

UC San Diego

UC San Diego Previously Published Works

Title

Assessment of a Split Homing Based Gene Drive for Efficient Knockout of Multiple Genes.

Permalink

<https://escholarship.org/uc/item/4vz2z6rs>

Journal

G3 (Bethesda, Md.), 10(2)

ISSN

2160-1836

Authors

Kandul, Nikolay P
Liu, Junru
Buchman, Anna
et al.

Publication Date

2020-02-01

DOI

10.1534/g3.119.400985

Peer reviewed

Assessment of a Split Homing Based Gene Drive for Efficient Knockout of Multiple Genes

Nikolay P. Kandul,* Junru Liu,* Anna Buchman,* Valentino M. Gantz,* Ethan Bier*,[†] and

Omar S. Akbari*,^{†,1}

*Section of Cell and Developmental Biology and [†]Tata Institute for Genetics and Society, University of California, San Diego, La Jolla, CA 92093

ORCID ID: 0000-0002-6853-9884 (O.S.A.)

ABSTRACT Homing based gene drives (HGD) possess the potential to spread linked cargo genes into natural populations and are poised to revolutionize population control of animals. Given that host encoded genes have been identified that are important for pathogen transmission, targeting these genes using guide RNAs as cargo genes linked to drives may provide a robust method to prevent disease transmission. However, effectiveness of the inclusion of additional guide RNAs that target separate genes has not been thoroughly explored. To test this approach, we generated a split-HGD in *Drosophila melanogaster* that encoded a drive linked effector consisting of a second gRNA engineered to target a separate host-encoded gene, which we term a gRNA-mediated effector (GME). This design enabled us to assess homing and knockout efficiencies of two target genes simultaneously, and also explore the timing and tissue specificity of Cas9 expression on cleavage/homing rates. We demonstrate that inclusion of a GME can result in high efficiency of disruption of both genes during super-Mendelian propagation of split-HGD. Furthermore, both genes were knocked out one generation earlier than expected indicating the robust somatic expression of Cas9 driven by *Drosophila* germline-limited promoters. We also assess the efficiency of ‘shadow drive’ generated by maternally deposited Cas9 protein and accumulation of drive-induced resistance alleles along multiple generations, and discuss design principles of HGD that could mitigate the accumulation of resistance alleles while incorporating a GME.

KEYWORDS

CRISPR
Cas9
Homing
split-HGD
Drosophila melanogaster
resistance allele

For standard Mendelian inheritance, any particular allele has a 50% chance in being transmitted to its offspring. While mechanisms of meiosis generally bias selection against violators of Mendel’s rules, there are many examples of naturally occurring selfish genetic elements (SGEs) that succeed in bypassing these rules. These SGEs enhance, or “drive” their transmission into subsequent generations, despite often times being harmful to the harboring individual (*i.e.*, imposing a fitness load). These include, for example, transposable elements (TEs), meiotic drivers, B chromosomes, post segregation

killers, heritable microbes, and homing endonuclease genes (Werren *et al.* 1988; Burt and Trivers 2006; Werren 2011; McLaughlin and Malik 2017). Drawing inspiration from these natural systems, strategies for exploiting drive to alter the genetics of wild pest populations have been proposed (Werren *et al.* 1988; Burt, 2003; Burt and Trivers 2006; Werren 2011; Esvelt *et al.* 2014; Champer *et al.* 2016; McLaughlin and Malik 2017), and some have even been experimentally tested in the laboratory, however none have been implemented in the field. For those tested in the laboratory, some examples include synthetic *Medea* elements (Chen *et al.* 2007; Akbari *et al.* 2014; Buchman *et al.* 2018a), engineered underdominance systems (Akbari *et al.* 2013; Buchman *et al.* 2018b), and those whose development was accelerated by the CRISPR revolution (Jinek *et al.* 2012; Cong *et al.* 2013; Mali *et al.* 2013) including toxin-antidote based systems (Oberhofer *et al.* 2019), and homing based gene drive systems (HGDs) (Esvelt *et al.* 2014; Gantz and Bier 2016; Champer *et al.* 2016; Marshall and Akbari 2018).

HGDs are perhaps the furthest along in development, and have already been tested in a broad range of organisms spanning

Copyright © 2020 Kandul *et al.*

doi: <https://doi.org/10.1534/g3.119.400985>

Manuscript received October 25, 2019; accepted for publication December 19, 2019; published Early Online December 27, 2019.

This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Supplemental material available at figshare: <https://doi.org/10.25387/g3.11449542>.

¹Corresponding author: 5101 TATA hall, 9500 Gilman Dr., University of California, San Diego, La Jolla, CA 92093. E-mail: oakbari@ucsd.edu

bacteria, yeast, insects, and mammals (Windbichler *et al.* 2011; Gantz and Bier 2015; DiCarlo *et al.* 2015; Gantz *et al.* 2015; Hammond *et al.* 2016, 2018; Champer *et al.* 2017, 2018; KaramiNejadRanjbar *et al.* 2018; Kyrou *et al.* 2018; Yan and Finnigan 2018; Li *et al.* 2019; Grunwald *et al.*, 2019, Valderrama *et al.*, 2019). They function by encoding the Cas9 endonuclease and an independently expressed guide RNA (gRNA) responsible for mediating DNA/RNA base pairing and cleavage at a predetermined site (Esvelt *et al.* 2014; Gantz and Bier 2015, 2016; Champer *et al.* 2016; Marshall and Akbari 2018). When the HGD is positioned within its target site in a heterozygote, double stranded DNA breaks (DSBs) on the opposite chromosome can result in the drive allele being used as a template (*i.e.*, donor chromosome) for DNA repair mediated by homologous recombination. This can result in copying, or “homing,” of the HGD into the broken chromosome (*i.e.*, receiver chromosome), thereby converting heterozygotes to homozygotes in the germline, which can bias Mendelian inheritance ratios and result in an increase in HGD frequency in a population.

Given the recent progress toward developing HGDs in pest species such as mosquitoes (Gantz *et al.* 2015; Hammond *et al.* 2016, 2018; Kyrou *et al.* 2018; Li *et al.* 2019), there is significant enthusiasm regarding their potential use to control wild populations. For example, given the enormous burden mosquitoes pose on humans, the release of HGDs linked with effector genes inhibiting mosquito pathogen transmission (Isaacs *et al.* 2011; Jupatanakul *et al.* 2017; Buchman *et al.* 2019a; b) may lead to replacement of disease-susceptible mosquitoes with disease-resistant counterparts resulting in reduced pathogen transmission (*i.e.*, population modification drive). Alternatively, HGDs targeting genes affecting the fitness of female mosquitoes could also spread, resulting in gradual population declines and potentially even elimination (*i.e.*, population suppression drive) (Windbichler *et al.* 2008, 2011; Kyrou *et al.* 2018). Given these features, both modification and suppression drives possess the potential to transform mosquito population control measures (Burt 2003; Esvelt *et al.* 2014; Gantz and Bier 2016; Champer *et al.* 2016), and therefore have excited significant ongoing discussions involving their potential usage, regulation, safety, ethics and governance (Oye *et al.* 2014; Akbari *et al.* 2015; National Academies of Sciences, Engineering, and Medicine *et al.* 2016; Adelman *et al.* 2017). Despite these exciting developments however, the elephant in the room persists - can a gene drive actually work in the wild? There are a number of open questions looming as to the efficiency of HGDs. For example, can a drive spread to fixation in the wild? Will it simply breakdown due to resistance? Will the linked anti-pathogen effector work efficiently given the expected diversity of parasites/virus genomes found in the wild? Can the pathogen evolve to become resistant to the anti-pathogen effector and perhaps even become more virulent (Marshall *et al.* 2019)? These are just a minority of legitimate concerns regarding the potential use of a gene drive that would need to be resolved prior to any release.

While many questions loom, there has been some effort to resolve these concerns safely in the lab. For example, with regard to the HGD breakdown due to resistance, multiple studies have explored design criteria attempting to suppress the effects of resistance alleles on drive propagation. For example, some studies have had some success using germline-restricted promoters to express Cas9 increasing rates of HDR, resulting in increased homing rates, as opposed to error-prone pathways such as non-homologous end joining (NHEJ) which results in the generation of resistance alleles (Hammond *et al.* 2018; Champer *et al.* 2018). Other studies have described (Esvelt *et al.* 2014; Champer *et al.* 2016; Marshall *et al.* 2017) and tested (Champer *et al.* 2018, 2019a; b; Oberhofer *et al.* 2018) multiplexed gRNAs in drives resulting

in moderate increases in drive efficacy. While others have had some success targeting highly conserved recessive fertility/viability genes whose homozygous mutants are inviable, or cannot reproduce, and therefore are expected to not affect the spread of HGDs (Hammond *et al.* 2016; KaramiNejadRanjbar *et al.* 2018; Kyrou *et al.* 2018; Oberhofer *et al.* 2018). However, despite these efforts, resistance alleles are still problematic, leaving open the question as to what is the best method to prevent their generation.

Here, to further explore this paramount issue of resistance to HGD we use *Drosophila melanogaster* as our model. We use a genetic safeguarded split-drive design as a safety feature and also encode a linked effector to the drive. This effector consisted of a second gRNA engineered to target a separate host encoded gene which we term a gRNA-mediated effector (GME) (Figure 1). Given that there are many host-encoded genes that are important for pathogen transmission (Cheng *et al.* 2016; Dong *et al.* 2018), one potential application of a HGD is to incorporate a cargo GME that targets a host encoded factor that is important for some aspect of pathogen transmission. If the GME is effective, then disruption of its target in the population should in principle occur as the drive spreads, thereby immunizing that population from pathogen transmission. Therefore, encoding a GME in a drive may be a useful feature going forward and worth further exploring. As a proof of concept to test the efficiency of a HGD linked GME, we designed both the drive and effector to target phenotypic genes which resulted in easily scorable recessive viable phenotypes. This novel drive architecture enabled us to test many germline Cas9 expressing promoters, while simultaneously measuring homing and cleavage efficiencies in both the germline and soma for both target genes over successive generations. While homing rates were modest, cleavage rates were high. For example, we determined that we can reproducibly achieve complete penetrance of somatic mosaic phenotypes for both target genes with up to 100% efficiency stemming from a combination of Cas9 maternal deposition and somatic expression. However, despite the robust cleavage efficiencies and impressive efficacy of the HGD linked GME, drive resistance alleles were still generated which would hinder spread. Given these results, alternative design principles are proposed that could potentially mitigate these issues while also incorporating a drive linked GME.

