoviruses from wild reservoir hosts to cultivated plants, as occurred in tomato crops in Brazil after the introduction of *Bt*MEAM1 in the mid-1990's (32, 33).

Specific associations between endemic populations of *B. tabaci* and indigenous begomoviruses have also led to the emergence of severe epidemic in crops (34, 35). Cassava

mosaic disease (CMD) is considered the most significant constraint to cassava production in Africa (36, 37). CMD is caused by viruses of the genus *Begomovirus* (family *Geminiviridae*), which are transmitted in a circulative manner by whiteflies of the *Bemisia tabaci* cryptic species complex (16). To date, nine cassava mosaic begomoviruses (CMBs) have been reported in association with CMD, seven of them in Africa and two in the Indian subcontinent (37-39). The emergence of CMD seems to have been the result of the transfer of indigenous begomoviruses from wild reservoir hosts to cassava, probably mediated by endemic populations of *B. tabaci* that have adapted to feed in cassava since its introduction from South America (34, 40). Although wild reservoir hosts and a possible ancestral progenitor of current begomoviruses causing CMD have not been found, the absence of cassava-infecting begomoviruses in the Americas supports an African origin for those viruses, and the presence of cassava-adapted *B. tabaci* species being restricted to Africa reinforces that hypothesis. The high species diversity and high level of molecular variation observed in viral populations causing CMD strongly suggests Africa as a diversification center for CMBs, with distinct CMBs recurrently emerging and evolving for a long time (41-44).

Even if CMD in Africa is caused by indigenous viruses, the fact that cassava in South America has not been affected by begomoviruses is puzzling (38, 45, 46). Carabali, Bellotti (47) suggested that the absence of cassava-infecting begomoviruses in the Americas would be due to lack of competent *B. tabaci* species that efficiently colonize cassava (34, 40). In Colombia, the inability of *B. tabaci* MEAM1 to colonize *M. esculenta* efficiently has been demonstrated under experimental conditions, reinforcing the above hypothesis (46). In addition, a recent study also from Colombia failed to detect any whitefly species of the *B. tabaci* complex in cassava (48).

Although whitefly diversity in Brazil has been surveyed extensively in recent years (49-54), no study has been carried out specifically to explore the composition of whitefly communities

colonizing cassava. Those studies carried out in other crops demonstrated that *B. tabaci* MEAM1 is the predominant species across Brazil in crops such as common bean, cotton, pepper, tomato and soybean. Furthermore, *B. tabaci* MED, which was recently introduced in Brazil, has quickly spread and currently is present in five states from the South and Southeast regions (52-54). A small number of whitefly samples from cassava were analyzed in those studies, with *B. tuberculata* and *Tetraleurodes acaciae* prevalent and detected exclusively in cassava (51, 54). A large survey addressing whitefly diversity in cassava in its domestication center could provide clues to understand the absence of a CMD-like disease in the Americas. Moreover, this knowledge would be useful to anticipate the potential of emergence of begomoviruses in the crop and to help anticipate a management strategy.

Given this context, the objective of this work was to evaluate whitefly diversity in cassava across Brazil to infer about the absence of begomovirus epidemic in cassava. Our results demonstrated that the most prevalent species in cassava were *T. acaciae* and *B. tuberculata*. In addition, we detected for the first time the presence of *Bt*MEAM1 colonizing cassava in Brazil. The possible implications of these findings are discussed considering the absence of CMD and the potential for its emergence in cassava fields in Brazil.

Methods

Whitefly and cassava samples

Whiteflies were collected exclusively from cassava (*M. esculenta*) plants across 12 Brazilian states representative of the five macroregions (North, Northeast, Midwest, Southeast and South; Figure 1) between March 2016 and February 2019 (Table 1). To gather evidence of whether a given species was colonizing cassava, adults and nymphs from the same field were collected

whenever possible (Table 1). Samples were obtained from commercial and non-commercial (subsistence) crops. Whitefly adults were sampled using a hand-held aspirator and nymphs were collected with the aid of a needle. Insects were preserved in 95% ethanol and stored at −20°C until being used for molecular identification of the species.

To verify the presence of begomoviruses infecting cassava, foliar samples were also collected at some sampled sites (Suppl. Table S1). The samples were collected randomly regardless of the presence of virus-like symptoms. The leaves were press-dried and stored at room temperature as herbarium-like samples until being used for DNA extraction.

Whitefly species identification

Whitefly species were identified using PCR-RFLP of the partial mtCOI fragment followed by sequencing, as previously described (54). When enough adults and nymphs were collected at a given sampled site, ten individuals from each stage were analyzed, and when only one stage was obtained, 20 individuals were tested (Table 1). When variation in the RFLP pattern was observed in the first screening, suggesting that more than one species could be present in that site, approximately five additional individuals for each stage were analyzed according to sample availability.

Total DNA was extracted from single individual whiteflies following a Chelex protocol (55). Briefly, adults or nymphs were ground in 30 µl of Chelex buffer (5% Chelex in 1x Tris-EDTA) using a toothpick in a 600 µl tube. Samples were vortexed for 30 seconds and incubated at 99°C for 8 min in a PTC-100 thermocycler (MJ Research). Next, the tubes were centrifuged at 14,000 g for 5 min and 20 µl of the supernatant was collected and transferred to a new tube. One microliter of the supernatant was used as a template for PCR amplification of a 800 bp fragment

of the mtCOI gene using primers C1-J-2195 and L2-N-3014 (56, 57). PCR was performed using 0.2 µM of forward and reverse primers in a final volume of 25 µl using GoTaq Colorless Master Mix (Promega), following the manufacturer's instructions. The PCR cycles consisted of an initial denaturing step at 95°C for 5 min, followed by 35 cycles at 95°C for 30 sec, 42°C for 45 sec and 72°C for 1 min, with a final extension at 72°C for 10 min. Amplified products were visualized in 0.8% agarose gels stained with ethidium bromide and directly used for RFLP analysis (58).

RFLP analysis of the amplicons consisted of 5 µl of each PCR product digested with 0.1 unit of *Taq*I (Promega) in a final volume of 20 µl. Reactions were performed at 65°C for 2 hours and visualized in 1.2% agarose gels stained with ethidium bromide. To verify whether the predicted mtCOI restriction pattern corresponded to a given species according to *in silico* prediction, a subset of PCR products from adults and nymphs representative of distinct patterns from different sampled sites were selected and sequenced. PCR products were precipitated with 100% ethanol and 3 M sodium acetate pH 5.2 (59) and sequenced commercially (Macrogen Inc.) in both directions using primers C1-J-2195/L2-N-3014.

For a small subset of samples that failed to yield a PCR product using primers C1-J-2195 and L2-N-3014, a second screening, using a recently described primer set with improved specificity for species of the *B. tabaci* complex and *B. afer* (2195Bt and C012/Bt-sh2), was performed (24). Samples that still failed to amplify or had unexpected RFLP pattern were analyzed with specific primers for *T. vaporariorum* (TvapF and Wfrev) (60).

Sequence comparisons and phylogenetic analysis

Nucleotide sequences were first checked for quality and assembled using Geneious v. 8.1 (61). mtCOI sequences were initially analyzed with the BLAST*n* algorithm (62) to determine the

209 whitefly species with which they shared greatest similarity. Pairwise comparisons between all
210 mtCOI sequences obtained here and those with higher similarities (as determined by the BLAST*n*
211 search) were performed with the program SDT v. 1.2 (63) using the MUSCLE alignment option
212 (64).

213 For phylogenetic analyses, the final dataset was composed of 142 sequences: 95 obtained
214 in this work and 47 sequences representative of species in the family Aleyrodidae. Sequences were
215 retrieved from GenBank and from the updated mtCOI reference dataset for species of the *Bemisia*
216 *tabaci* complex (65). Multiple sequence alignments were prepared using the MUSCLE option in
217 MEGA7 (66). Alignments were checked and manually adjusted when necessary. Phylogenetic
218 trees were constructed using Bayesian inference performed with MrBayes v. 3.0b4 (67). The
219 program MrModeltest v. 2.2 (68) was used to select the nucleotide substitution model with the best
220 fit in the Akaike Information Criterion (AIC). The analyses were carried out running 50,000,000
221 generations with sampling at every 1,000 generations and a burn-in of 25%. The convergence was
222 assumed when average standard deviation of split frequencies was lower than 0.001. Trees were
223 visualized and edited using FigTree (tree.bio.ed.ac.uk/software/figtree) and CorelDRAW X5,
224 respectively.

225

226 **Virus detection in foliar samples**

227 Total DNA was extracted as described (69) and used as a template for PCR using the DNA-
228 A universal primer pair PAL1v1978 and PAR1c496 (70). PCR was performed in a final volume
229 of 25 µl using *Taq* DNA Polymerase (Invitrogen) following the manufacturer's instructions. The
230 PCR cycles consisted of an initial denaturing step at 95°C for 5 min, followed by 35 cycles at 95°C

for 1 min, 52°C for 1 min and 72°C for 1 min, with a final extension at 72°C for 10 min. PCR products were visualized in 0.8% agarose gels stained with ethidium bromide

Diversity index and statical analysis

Simpson's index of diversity (1-D) was calculated to verify if there was any difference in whitefly diversity across macroregions. This index represents the probability that two randomly chosen individuals in a given sampled site will belong to distinct species (71). Simpson's index was chosen as its value increases with increasing diversity and assigns more weight to more abundant species in a sample. We assume that species colonizing cassava will be in abundance, whereas rare species that briefly visit the plant without colonizing it will be underrepresented. Simpson's index was calculated for each sampled site separately and then pooled according to macroregions. To assess the statistical significance of the differences in diversity among regions, the non-parametric Kruskal-Wallis test followed by *post hoc* multiple comparison test using Fisher's least significant difference was calculated, using the function kruskal implemented in the Agricolae package in R software (72). Non-parametric Spearman's rank correlation coefficient analysis was performed using the ggpubr package in R software (72).

Results

High whitefly species richness in cassava in Brazil

To verify the composition of whitefly communities colonizing cassava in Brazil, sampling was performed across the country, including the main agroecological zones. A total of 66 sites from 12 states were sampled (Figure 1; Table 1). Out of 1,385 individuals submitted to PCR-RFLP analysis, 58 adults and 37 nymphs from different locations and representing distinct restriction

254 patterns were sequenced. The combination of PCR-RFLP followed by sequencing showed
255 reliability and consistence for species identification without misidentification due to incongruence
256 between the two methods.

257 Based on pairwise comparisons and molecular phylogeny of the partial mtCOI gene, we
258 identified the presence of at least seven species comprising the whitefly community in cassava
259 (Figure 2; Table 1). Among them, *T. acaciae* and *B. tuberculata*, both previously reported in this
260 crop, were the most prevalent, representing over 75% of the analyzed individuals. In addition,
261 based on the criterion of 3.5% divergence to differentiate species within the *B. tabaci* complex,
262 three *B. tabaci* species were identified, with *Bt*MED previously reported, and *Bt*MEAM1 and
263 *Bt*NW identified for the first time in cassava fields in Brazil (Figure 2; Table 1). The species
264 *Bt*MEAM1 represented 18% of the total individuals analyzed, followed by *Bt*MED (1.6%) and
265 *Bt*NW (0.21%).

266 Furthermore, two putative new species were identified (Figure 2), provisionally named
267 whitefly new species 1 and 2 (WtNEW1 and WtNEW2). The WtNEW1 mtCOI sequence
268 (KY249522) showed highest identity (80.65%) and clustered close to the *T. acaciae* clade,
269 comprised of individuals reported here and three other previously reported sequences from cassava
270 in Brazil (Figure 2). For WtNEW2, two mtCOI sequences obtained from an adult (JX678666) and
271 a nymph (DQ989531) shared 97.81% among them and showed highest identity with *B. tabaci*
272 (adult: 82.11%; nymph: 81.68%) and clustered as a basal sister clade to the genus *Bemisia* (Figure
273 2). Although whitefly taxonomy is predominantly based on puparial characters (73) and there is
274 no taxonomic criterion established based in mtCOI sequences for most of the groups, as has been
275 proposed for the *B. tabaci* complex, the level of divergence between the two proposed new species
276 with the closest species is similar to the level of divergence observed between species already

277 described within the Aleyrodidae, as demonstrated in pairwise comparisons (data not shown) and
278 phylogenetic analysis (Figure 2). Nevertheless, further molecular and morphological
279 characterization should be performed. Together, these results indicate the existence of a high
280 whitefly species richness in cassava in Brazil.
281
282 **Both the prevalence and the capacity to colonize cassava differ among species**
283 Nymphs were collected for samples identified as *T. acaciae, B. tuberculata, Bt*MEAM1
284 and the two new putative species (Figure 3A), suggesting that these species may colonize cassava.
285 Nymphs were not obtained at two sites where *Bt*MED was prevalent (SP1 and SP12). Although it
286 could be suggested that this species has the potential to colonize cassava due to the high prevalence
287 of adults at these two sites, the lack of nymphs suggests otherwise. Moreover, at the sites PR4 and
288 MT6, *Bt*MEAM1 predominated among adults but 100% of the nymphs were *B. tuberculata*,
289 suggesting that the predominance at one stage does not necessarily mean predominance in another
290 stage. Indeed, correlation analysis between the number of adults and nymphs, performed for all
291 sites where both stages were sampled, showed no significant correlation between them (Supp.
292 Figure S1). Further sampling in those sites or free-choice experiments are necessary to confirm
293 the potential of *Bt*MED to colonize cassava. Considering the whole sampling, we detected only
294 three adults of *Bt*NW, suggesting an inability of this specie to colonize cassava.
295 To verify if prevalence differs among species across distinct developmental stages, the data
296 were separated according to stage and the proportions of individuals were compared for the three
297 most abundant species (Figure 3B, C). Considering the entire data set, *T. acaciae* was the prevalent
298 species, followed by *B. tuberculata* and *Bt*MEAM1 (x_2^2=152.63, P<2.2x10^{-16}). The same was true
299 according to stage, either adults (x_2^2=28.61, P<6.1x10^{-07}) or nymphs (x_2^2=169.44, P<2.2x10^{-16};

Figure 3B). However, caution is needed to interpret these results as only adults were sampled at some sites where *Bt*MEAM1 and *B. tuberculata* were prevalent (Figure 3A), which could bias the analysis, causing an underestimation of the number of nymphs for those species. Therefore, we also analyzed the data considering only those sites where both adults and nymphs were obtained. Again, *T. acaciae* was the predominant species followed by *B. tuberculata* and *Bt*MEAM1 considering either the entire data set (x_2^2=258.61, P<2.2x10^{-16}) or only nymphs (x_2^2=164.47, P<2.2x10^{-16}). When only adults were considered, *T. acaciae* was still predominant (x_2^2=113.52, P<2.2x10^{-16}) but no difference between *B. tuberculata* and *Bt*MEAM1 was observed (x_1^2=0.505, P=0.477; Figure 3C). Moreover, it could be argued that samples from Minas Gerais (MG) were overrepresented in our sampling (Figure 1C), which could also bias the results presented above due to the predominance of *T. acaciae* in this state (Figure 3A). To test this possibility, we analyzed the data excluding the samples from MG. In this case, when both stages were considered, *B. tuberculata* was predominant (x_2^2=62.09, P=3.3x10^{-14}) but no difference between *T. acaciae* and *Bt*MEAM1 was observed (x_1^2=1.91, P=0.166). When each stage was considered separately, *B. tuberculata* was predominant followed by *Bt*MEAM1 and *T. acaciae* (adults: x_2^2=43.94, P=2.9x10^{-10}; nymphs: x_2^2=84.19, P<2.2x10^{-16}). Together, these results indicate that the potential to colonize cassava differs among species, which could be due either to lower preference for the plant or to differences in the competitive ability among species during cassava colonization. In addition, they reinforce the low efficiency of *Bt*MEAM1 to colonize cassava.

Competitive interference does not explain the differences in prevalence

Interestingly, at least two species were detected co-occurring at 51% of the sampled sites (Figure 3A). To verify the possibility of competition among *T. acaciae*, *B. tuberculata* and

323 *Bt*MEAM1 to explain the observed differences in prevalence (instead of differences in host
324 preference), the competitive capacity of these three species was inferred based on the analysis of
325 predominance at the sites where they occurred together. Initially, we verified if there were any
326 differences in incidence, defined here as the number of sampled sites where at least one individual
327 belonging to one of the three species was detected (Figure 4A). The results demonstrate that there
328 were no differences in incidence among them (x_2^2=1.25, *P*=0.537; Figure 4A). In addition, no
329 differences were observed when the proportion of sites where whitefly species occurred alone or
330 in different combinations was compared (x_6^2=3.26, *P*=0.776; Figure 4B). However, when we
331 compared the occurrence between *Bt*MEAM1 and non-*B. tabaci* species at the sites where they
332 occur alone, the number of sites with non-*B. tabaci* species was higher (x_1^2 =6.53, *P*=0.011; Figure
333 4B). Thus, the competitive capacity was inferred based on the proportion of individuals from each
334 species at the fields where these species were detected co-occurring in different combinations
335 (Figure 4C). Interestingly, at the sites where *Bt*MEAM1 and *B. tuberculata* were sampled together,
336 *B. tuberculata* predominated over *Bt*MEAM1, suggesting higher competitive potential (Figure
337 4C). For all other species combinations, no evidence of differences in competitive capacity were
338 observed (Figure 4C). Together, these results suggest that, rather than competition, lower host
339 preference by *Bt*MEAM1 explains its non-prevalence compared to *T. acaciae* and *B. tuberculata*,
340 resulting in low colonization rate as indicated by the low number of *Bt*MEAM1 nymphs detected
341 in cassava (Figure 3).
342
343 **Composition and species diversity of whiteflies differ among Brazilian regions**
344 The predominance of species composing the whitefly community across macroregions
345 varied considerably. While *T. acaciae* predominated in the North, Southeast and Northeast, it was

not detected in the Midwest (Figure 5A). In addition, *B. tuberculata* was detected in all regions, and was prevalent in the South and Midwest. *Bt*MEAM1, although not prevalent in any of the regions, was also detected in all regions. Although the number of species detected was higher in the Southeast, where six species out seven were detected, whitefly diversity was significantly higher in fields in the Northeast according to Simpson's index of diversity (Figure 5B), with no differences among the other four regions.

No begomoviruses detected infecting cassava

To verify the presence of begomoviruses infecting cassava, we analyzed leaves sampled in some of the fields where whiteflies were collected (Supp. Table S1). Based on PCR detection using universal primers for begomoviruses, all plants were negative.

Discussion

Vectors play an essential role during the life cycle of plant viruses, directly affecting their ecology and evolution (74-76). Usually, a group of plant viruses establishes a very specific interaction with only one or a few related species of vectors, making virus ecology strongly dependent on that of its vector (74). It has been suggested that the natural host range of a virus is dependent on its vector's host range, as most plant viruses have greater specificity for the vector than for the plant host (77, 78). Indeed, the existence of a competent vector for transmission and able to colonize potential reservoir and recipient new hosts is a primary ecological factor driving host range expansion of viruses. Thus, vectors play an essential role during viral disease emergence and epidemics (12, 18, 77, 79). Understanding ecological factors, such as vector species dynamics

368 in crops, might provide important clues about historical and current events of emergence or re-
369 emergence of viral diseases, and even anticipate the potential for new ones to occur (80).

370 Although it could be suggested that there are no begomoviruses capable of infecting
371 cassava in the Americas, the high diversity of begomoviruses reported in a broad range of
372 cultivated and non-cultivated plants in several botanical families, including the Euphorbiaceae,
373 make this highly unlikely (33, 81-89). Thus, the absence of a competent vector able to colonize
374 cassava and transfer begomoviruses from wild plants to cassava, as previously suggested (47),
375 seems to be a more plausible hypothesis to explain the lack of begomovirus epidemics in this crop.

376 Our country-wide survey of whiteflies associated with cassava in Brazil uncovered a high
377 degree of species diversity and showed that *T. acaciae* and *B. tuberculata* are the prevalent species
378 across the country. Non-*B. tabaci* species, including *B. tuberculata*, have been shown to be
379 prevalent also in Colombia (48). In contrast, in Africa, endemic species of the *B. tabaci* complex
380 are prevalent in cassava (80, 90, 91). Previous studies surveying whitefly diversity in South
381 American countries failed to detect *T. acaciae* and *B. tuberculata* in crops other than cassava,
382 indicating a very narrow host range, which may in fact be restricted to cassava or at least to
383 cultivated plants (51, 54, 92).

384 *Bt*MEAM1 and *Bt*NW are reported here for the first time in cassava in Brazil. *Bt*MEAM1
385 was the third most prevalent species, representing 18% of the genotyped individuals, and with
386 similar incidence to *T. acaciae* and *B. tuberculata*. The failure of previous studies to detect
387 *Bt*MEAM1 in cassava may have been due to the small number of samples analyzed. The wide
388 distribution and prevalence of *Bt*MEAM1 in the main agroecological zones in Brazil has been well
389 established, mostly in association with annual crops such as soybean, cotton, common bean and
390 tomato (54). In these crops, *Bt*MEAM1 has a great reproductive capacity, rapidly increasing its

391 population. Interestingly, our data showed the higher prevalence of *Bt*MEAM1 to be in the
392 Midwest, where extensive agriculture predominates. The harvest of annual crops in the Midwest
393 might cause the migration of the insect to semiperennial hosts such as cassava, which could explain
394 why in some sites where *Bt*MEAM1 predominated among adults, it was not detected as nymphs
395 (e.g., sites MT5, MT6, PR4).

396 It will be important to monitor *Bt*MEAM1 populations in cassava over the next years, to
397 assess its possible adaptation to this host. The fact that we collected *Bt*MEAM1 nymphs at several
398 locations suggests that this process may already be under way. We also detected *Bt*MED, a
399 worrying result given the recent introduction of this species in the Brazil and its potential to
400 displace other species, including *Bt*MEAM1 (93-95). *Bt*MED has disseminated quickly in the
401 country, mainly in association with ornamental plants in greenhouses (54). Even though we detect
402 *Bt*MED associated to cassava, we cannot infer its potential to effectively colonize this host since
403 only adults were collected. The third species detected is the indigenous *Bt*NW. Although
404 *Bt*MEAM1 partially displaced *Bt*NW in Brazil, this species can still be sporadically detected,
405 mostly in association with non-cultivated hosts (50, 51, 54). It has been recurrently detected in
406 *Euphorbia heterophylla*, suggesting a potential to colonize other species in the family
407 Euphorbiaceae. However, the very low frequence with which it was detected and the absence of
408 nymphs indicate that *Bt*NW is poorly adapted to cassava.