MATERIALS AND METHODS

Design and assembly of constructs

The genetic assembly of the Gene Drive element (*GDe*) with two gRNAs and 3xP3-eGFP (Figure 1A) was previously described to generate a split *trans*-complementing Gene Drive system (Lopez del Amo *et al.* 2019). The assembly of BicC-Cas9 construct followed the same steps previously described for the other three Cas9 lines: *nos-Cas9*, *vas-Cas9*, and *Ubi-Cas9* (Kandul *et al.* 2019). The 2831 bases upstream of BicC-RA's start codon (*Bicaudal C*, CG4824) was PCR amplified with CGACGGTCACGGCGGGCA-TGTCGACGCGGCCGCATAATTATATAATAATAAACTGCATGC (BicC-F) and TCCGTCTGGTCTTATAGTCCATGTTTAAAC-TGTGGAATTCGGATGATGATGATGATC (BicC-R) from *Drosophila melanogaster* genome, and enzymatically assembled (Gibson *et al.* 2009) into *Ubi-Cas9* plasmid (addgene #112686) (Kandul *et al.* 2019) digested with NotI and XhoI.

Fly genetics and imaging

Flies were maintained under standard conditions at 25°. Embryo injections were carried at Rainbow Transgenic Flies, Inc.

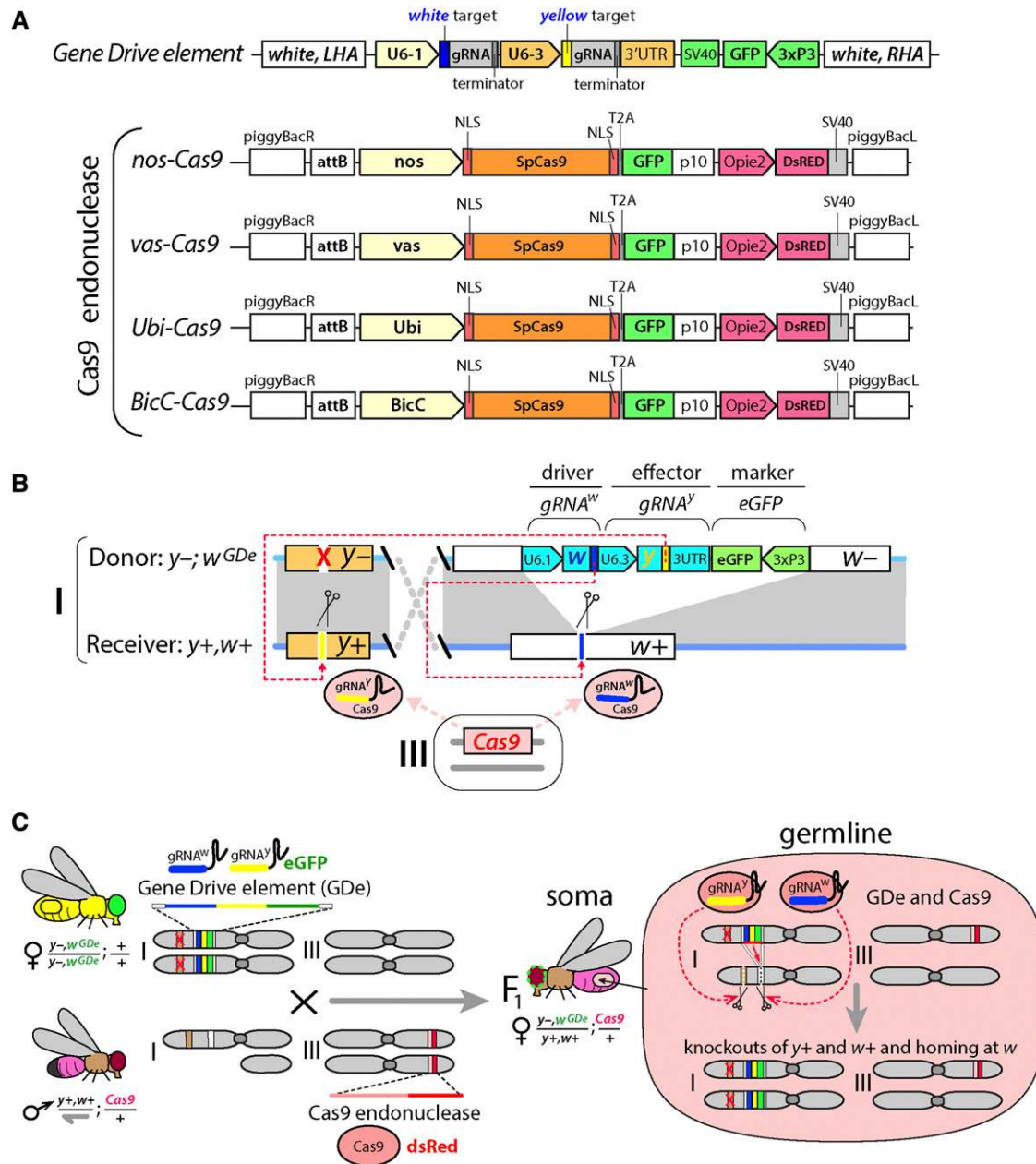


Figure 1 Development of the CRISPR/Cas9-mediated split-drive system. The homing gene drive (HGD) system was split into two components: Gene Drive element (GDe) and Cas9 endonuclease (Cas9). (A) Schematic maps (not to scale) of genetic constructs used to assemble split-HGD systems. The GDe contains two guide RNAs (gRNAs) targeting the DNA cleavage at *white* and *yellow* loci, and an eye-specific marker (3xP3-GFP) all surrounded by Left and Right Homology Arms (LHA and RHA) complementary to the *white* cut site. Four Cas9 constructs expressing SpCas9 (Cas9) in early germline cells with *nanos* (*nos*) and *vasa* (*vas*) promoters, in late germ cells with *Bicaudal C* (*BicC*) promoter, and in both germ and somatic cells with *Ubiquitin 63E* (*Ubi*) promoter carried the eGFP linked to the coding sequence of Cas9 via a self-cleaving T2A sequence and a body specific marker of transgenesis (*Opie2-DsRed*). (B) GDe was site-specifically inserted at *white* locus on the 1st chromosome (i.e., X chromosome) in *Drosophila* via HDR-mediated integration, w^{GDe} . The Cas9 constructs were inserted at the same site on the 3rd chromosome using ϕ C31-mediated integration. In the presence of Cas9, GDe direct cleavage at both w^+ and y^+ loci and can home at *white* locus from the w^{GDe} donor allele into the w^+ receiver allele via HDR in heterozygotes. (C) The genetic cross between the GDe and Cas9 homozygous lines generates trans-heterozygous $y^-; w^{GDe}/y^+, w^+; Cas9/+$ females. The germline Cas9 expression is expected to limit the activity of the split-drive system, y^+ and w^+ knockouts and w^{GDe} homing, to germ cells of the $y^-; w^{GDe}/y^+, w^+; Cas9/+$ females. The w^{GDe} allele cannot home in *Drosophila* males, because they have only one X chromosome, aka. hemizygous.

(<http://www.rainbowgene.com>). The BicC-Cas9 construct was inserted at the PBac[y⁺-attP-3B]KV00033 on the 3rd chromosome (Bloomington #9750) with ϕ C31-mediated integration (Groth 2004). Transgenic flies were balanced with Df(3L)R/TM6C,cu¹,Sb¹,Tb¹ (Bloomington

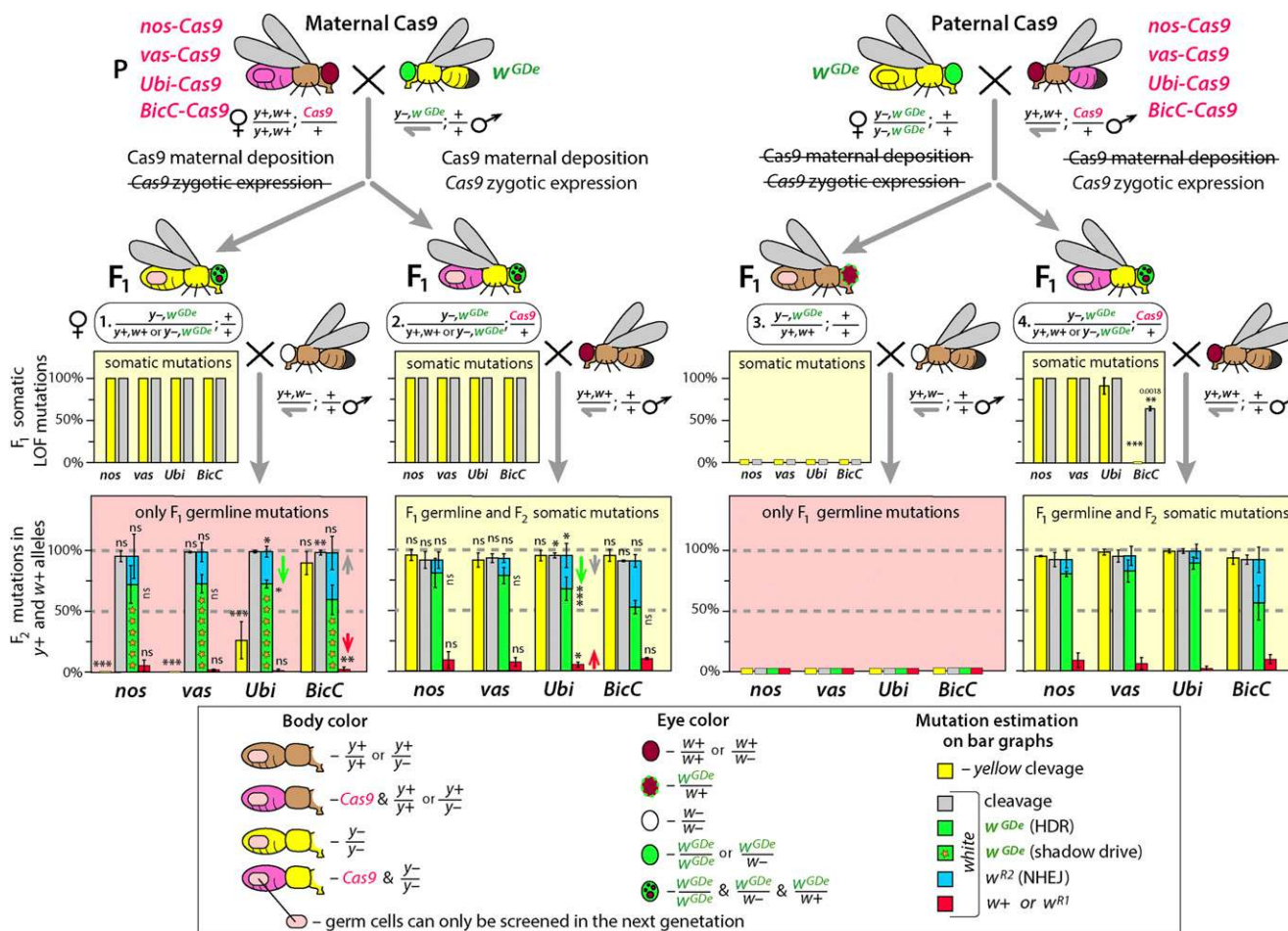
#57) and CxD,ry^{BM}/TM3,Sb¹,Ser¹ (Bloomington #1704) in the w^+ genetic background.