409 The identification of two putative new species highlights the remarkable genetic diversity
410 of whiteflies. Interestingly, one of the new species was collected in the state of Mato Grosso, which
411 corresponds to the region considered to be the domestication center of cassava (2-5). Further
412 studies are needed to explore plant biodiversity in this region (96, 97), which might reveal a similar
413 diversity of whiteflies which may be specifically adaptated to non-cultivated plant species due to

414 long term co-evolution. The close phylogenetic relationship of the new species with non-*B. tabaci*
415 whiteflies suggests that they are not virus vectors.

416 Whitefly species richness in cassava is just starting to be assessed, and may be greater than
417 reported here. Based on morphological characters, Alonso, Racca-Filho (98) reported the presence
418 of *Aleurothrixus aepim* and *Trialeurodes manihoti* colonizing cassava in the state of Rio de
419 Janeiro. Although we did not analyze samples from that region, the failure to detect these species
420 in other states suggests a restricted occurrence. Moreover, morphological characters alone are not
421 always sufficient to classify whiteflies at the species level, and additional studies using molecular
422 tools are needed to assess these molecularly uncharacterized whiteflies species (99).

423 Host suitability has been shown to be an important factor influencing the competitive
424 capacity among species of the *B. tabaci* complex (94, 95, 100). Watanabe, Bello (95) demonstrated
425 that displacement capacity between two invasive *B. tabaci* species was dependent on host
426 suitability. While *Bt*MEAM1 displaced *Bt*MED only on tomato, *Bt*MED displaced *Bt*MEAM1 on
427 sweet pepper and common bean. Luan, Xu (100) demonstrated that even in a host plant poorly
428 suitable for *Bt*MEAM1, it was able to displace an indigenous species challenger. These authors
429 demonstrated that even though host suitability may affect the speed of displacement, it may not
430 affect the direction, as *Bt*MEAM1 always won the challenge (100). Interestingly, two or more
431 species occurring sympatrically were detected in 51% of the fields analyzed in our study. In sites
432 where *Bt*MEAM1 and *B. tuberculata* co-occurred, *B. tuberculata* predominated, suggesting a
433 higher competitive capacity. Nonetheless, in all other combinations of co-occurring species, no
434 differences in prevalence were observed. Thus, competitive capacity is unlikely to explain the low
435 prevalence of *Bt*MEAM1, or the differences observed between *T. acaciae* and *B. tuberculata*.

436 Host adaptation may be a more important component affecting the low predominance of
437 *Bt*MEAM1 in cassava, as previously suggested (47). The inability of *Bt*MEAM1 and *Bt*MED to
438 colonize domesticated cassava efficiently has been demonstrated under experimental conditions
439 (45, 46, 102, 103). Carabali, Montoya-Lerma (45), evaluating the colonization potential of
440 *Bt*MEAM1 in three commercial cassava genotypes, demonstrated that only in one of them did
441 *Bt*MEAM1 complete its development cycle from eggs to adult, and even then, at very low rates
442 (0.003%). Using an electrical penetration graph assay, Milenovic, Wosula (103) demonstrated the
443 inability of *Bt*MED to feed in cassava plants. Adults of this species spent a very short time
444 ingesting cassava phloem sap compared to sap from a suitable host, suggesting that they would die
445 by starvation in the field. Furthermore, the low efficiency of whiteflies of the *Bt*MED
446 mitochondrial subgroups Q1 and Q2 in using cassava as a host has also been demonstrated (102).
447 Oviposition and adult survival rates were very low, and development from eggs to adults was not
448 observed. Although these studies were conducted under experimental conditions, the low
449 predominance of *Bt*MEAM1 and *Bt*MED shown here and in other field surveys in Africa (90, 91,
450 101) strongly indicates a low adaptation of these species to cassava.
451 Nevertheless, our results indicate an ongoing adaptation process of *Bt*MEAM1 to cassava,
452 with the detection of nymphs and adults in the same field. Interestingly, Carabali, Bellotti (47)
453 demonstrated a gradual increase in the rate of reproduction and development of *Bt*MEAM1 after
454 successive passages on plants phylogenetically related to the genus *Manihot* (*Euphorbia*
455 *pulcherrima* and *Jatropha gossypiifolia*), indicating the potential of this whitefly species to
456 become adapted to cassava through intermediate hosts. Furthermore, successful reproduction in
457 the wild relative *M. esculenta* ssp. *flabellifolia* indicates that this plant may constitute an
458 intermediate host leading to adaptation (46). This plant has been reported to be widely spread in

459 the Amazon basin and the Midwest region of Brazil (97). Interestingly, our data showed the higher
460 prevalence of *Bt*MEAM1 to be in the Midwest. Although we cannot establish a cause and effect
461 relationship, it is reasonable to speculate that *M. esculenta* ssp. *flabellifolia* could be acting as an
462 intermediate host mediating adaptation. A survey addressing whitefly diversity in this host should
463 be necessary to test this hypothesis.

464 In Brazil, cassava is predominantly grown as a subsistence crop, usually side by side with
465 other vegetables and with a high incidence of weeds. Growing cassava in a heterogenous
466 environment, especially in the presence of related plants, may increase the adaptation potential of
467 *Bt*MEAM1 and other species of the complex such as *Bt*MED, which we also detected in the open
468 field. A high diversity of plants in cassava fields may allow an overlapping of ecological niches
469 for distinct whitefly species, which under enough selection pressure may gradually adapt to new
470 hosts. The sympatric occurrence of *T. acaciae*, *B. tuberculata* and *Bt*MEAM1, supports the role of
471 botanical heterogeneity in shaping the composition of whitefly populations associated with
472 cassava. A similar pattern was observed in Colombia, with 66% of the surveyed sites showing at
473 least two species occurring sympatrically (48). Moreover, a predominance of one species in a given
474 developmental stage and a different one in another stage (e.g., nymphs *vs* adults) at the same site
475 suggests that other hosts may sustain reproduction and development, with adults migrating to
476 cassava.

477 *Euphorbia heterophylla* (family Euphorbiaceae) is an invasive weed widely spread across
478 Brazil and associated with several crops (89, 104). The presence of *E. heterophylla* plants in
479 association with cassava (Figure 1A) and the fact that it was the most suitable host for *Bt*MEAM1
480 in Brazil out seven tested (105) shows its potential to act as an intermediate host mediating
481 *Bt*MEAM1 adaptation. *E. heterophylla* has been frequently associated with the begomovirus

Euphorbia yellow mosaic virus (EuYMV) (89). Barreto, Hallwass (106) demonstrated that this plant is also a host of *Tomato severe rugose virus* (ToSRV), which even at a very low titer was transmitted to tomato plants, demonstrating the potential of *E. heterophylla* to act as a reservoir host. Surprisingly, considering that *E. heterophylla* and tomato belong to distinct botanical families, EuYMV is able to infect tomato (106). The closer botanical relationship between *E. heterophylla* and cassava may indicate a higher potential of EuYMV to infect cassava. The presence of EuYMV-infected *E. heterophylla* in cassava fields, as observed in this study (Figure 1A), its suitability as a host for *Bt*MEAM1, and the high efficiency of EuYMV transmission by *Bt*MEAM1 (107), suggest that EuYMV may have spillover potential to cassava. Experiments are ongoing in our laboratory to assess this spillover potential.

The emergence of begomoviruses in tomato crops in Brazil followed the introduction of *Bt*MEAM1 (32, 33), demonstrating the role of vector populations in promoting viral host range expansion and consequently epidemics. Thus, the adaptation of whiteflies to cassava could facilitate the emergence of begomoviruses in this crop. The establishment of management strategies to prevent or at least delaying the adaptation process is therefore necessary. *Bemisia tabaci* species may disperse across long distances international trade routes (108). Thus, preventing the introduction of cassava-adapted *B. tabaci* species from Africa should also be a priority.

Data accessibility

mtCOI sequences obtained in this study were deposited in GenBank under accession numbers: MT901081 to MT901172 and MT904381 to MT904382. For detailed information see Supplementary Table S2.

Competing interests

The authors declare that they have no competing interests

Author's contribution

CADX, FMZ and RKS contributed to the design and implementation of the study; CADX, AMN, VHB, LFMW, MAJ, LFB, JEABJ, AJB, RFC, ESG, JHJ, GK, GSAL, CAL, RNM, KFCP, FNS, RRS, ENS and JWPS collected whitefly and leaf samples; CADX, AMN, VHB and LFMW processed whitefly samples; CADX performed the analyses and drafted the manuscript; CADX and FMZ prepared the final version of the manuscript.

Acknowledgements

This work was funded by CAPES (Financial code 01), CNPq (409599/2016-6) and Fapemig (APQ-03276-18) grants to FMZ.

References

1. Montagnac JA, Davis CR, Tanumihardjo SA. Nutritional value of cassava for use as a staple food and recent advances for improvement. Comprehensive Reviews in Food Science and Food Safety. 2009;8(3):181-94.

2. Olsen KM, Schaal BA. Evidence on the origin of cassava: phylogeography of *Manihot esculenta*. Proceedings of the National Academy of Sciences, USA. 1999;96(10):5586-91.

3. Leotard G, Duputie A, Kjellberg F, Douzery EJ, Debain C, de Granville JJ, et al. Phylogeography and the origin of cassava: new insights from the northern rim of the Amazonian basin. Molecular Phylogenetics and Evolution. 2009;53(1):329-34.

4. Clement CR, Rodrigues DP, Alves-Pereira A, Mühlen GS, Cristo-Araújo Md, Moreira PA, et al. Crop domestication in the upper Madeira River basin. Boletim do Museu Paraense Emílio Goeldi Ciências Humanas. 2016;11:193-205.

5. Watling J, Shock MP, Mongeló GZ, Almeida FO, Kater T, De Oliveira PE, et al. Direct archaeological evidence for Southwestern Amazonia as an early plant domestication and food production centre. PloS one. 2018;13(7):e0199868.

6. Carter SE, Fresco LO, Jones PG, Fairbairn JN. Introduction and diffusion of cassava in Africa. IITA Research Guide no. 49. Ibadan, Nigeria: International Institute of Tropical Agriculture (IITA); 1997. 34 p.

7. Alves AAC, Setter TL. Response of cassava leaf area expansion to water deficit: cell proliferation, cell expansion and delayed development. Annals of Botany. 2004;94(4):605-13. Epub 2004/08/19.

8. El-Sharkawy MA. Cassava biology and physiology. Plant Molecular Biology. 2004;56(4):481-501.

9. Gleadow R, Pegg A, Blomstedt CK. Resilience of cassava (*Manihot esculenta* Crantz) to salinity: implications for food security in low-lying regions. Journal of Experimental Botany. 2016;67(18):5403-13. Epub 2016/08/09.

10. Herrera Campo BV, Hyman G, Bellotti A. Threats to cassava production: known and potential geographic distribution of four key biotic constraints. Food Security. 2011;3(3):329.

11. Vasquez-Ordonez AA, Hazzi NA, Escobar-Prieto D, Paz-Jojoa D, Parsa S. A geographic distribution database of the Neotropical cassava whitefly complex (Hemiptera, Aleyrodidae) and their associated parasitoids and hyperparasitoids (Hymenoptera). Zookeys. 2015(545):75-87. Epub 2016/01/23.

12. Navas-Castillo J, Fiallo-Olivé E, Sánchez-Campos S. Emerging virus diseases transmitted by whiteflies. Annual Review of Phytopathology 2011;49:219-48.

13. Njoroge MK, Mutisya DL, Miano DW, Kilalo DC. Whitefly species efficiency in transmitting cassava mosaic and brown streak virus diseases. Cogent Biology. 2017;3(1):1311499.