To assess the cleavage rates and homing efficiencies of the split-drive system, we genetically crossed the GDe line to four different

Cas9 lines in both directions. Two types of F₁ *trans*-heterozygous *y*⁻, *w*^{GDe}(*eGFP*)/*y*⁺, *w*⁺; *Cas9*(*RFP*)/+ females carrying either maternal or paternal *Cas9* (F₁ ♀ #2 or ♀ #4, respectively) and the F₁ heterozygous *y*⁻, *w*^{GDe}(*eGFP*)/*y*⁺, *w*⁺ females with either maternally or paternally deposited Cas9 protein were generated (F₁ ♀ #1 or ♀ #3, respectively; Figure 2). Their *yellow* and *white* LOF mutations and transgene markers were scored. To explore whether *yellow* and *white* loci were also mutated in the germ cells of the F₁ *trans*-heterozygous and heterozygous females, we genetically crossed them to *w*⁺, *y*⁺ and *w*⁻, *y*⁺ males, respectively, and examined their F₂ progeny. LOF *yellow* mutations were scored only in male progeny that inherited their single X chromosome from mothers. To explore the behavior of resistance alleles over multiple generations, the F₂ *trans*-heterozygous and heterozygous virgin female (♀ #6 or ♀ #5, respectively) progeny

of F₁ ♀ #2 were also collected, and genetic crosses and phenotype scoring were repeated for an additional generation, F₃. The above crossing schemes are depicted in Figure 2. To generate means and standard deviations for statistical comparisons, each genetic cross was set up in triplicate using 10♂ and 10♀ flies for each replicate cross. Cleavage and homing frequencies are presented as percentages of *y*⁺ and *w*⁺ alleles in heterozygous females, aka. they normalized to 50% (Table S1).

Flies were examined, scored, and imaged on the Leica M165FC fluorescent stereo microscope equipped with the Leica DMC2900 camera. To analyze Cas9 expression in ovaries of four homozygous *Cas9* lines, their ovaries were dissected in PBS buffer, examined, and imaged utilizing the same settings. The eGFP fluorescence was used as a proxy of Cas9 expression, since it was tagged to *Cas9* transgene via a T2A sequence (Figure S1).



Genotyping loci targeted with gRNAs

To explore the molecular changes that caused LOF and in-frame functional mutations in *yellow* and *white* loci, we PCR amplified the genomic regions containing target sites for *gRNA^w* and *gRNA^y*: GGCGATACTTGGATGCCCTGCCG and GGTTTTGGACACT-GGAACCGTGG, respectively. Single-fly genomic DNA preps were prepared by homogenizing a fly in 30 μ L of a freshly prepared squishing buffer (10mM Tris-Cl pH 8.0, 1mM EDTA, 25mM NaCl, 200 μ g/mL Proteinase K), incubating at 37° for 35 min, and heating at 95° for 2 min. 2 μ L of genomic DNA was used as template in a 40 μ L PCR reaction with LongAmp Taq DNA Polymerase (NEB). The 415bp PCR fragment of *white* target was amplified with CGTTAGGGAGCCGATAAAGAGGTCATCC (*w.sF*) and AAGAACGGTGAGTTTCTATTCGCAGTCGG (*w.sR*); and CACTCTGACCTATATAAACATGGACCGCAGTTTGG (*y.sF*) and CCAATTCATCGGCAAAATAGGCATATGCAT (*y.sR*) primers were used to amplify the 375bp PCR fragment of *yellow*. PCR amplicons were purified using QIAquick PCR purification kit (QIAGEN), and sequenced in both directions with Sanger method at Source BioScience. To characterize molecular changes at the targeted sites, sequence AB1 files were aligned against the corresponding reference sequences in SnapGene 4.

Statistical analysis

Statistical analysis was performed in JMP 8.0.2 by SAS Institute Inc. At least three biological replicates were used to generate statistical means for comparisons. To estimate the effect of Cas9 maternal deposition on homing efficiency, rates of cleavage, homing, and resistance allele formation in F₁ ♀ #4 with paternal Cas9 were compared to the corresponding values in F₁ ♀ #1 and ♀ #2 with maternally deposited Cas9 protein (Figure 2). To assess the significance of resistance allele accumulation and homing rate decline between F₂ and F₃ generations, rates of cleavage, homing, and resistance alleles in F₂ ♀ #5 and F₂ ♀ #6 (Figure 3A) were compared to the corresponding values in F₁ ♀ #1 and F₁ ♀ #2, respectively (Figure 2). *P* values were calculated for a two-sample Student's *t*-test with equal variance.

Gene drive safety measures

All crosses using gene drives genetics were performed in accordance to an Institutional Biosafety Committee-approved protocol from UCSD in which full gene-drive experiments are performed in a high-security ACL2 barrier facility and split drive experiments are performed in an ACL1 insectary in plastic vials that are autoclaved prior to being discarded in accord with currently suggested guidelines for laboratory confinement of gene-drive systems (Akbari *et al.* 2015; National Academies of Sciences, Engineering, and Medicine *et al.* 2016).

Ethical conduct of research

We have complied with all relevant ethical regulations for animal testing and research and conformed to the UCSD institutionally approved biological use authorization protocol (BUA #R2401).

Data availability

All data that are represented fully within the tables and figures. The *nos-*, *vas-*, *Ubi-Cas9* plasmids and the corresponding fly lines are deposited at Bloomington Drosophila Stock Center (#79004 – #79006) and AddGene.org (#112685 – #112687), respectively. The *BicC-Cas9* and *GDe* plasmids and fly lines will be made available upon request.

Supplemental material available at figshare: <https://doi.org/10.25387/figshare.11449542>.

RESULTS

Design of split-HGD encoding two gRNAs

To assess the feasibility and efficiency of utilizing a HGD to bias transmission while also expressing a GME, we designed a HGD that expressed two gRNAs (Lopez del Amo *et al.* 2019). The homing component of the split-HGD system, referred herein as a Gene Drive element (GDe), encodes a gRNA targeting *white* (*gRNA^w*, driver gRNA), a separate cargo GME targeting *yellow* (*gRNA^y*, effector), a *3xP3-eGFP* dominant marker, all together flanked by 1kb homology arms from the *white* target locus to direct targeted HDR mediated integration (Figure 1A). The *GDe* was integrated at the *white* locus (*w^{GDe}*) in *D. melanogaster* via HDR. In the presence of Cas9, the *GDe* directs cleavage at both *white* and *yellow*, both X-linked loci, and is also capable of homing into the *white* locus (Figure 1B-C). Importantly, in *D. melanogaster* homozygous loss-of-function (LOF) mutants of both *white* and *yellow* are viable and fertile with scorable recessive LOF phenotypes in the eye and body, respectively, enabling cleavage events to be directly quantified over successive generations. Additionally, males have only one X chromosome, and are therefore hemizygous for *white* and *yellow*, restricting the quantification of homing to heterozygous females (*y⁻,w^{GDe}/y⁺,w⁺*, Figure 1C).

High penetrance of F₁ somatic mutations generated by Cas9 through both maternal deposition and zygotic expression

We explored the effects of tissue specificity and timing of Cas9 expression on cleavage and homing in the germline by using four separate promoters with distinct expression profiles to express Cas9-T2A-GFP: *nanos* (*nos*) (Van Doren *et al.* 1998) and *vasa* (*vas*) promoters known for early germline-limited expression (Hay *et al.* 1988; Van Doren *et al.* 1998; Sano *et al.* 2002); *Bicaudal C* (*BicC*) promoter supporting later germline-limited expression (Saffman *et al.* 1998); and *Ubiquitin 63E* (*Ubi*) promoter with strong expression in both somatic and germ cells (Preston *et al.* 2006; Akbari *et al.* 2009). We controlled for variation in expression due to position effect (PE), by integrating each Cas9 construct (Figure 1A) into the same site on the 3rd chromosome using ϕ C31-mediated integration (Groth 2004). We confirmed germline expression by imaging the expression of a self-cleaving T2A-eGFP tag attached to the coding sequence of Cas9, and each promoter robustly expressed GFP in the ovaries (Lowest *nos-Cas9* < *vas-Cas9* < *Ubi-Cas9* < *BicC-Cas9* Highest) (Figure S1).

We quantified cleavage efficiencies by performing bi-directional crosses between hemizygous or homozygous *GDe* lines mated to heterozygous Cas9 lines (Figure 2). From these crosses we determined that maternally deposited Cas9 protein is sufficient to induce both *yellow* and *white* somatic LOF mutations in F₁ females heterozygous for the *GDe* both in presence (♀ #2; *y⁻,w^{GDe}/y⁺,w⁺*; *Cas9/+*) and in the absence (♀ #1; *y⁻,w^{GDe}/y⁺,w⁺*; Figure 2) of *Cas9* gene inheritance. To determine whether zygotic expression of *Cas9* can also induce somatic mutations, we scored *white* and *yellow* LOF somatic mutations in F₁ *trans*-heterozygous females inheriting *Cas9* exclusively from their fathers (*i.e.*, paternal *Cas9*). Unexpectedly, F₁ *trans*-heterozygous female progeny inheriting *Cas9* as a gene (♀ #4; *y⁻,w^{GDe}/y⁺,w⁺*; *Cas9/+*; Figure 2) from their fathers had mutations in both *white* and *yellow* with varying frequencies depending on which promoter drove *Cas9* expression. For example, *nos-Cas9* and *vas-Cas9* – induced 100% *white* and

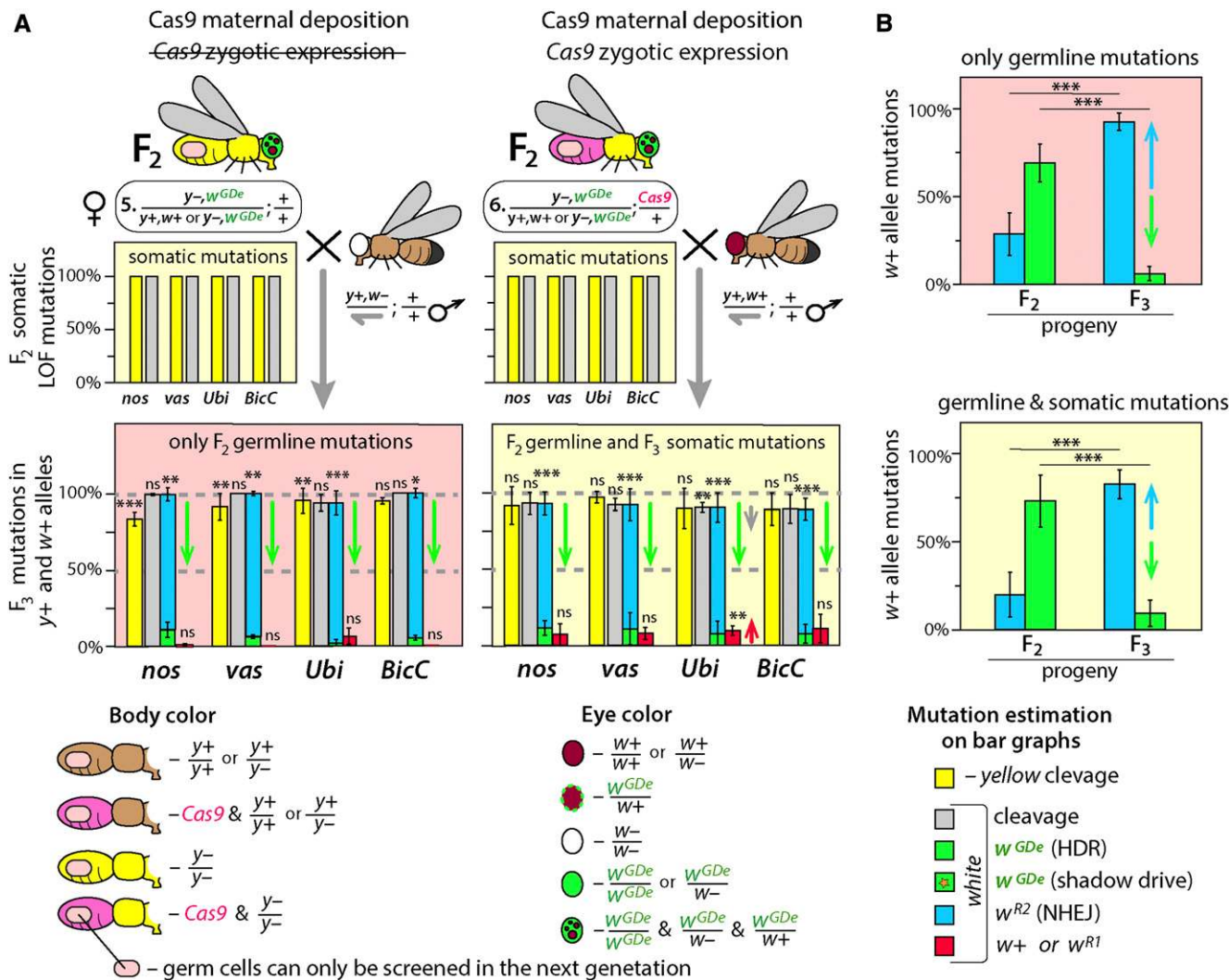


Figure 3 Resistance alleles accumulate over subsequent generations and restricts homing. Resistance alleles are expected to be immune to the further cleavage by the same Cas9/gRNA system and if their carrier is fertile can propagate at the expense of homing. (A) To explore this phenomenon, F₂ ♀ #5 and F₂ ♀ #6 collected among progeny of F₁ ♀ #2 were genetically crossed with *w*⁻ and *w*⁺ males, respectively, and their F₃ progeny were scored. While the cleavage rate in F₂ germ cells decreased only in F₂ ♀ #6 with *Ubi-Cas9* (red arrow) likely due to the rise of functional *w^{R1}* alleles, the homing frequency fell significantly for each tested split-drive system with and without Cas9 gene (green arrows). The fall of homing rate was accompanied by the accumulation of the *w^{R2}* alleles. (B) Accumulation of *w^{R2}* alleles resistant to cleavage by Cas9/gRNA^W restricted homing of GDe. Frequencies of homing and resistance alleles were averaged for all tested promoters and presented separately for progeny of heterozygous and *trans*-heterozygous females, F₂ ♀ #5 and F₂ ♀ #6, respectively. Resistance allele frequency increased from 28.5% or 19.9% to 92.6% or 82.6%, respectively, between F₂ and F₃ (blue arrows) and caused the dramatic decline in homing from 69.0% or 73.0% to 6.1% or 9.2%, respectively (green arrows). Notably, scoring of *w^{R2}* alleles in *w*⁻ recessive background resulted in the higher estimation of *white* LOF mutations alleles, since *w^{R2}* alleles were complemented by *w*⁺ alleles inherited from wild type males. Bar plots show the average ± SD over at least three biological replicate crosses. Statistical significance was estimated using a t-test with equal variance. ($P \geq 0.05$ ^{ns}, $P < 0.05$ *, $P < 0.01$ ***, and $P < 0.001$ ***).

yellow LOF somatic mutations in F₁ *trans*-heterozygous females, while *Ubi-Cas9* resulted in 100% of *white* and 91.3% ± 9.7% of *yellow* LOF somatic mutations, and *BicC-Cas9* resulted in only *white* LOF mutations in 64.3% ± 2.6% of the F₁ *y*⁻, *w^{GDe}*/*y*⁺, *w*⁺; *BicC-Cas9*/+ progeny (Figure 2). Interestingly however, 100% of F₁ heterozygous female progeny from the same fathers that did not inherit Cas9 as a gene (♀#3; *y*⁻, *w^{GDe}*/*y*⁺, *w*⁺; +/+; Figure 2) had wild type (*wt*) phenotypes, for both *white* (red eyes) and *yellow* (brown body), presumably resulting from lack of sufficient Cas9 protein deposited paternally to induce mutations in the zygote (Table S1). Taken together these data indicate that the Cas9 promoters

tested here are active both maternally and zygotically and can promote very high cleavage efficiency in somatic cells.

We assessed whether the *yellow* and *white* alleles were mutated by maternally deposited Cas9 in germ cells of F₁ *y*⁻, *w^{GDe}*/*y*⁺, *w*⁺ females by mating these females to *y*⁺, *w*⁻ males and scored recessive *yellow* phenotypes in resulting F₂ male progeny (*y*⁻) and recessive *white* phenotypes in resulting F₂ male and female progeny (*w*⁻/*w*⁻). We found that maternally deposited Cas9 protein expressed under *nos* and *vas* promoters did not induce *yellow* LOF mutations in germ cells of F₁ females, while expression from *Ubi* and *BicC* promoters resulted in 26% ± 15% and 89.4% ± 9.4% of *yellow* alleles being

mutated in germ cells of F₁ females (Figure 2), respectively, perhaps due to a stronger maternal deposition of Cas9 protein by these promoters (Figure S1) combined with possible preferential gRNA loading by Cas9. Despite the lack of LOF germline mutations in *yellow* by *nos* and *vas*, every tested Cas9 line provided a sufficient amount of maternally deposited Cas9 protein to knockout the *white* allele in 94.9% ± 4.5–98.8% ± 1.1% of F₁ germ cells (measured in F₂ progeny; Figure 2, Table S1). We explored whether the *w*⁺ alleles (1.2–5.1%) were cut by Cas9, and perhaps repaired into cleavage resistance alleles, by performing Sanger sequencing of PCR amplicons of the *white* target locus from individual male flies. Each tested F₂ male with red eyes (*w*⁺) indeed had a *wt w*⁺ allele, and we did not find any *white* in-frame functional resistance alleles in F₁ germ cells suggesting that these alleles likely remained uncut in the germline.

Maternally deposited Cas9 is sufficient to induce homing of GDe in germ cells

The Cas9/gRNA^w-induced DSBs at *white* locus can be repaired either by HDR resulting in homing of the *GDe* (*w*^{GDe}/*w*^{GDe}) or NHEJ incorporating *indel* mutations that can render the target locus unrecognizable by the Cas9/gRNA^w machinery, and when these mutations occur in germ cells they are referred to as resistance alleles (*w*^R): here LOF and in-frame functional resistance alleles are referred as *w*^{R2} and *w*^{R1}, respectively (Figure S2). To directly estimate the frequency of *w*^{GDe} homing and *w*^R generation in the absence of additional somatic mutations resulting from zygotic expression of *Cas9*, we analyzed *white* phenotypes in the F₂ progeny of the F₁ *w*^{GDe}/*w*⁺ females with maternally deposited Cas9 in a *w*⁻ recessive mutant background (Figure 2). Every tested Cas9 promoter provided a sufficient amount of maternally deposited Cas9 in the F₁ germ cells to enable the conversion of 59–72% of *w*⁺ alleles into *w*^{GDe} (*i.e.*, homing of *GDe*) in *y*⁻,*w*^{GDe}/*y*⁺,*w*⁺ females. This conversion which occurs in the presence of Cas9 protein, but absence of inheritance of the *Cas9* gene, was previously noted and termed “shadow drive” (Guichard *et al.* 2019). The remaining DSBs at *w*⁺ alleles were repaired by NHEJ and generated around 38–23% *w*^{R2} alleles (Figure 2). To explore molecular changes at *white* locus, we PCR amplified and Sanger sequenced *w*^{R2} alleles from individual F₂ male progeny and identified *indels* localized at the *white* cut site in each sequenced male (Figure S3A). The maternally deposited Cas9 by *BicC* promoter resulted in the lowest homing and the highest resistance allele rates (59.3% ± 12.3% and 38.7% ± 13.7%, respectively), though no significant difference was identified between *BicC* and other Cas9 promoters. Nevertheless, each tested promoter supplied Cas9 protein via mothers to the progeny that enabled shadow drive, thus resulting in super-Mendelian propagation of *w*^{GDe} to their grandchildren.