14. Maruthi MN, Jeremiah SC, Mohammed IU, Legg JP. The role of the whitefly, *Bemisia tabaci* (Gennadius), and farmer practices in the spread of cassava brown streak ipomoviruses. Journal of Phytopathology. 2017;165(11-12):707-17. Epub 2017/08/22.

15. Chandrashekar K, Rao A, Gorane A, Verma R, Tripathi S. *Aleurothrixus trachoides* (Back) can transmit begomovirus from Duranta to potato, tomato and bell pepper. J Biosci. 2020;45:36.

16. Zerbini FM, Briddon RW, Idris A, Martin DP, Moriones E, Navas-Castillo J, et al. ICTV Virus Taxonomy Profile: *Geminiviridae*. Journal of General Virology. 2017;98(2):131-3.

17. Whitfield AE, Falk BW, Rotenberg D. Insect vector-mediated transmission of plant viruses. Virology. 2015;480:278-89.

18. Gilbertson RL, Batuman O, Webster CG, Adkins S. Role of the insect supervectors *Bemisia tabaci* and *Frankliniella occidentalis* in the emergence and global spread of plant viruses. Annual Review of Virology. 2015;2:67-93.

19. Ghosh S, Kanakala S, Lebedev G, Kontsedalov S, Silverman D, Alon T, et al. Transmission of a new polerovirus infecting pepper by the whitefly *Bemisia tabaci*. Journal of virology. 2019;15:00488-19.

20. Costa TM, Inoue-Nagata AK, Vidal AH, Ribeiro SdG, Nagata T. The recombinant isolate of cucurbit aphid-borne yellows virus from Brazil is a polerovirus transmitted by whiteflies. Plant Pathology. 2020;69(6):1042-50.

21. Dinsdale A, Cook L, Riginos C, Buckley YM, De Barro P. Refined global analysis of *Bemisia tabaci* (Hemiptera: Sternorrhyncha: Aleyrodoidea: Aleyrodidae) mitochondrial cytochrome oxidase 1 to identify species level genetic boundaries. Annals of the Entomological Society of America. 2010;103(2):196-208.

22. De Barro PJ, Liu SS, Boykin LM, Dinsdale AB. *Bemisia tabaci*: A statement of species status. Annual Review of Entomology. 2011;56:1-19.

23. Lee W, Park J, Lee GS, Lee S, Akimoto SI. Taxonomic status of the *Bemisia tabaci* complex (Hemiptera: Aleyrodidae) and reassessment of the number of its constituent species. PloS one. 2013;8:e63817.

24. Mugerwa H, Seal S, Wang H-L, Patel MV, Kabaalu R, Omongo CA, et al. African ancestry of New World, *Bemisia tabaci*-whitefly species. Scientific reports. 2018;8(1):2734.

25. Vyskočilová S, Tay WT, van Brunschot S, Seal S, Colvin J. An integrative approach to discovering cryptic species within the *Bemisia tabaci* whitefly species complex. Scientific reports. 2018;8(1):10886.

26. Qin L, Pan L-L, Liu S-S. Further insight into reproductive incompatibility between putative cryptic species of the *Bemisia tabaci* whitefly complex. Insect Science. 2016;23(2):215-24.

27. De Marchi BR, Marubayashi JM, Favara GM, Yuki VA, Watanabe LFM, Barbosa LF, et al. Comparative transmission of five viruses by *Bemisia tabaci* NW2 and MEAM1. Tropical Plant Pathology. 2017;42(6):495-9.

28. Malka O, Santos-Garcia D, Feldmesser E, Sharon E, Krause-Sakate R, Delatte H, et al. Species-complex diversification and host-plant associations in *Bemisia tabaci*: A plant-defence, detoxification perspective revealed by RNA-Seq analyses. Molecular Ecology. 2018;27(21):4241-56.

29. Lefeuvre P, Martin DP, Harkins G, Lemey P, Gray AJA, Meredith S, et al. The spread of tomato yellow leaf curl virus from the Middle East to the world. PLoS Pathogens. 2010;6(10):e1001164.

30. Mabvakure B, Martin DP, Kraberger S, Cloete L, van Brunschot S, Geering AD, et al. Ongoing geographical spread of *Tomato yellow leaf curl virus*. Virology. 2016;498:257-64.

31. Pan H, Chu D, Yan W, Su Q, Liu B, Wang S, et al. Rapid spread of tomato yellow leaf curl virus in China is aided differentially by two invasive whiteflies. PloS one. 2012;7(4):e34817. Epub 2012/04/20.

32. Ribeiro SG, Ávila AC, Bezerra IC, Fernandes JJ, Faria JC, Lima MF, et al. Widespread occurrence of tomato geminiviruses in Brazil, associated with the new biotype of the whitefly vector. Plant Disease. 1998;82:830.

33. Rocha CS, Castillo-Urquiza GP, Lima ATM, Silva FN, Xavier CAD, Hora-Junior BT, et al. Brazilian begomovirus populations are highly recombinant, rapidly evolving, and segregated based on geographical location. Journal of virology. 2013;87(10):5784-99.

34. Fauquet C, Fargette D. African cassava mosaic virus: etiology, epidemiology and control. Plant Disease. 1990;74:404-11.

35. Pan LL, Cui XY, Chen QF, Wang XW, Liu SS. Cotton leaf curl disease: which whitefly is the vector? Phytopathology. 2018;108(10):1172-83.

36. Rey C, Vanderschuren H. Cassava mosaic and brown streak diseases: current perspectives and beyond. Annual Review of Virology. 2017;4(1):429-52. Epub 2017/06/25.

37. Jacobson AL, Duffy S, Sseruwagi P. Whitefly-transmitted viruses threatening cassava production in Africa. Current Opinion in Virology. 2018;33:167-76. Epub 2018/09/23.

38. Patil BL, Fauquet CM. Cassava mosaic geminiviruses: actual knowledge and perspectives. Plant Pathology. 2009;10(5):685-701.

39. Legg JP, Lava Kumar P, Makeshkumar T, Tripathi L, Ferguson M, Kanju E, et al. Cassava virus diseases: biology, epidemiology, and management. Advances in Virus Research. 2015;91:85-142. Epub 2015/01/17.

40. Legg J, Fauquet C. Cassava mosaic geminiviruses in Africa. Plant Molecular Biology. 2004;56(4):585-99.

41. De Bruyn A, Villemot J, Lefeuvre P, Villar E, Hoareau M, Harimalala M, et al. East African cassava mosaic-like viruses from Africa to Indian ocean islands: molecular diversity, evolutionary history and geographical dissemination of a bipartite begomovirus. BMC evolutionary biology. 2012;12:228. Epub 2012/11/29.

42. Ndunguru J, Legg J, Aveling T, Thompson G, Fauquet C. Molecular biodiversity of cassava begomoviruses in Tanzania: evolution of cassava geminiviruses in Africa and evidence for East Africa being a center of diversity of cassava geminiviruses. Virology Journal. 2005;2(1):21.

43. De Bruyn A, Harimalala M, Zinga I, Mabvakure BM, Hoareau M, Ravigne V, et al. Divergent evolutionary and epidemiological dynamics of cassava mosaic geminiviruses in Madagascar. BMC evolutionary biology. 2016;16:182.

44. Tiendrebeogo F, Lefeuvre P, Hoareau M, Harimalala MA, De Bruyn A, Villemot J, et al. Evolution of *African cassava mosaic virus* by recombination between bipartite and monopartite begomoviruses. Virol J. 2012;9:67.

45. Carabali A, Montoya-Lerma J, Bellotti AC. Development and reproduction of *Bemisia tabaci* "B" (Hemiptera : Aleyrodidae) on cassava (Manihot esculenta) genotypes. Revista Colombiana de Entomologia. 2008;34(1):28-32.

46. Carabali A, Belloti AC, Montoya-Lerma J. Biological parameters of *Bemisia tabaci* (Gennadius) biotype B (Hemiptera: Aleyrodidae) on *Jatropha gossypiifolia*, commercial (*Manihot esculenta*) and wild cassava (*Manihot flabellifolia* and *M. carthaginensis*) (Euphorbiaceae). Neotropical Entomology. 2010;39(4):562-7.

47. Carabali A, Bellotti AC, Montoya-Lerma J, Cuellar ME. Adaptation of *Bemisia tabac*i biotype B (Gennadius) to cassava, *Manihot esculenta* (Crantz). Crop Protection. 2005;24(7):643-9.

48. Gómez-Díaz JS, Montoya-Lerma J, Muñoz-Valencia V. Prevalence and diversity of endosymbionts in cassava whiteflies (Hemiptera: Aleyrodidae) from Colombia. Journal of Insect Science. 2019;19(3).

49. Rocha KCG, Marubayashi JM, Navas-Castillo J, Yuki VA, Wilcken CF, Pavan MA, et al. Only the B biotype of *Bemisia tabaci* is present on vegetables in Sao Paulo state, Brazil. Scientia Agricola. 2011;68(1):120-3.

50. Marubayashi JM, Yuki VA, Rocha KCG, Mituti T, Pelegrinotti FM, Ferreira FZ, et al. At least two indigenous species of the *Bemisia tabaci* complex are present in Brazil. Journal of Applied Entomology. 2013;137(1-2):113-21.

51. Marubayashi JM, Kliot A, Yuki VA, Rezende JA, Krause-Sakate R, Pavan MA, et al. Diversity and localization of bacterial endosymbionts from whitefly species collected in Brazil. PloS one. 2014;9(9):e108363.

52. Barbosa LF, Yuki VA, Marubayashi JM, De Marchi BR, Perini FL, Pavan MA, et al. First report of *Bemisia tabaci* Mediterranean (Q biotype) species in Brazil. Pest Management Science. 2015;71(4):501-4.

53. Moraes LA, Marubayashi JM, Yuki VA, Ghanim M, Bello VH, De Marchi BR, et al. New invasion of *Bemisia tabaci* Mediterranean species in Brazil associated to ornamental plants. Phytoparasitica. 2017;45(4):517-25.

54. Moraes LA, Muller C, Bueno R, Santos A, Bello VH, De Marchi BR, et al. Distribution and phylogenetics of whiteflies and their endosymbiont relationships after the Mediterranean species invasion in Brazil. Scientific reports. 2018;8(1):14589.

55. Walsh PS, Metzger DA, Higuchi R. Chelex 100 as a medium for simple extraction of DNA for PCR-based typing from forensic material. Biotechniques. 1991;10(4):506-13.

56. Simon C, Frati F, Beckenbach A, Crespi B, Liu H, Flook P. Evolution, weighting and phylogenetic utility of mitochondrial gene sequences and a compilation of conserved polymerase chain reaction primers. Annals of the Entomological Society of America. 1994;87(6):651-701.

57. Frohlich DR, Torres-Jerez II, Bedford ID, Markham PG, Brown JK. A phylogeographical analysis of the *Bemisia tabaci* species complex based on mitochondrial DNA markers. Molecular Ecology. 1999;8(10):1683-91.

58. Bosco D, Loria A, Sartor C, Cenis JL. PCR-RFLP identification ofBemisia tabaci biotypes in the Mediterranean Basin. Phytoparasitica. 2006;34(3):243.

59. Sambrook J, Russell DW. Standard ethanol precipitation of DNA in microcentrifuge tubes. CSH Protoc. 2006;1(1).

60. Scott IAW, Workman PJ, Drayton GM, Burnip GM. First record of *Bemisia tabaci* biotype Q in New Zealand. New Zealand Plant Protection. 2007;60:264-70.

61. Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, et al. Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. Bioinformatics. 2012;28(12):1647-9.

62. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. Basic local alignment search tool. Journal of molecular biology. 1990;215:403-10.

63. Muhire BM, Varsani A, Martin DP. SDT: A virus classification tool based on pairwise sequence alignment and identity calculation. PloS one. 2014;9:e108277.

64. Edgar RC. MUSCLE: a multiple sequence alignment method with reduced time and space complexity. BMC bioinformatics. 2004;5:1-19.

65. Boykin LM, Savill A, De Barro P. Updated mtCOI reference dataset for the *Bemisia tabaci* species complex. F1000Res. 2017;6:1835. Epub 2017/11/24.

66. Kumar S, Stecher G, Tamura K. MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. Molecular biology and evolution. 2016;33(7):1870-4.

67. Ronquist F, Huelsenbeck JP. MrBayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics. 2003;19(1):1572-4.

68. Nylander JAA. MrModeltest v2. Program distributed by the author Evolutionary Biology Centre, Uppsala University2004.

69. Doyle JJ, Doyle JL. A rapid DNA isolation procedure for small amounts of fresh leaf tissue. Phytochemical Bulletin. 1987;19:11-5.

70. Rojas MR, Gilbertson RL, Russell DR, Maxwell DP. Use of degenerate primers in the polymerase chain reaction to detect whitefly-transmitted geminiviruses. Plant Disease. 1993;77:340-7.

71. Morris EK, Caruso T, Buscot F, Fischer M, Hancock C, Maier TS, et al. Choosing and using diversity indices: insights for ecological applications from the German Biodiversity Exploratories. Ecology and Evolution. 2014;4(18):3514-24. Epub 2014/08/28.

72. R Development Core Team. R: A language and environment for statistical computing: R Foundation for Statistical Computing, Vienna, Austria; 2007.

73. Hodges GS, Evans GA. An iddntification guide to the whiteflies (Hemiptera: Aleyrodidae) of the southeastern United States. Florida Entomologist. 2005;88(4):518-34, 17.

74. Gallet R, Michalakis Y, Blanc S. Vector-transmission of plant viruses and constraints imposed by virus-vector interactions. Curr Opin Virol. 2018;33:144-50. Epub 2018/09/18.

75. Sacristan S, Malpica JM, Fraile A, Garcia-Arenal F. Estimation of population bottlenecks during systemic movement of Tobacco mosaic virus in tobacco plants. Journal of virology. 2003;77(18):9906-11.

76. Gutierrez S, Zwart MP. Population bottlenecks in multicomponent viruses: first forays into the uncharted territory of genome-formula drift. Curr Opin Virol. 2018;33:184-90. Epub 2018/11/25.

77. Elena SF, Fraile A, Garcia-Arenal F. Evolution and emergence of plant viruses. Advances in Virus Research. 2014;88:161-91. Epub 2014/01/01.

78. Dietzgen RG, Mann KS, Johnson KN. Plant virus-insect vector interactions: current and potential future research directions. Viruses. 2016;8(11):303.

79. Fereres A. Insect vectors as drivers of plant virus emergence. Current Opinion in Virology. 2015;10:42-6.

80. Legg JP, Sseruwagi P, Boniface S, Okao-Okuja G, Shirima R, Bigirimana S, et al. Spatio-temporal patterns of genetic change amongst populations of cassava Bemisia tabaci whiteflies driving virus pandemics in East and Central Africa. Virus Research. 2014;186:61-75.

81. Fernandes FR, Albuquerque LC, Giordano LB, Boiteux LS, Ávila AC, Inoue-Nagata AK. Diversity and prevalence of Brazilian bipartite begomovirus species associated to tomatoes. Virus Genes. 2008;36:251-8.

82. Fernandes FR, Albuquerque LC, Oliveira CL, Cruz ARR, Rocha WB, Pereira TG, et al. Molecular and biological characterization of a new Brazilian begomovirus, euphorbia yellow mosaic virus (EuYMV), infecting *Euphorbia heterophylla* plants. Archives of virology. 2011;156(11):2063-9.

83. Albuquerque LC, Inoue-Nagata AK, Pinheiro B, Resende RO, Moriones E, Navas-Castillo J. Genetic diversity and recombination analysis of sweepoviruses from Brazil. Virology Journal. 2012;9:241.

84. Albuquerque LC, Varsani A, Fernandes FR, Pinheiro B, Martin DP, Ferreira PTO, et al. Further characterization of tomato-infecting begomoviruses in Brazil. Archives of virology. 2012;157(4):747-52.

85. Macedo MA, Albuquerque LC, Maliano MR, Souza JO, Rojas MR, Inoue-Nagata AK, et al. Characterization of tomato leaf curl purple vein virus, a new monopartite New World begomovirus infecting tomato in Northeast Brazil. Archives of virology. 2017;163:737-43. Epub 2017/12/11.

86. Castillo-Urquiza GP, Beserra Jr. JEA, Bruckner FP, Lima ATM, Varsani A, Alfenas-Zerbini P, et al. Six novel begomoviruses infecting tomato and associated weeds in Southeastern Brazil. Archives of virology. 2008;153:1985-9.

87. Paz-Carrasco LC, Castillo-Urquiza GP, Lima AT, Xavier CA, Vivas-Vivas LM, Mizubuti ES, et al. Begomovirus diversity in tomato crops and weeds in Ecuador and the detection of a recombinant isolate of rhynchosia golden mosaic Yucatan virus infecting tomato. Archives of virology. 2014;159(8):2127-32.

88. Rodríguez-Negrete EA, Morales-Aguilar JJ, Domínguez-Duran G, Torres-Devora G, Camacho-Beltrán E, Leyva-López NE, et al. High-throughput sequencing reveals differential begomovirus species diversity in non-cultivated plants in Northern-Pacific Mexico. Viruses. 2019;11(7):594.

89. Mar TB, Xavier CAD, Lima ATM, Nogueira AM, Silva JCF, Ramos-Sobrinho R, et al. Genetic variability and population structure of the New World begomovirus *Euphorbia yellow mosaic virus*. Journal of General Virology. 2017;98(6):1537-51. Epub 2017/06/15.

90. Ghosh S, Bouvaine S, Maruthi MN. Prevalence and genetic diversity of endosymbiotic bacteria infecting cassava whiteflies in Africa. BMC Microbiology. 2015;15(1):93.

91. Tocko-Marabena BK, Silla S, Simiand C, Zinga I, Legg J, Reynaud B, et al. Genetic diversity of *Bemisia tabaci* species colonizing cassava in Central African Republic characterized by analysis of cytochrome c oxidase subunit I. PloS one. 2017;12(8).

92. Alemandri V, Vaghi Medina CG, Dumon AD, Arguello Caro EB, Mattio MF, Garcia Medina S, et al. Three members of the *Bemisia tabaci* (Hemiptera: Aleyrodidae) cryptic species complex occur sympatrically in Argentine horticultural crops. Journal of Economic Entomology. 2015;108(2):405-13.

93. Liu BM, Yan FM, Chu D, Pan HP, Jiao XG, Xie W, et al. Difference in feeding behaviors of two invasive whiteflies on host plants with different suitability: implication for competitive displacement. International Journal of Biological Sciences. 2012;8(5):697-706.

94. Sun D-B, Liu Y-Q, Qin L, Xu J, Li F-F, Liu S-S. Competitive displacement between two invasive whiteflies: insecticide application and host plant effects. Bulletin of Entomological Research. 2013;103(3):344-53. Epub 2013/03/05.

95. Watanabe LFM, Bello VH, De Marchi BR, Silva FBd, Fusco LM, Sartori MM, et al. Performance and competitive displacement of *Bemisia tabaci* MEAM1 and MED cryptic species on different host plants. Crop Protection. 2019;124:104860.

96. Nassar N. Cassava, *Manihot esculenta* Crantz and wild relatives: their relationships and evolution. Genetic Resources and Crop Evolution. 2001;48(5):429-36.

97. Olsen KM. SNPs, SSRs and inferences on cassava's origin. Plant Molecular Biology. 2004;56(4):517-26.

98. Alonso RdS, Racca-Filho F, Lima AFd. Occurrence of whiteflies (Hemiptera: Aleyrodidae) on cassava (*Manihot esculenta* Crantz) crops under field conditions in the state of Rio de Janeiro, Brazil. EntomoBrasilis. 2012;5(1):2. Epub 2012-04-02.

99. Dickey AM, Stocks IC, Smith T, Osborne L, McKenzie CL. DNA Barcode development for three recent exotic whitefly (Hemiptera: Aleyrodidae) invaders in Florida. Florida Entomologist. 2015;98(2):473-8, 6.

100. Luan J-b, Xu J, Lin K-k, Zalucki MP, Liu S-s. Species exclusion between an invasive and an indigenous whitefly on host plants with differential levels of suitability. Journal of Integrative Agriculture. 2012;11(2):215-24.

101. Tajebe LS, Boni SB, Guastella D, Cavalieri V, Lund OS, Rugumamu CP, et al. Abundance, diversity and geographic distribution of cassava mosaic disease pandemic-associated *Bemisia tabaci* in Tanzania. Journal of Applied Entomology. 2015;139(8):627-37.

102. Vyskočilová S, Seal S, Colvin J. Relative polyphagy of "Mediterranean" cryptic *Bemisia tabaci* whitefly species and global pest status implications. Journal of Pest Science. 2019;92(3):1071-88.

103. Milenovic M, Wosula EN, Rapisarda C, Legg JP. Impact of host plant species and whitefly species on feeding behavior of *Bemisia tabaci*. Frontiers in Plant Science. 2019;10(1):1.

104. Wilson AK. *Euphorbia heterophylla*: A review of distribution, importance and control. Tropical Pest Management. 1981;27:32-8.

105. Sottoriva LDM, Lourenção AL, Colombo CA. Performance of *Bemisia tabaci* (Genn.) biotype B (Hemiptera: Aleyrodidae) on weeds. Neotropical Entomology. 2014;43(6):574-81.

106. Barreto SS, Hallwass M, Aquino OM, Inoue-Nagata AK. A study of weeds as potential inoculum sources for a tomato-infecting begomovirus in central Brazil. Phytopathology. 2013;103(5):436-44. Epub 2013/03/16.

107. Mar TB, Mendes IR, Lau D, Fiallo-Olive E, Navas-Castillo J, Alves MS, et al. Interaction between the New World begomovirus *Euphorbia yellow mosaic virus* and its associated alphasatellite: effects on infection and transmission by the whitefly *Bemisia tabaci*. Journal of General Virology. 2017;98(6):1552-62. Epub 2017/06/08.

108. Hadjistylli M, Roderick GK, Brown JK. Global population structure of a worldwide pest and virus vector: Genetic diversity and population history of the *Bemisia tabaci* sibling species group. PloS one. 2016;11(11):e0165105.

824 **Table 1.** Sampled sites and whiteflies species detected in cassava in Brazil.

Sample	Date of collection	Location	Region	Geographical coordinates		Number of samples			Whiteflies species[&]	Reference
				Latitude	Longitude	Adults	Nymphs	Total		
AL1	July 2018	Arapiraca, AL	Northeast	09° 48' 27.66"S	36° 36' 40.68"W	15	15	30	Btu, Ta	This study
AL2	April 2018	TeotonioVilela, AL	Northeast	09° 53' 16.86"S	36° 23' 05.52"W	14	15	29	BtM, Btu, Ta	This study
AL3	April 2018	União dos Palmares, AL	Northeast	09° 11' 51.84"S	36° 01"51.90"W	16	15	31	BtM, Ta	This study
AL4	April 2018	Arapiraca, AL	Northeast	09° 47' 29.46"S	36° 25' 36.00"W	15	15	30	BtM, BtNW, Btu, Ta	This study
AL5	April 2018	Arapiraca, AL	Northeast	09° 43' 09.30"S	36° 40' 45.60"W	14	10	24	BtM, BtNW, Btu, Ta	This study
BA1	December 2017	Barra, BA	Northeast	11° 20' 52.37"S	43° 12' 57.44"W	28	0	28	BtM, Btu, Ta	This study
BA2	March 2018	LEM, BA	Northeast	12° 20' 10.00"S	45° 49' 12.00"W	13	0	13	BtM, Ta	This study
BA3	March 2018	Wanderley, BA	Northeast	12° 13' 42.04"S	43° 55' 50.00"W	10	10	20	BtM, Btu	This study
BA4	June 2018	Cristópolis, BA	Northeast	12° 13' 56.08"S	44° 23' 11.50"W	14	0	14	BtM, Ta	This study
DF1	March 2018	Planaltina, DF	Midwest	15° 30' 59.00"S	47° 16' 09.00"W	10	0	10	BtM	This study
DF2	March 2018	Planaltina, DF	Midwest	15° 28' 57.00"S	47° 20' 06.00"W	10	0	10	BtM	This study
DF3	March 2021	Planaltina, DF	Midwest	15° 31' 17.00"S	47° 21' 22.00"W	10	0	10	BtM	This study
ES1	January 2018	Sooretama, ES	Southeast	19° 06' 52.02"S	40° 04' 46.30"W	14	15	29	BtM, Ta	This study
ES2	January 2018	Marilândia, ES	Southeast	19° 24' 22.04"S	40° 32' 21.70"W	15	15	30	BtM, Btu, Ta	This study
ES3	January 2018	Pinheiros, ES	Southeast	18° 40' 83.20"S	40° 28' 63.40"W	11	11	22	Ta	This study
GO1	March 2018	Bela Vista, GO	Midwest	16° 59' 50.00"S	48° 57' 56.00"W	10	11	21	BtM	This study
GO2	March 2018	Itaberaí, GO	Midwest	15° 56' 58.00"S	49° 47' 07.00"W	15	15	30	BtM, Btu	This study
MG1	May 2018	Ouro Fino, MG	Southeast	22° 16' 44.00"S	46° 29' 33.00"W	12	10	22	Ta	This study
MG10	February 2018	Florestal, MG	Southeast	19° 52' 38.00"S	44° 25' 21.00"W	15	15	30	Btu, Ta	This study
MG11	March 2018	Florestal, MG	Southeast	19° 52' 38.00"S	44° 25' 21.00"W	15	15	30	Btu, Ta	This study
MG12	May 2018	Divinópolis, MG	Southeast	20° 06' 21.00"S	44° 55' 36.00"W	15	15	30	Btu, Ta	This study
MG13	August 2018	Viçosa, MG	Southeast	20° 46' 06.00"S	42° 52' 14.00"W	17	0	17	Btu, Ta	This study
MG14	March 2018	Piraúba, MG	Southeast	21° 16' 22.71"S	43° 02' 31.28"W	10	10	20	Ta	This study
MG15	March 2018	Descoberto, MG	Southeast	21° 28' 06.28"S	42° 58' 05.53"W	10	10	20	Ta	This study
MG16	March 2018	Mar de Espanha, MG	Southeast	21° 46' 07.26"S	43° 04' 26.62"W	15	15	30	BtM, Ta	This study

MG17	March 2018	Leopoldina, MG	Southeast	21° 33' 50.15"S	42° 40' 42.97"W	16	13	29	BtM, Btu, Ta	This study
MG18	March 2018	Dona Euzébia, MG	Southeast	21° 19' 18.83"S	42° 48' 37.07"W	15	14	29	Ta	This study
MG19	February 2018	Caparaó, MG	Midwest	20° 31' 48.00"S	41° 54' 00.36"W	15	15	30	BtM, Ta	This study
MG2	July 2018	Pouso Alegre, MG	Southeast	22° 15' 01.00"S	46° 58' 31.00"W	9	0	9	Ta	This study
MG3	February 2018	Careaçu, MG	Southeast	22° 04' 37.00"S	45° 41' 49.00"W	11	11	22	Ta	This study
MG4	June 2018	Lambari, MG	Southeast	21° 56' 05.00"S	45° 15' 49.00"W	15	15	30	Btu, Ta	This study
MG5	February 2018	Lima Duarte, MG	Southeast	21° 50' 35.00"S	43° 47' 01.00"W	10	10	20	Ta	This study
MG6	April 2018	RioPomba, MG	Southeast	21° 15' 50.00"S	43° 09' 59.00"W	10	0	10	WtNEW2, Btu	This study
MG7	February 2018	Florestal, MG	Southeast	19° 54' 13.00"S	44° 25' 48.00"W	15	15	30	BtM, Btu, Ta	This study
MG8	February 2018	Florestal, MG	Southeast	19° 51' 20.00"S	44° 23' 58.00"W	15	15	30	Btu, Btu-like	This study
MG9	March 2018	Florestal, MG	Southeast	19° 53' 39.00"S	44° 24' 55.00"W	14	11	25	BtM, BtNW, Btu, Ta	This study
MT1	December 2017	Canarana, MT	Midwest	13° 31' 16.00"S	52° 25' 03.00"W	15	16	31	WtNEW1	This study
MT2	December 2017	Canarana, MT	Midwest	13° 33' 47.00"S	52° 15' 53.00"W	0	20	20	Btu	This study
MT4	January 2018	Pedra Preta, MT	Midwest	16° 38' 29.00"S	54° 25' 41.00"W	10	10	20	Btu	This study
MT5	January 2018	Pedra Preta, MT	Midwest	16° 39' 07.00"S	54° 22' 27.00"W	10	10	20	BtM, Btu	This study
MT6	January 2018	Pedra Preta, MT	Midwest	16° 39' 28.00"S	54° 20' 20.00"W	10	10	20	BtM, Btu	This study
PA1	January 2018	Brasil Novo, PA	North	03° 12' 23.07"S	52° 30' 13.80"W	15	15	30	Btu, Ta	This study
PA2	January 2018	Vitória doXingu, PA	North	03° 04' 51.01"S	52° 10' 08.80"W	15	16	31	Btu, Ta	This study
PA3	August 2018	Altamira, PA	North	03° 18' 15.03"S	52° 07' 26.00"W	10	10	20	BtM	This study
PA4	January 2018	Altamira, PA	North	03° 09' 14.60"S	52° 07' 49.00"W	11	10	21	Ta	This study
PA5	January 2018	Belém, PA	North	01° 18' 20.00"S	48° 26' 46.00"W	10	12	22	Ta	This study
PI1	April 2018	Picos, PI	Northeast	07° 04' 79.50"S	41° 25' 57.80"W	0	29	29	Btu, Ta	This study
PI2	May 2018	Teresina, PI	Northeast	05° 02' 41.94"S	42° 47' 18.84"W	0	27	27	Btu, Ta	This study
PR1	March 2018	Santo Antôniodo Caiuá, PR	South	22° 41' 07.00"S	52° 19' 06.00"W	30	0	30	Btu, Ta	This study
PR2	March 2018	Santo Antôniodo Caiuá, PR	South	22° 49' 03.00"S	52° 21' 25.00"W	30	0	30	BtM, Btu	This study
PR3	March 2018	Paranavaí, PR	South	23° 06' 03.29"S	52° 29' 10.00"W	10	10	20	BtM, BtMED	This study
PR4	March 2018	Sertanópolis, PR	South	23° 02' 54.00"S	50° 59' 54.00"W	26	0	26	Btu, Ta	This study

SC2	March 2018	Agrônomica, SC	South	27° 34' 40.00"S	48° 32' 08.00"W	10	0	10	BtM	This study
SP1	July 2016	Holambra, SP	Southeast	22° 36' 26.00"S	47° 02' 50.00"W	10	0	10	Btu	Moraes et al., 2018
SP10	March 2016	Casa Branca, SP	Southeast	21° 49' 08.00"S	45° 58' 23.00"W	10	0	10	BtM	Moraes et al., 2018
SP10	July 2016	São Pedro do Turvo, SP	Southeast	22° 36' 32.00"S	49° 45' 29.00"W	10	0	10	BtMED	Moraes et al., 2018
SP11	July 2016	Oleo, SP	Southeast	22° 56' 32.00"S	49° 26' 15.00"W	10	0	10	Btu	Moraes et al., 2018
SP12	July 2016	Mogi Mirim, SP	Southeast	22° 28' 32.30"S	47° 00' 47.60"W	10	0	10	Btu	Moraes et al., 2018
SP2	July 2016	Mogi Mirim, SP	Southeast	22° 28' 08.00"S	46° 56' 25.00"W	10	0	10	Btu	Moraes et al., 2018
SP3	July 2016	Mogi Mirim, SP	Southeast	22° 24' 59.00"S	46° 59' 19.00"W	10	0	10	Btu	Moraes et al., 2018
SP4	February 2019	Mogi Mirim, SP	Southeast	22° 26' 44.00"S	47° 04' 11.00"W	5	5	10	Btu	This study
SP5	July 2017	Mogi Mirim, SP	Southeast	22° 27' 05.00"S	47° 04' 56.00"W	10	0	10	Btu	Moraes et al., 2018
SP6	January 2019	SãoPedro, SP	Southeast	22° 34' 08.00"S	48° 05' 22.00"W	10	0	10	BtMED	This study
SP7	July 2017	Montalvao, SP	Southeast	22° 02' 23.00"S	51° 19' 53.00"W	10	0	10	BtM	Moraes et al., 2018
SP8	July 2016	Pindamonhangaba, SP	Southeast	22° 56' 05.00"S	45° 26' 25.00"W	10	0	10	Btu	Moraes et al., 2018
SP9	January 2019	Casa Branca, SP	Southeast	21° 11' 32.00"S	47° 48' 44.00"W	4	10	14	BtM, Btu, BtMED	This study
Total						819	566	1385		

[*] Brazilian states where samples were collected: AL, Alagoas; BA, Bahia; DF, Distrito Federal; ES, Espírito Santo; GO, Góias; MG, Minas Gerais; MT, Mato Grosso; PA, Pará; PI, Piauí; PR, Paraná; SC, Santa Catarina; SP, São Paulo.

[&] Btu, *Bemisia tuberculata*; BtM, *Bemisia tabaci* MEAM1; BtMED, *B. tabaci* MED; BtNW, *B. tabaci* NW; Ta, *Tetraleuroides acaciae*; WtNEW2, whitefly new specie 1; WtNEW2, whitefly new specie 2.

Figure legends

Figure 1. A. Clockwise from top-left: Adults and nymphs of *Bemisia tuberculata* colonizing cassava in Mogi Mirim, São Paulo state. Growth of sooty mould fungus on the leaf surface due to the deposition of honeydew by whiteflies. Presence of begomovirus-infected *Blainvillea rhomboidea* (family Asteraceae) in a cassava field in Minas Gerais state. Presence of begomovirus-infected *Euphorbia heterophylla* (family Euphorbiaceae) in a cassava field in Minas Gerais state. **B.** Map of Brazil showing the locations where whiteflies samples were collected. The map is colored according to the regions as indicated in the legend. Blacks dots correspond to the sampled sites. Scale bar is only for Brazil map. **C.** Number of adults and nymphs analyzed from each sampled site according to state. **D.** Specie distribution according to region. AL, Alagoas; BA, Bahia; DF, Distrito Federal; ES, Espírito Santo; GO, Goiás; MG, Minas Gerais; MT, Mato Grosso; PA, Pará; Piauí; PR, Paraná; SC, Santa Catarina; SP, São Paulo.