Maternal deposition of Cas9 protein reduces the homing efficiency

Maternally deposited Cas9 can induce *white* cleavage and repair mediated by NHEJ as opposed to HDR in mitotically dividing germ cells which can result in a bias toward generating resistance alleles (*w*^{R2} and *w*^{R1}) at the expense of homing *w*^{GDe} (Lopez del Amo *et al.*, 2019). To explore this effect, we compared homing rates between F₁ *trans*-heterozygous females that inherited Cas9 either maternally (♀ #2; *y*⁻,*w*^{GDe}/*y*⁺,*w*⁺; *Cas9*/+) or paternally (♀ #4; *y*⁻,*w*^{GDe}/*y*⁺,*w*⁺; *Cas9*/+; Figure 2). For *nos-Cas9*, *vas-Cas9*, and *BicC-Cas9*, maternal deposition of Cas9 did not result in a significant bias in homing efficiencies. However, for *Ubi-Cas9* homing rates were significantly lower (67%) in the *trans*-heterozygous females that inherited *Cas9* maternally (♀ #2; *w*^{GDe}/*w*⁺; *Ubi-Cas9*/+) as compared to 88% for *trans*-heterozygous females inheriting *Ubi-Cas9* paternally (♀ #4;

w^{GDe}/*w*⁺; *Ubi-Cas9*/+). In addition to the lower homing rates for *Ubi-Cas9*, the rate of *w*^{R2} alleles was significantly higher with maternally deposited *Ubi-Cas9* as compared to paternally deposited *Ubi-Cas9*: 9.9% ± 5.7% vs. 27.3% ± 10.0%, *P* > 0.025 or 26.5% ± 4.4%, *P* > 0.029, respectively (Figure 2). Taken together, these results suggest that high levels of maternal deposition of Cas9 protein into developing oocytes can result in *white* cleavage in mitotic cells, prior to developmental stages where efficient HDR repair occurs, therefore leading to a higher frequency of *w*^R events.

Resistance alleles accumulate between F₂ and F₃ generations

Resistance alleles generated in germ cells are immune to subsequent cleavage by the Cas9/gRNA^w complex. *Drosophila white* and *yellow* LOF homozygotes are viable and fertile, as a result, the frequency of resistance alleles can potentially increase from generation to generation. We explored this possibility by crossing F₂ *trans*-heterozygous females (♀ #6, *w*^{GDe}/*w*⁺; *Ubi-Cas9*/+; Figure 3A) to *wt* (*y*⁺,*w*⁺) males, and scored their F₃ progeny for *yellow* and *white* phenotypes, as well as for inheritance of the *GDe*. Indeed, the frequency of *white* LOF mutations (*w*^{R2}) increased significantly between F₂ and F₃ progenies for each Cas9 promoter: 11.2% ± 6.2% vs. 81.7% ± 7.5% for *nos-Cas9*; 13.2% ± 5.6% vs. 82.4% ± 10.4% for *vas-Cas9*; 18.6% ± 12.0% vs. 84.6% ± 9.5% for *Ubi-Cas9*; and 36.7% ± 7.5% vs. 81.6% ± 7.1% for *BicC-Cas9*, *P* > 0.0001, respectively. This increased frequency of generating *w*^{R2} alleles negatively affected the homing rate, which dropped between F₂ and F₃ generations: from 80.0% ± 7.7% to 11.3% ± 4.8% for *nos-Cas9*; from 80.2% ± 7.4% to 10.8% ± 10.4% for *vas-Cas9*; from 78.0% ± 13.2% to 7.4% ± 8.4% for *Ubi-Cas9*; and from 53.9% ± 9.8% to 7.6% ± 6.2% for *BicC-Cas9* (♀ #6, *w*^{GDe}/*w*⁺; *Ubi-Cas9*/+; Figure 3A). To avoid any ambiguity caused by somatic expression of Cas9, the same analysis was repeated with the F₂ heterozygous females carrying maternally deposited Cas9 protein but lacking the *Cas9* gene resulting in similar conclusions (♀ #5, *y*⁻,*w*^{GDe}/*y*⁺,*w*⁺; Figure 3A). We assessed the accumulation of resistance alleles by comparing the mean frequencies of homing and resistance alleles between F₂ and F₃ generations. The frequency of resistance alleles rose from 28.5% ± 12.2% to 92.6% ± 5.0% in heterozygous females or from 19.9% ± 12.8% to 82.6% ± 8.2% in *trans*-heterozygous females, and decreased the homing rate from 69.0% ± 10.8% to 6.1% ± 4.2% and from 73.0% ± 14.6% to 9.2% ± 7.5%, respectively (*P* > 0.0001, Figure 3B). As expected, the frequency of LOF resistance alleles at *white* locus (*w*^{R1}) also increased from F₂ to F₃ generations and further restricted homing of the *GDe*. The frequency of in-frame functional *white* and *yellow* mutations (*w*^{R1} and *y*^{R1}) could also increase in the F₃ progeny, but unfortunately this effect could not be directly estimated. The frequency of cleavage at *white* significantly decreased in the F₃ progeny of F₂ *y*⁻,*w*^{GDe}/*y*⁺,*w*⁺; *Ubi-Cas9*/+ females, and could be explained by the increase of *w*^{R1} allele rate that were indistinguishable from *w*⁺ alleles phenotypically: from 3.4% ± 2.6% in F₂ to 9.7% ± 3.1% in F₃, *P* > 0.004 (Figures 2, 3A). We tested this hypothesis by Sanger sequencing F₃ *wt* males with red eyes and brown bodies, and identified in-frame *indels* and substitutions in the majority of tested males for each Cas9 promoter (*w*^{R1} and *y*^{R1} alleles, Figure S3). Therefore, many germ cells of F₂ *trans*-heterozygous and heterozygous with maternally deposited Cas9 females had *indel* mutations in the *white* and *yellow* loci (*y*⁻,*w*^{GDe}/*y*^{R1},*w*^{R1}) that were indeed resistant to further cleavage by Cas9/gRNA^w and Cas9/gRNA^y, respectively.

DISCUSSION

Homing based gene drives require efficient cleavage and copying in the germline in order to bias their transmission and are therefore

sensitive to both existing and induced target sequence variation. In fact, the NHEJ-mediated generation of resistance alleles in germ cells was previously identified as the major force opposing the spread of HGD into populations (Gantz *et al.* 2015; Champer *et al.* 2017; Hammond *et al.* 2017; Oberhofer *et al.* 2018). Here, we used a split-drive design to further explore the effect of timing and location of Cas9 expression on both homing and resistance allele formation. This experimental design enabled us to separate effects of somatic expression from maternal deposition of Cas9 on the *GDe* inheritance and mutagenesis of a targeted gene. Additionally, we linked a GME to the *GDe* to measure the efficacy of the knockout of an additional gene, *yellow*. Using this approach, we were able to draw several conclusions, including: i) in addition to germline expression, each tested Cas9 promoter (*nos*, *vas*, *BicC*, *Ubi*) directs significant expression in somatic tissues; ii) the maternal protein deposition or gene expression of Cas9 is sufficient for homing (shadow drive) in germ cells; iii) paternal Cas9 protein deposition in the sperm is insufficient for the mutagenesis of a target gene; iv) drive-induced resistance alleles accumulate over generations and are predicted to restrict the spread of the drive; and v) expression of a drive mediating gRNA in addition to a linked GME can result in 100% penetrance of both scorable LOF phenotypes. Below we discuss these conclusions further and also propose novel drive architectures to potentially overcome these issues.

Somatic expression of Cas9 results in high mutagenesis rates

The maternal protein deposition and gene expression of Cas9 in the presence of a gRNA transgene were previously reported to induce LOF mutations in some F₁ progeny from a cross using *nos*- or *vas*-driven Cas9 and U6-gRNA lines (Port *et al.* 2014; Lin and Potter 2016; Oberhofer *et al.* 2018; Kandul *et al.* 2019); however, the somatic nature of F₁ LOF mutations was not fully explored. This is in part due to the fact that when Cas9 and gRNA are linked together in a single-locus HGD, somatic and germline LOF mutations are not easily distinguishable from heritable mutations occurring in prior generations, which can result in overestimation of mutation rates. Therefore, unlinking these components enables a better method for methodically disentangling these events. Here, using a split-drive design, we were able to carefully assess the effects of timing, expression, and inheritance of Cas9 on both homing and cleavage efficiencies. As reported previously, we found that maternal Cas9 protein deposition was sufficient to induce homing in germ cells, aka. shadow drive (Champer *et al.* 2019c; Guichard *et al.* 2019), in addition to high rates of F₁ somatic LOF mutations (Port *et al.* 2014; Lin and Potter 2016; Oberhofer *et al.* 2018; Kandul *et al.* 2019). Interestingly, our estimations of homing rates by the shadow drive in *white* locus are notably higher than the previously reported in *yellow* locus: 59–72% (Figure 2) vs. 29–32% (Guichard *et al.* 2019) and 38% (Champer *et al.* 2019c), although the ratio of first generation drive to shadow drive frequencies is comparable in these systems (~50%). The differences may be due to the *yellow* locus being less accessible to cleavage and HDR than *white* locus, or perhaps the lower fitness of *yellow* LOF somatic mutations (see below) biases against their inheritance (Masse *et al.* 2019).

Rather unexpectedly, we found that the zygotic expression of Cas9 alone (paternal Cas9), without the maternally deposited Cas9 protein, was also sufficient to induce F₁ LOF somatic mutations. In fact, both *nanos* and *vasa* promoters, which were previously characterized to have early germline-limited expression (Van Doren *et al.* 1998; Sano *et al.* 2002; Kondo and Ueda 2013), in our system do support significant somatic expression of Cas9 which may stem from PE or perhaps the use of the P10 3'UTR. For example, F₁ progeny with both maternally

deposited Cas9 protein or with zygotically expressed Cas9 gene inherited from their fathers had *white* and *yellow* LOF somatic mutations with up to 100% efficiency (Figure 2). Consistent observations were reported in a recent work using a *trans*-complementing Gene Drive (tGD) system (Lopez del Amo *et al.* 2019) and for somatically induced lethality of Notch alleles driven by paternally provided Cas9 (Guichard *et al.*, 2019). Taken together, these data conclusively demonstrate that, in the context of the tested promoters, Cas9 somatic expression confounds the estimation of mutagenesis rates in germ cells and can result in the overestimation of homing rates for a single-locus HGD.

Resistance alleles accumulate over subsequent generations

Consistent with previous studies, we found that maternal deposition of Cas9 protein into embryos inheriting a *GDe* results in both resistance allele formation and homing in the germ cells (Champer *et al.* 2019c; Guichard *et al.* 2019). In addition to this observation, we also found that paternal Cas9 protein deposition was not sufficient to induce mutagenesis in target genes, presumably due to the low quantities of Cas9 carried by the sperm into the egg. Moreover, we determined that maternal deposition of Cas9 protein in *trans*-heterozygous females with *vas*- and *nos*-Cas9 does not induce more resistance alleles at the expense of a homing rate than those in the females that inherit *vas*- and *nos*-Cas9 paternally.

In the light of substantial somatic expression of Cas9 driven by common *Drosophila* germline promoters (Figure 2), the germline inheritance rate provides a better estimate of the rate of inducing resistance alleles in germ cells than the 'embryonic resistance allele' frequency used previously (Champer *et al.* 2019c). Consequently, our estimations of F₂ resistance allele formation are lower than those reported by Champer *et al.* as the embryo R² (LOF) resistance alleles for a split-gene drive system with the *nos*-Cas9, 11% ± 6% (Figure 2) vs. 74% ± 2% (Champer *et al.* 2019c).

The frequency of resistance alleles (*w^R*) increased dramatically between F₂ and F₃ generations and correlated with decreases in homing (Figure 3). Taken together, these results suggest that HDR-mediated homing and NHEJ-mediated formation of resistance alleles are integral outcomes of DSBs repair induced by Cas9/gRNA; and when resistance alleles do not cause lethality or sterility to their carrier, the accumulation of resistance alleles is predicted to impede the spread of the drive (Hammond *et al.* 2017; KaramiNejadRanjbar *et al.* 2018; Oberhofer *et al.* 2018).

gRNA-mediated effector (GME)

The CRISPR/Cas9 technology was previously used in the combination with multiple gRNAs to knock out or convert different genes simultaneously (*e.g.*, Cong *et al.* 2013; Lopez del Amo *et al.* 2019; Kandul *et al.* 2019; Guichard *et al.* 2019). Here we linked the second gRNA targeting *yellow* to the *GDe* inside the *white* locus, and demonstrated that both *yellow* and *white* were effectively knocked out in heterozygous females. In principle, the GME approach can be used to knock out multiple genes located on different chromosomes, such as multiple host factors required for mosquito infection with pathogens or repressors of mosquito anti-pathogen immune genes (Simões *et al.* 2018). Unlike the allelic drive (Guichard *et al.* 2019), the GME does not require the HDR-mediated conversion in germ cells; instead, it relies on the NHEJ-mediated *indel* formation in somatic tissues, and the widespread Cas9 somatic expression described here is expected to improve the penetrance of the GME-mediated knockout (Kandul *et al.* 2019). Robust knockout of host genes in somatic tissues may reduce the mosquito

fitness (Dong *et al.* 2018), but an efficient gene drive can spread its cargo genes in a population even if they are costly to their carriers (Kyrou *et al.* 2018). Therefore, our results suggest that the GME directing knock out of multiple mosquito genes to suppress pathogen infection in mosquitoes may be a viable strategy and should be further explored going forward.

Novel strategies for disarming resistance alleles in germ cells

The accumulation of drive resistance alleles reported here was in part due to the fact that *white* is recessive viable, enabling NHEJ-induced resistance alleles to accumulate. Given this accumulation, targeting non-essential genes using HGD may not be ideal. To avoid this issue, targeting essential genes would be a more appropriate design to ensure gene drive stability and spread. By targeting essential genes, it is possible that non-drive resistance alleles could be actively selected against using a phenomenon previously termed as lethal mosaicism (Kandul *et al.* 2019; Guichard *et al.* 2019) or by natural selection due

to increased fitness costs. Lethal mosaicism results in dominant biallelic knockouts of target genes throughout development, which could eliminate cleavage resistance alleles as they would be non-viable. We envision two novel drive design architectures that incorporate a GME and rely on lethal mosaicism to limit the generation of resistance alleles. First, haplo-sufficient genes essential for insect viability or fertility can be targeted by HGD designed to express a recoded version of the disrupted gene that is resistant to gRNA-mediated cleavage in addition to a linked GME (HGD+R+GME). This ensures that only the progeny that inherit the HGD+R+GME survive, while all progeny that inherit a cleaved allele perish due to non-rescued lethal mosaicism (Figure 4). Second, a Cleavage-only Gene Drive with Rescue could be designed that incorporates a GME (CGD+R+GME) which mechanistically relies exclusively on cleavage for biased inheritance and selection against drive resistance alleles (Figure 4) (Oberhofer *et al.* 2019). Both of these strategies would likely be effective in limiting the accumulation of drive resistance alleles. However, in-frame functional mutations (*R1* type) that confer resistance against the Cas9/gRNA and do

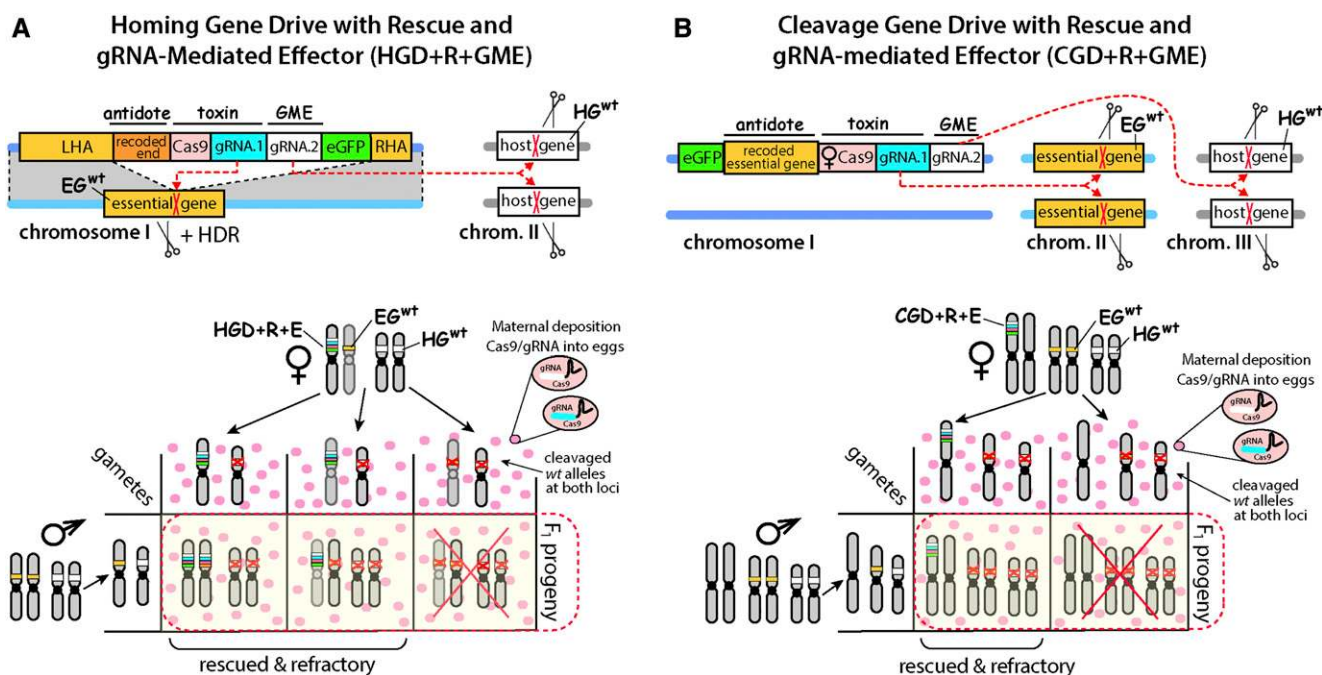


Figure 4 gRNA-mediated effector (GME) incorporated into two novel gene drive designs mechanistically based on lethal biallelic mosaicism. (A) Schematic of Homing Gene Drive targeting an essential gene with a recoded Rescue and GME (HGD+R+GME). The HGD+R+GME expresses Cas9 and two gRNAs targeting an essential gene (EG) and host gene (HG), a marker gene (*eGFP*), and the cleavage-resistant recorded portion of the essential gene that is being targeted by the gRNA/Cas9 complex (Rescue), which can rescue the knockout phenotype, flanked by Left and Right Homology Arms (LHA and RHA). Mechanistically, once HGD+R+GME is integrated precisely inside the EG it will direct cleavage of the EG^{wt} allele on a receiver chromosome, and induce knockout mutations that will either result in lethal biallelic mosaicism, or convert the receiver chromosome into EG^{HGD+R+GME} via homology directed repair (HDR). This ensures that only the progeny that inherit EG^{HGD+R+GME} survive, while all progeny that inherit a cleaved EG allele perish due to non-rescued lethal mosaicism. In addition, the HGD+R+GME induces knockout of HG located on another (or the same) chromosome, leading to desired phenotype (*i.e.*, pathogen resistance) to its carriers. The Punnett square below depicts the genetics of how HGD+R+GME achieves a 100% transmission rate and refractoriness in F₁ progeny. Female heterozygous for HGD+R+GME maternally deposits Cas9/gRNA complexes into every oocyte knocking out both EG and HG, and only zygotes that inherit the HDR+R+GME would survive as F₁ progeny. Notably, HDR will convert EG^{wt} alleles into EG^{HGD+R+GME} alleles and further increase numbers of surviving F₁ progeny and this non-Mendelian inheritance rate will depend on homing efficiencies. (B) Schematic of Cleavage-only Gene Drive targeting an essential gene with a recoded Rescue and GME (CGD+R+GME). The CGD+R+GME expresses Cas9 with multiple gRNAs targeting an ES (gRNA.1) and HG (gRNA.2), a marker gene (*eGFP*), and the cleavage-resistant recorded essential gene (Rescue) integrated at a separate genomic location from the target gene. Mechanistically, a CGD+R+GME drive relies exclusively on cleavage with no HDR required for biased inheritance. A Punnett square depicts the genetics of how CGD+R+GME achieves 100% transmission and infection resistance rates in F₁ progeny. The female heterozygous for CGD+R+GME deposits Cas9/gRNA complexes into every oocyte, only the half of the zygotes that inherits the CDR+R+GME in a Mendelian fashion survive as F₁ progeny, while the other half that do not inherit CDR+R+GME perishes due to lethal biallelic mosaicism.

not cause fitness costs to carriers may still be generated, which could still limit the spread of a drive, and generation of these R1 alleles could possibly be further minimized by inclusion of additional gRNAs that target the essential genes to mediate drive. To summarize, our results demonstrate that inserting a GME into a HGD, efficient knockouts of multiple genes can be achieved while simultaneously biasing GDe transmission rates into subsequent generations. However, resistance alleles were generated, and accumulated, which would limit the efficacy and spread of this system. To overcome these limitations, novel drive architectures are proposed and remain to be tested in future studies.