Figure 2. Bayesian phylogenetic tree based on partial nucleotide sequences of the mitochondrial cytochrome oxidase (mtCOI) gene of representative individuals of each whitefly species detected in this study and reference sequences retrieved from GenBank. The tree was rooted with the aphid *Aphis gossypii.* Bayesian posterior probabilities are shown at the nodes. The scale bar represents the number of nucleotide substitutions per site. Nodes with posterior probability values between 0.60 and 0.80 are indicated by empty circles and nodes with values equal to or greater than 0.81 are indicated by filled circles. Clades highlighted with different colors indicate the species detected in this study. Branches highlighted in red indicate the putative new species detected here.

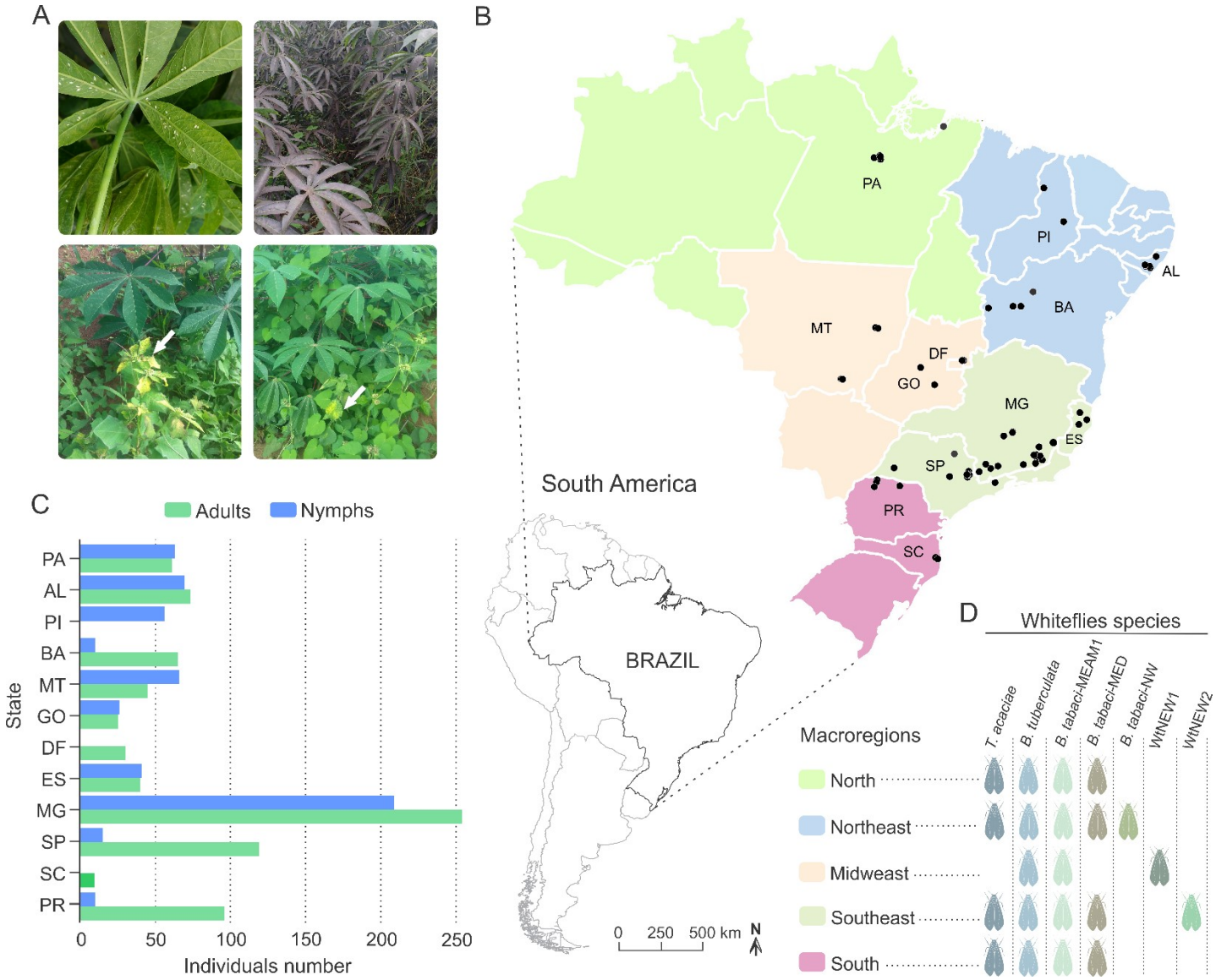
Figure 3. Composition of whitefly populations colonizing cassava in Brazil. **A.** Species composition at each sampled site according to stage of development (adult and nymphs). Asterisks

indicate that nymphs were not detected. **B, C, D.** Species distribution of the 1,385 individuals genotyped in this study considering the samples from all sites (**B**) or only samples from sites where both adults and nymphs were sampled (**C**) or without samples from Minas Gerais state (**D**).

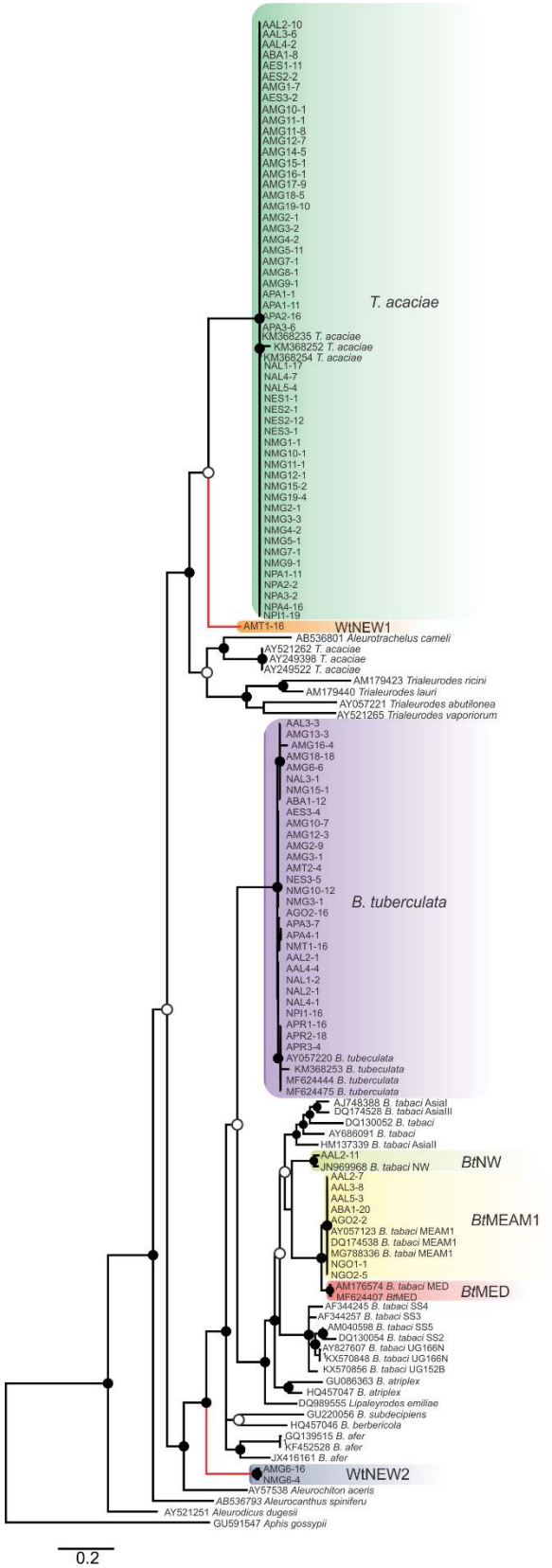
Figure 4. A. Incidence of *Trialeurodes acaciae*, *Bemisia tuberculata* and *B. tabaci* MEAM1 in cassava fields in Brazil, measured as the percentage of sampled sites where at least one individual belonging to each one of the three species was detected. Other species detected at low incidence are shown together as "others". **B.** Venn diagram showing the proportion of sites where each one of the three whitefly species occur alone or in different combinations. **C.** Competitive capacity inferred based on the prevalence of individuals from each of two species in fields where those two species were detected co-occurring. The horizontal line inside the box corresponds to the median. The asterisk indicates a significant difference according to the non-parametric Kruskal-Wallis test ($p<0.05$).

Figure 5. Composition and species diversity of whitefly populations differ among Brazilian regions. **A.** Pie charts represent the distribution of the 1,385 individuals genotyped in this study in the five geographic regions of Brazil. **B.** Boxplots correspond to Simpson's index of diversity (1-D) calculated for each geographic region. The index was first calculated for each sampled site and grouped by geographic region. Different letters indicate significant differences between groups according to the non-parametric Kruskal-Wallis test followed by *post hoc* multiple comparison test ($p<0.05$).

Figure 1

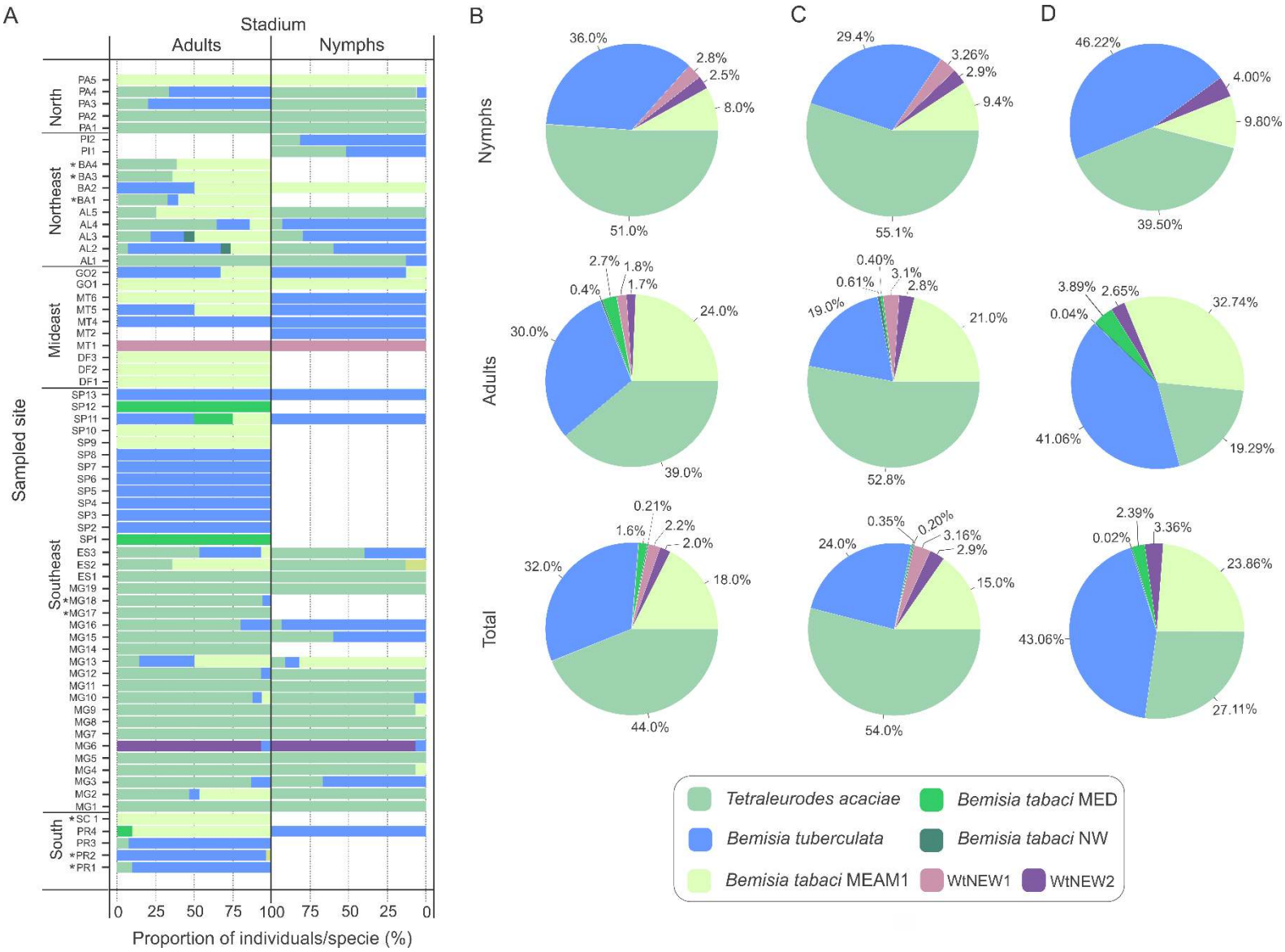

A
B
C
D
PA
PI
AL
BA
MT
DF
GO
MG
ES
SP
PR
SC
South America
BRAZIL
0 250 500 km
N
Adults
Nymphs
State
Individuals number
0
50
100
150
200
250
Whiteflies species
T. acaciae
B. tuberculata
B. tabaci-MEAM1
B. tabaci-MED
B. tabaci-NW
VvtNEW1
VvtNEW2
Macroregions
North
Northeast
Midweast
Southeast
South

872 **Figure 2**

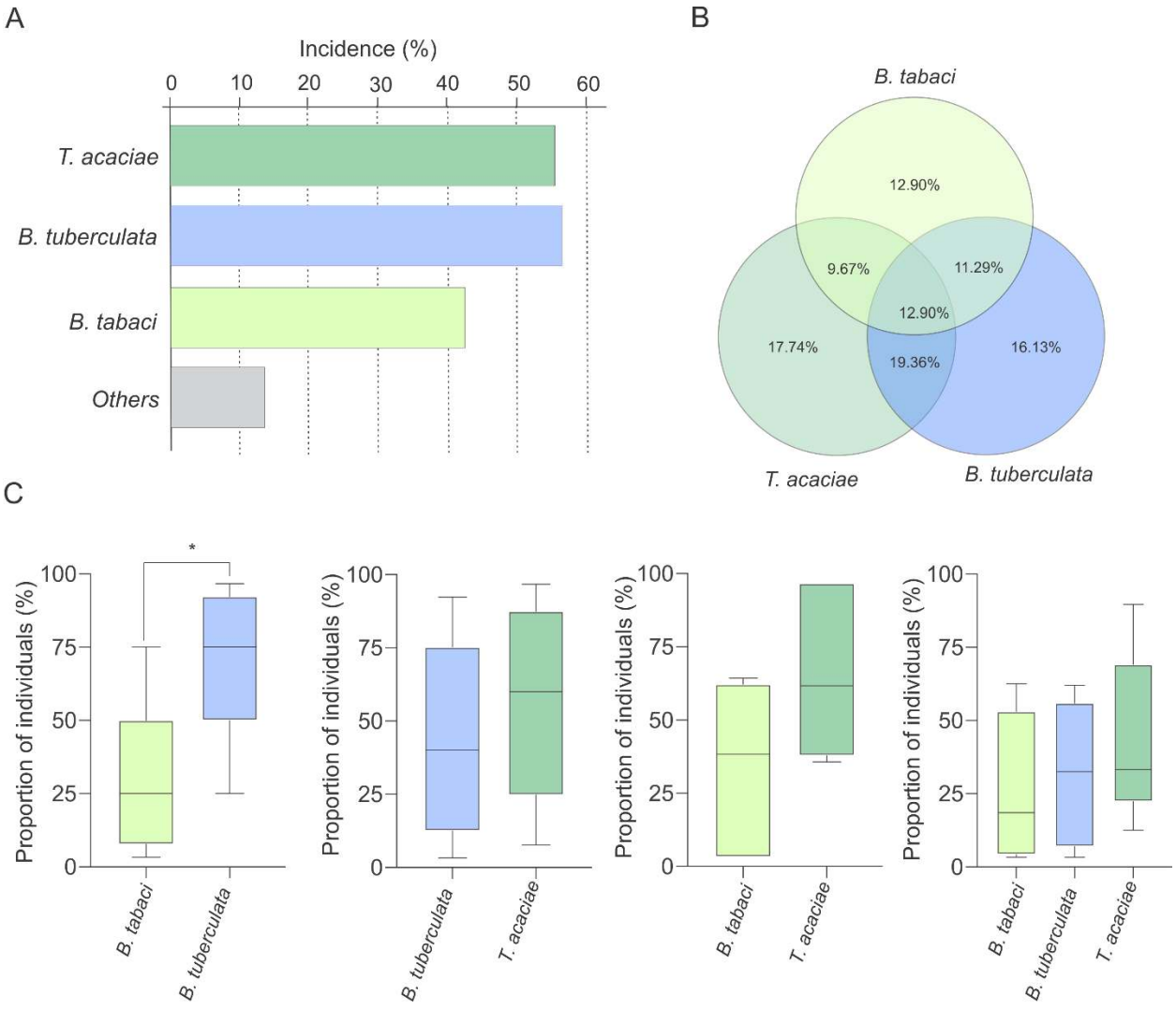


873

Figure 3

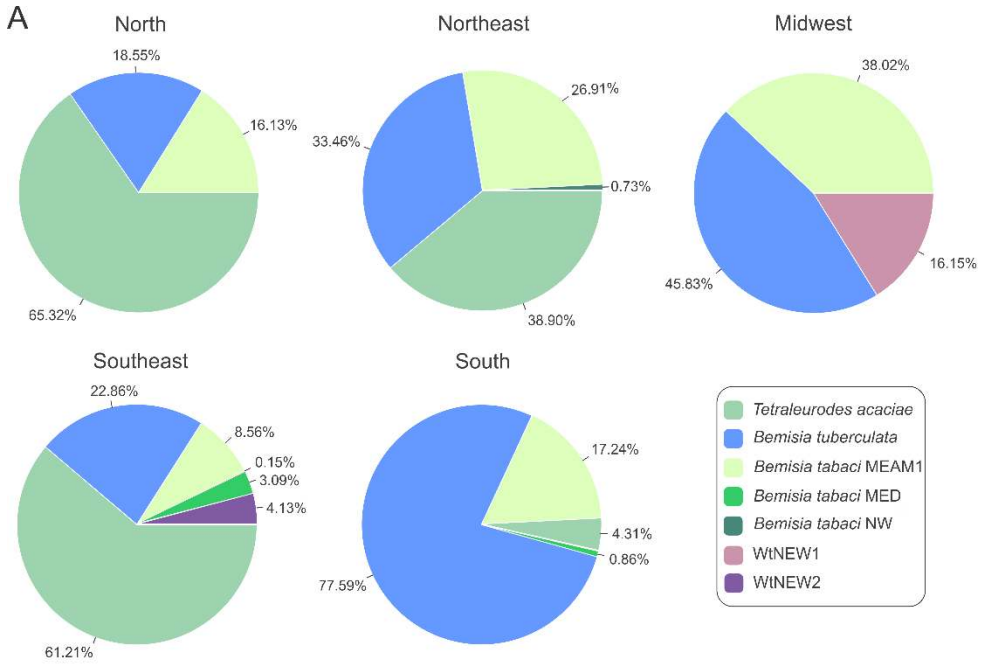

A
Stadium
Adults
Nymphs
Sampled site
North
Northeast
Mideast
Southeast
South
PA5
PA4
PA3
PA2
PA1
PI2
PI1
*BA4
*BA3
BA2
*BA1
AL5
AL4
AL3
AL2
AL1
GO2
GO1
MT6
MT5
MT4
MT2
MT1
DF3
DF2
DF1
SP13
SP12
SP11
SP10
SP9
SP8
SP7
SP6
SP5
SP4
SP3
SP2
SP1
ES3
ES2
ES1
MG19
*MG18
*MG17
MG16
MG15
MG14
MG13
MG12
MG11
MG10
MG9
MG8
MG7
MG6
MG5
MG4
MG3
MG2
MG1
*SC 1
PR4
PR3
*PR2
*PR1
0 25 50 75 100 75 50 25 0
Proportion of individuals/specie (%)
B
C
D
Nymphs
Adults
Total
36.0%
2.8%
2.5%
8.0%
51.0%
29.4%
3.26%
2.9%
9.4%
55.1%
46.22%
4.00%
9.80%
39.50%
2.7%
1.8%
0.4%
1.7%
24.0%
30.0%
39.0%
0.40%
3.1%
0.61%
2.8%
19.0%
21.0%
52.8%
3.89%
2.65%
0.04%
32.74%
41.06%
19.29%
0.21%
1.6%
2.2%
2.0%
32.0%
18.0%
44.0%
0.35%
0.20%
3.16%
24.0%
2.9%
15.0%
54.0%
2.39%
3.36%
0.02%
23.86%
43.06%
27.11%
Tetraleurodes acaciae
Bemisia tabaci MED
Bemisia tuberculata
Bemisia tabaci NW
Bemisia tabaci MEAM1
WtNEW1
WtNEW2

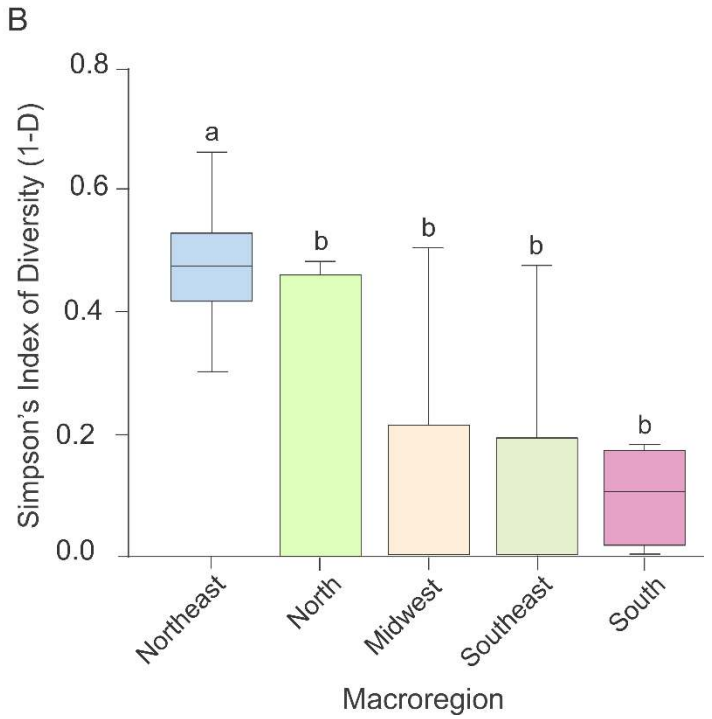
876 **Figure 4**



877

878 **Figure 5**





879