ACKNOWLEDGMENTS

This work was supported in part by the University of California, San Diego, Department of Biological Sciences, by funding from the Defense Advanced Research Project Agency (DARPA) under a “Safe Genes” Program Grant (HR0011-17-2-0047) awarded to O.S.A. and by the Office of the Director of the National Institutes of Health under award number DP5OD023098 awarded to V.M.G.

LITERATURE CITED

Adelman, Z., O. Akbari, J. Bauer, E. Bier, C. Bloss *et al.*, 2017 Rules of the road for insect gene drive research and testing. *Nat. Biotechnol.* 35: 716–718. <https://doi.org/10.1038/nbt.3926>

Akbari, O. S., D. Oliver, K. Eyer, and C.-Y. Pai, 2009 An Entry/Gateway cloning system for general expression of genes with molecular tags in *Drosophila melanogaster*. *BMC Cell Biol.* 10: 8. <https://doi.org/10.1186/1471-2121-10-8>

Akbari, O. S., K. D. Matzen, J. M. Marshall, H. Huang, C. M. Ward *et al.*, 2013 A Synthetic Gene Drive System for Local, Reversible Modification and Suppression of Insect Populations. *Curr. Biol.* 23: 671–677. <https://doi.org/10.1016/j.cub.2013.02.059>

Akbari, O. S., C.-H. Chen, J. M. Marshall, H. Huang, I. Antoshechkin *et al.*, 2014 Novel synthetic Medea selfish genetic elements drive population replacement in *Drosophila*; a theoretical exploration of Medea-dependent population suppression. *ACS Synth. Biol.* 3: 915–928. <https://doi.org/10.1021/sb300079h>

Akbari, O. S., H. J. Bellen, E. Bier, S. L. Bullock, A. Burt *et al.*, 2015 BIOSAFETY. Safeguarding gene drive experiments in the laboratory. *Science* 349: 927–929. <https://doi.org/10.1126/science.aac7932>

Buchman, A., J. M. Marshall, D. Ostrovski, T. Yang, and O. S. Akbari, 2018a Synthetically engineered Medea gene drive system in the worldwide crop pest *Drosophila suzukii*. *Proc. Natl. Acad. Sci. USA* 115: 4725–4730. <https://doi.org/10.1073/pnas.1713139115>

Buchman, A. B., T. Ivy, J. M. Marshall, O. S. Akbari, and B. A. Hay, 2018b Engineered Reciprocal Chromosome Translocations Drive High Threshold, Reversible Population Replacement in *Drosophila*. *ACS Synth. Biol.* 7: 1359–1370. <https://doi.org/10.1021/acssynbio.7b00451>

Buchman, A., S. Gamez, M. Li, I. Antoshechkin, S.-H. Lee *et al.*, 2019a Broad Dengue Neutralization in Mosquitoes Expressing an Engineered Antibody. *BioRxiv*. <https://doi.org/10.1101/645481>

Buchman, A., S. Gamez, M. Li, I. Antoshechkin, H.-H. Li *et al.*, 2019b Engineered resistance to Zika virus in transgenic *Aedes aegypti* expressing a polycistronic cluster of synthetic small RNAs. *Proc. Natl. Acad. Sci. USA* 116: 3656–3661. <https://doi.org/10.1073/pnas.1810771116>

Burt, A., 2003 Site-specific selfish genes as tools for the control and genetic engineering of natural populations. *Proc. Biol. Sci.* 270: 921–928. <https://doi.org/10.1098/rspb.2002.2319>

Burt, A., and R. Trivers, 2006 Genes in conflict: the biology of selfish genetic elements. Harvard University Press. 605p. <https://doi.org/10.4159/9780674029118>

Champer, J., A. Buchman, and O. S. Akbari, 2016 Cheating evolution: engineering gene drives to manipulate the fate of wild populations. *Nat. Rev. Genet.* 17: 146–159. <https://doi.org/10.1038/nrg.2015.34>

Champer, J., R. Reeves, S. Y. Oh, C. Liu, J. Liu *et al.*, 2017 Novel CRISPR/Cas9 gene drive constructs reveal insights into mechanisms of resistance allele formation and drive efficiency in genetically diverse populations. *PLoS Genet.* 13: e1006796. <https://doi.org/10.1371/journal.pgen.1006796>

Champer, J., J. Liu, S. Y. Oh, R. Reeves, A. Luthra *et al.*, 2018 Reducing resistance allele formation in CRISPR gene drive. *Proc. Natl. Acad. Sci. USA* 115: 5522–5527. <https://doi.org/10.1073/pnas.1720354115>

Champer, J., E. Yang, Y. L. Lee, J. Liu, A. G. Clark, *et al.*, 2019a Resistance is futile: A CRISPR homing gene drive targeting a haplolethal gene. *BioRxiv*. <https://doi.org/10.1101/651737>

Champer S. E., S. Y. Oh, C. Liu, Z. Wen, A. G. Clark, *et al.*, 2019b Computational and experimental performance of CRISPR homing gene drive strategies with multiplexed gRNAs. *BioRxiv*. <https://doi.org/10.1101/679902>

Champer, J., J. Chung, Y. L. Lee, C. Liu, E. Yang *et al.*, 2019c Molecular safeguarding of CRISPR gene drive experiments. *eLife* 8: e41439. <https://doi.org/10.7554/eLife.41439>

Chen, C.-H., H. Huang, C. M. Ward, J. T. Su, L. V. Schaeffer *et al.*, 2007 A synthetic maternal-effect selfish genetic element drives population replacement in *Drosophila*. *Science* 316: 597–600. <https://doi.org/10.1126/science.1138595>

Cheng, G., Y. Liu, P. Wang, and X. Xiao, 2016 Mosquito Defense Strategies against Viral Infection. *Trends Parasitol.* 32: 177–186. <https://doi.org/10.1016/j.pt.2015.09.009>

Cong, L., F. A. Ran, D. Cox, S. Lin, R. Barretto *et al.*, 2013 Multiplex genome engineering using CRISPR/Cas systems. *Science* 339: 819–823. <https://doi.org/10.1126/science.1231143>

DiCarlo, J. E., A. Chavez, S. L. Dietz, K. M. Esvelt, and G. M. Church, 2015 Safeguarding CRISPR-Cas9 gene drives in yeast. *Nat. Biotechnol.* 33: 1250–1255. <https://doi.org/10.1038/nbt.3412>

Dong, Y., M. L. Simões, E. Marois, and G. Dimopoulos, 2018 CRISPR/Cas9-mediated gene knockout of *Anopheles gambiae* FREP1 suppresses malaria parasite infection. *PLoS Pathog.* 14: e1006898. <https://doi.org/10.1371/journal.ppat.1006898>

Esvelt, K. M., A. L. Smidler, F. Catteruccia, and G. M. Church, 2014 Concerning RNA-guided gene drives for the alteration of wild populations. *eLife* 3: e03401. <https://doi.org/10.7554/eLife.03401>

Gantz, V. M., and E. Bier, 2015 Genome editing. The mutagenic chain reaction: a method for converting heterozygous to homozygous mutations. *Science* 348: 442–444. <https://doi.org/10.1126/science.aaa5945>

Gantz, V. M., N. Jasinskiene, O. Tatarenkova, A. Fazekas, V. M. Macias *et al.*, 2015 Highly efficient Cas9-mediated gene drive for population modification of the malaria vector mosquito *Anopheles stephensi*. *Proc. Natl. Acad. Sci. USA* 112: E6736–E6743. <https://doi.org/10.1073/pnas.1521077112>

Gantz, V. M., and E. Bier, 2016 The dawn of active genetics. *BioEssays* 38: 50–63. <https://doi.org/10.1002/bies.201500102>

Gibson, D. G., L. Young, R.-Y. Chuang, J. C. Venter, C. A. Hutchison, 3rd *et al.*, 2009 Enzymatic assembly of DNA molecules up to several hundred kilobases. *Nat. Methods* 6: 343–345. <https://doi.org/10.1038/nmeth.1318>

Groth, A. C., 2004 Construction of transgenic *Drosophila* by Using the Site-Specific Integrase From Phage C31. *Genetics* 166: 1775–1782. <https://doi.org/10.1534/genetics.166.4.1775>

Grunwald, H. A., V. M. Gantz, G. Poplawski, X.-R. S. Xu, E. Bier *et al.*, 2019 Super-Mendelian inheritance mediated by CRISPR–Cas9 in the female mouse germline. *Nature* 566: 105–109. <https://doi.org/10.1038/s41586-019-0875-2>

Guichard, A., T. Haque, M. Bobik, X.-R. S. Xu, C. Klanseck *et al.*, 2019 Efficient allelic-drive in *Drosophila*. *Nat. Commun.* 10: 1640. <https://doi.org/10.1038/s41467-019-09694-w>

Hammond, A., R. Galizi, K. Kyrou, A. Simoni, C. Siniscalchi *et al.*, 2016 A CRISPR-Cas9 gene drive system targeting female reproduction in the malaria mosquito vector *Anopheles gambiae*. *Nat. Biotechnol.* 34: 78–83. <https://doi.org/10.1038/nbt.3439>

Hammond, A. M., K. Kyrou, M. Bruttini, A. North, R. Galizi *et al.*, 2017 The creation and selection of mutations resistant to a gene drive

- over multiple generations in the malaria mosquito. *PLoS Genet.* 13: e1007039. <https://doi.org/10.1371/journal.pgen.1007039>
- Hammond A. M., K. Kyrou, M. Gribble, X. Karlsson, I. Morianou, *et al.*, 2018 Improved CRISPR-based suppression gene drives mitigate resistance and impose a large reproductive load on laboratory-contained mosquito populations. *BioRxiv*. <https://doi.org/10.1101/360339>
- Hay, B., L. Y. Jan, and Y. N. Jan, 1988 A protein component of *Drosophila* polar granules is encoded by vasa and has extensive sequence similarity to ATP-dependent helicases. *Cell* 55: 577–587. [https://doi.org/10.1016/0092-8674\(88\)90216-4](https://doi.org/10.1016/0092-8674(88)90216-4)
- Isaacs, A. T., F. Li, N. Jasinskiene, X. Chen, X. Nirmala *et al.*, 2011 Engineered resistance to *Plasmodium falciparum* development in transgenic *Anopheles stephensi*. *PLoS Pathog.* 7: e1002017. <https://doi.org/10.1371/journal.ppat.1002017>
- Jinek, M., K. Chylinski, I. Fonfara, M. Hauer, J. A. Doudna *et al.*, 2012 A Programmable Dual-RNA-Guided DNA Endonuclease in Adaptive Bacterial Immunity. *Science* 337: 816–821. <https://doi.org/10.1126/science.1225829>
- Jupatanakul, N., S. Sim, Y. I. Angleró-Rodríguez, J. Souza-Neto, S. Das *et al.*, 2017 Engineered *Aedes aegypti* JAK/STAT Pathway-Mediated Immunity to Dengue Virus. *PLoS Negl. Trop. Dis.* 11: e0005187. <https://doi.org/10.1371/journal.pntd.0005187>
- Kandul, N. P., J. Liu, H. M. Sanchez C., S. L. Wu, J. M. Marshall *et al.*, 2019 Transforming insect population control with precision guided sterile males with demonstration in flies. *Nat. Commun.* 10: 84. <https://doi.org/10.1038/s41467-018-07964-7>
- KaramiNejadRanjbar, M., K. Eckermann, H. M. M. Ahmed, H. M. Sánchez C., S. Dippel *et al.*, 2018 Consequences of resistance evolution in a Cas9-based sex conversion-suppression gene drive for insect pest management. *Proc. Natl. Acad. Sci. USA* 115: 6189–6194. <https://doi.org/10.1073/pnas.1713825115>
- Kondo, S., and R. Ueda, 2013 Highly improved gene targeting by germline-specific Cas9 expression in *Drosophila*. *Genetics* 195: 715–721. <https://doi.org/10.1534/genetics.113.156737>
- Kyrou, K., A. M. Hammond, R. Galizi, N. Kranjc, A. Burt *et al.*, 2018 A CRISPR–Cas9 gene drive targeting doublesex causes complete population suppression in caged *Anopheles gambiae* mosquitoes. *Nat. Biotechnol.* 36: 1062–1066. <https://doi.org/10.1038/nbt.4245>
- Li, M., T. Yang, N. P. Kandul, M. Bui, S. Gamez, *et al.*, 2019 Development of a Confinable Gene-Drive System in the Human Disease Vector, *Aedes aegypti*. *BioRxiv*. <https://doi.org/10.1101/645440>
- Lin, C.-C., and C. J. Potter, 2016 Non-Mendelian Dominant Maternal Effects Caused by CRISPR/Cas9 Transgenic Components in *Drosophila melanogaster*. *G3 (Bethesda)* 6: 3685–3691. <https://doi.org/10.1534/g3.116.034884>
- Lopez del Amo, V. A. L. Bishop, H. M. Sanchez C., J. B. Bennett, *et al.*, 2019 Split-gene drive system provides flexible application for safe laboratory investigation and potential field deployment. *BioRxiv*. <https://doi.org/10.1101/684597>
- Mali, P., L. Yang, K. M. Esvelt, J. Aach, M. Guell *et al.*, 2013 RNA-guided human genome engineering via Cas9. *Science* 339: 823–826. <https://doi.org/10.1126/science.1232033>
- Marshall, J. M., A. Buchman, H. M. Sánchez C, and O. S. Akbari, 2017 Overcoming evolved resistance to population-suppressing homing-based gene drives. *Sci. Rep.* 7: 3776. <https://doi.org/10.1038/s41598-017-02744-7>
- Marshall, J. M., and O. S. Akbari, 2018 Can CRISPR-Based Gene Drive Be Confined in the Wild? A Question for Molecular and Population Biology. *ACS Chem. Biol.* 13: 424–430. <https://doi.org/10.1021/acscchembio.7b00923>
- Marshall, J. M., R. Raban, N. P. Kandul, J. R. Edula, T. León *et al.*, 2019 Winning the tug-of-war between effector gene design and pathogen evolution in vector population replacement strategies. *Front. Genet.* 10: 1072. <https://doi.org/10.3389/fgene.2019.01072>
- Massey, J. H., D. Chung, I. Siwanowicz, D. L. Stern, and P. J. Wittkopp, 2019 The yellow gene influences *Drosophila* male mating success through sex comb melanization. *eLife* 8: e49388. <https://doi.org/10.7554/eLife.49388>
- McLaughlin, Jr., R. N., and H. S. Malik, 2017 Genetic conflicts: the usual suspects and beyond. *J. Exp. Biol.* 220: 6–17. <https://doi.org/10.1242/jeb.148148>
- National Academies of Sciences, Engineering, and Medicine, Division on Earth and Life Studies, Board on Life Sciences, and Committee on Gene Drive Research in Non-Human Organisms: Recommendations for Responsible Conduct, 2016 *Gene Drives on the Horizon: Advancing Science, Navigating Uncertainty, and Aligning Research with Public Values*. National Academies Press.
- Oberhofer, G., T. Ivy, and B. A. Hay, 2018 Behavior of homing endonuclease gene drives targeting genes required for viability or female fertility with multiplexed guide RNAs. *Proc. Natl. Acad. Sci. USA* 115: E9343–E9352. <https://doi.org/10.1073/pnas.1805278115>
- Oberhofer, G., T. Ivy, and B. A. Hay, 2019 Cleave and Rescue, a novel selfish genetic element and general strategy for gene drive. *Proc. Natl. Acad. Sci. USA* 116: 6250–6259. <https://doi.org/10.1073/pnas.1816928116>
- Oye, K. A., K. Esvelt, E. Appleton, F. Catteruccia, G. Church *et al.*, 2014 Regulating gene drives. *Science* 345: 626–628. <https://doi.org/10.1126/science.1254287>
- Port, F., H.-M. Chen, T. Lee, and S. L. Bullock, 2014 Optimized CRISPR/Cas tools for efficient germline and somatic genome engineering in *Drosophila*. *Proc. Natl. Acad. Sci. USA* 111: E2967–E2976. <https://doi.org/10.1073/pnas.1405500111>
- Preston, C. R., C. C. Flores, and W. R. Engels, 2006 Differential usage of alternative pathways of double-strand break repair in *Drosophila*. *Genetics* 172: 1055–1068. <https://doi.org/10.1534/genetics.105.050138>
- Saffman, E. E., S. Styhler, K. Rother, W. Li, S. Richard *et al.*, 1998 Premature translation of oskar in oocytes lacking the RNA-binding protein bicaudal-C. *Mol. Cell. Biol.* 18: 4855–4862. <https://doi.org/10.1128/MCB.18.8.4855>
- Sano, H., A. Nakamura, and S. Kobayashi, 2002 Identification of a transcriptional regulatory region for germline-specific expression of vasa gene in *Drosophila melanogaster*. *Mech. Dev.* 112: 129–139. [https://doi.org/10.1016/S0925-4773\(01\)00654-2](https://doi.org/10.1016/S0925-4773(01)00654-2)
- Simões, M. L., E. P. Caragata, and G. Dimopoulos, 2018 Diverse Host and Restriction Factors Regulate Mosquito–Pathogen Interactions. *Trends Parasitol.* 34: 603–616. <https://doi.org/10.1016/j.pt.2018.04.011>
- Van Doren, M., A. L. Williamson, and R. Lehmann, 1998 Regulation of zygotic gene expression in *Drosophila* primordial germ cells. *Curr. Biol.* 8: 243–246. [https://doi.org/10.1016/S0960-9822\(98\)70091-0](https://doi.org/10.1016/S0960-9822(98)70091-0)
- Valderrama, J. A., S. S. Kulkarni, V. Nizet, and E. Bier, 2019 A bacterial gene-drive system efficiently edits and inactivates a high copy number antibiotic resistance locus. *Nat. Commun.* 10: 5726. <https://doi.org/10.1038/s41467-019-13649-6>
- Werren, J. H., U. Nur, and C.-I. Wu, 1988 Selfish genetic elements. *Trends Ecol. Evol.* 3: 297–302. [https://doi.org/10.1016/0169-5347\(88\)90105-X](https://doi.org/10.1016/0169-5347(88)90105-X)
- Werren, J. H., 2011 Selfish genetic elements, genetic conflict, and evolutionary innovation. *Proc. Natl. Acad. Sci. USA* 108: 10863–10870. <https://doi.org/10.1073/pnas.1102343108>
- Windbichler, N., P. A. Papatianos, and A. Crisanti, 2008 Targeting the X Chromosome during Spermatogenesis Induces Y Chromosome Transmission Ratio Distortion and Early Dominant Embryo Lethality in *Anopheles gambiae*. *PLoS Genet.* 4: e1000291. <https://doi.org/10.1371/journal.pgen.1000291>
- Windbichler, N., M. Menichelli, P. A. Papatianos, S. B. Thyme, H. Li *et al.*, 2011 A synthetic homing endonuclease-based gene drive system in the human malaria mosquito. *Nature* 473: 212–215. <https://doi.org/10.1038/nature09937>
- Yan, Y., and G. C. Finnigan, 2018 Development of a multi-locus CRISPR gene drive system in budding yeast. *Sci. Rep.* 8: 17277. <https://doi.org/10.1038/s41598-018-34909-3>

Communicating editor: R. Kulathinal